

Re-establishment of the genus *Pseudalbizzia* (Leguminosae, Caesalpinioideae, mimosoid clade): the New World species formerly placed in *Albizia*

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Abstract

Following recent mimosoid phylogenetic and phylogenomic studies demonstrating the non-monophyly of the genus *Albizia*, we present a new molecular phylogeny focused on the neotropical species in the genus, with much denser taxon sampling than previous studies. Our aims were to test the monophyly of the neotropical section *Arthrosamanea*, resolve species relationships, and gain insights into the evolution

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of fruit morphology. We perform a Bayesian phylogenetic analysis of sequences of nuclear internal and external transcribed spacer regions and trace the evolution of fruit dehiscence and lomentiform pods. Our results find further support for the non-monophyly of the genus *Albizia*, and confirm the previously proposed segregation of *Hesperalbizia*, *Hydrochorea*, *Balizia* and *Pseudosamanea*. All species that were sampled from section *Arthrosamanea* form a clade that is sister to a clade composed of *Jupunba*, *Punjuba*, *Balizia* and *Hydrochorea*. We find that lomentiform fruits are independently derived from indehiscent septate fruits in both *Hydrochorea* and section *Arthrosamanea*. Our results show that morphological adaptations to hydrochory, associated with shifts into seasonally flooded habitats, have occurred several times independently in different geographic areas and different lineages within the ingoid clade. This suggests that environmental conditions have likely played a key role in the evolution of fruit types in *Albizia* and related genera. We resurrect the name *Pseudalbizia* to accommodate the species of section *Arthrosamanea*, except for two species that were not sampled here but have been shown in other studies to be more closely related to other ingoid genera and we restrict the name *Albizia* s.s. to the species from Africa, Madagascar, Asia, Australia, and the Pacific. Twenty-one new nomenclatural combinations in *Pseudalbizia* are proposed, including 16 species and 5 infraspecific varietal names. In addition to the type species *Pseudalbizia berteriana*, the genus has 17 species distributed across tropical regions of the Americas, including the Caribbean. Finally, a new infrageneric classification into five sections is proposed and a distribution map of the species of *Pseudalbizia* is presented.

Keywords

Arthrosamanea, hydrochory, monophyly, Neotropics, phylogeny, taxonomy

Introduction

The genus *Albizia* Durazz. has a complicated taxonomic history but has generally been treated as a pantropical genus with 120–140 species, of which 36 are endemic to Africa, with c. 30 species in Madagascar, of which c. 24 are endemic, c. 35 species in Asia, one in Australia, and 22 in tropical America (Lewis and Rico Arce 2005; Rico Arce et al. 2008). All species are woody, forming trees of variable stature and inhabit a wide range of lowland tropical biomes (Figs 1 and 2), including rain forests, seasonally dry tropical forests, and savannas, with one species, *Albizia julibrissin* Durazz., the type species of the genus, in subtropical and warm temperate forests in Asia. However, *Albizia* remains poorly defined; its delimitation remains one of the most challenging taxonomic problems in the legume family, and it is currently considered the main “dustbin” genus in tribe Ingeae (Koenen et al. 2020). In the past, the most problematic genus of tribe Ingeae was *Pithecellobium* Mart., but its taxonomy has been gradually clarified (Barneby and Grimes 1996). Resolution of the taxonomic status of *Albizia* has lagged behind that of *Pithecellobium* and only really started at the end of the twentieth century. For example, several new neotropical genera have been segregated from *Albizia*: *Balizia* Barneby & J.W. Grimes; *Hesperalbizia* Barneby & J.W. Grimes, and *Hydrochorea* Barneby & J.W. Grimes. Barneby and Grimes (1996) also re-established the genus *Pseudosamanea* Harms, which previously had been treated as a synonym within *Albizia* (Table 1). However, at the time they were established, the monophyly of these new and re-established genera had not been tested using phylogenetic analyses of molecular data.

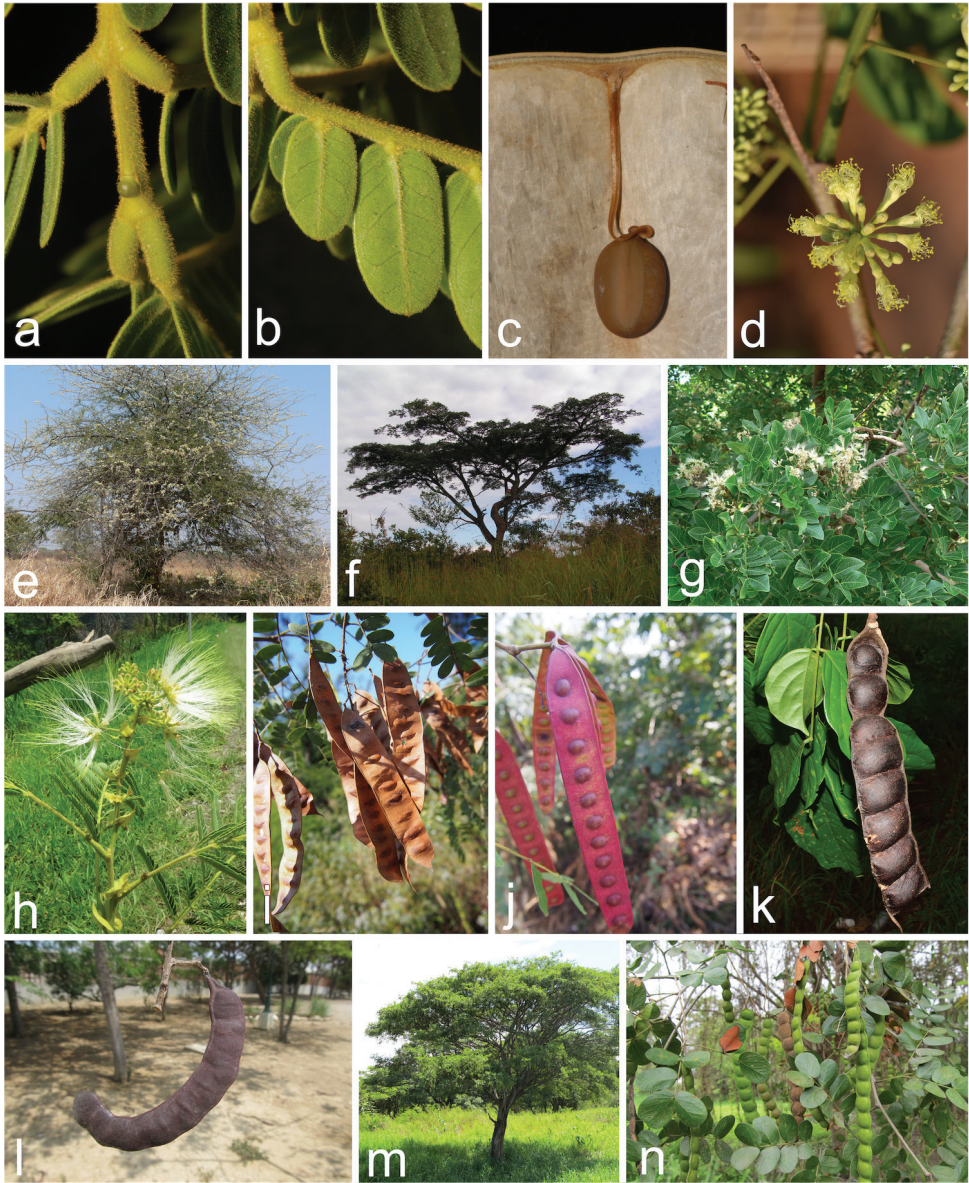


Figure 1. Morphology of *Albizia* s.l. showing selected members of the genera *Albizia* and *Pseudalbizzia* **a–c** *Albizia ferruginea* (Guill. & Perr.) Benth. in Congo **a** detail of leaf rachis and gland between terminal pinnae **b** detail of leaflets of a terminal pinna **c** seed and funiculus attached to the valve **d** *Albizia glaberrima* (Schumach. & Thonn.) Benth. in Malawi, detail of inflorescence **e** *Albizia anthelmintica* Brongn. in Malawi, habit **f** *Albizia adiantifolia* (Schumach.) W. Wight in Congo, habit **g** *Albizia glaberrima* in Malawi, branches and inflorescences **h** *Albizia chinensis* (Osbeck) Merr. in Thailand, inflorescences **i** *Albizia odoratissima* (L. f.) Benth. in Thailand, fruits **j** *Albizia procera* (Roxb.) Benth. in Thailand, fruits **k** *Albizia splendens* Miq. in Thailand, woody fruit **l** *Pseudalbizzia multiflora* var. *multiflora* in Ecuador, woody fruit **m, n** *Pseudalbizzia pistaciifolia* (Willd.) E.J.M Koenig & Duno in Ecuador **m** habit **n** woody fruit. Photos: **a, b** David J. Harris / With permission from RBG Edinburgh **c** Claude Boucher Chisale **d–f** Günter Baumann **g** Jos Stevens **h** Natcha Sutjaritjai **i–k** Prateep Panyadee **l–n** Xavier Cornejo.

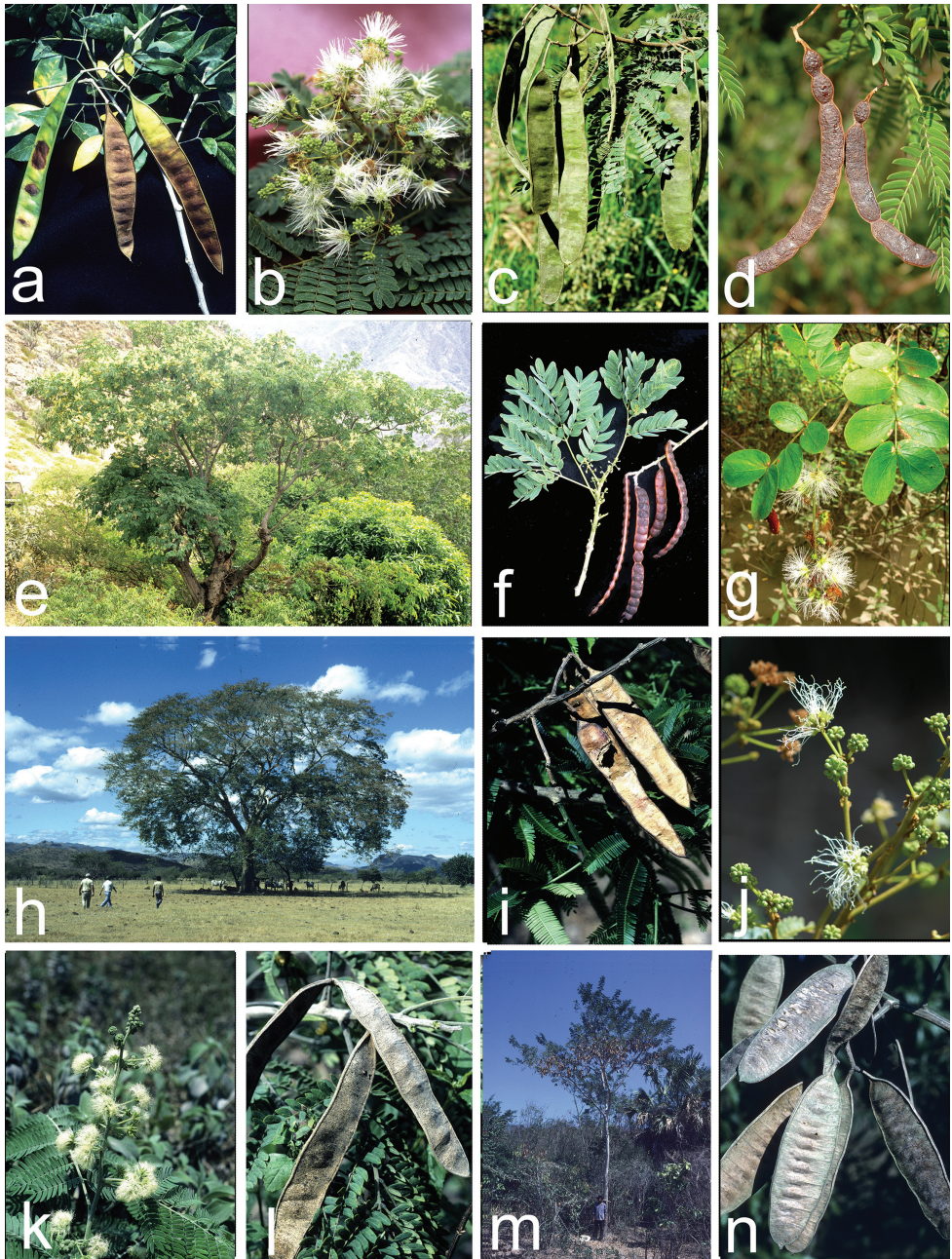


Figure 2. Habit, flower and fruit variation in the genus *Pseudalbizzia* **a** *P. adinocephala* pods (Hughes 1913) **b** *P. coripatensis* inflorescence (Hughes 2433) **c** *P. coripatensis* pods (Hughes 2433) **d** *P. inundata* pods (JRI Wood 26530) **e** *P. multiflora* habit (Hughes 2214) **f** *P. multiflora* leaves and pods (Hughes 2214) **g** *P. pistaciifolia* leaves and inflorescence (Cornejo 8426, GUAY) **h** *P. niopoides* habit (Hughes 419) **i** *P. niopoides* pods (Rivera 2245) **j** *P. polycephala* inflorescence (de Queiroz 15515) **k** *P. tomentosa* inflorescence (Hughes 1143) **l** *P. sinaloensis* pods (Hughes 1576) **m** *P. tomentosa* habit (Hughes 1335) **n** *P. tomentosa* pods (Hughes 1307). All photos by Colin Hughes except **g**, Xavier Cornejo.

Table 1. Main taxonomic changes related to *Albizzia*, Ingeae tribe [1981–2008]. Modified from Rico Arce et al. (2008).

Nielsen (1981)	Barneby and Grimes (1996)	Lewis and Rico Arce (2005)	Rico Arce et al. (2008)	Iganci et al. 2015
<i>Albizzia</i>	<i>Albizzia</i>	<i>Albizzia</i>	<i>Albizzia</i>	<i>Albizzia</i>
	<i>Balizzia</i>			<i>Balizzia</i>
<i>Cathormion</i>	<i>Cathormion</i>	<i>Cathormion</i>	*	*
	<i>Hydrochorea</i>		*	<i>Hydrochorea</i>
	<i>Hesperalbizzia</i>	<i>Hesperalbizzia</i>		
	<i>Pseudosamanea</i>	<i>Pseudosamanea</i>		

* Not explicitly mentioned in the study.

Barneby and Grimes (1996) placed the remaining New World species of *Albizzia* in their section *Arthrosamanea* (Britton & Rose) Barneby & J.W. Grimes. They characterized this section as forming a group that is homogeneous in most respects, but diverse in the late developmental stages of the fruit, which vary in: 1) fruit opening type: dehiscent, indehiscent, or breaking, 2) lateral shape: flat to conspicuously raised above the seed chambers, 3) texture and consistency of the valves: papery, chartaceous or woody, 4) longitudinal shape: straight to weakly falcate (Barneby and Grimes 1996) (Figs 1, 2 and 4). Within section *Arthrosamanea*, four series were proposed by Barneby and Grimes (1996): series *Paniculatae* with papery, plano-compressed, inertly dehiscent pods with continuous valves (13 species); series *Arthrosamanea* comprising 3 species with lomentiform, plano-compressed pods, where the ripe valves crack transversely between seeds but the wiry sutures persist at maturity; series *Multiflorae* (2 species) characterized by lomentiform fruits only reluctantly separating into articles, the thick-textured valves and sutural keels breaking transversely under pressure; and the monospecific series *Inundatae* which bears crypto-lomentiform pods, dehiscent through the sutures and with the valves differentiating into a continuous exocarp and a segmented endocarp separating into 1-seeded segments (Barneby and Grimes 1996).

Series *Paniculatae* is widespread across Mexico, Central and South America, occurring mainly in seasonally dry forests, grasslands, and less often in humid forests (in South America). All species of series *Paniculatae* have papyraceous, dehiscent fruits with one exception, *A. berteriana* (DC.) Fawc. & Rendle (the earlier combination *A. berteriana* (DC.) M. Gómez was invalidly published due to incorrect citation of the basionym, see Barneby and Grimes 1996), whose fruits are indehiscent and fall to the ground entire. In contrast, the other three series are distributed from Panama to South America and are most diverse in the Amazon basin (Barneby and Grimes 1996), and usually have more or less woody fruits, which are articulated and indehiscent, some dividing into monospermous segments through the grooves of the valves, considered to be an adaptation for hydrochory, i.e., seed dispersal in riparian and seasonally inundated forests (e.g., *A. inundata* (Mart.) Barneby & J.W. Grimes, *A. pistaciifolia* (Willd.) Barneby & J.W. Grimes, and *A. subdimidiata* (Splitg.) Barneby & J.W. Grimes).

The segregate genera established by Barneby and Grimes (1996) have not been universally accepted. For example, in the most recent taxonomic treatment of *Albizzia* for Mexico and Central America (Rico Arce et al. 2008), the genera *Balizzia*, *Hesperalbizzia*,

and *Pseudosamanea* were not recognized (Table 1). However, subsequent phylogenetic analyses have confirmed that these genera were rightfully segregated as distinct evolutionary lineages, *Hesperalbizia* being more closely related to *Lysiloma* Benth. (Duno de Stefano et al. 2021), and the closely related *Balizia* and *Hydrochorea* (the former reduced to synonymy of the latter, Soares et al. 2022) being placed as the sister-group of *Jupunba* Britton & Rose (Iganci et al. 2015; Soares et al. 2021, 2022). Phylogenomic analysis of the mimosoid clade, based on DNA sequences of 964 targeted nuclear genes confirmed these findings and, furthermore, showed that the species from the Americas (i.e., sect. *Arthrosamanea*) form a separate lineage from the African, Madagascan and Asian species (Koenen et al. 2020). Koenen et al. (2020) also showed that *Pseudosamanea*, although difficult to place in any clade, is not closely related to either *Albizia* s.s. or section *Arthrosamanea* and is perhaps most closely related to *Samanea* Merr. and *Chloroleucon* Britton & Rose ex Record.

While the data of Koenen et al. (2020) provided a robust phylogenomic backbone for the mimosoid and ingoid clades, and clearly demonstrated the non-monophyly of *Albizia* by sampling 25 species of that genus, only three of the 19 species from Central and South American sect. *Arthrosamanea* were included in that study, leaving doubts about the monophyly of that section and whether it should be segregated under the circumscription of Barneby and Grimes (1996), or whether there are further potential segregates, given the possibility that some of these species are more closely related to other neotropical genera. That possibility was suggested by the occurrence of lomentiform fruits in some species of section *Arthrosamanea* as reflected in the classification into separate series by Barneby and Grimes (1996). Similar lomentiform fruits also occur in *Hydrochorea* (Barneby and Grimes 1996; Soares et al. 2022) and *Albizia* s.s. (*Albizia dolichadena* (Kosterm.) I.C. Nielsen, *Albizia moniliformis* (DC.) F. Muell., *Albizia rosulata* (Kosterm.) I.C. Nielsen and *Albizia umbellata* (Vahl) E.J.M. Koenen), as well as a few other ingoid lineages (Barneby and Grimes 1996: 204; Koenen 2022a). For this reason, several species of neotropical *Albizia* have homotypic synonyms in *Arthrosamanea* Britton & Rose, *Samanea* or *Cathormion* Hassk., genera which previously had been recognized and defined based mainly on characters of fruit texture and dehiscence (Barneby and Grimes 1996). Barneby and Grimes (1996: 204) considered these fruit types to have arisen multiple times in parallel in different genera, and this was confirmed by subsequent phylogenetic (Iganci et al. 2015; Soares et al. 2021) and phylogenomic studies (Koenen et al. 2020), although the neotropical lomentiform *Albizia* species were not included in these studies, or remained unresolved.

Here we investigate whether *Albizia* sect. *Arthrosamanea* is monophyletic and thereby provide a more rigorous basis for recognizing its evolutionary distinctiveness from *Albizia* s.s. as a segregate genus. We infer a new phylogeny with emphasis on the neotropical species and make use of further insights offered by the phylogenomic analysis of Ringelberg et al. (2022). In addition, we use a tree topology inferred from data from the latter study to evaluate whether lomentiform fruits in *Albizia* sect. *Arthrosamanea* are independently derived from other lineages in which this fruit type occurs.

Based on our phylogenetic results and the recent findings of Koenen et al. (2020) and Ringelberg et al. (2022), we update the taxonomy of neotropical *Albizzia* by resurrecting the genus *Pseudalbizzia* Britton & Rose.

Materials and methods

We used the nuclear ribosomal External and Internal Transcribed Spacer (ETS and ITS) regions that previously been used to study sister-group relationships within tribe Ingeae (Brown et al. 2008; Iganci et al. 2015; Souza et al. 2016). Our combined dataset included 123 accessions, of which 50 are from Genbank and 73 are newly sequenced here, including 25 species of *Albizzia* s.l. sequenced for the first time. The outgroup, *Vachellia farnesiana* (L.) Wight & Arn., was designated to root the tree (Table 2). The plastid *trnK* region was initially explored but preliminary analyses suggested it is not sufficiently phylogenetically informative and these data were excluded from this study.

Fresh leaf material collected in the field plus herbarium material from the Jardín Botánico Regional Roger Orellana (CICY) were used for DNA extraction. Herbarium specimens used in these analyses came from AAU, CICY, FCME, MA, MEXU, and MO (acronyms as in Thiers 2016). Additional sequences were downloaded from GenBank (Table 2).

DNA from leaf fragments was obtained using the DNeasy Plant Mini Kit (QIAGEN Inc., Valencia, California) following the manufacturer's specifications. To assess concentration and relative quality of DNA, 3 μl of final volume plus 2 μl loading buffer were run for 30 minutes at 6 V cm^{-1} on a 1% agarose gel prepared with 0.5 \times TBE. The resulting gel was developed by immersion for 20–30 minutes in a 0.1 $\mu\text{g ml}^{-1}$ ethidium bromide solution and later observed in a DigiDoc-It Imaging System (version 6.7.1; UVP, Inc., Cambridge, UK) transilluminator. DNA purity and concentration were quantified with a NanoDrop 2000c. Afterwards, DNA samples were standardized to 10 $\text{ng } \mu\text{l}^{-1}$.

PCR amplifications were performed in an Applied Biosystems Veriti 96 Well Thermal Cycler. Volumes of reagents and conditions for the amplifications were as follows: **ITS**: 30 μL of mix containing 3 μl 10 \times Buffer, 2.5 μl MgCl_2 , 0.6 μl (\sim 10 ng) primer, 4 μl Q solution, 1 μl 1.25 mM L^{-1} dNTP, 0.2 μl (1 U) TAQ polymerase, 2 μl (\sim 10 ng) DNA, then completed to volume (approx. 16.1 μl) with ultra-pure water. PCRs were conducted under the following protocol: 94 $^{\circ}\text{C} \times 3 \text{ min}$ + 30 cycles (94 $^{\circ}\text{C} \times 1 \text{ min}$ + 60.5 $^{\circ}\text{C} \times 1 \text{ min}$ + 72 $^{\circ}\text{C} \times 2 \text{ min}$) + 72 $^{\circ}\text{C} \times 7 \text{ min}$. Primers were S3 (AACCTGCGGAAGGATCATTG) (Käss and Wink 1997), and 26S (TAGAATTCCTCCG-GTTCGCTCGCCGTTAC) (Sun et al. 1994). **ETS**: 30 μl of mix containing 3 μl 10 \times Buffer, 2.5 μl MgCl_2 , 0.6 μl (\sim 10 ng) primer, 4 μl Q solution, 1 μl 1.25 mM l^{-1} dNTP, 0.2 μl (1 U) TAQ polymerase, 2 μl (\sim 10 ng) DNA, then completed to volume (approx. 16.1 μl) with ultra-pure water. PCR amplifications were conducted under the following protocol: 94 $^{\circ}\text{C} \times 3 \text{ min}$ + 30 cycles (94 $^{\circ}\text{C} \times 1 \text{ min}$ + 60.5 $^{\circ}\text{C} \times 1 \text{ min}$ + 72 $^{\circ}\text{C} \times 2 \text{ min}$) + 72 $^{\circ}\text{C} \times 7 \text{ min}$. Primers used were 18S-IGS (5'-GAGACAAGCAT-ATGACTACTGGCAGGATCAACCAG-3') and 26S-IGS (5'-GGATTGTTACC-CACCAATAGGGAACGTGAGCTG-3') (Baldwin and Markos 1998).

Table 2. Voucher information of taxa included in the phylogenetic analysis with their corresponding GenBank accession numbers.

Accessions ITS	Accessions ETS
<i>Albizia adianthifolia</i> (Schumach.) W. Wight, MW699934, BGRO 001	<i>Albizia adianthifolia</i> , MW699372, BGRO 001
<i>Albizia amara</i> (Roxb.) Boivin, MW699936, BGRO 003	<i>Albizia amara</i> , MW699374, BGRO 003
<i>Albizia anthelmintica</i> Brongn., MW699937, BGRO 004	<i>Albizia anthelmintica</i> , MW699375, BGRO 004
<i>Albizia arenicola</i> R. Vig., MW699938, R. Randrianaivo 642, MO	<i>Albizia antunesiana</i> Harms, MW699376, S.H.C.P. 966, MO
<i>Albizia brevifolia</i> Schinz, MW699940, BGRO 005	<i>Albizia arenicola</i> , MW699377, R. Randrianaivo 642, MO
<i>Albizia glaberrima</i> Hutch. & Dalziel, MW699943, R.E. Gereau 6203, MA	<i>Albizia brevifolia</i> , MW699378, BGRO 005
<i>Albizia gummifera</i> (J.F. Gmel.) C.A. Sm., MW699944, J.E. Lawesson 5094, AAU	<i>Albizia chinensis</i> (Osbeck) Merr., MW69379, A. Ntemi & A. Athumani 478, MO
<i>Albizia harveyi</i> E. Fourn., MW699945, BGRO 006	<i>Albizia crasiramea</i> Lace, MW699380, K. Larsen et al. 46378, AAU
<i>Albizia julibrissin</i> Durazz., MW699946, BGRO 007	<i>Albizia ferruginea</i> (Guill. & Perr.) Benth., MW699382, C.H. Jongkind 2098, MA
<i>Albizia kalkora</i> (Roxb.) Prain, MW699947, E. Boufford 26356, MO	<i>Albizia glaberrima</i> , MW699383, R.E. Gereau 6203, MA
<i>Albizia lebbeck</i> (L.) Benth., MW699948, C. Chan 7539, CICY	<i>Albizia gummifera</i> , MW699384, J.E. Lawesson 5094, AAU
<i>Albizia petersiana</i> (Bolle) Oliv., MW699950, BGRO 008	<i>Albizia harveyi</i> , MW699385, BGRO 006
<i>Albizia procera</i> (Roxb.) Benth., MW699953, BGRO 009	<i>Albizia julibrissin</i> , MW699387, BGRO 007
<i>Albizia retusa</i> Benth., MW699954, K. Yasuda 1804, MO	<i>Albizia kalkora</i> , MW699388, E. Boufford 26356, MO
<i>Albizia tanganyicensis</i> Baker f., MW699956, BGRO 010	<i>Albizia lebbeck</i> (L.) Benth., MW699389, C. Chan 7539, CICY
<i>Albizia umbellata</i> (Vahl) E. J. M. Koenen, EF638182.1	<i>Albizia lebbekoides</i> (DC.) Benth., MW699390, H. Balslev 9333, AAU
<i>Balizia leucocalyx</i> (Britton & Rose) Barneby & J.W. Grimes, MW699959, S. Aguilar & F. Aguilar 1833, M	<i>Albizia lucidior</i> (Steud.) I.C. Nielsen ex H. Hara, MW699391, J.F. Maxwell 95–259, MO
<i>Lysiloma acapulcense</i> (Kunth) Benth., MW699960, H. Gómez D. 2003, MO	<i>Albizia petersiana</i> (Bolle) Oliv., MW699394, BGRO 008
<i>Lysiloma latisiliquum</i> (L.) Benth., MW699961, P. Simá 2287, CICY	<i>Albizia procera</i> , MW699396, BGRO 009
<i>Pseudalbizia adinocephala</i> (Donn. Sm.) E.J.M. Koenen & Duno, MW699935, BGRO 002	<i>Albizia retusa</i> , MW699397, K. Yasuda 1804, MO
MW699958, J.L. Linares 5406, FCME	<i>Albizia sabafariensis</i> Capuron, MW699398, R. Randrianaivo et al. 1387, MO
<i>Pseudalbizia berteriana</i> (Balb. ex DC.) Britton & Rose, MW699939, A. Jimenez 2113, MO	<i>Albizia tanganyicensis</i> , MW699400, BGRO 010
<i>Pseudalbizia edwallii</i> (Hoehne) E.J.M. Koenen & Duno, MW699942, J.M. Silva & L.M. Abe 4237, MEXU	<i>Albizia umbellata</i> , EF638157.1
<i>Pseudalbizia multiflora</i> (Kunth) E.J.M. Koenen & Duno, MW699949, X. Cornejo 1922, GUAY	<i>Balizia leucocalyx</i> , MW699403, S. Aguilar & F. Aguilar 1833, M
<i>Pseudalbizia pistaciifolia</i> (Willd.) E.J.M. Koenen & Duno, MW699951, X. Cornejo 5323, GUAY	<i>Balizia pedicellaris</i> (DC.) Barneby & J.W. Grimes, MW699404, P.R. House 1880, MA
<i>Pseudalbizia polycephala</i> (Benth.) E.J.M. Koenen & Duno, MW699952, L.P. Queiroz 9578, MEXU	<i>Havardia mexicana</i> , MW699405, S. Foldi s.n., CICY
<i>Pseudalbizia sinaloensis</i> (Britton & Rose) E.J.M. Koenen & Duno, MW699955, C.E. Hughes et al. 1576, FCME	<i>Hesperalbizia occidentalis</i> , MW699406, J.G. Hernandez Oria 21, FCME
<i>Pseudalbizia tomentosa</i> (Micheli) E.J.M. Koenen & Duno, MW699957, A. Dorantes et al. 165, CICY	<i>Lysiloma acapulcense</i> , MW699407, H. Gómez D. 2003, MO
<i>Pseudosamanea cubana</i> (Britton & P. Wilson) Barneby & J.W. Grimes, MW699941, GHBG 001	<i>Lysiloma latisiliquum</i> , MW699408, P. Simá 2287, CICY
<i>Pseudosamanea guachapele</i> (Kunth) Harms, MW699962, BGRO 011	<i>Paraserianthes lophantha</i> , MW699409, H. Balslev et al. 62450, AAU
<i>Zapoteca formosa</i> (Kunth) H.M. Hern., MW699963, R. Duno s.n. CICY. Additional accessions (ITS): <i>Acacia acadenia</i> F.Muell., AF487765.1	<i>Pithecellobium diversifolium</i> , MW699410, J.F.B. Pastore & R.M. Harley 2599 MO
<i>Acacia longifolia</i> (Andrews) Willd., HM007655.1	<i>Pithecellobium excelsum</i> , MW699411, G. P. Lewis et al. 2339, MO
<i>Acaciella angustissima</i> (Mill.) Britton & Rose, EF638169.1	<i>Pseudalbizia adinocephala</i> , MW699373, BGRO 002
<i>Balizia pedicellaris</i> (DC.) Barneby & J.W. Grimes, JX870657.1	MW699402, J.L. Linares 5406, FCME
<i>Calliandra dysantha</i> Benth., JX870684.1	<i>Pseudalbizia edwallii</i> , MW699381, J.M. Silva & L.M. Abe 4237, MEXU

Accessions ITS	Accessions ETS
<i>Calliandra foliosa</i> Benth., EF638181.1	<i>Pseudalbizzia inundata</i> (Mart.) E.J.M. Koenen & Duno, MW699386, H. Balslev et al. 97355, AAU
<i>Cojoba arborea</i> (L.) Britton & Rose, JX870758.1	<i>Pseudalbizzia multiflora</i> (Kunth) E.J.M. Koenen & Duno, MW699392, X. Cornejo & T. Andres 8705, GUAY
<i>Cojoba undulatmarginata</i> L. Rico, EF638187.1	<i>Pseudalbizzia niopoides</i> (Spruce ex Benth.) E.J.M. Koenen & Duno, MW699393, J.R. Grande 374, VEN
<i>Ebenopsis ebano</i> (Berland.) Barneby & J.W. Grimes, JX870759.1	<i>Pseudalbizzia polycephala</i> MW699395, L.P. Queiroz 9578, MEXU
<i>Enterolobium contortisiliquum</i> (Vell.) Morong, EF638190.1	<i>Pseudalbizzia sinaloensis</i> , MW699399, C.E. Hughes et al. 1576, FCME
<i>Enterolobium cyclocarpum</i> (Jacq.) Griseb., EF638191.1	<i>Pseudalbizzia tomentosa</i> , MW699401, A. Dorantes et al. 165, CICY
<i>Enterolobium timbouva</i> Mart., JX870760.1	<i>Pseudosamanea cubana</i> (Britton & P. Wilson) Barneby & J.W. Grimes, MW699412, BJ FTGH 2000
<i>Faidherbia albida</i> (Delile) A. Chev., EU812008.1	<i>Pseudosamanea guachapele</i> , MW699413, BGRO 011
<i>Havardia mexicana</i> (Rose) Britton & Rose, JX870762.1	<i>Samanea tubulosa</i> (Benth.) Barneby & J.W. Grimes, MW699414, G.A. Parada & V.D. Rojas 2480, MO. Additional accessions: <i>Acacia acradenia</i> , EF638116.1
<i>Havardia pallens</i> (Benth.) Britton & Rose, KF921656.1	<i>Acacia longifolia</i> , EF638115.1
<i>Hesperalbizia occidentalis</i> (Brandegee) Barneby & J.W. Grimes, EF638195.1	<i>Acaciella angustissima</i> EF638082.1
<i>Hyrochorea corymbosa</i> (Rich.) Barneby & J.W. Grimes, JX870763.1	<i>Pseudalbizzia adinocephala</i> EF638144.1
<i>Jupunba trapezifolia</i> (Vahl.) Moldenke, EF638166.1	<i>Albizia kalkora</i> EF638158.1
<i>Mariosousa coulteri</i> (Benth.) Seigler & Ebinger, EF638198.1	<i>Albizia lebbeck</i> EF638155.1
<i>Mariosousa dolichostachya</i> (S.F. Blake) Seigler & Ebinger, EF638199.1	<i>Albizia saponaria</i> (Lour.) Blume, EF638085.1
<i>Paraserianthes lophantha</i> (Willd.) I.C. Nielsen, EF638204.1	<i>Archidendropsis basaltica</i> (F. Muell.) I.C. Nielsen, EF638141.1
<i>Pithecellobium diversifolium</i> Benth., JX870768.1	<i>Archidendropsis thozetiana</i> (F. Muell.) I.C. Nielsen, EF638140.1
<i>Pithecellobium dulce</i> (Roxb.) Benth., EF638207.1	<i>Calliandra dysantha</i> EF638121.1
<i>Pithecellobium excelsum</i> (Kunth) Mart., EF638208.1	<i>Calliandra foliosa</i> EF638122.1
<i>Samanea saman</i> (Jacq.) Merr., JX870770.1	<i>Cojoba arborea</i> EF638095.1
<i>Samanea tubulosa</i> (Benth.) Barneby & J.W. Grimes, EF638212.1	<i>Cojoba undulatmarginata</i> EF638096.1
<i>Pseudosamanea guachapele</i> (Kunth) Harms, JX870769.1	<i>Ebenopsis confinis</i> (Standl.) Britton & Rose, EF638100.1
<i>Senegalia berlandieri</i> (Benth.) Britton & Rose, KY688777.1	<i>Ebenopsis ebano</i> EF638101.1
<i>Sphingia acatensis</i> (Benth.) Barneby & J.W. Grimes, EF638214.1	<i>Enterolobium contortisiliquum</i> EF638151.1
<i>Vachellia campechiana</i> (Mill.) Seigler & Ebinger, EF638215.1	<i>Enterolobium cyclocarpum</i> EF638149.1
<i>Vachellia farnesiana</i> (L.) Wight & Arn., EF638219.1	<i>Faidherbia albida</i> EF638163.1
<i>Viguieranthus ambongensis</i> (R. Vig.) Villiers, JX870773.1	<i>Havardia pallens</i> EF638146.1
<i>Viguieranthus densinervus</i> Villiers, JX870774.1	<i>Hesperalbizia occidentalis</i> EF638139.1
<i>Viguieranthus megalophyllus</i> (R. Vig.) Villiers, JX870776.1	<i>Hyrochorea corymbosa</i> EF638138.1
<i>Viguieranthus subauriculatus</i> Villiers, JX870778.1	<i>Jupunba trapezifolia</i> (Vahl.) Moldenke, EF638110.1
<i>Zapoteca tetragona</i> (Willd.) H.M. Hern., JX870784.1	<i>Mariosousa coulteri</i> (Benth.) Seigler & Ebinger, EF638124.1
	<i>Mariosousa dolichostachya</i> EF638084.1
	<i>Pararchidendron pruinatum</i> (Benth.) I.C. Nielsen, EF638129.1
	<i>Paraserianthes toona</i> (Bailey) I.C. Nielsen, EF638106.1
	<i>Pithecellobium dulce</i> EF638142.1
	<i>Pseudosamanea guachapele</i> EF638160.1
	<i>Samanea saman</i> EF638136.1
	<i>Samanea tubulosa</i> EF638135.1
	<i>Senegalia berlandieri</i> EF638162.1
	<i>Sphingia acatensis</i> EF638145.1
	<i>Vachellia farnesiana</i> EF638128.1
	<i>Viguieranthus ambongensis</i> KR997873.1
	<i>Viguieranthus densinervus</i> JX870891.1
	<i>Viguieranthus megalophyllus</i> KR997871.1
	<i>Viguieranthus subauriculatus</i> KR997076.1
	<i>Zapoteca formosa</i> EF638134.1
	<i>Zapoteca tetragona</i> EF638133.1.

The quality of the PCR products was evaluated by agarose electrophoresis (3 μ l of final volume plus 2 μ l of bromophenol blue, gel prepared with 0.5 \times TBE and 1% agarose, run at 120 volts and 25 amperes 30 min). PCR products were sequenced at MacroGen (<http://www.macrogen.com/eng/>) using the same amplification primers. The sequencing products were assembled and edited using the Sequencher v. 5.2.3. An initial automated alignment was conducted with MAFFT (Katoh et al. 2002) using the E-INS-i algorithm option, a 100PAM/k = 2 scoring matrix, a gap opening penalty of 1.3, and an offset value of 0.123. The alignments were visually inspected and manually edited for further improvement. The Akaike Information Criterion (AIC), implemented in jModeltest (Posada 2008) was used to select the best model of nucleotide substitution for each alignment. The selected models were TVM+I+G for ETS and GTR+I+G for ITS. Phylogenetic analyses were performed with MrBayes v.3.2.5 (Ronquist et al. 2012) separately for each dataset, and subsequently concatenated with each partition treated as independent and associated with its own evolutionary model. Analyses were performed using default parameters for 5 million generations. Two independent threads were run. Convergence was assessed with both MrBayes and Tracer (Rambaut et al. 2018). Posterior Probabilities (PP) \leq 0.95 were considered weakly supported whereas PP of 0.95–1.0 were deemed strongly supported (Alfaro et al. 2003).

To examine the evolution of fruit types within New World *Albizia* we utilize a phylogeny derived from a new analysis based on data of Ringelberg et al. (2022) for the Jupunba clade, as described in Soares et al. (2022), because it is based on a large set of 560 nuclear exons and flanking non-coding regions, and therefore shows enhanced resolution within this clade compared to the ITS + ETS phylogeny.

Results

Alignments of our combined datasets recovered by MAFFT required few manual adjustments. The ETS sequences had 381 bp and, once aligned, 52% of the data were informative. In the case of ITS, the sequences were slightly longer, 551 bp but only 34% were informative.

None of the molecular-based analyses (ETS, ITS, and ETS+ITS) using Bayesian inference recovered the genus *Albizia* as monophyletic. The combined ETS + ITS phylogeny (Fig. 3) is used as the basis for discussing the results in detail. The outgroup *Vachellia farnesiana*, plus *Acaciella angustissima* (Mill.) Britton & Rose, *Senegalia berlandieri* (Benth.) Britton & Rose and two species of *Mariosousa* Seigler & Ebinger form a paraphyletic grade subtending a fully supported clade (PP = 1) that includes all members of the tribe Ingeae as well as *Acacia* s.s.

The most relevant clade from the perspective of this study is highly supported (PP = 0.96) and includes all members of the genus *Albizia* and a few other genera of tribe Ingeae (clade A). The genus *Albizia*, as currently circumscribed, is non-monophyletic with species placed in two separate, strongly supported clades (Fig. 3). As in Koenen et al. (2020), species of *Albizia* sect. *Arthrosamanea* are placed in clade B (PP = 0.96),

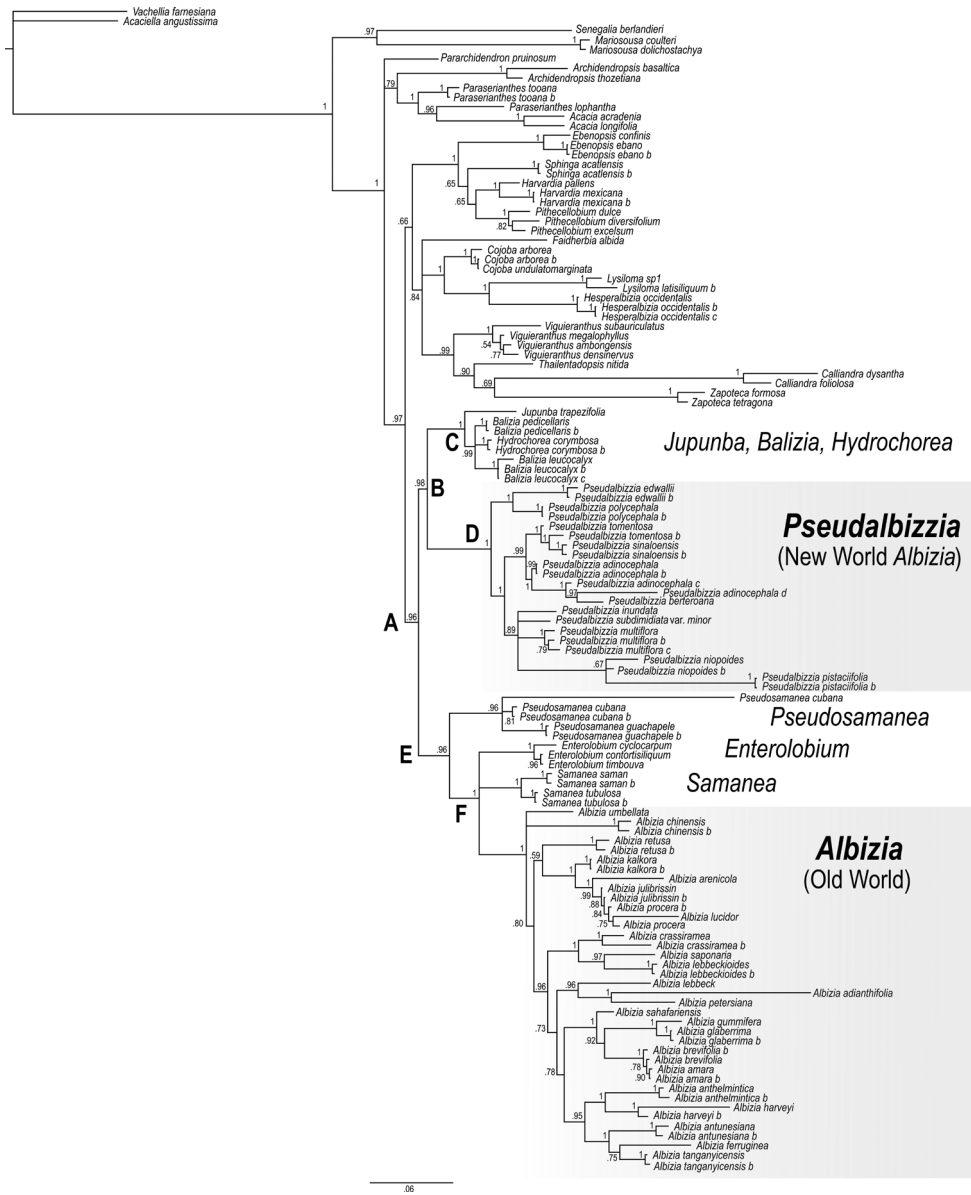
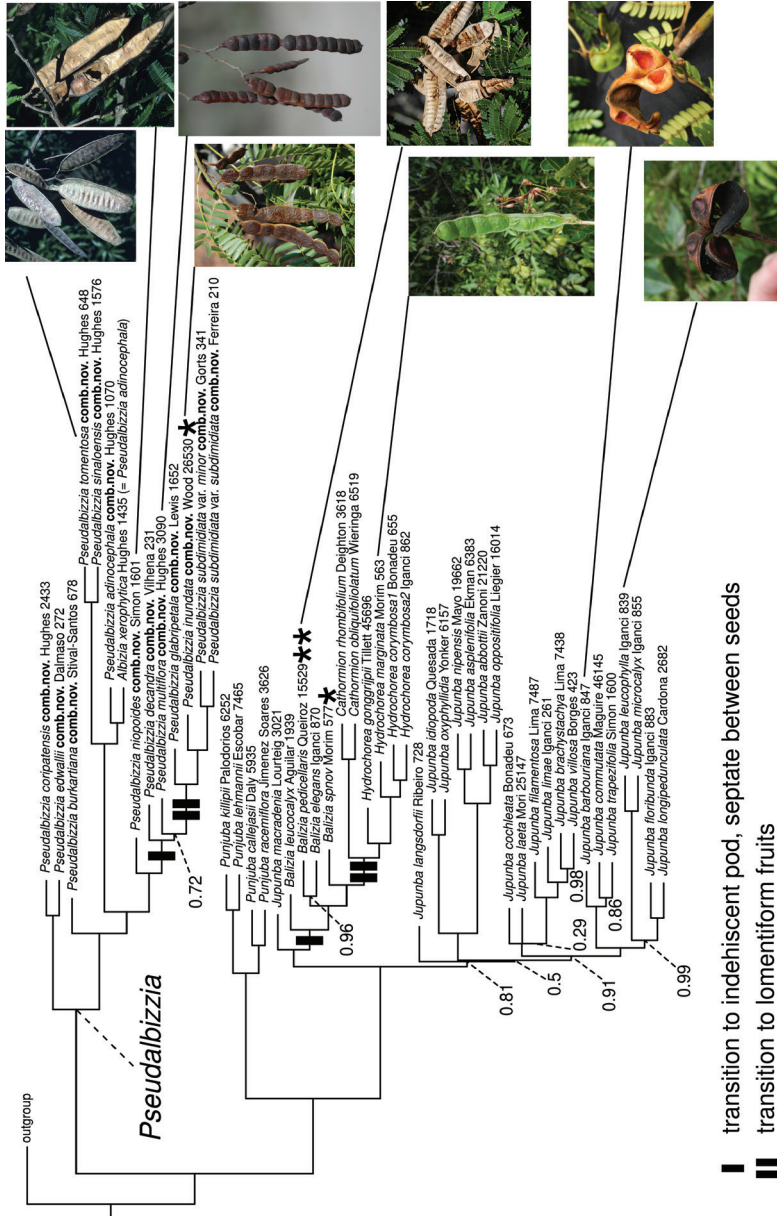


Figure 3. Phylogeny of the ingoid clade (sensu Koenen et al. 2020), i.e., the traditionally recognized tribes Ingeae + Acaciaeae (excl. *Vachellia*). Phylogram derived from Bayesian analysis in MrBayes of the combined ETS and ITS data for *Albizzia* and related genera. Main clades are labeled **A–F** (see text). Posterior support values are indicated above branches.

which equates to the *Jupunba* clade of Koenen et al. (2020). All species that were sampled from this section are included in this clade (Fig. 3 clade D), which received full support (PP = 1), and as in Koenen et al. (2020) this section is sister to clade C (PP = 1) comprising the genera *Jupunba*, *Balizia* and *Hydrochorea*.



- I transition to indehiscent pod, septate between seeds
- II transition to lomentiform fruits
- * cryptoloment, only the endocarp separating into 1-seeded articles
- ** follicular dehiscence

Figure 4. Phylogeny of the Jupunba clade redrawn from an ASTRAL species tree analysis by Soares et al. (2022) that utilizes data from Ringelberg et al. (2022) showing the evolutionary transitions from ancestrally papery, plano-compressed fruits to septate indehiscent fruits and subsequently to lomentiform hydrochorous fruits associated with species growing in seasonally inundated habitats in *Pseudalbizzia* and similar parallel transitions in *Balizia* and *Hydrochorea*. Photos of *Pseudalbizzia inun-data*, *P. multiflora*, *P. niopoides*, *P. tomentosa* and *Balizia pedicellaris*, by Colin Hughes, of *Hydrochorea marginata*, *Jupunba barbouriana* and *J. leucophylla*, by Erik Koenen.

Within sect. *Arthrosamanea*, three clades are well supported, one comprising *Albizzia polycephala* (Benth.) Killip ex Record and *Albizzia edwallii* (Hoehne) Barneby & J.W. Grimes of ser. *Paniculatae*, a second clade comprising species of ser. *Paniculatae* endemic to Mexico, Central America, and the Caribbean, and a third clade that includes *Albizzia niopoides* (Benth.) Burkart (also ser. *Paniculatae*) and the species from the other three series. The phylogeny of Ringelberg et al. (2022) presented here in Fig. 4 based on a new analysis of the Jupunba clade accessions from Soares et al. (2022), has greater resolution within sect. *Arthrosamanea* and shows that ser. *Paniculatae* forms a well-resolved paraphyletic grade in which the other three series are nested. The other two non-monospecific series also appear to be non-monophyletic, with the monospecific ser. *Inundatae* nested inside ser. *Arthrosamanea* and these together in turn nested in ser. *Multiflorae*, although support for the paraphyly of ser. *Multiflorae* is only 0.72 pp.

The Old World species of *Albizzia* form a monophyletic group (PP = 1) placed in clade E (PP = 0.96) (Fig. 3), with the genera *Enterolobium* Mart., *Samanea*, and *Pseudosamanea*. Within clade E *Pseudosamanea* (PP = 0.96) is sister to clade F which includes *Enterolobium*, *Samanea* and Old World *Albizzia*. All these clades have high support (PP = 1). These analyses also support the transfer of *Cathormion umbellatum* Kosterm., which is placed in the Old World *Albizzia* clade (Fig. 3), to *Albizzia*, as proposed by Koenen et al. (2020).

Our analyses also confirm that the monotypic genus *Hesperalbizzia*: *H. occidentalis* (Brandege) Barneby & J.W. Grimes is sister to *Lysiloma*, in the Cojoba clade (sensu Koenen et al. 2020), unrelated to either New World *Albizzia* (clade D) or Old World *Albizzia* (clade F), as previously shown by Duno de Stefano et al. (2021). Furthermore, *Pseudosamanea guachapele* (Kunth) Harms (previously *Albizzia guachapele* (Kunth) Dugand in Rico Arce et al. (2008)) is also unrelated to New World *Albizzia* but is instead a member of clade E, sister to clade F which includes *Enterolobium*, *Samanea* and Old World *Albizzia*.

In both *Albizzia* sect. *Arthrosamanea* and the closely related *Balizia* and *Hydrochorea*, these phylogenies suggest that lomentiform fruits were independently derived from indehiscent fruits that are septate between the seeds, as species with the latter fruit type form paraphyletic grades to the lomentiform species in both cases (Fig. 4). In turn, these indehiscent septate fruits are nested within paraphyletic assemblages of species with fruits that dehisce along one or both sutures in both groups. Interestingly, in both cases, a single species with crypto-lomentiform fruits is found, but it is not clear whether these were derived from the same ancestral fruit type or not. In *Hydrochorea* this crypto-lomentiform species appears as an intermediate between indehiscent and lomentiform species, while in *Albizzia* sect. *Arthrosamanea* the crypto-lomentiform-fruited *A. inundata* appears to be derived from a lomentiform-fruited ancestor. Another difference between these two groups is that the follicular dehiscence of *Balizia pedicellaris* (DC.) Barneby & J.W. Grimes fruits appears to be secondarily derived from indehiscent fruits, but we note that similar dehiscence is also found in a few species of *Jupunba*.

Discussion

This study addresses the non-monophyly of the genus *Albizia* and our results provide important insights into the evolutionary history of the neotropical species placed in sect. *Arthrosamanea*, with implications for their taxonomic classification. We show that sect. *Arthrosamanea*, with expanded taxon sampling relative to Koenen et al. (2020), and as also shown by the study of Ringelberg et al. (2022), is monophyletic with only two exceptions: *Albizia leonardii* Barneby & J.W. Grimes which is placed among taxa of the ‘senegalioid grade’ (Ringelberg et al. 2022; Terra et al. 2022) and *Albizia carbonaria* Britton that is more closely related to *Pseudosamanea* (Koenen 2022b; Ringelberg et al. 2022).

The geographically-based splitting of a large genus in tribe Ingeae, such as *Albizia*, which occupies a pantropical distribution, is not unprecedented, nor unexpected, especially given the lack of pantropical monographic synthesis or geographically widely sampled phylogenies for the mimosoid clade. For example, the genus *Pithecellobium*, once the largest genus of tribe Ingeae, has been progressively divided during the last 50 years into multiple genera (see Brown et al. 2008 for a general history of the tribe). Another example is the genus *Calliandra* Benth. for which the New World species of *Calliandra*. ser. *Laetevirentes* were segregated into *Zapoteca* H.M. Hern. (Hernández 1986), and almost all Old World species allocated progressively to other segregated genera: *Viguieranthus* Villiers (Villiers 2002), *Thailentadopsis* Kosterm. (Lewis and Schrire 2003), *Sanjappa* E.R. Souza & M.V. Krishnaraj (Souza et al. 2016) and *Afrocalliandra* E.R. Souza & L.P. Queiroz (Souza et al. 2013). All these taxonomic rearrangements were supported by morpho-anatomical and molecular phylogenetic analyses. Molecular data have also demonstrated that *Abarema* is polyphyletic (Iganci et al. 2015), prompting reinstatement of the genera *Jupunba* and *Punjuba* Britton & Rose (Soares et al. 2021). Finally, neither *Zygia* P. Browne nor *Marmaroxylon* Killip are monophyletic, although a new generic classification for those genera has yet to be proposed (Ferm et al. 2019). The non-monophyly of *Albizia* documented here and elsewhere (Koenen et al. 2020; Ringelberg et al. 2022) is thus not a surprise and reflects the state of flux surrounding generic delimitation in mimosoids, especially within the ingoid clade.

Some of the taxonomic proposals of Barneby and Grimes (1996) relative to the American segregates of *Albizia* s.l. are confirmed by our results. The genera *Balizia* and *Hydrochorea* form part of the *Jupunba* clade (sensu Koenen et al. 2020) (Figs 3 and 4) (Iganci et al. 2015), although neither *Hydrochorea* nor *Balizia* are monophyletic in our phylogeny (Figs 3 and 4, see Soares et al. 2022). Two neotropical species, included in *Albizia* by Rico Arce et al. (2008) are also placed outside New World *Albizia* (Fig. 3): *Hesperalbizia occidentalis* is closely related to *Lysiloma*, in agreement with previous results (Iganci et al. 2015; Duno de Stefano et al. 2021); similarly, *Pseudosamanea guachapele*, is also placed outside *Albizia* in our phylogeny, emerging, as expected, together with the other species *Pseudosamanea cubana* (Britton & Rose) Barneby & J.W. Grimes, although relationships within this clade are unresolved (Fig. 3).

Here we show that the dehiscent, papery, plano-compressed fruit type is ancestral within *Albizia* sect. *Arthrosamanea* (Fig. 4) and is associated with species growing predominantly in seasonally dry tropical forest and woodland, with successive shifts to septate indehiscent fruits and then lomentiform fruits with hydrochorous seed dispersal associated with species growing in seasonally inundated varzea forest, riverine habitats and low-lying margins of palm and white-sand savannas (Fig. 4). Interestingly, in the sister group of *Albizia* sect. *Arthrosamanea*, the mainly neotropical clade composed of *Jupunba*, *Punjuba*, *Balizia*, and *Hydrochorea*, a similar parallel evolutionary transition in fruit types is apparent. In *Jupunba* and *Punjuba*, fruits are always dehiscent, while a transition to septate indehiscent fruits occurred in *Balizia*, an exception being *Balizia pedicellaris* which has follicular dehiscence and a newly described species with crypto-lomentiform fruits (Fig. 4, and Soares et al. 2022). Nested within the paraphyletic *Balizia* is a clade comprising the genus *Hydrochorea* plus two African species of *Cathormion*, all species of which have indehiscent lomentiform fruits adapted for hydrochory and are found in riparian or other periodically flooded habitats in the Amazon basin, West Africa and the Congo basin (Fig. 4, and Soares et al. 2022).

Barneby and Grimes (1996) pointed out that a radiation of species with similarly heterogeneous fruit types to that seen in section *Arthrosamanea* occurs in Madagascan *Albizia* s.s. and that the association between lomentiform fruits, hydrochorous seed dispersal, and seasonally flooded habitats is also apparent in Old World *Albizia* s.s. For example, *Albizia dolichadena*, *A. moniliformis*, *A. rosulata*, and *A. umbellata* from Australasia also have lomentiform fruits and are distributed near streams or in riparian and swamp forests (Rico Arce et al. 2008). Furthermore, as indicated above and pointed out by Barneby and Grimes (1996), similar transitions to lomentiform fruits have occurred in parallel in several other lineages across the ingoid clade, including *Cathormion altissimum* (Hook.f.) Hutch. & Dandy (sometimes referred to as *Albizia altissima* Hook.f.; Koenen 2022a) and *Senegalia rostrata* (Humb. & Bonpl. ex Willd.) Seigler & Ebinger (syn. *Dugandia rostrata* (Humb. & Bonpl. ex Willd.) Britton & Killip, syn. *Manganaroa articulata* Speg.; Barneby and Grimes 1996: 204) in all cases apparently also closely associated with riparian and/or periodically inundated habitats. These repeated parallel derivations of similar, but not strictly homologous fruit types attest to the high evolvability of the mimosoid fruit more generally. In the light of phylogenetic evidence, it is now clear that these evolutionarily highly labile morphological adaptations of the fruit related to seed dispersal syndrome do not provide reliable characters for generic delimitation, supporting inclusion of the species that were placed in ser. *Arthrosamanea*, ser. *Inundatae* and ser. *Multi-florae* within *Albizia* sect. *Arthrosamanea* by Barneby and Grimes (1996), i.e., the clade of New World *Albizia* that is recovered in our analysis.

Taxonomic treatment

There are two validly published generic names – *Pseudalbizzia* of Britton and Rose (1928) and *Arthrosamanea* of Britton and Killip (1936) – that could be applied to the New World clade of *Albizia*. In accordance with Principle III of the International

Code of Nomenclature (Turland et al. 2018), we reinstate *Pseudalbizzia*, the earlier name associated with this clade, and provide the corresponding new combinations for its constituent species.

***Pseudalbizzia* Britton & Rose, N. Am. Fl. 23: 48. 1928.**

Type. *Pseudalbizzia berteriana* Britton & Rose.

Arthrosamanea Britton & Rose, in Britton & Killip, Ann. New York Acad. Sci. 35: 128, 1936. *Albizia* section *Arthrosamanea* (Britton & Rose) Barneby & J.W. Grimes, Mem. New York Bot. Gard. 74(1): 206. 1996. Type: *Arthrosamanea pistaciifolia* Britton & Rose.

Description. Unarmed **trees** with sympodial growth, up to 30 m, rarely small treelets of c. 3 m, microphyllidious to macrophyllidious; trunk 35–120(–150) cm dbh; young stems and all leaves and inflorescence-axes more or less densely tomentellous to pilosulous; stipules puberulent to glabrous, deltate, narrowly triangular, triangular-ovate, narrowly ovate, or narrowly lanceolate, veinless or faintly 3-veined, falling early to tardily, perhaps sometimes obsolete and/or lacking on mature leaves. **Leaves** bipinnate, not sensitive, (1–)2–15(–19) pairs of pinnae; leaflets (2–)16–52(–63) pairs per pinna; a nectary immediately below first pair of pinnae, near or well below mid-petiole, sometimes lacking or reduced to a minute pore, round, elliptic or vertically elongate, either shallow-cupular or almost plane, thick-rimmed, sometimes immersed in petiolar groove or even obsolete, much smaller nectaries at some distal pinnae, at the tip of most pinnae, and between 1–2 furthest pairs of leaflets; leaflets gently decrescent toward each end of the rachis or toward the base of the rachis or sub-equilong, the first pair of leaflets often reduced to paraphyllidia, sometimes minute, sometimes absent or perhaps falling early, the blades of the remaining leaflets elliptic, elliptic-ovate, oblong-elliptic, narrowly oblong-elliptic, lance-oblong to linear-lanceolate, base obliquely truncate to shallowly semi-cordate, apex deltately subacute, deltately acute to subacute, obtuse or apiculate, the larger ones (1.5–)2–4(–6) times as long as wide, margin strongly to slightly revolute; venation generally palmate, of 2–4(–5) veins from the pulvinule, the nearly straight main vein a little forwardly displaced and giving rise on each side to 2–13 major secondary veins, the inner of 2(–3) posterior primary veins incurved-ascending to anastomose slightly beyond mid-blade, the outer posterior vein and sometimes a faint anterior one very short and weak, all venation immersed on upper face. **Inflorescence** primary axis up to 30 cm long; peduncles (1–)2–8(–10) per node of the capitulate or corymbose-umbellate inflorescence, capitula 8–26(–40)-flowered; bracts heteromorphic or homomorphic, ovate, oblong-obovate or spatulate, linear-spatulate, falling early or persistent, sessile or shortly pedicellate, the flowers moderately to strongly dimorphic, the terminal ones generally longer. **Flowers** 5-merous, rarely 6-merous, glabrous to densely pubescent externally. Peripheral flowers: calyx campanulate, turbinate, turbinate-campanulate or narrowly campanulate, sessile or short pedicellate, lobes very short, depressed-deltate, ovate or triangular, glabrous or puberulent; corolla narrowly trumpet-shaped, erect or recurved, lobes ovate to lance-ovate; androecium with

9–30(–32) stamens, up to 20 mm long, united at the base forming a clear stemonozone, the staminal tube as long or longer than the stemonozone; ovary sessile or shortly stipitate, slenderly ellipsoid, conical at apex, glabrous or pubescent; style a little longer than the stamens, slightly dilated at the stigma. Terminal flowers: sessile or almost so, calyx shallowly campanulate to broadly campanulate, corolla tubular; androecium with 16–38(–42) stamens, 8.5–11.5(–13) mm long, united at the base forming a clear stemonozone, staminal tube equalling or longer than the stemonozone. **Fruits** solitary, or rarely 2–4 per capitulum, sessile, subsessile or cuneately contracted at base into a short pseudo-stipe, the body linear, linear-elliptic, narrowly elliptic-oblong, straight or nearly straight, sometimes decurved, plano-compressed, apex rounded but minutely apiculate to obtuse, (8–)13(–15)-seeded; valves papery, coriaceous, or grossly ligneous, olivaceous, castaneous, fuscous-greenish, or brown becoming tan-brown, closely transverse venulose, minutely puberulous, tomentulose, glabrescent to glabrous, framed by straight sutures or dilated, sometimes 3-angulate but not winged, transversely or horizontally, dehiscence tardy to very tardy, inert, through both sutures or dehiscence 0, in the latter, the pod crypto-lomentiform, incipiently lomentiform or lomentiform, then the whole fruit long persistent on the tree, commonly falling entire and breaking on the ground into 8–12 individually indehiscent segments, funicle apically sigmoid or ribbon-like (not sigmoid), lentiform; **seeds** obliquely ascending or straight, disciform, oblong-ellipsoid, elliptic, strongly compressed, the translucent, brownish or greyish testa produced as a peripheral wing, adherent to the embryo, which does not fill the testa-cavity, the pleurogram small, inversely U-shaped or U-shaped.

Notes. The genus forms a group that is homogeneous in most respects, but diverse in the late developmental stages of the fruit, including: 1) fruit opening type: dehiscent, indehiscent, or irregularly breaking, 2) lateral shape: flat to conspicuously raised over the seed chambers, 3) texture and consistency of the valves: papery, chartaceous to woody (Barneby and Grimes 1996). Figs 1, 2 and 4.

Pseudalbizzia (clade D) is the sister group of the *Jupunba-Punjuba-Balizia-Hydrochorea* clade (Fig. 3). *Jupunba* and *Punjuba* are markedly different morphologically, having spirally twisted dehiscent fruits with a red or ochre endocarp, reminiscent of the fruits of several other genera in tribe Ingeae (e.g., some *Pithecellobium* species, and some species of *Archidendron* F. Muell. and *Cojoba* Britton & Rose). The red or red-brown testa of the seeds of *Jupunba* and *Punjuba* are very distinctive, and are never black, and the embryo is nearly always aniline-blue due to the presence of delphinidin (an anthocyanidin). *Punjuba* is furthermore distinguished by its spicate inflorescences, which are not seen in *Pseudalbizzia*. *Balizia* has ligneous, indehiscent or tardily dehiscent pods, their seeds being released sometimes only after decay of the valves on the floor of terra firme forest, whereas in *Hydrochorea* the fruits are lomentiform, adapted to dispersal by water. The fruits of *Hydrochorea* recall some species of *Pseudalbizzia* adapted to similar riparian habitats. However, the species of *Pseudalbizzia* are markedly different in form of inflorescence, leaflet-venation, and shape of the ovary.

Two species previously placed in *Albizia* from the New World which were not included in our phylogenetic analysis, *Albizia carbonaria* and *A. leonardii*, have since been shown to be placed outside the New World *Albizia* clade (Ringelberg

et al. 2022; Koenen 2022b; Terra et al. 2022). Two other species, also not sampled here, nor by Ringelberg et al. (2022), are here tentatively included in *Pseudalbizzia*: *Albizia barinensis* L. Cárdenas and *Albizia buntingii* Barneby & J.W. Grimes (see below for discussion about the placement of these species). The genus *Pseudalbizzia* was published in the Flora of North America (Britton and Rose 1928) and included just a single species, *P. berteriana*. The original description of *Pseudalbizzia* closely matches *Albizia* and no characters distinguishing the two genera were discussed by Britton and Rose (1928). The generic name *Arthrosamanea* was also published by Britton & Rose, again with a single species, *A. pistaciifolia* (Willd.) Britton & Rose, in an account of the Mimosaceae and Caesalpiniaceae of Colombia (Britton and Killip 1936), but again no differences between the genus and *Albizia* or *Pseudalbizzia* were mentioned.

Pseudalbizzia as circumscribed here comprises 17 species and 5 varieties ranging in distribution from northwestern Mexico to northern Argentina and including the Greater Antilles (Figs 5 and 6). Full synonymy, detailed species descriptions, geographical distributions, representative samples of all species and keys for their identification can be found (under the name *Albizia*) in Barneby and Grimes (1996), Linares (2005) and Rico Arce et al. (2008). Finally, we propose a new sectional classification of

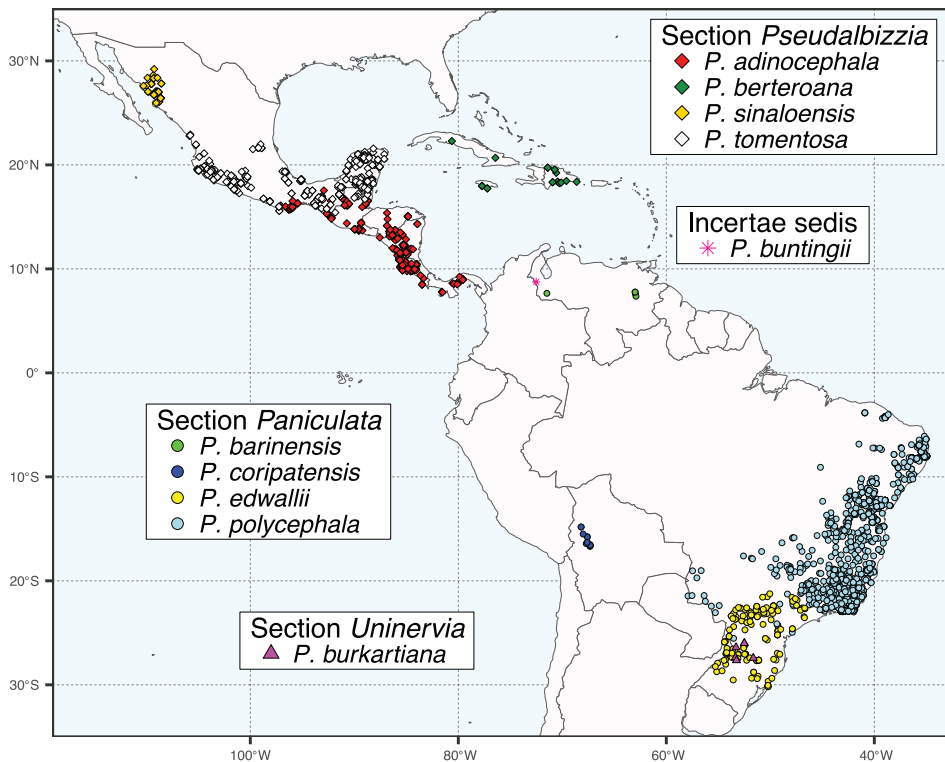


Figure 5. Distribution map of *Pseudalbizzia* sections *Paniculata*, *Pseudalbizzia*, *Uninervia* and *Pseudalbizzia buntingii* (incertae sedis), as per the legend.

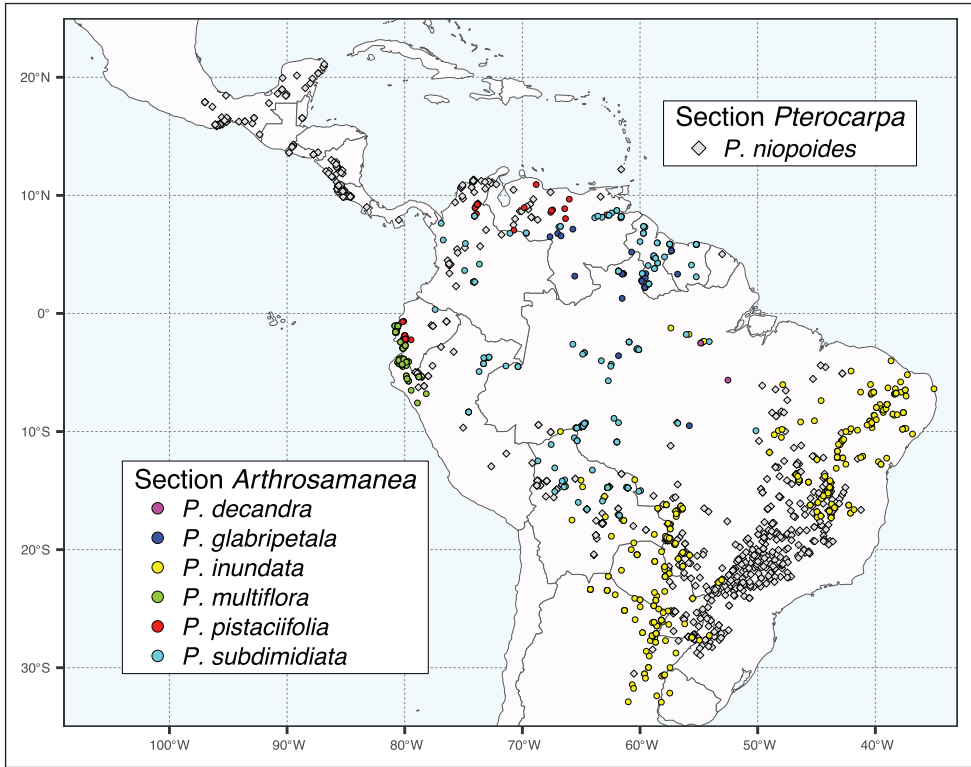


Figure 6. Distribution map of *Pseudalbizzia* sections *Arthrosamanea* and *Pterocarpa*, as per the legend.

Pseudalbizzia to account for the non-monophyly of the series of Barneby and Grimes (1996), based on the phylogenies (Figs 3 and 4) which sampled nearly all species. A key to the sections is provided.

Key to the sections of the genus *Pseudalbizzia*

- 1 Leaflets with a single vein from the pulvinule..... **sect. *Uninervia***
- Leaflets with 3–5 veins from the pulvinule..... **2**
- 2 Fruits with a narrowly winged margin, seeds oblique, foliage microphyllidious..... **sect. *Pterocarpa***
- Fruit margins not winged, or if winged, then foliage macrophyllidious and seeds straight..... **3**
- 3 Fruits indehiscent and septate or lomentiform **sect. *Arthrosamanea***
- Fruits dehiscent, plano-compressed, valves papery, not septate..... **4**
- 4 Micro- to mesophyllidious foliage, distributed in South America..... **sect. *Paniculata***
- Macro- or microphyllidious foliage, distributed in Mexico, Central America and the Caribbean **sect. *Pseudalbizzia***

***Pseudalbizzia* sect. *Paniculatae* (Benth.) E.J.M. Koenen & Duno, stat. nov. and sect. nov.**

urn:lsid:ipni.org:names:77303802-1

Pithecellobium sect. *Samanea* ser. *Paniculatae* Benth. pro parte, London J. Bot. 3: 219. 1844.
Albizia sect. *Arthrosamanea* ser. *Paniculatae* (Benth.) Barneby & J.W. Grimes pro parte, Mem. New York Bot. Gard. 74(1): 208. 1996. Type species (designated by Barneby and Grimes, Mem. New York Bot. Gard. 74(1): 208. 1996.): *Pithecellobium polycephalum* Benth. = *Pseudalbizzia polycephala* (Benth.) E.J.M. Koenen & Duno.
Pithecellobium sect. *Samanea* ser. *Parviflorae* [sic] Benth. pro parte, Trans. Linn. Soc. London 30: 591 (exclus. sp. 77). 1875 & in Martius, Fl. Bras. 15(2): 445. 1876. Type species (designated by Barneby and Grimes, Mem. New York Bot. Gard. 74(1): 208. 1996.): *Pithecellobium polycephalum* Benth. = *Pseudalbizzia polycephala* (Benth.) E.J.M. Koenen & Duno.

Type. *Pithecellobium polycephalum* Benth. = *Pseudalbizzia polycephala* (Benth.) E.J.M. Koenen & Duno.

Notes. Micro- to mesophyllidious trees with paniculate compound inflorescences of efoliate pseudoracemes and dehiscent plano-compressed papery fruits. Four species of humid, semi-deciduous and seasonally dry tropical and extratropical forests and woodland in South America (Fig. 5).

***Pseudalbizzia barinensis* (L. Cárdenas) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303803-1

Basionym. *Albizia barinensis* L. Cárdenas, Ernstia 21: 5, f. sn. 1983.

Type. Venezuela. Barinas, muy cerca de Punta de Piedra, 3 Apr 1976, L. Cardenas de Guevara 2273 (holotype: MY; isotypes: BM!, F! [F0093839F], K! [K000527984], NY! [NY00001781], RB! [RB00539860], US! [US00385615], VEN).

Notes. This species has not been included in any phylogenetic analysis, but its foliage, efoliate pseudoracemes and plano-compressed papery fruits leave little doubt that it should be placed in *Pseudalbizzia*. It is here included in section *Paniculata* based on these characters and its South American distribution.

***Pseudalbizzia coripatensis* (Rusby) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303804-1

Basionym. *Pithecellobium coripatense* Rusby, Bull. New York Bot. Gard. 4: 349. 1907.

Type. Bolivia. La Paz, Sur Yungas, at Coripata, 6 May 1894, M. Bang 2176 (holotype: NY! [NY00334642]; isotypes: BM! [BM000952433], G-2! [G00364414, G00364429], GH-2! [GH00064010, GH00064011], M! [M0218258], K! [K000527985], MINN, MO! [MO-954213], US).

***Pseudalbizzia edwallii* (Hoehne) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303805-1

Basionym. *Pithecellobium edwallii* Hoehne, Bol. Inst. Brasil. Sci. 2: 243. 1926.**Type.** Brazil, São Paulo, *G. Edwall 5608* (lectotype: SP, designated by Barneby and Grimes, Mem. New York Bot. Gard. 74(1): 209. 1996).***Pseudalbizzia polycephala* (Benth.) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303806-1

Basionym. *Pithecellobium polycephalum* Benth., London J. Bot. 3: 219. 1844.**Type.** Brazil, Rio de Janeiro, *J.B.E. Pohl 1420* (lectotype: K! (*herb. Bentham*) [K000528000], designated by Barneby and Grimes, Mem. New York Bot. Gard. 74(1): 208. 1996).***Pseudalbizzia* sect. *Uninervia* E.J.M. Koenen & Duno, sect. nov.**

urn:lsid:ipni.org:names:77303807-1

Type. *Albizia burkartiana* Barneby & J.W. Grimes = *Pseudalbizzia burkartiana* (Barneby & J.W. Grimes) E.J.M. Koenen & Duno.**Notes.** Microphyllidious trees with the inflorescences of section *Paniculata*, but with a single vein from the pulvinule at the base of the leaflets. A single, narrowly endemic species in Paraná pine woodland and the Southern Mata Atlantica of Brazil (Fig. 5).***Pseudalbizzia burkartiana* (Barneby & J.W. Grimes) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303808-1

Basionym. *Albizia burkartiana* Barneby & J.W. Grimes, Mem. New York Bot. Gard. 74(1): 211–212. 1996.**Type.** Brazil, Santa Catarina, Capinzal, on upper Rio Uruguai, 700 m, 21 Dec 1973, *P.R. Reitz & R. M. Klein 14359* (holotype: NY! [NY00001783]; isotype: US! [US00811452]).**Notes.** In the protologue the fruits were not described as these were not known at that time. This rare, locally endemic species has since been collected in fruit (*Stival-Santos 678*, BR), and we here provide a description of these. Fruits sessile but with a narrow pseudo-stipitate base, dehiscent along both slightly thickened sutures, the valves plano-compressed, papery in texture, light brown with finely prominent transverse veins, 6.5–12 × 1.2–1.6 cm, 7–12-seeded when well-fertilized.

***Pseudalbizzia* sect. *Pseudalbizzia*.**

Type as for the genus.

Notes. Trees with micro- or macrophyllidious foliage, inflorescences composed of efoliate pseudoracemes arising singly from a leaf axil or sometimes the capitula solitary or paired in the leaf axils, or the pseudoracemes combined into a terminal panicle, fruits plano-compressed with papery valves, dehiscent along both sutures or more rarely indehiscent (in *P. berteriana*), sometimes with a winged margin, seeds straight. Four species predominantly of seasonally dry tropical forests in Mexico, Central America and the Caribbean (Fig. 5).

***Pseudalbizzia adinocephala* (Donn. Sm.) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303809-1

Albizia xerophytica J. Linares, syn. nov., Revista Mex. Biodiversidad 76: 7. 2005. Type: Honduras. El Paraíso, Municipio Morocelí, orillas de Quebrada Grande c. 3.9 km al NE de Morocelí por el camino hacia El Plan. 2002. *J.L. Linares et al.* 5674 (holotype: MEXU! [MEXU01160777]; isotype: EAP).

Basionym. *Pithecellobium adinocephalum* Donn. Sm., Bot. Gaz. Crawfordsville. 57: 419. 1914.

Type. Costa Rica. San José, Ad fundum La Verbena prope Alajuelita, 100 m, Aug 1894, *A. Tonduz* 8932 (US-3); Dec 1894 (lectotype: *A. Tonduz* 9077 [US-212774]!; isolectotypes: BR-3! [BR0000005189519, BR0000005189182, BR0000005189847], G! [G00364416], designated by Barneby and Grimes, Mem. New York Bot. Gard. 74(1): 218. 1996).

Notes. *Albizia xerophytica* was described from material from dry forest habitats in southern Honduras based on minor differences in leaf and fruit morphology, but we do not consider these to be significantly different from the range of variation that is observed in *P. adinocephala* and prefer the broader concept of the species as described in Barneby and Grimes (1996: 218–220). The difference in habitat (i.e., lower rainfall regions) also appears to be minor, as some specimens from wetter sites have been identified as *A. xerophytica* (see map in Rico Arce et al. 2008) while specimens of *P. adinocephala* have been collected across the full range of drier and wetter sites. Finally, the distribution of *A. xerophytica* is entirely enclosed by the much wider range of *P. adinocephala*.

***Pseudalbizzia berteriana* (Balb. ex DC.) Britton & Rose, N. Amer. Fl. 23: 48. 1928.**

Basionym. *Acacia berteriana* Balb. ex DC., Prodr. 2: 470. 1825.

Type. Republica Dominicana, Sto. Domingo, *C.L.G. Bertero, herb. Balbis s.n.*, 1821 (holotype: G; isotype: M! [M0218254]).

***Pseudalbizzia sinaloensis* (Britton & Rose) E.J.M. Koenen & Duno, comb. nov.**
urn:lsid:ipni.org:names:77303810-1

Basionym. *Albizzia sinaloënsis* in Britton & Rose, *N. Amer. Fl.* 23(1): 45. 1928.

Type. Mexico. Sinaloa, vicinity of Fuerte, 26 March 1910, *J.N. Rose, P.C. Standley & Russell 13559* (holotype: NY! [NY00001775]; isotype: US! [US00000483]).

***Pseudalbizzia tomentosa* (M. Micheli) E.J.M. Koenen & Duno, comb. nov.**
urn:lsid:ipni.org:names:77303811-1

Basionym. *Pithecellobium tomentosum* M. Micheli, *Mém. Soc. Phys. Genève* 34: 285, t. 28. 1903.

Type. Mexico. Michoacán, rives de l'Espíritu Santo, 600 m, 19 April 1898 [*E. Langlassé*] 107 (G): Zimatango, 30 m, aout 1898, n 280 (G). (lectotype: *E. Langlassé* 107 G-385667!; isolectotypes: K! [K000082098], NY (fragm.)! [NY00001777], designated by Standley, *Contr. U.S. Natl. Herb.* 23: 396. 1922).

***Pseudalbizzia tomentosa* var. *nayaritensis* (Britton & Rose) E.J.M. Koenen & Duno, comb. nov.**
urn:lsid:ipni.org:names:77303812-1

Basionym. *Albizzia nayaritensis* Britton & Rose, *N. Amer. Fl.* 23: 47. 1928.

Type. Mexico. Nayarit; San Blas, La Palma, 20 m, 1923, *J. González Ortega 90N* (holotype: US! [US00918691]; isotypes: K! [K000082100], NY-2! [NY00001768, NY00001769]).

***Pseudalbizzia tomentosa* var. *purpusii* (Britton & Rose) E.J.M. Koenen & Duno, comb. nov.**
urn:lsid:ipni.org:names:77303813-1

Basionym. *Albizzia purpusii* Britton & Rose, *N. Amer. Fl.* 23: 45. 1928.

Type. Mexico. Veracruz, Rancho Remudadero, 19°15'N, 96°34'W, April 1922, *C.A. Purpus 8723* (holotype: NY! [NY00001773]; isotypes: GH! [GH00069252], MO! [MO-120564], UC! [UC214372], US! [US00000479]).

Pseudalbizzia tomentosa* var. *tomentosa***Pseudalbizzia* sect. *Pterocarpa* E.J.M. Koenen & Duno, sect. nov.**

urn:lsid:ipni.org:names:77303814-1

Type. *Pithecellobium niopoides* Spruce ex Benth. = *Pseudalbizzia niopoides* (Spruce ex Benth.) E.J.M. Koenen & Duno.

Notes. Microphyllidious trees with the inflorescence usually composed of axillary efoliate pseudoracemes, sometimes a partly or wholly terminal panicle (but not surpassing the foliage), the fruit with a narrowly winged margin and seeds oblique. A single widespread species found in deciduous seasonally dry forests, gallery forest, and evergreen forests in Mexico, Central and South America (Fig. 6).

***Pseudalbizzia niopoides* (Spruce ex Benth.) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303815-1

Basionym. *Pithecellobium niopoides* Spruce ex Benth., Trans. Linn. Soc. London 30: 591. 1875.

Type. Brazil, Pará, Santarem, Nov 1851, *R. Spruce 1088, Herb. Bentham* (holotype: K! [K000528013]).

***Pseudalbizzia niopoides* var. *colombiana* (Britton) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303816-1

Albizzia niopoides var. *colombiana* (Britton) Barneby & J.W. Grimes, Mem. New York Bot. Gard. 74(1): 222. 1996.

Basionym. *Albizzia colombiana* Britton, in Britton & Killip, Ann. New York Acad. Sci. 35: 131. 1936.

Type. Colombia. Magdalena, near Bonda, Santa Marta, 3 August 1899, *H.H. Smith 38* (holotype: NY! [NY00001784]; isotypes: BR! [BR0000005111176], E! [E00313853], K! [K000527990], NY!, U-2! [U0003354, U1253389]).

Pseudalbizzia niopoides* var. *niopoides***Pseudalbizzia* sect. *Arthrosamanea* (Britton & Rose) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303817-1

Arthrosamanea Britton & Rose, Ann. New York Acad. Sci. 35: 128, pro gen. 1936, *sensu stricto*. *Albizzia* sect. *Arthrosamanea* (Britton & Rose) Barneby & J.W. Grimes

pro parte, Mem. New York Bot. Gard. 74(1): 206. 1996. Type species: *Arthrosamanea pistaciifolia* (Willd.) Britton & Rose = *Mimosa pistaciifolia* Willd. = *Pseudalbizzia pistaciifolia* (Willd.) E.J.M. Koenen & Duno.

Albizzia sect. *Arthrosamanea* ser. *Multiflorae* Barneby & J.W. Grimes, Mem. New York Bot. Gard. 74(1): 234. 1996.

Albizzia sect. *Arthrosamanea* ser. *Inundatae* Barneby & J.W. Grimes, Mem. New York Bot. Gard. 74(1): 238. 1996.

Notes. Micro- or macrophyllidious trees, usually the efoliate pseudoracemes arising singly and only rarely arranged in panicles, fruits indehiscent and septate, or lomentiform, one species crypto-lomentiform. Six species of usually humid, often seasonally inundated forest or riparian habitats in South America (Fig. 6).

***Pseudalbizzia decandra* (Ducke) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303818-1

Basionym. *Pithecellobium decandrum* Ducke, Arch. Jard. Bot. Rio de Janeiro 5: 121. 1930.

Type. Brazil. Pará, habitat in silvis non inundatis civitatis Pará circa Óbidos, *A. Ducke* (Herb. Amaz. Mus. Pará 15.724, et H.J.B.R. 10.174) et loco Serra do Dedal ad lacum Faro, *A. Ducke* (H.J.B.R. 20.198), ubi florebat Januario 1927, *A. Ducke* (lectotype: *A. Ducke* 10174 RB!; isolectotypes: G! [G00364418], K-2!: [K000527990, K000527998], U-2! [U0003349, U0003350], designated by Barneby and Grimes, Mem. New York Bot. Gard. 74(1): 234. 1996.).

***Pseudalbizzia glabripetala* (H.S. Irwin) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303819-1

Basionym. *Pithecellobium glabripetalum* H.S. Irwin, in Mem. New York Bot. Gard. 15(1): 109. 1966.

Type. Guyana. Orealla, Corantyne River, Oct 1879, *G.S. Jenman* 364 (holotype: NY! [NY00334664]; isotypes: BM!, P!).

***Pseudalbizzia inundata* (Mart.) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303820-1

Basionym. *Acacia inundata* Mart., Spix & Mart. in Reise Bras. 1: 555. 1823.

Type. Brazil. Minas Gerais, Rio Sao Francisco, 1818, *C.F.P. von Martius* 1659 (holotype: M! [M0218478]; isotypes: K! [K000797598], NY!).

***Pseudalbizzia multiflora* (Kunth) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303821-1

Basionym. *Acacia multiflora* Kunth, Nov. Gen. Sp. (quarto ed.) 6: 277–278. 1823.**Type.** Peru. Cajamarca, Prov. Jaén, San Felipe, 980 m, *Aime Bonpland & F.W.H.A. von Humboldt 3562* (holotype: P! [P00679365]).***Pseudalbizzia multiflora* var. *multiflora******Pseudalbizzia multiflora* var. *sagasteguii* (Barneby & J.W. Grimes) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303822-1

Basionym. *Albizia multiflora* var. *sagasteguii* Barneby & J.W. Grimes, Mem. New York Bot. Gard. 74(1): 237–238. 1996.**Type.** Peru. Cajamarca, Prov. Contumazá, in a quebrada near San Benito, *A. Sagástegui 15410* (holotype: F! [F0042945F]; isotypes: MO! [MO-149743], NY!, US! [US00624358]).***Pseudalbizzia pistaciifolia* (Willd.) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303823-1

Basionym. *Mimosa pistaciaefolia* [sic] Willd., Sp. Pl. 4: 1028. 1806.**Type.** Venezuela. Caracas. *F. Bredemeyer s.n., herb. Willdenow* (holotype: B).***Pseudalbizzia subdimidiata* (Splitg.) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303824-1

Albizia subdimidiata (Splitg.) Barneby & J.W. Grimes, Mem. New York Bot. Gard. 74(1): 234. 1996.**Basionym.** *Acacia subdimidiata* Splitg. Tijdschr. Natuurl. Gesch. Physiol. 9: 112 (1842).**Type.** Suriname. “ad ripas fluminis Surinami superioris”, 27 April 1838. *Splitgerber 917* (holotype: L [L0018505]).***Pseudalbizzia subdimidiata* var. *minor* (Barneby & J.W. Grimes) E.J.M. Koenen & Duno, comb. nov.**

urn:lsid:ipni.org:names:77303825-1

Basionym. *Albizia subdimidiata* var. *minor* Barneby & J.W. Grimes, Mem. New York Bot. Gard. 74(1): 234. 1996.

Type. Guyana. Basin of Essequibo river, Kuyaliwak Falls, 1 Jan 1937, *A.C. Smith* 2156 (holotype: NY! [NY00001790]; isotypes: A! [A00069262], G! [G00364427], K! [K000528004], P, U! [U0003358]).

Pseudalbizzia subdimidiata var. *subdimidiata*

Incertae sedis

Pseudalbizzia buntingii (Barneby & J.W. Grimes) E.J.M. Koenen & Duno, comb. nov.

urn:lsid:ipni.org:names:77303826-1

Basionym. *Albizia buntingii* Barneby & J.W. Grimes, Mem. New York Bot. Gard. 74(1): 223. 1996.

Type. Venezuela. Zulia, alrededores de Casigua El Cubo, 100 m, al este del empalme de la via hacia Casigua con la carretera Machiques-La Fría, 25 Feb 1985, *G.S. Bunting* 13370 (holotype: NY! [NY00001782]).

Notes. Fruits of this species are unknown and the species is only known from the type locality (Fig. 5), but it is similar in leaf and inflorescence morphology to several South American species of *Pseudalbizzia*, as described in the protologue. Especially the efoliate pseudoracemes point to this species most likely being correctly accommodated in *Pseudalbizzia*. Collection of fruits and/or inclusion of the species in phylogenetic studies is needed to confirm its generic and sectional placements.

Non-native species

Some cultivated and sometimes naturalized Old World *Albizia* species are found in the New World, including: *A. procera* (Roxb.) Benth., *A. julibrissin*, *A. lebbek* (L.) Benth., and *A. chinensis* (Osbeck) Merr. For these species, Barneby and Grimes (1996) proposed *Albizia* section *Albizia*, now considered as *Albizia* s.s.

Author contributions

GAP, RR, GCFC, IVM, and RDD designed the study. GAP, LLCI, ELC, RDD contributed labwork. RDD, GCFC, IRM contributed data by supervising students in the lab. EJMK, RDD, XC, SM and CEH contributed taxonomic knowledge, JR contributed species distribution data and the maps. GAP, RDD, ITC, JRP, and RR undertook the phylogenetic analyses. EJMK, RDD, RR, GCFC, CEH and JR contributed to writing the manuscript.

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