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Specklinia dunstervillei, a New Species Long Confused with Specklinia endotrachys (Orchidaceae: Pleurothallidinae)

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Abstract

Specklinia dunstervillei is described as a new species in recognition of the distinctness of a Venezuelan species related to and confused with Specklinia endotrachys. It was illustrated for the first time by G. C. K. Dunsterville in 1963 from a plant collected in Trujillo on the Cordillera de Merida. The newly named species can be easily recognized by its small habit, short leaves and small reddish-orange flowers, the non-ascending dorsal sepal and the obtuse petals that are shortly apiculate. Specklinia dunstervillei is formally described and illustrated once again and compared morphologically and genetically with its closest relatives.

Introduction

In the systematic study of the *Specklinia endotrachys* species complex, Pupulin et al. [1] found that, aside from the latter, *Specklinia pfavii* (Rchb.f.) Pupulin & Karremans, *S. remotiflora* Pupulin & Karremans and *S. spectabilis* (Ames) Pupulin & Karremans could all be recognized as distinct species in the complex. *Specklinia endotrachys* (Rchb.f.) Pridgeon & M.W.Chase had traditionally been considered a widely distributed and highly variable albeit morphologically quite unique species, nevertheless, the four proposed taxa could be easily distinguished morphologically and ecologically from each other. At the time, the authors recognized that not all the available material labeled *S. endotrachys* could be adequately placed into one of those four species concepts, and suspected more taxa would be recognized in the complex in the future [1,2,3].

The fifth species to be added to the complex was the Guatemalan *Specklinia juddii* (Archila) Pupulin & Karremans, unknown at the time of the first publication and later placed amongst its relatives [2,3]. It had been published under the generic name *Empusella* (Luer) Luer, a monotypic genus typified by *S. endotrachys* and here regarded as a synonym of *Specklinia* Lindl. [4,5,6]. Similarly, the Venezuelan material labeled *S. endotrachys* and illustrated for the first time in Dunsterville and Garay [7], morphologically did not resemble the other five species in the complex and had remained a mystery to the authors [1,2,3]. Now with additional material at hand we are able to compare the Venezuelan material with the other species of this



complex, both morphologically (for all species currently recognized) and genetically (for all except *S. juddii*).

Materials and Methods

This study was conducted at Jardín Botánico Lankester (JBL) of the Universidad de Costa Rica and Naturalis Biodiversity Center—Leiden University, from 2012 to 2014. Plants were collected under the scientific permits handed by the Costa Rican Ministry of Environment (MINAE) to researchers at JBL. Individual plants were photographed, illustrated and preserved as DNA samples, herbarium specimens and spirit specimens in formaldehyde: acetic acid: ethanol [FAA (53% ethanol, 37% water, 5% formaldehyde and 5% glycerol)] (only including flowers) for future reference, deposited at JBL-spirit and L-spirit and the DNA bank of Naturalis Biodiversity Center. Taxon names mostly follow Pridgeon [8].

Photography

The Lankester Composite Digital Plate (LCDP) and color illustrations of the flowers were made using a Nikon D5100 digital camera with a AF-S VR Micro-NIKKOR 105mm f/2.8G IF-ED lens and an Epson Perfection Photo Scanner V600, and a Leica MZ9.5 stereomicroscope fitted with a Leica DFC295 digital microscope color camera with Leica FireCam 3.4.1 software.

Phylogenetic analysis

The data matrix included DNA sequences of 50 individuals (Table 1), 27 of which were produced in this study. The remaining data were obtained from GenBank [4,5,9]. Fresh leaf and flower cuttings of approximately 1 cm² were dried with silica gel. Samples (20 mg) were pulverized and extraction performed following the DNEasy procedure (Qiagen). The nuclear ribosomal internal transcribed spacer (nrITS) region was amplified using the methods and primers for sequencing and amplification described by Sun et al. [10], and Sanger sequencing was done commercially by Macrogen on a 96-capillary 3730xl DNA Analyzer automated sequencer (Applied Biosystems, Inc.) using standard dye-terminator chemistry (Macrogen, Inc.).

The Staden et al. [11] package was used for editing of the sequences. Contigs were exported as.fas files and opened in Mesquite v2.72 (Maddison & Maddison [12]), where they were checked for base calling errors, the matrix was aligned manually (S1 File: Sequence Matrix). The ends of each data set were trimmed to eliminate possible erroneous data, and gaps were regarded as missing data (filled with Ns). *Phloeophila peperomioides* was used as the outgroup, as it was found to be one of the most distantly related of all included species in this phylogenetic analysis (Pridgeon *et al.* [4]). The trees were produced with an analysis of the nrITS dataset of 43 sequences using BEAST v1.6.0. (Drummond & Rambaut [13]). Parameters were set to preset, except for substitution model GTR with 10 categories, clock model uncorrelated lognormal, tree prior Yule process, and number of generations 20,000,000. The resulting trees were combined using TreeAnnotator v1.6.0., where the first 3000 trees were used as burn-in. FigTree v1.3.1. (Rambaut [14]) was used to edit the resulting tree. Posterior probabilities are given for each node in decimal form.

A pairwise comparison of the ITS sequence of the accessions of *S. dunstervillei*, *S. endotrachys*, *S. montezumae* (as an outgroup), *S. pfavii*, *S. spectabilis* and *S. remotiflora* is presented in Table 2. Each different base was counted as an individual change, even when concurrent; insertions and deletions were counted as a single change regardless of length. All the mentioned accession of each species in Table 1 were combined and used for the comparison except for AF262859, a sequence labeled *S. endotrachys* by Pridgeon et al. [4] but which we suspect (based on DNA data) should be *S. remotiflora* or a closely related species.



Table 1. List of the 50 accessions used in the phylogenetic analysis. The vouchers, NCBI GenBank accession number and source are given. Scientific names follow Pridgeon 2005.

Taxon	Sequence Voucher	GenBank Accession Number	Sequence Source
Dryadella simula (Rchb. f.) Luer	Chase 1095	AF262825	Pridgeon et al. [4]
Dryadella susanae (Pabst) Luer	Chiron 11240	JQ306486	Chiron et al. [9]
Phloeophila peperomioides (Ames) Garay	None	AF275690	Pridgeon et al. [4]
Platystele compacta (Ames) Ames	Chase 5637	AF262822	Pridgeon et al. [4]
Platystele misera (Lindl.) Garay	Chase 5625	AF262823	Pridgeon et al. [4]
Platystele stenostachya (Rchb. f.) Garay	Chase 5618	AF262821	Pridgeon et al. [4]
Scaphosepalum grande Kraenzl.	Chase 1107	AF262819	Pridgeon et al. [4]
Scaphosepalum swertiifolium (Rchb. f.) Rolfe	Chase 1383	AF262818	Pridgeon et al. [4]
Scaphosepalum verrucosum (Rchb. f.) Pfitzer	Chase 1331	AF262820	Pridgeon et al. [4]
Specklinia absurda Bogarín, Karremans & Rincón	Bogarín 8711 (JBL-Spirit)	KC425827	Bogarín et al. [5]
Specklinia barbae (Schltr.) Luer	Karremans 4853	KC425771	This Study
Specklinia barbae (Schltr.) Luer	Karremans 3928	KC425769	This Study
Specklinia chontalensis (A.H.Heller & A.D.Hawkes) Luer (1)	Pupulin 6543	KC425776	This Study
Specklinia chontalensis (A.H.Heller & A.D.Hawkes) Luer (2)	Pupulin 6543	KF747799	This Study
Specklinia costaricensis (Rolfe) Pridgeon & M.W.Chase	Chase 5612	AF262862	Pridgeon et al. [4]
Specklinia digitalis (Luer) Pridgeon & M.W.Chase	Karremans 5737	KF747806	This Study
Specklinia dunstervillei	Karremans 5966	KP012456	This Study
Specklinia endotrachys (Rchb.f.) Pridgeon & M.W.Chase (1)	Blanco 961	KC425784	This Study
Specklinia endotrachys (Rchb.f.) Pridgeon & M.W.Chase (2)	Blanco 961	KF747810	This Study
Specklinia fuegi (Rchb.f.) Solano & Soto Arenas	Karremans 5600 (JBL-Spirit)	KC425786	Bogarín et al. [5]
Specklinia fulgens (Rchb.f.) Pridgeon & M.W.Chase	Chase 5630	AF262872	Pridgeon et al. [4]
Specklinia glandulosa (Ames) Pridgeon & M.W.Chase	Karremans 5501	KC425792	This Study
Specklinia glandulosa (Ames) Pridgeon & M.W.Chase	Karremans 3265	KC425791	This Study
Specklinia glandulosa (Ames) Pridgeon & M.W.Chase	Karremans 2945	KP012452	This Study
Specklinia glandulosa (Ames) Pridgeon & M.W.Chase	Karremans 5944	KP012453	This Study
Specklinia glandulosa (Ames) Pridgeon & M.W.Chase	Bogarín 2895	KP012454	This Study
Specklinia glandulosa (Ames) Pridgeon & M.W.Chase	Karremans 3268	KP012455	This Study
Specklinia grobyi (Bateman ex Lindl.) F.Barros	Chiron 04524	JQ306485	Chiron et al. [9]
Specklinia lanceola (Sw.) Lindl. (2)	Pridgeon s.n.	KC425838	Bogarín et al. [5]
Specklinia lanceola (Sw.) Lindl. (3)	Chase 1433	AF262861	Pridgeon et al. [4]
Specklinia lentiginosa (F.Lehm. & Kraenzl.) Pridgeon & M.W.Chase		AF275692	Pridgeon et al. [4]
Specklinia montezumae (Luer) Luer	Karremans 229 (JBL-Spirit)	KC425811	Bogarín et al. [5]
Specklinia montezumae (Luer) Luer	Karremans 5751	KF747816	This Study
Specklinia picta (Lindl.) Pridgeon & M.W.Chase	Van Den Berg 2146	JQ306384	Chiron et al. [9]
Specklinia pissina	Karremans 4797	KC425795	This Study
Specklinia pissina	Karremans 4839	KC425797	This Study
Specklinia pfavii (Rchb.f.) Pupulin & Karremans	Karremans 4825	KC425814	This Study
Specklinia pfavii (Rchb.f.) Pupulin & Karremans	Karremans 3656	KF747819	This Study
Specklinia pfavii (Rchb.f.) Pupulin & Karremans	JBL-11086	KF747820	This Study
Specklinia remotiflora Pupulin & Karremans (4)	Chase 1303	AF262859	Pridgeon et al. [4]
Specklinia remotiflora Pupulin & Karremans (1)	Karremans 4798a	KC425818	This Study
Specklinia remotiflora Pupulin & Karremans (2)	Karremans 4798b	KC425819	This Study
Specklinia remotiflora Pupulin & Karremans (3)	Karremans 4854	KC425820	This Study
Specklinia sp.	Karremans 6025	KP012457	This Study
Specklinia sp.	Pupulin 7709	KC425824	This Study
Specklinia spectabilis (Ames & C.Schweinf.) Pupulin & Karremans	Bogarín 7401	KC425830	This Study

(Continued)



Table 1. (Continued)

Taxon	Sequence Voucher	GenBank Accession Number	Sequence Source
Specklinia spectabilis (Ames & C.Schweinf.) Pupulin & Karremans	Karremans 5699	KC425828	This Study
Specklinia subpicta (Schltr.) F.Barros	Chiron 11046	JQ306389	Chiron et al. [9]
Specklinia succulenta Bellone & Archila	Bellone 680	JQ306383	Chiron et al. [9]
Specklinia tribuloides (Sw.) Pridgeon & M.W.Chase (1)	Chase 5615	AF262867	Pridgeon et al. [4]

Nomenclature

The electronic version of this article in Portable Document Format (PDF) in a work with an ISSN or ISBN will represent a published work according to the International Code of Nomenclature for algae, fungi, and plants, and hence the new names contained in the electronic publication of a PLOS ONE article are effectively published under that Code from the electronic edition alone, so there is no longer any need to provide printed copies.

Results

Photography

The color illustrations of species of the *Specklinia endotrachys* complex (<u>Fig 1</u>) shows a morphologically distinct entity, *Specklinia dunstervillei* (<u>Fig 1A and 1B</u>), recognized amongst others by the smaller flowers and shortly apiculate petals.

Phylogenetic analysis

The consensus gene tree (Fig 2) was obtained from a BEAST analysis of a matrix of 45 ITS sequences (Table 1), including 12 accessions belonging to 5 different species of the *Specklinia endotrachys* complex. The accessions of *S. dunstervillei*, *S. endotrachys*, *S. pfavii*, *S. spectabilis* and *S. remotiflora* are found in a highly supported monophyletic clade (P.P. = 0.99), sister to the accessions of *S. montezumae*.

The pairwise comparison of the ITS sequence of the accessions of the *S. endotrachys* species complex (Table 2) shows that the sequences of *Specklinia endotrachys* and *S. spectabilis* do not differ from each other, while *S. dunstervillei* differs in 2 bases from those species. *Specklinia pfavii* and *S. remotiflora* differ in 4 and 10 bases respectively, from the three before mentioned species. *Specklinia montezumae* differs in 9 to 11 bases from each of the members of the *S. endotrachys* species complex.

Table 2. Pairwise comparison of the number of single base differences amongst the nrITS sequences of *S. dunstervillei*, *S. endotrachys*, *S. montezumae*, *S. pfavii*, *S. spectabilis* and *S. remotiflora*.

	S. dunstervillei	S. endotrachys	S. pfavii	S. remotiflora	S. spectabilis	S. montezumae
Specklinia dunstervillei	-	2	4	10	2	10
Specklinia endotrachys	2	-	4	10	0	10
Specklinia pfavii	4	4	-	12	4	11
Specklinia remotiflora	10	10	12	-	10	9
Specklinia spectabilis	2	0	4	10	-	10
Specklinia montezumae	10	10	11	9	10	-



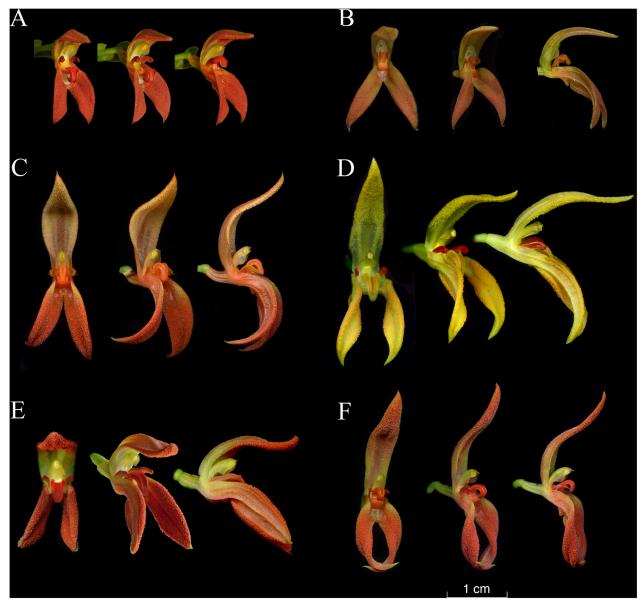


Fig 1. Comparison of flowers of species of the Specklinia endotrachys group. A. Specklinia dunstervillei (Karremans 5966). B. Specklinia dunstervillei (Karremans 5899). C. Specklinia endotrachys (Blanco 961). D. Specklinia pfavii (JBL-11086). E. Specklinia remotiflora (Bogarín 8181). E. Specklinia spectabilis (JBL-02641). All flowers shown in front, three-quarters side, and side views. Photographs by F. Pupulin (B-F) and R. van Vugt (A).

Discussion

Considering all the available evidence, including morphology, genetics, distribution and ecology, we find that the Venezuelan material labeled *Specklinia endotrachys*, actually belongs to an unnamed species, described here forth:

Specklinia dunstervillei Karremans, Pupulin & Gravendeel, sp. nov.

[urn:lsid:ipni.org:names: 77147597-1]



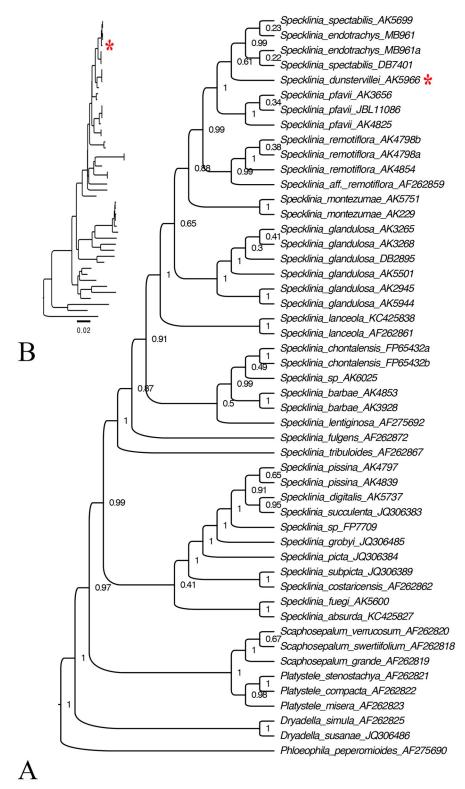


Fig 2. Phylogenetic relationship amongst the species of the Specklinia endotrachys complex. The trees were produced with an analysis of the nrITS dataset of 50 sequences using BEAST v1.6.0. Node values are posterior probabilities. The tree was edited using FigTree v.1.3.1. A. Tree with branches transformed to be of equal length. B. Branch lengths relative to relative number of changes.



Type. Venezuela. Without collecting data, cultivated by Jacobus Wubben in the Netherlands. Flowered in cultivation on March 29th 2013, *A.P. Karremans* 5966 & B. Gravendeel (holotype, JBL-spirit!; isotype, L-spirit!; Figs <u>1</u> and <u>3</u>).

The species is similar to Specklinia endotrachys but can be distinguished by the small habit, shorter leaves, smaller flowers, the flat, non-ascending dorsal sepal (vs. ascending), and the obtuse, shortly apiculate (vs. emarginate and long mucronate) petals.

Epiphytic, caespitose, erect herb to 10 cm tall, excluding the inflorescence. Roots fibrous, flexuous, glabrous. Stem abbreviated, terete-cylindric, to 1 cm long, monophyllous, completely concealed by papyraceous, subancipitous, acute sheaths to 1 cm long. Leaf narrowly obovateoblanceolate, 6-10 x 1.2-1.8 cm, minutely emarginate at apex, the mid-vein protruding abaxially into a small apicule, gradually tapering toward the base into a deeply conduplicate petiole with strongly ancipitous margins, subcoriaceous. Inflorescence borne laterally from the apex of the stem, without an annulus, a distichous, successively flowered raceme, with only one flower open at once, up to 12-13 cm long; peduncle flattened, ancipitous, to 11-12 cm long, with 2-3 amplectent, ancipitous, subacute bracts, 5-6 mm long. Floral bracts infundibuliform, broadly ovate, acute to subacuminate, 2–3 mm long. Pedicel cylindric, glabrous, 4 mm long, persistent. Ovary subclavate, 2 mm long, green tinted with orange. Flowers with reddish-orange sepals, petals and lip, the column greenish-yellow, lightly tinted orange. Sepals fleshy, densely papillose in the inner surface except at the base; dorsal sepal elliptic, 3-veined, acute, the base whitish semi-hyaline, flushed with orange along the veins, the distal two thirds densely papillose, 14- 16×5 –6 mm; lateral sepals narrowly elliptic-oblanceolate, subfalcate, 3-veined, 13– 15×4 mm, the base saccate, membranaceous-hyaline, the apex acute, gently twisting above the middle, the midvein strongly carinate abaxially. Petals small, ligulate-falcate, truncate, shortly apiculate, porrect, 3–4 × 1 mm, 1-veined, papillose-thickened toward the concave apex. Lip small, longitudinally arched-convex in natural position, thinly articulate with the column foot by a hyaline claw, narrowly elliptic-lanceolate when expanded, obtuse, the apex reflexed, appearing minutely retuse, the clawed base thickened, transversely minutely gibberose, $5-6 \times 2$ mm, provided with 2 slender keels, fringed-lacerate at the base, gently converging from the base of the lamina to near the apex. Column arched, terete-slender at the base, 4.5-5.0 mm long without the foot, provided with broad membranous wings serrulate along the margins, at the apex forming a deeply cucullate, serrulate clinandrium; column foot forward-projecting, stout, fleshy, incurved, 1.5 mm long. Anther cap deeply cucullate, ovate, 2-celled. Pollinia 2, obovatecomplanate, minutely hooked at the base, lacking caudicles.

Note: Only the specimens from Venezuela were used for the description (*Dunsterville 757* and *Karremans 5966*).

Etymology. The name honors G. C. K. Dunsterville, who prepared the first known illustration of the species.

Additional material examined. Costa Rica. Without collecting data, cultivated by Gerson Villalobos in Moravia, San José. Flowered in cultivation on September 1st 2013, *A.P. Karremans* 5899 (JBL-spirit!; Figs 4 and 5). Venezuela. Boconó-Guaramacal penetration road. About 8000 ft. in rain forest, *G. C. K. Dunsterville 757* (illustration of voucher in Dunsterville and Garay [7]!; Figs 6 and 7).

Other records. Colombia. Without collecting data, illustration of voucher in Ortiz and Uribe [15].

Distribution. The material collected by Dunsterville in Venezuela comes from the road between Boconó and Guaramacal in Trujillo, on the Cordillera de Merida, a branch of the Colombian Andes, at an elevation of about 2400 m. A few specimens that have been found in private collections in Costa Rica and Colombia lack precise location data.

Key to the species of the empusellous species of Specklinia.



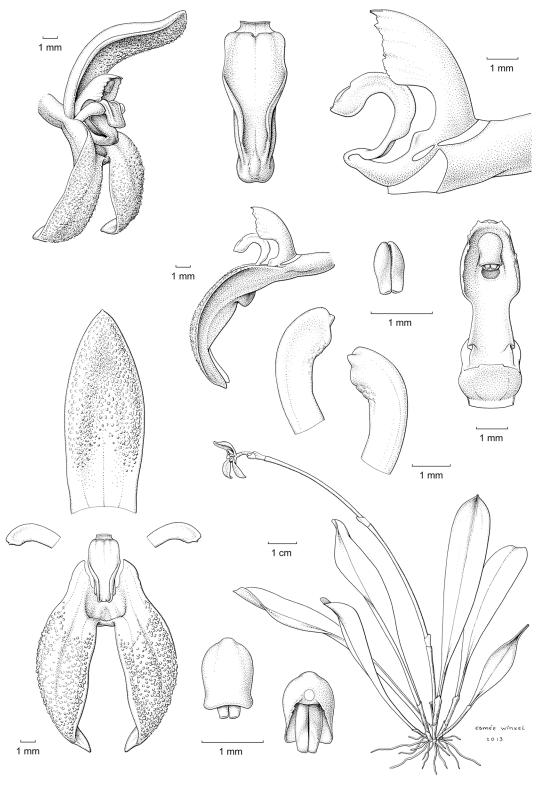


Fig 3. Specklinia dunstervillei Karremans, Pupulin & Gravendeel . A) Habit; B) Flower; C) Dissected perianth; D) Lateral view of the lip placement relative to lateral sepals; E) Column and lip, lateral view; F) Column, ventral view; G) Lip, extended; H) Petals; I) Anther cap with pollinia; J) Pollinia. Drawn from the holotype (Karremans 5966) by Esmée Winkel.





Fig 4. Acuarela of Specklinia dunstervillei Karremans, Pupulin & Gravendeel. By Sylvia Strigari, based on Karremans 5899 (JBL).

- 1. Flowers yellow with red petals; lip acute, yellow, with a central red line; column wings 1. Flowers reddish-orange; lip rounded to truncate, concolorous orange; column wings nar-

- 3. Plant repent, inflorescence lax; flowers bright orange in both outer and inner surfaces; col-



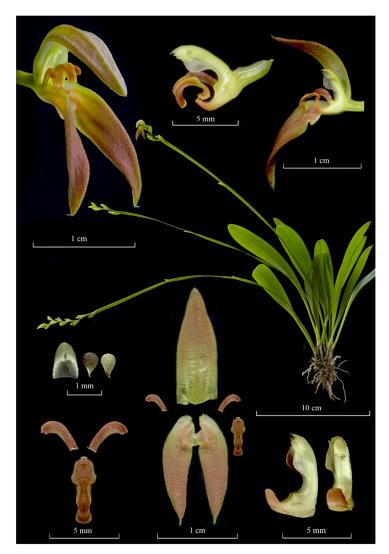
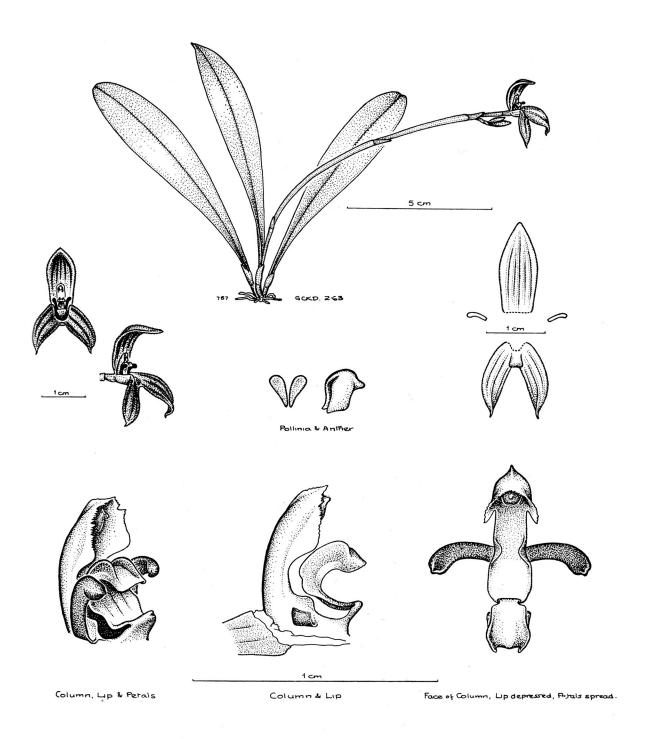


Fig 5. Lankester Composite Dissection Plate (LCDP) of Specklinia dunstervillei Karremans, Pupulin & Gravendeel. A. Habit. B. Flower. C. Transverse section of a whole flower. D. Dissected perianth. E. Column and lip, lateral view. F. Petals and lip. G. Column in ventral and lateral view. H. Pollinia and anther cap. Based on photographs of Karremans 5899 (JBL) by A.P. Karremans.

- 2. Flowers spreading, lateral sepals twisted and/or bent, petals apiculate to mucronate 4





PLEUROTHALLIS ENDOTRACHYS RCHB.F.

Fig 6. Specklinia dunstervillei Karremans, Pupulin & Gravendeel. Illustrated by G. C. K. Dunsterville in Dunsterville and Garay [7] from a plant found in Trujillo, Venezuela (Dunsterville 757). Reproduced with the kind permission of the Orchid Herbarium of Oakes Ames, the Harvard University Herbaria.





Fig 7. Specklinia dunstervillei Karremans, Pupulin & Gravendeel. Unpublished photographs by G. C. K. Dunsterville from the plant found in Trujillo, Venezuela (*Dunsterville* 757). A. The plant habit. B. Detail of the flower. Reproduced with the kind permission of the Orchid Herbarium of Oakes Ames, the Harvard University Herbaria.

Ecology. The typical form of *Specklinia dunstervillei* is that found in Colombia and Venezuela (Figs 1a, 3 and 6). The material illustrated by Dunsterville, which lacks an herbarium voucher (Gustavo Romero pers. comm.), was collected on the Cordillera de Merida at about 2400 m in elevation. That makes the new species the one with the most Easternly distribution in the complex and, together with *S. remotiflora*, the only one to grow in cloud forests at high elevations above 1800 m. A photograph of this species was published in Ortiz and Uribe [15] without precise locality, nonetheless it was definitively taken from a Colombian plant (Carlos Uribe V. pers. comm.). Considering that the Cordillera de Merida extends into Colombia, it is indeed not surprising that the species is also found there. Finally, we have chosen to regard the material found in Costa Rican private collections as *S. dunstervillei* as they are morphologically most similar to that species concept, nevertheless they do show quite some variation (Figs 1b, 4 and 5). The studied Costa Rican specimens lack specific collection data.

Like the other species of the *S. endotrachys* complex, *S. dunstervillei* has nectar secreting stomata placed on the apex of the warts that cover the surface of the sepals [1,2,3,6]. The released nectar gives the sepals a wet-glossy appearance, as noted by Dunsterville and Garay [7], and is given special attention by visiting fruit flies. At Lankester Botanical Garden the flowers were frequently visited by up to 6 individuals of *Drosophila* spp. The flies wander about on the sepals, sucking on the papillae rich surfaces, occasionally removing the pollinia, after stepping on the movable lip and being adpressed against the column (Fig.8).

Morphological recognition. *Specklinia dunstervillei* is morphologically similar to other members of the *S. endotrachys* complex. Within *Specklinia* they are recognized by the relatively large habit and flowers, the long, successive, flat, ancipitous inflorescence, the bright yellow to orange-red flowers, the verrucose sepals, the relatively minuscule petals and the highly sensitive strongly concave lip. Amongst its closest relatives, *S. dunstervillei* is recognized by the tiny habit, caespitose plants, short leaves (up to about 10 cm), the small flowers (dorsal sepal $14-16 \times 5-6$





Fig 8. Drosophila sp. adpressed by the lip against the column of Specklinia dunstervillei Karremans, Pupulin & Gravendeel (Karremans 5899). Photographed by A.P. Karremans.

mm), the flattening, non-ascending dorsal sepal, the lateral sepals that spread, slightly twisting downwards and the obtuse petals, which are shortly apiculate.

Genetic recognition. The nrITS sequence of the accession of *Specklinia dunstervillei* differs in two base pairs (out of just under 800bp) from the accessions of both *S. endotrachys* and *S. spectabilis*, which are identical to each other (Table 2). Considering that the species of the *S. endotrachys* complex differ in only 9 to 11 bases (between 1.1 and 1.4%) from an outgroup as morphologically distinct as *S. montezumae*, it becomes highly plausible that species with significant morphological and ecological differences within the complex itself, such as *S. endotrachys* and *S. pfavii*, can differ in just about 0.5% bases. Therefore it is also not unusual that more closely related species such as *S. endotrachys* and *S. dunstervillei* differ in only 2 bases, or that the sisters *S. endotrachys* and *S. spectabilis* do not differ at all in this particular DNA barcode.

The internal transcribed spacer (ITS) of nuclear ribosomal DNA has consistently shown high levels of discrimination among many species of plants [16] including Pleurothallidinae species, and are well suited for a broad range of phylogenetic studies [4,10,17,18]. However, in current literature DNA data are mostly used to support so called "cryptic" novel taxa, when the accession of a said taxon does not cluster with other accessions of the *a priori* believed same taxon [19,20,21,22]. When different accessions of a species complex cluster together authors seem to believe that there is evidence that they are a single species, however that is not only an illogical conclusion but also frequently untrue. Be it accessions of the same species, or sister species, or species of a single genus or the same family, any two accessions will cluster together with high support if they are relatively much more similar to each other than to the other sequences analyzed. The only measurable difference between the mentioned sister pairs would



be the branch length (as a measure of number of base pair changes). However, as there is no established similarity threshold above which one should consider two sequences to belong to two different species, or below which they should be considered a single species, it is at this time also not possible to infer from branch lengths alone if we are dealing with a single or more than one species. Meyer and Paulay [23] found that this especially counts for taxonomically understudied groups that are not yet thoroughly sampled.

In Pleurothallidinae, DNA data have been seldomly used to support novel species descriptions. Ramos-Castro et al. [24] used an nrITS based phylogeny to prove that their novel taxon Stelis zootrophionoides Castañeda-Zárate & Ramos-Castro belonged in that genus, however they did not venture into using DNA data to set their novelty apart from other taxa in the genus. In the phylogeny presented, the single accession of S. zootrophionoides clusters, as would be expected, with that of the morphologically similar S. nigriflora (L.O.Williams) Pridgeon & M.W.Chase. Therefore, their phylogeny by itself could not prove that the accessions belonged to two different species, and the authors relied exclusively on morphological differences to establish their novelty. A species labeled *Specklinia* sp. in Chiron et al. [9], of which the nrITS sequence clustered with that of Specklinia marginata (Lindl.) Pridgeon & M.W. Chase, was later published as a distinct species using mainly morphological characters. In both cases the authors could also have argued that the nrITS sequence of the accession of their novelty differed in about 1% from the sequence of the accession of its closest relative with which it clustered. A different approach was that of Meyer et al. [25], who used the presence of insertions/deletions in the trnH-psbA and ycf1 sequences of accessions of Dracula radiosa (Rchb.f.) Luer and close relatives to support the establishment of their novel species Dracula trigonopetala Gary Mey. & L. Baquero. In that case their novelty differed in less than 0.3% from its closest relatives (as an insertion/deletion counts for a single change), however, the 26 bp unique insertion in the trnH-psbA sequence of their novelty stands out in an otherwise quite conserved region and therefore easily sets it apart from its close relatives.

Conclusions

A common misconception amongst modern authors is that DNA data will in itself resolve taxonomic issues. In fact, that two sequences are identical in a particular region does not guarantee that they belong to the same species, nor does the fact that they differ guarantee that they belong to different species. DNA data albeit less subjective than morphological characterization, are nonetheless subject to many of the same pitfalls, which can basically be summarized as: 1) interpretation of the type specimens and correct application of names; 2) reduction or elimination of data reading mistakes; 3) adequate interpretation of the observed variation.

Considering all the evidence at hand we conclude that the specimen illustrated in Dunster-ville and Garay [7] from Venezuela and labeled *Specklinia endotrachys* actually represents an until now unnamed species. With the inclusion of the here proposed novelty, *Specklinia dunstervillei*, the *S. endotrachys* species complex now consists of six distinctly named species, instead of a single variable species. Its highest diversity is found in Costa Rica and Panama, but this might change as additional material becomes available from less sampled areas; we therefore do not reject the possibility of discovering other novelties in this group in the future.

Supporting Information

S1 File. Alignment matrix of 43 nrITS sequences used to infer the molecular phylogeny presented in Fig 2. The ends were trimmed and gaps were regarded as missing data. (NEX)



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Author Contributions

Conceived and designed the experiments: AK FP BG. Performed the experiments: AK FP. Analyzed the data: AK FP. Contributed reagents/materials/analysis tools: AK FP BG. Wrote the paper: AK.

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