Alpha thalassemia mental retardation X-linked

Acquired alpha-thalassemia myelodysplastic syndrome



### Schematic representation of the spectrum of *ATRX* mutations that have been described in boys with ATR-X syndrome with those found in ATMDS.

Mutations predicted to cause protein truncation (frameshifts and nonsense mutations) and null mutations. Amino acid changes, including in-frame insertions and deletions. Recurrent mutations in ATR-X syndrome are indicated by a number inside the circle, representing the number of families in whom the mutation has been identified.



**Figure 1** Hematological analysis in individuals with ATMDS. (**a**) Blood film from individual 1 showing hypochromia, anisocytosis, poikilocytosis and target cells. (**b**) Blood film of blood from individual 1 stained with brilliant cresyl blue showing the presence of cells with hemoglobin H ( $\beta_4$ ) inclusions. (**c**) Globin chain biosynthesis plot showing marked reduction in the ratio of  $\alpha/\beta$  globin synthesis in individual 2. (**d**) A comparison of the  $\alpha/\beta$  globin chair biosynthesis ratios for individual 2 (open circle) and the mean for normal controls (filled square).

- The chromatin-associated protein ATRX was originally identified because mutations in the ATRX gene cause a severe form of syndromal X-linked mental retardation associated with α-thalassemia.
- The disease-causing mutations fall into two groups: the majority affect buried residues and hence affect the structural integrity of the ADD domain; another group affects a cluster of surface residues, and these are likely to perturb a potential protein interaction site. The effects of individual point mutations on the folding state and stability of the ADD domain correlate well with the levels of mutant ATRX protein in patients, providing insights into the molecular pathophysiology of ATR-X syndrome

### **ATRX** mutations



Half of all of the disease-associated missense mutations cluster in a cysteine-rich region in the N terminus of ATRX. This region was named the ATRX-DNMT3-DNMT3L (ADD) domain, based on sequence homology with a family of DNA methyltransferases The positions of missense mutations are indicated with circles and the number of times (>1) the mutation has been identified in unrelated individuals is indicated within relevant circles. All of the circles drawn between the oblique lines above the bar refer to mutations within the ADD domain.



the structure of the ADD domain of ATRX consists of an N-terminal GATA-like zinc finger, a plant homeodomain finger (PHD), and a long C-terminal  $\alpha$ -helix that pack together to form a single globular domain. The  $\alpha$ -helix of the GATA-like finger is exposed and highly basic, suggesting a DNA-binding function for ATRX.  $\beta$ -Strands are labeled s1–s4 and helices h1–h4.

## Locations of mutations and secondary structural elements in the ADD domain



- Locations of mutations and secondary structural elements in the ADD domain. The N-terminal GATA-like zinc finger is indicated by a light green bar, the PHD finger by a mauve bar, and the C-terminal extension by a light blue bar. The conserved cysteine residues are marked as orange vertical bars.
- Missense mutations are highlighted in green (surface), blue (buried), and orange (cysteines); the insertion mutation is highlighted by an upward green arrow and the deletion by a downward blue arrow. Residues where there is homology across the whole family of ADD domain sequences (ATRX, DNMT3A, DNMT3B, and DNMT3L) are marked with filled circles (absolute conservation), gray circles (strong conservation), and open circles (weak conservation)



- Electrostatic potential and location of mutations in the structure of the ADD domain.
- (*a*) Surface electrostatic potential of the ADD domain. The helix in the GATA-like finger (h1) is solvent-exposed and basic, and the two helices within loop 2 of the PHD finger (h2 and h3) form another basic patch. The linker between the GATA-like and PHD fingers is highly acidic.
- (b) Ribbon structure of the ADD domain showing the locations of mutations found in patients with ATR-X syndrome. Mutations are classified as surface (green), buried (blue), or cysteine (orange) and are represented by using their side chains, except for the glycine mutation G249C/D and the glutamine insertion, which are represented by thickening the backbone. The surface mutations are individually labeled



- ATRX *in vivo* expression in EBV-transformed patient lymphocytes.
- (a) ATRX mRNA levels of patient mutations and normal controls as determined by quantitative RT-PCR. Patients are grouped according to the nature of their underlying mutation: cysteine mutations are orange, buried mutations are blue, and surface mutations are green. Values for normal individuals are represented by black circles. For each case, the ATRX mRNA level is expressed as the percentage of the average for 18 normal control individuals.
- (b) ATRX protein levels of patients and normal controls. ATRX protein levels are expressed as a percentage of the average value for seven normal control individuals.
- (c) Representative Western blots showing ATRX protein levels (including loading control). Lane 1 represents the ATRX protein level for a cysteine mutation, lanes 2–4 are buried mutations, lanes 5–7 are surface mutations, and lane 8 is a normal control.





ATRX colocalizes with HP1 at heterochromatin (in murine L929 cells).



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Figure 6-34a Molecular Cell Biology, Sixth Edition © 2008 W. H. Freeman and Company

## HP1

- structural protein that plays a role in heterochromatin formation, gene silencing
- three genes HP1Hs $\alpha$ , HP1Hs $\beta$  and HP1Hs $\gamma$ .
- distinct chromosomal localization patterns
- HP1 contain a chromo domain that binds methylated K9H3
- HP1 dimers establish a platform in which nuclear proteins interact
- Association of HP1 with a target gene causes alterations in chromatin structure and gene silencing

Table1 Chromatin modifications				
Mark*	Transcriptionally relevant sites†	Transcriptional role‡		
DNA methylation				
Methylated cytosine (meC)	CpG islands	Repression		
Histone PTMs				
Acetylated lysine (Kac)	H3 (9, 14, 18, 56), H4 (5, 8, 13, 16), H2A, H2B	Activation		
Phosphorylated serine/ threonine (S/Tph)	H3 (3, 10, 28), H2A, H2B	Activation		
Methylated arginine (Rme)	H3 (17, 23), H4 (3)	Activation		
Methylated lysine (Kme)	H3 (4, 36, 79) H3 (9, 27), H4 (20)	Activation Repression		
Ubiquitylated lysine (Kub)	H2B (123\$/120¶) H2A (119¶)	Activation Repression		
Sumoylated lysine (Ksu)	H2B (6/7), H2A (126)	Repression		
Isomerized proline (Pisom)	H3 (30-38)	Activation/ repression		

\*The modification on either DNA or a histone.

 $\dagger {\sf Well}{\rm -characterized sites with regard to genomic location for {\sf DNA methylation or residues within}$ histones for PTMs.

‡Whether the epigenetic mark is associated with activation or repression.

§Yeast (Saccharomyces cerevisiae).

¶Mammals.

### ATRX: funzioni

 Alteration of the regional distribution of heterochromatin by the complex or of recruitment of transcription factors that alter gene expression ?

## POSIZIONMENTO NUCLEOSOMA Rimodellamento Cromatina





### nucleosomal array remodeling activity of ATRX



Restriction enzyme accessibility assay, EtBr stain



#### nucleosomal array remodeling activity of ATRX





### ATRX: funzioni

• Alteration of the regional distribution of heterochromatin by the complex or of recruitment of transcription factors (directly or indirectly) that alter gene expression ?

epigenetic modifications of histone-associated DNA, leading to alterations in local chromatin conformation.

binding to DNA of transcription factors is affected by the ATRX complex

 ATRX is part of a multiprotein complex that uses the energy of ATP to remodel chromatin or its associated DNA in a way that affects transcriptional activity at euchromatic loci, including the -globin gene cluster.

#### RNA profiling in ATMD and **normal** granulocytes



Figure 2 Gene expression analysis in an individual with ATMDS. (a) A pc of the microarray: the pseudocolored signals for the two samples (ATMD aRNA in green and normal aRNA in red) superimposed to show their rela intensity. Yellow spots represent probes binding equivalent target cDNA both samples; red spots, normal target is more abundant than ATMDS ta green spots, ATMDS target is more abundant than normal target. One of three ATRX probes is shown. (b) Distribution plot showing the gene expression ratio between granulocytes from individual 1 with ATMDS an granulocytes from a mixed pool of normal individuals. Values in the rang 0–0.5 are boxed. (c) Magnification of the distribution plot in the range 0-0.5. The three probes representing ATRX are colored red. (d) Graph comparing real-time quantitative PCR data on ATRX expression in granulocytes from 7 normal controls, 13 individuals with MDS and individual 1 with ATMDS. Values are corrected so that the mean of the normal = 100%.

## METILAZIONE del DNA e ATRX



Bcll + Hpall or Mspl RibC7b5

#### Table 1 • The pattern of methylation in ATR-X syndrome<sup>a</sup>

Sequence	Hypomethylation	Hypermethylation	Unaffected
rDNA	+		
5SDNA			+
satellite 1 (oligo)			+
satellite 1 (DYZ2)		+	
satellite 2 (oligo)			+
satellite 3 (DYZ1)			+
α-satellite (oligo)			+
β-satellite (oligo)			+
Alu (consensus)			+
L1 (consensus)			+
TTAGGG (oligo) (telomeric	:)		+
Tel Bam 3.4 (subtelomeric)	)	(+)	
Tel Bam 11 (subtelomeric)			+
XIST <sup>b</sup>			+
D15S63 (PWS)			+
α-globin cluster <sup>d</sup>			+

• Mutations in the human methyl-CpGbinding protein gene *MECP2* cause the neurological disorder Rett syndrome

# • Interaction between chromatin proteins MECP2 and ATRX

Kriaucionis PNAS 2007;104;2709-2714.



### MeCP2 interacts with ATRX.

- (b) The ATRX-interaction region overlaps theMBDof MeCP2. The MeCP2 region that binds ATRX was identified by GST pulldown by using a deletion series of MeCP2-GST fusion proteins incubated with *in vitro*-translated [35S]-labeled ATRX fragment 1201–2190 (bracket; *a*). The input lane (*Top*) contained 25% of labeled protein used for the pull-down assay.
- (c) Summary of GST-pulldown showing presence or absence of an ATRX interaction leading to identification of an ATRX-interacting domain (AxID). Point mutations that inhibit ATRX binding (R133C, A140V, and R168X; see Fig. 5) are marked by arrows.





MeCP2 interacts with ATRX.

- (*a*) Yeast two-hybrid assays identified a region of ATRX that interacts with MeCP2. The diagram depicts ATRX and its domains.
- Symbols indicate the strength of the MeCP2 interaction. The MeCP2 interaction domain (MID) is marked.



MecP2 Interaction Domain



## ATRX localization in neurons of *Mecp2*-null mice

ATRX localization is disrupted in neurons of *Mecp2*-null mice



## Mutations in human MeCP2 cause mental retardation



Mutations in human in the MBD of human MeCP2 that cause mental retardation disturb the MeCP2–ATRX interaction without affecting methyl- CpG binding.



## MeCP2 mutant A140V target heterochromatic foci but cannot direct ATRX to heterochromatin



## disruption of the MeCP2–ATRX interaction leads to pathological changes that contribute to mental retardation.