

氧和二氧化碳慢性感知机制的免疫调节作用

田向向^{1,2}, 万延民³, 王万海^{2*}, 朱召芹^{1*}

(¹上海市公共卫生临床中心检验医学科, 上海 201508; ²郑州大学第一附属医院检验科, 河南省检验医学重点实验室, 郑州 450052; ³复旦大学附属华山医院感染科, 国家传染病医学中心, 上海市传染病与生物安全应急重点实验室, 上海 200040)

摘要: 感知氧和二氧化碳浓度变化并做出适当响应是机体维持正常生理功能的重要环节。目前已知的氧与二氧化碳感知信号通路主要分为急性感知(acute sensing)和慢性感知(chronic sensing)两类。通过由中枢和外周化学感受器介导的急性感知信号通路, 机体能够对氧或二氧化碳水平的突然变化做出快速响应, 即时调整通气量; 而当长时间处于氧或二氧化碳浓度异常环境时, 机体则不仅需要通过急性感知系统调整通气量, 还需要通过慢性感知通路调整组织、细胞的基因转录和代谢状态来适应异常的氧和二氧化碳浓度。慢性感知是由分布于各种细胞中的分子感受器介导的, 该通路对细胞功能具有广泛而复杂的影响。该文将在扼要回顾氧和二氧化碳感知机制的基础上, 着重阐述二者的慢性感知通路与机体免疫应答之间的关系。

关键词: 氧感知机制; 二氧化碳感知机制; 低氧诱导因子; 可溶性腺苷酸环化酶; 免疫应答

The role of oxygen and carbon dioxide chronic sensing pathways in immune regulation

TIAN Xiangxiang^{1,2}, WAN Yanmin³, WANG Wanhai^{2*}, ZHU Zhaoqin^{1*}

(¹The department of laboratory medicine, Shanghai Public Health Clinical Center, Shanghai 201508, China;

²Clinical Laboratory, The First Affiliated Hospital of Zhengzhou University, Key Laboratory of Laboratory Medicine of Henan Province, Zhengzhou 450052, China; ³Department of Infectious Disease of Huashan Hospital, National Medical Center for Infectious Diseases and Shanghai Key Laboratory of Infectious Diseases and Biosafety Emergency Response, Fudan University, Shanghai 200040, China)

Abstract: Sensing and responding to changes in oxygen and carbon dioxide concentrations are a vital part for the maintenance of normal physiological functions. There are two different mechanisms in oxygen and carbon dioxide sensing, including acute sensing and chronic sensing. Through the acute sensing pathway mediated by central and peripheral chemoreceptors, the body could respond quickly to sudden changes in oxygen or carbon dioxide levels and adjust ventilation instantly. While, if being exposed to abnormal oxygen or carbon dioxide concentration for a long time, the body will not only need to adjust ventilation through the acute sensory mechanism, but also need to modulate the genes transcription and the metabolism of tissues and cells through the chronic sensing pathway to adapt to the abnormal environment. Chronic sensing pathway is mediated by molecular sensor expressed in various cells, which has extensive and complex effects on cellular

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第一作者: E-mail: t0102@gs.zzu.edu.cn

*通信作者: 朱召芹, E-mail: zhaqinzh@163.com; 王万海, E-mail: wanghai0371@126.com

pathways. In this review, we briefly summarized the mechanisms of oxygen and carbon dioxide sensing and focused on the relationships between the chronic sensing pathways of O₂/CO₂ and immune responses.

Key Words: oxygen sensing mechanism; carbon dioxide sensing mechanism; HIF-1 α ; soluble adenylyl cyclase; immune response

氧气和二氧化碳是参与生命活动的两种至关重要的气体分子,由二者水平异常引起的病理效应(如低氧血症、高碳酸血症)严重时可危及生命^[1]。机体内氧气的消耗和二氧化碳的产生是相互关联的,因此在细胞和组织中二者的浓度存在密切的负相关关系^[2]。在正常生理状态下,机体能够通过分布于中枢和外周的感知反馈系统维持血液中氧气和二氧化碳含量的稳定:在氧气浓度下降或者二氧化碳浓度升高时,机体能够及时的通过调整通气量使之恢复正常水平^[3,4]。但在某些病理状态下,例如急性呼吸窘迫综合征、阻塞性睡眠呼吸暂停综合征^[5]、慢性阻塞性肺疾病^[6]、感染性肺炎等,氧气与二氧化碳的浓度维持机制被打破,二者可单独或同时出现异常,并可通过相应的细胞感受器触发一系列下游病理反应。概括而言,机体对氧气与二氧化碳的感知可以分为急性感知和慢性感知两类。其中,急性感知是指机体外周神经通过化学感受器探测氧与二氧化碳分压的变化,继而通过神经反馈调节换气节律;慢性感知是指机体通过分布于细胞表面或细胞内的受体分子探测氧与二氧化碳浓度的变化,继而调节细胞代谢和基因转录(图1)^[1]。本文将着重回顾氧气与二氧化碳的慢性感知机制,并扼要阐述其与免疫应答的关系。

1 细胞的氧浓度感知机制及其对机体免疫应答的影响

如前所述,机体能够通过急性和慢性感知通路感受体内氧浓度变化,急性感知可通过调节呼吸频率和深度从整体层面调节体内氧浓度;而慢性感知机制则可通过分布于细胞中的氧分子感受器来调节机体细胞的转录和代谢,帮助机体组织细胞适应氧浓度的变化。慢性感知机制主要通过调节基因转录得以实现,已发现的能被氧浓度调节的转录因子包括低氧诱导因子-1 α (hypoxia-inducible factor-1 α , HIF-1 α)、活化B细胞的核因子

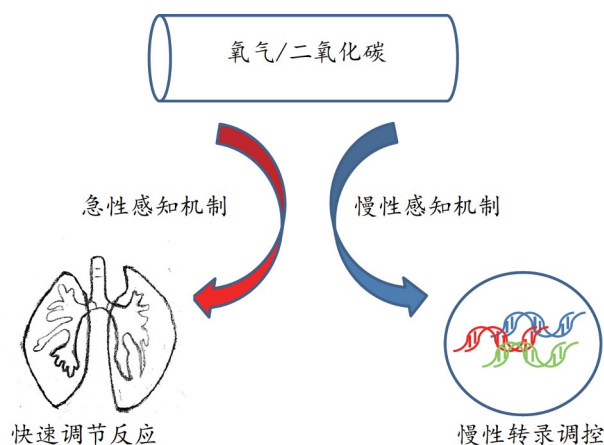


图1 氧和二氧化碳的急性与慢性感知^[1]

κ 轻链增强子(nuclear factor kappa-light-chain-enhancer of activated B cells, NF- κ B)、转录激活因子4(activating transcription factor 4, ATF4)、肿瘤蛋白P53(tumour protein p53, P53)等(表1)^[7,8]。这其中多数因子需要在特定细胞类型或特定细胞状态下发挥作用,而HIF通路的调节作用是广泛存在的,被认为是细胞氧浓度感知信号通路的总调节因子(master regulator)^[7]。

1.1 低氧诱导因子家族的结构和功能特征

HIFs家族在多细胞动物中广泛存在,该分子家族已知成员包括HIF-1、HIF-2和HIF-3。活化的HIFs是由HIF- α 和HIF- β 亚基构成的异源二聚体,前者是氧敏感的亚基,在常氧条件下极不稳定;后者是组成性表达亚基。与广泛表达的HIF-1 α 相比,HIF-2 α 则主要表达于癌组织、肺、内皮组织和颈动脉体,促进侵袭性肿瘤病情恶化^[9,10]。随着对HIF家族的研究逐渐深入,目前认为,低氧环境也可使HIF-3 α 表达增加,但此过程需要HIF-1 α 诱导^[11]。HIF-1 α 作为缺氧感受器被Semenza提出^[12],HIF-1 α 的转录和合成不受氧的影响,但是,在常氧状态下,HIF-1 α 合成后迅速降解,导致HIF-1 α 基本无法被检测到,其降解主要通过2个经典途径实现(图2)^[13]。(1)HIF-1 α 需氧降解区域(oxygen-

表1 能够感知氧浓度变化的转录因子^[7,8]

| 缩写 | 英文全名 | 中文全名 |
|---------------|--|---------------------------|
| HIF* | hypoxia-inducible factor | 低氧诱导因子 |
| NF-κB* | nuclear factor kappa-light-chain-enhancer of activated B cells | 活化B细胞的核因子κ轻链增强子 |
| ATF4 | activating transcription factor 4 | 转录激活因子4 |
| FOXO3a* | forkhead box O3a* | 叉形头转录因子O亚型3a |
| P53 | tumour protein p53 | 肿瘤蛋白P53 |
| CREB* | cyclic AMP response element binding protein* | 环一磷酸腺苷反应元件结合蛋白 |
| Myc | Myc avian myelocytomatosis viral oncogene homolog | Myc禽髓细胞瘤病毒癌基因同源物 |
| AP-1 | activating protein-1 | 激活蛋白1 |
| NF-IL6/C/EBPβ | CCAAT/enhancer binding protein(C/EBP)beta | CCAAT/增强子结合蛋白β |
| EGR-1 | early growth response-1 | 早期生长反应-1 |
| GATA-2 | GATA binding protein-2 | GATA结合蛋白-2 |
| Ets-1 | V-Ets avian erythroblastosis virus E26 oncogene homolog-1 | V-Ets禽成红细胞病病毒E26致癌基因同源物-1 |
| Dec1/2 | differentially expressed in chondrocytes 1/2 | 软骨细胞1/2中的表达差异 |
| RTEF-1 | related transcriptional enhancer factor-1 | 相关转录增强因子-1 |
| Purα | purine-rich binding protein alpha | 富含小嘌呤的结合蛋白α |
| STAT | signal transducer and activator of transcription | 信号传感器和转录激活因子 |
| GADD153 | growth arrest and DNA damage-153 | 生长停滞和DNA损伤-153 |
| SP1/SP3 | SP/XKLF | 特异性蛋白1/特异性蛋白3 |
| MASH2 | mammalian achaete-scute homologous protein-2 | 哺乳动物的同源蛋白-2 |

*表示能感知氧气和二氧化碳浓度变化的转录因子

dependent degradation domain, ODD)的第402和/或564位脯氨酸残基被脯氨酸羟化酶(the prolyl hydroxylase domain, PHD)羟基化。羟基化本质上是氧化反应, 脯氨酸羟化酶需要氧气作为底物, 缺乏氧气脯氨酸羟化酶就无法发挥作用。羟基化的HIF-1α可被希佩尔-林道蛋白(von Hippel-Lindau protein, pVHL)识别, 使得pVHL介导的E3泛素连接酶复合体与羟基化的HIF-1α结合, 导致HIF-1α的泛素化和蛋白酶体的降解^[14]。(2)通过HIF-1抑制因子(factor inhibiting HIF-1, FIH-1)使第803位天冬酰胺残基羟基化, 抑制cAMP反应元件结合蛋白(cyclic-AMP response binding protein, CBP)与腺病毒E1A相关的300 kDa蛋白(adenovirus E1A-associated 300 kDa protein, p300)的结合, 从而抑制HIF通路^[15]。由于PHDs发挥作用需要氧气作为底物, 因此在低氧条件下, PHDs无法发挥羟基化作用, 使得HIF-1α不被降解且能够入核与HIF-1β结合形成二聚体复合物, 促进下游靶基因的表达^[16]。HIF-1α能够导致很多靶基因的上调, 例如促红细胞生成素(erythropoietin, EPO)、血管生长

因子(vascular endothelial growth factor, VEGF)以及对心脏有保护作用的抗氧化酶。HIF-1α的作用有利有弊, 例如, 在缺氧条件下, 它会使VEGF的生成增加, 引发血管生成, 增加氧的利用率, 缓解组织和器官的缺氧程度, 避免细胞死亡^[17]。但与此同时, VEGF也可增加血管的通透性, 使免疫细胞更易迁移到组织中, 加重炎症程度^[18]。

1.2 HIF-1α与免疫应答的关系

生理或病理性原因均可导致组织微环境缺氧。生理性缺氧经常出现在细胞增殖快, 代谢需求高的组织部位, 例如细胞增殖活跃的免疫器官或组织^[19]。由病理因素导致的组织缺氧可见于肿瘤^[20]以及炎症部位^[21]。当血液富氧环境中的循环免疫细胞被招募到缺氧组织环境之中时, HIF能够帮助这些免疫细胞快速适应缺氧微环境^[22]。HIF在暴露于缺氧环境的天然免疫细胞和适应性免疫细胞中发挥重要的调节作用。在树突状细胞中, HIF-1α可调节其存活、迁移、抗原呈递、干扰素合成和分化^[23,24]。在T细胞中, HIF-1α不仅可调节其存活和分化, 还可调节其抗肿瘤能力^[13]。同样, HIF-

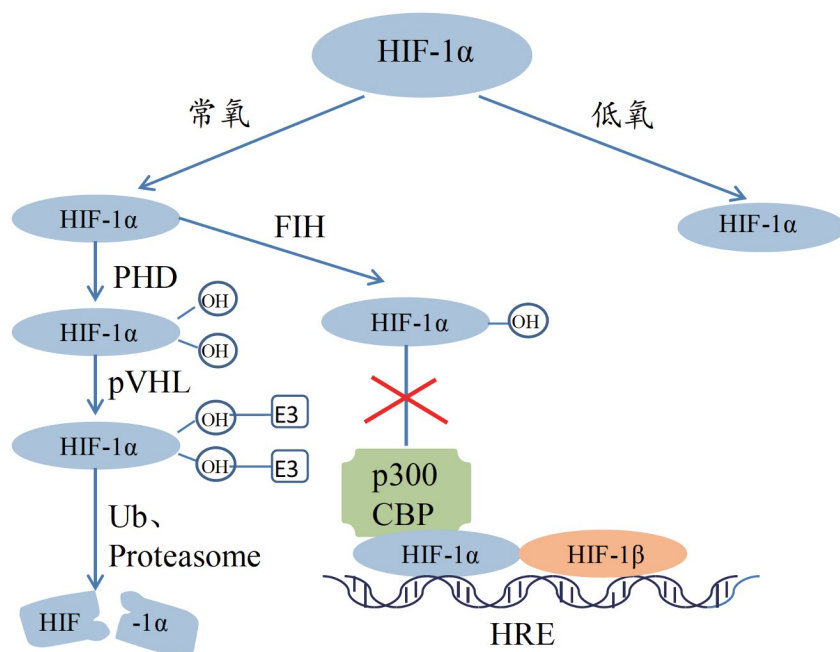


图2 HIF-1 α 在常氧条件下的调控方式^[13]

1 α 也能调节B细胞的存活^[25]。除此之外，HIF-1 α 也可通过代谢途径调节免疫细胞，例如其可以通过转录上调糖酵解相关基因的表达从而增加糖酵解的速率，糖酵解速率的增加反过来又可促进巨噬细胞、树突状细胞、T细胞和B细胞等免疫细胞的激活^[26]。与此同时，激活的免疫应答也能通过细胞因子或者代谢产物信号通路调节HIF-1 α 的表达水平或转录活性^[22]。例如，被炎症信号(细胞因子、细菌产物)激活的NF- κ B可促进HIF-1 α 的转录活性；免疫代谢物S-2-羟基戊二酸(S-2-hydroxyglutarate, S-2-HG)、气体介质(NO、H₂S)也可增强HIF-1 α 的转录活性。此外，HIF-1 α 与NF- κ B信号通路存在交互作用，HIF-1 α 能够促进肿瘤坏死因子- α (tumor necrosis factor- α , TNF- α)和白介素-6(interleukin-6, IL-6)等炎症因子的产生(图3A)^[27]，而白介素-1 β (interleukin-1beta, IL-1 β)等炎症因子也可以促进HIF-1 α 的表达^[28]，在参与炎症应

答的同时，HIF-1 α 还能够增强巨噬细胞和中性粒细胞的功能，提高其抵御细菌感染的能力(图3B)^[27]。不仅如此，HIF-1 α 也可促进非免疫细胞的炎症反应。例如，HIF-1 α 与脂肪细胞中炎症标志物(TNF- α 、IL-6等)的升高呈正相关^[29]；敲低急性期缺血性脑卒中小鼠小胶质细胞中的HIF-1 α ，可减少TNF- α 的产生，促进神经元的存活^[30]。

在病毒感染和抗感染免疫应答发生过程中，HIF-1 α 也被发现扮演重要角色。甲型流感病毒H5N1亚型感染猕猴后能够迅速上调肺组织中HIF-1 α 的表达水平^[31]。Uematsu等^[32]发现，敲除*Mint3*/*Apba3*基因能够显著减轻由H1N1感染导致的炎症应答，并改善小鼠肺炎症状。由于*Mint3*/*Apba3*敲除能够抑制HIF-1 α 的激活^[32,33]。因而这些发现间接提示了HIF-1 α 在流感病毒感染过程中具有促进炎症应答的作用。此外，体外实验证明HIF-1 α 在流感病毒诱导炎症反应过程中发挥关键的调控作

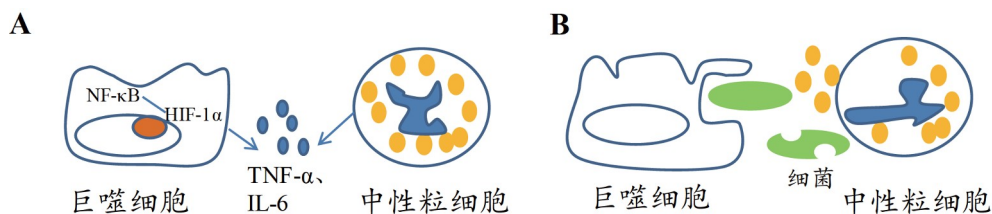


图3 HIF-1 α 能够促进炎症因子产生并提高巨噬细胞和中性粒细胞的功能^[27]

用, 抑制HIF-1 α 可显著降低TNF- α 和IL-6的表达^[34]。此外, HIF-1 α 缺陷的肺上皮细胞通过减少糖酵解和增强自噬, 促进了甲型流感病毒H1N1亚型的复制^[35]。这表明HIF-1 α 在甲型流感病毒感染中的病毒复制、细胞因子反应和肺部炎症中起重要作用。而且, 在新型冠状病毒导致的新冠状肺炎(corona virus disease 2019, COVID-19)中, 由病毒感染导致的HIF-1 α 在肺内累积可导致趋化因子释放并致使单核细胞、树突状细胞和中性粒细胞在肺部快速浸润。这种浸润会导致气管甚至毛细血管的阻塞, 这种阻塞会进一步加重缺氧, 迫使细胞进入厌氧糖酵解途径, 促进病毒复制^[36]。除病毒感染外, 脂多糖(lipopolysaccharide, LPS)在缺氧条件下也可诱导出更高水平的TNF和IL-6, 而将HIF-1 α 基因敲除后, LPS刺激后产生的TNF和IL-6减少^[37,38]。

2 细胞的二氧化碳浓度感知机制及其对机体免疫应答的影响

与氧的细胞感知机制研究进展不同, 在二氧化碳浓度感知信号通路上迄今为止尚未发现占据支配地位的主调控因子(master regulator)^[1,39], 但有一些转录因子(表2)^[8,40]已被证实参与由二氧化碳浓度变化引起的信号转导, 例如NF- κ B、热激转录因子1(heat shock transcription factor 1, HSF1)、叉形头转录因子O亚型3a(forkhead box O3a, Foxo3a)和环一磷酸腺苷反应元件结合蛋白(cyclic AMP response element binding protein, CREB)。NF- κ B参与炎症、免疫、细胞生存、凋亡和细胞周期等相关基因的调控, 体内pCO₂升高可伴随着NF- κ B信号通路发生改变, 但二者之间的因果关系仍然

未知^[41,42]。此外, 另有一些因子被发现能够间接感知体内二氧化碳浓度的变化, 例如可溶性腺苷酸环化酶(soluble adenylyl cyclase, sAC)可被HCO₃⁻激活。由于体内的CO₂、H⁺和HCO₃⁻浓度可在碳酸酐酶作用下达到瞬间平衡状态, 因此sAC通常也被认为是重要的CO₂浓度和pH值感受器^[43]。

2.1 sAC在二氧化碳慢性感知中的作用

近年来, sAC被认为可通过感知HCO₃⁻浓度的改变间接感知体内pCO₂的改变。sAC在20世纪70年代被发现^[44], 最初被认为只存在于哺乳动物精细胞中, 后被证实在多种组织中均有表达^[45,46]。腺苷酸环化酶(adenylyl cyclases, ACs)是将ATP转化为cAMP的一类酶家族, 哺乳动物细胞中存在两套独立调节cAMP的信号转导系统: 跨膜型腺苷酸环化酶(transmembrane adenylate cyclase, tmAC)和sAC。二者的细胞定位和调控方式均有不同。首先, tmAC只局限于细胞膜, 而sAC可分布于整个细胞, 包括线粒体、中心粒、有丝分裂纺锤体、中间体和细胞核等特定的亚细胞器中^[47]。其次, tmAC具有跨膜结构域, 可与G蛋白偶联受体(G-protein coupled receptor, GPCR)结合, 在细胞质中完成ATP向cAMP的转化, sAC则不与GPCR结合, 而是在碳酸酐酶的介导下, 被水合CO₂解离出的HCO₃⁻或通过Na⁺/HCO₃⁻离子转运体进入到细胞内的HCO₃⁻激活, 即机体环境中HCO₃⁻升高时, 在Ca²⁺的协同作用下, sAC活性增强, 使ATP转化为cAMP(图4)^[48]。sAC可不依赖于钙调素直接被Ca²⁺和HCO₃⁻激活, Ca²⁺降低了底物和ATP之间的K_m, 而HCO₃⁻则降低了底物抑制效应, 增加了最大反应速率^[49,50]。ACs均含有C1和C2催化结构域, 二者在交界处形成了一个无催化活性的“口袋”结构, 可

表2 能够感知二氧化碳浓度变化的转录因子^[8,40]

| 缩写 | 英文全名 | 中文全名 |
|-----------------|--|--------------------------|
| Phox2b | paired-like homeobox 2b | 配对同源异型盒蛋白2B |
| NF- κ B* | nuclear factor kappa-light-chain-enhancer of activated B cells | 活化B细胞的核因子 κ 轻链增强子 |
| FOXO3a* | forkhead box O3a* | 叉形头转录因子O亚型3a |
| CREB* | cyclic AMP response element binding protein* | 环一磷酸腺苷反应元件结合蛋白 |
| HIF* | hypoxia-inducible factor | 低氧诱导因子 |
| sAC | soluble adenylyl cyclase | 可溶性腺苷酸环化酶 |
| HSF-1 | heat shock transcription factor 1 | 热休克因子1 |

*表示能感知氧气和二氧化碳浓度变化的转录因子

使激活剂与之结合，激活ACs。图4为sAC的调控机制^[44]。

2.2 sAC信号通路对机体免疫应答的影响

与氧气类似，二氧化碳也可影响机体感染与免疫应答。已有研究证实，将细胞或小鼠暴露于二氧化碳含量高于5%的气体环境中，可造成细胞分泌的巨噬细胞炎症蛋白(macrophage inflammatory protein, MIP)-1、MIP-2、IL-8和IL-6等促炎因子增加，也诱导了更为严重的小鼠肺部炎症^[51]。此外，二氧化碳诱导小鼠肺部炎症与吸入高二氧化碳时间相关，在用LPS诱导小鼠前分别用5%的二氧化碳预处理小鼠10 min和60 min，结果显示，预先吸入10 min高浓度二氧化碳可降低小鼠肺部损伤^[52]。最近，有研究指出，CO₂在小鼠肺部免疫应答方面的影响，对小鼠鼻腔注入灰尘提取物或脂多糖并立即将其分别置于含有430 ppm(正常值)，1 000 ppm(建筑物推荐上限值)，5 000 ppm(极限值)的CO₂环境下，发现在CO₂浓度在1 000 ppm时，先天免疫会被抑制，而当CO₂浓度升高至5 000 ppm时，免疫反应增强^[53]。在此前已有研究证明，短暂吸入低浓度的CO₂对临床疾病的治疗是安全有效的基础上^[54-56]，最近的研究证明，对原代血管内皮细胞每3 h进行11 min的10% CO₂孵育，可有效降低因COVID-19导致的IL-6的升高，使其有可能成为治疗COVID-19的一种治疗措施^[57]。

AC是GPCR下游的关键信号分子，通过调节cAMP的合成调控多种细胞功能，从而参与机体的多种病理生理过程。哺乳动物细胞表达了9个tmAC亚型AC1~9和1个sAC，也称为AC10^[58]。以往研究已经表明，腺苷环化酶-cAMP-PKA信号通路具有重要免疫调节作用^[59]。截至到目前，虽然已有较多研究证明激活后的sAC能够上调cAMP^[60,61]，但是关于sAC直接调节免疫反应的报道极少，其对机体免疫应答的直接调节作用尚不明晰。鉴于cAMP-PKA信号通路在免疫调节中的重要作用^[62,63]，我们推测sAC对免疫系统可能具有一定的调节作用，后续我们还会开展相关的实验研究。

3 氧与二氧化碳慢性感知信号通路的交互作用

虽然以往研究已经对氧与二氧化碳的细胞感知机制进行比较深入的探索，但是两条感知通路之间是否存在交互调节目前仍然缺乏确切的机制证据。已有一些研究提示，二者之间存在联系，例如PKA与HIF-1 α 之间存在相互激活的关系。而PKA的活性可由pCO₂升高导致，提示这两条感知通路之间可能存在交互调节^[64,65]。此外，饲养于高二氧化碳环境的小鼠，HIF-1 α 被抑制，其血清中EPO水平显著降低^[2]；而将小鼠饲养于高氧和高二氧化碳的条件下三周，小鼠脑组织中HIF-1 α 、HIF-

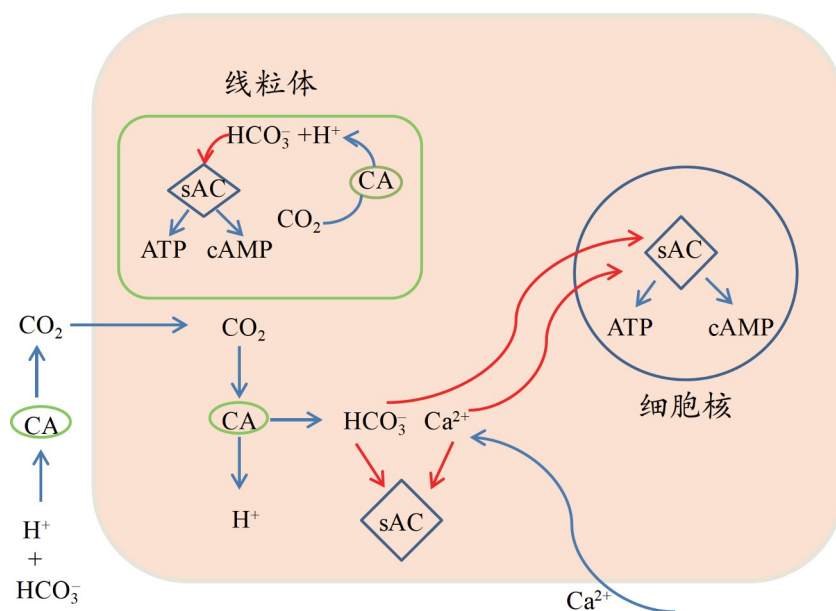


图4 sAC的调控机制

2 α 的累积及EPO的表达均显著增高^[66]。这些研究结果进一步说明, 二氧化碳感知通路可对氧感知通路造成影响, 但现象背后的分子机制仍有待阐明。

4 总结与展望

氧和高二氧化碳浓度异常是在呼吸系统疾病发生发展过程中经常出现的病理改变, 且这种异常往往与疾病预后相关^[67-72]。鉴于氧和二氧化碳慢性感知通路与免疫系统的密切关系, 我们认为二者浓度变化可能在肺部病毒感染病程中发挥重要作用, 深入阐明该问题有望为促进重症感染性肺炎的临床诊治研究提供新的依据和切入点。

参考文献

- [1] Cummins EP, Strowitzki MJ, Taylor CT. Mechanisms and consequences of oxygen and carbon dioxide sensing in mammals. *Physiol Rev*, 2020, 100(1): 463-488
- [2] Selfridge AC, Cavadas MAS, Scholz CC, et al. Hypercapnia suppresses the HIF-dependent adaptive response to hypoxia. *J Biol Chem*, 2016, 291(22): 11800-11808
- [3] De Vito E, Roncoroni A, Berizzo E, et al. Effects of spontaneous and hypercapnic hyperventilation on inspiratory effort sensation in normal subjects. *Am J Respir Crit Care Med*, 1998, 158(1): 107-110
- [4] Wilson DF, Roy A, Lahiri S. Immediate and long-term responses of the carotid body to high altitude. *High Altitude Med Biol*, 2005, 6(2): 97-111
- [5] Lévy P, Kohler M, McNicholas WT, et al. Obstructive sleep apnoea syndrome. *Nat Rev Dis Primers*, 2015, 1(1): 15015
- [6] Budweiser S, Jörres RA, Pfeifer M. Treatment of respiratory failure in COPD. *Int J Chron Obstruct Pulmon Dis*, 2008, 3(4): 605-618
- [7] Cummins EP, Taylor CT. Hypoxia-responsive transcription factors. *Pflugers Arch*, 2005, 450(6): 363-371
- [8] Cummins EP, Keogh CE. Respiratory gases and the regulation of transcription. *Exp Physiol*, 2016, 101(8): 986-1002
- [9] Ku JH, Park YH, Myung JK, et al. Expression of hypoxia inducible factor-1 α and 2 α in conventional renal cell carcinoma with or without sarcomatoid differentiation. *Urol Oncol-Semin Ori*, 2011, 29(6): 731-737
- [10] Ema M, Taya S, Yokotani N, et al. A novel bHLH-PAS factor with close sequence similarity to hypoxia-inducible factor 1 regulates the VEGF expression and is potentially involved in lung and vascular development. *Proc Natl Acad Sci USA*, 1997, 94(9): 4273-4278
- [11] Pasanen A, Heikkilä M, Rautavuoma K, et al. Hypoxia-inducible factor (HIF)-3 α is subject to extensive alternative splicing in human tissues and cancer cells and is regulated by HIF-1 but not HIF-2. *Int J Biochem Cell Biol*, 2010, 42(7): 1189-1200
- [12] Semenza GL, Wang GL. A nuclear factor induced by hypoxia via *de novo* protein synthesis binds to the human erythropoietin gene enhancer at a site required for transcriptional activation. *Mol Cell Biol*, 1992, 12(12): 5447-5454
- [13] McGettrick AF, O'Neill LAJ. The role of HIF in immunity and inflammation. *Cell Metab*, 2020, 32(4): 524-536
- [14] Thompson CB. Into thin air: how we sense and respond to hypoxia. *Cell*, 2016, 167(1): 9-11
- [15] Semenza GL. Hypoxia-inducible factor 1 (HIF-1) pathway. *Sci STKE*, 2007. doi: 10.1126/stke.4072007cm817925579
- [16] Ke Q, Costa M. Hypoxia-inducible factor-1 (HIF-1). *Mol Pharmacol*, 2006, 70(5): 1469-1480
- [17] Krenn K, Klepetko W, Taghavi S, et al. Vascular endothelial growth factor increases pulmonary vascular permeability in cystic fibrosis patients undergoing lung transplantation. *Eur J Cardio-Thoracic Surg*, 2007, 32(1): 35-41
- [18] Manalo DJ, Rowan A, Lavoie T, et al. Transcriptional regulation of vascular endothelial cell responses to hypoxia by HIF-1. *Blood*, 2005, 105(2): 659-669
- [19] Roman J, Rangasamy T, Guo J, et al. T-cell activation under hypoxic conditions enhances ifn- γ secretion. *Am J Respir Cell Mol Biol*, 2010, 42(1): 123-128
- [20] Emami Nejad A, Najafgholian S, Rostami A, et al. The role of hypoxia in the tumor microenvironment and development of cancer stem cell: a novel approach to developing treatment. *Cancer Cell Int*, 2021, 21(1): 62
- [21] Pham K, Parikh K, Heinrich EC. Hypoxia and inflammation: insights from high-altitude physiology. *Front Physiol*, 2021, 12: 676782
- [22] Taylor CT, Colgan SP. Regulation of immunity and inflammation by hypoxia in immunological niches. *Nat Rev Immunol*, 2017, 17(12): 774-785
- [23] Hammami A, Charpentier T, Smans M, et al. Irf-5-mediated inflammation limits cd8+ t cell expansion by inducing hif-1 α and impairing dendritic cell functions during leishmania infection. *PLoS Pathog*, 2015, 11(6): e1004938
- [24] Wobben R, Hüsecken Y, Lodewick C, et al. Role of hypoxia inducible factor-1 α for interferon synthesis in mouse dendritic cells. *Biol Chem*, 2013, 394(4): 495-505

- [25] Cho SH, Raybuck AL, Stengel K, et al. Germinal centre hypoxia and regulation of antibody qualities by a hypoxia response system. *Nature*, 2016, 537(7619): 234-238
- [26] Corcoran SE, O'Neill LAJ. HIF1 α and metabolic reprogramming in inflammation. *J Clin Invest*, 2016, 126(10): 3699-3707
- [27] Palazon A, Goldrath AW, Nizet V, et al. HIF transcription factors, inflammation, and immunity. *Immunity*, 2014, 41(4): 518-528
- [28] Jung YJ, Isaacs JS, Lee S, et al. IL-1 β mediated up-regulation of HIF-1 α via an NF κ B/COX-2 pathway identifies HIF-1 as a critical link between inflammation and oncogenesis. *FASEB J*, 2003, 17(14): 1-22
- [29] Santiago-Fernández C, Martín-Reyes F, Tome M, et al. Oxidized LDL increase the proinflammatory profile of human visceral adipocytes produced by hypoxia. *Biomedicines*, 2021, 9(11): 1715
- [30] Bok S, Kim YE, Woo Y, et al. Hypoxia-inducible factor-1 α regulates microglial functions affecting neuronal survival in the acute phase of ischemic stroke in mice. *Oncotarget*, 2017, 8(67): 111508-111521
- [31] Tolnay AE, Baskin CR, Tumpey TM, et al. Extrapulmonary tissue responses in cynomolgus macaques (*Macaca fascicularis*) infected with highly pathogenic avian influenza A (H5N1) virus. *Arch Virol*, 2010, 155(6): 905-914
- [32] Uematsu T, Fujita T, Nakaoka HJ, et al. Mint3/Apba3 depletion ameliorates severe murine influenza pneumonia and macrophage cytokine production in response to the influenza virus. *Sci Rep*, 2016, 6(1): 37815
- [33] Hara T, Mimura K, Abe T, et al. Deletion of the Mint3/Apba3 gene in mice abrogates macrophage functions and increases resistance to lipopolysaccharide-induced septic shock. *J Biol Chem*, 2011, 286(37): 32542-32551
- [34] Guo X, Zhu Z, Zhang W, et al. Nuclear translocation of HIF-1 α induced by influenza A (H1N1) infection is critical to the production of proinflammatory cytokines. *Emerging Microbes Infects*, 2017, 6(1): 1-8
- [35] Zhao C, Chen J, Cheng L, et al. Deficiency of HIF-1 α enhances influenza A virus replication by promoting autophagy in alveolar type II epithelial cells. *Emerging Microbes Infects*, 2020, 9(1): 691-706
- [36] Bhattacharya S, Agarwal S, Shrimali NM, et al. Interplay between hypoxia and inflammation contributes to the progression and severity of respiratory viral diseases. *Mol Aspects Med*, 2021, 81: 101000
- [37] Jantsch J, Chakravorty D, Turza N, et al. Hypoxia and hypoxia-inducible factor-1 α modulate lipopolysaccharide-induced dendritic cell activation and function. *J Immunol*, 2008, 180(7): 4697-4705
- [38] Köhler T, Reizis B, Johnson RS, et al. Influence of hypoxia-inducible factor 1 α on dendritic cell differentiation and migration. *Eur J Immunol*, 2012, 42(5): 1226-1236
- [39] Cummins EP, Selfridge AC, Sporn PH, et al. Carbon dioxide-sensing in organisms and its implications for human disease. *Cell Mol Life Sci*, 2014, 71(5): 831-845
- [40] Lu Z, Casalino-Matsuda SM, Nair A, et al. A role for heat shock factor 1 in hypercapnia-induced inhibition of inflammatory cytokine expression. *FASEB J*, 2018, 32(7): 3614-3622
- [41] Cummins EP, Oliver KM, Lenihan CR, et al. NF- κ B links CO₂ sensing to innate immunity and inflammation in mammalian cells. *J Immunol*, 2010, 185(7): 4439-4445
- [42] O'Toole D, Hassett P, Contreras M, et al. Hypercapnic acidosis attenuates pulmonary epithelial wound repair by an NF- κ B dependent mechanism. *Thorax*, 2009, 64(11): 976-982
- [43] Wiggins SV, Steegborn C, Levin LR, et al. Pharmacological modulation of the CO₂/HCO₃/pH-, calcium-, and ATP-sensing soluble adenylyl cyclase. *Pharmacol Ther*, 2018, 190: 173-186
- [44] Braun T, Dods RF. Development of a Mn-2+-sensitive, "soluble" adenylyl cyclase in rat testis. *Proc Natl Acad Sci USA*, 1975, 72(3): 1097-1101
- [45] Geng W, Wang Z, Zhang J, et al. Cloning and characterization of the human soluble adenylyl cyclase. *Am J Physiol Cell Physiol*, 2005, 288(6): C1305-C1316
- [46] Sinclair ML, Wang XY, Mattia M, et al. Specific expression of soluble adenylyl cyclase in male germ cells. *Mol Reprod Dev*, 2000, 56(1): 6-11
- [47] Zippin JH, Chen Y, Nahirney P, et al. Compartmentalization of bicarbonate-sensitive adenylyl cyclase in distinct signaling microdomains. *FASEB J*, 2003, 17(1): 82-84
- [48] Tresguerres M, Levin LR, Buck J. Intracellular cAMP signaling by soluble adenylyl cyclase. *Kidney Int*, 2011, 79(12): 1277-1288
- [49] Litvin TN, Kamenetsky M, Zarifyan A, et al. Kinetic properties of "soluble" adenylyl cyclase. *J Biol Chem*, 2003, 278(18): 15922-15926
- [50] Saalau-Bethell SM, Berdini V, Cleasby A, et al. Crystal structure of human soluble adenylyl cyclase reveals a distinct, highly flexible allosteric bicarbonate binding pocket. *ChemMedChem*, 2014, 9(4): 823-832
- [51] Abolhassani M, Guais A, Chaumet-Riffaud P, et al. Carbon dioxide inhalation causes pulmonary inflammation. *Am J Physiol Lung Cell Mol Physiol*, 2009, 296(4): L657-L665
- [52] Tang SE, Wu SY, Chu SJ, et al. Pre-Treatment with ten-minute carbon dioxide inhalation prevents lipopolysac-

- charide-induced lung injury in mice via down-regulation of toll-like receptor 4 expression. *Int J Mol Sci*, 2019, 20 (24): 6293
- [53] Schneberger D, Pandher U, Thompson B, et al. Effects of elevated CO₂ levels on lung immune response to organic dust and lipopolysaccharide. *Respir Res*, 2021, 22(1): 104
- [54] Ohlraun S, Wollersheim T, Weiß C, et al. Carbon Dioxide for the treatment of febrile seizures: rationale, feasibility, and design of the CARDIF-study. *J Transl Med*, 2013, 11 (1): 157
- [55] Szollosi I, Jones M, Morrell MJ, et al. Effect of CO₂ inhalation on central sleep apnea and arousals from sleep. *Respiration*, 2004, 71(5): 493-498
- [56] Tolner EA, Hochman DW, Hassinen P, et al. Five percent CO₂ is a potent, fast-acting inhalation anticonvulsant. *Epilepsia*, 2011, 52(1): 104-114
- [57] Galganska H, Jarmuszkiwicz W, Galganski L. Carbon dioxide inhibits COVID-19-type proinflammatory responses through extracellular signal-regulated kinases 1 and 2, novel carbon dioxide sensors. *Cell Mol Life Sci*, 2021, 78(24): 8229-8242
- [58] Flacke JP, Flacke H, Appukuttan A, et al. Type 10 soluble adenylyl cyclase is overexpressed in prostate carcinoma and controls proliferation of prostate cancer cells. *J Biol Chem*, 2013, 288(5): 3126-3135
- [59] Kammer GM. The adenylyl cyclase-cAMP-protein kinase A pathway and regulation of the immune response. *Immunol Today*, 1988, 9(7-8): 222-229
- [60] Chen Y, Cann MJ, Litvin TN, et al. Soluble adenylyl cyclase as an evolutionarily conserved bicarbonate sensor. *Science*, 2000, 289(5479): 625-628
- [61] Sun XC, Cui M, Bonanno JA. [HCO₃⁻]-regulated expression and activity of soluble adenylyl cyclase in corneal endothelial and Calu-3 cells. *BMC Physiol*, 2004, 4(1): 8
- [62] Raker VK, Becker C, Steinbrink K. The cAMP pathway as therapeutic target in autoimmune and inflammatory diseases. *Front Immunol*, 2016, 7: 123
- [63] Shu J, Zhang F, Zhang L, et al. G protein coupled receptors signaling pathways implicate in inflammatory and immune response of rheumatoid arthritis. *Inflamm Res*, 2017, 66(5): 379-387
- [64] Bullen JW, Tchernyshyov I, Holewinski RJ, et al. Protein kinase A-dependent phosphorylation stimulates the transcriptional activity of hypoxia-inducible factor 1. *Sci Signal*, 2016, 9(430): ra56
- [65] Lucia K, Wu Y, Garcia JM, et al. Hypoxia and the hypoxia inducible factor 1 α activate protein kinase A by repressing RII beta subunit transcription. *Oncogene*, 2020, 39(16): 3367-3380
- [66] Benderro GF, Sun X, Kuang Y, et al. Decreased VEGF expression and microvascular density, but increased HIF-1 and 2 α accumulation and EPO expression in chronic moderate hyperoxia in the mouse brain. *Brain Res*, 2012, 1471: 46-55
- [67] Zheng G, Wang Y, Wang X. Chronic hypoxia-hypercapnia influences cognitive function: a possible new model of cognitive dysfunction in chronic obstructive pulmonary disease. *Med Hypotheses*, 2008, 71(1): 111-113
- [68] West JB. Causes of and compensations for hypoxemia and hypercapnia. *Compr Physiol*, 2011, 1(3): 1541-1553.
- [69] López-Campos JL, Tan W, Soriano JB. Global burden of copd. *Respirology*, 2016, 21(1): 14-23
- [70] Nin N, Muriel A, Peñuelas O, et al. Severe hypercapnia and outcome of mechanically ventilated patients with moderate or severe acute respiratory distress syndrome. *Intensive Care Med*, 2017, 43(2): 200-208
- [71] Tiruvoipati R, Pilcher D, Buscher H, et al. Effects of hypercapnia and hypercapnic acidosis on hospital mortality in mechanically ventilated patients. *Crit Care Med*, 2017, 45(7): e649-e656
- [72] Nin N, Angulo M, Briva A. Effects of hypercapnia in acute respiratory distress syndrome. *Ann Transl Med*, 2018, 6(2): 37