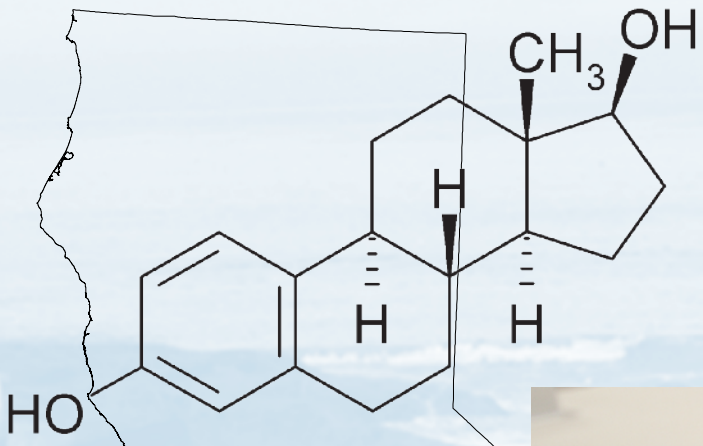
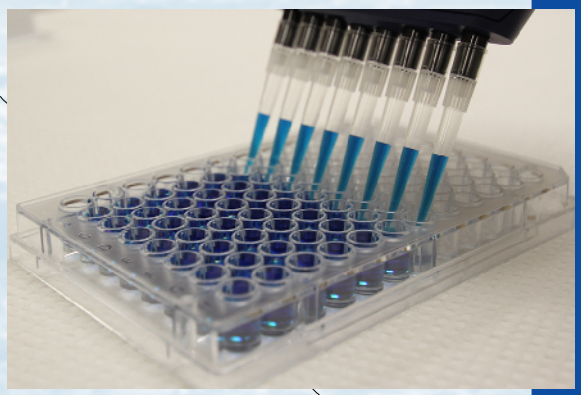
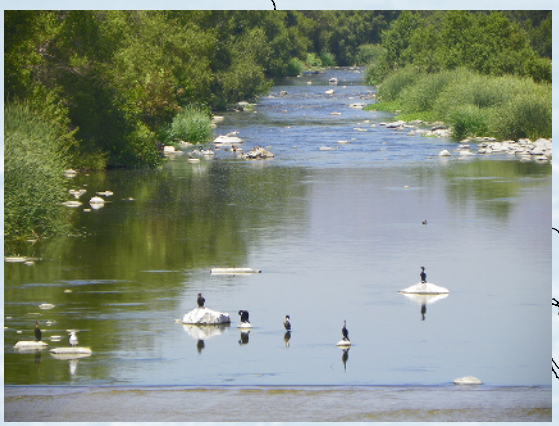
Monitoring of Constituents of Emerging Concern (CECs) in California's Aquatic Ecosystems - Pilot Study Design and QA/QC Guidance







Nathan G. Dodder
Alvine C. Mehinto
Keith A. Maruya

Southern California Coastal Water Research Project

SCCWRP Technical Report 854

Monitoring of Constituents of Emerging Concern (CECs) in California's Aquatic Ecosystems – Pilot Study Design and QA/QC Guidance

Nathan G. Dodder, Alvine C. Mehinto and Keith A. Maruya

Southern California Coastal Water Research Project Authority
Costa Mesa, CA

March 2015

Technical Report 854

Contents

1 INTRODUCTION

In October 2009, the State of California Water Resources Control Board (SWRCB) provided support for a scientific advisory panel to review existing scientific literature on constituents of emerging concern (CECs) in aquatic ecosystems; determine the state of the current scientific knowledge regarding the risks that CECs in freshwater and marine water pose to human health and aquatic ecosystems; and provide recommendations on improving the understanding of CECs for the protection of public health and the environment. Seven experts were vetted and convened as the CEC Ecosystems Panel (Panel) to provide information and recommendations on CECs[1] in coastal and marine ecosystems, which was subsequently tasked to expand the scope to include freshwater ecosystems. The Panel collaborated with stakeholders, who provided their perspective of the water quality issues and additional information during the development of their recommendations. In their final report, Monitoring Strategies for Chemicals of Emerging Concern (CECs) in California's Aquatic Ecosystems: Recommendations of a Science Advisory Panel, SCCWRP Technical Report 692, Anderson et al. (2012) recommended a risk-based screening framework to identify CECs for monitoring, applied the framework using existing information to three representative receiving water scenarios to identify a list of appropriate CECs for initial monitoring, developed an adaptive phased monitoring approach and suggested development of bioanalytical screening and predictive modeling tools to improve assessment of the presence of CECs and their potential risk to the environment.

Early in the process, the Panel was instructed by SWRCB staff to focus on ambient surface waters that receive discharge from sources regulated under the National Pollutant Discharge Elimination System (NPDES). As a result, permitted discharges from municipal wastewater treatment plants (WWTPs) and municipal separate storm sewer systems (MS4) were considered as the primary sources of CECs to receiving waters. Waterbodies that receive agricultural runoff were not considered.

1.1 Summary of Panel Recommendations

1.1.1 Adaptive Monitoring Strategy

The Panel recommended an adaptive monitoring approach with four sequential phases described below (**Fig. 1.1-1**) that is responsive to advances in assessment and monitoring technology.

PHASE 1 – PLANNING. The Panel met with scientists, managers and stakeholder groups representing local, regional and statewide interests, to learn about current CEC studies, regional and statewide monitoring programs, and NPDES permitted discharges that are relevant statewide. The Panel created a risk-based framework to identify high priority CECs based on available, peer-reviewed occurrence and toxicity information. In applying this framework, the Panel identified three exposure scenarios where WWTP and MS4 discharge could impact receiving water quality. These scenarios are (1) WWTP effluent dominated freshwater (rivers); (2) coastal embayments receiving both WWTP effluent and stormwater discharge; and (3) ocean discharge from large WWTP (> 100 million gallons per day) outfalls. The initial list of CECs was generated by comparing measured or predicted environmental concentrations (MECs or PECs) in aqueous, sediment and/or tissue to monitoring trigger levels (MTLs) based on biological effects thresholds that incorporated safety factors. CECs recommended for initial

[1] CECs may include a wide variety of substances including pharmaceuticals, flame retardants, newly registered contemporary use pesticides, commercial and industrial products, fragrances, hormones, antibiotics and nanoparticles that are not currently regulated in discharges to ambient waters across California.

monitoring exhibited a monitoring trigger quotient (MTQ = MEC/MTL) that exceeded unity (>1) and for which sufficiently robust analytical chemistry methods were available. The recommendations for Phase 1 were documented in the Panel's final report (Anderson et al. 2012).

PHASE 2 – DATA COLLECTION. The objectives of this phase are to: 1) verify the occurrence of high priority CECs in aqueous, sediment and tissue samples; 2) initiate compilation of a data set that characterizes their occurrence in source and receiving waters, and in appropriate matrices (i.e., water, sediment and tissue); 3) evaluate improved/supplemental methods and surrogate measures (e.g., bioanalytical screening tools); and 4) utilize, modify and/or initiate development of environmental fate models where appropriate. Screening-level mass balance models synthesize knowledge of CEC loading, and predict environmental compartment transfer and loss rates, as well as temporal CEC concentration trends. Through insight gained from these models, prioritization efforts in Phases 3 and 4 can subsequently focus on issues with the greatest potential risk.

PHASE 3 – INTERPRETATION. Using results from Phase 2, the list of CECs is re-evaluated and, if warranted, re-prioritized. Results of environmental fate modeling are evaluated to prioritize future monitoring and to conduct a preliminary review of the impacts of management actions.

PHASE 4 – ACTION PLAN TO MINIMIZE IMPACTS. If the assessment conducted during Phase 3 indicates certain CECs will persist and continue to present a concern, then during Phase 4 the Panel would develop guidance on the development and assessment of specific action plans for consideration by the SWRCB for implementation as part of their development of statewide policies, permits and/or guidance.

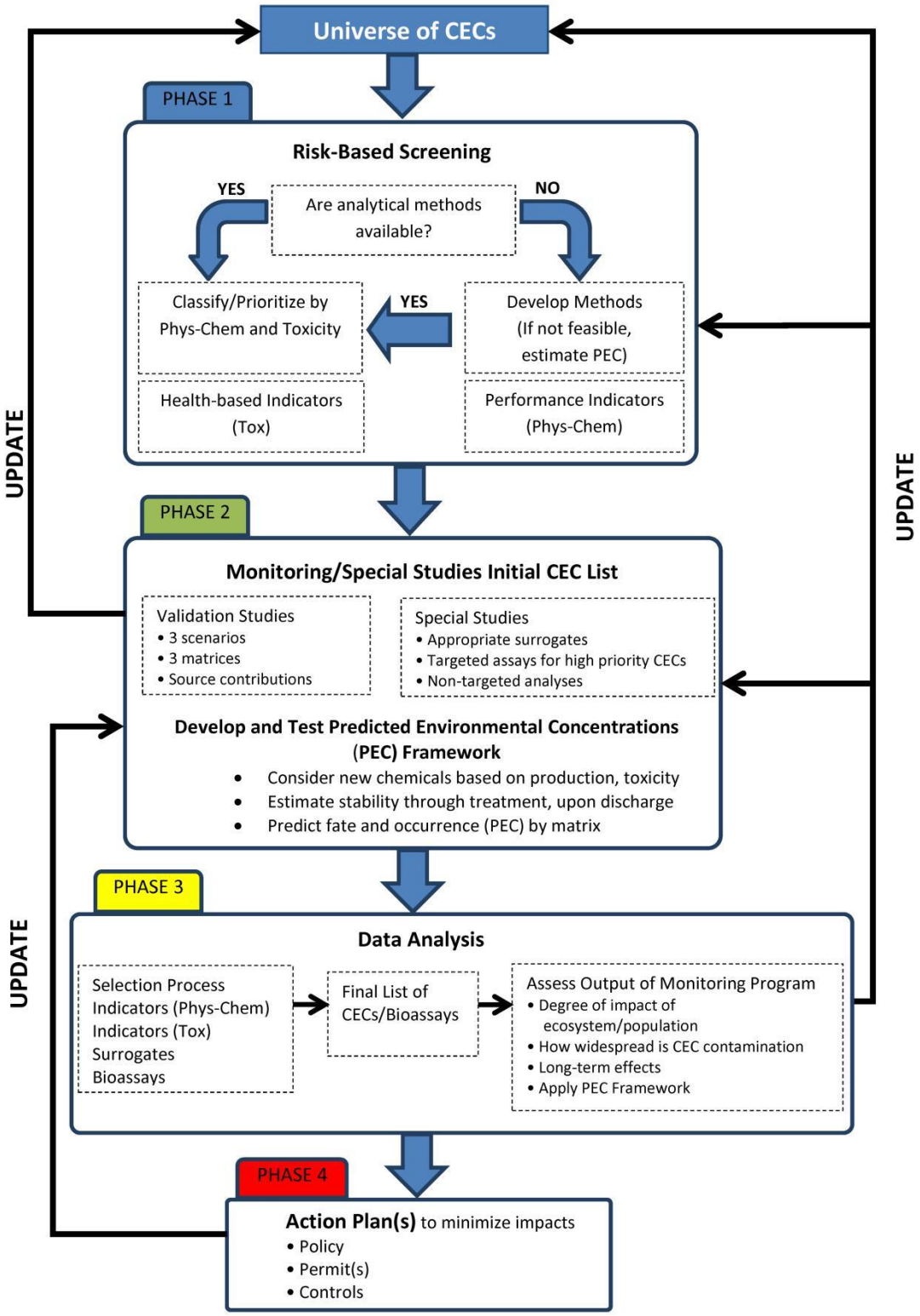


Figure 1.1-1. The adaptive monitoring strategy for constituents of emerging concern (CECs) developed by the Expert Panel convened to recommend CEC monitoring in California surface waters impacted by NPDES permitted discharges (i.e. treated wastewater effluent and stormwater runoff).

1.1.2 Discharge Scenarios

With guidance from the SWRCB and stakeholder community, the Panel identified three receiving water scenarios for which to provide CEC monitoring recommendations. These scenarios were selected based on the expected magnitude of CEC discharge from NPDES permitted sources and the severity of exposure to both human and ecological receptors.

1. Inland freshwaters where flow is dominated by treated WWTP effluent discharge (dry season).
2. Coastal embayments receiving treated WWTP effluent and stormwater (MS4) discharge (dry and wet seasons).
3. Offshore marine waters receiving treated effluent from large (>100 mgd) WWTPs.

These scenarios were considered separately because they have distinct differences in spatial and temporal source characteristics, fate and transport processes, and receptors of interest that define beneficial uses of the resource. A detailed description of relative CEC source contributions and exposure conditions for each of the three scenarios is provided in the Panel's final report (Anderson et al. 2012).

1.1.3 Initial List of CECs by Discharge Scenario ("Targeted Monitoring")

A total of 16 individual CEC analytes were recommended for chemical-specific (or "targeted") Phase 2 monitoring; however not all 16 CECs were selected for all scenarios (see **Appendix A, Table 8.1-1**). Due primarily to the limited degree of attenuation (e.g. by dilution), the number of CEC analytes recommended for monitoring was greatest for the WWTP effluent dominated inland freshwater (Scenario I). In contrast, the smallest number of CECs recommended was for sediment and tissue, due in large part to the paucity of MECs and MTLs available for these matrices compared with water (aqueous phase).

- The Panel was also charged to provide guidance on implementation of targeted CEC monitoring. Guidance on the type and number of waterbodies, spatial coverage and frequency of monitoring was developed to address the highest priority questions (see **Appendix A, Table 8.1-2**), e.g. what is the occurrence (magnitude, pervasiveness) of target CECs in waterbodies representing each scenario? What is the spatial and temporal variation in CEC occurrence in these scenarios?

1.1.4 Special Studies to Improve CEC Monitoring

One of the key limitations to the risk-based framework utilized by the Panel to identify CECs for targeted monitoring was the lack of robust monitoring/occurrence/toxicity data (i.e. MECs and MTLs) for the vast array of possible environmental contaminants. In recognition of this limitation, the Panel recommended a number of special studies using emerging technologies and/or methods that if successful, would provide a more comprehensive and efficient monitoring program for receiving waters (Anderson et al. 2012). These studies will complement and/or direct traditional targeted analytical methods while providing additional information on the occurrence of unknown CECs, and will be based on biological responses of aquatic organisms at the cellular (bioanalytical screening) and organism (*in vivo* testing) levels (see **Appendix A, Table 8.1-3**).

1.2 Pilot Monitoring (Phase 2) Design Guidance and Requirements

The objective of this document is to generate guidance, and where applicable, requirements for pilot monitoring and special studies for CECs that address elements described in Phase 2 of the Panel's adaptive monitoring strategy (Fig. 1.1-1). These elements are broadly classified into targeted (chemical-specific) monitoring and special studies. ***The intent of this effort is to translate the Panel's recommendations into guidance and, where applicable, requirements at a sufficient level of specificity and detail that can be directed and incorporated into local, regional and/or statewide workplans for future monitoring***.

To ensure relevance to the management decision-making process, the Panel emphasized the need for a purposive (i.e. question or hypothesis driven) approach to monitoring, offering several questions to be answered by the proposed pilot monitoring and special studies monitoring:

1. Which CECs are detected in freshwaters and depositional stream sediments, and in which large California watersheds are they detected?
2. Which CECs are detected in marine waters and sediments adjacent to WWTP and significant stormwater outfalls and how quickly do they attenuate?
3. Which CECs are detected in coastal embayment/estuarine water and sediments?
4. What is the relative contribution of CECs in WWTP effluent vs. stormwater?
5. What is the extent and magnitude of PBDE and PFOS contamination in tissues of aquatic wildlife across the State? Does tissue occurrence correspond with sediment occurrence?
6. What is the direction and magnitude of change in CEC concentrations (in water, sediment and tissues) over a multi- year time period?
7. How do the Panel's assumed relationships, based on the new CEC data (e.g., MEC or PEC, NOEC and MTL), change the estimated MTQs?
8. Does the new information (Question 7 above) modify the Panel's assumption regarding CEC potential risk and if so, does it trigger the need to evaluate CEC control efforts?
9. Which bioanalytical screening assays are effective to screen for target CECs in environmental samples?
10. How efficient are bioanalytical screening tools to detect unknown CECs?
11. What is the relationship between effects of CECs *in vitro* and toxicity observed *in vivo*?
12. What are the toxic effects of CECs on aquatic organisms?
13. Is there a relationship between the occurrence of antibiotics and antibiotic resistance patterns in effluent, surface waters and sediments?
14. Can passive samplers be used as a robust monitoring tool for CECs?

1.2.1 Targeted Monitoring

The design guidance to be specified for targeted monitoring for the CECs, scenarios and matrices listed in Tables 8.1-1 and 8.1-2, and as described in the project agreement, are:

1. List of target CEC analytes, preferred methods and desired reporting limits
2. List of candidate waterbodies that represent exposure scenarios identified by the Science Advisory Panel
3. List of target media (e.g. water, sediment, biological tissue), and candidate target species
4. Frequency, number, and location of sampling stations within each candidate waterbody
5. QA/QC goals for measurement of CECs for incorporation into the Project Supplemental Guidance for Quality Assurance/Quality Control document (see Task 5 in Contract)
6. List of appropriate monitoring questions for each exposure scenario
7. Data analysis and assessment methods for each exposure scenario
8. Data management plan
9. Strategy to coordinate with existing monitoring programs

The development of targeted monitoring requirements is addressed in Section 2 of this document.

1.2.2 Special Studies

The design guidance to be specified for special studies monitoring for the elements in Table 8.1-3, and as described in the project agreement, are:

1. List of target parameters, preferred methods and desired measurement goals
2. List of candidate waterbody(ies) for each special study
3. List of target media (e.g. water, sediment, biological tissue), and candidate target species
4. Frequency, number and location of sampling stations to be evaluated within each candidate waterbody
5. Quality assurance/quality control (QA/QC) goals for measurement of specific parameters
6. Rationale for exclusion/inclusion of studies that differ from the Panel's final recommendations

The development of special studies requirements is addressed in Section 3 of this document.

1.2.3 Supporting/Related Documentation

In addition to the design guidance specified herein, guidance for QA/QC is addressed in Section 10 of this document. This supplemental guidance provides criteria and guidelines to ensure that robust measurement of targeted monitoring and special study parameters is achieved.

1.3 Relevant Water Quality Monitoring Programs in California

1.3.1 SWAMP

The Surface Water Ambient Monitoring Program (SWAMP, (http://www.waterboards.ca.gov/water_issues/programs/swamp/about.shtml) was created to unify and coordinate all water quality monitoring conducted by the State and Regional Water Boards. The SWAMP mission is to provide resource managers, decision makers, and the public with timely, high-quality information to evaluate the condition of all waters across the State. SWAMP accomplishes this through the design and external review of monitoring programs, and by assisting others in generating comparable data for integrated assessments that provide answers

to current management questions. SWAMP monitoring programs are each designed to address one or more of the following assessment questions:

- Status: What is the overall quality of California's surface waters?
- Trends: What is the pace and direction of change in surface water quality over time?
- Problem Identification: Which water bodies have water quality problems and are at risk?
- Diagnostic: What are the causes and sources of water quality problems?
- Evaluation: How effective are clean water projects and programs?

Current SWAMP efforts focus on two critical assessment needs: human exposure via consumption of contaminated fish in fishable waters (Bioaccumulation Monitoring Program) and aquatic ecosystem health in streams and rivers (Bioassessment Monitoring Program and the Stream Pollution Trends Monitoring Program [SPoT]).

The Bioaccumulation Monitoring Program addresses whether fish found in California's streams, lakes and coastal areas are safe to eat by measuring contaminant concentrations in fish tissue. The Bioaccumulation Oversight Group (BOG) guides the implementation of the Bioaccumulation Monitoring Program. From 2007-2011, the program carried out statewide surveys of contaminants in sport fish from lakes and reservoirs, the coast, and rivers and streams. These surveys documented widespread, and in some cases severe, impact of bioaccumulative contaminants on the fishing beneficial use (Davis et al. 2013, 2014). Methylmercury is the contaminant that poses the greatest concern for consumers of fish caught in California water bodies. PCBs are the second greatest overall concern, but had a far lower rate of occurrence of concentrations exceeding consumption thresholds. Thus, recent studies have focused on methylmercury in lakes, including a study of exposure and risk to piscivorous wildlife in 2012-2013, and a sport fish survey of lakes with low concentrations in 2014. This effort will continue focusing on California lakes, asking why some lakes have higher methylmercury levels in sport fish than others (SWAMP 2014).

Initiated in 2008, SPoT measures contaminant concentrations and toxicity in sediments that accumulate in the lower reaches of large watersheds throughout California and relates contaminant concentrations to watershed land uses. Sediment samples are collected annually when streams return to base flow conditions after pollutant mobilization in runoff and during the wet season has abated. Each sample is analyzed for industrial compounds, pesticides, and metals, and is tested for toxicity to a resident aquatic crustacean, the amphipod *Hyalella azteca*. Results are compared across watersheds statewide, and pollutant concentrations are compared to land use and other human activities. In 2012, samples were collected from 100 of the nearly 200 major hydrologic units in California.

The most current SPoT summary report for the period 2008-12 provides evidence that pesticides are associated with ambient toxicity in California waters (Phillips et al. 2014). As a result, certain emerging pesticides are being prioritized for future SPoT monitoring. In 2013, fipronil was added as a SPoT analyte due to increasing use and the potential for surface water toxicity. Also, SPoT began collaborating with the California Department of Pesticide Regulation (DPR) to evaluate the effectiveness of new restrictions on the use of pyrethroid pesticides in urban applications. Four "intensive" monitoring sites were jointly sampled by SPoT and DPR to

determine whether new regulations result in reduced pyrethroid concentrations and associated effects.

SPoT has plans to continue its monitoring focus on emerging pesticides. In 2015, SPoT will add the additional indicator organism *Chironomus dilutus* to assess the effects of fipronil and degradates. SPoT is also exploring the possibility of incorporating water column monitoring for imidacloprid and other neonicotinoid pesticides beginning in 2016. In collaboration with DPR and SWAMP, a pilot monitoring project is measuring these pesticides in agricultural streams in 2014 and assessing their effect using *C. dilutus*. Legacy pesticides, PCBs, organophosphate pesticides and metals will be monitored every other year.

In addition to monitoring and assessment activities, SWAMP develops implements and maintains a monitoring infrastructure and associated tools. Key components of this infrastructure include Quality Assurance/Quality Control (QA/QC) protocols, database and data management tools, water quality indicators, methods, and standard operating procedures. These tools are available to SWAMP partners and other interested parties via the SWAMP website. SWAMP leverages limited resources by coordinating with other water quality monitoring efforts on a local, regional and statewide level. SWAMP works with partners to coordinate monitoring efforts among many groups and agencies, and to facilitate the use of data from many sources in statewide assessments.

1.3.2 Department of Pesticide Regulation

The California Department of Pesticide Regulation (DPR) is the lead agency for regulating the registration, sales and use of pesticides in California. This agency oversees pesticide monitoring programs in air, ground and surface waters across the State. The Surface Water Protection Program (SWPP) (http://www.cdpr.ca.gov/docs/emon/surfwtr/overvw.html) characterizes pesticide residues, identifies pesticide contamination sources (both agricultural and non-agricultural), determines the mobility of pesticides to surface water, and develops site-specific mitigation strategies. Investigations are done in consultation with other agencies, including the State and Regional Water Boards. In order to promote cooperation, DPR and the SWRCB signed a formal agreement and developed a companion document, "The California Pesticide Management Plan for Water Quality," to coordinate interaction, facilitate communication, promote problem solving, and ultimately assure the protection of water quality (http://www.cdpr.ca.gov/docs/emon/surfwtr/maaplan.html). Under this plan, DPR investigates pesticides of concern and develops recommended pesticide use practices designed to reduce or eliminate the impact of pesticides on surface water quality. Management practices designed to reduce contamination are usually implemented initially through voluntary and cooperative efforts. If such voluntary practices do not adequately mitigate impacts, DPR can invoke its regulatory authority to impose use restrictions, e.g. by establishing permit conditions to prevent excessive amounts of residues from reaching surface water. If such steps are not adequate, the State and Regional Water Boards may use their authorities to mitigate the adverse effects of pesticides.

To determine if mitigation is effective, the Environmental Monitoring Branch of DPR conducts monitoring studies on pesticides of concern. Two such studies planned for 2014-15 are focused on model watersheds in northern (Emsinger 2014) and southern (Budd 2014) California. Common to these regional studies are the measurement of target pesticides in water and sediment. Pyrethroids (including permethrin and bifenthrin), fipronil and degradates and

chlorpyrifos, identified as high priority CECs by the Panel, are included on DPR's analyte list. Sampling design for these studies focus on characterizing multiple events of dry and wet weather runoff into freshwater systems in suburban and urban neighborhoods.

In addition, DPR has conducted special investigations on the occurrence of pyrethroids in wastewater influent and effluent (Markle et al. 2014, Teerlink 2014). These data may reduce and/or obviate the need to monitor for pyrethroids in WWTP effluent as recommended by the Panel. A third DPR product that may serve useful in future prioritization and monitoring efforts is a model that predicts the mass of pesticides applied in urban landscapes that washoff and enter urban waterways (Luo 2014). Such models can estimate the occurrence of pesticides of concern (i.e. predicted environmental concentrations or PECs) where no measured data are available.

1.3.3 San Francisco Bay Regional Monitoring Program

The San Francisco Bay Regional Monitoring Program (RMP) (http://sfei.org/rmp) is a collaborative effort among the San Francisco Bay Regional Board, the regulated discharger community, and the coordinating entity, the San Francisco Estuary Institute (SFEI). The goal of the RMP is to collect data and communicate information about water quality in the Estuary to support management decisions. The RMP addresses five primary management questions (last refined in 2008), and which closely mirror those posed by SWAMP statewide.

1. Are chemical concentrations at levels of potential concern and are associated impacts likely?
2. What are the concentrations and masses of contaminants in the Estuary and its segments?
3. What are the sources, pathways, loadings, and processes leading to contaminant-related impacts?
4. Have the concentrations, masses, and associated impacts of contaminants increased or decreased?
5. What are the projected concentrations, masses, and associated impacts of contaminants?

More specific management questions under each of these five general categories, and for topics of particular interest, have also been articulated (SFEI 2014).

Status and Trends (S&T) monitoring in the RMP (http://www.sfei.org/content/status-trends-monitoring) is composed of the following elements:

1. long-term water, sediment, and bivalve monitoring
2. sport fish monitoring on a five year cycle
3. USGS hydrographic and sediment transport studies
    A. Factors Controlling Suspended Sediment in San Francisco Bay
    B. USGS Monthly Water Quality Data
4. triennial bird egg monitoring (cormorant and tern)

The RMP has investigated the occurrence and potential for impacts due to CECs since 2001[2]. Much of the pioneering work on flame retardants (e.g. PBDEs) and more recently, perfluorinated

[2] http://www.sfei.org/projects/chemicals-emerging-concern-strategy

compounds (PFCs) such as PFOS, have been conducted by the RMP as a result of recommendations made by the Emerging Contaminants Work Group (ECWG), a panel of stakeholders and internationally renowned scientists coordinated by the RMP. The role of the ECWG is to ensure the RMP is current with respect to CECs, and, as needed, to recommend, support and implement studies for consideration by the RMP Steering Committee. These studies have allowed for prioritization of these CECs using occurrence and toxicity data to determine the level of concern for individual contaminants in the Estuary.

The RMP recently synthesized the state of the science on occurrence of CECs in San Francisco Bay (Klosterhaus et al. 2013), including existing information on chemical usage, occurrence relative to other locations and toxicity. The RMP then developed a three-element CEC monitoring strategy (Sutton et al. 2013), which combines a) traditional targeted monitoring guided by a risk-based framework, similar to that proposed by Anderson et al. (2012), with b) review of the scientific literature and other CEC monitoring programs as a means of targeting new CECs, and c) non-targeted monitoring, including broad scan analyses of Bay biota samples and development of bioassays to identify estrogenic effects, both means of identifying previously unknown CECs present in the Bay. The major outcome of this effort is to provide updates on relevant information to the San Francisco Bay Regional Board and stakeholders including the ECWG, so that they may react and adapt to new information using a tiered risk-management action framework (Sutton et al. 2013).

RMP data, field operations and quality assurance/quality control (QA/QC) documentation can be accessed via on the SFEI website (http://www.sfei.org/programs/rmp-data). Results provided are updated as needed with reanalyzed results and corrections. In addition, a summary of the RMP CEC investigations (past and current) compared against the recommendations of the CEC Science Advisory Panel (Anderson et al. 2012) is contained in Appendix D.

1.3.4 Southern California Bight Regional Monitoring Program

Initiated in 1994 as a pilot study, the Southern California Bight Regional Monitoring Program (Bight) is currently conducted in five-year cycles and has involved over 100 different stakeholder organizations. Management of Bight activities is provided by SCCWRP (http://www.sccwrp.org). The goals of this program are to:

1. Establish regional reference conditions
2. Monitor trends over time
3. Develop new environmental assessment tools
4. Standardize regional data collection approaches
5. Provide a platform to support special studies, including those to prioritize CECs for future monitoring.

The monitoring approach utilizes a stratified random sampling design so that data can be statistically extrapolated to estimate conditions across the Bight. Subsections (strata) are selected to distinguish areas of interest such as the coastal ocean, ports, marinas, the Channel Islands, wastewater treatment plant locations, and land-based runoff locations. Each survey revisits some portion of sites sampled in previous Bight surveys in order to assess trends over the years. The Bight program includes inter-calibration exercises to standardize and improve data quality across participating organizations. An Information Management Committee oversees data structure and reporting requirements, and a centralized database model with a relational database structure was developed to provide easy data access to project scientists.

The current cycle (Bight '13) (http://sccwrp.org/ResearchAreas/RegionalMonitoring/Bight13RegionalMonitoring.aspx) has five components:

1. contaminant impact assessment (offshore sediment condition)
2. nutrient impact (water column condition)
3. microbiology (beach water quality condition)
4. marine protected areas (rocky reef condition)
5. debris assessment

Sampling and laboratory analyses were completed for approximately 400 sites. Hundreds of indicators were measured including sediment chemistry and toxicity; benthic infauna, fish, and invertebrates; contaminant bioaccumulation in bird eggs; trash and debris; physical water column characteristics; nutrients and algae; fecal indicator bacteria; and human pathogens. In 2008, PBDEs and pyrethroids were measured in sediments from at a subset of stations. The Bight Program does not currently target aqueous samples in inland freshwater systems (e.g. Scenario 1) or near marine outfalls (Scenario 3) in the manner specified herein.

The Bight '13 Contaminant Impact Assessment seeks to determine (1) the extent and magnitude of direct impact from sediment contaminants; (2) the trend in extent and magnitude of direct impacts from sediment contaminants; and (3) the indirect risk of sediment contaminants to seabirds. Per the Panel recommendations, new to Bight is the inclusion of PBDEs and PFOS as sediment analytes, and the sampling and analysis of eggs of multiple species of seabirds for contaminants, which includes CECs (PBDEs and PFOS) recommended by the Panel. Also included in the B'13 study are special studies that investigate the application of bioanalytical tools to screen for CECs in extracts of B'13 sediments, and trophic transfer of bioaccumulative compounds, including PBDEs, in the coastal Bight marine food web (B'13 CIA Committee 2013).

1.3.5 Bay Area Stormwater Management Agencies Association (BASMAA)

The Bay Area Stormwater Management Agencies Association (BASMAA) is a consortium of eight San Francisco Bay Area municipal storm water programs (http://www.basmaa.org). In addition, other agencies, such as the California Department of Transportation (Caltrans) and the City and County of San Francisco, participate in some BASMAA activities. Together, BASMAA represents more than 90 agencies, including 79 cities and 6 counties, and the bulk of the watershed immediately surrounding San Francisco Bay.

To comply with NPDES permit requirements for stormwater impacts to water quality, six BASMAA agencies collaborated to form the Regional Monitoring Coalition (RMC) and to develop, design and conduct a large scale monitoring and assessment program for Bay Area watersheds (SCVURPPP 2014). The current RMC work plan described 27 individual projects for FY2009-10 and FY2014-15, which are broken down into several primary topical areas, including Bay and Creek status monitoring; pollutant of concern (POC) loading; long term trends monitoring; and monitoring of emerging pollutants (i.e. CECs). Each of these components utilize a combination of probabilistic and targeted sampling design on selected or model watersheds/waterbodies and a schedule that is optimized for the parameter targeted.

The POC loading study is designed to identify those watersheds draining into the Bay that contribute the majority of mass loading of contaminants. A secondary objective is to determine

the effectiveness of management actions in reducing POC loads to the Bay. The current plan targets three of the CECs recommended by the Panel - PBDEs, fipronil and pyrethroids. Pyrethroids were implicated in toxicity observed in water samples tested using *H. azteca* in this study component (SCVURPPP 2014).

The long term trends monitoring component was integrated into monitoring of creeks performed under SPoT, which measures a number of trace metals and organic chemicals (PAH, organochlorine, pyrethroids and most recently, fipronil) in streams and rivers (see also 1.1.1 SWAMP). The initial projects for CECs will focus on characterization of loading and source identification for endocrine disrupting chemicals, PFCs and nonylphenols and their ethoxylates. In addition, piloting of bioanalytical screening tools consistent with the Panel recommendation is underway. Lastly, the RMC work plan calls for continuing collaboration and coordination with SWRCB efforts to fill data gaps on CECs in Bay receiving waters, e.g. as was recommended by the Panel, and reflected herein.

1.3.6 Southern California Stormwater Monitoring Coalition

The Southern California Stormwater Monitoring Coalition (SMC) was formed in 2001 by cooperative agreement of the Phase I municipal stormwater NPDES lead permittees, the NPDES regulatory agencies in southern California and SCCWRP (http://www.socalsmc.org/AboutUs.aspx). The original 11-member SMC renewed the cooperative agreement for five years commencing June 2008 and added three new member agencies, the California Department of Transportation, the City of Los Angeles and the SWRCB. The current list of SMC members include the stormwater management branches for Los Angeles, Orange, San Diego and Ventura counties, as well as inland empire and city agencies in the region. The SMC also has a cooperative Memorandum of Understanding with USEPA Office of Research and Development to facilitate the development of scientific and technical tools for stormwater program implementation, assessment, and monitoring. The SMC is managed by Steering Committee of its members that meets quarterly to review new projects and assess progress on ongoing projects. Annual reports are available online (http://www.socalsmc.org/Docs).

Despite the success of the SMC, numerous stormwater issues and unresolved problems persist. These remaining challenges, for example, identifying the causative stressor(s) for impacted stream biological communities and the paucity of data on the occurrence of and potential for impact due to CECs, have been especially difficult to address. As part of its 5 year strategic plan, the SMC convened a panel of experts to identify priority issues, which identified CECs as among their top priorities (Schiff et al. 2014). The proposed approach to CECs set forth by the panel was to identify, evaluate and incorporate bioanalytical screening tools to more comprehensively inform the need for more detailed toxicological monitoring. Once the appropriate tools are identified and optimized for stormwater applications, pilot scale evaluation in model MS4 watersheds are planned. The SMC recognizes the implications of SWAMP's CEC efforts (i.e. this pilot study plan), and pledges collaboration with SWAMP and the other monitoring programs described herein (e.g. BASMAA) to best inform SMC's future monitoring strategy for CECs.

1.3.7 Delta Regional Monitoring Program

The Delta Regional Monitoring Program (DRMP) is a new effort to collaboratively assess the water quality of the Sacramento-San Joaquin River Delta ecosystem. The primary agencies

coordinating this regional cooperative are the SWRCB (http://www.swrcb.ca.gov/centralvalley/water_issues/delta_water_quality/comprehensive_monitoring_program/), the Central Valley Regional Board, and SFEI (http://www.sfei.org/programs/delta-regional-monitoring-program). The goal of the DRMP is to better define water quality issues of regional concern and to improve the quality and efficiency of water quality monitoring. Four core management questions have been identified as guiding principles for the DRMP:

1. status and trends
2. sources, pathways and loadings
3. forecasting the impact of management actions on water quality
4. evaluating the effectiveness of management actions

Initial priorities are an improved understanding of the spatial and temporal distribution of prioritized water quality constituents (i.e. methylmercury, nutrients, pathogens, pesticides, and toxicity) in the Delta, improving the efficiency and usefulness of compliance monitoring and data reporting, and fostering large-scale collaborations. Monitoring is expected to begin in 2015.

1.3.8 Other Monitoring Efforts

Pilot and/or special studies on CECs have also been conducted at the regional and local scale in California. Stressor identification in coastal rivers and estuaries along the central California coast have focused on restricted and current use pesticides, including chlorpyrifos, pyrethroids, fungicides and at the current time, neonicotinoid insecticides (Worcester 2011). The Santa Ana Watershed Project Authority (SAWPA) is a collaborative among water agencies and the Santa Ana Regional Board that identifies and addresses water-related issues in the region. The Emerging Constituents Workgroup within SAWPA investigated the occurrence of pharmaceuticals and personal care products in the effluent dominated Santa Ana River watershed (SAWPA 2014). There is currently no known activity or future plans for CEC investigation by SAWPA. In recent years, the Los Angeles Regional Board has commissioned investigations to characterize the occurrence and fate of CECs, including those identified by the Panel, in effluent dominated waterways and their coastal transition zones (i.e. river mouths). These investigations started with water column occurrence (Sengupta et al. 2014) and are currently targeting priority CECs (e.g. PBDEs, PFOS) in sediment and fish tissue. To address recommendations coming out of this effort, the North Coast Regional Board has plans to conduct a CEC pilot study, focused on the contributions and impacts of WWTP and stormwater associated CECs discharged into the Russian River watershed. This study is tentatively scheduled to commence in 2015.

2 TARGETED CEC MONITORING PROGRAM DESIGN

2.1 Revisions and Addendums to Panel Recommendations

Subsequent to the Panel's final report (Anderson et al. 2012), the compilation of occurrence and toxicological data for fipronil, a phenylpyrazole insecticide whose application statewide increased during the period 2000-2010, was updated (**Tables 2.1-1 and -2**). The updated MTQs exceeded unity for the aqueous phase in inland freshwaters and coastal embayments (Scenarios 1 and 2). In addition, the MTQ exceeded unity for freshwater sediments, suggesting the need to monitor fipronil in inland freshwater (Scenario 1) sediments, a matrix that was not included for targeted CEC monitoring by the Panel. Since the parent compound is transformed in aquatic systems to several known metabolites, monitoring of these degradates (fipronil desulfinyl, fipronil sulfide, and fipronil sulfone) is also recommended.

It is also noted that the monitoring of pesticide analytes, i.e. fipronil and degradates, bifenthrin, permethrin (and other pyrethroids) and chlorpyrifos is currently planned for freshwater systems across California via existing SWAMP (SPoT) and DPR programs. The current designs for these programs carried into the initial 3-year pilot monitoring cycle will obviate the need for monitoring of these analytes as defined in Scenario 1 (Section 2.2.1) and MS4 (Section 2.2.4). Recommended monitoring trigger levels (MTLs) and reporting limits (RLs) for these scenarios are included in **Table 2.1-3**.

Table 2.1-1. Ecotoxicological data for fipronil.

	Aqueous Freshwater	Aqueous Saltwater	Sediment Freshwater	Sediment Saltwater
Reference	Weston & Lydy (2014)	USEPA (1996)	Maul et al. (2008)	Chandler et al. (2004a,b)
Organism	Chironomid	Mysids	Chironomid	Amphiascus
LC or EC	33 ng/L	<5 ng/L	0.90 ng/g dw	65 ng/g dw
Safety Factor	10	None	10	10
MTL	3.3 ng/L	5 ng/L	0.090 ng/g dw	6.5 ng/g dw

Table 2.1-2. Monitoring trigger quotients (MTQs) > 1 for fipronil by scenario and matrix. MEC - maximum measured environmental concentration. PEC - maximum predicted environmental concentration. The PECs for embayments (Scenario 2) were calculated assuming a 10-fold dilution factor of MECs representing inland fresh waterways (Scenario 1).

Scenario	Matrix	MEC or PEC	MTQ	Reference
1-Inland Freshwater	Aqueous	10,004 ng/L (MEC)	3000	Gan et al. (2012)
1-Inland Freshwater	Aqueous	2110 ng/L (MEC)	640	Ensminger et al. (2013)
1-Inland Freshwater	Sediment	1.1 ng/g dw (MEC)	12	Lao et al. (2010)
1- Inland Freshwater	Sediment	0.4 ng/g dw (MEC)	4.4	Delgado-Moreno et al. (2011)
2-Embayment	Aqueous	1000 ng/L (PEC)	200	Gan et al. (2012)
2-Embayment	Aqueous	211 ng/L (PEC)	42	Ensminger et al. (2013)

Table 2.1-3. Monitoring trigger levels (MTLs) and reporting limits (RLs) for pesticide analytes recommended for Scenario 1 and MS4 candidate waterways. Recommended RLs are derived from MTLs as reported by the CEC Ecosystems Panel.

Compound	Panel Freshwater MTL[1]	Recommended RL[2]
Aqueous Phase - Scenario 1 and MS4 (ng/L)		
Bifenthrin	0.40	0.20
Permethrin	1.0	0.50
Fipronil	42	21
Chlorpyrifos	5.0	2.5

[1] Monitoring Trigger Level established by CEC Ecosystems Panel (Anderson et al. 2012).
[2] Set at 50% of MTL.

2.1.1 Targeted Contaminants and Reporting Limits

Reporting limits for the target CECs are based on the MTLs recommended by the Panel. A goal of monitoring is to assess if the MTQ is greater than 1 (indicating it should continue to be monitored) or less than 1 (indicating it is not a high priority for future monitoring). Assuming variance in the measurement accuracy (typically 30%), the required reporting levels should extend below the MTL to ensure confidence the MTQ is greater or less than 1. Thus, the required reporting levels are set at ½ the MTL for each scenario and matrix (**Table 2.1.1-1)**. Reporting limits (RLs) for monitoring of WWTP effluent and in MS4 receiving waters are assumed to be the same as for Scenario 1 and 2 receiving waters, respectively.

It is also noted that the RLs for the pesticide analytes, in particular, fipronil and degradates, bifenthrin, permethrin (and other pyrethroids) and chlorpyrifos recommended herein may not be consistent with those reported for SWAMP (SPoT) and DPR programs that currently measure these analytes. In some cases, the RLs recommended herein (i.e. in Table 2.1.1-1) are lower than those currently reported by SWAMP and DPR.

Table 2.1.1-1. Monitoring trigger levels (MTLs) and reporting limits (RLs) by scenario, compound and matrix. Recommended RLs are derived from MTLs as reported by the CEC Ecosystems Panel. Achievable RLs reflect the current state of art for commercial services laboratories. Missing values indicate the achievable value is at or below the recommended RL. Recommended RLs for all CECs in wastewater treatment plant (WWTP) effluent and stormwater (MS4) influenced receiving waters are equivalent to Scenario 1 aqueous phase RLs; additional RLs for compounds that are otherwise measured only in sediment or tissues appear at the bottom of the table.

Compound	Panel Freshwater MTL[1]	Recommended RL[2]	Achievable RL[3]
Aqueous Phase - Effluent dominated inland waterways (Scenario 1) (ng/L)			
Estrone	6.0	3.0	
Ibuprofen	100	50	
Bisphenol A	60	30	
17-beta-estradiol	2.0	1.0	
Galaxolide (HHCB)	700	350	
Diclofenac	100	50	
Triclosan	250	125	
Sediment Phase - Effluent dominated inland waterways (Scenario 1) (ng/g dw)			
Fipronil	0.090	0.045	1.0
Aqueous Phase - Coastal embayments (Scenario 2) (ng/L)			
Bisphenol A	6.0	3.0	
Bifenthrin	0.040	0.020	0.2
Permethrin	0.10	0.050	0.5
Fipronil	5.0	2.5	
Chlorpyrifos	1.0	0.50	
Estrone	0.60	0.30	2.0
17-beta-estradiol	0.20	0.10	0.4
Galaxolide (HHCB)	70	35	
Sediment - Coastal embayments (Scenario 2) (ng/g dw)			
Bifenthrin	0.052	0.026	0.20
PBDE-47	0.030	0.015	
PBDE-99	0.030	0.015	
Permethrin	0.073	0.036	0.40
Fipronil	6.5	3.25	
PFOS[4]	NA	0.1	

Table 2.1.1-1 (cont.)

Compound	Panel Freshwater MTL[1]	Recommended RL[2]	Achievable RL[3]
Sediment - Ocean discharge (Scenario 3) (ng/g dw)			
Bis(2-ethylhexyl) phthalate (BEHP)	130	65	
p-nonylphenol	14	7.0	
PBDE-47	0.30	0.15	
PBDE-99	0.30	0.15	
Butylbenzyl phthalate (BBP)	6.3	3.15	
PFOS[4]	NA	0.1	
Tissues (All Scenarios) (ng/g dw)			
PBDE-47	28.9	14.5	
PBDE-99	28.9	14.5	
PFOS	1000	500	
WWTP Effluent and MS4 Receiving Water (ng/L) [5]			
Bis(2-ethylhexyl) phthalate (BEHP)			3.0
Butylbenzyl phthalate (BBP)			3.0
p-nonylphenol			22[6]
PBDE-47			0.10
PBDE-99			0.10
PFOS			1.0

[1] Monitoring Trigger Level established by CEC Ecosystems Panel (Anderson et al. 2012).
[2] Set at 50% of MTL.
[3] Minimum RL reported by commercial services laboratories. Missing values indicate the achievable value is at or below the recommended RL.
[4] PFOS was recommended for Scenario 2 and 3 sediment monitoring to obtain information on sediment-biota transfer, not based on MTLs. The recommended RL was based on typical values observed in the literature and attainable values by laboratories.
[5] RLs for analytes otherwise measured in sediment or tissues only (no MTL values available). For all other analytes, RLs for WWTP Effluent and MS4 receiving water samples are the same as the aqueous RLs for Scenario 1.
[6] Estimated from the sediment RL (7.0 ng/g), an estimated sediment-water partitioning coefficient, and assuming 1% organic carbon content of the sediment.

2.2 Design Requirements by Scenario

2.2.1 WWTP Effluent Dominated Inland Freshwater (Scenario 1)

Scenario 1 examines inland freshwater systems including rivers and lakes where the majority of the flow or volume during the dry season is WWTP effluent. Treated wastewater is expected to be the largest source of most CECs during this time period.

Monitoring Questions

1. Which CECs are detected in freshwaters and depositional stream sediments, and in which large California watersheds are they detected?
2. Can the CECs be shown to originate from the inland WWTP, or are they present at background concentrations?
3. How quickly (i.e., at what distance) do the CECs attenuate once discharged?
4. What are the concentrations and loadings of target CECs in the dry vs. wet seasons?
5. Do the new occurrence data change the estimated MTQs?

Design Considerations

The effluent of selected inland WWTPs and their corresponding waterways will be monitored. To determine the occurrence and attenuation of target CECs downstream of each identified WWTP (or series of upstream WWTPs), a minimum of 7 stations will be monitored: one station just downstream of the WWTP discharge location(s), five stations further downstream of the WWTP(s), and one background station located upstream of the WWTP(s) (**Figure 2.2.1-1**). To assess repeatability, duplicate field samples each will be collected at the WWTP and background stations. Both the wet and dry seasons will be monitored over a 3 year period (**Table 2.2.1-1**). For fipronil, annual sediment analysis at three stations (e.g., #1, #5, and background) during the dry season is also recommended based on Scenario 1 sediment MTQs > 1 (**Table 2.2.1-2**).

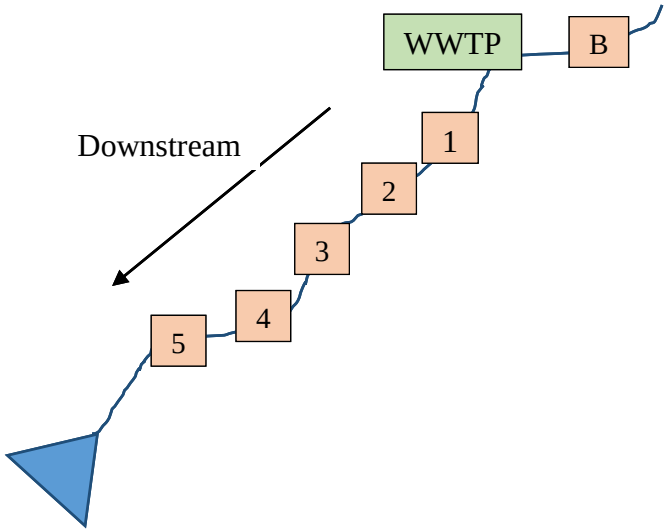


Figure 2.2.1-1. Design schematic for monitoring of CECs in Scenario 1.

Ideal candidates for this pilot study are waterways with well-characterized source and flow inputs. Examples of waterbodies that represent Scenario 1 in southern California are the Los Angeles, Santa Clara, San Gabriel, Santa Ana, and San Diego Rivers. The Los Angeles River and the Santa Clara River are proposed as candidates in southern California. In the Delta and Central Valley, proposed candidates are Alamo Creek downstream of the Vacaville Easterly WWTP and Pleasant Grove and Dry Creeks downstream of the City of Roseville Pleasant Grove and Dry Creek WWTPs, see map in **Appendix B**. No similar waterways have been identified in the San Francisco Bay region.

Table 2.2.1-1. Aqueous sampling frequency for Scenario 1.

Source	Receiving Water	Years	Waterways	Total Samples
WWTP effluent 1 station Wet and dry season 2 replicates Samples = 4/yr	Downstream 5 stations Wet and dry season Samples = 10/yr Background 1 station Wet and dry season 2 replicates Samples = 4/year 14 total samples/yr	3	4 (two each in SoCal and Delta/CV)	Effluent = 48 FW = 168

Table 2.2.1-2. Sediment sampling frequency for Scenario 1.

Waterway Sediment	Years	Waterways	Total Samples
3 stations Dry season Samples = 3/yr	3	4 (two each in SoCal and Delta/CV)	Sediment = 36

2.2.2 Coastal Embayment (Scenario 2)

Scenario 2 examines coastal embayments that receive CEC inputs at the land-ocean interface, which may originate from upstream WWTP discharge, direct WWTP discharge into the embayment, or stormwater runoff. As San Francisco Bay is by far the largest and most actively monitored coastal embayment in California, this scenario is based on monitoring in San Francisco Bay but may be extended to other coastal embayments across the State.

Monitoring Questions

1. Which CECs are detected in coastal embayment water and sediments?
2. Do CECs originate from the outfalls, or are embayment concentrations due to stormwater and other inputs?
3. Is there a sub-annual change in CECs discharged from WWTPs?
4. Do the new occurrence data change the estimated MTQs?

Design Considerations

The Panel's recommendation for Scenario 2 was a 2-D gradient (up to 6 stations) at each of five WWTPs within San Francisco Bay ("Bay"). Each station would consist of a sediment sample

and an overlying aqueous phase sample, since target compounds for this scenario may occur in both matrices. Monitoring was to be semi-annual over three years. The 2-D gradient design was recommended to measure spatial attenuation of the target contaminants.

Within the Bay, the Lower South Bay is most strongly impacted by effluent discharge due to its high population and correspondingly high WWTP discharges and lower oceanic dilution. This section of the Bay is the focus of Scenario 2 monitoring. Due to the multiple WWTP discharges with relatively close outfalls, tidal influences, and multi-directional currents that rapidly distribute contaminants throughout the Lower South Bay, however, the Panel's recommended design will likely not successfully measure stepwise decreases in contaminant concentration (attenuation) moving away from the zone of initial dilution (ZID) of a given outfall.

Instead, it is recommended that paired sediment/aqueous samples be collected at stations along the interior waters (aka the "spine") from the Lower South Bay to the Central Bay (n = 15 stations) (**Table 2.2.2-1**). This design will integrate influences from multiple WWTPs and will account for mixing. Sampling should take place during the dry season, when dilution from runoff is lowest, and concentrations can be expected to be at their highest. Paired effluent (n = 1) and ZID samples (n = 1 each for sediment and aqueous phase) from at least 5 major WWTPs in the South Bay should also be monitored, to characterize which contaminants, if any, originate from the outfall (**Table 2.2.2-2**). Sediment and receiving water sampling along the spine should occur annually over 3 years. Effluent and aqueous ZID sampling should be performed semi-annually (wet/dry season) over 3 years, and sediment ZID sampling annually over 3 years. Current RMP special studies will inform the selection of WWTPs, and effluent data for the target CEC should be provided.

The design guidance for interior waters can be applied to other coastal embayments across the state. The design guidance for WWTP effluent and ZID could be applied, with modification as necessary, to investigate the occurrence of CECs in the proximity of known or suspected sources of CECs or "hot spots", e.g. urban river mouths or industrial complexes.

Table 2.2.2-1. Aqueous and sediment sampling frequency for interior waters (Scenario 2).

Aqueous	Sediment	Years	Total Samples
15 stations Dry season Samples = 15/yr	15 stations Dry season Samples = 15/yr	3	Aqueous = 45 Sediment = 45

Table 2.2.2-2. WWTP effluent and ZID sampling frequency for Scenario 2.

Effluent	ZID Aqueous	ZID Sediment	Years	Total Samples
5 WWTPs Wet/Dry season Samples = 10/yr	5 aqueous Wet/Dry season Samples = 10/yr	5 sediment Dry season Samples = 5/yr	3	Effluent = 30 ZID Aqueous = 30 ZID Sediment = 15

2.2.3 WWTP Effluent Discharge to the Ocean (Scenario 3)

Scenario 3 examines WWTP effluent discharged by outfalls at mid-Continental Shelf depths (50-100 m). Discharged CECs are diluted by the ambient water, transformed into breakdown products and/or are transported away from the outfall by currents. This scenario is monitored exclusively at marine outfalls within the southern California Bight.

Monitoring Questions

1. Which CECs are detected in marine waters and sediments adjacent to WWTP outfalls, what are their concentrations, and how quickly do they attenuate?
2. Can the CECs be shown to originate from the outfalls, or are they present at background concentrations?
3. Is there a sub-annual change in discharged CECs?
4. Does the new occurrence data change the estimated MTQs?
5. What is the relative contribution of CECs in WWTP effluent vs. stormwater? (see also Section 2.2.4)

Design Considerations

The effluent and sediments at a minimum of two WWTP ocean outfalls will be monitored, with a grid of 8 sediment stations at each outfall (**Figure 2.2.3-1**). Observations of a stepwise decrease in concentrations away from the ZID verify the compounds originate from the outfall and are not at background concentrations due to other inputs. The exact locations will consider the oceanic conditions and historic depositional patterns at each candidate outfall and may be changed based on the results of initial monitoring. Three stations will be located down current from the zone of initial dilution (ZID), three will be located cross current, and one background station will be located up current of the outfall. The frequency of analysis is semi-annual (wet and dry) for the effluent and annual for the sediment (**Table 2.2.3-1**). Exact station locations may be assigned based on the results from the Bight '13 Special Study described in **Appendix C**.

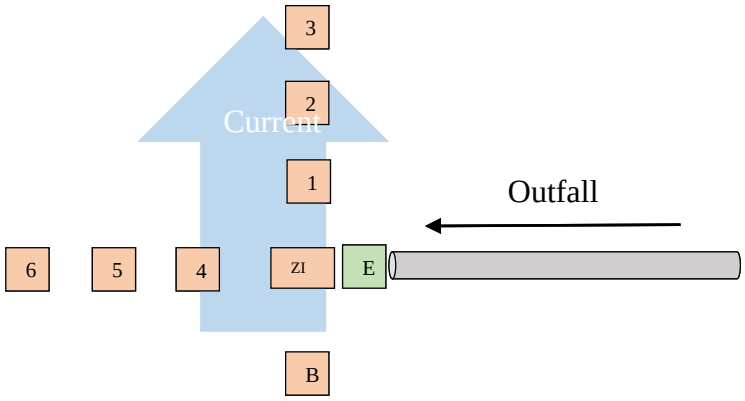


Figure 2.2.3-1. Design schematic for sampling of CECs in Scenario 3.

Table 2.2.3-1. Effluent and sediment sampling frequency for Scenario 3.

Source	Sediment	Years	WWTPs	Total Samples
WWTP effluent 1 station Wet and dry seasons 2 replicates Samples = 4/yr	Grid 7 stations Samples = 7/yr Background 1 station 2 replicates Samples = 2/yr 9 total samples/yr	3	2	Effluent = 24 Sediment = 54

2.2.4 Stormwater Discharge to Receiving Waters (MS4)

Unlike WWTP effluent, the vast majority of annual stormwater runoff and discharge occurs during the wet season (November through April) in all but the most arid regions of the State. Materials from various sources/surfaces (e.g. road dust, topsoil, sediments) are mobilized during wet weather events, transporting suspended particulates and associated contaminants, including some CECs, into receiving waters. Thus, annual loading (on a mass per year basis) of CECs into receiving waters is expected to be highly seasonal. Receiving water impacts resulting from such loading can be direct, e.g. release of pesticide residues from sediments transported into receiving waters resulting in invertebrate or fish toxicity, or indirect, e.g. bioaccumulation of sediment-associated CECs (e.g. PBDEs) by benthic organisms and subsequent trophic transfer into higher biota (e.g. fish and humans). During the dry season, in contrast, incidental runoff (e.g. due to excess irrigation of gardens and/or parks) may contain CECs (e.g. pesticides) at higher concentrations, since runoff volume and base flow to the receiving water are relatively small. Moreover, particulate loading is typically negligible under these conditions, directing attention to dissolved, aqueous phase (i.e. more water soluble) CECs. Thus, it is critical to address both short term toxicity and long term loading, as well as to take into account the distribution and fate of CECs for monitoring in MS4 watersheds.

Monitoring Questions

1. Which CECs are detected in waterways dominated by stormwater?
2. What are their concentrations and loadings in the dry vs. wet seasons?
3. What is the relative contribution of CECs in WWTP effluent vs. stormwater?
4. What is the spatial and temporal variability in loadings and concentrations (e.g. between storm variability during the wet season; in stream attenuation rate during low flow, dry season conditions)?

Design Considerations

Wet Weather. Since annual loading is the main concern during wet weather, a design that focuses on detection of target CECs, and estimating total loads for those detected into MS4 receiving waters are the primary goals. Current wet weather monitoring conducted by some programs relies on sampling at fixed mass emission (FME) or integrator stations located at the bottom of MS4 permitted watersheds. Integrator stations identified and monitored in other monitoring programs (e.g. RMC, SMC, SPoT, DPR) should be utilized for the candidate watersheds. Flow-weighted or time-interval sampling at FME stations for two storms per year per watershed will provide data to address monitoring questions 1-3 (**Table 2.2.4-1**). Ideally, the

storms sampled will include an early ("first flush") and late season event. A minimum of three watersheds statewide should be assessed over a 3-year pilot study period. Addressing question 4 will necessitate more intensive sampling during and/or between storm events, and, if warranted based on the results of the initial 3 year screening, should be planned during subsequent pilot study cycles. Non-filtered, whole water samples should be analyzed when addressing loading and for effects/toxicity evaluation. Sufficient sample size and analytical methods should be specified to meet target detectability of CECs (see also Sections 2.1.1 and 10 Supplemental Guidance for QA/QC).

Dry Weather. Since short term maximum concentrations resulting in acute toxicity is the main concern, a strategy that focuses on capturing worst case exposure conditions for a relevant endpoint/receptor of interest is the primary goal. A design that targets receiving water near known or suspected incidental runoff sources, e.g. culverts or sections that drain parks or golf courses, is needed to include worst case exposure scenarios. Depositional area sediments (river mouths, oxbows, retention basins) should be sampled at the start and end of the dry season to examine (1) what has been washed in during the previous wet season and (2) degree of attenuation occurring during the dry season (**Table 2.2.4-1**). Unless unexpectedly high total suspended solids (TSS) samples are encountered, non-filtered aqueous samples should be sufficient for monitoring and assessment during dry weather. To address chronic exposure of CECs, base flow conditions over longer time periods (weeks to months) can be assessed using emerging technology, e.g. passive sampling methods (PSMs) that provide a time-average concentration of CECs that have been pre-calibrated in the laboratory (see also Section 5). Such extracts are also amenable, without fortification, for toxicity screening.

Coordination with Special Studies

Samples collected for targeted chemistry will also be evaluated for toxicity parameters as specified in Section 3. Bioanalytical screening assays will be adapted and evaluated on organic extracts of water and sediment samples collected as part of this scenario. Targeted CEC monitoring that require RLs not readily achievable using conventional or commercially available methodology shall utilize PSMs, where such technology has been validated and is amenable for deployment (e.g. conditions and timing for continuous submerged conditions are available).

Candidate Watersheds

- San Francisco Bay: watersheds monitored by the RMC, SWAMP/SPoT and DPR, including Coyote Creek and the Guadalupe River (Santa Clara County) [1,3,4]; Grayson Creek (Contra Costa County)[4]; Arroyo de la Laguna (Alameda County) [4]
- Delta/Central Valley: watersheds monitored by the DRMP, SWAMP/SPoT and DPR, including Arcade Creek[4], Steelhead Creek, Morrison Creek, American River[3] and the Sacramento River at the Hood integration site[3] (Sacramento County); Pleasant Grove Creek (Placer County) [4] ; see map in **Appendix B**.
- Southern California: watersheds monitored by the SMC, SWAMP/SPoT and DPR, including Ballona Creek[2,3,4] and Bouquet Canyon Creek[3,4] (Los Angeles County); San Diego Creek[2,3] and Salt Creek[4] (Orange County); Chollas Creek[4] and San Diego River[2,3,4] (San Diego County).

[1] scheduled for monitoring by RMC (SCVURPPP 2014)
[2] scheduled for monitoring by SMC (SMC/BWG 2007)
[3] scheduled for monitoring of toxicity stressors by SPoT (Phillips et al. 2014)
[4] scheduled for monitoring of pesticides by DPR in 2014-15 (Emsinger 2014)

Table 2.2.4-1. Sampling matrix for MS4 watersheds. Monitoring of a minimum of 3 watersheds over a 3 year period is recommended.

Parameter	Sample Type	Stations	Frequency	Replication	Total Samples
Aqueous concentration, wet weather	Whole water (unfiltered)	1 (FME)	2 storms/yr	3	54
Aqueous concentration, dry weather	Whole water (unfiltered)	3 (source-related)	1/yr	1	27
Sediment concentration, dry weather	Whole (sieved) sediment	3 (depositional)	twice/yr	1	54

2.2.5 Tissue Monitoring

Wildlife living in receiving waters can be exposed to CECs by direct uptake via the aqueous phase and through ingestion of contaminated prey. Chemicals that are hydrophobic (log K_{ow} >3), remain un-ionized in either freshwater or saltwater environments, and that are persistent have the potential to bioaccumulate in aquatic biota. For CECs that biomagnify (e.g. PBDEs), an organism with a sub-critical body burden that comprises the majority of the diet of a higher level trophic receptor may pose an unacceptable risk to the predator organism if CEC concentrations exceed the predator-based critical body residue concentration.

While several of the CECs considered by the Panel have the potential to bioaccumulate, only two (PBDE and PFOS) have NOECs from which body burden-based MTLs could be derived. The Panel used studies on birds (adult Mallard and Bobwhite Quail) to set a PNEC of 1000 μg/kg for PFOS, and studies on the American Kestrel to set a NOEC of 289 μg/kg for the two PBDE congeners (47 and 99). The Panel was not able to identify allowable concentrations of PBDEs in fish for protection of marine mammals. The Panel believes such marine mammal-based MTLs could be derived in the future.

Monitoring Questions

1. What are the concentrations in tissues and do they exceed toxicity thresholds?
2. Do the new occurrence data change the recommendation to monitor?
3. Are concentrations of bioaccumulative CECs changing over time (annual to decadal time frames)?
4. Do bioaccumulative CECs occur in scenario-specific patterns?

Design Considerations

Toxicity Thresholds Based on Bird Eggs. Addressing changes in the MTQs requires analysis of bird eggs, since the thresholds for both PBDEs and PFOS were set using this matrix. Both the RMP and Bight programs are currently collecting these data. Since 2006, RMP has monitored bird eggs for PBDEs and PFCs every 3 years, addressing the temporal trend question. Bight is performing bird egg measurements on PBDEs and PFOS for the first time in 2014. Therefore,

data from the RMP and Bight programs may be used to re-assess tissue MTQs. Recommended species (where permitted) are the double-crested cormorant, western gull, and California, Caspian or Forster's least terns. Within the regional programs, we recommend bird egg temporal monitoring to continue in the future, particularly in key urban areas such as covered by the RMP and Bight. To our knowledge, bird egg monitoring does not currently occur in the Delta/Central Valley region, and is therefore recommended. A sample size of n = 10 egg composites for a single bird sentinel species is recommended over the 3-year pilot study cycle (**Table 2.2.5-1**). If the recommended target species listed above are not feasible for the Delta/Central Valley, alternate species as recommended by the DRMP or the Central Valley Regional Board can be substituted.

Marine Mammals. Marine mammals such as pinnipeds and cetaceans occupy high trophic positions and thus can have relatively high concentrations of bioaccumulative CECs (e.g. PBDEs). The Panel was unable to establish MTLs for marine mammals, but recognized the potential for risk associated with biomagnification and discussed possible future methods for determining marine mammal MTLs. Therefore, collection of occurrence data in marine mammals is warranted. Live-capture harbor seal blubber was measured for PBDEs in 2014 as part of a RMP special study, and PFCs will be measured in the blood. Although some specific studies have been carried out, contaminants in marine mammals are not routinely monitored in southern California, e.g., within the Bight program. It is recommended that southern California sea lions and/or bottlenose dolphins be measured for PBDEs (blubber) and PFOS (blood). A minimum sample size of n = 10 for each matrix (blood and blubber) that can be a composite total for both species, or of a single species, is recommended over the 3-year pilot study cycle (**Table 2.2.5-1**). As data exist for PBDEs in these two species, comparisons to current and future conditions can be made to obtain temporal trends (Meng et al. 2009; NOAA, unpublished). Live biopsies are recommended to obtain fresh tissue representative of a healthy population, however fresh dead strandings could be considered in the absence of access to tissues from live biopsies.

Fish and Bivalves. Compared with birds and marine mammals, some fish and all bivalves are more abundant and have higher site fidelity. These sentinels are therefore well suited to compare contaminants across scenarios, to assess temporal trends, to characterize exposure and to identify localized contamination sources. Bivalves in particular are sessile and there are substantial historical bivalve tissue data for comparison (Dodder et al. 2014; Klosterhaus et al. 2013; Sutton et al. 2014). However, these filter-feeding organisms indicate exposure to waterborne CECs, as opposed to bioaccumulation and/or biomagnification potential. For example, PFCs (including PFOS) were sporadically detected at low levels in California coastal mussels (*Mytilus* spp.) (Dodder et al. 2014), in direct contrast to elevated PFC concentrations in bird eggs (Sedlak and Greig 2012). Fish, on the other hand, occupy a higher trophic position and may have higher body burdens of target CECs. Therefore, monitoring of both bivalves (for PBDEs) and fish (for PBDEs and PFOS) is recommended. Sampling of fish and bivalves is recommended annually over the 3 year pilot study cycle (**Table 2.2.5-2**).

Candidate fish species will vary in availability by location. Species that exhibit high spatial fidelity and are suspected to accumulate relatively high levels of PBDEs and PFOS should be selected for monitoring. Candidate bivalve species are *Corbicula fluminea* (freshwater) and *Mytilus spp. (californianus or galloprovicialis)* for embayment and marine habitats. Fish may be individuals (provided enough sample mass is available) or composites, and bivalves should be composites. Only specimens of the same species should be composited together. Whole bodies

for small fish, and filets of larger fish should be analyzed. The final selection of sentinel species shall be made in coordination with SWAMP/BOG.

- For freshwater systems (e.g. Scenario 1 and MS4 monitoring), it is recommended that fish (PBDEs and PFOS) and bivalves (PBDEs) be sampled in one system each in the San Francisco Bay watershed, southern California and the Delta/Central Valley region. The selection of these systems can coincide with those identified for sediment and aqueous phase monitoring in Sections 2.2.1 and 2.2.4. Based on historical sampling and results from SWAMP/BOG, recommended fish species for freshwater systems are large and smallmouth bass, Sacramento or Santa Ana sucker, and channel catfish.
    - For Scenario 1, bivalves and fish should be collected from a location in close proximity to the WWTP outfall, during the period of highest effluent loading.
    - For MS4 watersheds, bivalves and fish should be in close proximity to FME/integrator stations (i.e. near the mouth of the watershed), where loadings are expected to be highest, during or near the end of the wet season.
- For San Francisco Bay (Scenario 2), the RMP measures PBDEs in bivalves every 2 years, and PBDEs and PFCs in sport fish every 5 years. Forage fish are not part of RMP Status and Trends monitoring. Therefore, embayment tissue monitoring can be carried out through RMP. Recommended fish species are shiner surfperch, white croaker, topsmelt, and California halibut.
- For marine outfall tissue monitoring (Scenario 3), it is recommended that fish be monitored for PBDEs and PFOS at two outfalls that are also monitored for sediment concentrations (n = 10 fish, each outfall). Species that have high site fidelity should be selected. The Bight program does not currently monitor fish for PBDEs and PFOS, therefore sampling is recommended annually over the 3 year pilot study cycle (Table 2.2.5-2). Recommended species include those collected in abundance historically at these outfalls, e.g. hornyhead turbot, Dover sole and scorpionfish.

Table 2.2.5-1. Recommended sampling of bird eggs and marine mammals for the 3-year pilot study cycle. Additional tissue samples are to be analyzed through regional programs, as noted in the text.

Sample	Region	Number per 3 yr cycle	Total Samples
Bird eggs	Delta/Central Valley	10 egg composites	10
Marine Mammals Blubber (PBDEs) Blood (PFOS)	Southern California Bight	5 sea lion 5 bottlenose dolphin	Blubber = 10 Blood = 10

Table 2.2.5-2. Fish and bivalve sampling frequency. Additional tissue samples are to be analyzed through regional programs, as noted in the text.

Sample	Scenario	Number per year	Locations	Years	Total Samples
Freshwater fish	Scenario 1 and MS4	5	3 Waterways ea. scenario	3	90
Marine fish	Scenario 3	5	2 WWTP outfalls	3	30
Bivalves	Scenario 1 and MS4	3	3 waterways ea. scenario	3	54

Non-Targeted Analysis**.** Targeted analytical methods will be used to quantify the Panel-recommended CECs. However, these methods are not designed to screen for new or unexpected contaminants; i.e., unknown CECs. The Panel recognized non-targeted analytical methods as of potential utility in periodically screening for unexpected contaminants, and in addition, as tool for toxicity identification evaluation (TIE) when responses and/or effects observed with in vitro, in vivo testing and/or in situ monitoring cannot be explained by targeted analytical chemistry. Non-targeted methods have recently been developed for analysis of bioaccumulative organic compounds in marine biota from the California coast (Hoh et al. 2012; Shaul et al. 2014). Application of non-targeted analysis to the tissue samples collected as part of this pilot study (this section) will establish baseline contaminant inventories and identify any high abundance compounds missed by targeted monitoring. In addition, the mass spectral libraries and retention time information generated by such periodic monitoring will allow for efficient identification of the contaminants in the future. Directly linking non-targeted mass spectrometry and in-vitro bioassays to identify contaminants contributing to the biological response is discussed as a research need in Section 5.2. (**Table 2.2.5-3**)

Table 2.2.5-3. Recommended non-targeted analysis of tissue samples collected for monitoring of PBDEs and PFOS.

Sample	Scenario/Region	Number per 3 yr cycle	Locations	Total Samples
Freshwater Fish	Scenario 1 and MS4	2	3 waterways ea. scenario	12
Marine mammal blubber	Scenario 2 (San Francisco Bay)	10	n/a	10
Marine fish	Scenario 3	5	2 WWTP outfalls	10
Marine mammal blubber (2 species)	Southern California Bight	5	n/a	10

3 SPECIAL STUDIES DESIGN REQUIREMENTS

3.1 Introduction

The Panel recommended that a number of special studies be conducted as part of a statewide CEC pilot monitoring program in order to evaluate and where possible, validate the methods evaluated in these studies prior to full implementation (Table 8.1-3). These studies largely address the potential for adverse effects of CECs in aquatic organisms (e.g. animal toxicity; microbial resistance) and will complement traditional targeted chemical monitoring (described in Section 2) by providing additional information on the occurrence of known and unknown CECs (e.g. bioanalytical screening assays).

Moreover, the special study bioassay components target and/or link the responses across increasingly complex levels of biological organization, and thus can be integrated in a multi-tiered interpretive framework (**Figure 3.1-1**). In Tier I, high-throughput *in vitro* bioassays (IVBs) are conducted to screen for the occurrence of chemicals, including CECs, in environmental samples based on their mode of action (MOA). *In vitro* assays are an efficient way to assess the ability of CECs to activate cellular receptors but stop short of predicting adverse outcomes at the organismal or population level. The Panel also recommended whole organism toxicity testing to determine if CECs present in aquatic ecosystems can have adverse effects at the organism level (Tier II), e.g. impaired reproduction in fish exposed to model chemicals, receiving water samples and/or WWTP effluent. In the case that samples of interest demonstrate effects in Tier II analyses that warrant further investigation, Tier III analyses focus on in situ evaluation, e.g. field collection of biological samples of sentinel organisms (e.g. invertebrates, fish, birds and/or mammals), specifically to investigate whether such MOAs identified using Tier 1 *in vitro* cell assays and adverse outcomes indicated by Tier II analyses are prevalent in the receiving water environment. Tier III tools/endpoints would incorporate both advanced molecular tools such as quantitative polymerase chain reaction (qPCR) or gene microarrays as well as more conventional in situ biomonitoring and assessment parameters (e.g. histology, species abundance/diversity).

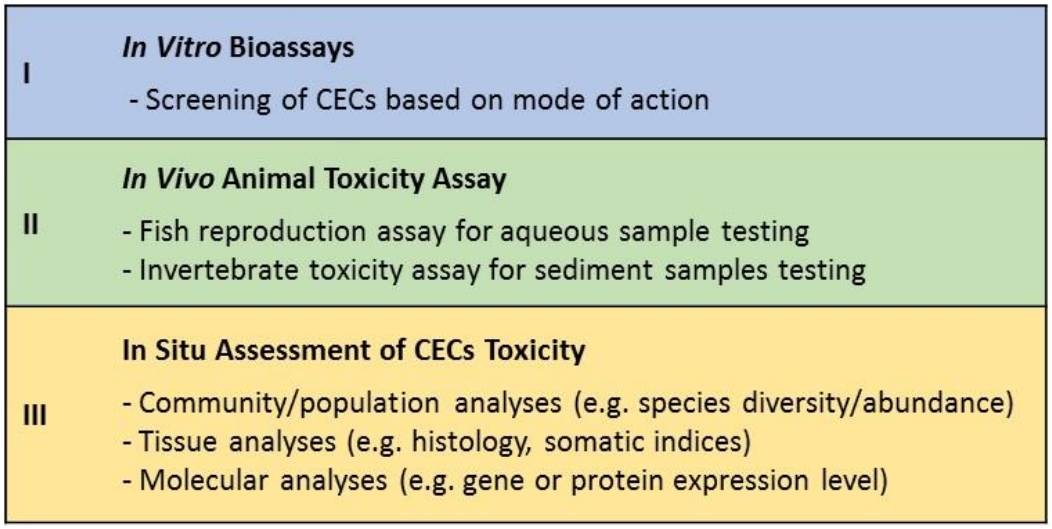


Figure 3.1-1. Proposed framework for biological assessment of CECs in aquatic ecosystems.

3.2 Tier I – Bioanalytical Screening Using High-Throughput *In Vitro* Assays

In vitro bioassays can be used to screen a large number of chemicals based on a MOA paradigm. Selected IVBs are currently being evaluated for screening of recycled and drinking water quality (Leusch et al. 2010; Escher et al. 2014), with encouraging results for the detection of endocrine disrupting CECs. To address the Panel's recommendations, a number of commercially available IVBs are proposed to assess the capability of environmental CECs to activate endocrine-related receptors, induce xenobiotic metabolism and cause cell damage (**Table 3.2-1**). Some chemicals are also known to suppress the activity of endocrine-related receptors causing adverse effects. For example, male fish exposed to anti-androgenic compounds or females exposed to anti-estrogenic compounds can cause reproductive impairment via alteration of plasma sex steroids levels and subsequent reduction in fertility and fecundity (Panter et al. 2004; Filby et al. 2007). To screen for these outcomes, estrogen receptor (ER) and androgen receptor (AR) assays will be conducted in agonist (receptor activation) as well as antagonist (inhibition of activity) mode.

Table 3.2-1. In vitro bioassays that screen for endocrine disruption, xenobiotic metabolism and general cell toxicity. ***Table adapted from Anderson et al. (2012).***

Endpoint	Response	Mode of Action	Potential Adverse Outcome
Estrogen Receptor Alpha (ERa)	Activation and inhibition	Estrogen signaling	Feminization of males. Impaired reproduction, cancer
Androgen Receptor (AR)	Activation and inhibition	Male sexual phenotype	Androgen insensitivity, masculinization of females, impaired reproduction
Glucocorticoid Receptor (GR)	Activation	Cortisol binding, regulation of gene transcription	Development, immune diseases, diabetes
Progesterone Receptor (PR)	Activation	Embryonic development, cell differentiation	Cancer, diabetes, hormone resistance syndrome
Aryl Hydrocarbon Receptor (AhR)	Activation	CYP1A metabolism induction	No known adverse outcome. Indicates exposure to dioxin-like chemicals
Cytotoxicity	-	General cell toxicity	Tissue damage, death

Two types of investigations are recommended. First, a battery of candidate IVBs will be evaluated to determine their response to the list of Panel recommended CECs at exposure concentrations of monitoring relevance (see Section 2). Second, the IVBs will be evaluated to determine the magnitude and range of response associated with real environmental samples and to assess the concordance with responses predicted using targeted analytical chemistry results. Because the output parameters resulting from bioassays are not directly comparable with individual chemical concentrations, translation of bioassay into equivalent concentrations, or bioassay equivalents (BEQs), is necessary (**Table 3.2-2**).

Table 3.2-2. Output parameters of *in vitro* assays.

	Parameter
Calibration	Dose response curve with reference toxicant
Concentration effect assessment	Relative Enrichment Factor (REF) (enrichment factor of extraction process and dilution of extract in the IVB)
Data analyses	Effect concentration (EC)
Output parameter	Bioassay equivalent concentration (BEQ)

3.2.1 *In Vitro* Screening of Targeted CECs

Questions to be addressed:

1. Which priority CECs are detectable at or below their respective monitoring trigger levels (MTLs) using the endocrine-related cell assays?
2. Which priority CECs are detectable at or below their respective MTLs using other relevant endpoints (e.g. AhR)?
3. What are the responses (additive or antagonist) of priority CECs mixtures using the selected cell assays?

Seventeen CECs (see Table 8.1-1) have been selected for target monitoring in water, sediment and/or tissue. The objective of this study is to identify the most robust cell assays to screen for priority CECs at environmentally relevant levels (**Table 3.2-3**). For each chemical, four concentrations will be selected including the lowest at or below its MTL (see Table 2.1.1-1). A mixture of the selected CECs will also be tested with individual concentrations at and above MTLs to determine if additive or antagonist effects may occur.

Table 3.2-3. *In vitro* assays for screening of priority CECs.

Endpoint	Priority CECs	Other environmental chemicals
ERa	BEHP and BBP[1] , galaxolide (Anti-ER)[2] , PFOS[3] 17-beta estradiol – known strong ER agonist Estrone – known moderate ER agonist BPA, nonylphenol – known weak ER agonists	Musks
AR	Galaxolide (Anti-AR)[2] No AR activation data for priority CECs of interest	
AhR	PBDE-47 and -99, chlorpyrifos[4]	PAHs, PCBs
GR	No GR activation data found for CECs of interest	Glucocorticoid steroids
PR	No PR activation data found for CECs of interest	Progestins (e.g. levonorgestrel)

[1]Harris et al. (1997), [2]Schreurs et al. (2005), [3]Kjeldsen and Bonefeld-Jorgensen (2013), [4]Long et al. (2003).

3.2.2 *In Vitro* Screening of Environmental Extracts

Questions to be addressed:

- How efficient are the candidate *in vitro* bioassays in detecting known and unknown CECs present in complex environmental mixtures (e.g. WWTP effluent and receiving water)?
- How do cell assay responses correlate with analytical chemistry data?

Aqueous environmental samples contain complex mixtures of CECs. *In vitro* screening assays can complement targeted chemistry and provide additional information on the chemicals present in these mixtures by integrating the response of all bioactive chemicals – both known and unknown - present in a water sample. Thus, it is important to evaluate the correlation between in vitro assay responses and chemistry data to understand the contribution of known (i.e. measurable) CECs. This pilot study will be conducted over a three-year period. Water samples will be collected, extracted and split on an annual schedule for targeted monitoring (see Section 2) and testing using the IVBs (**Table 3.2-4**). Prior to *in vitro* screening, the extracts will be solvent exchanged to dimethylsulfoxide (DMSO). Screening of sample extracts for cytotoxicity is performed prior to screening of the remaining candidate endpoints (or MOAs) (**Fig. 3.2-2**).

Table 3.2-4. Sampling locations and frequency for *in vitro* screening

	Sample Type	Location	Sampling Frequency	Waterways
Scenario 1 Freshwater	WWTP effluent	Outfall	2/year (wet & dry season)	2
	River water	Stations # B, 1, 3 and 5 (Section 2.2.1)	2/year (wet & dry season)	
Scenario 2 Embayment	WWTP effluent	Outfall	1/year	1
	Receiving water	Every third station for interior waters (Section 2.2.2)	1/year	
Scenario 3 Ocean	WWTP effluent	Outfall	1/year	3
	Receiving water	Stations # B, ZID, 3 and 6 (Section 2.2.3)	1/year	
Scenario 4 MS4	Watershed	1 FME	2 storms/year	3
		3 source-related (Section 2.2.4)	dry weather 1/year	

3.2.3 *In Vitro* Assay Parameters and Optimized Methods

A number of commercially available cell assays have been identified for screening CECs in environmental samples. Among those, the GeneBLAzer assays (Life Technologies) and the CALUX assays (BioDetection Systems) have shown promising results. It should be noted, however, that differences in operating procedures exist among the endpoints and manufacturers. Based on the performance of these assays in screening of potable and surface water samples (Escher et al. 2014), the minimum requirements for reference chemicals and enrichment (i.e. pre-

concentration) of aqueous samples relative to their collecting sample volume (denoted as REF) are provided in **Table 3.2-5**. Key cell bioassay conditions and QA/QC requirements are summarized in **Table 3.2-6**. Detailed procedures for conducting in vitro bioassays are available in the project QA/QC guidance document (Section 10).

Table 3.2-5. Aqueous sample enrichment requirements for candidate *in vitro* screening assays.

	Reference chemical	Relative enrichment factor (REF)
Estrogen receptor alpha (ERa)	17-beta estradiol (+) 4-hydroxy-tamoxifen (-)	5 to 20 X
Androgen receptor (AR)	flutamide (-)	20 to 50 X
Progesterone receptor (PR)	Levonorgestrel (+)	20 to 50 X
Glucocorticoid receptor (GR)	Dexamethasone (+)	10 to 50 X
Aryl hydrocarbon receptor (AhR)	PCB 126 (+)	TBD

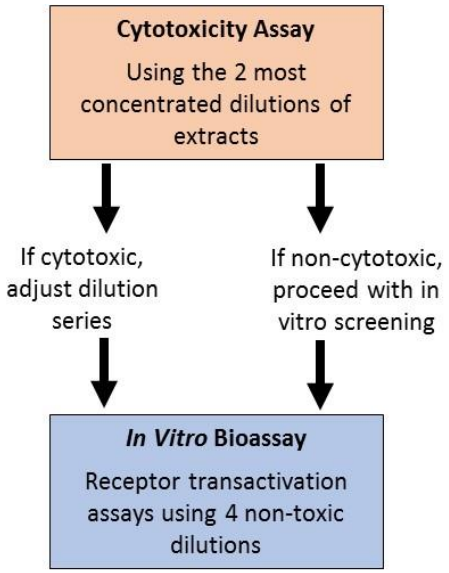


Figure 3.2-2. In vitro bioassay endpoints are sequenced to screen for cytotoxicity prior to testing for specific modes of action.

Table 3.2-6. Test conditions and QA/QC requirements for candidate *in vitro* screening assays

Parameters	***In Vitro* Bioassays Test Conditions**
Assay plates	96- or 384-well plates, black wall clear-bottom
Test samples	4 non-cytotoxic dilutions run in triplicate
Reference chemicals	Potent chemical used to calculate bioassay equivalent concentration (BEQ) - Initial calibration : 9 concentrations minimum within the dynamic range; analyzed in triplicate - Calibration verification: 5 concentrations minimum (in the lower end of the dynamic range) in duplicate
QA/QC	- Cell free media blank response – assay media only - Vehicle free response – cells in assay media - Vehicle blank response – cells with solvent vehicle - Matrix spike response
Acceptability criteria	Cytotoxicity assay- 80% or more survival compare to control Cell free blank response shall be less than 75% of the vehicle free response Vehicle blank response shall be within 15% RPD of the vehicle free response

3.3 Tier II – Toxicity Testing Using Whole Organisms

The Panel recommended that *in vivo* tests be conducted to evaluate the effects of environmental CECs on key biological processes such as development, reproduction and behavior in whole organisms. Toxicity testing using whole organisms will be implemented to (1) determine the levels of exposure to CECs and complex mixtures affecting sensitive organisms; and (2) to establish linkage between *in vitro* screening results and *in vivo* apical endpoints.

3.3.1 Linkage of In Vitro Responses with Effects on Fish Reproduction

Questions to be addressed:

1. What are the NOECs and LOECs of model compounds *in vivo*?
2. What is the relationship between *in vitro* assay responses and adverse effects on fish reproduction?

These studies will provide quantitative linkage between effects measured *in vitro* (i.e. induction/ suppression of receptor activity) and *in vivo* (i.e. reproductive output, sexual characteristics). The 21-day fathead minnow (*Pimephales promelas*) reproductive assay will be performed in accordance with USEPA (2007) and OECD (2012) guidelines, as summarized in the project QA/QC guidance document (Section 10). The toxicity of model compounds known to affect ER and AR receptors will be investigated. Specific parameters to be measured in this study are described in **Table 3.3-1**. Water samples should be collected directly from the exposure tanks and extracted and analyzed using the appropriate cell receptor assay and targeted chemistry.

Table 3.3-1. Key test parameters for linkage study of *in vitro* and *in vivo* responses to model compounds

	Test parameters - ER agonist
Chemicals	17-beta estradiol Solvent control (TEG or ethanol, less than 0.05%) Water control (no solvent)
In vitro endpoint	ER receptor transactivation
Fish assay endpoints	- % survival and changes in behavior relative to controls - No. eggs laid and fertilized - Levels of plasma steroids and vitellogenin (males) relative to controls - Reduction of the number of nuptial tubercles in males - Gonadosomatic index - Gonad histopathology (possible testis-ova in males) - qPCR (e.g. vtg, aromatase) and/or microarrays
	Test parameters - AR agonist
Chemicals	Trenbolone Solvent control (TEG or ethanol, less than 0.05%) Water control (no solvent)
In vitro endpoint	AR receptor transactivation
Fish assay endpoints	- % survival and changes in behavior relative to controls - No. eggs laid and fertilized - Levels of vitellogenin (in females) and plasma steroids relative to controls - Appearance of nuptial tubercles in females - Gonadosomatic index - Gonad histopathology (possible ovo-testis in females) - qPCR (e.g. vtg) and/or microarrays
	Test parameters - AR antagonist
Chemicals	Flutamide Solvent control (TEG or ethanol, less than 0.05%) Water control (no solvent)
In vitro endpoint	AR receptor activity inhibition
Fish assay endpoints	- % survival and changes in behavior relative to controls - No. eggs laid and fertilized - Levels of plasma steroids and vitellogenin (males) relative to controls - Reduction of the number of nuptial tubercles in males - Gonadosomatic index - Gonad histopathology (possible testis-ova) - qPCR and/or microarrays

3.3.2 Effects of CECs in Complex Environmental Matrices on Fish Reproduction

Questions to be addressed:

1. Do CECs present in complex mixtures effect fish physiology, behavior and reproduction?
2. What is the relationship between results of *in vitro* and *in vivo* assays?

The fish reproduction assay will be conducted using water samples from locations previously monitored by targeted chemical analyses and Tier I *in vitro* analyses (see **Table 3.2-1**), following the design in **Table 3.3-2**. The specific fish reproduction parameters to be measured in this study are described in **Table 3.3-1**.

Table 3.3-2. Aqueous test samples for fish reproduction assay

Scenario	Sample	Dilutions
Scenario 1 Freshwater	2 WWTP effluents Receiving river water Station #1 & 5 (Section 2.3.1)	1x – undiluted effluent 1x – undiluted samples
Scenario 2 Embayment*	2 WWTP effluents	1x – undiluted effluent 10x – worst case 100x – best case
Scenario 3 Oceans*	2 WWTP effluents	1x – undiluted effluent 50x – worst case > 1000x – best case

* Dilutions of WWTP effluent samples will be tested using the Fathead Minnow Assay until an estuarine/marine fish model is developed.

3.4 Tier III – In Situ Toxicity Assessment

In situ analyses will be conducted using fish species residing in the waterways previously monitored using targeted chemical analyses, Tier I (*in vitro* screening) and Tier II (*in vivo* laboratory exposures) assays.

The SWRCB has developed guidelines to sample and measure environmental chemicals (e.g. metals, PCBs, alkylphenols) in fish and invertebrates (Davis et al. 2014, SWAMP 2014). Tier III analyses will be conducted using the same fish species collected for tissue monitoring (Section 2.2.5). Recommended species include common carp, channel catfish, Sacramento sucker and largemouth bass for freshwater environments (scenario 1); topsmelt, white croaker, shiner surfperch and California halibut for coastal environments (scenario 2); white croaker, Dover sole, English sole, scorpion fish and hornyhead turbot (scenario 3). For in situ monitoring in the Delta, largemouth bass can serve as a sentinel fish species. For each waterway, a minimum of 2 species and 5 fish per species (n = 10 fish minimum) will be collected. Liver-somatic (LSI) and gonadosomatic (GSI) indexes will be evaluated. Gonads and liver will then be preserved for histopathological analyses.

4 STATEWIDE CEC MONITORING PROGRAM FRAMEWORK

4.1 Relationship Between Biological and Chemical Monitoring

A comprehensive monitoring strategy for aquatic ecosystems combines biological and chemical monitoring elements in a multi-tiered framework to determine if beneficial uses are compromised and intervening management action is needed (**Figure 4.1-1**). In Tier I, *in vitro* transactivation bioassays (see Section 3) screen for known and unknown CECs in concert with conventional targeted chemical analysis (see Section 2). Because all relevant MOAs and/or effects at the organism level are not addressed by currently available IVBs, periodic *in vivo* testing is also recommended in Tier I. If, however, screening level IVB results are below pre-established thresholds deemed protective, the frequency of *in vivo* testing in Tier I can be reduced. Should IVB results exceed thresholds, Tier II diagnostic evaluation using appropriate sentinel species and non-targeted chemical analysis (NTA) are undertaken to determine the likelihood and severity of impact, as well as to broaden the scope of pollutants targeted by chemical analysis in identifying likely causative stressors. If Tier II *in vivo* testing indicates a level of toxicity that is of concern, confirmatory monitoring (Tier III) is accelerated to determine if resources *in situ* are being impacted. Tier III monitoring is also necessary as an additional safeguard because Tier I and II monitoring tools are not entirely fail safe. The monitoring tools in Tiers I and II can also be utilized to identify MOAs and apical endpoints as well as chemical stressors in the case that *in situ* monitoring reveals an unacceptable level of impact.

4.2 Adaptive Management

The state of knowledge on CEC sources, fate and effects in aquatic ecosystems is continually evolving. To keep pace with new information and availability of new tools, the four-step adaptive process recommended by the Panel (**Figure 1.1-1**) is key to maintaining an up-to-date, relevant monitoring approach. Phase I sets the expectations of the pilot study, identifying and translating the most pressing management questions into fundamental, focused questions that subsequent monitoring will address. Phase II constitutes the data gathering step, as described in this 3-year pilot study plan, in this cyclical process. Plans should be made in Year 4 of this 5-year cycle for the subsequent evaluation of monitoring data and the efficacy of new monitoring tools and models that predict occurrence, effects and the linkage between *in vitro* and *in vivo* endpoints (Phase III). This evaluation should include a review and modification, as necessary, of the:

1. Updated monitoring trigger quotients (MTQs)
2. Scenarios and model watersheds sampled
3. Sampling design (sample size, frequency, spatial coverage)
4. CEC analyte list and matrix specific RLs
5. Performance of tools evaluated as part of the special studies, e.g. bioanalytical screening assays, non-targeted chemical analysis

The final year of the 5-year cycle (Phase IV) should be devoted to initiating management actions, as needed and as informed by the monitoring data. This step also provides an opportunity to revisit and revise, as necessary, the management and monitoring questions of importance regarding CECs, in preparation for initiation of the next monitoring cycle (Phase I).

4.2.1 Statewide Coordination

Convening of a management coordination team for statewide CEC pilot monitoring is recommended to capture the ever-changing scientific, regulatory and resource management landscape. Key functions for the management team include:

- Revisit, revise (as needed) and translate management questions into pilot study questions
- Review literature for updating benchmarks, thresholds and methodologies
- Set expectations for pilot study and generate minimum designs to achieve goals
- Compile, evaluate and analyze monitoring and modeling data
- Build consensus on interpretation of data
- Facilitate technology transfer for new, successful monitoring methods and models
- Foster communication with other CEC monitoring entities

The composition of a coordinated management team should consist of key representatives of the following (type) of organizations:

- State Water Board (e.g. SWAMP and SpOT coordinators)
- Regional Monitoring Agencies (SFEI, SCCWRP, Delta RMP, DPR)
- Stakeholders (CASA/Tri-TAC, CASQA, NGOs)
- Independent Science Advisory Panel

The coordination team should meet once a year, as a minimum, to perform the functions described above, e.g. a review of interim pilot study results and progress after the first year of a 3-year data collection cycle (Phase II). At the end of each 5-year pilot study cycle, the coordination team should hold a state-of-the-CEC monitoring symposium to reach consensus on the interpretation of pilot study information, discuss lessons learned, and chart a direction for future pilot monitoring cycles.

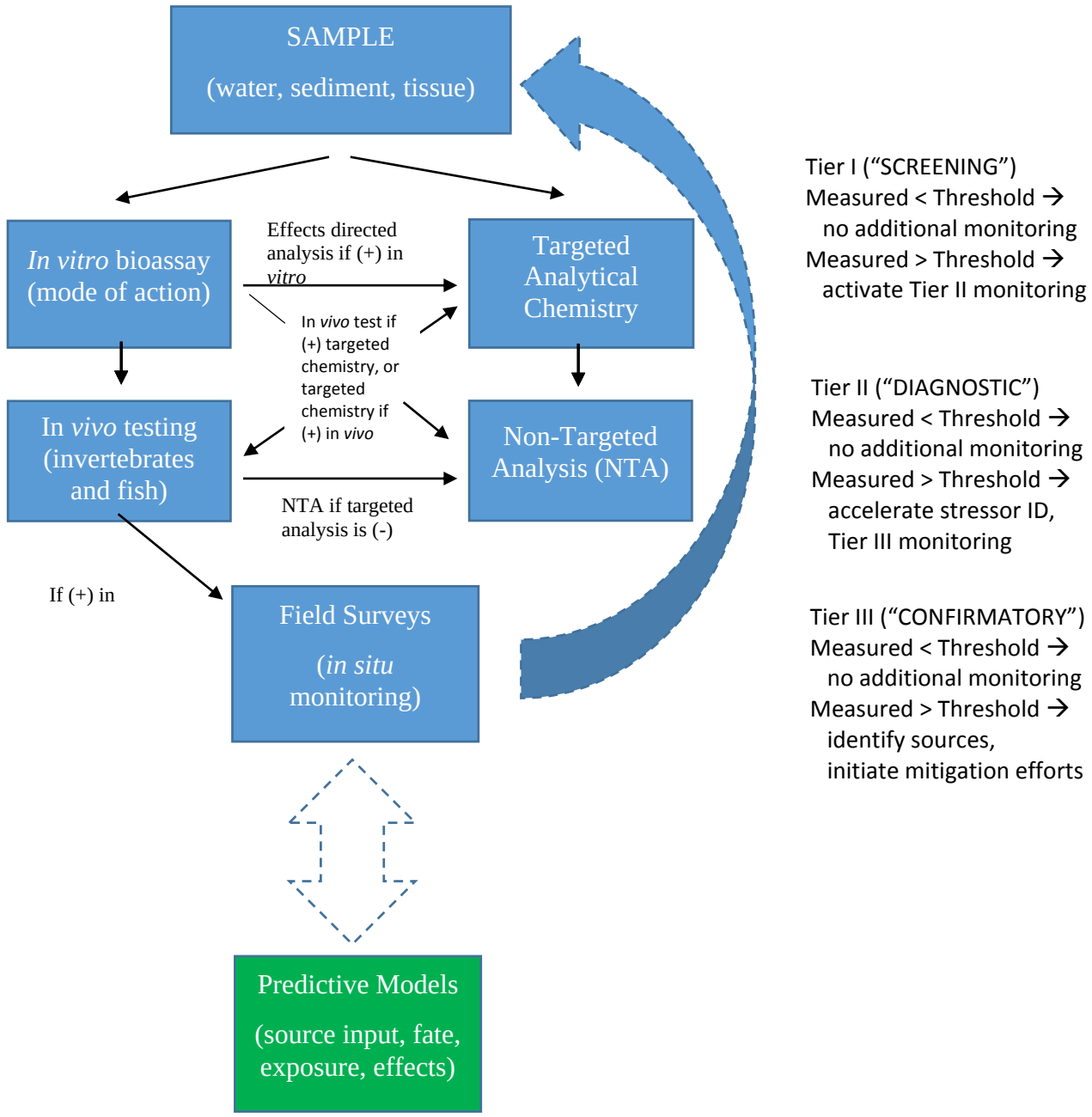


Figure 4.1-1. A comprehensive CEC management framework utilizes the results of tiered biological and chemical monitoring of increasing focus, complexity and relevance to efficiently screen for CECs and identify potential causative agents when cell-based, whole organism and field-scale impacts are observed, coupled with models that predict the potential for impact and that inform management on the effectiveness of corrective actions.

5 RESEARCH NEEDS

5.1 Toxicity Testing

Development of *in vivo* test species across habitats (fresh, marine, water column, sediment). The Panel recommended that whole organism toxicity tests focused on reproductive and/or developmental endpoints be conducted for all scenarios (except MS4) and matrices. The fathead minnow reproductive assay, proposed and described in Section 3, can only be applied to evaluate aqueous freshwater samples. Toxicity assays must be optimized and validated for other scenarios and matrices (**Tables 5.1-1, 5.1-2** and **5.1-3**).

Development of *in vitro* assays for all relevant modes of action. For effective bioanalytical monitoring, a comprehensive suite of *in vitro* endpoints is warranted. *In vitro* assays recommended for pilot CEC monitoring are commercially available and screen mostly for endocrine disrupting chemicals. Other environmentally relevant endpoints exist and need to be optimized for CEC monitoring (**Table 5.1-4**).

Table 5.1-1. Candidate fish species for estuarine/marine aqueous toxicity testing.

	Sheepshead minnow ***Cyprinodon variegatus***	**Atlantic killifish** ***Fundulus heteroclitus***	**Inland silverside** ***Menidia beryllina***
Test duration	180 days	15 days	15 – 20 days
Endpoints	- Fecundity, fertility, GSI - Plasma sex steroids and vitellogenin - Hatching success - Larval morphology	- Plasma sex steroid - Vitellogenin - GSI	- Fecundity, fertility - Molecular markers - Hatching success - Gonad histology
Strengths	- EPA validated protocol	- Killifish species are widespread	- EPA validated species - found in state waters
Limitations	- Long test duration - Less responsive to CECs than other fish	- Adapted to polluted environments - No egg output endpoint	- Reproductive endpoints have not been validated
References	Raimondo et al. (2009)	MacLatchy et al. (2003)	Personal communication (S. Brander, UNCW)

Table 5.1-2. Candidate invertebrate models for freshwater sediment toxicity testing.

	California blackworm ***Lumbriculus variegatus***	**Amphipod** ***Hyalella azteca***	**Midge** ***Chironomus species***
Test duration	28 days	42 days	44 days (*C. riparius*) 65 days (*C. tentans*)
Endpoints	- No. surviving worms - Growth (biomass) - Behavior (e.g. sediment avoidance)	- No. offspring/female - No. surviving adults - Sex ratio of surviving adults	- Development rate - Adult survival - Sex ratio of emerging adults - Fecundity and fertility
Comments	Asexual reproduction by regeneration	USEPA protocol currently optimized to include guidance on feeding and water quality	Shorter 28-day test is available with developmental endpoints
References	USEPA (2000), OECD (2007)	USEPA (2000)	OECD (2010)

Table 5.1-3. Candidate invertebrate models for estuarine/marine sediment toxicity testing.

	Polychaete ***Neanthes arenaceodentata***	**Amphipod** ***Leptocheirus plumulosus***	**Copepod** ***Amphiascus tenuiremis***
Test duration	28 days	28 days	16-17 days
Endpoints	- Survival - Growth - Bioaccumulation	- Survival - Growth rate - No. offsprings/adult - Behavior (sediment avoidance)	- Growth - Survival - Sex ratio - Fertility
Comments	No egg output endpoint	High variability often reported for reproduction	Patent rights on lab-cultured test organism
References	Farrar and Bridges (2011)	USEPA (2001), ASTM (2010)	Chandler et al. (2004b)

Development of *in situ* endpoints. In situ analyses conducted during routine environmental monitoring programs often focus on bioaccumulation of chemicals in tissues and the damages caused in tissues (histopathology). Special studies have also investigated the effects of environmental pollution on the population, but these studies can be expensive and time-consuming. Additional *in situ* endpoints indicative of early signs of exposure and toxicity should be developed. New molecular technologies measuring changes in gene expression (qPCR, microarrays, direct sequencing), protein levels (proteomics) and metabolite levels (metabolomics) have shown promising results (Biales et al. 2013; Martinovic-Weigelt et al. 2014; Skelton et al. 2014). Further research should be conducted using resident organisms to identify sensitive and reliable molecular endpoints.

Table 5.1-4. *In vitro* assays to develop for CEC monitoring

Endpoint	Mode of Action/ Adverse outcome
P53 or Umu	Genotoxicity
Peroxisome proliferator activated receptor (PPARa and PPARg)	Fatty acid storage, glucose metabolism
Acetylcholine receptor	Neurotoxicity
*Thyroid receptor (TR)**	*Metabolism, growth*

* Commercial assays exist but performance is highly variable.

5.2 Effect Directed Chemical Analysis

Environmental chemical mixtures inducing an *in vitro* assay response can be elucidated with a combination of targeted and non-targeted analysis. Targeted priority chemicals may explain a portion of the assay response, with the remaining unknown but responsible compounds identified through non-targeted analysis. This application is essentially a TIE methodology designed around the IVBs that utilizes recent advances in analytical instrumentation for non-targeted screening. Either gas-chromatography based (for hydrophobic compounds, e.g., GCxGC-TOF) or liquid chromatography based (for aqueous phase compounds, (e.g., LC-Q/TOF) non-targeted methods may be applied to the identification of bioactive compounds. The two primary research lines that must be addressed prior to implementing are the development of (1) libraries containing mass spectra and retention time information of chemicals with known *in vitro* and *in vivo* responses and (2) effects directed analytical methods that directly link bioassay response with chemical fractionation, which reduces mixture complexity and informs analytical method choice.

5.3 Passive Sampling Methods

As new science pushes monitoring thresholds lower, conventional environmental sampling and analytical methods become antiquated, incapable and cost-ineffective in concentrating high priority CECs from environmental media. Passive sampling methods (PSMs) show promise in sampling chemical constituents at very low occurrence in water, sediment and even biological tissue (sub-parts per billion concentrations). For hydrophobic CECs (e.g. PBDEs), PSMs that employ low density polyethylene films or polysiloxane (silicone) thin film coatings supported on hollow glass fibers or jars can pre-concentrate target analytes from freshwater, seawater, sediment and lipid-poor fish tissue. PSMs that employ sorbents that can concentrate both hydrophobic and hydrophilic CECs have been utilized is freshwater and coastal marine environments, however calibration of such samplers for estimation of concentration is incomplete. As the science on PSMs matures, and new approaches are developed and validated, these methods should be considered for future CEC monitoring programs in California water bodies.

5.4 Antibiotic Resistance

As identified by the Panel, antibiotics may adversely affect bacteria resulting in death at high clinical, therapeutic doses whereas at lower doses bacteria may survive and adapt to exposure by mutations which may result in development of antibiotic resistance (ABR). It remains unknown whether ABR in receiving waters of California is widespread, and if so, what implications for environmental quality and protection of beneficial uses would result from such occurrence. This is in large part due to the lack of definitive methods to quantify ABR in environmental media. Previous studies (Auerbach et al. 2007; FIWG-PIE 2009; Kummerer 2009; NOAA 2011; Pellegrini et al. 2011; Rosenblatt-Farrell 2009; Szczepanowski et al. 2004, 2009; USGS 2002; Uyaguari et al. 2009, 2011; Van Dolah et al. 2000) in other parts of the US have documented the high levels of ABR in WWTPs, confined animal feeding operations (CAFOs) and on golf courses receiving secondary treated effluent as irrigation. Antibiotic resistance can be initiated by low level exposure at concentrations below the Minimum Inhibitory Concentrations (MIC) for most antibiotics which may lead to the development of plasmids containing resistant genes which may be discharged into the environment (Bennett 2008; Garriss et al. 2009; Kummerer 2009; Pellegrini et al. 2011; Rosenblatt-Farrell 2009; Szczepanowski et al. 2004, 2009; Uyaguari et al. 2011). Distinct ABR patterns have been found within WWTPs and CAFOs which are related to the extent and magnitude of antibiotic use in humans and livestock. The panel felt that given the complexities for development of ABR it was important to focus on ABR monitoring on WWTP effluent and evaluate the ABR within indicator bacteria at each site initially to define the extent and magnitude of ABR within major point source discharges within these effluent dominated inland waterways. Based upon those results it would be imperative to develop more robust ABR assessment methods

Thus, development of standardized biological screening assays for quantitation of ABR in receiving water samples (water, sediment and tissue) for antibiotics that have been measured in monitoring studies conducted in California and throughout the US is recommended. To determine what risks due to ABR are plausible in California receiving waters, it is recommended that the SWRCB convene an expert panel of microbiologists, microbial ecologists, aquatic ecotoxicologists and water quality scientists, to define such risks, and to provide advice and oversight on the development and implementation of the ABR methods that can be employed in future monitoring studies. Specific focus of this workshop would include:

1. Identification of new/novel methods and approaches for assessing the extent and magnitude of of ABR beyond the current custom ABR panels which can currently address only the number and intensity (> MIC) of the ABR by individual antibiotics within the panel.
2. Identification of ABR genes which may pose the greatest risks to humans and wildlife (i.e. BLASTm-1 gene and genes that may cause Methicillin Resistant Staph. Aureus (MRSA)
3. The potential for lateral ABR gene transfer among microbial species including pathogens such as Vibrio bacteria and other species commonly found in wound infections.

5.5 Model Development

In addition to the collection of monitoring data, key data gaps on source contribution, occurrence and toxicity of CECs should be addressed through the development and application of environmental fate and effects sub-models (Anderson et al. 2012). Many such sub-models have been developed for various exposure scenarios, including WWTP discharge into rivers and coastal embayment box models that consider contaminant input from multiple sources. At the federal level, USEPA is developing a comprehensive modeling strategy that combines predictions of exposure (Expocast; http://www.epa.gov/ncct/expocast/) and toxicity (ToxCast; http://www.epa.gov/ncct/toxcast/) for thousands of current use and high production chemicals. EPA's effort is currently focused on human health, but plans are to eventually address ecological receptors as well. The development and calibration of such sub-models using pilot monitoring data, and subsequent integration of modular modeling components that characterize source input, fate, exposure and effects into a comprehensive management "on-ramp" tool will be useful in assessing the impact of management actions, e.g. best management practices (BMPs), implemented or proposed to reduce the potential for impact by CECs. Specific recommendations include:

1) Improve and expand the application of conceptual models to estimate occurrence, distribution among aqueous, particulate, sediment and biological compartments, to assist design monitoring efforts and to evaluate CEC control measures. These models should also be used to refine screening evaluations on CEC sources and indirect exposure routes for hydrophobic CECs presented in this document. This work should be sequenced according to the complexity of exposure scenarios, e.g. effluent dominated waterways (Scenario 1) would represent the simplest starting scenario.

2) Develop a screening-level mass-based model to estimate the predicted environmental concentrations (PECs) in effluents and stormwater runoff coupled with structure-based toxicity assessments.

3) Tailor the construct and outputs from EPA's Expocast and Toxcast to address scenarios of highest importance for CECs in California receiving waters.

4) Integrate calibrated sub-models addressing source input, fate, exposure and effects into a comprehensive management CEC impact or "on-ramp" model.

5) Generate credible values (or ranges thereof) for critical model parameters, including
   a) bioaccumulation and trophic transfer factors for high priority bioaccumulative CECs, including PFOS and PBDEs, for freshwater, estuarine and marine food webs.
   b) measured or predicted half-lives and/or clearance rates of high priority CECs in aqueous (fresh and seawater), sediment and tissue.
   c) relative potency factors for CECs that link molecular initiating events (e.g. positive IVB response) and whole organism apical effects (e.g. reduced fecundity).

6 REFERENCES

Alvarez, D.A., Maruya, K.A., Dodder, N.G., Lao, W., Furlong, E.T., Smalling, K.L. 2013. Occurrence of contaminants of emerging concern along the California coast (2009-10) using passive sampling devices. *Mar Pollut Bull* 81:347-354.

Anderson, P.D., Denslow, N.D., Drewes, J.E., Olivieri, A.W., Schlenk, D., Scott, G.I., Snyder, S.A. 2012. Monitoring strategies for chemicals of emerging concern (CECs) in California's aquatic ecosystems. Recommendations of a Science Advisory Panel. Final Report. Costa Mesa (CA), USA: Southern California Coastal Water Research Project Authority, Technical Report 692, 215 pp.

American Society for Testing and Materials (ASTM). 2010. Standard method for measuring the toxicity of sediment-associated contaminants with estuarine and marine invertebrates (E1367). pp. 400-461, 2010 Annual Book of ASTM Standards, Vol. 11.06. American Society for Testing and Materials. West Conshohocken, PA.

Auerbach, E.A., Seyfried, E.E., McMahon, K.D. 2007. Tetracycline resistance genes in activated sludge wastewater treatment plants. *Water Res* 41:1143-1151.

Bennett, P. M. 2008. Plasmid encoded antibiotic resistance: acquisition and transfer of antibiotic resistance genes in bacteria. *Br J Pharmacol* 153 Suppl 1:S347-57.

Biales, A.D., Kostich, M., Burgess, R.M., Ho, K.T., Bencic, D.C., Flick, R.L., Portis, L.M., Pelletier, M.C., Perron, M.M., Reiss, M. 2013. Linkage of genomic biomarkers to whole organism end points in a Toxicity Identification Evaluation (TIE). *Environ Sci Technol* 47:1306-1312.

Bight'13 Contaminant Impact Assessment Committee. 2013. Southern California Bight 2013 regional marine monitoring survey: Contaminant impact assessment workplan, Southern California Coastal Water Research Project Authority, Costa Mesa, CA, 80 pp.

Budd, R. 2014. Study 270 (2014-15) – Ambient and mitigation monitoring in urban areas of southern California during fiscal year 2014-15. Department of Pesticide Regulation, Sacramento, CA, 22 pp.

Chandler, G.T., Cary, T.L., Bejarano, A.C., Pender, J., Ferry, J.L. 2004a. Population consequences of fipronil and degradates to copepods at field concentrations: an integration of life cycle testing with leslie matrix population modeling. *Environ Sci Technol* 38:6407–6414.

Chandler, G.T., Cary, T.L., Volz, D.C., Walse, S.S., Ferry, J.L., Klosterhaus, S.L. 2004b. Fipronil effects on estuarine copepod (*Amphiascus tenuiremis*) development, fertility, and reproduction: a rapid life-cycle assay in 96-well microplate format. *Environ Toxicol Chem* 23:117–124.

Davis, J.A., Ross, J.R.M., Bezalel, S.N., Hunt, J.A., Ichikawa, G., Bonnema, A., Heim, W., Crane, D., Swenson, S., Lamerdin, C. 2013. Contaminants in fish from California Rivers and Streams 2011. A report of the Surface Water Ambient Monitoring Program (SWAMP), State Water Resources Control Board, Sacramento, CA, 54 pp.

Davis, J.A., Ross, J.R.M., Bezalel, S.N., Hunt, J.A., Melwani, A.R., Allen, R.M., Ichikawa, G., Bonnema, A., Heim, W., Crane, D., Swenson, S., Lamerdin, C., Stephenson, M., Schiff, K. 2014. Contaminants in fish from the California coast. 2009-2010: summary report on a two-year screening survey. A report of the Surface Water Ambient Monitoring Program (SWAMP), State Water Resources Control Board, Sacramento, CA, 87 pp.

Delgado-Moreno, L., Lin, K., Veiga-Nascimento, R., Gan, J. 2011. Occurrence and toxicity of three classes of insecticides in water and sediment in two southern California coastal watersheds. *J Agric Food Chem* 59:9448–9456.

Dodder, N.G., Maruya, K.A., Lee Ferguson, P., Grace, R., Klosterhaus, S., La Guardia, M.J., Lauenstein, G.G., Ramirez, J. 2014. Occurrence of contaminants of emerging concern in mussels (*Mytilus* spp.) along the California coast and the influence of land use, storm water discharge, and treated wastewater effluent. *Mar Pollut Bull* 81:340–346.

Ensminger, M.P., Budd, R., Kelley, K.C., Goh, K.S. 2013. Pesticide occurrence and aquatic benchmark exceedances in urban surface waters and sediments in three urban areas of California, USA, 2008-2011. *Environ Monit Assess* 185:3697–3710.

Emsinger, M.P. 2014. Study 269 (2014-15) – Ambient and mitigation monitoring in urban areas of northern California during fiscal year 2014-15. Department of Pesticide Regulation, Sacramento, CA, 19 pp.

Escher, B.I., Allinson, M., Altenburger, R., Bain, P.A., Balaguer, P., Busch, W., Crago, J., Denslow, N.D., Dopp, E., Hilscherova, K., Humpage, A.R., Kumar, A., Grimaldi, M., Jayasinghe, B.S., Jarosova, B., Jia, A., Makarov, S., Maruya, K.A., Medvedev, A., Mehinto, A.C., Mendez, J.E., Poulsen, A., Prochazka, E., Richard, J., Schifferli, A., SchlenK, D., Scholz, S., Shiraishi, F., Snyder, S., Su, G., Tang, J.Y., Burg, B.V., Linden, S.C., Werner, I., Westerheide, S.D., Wong, C.K., Yang, M., Yeung, B.H., Zhang, X., Leusch, F.D. 2014. Benchmarking organic micropollutants in wastewater, recycled water and drinking water with in vitro bioassays. *Environ Sci Technol* 48:1940-1956.

Farrar, J.D., Bridges, T.S. 2011. 28-day chronic sublethal test method for evaluating whole sediments using an early life stage of the marine polychaete *Neanthes arenaceodentata*. ERDC TN-DOER-R14. US Army Corps of Engineers. Vicksburg, MS.

Federal Interagency Working Group – Pharmaceuticals in the Environment (FIWG-PIE). 2009. Pharmaceuticals in the Environment – An Interagency Research Strategy. U.S Congressional Committee on Environmental and Natural Resources Report, Washington, DC, 61 pp.

Filby, A.L., Thorpe, K.L., Maack, G., Tyler, C.R. 2007. Gene expression profiles revealing the mechanisms of anti-androgen- and estrogen-induced feminization in fish. *Aquat Toxicol* 81:219-231.

Gan, J., Bondarenko, S., Oki, L., Haver, D., Li, J.X. 2012. Occurrence of fipronil and its biologically active derivatives in urban residential runoff. *Environ Sci Technol* 46:1489–1495.

Garriss, G., Waldor, M.K., Burrus, V. 2009. Mobile antibiotic resistance encoding elements promote their own diversity. *PLoS Genet* 5:e1000775.

Harris, C.A., Henttu, P., Parker, M.G., Sumpter, J.P. 1997. The estrogenic activity of phthalate esters in vitro. *Environ Health Perspect* 105:802-811.

Hoh, E., Dodder, N.G., Lehotay, S.J., Pangallo, K.C., Reddy, C.M., Maruya, K.A. 2012. Nontargeted comprehensive two-dimensional gas chromatography/time-of-flight mass spectrometry method and software for inventorying persistent and bioaccumulative contaminants in marine environments. *Environ Sci Technol* 46:8001–8008.

Kjeldsen, L.S., Bonefeld-Jorgensen, E.C. 2013. Perfluorinated compounds affect the function of sex hormone receptors. *Environ Sci Pollut Res* 20:8031-8044.

Klosterhaus, S., Yee, D., Sedlak, M., Wong, A., Sutton, R. 2013. Contaminants of emerging concern in San Francisco Bay: a summary of occurrence data and identification of data gaps. RMP Contribution 698, San Francisco Estuary Institute, Richmond, CA, 121 pp.

Kummerer, K. 2009. Antibiotics in the aquatic environment--a review--part I. *Chemosphere* 75:417-34.

Lao, W., Tsukada, D., Greenstein, D.J., Bay, S.M., Maruya, K.A. 2010. Analysis, occurrence, and toxic potential of pyrethroids, and fipronil in sediments from an urban estuary. *Environ Toxicol Chem* 29:843–851.

Leusch, F.D.L., De Jager, C., Levi, Y., Lim, R., Puijker, L., Sacher, F., Tremblay, L.A., Wilson, V.S., Chapman, H.F. 2010. Comparison of five in vitro bioassays to measure estrogenic activity in environmental waters. *Environ Sci Technol* 44:3853-3860.

Long, M., Laier, P., Vinggaard, A.M., Anderson, H.R., Lynggaard, J., Bonefeld-Jorgensen, E.C. 2003. Effects of currently used pesticides in the AhR-CALUX assay: comparison between the human TV101L and the rat H4IIE cell line. *Toxicol* 194:77-93.

Luo, Y. 2014. Review of modeling approaches for pesticide washoff from impervious surfaces. ACS Symposium. October 3, 2014. 20 pp.

MacLatchy, D.L., Courtenay, S.C., Rice, C.D., Van Der Kraark, G.J. 2003. Development of a short-term reproductive endocrine bioassay using steroid hormone and vitellogenin endpoints in the estuarine mummichog (*Fundulus heteroclitus*). *Environ Toxicol Chem* 22:996-1008.

Markle, J.C., van Buuren, B.H., Moran, K., Barefoot, A.C. 2014. Pyrethroid pesticides in municipal wastewater: A baseline survey of publicly owned treatment works facilities in California in 2013. In: Describing the behavior and effects of pesticides in urban and agricultural settings, ACS Symposium Series. American Chemical Society, pp. 177–194.

Martinovic-Weigelt, D., Mehinto, A.C., Ankley, G.T., Denslow, N.D., Barber, L.B., Lee, K.E., King, R.J., Schoenfuss, H.L., Schroeder, A.L., Villeneuve, D.L. 2014. Transcriptomic effects-based monitoring for endocrine active chemicals: assessing relative contribution of treated wastewater to downstream pollution. *Environ Sci Technol* 48:2385-2394.

Maul, J.D., Brennan, A.A., Harwood, A.D., Lydy, M.J. 2008. Effect of sediment-associated pyrethroids, fipronil, and metabolites on *Chironomus tentans* growth rate, body mass, condition index, immobilization, and survival. *Environ Toxicol Chem* 27:2582–2590.

Meng, X.-Z., Blasius, M.E., Gossett, R.W., Maruya, K.A. 2009. Polybrominated diphenyl ethers in pinnipeds stranded along the southern California coast. *Environ Pollut* 157:2731–2736.

Organization for Economic Co-operation and Development (OECD). 2007. Sediment-water *Lumbriculus* toxicity test using spiked sediment. Test guideline No. 225, Guidelines for the testing of chemicals. OECD, Paris.

OECD. 2010. Sediment-water chironomid life-cycle toxicity test using spiked water or spiked sediment. Test guideline No. 233, Guidelines for the testing of chemicals. OECD, Paris.

Panter, G.H., Hutchinson, T.H., Hurd, K.S., Sherren, A., Stanley, R.D., Tyler, C.R. 2004. Successful detection of (anti-) androgenic and aromatase inhibitors in pre-spawning adult fathead minnows (*Pimephales promelas*) using easily measured endpoints of sexual development. *Aquat Toxicol* 70:11-21.

Pellegrini, C., Celenza, G., Segatore, B., Bellio, P., Setacci, D., Amicosante, G., Perilli, M. 2011. Occurrence of class 1 and 2 integrons in resistant Enterobacteriaceae collected from a urban wastewater treatment plant: first report from central Italy. *Microb Drug Resist* 172): 229-234.

Phillips, B.M., Anderson, B.S., Siegler, K., Vorhees, J., Tadesse, D., Webber, L., Breuer, R. 2014. Trends in chemical contamination, toxicity and land use in California watersheds: Stream Pollution Trends (SPoT) monitoring program third report – Five-year trends 2008-2012. SWAMP-MR-SB-2014-001, State Water Resources Control Board, Sacramento, CA.

Raimondo, S., Hemmer, B.L., Goodman, L.R., Cripe, G.M. 2009. Multigenerational exposure of the estuarine sheepshead minnow (*Cyprinodon variegatus*) to 17β-estradiol. II. Population-level effects through two life cycles. *Environ Toxicol Chem* 28:2409–2415.

Rosenblatt-Farrell N. 2009. The Landscape of Antibiotic Resistance. *Environ Health Perspect* 117:A244-A250.

San Francisco Estuary Institute (SFEI). 2014. RMP Update 2014. SFEI Contribution 728. San Francisco Estuary Institute, Richmond, CA.

Santa Ana Watershed Project Authority (SAWPA). 2014. 2013 sampling report for emerging constituents in the Santa Ana Region. SAWPA, Riverside, CA, 54 pp.

Santa Clara Valley Urban Runoff Pollution Prevention Program (SCVURPPP). 2014. Watershed monitoring and assessment program. Integrated Monitoring Report – Part A. Water Quality Monitoring Years 2012 and 2013. 364 pp.

Schiff, K., Stein, E.D., Aminzadeh, S., Boehm A., Hildebrand, G., Honeybourne, L., Nasseri, I., Ode, P., Taylor. S., Senn, D., Smith, J., Sommers, C., Strecker, E. 2014. Southern California Stormwater Monitoring Coalition 2014 Research Agenda. Technical Report 828. Southern California Coastal Water Research Project Authority, Costa Mesa, CA. 33 pp.

Schreurs, R., Sonneveld, E., Jansen, J.H.J., Seinen, W., van der Burg, B. 2005. Interaction of polycyclic musks and UV filters with the estrogen receptor (ER), androgen receptor (AR), and progesterone receptor (PR) in reporter gene bioassays. *Toxicol Sci* 83, 264-272.

Sedlak, M.D., Greig, D.J., 2012. Perfluoroalkyl compounds (PFCs) in wildlife from an urban estuary. *J Environ Monit* 14:146–154.

Sengupta, A., Lyons, J.M., Smith, D.J., Heil, A., Drewes J.E., Snyder, S.A., Maruya, K.A. 2014. The occurrence and fate of chemicals of emerging concern (CECs) in coastal urban rivers receiving discharge of treated municipal wastewater effluent. E*nviron Toxicol Chem* 33:350-358.

Shaul, N.J., Dodder, N.D., Aluwihare, L.I., Mackintosh, S.A., Maruya, K.A., Chivers, S.J., Danil, K., Weller, D.W., Hoh, E., 2014. Nontargeted biomonitoring of halogenated organic compounds in two ecotypes of bottlenose dolphins (*Tursiops truncatus*) from the Southern California Bight, *Environ Sci Technol* DOI:10.1021/es505156q.

Skelton, D.M., Ekman, D.R., Martinovic-Weigelt, D., Ankley, G.T., Villeneuve, D.L., Teng, Q., Collette, T.W. 2014. Metabolomics for in situ environmental monitoring of surface waters impacted by contaminants from both point and nonpoint sources. *Environ Sci Technol* 48:2395-2403.

Stormwater Monitoring Coalition/Bioassessment Working Group (SMC/BWG). 2007. Regional monitoring of southern California's coastal watersheds. Technical Report 539, Southern California Coastal Water Research Project Authority, Costs Mesa, CA, 33 pp.

Sutton, R., Sedlak, M., Yee, D. 2013. Contaminants of emerging concern in San Francisco Bay: a strategy for future investigations. RMP Contribution 700, San Francisco Estuary Institute, Richmond, CA, 56 pp.

Sutton, R., Sedlak, M., Davis, J. 2014. Polybrominated Diphenyl Ethers (PBDEs) in San Francisco Bay: A summary of occurrence and trends. Contribution Number 713, San Francisco Estuary Institute, Richmond, CA.

Surface Water Ambient Monitoring Program (SWAMP). 2014. Sampling and analysis plan for a study of lakes and reservoirs with low concentrations of contaminants in sport fish (Final). State Water Resources Control Board, Sacramento, CA.

Szczepanowski, R., Krahn, I., Linke, B., Goesmann, A., Puhler, A., Schluter, A. 2004. Antibiotic multiresistance plasmid pRSB101 isolated from a wastewater treatment plant is related to plasmids residing in phytopathogenic bacteria and carries eight different resistance determinants including a multidrug transport system. *Microbiology* 150:3613-30.

Szczepanowski, R., Linke, B., Krahn, I., Gartemann, K.H., Gutzkow, T., Eichler, W., Puhler, A., Schluter, A. 2009. Detection of 140 clinically relevant antibiotic-resistance genes in the plasmid metagenome of wastewater treatment plant bacteria showing reduced susceptibility to selected antibiotics. *Microbiology* 155:2306-19.

Teerlink, J. 2014. Pyrethroids in Wastewater: Review of potential sources, occurrence and Fate. Department of Pesticide Regulation, Sacramento, CA, 6 pp.

U.S. Environmental Protection Agency (USEPA). 1996. Pesticide fact sheet: Fipronil. EPA-737-F-96-005. USEPA Office of Prevention, Pesticides, and Toxic Substances. Washington, DC.

USEPA. 2000. Methods for measuring the toxicity and bioaccumulation of sediment associated contaminants with freshwater invertebrates. Second edition. EPA 600/R-99/064. Revision to the first edition dated June 1994.

USEPA. 2001. Methods for assessing the chronic toxicity of marine and estuarine sediment-associated contaminants with the amphipod *Leptocheirus plumulosus*. EPA/600/R-01/020. Washington, DC.

USEPA (2007) Validation of the Fish Short-Term Reproduction Assay: Integrated Summary Report, December 2007, accessed 9/29/2014, at http://www.epa.gov/endo/pubs/att-f_fish_assay_protocol.pdf.

U.S. Geological Survey (USGS). 2002. Water-quality data for pharmaceuticals, hormones, and other organic wastewater contaminants in U.S. streams, 1999-2000" (USGS Open-File Report 02-94)1999-2000 Survey of 139 US Waterways. USGS, Washington, DC.

Uyaguari, M.**,** Key, P.B., Gooch, **J.,** Jackson, K., Scott. G.I. 2009. Acute effects of the antibiotic oxytetracycline on the bacterial community of the grass shrimp, *Palaemonetes pugio. Environ Toxicol Chem* 28:2715-2724.

Uyaguari, M., Norman, S., Gooch**,** J.**,** Jackson, K., Scott, G.I. 2011. The discovery of novel bacterial antibiotic resistance genes in activated sludge using a metagenomic approach. *J Appl Environ Microbiol* submitted.

Van Dolah, F., Van Dolah, R.F., Chestnut, D.E., Scott, G.I. 2000. A baseline assessment ofenvironmental and biological conditions in Broad Creek and the Okatee River, Beaufort County, South Carolina. Final Report to the Beaufort County Council, 281p.

Weston, D. P., Lydy, M.J. 2014. Toxicity of the insecticide fipronil and its degradates to benthic macroinvertebrates of urban streams. *Environ Sci Technol* 48:1290-1297.

Worcester, K. 2011. Assessment of surface water quality and habitat in agricultural areas of the Central Coast of California, and associated risk to the marine environment. SWAMP, State Water Resources Control Board, Sacramento, CA, 92 pp.

7 GLOSSARY OF TERMS

ABR	Antibiotic Resistance
AhR	Aryl hydrocarbon receptor
AR	Androgen Receptor
BASMAA	Bay Area Stormwater Management Agencies Association
BBP	Butylbenzylphthalate
BEHP	Bis(2-ethylhexyl)phthalate
BEQ	Bioassay equivalent concentration
BOG	Bioaccumulation Oversight Group
CECs	Chemicals of Emerging Concern
DDT	Dichlorodiphenyltrichloroethane
DMSO	Dimethylsulfoxide
DPR	Department of Pesticide Regulation
DRMP	Delta Regional Monitoring Program
Dw	Dry weight
E2	17β-estradiol
EDC	Endocrine Disrupting Chemical
ECWG	Emerging Contaminants Work Group
FME	Fixed mass emission
GC-MS	Gas Chromatography-Mass Spectrometry
GCxGC/TOF-MS	Two Dimensional Gas Chromatography-Time of Flight Mass Spectrometry
GR	Glucocorticoid Receptor
IVB	In vitro bioassay
LC-MS	Liquid Chromatography-Mass Spectrometry
LOEC	Lowest Observed Effect Concentration
MEC	Measured Environmental Concentration
mgd	Million gallons per day
MOA	Mode of Action
MS4	Municipal Separate Storm Sewer System
MTL	Monitoring Trigger Level
MTQ	Monitoring Trigger Quotient
NIST	National Institute of Standards and Technology
NOEC	No Observed Effect Concentration
NPDES	National Pollutant Discharge Elimination System
NTA	Non-targeted chemical analysis
PAH	Polycylic Aromatic Hydrocarbon
PBDE	Polybrominated Diphenyl Ether
PCB	Polychlorinated Biphenyl

PEC	Predicted Environmental Concentration
PFC	Perfluorinated Compound
PFOS	Perfluorooctane Sulfonate
PNEC	Predicted No Effect Concentration
POC	Pollutant of concern
POTW	Publicly Owned Treatment Works
PR	Progesterone Receptor
PSD	Passive sampling device
PSM	Passive sampling method
QA/QC	Quality Assurance/Quality Control
QAPP	Quality Assurance Project Plan
QSAR	Quantitative Structure Activity Relationship
REF	Relative enrichment factor
RL	Reporting limit
RMC	Regional Monitoring Coalition
RMP	Regional Monitoring Program
RW	Receiving Water
RWQCB	Regional Water Quality Control Board
SCCWRP	Southern California Coastal Water Research Project
SFEI	San Francisco Estuary Institute
SMC	Stormwater Monitoring Coalition
SPoT	Stream Pollution Trends Monitoring Program
SRM	Standard Reference Material
S&T	Status and Trends
SWAMP	California Surface Water Ambient Monitoring Program
SMC	Stormwater Monitoring Coalition
SWPP	Surface Water Protection Program
SWRCB	State Water Resources Control Board
TIE	Toxicity Identification Evaluation
TSS	Total Suspended Solids
USGS	United States Geological Survey
USEPA	United States Environmental Protection Agency
VTG	Vitellogenin
WET	Whole Effluent Testing
WWTP	Wastewater Treatment Plant

8 APPENDICES

8.1 Appendix A: Summary of CEC Expert Panel Recommendations

Table 8.1-1. Constituents of emerging concern (CECs) recommended for pilot (Phase 2) monitoring by the CEC Ecosystems Panel. Each column lists exposure scenarios (E = coastal embayment; F = inland freshwater, O = ocean) and matrices of interest (i.e., aqueous, sediment, tissue). M = monitor; NA = not applicable. WWTP – municipal wastewater treatment plant.

Scenario	Source: WWTP Effluent	Source: Storm Water (MS4)	Scenario 1 Effluent Dominated Inland Freshwater	Scenario 2 Embayment		Scenario 3 Ocean	All Scenarios
Matrix	**Aqueous**	**Aqueous, Sediment**	**Aqueous**	**Aqueous**	**Sediment**	**Sediment**	**Tissue**
Additional Information in Panel Report			Tables 6.1 & 6.6	Table 6.2	Table 6.3	Table 6.4	Table 6.5
Bis(2-ethylhexyl) phthalate (BEHP)	O	NA	NA	NA	NA	M	NA
Butylbenzyl phthalate (BBP)	O	NA	NA	NA	NA	M	NA
p-Nonylphenol	O	NA	NA	NA	NA	M	NA
Bifenthrin	E F	M	M	M	M	NA	NA
Permethrin	E F	M	M	M	M	NA	NA
Chlorpyrifos	E F	M	M	M	NA	NA	NA
Estrone	E F	M	M	M	NA	NA	NA
17-beta estradiol	E F	M	M	M	NA	NA	NA
Galaxolide (HHCB)	E F	M	M	M	NA	NA	NA
Bisphenol A	E F	M	M	M	NA	NA	NA
Ibuprofen	F	M	M	NA	NA	NA	NA
Diclofenac	F	M	M	NA	NA	NA	NA
Triclosan	F	M	M	NA	NA	NA	NA
PBDE -47 and -99	E F O	M	NA	NA	M	M	M
PFOS	E F O	M	NA	NA	M	M	M

Table 8.1-2. Preliminary design guidance for pilot monitoring of CECs (Phase 2) in each of the three receiving water scenarios and for stormwater (MS4) discharge. F = freshwater; M = monitor; NA = not applicable; RW = receiving water.

	Source	Scenario 1	Scenario 2	Scenario 3
General Monitoring Design Parameters	**Stormwater (MS4) Discharging to Receiving Water[a]**	**WWTP Discharging to Inland Freshwater[b]**	**WWTP Discharging to Coastal Embayment[c]**	**WWTP Discharging to Ocean[d]**
Spatial coverage – Receiving Water (RW)	1-D gradient (up to 6 sites for each location)	1-D (up to 6 sites for each location)	2-D gradient (up to 7 sites in estuary)	2-D grid (up to 7 sites each location)
Number of POTW and/or FW Locations	Two large FW streams and the Delta	Two POTWs and RW	Five POTWs in one estuary/embayment	Two POTWs and corresponding RWs
Frequency	Wet and Dry Season over three years	Wet and Dry Season over three years	Semi-annual (aqueous) or annual (sediment, tissue) over three years	Semi-annual (aqueous) or annual (sediment, tissue) over three years
Background	M	M	M	M
Aqueous (non-filtered)	M	M	M	NA
Sediment (top 5 cm)	M	M	M	M
Tissue [e]	M	M	M	M

a - Potentially conduct pilot investigation for one stream in the San Francisco Bay Area; one stream in Southern California, and one stream in the Sacramento-San Joaquin Delta.

b - Potentially conduct pilot investigation in Southern California.

c - Daily discharge <100 mgd; potentially conduct pilot investigation in San Francisco Bay.

d - Daily discharge $\geq$100 mgd; potentially conduct pilot investigation in southern California.

e - Identify appropriate species and tissues (e.g., bivalve and fish tissue for PBDEs; bird eggs for PFOS).

Table 8.1-3. Special studies recommended for pilot evaluation (Phase 2) to improve CEC monitoring in aquatic ecosystems. WWTP – municipal wastewater treatment plant.

Special Study	WWTP Discharging to Inland Freshwater (Scenario 1)	WWTP Discharging to Coastal Embayment (Scenario 2)	WWTP Discharging to Ocean (Scenario 3)	Stormwater (MS4) Discharging to Receiving Water
Bioanalytical Screening Assays[a]	yes	yes	yes	yes
Toxicity[b]	yes	yes	yes	no
Antibiotic Resistance[c]	yes	yes	no	no
Passive Sampling Devices (PSDs)[d]	yes	no	yes	no

a – Conduct evaluation and validation of bioanalytical screening methods in combination with targeted and non-targeted chemical analyses to identify bioactive substances using a toxicity identification evaluation (TIE) process.

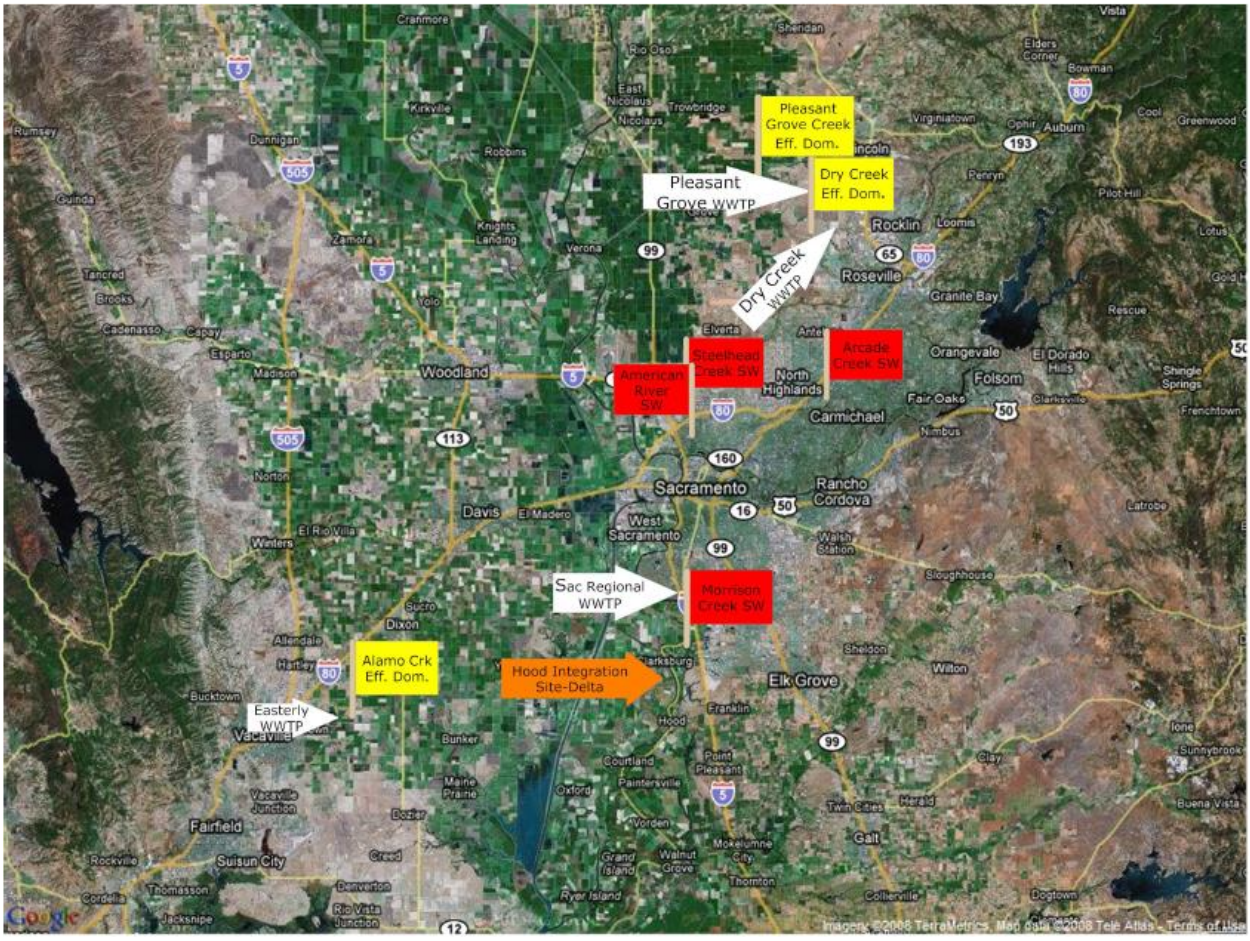
b – e.g. 21 d fathead minnow recrudescence assay for freshwater matrices. Implement periodic reproduction assessments using appropriate fish and invertebrate species. Coordinate efforts with NPDES WET and bioassessment monitoring. This assay should be used for investigative purposes.

c -- Conduct a pilot investigation using a bioassay to screen for antibiotic resistance in effluent, water and/or sediment.

d – Conduct a pilot investigation using PSDs that provide adequate capacity to concentrate the CECs in the priority list. These devices should have demonstrated acceptable performance in laboratory or field validation studies, and published guidance on translation of results.

8.2 Appendix B: Delta Station Map

Candidate northern California Delta Scenario 1 WWTP (white) and Stormwater (red) station locations.



8.3 Appendix C: Bight '13 Outfall Special Study

Southern California Bight 2013 Targeted CEC Survey

A Bight '13 Special Study was implemented to address Scenario 3 monitoring. This study is intended as a pilot project, and future surveys may be modified based on the results of this initial monitoring. The design addresses Scenario 3 questions regarding marine outfall discharge, as also compares marine outfall receiving stations with storm water receiving stations. All samples are sediments.

Aim 1. Compare CEC sediment concentrations impacted by the three sources (marine outfalls, storm water, and inland waste water). Only marine outfall zone-of-initial-dilution (ZID) stations will be used for this purpose. Outfall contaminant concentrations are expected to be highest in the ZID and are potentially more variable than stations further out. To account for this potential variability, three sub-stations within the ZID were be sampled, and the composite will be analyzed as a single sample.

Aim 2. Verify CECs originate from the outfalls and are not simply at background concentrations. Decreasing CEC concentrations down-current away from the outfall will indicate the compounds originate at the outfall. Also, stations up current (presumably at background), and cross-current station will indicated if the outfall is the source. Outfall stations were assigned in consultation with the dischargers and based on 1) the predominant current direction throughout the year, and 2) spatial trends of legacy contamination. The main gradient direction relative to the outfall varied among locations. For example, the LACSD outfall is perpendicular to the current in that region, but the OCSD outfall is parallel the current. The selected station distance is expected to show a decrease in CEC concentrations away from the outfall, based on legacy data.

Target Compounds

The four analyte classes are alkylphenols (APs), perfluorinated compounds (PFCs), pyrethroids/fipronil, and polybrominated diphenyl ethers (PBDEs). They will be measured at all stations in the survey. Phthalates, recommended by the Panel for Scenario 3 monitoring, will not be measured due to resource limitations.

Survey Design

Fifteen river-mouth samples throughout southern CA were obtained as part of the regular Bight '13 sediment survey (sampled July – September 2013). There was 1 station per river-mouth. Ten stations receive storm water and 5 receive both storm water and waste water discharge.

The 5 outfalls were City of LA Hyperion (CLA), LA County Sanitation District's outfall off Palos Verdes (LACSD), Orange County Sanitation District (OCSD), and the two City of San Diego (CSD) outfalls Point Loma and South Bay. There are 5 stations at each outfall, and three sub-stations within the ZID station. Samples were collected in January 2014.

Relationship to the Panel's original marine outfall design. For this pilot survey we expanded the number of outfalls from 2 in the original design to 5. This required a reduction in the number of stations per outfall from 7 to 5. Increasing the number of outfalls provides more ZID stations for comparison to the river-mouth concentrations (see Aim 1), and provides information on CEC occurrence at all major ocean outfalls in the region.

8.4 Appendix D: Summary of RMP CEC Investigations

San Francisco Bay RMP CEC Monitoring Activities: Receiving Waters, Sediment, Tissue

Compound*	SWRCB Panel Guidance: Embayments	RMP SF Bay Risk Tier (1)	RMP Status & Trends Monitoring	RMP Approach	RMP References for Existing Bay Data
Flame Retardants					
Alternative (non-PBDE) Flame Retardants	not evaluated	Possible (I)		2014 Special Study to build upon previous special studies, other data detecting flame retardants in ambient water (phosphates, qualitative), sediment and biota.	1-4
PBDEs (BDE-47 and 99)	sediment, tissue	Moderate (III)	sediment, tissue (bivalves, sport fish, bird eggs); water discontinued	Analyzed extensively in water, sediment and tissue. Concentrations declining in multiple species and sediment. Prepared summary report on ten years of RMP data.	1,5
Hormones					
17-beta estradiol	water			No Bay data. Bioanalytical tools project will characterize single receiving water sample.	6
Estrone	water			No Bay data. Bioanalytical tools project will characterize single receiving water sample.	6
Pesticides					
Bifenthrin (Pyrethroid)	water, sediment	Low (II)	sediment	Hydrophobic; based on Bay sediment concentrations, expect ND in water.	1
Fipronil	water, sediment	Moderate (III)	sediment	ND in pilot water study; continue sediment monitoring.	1
Permethrin (Pyrethroid)	water, sediment	Low (II)	sediment	Hydrophobic; based on Bay sediment concentrations, expect ND in water.	1

Compound*	SWRCB Panel Guidance: Embayments	RMP SF Bay Risk Tier (1)	RMP Status & Trends Monitoring	RMP Approach	RMP References for Existing Bay Data
PPCPs & Plastic Additives					
Bis(2-ethylhexyl) phthalate (DEHP)	NA	Possible (I)		Widely detected at low level in surface water, tissue and sediment. Below available effects thresholds for sediment. Uncertainty regarding the applicability of thresholds to Bay data.	1
Bisphenol A	water	Possible (I)		ND samples; DL high. Bioanalytical tools project will characterize single receiving water sample. Draft RMP review of potential PPCP targets suggests this analyte may be appropriate for future special studies.	1,6,7
Butylbenzyl phthalate	NA	Possible (I)		Exceed low apparent effects threshold values in sediment but high uncertainty regarding the application of these thresholds to the Bay. ND in mussel tissue. Draft RMP review of potential PPCP targets suggests this analyte may be appropriate for future special studies.	1,7
Diclofenac	NA			No Bay data. Draft RMP review of potential PPCP targets suggests this analyte is unlikely to be a concern in the Bay.	7
Galaxolide (HHCB)	water	Low (II)		Detected at low levels in Bay samples from 1999-2000 and in later Bay POCIS passive sampling study. Bioanalytical tools project will characterize single receiving water sample. Draft RMP review of potential PPCP targets suggests this analyte is unlikely to be a concern in the Bay.	1,6,7
Ibuprofen	NA	Low (II)		Mostly ND in pilot studies.	1,8,9

Compound*	SWRCB Panel Guidance: Embayments	RMP SF Bay Risk Tier (1)	RMP Status & Trends Monitoring	RMP Approach	RMP References for Existing Bay Data
p-Nonylphenol	NA	Moderate (III)		Detected in water, sediment and tissue. Bioanalytical tools project will characterize single receiving water sample.	1,6,9,10
Triclosan	NA	Low (II)		Low to ND in sediment. ND in water and mussels.	1,11
PFASs					
PFOS	sediment, tissue	Moderate (III)	tissue (sport fish, bird eggs)	Detected in elevated concentrations in seals and bird eggs. Continue monitoring in tissue. Other studies have detected PFOS in Bay sediment; RMP will consider monitoring this matrix.	1,12,13

San Francisco Bay RMP CEC Monitoring Activities: WWTP Effluent

Compound*	SWRCB Panel Guidance: WWTP Effluent	RMP SF Bay Risk Tier (1)	RMP Status & Trends Monitoring	RMP Approach	RMP References for Existing Bay Data
Flame Retardants					
Alternative (non-PBDE) Flame Retardants	not evaluated	Possible (I)		2014 Special Study to characterize three effluent samples. TCEP detected in effluent from single POTW in past study; phosphates detected in biosolids.	1,3,4,14
PBDEs (BDE-47 and 99)	effluent	Moderate (III)		Effluent discharges have been characterized in the past. Declining concentrations in Bay; not a high priority for monitoring given use restrictions.	1,5,15
Hormones					
17-beta estradiol	effluent			No Bay data. Bioanalytical tools project will characterize single effluent sample.	6
Estrone	effluent			Detection in single POTW effluent. Bioanalytical tools project will characterize single effluent sample.	6,16
Pesticides					
Bifenthrin (Pyrethroid)	effluent	Low (II)		Effluents from 32 facilities have been monitored for pyrethroids.	1,17
Fipronil	NA	Moderate (III)		2015 Special Study proposal to characterize up to eight effluents.	1,18
Permethrin (Pyrethroid)	effluent	Low (II)		Effluents from 32 facilities have been monitored for pyrethroids.	1,17
PPCPs & Plastic Additives					
Bis(2-ethylhexyl) phthalate (DEHP)	NA	Possible (I)		Detected in effluent from single POTW in past study.	14

Compound*	SWRCB Panel Guidance: WWTP Effluent	RMP SF Bay Risk Tier (1)	RMP Status & Trends Monitoring	RMP Approach	RMP References for Existing Bay Data
Bisphenol A	effluent	Possible (I)		Detected in effluent from single POTW in past study. Draft RMP review of potential PPCP targets suggests this analyte may be appropriate for future special studies.	7,14
Butylbenzyl phthalate	NA	Possible (I)		Detected in effluent from single POTW in past study. Draft RMP review of potential PPCP targets suggests this analyte may be appropriate for future special studies.	7,14
Diclofenac	NA			No Bay effluent data. Draft RMP review of potential PPCP targets suggests this analyte is unlikely to be a concern in the Bay.	7
Galaxolide (HHCB)	effluent	Low (II)		No Bay effluent data. Bioanalytical tools project will characterize single effluent sample. Draft RMP review of potential PPCP targets suggests this analyte is unlikely to be a concern in the Bay.	1,6,7
Ibuprofen	NA	Low (II)		Not detected in one pilot study, detected in another.	1,8,16
p-Nonylphenol	NA	Moderate (III)		Not detected in effluent from single POTW in past study; ethoxylates may be better targets. Bioanalytical tools project will characterize single effluent sample.	6,14
Triclosan	NA	Low (II)		Detected in effluent from two POTWs in past studies.	14,16
PFASs					
PFOS	effluent	Moderate (III)		2015 Special Study proposal to characterize up to eight effluents.	1,18

*Chlorpyrifos not included in monitoring - see SWRCB Panel September 2013 meeting notes and rationale.

San Francisco Bay RMP CEC Monitoring Activities: Urban Creeks (Stormwater)

Compound*	SWRCB Panel Guidance: Receiving Water	RMP SF Bay Risk Tier (1)	RMP Status & Trends Monitoring	RMP Approach	RMP References for Existing Bay Data
Flame Retardants					
Alternative (non-PBDE) Flame Retardants	not evaluated	Possible (I)		2014 Special Study to characterize stormwater discharges from two sites.	4
PBDEs (BDE-47 and 99)	stormwater	Moderate (III)	stormwater	Ongoing monitoring in stormwater from a variety of sites.	1,5
Hormones					
17-beta estradiol	stormwater			No Bay stormwater data.	
Estrone	stormwater			No Bay stormwater data.	
Pesticides					
Bifenthrin (Pyrethroid)	stormwater	High (IV)**	stormwater	Ongoing monitoring in stormwater from a variety of sites.	1
Fipronil	stormwater	Moderate (III)	stormwater	Ongoing monitoring in stormwater from a variety of sites.	1
Permethrin (Pyrethroid)	stormwater	High (IV)**	stormwater	Ongoing monitoring in stormwater from a variety of sites.	1
PPCPs & Plastic Additives					
Bis(2-ethylhexyl) phthalate (DEHP)	NA	Possible (I)		No Bay stormwater data.	
Bisphenol A	stormwater	Possible (I)		Detected in 3/4 samples; unpublished data.	
Butylbenzyl phthalate	NA	Possible (I)		No Bay stormwater data.	
Diclofenac	stormwater			Detected in four samples; unpublished data.	
Galaxolide (HHCB)	stormwater	Low (II)		No Bay stormwater data.	

Compound*	SWRCB Panel Guidance: Receiving Water	RMP SF Bay Risk Tier (1)	RMP Status & Trends Monitoring	RMP Approach	RMP References for Existing Bay Data
Ibuprofen	stormwater	Low (II)		Detected in 3/4 samples; unpublished data.	
p-Nonylphenol	NA	Moderate (III)		No Bay stormwater data.	
Triclosan	stormwater	Low (II)		Not detected in four samples; unpublished data.	
PFASs					
PFOS	stormwater	Moderate (III)		Past monitoring data available.	19

*Chlorpyrifos not included in monitoring - see SWRCB Panel September 2013 meeting notes and rationale.

**Classified as High Concern for Bay tributaries, but Low Concern for ambient Bay water - see RMP. 2013. Pulse of the Bay: Contaminants of Emerging Concern. A Report of the Regional Monitoring Program for Water Quality in San Francisco Bay.

9 REFERENCES

(1) Klosterhaus, S., Yee D., Sedlak, M., Wong, A., Sutton, R. 2013a. Contaminants of Emerging Concern in San Francisco Bay: A Summary of Occurrence Data and Identification of Data Gaps. RMP Contribution 698. San Francisco Estuary Institute, Richmond, CA. 121 pp.

(2) Klosterhaus, S.L., H.M. Stapleton, M.J. La Guardia, D.J. Greig. 2012a. Brominated and chlorinated flame retardants in San Francisco Bay sediments and wildlife. Environment International, 47, 56-65.

(3) Klosterhaus, S.L., Davis, E.F., H.M. Stapleton. 2009. Organophosphate flame retardants in San Francisco Bay biosolids and sediments. Society for Environmental Toxicology and Chemistry North America Conference 2009. New Orleans, Louisiana.

(4) Sutton R., Sedlak M. 2013. Monitoring alternative flame retardants in SF Bay water, sediment, and biota. Proposal. June 2013. *and* Sutton R., Sedlak M. 2013. Monitoring alternative flame retardants in SF Bay water, sediment, and biota: Pathway characerization – wastewater and stormwater. Proposal addendum requested by the RMP Steering Committee. September 2013.

(5) Sutton, R., Sedlak, M., Davis, J. (2014). Polybrominated Diphenyl Ethers (PBDEs) in San Francisco Bay: A Summary of Occurrence and Trends. RMP Contribution No. 713. San Francisco Estuary Institute, Richmond, California. 62pp.

(6) Denslow N., Maruya K., Bay S. 2012. Linkage of In Vitro Assay Results with In Vivo End Points. Proposal for the RMP Emerging Contaminants Workgroup.

(7) Eastman, R. 2013. RMP PPCP Review 2013. Draft.

(8) Harrold, K.H., D. Yee, M. Sedlak, S.L. Klosterhaus, J.A. Davis, M. Woudneh, P. Riley. 2009. Pharmaceuticals and Personal Care Products in Wastewater Treatment Plant Influent and Effluent and Surface Waters of Lower South San Francisco Bay. San Francisco Estuary Institute. Oakland, CA.

(9) Klosterhaus, S., Grace, R., Hamilton, C., D. Yee. 2013b. Method validation and reconnaissance of pharmaceuticals, personal care products, and alkylphenols in surface waters, sediments, and mussels in an urban estuary. Environment International, 54, 92-99.

(10) Klosterhaus, S., R.M. Allen, J.A. Davis. 2012b. Contaminants of Emerging Concern in the San Francisco Estuary: Alkylphenol Ethoxylates. RMP Contribution #657. San Francisco Estuary Institute. Richmond, CA.

(11) Klosterhaus, S., R. Allen, J. Davis. 2011. Contaminants of EmergingConcern in the San Francisco Estuary: Triclosan and Triclocarban. A Report of the Regional Monitoring Program for Water Quality in the San Francisco Estuary. SFEI Contribution #627. Final Report. San Francisco Estuary Institute, Oakland, CA.

(12) Sedlak, M.D., D.J. Greig. 2012. Perfluoroalkyl compounds (PFCs) in wildlife from an urban estuary. Journal of Environmental Monitoring, 14, 146-154.

(13) Higgins, C.P., J.A. Field, C.S. Criddle, R.G. Luthy. 2005. Quantitative determination of perfluorochemicals in sediments and domestic sludge. Environmental Science and Technology, 39, 3946-3956.

(14) Jackson J., Sutton, R. 2008. Sources of endocrine-disrupting chemicals in urban wastewater, Oakland, CA. Science of the Total Environment, 405, 153-160.

(15) North, K. 2004. Tracking polybrominated diphenyl ether releases in a wastewater treatment plant effluent, Palo Alto, California. Environmental Science and Technology, 28, 4484-4488.

(16) Dunlavey E., Ervin J., Tucker D. (2010) Environmental Fate and Transport of Microconstituents. Water Environment and Technology 22, 2-5.

(17) Markle, J.C., van Buuren, B.H., Moran, K.D., Barefoot, A.C. 2014. Pyrethroid pesticides in municipal wastewater: A baseline survey of publicly owned treatment works facilities in California in 2013. https://bacwa.app.box.com/s/jxhrd2lte3o1aquy7abf

(18) Sutton R., Sedlak M. 2014. Evaluating emerging contaminant pathways: Wastewater discharges. Proposal. April 2014.

(19) Houtz, E.F., Sedlak, D.L. 2012. Oxidative conversion as a means of detecting precursors to perfluoroalkyl acids in urban runoff. Environmental Science and Technology, 46, 9342-9349.

10 QA/QC GUIDANCE

10.1 Introduction

In 2009, the State of California Water Resources Control Board (SWRCB) tasked a scientific advisory panel ("Panel") to assess current scientific knowledge of the risks posed by CECs to freshwater, coastal and marine ecosystems, and to provide recommendations for CEC monitoring that will protect beneficial uses in these ecosystems. In their final report, the Panel utilized a risk-based screening framework to identify a list of CECs for monitoring in three representative receiving water scenarios, and recommended development of better CEC monitoring and assessment tools, including bioanalytical screening methods (Anderson et al. 2012).

In response to these recommendations, SWRCB staff tasked the Southern California Coastal Water Research Project Authority (SCCWRP) to generate a study plan to perform pilot monitoring of CECs statewide. The major elements of this pilot investigation are to (1) measure occurrence of CECs identified by the Panel in source and receiving waters and in appropriate matrices (i.e. discharged wastewater treatment plant (WWTP) effluent, waters receiving WWTP effluent and stormwater runoff, sediment, and/or tissue) and (2) evaluate alternative monitoring methods, including bioanalytical screening tools and whole organism toxicity tests that better target biological responses associated with CECs. A full description of the study plan elements is documented elsewhere (SCCWRP 2015).

By definition, CECs are not widely regulated and thus not routinely monitored. As a result, there is a likelihood of larger variation in data quality among laboratories, since available analytical methods may not be as robust as for historical (priority) pollutants. Statewide monitoring will include participation by multiple agencies, field crews, and laboratories; therefore, ensuring that results are comparable among different groups by maintaining consistency in field and laboratory operations is critical to success.

10.1.1 Scope

Since integrated statewide CEC pilot monitoring is not currently in the implementation phase, the level of detail available at the time of writing fall short of the information required in a Quality Assurance Project Plan (QAPP). In lieu of a QAPP, this document describes currently available QA/QC related information which should be used as guidance in generating a QAPP when the appropriate level of detail is made available for project implementation. A description of the necessary information is included in Section 10.7.

10.1.2 Objectives

The goal of this document is to ensure data quality and comparability among participating agencies, field crews, and laboratories, and to ensure data can confidently be compared to other surveys. Ensuring data quality consists of two distinct but related activities: quality assurance and quality control.

Quality assurance (QA) includes design, planning, and management actions conducted prior to field sampling to ensure appropriate types and quantities of data are collected. The goals of QA are to ensure that: 1) sample transport and processing, and laboratory analytical techniques will be applied consistently and correctly; 2) the number of lost, damaged, and uncollected samples will be minimized; 3) the integrity of the data will be maintained and documented from sample collection to entry into the data record; 4) data will be comparable; and 5) measurements can be reproduced. This will be achieved by:

1. Evaluation of laboratories' ability to conduct the analyses based on prior data, and the establishment of reporting levels (RLs),
2. Development of the project quality control procedures described below,
3. Evaluation of the comparability of analytical and bioassay methods through inter-laboratory evaluations, and
4. Development of a data management plan.

Quality control (QC) activities are implemented during the data collection phase of the project to evaluate the effectiveness of the QA procedures. These activities ensure that measurement error and bias are identified, quantified, and either accounted for or eliminated. This will be achieved by:

1. Standard procedures for sample collection and recording of field observations,
2. Standard procedures for sample shipment and storage, and
3. Adherence to a common set of measurement quality objectives (MQOs). The MOQ defines acceptance criteria based on calibration of the instrument, evaluation of blank concentrations, repeated measurements to establish method precision, and use of test samples to establish method accuracy.

10.2 Sample Collection, Handling and Preservation

Field personnel must strictly adhere to established protocols to insure the collection of representative, uncontaminated pilot study samples. Guidelines for sample storage are provided in **Table 10.2-1**. Changes and/or additions to these guidelines may be proposed by project participants if proper justification is provided.

- Field personnel must be thoroughly trained
    - in the proper use of sample collection gear,
    - in distinguishing acceptable versus unacceptable samples in accordance with pre-established criteria,
    - to recognize and avoid potential sources of sample contamination.

- Sampling equipment and utensils that come in direct contact with the sample should be made of non-contaminating materials and should be thoroughly cleaned between sampling stations.
- Sample storage containers should be of the recommended type and must be free of contaminants.
- Conditions for sample collection, preservation and holding times should be followed, and relevant field observations should be recorded.

On the day of sampling, field personnel should avoid contact with or consumption of products that contain the target analytes. This may include soaps, detergents, fragrances, sunscreen, and pharmaceuticals. Storage containers with Teflon should not be used to store samples that are slated for analysis of perfluorinated compounds (PFCs).

Table 10.2-1. Sample collection and holding time conditions.

Matrix	Container Type	Container size (mL)	Preservation Requirements	Maximum Holding Time
Aqueous	Pre-cleaned amber glass	1000 (100% full)	Cold (4 °C), with preservative added as required	2 weeks
Sediment	Pre-cleaned amber glass	250 or 125 (80% full)	Frozen (-20 °C)	1 year
Tissue	Pre-cleaned amber glass	250 or 125 (80% full)	Frozen (-20 °C)	1 year

10.3 Laboratory Documentation of General Practices

All laboratories performing measurement of parameters specified in the pilot study plan (SCCWRP 2015) and as delineated in Sections 10.4-10.6 herein must have the appropriate facilities to store and prepare samples, and appropriate instrumentation and staff to provide data of the required quality within the specified time period. Laboratories are expected to conduct operations using good laboratory practices, including:

- A program of scheduled maintenance of analytical balances, laboratory equipment, static and flow through exposure apparatuses, and instrumentation.
- Checking and recording the composition of fresh calibration standards against the previous lot.
- Checking and recording of water or sediment quality parameters in toxicity tests.
- Monitoring and recording temperatures within exposure rooms, storage areas and freezer units.
- Acquisition of solvents, test cell lines/kits and other consumables of suitable quality.
- Dating and storing all samples safely upon receipt and use of a laboratory information management system to track the location and status of any sample.

Personnel shall be well versed in good laboratory practices, including standard safety procedures. It is the responsibility of the laboratory to ensure that safety training is mandatory for all personnel. The laboratory is responsible for maintaining a current safety manual in compliance with the Occupational Safety and Health Administration (OSHA), or equivalent state or local regulations. The safety manual should be readily available to laboratory personnel. Best safety practices should be followed at all times, including proper storage, handling, and disposal of chemicals; verification of fume hood operation; and use of supplies/equipment to prevent potential health hazards.

Laboratories shall be able to provide documentation of their ability to conduct analyses with the level of data quality specified herein. Specifically, the following documents and information must be available upon request:

- QA Plan: Policies and protocols specific to a particular laboratory including personnel responsibilities, procedures for determining the acceptability of results, and procedures for release of the data.
- Standard Operating Procedures (SOPs): Step-by-step instructions describing in detail implementation of the method, specific for the particular equipment and instruments used.
- Instrument performance information: Laboratories should collect ongoing data on instrument baseline noise, calibration standard response, detection limits, and laboratory blanks.

10.4 Analysis of Chemical Contaminants

10.4.1 General Approach

A performance-based approach to QA/QC is recommended. In this format, specific analytical methods are not prescribed, rather each laboratory may use methods of their choice as long as QA/QC requirement are met and acceptable performance is demonstrated. For CECs in particular, mass spectrometry based methods shall be used; e.g., gas chromatography/mass spectrometry (GC/MS) or liquid chromatography/tandem mass spectrometry (LC-MS/MS). Also, these methods shall employ spiked surrogate or internal standards to generate calibration curves. Standard addition methods shall not be used. Detailed criteria based on QA/QC guidelines adopted by the Southern California Bight Program (SCCWRP 2013), the Surface Water Ambient Monitoring Program (SWAMP 2008) and the U.S. Geological Survey (USGS 2004), are described in the following subsections:

Sec 10.4.2 Target, matrix and scenario specific reporting limits (RLs)

Sec 10.4.3 Performance in inter-laboratory comparison exercises

Sec 10.4.4 Sample completeness

Sec 10.4.5 Measurement quality objectives (MQOs)

10.4.2 Reporting Limits (RLs)

Recommended reporting limits (RLs) for pilot study CECs were set at 50% of monitoring trigger levels (MTLs) established by the Panel (Anderson et al. 2012) in order to allow for the collection of data that will be useful in evaluating CEC risk (SCCWRP 2015). These RLs are specified for each target compound (i.e. CEC), matrix and scenario, and thus may differ among scenarios (**Table 10.4-1**). In some cases, the Panel recommended RL is lower than what commercial services labs currently offer. As methods continue to improve and evolve, participating labs shall strive to achieve the recommended RLs, and shall in all cases meet the minimum achievable RLs.

10.4.3 Inter-Laboratory Comparison Exercise

All laboratories contributing analytical chemistry data for the pilot study shall participate in an inter-laboratory exercise to demonstrate comparability with all participants, including those considered as referee labs. The recent advent of commercially available services for many of the target CECs, coupled with the extremely low RLs required, necessitate an assessment of data comparability among participating labs. The inter-laboratory comparison will provide an opportunity to revise project MQOs, if warranted, based on group consensus. Additional value in participating in inter-laboratory exercises are: 1) laboratories not passing minimum performance criteria are made aware of methodological issues and can work with referee labs to resolve these issues, and 2) a quantitative assessment of among-laboratory variability will provide context for managers when comparing results to other CEC-related projects.

Table 10.4-1. Monitoring trigger levels (MTLs) and reporting limits (RLs) by scenario, compound and matrix. Recommended RLs are derived from MTLs as reported by the CEC Science Advisory Panel. Achievable RLs reflect the current state of art for commercial services laboratories. Recommended RLs for all CECs in wastewater treatment plant (WWTP) effluent and stormwater (MS4) influenced receiving waters are equivalent to Scenario 1 aqueous phase RLs; additional RLs for compounds that are otherwise measured only in sediment or tissues appear at the bottom of the table.

Compound	Panel Freshwater MTL[1]	Recommended RL[2]	Achievable RL[3]
Aqueous Phase - Effluent dominated inland waterways (Scenario 1) (ng/L)			
Bifenthrin[4]	0.40	0.20	
Permethrin[4]	1.0	0.50	
Fipronil[4]	42	21	
Chlorpyrifos[4]	5.0	2.5	
Estrone	6.0	3.0	
Ibuprofen	100	50	
Bisphenol A	60	30	
17-beta-estradiol	2.0	1.0	
Galaxolide (HHCB)	700	350	
Diclofenac	100	50	
Triclosan	250	125	
Sediment Phase - Effluent dominated inland waterways (Scenario 1) (ng/g dw)			
Fipronil	0.090	0.045	1.0
Aqueous Phase - Coastal embayments (Scenario 2) (ng/L)			
Bisphenol A	6.0	3.0	
Bifenthrin	0.040	0.020	0.2
Permethrin	0.10	0.050	0.5
Fipronil	5.0	2.5	
Chlorpyrifos	1.0	0.50	
Estrone	0.60	0.30	2.0
17-beta-estradiol	0.20	0.10	0.4
Galaxolide (HHCB)	70	35	
Sediment - Coastal embayments (Scenario 2) (ng/g dw)			
Bifenthrin	0.052	0.026	0.20
PBDE-47	0.030	0.015	
PBDE-99	0.030	0.015	
Permethrin	0.073	0.036	0.40
Fipronil	6.5	3.25	
PFOS[5]	NA	0.1	

Sediment - Ocean discharge (Scenario 3) (ng/g dw)		
Bis(2-ethylhexyl) phthalate (BEHP)	130	65
p-nonylphenol	14	7.0
PBDE-47	0.30	0.15
PBDE-99	0.30	0.15
Butylbenzyl phthalate (BBP)	6.3	3.15
PFOS[5]	NA	0.1
Tissues (All Scenarios) (ng/g dw)		
PBDE-47	28.9	14.5
PBDE-99	28.9	14.5
PFOS	1000	500
WWTP Effluent and MS4 Receiving Water (ng/L) [6]		
Bis(2-ethylhexyl) phthalate (BEHP)		3.0
Butylbenzyl phthalate (BBP)		3.0
p-nonylphenol		22 [7]
PBDE-47		0.10
PBDE-99		0.10
PFOS		1.0

[1] Monitoring Trigger Level established by CEC Science Advisory Panel (Anderson et al. 2012).
[2] Set at 50% of MTL.
[3] Minimum RL reported by commercial services laboratories. Missing values indicate the achievable value is at or below the recommended RL.
[4] Scenario 1 pesticides are currently monitored by other programs. The recommended RLs are listed here for comparison purposes only.
[5] PFOS was recommended for Scenario 2 and 3 sediment monitoring to obtain information on sediment-biota transfer, not based on MTLs. The recommended RL was based on typical values observed in the literature and attainable values by laboratories.
[6] RLs for analytes otherwise measured in sediment or tissues only (no MTL values available). For all other analytes, RLs for WWTP Effluent and MS4 receiving water samples are the same as the aqueous RLs for Scenario 1.
[7] Estimated from the sediment RL (7.0 ng/g), an estimated sediment-water partitioning coefficient, and assuming 1% organic carbon content of the sediment.

A referee laboratory will be assigned to prepare reference materials representing the matrix and target analytes of interest (**Table 10.4-2**). Either materials with native levels of target CECs or representative matrices spiked with target CECs at concentrations at or above RLs will be used as reference materials. After division of the spiked reference material into multiple aliquots, the referee laboratory should verify the concentrations of the target analytes and establish sample homogeneity through within jar and between jar analyses. Participating laboratories should not have prior knowledge of target CEC concentrations, and should make repeated (e.g., triplicate) measurements of the reference material to assess within-laboratory variability. Standard reference materials (SRMs) that contain target CECs or analogs thereof, if available, may also be analyzed to test accuracy of methods employed for the reference material. The exact performance criteria should be decided by project participants based on their measurement knowledge for each analyte. However, laboratories should be assessed by comparing their results to a "target" value (e.g. ± 40% the group mean).

Table 10.4-2. Inter-laboratory comparison reference materials.

Reference Material	Covers Scenario
Freshwater	Scenario 1, Scenario 2, and Stormwater (MS4)
Effluent	WWTP effluent
Sediment	Scenario 2 and Scenario 3
Tissue	All scenarios

10.4.4 Completeness

Completeness is defined as the proportion of samples that are successfully collected, analyzed and that pass quality control (QC) validation. Losses may occur as a result of field conditions, logistical difficulties, or failure to achieve QC criteria. The MQO for completeness is 90% for each analyte. To achieve this criteria, the sampling design for the pilot study shall be sufficiently redundant to absorb the loss of up to 10% of the samples/analytes without compromising the pilot study goals, provided that the losses are not concentrated in a single subpopulation of interest.

10.4.5 Measurement Quality Objectives (MQOs)

The measurement quality objectives (MQOs) delineated in the following sections are intended to provide a common foundation for laboratory performance and should be considered as the minimum requirements for analyzing CECs in pilot study samples. Additional MQOs may be instituted by participating labs, as long as the MQOs presented herein are satisfied. Aqueous sample concentrations shall be reported using specific units (e.g., ng/L). Sediment sample concentrations shall be reported on a dry weight basis with the percent moisture of the corresponding sample also reported. Tissue sample concentrations should be reported on a wet weight basis, with percent moisture and percent lipid of the corresponding sample also reported. The methods for measuring percent moisture and percent lipids should be standardized among the participating laboratories.

10.4.5.1 Measurement Range and Sensitivity

Prior to the commencement of sample analysis, each laboratory should establish the working calibration range, and determine nominal method detection and reporting limits (MDLs and RLs, respectively) on an analyte- and matrix-specific basis. These steps are detailed in the following sections.

Calibration Range

The working calibration range for each target CEC must be established using a minimum of five concentrations, and acceptable performance should be demonstrated on an accuracy-based material (e.g., reference material described in 10.4.3). Only data resulting from quantification within the working calibration range may be reported by a laboratory without annotation. Samples with measured concentrations above or below the calibration range should be reanalyzed using appropriate sample mass and/or volume.

Reporting Level

The RL is the minimum concentration that can be reliably measured, and is also the minimum target concentration at which laboratories shall report data. By default, the RL is the lowest concentration in the calibration curve. If an alternate definition for RL is used, this shall be submitted to and approved by the Data Management Team (Section 10.7) prior to sample analysis.

Method Detection Limits (MDLs)

The method detection limit (MDL) represents a quantitative estimate of low-level response detected at the maximum sensitivity of a method. This level should be a concentration below the RL, and laboratories should describe the method they used to determine the MDL.

10.4.5.2 Ongoing Measurement Objectives

Following a successful setup phase, each laboratory must demonstrate maintenance of performance by repeating analysis of QC samples within each analytical batch. Descriptions of the QC samples are in the following sections, with the corresponding MQOs in **Table 10.4-3**. If control limits for any objective are not met, the laboratory shall take action to find and eliminate the problem before continuing with sample analysis. If a major unresolvable flaw is found, it may be necessary to repeat the analysis of the affected batch of samples.

Based on laboratory participant and project management consensus and the results of the inter-calibration exercise, it may be necessary to revise the MQOs for specific analytes prior to the collection of field data. The MQO criteria listed here should be viewed as a starting point for discussion with participation laboratories.

Initial Calibration and Continuing Calibration Verification

A new response factor or calibration curve should be established for each instrumental batch. A continuing calibration verification standard shall be analyzed at specified intervals (every 10 samples or 8 hours) to monitor temporal variability in the instrument. The continuing calibration verification standard should be at the mid-range calibration concentration, and must be within ±20% of the initial calibration response. An instrument blank should be included in the calibration curve to verify that the instrument is free of contamination or carryover.

Method Blanks

Method blanks assess laboratory contamination during sample preparation and analysis. One method blank should be run in each sample preparation batch, and it should be processed and analyzed using the same protocol used for samples. Blanks exceeding the MQO require corrective action to bring subsequent blanks into compliance. This may involve performing equipment maintenance, changing reagents and/or, as a last resort, modifying SOPs. Although acceptable laboratory blanks are important, improvements in analytical sensitivity and the pervasiveness of some contaminants result in situations where detection in laboratory blanks is unavoidable. The magnitude of the blank concentrations must be evaluated against the sample concentrations and the MQOs (see Table 10.4-3). Blank subtraction is allowed if the blank concentration is < 30% of the analyte concentration in the same batch.

Sample Duplicates

Analysis of sample duplicates is used to assess the precision of an analytical method and to check for sample heterogeneity. At least one sample per batch of 20 samples should be analyzed in duplicate (Table 10.4-3).

Matrix Spikes and Matrix Spike Duplicates

Matrix spike and matrix spike duplicates (MS/MSD) are laboratory-prepared samples of the matrix spiked with known levels of the target analytes, used to evaluate the effect of the sample matrix on analyte recovery, and additionally, to provide an estimate of analytical precision. The material to be spiked should represent the matrix of interest, i.e. be as similar as possible to the sample being analyzed. A minimum of one MS/MSD pair should be analyzed for every batch of 20 samples. The matrix spike solution should contain all the analytes of interest. The final spiked concentration of each analyte should be at least 3 times the RL. If the unspiked matrix contains background concentrations of any target analyte, the sample should be spiked with one to five times the preexisting concentration in the sample. Acceptance criteria for recovery of spiked analytes are provided in Table 10.4-3.

Standard Reference Materials or Laboratory Control Samples

Method accuracy is evaluated through the analysis of standard reference materials (SRMs) or laboratory control samples (LCS). Analyses of SRMs must yield values within the specified range of the certified (or reference) values provided by the supplier. Certified values have lower uncertainty than reference values, but in the absence of certified values, reference values are acceptable for assessment of accuracy. Due to the inherent variability in analyses near the MDL, criteria for accuracy will only apply to analytes having certified values that are >3 times the RL established by the laboratory. If a SRM for all target analytes is unavailable, an LCS can be substituted. An LCS is prepared by the laboratory using contaminant free water or an appropriate inert solid material spiked with the target analyte at a known concentration within the calibration curve. A minimum of one SRM or LCS should be analyzed per batch of 20 samples. Acceptance criteria for accuracy of target CECs are provided in Table 10.4-3.

Standards and Standard Recovery

Quantification standards are isotope-labeled or structurally similar analogs to the target analytes. Laboratories may refer to them as internal-, surrogate-, and/or isotope dilution standards, but the

exact definition of these terms is inconsistently applied in the literature. These standards are used to generate calibration curves and are added at known levels to field samples to monitor and adjust for extraction efficiency, sample losses, retention time shifts, instrumental drift, and ion suppression. The percent recovery of standards added prior to extraction and accounting for extraction and sample losses must be within control limits specified in Table 10.4-3.

Table 10.4-3. Ongoing Measurement Quality Objectives (MQOs) for target analytes in all matrices.

Measurement	Frequency	Control Limit
Initial Calibration	A new response factor or calibration curve should be established for each instrumental batch.	Relative standard deviation (RSD) of the response factor ≤ 25% Coefficient of determination r^2 ≥ 0.990 for linear and non-linear curves. First or second order curves allowed. Minimum of 5 points per curve.
Continuing Calibration Verification	Every 10 samples or 8 hours	Expected concentration ± 20%.
Method Blank	5% of total no. samples (1 per batch of 20 samples)	Less than the RL for target analytes.
Sample Duplicate	5% of total no. samples (1 per batch of 20 samples)	RPD ≤ 35%.
Certified Reference Material or Laboratory Control Sample	5% of total no. samples (1 per batch of 20 samples)	70-130% recovery if certified; otherwise, 50-150% recovery.
Matrix Spike/Matrix Spike Duplicate Pair	5% of total no. samples (1 per batch of 20 samples)	50-150% or based on historical laboratory control limits; RPD ≤ 25%.
Spiked Standard Recovery	All field and QC samples	50-150% or based on historical laboratory control limits.

10.5 Biological Testing

10.5.1 Bioanalytical Screening Tools

10.5.1.1 General Approach

The QA/QC criteria for these new monitoring tools were based on technical reports from EPA's Endocrine Disruptor Screening Program (USEPA 2013), and recently completed research projects on adapting in vitro bioassays (IVBs) for water quality screening (SCCWRP 2014; WRRF 2014). A performance-based approach is adopted where each laboratory may use their method of choice. General requirements are described in the following subsections:

Sec 10.5.1.2 In vitro bioassay (IVB) endpoints

Sec 10.5.1.3 Selection of reference toxicants

Sec 10.5.1.4 Measurement quality objectives (MQOs)

Sec 10.5.1.5 Performance in inter-laboratory comparison exercises

10.5.1.2 In Vitro Bioassay (IVB) Endpoints

Cellular (*in vitro*) bioassays will be used to screen chemicals and to determine their potential toxic effects. These tools will be applied for all four scenarios using water, sediment and tissue samples. The IVB endpoints described in the pilot study plan (SCCWRP 2015) and listed in **Table 10.5-1** can screen for endocrine disrupting chemicals (e.g. estrogens, androgens, progestins and glucocorticoid steroids) as well as dioxin-like chemicals.

Commercial Suppliers

In vitro bioassays selected for CEC monitoring are all commercially available. The existing suppliers are specified in Table 5.1.

Table 10.5-1. Recommended commercial suppliers for in vitro biossays (IVBs).

Endpoints	Bioassay, Supplier
Estrogen Receptor (ER)	GeneBLAzer ERα Division Arrested Assay, Life Technologies[1] ERα CALUX, BioDetection Systems[2]
Androgen Receptor (AR)	GeneBLAzer AR Division Arrested Assay, Life Technologies[1] AR CALUX, BioDetection Systems[2]
Glucocorticoid Receptor (GR)	GeneBLAzer GR Division Arrested Assay, Life Technologies[1] GR CALUX, BioDetection Systems[2]
Progesterone Receptor (PR)	GeneBLAzer PR Division Arrested Assay, Life Technologies[1] PR CALUX, BioDetection Systems[2]
Aryl Hydrocarbon Receptor (AhR)	AhR CALUX, BioDetection Systems[2]

[1] Madison, WI (USA); [2] Amsterdam, The Netherlands

Sample Processing

Samples to be screened by IVBs shall be collected and preserved following the methods described in Section 10.2. Samples will be extracted following the same protocols used for analytical chemistry *with one critical modification.* To prevent non-sample related interference in bioassay response, addition, fortification or spiking of chemicals of any kind (e.g. internal

standards or recovery surrogates per section 10.4), except those specifically identified to evaluate IVB performance, shall not be performed.

10.5.1.3 Reference Toxicants

Reference toxicants used in the IVBs shall meet the following requirements:

- High affinity for the endpoint of interest
- Linear dose response shall have a dynamic range of 5-fold minimum
- Endpoint specific sensitivity thresholds reported in Table 10.5-1 shall be attained

Since there is limited information on the performance of alternative reference toxicants, it is recommended that all laboratories employ the reference toxicants listed in **Table 10.5-2**. The performance of these chemicals has been evaluated in recent studies that adapted bioassay protocols for water quality measurement (SCCWRP 2014; Escher et al. 2014).

Table 10.5-2. Recommended reference toxicants for in vitro bioassays (IVBs). Agonist mode (+); antagonist mode (-).

Endpoints	Reference Toxicant	Sensitivity Threshold (ng/L)
Estrogen Receptor (ER)	17-beta estradiol (+) 4-hydroxy-tamoxifen (-)	0.5
Androgen Receptor (AR)	Flutamide (-)	20
Glucocorticoid Receptor (GR)	Dexamethasone (+)	50 (TBR)
Progesterone Receptor (PR)	Levonorgestrel (+)	50 (TBR)
Aryl Hydrocarbon Receptor (AhR)	3,3',4,4',5-Pentachlorobiphenyl (PCB 126)(+)	50 (TBR)

TBR - to be resolved

10.5.1.4 Measurement Quality Objectives (MQOs)

The MQOs delineated in **Table 10.5-3** are intended to provide a common foundation for laboratory performance and should be considered as the minimum requirements for bioanalytical screening of pilot study samples. Additional MQOs may be instituted by participating laboratories, as long as the MQOs presented herein are satisfied. In vitro bioassay results shall be reported as bioassay equivalent concentrations (BEQs) in units of ng/L (as reference toxicant).

Table 10.5-3. Measurement quality objectives (MQOs) for *in vitro* bioassays (IVBs)

Measurement Parameter	Frequency of Analysis	Control Limits
Extract Cytotoxicity	Per sample extract	Dilutions of the extract shall not cause > 20% cell mortality (corrected for background).
Cell-Free Media Blank	Per assay plate	Average response for cell free blank (media only) shall be less than 75% of the solvent vehicle free blank response (cells and media). RSD of replicate wells shall be < 20%.
Vehicle Blank Response	Per assay plate	Average response of cells exposed to the solvent vehicle shall be within 15% RSD of the vehicle free response.
Initial Calibration	Per bioanalytical batch	Linear dose-response curve for reference toxicant; $r^2 > 0.95$. Minimum of 9 points per curve (one of them at or below sensitivity threshold (Table 5.2).
Calibration Verification	Per subsequent assay plates within a bioanalytical batch	Continuing calibration shall remain within 15% of mean response for initial calibration.
Spiked Sample	Per extraction batch	Assay response of sample spiked with reference toxicant shall be within 70 to 130% of expected response.
Reproducibility	Per sample	Differences among replicate bioassay responses shall be less than 20% RPD within and among laboratories.

10.5.1.5 Inter-laboratory Comparisons

All laboratories conducting IVBs shall participate in an inter-laboratory comparison exercise prior to sample testing. This exercise will include the analysis of spiked samples prepared by a referee laboratory, and un-spiked pilot study samples for each endpoint undertaken. Samples will be distributed blindly to the participating laboratories and analyzed in triplicate. Successful completion of this exercise will be evaluated based on attainment of MQOs (see sec 10.5.1.4), and data comparability among laboratories.

Data comparability will be based on the following acceptance criteria:

- Intra-laboratory reproducibility shall be 20 % relative percent difference (RPD)
- Percentage difference from the BEQ target value for each spiked sample shall be <30%
- Sensitivity and dose-response curve of reference toxicants shall be in accordance with MQOs (see Tables 10.5-2 and 10.5-3)

Laboratories unable to successfully complete the inter-laboratory comparison exercise will be asked to review their test procedures, make suggested changes, and retest the comparison samples. Failure to meet the inter-laboratory comparison criteria will result in the addition of a cautionary data qualifier flag to that laboratory's data or exclusion from testing during the monitoring program. However, by participating in these exercises, laboratories not passing minimum performance criteria will be informed of methodological issues and shall be able to work with referee labs to resolve issues. In addition, the quantitative assessment of among-laboratory variability afforded by these exercises will provide context for managers when comparing results to other CEC-related projects.

10.5.2 In Vivo Toxicity Testing

10.5.2.1 General Approach

For each scenario (freshwater, embayment, ocean and stormwater), the toxicity of water and/or sediment samples will be evaluated using a whole organism (*in vivo*) test that include reproductive or developmental endpoints. To date, the 21-day reproduction test using fathead minnow (*Pimephales promelas*) is one of the most promising assays for detecting the effects of endocrine disrupting CECs. Thus, it is the only *in vivo* test to be evaluated in the pilot study at this time. This test will be conducted using aqueous freshwater samples (e.g. WWTP effluent, river water). Tests for other scenarios and matrices may be optimized and added to the pilot study plan at a later date.

10.5.2.2 Toxicity of freshwater samples using fathead minnow (Pimephelas promelas)

A short-term reproduction assay using *P. promelas* will be conducted on aqueous freshwater samplesaccording to USEPA (2007) and OECD (2012) guidelines. This test consists of a three to four week acclimation period, followed by a two week (minimum) pre-exposure period and a 21-day exposure to the test samples.

Clean water controls and freshwater samples will be tested in quadruplicate vessels under flow through conditions. Each test vessel will contain two males and four females fed daily with frozen blood worms.

The following test criteria are from the EPA and OECD fish reproduction protocols. These documents should be consulted for additional information on exposure conditions.

Selection of Organisms

Reproductively mature fish (namely, with visible secondary sexual characteristics) capable of actively spawning will be used. Fathead minnows should be preferably five to seven months old, and selected from a single laboratory population that has been cultured at 25 ± 2°C. If possible, the range of individual weights by sex should be kept within 20% of the mean weight of the same sex. For inter-calibration exercises or multi-laboratories studies, it is recommended a common supplier be identified to supply fish within a defined size (e.g. based on mass) range.

During the acclimation period, fish mortalities must be recorded and the following criteria applied:

- Mortalities less than 5% of fish population in seven days: accept the batch
- Mortalities greater than 10% of population in seven days: reject the entire batch
- Mortalities between 5 and 10% of population: acclimate for seven additional days; if more than 5% mortality during second seven days, reject the entire batch

Fish will not be treated for any disease during the holding period, pre-exposure period, or exposure period.

Toxicity Endpoints

Toxicity of test samples will be determined relative to the responses measured in the control vessels. The following endpoints will be measured over the course of the exposure or at termination of the test:

- Survival: Daily assessment. Dead fish will not be replaced in either control or treatment vessels.
- Behavior: Daily qualitative observations of changes in behavior such as uncoordinated swimming, loss of equilibrium, atypical feeding, and hyperventilation.
- Appearance: Daily qualitative assessment of secondary sex characteristics (e.g. size of males' fatpad, number and prominence of nuptial tubercles) and fish coloration conducted daily. Secondary sex characteristics are often affected by the presence of endocrine active chemicals.
- Egg production (fecundity): Number of eggs laid per surviving female per reproductive day.
- Fertilization success: Percentage of fertilized eggs, calculated as the number of embryos/ number of eggs x 100%.
- Vitellogenin (vtg) concentration: Vtg measurements in the plasma will be performed using a validated enzyme-linked immunosorbent assay (ELISA) method capable of detecting vtg in the low ng/mL range.
- Gonad condition measured as the gonadosomatic index (GSI; gonad weight/ body weight x 100%). Typical GSI values are 8 to 13% for reproductive females and 1 to 2% for reproductive males. CECs that affect egg production will also cause a reduction of the GSI in one or both sexes.
- Gonad histopathology (optional): Toxicity responses include intersex, decreased yolk formation, oocyte atresia, testicular degeneration, and hyperplasia.

Measurement Quality Objectives (MQOs)

Measurement quality objectives (MQOs) for the fathead minnow reproduction assay are summarized in **Table 10.5-4**.

Table 10.5-4. Measurements quality objectives (MQOs) for fathead minnow assay

Parameter	Acceptance Criteria
Survival	≥ 90% survival in clean water and/or solvent control vessels at the end of the exposure.
Egg Production	Spawning of 50 to 250 eggs every 4 days minimum during the pre-exposure. Parameters shall be maintained in the control vessels during the exposure.
Fertilization Success	Control fertilization shall be ≥ 95%.
Vitellogenin Concentration	Calibration curve with 6 points minimum, $r^2 \geq 0.98$). Absorbance of duplicate blank samples shall be ≤ 5% of the maximum calibration standard absorbance with a RPD < 20%.
Water Chemistry	≥ 60% air saturation; temperature 25°C± 1°C
Spiked Chemical Exposure	Concentrations shall be maintained within ± 20% of the mean measured value throughout the exposure period.

Note: Water chemistry parameters (e.g. temperature, dissolved oxygen, pH, conductivity) should be recorded daily and reported with the test results. If a parameter falls outside of the MQO for one replicate on a given day, best professional judgment should be used to determine the validity of the test.

10.6 Data Management Plan

The following sections describe the roles and responsibilities, formatting, verification and quality assessment, and reporting requirement for all pilot study monitoring data.

10.6.1 Roles and Responsibilities

Pilot study data shall be submitted by all participating entities to a single Data Management Team (DMT). The DMT is responsible for coordinating receipt of the data, developing and maintaining a data repository for the project, verifying data quality, and providing information to stakeholders and the State data repository. The DMT is responsible for coordinating the development of a common submission format.

10.6.2 Data Submission Format

The data submission formatting will align as closely as possible to the California Environmental Data Exchange Network (CEDEN) data submission templates, with additional fields to include project specific information as needed. Ultimately, the complete data set must be submitted to CEDEN by the DMT.

10.6.3 Data Submission

Data will be submitted by electronic spreadsheet, with an accompanying narrative describing any issues that should be brought to the attention of the DMT. Upon receipt and evaluation by the DMT, the analytical laboratory must be notified of any additional information or corrective actions deemed necessary. Following satisfactory resolution of all "corrective action" issues, the final action is to notify the laboratory in writing that the submitted results have been officially accepted as complete. Evaluation of the data by the DMT should begin as soon as possible following its receipt, since delays increase the chance that information may be lost. The following steps are to be followed and documented: 1) checking data completeness, 2) assessing data quality, and 3) QC reporting. All instrumental data and calculations leading to the submitted results should be retained by the laboratories in case a detailed inspection is required.

10.6.4 Data Completeness

Upon receipt of data, the DMT will verify it has been supplied in the correct format and enter it in to the repository. Checks will be performed to verify results have been reported for all expected stations, samples, and analytes, and all QC data has been included. The field crew or laboratory will be contacted to request any missing data. Significant revisions may require resubmission of the entire data set. Raw data (e.g., chromatograms or original quantitation reports) are not required for submission but must be maintained by the laboratories and made available if requested.

10.6.5 Assessing Data Quality

Data quality will be validated by the DMT as follows:

1. A check to verify that all reporting units and number of significant figures are correct.
2. A check to verify that all calculated percent recovery values and relative percent difference values are correct.
3. All QC data should be compared against the established MQO criteria.

There are several possible courses of action to be taken if the reported data are deficient during the assessment of data quality. First, the laboratory's narrative explanation should be consulted to determine if the problems were satisfactorily addressed. If there were minor MQO criteria exceedances in isolated cases, then it is appropriate for the laboratory to report the results along with appropriate qualifiers for those cases. Pervasive violations of MQO criteria, however, will result in one of the following courses of action. 1) All associated results will be qualified as estimated values. For example, if an analyte had minor QC violations in 3 of 5 analytical batches, the results from all 5 batches may be qualified as estimated. 2) In the most extreme situation, all associated data will be rejected and deleted from the repository.

Because some degree of expert judgment and subjectivity is typically necessary to evaluate QA/QC results and assign data qualifiers, validation will be conducted only by qualified personnel. Data which are qualified as estimates because of minor MQO violations are still usable for most assessment and reporting purposes. However, all QA/QC data will be available in the repository, so interested users may make their own determination of data quality.

10.6.6 Reporting

The DMT will produce reports documenting the results of QC reviews. These documents will summarize all conclusions concerning data acceptability and should note all significant quality assurance problems. These reports provide data users with a written record of QC concerns and a documented rationale for why certain data were accepted as estimates or were rejected. The following items should be addressed in the QA report:

1. A statement on the completeness of the data set relative to the original objectives.
2. A summary of overall data quality, including a description qualified data and rationales
3. Brief descriptions of analytical methods and the method(s) used to determine reporting and detection limits.

10.7 Additional Information

Information in this document is intended to be serve as guidance in generating a project QAPP for statewide CEC pilot monitoring data collection. This QAPP should follow EPA Guidance for Quality Assurance Project Plans (EPA QA G-5), which requires information that can only be known in the implementation phase of the project, i.e. once the organization and scope of the various project components are finalized. This includes 1) project management information such as the names of key personnel, 2) data generation information such as exact sampling and analytical methods, and 3) an assessment plan to ensure the QA Project Plan is being implemented as approved.

10.8 QA/QC Guidance References

Anderson, P.D., Denslow, N.D., Drewes, J.E., Olivieri, A.W., Schlenk, D., Scott, G.I., Snyder, S.A. 2012. Science Advisory Panel for Southern California Coastal Water Research Project (SCCWRP). Monitoring Strategies for Chemicals of Emerging Concern (CECs) in California's Aquatic Ecosystems, Technical Report 692, Southern California Coastal Water Research Project Authority, Costa Mesa, CA, 215 pp.

Escher, B.I., Allinson, M., Altenburger, R., Bain, P.A., Balaguer, P., Busch, W., Crago, J., Denslow, N.D., Dopp, E., Hilscherova, K., Humpage, A.R., Kumar, A., Grimaldi, M., Jayasinghe, B.S., Jarosova, B., Jia, A., Makarov, S., Maruya, K.A., Medvedev, A., Mehinto, A.C., Mendez, J.E., Poulsen, A., Prochazka, E., Richard, J., Schifferli, A., Schlenk, D., Scholz, S., Shiraishi, F., Snyder, S., Su, G., Tang, J.Y., Burg, B.V., Linden, S.C., Werner, I., Westerheide, S.D., Wong, C.K., Yang, M., Yeung, B.H., Zhang, X., Leusch, F.D. 2014. Benchmarking organic micropollutants in wastewater, recycled water and drinking water with in vitro bioassays. *Environ Sci Technol* 48:1940-1956.

Organization for Economic Cooperation and Development (OECD). 2012. Fish Short Term Reproduction Assay. OECD Environmental Health and Safety Publications, Series on Testing and Assessment No. 229, October 2, 2012, OECD, Paris.

Southern California Coastal Water Research Project (SCCWRP). 2013. Bight '13 Contaminant Impact Assessment Committee, Southern California Bight 2013 Regional Marine Monitoring Survey (Bight '13) Quality Assurance Manual, Southern California Coastal Water Research Project Authority, Costa Mesa, CA.

SCCWRP. 2014. Development of Bioanalytical Techniques for Monitoring of Constituents/Chemicals of Emerging Concern (CECs) in Recycled Water Applications for the State of California, Final Report, Agreement 10-096-250, Southern California Coastal Water Research Project Authority, Costa Mesa, CA, 281 pp.

SCCWRP. 2015. Monitoring of Constituents of Emerging Concern (CECs) in Aquatic Ecosystems – Pilot Study Guidance, Southern California Coastal Water Research Project Authority, Costa Mesa, CA, 72 pp.

SWAMP Quality Assurance Team. 2008. Surface Water Ambient Monitoring Program Quality Assurance Program Plan, Version 1.0, September 1, 2008.

U.S. Environmental Protection Agency (USEPA). 2007. Validation of the Fish Short-Term Reproduction Assay: Integrated Summary Report, December 2007, accessed 9/29/2014, at http://www.epa.gov/endo/pubs/att-f_fish_assay_protocol.pdf.

USEPA. 2013. Endocrine Disruptor Screening Program. Sap Review of EDSP Tier 1 Screening: Assay and Battery Performance, May 21-23, 2013.

WateReuse Research Foundation (WRRF). 2014. Development of Bioanalytical Techniques to Assess the Potential Human Health Impacts of Recycled Water, WRRF, Arlington, VA, 316 pp.

U.S. Geological Survey (USGS). 2004. Techniques of Water-Resources Investigations, book 9, chapter A5, Wilde, F.D., Radtke, D.B., Gibs, Jacob, Iwatsubo, R.T., eds., Processing of water samples (ver. 2.2), April 2004, accessed 9/3/2014, at http://pubs.water.usgs.gov/twri9A5/.

10.9 QA/QC Guidance Abbreviations

AhR – aryl hydrocarbon receptor
AR – androgen receptor
BEQ – bioassay equivalent concentration
CEC - constituents of emerging concern
CEDEN - California Environmental Data Exchange Network
DMT – data management team
ER – estrogen receptor
GC/MS – gas chromatography/mass spectrometry
GR – glucocorticoid receptor
GSI – gonadosomatic index
IVB – in vitro bioassay
LC-MS/MS – liquid chromatography/tandem mass spectrometry
LCS – laboratory control sample
MDL – method detection limit
MQO – measurement quality objective
MS/MSD – matrix spike/matrix spike duplicate
MS4 – municipal separate stormwater sewer system
MTL – monitoring trigger level
OECD – Organization for Economic Cooperation and Development
OSHA – Occupational Health and Safety Administration
PBDE – polybrominated diphenyl ether
PFC – perfluorinated compound
PR – progesterone receptor
QA – quality assurance
QC – quality control
QAPP – Quality Assurance Project Plan
RL – reporting limit
RPD – relative percent difference
RSD – relative standard deviation
SCCWRP – Southern California Coastal Water Project Authority
SOP – standard operating procedure
SRM – standard reference material
SWAMP – Surface Water Ambient Monitoring Program
SWRCB – State Water Resources Control Board
TBD - to be determined
TBR - to be resolved
Vtg – vitellogenin
WWTP – wastewater treatment plant