1	Differences in the reproductive behaviour and larval development
2	of two Canthon rutilans subspecies reinforce their thermal
3	regional segregation
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1 Abstract

- 2 Two dung beetle subspecies of *Canthon rutilans* Castelnau (Coleoptera: Scarabaeidae,
- 3 Scarabaeinae), C. rutilans rutilans and C. rutilans cyanescens, inhabit different environmental
- 4 temperature conditions in southern Brazil. We developed a laboratory breeding experiment
- 5 with 60 pairs of individuals of the two subspecies at five temperature conditions (from 15 to
- $6 \quad 35 \text{ °C}$) to compare the influence of temperature on the behaviour of adults and the
- 7 development of larvae. The behavioural patterns of the adults in both subspecies differ
- 8 according to temperature. The size of food balls was smaller and lighter in *C. rutilans*
- 9 *cyanescens*. Although temperature did not influence the feeding behaviour (measured as the
- 10 number of food balls made), reproductive behaviour (measured as the number of brood balls
- 11 made by pair) was significantly lower at 15 °C for both subspecies. Besides that, brood balls
- 12 from *C. rutilans rutilans* were bigger and heavier at lower temperatures. The number of
- 13 offspring and the time of emergence depend on temperature too. However, the weight of the
- 14 offspring and the longevity of adults depend on the subspecies and temperature treatment.
- 15 These results demonstrate that both subspecies have different thermal adaptations, where *C*.
- 16 *rutilans rutilans* has reproductive behaviour adapted to living under colder and broader
- 17 conditions than *C. rutilans cyanescens*.
- 18

19 Abbreviated abstract (2-3 sentences, max. 80 words)

Two spatially segregated dung beetle subspecies, *Canthon rutilans rutilans* and *C. rutilans cyanescens* (Coleoptera: Scarabaeidae), were tested under various experimental thermal conditions. Reproductive behaviour, larval development, and longevity of adults differed depending on the temperature, where *C. rutilans rutilans* has reproductive behaviour adapted to living under colder and broader conditions than *C. rutilans cyanescens*. These behavioural and developmental adaptations support but not confirm that these two taxa can be considered different species.

27

1 Introduction

2 Insects are exposed to climatic variation in temperature, humidity, atmospheric pressure,

3 wind, and precipitation, that may involve changes in their productivity, competition, and

4 predation (Rahbek, 1995; Lieberman et al., 1996; Lomolino, 2001). Temperature conditions

5 during development can affect the adults of many ectotherms (Atkinson, 1994; Atkinson &

6 Sibly, 1997). Insects have response mechanisms at non-freezing low temperatures (Chown &

7 Nicholson, 2004). The endothermic capacity for flight in some insects modifies their

8 geographical distribution, such as habitat selection (Blouin-Demers & Weatherhead, 2002;

9 Chown & Nicholson, 2004). Depending on their physiological limits, an insect species has its

10 own thermal niche, which limits its actual and potential range and which can be used to

11 explain species distribution (Verdú et al., 2006).

Dung beetle species (Coleoptera: Scarabaeidae, Scarabaeinae) are preferentially diversified 12 in tropical regions (Halffter, 1991) as temperature is a limiting factor for the development of 13 dung beetles inhabiting cold-temperate areas (Hanski, 1991). Similarly, the richness of 14 species decreases with elevation (Escobar et al., 2007; da Silva et al., 2018). At higher 15 elevations, the annual and daily thermal ranges are greater and dung beetle species have a 16 17 larger range of thermal tolerance than species at lower elevations (Gaston & Chown, 1999). Species with capacity to regulate their body temperature have a higher resistance to low 18 temperatures (Verdú et al., 2006). Other adaptations to low temperatures include the use of 19 20 alternative trophic resources; e.g., the dung beetle Thorectes lusitanicus (Jekel) can also eat acorns (Quercus spp.), making it more resistant to low temperatures (Pérez-Ramos et al., 21 2007; Verdú et al., 2010). However, the dung beetle's most common trophic resource is 22 vertebrate faeces, which is used by both juveniles and adults (Halffter & Edmonds, 1982). 23 Recently, it was proposed that two subspecies of the Neotropical species Canthon 24 (Francmonrosia) rutilans Castelnau - the red Canthon rutilans rutilans Castelnau and the 25 blue Canthon rutilans cyanescens Harold (Figure 1) - may be two species, considering their 26 differing thermal preferences found in a regional study (Hensen et al., 2018). In southern 27 Brazil, these subspecies do not seem to co-occur: C. rutilans rutilans occurs in cool and 28 29 subtropical montane regions at more than 1 000 m elevation and adults are active at mean environmental temperatures of 14.4-18.6 °C, whereas C. rutilans cyanescens occurs in 30 regions under 1 000 m and adults are active at 15.3-24.0 °C (Hensen at al., 2018). These two 31 subspecies are rollers (telecoprids), i.e., they make dung balls that are buried for feeding or 32 nesting. When nesting, females transform the ball to a 'water drop' shape and put one egg in it 33

(Halffter & Edmonds, 1982; Hernández et al., 2020). Males and females cooperate in parental
 care of brood balls (Halffter & Edmonds, 1982).

After collecting in a field station, specimens of both subspecies were bred in the laboratory 3 at different temperatures. As is often the case, different populations of both subspecies may 4 5 have a specific but plastic thermal response (Chown & Terblanche, 2006). In this study, we assume that the specific thermal responses of the selected individuals will become apparent 6 7 under the conditions used along an experimental gradient. The main objective of this study was to compare the influence of temperature on the behaviour, dung ball production, larval 8 9 development, and adult longevity of C. rutilans rutilans and C. rutilans cyanescens. We hypothesize that the adult behaviour and development of the specimens differ between the 10 subspecies in agreement with their thermal preferences derived from field data. 11

12

13 Materials and methods

14 Dung beetle capture for experiments

15 Dung beetles were captured in two areas of the Santa Catarina State, southern Brazil, as these

subspecies do not seem to co-occur (Hensen at al., 2018). The specimens of C. rutilans

17 *rutilans* were collected during the summer (end of December 2016) at 1 360 m above sea

18 level in São Joaquim National Park (28°8'28"S, 49°38'13"W), in the municipality of Urubici.

19 The individuals of *C. rutilans cyanescens* were also collected during the summer (December

20 2017 and January 2018) at 250 m a.s.l. in Serra do Tabuleiro State Park (27°44'01"S,

48°48'33"W), in the municipality Santo Amaro da Imperatriz. Although the specimens of the

22 two subspecies were collected in different years for logistical reasons, we assume that the

23 effect of this difference is negligible compared with effects of factors studied in the

24 experiments. Dung beetles were sampled several times until we collected enough individuals

to conduct laboratory experiments. All unnecessary specimens (from other species) were

26 released in the same location where they had been collected. We used pitfall traps baited with

27 dog faeces to collect twice a day (morning and afternoon) the studied individuals because this

bait has proven to be effective in collecting dung beetles (Batilani-Filho & Hernández, 2017).

29 These traps were plastic containers (15 cm diameter, 20 cm deep) buried in the ground up to

30 the rim and closed with a plastic lid containing a hole allowing the entry but preventing the

31 escape of individuals. The traps were protected from rain with a plastic cover approximately

32 10 cm above the container. The traps contained local soil and a piece of dog faeces (ca. 30 g)

in the centre. The faeces were previously obtained from dogs not submitted to any medical

1 treatment and were kept under refrigeration at the Biotério Central (Bioterium) of the

- 2 Universidade Federal de Santa Catarina.
- 3

4 Rearing and maintenance under different temperatures

The beetles were transported to the Laboratory of Terrestrial Animal Ecology 5 (LECOTA/UFSC) and maintained at 26 ± 1 °C and L12:D12 photoperiod. They were stored 6 7 in 5-1 plastic pots with soil from the location and were weekly fed with dog faeces until the beginning of the experiments. The individual of C. rutilans rutilans (Figure 1A) and C. 8 9 rutilans cyanescens (Figure 1B) were individually identified, weighed, sexed, and classified into five age categories (very young, young, mature, old, and very old) according to the tibia 10 degree and clypeus erosion (Tyndale-Biscoe, 1984). Only those specimens classified as young 11 or mature (some tibia spines missing and some erosion in the tarsal spur and clypeus) were 12 13 used in the experiments to guarantee the sexual maturity of the individuals.

Six pairs from each subspecies were allocated per treatment. Each pair is composed of 14 15 specimens with similar body size (heavy, medium, or light) to minimize the possible effect of body size differences on the measured behavioural and developmental variables. Thus, we 16 organized the 30 pairs by weight, with male and female from each pair with similar weight, 17 and then they were distributed randomly among the treatments. Each pair was kept in a clear 18 plastic container (13 cm diameter, 11 cm deep) with 500 g of soil from the location where 19 each subspecies was collected, as suggested by Villalobos et al. (1998). The soil was 20 previously sifted and heated for 90 s in a microwave at maximum power in order to remove 21 22 possible pathogenic bacteria and fungi. The containers had a plastic lid with a hole in the middle covered by a tissue that allowed air entry. During the experiment, the soil in the 23 containers was constantly kept moist by periodically spraying it with water. All pairs were fed 24 weekly with 10 g of dog faeces due to their easy collection in the Bioterium, where the dogs 25 26 are being fed in a standardized and safe way. Further, this type of resource has proven to be a suitable trophic resource for laboratory rearing of dung beetles when compared with the dung 27 28 of several native mammal species (Hernández et al., 2020).

Pairs included in the containers were maintained at five thermal conditions or treatments (15, 20, 25, 30, and 35 °C), and each treatment was repeated 6×; thus, we used a total of 120 individuals (2 subspecies × 2 individuals × 6 repetitions × 5 thermal treatments) and 30 experimental units per subspecies (6 repetitions × 5 thermal treatments). We selected the minimum temperature (15 °C) considering the lowest environmental mean temperature experienced by the active adults of these two subspecies (Hensen et al., 2018). Beginning with

15 °C, treatments increased by 5 °C until a maximum of 35 °C, a value close to the 1 2 temperature in which one can find active adults of C. rutilans rutilans (33.5 °C) and C. rutilans cyanescens (32.3 °C). 15 °C was maintained in the laboratory with air condition, 3 whereas the four higher temperatures were maintained by placing the containers in four 4 independent electric Titã ACL-5810D showcases ($77 \times 36.5 \times 44.5$ cm). Although the 5 experiments for each subspecies were carried out in different years, they started in the same 6 7 period during the summer: for C. rutilans rutilans the experiment started in January 2017 and for C. rutilans cyanescens in February 2018. After the experiment (7 weeks of data), we 8 9 maintained the pairs under the experimental conditions until 90% of the individuals of each subspecies died (around 30 weeks or approximately 4× the experimental period). Thus, the 10 experiment of C. rutilans rutilans ended in November 2017 and for C. rutilans cyanescens in 11 12 June 2018.

13

14 Behavioural activity

15 Photographs of each pair/container were taken the day the pair was fed. The clear plastic container lids were removed and replaced with glass lids. A Canon T3 camera connected to a 16 17 laptop was attached to a tripod to take pictures from above. Both subspecies have a diurnal behaviour and were maintained in the laboratory in a L12:D12 day/night cycle (day from 18 07:00 to 19:00 hours). The camera took pictures every 5 min for 3 h (time lapse), beginning at 19 15:00 and finishing at 18:00 hours. Each pair was photographed only 1 day in the experiment, 20 with 40 photos per pair totalizing 1 200 photos (40 photos/pair \times 6 pairs/temperature \times 5 21 temperature treatments). Because five pairs of C. rutilans cyanescens died before they could 22 be photographed, we only analysed 1 000 photographs for this subspecies. Thus, the data 23 finally analysed came from 30 pairs of C. rutilans rutilans and 25 pairs of C. rutilans 24 25 cyanescens.

Sequential images allow us to detect each one of the movements and behavioural patterns 26 of all the individuals. In total 80 responses per pair were analysed. As there were two 27 28 individuals per container, we registered in each photo what each individual was doing. Thus, we have two behavioural activities per photo except in the case of sexual behaviours. Based 29 30 on these observations we described the behaviour and classified the behavioural patterns into 31 four behavioural categories: sexual (copulation and parental care), locomotion (move on the surface), alimentation (feed and stay on the food ball), and others (buried) (Table 1). The data 32 of each beetle were divided into six behavioural patterns: copulation (COP), parental care 33 34 (PCARE), movement on the surface (MOVE), feeding (FEED), staying on the food ball

- (STAY), and burying (BURY). 1
- 2

Dung ball production 3

The 30 experimental units per subspecies were followed for 8 weeks (7 weeks of data) by 4 5 counting weekly the food and brood dung balls made by each beetle pair. Additionally, we weighed the balls with a precision balance and measure their size (diameter) with a digital 6 7 caliper. Food and brood dung balls can be clearly distinguished due to the 'water drop' shape of brood balls. The mean weight and diameter of the balls made by each pair were used in the 8 9 statistical analyses as individual identification of each ball was not possible (balls can be destroyed by a pair and new balls can be built between observation periods). We consider that 10 the data obtained from each pair are statistically independent because their own behaviour 11 12 does not influence the behaviour of the other pairs (Schank & Koehnle, 2009). 13

Emergence and longevity 14

15 The newly emerged specimens in each container were counted and individuals were weighed

- in order to assess possible differences due to temperature treatments and subspecies identity. 16
- 17 Finally, we calculated the number of weeks elapsed until the death of each individual of each
- 18 subspecies (longevity) to examine whether the average lifetime of the two subspecies differed
- due to temperature treatments. Although longevity will depend on the age of the individuals at 19
- 20 the moment of their capture, we assumed that the random distribution of individuals along the
- temperature experiments minimized the role of this possible bias. 21
- 22

23 **Statistical analysis**

A full factorial ANOVA design in General Linear Models was used to estimate the statistical 24

significance of the dependent variables, including temperature treatments (five levels) and 25

- 26 subspecies identity (two levels) as explanatory factors. The two main effects (temperature and
- subspecies) and their interaction were tested ($\alpha = 0.05$), but if the interaction was statistically 27
- significant, the main effects were ignored in the interpretation of the results. We used Tukey 28
- honestly significant differences (HSD) post-hoc tests after Bonferroni corrections to test for 29
- differences in mean values among factor levels. These calculations were done with the 30
- software R v.3.2.3 and histogram tools ggplot2 (Wickham & Chang, 2016). 31
- 32

Results 33

1 Body weight

- 2 The subspecies identity factor was significant for body weight differences ($F_{1,116} = 6.98$, P =
- 3 0.009): where the individuals of *C. rutilans rutilans* are heavier than those of *C. rutilans*
- 4 *cyanescens* (mean \pm SE = 0.171 \pm 0.039 vs. 0.153 \pm 0.035 g, both n = 60). The sex factor does
- 5 not significantly explain body weight differences ($F_{1,116} = 0.61$, P = 0.44).
- 6

7 Ethogram

- 8 The predominant behaviour observed at all temperature treatments in both subspecies was
- 9 staying buried under the ground (BURY), which was observed in more than half of all
- 10 occasions, and in more than 80% of occasions at 20 °C, independent of subspecies identity
- 11 (Table 2). Individuals moving on the ground (MOVE) or feeding in the dung (FEED) were
- 12 observed in ca. 21 and 8% of the occasions, respectively. The other behavioural patterns
- 13 occurred much less frequently (Table 2). The few statistically significant relationships suggest
- 14 that C. rutilans cyanescens would move more on the soil surface, regardless of temperature,
- and that all the specimens feed more at higher temperatures (Table 2).
- 16

17 Food and brood ball production

- 18 The number of food balls per pair oscillates from 0 to 37 (mean \pm SE = 12.5 \pm 2.0), whereas
- 19 the number of brood balls is lower (6.2 ± 4.4 ; range: 0-24). The number of food balls did not
- 20 differ statistically between the two subspecies or between temperatures (Table S1). However,
- temperature was found to have a significant effect on the number of brood balls (Table S1).
- Tukey's HSD tests indicated that the number of brood balls at 15 °C is lower (P = 0.02) than
- the numbers at higher temperatures (30°C) independently of the subspecies (Figure 2)..

24

25 Weight and size of food and brood balls

- The correlation between the weight and diameter of food and brood balls is very high (both *r* = 0.95, P<0.001). Weight and diameter of food balls differ between temperatures and subspecies (Table S2). Food balls are smaller and lighter for *C. rutilans cyanescens*, as well as in extreme temperatures (15 and 35°C; Tukey's HSD tests: 0.02>P>0.002; Figure 3A,B). For brood balls the temperature*subspecies interaction is always statistically significant (Table S2), indicating that the responses of weight and diameter of the dung balls to temperature
- 32 treatments depends on the subspecies (Figure 3C,D). Interestingly, *C. rutilans rutilans* brood
- balls seem to be bigger and heavier at lower temperatures, whereas those of *C. rutilans*
- 34 *cyanescens* remain practically unchanged along the temperature treatments.

1

2 **Emerged adults and longevity** A total of 28 new adults emerged from the brood balls of C. rutilans rutilans and 15 from C. 3 rutilans cyanescens. The first C. rutilans rutilans emerged during the 7th week at 30 °C 4 5 whereas the last one emerged in the 16th week at 15 °C. The first C. rutilans cyanescens emerged during the 5th week at 30 °C, the last during the 12th week at 25 °C. The number of 6 7 newly emerged individuals did not differ between subspecies, but was highly dependent on temperature (Table S3). In general, the emergence of new individuals increased up to 30 °C, 8 9 and decreased drastically at 35 °C, regardless of the subspecies (Figure 4A). The weight of the newly emerged individuals differed among temperatures and was highly 10 dependent on subspecies (Table S3); new adults of C. rutilans rutilans were heavier than 11 those of C. rutilans cyanescens. Tukey's HSD tests indicated that the emerged individuals 12 13 were significantly heavier at 20 °C than at higher temperatures (Figure 4B). The time required for the emergence of new adults differed between temperature 14 15 treatments (Table S3) but not between subspecies. Thus, new adults need more time to emerge under low temperatures (15 and 20 °C; Tukey's HSD tests: P<0.001; Figure 4C). 16 17 Finally, the mean (\pm SE) longevity of both subspecies is 11.0 ± 1.3 weeks, oscillating from 1 to 30 weeks. Interestingly, the temperature*subspecies interaction is highly significant in 18 explaining the longevity of wild-caught adults (Table S3); C. rutilans rutilans live longer than 19 C. rutilans cyanescens at all the temperatures, except 35 °C (Figure 4D). Thus, the longevity 20 of C. rutilans rutilans decreases with increasing temperature, whereas longevity of C. rutilans 21 22 cyanescens seems to be temperature independent (Figure 4D). 23

24 **Discussion**

Our results indicated that the two subspecies differ in their reproductive strategy depending on 25 the environmental temperature, as well as in their larval development and longevity, matching 26 27 their regional occurrences. We found no significant behavioural differences between the subspecies, perhaps due to the low sample sizes. Below 25 °C, the number of dung balls for 28 29 breeding in C. rutilans cyanescens was low. This is in agreement with this subspecies' 30 distribution – it occurs under warmer conditions (Hensen et al., 2018). Furthermore, at low temperatures the weight and diameter of the brood balls were significantly higher in C. 31 rutilans rutilans than in C. rutilans cyanescens, as were the weight (but not the number) of 32 newly emerged adults and the longevity of parental specimens at the lowest temperature 33

considered. All these results support thermal segregation between the two subspecies,
 reinforcing the likelihood that they could be considered as different species (Hensen et al.
 2018). Considering that genetic isolation is the main factor generating the differences of any
 kind able to classify them as different species (De Queiroz, 2007), further genetic analyses
 may confirm their separation as different species.

6 Our results also corroborate the role of temperature in determining body size, 7 developmental time, or brood ball weight. In ectotherms, temperature affects the rate of 8 development and the age at which an individual reaches its body size (Atkinson & Sibly, 9 1997; Angilletta & Dunham, 2003). Our results show that C. rutilans rutilans adults are larger than those of C. rutilans cyanescens. In a revision, Atkinson (1994) showed that in many 10 cases the body size of insects is reduced with an increase in temperature (examples in Diptera, 11 Hemiptera, Hymenoptera, and Lepidoptera), but this pattern is variable in Coleoptera. Simple 12 13 explanations trying to establish a general temperature-size rule in ectotherms are insufficient, as in many special cases the mechanisms involved may be complex (Atkinson & Sibly, 1997; 14 15 Angilletta & Dunham, 2003).

In addition, the behaviour of the two subspecies may be unequally influenced by 16 17 temperature. Canthon rutilans rutilans adults remained buried in the ground longer than adults of C. rutilans cyanescens, which moved on the ground more. This difference may be 18 related to environmental temperature, as the soil has a temperature-buffering capacity and C. 19 rutilans rutilans, the characteristically 'cold' subspecies, may avoid low temperatures by 20 staying underground (Lobo et al., 2019). Temperature variation did not influence the feeding 21 22 patterns in the two subspecies in contrast to the influence of temperature on reproductive behaviour. The production of brood balls depends on temperature in both subspecies. Canthon 23 rutilans rutilans produce larger and bigger brood balls (more faeces inside) at lower 24 25 temperatures. This may be another adaptation to living at lower temperatures as larvae may 26 need more time to develop under these conditions. Dung beetles can adjust the brood ball size according to dung quality (Kishi, 2014) and adult body size (Hunt & Simmons, 2000, 2002). 27 28 Females of C. rutilans rutilans produced fewer brood balls at lower temperatures than at higher temperatures, but they were bigger. The new adults were heavier at lower 29 30 temperatures. Temperature is also known to influence larval development time: at lower temperatures the larvae develop slower. 31 32 The first individual of both subspecies emerged at the same temperature. However, new

adults of *C. rutilans rutilans* needed 7 weeks to complete their life cycle and new adults of *C. rutilans cyanescens* only 5 weeks. The last individual to emerge was after 4 months at the

lowest temperature for C. rutilans rutilans, and after 3 months at 25 °C for C. rutilans 1 cyanescens, reinforcing the adaptation of C. rutilans rutilans towards cold conditions. Both 2 subspecies produced brood balls at the lowest temperature, although fewer, but only the larvae 3 of C. rutilans rutilans were able to complete their development.. 4 Some studies with C. rutilans cyanescens showed that adults, feeding on dog faeces at 25 5 °C, had a F1 with a larval development cycle that lasts 33 ± 4 days on average (Hernández et 6 al., 2020). The most studied species of this genus is Canthon cyanellus cyanellus LeConte, 7 whose ideal breeding temperature under laboratory conditions has been found to be 27 ± 1 °C 8 in Mexico (Favila et al., 2005). The development time increases considerably at low 9 temperatures in most insects (Ratte, 1984), and there are no differences between the number 10 of larval stages at different temperatures, only the time that they need to develop and to reach 11 the same size changes (Sehnal, 1991). The survival of males from C. rutilans cyanescens 12 13 reaches more than three months (Wuerges & Hernández, 2020), but we observed that temperature did not affect its survival. Nevertheless, the survival of C. rutilans rutilans was 14 15 almost three times less at high temperatures. This result demonstrates the relationship between temperature and mortality due to aging; with an increase in temperature there is greater 16 17 molecular damage, and therefore mortality will increase (Calow, 1978). 18 Differential adaptations to the climatic conditions along elevational gradients can lead to allopatric speciation (Janzen, 1967; Ghalambor et al., 2006; Mittelbach et al., 2007). 19 Currently, C. rutilans rutilans and C. rutilans cyanescens are considered two subspecies due 20 to their adult morphological similarity. A recent study (Hensen et al., 2018) showed that the 21 22 two subspecies are spatially segregated, and also that they inhabit under conditions that differ in environmental temperature. In the current study, we showed that environmental 23 temperature influenced both subspecies differently: C. rutilans rutilans, occurring at high 24 25 elevations, has a more variable response to environmental temperature than C. rutilans 26 cyanescens, which occur under warmer conditions. Thus, our results support that these two taxa may be considered different species. Further genetic analyses would help to confirm this 27 hypothesis. 28

29

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6	
7	
8	Figure captions
9	Figure 1 Adults of Canthon rutilans subspecies: (A) characteristically red C. rutilans rutilans
10	and (B) blue C. rutilans cyanescens.
11	
12	Figure 2 Number of (A) food balls and (B) brood balls made by pairs of Canthon rutilans
13	rutilans (white) and C. rutilans cyanescens (grey) during 7 weeks of laboratory experiments
14	at 15-35 °C. The star and the central line in each box represent the mean and median,
15	respectively. The boxes represent the first and third quartile values, the vertical lines represent
16	the 5th and 95th percentiles, and the dots are outliers.
17	
18	Figure 3 (A, B) Weight and (C, D) diameter of (A, C) food balls and (B, D) brood balls made
19	by pairs of Canthon rutilans rutilans (white) and C. rutilans cyanescens (grey), during 7
20	weeks of laboratory experiments at 15-35 °C. The star and the central line in each box
21	represent the mean and median, respectively. The boxes represent the first and third quartile
22	values, the vertical lines represent the 5th and 95th percentiles, and the dots are outliers
23	
24	Figure 4 (A) Number, (B) weight (g), and (C) time of emergence (weeks) of F1 individuals,
25	and (D) adult longevity (weeks) of wild-caught individuals of Canthon rutilans rutilans
26	(white) and C. rutilans cyanescens (grey) at 15-35 °C. The star and the central line in each
27	box represent the mean and median, respectively. The boxes represent the first and third
28	quartile values, the vertical lines represent the 5th and 95th percentiles, and the dots are
29	outliers In panels B and C, the boxplot at 15 °C belongs to C. rutilans rutilans.
30	
31	
32	
33	
34	

1 Supporting Information

- 2 Additional Supporting Information may be found in the online version of this article.
- 3
- 4 **Table S1** Results from a GLM full factorial design accounting for the number of food and

5 brood balls, including temperature treatments (five levels) and subspecies identity (two levels)

- 6 as explanatory factors. $R2 \times 100\%$ is the percentage of total variability explained by the
- 7 complete model
- 8
- 9 **Table S2** Results from a GLM full factorial design for the weight and diameter of food and
- brood balls, including the treatments of temperature (five levels) and subspecies identity (two brood balls, including the treatments of temperature of tests and subspecies identity (two
- 11 levels) as explanatory factors. $R^2 \times 100\%$ is the percentage of total variability explained by
- 12 the complete model
- 13
- 14 **Table S3** Results from a GLM full factorial design for the number, weight, and emergence
- 15 time of F1 and for adult longevity, including the temperature treatments (five levels) and
- subspecies identity (two levels) as explanatory factors. $R^2 \times 100\%$ is the percentage of total
- 17 variability explained by the complete model



1 cm

- 2

Figure 1









-all lettering in figures should be ARIAL

- 5 -move the text labels along the vertical axes a bit away from the axes please
- 6 -remove 'Temperature (°C)' from the top panel
- 7 -remove the legend <info is already in the caption>
- 8 Put caps on the whiskers, about half the width of the box



- 3 -all lettering in figures should be ARIAL
- 4 -move the text labels along the vertical axes a bit away from the axes please
- 5 -remove 'Temperature (°C)' from the top two panels; move the lower two panels a bit up
- 6 -remove the four legends <info is already in the caption>
- 7 Put caps on the whiskers, about half the width of the box
- 8



- 3 -all lettering in figures should be ARIAL
- 4 -move the text labels along the vertical axes a bit away from the axes please
- 5 -remove 'Temperature (°C)' from the top two panels; move the lower two panels a bit up
- 6 -remove the four legends <info is already in the caption>
- 7
- 8 Put caps on the whiskers, about half the width of the box

Table 1 Ethogram of behavioural categories observed in *Canthon rutilans* under laboratory

2 conditions

Behavioural	Behavioural patterns	Description of behaviour
categories		
Sexual	Copulation (COP)	Male adheres to the female in the dorsal part
	Parental care (PCARE)	Individual is above the brood ball, dislocates/rolls the
		brood ball, buries the brood ball
Locomotion	Move on the surface (MOVE)	Individual walks along the side of the case, tries to climb
		on the case wall, is on the soil surface
Alimentation	Feed (FEED)	Individual feeds on faeces or food balls
	Stay on the food ball (STAY)	Individual is on the food ball
Others	Buried (BURY)	Individual is hidden in the soil

1 Table 2 Behavioural patterns (% of time spent) in pairs of the two subspecies of *Canthon*

Subspecies	T (°C)	СОР	PCARE	MOVE	FEED	STAY	BURY
C. rutilans rutilans	35	6.67	3.33	19.17	19.58	4.37	46.88
	30	0	4.37	10.00	12.92	14.38	58.33
	25	0	12.92	8.54	3.54	6.25	68.75
	20	0	0	11.67	5.21	0	83.12
	15	0.83	0.21	21.88	2.92	10.83	63.33
C. rutilans cyanescens	35	0	0.25	21.50	15.00	8.25	55.00
	30	3.50	15.00	31.50	5.25	18.00	26.75
	25	0	14.25	32.25	5.75	0	47.75
	20	0	0	13.12	0.63	0.31	85.94
	15	0.83	0	42.71	5.83	0	50.63
Temperature (F _{4,45})		1.27 ns	2.29 ns	1.94 ns	3.40*	2.11 ns	4.11**
Subspecies (F _{1,45})		0.33 ns	0.22 ns	9.17**	0.68 ns	0.25 ns	2.65 ns
Temperature*subspecies (F _{4,45})		2.25 ns	0.40 ns	1.13 ns	0.56 ns	0.65 ns	1.20 ns

2 *rutilans* in the laboratory at 15-35 °C and results of two-way-ANOVA

3 COP, copulation; PCARE, parental care; MOVE, movement on the surface; FEED, feeding;

4 STAY, staying on the food ball; BURY, burying in the soil.

5 *0.01<P≤0.05; **P≤0.01.

1 Supporting Information

- 2 Table S1 Results from a GLM full factorial design accounting for the number of food and
- 3 brood balls, including temperature treatments (five levels) and subspecies identity (two levels)
- 4 as explanatory factors. $R^2 \times 100\%$ is the percentage of total variability explained by the
- 5 complete model

	Temperature	Subspecies	Temperature*subspecies	$R^2 \times 100\%$
No. food balls	$F_{4,50} = 1.58$	$F_{1,50} = 0.27$	$F_{4,50} = 1.04$	17.7
	P = 0.19	P = 0.61	P = 0.40	
No. brood balls	$F_{4,50} = 2.92$	$F_{1,50} = 1.93$	$F_{4,50} = 2.48$	32.0
	P = 0.03	P = 0.17	P = 0.06	

- 1 **Table S2** Results from a GLM full factorial design for the weight and diameter of food and
- 2 brood balls, including the treatments of temperature (five levels) and subspecies identity (two
- 3 levels) as explanatory factors. $R^2 \times 100\%$ is the percentage of total variability explained by
- 4 the complete model

	Temperature	Subspecies	Temperature*subspecies	R ² ×100%
Weight food balls	$F_{4,48} = 7.90$	$F_{1,48} = 27.73$	$F_{4,48} = 1.59$	57.8
	P<0.001	P<0.001	P = 0.19	
Weight brood balls	$F_{4,30} = 15.14$	$F_{1,30} = 30.78$	$F_{4,30} = 3.01$	77.5
	P<0.001	P<0.001	P = 0.03	
Diameter food balls	$F_{4,48} = 5.95$	$F_{1,48} = 38.50$	$F_{4,48} = 2.36$	59.9
	P<0.001	P<0.001	P = 0.07	
Diameter brood balls	$F_{4,30} = 15.35$	$F_{1,30} = 33.33$	$F_{4,30} = 2.80$	77.9
	P<0.001	P<0.001	P = 0.04	

- 1 Table S3 Results from a GLM full factorial design for the number, weight, and emergence
- 2 time of F1 and for adult longevity, including the temperature treatments (five levels) and
- subspecies identity (two levels) as explanatory factors. $R^2 \times 100\%$ is the percentage of total

Parameters	Temperature	Subspecies	Temperature*subspecies	$R^2 \times 100\%$
No. F1	$F_{4,50} = 4.62$	$F_{1,50} = 2.39$	$F_{4,50} = 1.01$	33.3
	P = 0.003	P = 0.13	P = 0.41	
Weight of F1	$F_{3,35} = 4.49$	$F_{1,35} = 30.09$	$F_{2,35} = 1.33$	56.9
	P = 0.009	P<0.001	P = 0.28	
Time emergence of F1	$F_{3,36} = 17.70$	$F_{1,36} = 3.32$	$F_{2,36} = 1.22$	62.0
	P<0.001	P = 0.08	P = 0.31	
Longevity	$F_{4,101} = 7.33$	$F_{1,101} = 181.72$	$F_{4,101} = 7.01$	70.3
	P<0.001	P<0.001	P<0.001	

4 variability explained by the complete model