

## Morphology of the first zoeal stages of eleven Sesarmidae (Crustacea, Brachyura, Thoracotremata) from the Indo-West Pacific, with a summary of familial larval characters

JOSÉ A. CUESTA<sup>1\*</sup>, GUILLERMO GUERAO<sup>2</sup>, HUNG-CHANG LIU<sup>3</sup> and CHRISTOPH D. SCHUBART<sup>4</sup>

<sup>1</sup>Instituto de Ciencias Marinas de Andalucía, CSIC, Avenida República Saharaui, 2, 11510 Puerto Real, Cádiz, Spain  
Tel. +34 (956) 83612; Fax: +34 (956) 834701; email: jose.cuesta@icman.csic.es

<sup>2</sup>Departament de Biología Animal (Artròpodes), Facultat de Biología (U.B.), Av. Diagonal, 645, 08028 Barcelona, Spain

<sup>3</sup>Research Center for Biodiversity, Academia Sinica, Nankang, Taipei 115, Taiwan

<sup>4</sup>Biologie I, Universität Regensburg, D-93040 Regensburg, Germany

Received 29 September 2005; Accepted 22 February 2006

### Summary

The first zoeal stages of eleven species of Sesarmidae from the Indo-West Pacific were obtained from ovigerous females. Those of *Labuanium scandens*, *L. rotundatum*, *L. trapezoideum*, *L. politum*, *Metasesarma Aubryi*, *Pseudosesarma crassimanum*, *Stelgistra stormi*, and *Sesarmops impressum*, are described for the first time, while the first zoeal stages of *Clistocoeloma menguiense*, *Metasesarma obesum* and *Sesarmops intermedium* are re-described. Larval characters of all these species are compared with previously described ones for the family and morphological features are re-evaluated. Minute spines on the telson of the zoeae are described as a new larval character in Sesarmidae and their presence or absence in other grapsoid groups is discussed. The results demonstrate that a recurrent combination of reliable larval characters distinguishes zoeae and megalopae of the examined sesarmids from the rest of the Grapoidea. This appears to be consistent with recent studies that redefine the Sesarmidae.

**Keywords:** Sesarmidae, Thoracotremata, larval morphology, zoea, megalopa, taxonomy, Indo-West Pacific

### Introduction

The larval descriptions of 54 species of Sesarmidae are known from 53 publications. About 230 species are assigned to this grapsoid family and these are mostly intertidal to semi-terrestrial, which allows relatively easy collecting. Furthermore, sesarmid larvae can be effortlessly reared in laboratory due to the large size of early zoeal stages. In contrast, Grapsidae have small first stage zoeae that do not feed on *Artemia nauplii* and this

makes their rearing difficult. Consequently the complete larval development is only known for *Metopograpsus*.

Larval morphology has become an increasingly useful tool for grapsoid systematics. For example, Wear (1970), Green and Anderson (1973), Terada (1976), Fielder and Greenwood (1983), and Krishnan and Kannupandi (1989) suggested that the assignment of *Chasmagnathus*, *Cyclograpsus*, *Helice*, *Helograpsus*, *Metaplagus* and *Paragrapsus* to the then Sesarminae was

\*Corresponding author.

Presented at the Fourth Crustacean Larval Conference, Glasgow, Scotland, UK, July 18–22, 2005.

anomalous. However no formal changes to the classification were proposed. Recently, all grapsoid families, including the Sesarmidae, were redefined using molecular and adult data as well as larval morphology (Schubart et al., 2000; Cuesta et al., 2001; Schubart et al., 2002) and the above six genera have been excluded from among the taxa now assigned to the Sesarmidae.

The first zoeal stages of eight sesarmid species from the Indo-West Pacific including *Labuanium scandens* Ng and Liu, 2003; *L. rotundatum* (Hess, 1865); *L. trapezoideum* (H. Milne Edwards, 1837); *L. politum* (de Man, 1888); *Metasesarma aubryi* A. Milne-Edwards, 1869; *Pseudosesarma crassimanum* (de Man, 1887); *Stelgistra stormi* (de Man, 1895); and *Sesarmops impressum* (H. Milne-Edwards, 1837) are described and illustrated for the first time. Previous accounts for the first zoeae of *Clistocoeloma merguiense* de Man, 1888; *Metasesarma obesum* (Dana, 1851) and *Sesarmops intermedium* (de Haan, 1835) have been reported, but they are here re-described to include new characters previously overlooked.

The aim of this study is to review the known sesarmid larval descriptions and incorporate the above new data in order to summarise the distinctive characters that now define this family.

## Materials and Methods

The first zoeal stages of *Labuanium scandens*, *L. trapezoideum*, *L. rotundatum*, *L. politum*, *Metasesarma aubryi*, *M. obesum* (previously known as *M. rousseauxii*, see Ng and Schubart, 2003), *Pseudosesarma crassimanum*, *Stelgistra stormi*, *Clistocoeloma merguiense*, *Sesarmops impressum*, and *Sesarmops intermedium* were obtained from ovigerous crabs collected from the Indo-West Pacific (for details see Table 1).

All zoeae with active natatory behaviour were fixed in 4% formaldehyde-seawater. Dissections were carried out under a Wild MZ6 stereo microscope and drawings and measurements were made using a Zeiss Axioskop compound microscope equipped with a *camera lucida*. All measurements were made with a calibrated ocular micrometer. Drawings were based on 5 larvae and morphometrics on 10 larvae per stage. Measurements were made of the following: rostro-dorsal length (rdl) from the tip of the rostral spine to the tip of the dorsal spine; carapace length (cl) from the base of the rostrum to the posterior margin; exopod length (el) from the base of the antennal exopod to the distal margin (without setae); protopodal process length (pl) from the base of the antennal exopod to the tip of the protopodal process; furcal length (fl) from an imaginary line across the base

Table 1. List of the first zoeal stages described in the present study, indicating localities and date of collection, collectors, and accession numbers. Zoological Reference Collection, National University of Singapore, Singapore: ZRC; Colecciones Biológicas de Referencia, Instituto de Ciencias del Mar de Barcelona, Spain: ICM; Institute of Zoology, Academia Sinica, Nankang, Taiwan: ASIZ

Species	Locality	Date	Collector	Accession number
<i>Labuanium scandens</i>	Pingtung, Taiwan	10 August 1998	H-C Liu	ZRC 2005.0127
<i>Labuanium trapezoideum</i>	Taitung, Taiwan	14 November 2001	H-C Liu	ZRC 2005.0128
<i>Labuanium rotundatum</i>	Pago Bay, Guam	4 August 2001	H-C Liu and P.K.L. Ng	ZRC 2005.0129
<i>Labuanium politum</i>	Loboc River, Philippines	5 March 2003	H-C Liu and P.K.L. Ng	ASIZ 72932
<i>Metasesarma aubryi</i>	Pingtung, Taiwan	18 October 1998	H-C Liu	ZRC 2005.0130 / ICMD 2/2006
<i>Metasesarma obesum</i>	Malotoy, Sulawesi, Indonesia	17 January 2000	Schubart et al.	ZRC 2005.0131 / ICMD 10/2006
<i>Metasesarma obesum</i>	Pingtung, Taiwan	26 August 1999	H-C Liu	ZRC 2005.0132 / ICMD 11/2006
<i>Stelgistra stormi</i>	Hengchun, Taiwan	12 July 1999	H-C Liu	ZRC 2005.0133 / ICMD 7/2006
<i>Pseudosesarma crassimanum</i>	Sungei Benut, Malaysia	30 September 1999	Schubart et al.	ZRC 2005.0134 / ICMD 3/2006
<i>Clistocoeloma merguiense</i>	Mandai, Singapore	10 September 1999	Schubart and Sivasothi	ZRC 2005.0135 / ICMD 6/2006
<i>Sesarmops impressum</i>	Pingtung, Taiwan	22 September 1998	H-C Liu and C-H Wang	ZRC 2005.0136 / ICMD 8/2006
<i>Sesarmops intermedium</i>	Harbor Bay, Taiwan	3 September 1999	H-C Liu	ZRC 2005.0137 / ICMD 9/2006

of the outer seta on the posterior margin of the telson to the furcal tip; and basal telson length (bt), from a line across the anterior margin to the posterior margin of the telson (base of the outer seta). Plumose natatory setae of the maxillipeds' exopods are drawn truncated in Fig. 3. Descriptions and figures are arranged according to the standard proposed by Clark et al. (1998).

The morphology and setation of the mouthparts of all described sesarmid species is similar and does not show any variation between species. Therefore, these appendages are described and illustrated only for *Labuanium scandens*.

Maternal females and samples of the first zoeal stage of all species were deposited at the Zoological Reference Collection (ZRC), National University of Singapore, Singapore, while another set of the first zoeae was deposited at the Colecciones Biológicas de Referencia, Instituto de Ciencias del Mar de Barcelona (ICM), Spain (for catalog numbers see Table 1).

## Results — Zoea I descriptions

### *Labuanium scandens* Ng and Liu, 2003 (Figs. 1A–3B)

Dimensions: rdl:  $0.77 \pm 0.014$  mm; cl:  $0.52 \pm 0.01$  mm.

Carapace (Fig. 1A). Globose, smooth and without tubercles; dorsal spine present, short and curved; rostral spine present, straight and similar in length to dorsal spine; lateral spines absent; pair of setae on anterodorsal and posterodorsal regions; posterior and ventral margin without setae; eyes sessile.

Antennule (Fig. 1B). Uniramous; endopod absent; exopod unsegmented with 4 terminal aesthetascs (3 long, 1 shorter and thin) and 1 terminal seta.

Antenna (Fig. 1C). Well developed protopod almost reaching the tip of the rostral spine and bearing two unequal rows of 11–12, and 15–16 spines of different size, respectively; endopod absent; exopod elongated,

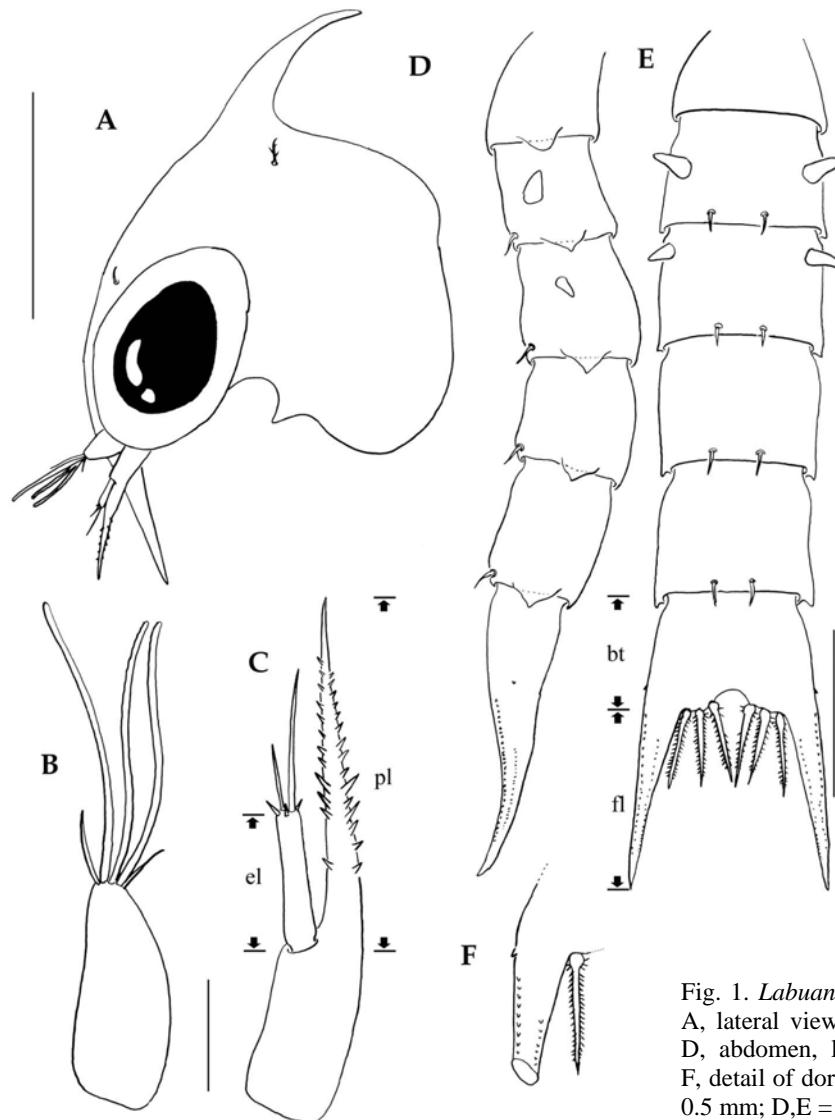


Fig. 1. *Labuanium scandens* Ng and Liu, 2003, zoea I. A, lateral view of carapace; B, antennule; C, antenna; D, abdomen, lateral view; E, abdomen, dorsal view; F, detail of dorsal area of furcal arm. Scale bars, A–C = 0.5 mm; D,E = 0.1 mm.

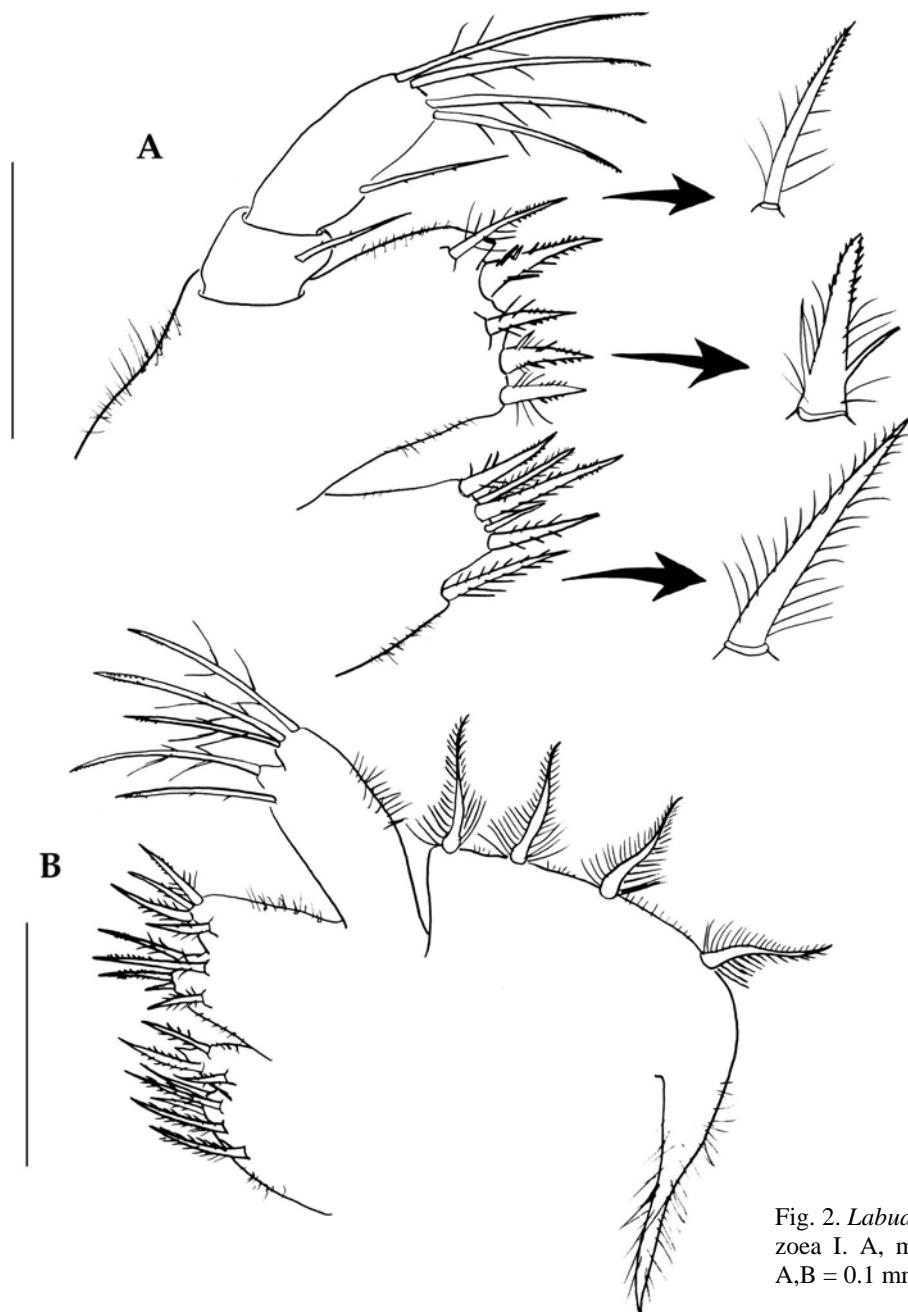


Fig. 2. *Labuanium scandens* Ng and Liu, 2003, zoea I. A, maxillule; B, maxilla. Scale bars: A,B = 0.1 mm.

more than 2/3 of the protopod length, with 2 terminal setae (1 long but not reaching the tip of protopod, 1 shorter) and 3 small terminal spines; pl/el = 2.2–2.3.

Mandible. Palp absent; molar and incisor processes well developed.

Maxillule (Fig. 2A). Coxal endite with 6 plumose setae; basial endite with 5 setae (2 cuspidate and 3 plumodenticulate) and 2 teeth (setal buds); endopod 2-segmented with 1 simple seta on the proximal segment and 5 (1 subterminal and 4 terminal) plumodenticulate setae on the distal segment; exopod seta absent; episod seta absent.

Maxilla (Fig. 2B). Coxal endite bilobed with 5+3 (plus a marginal spine) plumodenticulate setae; basial endite bilobed with 5+4 plumodenticulate setae; endopod unsegmented, bilobed with 2+3 long plumodenticulate setae on the inner and outer lobe respectively; scaphognathite (exopod) with 4 plumose marginal setae and a long setose posterior process.

First Maxilliped (Fig. 3A). Coxa with 1 sparsely plumose seta; basis with 10 inner setae arranged 2 sparsely plumose + 2 sparsely plumose + 3 simple + 3 simple, and a mat of long dorsobasal microtrichiae on the outer side; endopod 5-segmented with 2 (1 simple, 1



Fig. 3. *Labuanium scandens* Ng and Liu, 2003, zoea I. A, first maxilliped; B, second maxilliped. Scale bar: A,B = 0.1 mm.

plumodenticulate), 2 (1 simple, 1 plumodenticulate), 1 plumodenticulate, 2 plumodenticulate, and 5 (1 plumose subterminal + 4 plumodenticulate terminal) setae; exopod 2-segmented, distal segment with 4 long terminal plumose natatory setae.

**Second Maxilliped** (Fig. 3B). Coxa without setae; basis with 4 medial setae arranged 1 plumodenticulate + 1 simple + 1 simple + 1 simple; endopod 3-segmented with 0, 1 denticulate, 6 [3 (1 denticulate, 1 sparsely plumose, 1 simple) subterminal + 3 (2 sparsely plumose, 2 simple) terminal] setae; exopod 2-segmented, distal segment with 4 long terminal plumose natatory setae.

**Third Maxilliped.** Absent.

**Pereiopods.** Absent.

**Abdomen** (Fig. 1D, E). Five abdominal somites; somites 2 and 3 with a pair of dorsolateral processes; somites 3-5 with small posterolateral processes of subtriangular shape; somites 2-5 with a pair of postero-dorsal simple setae; pleopods absent.

**Telson** (Fig. 1F). Bifurcated with 3 pairs of serrulate setae on posterior margin; mid-internal side of inner pair without spines; dorsal part of each furcal arm with two rows of spines and outer part with shorter row of minute spines and a minute scale-like spine; fl/bt = 1.5–1.6.

#### *Labuanium rotundatum* (Hess, 1865) (Figs. 4A-E)

Dimensions: rdl:  $0.80 \pm 0.02$  mm; cl:  $0.46 \pm 0.03$  mm.

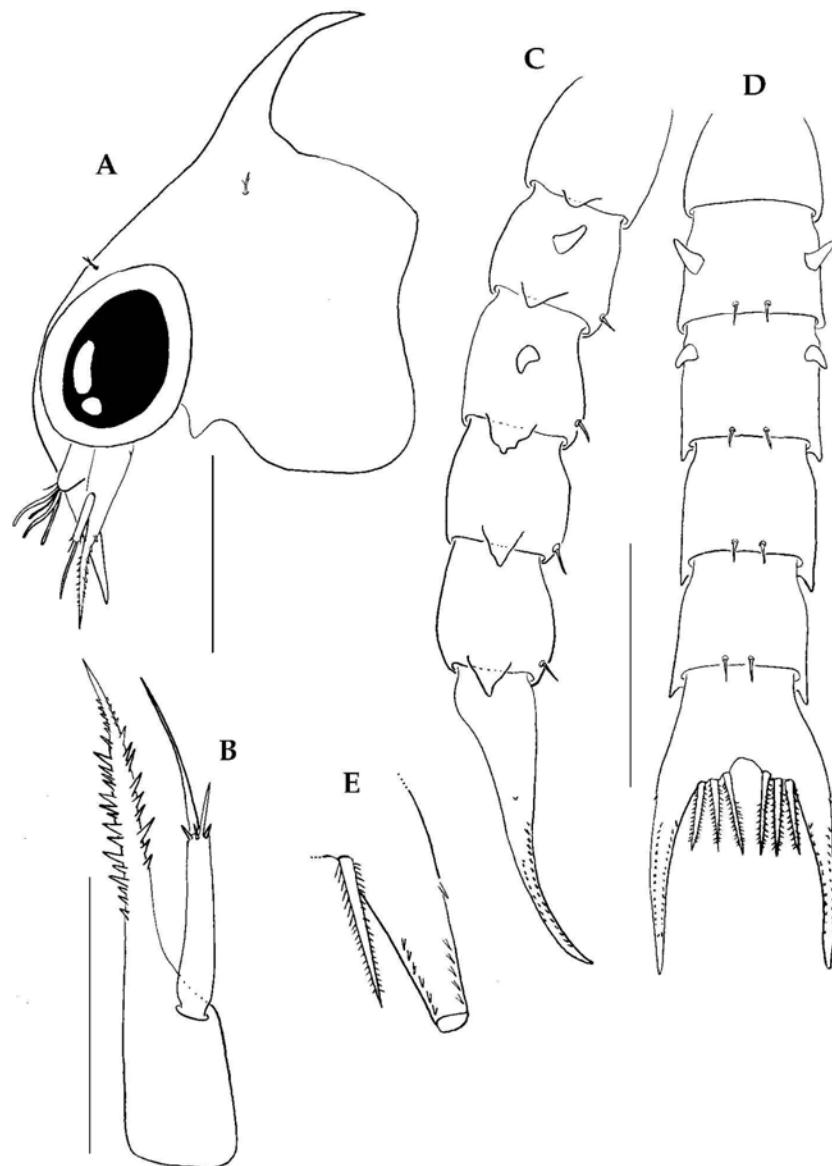


Fig. 4. *Labuanium rotundatum* (Hess, 1865), zoea I. A, lateral view of carapace; B, antenna; C, abdomen, lateral view; D, abdomen, dorsal view; E, detail of dorsal area of furcal arm. Scale bars: A,C,D = 0.25 mm; B = 0.125 mm.

Carapace (Fig. 4A). Smooth and without tubercles; dorsal spine present, short and curved; rostral spine present, straight and similar in length to dorsal spine.

Antenna (Fig. 4B). Protopod longer than rostral spine and bearing two unequal rows of 13–14 and 20–21 spines of different size, respectively; exopod elongated, more than 2/3 of the protopod length, with 2 terminal setae (1 long, 1 shorter) and 3 small terminal spines; pl/el = 2.0–2.1.

Abdomen (Fig. 4C,D). Somites 2 and 3 with a pair of dorsolateral processes; somites 3–5 with small postero-lateral processes of subtriangular shape, more developed than those of *L. scandens*; somites 2–5 with a pair of posterodorsal simple setae.

Telson (Fig. 4E). Mid-internal side of inner pair without spines; dorsal part of each furcal arm with two

rows of spines and outer part with a shorter row of minute spines and a minute scale-like spine; fl/bt = 1.6–1.7.

#### *Labuanium trapezoideum* (H. Milne Edwards, 1837) (Figs. 5A–E)

Dimensions: rdl:  $0.81 \pm 0.03$  mm; cl:  $0.49 \pm 0.02$  mm.

Carapace (Fig. 5A). Smooth and without tubercles; dorsal spine present, short and strongly curved; rostral spine present, straight and similar in length to dorsal spine.

Antenna (Fig. 5B). Protopod slightly longer than rostral spine and bearing two unequal rows of 11 and 14 spines of different size, respectively; exopod elongated, more than 2/3 of the protopod length, with 2 terminal

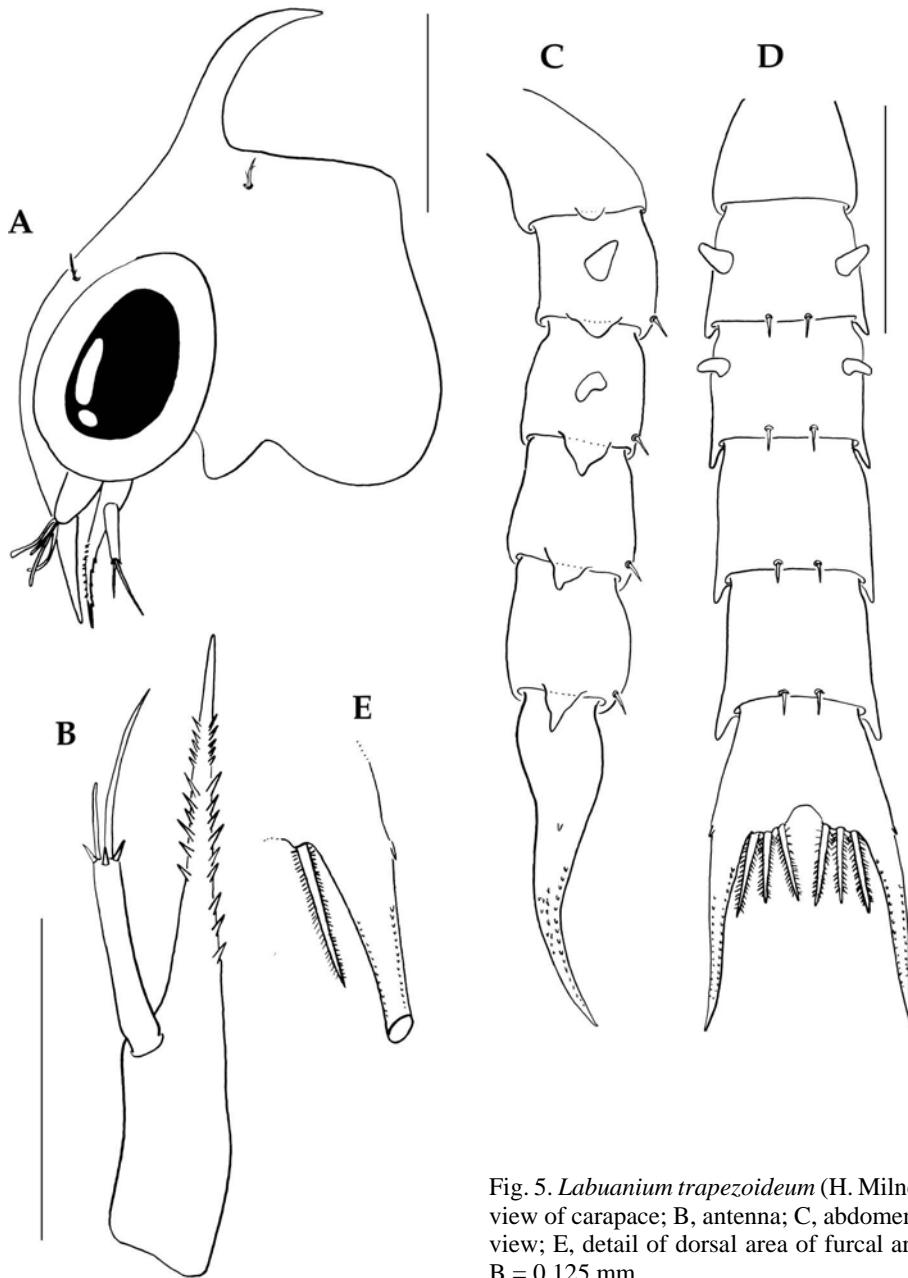


Fig. 5. *Labuanium trapezoideum* (H. Milne Edwards, 1837), zoea I. A, lateral view of carapace; B, antenna; C, abdomen, lateral view; D, abdomen, dorsal view; E, detail of dorsal area of furcal arm. Scale bars, A,C,D = 0.25 mm; B = 0.125 mm.

setae (1 long almost reaching the tip of protopod, 1 shorter) and 3 small terminal spines; pl/el = 2.0–2.1.

Abdomen (Fig. 5C, D). Somites 2 and 3 with pair of dorsolateral processes; somites 3–5 with small postero-lateral processes of subtriangular shape, more developed than those of *L. scandens*; somites 2–5 with a pair of posterodorsal simple setae.

Telson (Fig. 5E). Mid-internal side of inner pair sparsely spinulate; dorsal part of each furcal arm with two rows of spines and outer part with a shorter row of minute spines and a minute scale-like spine; fl/bt = 1.4–1.5.

#### *Labuanium politum* (de Man, 1888) (Figs. 6A–E)

Dimensions: rdl:  $0.85 \pm 0.03$  mm; cl:  $0.49 \pm 0.04$  mm.

Carapace (Fig. 6A). Smooth and without tubercles; dorsal spine present, straight; rostral spine present, straight and similar in length to dorsal spine.

Antenna (Fig. 6B). Protopod almost reaching to the tip of the rostral spine and bearing two unequal rows of 6–7, and 12–13 spines of different size, respectively; exopod elongated, more than 2/3 of the protopod length, with 2 terminal setae (1 long, 1 shorter) and 3 small terminal spines; pl/el = 2.05–2.10.

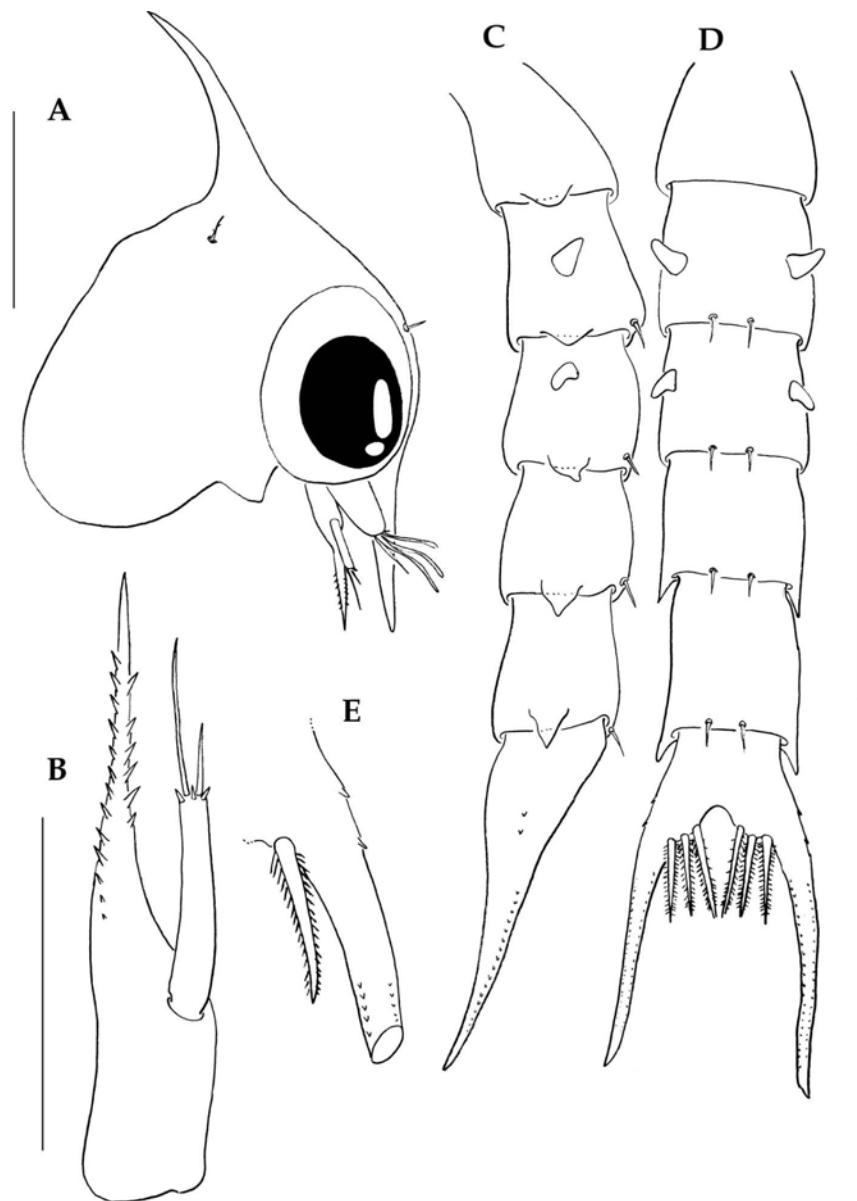


Fig. 6. *Labuanium politum* (de Man, 1888), zoea I. A, lateral view of carapace; B, antenna; C, abdomen, lateral view; D, abdomen, dorsal view; E, detail of dorsal area of furcal arm. Scale bars: A,C,D = 0.25 mm; B = 0.125 mm.

Abdomen (Fig. 6C, D). Somites 2 and 3 with pair of dorsolateral processes; somites 3–5 with small postero-lateral processes with subtriangular shape; somites 2–5 with a pair of posterodorsal simple setae.

Telson (Fig. 6E). Mid-internal side of inner pair sparsely spinulate; dorsal part of each furcal arm with two rows of minute spines and outer part with two minute scale-like spine; fl/bt = 2.50–2.65.

***Metasesarma aubryi* A. Milne Edwards, 1869  
(Figs. 7A–E)**

Dimensions: rdl:  $0.87 \pm 0.04$  mm; cl:  $0.60 \pm 0.01$  mm.

Carapace (Fig. 7A). Smooth and without tubercles; dorsal spine present, short and curved; rostral spine present, straight and similar in length to dorsal spine.

Antenna (Fig. 7B). Protopod almost reaching to the tip of the rostral spine and bearing two unequal rows of 15–16, and 24–26 spines of different size, respectively; exopod elongated, more than 2/3 of the protopod length, with 2 terminal setae (1 long, 1 shorter) and 3 terminal spines; pl/el = 2.1–2.2.

Abdomen (Fig. 7C, D). Somites 2 and 3 with a pair of dorsolateral processes; somites 3–5 with small posterolateral processes of subtriangular shape; somites 2–5 with a pair of posterodorsal simple setae.

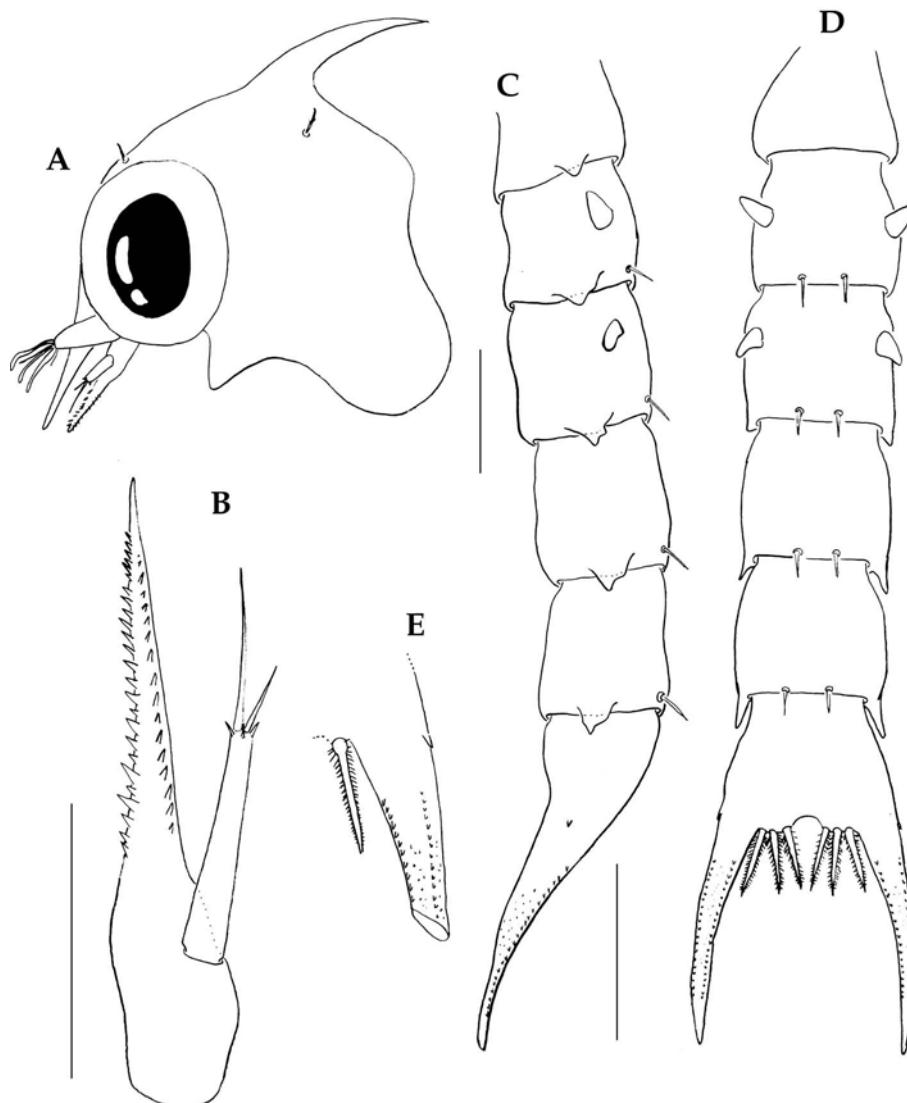


Fig. 7. *Metasesarma aubryi* A. Milne-Edwards, 1869, zoea I. A, lateral view of carapace; B, antenna; C, abdomen, lateral view; D, abdomen, dorsal view; E, detail of dorsal area of furcal arm. Scale bars: A,C,D = 0.2 mm; B = 0.1 mm.

Telson (Fig. 7E). Mid-internal side of inner pair without spines; dorsal part of each furcal arm with two rows of spines and outer part covered with minute spinules and a minute scale-like spine; fl/bt = 2.0–2.1.

#### *Metasesarma obesum* (Dana, 1851) (Figs. 8A–E)

Rajabai (1961: 160–162, Fig. III, 1–12), prezoea, zoea I (as *M. rousseauxii*).

Dimensions: rdl:  $0.69 \pm 0.02$  mm; cl:  $0.44 \pm 0.02$  mm (Taiwan).

Dimensions: rdl:  $0.63 \pm 0.02$  mm; cl:  $0.42 \pm 0.02$  mm (Sulawesi).

Carapace (Fig. 8A). Smooth and without tubercles; dorsal spine present, strongly curved (almost  $90^\circ$  angle); rostral spine present, straight shorter than dorsal spine.

Antenna (Fig. 8B). Protopod almost reaching to the tip of the rostral spine and bearing two unequal rows of 10–12, and 13–16 spines of different size, respectively; exopod elongated, more than 2/3 of the protopod length, with 2 terminal setae (1 long almost reaching to the tip of protopod, 1 shorter) and 2 terminal spines; pl/el = 2.10–2.26.

Abdomen (Fig. 8C, D). Somites 2 and 3 with pair of dorsolateral processes; somites 3–5 with small postero-lateral processes of subtriangular shape; somites 2–5 with a pair of posterodorsal simple setae.

Telson (Fig. 8E). Mid-internal side of inner pair sparsely spinulate; dorsal part of each furcal arm with two rows of spines and covered with spinules, and outer part with 2 minute scale-like spines; fl/bt = 1.3–1.5.

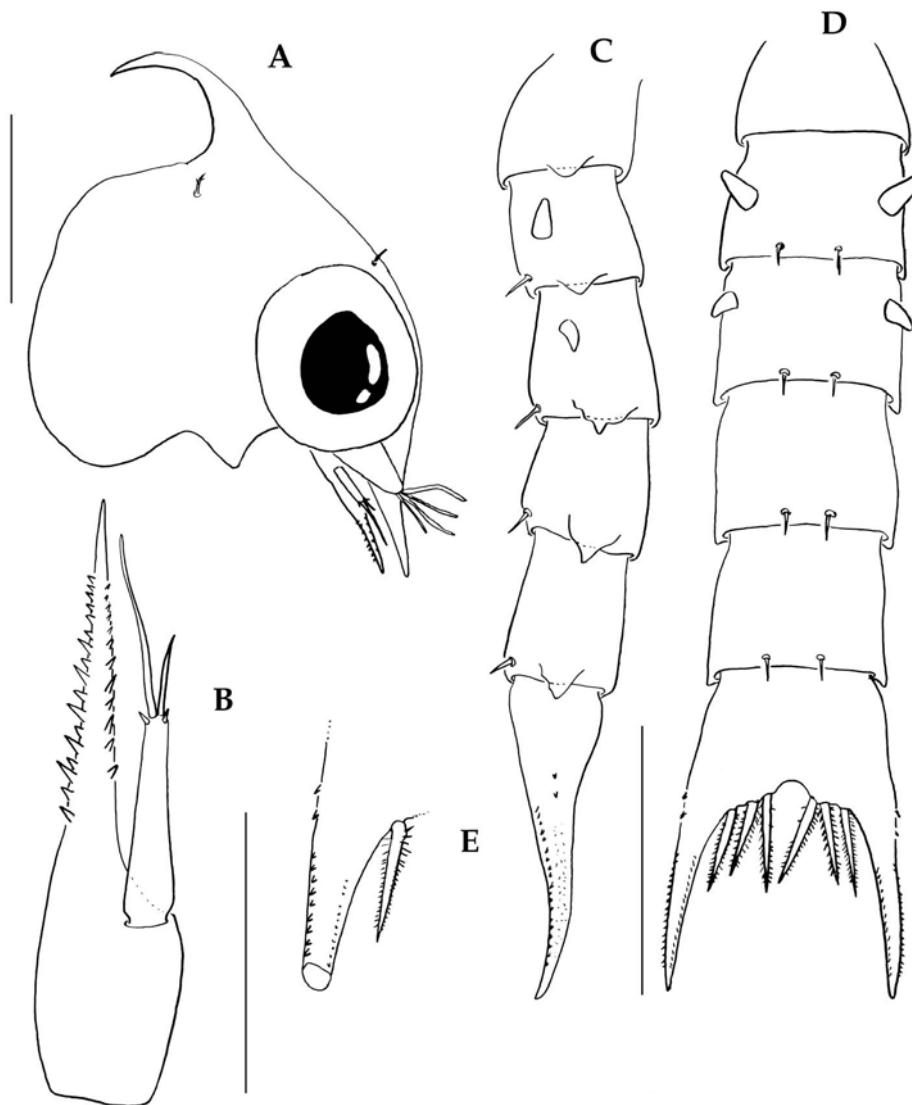


Fig. 8. *Metasesarma obesum* (Dana, 1851), zoea I. A, lateral view of carapace; B, antenna; C, abdomen, lateral view; D, abdomen, dorsal view; E, detail of dorsal area of furcal arm. Scale bars: A = 0.2 mm; C,D = 0.25 mm; B = 0.1 mm.

***Pseudosesarma crassimanum* (de Man, 1887) (Figs. 9A–E)**

Dimensions: rdl:  $0.72 \pm 0.03$  mm; cl:  $0.43 \pm 0.02$  mm.

Carapace (Fig. 9A). Smooth and without tubercles; dorsal spine present, almost strait; rostral spine present, straight and similar in length to dorsal spine.

Antenna (Fig. 9B). Protopod almost reaching to the tip of the rostral spine and bearing two unequal rows of 4–5, and 6 spines of different size, respectively; exopod elongated, more than 2/3 of the protopod length, with 2 terminal setae (1 long, 1 shorter) and 3 small terminal spines; pl/el = 2.18–2.42.

Abdomen (Fig. 9C, D). Somites 2 and 3 with pair of dorsolateral processes; somites 3–4 with posterolateral processes of subtriangular shape, and posterolateral

processes on somite 5 long and acute distally; somites 2–5 with a pair of posterodorsal simple setae.

Telson (Fig. 9E). Mid-internal side of inner pair sparsely spinulate; dorsal part of each furcal arm with two rows of spines and outer part without spines, only scattered minute spinules; fl/bt = 1.7–1.8.

***Stelgistra stormi* (de Man, 1895) (Figs. 10A–E)**

Dimensions: rdl:  $0.89 \pm 0.01$  mm; cl:  $0.63 \pm 0.03$  mm.

Carapace (Fig. 10A). Smooth and without tubercles; dorsal spine present, short and curved; rostral spine present, straight and longer than dorsal spine.

Antenna (Fig. 10B). Protopod almost reaching to the tip of the rostral spine and bearing two unequal rows of 13–14, and 23–24 spines of different size, respectively;

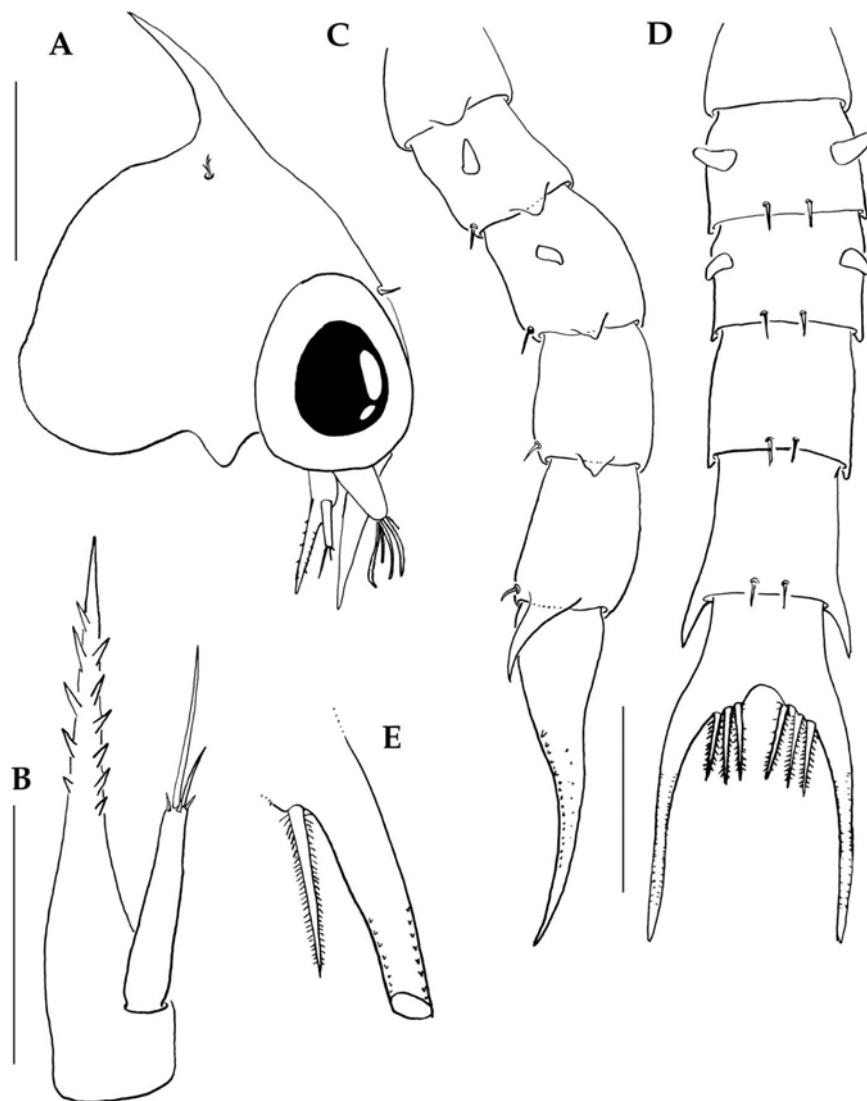


Fig. 9. *Pseudosesarma crassimanum* (de Man, 1887), zoea I. A, lateral view of carapace; B, antenna; C, abdomen, lateral view; D, abdomen, dorsal view; E, detail of dorsal area of furcal arm. Scale bars: A,C,D = 0.2 mm; B = 0.125 mm.

exopod elongated, more than 2/3 of the protopod length, with 2 terminal setae (1 long, 1 shorter) and 3 small terminal spines; pl/el = 2.0–2.2.

Abdomen (Fig. 10C, D). Somites 2 and 3 with pair of dorsolateral processes; somites 3–5 with small rounded posterolateral processes; somites 2–5 with a pair of posterodorsal simple setae.

Telson (Fig. 10E). Mid-internal side of inner pair sparsely spinulate; dorsal part of each furcal arm with two rows of minute spines and outer part without spines; fl/bt = 1.2–1.3.

#### *Clistocoeloma merguiense de Man, 1888 (Figs. 11A-E)*

Saba (1972: 25, Figs. 1, 2a, 3a, 4a).

Dimensions: rdl:  $0.61 \pm 0.01$  mm; cl:  $0.39 \pm 0.04$  mm.

Carapace (Fig. 11A). Smooth and without tubercles; dorsal spine present, short and curved; rostral spine present, straight and shorter in length to dorsal spine.

Antenna (Fig. 11B). Protopod longer than the rostral spine and bearing two unequal rows of 6, and 11–12 spines of different size, respectively; exopod elongated, more than 2/3 of the protopod length, with 2 terminal setae (1 long almost reaching to the tip of protopod, 1 shorter) and 3 small terminal spines; pl/el = 2.0–2.1.

Abdomen (Fig. 11C, D). Somites 2 and 3 with pair of dorsolateral processes; somites 3–5 with small posterolateral processes of subtriangular shape; somites 2–5 with a pair of posterodorsal simple setae; pleopods absent.

Telson (Fig. 11E). Mid-internal side of inner pair

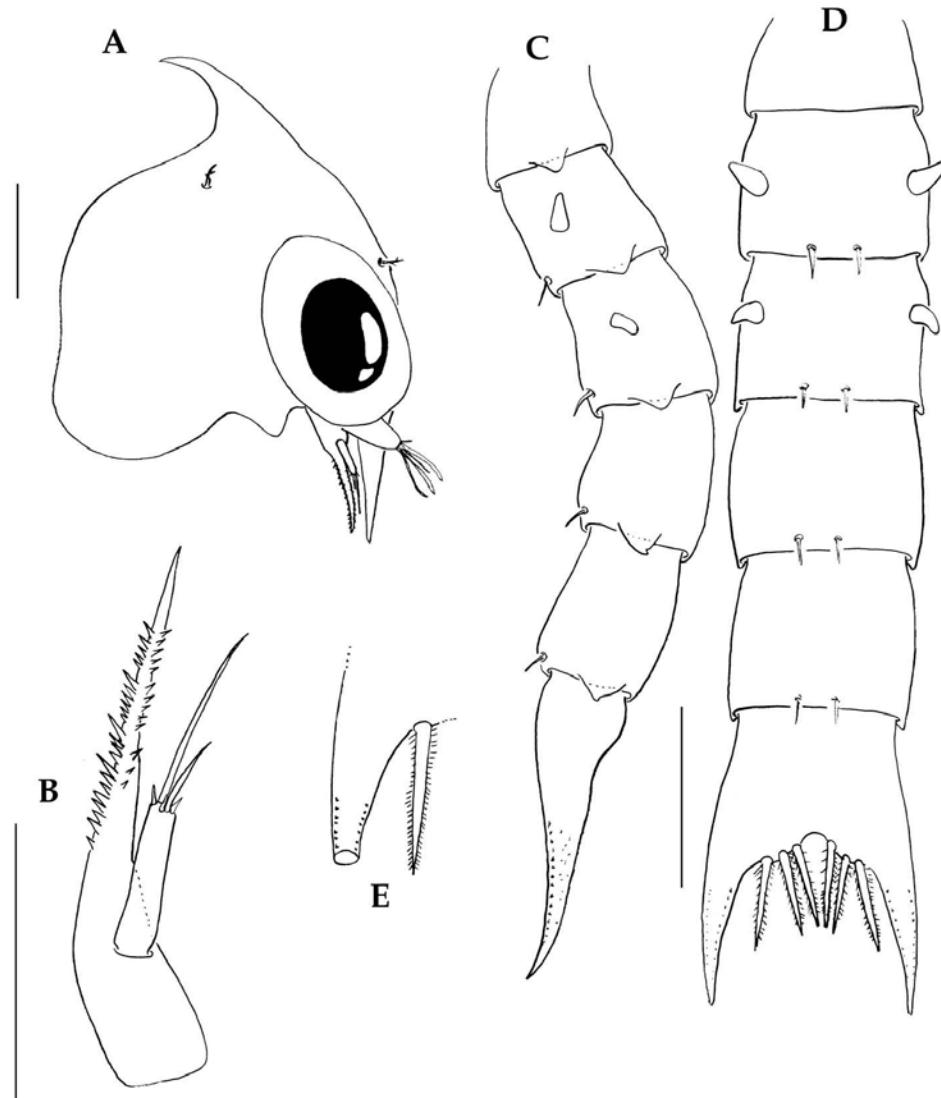


Fig. 10. *Stelgistra stormi* (de Man, 1895), zoea I. A, lateral view of carapace; B, antenna; C, abdomen, lateral view; D, abdomen, dorsal view; E, detail of dorsal area of furcal arm. Scale bars: A,C,D = 0.2 mm, B = 0.125 mm.

without spines; dorsal part of each furcal arm with two rows of spines and outer part with a shorter row of minute spines and a minute scale-like spine; fl/bt = 1.5–1.6.

*Sesarmops impressum* (H. Milne Edwards, 1837) (Figs. 12A–E)

Dimensions: rdl:  $0.80 \pm 0.01$  mm; cl:  $0.45 \pm 0.01$  mm.

Carapace (Fig. 12A). Smooth and without tubercles; dorsal spine present, straight; rostral spine present, straight and similar in length to dorsal spine.

Antenna (Fig. 12B). Protopod, slightly longer than rostral spine and bearing two unequal rows of 5–6, and 7–8 spines of different size, respectively; exopod elongated, more than 2/3 of the protopod length, with 2

terminal setae (1 long reaching the tip of protopod, 1 shorter) and 3 small terminal spines; pl/el = 2.8–2.9.

Abdomen (Fig. 12C, D). Somites 2 and 3 with pair of dorsolateral processes; somites 3–5 with small postero-lateral processes of subtriangular shape; somites 2–5 with a pair of posterodorsal simple setae.

Telson (Fig. 12E). Mid-internal side of inner pair without spines; dorsal part of each furcal arm with two rows of spines and outer part with a shorter row of minute spines and a minute scale-like spine; fl/bt = 1.5–1.7.

*Sesarmops intermedium* (de Haan, 1835) (Figs. 13A–E)

Baba and Fukuda (1975: 63–64, Fig. 1) (as *Sesarma*

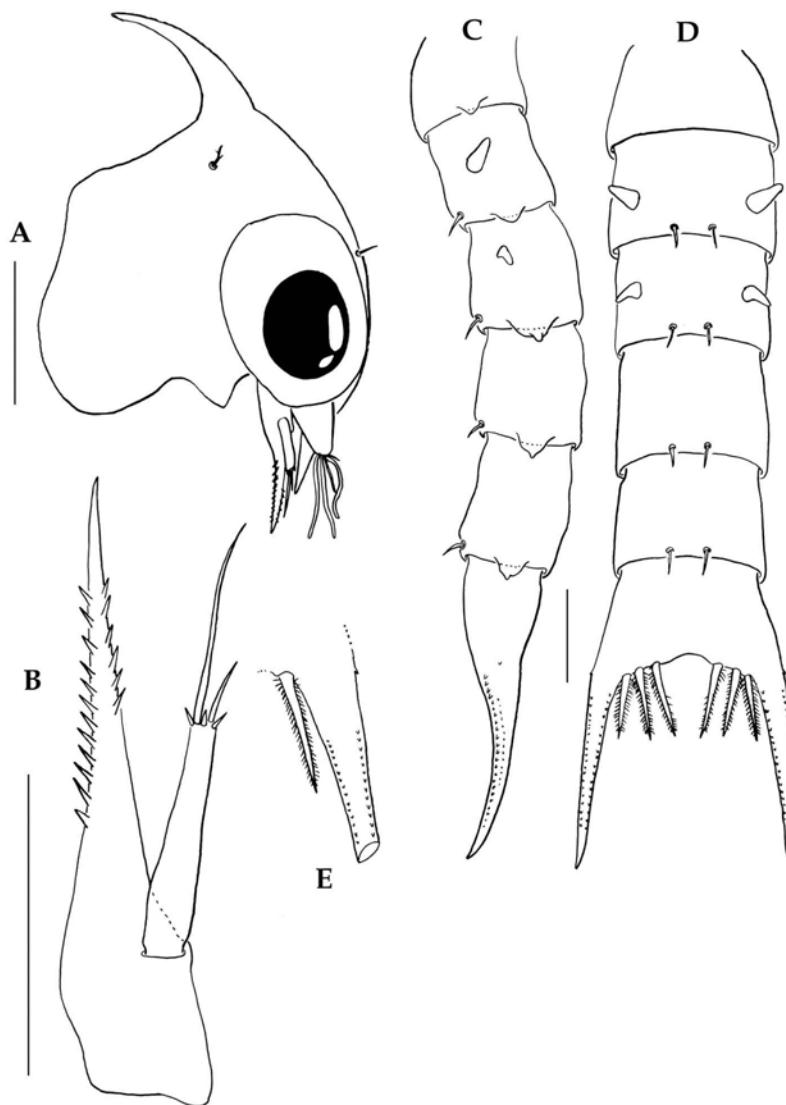


Fig. 11. *Clistocoeloma merguiense* de Man, 1888, zoea I. A, lateral view of carapace; B, antenna; C, abdomen, lateral view; D, abdomen, dorsal view; E, detail of dorsal area of furcal arm. Scale bars: A = 0.2 mm; C,D = 0.1 mm; B = 0.125 mm.

intermedium); Fukuda and Baba (1976: 63, Fig 5f) (as *Sesarmops intermedius*); Terada (1976: 143–148, Figs. 5A1, B1, C1, D1, 6E1, F1, G1, H1) (as *Sesarmops intermedia*).

Dimensions: rdl:  $0.75 \pm 0.05$  mm; cl:  $0.42 \pm 0.02$  mm.

Carapace (Fig. 13A). Smooth and without tubercles; dorsal spine present, slightly curved; rostral spine present, straight and similar in length to dorsal spine.

Antenna (Fig. 13B). Protopod almost reaching to the tip of the rostral spine and bearing two unequal rows of 4–5, and 7–8 spines of different size, respectively; exopod elongated, less than 1/4 of the protopod length, with 2 terminal setae (1 long reaching the tip of protopod, 1 shorter) and 2 small terminal spines; pl/el = 4.9–5.0.

Abdomen (Fig. 13C, D). Somites 2 and 3 with a pair of dorsolateral processes; somites 3–5 with postero-lateral processes of subtriangular shape, those of somites 4 and 5 more developed; somites 2–5 with a pair of postero-dorsal simple setae.

Telson (Fig. 13E). Mid-internal side of inner pair without spines; dorsal part of each furcal arm with two rows of spines and outer part with scattered minute spines and a minute scale-like spine; fl/bt = 1.6–1.7.

## Discussion

The first zoeal stage morphology of the eleven species from the present study and the previously described sesarmids is similar. Larval morphology of

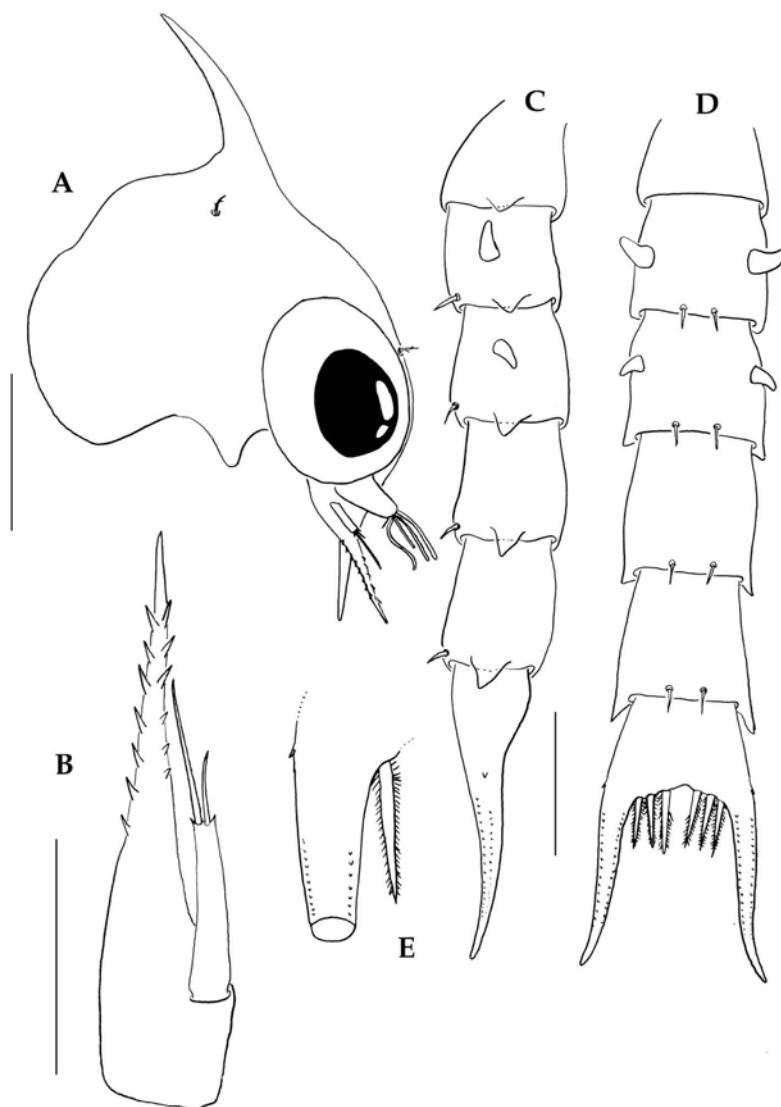


Fig. 12. *Sesarmops impressum* (H. Milne Edwards, 1837), zoea I. A, lateral view of carapace; B, antenna; C, abdomen, lateral view; D, abdomen, dorsal view; E, detail of dorsal area of furcal arm. Scale bars: A,C,D = 0.2 mm; B = 0.125 mm.

the Sesarmidae in general presents little variation between species and genera; the carapace, antennule, mouthparts, and abdomen do not vary in number of setae. The main differences were only found in carapace spinulation, antennal type, abdomen and telson morphology and armature (see Schubart and Cuesta, 1998; Cuesta et al., 1999; Figs. 1, 4–13 of present study). These differences allow, in some cases, identification of first zoeal stages to species, but the homogeneity of larvae often renders it difficult to characterise them to genus level.

In this study, the first zoeal stage of species belonging to *Labuanium*, *Stelgistra*, and *Pseudosesarma* are described for the first time. They do not present any diagnostic characters that would facilitate their

identification to genus level. In this study, the first zoeae of *Clistocoeloma merguiense*, *Metasesarma obesum* and *Sesarmops intermedium* are re-described and new characters are presented. A comparison with previous descriptions are summarised in Table 2. In Table 3, a comparison among the eleven first zoeal stages described in this study time is presented. Differences are mostly found in the antennal spinulation and exopod length (ratio pl/el), dorsal carapace spine morphology (from strongly curved to almost strait), and telson armature and furcal length (measured as ratio fl/bt). Among the observed differences is the size of the antennal exopod of *Sesarmops intermedium*, which is highly reduced when compared to that of *S. impressum* (pl/el 4.9–5 vs. 2.8–2.9). This difference surpasses the

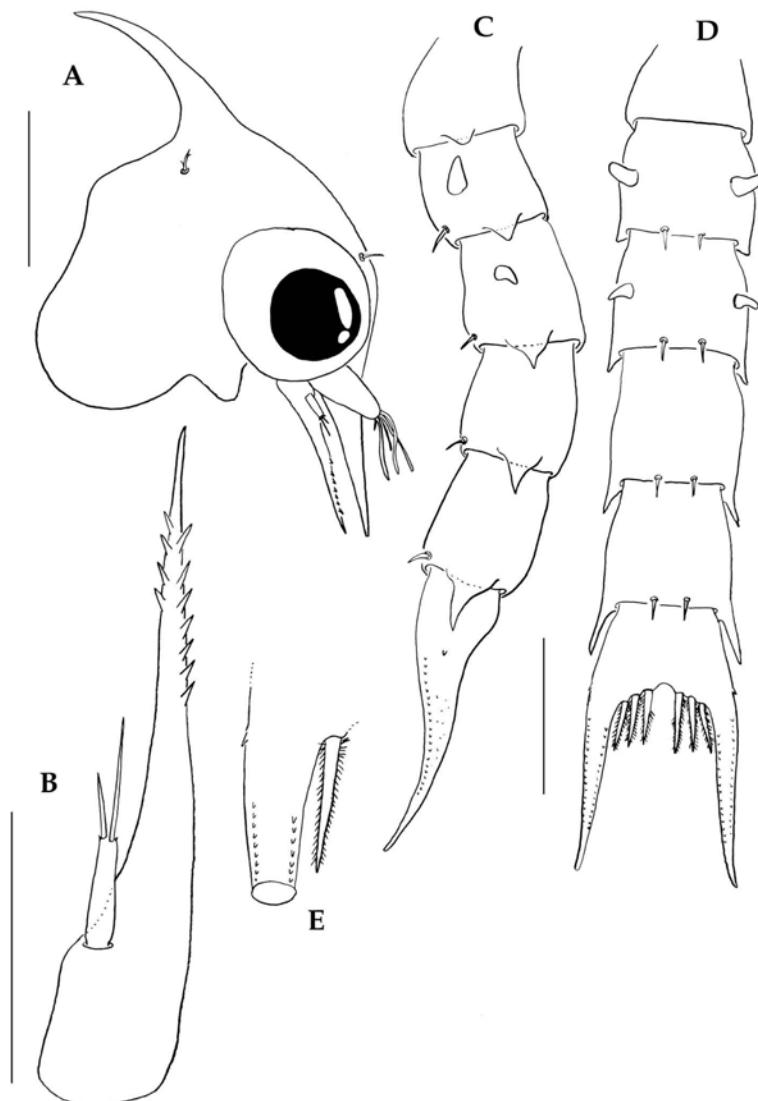


Fig. 13. *Sesarmops intermedium* (de Haan, 1835), zoea I. A, lateral view of carapace; B, antenna; C, abdomen, lateral view; D, abdomen, dorsal view; E, detail of dorsal area of furcal arm. Scale bars: A,C,D = 0.2 mm; B = 0.125 mm.

expected variation for congeneric species and questions the placement of these two species within the same genus. This confirms the unpublished findings of Schubart and Cuesta based on larval morphology and mitochondrial DNA, which suggested that *S. intermedium* was closer to *Chiromantes* than to *Sesarmops impressum* (type species of the genus).

This study discloses a character never previously described for sesarmid zoea: the presence of one or two scale-like outer minute spines on the telson furcae. These minute spines have been observed in all studied species with the exception of *Pseudosesarma crassimanum* and *Stelgistra stormi*. Careful re-examination of previously described sesarmids also revealed the

presence of these spines in all zoeal stages of *Aratus pisonii* (Cuesta et al., 2006), *Perisesarma fasciatum*, *Armases angustipes*, and *A. miersii*, and in zoeal stages of *Armases ricordi* (Z I–III), *A. cinereum* (Z I), *A. rubripes* (Z I), *Sesarma rectum*, *S. reticulatum*, *S. rhizophorae*. These spines were not present in *Sesarma curacaoense*, a species with abbreviated development. In zoea I of *Sesarma rubinofforum* and *Scandarma lintou*, the spines could not be observed clearly, so we cannot confirm their presence or absence. Due to the minute size of these spines, new studies with scanning electron microscopy will be needed to clarify the detailed morphology of these structures and to confirm their presence in other species.

Table 2. Comparison of morphological and meristic features between previous and present descriptions of the first zoeal stage of re-described species, *Clistocoeloma merguiense* (CLMER), *Metasesarma obesum* (MEOBE), and *Sesarmops intermedium* (SEINT). Abbreviations: ps, present study; a, aesthetascs; s., setation, s, seta/e; sp, spine/s; pp, posterior process, (-) equal to present description

	CLMER (Saba, 1972)	CLMER (ps)	MEOBE (Rajabai, 1961)	MEOBE (ps)	SEINT (Baba and Fukuda, 1975)	SEINT (Fukuda, and Baba, 1976)	SEINT (Terada, 1976)	SEINT (ps)
<b>Carapace</b>								
Anterodorsal s.	No data	1 pair	No data	1 pair	No data	No data	No data	1 pair
Dorsolateral s.	No data	1 pair	No data	1 pair	No data	No data	No data	1 pair
<b>Antennule</b>								
Protopod	3a	4a. 1s	3a	4a, 1s	4a	4a	-	4a, 1s
<b>Antenna</b>								
Exopod	2s	2s, 3 sp	2s	2s, 3 sp	1s	2s	2s	2s, 2 sp
<b>Maxillule</b>								
Coxal endite s.	No data	6	3	6	-	5	5	6
Basial endite s.	No data	5	2	5	-	-	-	-
Endopod	No data	1, 1+4	-	-	-	-	-	-
<b>Maxilla</b>								
Coxal endite s.	No data	5 + 3	2 + 2	5 + 3	5	4 + 3	-	5 + 3
Basial endite s.	No data	5 + 4	4 + 4	5 + 4	-	-	-	-
Endopod s.	No data	2 + 3	2 + 2	2 + 3	-	-	-	-
Scaphognathite	No data	4 s + pp	-	-	-	-	-	-
<b>First maxilliped</b>								
Basis s.	No data	2+2+3+3	1+1+2+2	2+2+3+3	No data	-	-	2+2+3+3
Endopod	No data	2,2,1,2,5	0,1,1,2,4	2,2,1,2,5	-	-	-	-
<b>Second maxilliped</b>								
Basis s.	No data	1+1+1+1	-	-	No data	1+1+1+1	-	1+1+1+1
Endopod	1, 0, 5	0, 1, 6	0, 1, 4	0, 1, 6	-	-	-	-
<b>Abdomen</b>								
Dorsolateral processes	somite 2	somites 2–3	-	-	-	-	-	-
<b>Telson</b>								
Outer minute spines	No data	1 pair	No data	2 pairs	No data	No data	No data	1 pairs

Table 3. Comparison of morphological and meristic features of the first zoeal stages described in the present study. Abbreviations: Plp, posterolateral process; fl, furcal length; bt, base of telson length. In brackets the variation in the number of spines for each row on antennal protopod

Species	Antennal protopod # spines	Abdomen Plp 5th somite	Telson	
			# scale-like spines	fl/bt
<i>Labuanium scandens</i>	(11–12) (15–16)	Not elongated	1	1.5–1.6
<i>Labuanium rotundatum</i>	(13–14) (20–21)	Elongated	1	1.6–1.7
<i>Labuanium trapezoideum</i>	(11) (14)	Elongated	1	1.4–1.5
<i>Labuanium politum</i>	(6–7) (12–13)	Not elongated	2	2.5–2.65
<i>Metasesarma aubryi</i>	(15–16) (24–26)	Not elongated	1	2–2.1
<i>Metasesarma obesum</i>	(10–12) (13–16)	Not elongated	2	1.3–1.5
<i>Pseudosesarma crassimanum</i>	(4–5) (6)	Elongated	0	1.7–1.8
<i>Stelgistra stormi</i>	(13–14) (23–24)	Not elongated	0	1.2–1.3
<i>Clistocoeloma merguiense</i>	(6) (11–12)	Not elongated	1	1.5–1.6
<i>Sesarmops impressum</i>	(5–6) (7–8)	Not elongated	1	1.5–1.7
<i>Sesarmops intermedium</i>	(4–5) (7–8)	Not elongated	1	1.6–1.7

Table 4. Sesarmidae species for which larvae have been described, including the number of described larval stages and authors. Abbreviations: \*, papers not examined; (-), data not accessible

Species	Larval stages described	Authors
<i>Aratus pisonii</i>	Prezoea–Zoea I Zoea I–IV–megalopa Zoea I Zoea I–IV–megalopa	Hartnoll, 1965 Warner, 1968 Fransozo et al., 1998 Cuesta et al., 2006
<i>Armases angustipes</i>	Zoea I–IV–megalopa Zoea I–IV–megalopa	Kowalcuk, 1994 Cuesta and Anger, 2001
<i>Armases cinereum</i>	Zoea I Zoea I–IV–megalopa	Hyman, 1924 Costlow and Bookhout, 1960
<i>Armases magdalenense</i>	Megalopa	Rathbun, 1923
<i>Armases miersii</i>	Zoea I–III–megalopa	Cuesta et al., 1999
<i>Armases ricordi</i>	Zoea I–IV–megalopa	Díaz and Ewald, 1968
<i>Armases rubripes</i>	Zoea I Zoea I–IV–megalopa Zoea V Zoea I	Schubart and Cuesta, 1998 Díaz and Ewald, 1968 Montú et al., 1990 Schubart and Cuesta, 1998
<i>Beanius andersoni</i>	Zoea I–IV	Vijayakumar and Kannupandi, 1986
<i>Bresedium brevipes</i>	Zoea I–IV–megalopa	Fielder and Greenwood, 1983
<i>Chiromantes dehaani</i>	Zoea I–IV <sup>(1)</sup> –megalopa (–) Zoea I–IV–megalopa Zoea I	Yatsuzuka, 1957* Terada, 1974* Baba and Miyata, 1971 Muraoka, 1979a*
<i>Chiromantes eulimene</i>	Zoea I–V Zoea I	Pereyra Lago, 1993a Flores et al., 2003
<i>Chiromantes haematocheir</i>	Zoea I–V–megalopa Zoea I–V <sup>(2)</sup> –megalopa Zoea I Megalopa	Terada, 1974 Fukuda and Baba, 1976 Muraoka, 1979a* Muraoka, 1980*
<i>Clistocoeloma lanatum</i>	Zoea I–IV–megalopa	Kakati and Sankolli, 1975
<i>Clistocoeloma merguiense</i>	Zoea I–III Zoea I	Saba, 1972 present study
<i>Episesarma lafondi</i>	Zoea I–IV–megalopa	Islam et al., unpublished data
<i>Episesarma mederi</i>	(–)	Selvakumar, 1988*
<i>Geosesarma notophorum</i>	Direct development	Ng and Tan, 1995
<i>Geosesarma peraccae</i>	Zoea I–II–“megalopa”	Soh, 1969
<i>Labuanium politum</i>	Zoea I	Present study
<i>Labuanium rotundatum</i>	Zoea I	Present study
<i>Labuanium scandens</i>	Zoea I	Present study
<i>Labuanium trapezoideum</i>	Zoea I	Present study
<i>Metasesarma aubryi</i>	Zoea I	Present study
<i>Metasesarma obesum</i>	Prezoea–zoea I Zoea I	Rajabai, 1961 Present study
<i>Metopaulias depressus</i>	Zoea I–II–megalopa	Hartnoll, 1964
<i>Muradium tetragonum</i>	Prezoea–zoea I (–)	Rajabai, 1961 Sundaramoorthy, 1987*
<i>Nanosesarma gordoni</i>	Zoea I–V Zoea I Zoea I–V	Fukuda, 1978 Muraoka, 1979b*
<i>Nanosesarma minutum</i>	Zoea I	Terada, 1982
<i>Neosarmatium indicum</i>	Zoea I–V–megalopa	Fukuda, 1978
<i>Neosarmatium meinerti</i>	Zoea I–V–megalopa	Islam et al., 2002
<i>Neosarmatium trispinosum</i>	Zoea I Zoea I–V–megalopa	Pereyra Lago, 1989 Flores et al., 2003
<i>Parasesarma acis</i>	Zoea I	Greenwood and Fielder, 1988 Islam et al., 2004
		Baba and Fukuda, 1975

Table 4, continued

Species	Larval stages described	Authors
<i>Parasesarma acis</i>	Zoea I–IV–megalopa Megalopa	Terada, 1976 Muraoka, 1980*
<i>Parasesarma batavicum</i>	(–)	Selvakumar, 1988*
<i>Parasesarma catenatum</i>	Zoea I–IV–megalopa Zoea I	Pereyra Lago, 1987 Flores et al., 2003
<i>Parasesarma erythrodactylum</i>	Zoea I Zoea I–V–megalopa	Green and Anderson, 1973 Greenwood and Fielder, 1988
<i>Parasesarma leptosoma</i>	Zoea I	Flores et al., 2003
<i>Parasesarma pictum</i>	(–)	Terada, 1974*
<i>Parasesarma plicatum</i>	Zoea I–IV–megalopa Zoea I Megalopa	Pasupathi and Kannupandi, 1987 Muraoka, 1979a* Muraoka, 1980
<i>Perisesarma bidens</i>	Zoea I–IV–megalopa Zoea I–V–megalopa	Baba and Fukuda, 1975 Fukuda and Baba, 1976
<i>Perisesarma fasciatum</i>	Zoea I–IV–megalopa	Selvakumar, 1999
<i>Perisesarma guttatum</i>	Zoea I–IV–megalopa Zoea I	Terada, 1976 Krishnan and Kannupandi, 1987
<i>Perisesarma messa</i>	Zoea I	Islam and Shokita, 2000
<i>Pseudosesarma crassimanum</i>	Zoea I	Guerao et al., 2004
<i>Sarmatium crassum</i>	Zoea I	Pereyra Lago, 1993b
<i>Scandarma lintou</i>	Zoea I	Flores et al., 2003
<i>Selatiuum brockii</i>	Zoea I–IV–megalopa	Schubart et al., 2003
<i>Sesarma aequatoriaile</i>	Zoea I	Vijayakumaran and Kannupandi, 1987
<i>Sesarma bidentatum</i>	Prezoea–Zoea I	Schubart and Cuesta, 1998
<i>Sesarma curacaoense</i>	Zoea I–II–megalopa	Hartnoll, 1964
<i>Sesarma rectum</i>	Zoea I–III–megalopa	Anger et al., 1995
<i>Sesarma reticulatum</i>	Zoea I	Schubart and Cuesta, 1998
<i>Sesarma rhizophorae</i>	Zoea I–III–megalopa	Fransozo and Hebling, 1986
<i>Sesarma rubinofforum</i>	Zoea I	Hyman, 1924
<i>Sesarma sp.</i>	Zoea I	Costlow and Bookhout, 1962
<i>Sesarmops impressum</i>	Zoea I	Schubart and Cuesta, 1998
<i>Sesarmops intermedium</i>	Zoea I	Schubart and Cuesta, 1998
<i>Stelgistra stormi</i>	Zoea I–V–megalopa Zoea I–V–megalopa Zoea I Zoea I Zoea I	Aikawa, 1929 Present study Baba and Fukuda, 1975 Terada, 1976 Fukuda and Baba, 1976
	Zoea I	Present study
	Zoea I	Muraoka, 1979a*
	Zoea I	Present study

Notes: (1) sometimes presents a zoeal V stage; (2) zoea IV and zoea V can moult to megalopa; (3) sometimes presents a zoeal VI stage, but do not moult to megalopa.

This type of telson spinulation has also been observed in other Grapoidea, viz. Glyptograpsidae (*Glyptograpsus impressus* and *Platychirograpsus spectabilis*) (see Schubart et al., 2002) and in two genera of the Gecarcinidae (*Cardisoma armatum*, *C. carnifex*, *C. guanhumi* and *Discoplax hirtipes*) (see Cuesta et al.,

2002; Flores et al., 2003; Cuesta and Anger, 2005). A possible relationship of Sesarmidae with *Cardisoma* and *Discoplax* has been suggested previously by Cuesta et al. (2002) based on the following shared larval features: antennal and telson morphology, and maxillar setation (2 + 3), but differing in second maxilliped endopod

Table 5. Zoeal morphological and meristic characters of the families Sesarmidae, Varunidae, Grapsidae, Plagusiidae, Glyptograpsidae and Gecarcinidae. Abbreviations: (−), absent; (+), present; Antenna type A refers to exopod equal to 1/4–2/3 of protopod length, with 2 unequal-sized simple terminal setae and 2–5 terminal short spines; B refers to exopod less than 1/2 of protopod length, with 2 simple terminal setae; C refers to exopod well developed, more than 1/2 protopod length with 0–2 medial setae; and D refers to exopod absent or reduced to a small protuberance with one terminal simple seta. Telson type A refers to a furca with or without minute outer spines, but always with 3 pairs of posterior processes throughout development; B refers to furca with or without minute outer spines, but with the number of pairs of posterior processes increasing throughout development

	Carapace lateral spines	Antenna type	Maxilla endopod	First maxilliped		Second maxilliped endopod	Telson type
				Basis	Endopod 1st seg. s.		
Gecarcinidae Macleay, 1838	(+)	A	2, 2/2, 3	2, 2, 3, 3	2	1, 1, 6	B
Glyptograpsidae Schubart, Cuesta and Felder, 2002	(+)	A	1, 2	2, 2, 3, 3	2	0, 1, 6 / 1, 1, 6	B
Grapsidae Macleay, 1838	(−)/(+)	D	2, 2	2, 2, 2, 2	1	0, 1, 5	B
Plagusiidae Dana, 1851	(+)	B	2, 3	2, 2, 2, 2	2	1, 1, 6	B
Sesarmidae Dana, 1851	(−)	A	2, 3	2, 2, 3, 3	2	0, 1, 6	A
Varunidae H. Milne Edwards, 1853	(−)/(+)	C	2, 2	2, 2, 3, 3	2	0, 1, 6	B

Table 6. Megalopal morphological and meristic characters of the families Gecarcinidae, Grapsidae, Plagusiidae, Sesarmidae, and Varunidae. Abbreviations: (−), absent; (+), present; m, marginal setae; l, lateral setae (presented as anterior-posterior)

	Antennule endopod	Antenna no. of segments	Mandible palp setation	Maxilla scaphognathite setation	Second maxilliped epipodite	Pleopod cincinnuli	Uropod setation
Gecarcinidae	(+)	9–10	0–2, 7–11	(60–85) m, (3/4–0/1/2) 1	(−)	3–4	0–5, 12–17
Grapsidae	(+)	11	0, 7–15	(58–86) m, (3–1) 1	(+)	3–6	0–3, 13–23
Plagusiidae	(+)	11	0, 1–4, 15–30	(73–134) m, (5–6) 1	(+)	8–17	5–9, 27–33
Sesarmidae	(−)	8–9	0, 4	(25–50) m, (2–0/1) 1	(−)	2	1, 5–7
Varunidae	(+)	10	0, 5–13	(39–90) m, (3–2) 1	(+)	3	1, 8–13

setation (0, 1, 6 vs 1, 1, 6) and lateral carapace spines (absent vs present). Close phylogenetic relationship of these taxa has not been confirmed to date by molecular systematics, though a possible sister group relationship between Glyptograpsidae and Sesarmidae was weakly supported in the phylogeny of Schubart et al. (2000), but not in Schubart et al. (2002).

The Varunidae do not present outer spines on the telson (Cuesta et al., 2000), but in Plagusiidae, Grapsidae, and *Gecarcoidea lalandii*, *Epigrapsus notatus*, *E. politus* (Gecarcinidae) they are well developed.

Complete or partial larval development is now known for only 23 Sesarmidae genera (ca. 75% of the sesarmid genera) and 54 species (ca. 25% of the species) (see Table 4). This is the case for zoeal stages, but megalopae are known from only 12 genera (ca. 40% of sesarmid genera). However, although no larval descriptions are available for *Episesarma*, *Haberma*, *Metagrapsus*, *Namlacium*, *Neosesarma*, *Sesarmoides*, and *Tiomanum* it is possible to propose a suite of

characters that define sesarmid larvae similar to the general larval characters established for other grapsoid families (Cuesta et al., 1997, Cuesta and Schubart, 1997, 1999, Cuesta et al., 2000, Schubart et al., 2002, Cuesta et al., 2002; Cuesta and Anger, 2005).

These sesarmid zoeal characters are:

(1) Carapace without lateral spines. Zoa I with a pair of anterodorsal setae.

(2) Antennal exopod with terminal small spines and setae of different size. Exopod with variable length, commonly between 1/4 and 2/3 of the protopod length. Protopod with well developed spines distributed in two rows, normally with unequal number of spines.

(3) Maxilliped basis with 2+2+3+3 setae.

(4) First maxilliped basis with 2+2+3+3 setae. Endopod setation 2, 2, 1, 2, 5 in the first zoea. Through development the segments 2 and 5 acquire one seta each, and likewise another one in segment 3. The last zoeal stages present a setation 2, 2, 2, 2, 6 or more frequently 2, 3, 2, 2, 6.

(5) Second maxilliped basis with 1+1+1+1 setae. Endopod setation 0, 1, 6.

(6) Abdomen of first zoeal stage with 5 somites and last zoeal stage with 6 somites. Dorsolateral processes only on somites 2 and 3. In last stage, somite 1 presents 3 middorsal setae.

(7) Telson with 3 serrulate setae on posterior margin throughout development. Furcal arms with two dorsal rows of spinules of varying size.

Sesarmid megalopal characters:

(1) Carapace longer than broad. Rostrum ventrally deflected (approximately 70°) with a medial cleft.

(2) Antennule endopod absent.

(3) Antennal peduncle 3-segmented and flagellum with 5 or 6 segments.

(4) Mandibular palp 2-segmented with 4 setae on distal segment.

(5) Maxillar scaphognathite with less than 40 marginal plumose setae, and with 2 anterior and 1 posterior lateral setae.

(6) First maxilliped epipod with a maximum of 7 setae.

(7) Second maxilliped epipod absent or in a few cases rudimentary and without setae.

(8) Pleopod exopods with 7–14 plumose setae and two cincinuli on endopod.

(9) Uropods with setation 1,5, 1,6 or 1,7.

The number of zoeal stages in sesarmid species varies from 5 as in *Neosarmatium trispinosum* and *Perisesarma guttatum*, to 4 (typical) as in *Aratus pisonii*, *Armases angustipes* and *Parasesarma catenatum*, 3 as in *Sesarma reticulatum* and *Armases miersii*, 2 as in *Metopaulias depressus* and *Sesarma curacaoense*, and direct development as in *Geosesarma notophorum* (see Table 4). A similar case of variability in the number of zoeal stages has been described by Clark (2005) for the Pilumninae.

All grapsoid larval stages can be identified to family level by the combination of the zoeal and megalopal features listed in Tables 5 and 6 with the exception of Glyptograpsidae for which the megalopae are unknown. Although sesarmid zoeae tend to be larger than those of other families, their megalopae are the smallest ones among the Grapoidea. Decrease in megalopa size goes along with the reduction in the appendage setation and the reduction and loss of structures such as antennular endopod or the second maxilliped epipodite (see Table 6), but not the reduction of segmentation (i.e. antennal flagellum).

### Acknowledgements

José A. Cuesta was funded by a research contract Ramón y Cajal from Ministerio de Educación y Ciencia,

Spain. Christoph D. Schubart was supported by a research fellowship of the Raffles Museum Singapore, kindly facilitated by Peter K.L. Ng, which also allowed him to travel to Taiwan and Sulawesi. Hung-Chang Liu was invited by P.K.L. Ng to participate in collecting activities in Guam and the Philippines. The knowledgeable and pleasant company of Siva in the mangroves of Singapore will always be remembered. This contribution was presented as a poster at the Fourth Crustacean Larval Conference, hosted as a Symposium of the Sixth International Crustacean Conference held at Glasgow, Scotland, UK (July 18–22, 2005). We want to thank Paul Clark for his effort as organizer of the 4CLC and editor of the proceedings volume of this Conference. Thanks are also due to Ngan Kee Ng and two anonymous referees for their suggestions and criticism that clearly improved an earlier version of the manuscript.

### References

- Aikawa, H., On larval forms of some brachyura. Rec. Oceanogr. Works, Japan, 2 (1929) 17–55.
- Anger, K., Schreiber, D. and Montú, M., Abbreviated larval development of *Sesarma curacaoense* (Rathbun, 1897) (Decapoda: Grapsidae) reared in the laboratory. Nauplius, 3 (1995) 127–154.
- Baba, K. and Fukuda, Y., Newly obtained first zoeae of three species of *Sesarma* (Crustacea, Brachyura). Mem. Fac. Educ. Kumamoto Univ., 24 (1975) 63–68.
- Baba, K. and Miyata, K., Larval development of *Sesarma (Holometopus) dehaani* H. Milne Edwards (Crustacea, Brachyura) reared in the laboratory. Mem. Fac. Educ. Kumamoto Univ., 19 (1971) 54–64.
- Clark, P.F., The evolutionary significance of heterochrony in the abbreviated zoeal development of pilumnine crabs (Crustacea: Brachyura: Xanthoidea). Zool. J. Linn. Soc., 143 (2005) 417–446.
- Clark, P.F., Calazans, D.K. and Pohle, G.W., Accuracy and standardization of brachyuran larval descriptions. Invert. Reprod. Develop., 33 (1998) 127–144.
- Costlow, J.D. Jr. and Bookhout, C.G., The complete larval development of *Sesarma cinereum* (Bosc) reared in the laboratory. Biol. Bull., 118 (1960) 203–214.
- Costlow, J.D. Jr. and Bookhout, C.G., The larval development of *Sesarma reticulatum* Say reared in the laboratory. Crustaceana, 4 (1962) 281–294.
- Cuesta, J.A. and Anger, K., Larval morphology of the sesarmid crab *Armases angustipes* (Dana, 1852) (Crustacea, Decapoda, Grapoidea) reared under laboratory conditions. J. Crust. Biol., 21(3) (2001) 821–838.
- Cuesta, J.A. and Anger, K., Morphology and salinity tolerance of the larval stages of *Cardisoma armatum* (Brachyura: Grapoidea: Gecarcinidae) reared in the laboratory. J. Crust. Biol., 25(4) (2005) 640–654.
- Cuesta, J.A., Diesel, R. and Schubart, C.D., Re-examination of zoeal morphology of *Chasmagnathus granulatus*,

- Cyclograpus lavauxi*, *Hemigrapsus crenulatus*, and *H. edwardsi* confirms consistent chaetotaxy in Varunidae (Decapoda: Brachyura). *Crustaceana*, 74(9) (2001) 895–912.
- Cuesta, J.A., González-Gordillo, J.I. and Rodríguez, A., First zoeal stages of *Grapus adscensionis* (Osbeck) and *Planes minutus* (Linnaeus) (Brachyura, Grapsidae) hatched in the laboratory, with notes on larval characters of the subfamily Grapsinae. *J. Nat. Hist.*, 31(6) (1997) 887–900.
- Cuesta, J.A., Liu, H.-C. and Schubart, C.D., First zoeal stages of *Epigrapsus politus* Heller and *Gecarcinoides lalandii* H. Milne-Edwards, with remarks on zoeal morphology of the Gecarcinidae Macleay (Crustacea: Brachyura). *J. Nat. Hist.*, 36(14) (2002) 1671–1685.
- Cuesta, J.A., Rodríguez, A., García-Guerrero, M.U. and Hendrickx, M.E., Larval morphology of the sesarmid crab *Aratus pisonii* (H. Milne Edwards, 1837) (Decapoda, Brachyura, Grapoidea) from laboratory-reared material. *Crustaceana*, 79(2) (2006) 175–196.
- Cuesta, J.A. and Schubart, C.D., The first zoeal stage of *Glyptograpsus impressus*, with comments on the subfamilial arrangement of Grapsidae (Crustacea: Brachyura). *Cah. Biol. Mar.*, 38 (1997) 291–299.
- Cuesta, J.A. and Schubart, C.D., First zoeal stages of *Geograpsus lividus* and *Goniopsis pulchra* from Panama confirm consistent larval characters within the subfamily Grapsinae (Brachyura: Grapsidae). *Ophelia*, 51(3) (1999) 163–176.
- Cuesta, J.A., Schubart, C.D. and Rodríguez, A., Larval development of *Brachynotus sexdentatus* Risso (Crustacea, Decapoda, Varunidae) reared in the laboratory, with notes on larval characters of the Varunidae. *Invert. Reprod. Develop.*, 38(3) (2000) 207–223.
- Cuesta, J.A., Schubart, C.D., Schuh, M. and Diesel, R., Abbreviated development of *Armases miersii* (Grapsidae: Sesarminae), a crab that breeds in supralittoral rock pools. *J. Crust. Biol.*, 19(1) (1999) 26–41.
- Dana, J.D., Conspectus crustaceorum quae in orbis terrarum circumnavigatione, Carolo Wilkes e classe Reipublicae Foederatae Duce, lexit et descripsit J. D. Dana. *Crustacea Grapoidea*, (Cyclometopa, Edwardsii). *Proc. Acad. Nat. Sci., Philadelphia*, 5 (1851) 247–254.
- Díaz, H. and Ewald, J.J., A comparison of the larval development of *Metasesarma rubripes* (Rathbun) and *Sesarma ricordi* H. Milne-Edwards (Brachyura, Grapsidae) reared under similar laboratory conditions. *Crustaceana* (Suppl. II) (1968) 225–248.
- Fielder, D.R. and Greenwood, J.G., The zoeal stages and megalopa of *Bresedium brevipes* (de Man, 1899) (Crustacea: Decapoda: Grapsidae), reared in the laboratory. *J. Plankton Res.*, 5 (1983) 585–598.
- Flores, A.A.V., Paula, J. and Dray, T., First zoeal stages of grapsoid crabs (Crustacea: Brachyura) from the East African coast. *Zool. J. Linn. Soc.*, 137 (2003) 355–383.
- Fransozo, A. and Hebling, N.J., Desenvolvimento larval de *Sesarma (Holometopus) rectum* Randall, 1840 (Decapoda, Grapsidae) em laboratório. *Rev. Brasil. Biol.*, 46(2) (1986) 353–364.
- Fransozo, A., Cuesta, J.A. and Negreiros-Fransozo, M.L., First zoeal stage of two species of Grapsidae (Decapoda, Brachyura) and a key to such larvae from the Brazilian coast. *Crustaceana*, 71(3) (1998) 331–343.
- Fukuda, Y., Preliminary notes on recently obtained larvae of brachyuran Crustacea of the sea around the Aitsu Marine Biological station. *Calanus*, 6 (1978) 10–16.
- Fukuda, Y. and Baba, K., Complete larval development of the sesarmid crabs, *Chromantes bidens*, *Holometopus haematocheir*, *Parasesarma plicatum* and *Sesarmops intermedius*, reared in the laboratory. *Mem. Fac. Educ. Kumamoto Univ.*, 25 (1976) 61–75.
- Green, P.A. and Anderson, D.T., The first zoea larvae of the estuarine crabs *Sesarma erythrodactyla* Hess, *Helograsus haswellianus* (White-Legge) and *Chasmagnathus laevis* Dana (Brachyura, Grapsidae, Sesarminae). *Proc. Linn. Soc. N.S.W.*, 98 (1973) 13–28.
- Greenwood, J.G. and Fielder, D.R., Larval development of three species of *Sesarma* (Crustacea, Brachyura, Grapsidae) from Eastern Australia. *Micronesia*, 21 (1988) 71–91.
- Guerao, G., Anger, K., Nettelmann, U. and Schubart, C.D., Complete larval and juvenile development of the mangrove crab *Perisesarma fasciatum* (Crustacea: Brachyura: Sesarmidae) from Singapore, with a larval comparison of *Parasesarma* and *Perisesarma*. *J. Plankton Res.*, 26 (2004) 1389–1408.
- de Haan, W. Crustacea. Fauna Japonica, sive Descriptio animalium, quae in itinere per Japoniam, jussu et auspiciis superiorum, qui sumnum in India Batava imperium tenent, suscepto, annis 1823–1830 collegit, notis, observationibus et adumbrationibus illustravit P.F. de Siebold. *Conjunctis studiis* C.J. Temminck et H. Schlegel pro Vertebratis atque W. de Haan pro Invertebratis elaborata Regis aupicus edita. I. P. F. v. Siebold. Leiden, Lugundi-Batavorum. Decas II (1835) 25–64, pls 9–15, 17, C, D. (For dates see Sherborn and Jentink, 1895; Holthuis, 1953 and Holthuis and T. Sakai, 1970).
- Hartnoll, R.G., The freshwater grapsid crabs of Jamaica. *Proc. Linn. Soc. Lond.*, 175 (1964) 145–169.
- Hartnoll, R.G., Notes on the marine grapsid crabs of Jamaica. *Proc. Linn. Soc. Lond.*, 176 (1965) 113–147.
- Hess, W., Beiträge zur Kenntnis der Decapoden-Krebse Ost-Australiens. *Archiv für Naturgeschichte*, 31 (1865) 127–173.
- Hyman, O.W., Studies on larvae of crabs of the family Grapsidae. *Proc. U.S. Nat. Mus.*, 65 (1924) 1–8.
- Islam, M.S., Rahman, M.A. and Shokita, S., Larval development of the mangrove crab *Neosarmatium trispinosum* (Brachyura: Grapoidea) described from laboratory-reared material. *J. Crust. Biol.*, 24 (2004) 356–371.
- Islam, M.S. and Shokita, S., Larval development of the mangrove crab *Perisesarma bidens* (De Haan) (Crustacea: Brachyura: Sesarminae). *Bangladesh J. Fish. Res.*, 4 (2000) 43–56.
- Islam, M.S., Shokita, S. and Shikatani, N., Larval development of the mangrove crab *Neosarmatium indicum* (Brachyura: Grapoidea) described from laboratory-reared material. *J. Crust. Biol.*, 22 (2002) 916–937.
- Kakati, V.S. and Sankolli, K.N., Larval culture of an estuarine crab, *Sesarma lanatum* Alcock in the laboratory

- (Brachyura, Grapsidae). Bull. Dept. Mar. Sci. Univ. Cochim, 7(2) (1975) 389–401.
- Kowalcuk, V.G., Estructura populacional de *Armases angustipes* (Dana, 1852) (Decapoda. Brachyura: Grapsidae) da Ilha do Farol, Caiobá, PR e seu desenvolvimento pós-embrionario sob condições de laboratório. Master Thesis, University of Curitiba, 1994.
- Krishnan, T. and Kannupandi, T., Larval development of the mangrove crab *Sesarma bidens* (De Haan, 1853) in the laboratory (Brachyura: Grapsidae: Sesarminae). Mahasagar, 20 (1987) 171–181.
- MacLeay, W.S., On the Brachyurous Decapod Crustacea. Brought from the Cape by Dr. Smith. Illustrations of the Zoology of South Africa; consisting chiefly of figures and descriptions of the objects of natural history collected during an expedition into the interior of South Africa, in the years 1834, 1835, and 1836; fitted out by “The Cape of Good Hope Association for Exploring Central Africa;” together with a summary of African Zoology, and an inquiry into the geographical ranges of species in that quarter of the globe, Published under the Authority of the Lords Commissioners of Her Majesty's Treasury, Invertebratae. A. Smith. London, Smith, Elder and Co.: (1838) [1849] IV: 53–71, pls 2, 3. [For dates of publication see Waterhouse 1880: 489–491].
- Man, J.G. De, Uebersicht der Indo-Pacificischen Arten der Gattung *Sesarma* Say nebst einer Kritik der von W. Hess und E. Nauck in den Jahren 1865 und 1880 beschriebenen Decapoden. Zool. Jahrb., Abth. f. Syst., 2(3–4) (1887) 639–722, pl. 17.
- Man, J.G. De, Report on the podophthalmous Crustacea of the Mergui Archipelago, collected for the Trustees of the Indian Museum, Calcutta, by Dr. John Anderson, F.R.S., Superintendent of the Museum. J. Linn. Soc., 22 (1888) 1–312, pl. 1–19.
- Man, J.G. De, Bericht über die von Herrn Schiffscapitän Storm zu Atjeh, an den westlichen Küsten von Malakka, Borneo und Celebes sowie in der Java-See gesammelten Decapoden und Stomatopoden. Zool. Jahrb., Abth. f. Syst., 8(4) (1895) 485–609; 9(5): pl. 12–14.
- Milne-Edwards, A., Note sur quelques nouvelles espèces du genre *Sesarma* (Say). Nouv. Archs Mus. Hist. nat. Paris, 5(4) (1869) 25–31.
- Milne Edwards, H., Histoire naturelle des Crustacés, comprenant l'anatomie, la physiologie et la classification de ces animaux, Vol. 2, 1837, Librairie Encyclopédique de Roret, Paris, 532 pp. Atlas 32 pp.
- Milne Edwards, H., Mémoires sur la famille des Ocyptodiens. Annls Sci. nat. série 3 (Zoologie), 20 (1853) 163–228, pls 6–11.
- Montú, M., Anger, K. and De Bakker, C., Variability in the larval development of *Metasesarma rubripes* (Decapoda, Grapsidae) reared in the laboratory. Neritica, 5 (1990) 113–128.
- Muraoka, K., Brachyuran larvae (Crustacea, Decapoda) – 3. Aquabiology, 1 (1979a) 48–49.
- Muraoka, K., Brachyuran larvae (Crustacea, Decapoda) – 4. Aquabiology, 1 (1979b) 54–55.
- Muraoka, K., Brachyuran larvae (Crustacea, Decapoda) – 5. Aquabiology, 2 (1980) 60–61.
- Ng, P.K.L. and Liu, H.-C., On a new species of tree-climbing crab of the genus *Labuanium* (Crustacea: Decapoda: Brachyura: Sesarmidae) from Taiwan. Proc. biol. Soc. Wash., 116 (2003) 601–616.
- Ng, P.K.L. and Schubart, C.D., On the identities of *Sesarma obesum* Dana, 1851, and *Sesarma eydouxi* H. Milne Edwards, 1853 (Crustacea: Decapoda: Brachyura: Sesarmidae). Zoosistema, 25(3) (2003) 425–437.
- Ng, P.K.L. and Tan, C.G.S., *Geosesarma notophorum* sp. nov. (Decapoda, Brachyura, Grapsidae, Sesarminae), a terrestrial crab from Sumatra, with novel brooding behaviour. Crustaceana, 68(3) (1995) 390–395.
- Pasupathi, K. and Kannupandi, T., Laboratory culture of a mangrove crab, *Sesarma pictum* De Haan, 1853 (Brachyura: Grapsidae). In: Proc. Fifth Indian Symp. Invertebrate Reproduction, S. Palanichamy (ed.), 1987, pp. 294–307.
- Pereyra Lago, R., Larval development of *Sesarma catenata* Ortmann (Brachyura, Grapsidae, Sesarminae) reared in the laboratory. S. Afr. J. Zool., 22 (1987) 200–212.
- Pereyra Lago, R., The larval development of the red mangrove crab *Sesarma meinerti* de Man (Brachyura, Grapsidae) reared in the laboratory. S. Afr. J. Zool., 24 (1989) 199–211.
- Pereyra Lago, R., The zoeal development of *Sesarma eulimene* de Man (Decapoda, Brachyura, Grapsidae), and identification of larvae of the genus *Sesarma* in South African waters. S. Afr. J. Zool., 28 (1993a) 173–181.
- Pereyra Lago, R., Larval development of *Sesarma guttatum* A. Milne Edwards (Decapoda. Brachyura. Grapsidae) reared in the laboratory, with comments on larval generic and familial characters. J. Crust. Biol., 13 (1993b) 745–762.
- Rajabai, K.G., Studies on the larval development of Brachyura VII. Early development of *Metopograpsus messor* (Forskal), *Plagusia depressa squamosa* (Herbst) *Metasesarma rousseuxii* A. M. Edwards and *Sesarma tetragonum* (Fabricius) of the family Grapsidae. J. Zool. Soc. India, 13 (1961) 154–165.
- Rathbun, M.J., The Brachyuran crabs collected by the U.S. Fisheries Steamer “Albatross” in 1911, chiefly on the West Coast of Mexico. Bull. Am. Mus. nat. Hist., 48 (1923) 619–637.
- Saba, M., “Umore-benkeigani” (*Clistocoeloma merguiense* de Man) no koukihassei. Mie-seibutsu, 22 (1972) 25–29.
- Schubart, C.D. and Cuesta, J.A., First zoeal stages of four *Sesarma* species from Panama, with identification keys and remarks on the American Sesarminae (Crustacea: Brachyura: Grapsidae). J. Plankton Res., 20 (1998) 61–84.
- Schubart, C.D., Cuesta, J.A., Diesel, R. and Felder, D.L., Molecular phylogeny, taxonomy, and evolution of non-marine lineages within the American Grapoidea (Crustacea: Brachyura). Mol. Phylogen. Evol., 15 (2000) 179–190.
- Schubart, C.D., Cuesta, J.A. and Felder, D.L., Glypto-grapsidae, a new brachyuran family from Central America: larval and adult morphology, and a molecular phylogeny of the Grapoidea. J. Crust. Biol., 22 (2002) 28–44.

- Schubart, C.D., Liu, H.-C. and Cuesta, J.A., A new genus and species of tree-climbing crab (Crustacea: Brachyura: Sesarmidae) from Taiwan with notes on its ecology and larval morphology. *Raffles Bull. Zool.*, 51 (2003) 49–63.
- Selvakumar, S., Life history salinity tolerance and bioassay studies on the larvae of some grapsid and xanthid crabs of Portonovo coast (Brachyura: Decapoda: Crustacea). PhD. Thesis, Annamalai University, 1988.
- Selvakumar, S., The complete larval development of *Parasesarma plicatum* (Latreille, 1806) (Decapoda: Brachyura: Grapsidae) reared in the laboratory. *Raffles Bull. Zool.*, 47 (1999) 237–250.
- Soh, C.L., Abbreviated development of a non-marine crab, *Sesarma (Geosesarma) perracae* (Brachyura, Grapsidae), from Singapore. *J. Zool., Lond.*, 158 (1969) 357–370.
- Sundaramoorthy, S., Taxonomical, developmental and larval bioassay studies in the mangrove crab *Neoepisesarma (Muradium) tetragonum* Fabricius (Grapsidae, Brachyura, Decapoda). PhD. Thesis, Annamalai University, 1987.
- Terada, M., Studies on the post-embryonic development in some crabs of the family Grapsidae (subfamilies Varuninae and Sesarminae). Futamata High School, Shizuoka Prefecture, 1974, 52 pp., 18 figs.
- Terada, M., Comparison of the larval development of nine crabs belonging to the subfamily Sesarminae. *Res. Crustacea*, 7 (1976) 138–169.
- Terada, M., The zoeal development of *Nanosesarma gordoni* (Shen) (Brachyura, Sesarminae) in the laboratory. *Proc. Jap. Soc. syst. Zool.*, 22 (1982) 35–45.
- Vijayakumar, G. and Kannupandi, T., Zoeae and megalopa of the mangrove crab *Sesarma andersoni* De Man reared in the laboratory. *Mahasagar*, 19 (1986) 245–255.
- Vijayakumar, G. and Kannupandi, T., Laboratory-reared zoeae and megalopa of the mangrove crab *Sesarma brockii* De Man. *Ind. J. Fish.*, 34 (1987) 133–144.
- Warner, G.F., The larval development of the mangrove tree crab *Aratus pisonii* (H. Milne-Edwards) reared in the laboratory (Brachyura, Grapsidae). *Crustaceana, Suppl.* 2 (1968) 249–258.
- Wear, R.G., Life-history studies on New Zealand Brachyura. 4. Zoea larvae hatched from crabs of the family Grapidae. *N.Z. J. mar. Freshw., Res.*, 4 (1970) 3–35.
- Yatsuzuka, K., Study of brachyuran zoea, artificial rearing and development. *Suisangaku Shusei*, (1957) 571–590.