# EPIPHYTIC LICHEN DIVERSITY IN BROAD-LEAVED TREE FORESTS IN LATVIA

# Kristīne Štikāne, Guntis Brūmelis, Alfons Piterāns, Rolands Moisejevs

Štikāne K., Brūmelis G., Piterāns A., Moisejevs R. 2017. Epiphytic lichen diversity in broadleaved tree forests in Latvia. *Acta Biol. Univ. Daugavp.*, *17 (1): 123 – 132*.

Broad-leaved tree forests host a large diversity of epiphytic lichens, but this forest type is rare in Latvia. Epiphytic lichen diversity of broad-leaved trees mostly has been studied in protected areas and there is insufficient knowledge of the general distribution of epiphytic lichens outside the protected areas. The aims of this study were to compare epiphytic lichen diversity among tree species and between broad-leaved tree forest in protected and non-protected areas in Latvia.

Epiphitic lichen diversity in broad-leaved tree woodland was studied in two regions of Latvia: Kurzeme and Zemgale. Altogether 67 lichen species were recorded on 160 sampled trees of 11 tree species in 19 forest stands. Eight of the recorded lichens are listed in the Latvian protected species list, of which 4 are species for which microreserves can be established. Only two were listed in the Latvian Red Data Book. Protected species were found in 18 of the studied 19 stands, but only 6 of these stands occurred in protected areas. One recorded lichen species *Opegrapha niveoatra* is reported as new to Latvia. *Quercus robur* had the highest number of lichen species on the basal trunk. 53.85% of recorded occurences (21 of 39 records) of protected lichen species and 60.42% of recorded occurences (29 of 48 records) of Woodland Key Habitat species occurred outside of protected areas.

Key words: epiphytic lichens, species diversity, broad-leaved forests, fragmentation, protected species, protected areas.

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### **INTRODUCTION**

Biological diversity is the foundation of functioning of ecosystems (Kouki 1994), and involves diversity of species, structures and processes, at various spatial scales (Peterken 1996, Esseen et al. 1997). Broadleaved tree (*Fraxinus excelsior*, *Ulmus glabra*, *Tilia cordata*, *Quercus robur*) forests are associated with high epiphytic lichen diversity (Esseen et al. 1997, Jüriado et al. 2003), particularly in boreo-nemoral broadleaved tree woodland compared to boreal coniferous forest (Barkman 1958, Jüriado et al. 2003). Forest age and continuity are two main factors affecting epiphytic lichen diversity (Lesica et al. 1991, Esseen et al. 1996, Esseen et al 1997, McCune 1993, Jüriado et al. 2003). Tree diversity of a woodland is known to promote epiphytic lichen diversity (Fahselt, Krol 1989, Kuusinen 1996a, Kuusinen 1996b, Mežaka et al. 2012).

Greater richness of epiphytic bryophytes and lichens occurs on deciduous in comparison with coniferous trees (Berg et al. 1994). The main factors affecting epiphytic lichen growth are moisture, temperature and light (Hale 1973, Fahselt, Krol 1989, McCune 1993, Esseen et al. 1997), which differ between forest site types.

The impact of man on forests in the temperate zone has been profound (Jones 1945, Esseen et al. 1997), and in Europe natural undisturbed woodland is almost non-existent (Peterken 1981).

In Latvia, which occurs in the transition boreonemoral zone between temperate and boreal forest zones, where mixed forest with broadleaved and coniferous trees is common (Sjors 1963), the former area of broadleaved forest has almost disappeared, and *Quercus robur* woodland now covers only about 1.0% of the total forest area (LMD 2017). The broadleaved forests in Latvia are highly fragmented by agricultural land and intensively managed forest, which might be expected to limit successful lichen dispersal (Ellis & Coppins 2007).

However, it is predicted than global change will lead to an increase in area of *Quercus robur* in northern Europe at the expense of coniferous tree species (Hanewinkell et al. 2012).

In Latvia, epiphytic lichen communities have been studied mostly in protected territories and Woodland Key habitats (Mežaka et al. 2008), but publications on the distribution of lichens outside of these areas is lacking. Study is needed to understand the affects of many factors like spatial and temporal continuity on epiphytic lichen diversity and indicators species (Norden & Appelqvist 2001). In this regard, knowledge is needed on occurrence of protected and rare lichen species outside of protected areas, to enable development of conservation strategies (Marmor et al. 2011). The aim of the study was determine the occurrence of lichen species in relation to tree species and protection status of stands in broadleaved forests in two regions of Latvia.

### **MATERIAL AND METHODS**

### Climate

All selected stands were located in South-Western region of Latvia. The climate of the South-Western region of Latvia is mainly attributed to Atlantic cyclones that bring air masses and precipitation from the west and northwest. The average temperature ranges from -3 to -6°C in January, and from 16.5 to 17°C. in July. Annual precipitation ranges from 600 to 850 mm (Turlājs 2011).

### Site description

The stands were randomly selected from the Forest Register of the State Forest Service with criteria of broad-leaved trees composition at least 70% (Fig 1). Model territories selected were the Kurzeme and Zemgale regions, which historically had the largest area of broad-leaved tree stands. Three stands (PAD2, PAD12, and MEZ5) of 19 complied with criteria of protected habitats of the European Community 9160 "Sub-Atlantic and medio-European oak or oak-hornbeam forests of the Carpinion betuli" and are included in the Natura2000 network (Tab. 1) (Auniņš et al. 2013). Two of these (PAD2 and PAD12) are part of the nature reserve "Tebras ozolu meži" and one (MEZ5) is in the microreserve "Ozoldārzs" (Anonymous 2017). In this study, old wooded city parks (ZAL1, ZAL2, and DOB5) were grouped together with protected areas, as these forests have a history of protection as manor parks (Table 1).

### Field study

The study was conducted in August to November 2016. Epiphytic lichens were sampled in each

Epiphytic lichen diversity in broad-leaved tree forests in Latvia

ID	Studied area	Name of plot	X coordinates*	Y coordinates*	Nr.of Sampled trees	Mean diameter	Minimum diameter
1	Dobele	DOB5***	456121	273451	12	45.71	25.00
2	Dzelda	DZD1	364660	272521	9	37.78	20.00
3	Kazdanga	KAZ3	363704	285176	12	33.63	19.00
4	Ķikuri	KIK1	355279	296965	9	46.11	20.00
5	Mežotne	MEZ1	506418	259605	3	29.67	23.00
6	Mežotne	MEZ2	504951	255603	7	70.07	45.00
7	Mežotne	MEZ3	507937	255941	9	34.72	18.50
8	Mežotne	MEZ4	498488	254948	9	31.89	11.50
9	Mežotne	MEZ5**	502092	255739	6	67.25	19.00
10	Padure	PAD12**	351422	293116	13	47.27	26.00
11	Padure	PAD2**	350890	293087	9	39.00	32.00
12	Padure	PAD9	348218	296687	12	41.17	14.00
13	Platone	PLAT1	477788	251336	8	50.19	29.00
14	Platone	PLAT2	474592	258436	6	34.08	16.00
15	Vītiņi	VIT1	440603	251127	9	49.11	26.00
16	Vītiņi	VIT2	439148	254595	6	55.08	11.00
17	Vītiņi	VIT3	438710	254196	4	55.75	36.50
18	Zaļenieki	ZAL1***	470080	265518	9	51.83	25.00
19	Zaļenieki	ZAL2***	470573	265939	9	36.06	18.00

Table 1. Location and characteristics of the studied forest stands

\*Geographical system of coordinates: LKS 1992 Latvia TM.

\*\*Protected territory (LĪADT 1993).

\*\*\* Old wooded city park.



Fig. 1. Location of studied forest stands.

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Tuble 2. Concered signal epipilytic nehen species proportion within studied areas																
Species status*	WKH	WKH, P, MR	WKH, P	WKH, P	WKH, P	WKH	WKH	WKH	WKH, P, MR, R	WKH, P, MR	P, R	WKH	WKH, P, MR	WKH	P, MR	P, WKH, MR
Lichen species	Acrocordia gemmata	Arthonia byssacea	Arthonia leucopellaea	Arthonia spadicea	Arthonia vinosa	Bacidia rubella	Graphis scripta	Lecanactis abietina	Menegazzia terebrata	Opegrapha vermicellifera	Pertusaria hemisphaerica	Phlyctis agelaea	Sclerophora peronella	At least one WKH species	At least one P or MR	Signal species (P, MR or WKH species)
Number of protected stands where recorded	2	3	3	1	1	3	3	0	0	0	0	1	0	5	6	6
Number of non- protected stands where recorded	7	4	5	7	1	3	6	4	1	1	1	3	1	11	12	12
Proportion of protected stands where recorded, %	33.33	50.00	50.00	16.67	16.67	50.00	50.00	0	0	0	0	16.67	0	83.33	100.00	100.00
Proportion of non- protected stands where recorded, %	53.85	30.77	38.46	53.85	7.69	23.08	46.15	30.77	7.69	7.69	7.69	23.08	7.69	84.62	92.31	92.31

Table 2. Collected signal epiphytic lichen species proportion within studied areas

\*WKH-Woodland Key Habitat species (Auniņš et al. 2013), P – Latvian protected species list (Anonymous 2000), MR – microreserves can be established (Anonymous 2013), R – listed in the Latvian Red Data Book (Andrušaitis et al. 1996).

stand from three randomly selected trees (from the largest ones) for each tree species in the stand. Trees less than 10 cm diameter were not sampled. All lichens present at stem height 5 to 180 cm were sampled. In some cases when less than three trees per species were found, less were sampled. Diameter at breast height (130 cm) was measured for each sampled tree.

#### Data analysis

The collections were determined using the routine lichenological methods (Smith et al., 2009). Spottest reactions were checked with 10% KOH (K),

sodium hypochlorite (C), para-phenylenediamine in ethanol (Pd), and Lugol's solution (I). The nomenclature of lichenized fungi taxa follows Nordin et al. (2011).

Differences in epiphytic lichen occurrence and richness were examined in relation to tree species and forest stand. Relationship between species richness and stem diameter by tree species was determined using Pearson or Spearman correlation analysis depending on data distribution, using R version 3.2.5 (R Development Core Team 2011).

Tree species	Total richness	Mean richness	
Populus tremula L., 1753	19	0.63	
Betula pendula Roth, 1788	11	0.37	
Salix caprea L., 1753	4	0.09	
Picea abies (L.) H.Karst., 1881	19	1.07	
Ulmus glabra Huds., 1762	11	0.18	
Acer platanoides L., 1753	15	0.37	
Tilia cordata Mill., 1768	25	1.16	
Fraxinus excelsior L., 1753	12	0.32	
Quercus robur L., 1753	43	2.38	
Pinus sylvestris L., 1753	5	0.12	
Quercus rubra L., 1753	3	0.12	

Table 3.	Total and and mean lichen species
richness	for each tree species

Mean lichen richness and total richness were calculated for each tree species. Tree and lichen richness were determined for each stand. Gradient analysis of epiphytic lichen communities in forest stands was conducted using NMS (Non-metric Multidimensional Scaling) with Sorensen-Bray Curtis distance measure (McCune et al. 2002).

# RESULTS

### Lichen diversity

A total of 67 lichen species (Appendix 1) were recorded on 161 individuals of 11 tree species. Of the studied 19 forest stands in Southwest part of Latvia, of which three were in Natura 2000 teritories and three were old manor parks, only one stand (VIT2) lacked valuable lichen species that were either protected or Woodland Key Habitat indicators. Eighteen of the nineteen stands contained at least one protected species and fourteen contained at least one Woodland Key Habitat (WKH) species (Table 2). Only one of the stands lacked both protected and WKH species.

*Opegrapha niveoatra* was recorded in Latvia for the first time, in stand VIT1 (Table 1) in Zemgale, on a *Populus tremula* with diameter 37.5 cm, on bark. This stand also contained WKH species like *Acrocordia gemmata* and *Lecanactis abietina* and protected species like *Arthonia leucopellaea* and *Arthonia spadicea*.

Of the examined trees in stands, 58 were in protected stands and 103 in non-protected stands. A total of 58 lichen species were recorded in protected stands and 58 in non-protected stands. The largest number of lichen species (43 of 67 species) were observed on Quercus robur stems (Table 3). Eight species were included in the list of protected species in Latvia (Anonymous 2000), of which four (Arthonia byssacea, Menegazzia terebrata, Opegrapha vermicellifera, and Sclerophora peronella were in the list of protected species for which microreserves can be established (Table 2). Two of the protected species (Menegazzia terebrata and Pertusaria hemisphaerica are included in the Red Data List for Latvia (Andrušaitis et al. 1996). Acrocordia gemmata (WKH indicator species) and Arthonia spadicea (protected species) were found in 53.85% (7 no 13) of the non-protected stands. Arthonia byssacea, a species for which microreserves can be established, was found in four non-protected stands and Arthonia leucopellaea, a protected species, was found in five non-protected stands. All records of Menegazzia terebrata, Opegrapha vermicellifera, Pertusaria hemisphaerica, Arthonia vinosa and Sclerophora peronella, were from non-protected stands, but each of these species was found in only one stand (Table 2).

Highest total and mean richness of lichen species was found on *Quercus robur* (43 species) and *Tilia cordata* (25 species) and the lowest on *Pinus sylvestris* (5 species) and *Salix caprea* (4 species).

Relationship between tree stem diametar and lichen species richnes was determined for trees with sample size at least 15 trees (*Quercus robur*, *Populus tremula*, *Tilia cordata* and *Picea abies*), but all correlations were non-significant. A significant correlation (R=0.6521, p<0.001) was found between lichen and tree species diversity in plots.

# DISCUSSION

A total of 67 epiphytic lichen species were found in the study. A similar number of lichen species (75) was found in a previous study of WKH broadled forest of ravines and screes (Mežaka et al. 2012). Of the four protected species found, for whch microreserves can be established, three (*Menegazzia terebrata, Opegrapha vermicellifera* and *Sclerophora peronella* were found outside of protected areas.

*Opegrapha niveoatra* ((Borrer) J.R. Laundon, 1963), which was reorded for the first time in Latvia, also was found outside of protected areas, on a *Populus tremula*. Surprisingly, only two (*Menegazzia terebrata* and *Opegrapha vermicellifera*) of the recorded protected lichen species are included in the Red Data List for Latvia (Andrušaitis et al. 1996), indicating that this list needs to be updated.

Three of the 19 studied forest stands (PAD2, PAD12, and MEZ5) had been previously identified as EU protected habitat 9160 Sub-Atlantic and medio-European oak or oakhornbeam forests of the Carpinion betuli and were found in protected areas. These stands contained the protected species Arthonia byssacea, Arthonia leucopellaea and Arthonia vinosa. However, the epiphytic lichen species that are WKH indicator species were lacking in some protected areas (PAD12, PAD2, and ZAL2), while common in others (for example, on 5 of 6 studied trees in stand MEZ5). Regarding the studied old manor parks, the WKH species Bacidia rubella and Graphis scripta were found in stand ZAL1 and Acrocordia gemmata, Bacidia rubella and Phlyctis agelaea in DOB5. It can be assumed that old manor parks were important in maintaining continuity of substrate for epiphytes requiring broad-leaved tree species, and that light conditions were variable and promoting high diversity. It is possible that some of the other sites were part of an ancient wood meadow landscape around manors, with spatial and temporal continuity of broad-leaved trees. Certainly, the presence of some oak with large diameter in some stands suggests past land-use and wooded meadow. Thus, the legacy of previous land-use probably contributed the present high richness of protected and DMB species outside of protected areas. It is likely that some of the studied stands complied with EU protected Habitats, but had not been previously identified as such. This indicates the necessity for further study of the epiphytic lichens in relation to past stand history and spatial continuity of suitable substrate.

Higher lichen richness is associated with *Acer* platanoides, Fraxinus excelsior, Quercus robur, *Tilia cordata*, and *Ulmus glabra*, compared with *Betula sp.* and coniferous trees (Barkman 1958, Berg et al. 1994, Cieśliński et al. 1995). The high lichen richness on *Quercus robur* and other broad-leaved tree species can be explained by characteristics of bark like porousness (Rose 1974), pH and crevice depth (Mežaka et al. 2012). In comparison with the broad-leaved tree species, 19 lichen species were found on spruce and 19 on birch, only slightly less than on *Tilia cordata*.

While the relationship between tree diameter and epiphytic lichen richness is known (McCune 1993, Esseen et al. 1996, Peterken 1996, Jüriado et al. 2003), in our study we found no such relationship for the tree species investigated, which migh be explained by low sample size and effect of other factors like light, humidity and temperature (Hale 1973, Fahselt, Krol 1989, McCune 1993, Esseen et al. 1997) as well as anthropgenic effect (Hale 1983, Nash 1996, Conti, Cecchetti 2001).

The observed correlation between tree and lichen species diversity can be explained by higher substrate and niche diversity (Fahselt, Krol 1989, Lesica et al. 1991, Kuusinen 1996a, Kuusinen 1996b, Jüriado 2003, Fritz 2009, Mežaka et al. 2012). This preliminary study of 19 stands did not allow objective analysis of lichen species composition in relation to tree species. However, differences were obvious. For example, *Lecanactis abietina* is a common indicator of old *Picea abies* stands (Piterāns 2001) and *Arthonia byssacea, Arthonia vinosa, Arthonia spadicea,* and *Opegrapha vermicellifera* require broadleaved tree species or *Populus tremula* bark as a substrate (Piterāns 2001). Further study in this direction is being planned.

Some of the stands, like DZD1, are not protected, but presence of species Menegazzia terebrata and Opegrapha vermicellifera indicate a need to establish new microreserves. A similar stand is MEZ5, where two species for which microreserves can be established (Arthonia byssacea and Arthonia spadicea and 4 WKH indicator species (Acrocordia gemmata, Bacidia rubella, Phlyctis agelea and Graphis scripta) were found. The study should be expanded to a greater number of stands, particularly ensuring comparable number of protected and non-protected areas, and with different levels of forest fragmentation and management legacies. It might be expected that after future invesistigation of species distributions, the protection status of some species will change.

# CONCLUSIONS

1. Higher tree species diversity can ensure higher lichen species diversity.

2. Higher richness of protected and WKH species has been supported by past land-use as manor parks and wooded meadow landscape.

3. Broad-leaved tree species like *Quercus robur* un *Tilia cordata* and also the pioneer species *Populus tremula* support higher lichen richeness than coniferous species.

# ACKNOWLEDGEMENTS

Appreciation is extended to Latvian State Forest Research Institute "Silava", and particularly to Dr. silv. Dagnija Lazdiņa, and also the State Research Programme EVIDENT for financial support. We appreciate the help of Zigmārs Rendenieks in stand selection.

### REFERENCES

- Andrušaitis G., Vimba E., Piterāns A. 1996. Latvian Red Data Book. Rare and extinct plant and animal species. Volume 1. Mushrooms and lichens. Rīga: Latvijas Zinātņu akadēmija, Bioloģijas institūts, Pp. 202.
- Anonymous. 2000. Regulations of Chamber of Ministers Nr. 396. Regulation about specially protected species. Ministru kabineta noteikumi Nr.396. Noteikumi par īpaši aizsargājamo sugu un ierobežoti izmantojamo īpaši aizsargājamo sugu sarakstu. https://likumi.lv/doc.php?id=12821 [25.03.2017.].
- Anonymous. 2013. Regulations of Chamber of Ministers Nr. 940. Regulation about microreserves. Ministru kabineta noteikumi Nr.940. Noteikumi par mikroliegumu izveidošanas un apsaimniekošanas kārtību, to aizsardzību, kā arī mikroliegumu un to buferzonu noteikšanu. https://likumi.lv/doc. php?id=253746 [25.03.2017.].
- Anonymous. 2017. Nature data conservation system of Mature Conservation Agency OZOLS. [12.04.2017.].
- Auniņš, A., Lārmanis, V., Rove, I., Rūsiņa, S., Laime, B. (eds). 2013. European Union Protected Habitats in Latvia. Interpretation Manual. Riga, Latvian Fund for Nature, Ministry of Environmental Protection and Regional Development. Pp. 320.
- Barkman J. J. 1958. *Phytosociology and Ecology* of Cryptogamic Epiphytes. Assen: Van Gorcum & Company, Pp. 628.
- Berg A., Ehnström B., Gustafsson L., Hallingbäck T., Jonsell M., Weslien J. 1994. Threatened plant, animal, and fungus species in Swedish forests: distribution and habitat associations. *Conservation Biology*, 8 (3): 718 – 731.

- Cieśliński S., Czyżewska K., Glane K. 1995. Lichenes. *Phytocoenosis*, 7: 75 – 86.
- Ellis C. J. & Coppins B. J. 2007. 19th Century woodland structure controls stand-scale epiphyte diversity in present-day Scotland. *Diversity and Distribution*, 13: 84 – 91.
- Esseen P. A., Renhorn K. E., Pettersson R. B. 1996. Epiphytic lichen biomass in managed and old-growth boreal forest. *Ecological Applications* 6 (1): 228 – 238.
- Esseen P. A., Ehnström B., Ericson L., Sjöberg K. 1997. Boreal forests. *Ecological Bulletins*, 46: 16 – 47.
- Fahselt D., Krol M. 1989. Biochemical comparison of two ecologically distinctive forms of *Xanthoria elegans* in the Canadian High Arctic. *Lichenologist*, 21: 135 – 145.
- Fritz O. 2009. Tree age is a key factor for the conservation of epiphytic lichens andbryophytes in beech forests. *Applied Vegetation Science*, 12: 93–106.
- Hale M. E. 1973. *The Lichens. Chapter 14: Growth.* New York: Academic Press, Pp. 473 – 492.
- Hale M. E. 1983. *The Biology of Lichens*. London: Baltimore, Edward Arnold, Pp.190.
- Hanewinkell M., Cullmann D. A., Schelhaas M.
  J., Nabuurs G. J., Zimmermann N. E. 2012.
  Climate change may cause serve loss in the economic value of European forest land. *Nature Climate Change*, 3: 203 207.
- Jones E. W. 1945. The structure and reproduction of the virgin forest of the North temperate zone. *The New Phytologist*, 44 (2): 130 – 148.
- Jüriado I., Paal J., Liira J. 2003.Epiphytic and epixylic lichen species diversity in Estonian natural forests. *Biodiversity and Conseration*, 12: 1587 – 1607.

- Kouki J. 1994. Biodiversity in the Fennoscandian boreal forests: natural variation and its management. *Annales Zoologici Fennici*, 31: 1 - 217.
- Kuusinen M. 1996a. Epiphyte flora and diversity on basal trunks of six old-growth forest tree species in southern and middle boreal Finland. *Lichenologist*, 28 (5): 443 – 463.
- Kuusinen M. 1996b. Cyanobacterial macrolichens on *Populus tremula* as indicators of forest continuity in Finland. *Biological Conservation*, 75 (1): 43 – 49.
- Lesica P., McCune B., Cooper S.V., Hong W.S. 1991. Differences in lichen and bryophyte communities between old-growth and managed second-growth forests in the Swan Valley, Montana. *Canadian Jounal of Botany*, 69: 1745 - 1755.
- LĪADT 1993. Law of Republic of Latvia. About specially protected nature territories. Latvijas Republikas likums. Par īpaši aizsargājamām dabas teritorijām. https://likumi.lv/ta/ id/59994-par-ipasi-aizsargajamam-dabasteritorijam [1.04.2017.].
- LMD 2017. Homepage of Latvian Forest department. Latvijas Meža dienesta mājaslapa. http://www.lmd.lv/par-latvijasmeziem [03.05.2017.].
- Marmor L., Tōrra T., Saag L., Randlane T. 2011. Effects of forest continuity and tree age on epiphytic lichen biota in coniferous forests in Estonia. *Ecological Indiactors*, 11: 1270 – 1276.
- McCune B. 1993. Gradients of epiphyte biomass in three *Pseudotsuga-Tsuga* forests of different ages in western Oregon and Washington. *Bryologist*, 96: 405 - 411.
- McCune B., Grace J. B., Urban D. L. 2002. Analysis of ecological communities. U.S.A. Gleneden Beach, Oregan: MjM Software Design, Pp. 304.

- Mežaka A., Brūmelis G., Piterāns A. 2008. The distribution of epiphytic bryophyte and lichen species in relation to phorophyte characters in Latvian natural old-growth broad leaved forests. *Folia Cryptogamica Estonica*, 44: 89–99.
- Mežaka A., Brūmelis G., Piterāns A. 2012. Tree and stand-scale factors affecting richness and composition of epiphytic bryophytes and lichens in deciduous woodland key habitats. *Biodiversity and Conservation*, 21 (12): 3221–3241.
- Nash T. H. 1996, 2008. *Lichen Biology. Second Edition.* New York: Cambridge University Press, Pp. 486 pp.
- Norden B. & Appelqvist T. 2001. Conceptual problems of ecological continuity and its bioindicators. *Biodiversity and Conservation*, 10: 779 – 791.
- Nordin A., Moberg R., Tønsberg T., Vitikainen O., Dalsätt Å., Myrdal M., Snitting D., Ekman S. 2011. Santesson's Checklist of Fennoscandian Lichen-forming and Lichenicolous Fungi. Ver. 29 April 2011. http://130.238.83.220/santesson/home.php [30.03.2015].
- Peterken G. F. 1981. *Woodland Conservation and Management*. Great Britain: Springer US, Pp. 337.

- Peterken G. F. 1996. Natural woodland. Ecology and conservation in Northern temperate regions. Great Britain: Cambridge University Press, Pp. 540.
- Piterāns A. 2001. Summary of Latvian lichens. Latvijas ķērpju konspekts. *Latvijas* Veģetācija, 3: 5 - 46.
- R Development Core Team 2011. *R: A Language and Environment for Statistical Computing.* The R Foundation for Statistical Computing, Vienna, Austria.
- Rose F. 1974. The epiphytes of oak. *The British* Oak. Its History and Natural History, 250 - 273.
- Sjors H. 1963. Amphi-Atlantic zonation nemoral to Arctic. In: Love A., Love D (eds). North Atlantic biota and their history. The Macmillan Company, New York, Pp. 109–125.
- Turlājs, J. 2011. Latvijas ģeogrāfijas atlants [Geographical atlas of Latvia]. Rīga: Publishing house "Jāņa sēta".
- Smith C.W., Aptroot A., Coppins B.J., Fletcher A., Gilbert O.L., James P.W., Wolseley P.A., 2009: *The lichens of Great Britain and Ireland*. London, Pp. 1046.

#### **Appendix 1**

#### List of taxa of epiphytic lichens recorded during the researh in specific forest stands (Number I D in Table 1)

Acrocordia gemmata (Ach.) A.Massal., 1854 :	7, 9, 15, 16, 17)
(1, 0, 9, 12, 13, 13, 10)	Artinonia paleitalia Nyl., 1905. (1, 10)
Amanainea punctata (Hoffm.) Coppins &	Arthonia radiata (Pers.) Ach., 1808 : (3, 7, 9,
Scheid., 1993 : (6)	12, 16, 18)
Arthonia atra (Pers.) A. Schneid., 1898 : (15)	Arthonia sp. Zahlbr., 1903 : (2, 13)
Arthonia byssacea (Weigel) Almq., 1880 : (1, 9,	Arthonia spadicea Leight., 1854 : (2, 7, 8, 12,
16, 17, 18, 19)	13, 14, 16, 17)
<i>Arthonia dispersa</i> (Schrad.) Nyl., 1861 : (1, 7)	Arthonia vinosa Leight., 1856 : (4, 10, 11)
Arthonia leucopellaea (Ach.) Almq., 1880 : (1,	Bacidia rubella (Hoffm.) A.Massal., 1852 : (1,

7, 12, 14, 18) Bacidina arnoldiana (Körb.) V.Wirth & Vězda, 1994 : (1, 9, 13, 14) Buellia griseovirens (Turner & Borrer ex Sm.) Almb., 1952 : (1, 4, 8, 9, 10, 12, 13, 15) *Calicium salicinum* Pers., 1794 : (4) Calicium viride Pers., 1794 : (18) Candelariella xanthostigma (Pers. ex Ach.) Lettau, 1912 : (7) Chaenotheca ferruginea (Turner ex Sm.) Mig., 1930 : (1, 15, 16, 19) Chaenotheca stemonea (Ach.) Müll.Arg., 1862 :(17) Cladonia chlorophaea (Flörke ex Sommerf.) Spreng., 1827 : (2, 3, 10, 12) Cladonia coniocraea (Flörke) Spreng., 1827 : (2, 3, 10, 11, 12)Cladonia cornuta (L.) Hoffm., 1791 : (10) *Cladonia sp.* : (4, 10, 11, 19) Dimerella lutea (Dicks.) Trevis., 1880 : (3) *Dimerella pineti* (Ach.) Vězda, 1975 : (1, 2, 11) *Evernia prunastri* (L.) Ach., 1810 : (6, 9) Graphis scripta (L.) Ach., 1809 : (5, 6, 7, 8, 9, 12, 13, 18, 19) Hypocenomyce scalaris (Ach.) M.Choisy, 1951 :(1) Hypogymnia physodes (L.) Nyl., 1896 : (2, 3, 4, 12) Lecanactis abietina (Ehrh. ex Ach.) Körb., 1855 : (2, 10, 15, 17)*Lecanora chlarotera* Nyl., 1872 : (1, 5, 6, 7, 13) Lecidella elaeochroma (Ach.) M.Choisy, 1950 : (9) Lecidella euphorea (Flörke) Hertel, 1980: (1, 5, 6, 7, 8, 9, 13, 15, 16, 18) Lecidella stigmatea (Ach.) Hertel & Leuckert, 1969:(7)Lepraria elobata Tønsberg, 1992: (1, 2, 4, 11, 14) Lepraria incana (L.) Ach., 1803 : (1, 2, 3, 4, 6, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19) Lepraria jackii Tønsberg, 1992 : (2) *Lepraria lobificans* Nyl., 1873 : (15, 16, 17) Lepraria neglecta (Nyl.) Erichsen., 1958 : (16) Melanelixia glabra (Lamy) Sandler & Arup, 2011 : (12) Melanelixia glabratula (Lamy) Sandler & Arup, 2011:(1,3,13,19)Melanohalea olivacea (L.) O.Blanco et al., 2004:(1)

Menegazzia terebrata (Hoffm.) A. Massal., 1854 : (15) Opegrapha niveoatra (Borrer) J.R. Laundon, 1963:(2)Opegrapha vermicellifera (Kunze) J.R.Laundon, 1963:(2)Opegrapha vulgata (Ach.) Ach., 1803 : (10, 13, 14,15, 18) Parmelia sulcata Taylor, 1836 : (1, 13) Peltigera praetextata (Flörke) Vain., 1899 : (4, 12) Pertusaria albescens (Huds.) M.Choisy & Werner, 1932 : (2, 3, 12) Pertusaria amara (Ach.) Nyl., 1872 : (2, 4, 10, 12, 17) Pertusaria coccodes (Ach.) Nyl., 1857 : (12) Pertusaria hemisphaerica (Flörke) Erichsen, 1932 : (3) Phaeophyscia orbicularis (Neck.) Moberg 1977 :(18)*Phlyctis agelaea* (Ach.) Flot., 1850 : (7, 8, 12) Phlyctis argena (Ach.) Flot., 1850 : (1, 2, 3, 4, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19) *Physcia dubia* (Hoffm.) Lettau, 1912 : (7) *Physcia tenella* (Scop.) DC., 1805 : (7) Physconia distorta (With.) J.R.Laundon, 1984 :(7) Physconia grisea (Lam.) Poelt, 1965 : (12) Platismatia glauca (L.) W.L.Culb. & C.F.Culb., 1968:(2,4)Pseudoschismatomma rufescens (Pers.) Ertz & Tehler, 2014 : (1, 7, 8, 13, 14, 17) *Pyrenula nitida* (Weigel) Ach., 1814 : (2, 17) Ramalina pollinaria (Westr.) Ach., 1810 : (6) Sclerophora peronella (Ach.) Tibell, 1984 : (7) Zwackhia viridis (Ach.) Poetsch & Schied., 1872 : (18)

> Received: 31.04.2017. Accepted: 02.10.2017.