

# Soil seed bank characteristics in relation to different shrub species in semiarid regions

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## Abstract

Little information is available about the effects of different species of shrubs on the composition of the soil seed bank (SSB) in semiarid regions. We determined the role of three dominant shrub species on SSB characteristics and evaluated their potential for their possible use in rangeland restoration projects. Fifteen sites, each containing three shrub species (*Amygdalus scoparia*, *Daphne mezereum* and *Ebenus stellata*) and a herbaceous patch (control) in close proximity, were sampled and their SSB density, species richness and diversity at 0-10 cm depth were determined. The results showed that the density of the SSB was highest under *A. scoparia* (1133 seeds per m<sup>2</sup>) and lowest in herbaceous vegetation (110 seeds per m<sup>2</sup>). Species richness and diversity of the SSB was significantly greater under *E. stellata* than under the other shrubs and control. This study revealed that the extent to which vegetation affected SSB characteristics did not only depend on the presence of shrubs, but also on the species of shrub. These different roles of different species of shrubs on SSB are advised to be considered in the restoration of degraded areas through planting of shrubs in semiarid regions. Planting and the extension of *E. stellata* cover in degraded sites could be of priority due to its prominent role in herbaceous SSB reservoir and species diversity and richness.

## Introduction

Arid and semi-arid ecosystems occupy 36% of the land area of the globe and shrubs function as foundation species within these ecosystems (Yang and Williams, 2015). A foundation species in ecology was described as species with significant impacts on the structure and functioning of an ecosystem (Lortie et al., 2017). It has been frequently called shrubs as fertile islands, since, they have significant influences on habitat conditions. Shrubs have been shown to increase soil microbial functions (Chandregowda et al., 2018), enhance mycorrhizal colonization (Armenta Calderón et al., 2019), alter runoff and sediment yields (García Ruiz et al., 2013; Keesstra et al., 2016; Lu et al., 2019) and affect the soil seed bank (SSB) (Niknam et al., 2018; Funk et al., 2019). They increase SSB under their canopies by trapping seeds or increasing seed production by sub-canopy plants through ameliorating the environment (García-Sánchez et al., 2012; Mussa et al., 2016).

Study on SSB is important, since, it is one of the most important functional parts of any plant community and can be significant components in the process of rehabilitating degraded lands (Mohammed and Denboba, 2020). Bakker (1989) identified SSBs as non-mature seeds buried in soil that can replace existing vegetation when they are degraded.

In semiarid regions, shrubs are able to change SSB characteristics. According to some reports SSB density was much higher under the shrubs than the surrounding areas (Pugnaire and Lázaro, 2000; Marone et al., 2004). In overgrazing sites, particularly, shrubs accumulated large and diverse SSBs beneath their canopy which were different in composition from seed banks of the open matrix (Dreber and Esler, 2011). This significant effect are induced by the ability of shrubs in seed trapping and providing suitable microclimate

and conditions for seed production by other plant species (Erfanzadeh et al., 2014). However, different species of shrubs are different in their canopy architectures. Some shrub species have raised stems and some are attached to the ground. The canopy is dense in some and open in others. Therefore, it can be supposed that shrubs may have different performances in trapping seeds and could affect differently seed production by smaller species in their sub-canopy. As a result, different species of shrubs may alter SSB characteristics, differently. Nevertheless, our literature review showed that none of the studies has compared the effect of different species of shrubs on SSBs in semiarid regions.

In this study, we compared the effect of three dominant species of shrubs on sub-canopy SSB characteristics. Since the selected shrubs were different in their canopy traits, we supposed that they affect SSB characteristics, differently. In restoration perspective, we need to know whether these shrub species can recover and consistently facilitate the abundance and diversity of other plants through SSB and whether this effect is different between different species of shrubs with different crown features and architectures. Unfortunately, planting of exotic shrubs (e.g. *Atriplex canescens*) has been extensively occurred in arid and semiarid rangelands in Iran for restoration goals. Before starting the restoration activities in degraded sites by planting native shrubs, it is important to us to know different potentials of different shrub species in facilitation of herbaceous plant recovery through SSB. We supposed that dwarf and procumbent shrubs have higher ability to trap seeds comparing with erect stem and free canopies. In this study, three dominant shrubs with different features and architectures in the canopy were selected, i.e. *Amygdalus scoparia* and *Daphne mezereum* with single-elongated main stems and, *Ebenus stellata* with procumbent canopy and multiple stems. We hypothesized that the density and species richness and diversity of SSB under the canopy of *E. stellata* would be higher than *A. scoparia* and *D. mezereum*.

## Materials and Methods

### Study area

This study was conducted in the rangelands of Chenarnaz, Yazd province, Iran (30° 03' 51" N - 30deg 05' 89" N; 53deg 00' 16" E - 54deg 01' 23" E) (Fig. 1). The average altitude is 2200 m asl. The average annual temperature is 17.5degC and the average rainfall is 250 mm, which has a semiarid climate based on Domarten index.

Sheep and goats are the dominant grazers in the area (ca. four heads of sheep and/or goats per ha) during the year. Intensive grazing together with recently drought have led to partly exposed soil and created empty gaps in the AGV (Gravand et al., 2016). Therefore, herbaceous revegetating bare soil and restoration of degraded sites using native plants is a priority. SSB has been found one of the important potential that helps the restoration of degraded sites (Shang et al., 2016). Therefore, this study was conducted to quantify the potential of different species of shrubs to enhance the SSB associated with them and thus their potential use for restoration. Therefore, it is important to identify firstly shrubs that associated with larger and richer SSB and secondly, consider restoring shrub species which have higher potential as SSB reservoir accompany with higher palatability for grazing. Thereupon, three shrubs together with surrounding herbaceous vegetation (hereafter called control) were selected:

A) *Amygdalus scoparia* Spach (Rosaceae family) is a wild species of almond that occupies large areas in many parts of central Iran and its neighbouring countries. The extraction and use of the oil from the *A. scoparia* is of interest due to their fatty acids composition that is comparable to those of olive oil (Sorkheh et al., 2016). The plant is attractive for grazing animals due to its shade, fruits and high palatability of leaves. It is a deciduous large shrub that grows to a height of up to 6 m, having a single-elongate main stem. It produces numerous long and green branches. Fruits are drupes and are 1 to 1.5 cm long and 0.5 cm wide. They are ripened and dehiscent at the end of July (Mozaffarian, 2012).

B) *Daphne mezereum* L. (Thymelaeaceae family) is a rounded-upright deciduous shrub with an erect and bushy habit that typically grows to 1.5 m tall. All parts of this plant are poisonous to humans if ingested, especially the fruits, sap and bark. Therefore, this shrub is unpalatable for grazing animals. Nevertheless, fruits are attractive to birds with no resulting ill effects (Mozaffarian, 2012). This species is found globally

in dry and semi-dry areas and, in the provinces located in the central Iran.

C) *Ebenus stellata* Boiss. (Fabaceae family) is a thorny shrub with a height of 30-120 cm, having short and oblong-leafy branches and ternate leaves that are alternate and covered with dense silk flakes. This species grows in large part of Iran including Kerman, Yazd, Esfahan, Fars and Hormozgan provinces and some dry and semi-dry regions of world (its native range is Oman and Iran to India). The canopy structure is open with compact thorny branches that it is difficult to graze livestock (Mozaffarian, 2012) (Fig. 2).

### Soil sampling and greenhouse experiments

Soil samples were collected in the early autumn after the ending of the growing season and seed dispersion. Thus, the SSB contained transient and persistent seeds. After a field survey, fifteen sampling sites were randomly selected, each site containing the three species of shrubs together with a herbaceous control area (outside the shrub canopies) in close proximity to each other (Fig. 2). The distance between any two sampling sites was at least 100 m to exclude spatial autocorrelation. In each sampling site, beneath each shrub individual (patch), after removing coarse litter ( $> 2$  cm) 10 soil cores (subsamples) were randomly collected, to a depth of 10 cm, with a 5 cm diameter auger and then the subsamples were pooled for each patch (totally 60 soil samples were collected). Then, the soil samples were stored at 4 oC to 5 oC for cold stratification (Dreber, 2011) for a period of 25 days. Subsequently, each soil sample was distributed evenly over a mix of sterilised potting soil and sand in the trays of 25 cm x 35 cm (60 trays in total). The sterilised soil layer was 3 cm and the field collected soil layer was maximum 2 cm in the thickness in the trays. The germination trays were labelled and distributed randomly on benches in the greenhouse with natural light and temperature conditions (varied between 15 degC and 26 degC ) and irrigated every second day (Niknam et al., 2018). In addition, six control trays containing only sterile material were randomly placed between the sample trays to test for seed contamination.

Identifiable germinated seedlings were counted and removed from the trays every week. The seedlings were identified to species level. Seedlings that could not be identified were transplanted to pots to allow further growth until identification was possible.

After a period of seven months, no further seedlings were emerged. Therefore, the trays were left to dry for two weeks and then the samples were reirrigated for another one month to help seed dormancy breaking.

### Soil seed bank characteristics measurements

Using the greenhouse data, the number of seeds per  $m^2$  was calculated (SSB density) and the number of species for each soil sample was considered as SSB richness.

In addition, SSB species diversity indices were calculated for each individual under-shrub. The Shannon index is most frequently used to characterize the diversity of communities; it is sometimes referred to as the Shannon-Wiener index (Equation 1).

$$\text{Equation 1 } H' = \sum_{i=1}^s p_i \log p_i$$

Where  $p_i$  is the relative abundance of SSB of the  $i$ th species in a soil sample, and  $S$  is the number of detected species in SSB in that sample (Chernov et al., 2015). Another diversity index frequently used in ecology is the Simpson index, which is frequently determined as the probability of belonging to different taxa for two plant species randomly selected from an indefinitely large community. The Simpson index was calculated from the equation 2 (Chernov et al., 2015).

$$\text{Equation 2 } S_I = \sum_{i=1}^s \frac{n_i (n_i - 1)}{N(N-1)}$$

Where  $n_i$  is the individual number of each plant species in the SSB in a sample, and  $N$  is the total number of all germinants of all plant species in SSB in that sample.

The diversity indices were calculated using the Past software.

In addition, during the growth season, we recorded the presence of all plant species within each of the patches sampled for SSB. Species abundance of the AGV was not estimated because it was not possible to place a sampling frame beneath the shrubs and we used presence-absence data for AGV in the analyses. Qualitative similarity between the species composition of the AGV and the SSB was assessed using the Jaccard similarity index (Kent and Coker 1994) in each patch using equation 3.

$$\text{Equation 3 } IS_j = \left[ \frac{C}{(C+A+B)} \right] \times 100$$

Where C is the number species common between AGV and SSB, A, the number of species found only in the AGV and B, the number of species found only in the SSB.

We also estimated the mean canopy surface of our shrubs on ground and the height of each individual shrub using metal tape measure.

### Data statistical analysis

Firstly, normality of data (SSB density, species richness, diversity indices and similarity between AGV and SSB) was examined using the Kolmogorov-Smirnov test and homogeneity of variance using Levene's test. Total seed density was transformed to meet the normal distribution. To evaluate the effect of shrub species on SSB properties one-way ANOVA and LSD mean comparison tests were used. All statistical analyses were performed in the SPSS software ver. 16.

## Results

### Soil seed bank composition

A total of 118 species were found in the SSB and AGV. 55 species were present in SSB while they were absent in the AGV and 53 species were present in the AGV while absent in the SSB (Appendix 1).

In total, 2316 seedlings of 67 species (22 families) were germinated in soil samples: 910 seedlings belonging to 28 species in *A. scoparia* patch, 661 seedlings belonging to 30 species in *D. mezereum* patch, 637 seedlings belonging to 45 species in *E. stellata* patch and 108 seedlings belonging to 23 species in herbaceous patch (control). There were 12 species, common in four patches. The germinated seeds of *A. scoparia* shrub was observed in the greenhouse while seeds of *D. mezereum* and *E. stellata* shrubs were absent in the SSB. Most observed species in SSB belonged to Asteraceae (12 species, 17.91% of total species), Poaceae (9 species, 13.43% of total species) and Lamiaceae (9 species, 13.43% of total species), respectively (Appendix 1).

### Variation of soil seed bank characteristics under the shrubs

The ANOVA results showed that the highest and lowest values of SSB densities were found under *A. scoparia* (1133 seeds /m<sup>2</sup>) and control (110 seeds /m<sup>2</sup>), respectively (df = 3, F = 3.56 and p<0.05) (Fig. 3). The highest and lowest species number of SSB were observed under *E. stellata* (8.26 species per samples) and control (3.13 species per samples), respectively (df = 3, F = 6.41 and P<0.01) (Fig. 4). In addition, the results showed that the highest and lowest values of Shannon-Wiener diversity index were observed beneath of *E. stellata* and control with 1.06 and 0.83, respectively (df = 3, F = 3.32 and P<0.05) (Fig. 5). The highest and lowest values of Simpson diversity indices were found under canopy of *E. stellata* (0.75) and the control (0.44), respectively (df = 3, F = 5.02 and P<0.01) (Fig. 5). The highest values of similarity between SSB and AGV were found under three shrubs (18% to 19%) and the lowest was observed in the control (8%) (df = 3, F = 15.11 and P<0.01) (Fig. 6).

In addition, the mean surface of shrub canopies on ground was ca. 7.5 m<sup>2</sup>, 5.5 m<sup>2</sup> and 4.00 m<sup>2</sup> for *A. scoparia*, *D. mezereum* and *E. stellata*, respectively, and amongst three shrubs, *A. scoparia* had the highest mean height with ca. 3.5 m comparing with *D. mezereum* and *E. stellata* with ca. 2.60 m and ca. 1.83 m, respectively

## Discussion

Our results showed that among 22 plant families in SSB, the highest number of species belonged to Asteraceae. Previous studies showed that these plants were also widely present in the SSB (e.g. Gomaa et al., 2012). One of the possible reasons for the increase of these plants in SSB is the abundant seed production and morphological characteristics of the seeds. Species of Asteraceae with small seed size and wing shape, light and easy dispersal provide conditions for the presence of their seeds in the SSB (Harper, 1977; Hong et al., 2012). Forbs were the most abundant plants in the SSB composition. Our results showed that number of forb species was higher (33, 27, 24 and 17 species beneath *E. stellata*, *D. mezareum*, *A. scoparia* and control, respectively) than grasses (6, 7, 5 and 5 species beneath *E. stellata*, *D. mezareum*, *A. scoparia* and control, respectively) in the study area. In accordance with the results of our study, Bertiller et al. (2011), Parlak et al. (2011) and Tessema et al. (2012) reported that forbs had the highest number of species in SSB. Higher number of forbs in the AGV might be a reason for increasing the seeds of these species in the SSB. In contrast, woody plant species (trees and shrubs) were scarcely found in the greenhouse. Although, Teketay and Granstrom (1997) and Chaideftou et al. (2009) attributed the lack of woody species in the SSB to the lack of mature species in the AGV, in our study, this cannot be the reason because woody species in the AGV were frequent. Many factors are involved in reducing the density and richness of woody species in the SSB in an area. These could include the larger size of the seeds, the higher amount of predations and seed dormancy (Esmailzadeh et al., 2011). Some studies have shown that breaking seed dormancy of woody plants requires special conditions and if these conditions are not provided, these plants will eventually be removed from the SSB (Chaideftou et al., 2009).

In general, this study showed that the SSB density and species richness and diversity under the shrubs was higher than control and this differentiation was more pronounced for forbs. Previous studies (e.g. Marone et al., 2004) showed that the seed density of forbs were often higher under woody plants and positively correlated with the cover of woody vegetation, whereas the seed density of grasses were less associated by woody vegetation. Our results are consistent with some previous studies (e.g. Erfanzadeh et al., 2014) and disagree with others (e.g. Mdela et al., 2020). Positive effects of shrubs on SSB are exerted through direct and indirect ways. They increase buried seeds in soil by directly trapping seeds or by indirect mechanisms through an intermediary animal or plant species (Bullock and Moy, 2004; Giladi et al., 2013). Shrubs significantly influence the movement wind or water around their canopy (Hoffman et al., 2013) and thus can trap seeds or act as barrier for movement (Giladi et al., 2013). Shrubs can indirectly facilitate seed arrival by acting as a perching site for seed-carrying birds (Debussche and Isenmann, 1994) or as cache for granivorous rodents (Beck and van der Wall, 2010) and ants (Vergara-Torres et al., 2018). Additionally, shrubs can indirectly increase SSB by facilitating the plants that are able to increase seed production or viability and vigourity of produced seeds (Pugnaire and Lazaro, 2000). Shrubs provide a suitable conditions for growing, flowering and seeding of herbaceous plants under their canopies through modifying the physical and chemical properties of soil with litter and root exudation, improving soil micro-relief, decreasing direct sunlight, increasing soil moisture, protecting the surface soil from erosion and adding organic matter into the soil (Ruiz et al., 2008; Barness et al., 2009; Olvera-Carrillo et al., 2009; Li et al., 2011; Sylvain and Wall, 2011; Garcia-Sanchez et al., 2012).

However, different effects of different shrubs on SSB characteristics were observed in increasing SSB density under the canopy of *A. scoparia* and species richness and diversity under the canopy of *E. stellata*. Previous studies showed that the size of a shrub could impact the arrival of seeds (Pugnaire and Lazaro, 2000) because larger shrubs can provide greater facilitative effects. Larger shrubs can intercept more solar radiation (Maestre and Cortina, 2005), have higher soil nutrients (Zhang et al., 2015), or lower evapotranspiration (Kidron and Gutschick, 2013) creating a favourable microclimate for seed production, particularly by annuals (Filazzola et al., 2019). As a result, comparing to the other shrubs, taller and larger canopy in *A. scoparia* might increase SSB density through higher seed production by plants. The seeds of some annuals were found at strong frequent under *A. scoparia*, i.e. *Bromus tectorum*, *Galium aparine* and *Veronica anagalis*. However, procumbent canopy in *E. stellata* might increase species diversity and richness of SSB. It might be that attached crown cover to the ground in *E. stellata* physically obstruct more seeds and enhance species diversity and richness in SSB. Our results showed that the seeds of many species were found under *E.*

*stellata*, while they were absent in control or under other shrubs, e.g. *Polygonum dumosum*, *Poa sinaica*, *Tragopogon jezdiianus* and *Dianthus orientalis*, *Tianthus crinitus*. Briefly, indirect effect of *A. scoparia* on seed production of few plant species (some annuals) and direct effect of *E. stellata* on seed trapping of many species resulted these significant differences of SSB density, species richness and diversity between shrubs.

Similarity between the seed bank and the AGV was generally low in three shrubs and control. The low similarity between the AGV and the SSB in our and other studies is usually due to the fact that some species were present in the vegetation, while they were absent from the seed bank, and vice versa (e.g. Valkoa et al., 2014; Erfanzadeh et al., 2016). However, the similarity between the AGV and the SSB was lowest in the control. In this area, many species, such as *Acantholimon scorpius* and *Ebenus stellata*, were absent from the seed bank while they were present in the AGV. Most of these species were perennial, and these, especially shrubby ones, are well-known for their transient seed bank (Thompson et al., 1997). Moreover some annuals such as *Bromus tectorum* and *Galium aparine* were found in the SSB in control and under three shrubs. These species were present in the AGV under the shrubs while absent in the AGV of control. At the sampling time, some annuals in the AGV might be grazed or dried and ended their phenological stages in the control due to higher availability to grazers or solar radiation, temperature and wind speed comparing with under the canopies of shrubs.

## Conclusions

AGV in the present study area suffers from human activities such as over grazing. This may lead to habitat degradation and vegetation destruction in some part of the area. Knowledge of the SSB and its temporal and spatial variation is a useful tool for conservation and restoration efforts. This study showed that shrubs, overall, played an important role in reserving of herbaceous species seeds under their canopies. However, the extent to which shrubs affects SSB characteristics is dependent on the species of shrub. These different roles of shrubs on SSB are advised to be considered in restoration of areas through conservation of endemic shrubs in the semiarid regions. Although, *E. stellata* has a low or intermediate palatability for grazers, it can be of priority for rangeland improvement and, restoration of degraded sites if plant diversity increase is of priority. However, highest SSB density under the canopy of *A. scoparia* with its potential in medicinal and grazing uses may be considered for recovery of degraded site through planting of this shrub as second priority.

## References

- Armenta Calderon AD, Moreno-Salazar SF, Furrázola Gomez E, Ochoa-Meza A. 2019. Arbuscular mycorrhiza, carbon content and soil aggregation in Sonoran Desert plants. *Spanish Journal of Soil Science* **9**:42-53.
- Bakker HG. 1989. Ecology of Soil Seed Banks. Academic Press, Inc. San Diego. 462 pp.
- Barnes G, Zaragoza SR, Shmueli I, Steinberger Y. 2009. Vertical distribution of a soil microbial community as affected by plant ecophysiological adaptation in a desert system. *Microbial Ecology* **57**:36-49.
- Beck MJ, van der Wall SB. 2010. Seed dispersal by scatter-hoarding rodents in arid environments. *Journal of Ecology* **98**:1300-1309.
- Bertiller M B, Ares J. 2011. Does Sheep Selectivity Along Grazing Paths Negatively Affect Biological Crusts and Soil Seed Banks in Arid Shrublands? A Case Study in the Patagonian Monte, Argentina. *Journal of Environmental Management* **92**:2091-2096.
- Bullock JM, Moy IL. 2004. Plants as seed traps: inter-specific interference with dispersal. *Acta Oecologica* **25**:35-41.
- Chaideftou E, Thanos CA, Bergmeier E, Kallimanis A, Dimopoulos P. 2009. Seed Bank Composition and Above-Ground Vegetation in Response to Grazing in sub-Mediterranean Oak Forests (NW Greece). *Plant Ecology* **201**:255-265.

- Chandregowda MH, Murthy K, Bagchi S. 2018. Woody shrubs increase soil microbial functions and multifunctionality in a tropical semi-arid grazing ecosystem. *Journal of Arid Environments* **155** :65-72.
- Chernov T I, Tkhakakhova AK, Kutovaya OV. 2015. Assessment of diversity indices for the characterization of the soil prokaryotic community by meta-genomic analysis. *Eurasian Soil Science* **48** :410-415.
- Debussche M, Isenmann P. 1994. Bird-dispersed seed rain and seedling establishment in patchy Mediterranean vegetation. *Oikos* **69** :414-426.
- Dreber N, Esler KJ. 2011. Spatial-Temporal Variation in Soil Seed Banks under Contrasting Grazing Regimes Following Low and High Seasonal Rainfall in Arid Namibia, *Journal of Arid Environments* **13** :174-184.
- Dreber N. 2011. How best to quantify soil seed banks in arid rangelands of the Nama Karoo? *Environmental Monitoring Assessment* **173** :813-824.
- Erfanzadeh R, Kamali P, Ghelichnia H, Petillon J. 2016. Effect of grazing removal on aboveground vegetation and soil seed bank composition in sub-alpine grasslands of northern Iran. *Plant Ecology and Diversity* **9** :309-320.
- Erfanzadeh R, Shahbazian R, Zali H. 2014. Role of plant patches in preserving flora from the soil seed bank in an overgrazed high-mountain habitat in northern Iran. *Journal of Agricultural Sciences and Technology* **16** :229-238
- Esmailzadeh O, Hosseini S, Tabari M. 2011. Relationship Between Soil Seed Bank and Above-Ground Vegetation of a Mixed-deciduous Temperate Forest in Northern Iran. *Journal of Agricultural Science and Technology* **2** :399-409.
- Funk FA, Loydi A, Peter G, Distel RA. 2019. Effect of grazing and drought on seed bank in semiarid patchy rangelands of northern Patagonia, Argentina. *International Journal of Plant Sciences* **180** :337-344
- Garcia-Ruiz JM, Nadal-Romero E, Lana-Renault N, Begueria S. 2013. Erosion in Mediterranean landscapes: changes and future challenges. *Geomorphology* **198** :20-36
- Garcia-Sanchez R, Camargo-Ricalde SL, Garcia-Moya E, Luna-Cavazos M, Romero-Manzanares A, Manuel Montano N. 2012. *Prosopis laevigata* and *Mimosa biuncifera* (Leguminosae), jointly influence plant diversity and soil fertility of a Mexican semiarid ecosystem. *Revista de Biología Tropical* **60** :87-103
- Giladi I, Segoli M, Ungar ED. 2013. Shrubs and herbaceous seed flow in a semi-arid landscape: dual functioning of shrubs as trap and barrier. *Journal of Ecology* **101** :97-106.
- Gomaa NH, 2012. Soil Seed Bank in Different Habitats of the Eastern Desert of Egypt. Saudi. *Journal of Biological Sciences* **19** :211-220.
- Gravand J, Hosseini SM, Ahmadi K, Avoili A, Ahmadi AR. 2016. Investigation of the structure of *Pistacia atlantica* masses in enclosed and non-enclosed areas (Protected Region of Bagh Shad in Yazd). *Natural Ecosystems of Iran* **7** :89-102.
- Harper JV. 1997. Population Biology of Plant. Academic Press, Ny, pp.83-110.
- Hoffman O, Yizhaq H, Boeken BR. 2013. Small-scale effects of annual and woody vegetation on sediment displacement under field conditions. *Catena* **109** :157-163.
- Hong J, Liu Sh, Shi G, Zhang Y. 2012. Soil Seed Bank Techniques for Restoring Wetland Vegetation Diversity in Yeyahu Wetland. *Ecological Engineering* **5** : 192-202.
- Keesstra S, Pereira P, Novara A, Brevik EC, Azorin-Molina C, Parras-Alcantara L, Jordan A, Cerda A. 2016. Effects of soil management techniques on soil water erosion in apricot orchards. *Science of the Total Environment* **551** :357-366
- Kent M, Coker P. 1994. Vegetation description and analysis. A practical approach. Wiley, Chichester. UK.

Kidron GJ, Gutschick VP. 2013. Soil moisture correlates with shrub–grass association in the Chihuahuan Desert. *Catena***107** :71-79.

Li C, Li Y, Ma J. 2011. Spatial heterogeneity of soil chemical properties at fine scales induced by *Haloxylon ammodendron*(Chenopodiaceae) plants in a sandy desert. *Ecological Research***26** :385-394

Lortie CJ, Gruber E, Filazzola A, Noble T, Westphal M. 2017. The Groot effect: plant facilitation and desert shrub regrowth following extensive damage. *Ecology and Evolution*. **8** :706-715.

Lu R, Liu YF, Jia C, Huang Z, Liu Y, Hea H, Liu BR, Wang ZJ, Zhenga J, Wua GL. 2019. Effects of mosaic-pattern shrub patches on runoff and sediment yield in a wind-water erosion crisscross region. *Catena***174** :199-205.

Maestre FT, Cortina J. 2005. Remnant shrubs in Mediterranean semi-arid steppes: effects of shrub size, abiotic factors and species identity on understorey richness and occurrence. *Acta Oecologica***27** :161-169.

Marone L, Cueto VR, Milesi FA, Lopez de Casenave J. 2004. Soil Seed Bank Composition Over Desert Microhabitats, Patterns and Plausible Mechanisms, *Canadian Journal of Botany***20** :1809-1816.

Mndela M, Madakadze CI, Nherera-Chokuda F, Dube S. 2020. Is the soil seed bank a reliable source for passive restoration of bush-cleared semi-arid rangelands of South Africa? *Ecological Processes***9** :1.<https://doi.org/10.1186/s13717-019-0204-6>.

Mohammed SA, Denboba MA. 2020. Study of soil seed banks in ex-closures for restoration of degraded lands in the central Rift Valley of Ethiopia. *Scientific Reports* **10** :956. <https://doi.org/10.1038/s41598-020-57651-1>.

Mozafarian M. 2012. Recognition of Medicinal and Aromatic Herbs in Iran, Moaser Publisher, Iran.

Mussa M, Ebro A, Nigatu L. 2016. Impact of woody plants species on soil physico-chemical properties along grazing gradients in rangelands of eastern Ethiopia. *Tropical and Subtropical Agroecosystems***19** :343-355.

Niknam P, Erfanzadeh R, Ghelichnia H, Cerda A. 2018. Spatial Variation of Soil Seed Bank under Cushion Plants in a Subalpine Degraded Grassland. *Land Degradation and Development* **29** :4-14.

Olvera-Carrillo Y, Mendez I, Sanchez-Coronado ME, Marquez-Guzman J, Barradas VL, Huante P, Orozco-Segovia A. 2009. Effect of environmental heterogeneity on field germination of *Opuntia tomentosa*(Cactaceae, Opuntioideae) seeds. *Journal of Arid Environments***73** :414-420.

Parlak AO, Gokkus A, Demiray HC. 2011. Soil Seed Bank and Aboveground Vegetation in Grazing Lands of Southern Marmara, Turkey. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* **39** :96.

Pugnaire F, Lazaro R. 2000. Seed bank and understorey species composition in a semi-arid environment: the effect of shrub age and rainfall. *Annals of Botany* **86** :807-813.

Ruiz TG, Zaragoza SR, Cerrato RF. 2008. Fertility islands around *Prosopis laevigata* and *Pachycereus hollianus* in the drylands of Zapotitlan Salinas, Mexico. *Journal of Arid Environments* **72** :1202-1212

Shang ZH, Yang SH, Wang YL, Shi JJ, Ding LM, Long RJ. 2016. Soil seed bank and its relation with above-ground vegetation along the degraded gradients of alpine meadow. *Ecological Engineering***90** :268-277.

Sorkheh K, Kiani S, Sofo A. 2016. Wild almond (*Prunus scoparia*L.) as potential oilseed resource for the future: Studies on the variability of its oil content and composition. *Food Chemistry***212** :58-64.

Sylvain ZA, Wall DH. 2011. Linking soil biodiversity and vegetation: implications for a changing planet. *American Journal of Botany***98** :517-527.

Teketay D, Granstrom A. 1997. Seed Viability of Afromontane Tree Species in Forest Soils. *Journal of Tropical Ecology* **13** :81-95.



Tessema ZK, de Boer WF, Baars RM, Prins HH. 2012. Influence of grazing on soil seed banks determines the restoration potential of aboveground vegetation in a semi-arid Savanna of Ethiopia. *Biotropica* **44** :211-219.

Thompson K, Bakker JP, Bekker RM. 1997. The soil seed bank of North West Europe: methodology, density and longevity. CUP, Cambridge University Press.

Valkoa O, Tothmereszb B, Kelemen A, Simon E, Miglecz T, Lukacs BA, Torok P. 2014. Environmental factors driving seed bank diversity in alkali grasslands. *Agriculture, Ecosystems and Environment* **182** :80-87.

Vergara-Torres CA, Corona-Lopez AM, Diaz-Castelazo C, Toledo-Hernandez VH, Flores-Palacios A. 2018. Effect of seed removal by ants on the host-epiphyte associations in a tropical dry forest of central Mexico. *AoB PLANTS* **10**: ply056; doi: 10.1093/ aobpla/ply056

Yang, X., Williams M. 2015. Landforms and processes in arid and semi-arid environments. *Catena* **134** :1-3.

Zhang G, Yang Q, Wang X, Zhao W. 2015. Size-related change in *Nitraria sphaerocarpa* patches shifts the shrub-annual interaction in an arid desert, northwestern China. *Acta Oecologica* **69** : 121-128.

## Figures Captions

**Fig. 1.** Geographical location of the study area and fifteen sites in which all three shrub species were found closed to each other in each site.

**Fig. 2.** Sampling areas containing *Amygdalus scoparia* (A), *Daphne mezereum* (B) and *Ebenus stellata* (C) that formed woody patches in the surrounding herbaceous vegetation, used as control (D) for comparing soil seed bank characteristics, Chenarnaz rangelands, Yazd province, Iran (30deg 03' 51" N - 30deg 05' 89" N; 53deg 00' 16" E - 54deg 01' 23" E).

**Fig. 3.** Mean densities (+SE) of seeds that germinated under three shrubs (*Amygdalus scoparia* , *Daphne mezereum* and *Ebenus stellata* ) and herbaceous vegetation, Chenarnaz Rangelands, Yazd province, Iran. Lower case letters indicate statistically significant differences ( $P < 0.05$ ) among patch types.

**Fig. 4.** Mean species richness (+ SE) of germinants under three shrubs (*Amygdalus scoparia* , *Daphne mezereum* and *Ebenus stellata* ) and herbaceous vegetation, Chenarnaz Rangelands, Yazd province, Iran. Lower case letters indicate statistically significant differences ( $P < 0.05$ ) among patch types.

**Fig. 5 .** Mean (+ SE) Shannon and Simpson diversity indices under three shrubs (*Amygdalus scoparia* , *Daphne mezereum* and *Ebenus stellata* ) and herbaceous vegetation, Chenarnaz Rangelands, Yazd province, Iran. Lower case letters indicate statistically significant differences ( $P < 0.05$ ) among patch types.

**Fig. 6.** Mean (+ SE) Jaccard similarity index between soil seed bank and above-ground vegetation under three shrubs (*Amygdalus scoparia* , *Daphne mezereum* and *Ebenus stellata* ) and herbaceous vegetation, Chenarnaz Rangelands, Yazd province, Iran. Lower case letters indicate statistically significant differences ( $P < 0.05$ ) among patch types.

**Appendix. 1.** Average soil seed bank density of each species under the canopy of each shrub (the digits). \*presence of the species in the above-ground vegetation.

Species	Family	Growth habit	Seed density (seeds per m <sup>2</sup> )	
			<i>Amygdalus Scoparia</i>	<i>Daphne Meza</i>
<i>Acantholimon</i> sp.	Plumbaginaceae	Shrub	0*	0*
<i>Acantholimon scorpius</i> L.	Plumbaginaceae	Shrub	0*	0*
<i>Acanthophyllum spinosum</i> C. A. May	Caryophyllaceae	Shrub	0*	0*
<i>Aegopordon berardioides</i> Boiss.	Asteraceae	Forb	0*	0*
<i>Allium inutiflorum</i> Regel.	Liliaceae	Forb	0	0*
<i>Allium</i> sp.	Liliaceae	Forb	74.71*	93.39*
<i>Alkanna</i> sp.	Boraginaceae	Forb	0	0

Species	Family	Growth habit	Seed density (seeds per m <sup>2</sup> )	Seed density
<i>Alyssum marginatum</i> L.	Brassicaceae	Forb	0*	0*
<i>Alyssum minus</i> (L.) Rothm.	Brassicaceae	Forb	0*	18.67*
<i>Alyssum</i> sp.	Brassicaceae	Forb	0*	0
<i>Amygdalus lycioides</i> Spach.	Rosaceae	Shrub	0*	0*
<i>Amygdalus scoparia</i> Spach.	Rosaceae	Tree	149.42*	18.67*
<i>Angelonia</i> sp.	Plantaginaceae	Forb	0	0*
<i>Arrhenathrum kotschy</i> Boiss	Poaceae	Grass	0	18.67
<i>Artemisia aucheri</i> Boiss.	Asteraceae	Shrub	74.71*	74.71*
<i>Astragalus albispinus</i> Sirj & Born.	Papilionaceae	Forb	0*	0*
<i>Astragalus</i> sp.	Papilionaceae	Forb	0*	37.35*
<i>Astragalus spachianus</i> Boiss.	Papilionaceae	Forb	0*	0*
<i>Astragalus terrestris</i> Kitam.	Papilionaceae	Forb	0*	0*
<i>Asperula orientalis</i> Boiss. & Hohen.	Rubiaceae	Grass	56.03	18.67
<i>Brassica</i> sp.	Brassicaceae	Forb	0	0*
<i>Bromus danthonia</i> (L.) DC.	Poaceae	Grass	0	0*
<i>Bromus tectorum</i> L.	Poaceae	Grass	11057.77*	5360.77*
<i>Bromus scoparius</i> L.	Poaceae	Grass	0*	0*
<i>Carex</i> sp.	Cyperaceae	Forb	56.03	18.67
<i>Carthamus glaucus</i> M.Bieb.	Asteraceae	Forb	74.71	224.144
<i>Centaurea virgate</i> Lamarck.	Asteraceae	Forb	0*	0*
<i>Crepis</i> sp.	Asteraceae	Forb	0	0
<i>Cicer oxyodon</i> Boiss & Hohen.	Fabaceae	Forb	0	18.67
<i>Clypeola aspera</i> (Grauer) Turrill.	Brassicaceae	Forb	0	0
<i>Convolvulus fruticosus</i> L.	Convolvulaceae	Forb	0	0*
<i>Crepis sancta</i> L.	Asteraceae	Forb	0*	0
<i>Daphne mezereum</i> L.	Thymelaeaceae	Shrub	0*	0*
<i>Dianthus crinitus</i> Sm.	Caryophyllaceae	Forb	0	0
<i>Dianthus orientalis</i> Beitr.	Caryophyllaceae	Forb	0	0
<i>Dichanthiu mannulatum</i> (Forssk.) Stapf.	Poaceae	Grass	0	280.18
<i>Ebenus stellata</i> Bioss.	Fabaceae	Shrub	0*	0*
<i>Echinophora platyloba</i> DC.	Thymelaeaceae	Forb	0*	0*
<i>Erymopyrum distans</i> (Ledeb) Jaub	Poaceae	Grass	653.75	541.68
<i>Erysimum</i> sp.	Brassicaceae	Forb	74.71	0
<i>Erodium cicutarium</i> L.	Geraniaceae	Forb	0*	0
<i>Erodium</i> sp.	Geraniaceae	Forb	0*	0*
<i>Eryngium</i> sp.	Umbelliferae	Forb	0*	0
<i>Eryngium bangai</i> Bioss.	Umbelliferae	Forb	0*	0*
<i>Festuca ovina</i> L.	Poaceae	Grass	597.71	280.18
<i>Galium aparine</i> L.	Rubiaceae	Forb	859.21*	1064.68*
<i>Hertia angustifolia</i> (DC.) Kuntze	Asteraceae	Forb	0*	0*
<i>Geranium</i> sp.	Geraniaceae	Forb	0*	0*
<i>Isatis</i> sp.	Brassicaceae	Forb	0	0
<i>Juncus inflexus</i> L.	Juncaceae	Forb	37.35	0
<i>kochia prostrata</i> (L.) Schrad	Chenopodiaceae	Forb	0	18.67
<i>Lactuca glaucifolia</i> Boiss.	Asteraceae	Forb	0	37.35
<i>Lappula microcarpa</i> (Ledebour) En & Pr	Boraginaceae	Forb	0	0
<i>Linum</i> sp.	Linaceae	Forb	37.35	18.67
<i>Lactuca lanceolate</i> L.	Asteraceae	Forb	0*	0*
<i>Lactuca orientalis</i> Boiss.	Asteraceae	Forb	0*	0
<i>Lactuca serriola</i> L.	Asteraceae	Forb	0	0

Species	Family	Growth habit	Seed density (seeds per m <sup>2</sup> )	Seed density
<i>Lolium perenne</i> L.	Poaceae	Grass	0*	0
<i>Lolium</i> sp.	Poaceae	Grass	0*	0
<i>Loranthus grewinkii</i> Boiss & Buhse	Loranthaceae	Forb	0	0*
<i>Medicago radiata</i> L.	Fabaceae	Forb	0*	0*
<i>Micropus</i> sp.	Asteraceae	Forb	0*	0*
<i>Myosotis</i> sp.	Boraginaceae	Forb	0	0*
<i>Mentha longifolia</i> (L.) Huds.	Lamiaceae	Forb	0	18.67
<i>Minuartia decipiens</i> (Fenzl) Bornm.	Caryophyllaceae	Forb	18.67	0
<i>Marrubium vulgare</i> L.	Lamiaceae	Forb	18.67*	0
<i>Marrubium</i> sp.	Lamiaceae	Forb	0	0
<i>Medicago sativa</i> L.	Fabaceae	Forb	0	0
<i>Medicago</i> sp.	Fabaceae	Forb	18.67	0
<i>Nepeta pungens</i> (Bunge) Benth., Lab. Gen.	Lamiaceae	Forb	0	56.03
<i>Nonea mucronata</i> Forssk.	Chenopodiaceae	Shrub	18.67*	0
<i>Onopordon</i> sp.	Asteraceae	Forb	0*	56.03*
<i>Papaver</i> sp.	Papaveraceae	Forb	0*	0
<i>Paracaryum</i> sp.	Boraginaceae	Forb	0*	0
<i>Peganum harmala</i> L.	Zygophyllaceae	Forb	56.03	0
<i>Pistacia atlantica</i> Desf.	Anacardiaceae	Tree	0	112.07
<i>Pimpinella affinis</i> L.	Apiaceae	Forb	93.39	0
<i>Phlomis olivieri</i> Benth.	Lamiaceae	Forb	37.35	37.35
<i>Phlomis aucheri</i> Boiss.	Lamiaceae	Forb	0	0
<i>Polygonum erectum</i> L.	Polygonaceae	Forb	18.67	18.67
<i>Polygonum</i> sp.	Polygonaceae	Forb	18.67	0
<i>Polygonum dumosum</i> Boiss	Polygonaceae	Forb	0	0
<i>Poa annual</i> L.	Poaceae	Grass	0*	0
<i>Poa sinaica</i> Steud.	Poaceae	Grass	0	0
<i>Psathyrostachys</i> sp.	Poaceae	Grass	0*	0*
<i>Ribes iebersteinii</i> Berland. ex DC.	Grossulariaceae	Tree	18.67	0
<i>Scariola paradoxa</i> L.	Asteraceae	Forb	0	18.67
<i>Scoriola orientalis</i> (Boiss.) Sojak.	Asteraceae	Forb	0	18.67
<i>Scandix aucheri</i> Boiss.	Apiaceae	Forb	18.67	37.35
<i>Scorzonera mucida</i> L.	Asteraceae	Forb	37.35	18.67
<i>Senecio destontainei</i> L.	Asteraceae	Forb	18.67	0
<i>Saussurea heteromalla</i> DC.	Asteraceae	Forb	0	18.67
<i>Silene spergulifolia</i> (Willd.) M. Bieb.	Caryophyllaceae	Forb	224.14	448.28
<i>Silene</i> sp.	Caryophyllaceae	Forb	149.42	242.82
<i>Sinapis</i> sp.	Brassicaceae	Forb	0	18.67
<i>Solanum nigrum</i> L.	Solanaceae	Forb	168.10	0
<i>Scabiosa olivieri</i> L.	Caprifoliaceae	Forb	0*	0
<i>Schismus arabicus</i> Ness.	Poaceae	Forb	0	0*
<i>Scirpoides holoschoenus</i> L.	Cyperaceae	Forb	0*	0
<i>Scorzonera</i> sp.	Asteraceae	Forb	0	0*
<i>Senecio</i> sp.	Asteraceae	Forb	0*	0
<i>Stachys inflata</i> Benth.	Lamiaceae	Forb	0*	18.67
<i>Stellaria blatterii</i> Mattf.	Caryophyllaceae	Forb	0	0
<i>Stipagrostis plumose</i> (Linn.)	Poaceae	Forb	0	298.85
<i>Stipa arabica</i> Trin & Ru.	Poaceae	Grass	933.93	2110.69
<i>Sterigmostemum longistylum</i>	Brassicaceae	Forb	0*	0
<i>Stipa barbata</i> Desf.	Poaceae	Grass	0*	0*

Species	Family	Growth habit	Seed density (seeds per m <sup>2</sup> )	Seed density
<i>Stipa parviflora</i> Desf.	Poaceae	Grass	0*	0
<i>Taraxacum montanum</i> (C.A. Mey.)	Asteraceae	Forb	242.82	485.64
<i>Thymus trnascausicus</i> Ronniger.	Lamiaceae	Shrub	112.07	149.42
<i>Tragopogon jezidianus</i> L.	Asteraceae	Forb	0	0
<i>Valerianella oxyrhynchus</i>	Valerianaceae	Forb	0*	0*
<i>Veronica anagallis</i> L.	Plantaginaceae	Forb	971.29	0
<i>Ziziphora clinopodioids</i> Lam.	Lamiaceae	Shrub	18.67	0
<i>Ziziphora tenuior</i> L.	Lamiaceae	Forb	0*	0*





