

Phylogenetic analysis of the Red Sea tiger snake eel *Myrichthys maculosus* (Family Ophichthidae) by using 16S rRNA

Zeinab A. Mar'ie^{1,2}

- 1- Zoology Department, Faculty of Science, South Valley University, Egypt.
- 2- Science Department, Faculty of Education, Hurghada, South Valley University, Egypt.

zainab.khalek@sci.svu.edu.eg

ARTICLE INFO

Article History:

Received: Sept. 18, 2021
Accepted: Sept. 28, 2021
Online: Oct. 7, 2021

Keywords:

Ophichthidae
Myrichthys maculosus
16S rRNA
Phylogenetic analysis

ABSTRACT

The Red Sea tiger snake eel *Myrichthys maculosus* belongs to the family Ophichthidae, order Anguilliformes, represented with 11 relative species and two outgroups for phylogenetic analysis by mean of large subunit 16S rRNA sequence. *M. maculosus* contains a high concentration of AT (53.8%) more than GC (46.1%), higher also in other understudying Ophichthidae species. The genetic distance values among *M. maculosus* and other related species ranged from 0.0006 to 0.029, while among all species ranged from 0.006 to 0.037%. Overall the distance value was 0.167%. The smallest genetic distance (0.006) was between *M. maculosus* (MW435681) and the same one under the accession number of DQ645692, while the largest distance (0.036%) was between *Scolecenchelys breviceps* and *Ophichthus zophochir* & *O. apicalis*. The three phylogenetic trees Maximum likelihood (ML), Neighbour Joining (NJ), and Minimum Evolution (ME) applied here showed the same relations with slightly different in support values.

INTRODUCTION

Ophichthidae or snake eels is a family of order Anguilliformes, which comprise 19 families, 159 genera and nearly 938 species, most of them are marine, but some species entering rivers (Nelson *et al.*, 2016). The majority of Ophichthidae spends their time buried in sand and applies their sense of smell in hunting small fishes and crustaceans (Nelson, 1994). Comparing with morays snake eels is much less seen. The tiger eel (spotted snake eel) may reach to 1 m long or more, generally lives at depths between 1 and 25 m or more down, mostly in sandy areas by reefs (Debelius, 2011 ; Lieske and Myers, 2012).

Recently the phylogenetic relationship of eel species and revised taxonomic hierarchy, evaluated by short fragments of mitochondrial and also nuclear gene (Jamandne *et al.*, 2007; Tang and Fietitz 2012; Peninal *et al.*, 2017). The variation of eel species and evidenced the major lineages originated between the end of Cretaceous

and Early Eocene revealed by molecular studies as demonstrated by (Santini *et al.*, 2013). In 2018, Laskar *et al.* confirmed that molecular data are very strong tool in species identification.

16S rRNA sequencing able to reclassifying the organism in to another new species (Weisburg *et al.*, 1991). The large subunit 16S ribosomal RNA composed of 30 S small subunit of the mitochondrial ribosome in vertebrate has been reported 1640 bp long in fish (Naock *et al.*, 1996). In 2005 Vences *et al.* improved the role of 16S rRNA gene sequence in the bar coding of vertebrates. The mitochondrial (16S) gene extensively used in phylogenetic linkages of fishes at different taxonomic levels (Faddagh *et al.*, 2012; Mar'ie and Allam, 2019). Between species it is variable, but stable within same species Yang *et al.* 2014. The mitochondrial DNA has a circular structure thus allows more efficient amplification than the nuclear DNA (Panday *et al.*, 2014).

The point of this work is to confirm the strong role of large subunit "16S rRNA" sequencing in clarify the phylogenetic relationships of family Ophichthidae. Three phylogenetic methods; Maximum likelihood (ML), Neighbour Joining (NJ) and Minimum Evolution (ME) applied here to compare results with more widely used methods.

MATERIALS AND METHODS

Collection of the samples:

Tiger spotted eel collected by the helpful of fishermen in Hurghada , Red Sea in Egypt, then brought to the laboratory for the morphologically identification according to Randall (1982). The muscles tissues were individually isolated and preserved in -20°C until genomic DNA extraction.

DNA Extraction:

The genomic DNA was extracted from the muscles tissues using the DNA extraction method of QIAamp DNA Mini kit (Qiagen, Hidden, Germany) depending on the manufacturer's instructions.

PCR Conditions:

Polymerase chain reaction (PCR) amplification of genomic DNA was performed using forward (16sar: 3'-CGCCTGTTTAACAAAAACAT-5') and reverse (16sbr: 5'-CGCCTGTTTAACAAAAACAT-3') primers according to (Simon *et al.*, 1991). The PCR reactions were carried out with 10 pmoles of each primer, ~100 ng of genomic DNA and 12.5 µL PCR master mix (OnePCR™ ready-to-use, Catalogue Number: MB203-0100, GeneDireX, Miaoli County, Taiwan) in a final reaction volume of 25 µL. PCR reaction was carried out with, an initial denaturation for 2 minutes at 95°C, followed by 35 cycles for 30s at 95°C, annealing: for 30s at 50°C and an extension at 72 °C for 10 min. The PCR products were run on 1.3% agarose gel stained with ethidium bromide.

The Sequencing of PCR Products:

PCR amplification and agarose gel electrophoresis resulted in a single band of (581 bp.). DNA sequencing was achieved by Macrogen (Seoul, South Korea). The sequenced region of 16S rRNA in *Myrichthys maculosus* was submitted in the (GenBank/NCBI) under accession number (MW435681). using the same primer used for amplification Sequence was subjected to BLAST/N at the National Centre for Biotechnology Information (NCBI). Eleven related Ophichthidae species were selected in addition to two species as out group (Table 1).

Table (1): details of the present study tiger eel *Myrichthys maculosus* and its related species (Ophichthidae) with two out group (Congridae) from the GenBank/ NCBI based on (16S) genes sequences

No.	Species	Accession umber	Size of the amplicons bp	Similarity	Query coverage (%)
1	<i>Myrichthys maculosus</i>	MW435681.1	581	100%	100%
2	<i>Myrichthys maculosus</i>	DQ645692.1	991	99.83%	100%
3	<i>Ophichthus celebicus</i>	KX426295.1	645	91.11%	100%
4	<i>Callechelys catostoma</i>	JX242987.1	645	90.61%	100%
5	<i>Brachysomophis crocodilinus</i>	DQ645689.1	994	90.12%	100%
6	<i>Myrichthys breviceps</i>	AF455777.1	549	91.47%	94%
7	<i>Ophichthus puncticeps</i>	AF455781.1	549	91.34%	94%
8	<i>Echelus myrus</i>	DQ645690.1	989	88.96%	100%
9	<i>Scolecenchelys breviceps</i>	DQ645691.1	996	87.61%	100%
10	<i>Ophichthus zophochir</i>	AY952487.1	528	88.18%	91%
11	<i>Ophichthus shaoi</i>	LC506439.1	511	89.19%	88%
12	<i>Ophichthus apicalis</i>	KX426283.1	638	89.59%	100%
13	<i>Gnathophis longicauda</i>	DQ645704.1	990	87.29%	100%
14	<i>Gnathophis bathytapos</i>	JX242952.1	641	86.82%	100%

Sequence Alignments:

Sequences of (16S rRNA) were aligned with homologous sequences of related species from the GenBank database. Phylogenetic analyses were performed with MEGA version 7.0 18 (Kumar *et al.* 2016) using Maximum likelihood (ML), Neighbour Joining (NJ) and Minimum Evolution (ME) methods of trees construction and using 1000 bootstrap iterations (Felsenstein 1985). Sequence divergences were calculated using Kimura 2-parameter distances (Kimura 1980) to provide a graphical representation of divergence between species.

RESULTS

Sequence variation using 16S:

Genetic distance:

It's pretty obvious from the data in table (2) that *Scolecenchelys breviceps* has the longest DNA sequences (994 pb), whereas *Ophichthus shaoi* has the smallest DNA sequences (511 pb). The A+T content is higher than C+G content in *Myrichthys*

maculosus (53.8% and 46.1%, respectively) and in all other species. The A nucleotide got the largest average with 34.9.

Table 2: Nucleotide frequencies and its average of 16S rRNA sequence in *Myrichthys maculosus* and its related family (Ophichthidae) species with two out groups (Congridae).

Species	Base pair length	Nucleotide Number %				A+T content %	C+G content %
		T %	C%	A%	G%		
<i>Myrichthys maculosus</i>	581	21.3	24.4	32.5	21.7	53.8	46.1
<i>Myrichthys maculosus</i>	991	19.6	23.6	37.3	19.4	56.9	43.0
<i>Ophichthus celebicus</i>	645	20.1	25.8	33.0	21.0	53.1	46.8
<i>Callechelys catostoma</i>	645	22.0	23.8	34.4	19.8	56.4	43.6
<i>Brachysomophis crocodilinus</i>	994	19.4	23.8	35.6	21.2	55.0	45.0
<i>Myrichthys breviceps</i>	549	22.8	22.8	31.3	23.1	54.1	45.9
<i>Ophichthus puncticeps</i>	549	22.4	22.8	33.5	21.3	55.9	44.1
<i>Echelus myrus</i>	989	19.8	21.7	38.6	20.0	58.4	41.7
<i>Scolecenchelys breviceps</i>	996	20.3	23.4	36.7	19.6	57.0	43.0
<i>Ophichthus zophochir</i>	528	22.2	22.3	35.6	19.9	57.8	42.2
<i>Ophichthus shaoi</i>	511	21.5	24.3	33.9	20.4	55.4	44.7
<i>Ophichthus apicalis</i>	638	20.7	24.6	33.6	21.2	54.3	23.8
<i>Gnathophis longicauda</i>	990	18.9	23.1	37.2	20.8	56.1	43.9
<i>Gnathophis bathytopos</i>	641	20.7	25.0	34.7	19.6	55.4	44.6
Avg.		20.8	23.7	34.9	20.6		

The genetic distance values among *Myrichthys maculosus* and its related species of the same family ranged from 0.0006 to 0.029%, while among all species ranged from 0.006 to 0.037%. Overall the distance value among all current fish species was 0.167%. The smallest genetic distance was between *M. maculosus* and the same one under accession number of DQ645692, while the largest distance (0.036%) was between *Scolecenchelys breviceps* and *Ophichthus zophochir* & *O. apicalis* (Table 3).

Phylogenetic inference:

The Red Sea tiger snake eel "*Myrichthys maculosus*" was submitted to phylogenetic analysis by (16S) sequence together with 11 of other related species from GenBank/NCBI, representing the same species under accession number DQ645692, the same genera (*M. breviceps*), five species of the same genera *Ophichthus* (*O. celebicus*, *O. puncticeps*, *O. zophochir*, *O. shaoi*, and *O. apicalis*) and one species of genera; *Callechelys*, *Brachysomophis*, *Echelus*, and *Scolecenchelys*, in addition to 2 species as out group from family Congridae (*Gnathophis longicauda* & *G. bathytopos*).

Table 3: Pairwise distances based on (16S rRNA) sequence using Kimura 2- parameter among *Myrichthys maculosus* and its related family species additional to the out group.

		1	2	3	4	5	6	7	8	9	10	11	12	13	14
1	MW435681.1_Myrichthys maculosus		0.006	0.024	0.023	0.029	0.023	0.022	0.026	0.029	0.027	0.025	0.025	0.030	0.029
2	DQ645692.1_Myrichthys maculosus	0.009		0.025	0.023	0.029	0.024	0.023	0.028	0.031	0.027	0.027	0.025	0.031	0.030
3	KX426295.1_Ophichthus celebicus	0.118	0.129		0.027	0.034	0.034	0.027	0.031	0.034	0.030	0.014	0.026	0.029	0.030
4	JX242987.1_Callechelys catostoma	0.108	0.113	0.136		0.032	0.028	0.024	0.030	0.030	0.027	0.026	0.025	0.030	0.030
5	DQ645689.1_Brachysomophis crocodilinus	0.153	0.159	0.195	0.177		0.030	0.028	0.028	0.032	0.031	0.035	0.032	0.034	0.035
6	AF455777.1_Myrichthys breviceps	0.103	0.109	0.193	0.141	0.170		0.029	0.033	0.034	0.030	0.032	0.034	0.037	0.036
7	AF455781.1_Ophichthus puncticeps	0.097	0.108	0.141	0.120	0.141	0.146		0.025	0.025	0.027	0.026	0.031	0.029	0.028
8	DQ645690.1_Echelus myrus	0.133	0.144	0.185	0.168	0.145	0.179	0.117		0.026	0.032	0.032	0.030	0.029	0.029
9	DQ645691.1_Scolecenchelys breviceps	0.162	0.173	0.203	0.174	0.191	0.209	0.123	0.134		0.036	0.032	0.036	0.033	0.032
10	AY952487.1_Ophichthus zophochir	0.130	0.135	0.170	0.141	0.174	0.163	0.141	0.173	0.223		0.029	0.026	0.033	0.033
11	LC506439.1_Ophichthus shaoi	0.130	0.141	0.047	0.125	0.196	0.175	0.136	0.192	0.185	0.165		0.028	0.028	0.030
12	KX426283.1_Ophichthus apicalis	0.129	0.128	0.134	0.119	0.176	0.192	0.169	0.173	0.215	0.140	0.152		0.030	0.032
13	DQ645704.1_Gnathophis longicauda	0.178	0.184	0.166	0.173	0.203	0.233	0.161	0.161	0.201	0.196	0.166	0.178		0.015
14	JX242952.1_Gnathophis bathytopos	0.166	0.172	0.178	0.167	0.210	0.220	0.155	0.167	0.189	0.202	0.184	0.190	0.047	

Three phylogenetic methods; Maximum likelihood (ML), Neighbour Joining (NJ), and Minimum Evolution (ME) based on 16S rRNA gene were applied to confirm the relations among current Ophichthidae species. The three methods showed the same relations with slightly different in support values.

Myrichthys maculosus with the eleven related Ophichthyidae members showed distinct clustering, as follow: 1) *Myrichthys maculosus* with its identical DQ645692 clustering and cladded as sister species of *M. breviceps*. 2) *O. celebicus* & *O. shaoi* identical cladded together as a sister species of *C. catostoma* (single clad) and with *O. zophochir* and *O. apicalis* (sister clad). 3) *E. myrus* and *S. breviceps* clustering together as sister species of *B. crocodilinus* (single clad) and *O. puncticeps* (single clad). 4) The two out group family Congridae (formed a separate cluster from the rest (Figs. 1-3).

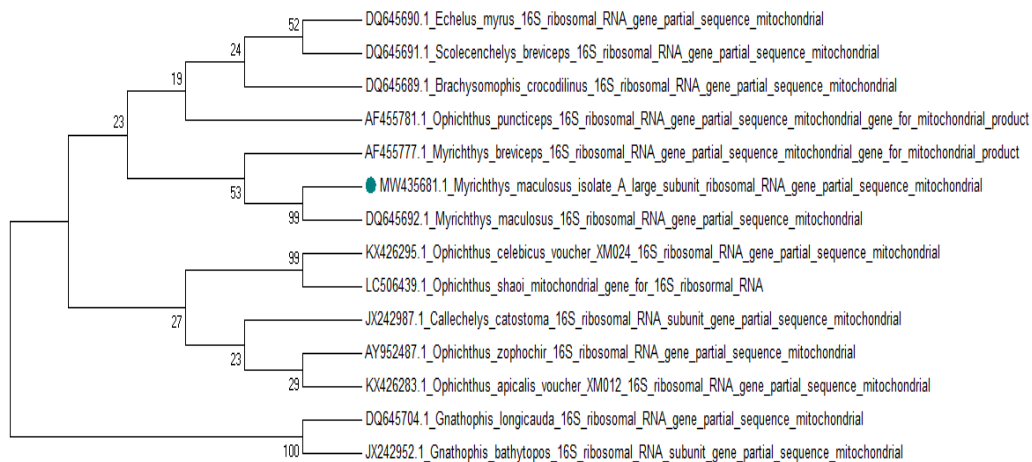
**Fig. 1.** Molecular phylogenetic tree by maximum likelihood method among tiger snake eel *Myrichthys maculosus* and related species of family Ophichthidae based on (16S) including the out group species.



Fig. 2. Molecular phylogenetic tree by minimum evolution method among tiger snake eel *Myrichthys maculosus* and related species of family Ophichthidae based on (16S) including the out group species.

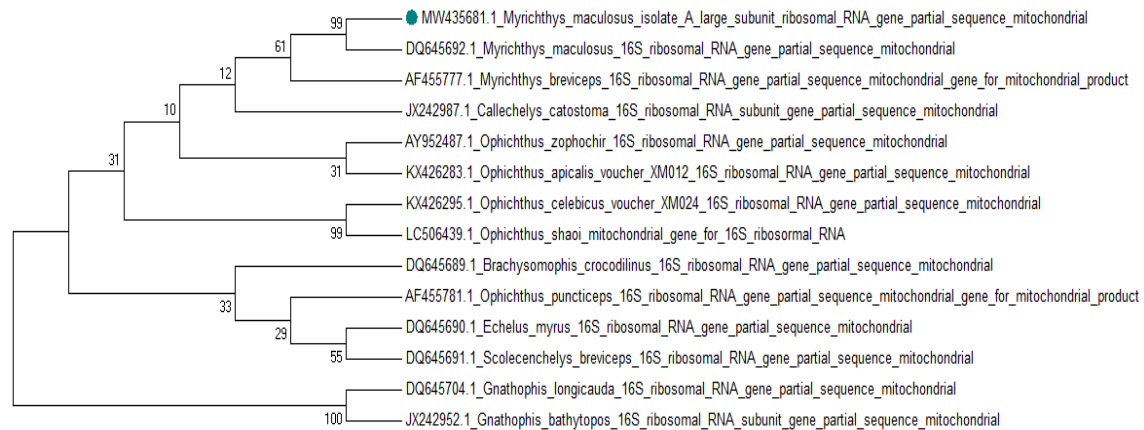


Fig. 3. Molecular phylogenetic tree by neighbour Joining method among tiger snake eel *Myrichthys maculosus* and related species of family Ophichthidae based on (16S) including the out group species

DISCUSSION

In 2009, Rojas *et al.* indicated that for species identification, the 16S rRNA gene which one of the mitochondrial gene considered a good marker. In the same vein, Yang *et al.* (2014) recorded that the genes of rRNA participate the homologous structures in species of organisms, from bacteria to humans due to similarity of several nucleotides, even though the genes exhibit inter and intra-specific nucleotide variations.

The present study employed the large subunit “16S rRNA” sequencing in clarify the phylogenetic relationships of family Ophichthidae, which utilized before by Miglietta *et al.* (2009); Moura *et al.* (2011) and Saad (2019) due to its facility to amplify in fish identification.

The concentration of A+T (53.8%) in *Myrichthys maculosus* was higher than the C+G (46.1%), like many pre-investment in fishes (**Perna and Kocher, 1995; Saccone et al., 1999 and 2000 and Mar'ie and Allam 2019**). The differences in CG amount among organism could affect reconstructions of evolutionary history because tree building techniques assigned unrelated species with similar GC content to the same group (**Mooers and Holmes, 2000**). In **2019, Saha et al.** illustrated the polymorphic loci analysis of 16S rRNA of the Bombay duck (*Harpadon nehereus*) order Aulopiformes, Family Synodontidae, and found that GC more than AT.

The smallest genetic distance reported here (0.006) was between the current tiger snake eel *M. maculosus* and the same one with accession number (DQ645692), whereas the furthest was *Scolecenchelys breviceps* (0.029). By using 12S & 16S rRNA, and with no support to the lineage based on morphology, **Lo'pezet al. (2007)** proposed a close relation between Colocongridae and Derichthyidae (Order Anguilliformes). **Song and Tang (2017)** investigated the complete mitochondrial genome of *Ophichthus rotundus* and found the AT>GC, and the high relation to *M. maculosus* by using NJ tree.

CONCLUSION

The application of 16S rRNA sequence analysis helps to clarify the interrelationship among species of the same family. The author suggested more comprehensive studies of Red Sea eels by using different molecular data.

ACKNOWLEDGMENT

The author is on much indebted Dr. Mohammad Allam Lecturer of molecular genetics, Department of Zoology, Faculty of science South Valley University for his ever-ready help and guidance throughout the entire breath- breaking stages of the work.

REFERENCES

- Debelius, H.** (2011). Red Sea Reef Guide, 5th Revised Edition. Hollywood Import & Export Inc.
- Faddagh, M. S.; Husain, N. A. and Al-Badran, A. I.** (2012). Usage Mitochondrial 16S rRNA Gene as Molecular Marker in Taxonomy of Cyprinid Fish Species (Cyprinidae: Teleostei). Journal of King Abdulaziz University, Marine Science, 23 (1): 39-49.
- Felsenstein, J.** (1985). Confidence limits on phylogenies: An approach using the bootstrap. Evolution, 39 (4): 783-791.

-
- Jamandre, B. W. D.; Shen, K. N.; Yambot, A. V. and Tzeng, W. N.** (2007). Molecular phylogeny of Philippine freshwater eels *Anguilla* spp. (Actinopterygii: Anguilliformes: Anguillidae) inferred from mitochondrial DNA. *Raffles B Zool.* 14:51–59.
- Kimura, M.** (1980). A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, 16 (2): 111-120.
- Kumar, S.; Stecher, G. and Tamura, K.** (2016). MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Molecular Biology and Evolution*, 33 (7): 1870-1874.
- Laskar, B. A.; Kumar, V.; Kundu, S.; Tyagi, K. and Chandra, K.** (2018). Taxonomic quest: validating two mahseer fishes (Actinopterygii: Cyprinidae) through molecular and morphological data from biodiversity hotspots in India. *Hydrobiologia*. 815:113–124.
- Lieske, E. and Myers, R. F.** (2012). *Coral Reef Guide Red Sea*. HarperCollins Publishers London, United Kingdom.
- Lo'pez, J. A.; Westneat, M. W. and Hanel, R.** (2007). The phylogenetic affinities of the mysterious anguilliform genera *Coloconger* and *Thalassenchelys* as supported by mtDNA sequences. *Copeia* 2007, pp.959–966.
- Mar'ie, Z. A. and Allam, M.** (2019). Molecular Phylogenetic Linkage for Nile and Marine Puffer Fishes Using Mitochondrial DNA sequences of Cytochrome b and 16S rRNA. *Egyptian Journal of Aquatic Biology & Fisheries* vol. 23(5): 67- 80.
- Miglietta, M. P.; Schuchert, P. and Cunningham, C.W.** (2009). Reconciling genealogical and morphological species in a worldwide study of the family Hydractiniidae (Cnidaria, Hydrozoa). *Zoologica Scripta*, 38 (4): 403-430.
- Mooers, A. Ø. and Holmes, E. C.** (2000). The evolution of base composition and phylogenetic inference, *Trends Ecol. Evol.*, 15: 365– 369.
- Moura, C. J.; Cunha, M. R.; Porteiro, F. M. and Rogers, A. D.** (2011). The use of the DNA barcode gene 16S mRNA for the clarification of taxonomic problems within the family Sertulariidae (Cnidaria, Hydrozoa). *Zoologica Scripta*, 40 (5): 520-537.
- Naock, K.; Zardoya, R. and Meyer, A.** (1996). The complete mitochondrial DNA sequence of the Bichir (*Polypterus ornatipinnis*), a basal ray-finned fish: Ancient establishment of the consensus vertebrate gene order. *Genetics*, 144 (3): 1165-1180.
- Nelson, J. S.** (1994). *Fishes of the world*. Third edition. John Wiley & Sons, Inc., New York. 600 pp.

- Nelson, J. S.; Grande, T. and Wilson, M.V. H.** (2016). Fishes of the world. 5th. edition. John Wiley & Sons, Inc., Hoboken, New Jersey and Canada.
- Panday, R.; Jha, D. K.; Thapa, N.; Pokharel, B. R. and Aryal, N. K.** (2014). Forensic Wild life parts and their product identification and individualization using DNA Barcoding. The open Forensic Science Journal 7: 6-13.
- Peninal, S.; Subramanian, J.; Elavarasi, A. and Kalaiselvam, M.** (2017). Genetic identification of marine eels through DNA barcoding from Parangipettai coastal waters. Genomics Data. 11:81–84.
- Perna, N. T. and Kocher, T. D.** (1995). Patterns of nucleotide composition at fourfold degenerate sites of animal mitochondrial genomes, J. Mol. Evol., 41: 353–358.
- Randall, J. E.** (1982). The Diver's Guide to Red Sea Reef Fishes. Publishing limited 20 Berkeley Street, Berkeley square London Wix 5AE, 96 pp.
- Rojas, M.; Gonzalez, I.; Fajarrdo, V.; Martin, I.; Hernandez, P. E.; Garcia, T. and Martin, R.** (2009). Authentication of meats from quail (*Coturnix coturnix*), pheasant (*Phasianus colchicus*), partridge (*Alectoris spp.*) and guinea fowl (*Numida meleagris*) using polymerase chain reaction targeting specific sequences from the mitochondrial 12S rRNA gene. Food Control 20, 896-902.
- Saad, Y. M.** (2019). Analysis of 16S mitochondrial ribosomal DNA sequence variations and phylogenetic relations among some Serranidae fishes. South African Journal of Animal Science, 49 (1): 80-89.
- Saccone, C.; Giorgi, C. D.; Gissi, C.; Pesole, G. and Reyes, A.** (1999). Evolutionary genomics in Metazoa: the mitochondrial DNA as a model system, Gene, 238, 195-209. **Saccone, C.; Gissi, C.; Lanave, C.; Larizza, A.; Pesole, G. and Reyes, A.** (2000). Evolution of the mitochondrial genetic system: an overview, Gene, 261: 153–159.
- Saha, S.; Ferdous, Z.; Jahan, H.; Khandaker, A. M.; Shahjahan, R. D. and Begum, R. A.** (2019). Polymorphic loci analysis of 16S rRNA ribosomal RNA gene of Bomby duck (*Harpadan nehereus*). Bangladesh J. zool. 47(1): 49-57.
- Santini, F.; Kong, X.; Sorenson, L.; Carnevale, G.; Mehta, R. S. and Alfaro, M. E.** (2013). A multi-locus molecular timescale for the origin and diversification of eels (Order: Anguilliformes). Mol Phylogenet Evol. 69(3): 884–894.
- Simon, C.; Franke, A. and Martin, A.** (1991). The polymerase chain reaction: DNA extraction *and amplification*, In Molecular Techniques in taxonomy, Eds. G.M. Hewitt, A.W.B. Johnston and J.P.W. Young. NATO AS1 series H 57: 329-355, 57: 329-355.

- Song, X. and Tang, W.** (2017). Complete mitochondrial genome of *Ophichthus roundus* (Anguilliformes: Ophichthidae). *Mitochondrial DNA PART B*: 2 (1): 176-177.
- Tang, K. L. and Fielitz, C.** (2012). Phylogeny of moray eels (Anguilliformes: Muraenidae), with a revised classification of true eels (Teleostei: Elopomorpha: Anguilliformes). *Mitochondrial DNA*. 24(1):55–66.
- Vences, M.; Thomas, M.; Vander, M. A.; Chiari, Y. and Vieites, D.R.** (2005). Comparative performance of the 16S rRNA gene in DNA barcoding of amphibians. *Frontiers in Zoology* 2: 5-6.
- Weisburg, W. G.; Barns, S. M.; Pelletier, D. A. and Lane, D. J.** (1991). 16S ribosomal DNA amplification for phylogenetic study. *Journal of Bacteriology* 173: 697-703.
- Yang, S. L.; Lin, R. Y.; Xu, L. X. and Cheng, L.** (2014). Analysis of polymorphisms of mitochondrial DNA D-loop and Mc1R gene in Chinese Wuchuan Black cattle. *J. Appl. Anim. Res.*, 42: 487–491.