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College of Tropical Agriculture and Human Resources  
Founding College of the University of Hawai'i  
Office of the Dean and Director of Research and Cooperative Extension

February 9, 2023

Mary Alice Evans, Director  
State Office of Planning and Sustainable Development  
Environmental Review Program  
235 South Beretania Street, Room 702  
Honolulu, HI 96813

Dear Ms. Evans:

Subject: Final Environmental Assessment and Finding of No Significant Impact for the Proposed Statewide Release of *Phymastichus coffea*

With this letter, the College of Tropical Agriculture and Human Resources (CTAHR), University of Hawai'i at Mānoa, transmits the Final Environmental Assessment and Finding of No Significant Impact (EA-FONSI) for the proposed statewide release of *Phymastichus coffea*, a biological control agent of coffee berry borer, *Hypothenemus hampei*, for publication in the next edition of the Environmental Notice.

The following documents are uploaded to the Environmental Review Program online submission system: publication form; EA-FONSI; published data demonstrating environmental safety of the proposed biological control agent; a cultural impact assessment; and, public comments on the Draft EA.

If any questions arise regarding this submission, please contact Dr. Mark G. Wright, Department of Plant and Environmental Protection Sciences, CTAHR at (808) 271-2037, or [markwrig@hawaii.edu](mailto:markwrig@hawaii.edu).

Yours sincerely

Dr. Ania M. Wiczorek  
Interim Dean and Director

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**From:** webmaster@hawaii.gov  
**Sent:** Friday, February 10, 2023 10:28 AM  
**To:** DBEDT OPSD Environmental Review Program  
**Subject:** New online submission for The Environmental Notice

**Follow Up Flag:** Follow up  
**Flag Status:** Flagged

<b>Action Name</b>
Final EA FONSI coffee berry borer biocontrol, <i>Phymastichus coffea</i>
<b>Type of Document/Determination</b>
Final environmental assessment and finding of no significant impact (FEA-FONSI)
<b>HRS §343-5(a) Trigger(s)</b>
<ul style="list-style-type: none"><li>(1) Propose the use of state or county lands or the use of state or county funds</li></ul>
<b>Judicial district</b>
Statewide
<b>Tax Map Key(s) (TMK(s))</b>
/Users/markwright/Dropbox (Wright lab)/Phymastichus permit revisions/TMK files UH farms Oahu.zip
<b>Action type</b>
Agency
<b>Other required permits and approvals</b>
USDA APHIS; Hawaii Dept of Agriculture
<b>Proposing/determining agency</b>
University of Hawaii, CTAHR
<b>Agency contact name</b>
Mark Wright
<b>Agency contact email (for info about the action)</b>
<a href="mailto:markwrig@hawaii.edu">markwrig@hawaii.edu</a>
<b>Email address or URL for receiving comments</b>
<a href="mailto:markwrig@hawaii.edu">markwrig@hawaii.edu</a>
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(808) 271-2037
<b>Agency address</b>
3050 Maile Way Rm 310, Honolulu, HI 96822 United States <a href="#">Map It</a>
<b>Was this submittal prepared by a consultant?</b>
No
<b>Action summary</b>

Field Release of *Phymastichus coffea* (Hymenoptera: Eulophidae) for the Biological Control of Coffee Berry Borer, *Hypothenemus hampei* (Coleoptera: Scolytinae) in Hawaii.

**Reasons supporting determination**

Clear demonstration of host specificity of the proposed biological control agent, with minimal likelihood of negative environmental impacts.

**Attached documents (signed agency letter & EA/EIS)**

- [FONSI-EA-Phymastichus-coffea.pdf](#)
- [DONE.SIGN-CTAHR-FinalEA-FONSI-submisison-cover-letter.pdf](#)

**Shapefile**

- The location map for this Final EA is the same as the location map for the associated Draft EA.

**Action location map**

- [TMK-files-UH-farms-Oahu.zip](#)

**Authorized individual**

Mark G Wright

**Authorization**

- The above named authorized individual hereby certifies that he/she has the authority to make this submission.

## Final Environmental Assessment

### Field Release of *Phymastichus coffea* (Hymenoptera: Eulophidae) for the Biological Control of Coffee Berry Borer, *Hypothenemus hampei* (Coleoptera: Scolytinae) in Hawaii



Peter A. Follett  
USDA-ARS  
U.S. Pacific Basin Agricultural Research  
Center  
64 Nowelo St.  
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Department of Plant and Environmental  
Protection Sciences  
University of Hawaii at Manoa  
3050 Maile Way,  
Honolulu, HI 96822

#### **Agency Contact**

Mark G. Wright  
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email: [markwrig@hawaii.edu](mailto:markwrig@hawaii.edu)

**February 2023**

## **Project Summary Sheet**

**Project Name:** Field Release of *Phymastichus coffea* (Hymenoptera: Eulophidae) for the Biological Control of Coffee Berry Borer, *Hypothenemus hampei* (Coleoptera: Scolytinae) in Hawaii.

**Proposing agency:** University of Hawaii at Manoa, State of Hawaii

**Project location:** Statewide

**Property Owner:** State of Hawaii

**State Land Use Classification:** Not applicable

**Agency Determination:** Finding of No Significant Impact (FONSI)

### **Agencies, Organizations, and other Stakeholders Consulted:**

#### **FEDERAL AGENCIES**

US House of Representatives, Representative Tulsi Gabbard

US Senate, Senator Maizie Hirono

US Forest Service

US Fish and Wildlife Service

USDA-APHIS

Smithsonian Institution, Dr. Lourdes Chamorro

#### **STATE AGENCIES**

DLNR Division of Forestry and Wildlife

Hawaii Department of Agriculture

#### **ORGANIZATIONS**

Hawaii Farm Bureau

Big Island Invasive Species Committee

Hawaii Invasive Species Committee

Kona Coffee Growers Association

Hawaii Coffee Growers Association

## **Addressing 13 Administrative Criteria for Significance from the Guide to the implementation and Practice of the Hawaii Environmental Policy Act 2012 edition:**

Each criterion as listed in the above guide is addressed below:

1. Involves an irrevocable commitment to loss or destruction of any natural or cultural resource

The release of *Phymastichus coffea* for the biological control of coffee berry borer will not result in any destruction or other negative impacts on natural or cultural resources in Hawaii. The draft EA details the results of work conducted to determine whether this prospective biological control agent will pose any environmental threats in Hawaii, showing that no negative impacts are expected. No native beetles in the subfamily Scolytinae were parasitized by *P. coffea* under no-choice testing conditions, providing high-confidence evidence that non-target impacts on native species are highly unlikely. This is addressed extensively in the attached DEA.

2. Curtails the range of beneficial uses of the environment

The release of *P. coffea* for biological control of coffee berry borer will not curtail beneficial uses of the environment in any way. On the contrary, it will increase the viability of coffee farming in Hawaii, thus sustaining the range of beneficial uses of the environment.

3. Conflicts with the state's long-term environmental policies or goals and guidelines as expressed in [Chapter] 344, HRS, and any revisions thereof and amendments thereto, court decisions, or executive orders

There is no conflict with the State's long-term environmental policies or goals.

4. Substantially affects the economic or social welfare of the community or State

No negative impacts on the social welfare of communities is anticipated. The biological control of coffee berry borer will likely increase the economic and social welfare of communities. These benefits will primarily be realized through financial savings resulting from effective invasive pest management using options other than pesticide applications.

5. Substantially affects public health

No negative impacts on public health are expected or likely. The biological control agent is a small non-stinging Hymenoptera species, restricted to using coffee berry borer, and possibly some closely related invasive insect species, as hosts. The biological control agent poses no potential risk to human health.

6. Involves substantial secondary impacts, such as population changes or effects on public facilities

The release of this biological control agent has no potential to impact human populations or facilities. No secondary impacts will occur.

7. Involves a substantial degradation of environmental quality

No degradation of environmental quality will occur.

8. Is individually limited but cumulatively has considerable effect upon the environment or involves a commitment for larger actions

No negative environmental impact is anticipated based on data showing that *P. coffea* is limited to the genus *Hypothenemus* as hosts. There are no native *Hypothenemus* species in Hawaii.

9. Substantially affects a rare, threatened, or endangered species, or its habitat

No threatened, rare, or endangered species, or their habitat, will be threatened by this biological control agent. No native species of in the subfamily Scoytinae were parasitized during careful screening of a broad selection of beetle species present in Hawaii. The data showed that only closely related species in the non-native genus *Hypothenemus* could were parasitized by *P. coffea*. The wasps showed no behavioral response to native beetles and where unable to utilize them as hosts. These results were published as:

Yousuf, F., Follett, P.A., Gillett, C.P.D.T., Honsberger, D., Chamorro, L., Johnson, T.M., Jaramillo, M.G., Machado, P.B. & Wright, M.G. 2021. Limited host range in the idiobiont parasitoid *Phymastichus coffea*, a prospective biological control agent of the coffee pest *Hypothenemus hampei* in Hawaii. *Journal of Pest Science* 94: 1183-1195.  
<https://doi.org/10.1007/s10340-021-01353-8>

10. Detrimentally affects air or water quality or ambient noise levels

No negative impacts on water quality or noise levels will occur.

11. Affects or is likely to suffer damage by being located in an environmentally sensitive area such as a flood plain, tsunami zone, beach, erosion-prone area, geologically hazardous land, estuary, fresh water or coastal waters

No effects on environmentally sensitive areas will occur.

12. Substantially affects scenic vistas and view planes identified in county or state plans or studies

No impacts on scenic vistas or view planes will occur.

13. Requires substantial energy consumption

This project does not require substantial energy consumption.



## **COMMENTS on DRAFT EA, INCLUDING RESPONSES TO SUBMITTERS**

Comments received on the draft EA, and responses are attached on the next four pages. No edits were required on the draft EA for the final EA. All comments were received by email, these are attached below. Two comments were received expressing concern about the proposed release of *Phymastichus coffea*, and two in support of the release. My responses to those expressing concern are included.

**From:** Mark Wright markwrig@hawaii.edu  
**Subject:** Re: JADAM  
**Date:** December 16, 2022 at 4:09 PM  
**To:** Dana Melina Keawe danakeawe@gmail.com

---



Dear Ms. Keawe

Thank you for your comment. We are strongly supportive of a range of options for CBB management, including potential JADAM solutions.

We see biological control as a safe and sustainable tool to include in the integrated CBB management options available to Hawaii growers.

Aloha, Mark

Mark G Wright Ph.D.  
Professor and Extension Entomologist  
Entomology Section  
Department of Plant and Environmental Protection Sciences  
University of Hawaii at Manoa

Lab website: <http://wrightlabuh.weebly.com>

On Dec 15, 2022, at 5:49 PM, Dana Melina Keawe <[danakeawe@gmail.com](mailto:danakeawe@gmail.com)> wrote:

JADAM is the best way to combat CBB. You can subscribe to their Utube channel to see their research project which proved JADAM works to combat CBB. And also get the recipe solution.

Introducing a wasp to Hawai'i is NOT a good idea.

Dana Keawe

**From:** Mark Wright markwrig@hawaii.edu  
**Subject:** Re: comment on field release of *Phymastichus coffea*  
**Date:** December 16, 2022 at 4:14 PM  
**To:** Peter Mathews petermathews@yahoo.com



Dear Mr. Mathews

Thank you for your comment and opinion.

We share your concerns about the native insect fauna, and have taken significant measures to ensure that our proposed release will not have any negative impacts.

Long-term data shows that biological control introductions to Hawaii made subsequent to 1970 have no negative impacts on native species. This is due in substantial part to the rigorous non-target screening conducted since 1970, and becoming increasingly rigorous through the latter 1900s.

We provide considerable evidence in the DEA based on non-target screening assays and target insect phylogenies showing that impacts on native species from *Phymastichus coffea* are highly unlikely.

Yours sincerely,

Mark

Mark G Wright Ph.D.  
Professor and Extension Entomologist  
Entomology Section  
Department of Plant and Environmental Protection Sciences  
University of Hawaii at Manoa

Lab website: <http://wrightlabuh.weebly.com>

On Dec 15, 2022, at 8:38 PM, Peter Mathews <[petermathews@yahoo.com](mailto:petermathews@yahoo.com)> wrote:

Mr. Wright,

I am writing to express my concerns regarding release of the above wasp through the State for the purpose of controlling the coffee berry borer.

I understand this has been successful elsewhere, and a study was performed at Volcano.

There is a long history of unintended consequences when introducing one species to counter another. I do not support this release. Hawaii's endemic wildlife is already at risk from many threats, and introducing yet another organism into the ecosystem in a hopes to achieve a specific effect is misguided and potentially dangerous.

Sincerely,

Peter Mathews

**From:** HC Bittenbender hcbitt@hawaii.edu  
**Subject:** Proposed release of *Phymastichus coffea* for control of the Coffee Beery Borer  
**Date:** December 22, 2022 at 3:17 PM  
**To:** Mark G. Wright markwrig@hawaii.edu  
**Cc:** Ania Wieczorek ania@hawaii.edu, Sharon Hurd sharon.k.hurd@hawaii.gov

SB

Aloha Mark Wright,

I am submitting my comments in strong support of the proposed release of the parasitic wasp *Phymastichus coffea* to control the Coffee Berry Borer (*Hypothenemus hampei*) aka CBB.

As a retired CTAHR extension specialist responsible for coffee (1986- 2018) and macadamia (1986-1995) I am familiar with the serious crop loss of green coffee beans caused by CBB.

In addition I am familiar crop losses caused by related *Hypothenemus* spp. The Black Twig Borer (*H. seriatus*) began attacking anthuriums in the 1960s and was later found attacking coffee trees in Kona, this was reported by the late CTAHR entomologist Arnold Hara. Until now our only control for Black Twig borer on coffee has been pruning the damaged branches and destroying the prunings to kill the adult borer, larvae and eggs. In the 1986 I referred damaged macadamia nuts to the late CTAHR entomologist Jack Beardsley, who identified the insect in the nuts as *H. obscurus* initially named the macadamia shot hole borer later the Tropical nut borer. To date our only control measure has been harvesting the nuts from the ground within 3 weeks of dropping and drying immediately; this is much earlier than the previous six week harvest interval prior to the arrival Tropical Nut Borer.

## **Draft Environmental Assessment: Field Release of *Phymastichus coffea* (Hymenoptera: Eulophidae) for the Biological Control of Coffee Berry Borer, *Hypothenemus hampei* (Coleoptera: Scolytinae) in Hawaii**

section 3.8 Table 1. 'Previous reports of parasitism of Scolytinae species by *Phymastichus coffea* in no-choice laboratory assays' states that *P. coffea* also parasitizes the Tropical Nut Borer and Black Twig Borer.

Releasing *P. coffea* provides Hawaii farmers an opportunity to biologically control three *Hypothenemus* species for which the current recommended control practices add substantial labor and materials costs to our farmers.

Seldom does the opportunity arise to control, if only partially, three pests at minimal costs to farmers. This is a great contribution to Hawaii agriculture.

If I should direct my letter of support to another agency or individual please let and I will do so.

H.C. "Skip" Bittenbender, Ph.D.  
Emeritus Extension Specialist for Cacao, Coffee and Kava

Tropical Plant and Soil Sciences  
CTAHR-College of Tropical Agriculture and Human Resources



**Maui Coffee Association, P.O. Box 1089, Kula, HI. 96790**

Aloha Mark,

Thank you very much for making the draft environmental review on the introduction of a biologic control for CBB available. Thank you also for taking the initiative to get this going. We farm coffee on Maui and we have many small farmers all over the island. CBB has caused a tremendous amount of damage leading some farms to abandon their orchards completely. On our farm we have used Mycotrol and pain staking sanitation practices (removal of all cherries at the end of the season) to control CBB infestation with very limited success. The problem of using Mycotrol/Botanigard is the timing of application relative to CBB activity and having the right environmental conditions e.g. no wind and no post-application rainfall. This results in low efficiency.

I am cautiously optimistic that the introduction of the host-specific wasp will help us immensely. In particular it may help keep populations in check not only on farms but in the widespread feral populations of coffee which provide an nearly limitless reservoir of CBB such that even if we get our farms clean their is always a healthy population ready to move in. The feral populations are completely unmanaged (a similar problem is present in Kona) and unless we have a control mechanism that works well on both managed and unmanaged coffee trees we will never get a leg up. We are seeing high infestation rates on our farm which has caused 30% reduction in yield AND requires extensive hand sorting in parchment stage to maintain bean quality prior to the arrival of CBB. It is a desperate state and we are looking at stump pruning all of our coffee this year to try and break the cycle of CBB...other farms have adopted similar approaches.

The host-specific wasp seems like a viable solution. A quick look at scientific articles on the web showed studies in Mexico where CBB was present, the introduction of the wasp reduced losses from 60% to less than 5% with no impacts on unintended targets. Maui Coffee Association is very supportive of this initiative and appreciate your work to get this biocontrol introduced and your work to determine thoroughly the lack of effects on unintended species.

Yours sincerely,

Gerald M. Ross, president, Maui Coffee Association

End of comments received on draft EA

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## APPENDICES

**Appendix A:** Yousuf, F., Follett, P.A., Gillett, C.P.D.T., Honsberger, D., Chamorro, L., Johnson, T.M., Jaramillo, M.G., Machado, P.B. & Wright, M.G. 2021. Limited host range in the idiobiont parasitoid *Phymastichus coffea*, a prospective biological control agent of the coffee pest *Hypothenemus hampei* in Hawaii. *Journal of Pest Science* <https://doi.org/10.1007/s10340-021-01353-8>

**Appendix B:** Cultural Impact Assessment for Proposed Statewide Release of *Phymastichus Coffea* to Control Coffee Berry Borer

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## 1. Summary

The coffee berry borer (CBB), *Hypothenemus hampei* (Ferrari), (Coleoptera: Curculionidae: Scolytinae) is the most destructive insect pest of coffee globally. Though endemic to Central Africa, CBB is now found in almost every coffee-producing country in the world. In 2010, it first invaded the island of Hawai'i where high quality coffee is the second largest cash crop, valued at more than \$55 million during the 2020-21 season. Coffee berry borer has since invaded coffee on the islands of Oahu, Maui and Kauai. Coffee crop loss due to CBB is estimated at \$7.7 million. CBB has had the effect of making coffee farming more intensive and less profitable: damage causes significant losses in yield and alters the flavor profile of salvageable coffee beans. If left unmanaged, CBB can damage >90% of the crop.



Figure 1: CBB gallery inside bean, with visible eggs and larvae

The primary means of control in Hawaii is using the microbial insecticide *Beauveria bassiana* and sanitation (removal of all coffee berries after harvest). Biological control of CBB using parasitoids has been conducted in many countries around the world, especially in Latin America (Mexico south to Brazil) and has potential for Hawaii. One of the most promising agents is a parasitoid wasp, *Phymastichus coffea* LaSalle (Hymenoptera: Eulophidae). *Phymastichus coffea* is a primary, gregarious, idiobiont endoparasitoid of CBB adult females. After being parasitized by *P. coffea*, females stop oviposition and usually die after 4-12 days. Therefore, *P. coffea* was chosen as a potential biological control agent and was brought from Colombia into a quarantine containment facility in Volcano, Hawaii for host range testing to determine whether the parasitoid might attack non-target species and thereby pose a risk to the environment. Using no-choice tests, 43 different species of Coleoptera were tested, including 23 scolytines (6 *Hypothenemus* species, 7 native *Xyleborus* species, and 10 others), and 4 additional Curculionidae. *P. coffea* was only able to parasitize the target host *H. hampei* and 4 other adventive species of *Hypothenemus*: *H. obscurus*, *H. seriatus*, *H. birmanus* and *H. crudiae*. *Hypothenemus hampei* had the highest parasitism rate and shortest parasitoid development time of the five parasitized *Hypothenemus* spp. Parasitism and parasitoid

emergence decreased with decreasing phylogenetic relatedness of the *Hypothenemus* spp. to *H. hampei*, and the most distantly related species, *H. eruditus*, was not parasitized. There are no native Hawaiian species in the genus *Hypothenemus*. *Phymastichus coffea* appears to be host-specific at the genus level, and only able to survive on species closely related to *H. hampei*. Therefore, release of *P. coffea* for control of CBB in Hawaii coffee should cause no harm to the environment.

This Draft Environmental Assessment (DEA) was prepared by UH and the USDA-ARS (Hilo, Hawaii) for HDOA Plant Quarantine Branch and submitted to the Office of Environmental Quality Control (OEQC), Department of Health, State of Hawaii, to comply with the provisions of Hawaii Revised Statutes, HAR Chapter 11-200.1, Environmental Impact Statements.

## **I. Proposed Action**

An application was submitted by the USDA-ARS, Hilo, Hawaii, to the HDOA Plant Quarantine Branch, 1849 Auiki Street, Honolulu, Hawaii 96819, for a permit to introduce *Phymastichus coffea* LaSalle (Hymenoptera: Eulophidae) into the State of Hawaii under the provisions of Hawaii Revised Statutes, Chapter 141, Department of Agriculture, and Chapter 150A, Plant and Non-Domestic Animal Quarantine. *Phymastichus coffea* will be used to control the coffee berry borer, *Hypothenemus hampei* (Ferrari) (CBB) (Coleoptera: Scolytinae), a serious invasive pest of coffee in Hawaii.

### **1.1 Purpose of release**

The USDA-ARS proposes to introduce the parasitoid wasp, *Phymastichus coffea* from containment into the natural environment of the State of Hawaii as a biological control agent to suppress infestations of the coffee berry borer, *Hypothenemus hampei*. Host specificity studies have been completed in the USDA Forest Service quarantine facility at Hawaii Volcanoes National Park. In addition to its natural host, coffee berry borer, *P. coffea* was found to attack four other species in the genus *Hypothenemus*. The parasitoid did not attack any of the native and beneficial beetles tested. It is expected that *P. coffea* will become established as a classical biological control agent, providing sustained population suppression of CBB in Hawaii. If establishment of *P. coffea* is variable or unsuccessful in some areas, additional releases will be made, or augmentative releases might be considered in some locations.

### **1.2 Need for release**

The coffee berry borer is the most serious pest of coffee in most coffee producing countries. In Hawaii, coffee berry borer was first reported in 2010 from South Kona and soon spread throughout Hawaii island coffee farms and to the other islands. The coffee berry borer severely affects the yield and quality of the coffee and it is an important constraint on production and development of the crop. The current crop losses of coffee due to the coffee berry borer infestation in Hawaii is estimated at \$7.7 million (HDOA 2019). If left uncontrolled coffee berry borer can infest >90% of coffee berries. The control of this pest with pesticides is expensive and has limited success if the borer has reached the endosperm of the seeds (Vega et al., 2015). Biological control is a sustainable option to manage the coffee berry borer. *Phymastichus coffea* has proven to be an effective biological control agent of coffee berry borer in other coffee growing

regions in the world (Escobar-Ramirez et al., 2019). Furthermore, *P. coffea* is the only parasitoid tested thus far that has been shown to reduce yield loss from CBB damage (Infante et al., 2013). *Phymastichus coffea* has the potential to be an effective biological control agent against the coffee berry borer in Hawaii.



Figure 2: Adult CBB as found inside a green berry

### 1.3 Reasons for choice of entomophagous biological control agent

The parasitoids, *Cephalonomia stephanoderis* Betrem, *C. hyalinipennis* Ashmead, *Prorops nasuta* Waterston (Hymenoptera:Bethylidae), *Heterospilus coffeicola* Schneideknecht (Hymenoptera: Braconidae), and *Phymastichus coffea* LaSalle (Hymenoptera: Eulophidae), all of African-origin, have been introduced in many coffee producing countries, particularly in Central and South America (Klein-Koch et al. 1988; Barrera et al. 1990; Baker 1999; Jaramillo et al. 2005; Portilla and Grodowitz 2018), but none have been released in Hawaii.

*Phymastichus coffea* was chosen as the best candidate parasitoid in Hawaii because of its previously reported high host specificity and ability to significantly reduce and regulate *H. hampei* populations in the field (Gutierrez et al. 1998; López-Vaamonde and Moore 1998; Castillo et al. 2004a,b; Rodríguez et al. 2017). In field cage studies in Mexico and Costa Rico, *P. coffea* proved to be the most promising biological control agent against *H. hampei* with parasitism rates as high as 95% (Espinoza et al. 2009; Infante et al. 2013).



Figure 3: *Phymastichus coffea* parasitizing CBB in berry. Photo courtesy of Cenicafé.

To date, *P. coffea* has been released in 12 countries as a classical biological control agent (Bustillo et al. 1998; Damon 2000; Jaramillo et al. 2005; Vega et al. 2015). *P. coffea* is native to Africa and present in most coffee producing countries on that continent. It is a primary, gregarious, idiobiont endoparasitoid of adult *H. hampei* females with a high capacity for host-discrimination (Feldhege 1992; Infante et al. 1994; López-Vaamonde and Moore 1998; Castillo et al. 2004). Two laboratory studies reported that in addition to *H. hampei*, *P. coffea* parasitizes other *Hypothenemus* spp. such as *H. seriatus* and *H. obscurus* (López-Vaamonde and Moore, 1998), and *H. eruditus* Westwood and *H. crudiae* (Panzer) (Castillo et al. 2004a,b). However, parasitism of closely related species in the field has not been reported (Escobar-Ramírez et al. 2019).

#### **1.4 Specific location of rearing/containment facilities and name of qualified personnel operating the facility**

*Phymastichus coffea* was obtained from Cenicafé in Colombia under USDA APHIS PPQ, permit no. P526P-18-00696 and brought into a fully certified quarantine insect containment facility managed by the USDA Forest Service at Hawaii Volcanoes National Park, Volcano, Hawaii, for host-specificity testing. The director and primary user of this facility is Dr. M. Tracy Johnson of the USDA Forest Service, Institute for Pacific Island Forestry.

#### **1.5 Timing of the release as well as factors that affect the timing of release**

If *Phymastichus coffea* is approved for release, Cenicafé (Colombia) will supply wasps for the initial releases. Cenicafé is currently mass rearing *P. coffea* on field-collected CBB and can provide *P. coffea* at any time of year. *P. coffea* will be released in coffee on all islands where CBB occurs (Hawaii, Oahu, Maui, Kauai). *Phymastichus coffea* will be released and monitored for establishment in a classical biological control program. In the future, augmentative releases of *P. coffea* from Cenicafé may be possible if documentation and certification of their rearing process and facility demonstrates that the colony is pure and quality control ensures there will be no contamination. Currently, trapping and sampling of infested coffee fruits is conducted to monitor

*H. hampei* flights and optimize timing of *Beauveria bassiana* applications for control (Aristizabal et al. 2016). After *H. hampei* bores into the coffee berries it is protected and difficult to control with biopesticides or conventional insecticides. To achieve maximum *P. coffea* parasitism in the field, releases should be made at times when *H. hampei* adults are active (e.g. when trap catches are high, or female *H. hampei* are actively boring into fruits) and the coffee crop is at a susceptible stage. Optimal timing of releases may differ for different elevations due to *H. hampei* population dynamics (Hamilton et al. 2019). Studies suggest *P. coffea* may be susceptible to *B. bassiana*, however (Barrera 2005; Castillo et al. 2009; Ruiz et al. 2011), so releases should be timed to avoid *B. bassiana* applications or used in alternation with *B. bassiana* against *H. hampei*. If *P. coffea* is highly effective, then dependence on *B. beauveria* applications could be reduced dramatically.

### **1.6 Location of planned first release**

First releases will be made in the South Kona district of the Big Island of Hawaii in the main coffee growing region as it is close to the USDA ARS laboratory and University of Hawaii experiment station which will facilitate monitoring. Other sites may also be selected depending on the number of parasitoids available.

According to the simulation model output, *P. coffea* is predicted to provide feasible control of coffee berry borers in areas where flowering periods are frequent throughout the year (Rodríguez et al. 2017). In Hawaii, Maui and Oahu due to relatively constant temperatures with abundant rainfall, coffee flowering and harvesting seasons may be irregular. However, Kona is different with more pronounced seasonal conditions. So, depending on the flowering season, releases of *P. coffea* will be made approximately 70 and 170 days after flowering periods (when coffee berries have >20% dry matter content), or at times when CBB adults are active (e.g. trap catches are high) and the crop is at a susceptibility stage.

*P. coffea* may be sensitive to *Beauveria bassiana*, the fungal biopesticide used against the coffee berry borer and to other insecticides (Castillo et al.2009; Barrera 2005; Gómez et al. 2011). Therefore, it is important to make sure that the parasitoids are not released just before or just after or concurrently with pesticides to prevent any negative effects on survivorship and establishment.

### **1.7 Methods to be used after agent importation**

Newly emerged female *P. coffea* will be collected into plastic containers covered with muslin impregnated with a 50% honey-water solution. The containers will be placed in a cool box and transported to the field. The parasitoids will be released in the center of the coffee field. A ratio of 1 parasitoid per 10 hosts (determined from random field sampling for infested coffee berries) or less would be ideal (Espinoza et al. 2009). Once the parasitoids are released, they will disperse naturally to search for new coffee berry borer hosts to parasitize.

### **1.8 Methods to be used for disposing of any host material, pathogens, parasites, parasitoids, and hyperparasitoids accompanying an import**

Because of its short life span (2-4 days), *P. coffea* will be shipped from Cenicafé as parasitized adult CBB into quarantine containment and reared through a generation to ensure that no hyperparasitoids. A sample of parasitized CBB will be tested for plant pathogens, e.g.

coffee leaf rust, by USDA ARS scientists. Parasitized CBB adults shipped to Hawaii for host range testing exhibited low *P. coffea* emergence (5-20%). Studies are underway with Cenicafé to optimize shipping conditions for improved parasite emergence. No pathogens or hyperparasitoids have been observed at Cenicafé on *P. coffea*-parasitized CBB. *P. coffea* shipments will not contain any plant material, e.g. coffee berries. Parasitized CBB may be shipped on artificial diet, which will be autoclaved after parasitoid emergence.

### 1.9 Agencies or individuals that will be involved in the release and monitoring

USDA-ARS (Peter Follett, Melissa Johnson), University of Hawaii (Mark Wright, Andrea Kawabata, graduate students), and the Hawaii Department of Agriculture (Mohsen Ramadan, Juliana Yalamar) will be involved in the release and evaluation of *P. coffea* (establishment, dispersal, parasitism rates, behavior, integration with coffee IPM practices, nontarget effects).

## 2. Target Pest Information

### 2.1 Taxonomy: scientific name, full classification, synonymy, common name and sufficient characterization to allow unambiguous recognition

Order: Coleoptera

Family: Curculionidae

Subfamily: Scolytinae

Genus: *Hypothenemus*

Species: *H. hampei*

Common name: coffee berry borer (CBB)

Binomial name: *Hypothenemus hampei* (Ferrari, 1867)

Synonyms

*Cryphalus hampei* Ferrari, 1867

*Stephanoderes hampei* Ferrari, 1871

*Stephanoderes coffeae* Hagedorn, 1910

*Xyleborus coffeivorus* Van der Weele, 1910

*Xyloborus coffeicola* Campos Novaes, 1922

*Hypothenemus coffeae* (Hagedorn)

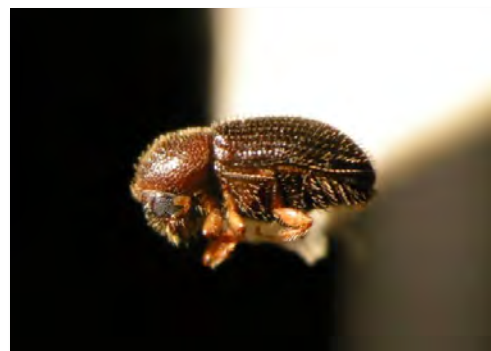


Figure 4: CBB. Courtesy HDOA

The genus *Hypothenemus* is one of the most speciose in the Scolytinae and common in all tropical and subtropical areas. The taxonomic characters useful in identifying *Hypothenemus hampei* and related members of the genus is presented in Vega et al. 2015 (“The Genus *Hypothenemus*, with emphasis on *H. hampei*, the coffee berry borer” pp. 427-494, In *Bark Beetles: Biology and Ecology of Native and Invasive Species* [F. E. Vega and R. W. Hofstetter, Eds.], Academic Press, San Diego). The information below is excerpted from this book chapter.

Most *Hypothenemus* species are very small (<2 mm long), poorly described, and difficult to distinguish. Several species are globally distributed, undoubtedly aided by human activities.

Although the vast majority of *Hypothenemus* species live innocuously in twigs, some have become important pests, most notably the coffee berry borer *Hypothenemus hampei* (Ferrari), which lives inside the coffee berry and consumes the seeds, and the tropical nut borer *Hypothenemus obscurus* (F.), which attacks a range of seeds and fruits.

The frons of *H. hampei* may have a broad, indistinct frontal groove, or no groove at all. There are usually four marginal asperities. The setae on the pronotum are mixed, with some slightly flattened. The shape of the pronotum, viewed from above, is slightly more narrowly rounded (i.e., more triangular) than the similar *Hypothenemus* species. The elytral declivity of *H. hampei* is much more broadly rounded than in the similar species, without a distinct transition from the elytral disc. When viewed laterally, the declivity takes up more than half of the length of the elytra, whereas in the similar species, the elytral disc takes up more than half of the length. As with most *Hypothenemus*, the interstitial bristles are prominent and in almost perfectly uniseriate rows. The shape of the interstitial bristles, however, is distinctive, and differentiates the coffee berry borer from most other *Hypothenemus* species. The bristles are long, narrow, and slightly flattened. The tip of each bristle is square, and not much wider than the rest of its length. The bristles on the elytral disc are not much shorter than those on the declivity. Males are smaller with reduced eyes. The interstitial bristles are relatively long, and often not in distinct rows.

Phylogenetically, *H. hampei* is in a clade distantly related to native Hawaiian Scolytinae species, which are all within the Tribe Xyleborini (Johnson et al. 2018). There are other *Hypothenemus* species in Hawaii, all adventive. While there are anecdotal reports of *H. hampei* feeding on plants other than coffee (e.g. *Leucaena leucocephala*), there is no indication that they could complete their life cycle in those hosts. No native Scolytinae are known to utilize those plants.

## **2.2 Economic impact and benefits of the target pest: *Hypothenemus hampei***

The coffee berry borer (CBB), *Hypothenemus hampei* (Ferrari), (Coleoptera: Curculionidae: Scolytinae) is the most destructive insect pest of coffee globally, inflicting economical losses of over US\$500 million annually. Though endemic to Central Africa, CBB is now found in almost every coffee-producing country in the world. In 2010, it first invaded the island of Hawai'i where high quality coffee is the third largest cash crop, valued at more than \$43 million during the 2017-18 season. Coffee berry borer has since invaded coffee on the islands of Oahu and Maui and most recently Kauai. Coffee crop loss due to CBB is estimated at \$7.7 million. CBB has had the effect of making coffee farming more intensive and less profitable: damage causes significant losses in yield and alters the flavor profile of salvageable coffee beans. If left unmanaged, CBB can damage >90% of the crop.

CBB has been found on several incidental non-crop host plants in Hawaii such as haole koa (*Leucaena leucocephala*), black wattle (*Acacia decurrens*), and red fruit passionflower or love-in-a-mist (*Passiflora foetida*). However, to date researchers have found only a very low incidence of CBB in any of these other plants, and no signs of CBB reproduction in any of them. Wild (uncultivated) coffee plants are a significant reservoir for CBB populations (Messing 2012).



### **2.3 Biology and reproductive potential of the target pest**

*Hypothenemus hampei* attacks coffee berries when the dry matter content of the endosperm, which increases with age, exceeds 20% (Jaramillo et al. 2005). After finding a suitable berry host, *H. hampei* bores into the coffee fruit through the central disc and excavates galleries where it lays eggs. The offspring develop inside the seeds and feed on the endosperm tissue (Damon 2000), reducing both coffee yield and quality. *H. hampei* feeding damage can also cause premature fall of berries younger than 80 days (Decazy 1990). *H. hampei* adults boring into the berry may remain in the 'A' position (Jaramillo et al. 2006) with the abdomen half exposed outside the berry potentially for weeks waiting for the dry matter content to reach 20% (Jaramillo et al. 2005). Females are synovigenic and lay eggs in batches of 2-3 eggs beginning three days after penetration into the seed. About 31-119 eggs are laid within a single berry over a period of 3 weeks. Soon after egg laying commences wing muscles of the female degenerate, preventing the colonization of other berries (Ticheler 1963). Multiple generations may occur in the coffee berry under Hawaii conditions. Waterhouse and Norris (1989), suggested that females may leave the berry when all the seed tissue is consumed or deteriorated in some way, or when her progeny begin to emerge, in order to continue egg-laying in another berry. After *H. hampei* bores into the coffee berries it is protected and difficult to control with biopesticides or conventional insecticides.

### **2.4 Global distribution of the target pest**

The coffee berry borer (CBB), *Hypothenemus hampei* (Ferrari), (Coleoptera: Curculionidae: Scolytinae) is the most destructive insect pest of coffee globally. Though endemic to Central Africa (likely the Ethiopian Highlands), CBB is now found in almost every coffee-producing country in the world. Coffee berry borer was first discovered in 1867 in France in coffee seeds traded from unknown origin (Waterhouse and Norris 1989), and in Africa it was reported in 1901 from Gabon (Le Pelley 1968) and in 1903 from Zaire (Murphy and Moore 1990). The beetle is endemic to central Africa, but the exact origin of the pest is still not clear (Damon 2000).

### **2.5 Economically, ecologically important (e.g. keystone, endangered) species in North America (introduced and native) that are phylogenetically related or occur in the same habitat as the target pest**

We test the hypothesis that *P. coffea* is monophagous, with a physiological host range limited to its natal host, *H. hampei*. There are 11 species of adventive *Hypothenemus* (Tribe Cryphalini) recorded in Hawaii (Nishida 2002). There are no records of native Hawaiian *Hypothenemus* spp. except for a questionable old record (1913) of *H. ruficeps* (Swezey 1954), which has never been collected or reported since and is likely a synonym with *H. eruditus* or *H. crudiae* (C. Gillett, unpublished), which are adventive. There are also no known native species of the Tribe Cryphalini in Hawaii. There are, however, many native species in another scolytine genus, *Xyleborus* (Tribe Xyloborini) (Samuelson 1981; Gillett et al. 2019), which may potentially be impacted by release of an exotic parasitoid against a scolytine pest such as *H. hampei*. The Xyloborini are phylogenetically distant from the adventive Cryphalini species in Hawaii (Johnson et al. 2018). Our host range testing in quarantine included 6 of the 11 species of *Hypothenemus* (all adventive species) and 7 of the 28 species of native *Xyleborus* as well as an additional 10 exotic scolytine species in related genera (*Xyleborinus*, *Xylosandrus*, *Xyleborus*, *Euwallacea*, others).

## **2.6 Regulatory or pest status of the target pest in the state, provincial or federal law**

*Hypothenemus hampei* is established on all the Hawaiian Islands growing coffee and considered a significant pest that is actively being controlled.

## **2.7 Knowledge of status of other biological control agents (indigenous or introduced) that attack the pest**

No biocontrol agents were previously released in Hawaii against *H. hampei*. Two exotic predatory beetles, *Cathartus quadricollis* and *Leptophloeus* sp., are commonly found in overripe and dried coffee berries naturally preying on the immature stages of *H. hampei* in Hawaii (Follett et al. 2016; Brill et al. 2020). Our host testing in quarantine showed that *P. coffea* will not parasitize these beetles, and that the beetles did not predate on the parasitoids. Also, these predators attack eggs, larvae and pupae of *H. hampei* in overripe and dried berries (left after harvesting), whereas *P. coffea* attacks adult female *H. hampei* primarily in developing green berries at an earlier stage of crop maturity.

*Beauveria bassiana*, formulated as BotaniGard<sup>®</sup>, is sprayed frequently for *H. hampei* control. Repeated applications reduce coffee berry borer damage, but are costly, and efficacy varies depending on local conditions (Greco et al. 2018).

## **2.8 Life stage of the pest that is vulnerable to the biological control agent**

*Phymastichus coffea* is a primary, gregarious, idiobiont endoparasitoid of adult *H. hampei* females. The beetles are parasitized by *P. coffea* while actively boring into coffee fruits with the abdomen exposed, which can be a prolonged process depending on the ripeness of the fruits. This is unique behavior among Scolytinae, which typically bore into wood.

# **3. Biological Control Agent Information**

## **3.1 Taxonomy: scientific name (order, family, genus, species, scientific authority)**

*Phymastichus coffea* LaSalle (Hymenoptera: Eulophidae). It has no common name. *Phymastichus coffea* was collected in Togo in 1987 and described by LaSalle in 1990. The parasitoid wasp belongs to the family Eulophidae, one of the largest in the Hymenoptera, with nearly 4000 described species. The sub family Tetrastichinae to which the parasitoid belongs has 42 genera and is most widespread of all parasitic groups. Tetrastichinae has an extraordinarily wide host range attacking over 100 families of insects in 10 different orders, as well as mites, spider eggs, and even nematodes (LaSalle 1994). *Phymastichus* can be distinguished from other Tetrastichinae by the presence of distinctively swollen parastigma and lack the presence of a sensory plaque on the ventral edge of the male scape (LaSalle 1990). There are only two known species in this genus, (i) *Phymastichus coffea* and (ii) *P. xylebori*. Both species have potential value in biological control programs against scolytines. *Phymastichus coffea* attacks mainly adult *H. hampei* (CBB) whereas, *P. xylebori* attacks adults of the highly polyphagous island pinhole borer, *Xyleborus perforans* (Wollaston). A third species, *Phymastichus* sp. nova (D. Honsberger pers. comm.) is currently being described from Hawaii. The latter does not parasitize *H. hampei*.

### **3.2 Methods used to identify the biological control agent**

*Phymastichus coffea* was imported into Colombia at Cenicafé, where it has been mass reared in pure culture on CBB-infested coffee since its importation.

### **3.3 Location of reference specimens**

Voucher specimens are deposited at Cenicafé (Manizales, Colombia), at the USDA-ARS laboratory in Hilo, Hawaii, and at the University of Hawaii at Manoa.

### **3.4 Natural geographic range, other areas where introduced, and expected attainable range in Hawaii (also habitat preference and climactic requirements of the biological control agent)**

To date, *P. coffea* has been released in 12 countries as a classical biological control agent (Bustillo et al. 1998; Damon 2000; Jaramillo et al. 2005; Vega et al. 2015). *Phymastichus coffea* is native to Africa and present in most coffee producing countries on the continent. According to the CABI Invasive Species Compendium, *P. coffea* occurs in Kenya, Togo, and Mexico. Kenya and Togo are presumably within the native range, whereas it may have established in Mexico after release as a biological control agent against coffee berry borer. Hawaii is characteristically tropical but with moderate temperatures and humidity due to the influence of north and eastern trade winds. The climate at the elevations where coffee is grown should allow survival of *P. coffea* year-round.

### **3.5 Source of the biological control agent**

Centro Nacional de Investigaciones de Café - CENICAFÉ, Manizales, Colombia.

### **3.6 Host/biological control agent interactions**

*Phymastichus coffea* is an idiobiont, gregarious endoparasitoid of adult coffee berry borer, commonly laying two eggs (a male and a female) per host (Lopez-Vaamonde and Moore 1998). Both male and female develop in a single host, the female in the abdomen and the male in the prothorax (Espinoza et al., 2009), although a single female parasitoid is sometimes found living solitarily in the abdomen of the host. The parasitoid develops through four major life stages—egg, larva (three instars lasting ~21 days), pupa (~9 days) and adult. The complete development (egg to adult) occurs over 30-43 days depending on temperature and condition of the CBB host mummies. For example, at 23°C the life cycle of *P. coffea* is 43 days. The parasitoid emerges by cutting an opening in the host's integument (Feldhege, 1992).



Figure 5: Parasitized CBB with *Phymastichus pupa* in abdomen.

The average lifespan of the parasitoid is 1-2 days for males and 3-4 days for females (Espinoza et al., 2009). Longevity can be prolonged with 50% honey-water solution as food and if the temperature is decreased (F. Yousuf unpublished). On emergence, female parasitoids can have up to 10 eggs in the ovarioles, but more eggs are formed throughout her lifetime (synovigenic strategy) (Lopez-Vaamonde and Moore, 1998). There is no preoviposition period and the adult female parasitoids can parasitize the coffee berry borer adults immediately after emergence (Infante et al., 1994). It has been shown that *H. hampei* is attracted to semiochemicals released from coffee fruits (Mendesil et al. 2009); semiochemicals released during *H. hampei* feeding on fruits have been shown to attract *P. coffea* (Cruz-Lopez et al. 2016), and may play also a significant role in mediating the host specificity of their parasitoids under field conditions.

### **3.7 Biology and reproductive potential (including dispersal capability and damage inflicted on the target pest.)**

Gravid *P. coffea* females start to search for their hosts immediately after emerging from the adult female host and parasitism occurs within the first hours after emergence (Infante et al. 1994). *Phymastichus coffea* has an extremely short life span as an adult; the longevity of males ranges from 8-48 h and females from 16-72 h (Vergara et al. 2001; Portilla and Grodowitz 2018). *Phymastichus coffea* commonly lays two eggs (a male and a female) (López-Vaamonde and Moore 1998) in an *H. hampei* adult female at the time she is initiating fruit perforation, which causes paralysis and prevents further damage to the coffee berry. Both male and female develop in a single host, the female in the abdomen and the male in the prothorax (Espinoza et al. 2009). The parasitized *H. hampei* usually dies within 4-12 days after parasitism (Infante et al. 1994). The life cycle (egg to adult) of *P. coffea* varies from 30-47 days depending on the environmental conditions (temperature and humidity). Females are ~1 mm long, whereas males are half that size (LaSalle 1990). *P. coffea* can parasitize multiple hosts during its short lifespan. High levels of parasitism have been recorded in previous studies under cage and field conditions.

### 3.8 Known host range based on the scientific literature, host data from museum specimens, and unpublished records

The parasitoid has been described as a primary, gregarious, endoparasitoid of adult females of coffee berry borer (Feldhege 1992). To the best of our knowledge, no reports of parasitism by *P. coffea* on other hosts under field conditions exist. However, based on the results of no choice laboratory assays, two papers have reported *P. coffea* as oligophagous i.e. attacking other non-target scolytine hosts in addition to its primary host (Table 1) (López-Vaamonde and Moore 1998; Castillo et al. 2004).

**Table 1.** Previous reports of parasitism of Scolytinae species by *Phymastichus coffea* in no-choice laboratory assays.

Scolytinae species	Parasitism (%)	Parasitoid emergence (%)	Reference
<i>Hypothenemus hampei</i>	67.3, 64	48, 54	López-Vaamonde and Moore 1998, Castillo et al., 2004
<i>Hypothenemus obscurus</i>	83.3	15	López-Vaamonde and Moore 1998
<i>Hypothenemus seriatus</i>	76.6	12	López-Vaamonde and Moore 1998
<i>Hypothenemus eruditus</i>	6	4	Castillo et al., 2004
<i>Hypothenemus crudiae</i>	14	14	Castillo et al., 2004
<i>Hypothenemus plumeriae</i>	0	0	Castillo et al., 2004
<i>Araptus</i> sp.	70	18	López-Vaamonde and Moore 1998
<i>Araptus fossifrons</i>	0	0	Castillo et al., 2004
<i>Scolytodes borealis</i>	0	0	Castillo et al., 2004
<i>Tomicus piniperda</i>	0	0	Castillo et al., 2004
<i>Dendroctonus micans</i>	0	0	López-Vaamonde and Moore 1998

As shown in Table 1, although the parasitoid attacked other scolytines, it was restricted to species belonging to the same genus as its natural host, *Hypothenemus*, mostly. Two *Araptus* species were also tested by López-Vaamonde and Moore (1998), and Castillo et al. (2004) but only one showed positive parasitism. Castillo et al. (2004) report that *P. coffea* did not complete its life cycle in *Araptus*, despite relatively high numbers of parasitism attempts in laboratory exposures, while López-Vaamonde and Moore (1998) reported 70% parasitism, and 10-15% emergence of parasitoids, with high parasitoid mortality. No other records of the parasitoid attacking *Araptus* species are available in the literature.

### 3.9 History of past use of the biological control agent

The parasitoids, *Cephalonomia stephanoderis* Betrem, *C. hyalinipennis* Ashmead and *Prorops nasuta* Waterston (Hymenoptera: Bethyridae), *Heterospilus coffeicola* Schneideknecht (Hymenoptera: Braconidae) and *Phymastichus coffea* LaSalle (Hymenoptera: Eulophidae), all of African origin, have been introduced in many coffee producing countries, particularly in Central and South America (Klein-Koch et al. 1988; Barrera et al. 1990; Baker 1999; Jaramillo et al. 2005; Portilla and Grodowitz 2018), but none have been released in Hawaii. To date, *P. coffea* has been released in 12 countries as a classical biological control agent (Bustillo et al. 1998; Damon 2000; Jaramillo et al. 2005; Vega et al. 2015). Cenicafé (Colombia) recently released

~800,000 *P. coffea* (Feb-Jun 2021) in 40 ha of coffee to examine parasitism rates and the potential for inundative releases of mass reared parasitoids for *H. hampei* control (P. Benevides, pers. comm.).

### **3.10 Pathogens, parasites, parasitoids and hyperparasitoids (order, family, genus, species, scientific authority) of the agent and how they will be eliminated from the imported culture of the agent.**

Imported *P. coffea* will be reared for a generation in quarantine before release to inspect for hyperparasitoids or other insect contaminants. A sample of *P. coffea*-parasitized CBB will be tested for the presence of plant pathogens, e.g. coffee leaf rust, by USDA ARS scientists.

### **3.11 Procedures stating how the biological control agent will be handled in containment (e.g. scaling up for release)**

*Phymastichus coffea* will be obtained from an established stock maintained at the National Coffee Research Center-Cenicafé, Manizales (Caldas) Colombia, which was started from *P. coffea* collected in Kenya and shipped to Colombia in 1996 and has been maintained in colony in large numbers since that time (Orozco and Aristizábal 1996). *Phymastichus coffea* has been mass reared by Cenicafé on wild-caught CBB for field releases on multiple occasions and the colony receives frequent infusions of field collected material. For nontarget testing, *Phymastichus coffea* was shipped from Cenicafé in its larval stage in parasitized *H. hampei* hosts under USDA APHIS PPQ, permit no. P526P-18-00696 to a certified quarantine insect containment facility managed by the USDA Forest Service at Hawaii Volcanoes National Park, Volcano, Hawaii. Parasitized *H. hampei* were incubated in controlled climate chambers at  $25^{\circ} \pm 1^{\circ}\text{C}$ ,  $75 \pm 10\%$  relative humidity, and 8:16 h light:dark photoperiod at the quarantine containment facility. In the future, we hope that USDA APHIS and HDOA will allow the shipment of *P. coffea* from Cenicafé to Hawaii for release directly in the field without containment. Cenicafé is developing a new rearing system on diet rather than infested coffee beans to improve quality control and reduce the risk of contaminants.

### **3.12 Closely related genera, sibling species, cryptic species and ecologically similar species of the biological control agent in Hawaii, when they occur**

The eulophid genus *Phymastichus* contains two described species: *P. coffea* and *P. xylebori*. The candidate biological control agent *Phymastichus coffea* is not known to occur in Hawaii. *Phymastichus xylebori* is adventive in Hawaii and has been found on the Big Island parasitizing *Xyleborus perforans*; *P. xylebori* has not been found in coffee parasitizing *H. hampei* in Hawaii.

## **4. Host Specificity Testing**

### **4.1 Selection of nontarget test arthropods**

The selection of non-target hosts in Hawaii was based on phylogenetic relatedness to the target host (Johnson et al. 2018), sympatry of target- and non-target species, and size. Coleoptera species commonly occurring in the coffee landscape and species in culture at USDA-ARS in Hilo,

Hawaii were also tested; these were species not phylogenetically close to the target host but could provide insights into unexpected host use. There are 21 native and 38 non-native scolytine species in Hawaii (Samuelson 1981; Nishida 2002; Cognato and Rubinoff 2008). Because of the relatively large native scolytine fauna in Hawaii, and their remote or poorly studied habitats, only a subset of these species could be tested for their suitability as hosts to *P. coffea*. Exotic and native scolytine species were collected from coffee and macadamia farms and their surrounding habitats, and extensive searches from native forests from different Hawaiian Islands (Hawaii Island, Oahu, Maui, Molokai, Lanai and Kauai) (Gillett et al. 2020a). We investigated the host selection and parasitism response of *P. coffea* adult females to 43 different species of Coleoptera, including 23 Scolytinae (six *Hypothenemus* species and 17 others), and four additional Curculionidae (Yousuf et al. 2021). The list included Hawaiian endemic species (several Scolytinae in the genus *Xyleborus* and *Nesotocus giffardi*, a curculionid weevil), exotic pest species (e.g. the scolytines *Hypothenemus obscurus* [tropical nut borer] and *Xylosandrus compactus* [black twig borer], and the curculionids *Sitophilus oryzae* [rice weevil] and *Cylas formicarius* [sweetpotato weevil]), and beneficial species (e.g. a weed biocontrol agent *Uroplata girardi* from lantana, several coccinellids, and two flat bark beetle predators of *H. hampei*, *Cathartus quadricollis* and *Leptophloeus* sp.) (Tables 2-5) (Appendix A: Yousuf et al. 2021). All beetles used in host specificity tests were collected live and later preserved in 75% alcohol or pinned for identification by taxonomists with expertise in the respective taxa. The body size of the collected species ranged from 1-7 mm but the majority of species were similar in size to *H. hampei* which is 1.5-2.0 mm in length. Beetles were collected using Lindgren funnels or bucket or Broca traps baited with denatured ethanol only or ethanol + methanol + ethylene glycol lures, or collected directly from infested plant material (fruits, pods, stems, bark and seeds), or reared from infested wood in the laboratory (Gillett et al. 2020b). All non-target testing was conducted at the USDA Forest Service quarantine containment facility at Hawaii Volcanoes National Park, Volcano, Hawaii.

## 4.2 Laboratory tests

### No-choice tests

We used no-choice tests because these would reflect physiological host range and the most conservative assessment of potential for parasitism in the field, rather than choice tests (Van Driesche and Murray 2004). Choice tests that include the target host may mask the acceptability of lower ranked hosts, thereby producing false negative results (Withers and Mansfield 2005). Twenty individuals of each test species were placed in a sterilized glass Petri dish (80 mm in diameter) lined with filter paper and immediately afterwards four *P. coffea* females (<12h old) that had not been exposed to adult hosts prior to the experiments were introduced. Therefore, when ample hosts were available, each replicate consisted of 20 hosts and four parasitoids for a 5:1 host:parasitoid ratio. However, due to difficulties in finding certain species live in adequate numbers, e.g. native scolytine bark beetles, and difficulties synchronizing parasitoid emergence with field collection or emergence from wood of live beetles, the host:parasitoid ratio and numbers of replicates were adjusted as needed. For example, if only 10 non-target beetles were available for screening, then two replicates each with 5 beetles and 1 parasitoid (maintaining the 5:1 host:parasitoid ratio) were performed. In all non-target host screening tests, *H. hampei* was

included as a positive control to confirm parasitoid viability. The host:parasitoid ratio of the *H. hampei* controls was adjusted to match the nontarget species in the test, whether it was 5:1 or otherwise. The generalized behavioral response of the parasitoids towards target and non-target hosts was also determined for a subset of parasitoids by visual observation and video recording of parasitoid behavior, e.g. any contact with the host by landing on the host or antennation, and/or walking on the host. Host acceptance was noted when the parasitoid adopted a characteristic oviposition position on top the elytra of the host (Lopez-Vaamonde and Moore 1998).

After *P. coffea* exposure, *H. hampei* and all other non-target species were incubated at  $25 \pm 1^\circ\text{C}$ ,  $75 \pm 10\%$  RH and 24:0 (L:D) photoperiod for 72h. After 72h, parasitoids and filter paper linings were removed and the beetles were provided with a small cube (2 x 2 x 2 cm) of general beetle diet (F. Yousuf, unpublished). The beetles were again incubated at the same environmental conditions but now at 0:24 (L:D). After 10 days all the remaining diet and frass was removed (without disturbing the parasitized beetles) to avoid fungal contamination. Parasitized beetles typically became paralyzed and eventually died within 4-12 days after parasitoid oviposition. Beetles were held for a total of ~5-6 weeks for parasitoid emergence. Beginning after 25 days incubation, *H. hampei* mummies were inspected daily for adult wasp emergence. Parasitism was assessed based on observation of emergence of parasitoid progeny (F1 adult wasps) from the parasitized beetles, by inspection for exit holes on cadavers, or by dissection. Beetles with no exit holes were dissected (by separating the thorax from the abdomen) under a stereomicroscope using fine forceps and entomological pins at 20-100X magnification for evidence of parasitism, i.e., presence of *P. coffea* immature life stages (eggs, larvae or pupae), or unemerged adults. The number of unemerged life stages was recorded for each dissected beetle. After 5-6 weeks of incubation, dead beetle specimens sometimes became very dry and searching for the presence of eggs and early instar larvae was difficult. In such cases, beetles were dissected and examined under a compound microscope at 200X to seek unemerged *P. coffea*. The sex of emerged adult *P. coffea* offspring was determined by examination using a stereomicroscope. In most cases, two parasitoids (one male and one female) emerged per beetle host. To confirm this the sum of the emerged male and female parasitoids in each replicate was divided by two and compared to the number of parasitized hosts with exit holes. The sex of unemerged parasitoids was not determined. For data on parasitism, life stages, sex ratio, and development time, averages were calculated for each replicate (per Petri dish) for each species and used in statistical analysis. Grand means of all the replicates for each of the five *Hypothenemus* species are presented in figures and tables.

### Statistical analysis

Parasitism rate was calculated by dividing the number of parasitized hosts by the total number of hosts exposed to the parasitoids in each replicate. Parasitism included both emerged and unemerged wasps. Emergence rate was calculated by dividing the number of beetles with exit holes by the total number of parasitized hosts (emerged plus unemerged wasps). The sex ratio of the parasitoid progeny was calculated by dividing the number of emerged female parasitoids (F) by the total number of emerged male (M) and female (F) parasitoids  $[F / (F+M) \times 100]$ . The Shapiro–Wilk test (Shapiro and Wilk 1965; Razali and Wah 2011), numerical approaches (skewness and kurtosis indices), and the normal Q-Q plot-based graphical method were used to check the distribution of the data and showed that the data were not normally distributed. Generalized linear models (GLM) were therefore used to analyze the data, with



appropriate distribution function links. Parasitism and emergence rates of the parasitoids, and the percentage of different life stages (larvae, pupae and adults) in parasitized beetles with unemerged parasitoids were analyzed using GLM with a binary logistic function and sex ratio with a gamma log link function. Developmental time of the F1 offspring (egg to adult) was analyzed using GLM with a negative binomial log link function because data were overdispersed (i.e. variance > mean). Wald  $\chi^2$  approximations are reported. All analyses were performed using IBM SPSS statistics software.

## Results

Out of 43 total coleopteran species tested, including 23 scolytines, *P. coffea* oviposited and completed developed only in the target *Hypothenemus hampei* and four other species of *Hypothenemus*: *H. obscurus*, *H. seriatus*, *H. birmanus* and *H. crudiae* (Tables 2-5). Mean percentages of parasitism and emergence for the *Hypothenemus* spp. tested are shown in Figure 6. Parasitism ( $\chi^2 = 65.13$ ,  $df = 4$ ,  $p = 0.0001$ ) and emergence ( $\chi^2 = 23.20$ ,  $df = 4$ ,  $p = 0.0001$ ) were significantly higher in *H. hampei* than all other *Hypothenemus* species. *Hypothenemus hampei* had the highest percentage emergence of *P. coffea* at 70.4%, whereas *H. crudiae* had the lowest at 16.7% (Figure 6). In *H. crudiae*, out of five parasitized hosts only one had emergence. Although *P. coffea* only parasitized *Hypothenemus* spp., it did inspect three other non-target scolytine hosts, *Hypothenemus eruditus*, *Xyleborus kauaiensis* and *Xyleborus ferrugineus*, but left hosts without initiating oviposition (i.e. no parasitism found). The phylogenetic relationship of five *Hypothenemus* species included in our tests, extracted from Johnson et al. (2018), is also shown in Figure 6; *H. crudiae* is not included in the phylogeny because it was not included in Johnson et al (2018). Both parasitism and emergence in our tests decreased across *Hypothenemus* species with decreasing phylogenetic relatedness to *H. hampei*. *Hypothenemus eruditus*, the most distantly related species tested from *H. hampei* according to Johnson et al. (2018) was not parasitized (Figure 6).

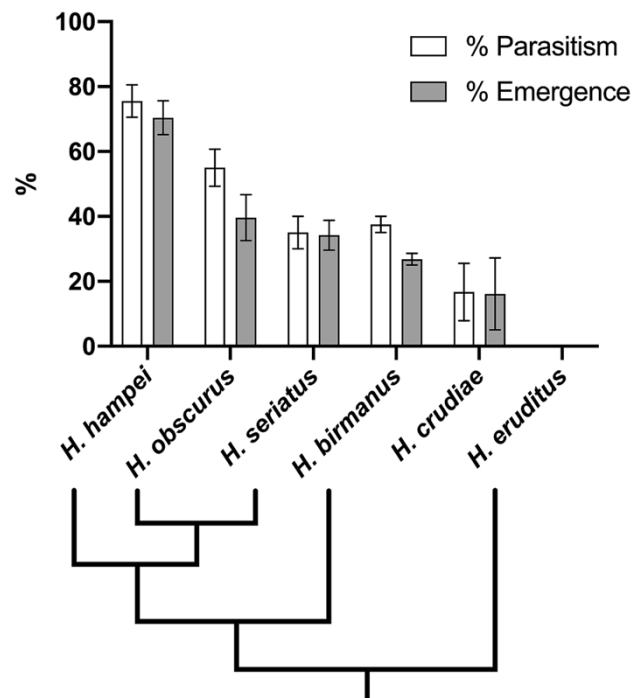


Figure 6: Percentage parasitism and emergence (mean  $\pm$  SE) of adult *Phymastichus coffea* parasitoids from *Hypothenemus* spp. Inferred from Johnson et al (2018).

Both parasitism and emergence in our tests decreased across *Hypothenemus* species with decreasing phylogenetic relatedness to *H. hampei*. *Hypothenemus eruditus*, the most distantly related species tested from *H. hampei* according to Johnson et al. (2018) was not parasitized (Figure 6).

Parasitoid development time among the three different *Hypothenemus* spp. did not differ significantly compared with *H. hampei* ( $\chi^2 = 0.17$ ,  $df = 4$ ,  $p = 0.997$ ), but did differ with *H. crudiae* (Table 2). The mean development time of *P. coffea* from oviposition to adult emergence was

shortest in *H. hampei* ( $32.2 \pm 0.5$  days, mean  $\pm$  SE), longest in *H. crudiae* ( $41.0 \pm 0.0$  days) and intermediate in the other three *Hypothenemus* spp. (Table 2), which generally agrees with the phylogenetic pattern observed for parasitism and emergence (Figure 1). The percentage of female versus male *P. coffea* emerging from parasitized *H. hampei* was  $50.8\% \pm 0.4$  (mean  $\pm$  SE), which was significantly different ( $\chi^2 = 27.3$ ,  $df = 4$ ,  $p = 0.0001$ ) from *H. seriatus* and *H. birmanus* (Table 2). *Hypothenemus eruditus* was not parasitized by *P. coffea* and hence was not included in any statistical analyses.

**Table 2.** Development time and sex ratio of *Phymastichus coffea* in no-choice in vitro non-target host selection screening of *Hypothenemus* species, including *H. hampei* as a control species.

Species	Insect status	Total beetles exposed	Development time (days $\pm$ SE)	Sex ratio (mean % females $\pm$ SE)
<i>Hypothenemus hampei</i> (control)	Exotic/Pest	170	$32.2 \pm 0.5$	$50.8 \pm 0.4$
<i>Hypothenemus obscurus</i>	Exotic/Pest	80	$35.0 \pm 0.9$	$54.8 \pm 1.6^*$
<i>Hypothenemus seriatus</i>	Exotic	60	$38.0 \pm 1.0$	$51.1 \pm 1.1$
<i>Hypothenemus birmanus</i>	Exotic	40	$37.0 \pm 1.0$	$57.7 \pm 3.8^*$
<i>Hypothenemus crudiae</i>	Exotic	30	$41.0 \pm 0.0^*$	50.0
<i>Hypothenemus eruditus</i>	Exotic	80	-	-

\* significantly different from *Hypothenemus hampei* (control),  $p < 0.05$ .

Parasitized *H. hampei* had the lowest percentage of unemerged parasitoids compared to the other four *Hypothenemus* species (Figure 7), indicating that *H. hampei* is a superior host for *P. coffea* development. For each parasitized host beetle with unemerged parasitoids, invariably two parasitoids were present, and the parasitoids were of the same life stage (larva, pupa, or adult). The frequency of the different life stages for parasitized hosts with unemerged parasitoids differed among *Hypothenemus* species (Figure 7). Parasitized *H. hampei* had a significantly lower percentage of larval ( $\chi^2 = 15.10$ ,  $df = 3$ ,  $p = 0.001$ ) and higher percentage of adult parasitoids that were unemerged ( $\chi^2 = 18.36$ ,  $df = 3$ ,  $p = 0.0001$ ) compared to the other *Hypothenemus* species. The higher percentage of unemerged parasitoids developing to the adult stage again indicates that *H. hampei* is a superior developmental host than the other *Hypothenemus* spp. The percentage of unemerged pupae found in parasitized *H. hampei* was not significantly different from *H. obscurus*, *H. seriatus* and *H. birmanus*, but *H. crudiae* had a significantly higher percentage of pupae than *H. hampei* ( $\chi^2 = 95.40$ ,  $df = 4$ ,  $p = 0.0001$ ) (Figure 7). No eggs were found in any of the parasitized *Hypothenemus* hosts.

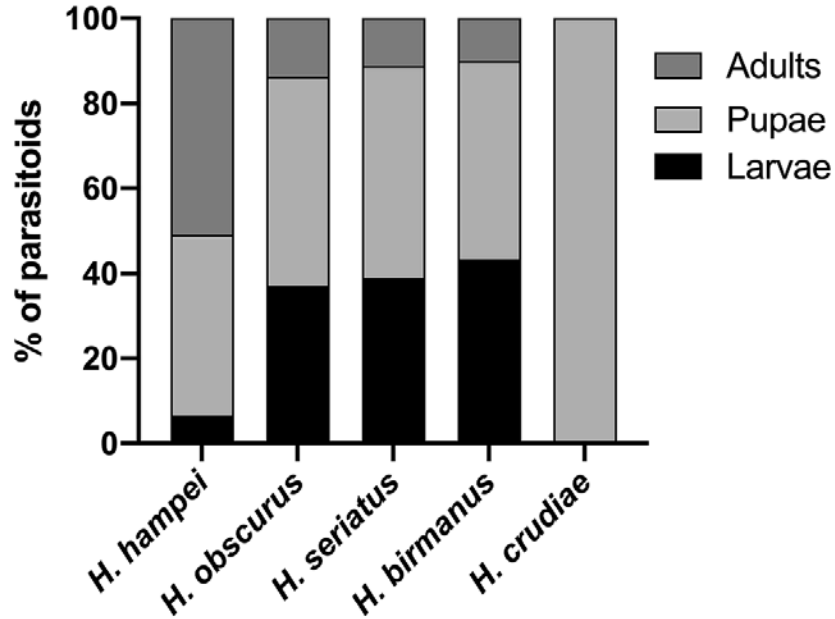


Figure 7: Fate of unemerged *Phymastichus coffea* parasitoids from parasitized *Hypothenemus* spp. in no-choice in vitro non-targeted selection screening. Parasitized *Hypothenemus* beetles with unemerged parasitoids were dissected to identify life stages (larva, pupa, adult)

#### Summary of laboratory tests in quarantine

The candidate biological control agent *Phymastichus coffea* was brought from Colombia into a Hawaii quarantine containment facility for host range testing to determine whether the parasitoid might attack non-target species in addition to the target host *H. hampei* and thereby pose a risk to Hawaiian endemic species. Using no-choice tests, 43 different species of Coleoptera were exposed to *P. coffea* in vitro, including 23 scolytines (six natives, 17 non-native species including *H. hampei* as seen in Table 3), six beneficial species (Table 4) and 12 other species including one native weevil (*N. giffardi*) (Table 5). Only five species from the genus *Hypothenemus* were parasitized by *P. coffea*, including the two pest species *H. hampei* (coffee berry borer) and *H. obscurus* (tropical nut borer, a macadamia nut pest), and three other exotic species *H. seriatus*, *H. birmanus*, and *H. crudiae* (Figure 6). Thus, *P. coffea* appears to be host specific at the genus level and should pose no harm to endemic species if released in Hawaii coffee for classical biological control of *H. hampei*. Nevertheless, no level of host specificity testing can ensure zero risk to non-target organisms when introducing a natural enemy in a new habitat (Louda et al. 2003).

**Table 3.** Parasitism and parasitoid emergence rates in no-choice in vitro non-target host acceptance screening of *Phymastichus coffea* exposed to various Scolytinae (Hawaii native and non-native) species.

Family	Species	Insect status	Total beetles exposed	Parasitism (%) (Mean ± SE)	Parasitoid emergence (%) (Mean ± SE)
Curculionidae:	<i>Xylosandrus compactus</i>	Exotic/Pest	80	0	0
Scolytinae	<i>Xylosandrus crassiusculus</i>	Exotic	80	0	0
	<i>Xyleborinus saxeseni</i>	Exotic	80	0	0
	<i>Xyleborinus andrewesi</i>	Exotic	60	0	0
	<i>Xyleborus ferrugineus</i>	Exotic	60	0	0
	<i>Euwallacea fornicatus</i>	Exotic	60	0	0
	<i>Euwallacea interjectus</i>	Exotic	60	0	0
	<i>Hypochryphalus sp.</i>	Exotic	60	0	0
	<i>Chryphalus sp.</i>	Exotic	80	0	0
	<i>Ptilopodius pacificus</i>	Exotic	80	0	0
	<i>Xyleborus molokaiensis</i>	Native	30	0	0
	<i>Xyleborus mauiensis</i>	Native	15	0	0
	<i>Xyleborus simillimus</i>	Native	18	0	0
	<i>Xyleborus hawaiiensis</i>	Native	9	0	0
	<i>Xyleborus lanaiensis</i>	Native	19	0	0
	<i>Xyleborus obliquus</i>	Native	3	0	0
<i>Xyleborus kauaiensis</i>	Native	35	0	0	

**Table 4.** Parasitism and parasitoid emergence rates in no-choice in vitro non-target host acceptance screening of *Phymastichus coffea* on beneficial Coleoptera species.

Family	Species	Insect status	Total beetles exposed	Parasitism (%)	Parasitoid emergence (%)
Chrysomelidae: Cassidinae	<i>Uroplata girardi</i>	Exotic	60	0	0
Coccinellidae	<i>Scymnodes lividigaster</i>	Exotic	40	0	0
Coccinellidae	<i>Rhyzobius forestieri</i>	Exotic	60	0	0
Coccinellidae	<i>Halmus chalybeus</i>	Exotic	40	0	0
Laemophloeidae	<i>Leptophloeus sp.</i>	Unknown	60	0	0
Silvanidae	<i>Cathartus quadricollis</i>	Exotic	80	0	0

**Table 5.** Parasitism and parasitoid emergence rates in no-choice in vitro non-target host acceptance screening of *Phymastichus coffea* on Hawaiian native and introduced coleopteran species from families and subfamilies other than Curculionidae:Scolytinae.

Family	Species	Insect status	Total beetles exposed	Parasitism (%)	Parasitoid emergence (%)
Anthribidae	<i>Araecerus simulatus</i> or <i>A. levipennis</i>	Unknown	6	0	0
Anthribidae	<i>Araecerus</i> sp. near <i>varians</i>	Unknown	15	0	0
Brentidae:Brentinae	<i>Cylas formicarius</i>	Exotic/Pest	80	0	0
Chrysomelidae:Bruchinae	<i>Acanthoscelides macrophthalmus</i>	Unknown	10	0	0
Curculionidae:Cossoninae	<i>Phloeophagosoma tenuis</i>	Unknown	8	0	0
Curculionidae:Cossoninae	<i>Nesotocus giffardi</i>	Native	12	0	0
Curculionidae:Curculioninae	<i>Sigastus</i> sp.	Exotic/Pest	6	0	0
Curculionidae:Platypodinae	<i>Crossotarsus externedentatus</i>	Exotic	60	0	0
Dryophthoridae:Dryophthorinae	<i>Sitophilus oryzae</i>	Exotic/Pest	60	0	0
Dryophthoridae:Dryophthorinae	<i>Sitophilus linearis</i>	Exotic	40	0	0
Nitidulidae:Carpophilinae	<i>Carpophilus dimidiatus</i>	Exotic	10	0	0
Nitidulidae:Carpophilinae	<i>Carpophilus zeaphilus</i>	Exotic	60	0	0
Tenebrionidae	<i>Tribolium castaneum</i>	Exotic/Pest	21	0	0
Tenebrionidae	<i>Hypophloeus maehleri</i>	Exotic	60	0	0

### 4.3. Information on the biological control agent from the area of origin based of field surveys or experimental field manipulation

In field cage studies in Mexico and Costa Rico, and also in Colombia (P. Benevides, pers. comm.), parasitism by introduced *P. coffea* was as high as 95% (Espinoza et al. 2009; Infante et al. 2013).

## 5. Environmental and Economic Impacts of the Proposed Release

### 5.1 Known impact of the biological control agent on humans and vertebrates

None.

### 5.2. Expected benefits of releasing this biological control agent

*Phymastichus coffea* is a potentially effective biological control agent for *H. hampei* and could be incorporated into existing IPM programs in Hawaii. To achieve maximum *P. coffea* parasitism in the field, releases should be made at times when *H. hampei* adults are active (e.g., when trap catches are high or female *H. hampei* are actively boring into fruits) and the coffee crop is at a susceptible stage. Studies suggest *P. coffea* may be susceptible to *B. bassiana*, however (Barrera 2005; Castillo et al. 2009; Ruiz et al. 2011), so releases should be timed to avoid *B. bassiana* applications or used in alternation with *B. bassiana* against *H. hampei*. If *P.*

*coffea* is highly effective, then dependence on *B. beauveria* applications could be reduced dramatically.

### **5.3 Direct impact of the biological control agent on target and non-target species.**

*Phymastichus coffea* is expected to help suppress *H. hampei* populations in coffee and may also provide a level of suppression of *H. obscurus* in macadamia nut farms which are relatively close to coffee growing areas or interspersed with coffee farms in some cases. Using a no-choice laboratory bioassay, we demonstrated that *P. coffea* was only able to parasitize the target host *H. hampei* and four other adventive species of *Hypothenemus*: *H. obscurus*, *H. seriatus*, *H. birmanus* and *H. crudiae* (Figure 6; Yousuf et al. 2021). *Hypothenemus hampei* had the highest parasitism rate and shortest parasitoid development time of the five parasitized *Hypothenemus* spp. Parasitism and parasitoid emergence decreased with decreasing phylogenetic relatedness of the *Hypothenemus* spp. to *H. hampei*, and the most distantly related species included in the trials, *H. eruditus*, was not parasitized. No species in any of the other genera tested were parasitized. These results suggest that the risk of harmful non-target impacts is minimal because there are no native species of *Hypothenemus* in Hawaii, and *P. coffea* could be safely introduced for classical biological control of *H. hampei* in Hawaii. Furthermore, as *P. coffea* is attracted to semiochemicals released from coffee fruit damaged by *H. hampei*, it is likely that under field conditions they will not be attracted to non-target species on different host plants lacking those cues.

### **5.4 Indirect impacts**

Potentially, *P. coffea* might interfere with two resident predators, *Cathartus quadricollis* and *Leptophloeus* sp., that naturally occur in coffee and attack CBB, or vice versa. However, these predators are mainly found in overripe and dried coffee berries naturally preying on the immature stages of *H. hampei* in Hawaii (Follett et al. 2016; Brill et al. 2020). Our host testing in quarantine showed that *P. coffea* will not parasitize these beetles, and that the beetles did not predate on the parasitoids. Also, these predators attack eggs, larvae and pupae of *H. hampei* in overripe and dried berries (left after harvesting), whereas *P. coffea* attacks adult female *H. hampei* primarily in developing green berries at an earlier stage of crop maturity. The biopesticide *Beauveria bassiana* also has the potential to interfere with *P. coffea* parasitism of CBB and survival. Indeed, studies suggest *P. coffea* may be susceptible to *B. bassiana* (Barrera 2005; Castillo et al. 2009; Ruiz et al. 2011). Therefore, releases of *P. coffea* should be timed to avoid *B. bassiana* applications or used in alternation with *B. bassiana* against *H. hampei*. If *P. coffea* is highly effective, then dependence on *B. bassiana* applications could be reduced dramatically.

### **5.5 Possible direct or indirect impact on threatened or endangered species in Hawaii**

Only five species from the genus *Hypothenemus* were parasitized by *P. coffea*, including the two pest species *H. hampei* (coffee berry borer) and *H. obscurus* (tropical nut borer, a macadamia nut pest), and three other exotic species *H. seriatus*, *H. birmanus*, and *H. crudiae* (Figure 1). Thus, *P. coffea* appears to be host specific at the genus level, on beetles relatively closely related to *H. hampei*, and, as there are no native Hawaiian species of *Hypothenemus*, should pose no harm to endemic species if released in Hawaii coffee for classical biological

control of *H. hampei*. However, no level of host specificity testing can ensure zero risk to non-target organisms when introducing a natural enemy in a new habitat (Louda et al. 2003).

## **5.6 Impact of biological control agent on physical environment**

None anticipated (see attached cultural impact assessment)

## **5.7 Proposed contingency plan to mitigate undesired environmental impacts**

Release of *P. coffea* will be a permanent, non-reversible action. *P. coffea* is not expected to attack any native Hawaiian species or disrupt native ecosystems given its high host specificity and short life span. Therefore, undesired environmental impacts are not anticipated.

# **6. Post-release Monitoring**

## **6.1 Biological control agent establishment and spread**

Once permits for release of *P. coffea* are obtained, field releases will begin on commercial coffee farms. In selected locations, data will be taken on establishment, dispersal from release points, parasitism rates, coffee berry infestation rates, and crop damage. Non-release sites will be used as controls initially to determine spread. Establishment is not certain and repeated releases may be required. *P. coffea* could not be found 8-12 months after release in Mexico and it also did not establish in coffee in Colombia after several years of mass releases. In Colombia and Mexico, coffee growers can effectively clean-pick their plantations. This may result in a dearth of hosts for the parasitoids, impacting their ability to establish. In Hawaii, there are widespread feral coffee stands, unmanaged coffee farms, and clear picking is seldom a viable option for various reasons. The year-round presence of hosts is expected to facilitate establishment of *P. coffea*. After release in Hawaii, regular surveys will be conducted to recover *P. coffea* in release areas. Adult *H. hampei* will be collected from fruit and returned to the laboratory for to determine whether they are parasitized. Diapause has not been investigated previously in *P. coffea* but it has been suggested that diapause may be the survival mechanism for the parasitoids between for the period when hosts are rare (McClay 1993). Overripe and drying coffee berries will be collected from release sites during the off-season and held to determine whether parasitoids emerge over time, possibly from a diapause state.

## **6.2 Biological control agent and target pest densities and distribution over time**

Coffee berry borer densities in Hawaii coffee are variable from year to year depending on climatic conditions and control measures (sanitation, *Beauveria bassiana* applications). *P. coffea* releases will be made on farms where USDA-ARS maintains CBB population monitoring and crop loss assessment activities as part of a long-term area-wide program. Data will be taken on percentage parasitism 1 week after *P. coffea* release and adult CBB will be held for parasitoid emergence. Coffee is a 7-month crop from the time of flowering to harvest. *P. coffea* releases will be made when trapping indicates peak flights of adult CBB and field sampling shows CBB adults boring into coffee berries, the time at which adult CBB are most susceptible to parasitism. Samples will be collected over a range of distances from release sites to assess dispersal of the parasitoids within and among coffee plantations over time. After harvest, samples will be collected

from residual fruits on coffee trees and from fallen fruits that lie beneath plants and sustain *H. hampei* reservoirs. The abundance of adult *H. hampei* available as hosts to *P. coffea* will decline during the months between harvest and the fruit set, a period of 4-5 months depending on location. We will investigate the potential for *P. coffea* to enter diapause during this period, allowing them to survive within *H. hampei* in desiccating fruit on trees or on the ground. Possible diapause will be detected by collecting desiccated fruits from the ground and overripe fruit remaining on trees, and holding them to determine if parasitoids emerge over a prolonged period. Laboratory trials will be conducted to assess whether diapause can be induced in *P. coffea* under controlled conditions.

The above studies will measure dispersal of *P. coffea*, as well as inter-seasonal survival of the wasps, thus whether wide-spread establishment occurred. We will simultaneously commence measuring the intergenerational impact of *P. coffea* on *H. hampei* populations. Cohorts of *H. hampei* will be monitored commencing when newly developed coffee fruit become susceptible in the field. Using life table analyses, the contribution of *P. coffea* to *H. hampei* generational mortality will be quantified and compared with other mortality factors that may be acting on the beetle population. These analyses will provide an accurate assessment of the impact of the biological control agent on the target pest densities over time since introduction of the natural enemy.

### **6.3 Impact on selected non-target species for which potential impacts are identified**

Preliminary data will be collected on semiochemical attraction of *Phymastichus coffea* to different *Hypothenemus* and other Scolytinae spp. *in vitro.*, to investigate the potential for developing methods to screen parasitoids for non-target effects based on responses to semiochemical diversity. We will compare *P. coffea* responses to chemical signals from Scolytinae species of varying host-specificity and compare this with two other *Phymastichus* species in Hawaii, *Phymastichus xylebori* LaSalle and *Phymastichus* sp. nova. *P. xylebori* parasitizes *Xyleborus perforans*, while *Phymastichus* sp. nova has been recorded from at least five host beetles (D. Honsberger pers. comm.). These comparisons will provide insights into the cues used by *Phymastichus* to locate hosts, and potentially the extent to which host specificity is mediated by parasitoid-host chemical interactions.

Various scolytines in the vicinity of release sites will be sampled periodically to determine whether any non-target parasitism occurs. While no non-target host use is predicted in Hawaii, this will serve as a test of the quarantine host-range testing predictions. This information will contribute to our overall understanding of and ability to predict zero impact on nontarget species.

## **7. Pre-release compliance**

### **7.1 Reference specimens**

*Phymastichus coffea* specimens in vials with alcohol have been deposited at multiple locations including Cenicafé, USDA ARS in Hilo, Hawaii, and the University of Hawaii at Manoa. Hundreds of specimens are available for DNA extraction. All specimens were reared at Cenicafé



in Colombia and shipped to Hawaii during host specificity testing in quarantine. A smaller number of pinned specimens is also available.

## **7.2 Planned location and timing of first release**

The planned site for the first release is Greenwell Farms (Kealahou, HI) in Kona, Big Island. The owner, Tom Greenwell, is a long-time cooperater with one of the largest coffee farms on the island. Interest is high across the coffee industry and among individual growers to have *P. coffea* releases. The number and timing of releases will be partly dictated by the number of *P. coffea* available. A letter confirming the release dates and locations will be submitted to USDA APHIS within 3 months after release.

## **8. List of Agencies and Persons Consulted**

Dr. Tracy Johnson, Research Entomologist, U.S. Forest Service, Institute of Pacific Islands Forestry, and director of the Hawaii Volcanoes National Park Quarantine Facility, Volcano, Hawaii.

Dr. Pablo Benavides Machado, Scientific Investigator III, Entomology, National Coffee Research Center-Cenicafé, Manizales (Caldas) Colombia. Provided *Phymastichus coffea* for testing.

Dr. Marisol Giraldo Jaramillo, Scientific Investigator I, Entomology, National Coffee Research Center-Cenicafé, Manizales (Caldas) Colombia. Provided *Phymastichus coffea* for testing.

Dr. Maribel Portilla, Research Entomologist, USDA-ARS Southern Insect Management Research Unit, Stoneville, Mississippi. Provided training on *Phymastichus coffea* rearing.

Dr. Conrad P.D.T. Gillett, Postdoctoral Research Fellow, University of Hawai'i Insect Museum Department of Plant and Environmental Protection Sciences, Entomology, College of Tropical Agriculture and Human Resources, University of Hawai'i at Mānoa, Honolulu, Hawaii. Confirmed identification of Scolytinae.

Dr. Lourdes Chamorro, Research Entomologist/Curator of Curculionoidea, Systematic Entomology Laboratory - ARS, USDA, c/o Smithsonian Institution - National Museum of Natural History. Provided identification of Curculionidae other than Scolytinae.

Tabetha Block, HETF Resource Associate, Forest Service Contractor, Institute of Pacific Islands Forestry, Hilo. HETF permit issuer.

Jay Hatayama, Forest Management Supervisor II, State of Hawaii, Division of Forestry and Wildlife, Hilo, Hawaii. DNLR permit issuer.

Other: Various coffee and macadamia growers on Hawaii Island.

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## Appendix A

Yousuf, F., Follett, P.A., Gillett, C.P.D.T., Honsberger, D., Chamorro, L., Johnson, T.M., Jaramillo, M.G., Machado, P.B. & Wright, M.G. 2021. Limited host range in the idiobiont parasitoid *Phymastichus coffea*, a prospective biological control agent of the coffee pest *Hypothenemus hampei* in Hawaii. *Journal of Pest Science*  
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# Limited host range in the idiobiont parasitoid *Phymastichus coffea*, a prospective biological control agent of the coffee pest *Hypothenemus hampei* in Hawaii

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## Abstract

*Phymastichus coffea* LaSalle (Hymenoptera:Eulophidae) is an adult endoparasitoid of the coffee berry borer, *Hypothenemus hampei* (Ferrari) (Coleoptera:Curculionidae:Scolytinae), which has been introduced in many coffee producing countries as a biological control agent. To determine the effectiveness of *P. coffea* against *H. hampei* and environmental safety for release in Hawaii, we investigated the host selection and parasitism response of adult females to 43 different species of Coleoptera, including 23 Scolytinae (six *Hypothenemus* species and 17 others), and four additional Curculionidae. Non-target testing included Hawaiian endemic, exotic and beneficial coleopteran species. Using a no-choice laboratory bioassay, we demonstrated that *P. coffea* was only able to parasitize the target host *H. hampei* and four other adventive species of *Hypothenemus*: *H. obscurus*, *H. seriatus*, *H. birmanus* and *H. crudiae*. *Hypothenemus hampei* had the highest parasitism rate and shortest parasitoid development time of the five parasitized *Hypothenemus* spp. Parasitism and parasitoid emergence decreased with decreasing phylogenetic relatedness of the *Hypothenemus* spp. to *H. hampei*, and the most distantly related species, *H. eruditus*, was not parasitized. These results suggest that the risk of harmful non-target impacts is low because there are no native species of *Hypothenemus* in Hawaii, and *P. coffea* could be safely introduced for classical biological control of *H. hampei* in Hawaii.

**Keywords** Coffee berry borer · Host specificity testing · Non-target · Biocontrol · Endoparasitoid · Scolytinae

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## Key message

- *Phymastichus coffea* is an idiobiont adult parasitoid of the coffee pest *Hypothenemus hampei*.
- In host range testing, *P. coffea* parasitized only five *Hypothenemus* spp.
- The parasitism rate was highest and parasitoid development time was shortest in *H. hampei*.
- No Hawaiian native species was parasitized by the parasitoid.
- *Phymastichus coffea* can be introduced safely for biocontrol of coffee berry borer in Hawaii.



## Introduction

The coffee berry borer, *Hypothenemus hampei* (Ferrari) (Coleoptera:Curculionidae:Scolytinae), native to Central Africa, is the most damaging insect pest of coffee worldwide, inflicting economical losses of over US \$500 million annually (Vega et al. 2015). In Hawaii, *H. hampei* was first recorded in Kona, Hawaii island, in 2010 (Burbano et al. 2011) and is now widespread throughout all the coffee-growing areas of Hawaii. Coffee is the third largest cash crop in the state of Hawaii, valued at more than \$43 million (USDA-NASS 2018). *Hypothenemus hampei* has had the effect of making coffee farming more intensive and less profitable, which is a major economic challenge to small-scale coffee production like that in Hawaii (Johnson et al. 2020). If left unmanaged, *H. hampei* can damage > 90% of the crop.

*Hypothenemus hampei* attacks coffee berries when the dry matter content of the endosperm, which increases with age, exceeds 20% (Jaramillo et al. 2005). After finding a suitable berry host, *H. hampei* bores into the coffee fruit through the central disk and excavates galleries where it lays eggs. The offspring develop inside the seeds and feed on the endosperm tissue of the berries (Damon 2000), reducing both coffee yield and quality. *Hypothenemus hampei* feeding damage can also cause premature fall of berries younger than 80 days (Decazy 1990). *Hypothenemus hampei* adults boring into the berry may remain in the 'A' position (Jaramillo et al. 2006) with the abdomen half exposed outside the berry potentially for weeks waiting for the dry matter content to reach 20% (Jaramillo et al. 2005).

Strategies to control *H. hampei* include mechanical, chemical and biological controls (Infante 2018). Sanitation and biological control (using parasitoids, predators and entomopathogenic microorganisms) are the most sustainable, environmentally friendly and widely used non-chemical control methods. The parasitoids, *Cephalonomia stephanoderis* Betrem, *C. hyalinipennis* Ashmead and *Prorops nasuta* Waterston (Hymenoptera:Bethylidae), *Heterospilus coffeicola* Schneideknecht (Hymenoptera: Braconidae) and *Phymastichus coffea* LaSalle (Hymenoptera: Eulophidae), all of African origin, have been introduced in many coffee producing countries, particularly in Central and South America (Klein-Koch et al. 1988; Barrera et al. 1990; Baker 1999; Jaramillo et al. 2005; Portilla and Grodowitz 2018), but none have been released in Hawaii. In Hawaii, the primary methods for controlling *H. hampei* are sanitation (frequent harvests and removal of all left over coffee berries after harvest) and applications of the biopesticide *Beauveria bassiana* (Ascomycota:Hypocreales), an entomopathogenic fungus (Aristizábal et al. 2016). Two generalist predators, *Leptophloeus* sp. and *Cathartus*

*quadricollis* (Coleoptera:Laemophloeidae and Silvanidae, respectively), occur naturally in Hawaii coffee and have been shown to feed on immature stages of *H. hampei* in overripe and dried berries (Follett et al. 2016; Brill et al. 2020), but are not very efficient in preventing damage in the first place.

Most of the studies on biological control of *H. hampei* have been conducted outside Hawaii, but in similar coffee production systems. In field-cage studies conducted in Mexico and Costa Rica, *P. coffea* proved to be the most promising biological control agent against *H. hampei* with parasitism rates as high as 95% (Espinoza et al. 2009; Infante et al. 2013). To date, *P. coffea* has been released in 12 countries as a classical biological control agent (Bustillo et al. 1998; Damon 2000; Jaramillo et al. 2005; Vega et al. 2015). *Phymastichus coffea* is native to Africa and present in most coffee producing countries on that continent. It is a primary, gregarious, idiobiont endoparasitoid of adult *H. hampei* females with a high capacity for host discrimination (Feldhege 1992; Infante et al. 1994; López-Vaamonde and Moore 1998; Castillo et al. 2004). Two laboratory studies reported that in addition to *H. hampei*, *P. coffea* parasitizes other *Hypothenemus* spp. such as *H. seriatus* and *H. obscurus* (López-Vaamonde and Moore 1998), and *H. eruditus* Westwood and *H. crudiae* (Panzer) (Castillo et al. 2004). However, parasitism of closely related species in the field has not been reported (Escobar-Ramírez et al. 2019). Gravid *P. coffea* females start to search for their hosts immediately after emerging from the adult female host and parasitism occurs within the first hours after emergence (Infante et al. 1994). *Phymastichus coffea* has an extremely short life span as an adult; the longevity of males ranges from 8 to 48 h and females from 16 to 72 h (Vergara et al. 2001; Portilla and Grodowitz 2018). *Phymastichus coffea* generally lays two eggs (into the abdomen, thorax, or between the thorax and abdomen) in an *H. hampei* adult female at the time she is initiating fruit perforation, which causes paralysis and prevents further damage to the coffee berry. The parasitized *H. hampei* usually dies within 4–12 days after parasitism (Infante et al. 1994). The life cycle (egg to adult) of *P. coffea* varies from 30 to 47 days depending on the environmental conditions (temperature and humidity). Females are ~ 1 mm long, whereas males are half that size (LaSalle 1990).

Earlier studies have shown the high host specificity of *P. coffea* and its ability to significantly reduce and regulate *H. hampei* populations (Gutierrez et al. 1998; López-Vaamonde and Moore 1998; Castillo et al. 2004; Rodríguez et al. 2017). Therefore, we decided to consider *P. coffea* as a biological control agent of *H. hampei* in Hawaii. A critical step was to determine its host specificity and assess possible risks to the Hawaii environment though impacts on endemic and other non-target species (Follett and Duan 1999; Messing and Wright 2006). Greatest non-target species impacts from

introduced biological control agents are likely to occur on species closely related to the target pest species (Van Drieseche and Murray 2004), but not always (Messing 2001), and thus, phylogenetically closely and distantly related species should be included in non-target screening efforts. This is an important element of biological control, particularly in Hawaii, where classical biological control may have had significant negative impacts on native species in the past (e.g., Howarth 1991; Henneman and Memmott 2001). While some studies have suggested that this is true (see references in Messing and Wright 2006), a number of carefully crafted field studies of population level impacts on non-target species have suggested that introduced parasitoids have had minimal, or sometimes moderate, impacts on endemic species (Johnson et al. 2005; Kaufman and Wright 2009). Where higher impacts have been detected, they are typically from accidentally introduced parasitoid species, and host insects in disturbed habitats are most susceptible to these impacts (Kaufman and Wright 2011). However, the potential for non-target impacts must be carefully considered, and outcomes of exposures of unintended hosts to prospective biological control agents can provide insights into host range patterns and determinants.

In this paper, we present new insights into the host specificity of *P. coffea*, a prospective biological control agent of *H. hampei* in Hawaii, by testing it against 43 different species of Coleoptera. Non-target testing included Hawaiian endemic, exotic and beneficial coleopteran species. There are currently no records of native Hawaiian *Hypothenemus* spp. except for an old record (1913) of *H. ruficeps* (Swezey 1954), which has never been collected or reported since and is possibly a synonym with the adventive species *H. eruditus* or *H. crudiae* (C. Gillett, unpublished). There are, however, many native species in another scolytine genus, *Xyleborus* (Samuelson 1981; Gillett et al. 2019), which may potentially be impacted by release of an exotic parasitoid against a scolytine pest such as *H. hampei*. We test the hypothesis that *P. coffea* is host specific and will not attack native Hawaiian Scolytinae species.

## Materials and methods

### Parasitoid, *Phymastichus coffea*

*Phymastichus coffea* used in this study were obtained from an established stock maintained at the National Coffee Research Center-Cenicafé, Manizales (Caldas) Colombia, which was started from *P. coffea* collected in Kenya and shipped to Colombia in 1996 and has been maintained in colony in large numbers since that time (Orozco-Hoyas and Aristizábal 1996). *Phymastichus coffea* has been mass reared by Cenicafé for field releases on multiple occasions

and the colony receives frequent infusions of field-collected material. *Phymastichus coffea* was shipped from Cenicafé in its larval stage in parasitized *H. hampei* hosts under USDA APHIS PPQ, permit no. P526P-18-00,696 to a certified quarantine insect containment facility managed by the USDA Forest Service at Hawaii Volcanoes National Park, Volcano, Hawaii. Parasitized *H. hampei* were incubated in controlled climate chambers at  $25^{\circ} \pm 1^{\circ} \text{C}$ ,  $75 \pm 10\%$  relative humidity and 8:16 h light:dark photoperiod at the quarantine containment facility.

Emerged male and female parasitoid adults were collected using a manual aspirator into a clean glass container. Parasitoids were held for mating and oocyte maturation and provided with 50% (w/v) honey (raw organic) solution for ~2 h before being used in the experiments (López-Vaamonde and Moore 1998). Infante et al. (1994) reported that *P. coffea* does not go through a preoviposition period and exhibits facultative arrhenotokous-type parthenogenesis, where the female parasitizes its host before or after copulation, producing haploid males (Portilla and Grodowitz 2018). Feldhege (1992) reported a preoviposition period of between 5 min and 4 h. The adult parasitoids are very short-lived: males (~8–48 h) and females (~16–72 h) (Vergara et al. 2001; Rojas et al. 2006; Espinoza et al. 2009; Portilla and Grodowitz 2018). The ability to parasitize hosts decreases with age, so it was important to use freshly emerged parasitoids (<12 h old) in all experiments.

### Coffee berry borer, *Hypothenemus hampei*

Field-collected *H. hampei* were used in all no-choice host specificity experiments. *Hypothenemus hampei*-infested coffee berries were collected from coffee trees (*Coffea arabica*) at OK Coffee Farm in Hilo, Hawaii (19.727583, -155.111186, elevation 156 m). These collections were transported in cold boxes to the USDA-ARS laboratory and placed in a custom-made extraction unit lined with tissue paper (Tech wipes 1709/7052, Horizon) to absorb condensation and prevent mold growth. Adult *H. hampei* were collected directly from the infested coffee berries by dissecting the berries or from the extraction unit using an aspirator. All the collected *H. hampei* were provided with artificial diet (modified from Brun et al. 1993) until use in the experiments.

### Collection of non-target coleopteran species

The selection of non-target hosts was based on phylogenetic relatedness to the target host, sympatry of target and non-target species, and size. Species commonly occurring in the coffee landscape and species in culture at USDA-ARS in Hilo, Hawaii, were also tested. There are 21 native and 38 non-native scolytine species in Hawaii (Samuelson 1981;

Nishida 2002; Cognato and Rubinoff 2008). Because of the relatively large native scolytine fauna in Hawaii, and their remote or poorly studied habitats, only a subset of these species could be tested for their suitability as hosts to *P. coffea*. Exotic and native scolytine species were collected from coffee and macadamia farms and their surrounding habitats, and from native forests from different islands (Hawaii Island, Oahu, Maui, Molokai and Kauai) in Hawaii (Gillett et al. 2020a). Host specificity tests were conducted with a total of 43 species from seven different coleopteran families including Hawaiian endemic species (several Scolytinae in the genus *Xyleborus* and *Nesotocus giffardi*, a curculionid weevil), exotic pest species (e.g., the scolytines *Hypothenemus obscurus* [tropical nut borer] and *Xylosandrus compactus* [black twig borer], and the curculionids *Sitophilus oryzae* [rice weevil] and *Cylas formicarius* [sweetpotato weevil]), and beneficial species (e.g., a weed biocontrol

agent *Uroplata girardi* from lantana, several coccinellids, and two flat bark beetle predators of *H. hampei*, *Catharus quadricollis* and *Leptophloeus* sp.) (Tables 1, 2, 3, 4). All beetles used in host specificity tests were collected live and later preserved in 75% alcohol or pinned for identification by taxonomists with expertise in the respective taxa. The body size of the collected species ranged from 1 to 7 mm, but the majority of species were similar in size to *H. hampei* which is 1.5–2.0 mm in length. Beetles were collected using Lindgren funnels or bucket or Broca traps baited with denatured ethanol only or ethanol + methanol + ethylene glycol lures or collected directly from infested plant material (fruits, pods, stems, bark and seeds) or reared from infested wood in the laboratory (Gillett et al. 2020b). All non-target testing was conducted at the USDA Forest Service quarantine containment facility at Hawaii Volcanoes National Park, Volcano, Hawaii.

**Table 1** Development time and sex ratio of *Phymastichus coffea* in no-choice in vitro non-target host selection screening of *Hypothenemus* species, including *H. hampei* as a control species

Species	Insect status	Total beetles exposed	Development time (days ± SE)	Sex ratio (mean % females ± SE)
<i>Hypothenemus hampei</i> (control)	Exotic/pest	170	32.2 ± 0.5	50.8 ± 0.4
<i>Hypothenemus obscurus</i>	Exotic/pest	80	35.0 ± 0.9	54.8 ± 1.6*
<i>Hypothenemus seriatus</i>	Exotic	60	38.0 ± 1.0	51.1 ± 1.1
<i>Hypothenemus birmanus</i>	Exotic	40	37.0 ± 1.0	57.7 ± 3.8*
<i>Hypothenemus crudiae</i>	Exotic	30	41.0 ± 0.0*	50.0
<i>Hypothenemus eruditus</i>	Exotic	80	–	–

\*significantly different from *Hypothenemus hampei* (control),  $p < 0.05$

**Table 2** Parasitism and parasitoid emergence rates in no-choice in vitro non-target host acceptance screening of *Phymastichus coffea* exposed to various Scolytinae (Hawaii native and non-native) species

Family	Species	Insect status	Total beetles exposed	Parasitism (%) (Mean ± SE)	Parasitoid emergence (%) (Mean ± SE)
Curculionidae:Scolytinae	<i>Xylosandrus compactus</i>	Exotic/pest	80	0	0
	<i>Xylosandrus crassiusculus</i>	Exotic	80	0	0
	<i>Xyleborinus saxeseni</i>	Exotic	80	0	0
	<i>Xyleborinus andrewesi</i>	Exotic	60	0	0
	<i>Xyleborus ferrugineus</i>	Exotic	60	0	0
	<i>Euwallacea fornicatus</i>	Exotic	60	0	0
	<i>Euwallacea interjectus</i>	Exotic	60	0	0
	<i>Hypochryphalus</i> sp.	Exotic	60	0	0
	<i>Chryphalus</i> sp.	Exotic	80	0	0
	<i>Ptilopodius pacificus</i>	Exotic	80	0	0
	<i>Xyleborus molokaiensis</i>	Native	30	0	0
	<i>Xyleborus mauianus</i>	Native	15	0	0
	<i>Xyleborus simillimus</i>	Native	18	0	0
	<i>Xyleborus hawaiiensis</i>	Native	9	0	0
	<i>Xyleborus lanaiensis</i>	Native	19	0	0
	<i>Xyleborus obliquus</i>	Native	3	0	0
	<i>Xyleborus kauaiensis</i>	Native	35	0	0

**Table 3** Parasitism and parasitoid emergence rates in no-choice in vitro non-target host acceptance screening of *Phymastichus coffea* on beneficial Coleoptera species

Family	Species	Insect status	Total beetles exposed	Parasitism (%)	Parasitoid emergence (%)
Chrysomelidae:Cassidinae	<i>Uroplata girardi</i>	Exotic	60	0	0
Coccinellidae	<i>Scymnodes lividigaster</i>	Exotic	40	0	0
Coccinellidae	<i>Rhyzobius forestieri</i>	Exotic	60	0	0
Coccinellidae	<i>Halmus chalybeus</i>	Exotic	40	0	0
Laemophloeidae	<i>Leptophloeus</i> sp.	Unknown	60	0	0
Silvanidae	<i>Cathartus quadricollis</i>	Exotic	80	0	0

**Table 4** Parasitism and parasitoid emergence rates in no-choice in vitro non-target host acceptance screening of *Phymastichus coffea* on Hawaiian native and introduced coleopteran species from families and subfamilies other than Curculionidae:Scolytinae

Family	Species	Insect status	Total beetles exposed	Parasitism (%)	Parasitoid emergence (%)
Anthribidae	<i>Araecerus simulatus</i> or <i>A. levipennis</i>	Unknown	6	0	0
Anthribidae	<i>Araecerus</i> sp. near <i>varians</i>	Unknown	15	0	0
Brentidae:Brentinae	<i>Cylas formicarius</i>	Exotic/Pest	80	0	0
Chrysomelidae:Bruchinae	<i>Acanthoscelides macropthalmus</i>	Unknown	10	0	0
Curculionidae:Cossoninae	<i>Phloeophagosoma tenuis</i>	Unknown	8	0	0
Curculionidae:Cossoninae	<i>Nesotocus giffardi</i>	Native	12	0	0
Curculionidae:Curculioninae	<i>Sigastus</i> sp.	Exotic/Pest	6	0	0
Curculionidae:Platypodinae	<i>Crossotarsus externedentatus</i>	Exotic	60	0	0
Dryophthoridae:Dryophthorinae	<i>Sitophilus oryzae</i>	Exotic/Pest	60	0	0
Dryophthoridae:Dryophthorinae	<i>Sitophilus linearis</i>	Exotic	40	0	0
Nitidulidae:Carpophilinae	<i>Carpophilus dimidiatus</i>	Exotic	10	0	0
Nitidulidae:Carpophilinae	<i>Carpophilus zeaphilus</i>	Exotic	60	0	0
Tenebrionidae	<i>Tribolium castaneum</i>	Exotic/Pest	21	0	0
Tenebrionidae	<i>Hypophloeus maehleri</i>	Exotic	60	0	0

## No-choice tests

In this study, we used no-choice tests because these would reflect physiological host range and the potential for parasitism in the field more accurately than choice tests (Van Driesche and Murray 2004). Choice tests that include the target host may mask the acceptability of lower ranked hosts, thereby producing false negative results (Withers and Mansfield 2005). Twenty individuals of each test species were placed in a sterilized glass Petri dish (80 mm in diameter) lined with filter paper and immediately afterward four *P. coffea* females (< 12 h old) that had not been exposed to adult hosts prior to the experiments were introduced. Therefore, when ample hosts were available, each replicate consisted of 20 hosts and four parasitoids for a 5:1 host–parasitoid ratio. However, due to difficulties in finding certain species live in adequate numbers, e.g., native scolytine bark beetles, and difficulties synchronizing parasitoid emergence with field collection or emergence from wood of live beetles, the host–parasitoid ratio and numbers of

replicates were adjusted as needed. For example, if only 10 non-target beetles were available for screening, then two replicates each with 5 beetles and 1 parasitoid (maintaining the 5:1 host–parasitoid ratio) were performed. In all non-target host screening tests, *H. hampei* was included as a positive control to confirm parasitoid viability. The host–parasitoid ratio of the *H. hampei* controls was adjusted to match the non-target species in the test, whether it was 5:1 or otherwise. The generalized response of the parasitoids toward target and non-target hosts was also determined for a subset of parasitoids by visual observation and video recording of parasitoid behavior, e.g., any contact with the host by landing on the host or antennation, and/or walking on the host. Host acceptance was noted when the parasitoid adopted a characteristic oviposition position on top the elytra of the host (Lopez-Vaamonde and Moore 1998).

After *P. coffea* exposure, *H. hampei* and all other non-target species were incubated at  $25 \pm 1$  °C,  $75 \pm 10\%$  RH and 24:0 (L–D) photoperiod for 72 h. After 72 h, parasitoids and filter paper linings were removed and the beetles were

provided with a small cube (2×2×2 cm) of general beetle diet (FY, unpublished). The beetles were again incubated at the same environmental conditions, but now at 0:24 (L–D). After 10 days, all the remaining diet and frass was removed (without disturbing the parasitized beetles) to avoid fungal contamination. Parasitized beetles typically become paralyzed and eventually die within 4–12 days after parasitoid oviposition. Beetles were held for a total of ~5–6 weeks for parasitoid emergence. Beginning after 25-day incubation, *H. hampei* mummies were inspected daily for adult wasp emergence. Parasitism was assessed based on observation of emergence of parasitoid progeny (F1 adult wasps) from the parasitized beetle, by inspection for exit holes on cadavers or by dissection. Beetles with no exit holes were dissected (by separating the thorax from the abdomen) under a stereomicroscope using fine forceps and entomological pins at 20–100X magnification for evidence of parasitism, i.e., presence of *P. coffea* immature life stages (eggs, larvae or pupae), or unemerged adults. The number of unemerged life stages was recorded for each dissected beetle. After 5–6 weeks of incubation, dead beetle specimens sometimes became very dry and searching for the presence of eggs and early instar larvae was difficult. In such cases, beetles were dissected and examined under a compound microscope at 200X to seek unemerged *P. coffea*. The sex of emerged adult *P. coffea* offspring was determined by examination using a stereomicroscope. In most cases, two parasitoids (one male and one female) emerged per beetle host. To confirm this, the sum of the emerged male and female parasitoids in each replicate was divided by two and compared to the number of parasitized hosts with exit holes. The sex of unemerged parasitoids was not determined. For data on parasitism, life stages, sex ratio and development time, averages were calculated for each replicate (per Petri dish) for each species and used in statistical analysis. Grand means of all the replicates for each of the five *Hypothenemus* species are presented in figures and tables.

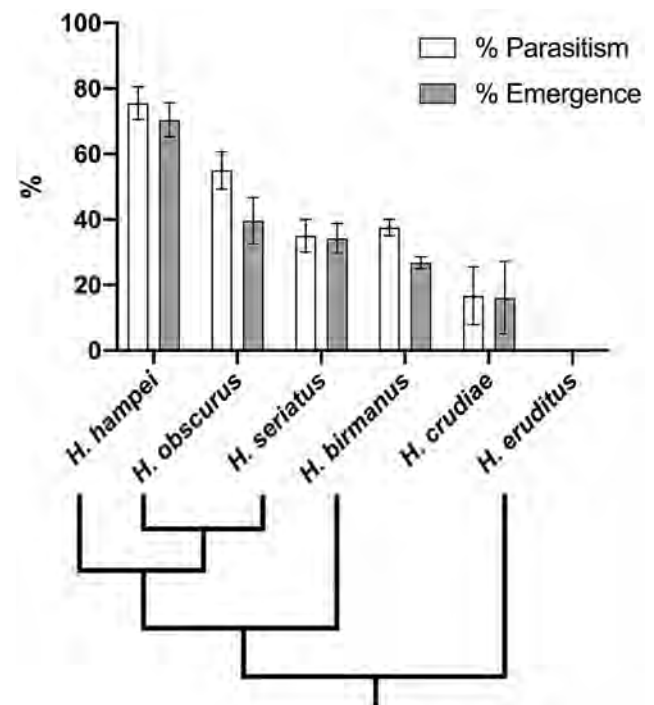
### Statistical analysis

Parasitism rate was calculated by dividing the number of parasitized hosts by the total number of hosts exposed to the parasitoids. Parasitism included both emerged and unemerged wasps. Emergence rate was calculated by dividing the number of beetles with exit holes by the total number of parasitized hosts (emerged plus unemerged wasps). The sex ratio of the parasitoid progeny was calculated by dividing the number of emerged female parasitoids (F) by the total number of emerged male (M) and female (F) parasitoids [ $F/(F+M) \times 100$ ]. The Shapiro–Wilk test (Shapiro and Wilk 1965; Razali and Wah 2011), numerical approaches (skewness and kurtosis indices) and the normal Q–Q plot-based graphical method were used to check the distribution of the

data and showed that the data were not normally distributed. Generalized linear models (GLM) were therefore used to analyze the data, with appropriate distribution function links. Parasitism and emergence rates of the parasitoids, and the percentage of different life stages (larvae, pupae and adults) in parasitized beetles with unemerged parasitoids were analyzed using GLM with a binary logistic function and sex ratio with a gamma log link function. Developmental time of the F1 offspring (egg to adult) was analyzed using GLM with a negative binomial log link function because data were overdispersed (i.e., variance > mean). Wald Chi-squared approximations are reported. All analyses were performed using IBM SPSS statistics software.

### Results

Out of 43 total coleopteran species tested, including 23 scolytines, *P. coffea* oviposited and completed developed only in the target *Hypothenemus hampei* and four other species of *Hypothenemus*: *H. obscurus*, *H. seriatus*, *H. birmanus* and *H. crudiae*. Mean percentages of parasitism and emergence for the *Hypothenemus* spp. tested are shown in Fig. 1. Parasitism ( $\chi^2 = 65.13$ ,  $df = 4$ ,  $p = 0.0001$ ) and emergence ( $\chi^2 = 23.20$ ,  $df = 4$ ,  $p = 0.0001$ ) were significantly higher in *H. hampei* than all other *Hypothenemus*

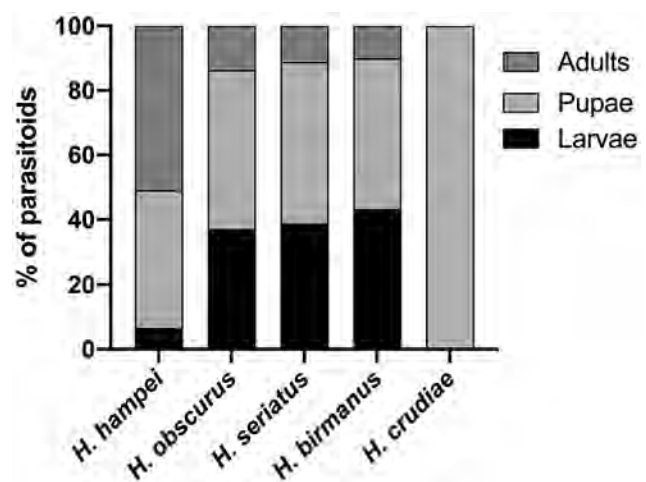


**Fig. 1** Percentage parasitism and emergence (mean ± SE) of adult *Phymastichus coffea* parasitoids from *Hypothenemus* spp. The phylogeny below the graph for the species included in the study (except *H. crudiae*) was inferred from Johnson et al. (2018)

species. *Hypothenemus hampei* had the highest percentage emergence of *P. coffea* at 70.4%, whereas *H. crudiae* had the lowest at 16.7% (Fig. 1). In *H. crudiae*, out of five parasitized hosts only one had emergence. Although *P. coffea* only parasitized *Hypothenemus* spp., it did inspect three other non-target scolytine hosts, *Hypothenemus eruditus*, *Xyleborus kauaiensis* and *Xyleborus ferrugineus*, but left hosts without initiating oviposition (i.e., no parasitism found). The phylogenetic relationship of five *Hypothenemus* species included in our tests, extracted from Johnson et al. (2018), is also shown in Fig. 1; *H. crudiae* is not included in the phylogeny because it was not included in Johnson et al. (2018). Both parasitism and emergence in our tests decreased across *Hypothenemus* species with decreasing phylogenetic relatedness to *H. hampei*. *Hypothenemus eruditus*, the most distantly related species from *H. hampei* according to Johnson et al. (2018), was not parasitized (Fig. 1).

Parasitoid development time among the three different *Hypothenemus* spp. did not differ significantly compared with *H. hampei* ( $\chi^2 = 0.17$ ,  $df = 4$ ,  $p = 0.997$ ), but did differ with *H. crudiae* (Table 1). The mean development time of *P. coffea* from oviposition to adult emergence was shortest in *H. hampei* ( $32.2 \pm 0.5$  days, mean  $\pm$  SE), longest in *H. crudiae* ( $41.0 \pm 0.0$  days) and intermediate in the other three *Hypothenemus* spp. (Table 1), which generally agrees with the phylogenetic pattern observed for parasitism and emergence (Fig. 1). The percentage of female versus male *P. coffea* emerging from parasitized *H. hampei* was  $50.8\% \pm 0.4$  (mean  $\pm$  SE), which was significantly different ( $\chi^2 = 27.3$ ,  $df = 4$ ,  $p = 0.0001$ ) from *H. seriatus* and *H. birmanus* (Table 1). *Hypothenemus eruditus* was not parasitized by *P. coffea* and hence was not included in any statistical analyses.

Parasitized *H. hampei* had the lowest percentage of unemerged parasitoids compared to the other four *Hypothenemus* species (Fig. 1), indicating that *H. hampei* is a superior host for *P. coffea* development. For each parasitized host beetle with unemerged parasitoids, invariably two parasitoids were present, and the parasitoids were of the same life stage (larva, pupa or adult). The frequency of the different life stages for parasitized hosts with unemerged parasitoids differed among *Hypothenemus* species (Fig. 2). Parasitized *H. hampei* had a significantly lower percentage of larval ( $\chi^2 = 15.10$ ,  $df = 3$ ,  $p = 0.001$ ), and higher percentage of adult parasitoids that were unemerged ( $\chi^2 = 18.36$ ,  $df = 3$ ,  $p = 0.0001$ ) compared to the other *Hypothenemus* species. The higher percentage of unemerged parasitoids developing to the adult stage again indicates that *H. hampei* is a superior developmental host than the other *Hypothenemus* spp. The percentage of unemerged pupae found in parasitized *H. hampei* was not significantly different from *H. obscurus*, *H. seriatus* and *H. birmanus*, but *H. crudiae* had a significantly higher percentage of pupae than *H. hampei*



**Fig. 2** Fate of unemerged *Phymastichus coffea* parasitoids from parasitized *Hypothenemus* spp. in no-choice in vitro non-target host selection screening. Parasitized *Hypothenemus* beetles with unemerged parasitoids were dissected to identify life stages (larva, pupa, adult)

( $\chi^2 = 95.40$ ,  $df = 4$ ,  $p = 0.0001$ ) (Fig. 2). No eggs were found in any of the parasitized *Hypothenemus* hosts.

## Discussion

*Phymastichus coffea* is a potential biological control agent of *H. hampei* and was brought from Columbia into a quarantine containment facility in Hawaii for host range testing to determine whether the parasitoid might attack non-target species and therefore pose a risk to Hawaiian endemic species. Using no-choice tests, 43 different species of Coleoptera were exposed to *P. coffea* in vitro, including 23 scolytines (six natives, 17 non-native species including *H. hampei*), six beneficial species and 12 other species including one native weevil (*N. giffardi*). Only five species from the genus *Hypothenemus* were parasitized by *P. coffea*, including the two pest species *H. hampei* (coffee berry borer) and *H. obscurus* (tropical nut borer, a macadamia nut pest), and three other exotic species *H. seriatus*, *H. birmanus* and *H. crudiae* (Fig. 1). Thus, *P. coffea* appears to be host specific at the genus level and should pose no harm to endemic species if released in Hawaii coffee for classical biological control of *H. hampei*. Nevertheless, no level of host specificity testing can ensure zero risk to non-target organisms when introducing a natural enemy in a new habitat (Louda et al. 2003).

We observed that once the host and parasitoids were exposed in the Petri dish arena that *P. coffea* inspected *H. hampei* and other *Hypothenemus* spp. hosts by antennation before proceeding to oviposition or rejection. *Phymastichus coffea* did not show any oviposition response to other non-target hosts. This could be dependent on several factors

because parasitoids may search and decide host suitability by using a broad spectrum of different stimuli such as plant–host complex volatiles, host feces volatiles, host sex pheromones, and tactile and visual cues (Chiu-Alvarado and Rojas 2008; Yang et al. 2008). Host habitat and host diet may influence the volatile composition emitted by the potential host insect, which can either deter or attract parasitoids from a distance. To minimize the effect of diet, we provided a general beetle diet to all the field-collected coleopteran hosts during the experiments. Parasitism of non-target hosts in the field may not be the same as our *in vitro* test results because of various factors related to the host's natural habitat. Most of the coleopteran species tested in our study are normally found tunneling in seeds, decomposing wood (under the bark and/or in sapwood) or decaying fruits. This cryptic behavior would likely provide protection from *P. coffea* which is accustomed to searching for *H. hampei* adult females, while they are exposed on the surface of coffee berries.

*Phymastichus coffea* was attracted to and parasitized only four species of *Hypothenemus* in addition to its target host *H. hampei*. This is consistent with studies reported by López-Vaamonde and Moore (1998), and Castillo et al. (2004). Combining information from our study and previous studies, seven species of beetles are now known to be able to serve as hosts in captive exposure studies for *P. coffea*: *H. hampei*, *H. obscurus*, *H. seriatus*, *Araptus* sp. (Lopez-Vaamonde and Moore 1998), *H. crudiae* and *H. eruditus* (Castillo et al. 2004), in addition to *H. birmanus* (this study). Parasitism of the scolytine *Araptus* sp. seems to be an outlier, but this genus does not occur in Hawaii. Aside from *Araptus*, *P. coffea* appears to be genus specific attacking closely related, but not all *Hypothenemus* species, given that species from closely related genera were not parasitized under no-choice test conditions. In our study, *P. coffea* did not attack *H. eruditus*. We believe that *H. eruditus* may not be a suitable host for the parasitoid because of its small size ( $\leq 1$  mm); *Phymastichus coffea* usually lays two eggs per host (1 male and 1 female), and in such a small host, successful development would be unlikely due to the limited availability of resources within the host. Host size is an important variable on which the survival and growth of parasitoid progeny depends. Females of most parasitoids preferentially lay eggs on larger hosts (Fox and Mousseau 1995). Also, *H. eruditus* is phylogenetically distant from *H. hampei* (Fig. 1) which is addressed below.

Our results also showed that *H. hampei* had the lowest numbers of unemerged parasitoids when compared with the other four *Hypothenemus* species (Fig. 2). The number of larvae and pupae were lower, and adults were higher in parasitized *H. hampei* with unemerged parasitoids. Similarly, in other three *Hypothenemus* spp. (*H. obscurus*, *H. seriatus* and *H. birmanus*) many unemerged parasitoids could not complete their development and died in their larval or pupal

stage with only a few reaching to the adult stage. In parasitized *H. crudiae* with unemerged parasitoids, most apparently could not reach the adult stage. Although the rate of completing the life cycle differed among *Hypothenemus* species, eggs did hatch in all parasitized species. Many factors can be responsible for suitability of the host for parasitoid development (Pennacchio and Strand 2006). Factors such as host physiology (e.g., presence of endosymbiotic bacteria), behavior (e.g., feeding habitat-sequestering secondary metabolites) and ecology (e.g., spatial/temporal overlap) may influence host acceptance by parasitoids and successful development (Desneux et al. 2009). All the non-target species used in the experiments were freshly collected from the field and may have carried toxins (accumulated from plant feeding) that may have interfered with the successful development of immature parasitoids within the hosts due to the ingestion of unsuitable food (e.g., see Desneux et al. 2009).

*Phymastichus coffea* also did not successfully parasitize any of the non-*Hypothenemus* species tested, including both native (*Xyleborus*) and exotic (*Xyleborinus*, *Xylosandrus*, *Xyloborus*, *Euwallacea*, others) Scolytinae, and other curculionid species from subfamilies other than Scolytinae, including the native weevil, *N. giffardi*. We did not find any *P. coffea* life stages (eggs, larvae, pupae, adults) after dissection in any of the non-*Hypothenemus* non-target species tested (Tables 2, 3, 4). Host specialization is relatively common in parasitic Hymenoptera and can be related to phylogeny, ecology and life histories (Price 1980; Stireman et al. 2006). It appears that at least host phylogeny was an important factor in host selection for *P. coffea* under our laboratory conditions.

Host range of idiobiont parasitoids is typically broader than koinobiont species (Askew and Shaw 1986; Hawkins et al. 1992), and it would hypothetically be reasonable to expect that *P. coffea* would follow this pattern. However, our results show that *P. coffea* was unable to successfully parasitize any species outside of the genus *Hypothenemus* and, even within the genus, was only moderately successful on species even closely related to *H. hampei*. While parasitism of *H. hampei* and subsequent parasitoid emergence was relatively high, both were significantly lower in *H. obscurus* and *H. seriatus*, sister species to *H. hampei*; *H. eruditus*, in a sister clade to the other species (Johnson et al. 2018), had zero parasitism. This demonstrates decreasing susceptibility to *P. coffea* with increasing phylogenetic distance among the *Hypothenemus* spp. exposed to the parasitoids in this study. Among the *Hypothenemus* spp. included in the phylogenetic reconstruction published by Johnson et al. (2018), *H. hampei* is the only species that has undergone a reversal in host range breadth, to become monophagous on coffee, while the other *Hypothenemus* spp. have retained a host generalist biology. *Hypothenemus hampei* has developed a unique association with *Pseudomonas* bacterial endosymbionts to

facilitate detoxification of caffeine, permitting it to exploit *Coffea arabica* seeds as their host (Ceja-Navarro et al. 2015), and potentially other physiological adaptations to its unique host, possibly providing adaptive challenges to parasitoids, and mediating host specificity of *P. coffea*. Messing (2001) questioned the practicality of applying centrifugal phylogeny approaches to selecting species to examine in non-target studies of potential biological control agents, particularly parasitoids. Our results support the predictions of the latter approach, with more distantly related *Hypothenemus* species less susceptible to *P. coffea* attack and more distantly related genera (e.g., *Xyleborus* spp.) not attacked at all. However, Messing (2001) emphasized the fact that interactions between the host insect and its host plant may override host phylogenetic patterns, by providing the stimuli for parasitoids to attack hosts, a consideration which may play a role in this study system. If this is the case, it is possible that *P. coffea* will produce even higher levels of parasitism than recorded in the artificial environment we used in our study, when attacking wild *H. hampei* boring into coffee fruits, producing the full range of cues stimulating parasitism, and lower field parasitism of the non-target *Hypothenemus* spp. included here.

Among all the parasitized *Hypothenemus* species, *H. hampei* had the highest rate of parasitoid emergence. The total developmental time (from egg to adult) of *P. coffea* was shortest in *H. hampei* (32 days); parasitism of *H. crudiae* resulted in the longest developmental time (41 days). Another study reported a similar development time of the *P. coffea* in *H. hampei*, 38–42 days at 23 °C and 66% RH (Rafael et al. 2000). Castillo et al. (2004) reported a *P. coffea* development time of 42.6 days for *H. hampei* and 40 days for *H. crudiae* at  $26 \pm 2$  °C and 70–80% RH. Total developmental time is directly related to the temperature. For example, the total development period of *Diglyphus isaea* (Hymenoptera:Eulophidae) decreased with increasing temperature between 15 and 35 °C and no development was found at 10 and 40 °C (Haghani et al. 2007). Temperature is a critical abiotic factor influencing the physiology and dynamics of insects. Therefore, in this study we selected a temperature for our no-choice assays which reflects the ambient field temperature the insects are expected to experience. In addition to temperature, age of the parasitoids and host play an important role in the subsequent development of parasitoid offspring (Pizzol et al. 2012). Hence, we used uniformly aged parasitoids and hosts throughout our experiments to minimize any impact on host parasitism and parasitoid development.

*Phymastichus coffea* commonly lays two eggs (a male and a female) per host (López-Vaamonde and Moore 1998). Both male and female develop in a single host, the female in the abdomen and the male in the prothorax (Espinoza et al. 2009). In this study, slightly fewer

male parasitoids emerged as compared to females from parasitized hosts. The proportion of females emerging from *H. hampei* was 50.8% which is consistent with the results obtained by López-Vaamonde and Moore (1998) and Rafael et al. (2000). Likewise, sex ratios of *P. coffea* emerging from *H. obscurus* 54.8%, *H. seriatus* 51.1% and *H. crudiae* 50.0% were consistent with the sex ratio results reported by (López-Vaamonde and Moore 1998; Castillo et al. 2004) of 1.25:1, 1:1 and 1:1 (female–male), respectively, for these species. In our study, the proportion of females emerging from parasitized *H. birmanus* 57.7%, was the highest among all other *Hypothenemus* species tested. The slightly fewer males produced per host in our study could be due to either to some parasitoid's preference to lay one egg per host (Feldhege 1992) or the lower survivorship of male eggs or larvae. Preference to lay female eggs over male can be dependent on several factors such as host quality, host age, immune response, genetic factors, photoperiod and relative humidity, host density or host-related volatile composition (King 1987).

All the above tests were conducted in a quarantine laboratory with no field studies. We conducted no-choice tests because they may provide more accurate and conservative information on host preferences and physiological host range than choice tests because of lower levels of interference due to unexpected responses to multiple host cues (Van Driesche and Murray 2004). Sands (1997) showed that laboratory studies often overestimate the host range of the parasitoid and realized ranges under field conditions may be substantially less than predicted from no-choice tests, but they are necessary to give a worst-case prediction of the number of hosts at risk of being attacked in the field (Avilla et al. 2016). *Phymastichus coffea* attacked other non-target *Hypothenemus* species in our no-choice trials, but this does not necessarily mean that those species will be attacked in the field. For example, an idiobiont braconid wasp, *Bracon hebetor* is reported to parasitize a wide variety of moths within and outside in Phycitinae (Lepidoptera:Noctuidae) in the laboratory, but in the field it is restricted to only larvae of *Plodia interpunctella* (Lepidoptera:Noctuidae) (Antolin et al. 1995). This is because in the field, parasitoids use a spectrum of long- and short-range cues (chemical, visual, vibrational and tactile signals) to locate hosts (Strand and Pech 1995). Chemical cues (infochemicals) can play an important role in host location. A study conducted by Rojas et al. (2006) showed that *P. coffea* can distinguish between *H. hampei*-infested and uninfested coffee berries, and were highly attracted to the dust/frass originating from *H. hampei* infested berries, but showed no response to the dust/frass originated from the closely related non-target host, *H. crudiae*. This behavior depending on plant and host cues suggests that it is very unlikely that *P. coffea* will have any



negative effects on non-target scolytids, or any other beetles, under field conditions.

No biocontrol agents were previously released in Hawaii against *H. hampei*. Two exotic predatory beetles, *Cathartus quadricollis* and *Leptophloeus* sp., are commonly found in overripe and dried coffee berries predated on the immature stages of *H. hampei* (Follett et al. 2016; Brill et al. 2020). Our host testing in quarantine showed that *P. coffea* will not parasitize these beetles and that the beetles did not predate on the parasitoids. Also, these predators attack eggs, larvae and pupae of *H. hampei* in overripe and dried berries (left after harvesting), whereas *P. coffea* only attacks adult female *H. hampei* at an earlier stage of crop maturity. The other four *Hypothenemus* species that were attacked by *P. coffea* have very different field habitats, but might serve as useful transitory hosts for *P. coffea* at times when, or in areas where, *H. hampei* populations are at low densities, such as between coffee seasons. For example, macadamia nut farms are often located close to coffee farms in Hawaii and may provide a year-round source of *H. obscurus*, a pest of macadamia nut. Feral coffee in Hawaii could also serve as a continuous source of *H. hampei* throughout the year.

*Phymastichus coffea* is a potentially effective biological control agent for *H. hampei* and could be incorporated into existing IPM programs in Hawaii. *Phymastichus coffea* may be simply released and monitored for establishment in a classical biological control program, or it may be mass reared for inundative releases. Currently, trapping and sampling of infested coffee fruits is conducted to monitor *H. hampei* flights and optimize timing of *Beauveria bassiana* applications for control (Aristizabal et al. 2016). After *H. hampei* bores into the coffee berries, it is protected and difficult to control with biopesticides or conventional insecticides. To achieve maximum *P. coffea* parasitism in the field, inundative releases should be made at times when *H. hampei* adults are active (e.g., when trap catches are high or female *H. hampei* are actively boring into fruits) and the coffee crop is at a susceptible stage. Optimal timing of inundative releases may differ for different elevations due to *H. hampei* population dynamics (Hamilton et al. 2019). Studies suggest *P. coffea* may be susceptible to *B. bassiana*, however (Barrera 2005; Castillo et al. 2009; Ruiz et al. 2011), so inundative releases should be timed to avoid *B. bassiana* applications or used in alternation with *B. bassiana* against *H. hampei*. If *P. coffea* is highly effective, then dependence on *B. beauveria* applications could be reduced dramatically.

## Author contributions

FY designed methodology, conducted the experiments and wrote the manuscript; PF designed the experiments; PF and MW provided overall project management and manuscript

editing; FY, CG and DH conducted field surveys and collected live beetles for testing. FY, CG and LC identified beetle species; MGJ and PBM reared and supplied *Phymastichus coffea*. All authors read and gave final approval for publication.

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## Declarations

**Conflict of interest** The authors have declared that no conflict of interest exists.

**Informed consent** Informed consent was obtained from all individual participants included in the study.

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## **Appendix B**

Cultural Impact Assessment for Proposed Statewide Release of *Phymastichus Coffea* to Control Coffee Berry Borer

# Cultural Impact Assessment for Proposed Statewide Release of *Phymastichus Coffea* to Control Coffee Berry Borer



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## Introduction

At the request of the University of Hawai'i and the United States Department of Agriculture Agricultural Research Service, the Synergistic Hawai'i Agriculture Council (SHAC) conducted a Cultural Impact Assessment (CIA) for the proposed statewide release of *Phymastichus coffea*. Used as a biocontrol in coffee, *P. coffea* is a tiny wasp that targets and parasitizes the coffee berry borer beetle (CBB) *Hypothenemus hampei* (Coleoptera: Curculionidae).

This CIA and its interviews were designed to identify any utilization of coffee for cultural practices or community concerns about environmental impacts from the release of *P. coffea*. It is a companion document to an Environmental Assessment drafted by USDA and was prepared in adherence with the Office of Environmental Quality Control (OEQC) *Guidelines for Assessing Cultural Impact*, adopted by the Environmental Council, State of Hawai'i, on November 19, 1997 and pursuant to Chapter 343 of the Hawaii Revised Statutes as well as the 2019 revisions to HAR Chapter 11-200.1.

## Proposed Action

Biological control (biocontrol) is a component of an integrated pest management strategy. It is defined as the reduction of pest populations by natural enemies and typically involves an active human role (Flint, 1998). Classical biocontrol is the selection and introduction of a natural enemy of an invasive plant or insect pest, and then “reuniting” of this natural enemy with the invasive pest to provide long-term, cost-effective, and sustainable pest management. Both State and Federal agencies have been cooperating on biocontrol activities to minimize the threat of invasive pests in Hawai'i's natural environment. Selection of a biocontrol for potential release undergoes a multi-step regulatory process to ensure native plants, insects, or traditional and customary practices are not impacted by the introduction.

## *Coffea arabica* and *Phymastichus coffea*

Coffee (*Coffea arabica*) is an introduced plant to Hawai'i, and familiar to most people. Thought to be native to Ethiopia, the intensive cultivation of coffee in Northern Africa (and beyond) began as early as the 16<sup>th</sup> century. Thriving in subtropical climate zones, there is now a “coffee belt” between the Tropics of Cancer and Capricorn, where some 70 countries grow and export the bean. Early traders noticed Hawai'i's place on the belt, and began to import seeds in the 19<sup>th</sup> century.

*Coffea* favors a tropical climate with a distinct wet and dry season. Despite this preference, the well-draining cinder soils of Hawai'i can support the plant even in extremely wet locations.



Feral coffee is readily found in Maui's 'Iao Valley and in the gulches of Hilo. The plant does not tolerate extreme heat, nor frost, and is typically grown commercially between 400 ft (122 m) and 2,400 ft (732) in elevation (Bittenbender, 1999). At the time of its arrival in Hawai'i, the *Coffea* plant had been hybridized by commercial breeders. The first varietal to take root commercially, and still grown extensively in Kona, was Typica. Subsequent introductions included the hybrid Caturra, Catuai and Mokka varietals, each suited to a slightly different climate.



*Figure 1: Typica tree in flower, Haiku, Maui*

In 1842, the Kingdom of Hawai'i recognized the potential importance of Hawaiian coffee and taxed any foreign coffees brought into the islands. Coffee continues to be an economically important crop, with a farmgate green bean value of \$102.9 million, likely top in the state when considering roasted valuations (NASS, 2021). It is also a familiar plant found growing wild in every county statewide. While it is not a traditional Hawaiian crop, coffee has contributed

greatly to the post-contact agricultural history of the State. There are almost 1,500 coffee farmers in the state. The majority of commercial growers are smallhold, operating less than five acres of land, and are considered socially-disadvantaged by the USDA (NASS, 2017).



Figure 2: Commercial coffee field in Pahala, Ka'u District, Island of Hawai'i

CBB is the most devastating invasive insect pest in coffee plantations and is estimated to cause more than \$500 million in damage around the world (Vega 2020). The *Hypothenemus* is a genus of over 200 described oriental bark beetles within the Curculionidea family (Johnson et al., 2020). CBB was first reported in coffee plantations during an 1897 survey of the West African nation of Liberia (Hopkins, 1915). The pest is notably distinguished from all 850 other insect species that can feed on parts of the coffee plant in that it is the only one able to feed and complete its life cycle in the coffee bean itself. The female beetle bores a small hole into the developing fruit and lays up to 100 eggs in the bean (Jaramillo, 1997). Larvae subsequently feed on the bean, and create cavities, greatly reducing quality and impacting market value. Because the lifecycle occurs largely within the protection of the bean, once the insect penetrates the bean, she and her progeny are relatively protected from insecticides or other conventional control measures. The insect rapidly propagates in Hawai'i, with a mean life cycle of approximately 51 days, totaling more than 7 generations per year (Hamilton 1999).



*Figure 3: Bore holes indicate CBB activity in ripening coffee fruit*

The arrival of the invasive CBB pest created a significant challenge for growers. First reported in the South Kona region of the Big Island in 2010, the beetle quickly spread to Ka'u and on to the neighbor islands of O'ahu (2014), Maui (2016), Kua'i (2020), and Lanai (2020), causing widespread damage and economic loss. The negative consequences of this invasion continue to be felt by growers, processors, buyers, and consumers. The estimated economy-wide impact of CBB for the crop years 2011/12 and 2012/13 was a \$12.7M loss in crop value, a \$25.7M loss in sales, a \$7.6M loss in household earnings, and a loss of more than 380 jobs (Lueng, 2013). The added production costs of CBB are significant, and have the potential to drive small farms out of business (Woodill, 2014). CBB found in unmanaged and feral coffee trees can be a source of ongoing infestations for neighbor farms (Johnson et al., 2020).

Mitigation and containment are possible, whereas eradication has not proven feasible. Current control methods are laborious and costly, involving hand removal of beans as well as repeated

applications of chemical insecticides. In the years following, detection, multiple programs, and resources were directed at the problem of CBB in Hawai'i, including pest subsidies, grower education programs and a relaxation of the Hawai'i Department of Agriculture quality standards (Johnson et al., 2020). Despite these efforts, CBB remains an intractable issue for growers due to high labor costs and the unsuitability of control through chemical pesticides. Thus, management strategies that limit human labor, such as biocontrol, are identified by farmers as a major need.



Figure 4: Live beetle and damage inside the bean. Adult CBB is approximately 1.8 mm long

There are three natural enemies to CBB that are indigenous to Africa. One of these, *Phymastichus coffea* is an endoparasitoid that attacks CBB adults and is found widespread in African coffee regions. Females, under 1mm in length, oviposit in the abdomen of the CBB adults, laying a single male and a single female egg, which hatch and feed on the internal tissues of the host. Host CBB that are parasitized by *P. coffea* die within 15 days (Espinoza, 2009). This species of parasitoid is considered ideal for use as a CBB biocontrol agent because

of its highly discriminatory nature and its ability to enter the bean itself. It has been released in at least 12 countries to date. Although it is short-lived (2–3 days), it can be released any time after fruit colonization, with studies showing successful parasitization up to seven days after CBB have initiated berry entry.

The Espinoza (2009) study results demonstrate that using *P. coffea* at a density of 1 parasitoid per 10 hosts results in a 3- to 5.6-fold decrease in CBB damage to the coffee beans when compared to the control. This is due to the fact that individuals parasitized by *P. coffea* drastically changed their behavior, stopped reproducing and died before they damaged the coffee bean.

Unlike other CBB-management methods, release of *P. coffea* does not require trespassing on private land to treat unmanaged coffee trees or application of chemicals to feral coffee in public areas as the wasp can fly up to 70 meters from its release site. Using *P. coffea* to control CBB is an opportunity to reduce the collateral impacts of areawide control activities.

## Hawai'i – Historical and Cultural Background

### PRE-SETTLEMENT

#### GEOGRAPHICAL

The Hawaiian Islands lie in the middle of the vast Pacific Ocean located approximately 2,500 miles from the nearest continent on the Earth. Islands rose individually to the surface as the Pacific Plate drifted north-northwest over a lava hot spot creating these new land masses. The youngest and most southern island in the chain, Hawai'i, is thought to be about 400,000 years old. Ni'ihau and Kauai to the northwest end of the main Hawaiian islands are approximated to be about 3-5 million years old. The newest formation south of Hawai'i island, Loihi, will most likely reach the surface in 50,000 years. Kure Atoll to the far northwest of the archipelago is one of the atolls still above water, close to 30 million years old (Olson, 2004). The islands are host to many diverse climate zones and the largest mountain on the planet, Mauna Kea on Hawai'i island, standing at 39,000 feet (14,000 metres) if taken from measurement at its sub-surface base to its summit (Wylie, 2015).

The main and most populated islands in the Hawaiian-Emperor Chain are Hawai'i, Maui, Kaho'olawe, Lāna'i, Moloka'i, O'ahu, Kauai and Ni'ihau. The Papahānaumokuākea Marine National Monument, established in 2006, extends from Nihoa northwest to Kure Atoll. Stretching over 1,350 miles and covering 582,578 square miles, it is one of the largest marine conservation areas on Earth, offering both environmental and Native Hawaiian cultural protections (Papahānaumokuākea Marine National Monument).

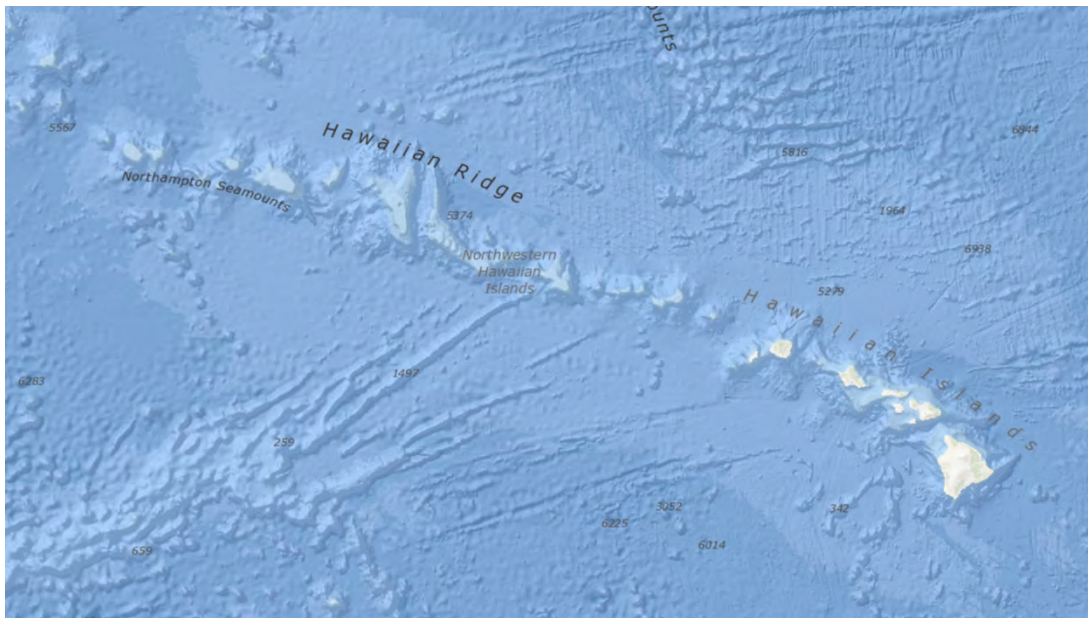


Figure 5: Map of the Hawaiian Archipelago  
NOAA

## PLANTS AND ANIMALS

The position of these islands on the planet created space in which flora and fauna developed unimpeded and unchallenged. Various birds, trees, plants, and creatures of the sea and land made their way by air or water here to thrive on the shores and slopes of this volcanic chain, creating an abundance of life (Olson, 2004). This life would eventually come to support the Polynesians who made their way across the Pacific to the many island groupings in one of the most rapid settlement excursions known to humans.

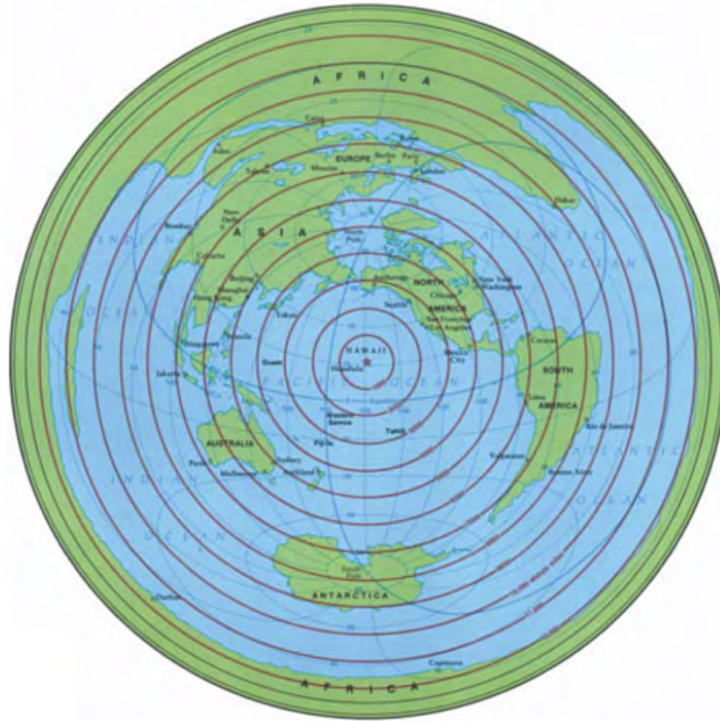


Figure 6: Azimuthal equidistant projection map Hawaii (Armstrong, 1983)

Prior to the arrival of the Polynesians, Hawaii lay untouched except for the natural forces of tsunami, earthquakes, hurricanes, drought and even blizzards atop the peaks of its highest mountains. The plants, animals and insects that made their way here established themselves and became some of the most unique species on the planet. Although similarities can be seen with their counterparts on the continents, many developed interesting new characteristics. Typical protective defense systems in place in these organisms on the continents were lost over time as there were no predators nor competitors to challenge them (Olson, 2004). Stinging nettles on the mainland of North America, for example, has a relative here in Hawai'i known as māmaki (*Pipturus albidus*). Māmaki has lost the stinging leaf its mainland relative is known for, however still carries the same usages in medicinal remedies (Bishop Museum, 2021).

Early examples of these pre-settlement species include fern spores, koa, pōhuehue (beach morning glory), snails, and insects most likely from North America. Tradewinds that prevail from the Northeast and storms from the South most likely helped propel them to the Hawaiian islands (Dunford et. al, 2013). Once here, as mentioned above, many lost their natural defenses due to lack of predation and continually diversified, adapting to the wetlands and drylands of the islands.

Plants existing pre-settlement:

Koa, pūkiawe, māmaki, 'a'ali'i, olonā, 'uki'uki, kauila, 'ōlapa, 'ākala, maile, māmane, 'ōhelo, 'ūlei, hāpu'u, 'ilima, alahe'e, alani, 'ōhi'a lehua, mokihana and wiliwili (Dunford, et. al. 2013).

## SETTLEMENT & PRE-EUROPEAN CONTACT

There is dispute as to the actual dates of arrival of the Polynesians who settled the Hawaiian islands. Current archaeological carbon dating points to 1000 CE as the approximate date of first settlement in the islands although ranges from 800-1200 CE are possible (Kirch, 2011 and Cordy 2000). Two possible sources for the voyagers who made their way to Hawai'i are the Marquesas (Nu'uhiwa) c. 900 CE and Tahiti (Kahiki) c. 1200 CE (Dunford et. al. 2013).

Polynesian settlers sailed with many plants and animals on their wa'a (canoes). The history of settlement is also the history of agriculture, and of species introduction. During the pre-contact era up to about 1450 CE, when migration seems to have slowed perhaps due to the Little Ice Age (Dunford, et al. 2013), several species were introduced.

Species introduced by Polynesians:

pua'a (pig), moa (chicken), 'īlio (dog), 'iole (rat)

kō (sugar cane), 'ohe (bamboo), niu (coconut palm), kalo (taro),  
kī (ti), pia (Polynesian arrowroot, ), uhi (yam)  
Pi'a (Five-Leafed yam), mai'a (banana), 'ōlena (turmeric)  
'awapuhi (wild ginger), 'awa (kava), 'ulu (breadfruit)  
wauke (paper mulberry), pa'ihi (nasturtium), auhuhu (Fish Poison  
plant), kukui (candlenut tree), hau (hibiscus), milo (Portiatree)  
kamani (Alexandrian laurel), 'ōhi'a 'ai (mountain apple)  
'uala (sweet potato), kou (Cordia wood), noni (Indian mulberry)  
ipu (Bottle gourd) (Dunford, et. al. 2013 and St. John et. al 1980).

The introduction of these new species provided great sustenance for the kanaka maoli (Hawaiians) (Dunford et. al. 2013). These species, however, also began to encroach upon the endemic pre-settlement species. Pua'a dug up rooted vegetables and "the main source of destruction of the native forests was the introduction of the Polynesian rat, *Rattus exulans*" (Athens et. al, 2002). Prehistoric avian species also suffered from the rat but also from human settlement as initially forests where the birds resided were burned and cleared for agricultural development by the settlers.



## LAND DIVISIONS AND SOCIETAL STRUCTURE

***“Hawaiian integrated farming systems evolved and proliferated within a unique socio-cultural context” (Costa-Pierce, 1987).***

### AHUPUA‘A

Islands in the Hawaiian language (‘ōlelo Hawai‘i) were called mokupuni. Mokupuni were divided into moku (districts) and within these moku were created smaller areas called ahupua‘a (Williams, 1997). In some ahupua‘a there were even smaller areas: ‘ili kūpono and ‘ili ‘āina (Dunford et.al, 2013 and Cordy, 2000). Most important, however, were the ahupua‘a.

Ahupua‘a usually ran from mauka to makai (mountain to ocean) with possible smaller ones that didn’t have this feature. Residents worked and gathered within their ahupua‘a which were designed to provide resources for them from upland crops to ocean provisions (William,1997).

There were three distinct areas within these ahupua‘a: uka, which included mountain and upland areas; kula, the flat and sloping plains and fields; and kai, the seashore and sea environment sometimes up to a mile offshore (Williams, 1987). Frequently the uka and kula zones would be terraced cross-slope to retain soil and prevent erosion. However, this pattern was notably different in the dry Kona region, where kua‘iwi, or stone ridges, ran mauka-makai in a diverse matrix of crops (Lincoln, 2014).

The Kona Field System was considered a marvel by early European visitors, and was indicative of the intensive agricultural activity and horticultural expertise of Hawaiian farmers. Archibald Menzies, a botanist who traveled with Captain George Vancouver, wrote in 1794:

“On leaving this station, we soon lost sight of the vessels, and entered their breadfruit plantations...The size of the trees, the luxuriance of their crops and foliage, sufficiently show they thrive equally well...The space between the trees did not lay idle. It was chiefly planted with sweet potatoes and rows of cloth plant (wauke). As we advanced beyond the breadfruit plantations, the country became more and more fertile, being in a high state of cultivation...In clearing the ground, the stones were heaped up in ridges between the little



*Figure 7: Kua'iwi mauka-makai wall in a Hōnaunau field.  
Height is 2 ft, width is 12 feet.*

fields and planted on each side, either with a row of sugar cane or the sweet root of these island (ti)...so that even these stony uncultivated banks are by this means made useful to the proprietors, as well as ornamental to the fields they intersect. The product of these plantations, besides the above mentioned, are the cloth plant, taro, and sweet potatoes...The whole field is generally covered with a thick layer of hay, made from the long coarse grass or the tops of sugar cane, which continually preserves a certain degree of moisture in the soil that would otherwise be parched by the scorching heat of the solar rays...Their fields in general are productive of good crops that far exceed in point of perfection the produce of any civilized country within the tropics.”

The kua’iwi system is still evident today, and forms the backbone of land in use for agriculture and coffee in South Kona.

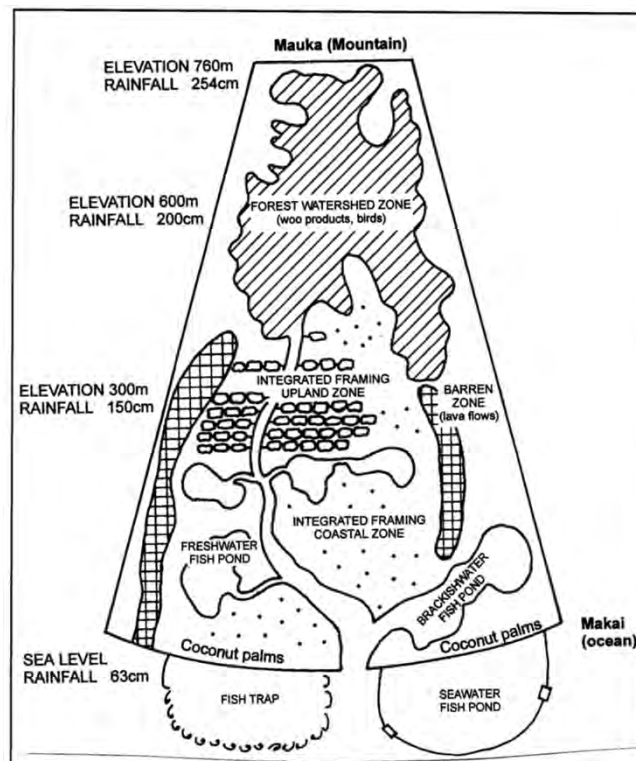


Figure 8: Example of individual ahupua'a configuration (Davidson-Hunt, 2021)  
Adapted from Costa-Pierce (1987)

Within each ahupua'a area, crops were cultivated for specific microclimate zones. Uka provided trees and plants used for canoe-building, weaponry, tools, cloth (kapa), cordage, lei and feathers for ali'i clothing collected from the native birds in these upland forests. The kula plains grew most of the food plants including mai'a (at the fringes of uka), kalo, 'ulu, 'uala and uhi. Kukui for oil, ipu for gourds, ki for capes and pili grass for thatched roofing were also grown in the kula areas. Finally, kai was where Hawaiians resourced fish (i'a), salt (pa'akai),

limu (seaweed), coconut, hau, hala and noni. The kai sections, especially in leeward areas where the water was calm and shallow, sometimes were host to the loko i'a (fish pond). These loko i'a housed Hawaiian fish farms which are being revitalized even today (Dunford et. al, 2013 and Williams, 1997).

Governance of these ahupua'a followed a distinct chain of command. Mokupuni were led by an ali'i nui (high chief). Each moku, or district, within the mokupuni was governed by an ali'i 'ai moku (lesser chief). Ahupua'a divisions within a moku were controlled by the ali'i 'ai ahupua'a who in turn had konohiki (headmen) to oversee the people (maka'ainana) farming and caretaking the lands. Sometimes the ali'i 'ai ahupua'a and konohiki were the same person (Dunford et. al, 2013).

## LAND TENURESHIP

Most of the population chose to live in small villages on non-agricultural land near the shore or clustered around bays where the air was warm and dry (Dixon, 1789). Hawaiian settlements developed around not just the environmental landscape, but also in accordance to societal organization of the ali'i, konohiki and maka'ainana (Kirch, 2011). Farming was usually done by a family unit known as an 'ohana. These family relationships were core to the pre-contact farming practices and of great significance to the Hawaiians (Costa-Pierce, 1987). 'Ohana created and maintained complex agricultural systems "that connected agricultural watersheds to oceanic environments" (Costa-Pierce, 1987).

The traditional management system for the early Hawaiians was based on strict kapu, laws meant to preserve societal order. These kapu pertained to aspects of daily life which included practices in religion, ways of eating, areas one was allowed to enter and times of harvest and gathering to name just a few. Some of these kapu were so strict they carried the penalty of death (Dunford et. al, 2013). In general practice, the 'auhau (taxes) were gathered during the Makahiki (gathering time for collecting taxes with focus on more celebratory aspects of life versus war) (I'i, 1959).

The concept of land ownership viewed through Western culture is far different from the Hawaiian socio-cultural understanding of ownership. The maka'ainana worked the land for the ali'i 'ai moku who oversaw the district in turn for the ali'i nui. In essence, it was a system of feudal tenureship with freedom to move within the ahupua'a and with the responsibility to pay your taxes in the form of food and animals once a year to the ali'i (Handy & Pukui, 1998).

This idea and practice of tenureship is what would help contribute to the downfall of the Hawaiian farming practices. It would also provide the opening for Westerners, post-contact, to permanently change the landscape and traditional lifestyle and welfare of the Hawaiian people.

## EUROPEAN CONTACT & THE HISTORIC PERIOD

***“With the general demise of native Hawaiian society, the majority of Hawaiian integrated farming systems fell into disuse and disrepair” (Costa-Pierce, 1987).***

The arrival of Captain James Cook to the islands in 1778 CE heralded immense change for the Hawaiian people who had lived for approximately a millenia without contact except from other occasional Polynesian voyagers (Kirch, 1998).

The next most significant person in the initial contact years was Captain George Vancouver who had served as an officer to Cook. Returning in 1791 leading the second British expedition, he made several trips to the islands bringing cattle (pipi), goats, geese, sheep and oranges (Speakman & Hackler 1989 and Hawai'i Dept. Of Agriculture). Eventually, mangoes, papaya, plumeria, coffee and lychee would also be introduced in the early nineteenth century (Dunford et. al, 2013).

After Cook's arrival to Hawai'i, the islands become a stopping point and eventual base for Western political and economical expansion into the Pacific and Asia. Landscape and cultural changes sailed in with the explorers, New England whaling industry and the missionaries who arrived in its wake. Over time, the raising of the new crops and animals they introduced to Hawai'i would contribute to the undermining of the traditional farming practices (Lâm, 1989). Development of imported agricultural in the Hawaiian islands increased rapidly during the early nineteenth century. The increase in the foreign population and creation of whaler ports on several of the islands produced a new supply and demand chain that would forever alter the islands.

'Iliahi (sandalwood) became a major commodity in 1810 heralding the increased economic investment by foreigners. Eventually when the sandalwood trade waned, the damage to the traditional subsistence economy had been done. The whaling industry as well now had a foothold in the islands and the ali'i had incurred massive debt to the foreign investors. By 1826, the first gunboat incidence occurred when the U.S. Navy moored in Honolulu harbor attempting to forcefully collect on these ali'i debts.

The whaling industry impacted traditional Hawaiian lifestyles in many areas. The cash economy began to supplant the previous subsistence economy. Hawaiians began to relocate to the now town and city centers for work, with many men signing on to the whaling ships. Agriculture turned to growing crops to be sold to the peoples inhabiting these areas and to provision all the trade and merchant vessels at port. Disruption of the agricultural farming systems that had served Hawai'i for a millenia seriously impacted the traditional socio-cultural basis for the kanaka maoli. It would pave the way for the end of land tenureship and the evolution of private property rights especially to be held by foreign entities (Kent, 1993).

## THE GREAT MAHELE OF 1848 and THE KULEANA ACT OF 1850

Foreign economic disruption of the traditional subsistence trade practices led to a cultural clash related to the concept of land ownership. Hawaiians' utilization of a method of tenureship approach to the land was in opposition to and undermined the Western cultures' idea of right to privately own land which placed great value both economically and politically on this type of usage.

Between 1839 and 1845, major shifts occurred within the Hawaiian political system in response to decades of foreign influence. Hawai'i was recognized as a consitutional monarchy by France, Belgium and Great Britain; the Bill of Rights was drawn up, and a constitution was signed in 1840 (Kamakakau, 1992). Several other pieces of legislation followed which would lead to the privatization of land ownership. The Act to Quiet Lands Titles was the first in 1844, initiating ten years of land ownership transformation. The Act created a Board of Commissioners to oversee the process of the division of lands between the king and his subjects. It also opened up the potential, perhaps not intentionally, for foreign buyers to gain a foothold into land ownership in Hawai'i.

The Great Māhele spanned the years of 1845 to 1855 culminating in The Great Māhele Act of 1848 and the Kuleana Act of 1850. The 1848 act relocated one third of the lands to the king, which would be known as crown lands, another one third to the konohiki or chiefs and the last third to the maka'āinana. Importantly, the initial Māhele did not change the tenureship concept for the maka'āinana (Lâm,1989).

The Resident Alien Land Ownership Act of July, 1850 and the Kuleana Act of August, 1850 would effectively be the instruments to commit the final severance. The Resident Alien Act gave foreigners the right to own land privately. The Kuleana Act gave Hawaiians two years to pay for and complete surveys on land that they were currently using but only up to 0.25 acre. Most Hawaiians did not understand nor took advantage of, nor perhaps weren't financially able, to take advantage of this process. At the end of the two years only 8,200 kuleana parcels were recognized and awarded which amounted to less than 1% of the lands (Lâm 1989). Combined, these two acts, whether good-intentioned or not, effectively ended traditional land use in the Hawaiian islands.

The rise of the plantation in coordinance with the sugar trade was a direct result of these processes. Labor and land were restructured to maximize profits in the hands of the owners of these plantations. These owners would eventually play a large part in the overthrow of the Hawaiian kingdom in the late nineteenth century (Kent 35-6).

## IMPACT OF INTRODUCED DISEASES

Prior to outside contact, Hawaiians had already suffered greatly from warfare, famine and infant mortalities. However, the economic and socio-cultural changes brought upon Hawai'i

were only part of the process of the change in society. For a long period of time, Hawai'i enjoyed the separation from the outside world and along with that, freedom from newly transmitted diseases. That changed with the arrival of Cook in 1778 and led to a steep decrease in Hawaiian population over the next century (Bushnell,1993 ).

Sailors on the voyaging ships introduced several venereal diseases, followed by tuberculosis in 1786. By 1804, Hawai'i saw its first large epidemic of what was most likely typhoid fever. Leprosy made its way to the islands by 1823 (Kamakau, 1992). There were continual outbreaks from 1826-57 derived from insect-borne disease, venereal disease and epidemics from inbound ships. An American warship brought in measles to Hilo in 1848 killing off 1/3 of the population. Several outbreaks of colds and flus occurred and by 1853 smallpox had arrived.

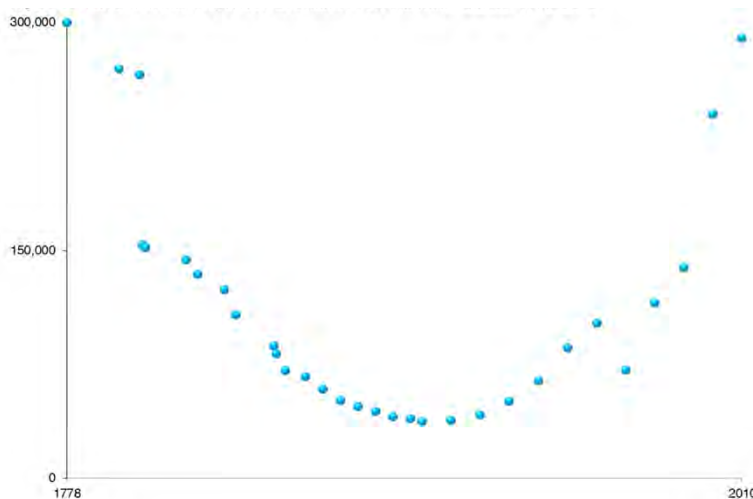


Figure 9: Map showing population decrease  
(Office of Hawaiian Affairs, 2017)

Decimation of the native Hawaiian population in the nineteenth century along with changes in the laws governing land ownership, created a space into which foreign investment and eventual political policy would lay the foundations for the modern era in Hawai'i.

## AGRICULTURE IN THE POST-CONTACT ERA

The rise of foreign influences and trading ports saw a divergence in the agricultural production of each island.

O'ahu, Maui and Kauai followed similar paths during the period from the late 1790's through the 1850's. Whaling ports were the main drivers for change on these three islands and Honolulu, Lahaina and Kōloa Harbors became major resupply points for ships.

## O'AHU

The first half of the nineteenth century saw a diversification of imported food crops, supplanting the traditional crops that had been grown by the Native Hawaiians. As was similar on the other islands, imported crops were grown to resupply the visiting ships and cater to changing tastes in a rapidly diversifying population. The rise in a cash economy supplanted the traditional subsistence and 'ohana-based structure.

The sugar industry was king during the mid-1800's but as the twentieth century fast approached, sugar began to wane economically. Other potential crops were explored for both local use and exportation. Specific to O'ahu, the plains of Wahiawa had developed an irrigation system and American homesteaders experimented with several crops. These included banana, papaya, fig, olive, orange, mango, pineapple and also coffee and vegetable oils, with pineapple and coffee eventually becoming the focal crops (evols.library).

Modern agriculture on O'ahu includes more than 40 different crops including pineapple, tropical flowers, coffee, melons, papayas, pumpkins, and bananas. O'ahu is also home to University of Hawai'i and the College of Tropical Agriculture and Human Resources (CTAHR). CTAHR is engaged in the study of and promulgation of agriculture throughout the Hawaiian islands.

## KAUA'I

Waimea was the first point of contact on Kaua'i for Captain James Cook in January of 1778. The south shore of the island would eventually host the whaling and sugar industries for the better part of the nineteenth century. Kōloa Village and Landing were the main point for distribution of products like sugar, molasses, beef and sweet potatoes to the ships (kauai.gov). Commercial pineapple as an industry navigated from O'ahu to the neighbor islands, especially Kaua'i and Maui as the previously-established sugar plantation farming methodology supported the growing and harvesting infrastructure for pineapples (Bartholomew).

Modern crops include papaya, tropical flowers, large kalo (taro) lo'i or ponded fields, and GMO biotech seed crops. GMO corn research fields were implemented on Kaua'i as early as the late 1960's and remain in rotation. The largest coffee plantation in the state is located in Kalaheo.

## MAUI

Like Kaua'i, Maui's agricultural history followed the whaling industry's needs from the 1820's to the 1850's. Crops shifted from traditional Hawaiian foods to those desired for the ships' stores. Lahaina on the west side was the main harbor used for the export of goods. Towards the

advent of the twentieth century, pineapple became a staple crop and eventually canneries were started on the island (Bartholomew et al., 2002).

Modern agriculture now includes a thriving coffee industry, cattle, pineapple, onions, papayas, tropical flowers, raw sugar, and the GMO biotech seed industry (mauicounty.gov).

## LĀNA'Ī

Lāna'ī has a uniquely different history of agricultural development than the other islands. The population had been decimated by wars within the Hawaiian kingdom's expansion under Kamehameha I and remained sparsely populated with subsistence farmers and fishermen. It wasn't until Walter Gibson arrived in the 1860's and acquired private land that agriculture shifted to more modern crops. Gibson brought ranching to the island which was followed by sugar from 1899-1921. The first pineapples were grown during the latter period of that time, and in 1921, James Dole acquired the island under private ownership. Soon Lāna'ī became known as the pineapple island (lanaichc.org). Pineapple was phased out of production by 1992, due to high labor and land costs. Today, with 91% of the island in private ownership, the focus is increasingly on tourism and resort development instead of major agricultural crops (Land Use Baseline).

## MOLOKA'Ī

Aquaculture and ranching were mainstays of the transitional agricultural landscape on Moloka'ī. When the Hawaiian Homes Act of 1920 was established, many homesteads were created on the north shore in Ho'olehua. Initially, land was leased out for pineapple production but moved into diversified crops as did the other islands, just at a later rate of change (hdoa.hawaii.gov).

Moloka'ī's strong winds and lack of water prevented the larger crop systems from maintaining economic sustainability. Pineapple companies left in the 1970's, as did a large portion of the population dependent on their income. Today, Moloka'ī is predominantly Hawaiian by population and the residents do not cater highly to tourism. In the homestead area, foodcrops such as banana, papaya, taro, sweet potatoes and onions are grown (molokai.org). There is a large commercial coffee farm in the Kualapu'u village area.

The GMO biotech seed companies comprise more than 50% of the crop production on the island and as with other islands, has become a controversial land use issue (molokai.org). The only true port on the island is Kaunakakai on the south shore.

## HAWAI'Ī

Hawai'ī island has a rich history in agricultural development, both pre- and post-contact. A variety of ethnographic materials exist for West Hawai'i, primarily because it was the ancestral seat of a powerful line of hereditary chiefs, including Kamehameha. The early European visitors



paying their respects to the ruling power in the islands left behind journals and logs as they investigated the Kona and Kohala districts (Greene 1993).

As the largest of the Hawaiian islands, it also is home to an abundance of climate zones and can sustain a wide diversity of crops. About half of the state's commercial farms are located here (NASS Census, 2017).

Hawai'i Island's forests were host to the majority of the 'iliahi (sandalwood) growth. Kamehameha I controlled much of the trade, but on his death, the trade (and subsistence agriculture as a whole) began to fall apart for the Hawaiians. His kapu on felling young trees collapsed, and the mountains were eventually stripped of most of these trees. His son, Kamehameha II sank into debt as the crop declined and the industry had collapsed by 1830 (hawaiihistory.org). Kamehameha III banned the collection of sandalwood in 1839. This rare and expensive crop is still propagated and harvested on the upland slopes of the west side, albeit in very small quantities (nativeplants.hawaii.edu).

Many varieties of crops were introduced to the island, concurrent with other islands (nativeplants.hawaii.edu). This included oranges and cattle in the 1790's followed by pineapple and coffee by 1810. Commercial crops of mango, rice, eucalyptus and macadamia nuts were all introduced before the turn of the century. Sugar was primarily farmed in the south and east sides of Hilo, Hamakua and Puna until its economic collapse on the island in the 1990's.

On the Kona side, coffee production moved to the forefront during the mid-1800's. The ease of exporting the raw bean by sea trade allowed the crop to rise in prominence. The districts of North and South Kona were granted special labeling rules by the Department of Agriculture. With the closure of the last sugar mills in Hamakua (1994) Puna (1995) and Pahala (1996), a nascent coffee industry began to grow in these regions. Ka'u coffee (Pahala) rapidly grew in size and reputation.

There are many individual small farms focused on a large variety of crops including chocolate, honey, avocado, tropical fruits and flowers, sweet potato, and kalo. The GMO biotech seed crops also have a presence, mostly on the east side. Parker Ranch, in Kamuela, is one of the largest cattle ranches in the United States. Large macadamia nut farms are located in the Hilo and Ka'u districts.

## MODERN ERA

The Hawai'i of today is a far cry from what it was pre-contact. There are no illusions that life pre-contact was a perfect utopia. However, a Hawaiian such as Kamehameha I might be hard-pressed to see any familiarities of his time in the current era.

At this moment, the islands face many challenges. Hawai'i is deeply dependent on a tourist-based economy, which proved fragile during COVID-19 quarantines. Home ownership is

virtually impossible for many Kānaka Maoli as housing prices have risen well beyond what is affordable to many residents in a service-based economy. Even the neighbor islands of Hawai'i, Maui, Moloka'i, Lāna'i and Kauai have seen housing prices rise close to equal of those on O'ahu. This has led to an exodus of Hawaiians to the mainland United States in search of better jobs and housing opportunities.

On the upside, there is a nascent effort in smaller communities to restructure the economy. The focus is on industries that serve and benefit the community especially in the areas of economic, social and mental welfare. Agriculture is one of the industries that could help alleviate the reliance on tourism. Coffee, avocados, kalo, bananas, papayas, mangoes and pineapples are just a few of these crops that are produced locally. Perhaps with strong support to these farming endeavors, Hawai'i can reclaim its inherent agricultural proficiency in order to support a healthier economic base for its social and cultural communities.

## Community Interviews

To gain deeper understanding of the project area, a variety of stakeholders was interviewed for their knowledge of cultural practices within the coffee-growing Hawaiian islands: Oahu, Maui, Molokai, and Hawai'i Island. In keeping with the *Guidelines for Assessing Cultural Impacts* from the State's Department of Health - Office of Environmental Quality Control, interviews concerned not just coffee on these islands, but larger areas and cultural practices that could be affected by the release of *Phymastichus coffea*.

SHAC staff contacted eight community members for these interviews via telephone and email. Two declined, while six others agreed to be interviewed in May 2021. Each person contacted fits into one or more of the following categories: 1) Native Hawaiian cultural practitioner, 2) coffee farmer in Hawai'i, or 3) conservationist managing lands planted with Hawaiian coffee. To solicit additional feedback from members of the public who fit these criteria, a public notice was published on June 1 in Ka Wai Ola, the Office of Hawaiian Affairs newspaper and on their website at <https://kawaiola.news/hoolahalehulehu/public-notice-june-2021/>. No responses were received.

Following is the list of interviewees and the method of each interview:

Name of Interviewee	(Island) Title, Organization	Method of Interview
Shalan Crysdale	(Ka'u District, Hawai'i Island; Molokai) Hawai'i Island Program Director, The Nature Conservancy	Zoom
Hi'ilani Shibata	(Oahu) Co-owner, Ka Mahina Project; Lead Cultural Trainer of the Native Hawaiian Hospitality Association	Zoom
Kimokeo Kapahulehua	(Maui) President, Kimokeo Foundation	Facetime
Bryce Nakamura	(Kona District, Hawai'i Island) Third generation Kona coffee farmer	Zoom
Chuck Leslie	(Kona District, Hawai'i Island) Third-generation Nāpō'opo'o fisherman	Zoom
Wally Young	(Ka'u District, Hawai'i Island) Ka'u coffee farmer	In person

Each interview started with a short introduction to *P. coffea*, including photos of the parasitoid wasp laying eggs in a coffee berry borer (CBB) beetle as it entered a coffee cherry. All interviewees already were aware of CBB and its threat to Hawai'i-grown coffee. Points emphasized included the following:

- *P. coffea* originally is from Africa, and it has been introduced to coffee producing countries, such as Colombia, Guatemala, Honduras, Jamaica, El Salvador, Ecuador, India, Brazil, and Mexico for biological control purposes.
- *P. coffea* was brought from Colombia into a quarantine containment facility in Volcano for 1.5 years of tests. PBARC scientist Peter Follett aimed to determine whether the wasp might attack other beetle species and thereby pose a risk to the environment.
- During the USDA's tests, the wasp did not impact any native species. Only 5 insect species were parasitized by *P. coffea*, including CBB and Tropical Nut Borer, a macadamia nut pest. *Phymasticus coffea* appears to be very specific in what it attacks and should cause no harm to the environment if released for CBB control.

- The wasp cannot sting humans or animals.

Subsequent questions focused on four areas: 1) each individual's background and cultural practices, as well as experiences with pests and plant diseases that impact their cultural practices; 2) their knowledge about coffee production and Hawaiian agriculture; 3) their views about proper methods of pest control; and 4) any additional comments and concerns. SHAC staff prepared draft summaries of participants' interviews for them to review and add revisions. Below are the approved summaries of each interview:

### Shalan Crysdale, The Nature Conservancy

Since 2009, Crysdale has been working on Hawai'i Island for The Nature Conservancy (TNC). He began his tenure with TNC as the field coordinator for the Ka'u Preserve, was promoted to natural resource manager, and is now the Hawai'i Island forest program director. As such, he is directly responsible for three units of TNC-owned lands: Ka'u Preserve, Kona Hema in South Kona, and Kamehame in Ka'u District.

Of these three, Kona Hema has a few patches of naturalized planted coffee. Situated on old terraces, these thick patches of coffee may date to the turn of the 20th century, Crysdale says. In addition, Kona Hema has an experimental, high-elevation strand of macadamia nuts planted by longtime agribusiness developer Sally Rice, who currently co-owns consultancies Agricon Hawaii and Agro Resources Hawaii.

Another TNC-owned unit, Pelekunu on North Molokai, is the site of a long-gone village that once grew coffee. Some coffee trees still exist there, Crysdale says.

Like others interviewed for this CIA, Crysdale doesn't know of any traditional Hawaiian cultural practices utilizing the coffee plant, fruit or seeds. Instead, coffee was a cash crop that many Japanese families depended on at the turn of the 20th century. Crysdale recalls hearing stories about agricultural workers who declined to renew their contracts as sugarcane workers, choosing instead to grow coffee on the Kona side of Hawai'i Island. For decades, those farms have provided harvesting jobs for new arrivals to the island. Coffee picking, Crysdale says, "is an entry point to Hawai'i living."

One hundred years ago, the farmers had limited themselves to the best areas for growing coffee. But as the popularity of Kona coffee grew, Crysdale increasingly saw native forests and more marginal lands converted to coffee farms. With the addition of more farms came an increased reliance on herbicide.

"In the long run, that's a negative," he says. We don't want to see that show up in our water table."

Crysdale himself has some experience with tending coffee. This past season, from trees surrounding his home, Crysdale's family had a small harvest that resulted in about 25 pounds of roasted coffee. While this is a small amount, it was enough for Crysdale to see firsthand the unfortunate impacts of CBB on his own crop.

For Crysdale, protecting agriculture and ecosystems from introduced pests is "very top priority. What we're dealing with in Hawai'i is a rate of extinction that's unparalleled anywhere else in the world. It was like a flatline of species lost until these last 200 years."

He pointed out a few pests that have impacted his conservation work: 1) Rats are the number one pest in the forest, especially for forest birds with low-lying nests. 2) Mosquitoes carry avian malaria. 3) Invasive plants, such as strawberry guava and Christmas berry, grow prolifically and crowd out native plants.

There have been advancements in controlling these pests, Crysdale says. Automatic rat traps reset themselves and release just a little non-toxic bait, preventing the accidental poisoning of native birds. Sterilized male mosquitoes mate with female mosquitoes and leave them barren. *Tectococcus ovatus* is a biological control for strawberry guava. But to Crysdale, the best cure is prevention. He would like to see the State invest in more robust inspections and severe penalties.

Crysdale generally is supportive of insect biocontrols because of the success of *Eurytoma erythrinae*, a parasitoid wasp of the Erythina Gall Wasp (*Quadrastichus erythrinae*). Before the release of *E. erythrinae* as a biocontrol, the Erythina Gall Wasp was unchecked in laying its eggs in the leaves and stems of *wiliwili* trees, a dryland forest species native to Hawai'i. Crysdale recalls *wiliwili* trees with gnarled new growth. Severe infestations resulted in defoliation, or even death.

The release of *E. erythrinae* had an "instantaneous" effect, Crysdale says. The difference was like "night and day." New growth looked normal again, and *wiliwili* trees started growing at South Point on Hawai'i Island for the first time in years.

He hopes *P. coffea* would have a similar effect on CBB, for the sake of coffee and other host plants. Crysdale wondered whether CBB also infests any native plants. If so, *P. coffea* would benefit those as well.

"The idea that we are going to fence and spray ourselves out of these [pest] problems is too hard," Crysdale says. "Biocontrol is better. Let nature be a solution to nature. This is a very effective tool."

Hi'ilani Shibata, Ka Mahina Project and Native Hawaiian Hospitality Association

Originally from Hilo and now living on Oahu, Shibata is a longtime educator of Native Hawaiian cultural practices and history. She is co-owner of the Ka Mahina Project, which promotes a healthier life through traditions that honor Hina, the Hawaiian moon goddess. Shibata also is lead cultural trainer for the Native Hawaiian Hospitality Association. Previously, she spent 14 years as education manager at the Bishop Museum in Honolulu. Shibata's own cultural practices include *lomilomi* and traditional *ho'oponopono*.

She also has conducted farmer education, based on her own family's experience with small-scale agriculture. Her husband had a two-acre farm that grew crops such as taro, *'ulu*, sugarcane and bananas -- just enough to feed family and friends. (They are looking for another plot of land to resume farming.) Over the years, she has seen growth in the number of Hawai'i's small and large farms. She hopes to see the establishment of more small ones.

Shibata's family doesn't grow coffee, but she has participated in coffee harvesting and processing at the Hawai'i Agricultural Research Center. She has noticed a difference in flavor between coffee produced on Hawai'i Island, versus coffee grown on the other Hawaiian islands. It's a variance she attributes to Hawai'i Island's younger volcanic soils.

Kona coffee has contributed much to the history of Hawai'i, especially since it's known globally, Shibata says. As coffee is not a traditional Hawaiian plant, she doesn't know of Native Hawaiian cultural practices that incorporate it. "It's not like they rejected it," she says of the Polynesian pioneers to Hawai'i. "I just don't think it's something they had."

Shibata has seen invasive pests affect both agriculture and plants important to Native Hawaiian culture. *'Uala* and taro are targeted by sweet potato weevil and apple snails, respectively. On her husband's farm, they noticed longneck turtles, poisonous dark frogs, and Japanese eels -- all non-native species, Shibata says. *Wiliwili* trees have been harmed by the Erythrina Gall Wasp. And the leaves of the *hala* tree, used by lauhala weavers, suffer from hala scale.

"Any time a native plant is affected negatively, it will have multiple effects on our culture," Shibata says. Since shipping introduces invasive species, she hopes more local agriculture would reduce imports. Shibata also would like to see more inspectors looking for invasive pests: "Protecting agriculture and ecosystems, it's really important. And it's really hard, because there's very little money."

When it comes to controlling pests, Shibata prefers physical and biological controls. She has participated in removing invasive miconia trees. And she's in favor of parasitoids -- as long as they are researched as extensively as the one that saved the *wiliwili* trees from the Erythrina Gall Wasp. "I'm not into chemicals because they go into our water systems," she says.

Kimokeo Kapahulehua, Kimokeo Foundation

Born in Lihue, Kauai and now living in Kihei, Maui, 73-year-old Kapahulehua is a cultural educator with a long history of spreading Native Hawaiian traditions. The organization he founded, the Kimokeo Foundation, describes Kapahulehua's work this way: "His accomplishments are vast, spanning from being heavily involved in Hawaiian outrigger canoe paddling and voyaging, to the preservation of Native Hawaiian forests, to the revitalization of an ancient Hawaiian fishpond to educating thousands of youth about the Hawaiian culture and its practices, to raising money for cancer survivors."

For all of his life, Kapahulehua's cultural practices have involved the sea. He grew up in a fishing family that used both nets and spears to catch their prey. By the time he was about eight years old, he was paddling the outrigger canoes called *wa'a*.

Kapahulehua eventually became Maui Island's Gray Line tour manager. As an adult, his most ambitious project was to travel by *wa'a* along the length of the entire Hawaiian island chain. Starting with Hawai'i Island and ending with the Kure Atoll, the voyage took his team six years and spanned 1,750 miles.

Kapahulehua also has led teams in harvesting logs to make *wa'a*. In 2000, his team harvested a log for a canoe at Haleakalā, Maui. It was the first time in 64 years that a log was cut down for this purpose. In 2010, his team went to Mauna Kea and harvested eight logs to make canoes. Each time, they followed protocols established by their ancestors.

Kapahulehua points out that Native Hawaiians originally didn't have coffee, so there were no traditional cultural practices with it. "When I was brought up, there was only one place to get coffee," he says. "That was in Kona." He recalls seeing family members in Kona harvesting red coffee cherries, as well as coffee beans drying on platforms known as hoshidanans.

As coffee cultivation spread to areas such as Ka'u, Molokai and Maui, it became "a significant way of life for our people," Kapahulehua says. Coffee farming supported a lot of families and is now as integral to Hawai'i's agricultural history as sugarcane, pineapple, papaya, banana, ti leaves, anthuriums, bird of paradise and other tropical flowers, he adds.

For all of the above reasons, he considers it very important to control coffee berry borer (CBB) and other pests. "We're the number one state in growing coffee," he says. "That's a greater concern -- for us to maintain that industry."

Kapahulehua has seen pests damage other industries and ecosystems. The *ta'ape*, a yellow-skinned snapper introduced to Hawai'i by what was known as the Division of Fish and Game in the 1950s and 1960s, eats the eggs of native fish, he says. Mongolian seaweed, an invasive species, has overtaken native Hawaiian seaweed in some ocean areas. The US Fish & Wildlife Service now requires inspections of canoe hulls and boat hulls to prevent the spread of this invasive species.

To protect both farms and natural ecosystems, Kapahulehua prefers biocontrol methods over chemical sprays as long as they are tested properly. Such a process would involve scientists studying flora and fauna, in addition to entomologists, he says. With regards to *P. coffea*, the parasitoid wasp that would kill CBB, Kapahulehua questions what percentage of native insect species in Hawai'i were tested against it. His concern is adequate testing to ensure the protection of Hawai'i's endangered insects -- such as moths, as well as of Hawai'i's native plants and fruits.

### Bryce Nakamura, Kona coffee farmer

Nakamura, 67, is a third-generation Kona coffee farmer. He is descended from Japanese immigrant laborers for Hawai'i's sugar industry. His great-grandfather established the family farm on 30 acres of Bishop Estate (now Kamehameha Schools) land overlooking Kealahou Bay. The family's first crop was tobacco, followed by coffee.

Coffee's importance to his family is economic. Before tourism grew, agriculture was the main industry in Hawai'i. And back then, anyone who leased Bishop land was required to improve it with agriculture, Nakamura says. In subsequent decades, the Kona coffee brand helped build more farms.

Watching his father work so hard on the farm convinced Nakamura to become a pharmacist. He spent 29 years working at Kona Community Hospital before retiring. "I went to school to run away from coffee," he says. But now that Nakamura's father has died and his mother is in her 90s, the responsibility for tending the fields rests on him.

Granted, the acreage isn't as much as it used to be. Nakamura's father sold off most of the farm in the early 2000s, leaving 5.5 acres of Bishop Estate land under the family's control. Two acres are planted with interspersed macadamia nut trees and coffee trees. A separate 1-acre plot is planted with only coffee.

When asked if he knew of Native Hawaiian cultural practices that involve coffee, Nakamura couldn't think of any. His family's own Japanese cultural practices consisted of pounding mochi with a rock his great-grandfather found in Waipio Valley and crafted into a mochi pounding bowl, as well as going to Obon dances. None of these activities have been affected by pests, but his farm certainly has been.

Nakamura knows firsthand what it's like to battle coffee berry borer (CBB). It's recommended that farmers spray *Beauveria bassiana*, the fungus that desiccates the beetles upon contact, every three weeks in his area. But the CBB population is high in nearby wild coffee stands and poorly-tended neighboring farms -- which means Nakamura must spray every two weeks to control the beetles on his own farm. He sees a difference between the CBB populations in his two fields: On the one acre planted only with coffee, the CBB infestation stays under 5%. But in



the field that is macadamia nut trees interspersed with coffee trees, the CBB infestation stubbornly stays at about 15%. If he didn't spray *B. bassiana* at all, the infection rate in both fields would shoot up to 70%-80%, he says.

In addition to CBB in his coffee trees, Nakamura has a pest problem on his macadamia nut farm. Beetles are boring into the trunks of his macadamia nut trees, which releases resin and allows a fungus to enter. So far, he says he has lost 50%-60% of his trees to this fungus. Last season, his farm's nut production decreased by 70%. Instead of replanting macadamia nut trees, Nakamura slowly is letting the coffee take over that section of the farm.

Based on these experiences, Nakamura says it's a good idea to protect agriculture from pests. He considers both sprays and insect biocontrols to be important in this goal. Subsidies for farmers -- such as the federal and state programs that reimbursed coffee farmers for *B. bassiana*, are the best way to motivate growers to use these pest-control methods. Critical to these programs' success, however, is ease of applying and record keeping. He personally found the federal reimbursement program for *B. bassiana* easier to use.

As for the parasitoid wasp *P. coffea*, Nakamura was heartened to hear that it didn't attack humans or animals. As long as *P. coffea* doesn't harm native species or populations of beneficial insects, "I have no reservations here," he says. One concern is whether he can maintain his schedule of spraying *B. bassiana* every two weeks without hurting the parasitoid wasp.

In general, Nakamura would like to see additional biocontrols similar to *P. coffea*. "Anytime you can get nature to work with you, it's better," he says.

## Chuck Leslie, Third-generation Nāpō'opo'o fisherman

Charles "Chuck" Kealoha Leslie, 80, is one of the few remaining Native Hawaiian fishermen of 'opelu, a type of mackerel. He grew up along Hawai'i Island's Kealakekua Bay, where his father started teaching him how to hand weave fishing nets at the age of five. Now, Leslie is the last in his lineage still fishing at Kealakekua Bay with traditional nets. The ones he makes are particularly good at catching larger volumes of fish. Leslie is training a younger generation in net weaving, before the art is lost.

Leslie's family also has a long history in Kona coffee. His great-grandfather, John Gaspar, built Hawai'i's first coffee mill in 1880. Leslie himself spent four seasons working in the Captain Cook Coffee Mill. He recalls days of hauling 1,600-1,800 heavy bags of fresh coffee cherries, loading them into a pulper to strip off their skins, then spreading their seeds to dry on covered platforms called *hoshidanas* or in mechanical dryers.

While growing up, Leslie harvested coffee with his siblings and parents. The farmers they worked for included Bryce Nakamura's father. (Nakamura also was interviewed for this Cultural

Impact Assessment.) Even though coffee was much cheaper when Leslie was a child compared to now, there was still money to be made, he says.

Like the other interviewees, Leslie doesn't recall any Native Hawaiian cultural traditions around coffee. Interestingly, peak *'ope'lu* fishing season coincides with peak ripeness of the coffee harvest in Kona -- a parallel that has remained true over the decades despite variations in harvest season from year to year, he says.

Leslie is familiar with coffee berry borer (CBB) and its damaging impact on Hawai'i's coffee industry. He believes in quickly protecting agriculture and ecosystems from invasive species. "If you know it's gonna be a pest, get rid of it as soon as possible," he says.

To Leslie, past introductions of non-native species offer cautionary tales. Roi, a type of grouper that was brought from French Polynesia to Hawai'i in the 1950s, has since spread to coral reefs throughout the State. In addition to eating native fish, roi can harbor the toxin that causes ciguatera fish poisoning. Leslie says a number of his friends have fallen ill with ciguatera -- sometimes from roi, and sometimes from other species. Like Kimokeo Kapahulehua (see interview above), Leslie points out that the yellow-skinned snapper called *ta'ape* eats the eggs of native fish. It was introduced to Hawai'i by the Division of Fish and Game in the 1950s and 1960s. *To'au*, an invasive blacktail snapper, hurts coral reefs. And gorilla ogo, a seaweed that was introduced to Hawai'i with the aim of producing agar, spreads quickly and overruns fishponds.

Some of these fish and seaweed species were introduced in an uncontrolled manner. After seeing their effects on oceans surrounding Hawai'i, Leslie is glad that potentially beneficial species are far more rigorously tested than they were before. Regarding the USDA assessment for *P. coffea*, he wonders what would happen if the parasitoid wasp is successful in eliminating CBB in Hawai'i. Would *P. coffea* also eliminate the other three species that the USDA identified as parasite targets? Or are there other environmental factors that would prevent their elimination?

He stressed the importance of using biocontrols such as *P. coffea* instead of chemical sprays, provided the proper research is conducted. Says Leslie: "If the wasp can control the CBB, that's better."

## Wally Young Sr., Ka'u Coffee Farmer

Wally Young, 79, was born in the Ka'u District's town of Waiohinu and has lived there almost his entire life. His father moved the family from Kona to Ka'u to join the sugarcane industry. At the time, there were two separate sugar mills in Ka'u: Hawaiian Agricultural Company in Pahala and Hutchinson Plantation in Na'alehu.

Young is one of 10 brothers and sisters who learned Native Hawaiian traditions from their mother. Her flower garden produced blossoms for lei making. She also gathered leaves from hala trees to weave hats, mats and lauhala baskets for harvesting coffee. "In Kona, that's how they made money," he said.

Young's family has an agricultural background. In the 1950s, his uncle ran the AC Young Farm in Kiola Ka'a, about two miles away from Waiohinu. The farm grew vegetables such as taro, cabbage, tomatoes and cucumbers. Young's father helped out, and also used some of the field to grow vegetables for his own family.

Cooking by Young's mother upheld Native Hawaiian traditions. She pounded poi -- both from the taro her husband grew and 'ulu from a tree in their yard. They would gather kukui nuts to make 'inamona, a condiment of pounded nuts, Hawaiian sea salt and chili pepper. Young's parents also taught him how to cook kahlua pig in an imu, a skill he has passed onto his children.

Young remembers five Ka'u coffee farmers in the 1940s and 1950s -- three in Waiohinu and two in Pahala. As a child, he used to pick for one farmer and would sometimes help in the wet mill. At the time, the Ka'u farmers used to sell their coffee as cherry or parchment to Kona operations.

Native Hawaiians didn't have traditional cultural uses for coffee or the coffee plant. Instead, the crop was important because it economically supported people, Young said. Japanese growers owned the early Kona coffee farms, and they hired the Native Hawaiians to harvest for them.

After serving in the Army for three winters in Germany, Young returned to Waiohinu and worked in construction. Eventually, the two sugar mills merged into a single company, called Ka'u Agriculture. Then he worked in Ka'u's sugar industry for 33 years, a job that allowed him to purchase property in Waiohinu. "When they brought down the last load of the sugar cane, it was sad," he said.

After the closure of Ka'u Agriculture in 1996, the sugarcane plantations were transformed into coffee fields by the former sugar-mill employees. Young was one of the first to start his coffee farm in 1997, with five acres of land. With the launch of the Ka'u coffee industry, Native Hawaiians, Portuguese and Filipinos started growing coffee in larger numbers, he said.

In the early years, making money from Ka'u coffee was difficult, Young says. His farm initially was a side business, while his main income was as an auto mechanic. As sales of coffee improved, Young was able to sell his mechanic business to his son and expand his farm to 16 acres.

Coffee has positively contributed to Hawai'i's larger cultural history, Young says. It's now one of the top agricultural industries in Hawai'i. The biggest negatives are the arrival of CBB, and now coffee leaf rust.

Young believes in attacking pests as soon as they arrive in Hawai'i. As a conventional farmer, he sees the benefits of both chemical and biocontrol methods for pests. "If it's invasive, I think you should control it right away," he said.

Young had no specific recommendations for testing the effectiveness of new pest controls. He trusts that the process is more stringent than in the 1950s, when the *ta'ape*, an invasive fish, was released into Hawai'i. "Now, they check 'em out real good and make sure it doesn't screw up the environment," Young said.

He welcomes *P. coffea* as an additional tool in the fight against CBB. But he wonders about its resiliency against various sprays on the farm: the fungus *B. bassiana*, a biocontrol for CBB; anti-fungal copper sprays for coffee leaf rust; herbicides and pesticides. Just *B. bassiana* alone requires spraying every two weeks. At the time of this writing, one local vendor was selling a gallon of this fungus for \$198.00, with a price increase expected soon. "It's really expensive," Young said.

He worries about killing *P. coffea*, but he can't reduce spraying to accommodate the wasp -- unless *P. coffea* demonstrates a strong ability to kill the CBB on its own.

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NASS: USDA National Agriculture Statistics Service  
[https://www.nass.usda.gov/Statistics\\_by\\_State/Hawaii/Publications/Fruits\\_and\\_Nuts/Coffee-01-26-2021.pdf](https://www.nass.usda.gov/Statistics_by_State/Hawaii/Publications/Fruits_and_Nuts/Coffee-01-26-2021.pdf)

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### CULTURAL IMPACT ASSESSMENT: INVASIVE COFFEE BERRY BORER BEETLE

At the request of the University of Hawaii, the Synergistic Hawaii Agriculture Council is preparing a Cultural Impact Assessment for the statewide release of a wasp (*Phymastichus coffeae*) to control the invasive Coffee Berry Borer beetle. The wasp is harmless to humans. Please contact Suzanne Shriner at 808-365-9041 or [suzanne@shachawaii.org](mailto:suzanne@shachawaii.org) to share your mana'o about any cultural or historical resources relating to the lands now in use for coffee growing or any other information you feel is relevant. This could include mo'olelo, history, or knowledge of traditional and customary practices (both past and present). Letters can be sent to 190 Keawe St, Suite 25, Hilo, 96720. ■