



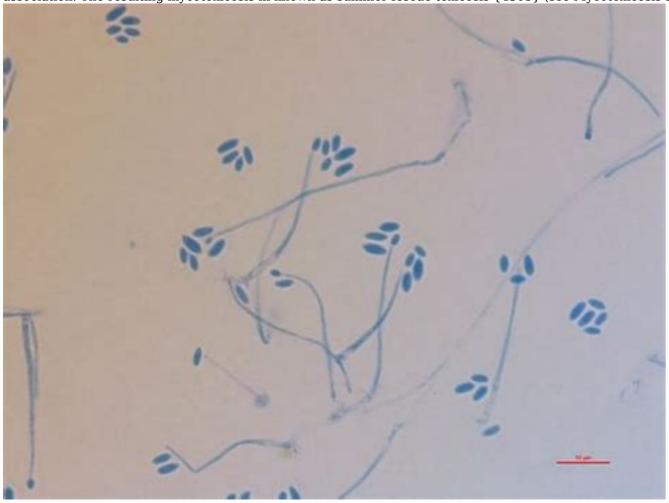
Acremonium spp pdf

Acremonium spp.

Kingdom Fungi Order Hypocreales Phylum Ascomycota Family Hypocreaceae Class Euascomycetes Genus Acremonium The Acremonium genus was formerly known as Cephalosporium. Depending on the different authors, the genusAcremonium currently contains between 20 and 200 recognised named species {814; 813; 3971}. The taxonomic fungal database administered by the International Mycological Association has 205 registered strains of which 197 are named species {3971}. This genus comprises moulds that lack any known sexual state or teleomorph forms and thus belongs to the Fungi Imperfecti group. However, because it possesses structural characteristics similar to those of the Ascomycota group, it is often included in the Ascomycota phylum. Furthermore, some authors have proposed that certain fungi could be sexual stages of different Acremonium species, amongst which include Acanthonitschkea, Albertiniella, Allantonectria, Allescheria, Atkinsonella , Bulbithecium and Bysostilbe {3842}. Habitat/Ecology Acremonium tax a world-wide distribution: it is commonly found in the environment in soil and on dead plant material as well as in hay and rotting mushroomics; some species are pathogenic to plants and humans {813; 2114}. The mode of dissemination of wet spores is by insects, water droplets or by wind {2204; 813}. In aerobiological counts, Acremonium species of Acremonium species of the air and its occurrence may be quite low {1973}. It is reported as one of the less prevalent genera in air samples. Alternatively, some species are widespread indoor faltafor faltafor; 468}: species of Acremonium species of Acremonium species of species for and are thus only discharged into the air in low concentrations {1973}. In northern climates, concentrations in the outdoor air vary seasonally and comprise a small proportion of the natural aerosolised fungal flora. Spore levels of Acremonium appear to be higher in rural as opposed to urban environments {854}.

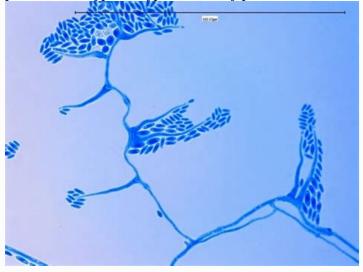
This fungus is isolated in rural environments in cow barns, farm houses and rural outdoors but is rarely observed in urban apartments. One study {854} identified A. murorum and A. strictum in swineries and grain mills albeit in very low concentrations (less than 1.7% of total isolates). In a study of tap water, Acremonium was found among the highest of total fungal counts {1733}. However, at ambient temperature, none of the fungi isolated could be associated epidemiologically with human pathogenicity per se. These fungi may nevertheless be involved in the production of certain tastes and odours in water. Growth requirements Acremonium requires very wet conditions, e.g. building materials with high water affinity. Consequently it is often seen growing in combination with Stachybotrys, a tertiary coloniser.

Most species do not grow at 37°C {813}. When found indoors, it usually originates either from outdoor air or from contaminated building materials following an incidence of water damage. However, the spores of this fungus are formed in a slimy mass resulting in limited aerosolisation: therefore its prevalence may be low in air samples {1973}. Acremonium has been found indoors growing on building materials, such as acoustic and thermal fibreglass insulation used in heating ventilation and air conditioning systems. Because of its high water affinity, it is often isolated from cooling coils, drain pans, window seals and water from humidifiers. More details Acremonium has often been reported associated with contamination by Stachybotry's chartarum, as this fungus is also favoured by very wet circumstances. A stench in the air may be associated with the presence of this fungus {431}. In indoor investigations, Acremonium occupied a middle to low rank among the dominant genera in winter and summer in Argentina {1584}. The same can be said of homes in Southern California, where Acremonium was not among those most frequently isolated, although it was found in 35% of homes tested, with a mean viable spore concentration of 30.6 CFU/m3 {1824}. Conversely, a one year survey of outdoor air of a Brazilian city showed a low prevalence of Acremonium (0.2%), ranking 19th rank among the 25 genera identified {1973}. In an investigation of the presence of indoor air fungi in eight houses in Finland, Airaksinen et al. {2104} foundAcremonium in a majority of dwellings with a mean concentration of 160 CFU/m3. Moreover, higher concentrations were identified in crawl spaces (mean concentration 2 240 CFU/m3) hence enabling the authors to establish a clear link between fungal spores in the indoor air and contaminated crawl space. In another study, among 53 species, Acremonium strictum was found in 25% of air samples (but not in water) of water closet environments and thereby considered as one of the most prevalent species {2193} in this study. Patovirta et al. {199} performed a follow-up study after an extensive mould remediation in school buildings; Acremoniumsp. was found in the air and on building materials prior to and shortly after remediation but not after an extended period of 3 years. Elsewhere, Acremonium was identified in 14 out of 72 samples of materials from 23 mould-infested buildings and was ranked 7th among the most frequent genera isolated {725; 605}. As with other fungi, Acremonium was most often found on wood {605}. Acremonium strictum and Acremonium sp. were also found in carpet dust samples in Belgian homes, at concentrations of 104 CFU/g of dust {3972}. In a controlled study on mould growth on wet gypsum wallboard in an indoor environment, Acremonium was among early colonisers, along with Cladosporium and Penicillium; it was detected three weeks after immersing the building material in water {4080; 587}. Normal laboratory precautions should be exercised in handling cultures of this species within 5 days. Colonies may be slow to moderately rapid when grown at 25 °C; colonies are often compact, reaching less than 2.5 cm in diameter in 5-10 days {3283; 1056; 415}. Colony texture is at first glabrous, compact, sometimes moist, flat or folded, then subsequently felt-like and, with age, becomes overgrown with loose hyphae: at this stage, the colonies appear powdery in texture, suede-like or floccose. Colonies may be white, pale grey, pink, rose or orange in color {814; 813; 412; 1056}. The reverse side is either uncoloured or tinted by a pink to rose coloured pigment {4080; 3283; 415}. Microscopic morphology The Acremonium septate hyphae are hyaline and very delicate. Vegetative hyphae often form hyphal ropes. The conidiogenous cells are phialides. The phialides are solitary, aculeate, mostly awl-shaped, simple, erect, and arise from the substrate mycelium or from bundled aerial hyphae by a septum and taper towards their apices. When conidiophores occur, their branches are only restricted to the basal part. Conidia are oblong (1-3 x 4-8 mm), usually one-celled, hyaline or pigmented. They usually appear in clusters, in balls or rarely as fragile chains; the conidia may be aggregated in slimy heads at the apex of each phialides, bound by a gelatinous material {814; 812; 1056}. More details The conidia are mainly single-celled or rarely multicellular (2- or 3celled conidia), fusiform with a slight curve or may resemble a shallow crescent. These structural properties of conidia vary depending on the species; for example, Acremonium spp., as most fungi, produce various hydrocarbons, alcohols, ketones, esters and terpenes, in nature as well as on building materials {594; 2076}. Production of microbial volatile organic compounds (MVOCs) is influenced by both medium and species. One study was able to detect cyclotrisiloxane, limomene, pentane, arsenous acid and benzene on fibreglass experimentally colonised with Acremonium {2051}. More details A number of organic compounds, including volatile organic compounds (VOCs), have been identified in indoor air in damp buildings contaminated by fungi; these VOCs are thought to contribute to various indoor air problems. However, most of the identified metabolites are non-reactive and found in low concentration in indoor air {594}. Some species have a defined MVOC profile that may be subject to considerable modification in response to external factors such as cultivation on different substrata. These differing substrata change both the number and concentration of MVOCs {2968; 2809; 1148} whereas other volatile metabolites are specific for single species {2809}. Mycotoxins Some species of Acremonium produce mycotoxins. Among the best studied mycotoxins produced by Acremonium spp. are citrinin, cephalosporin, ergovaline {2109} and lolitrem A and B. For instance, lolitrems produced by Acremonium lolii are potent trichothecene mycotoxins under certain conditions {431}; to date, this toxin production has been associated solely with substrates of contaminated food, fodder and pasture. Most Acremonium toxins are associated with rural settings. More details A novel mycotoxin named acrebol, consisting of two closely similar peptaibols, was isolated from an indoor strain of Acremonium exuviarum {4336}. Acrebol induces necrosis-like cellular death in mouse insulinoma MIN-6 cells. Acremonium coenophialum can produce directly by the fungus alone or whether it is the product of a symbiotic fungus-plant association. The resulting mycotoxicosis in known as summer fescue toxicosis {4305} (see Mycotoxicosis section). A novel Acremonium metabolite, named FR235222, possesses potent immunosuppressive properties on mouse cell lines. The potential clinical use of this molecule is currently under study {2111}.



No specific irritation or inflammation symptoms have been attributed specifically to Acremonium spp. Many substances common to most moulds, such as glucans, may cause irritation and inflammation in the exposed subject.

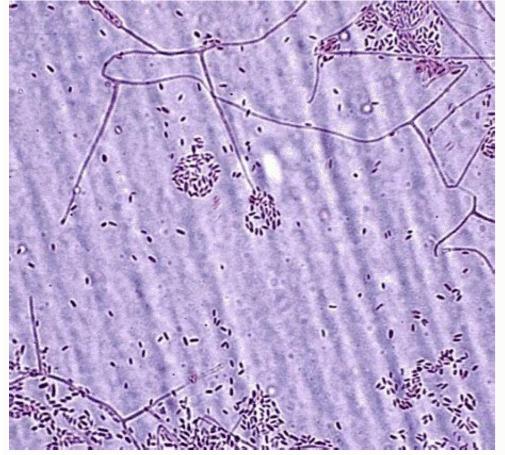
However, no particular substance has been reported in relation to Acremonium. More details Generally speaking, all moulds contain substances that are irritants and promote inflammation to some degree. Some VOCs produced by moulds in the indoor setting on damp building materials are thought to contribute to various health problems such as eye irritation, irritation of the nose and throat, lethargy and headache {594} (1-->3)-Beta-D-glucans are non-specific and non-allergenic structural cell wall components present in most fungi and have been suggested to play a causal role in the development of respiratory symptoms associated with indoor fungal exposure {1346}. Allergic reactions Some species of Acremonium have been reported to be allergenic and have been linked to Type I allergies (hay fever and asthma) {3286; 2342; 3095; 388}. However the few aerobiological surveys that have detected Acremonium indoors report a low prevalence from 4.6 % {4321} to slightly above 10% {4274}; this fungus rarely appears in large quantities. Consequently, the contribution of Acremonium to fungal allergies is likely low. Allergic components and mechanism No specific allergenic fraction of Acremonium is listed with either the Biological Product registry of the Federal Drug Administration (FDA) {3285} or with the International Union of Immunological Societies. Hypersensitivity pneumonitis Type III hypersensitivity pneumonitis due to Acremonium sp. has been reported in a few instances, mostly associated with contaminated home humidifiers {2761; 4322; 1407}.



In one study, hypersensitivity pneumonitis associated with contaminated home ultrasonic humidifiers was diagnosed in five patients. Tests for precipitating antibodies against an extract of the humidifier water yielded strong positive reactions in all patients tested; all patients also tested positive for precipitins against Cephalosporium acremonium (syn. Acremonium chrysogenum) {1407}. Two other cases of hypersensitivity lung disease apparently caused by home environment contamination by Cephalosporiumhave been identified {4322}; one case was presumptively in a home contaminated by sewage flooding and the other as a result of exposure to humidifier contaminated water. Toxic effects (mycotoxicosis) Many strains of Acremonium are active producers of toxins under given sets of growth conditions. Toxic effects due to ingested Acremonium toxins include cytotoxic, nephrotoxic and tremorgenic effects. These pathologies are well known to occur in livestock and other animals {3182; 3184; 3185; 4305} but no cases of human mycotoxicosis have yet been reported. Animal cases of neurotoxic mycotoxicosis have often been reported associated with Acremonium contamination of grasses and feed {2192}.

In particular, Acremonium infected tall fescue grasses may be responsible for toxicosis reported in both sheep and cattle, including Staggers disease as well as failure to thrive in cattle {4327}. More details Clinical signs of Staggers disease are transient ataxia, which is aggravated by stimulation; nearly complete recovery occurs after removal of ryegrass as the primary forage. Morbidity associated with these toxicosis or fescue poisoning is a disease caused by ingestion of Acremonium contaminated tall fescue grass; it produces a dry gangrenous lesion in affected livestock (rarely sheep) similar to that found in ergot poisoning {4305; 2189}. For many years, endophytic mycotoxin intoxications have been limited to the United States and New Zealand. However, these mycotoxicoses have since been reported in Western Europe {4324}.

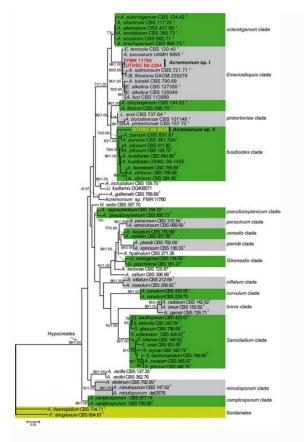
Experimentally, injections of ryegrass extracts containing the tremorgenic mycotoxin lolitrem-B into mice induce signs of toxicosis {2185; 4326; 2192; 4327}. Infections and colonisations Acremonium species are infrequent pathogens in humans {2152; 2131}. However, they can cause a spectrum of infections, ranging from mycotic keratitis and mycetoma in the normal host, to fungemia, disseminated infections and cutaneous infections in immunocompromised subjects {2105; 316}. Opportunistic infections, infections of the cornea and nails {2134} in individuals with weak immune systems have been occasionally reported. Life-threatening unusual pulmonary mycoses caused by A. strictum have been reported in patients with haematological diseases {2139}. Acremonium mycetomas and other rare hyalohyphomycoses infections have been reported in immunodeficient patients and in immunodeficient patients {2085}.



About 10 species of Acremonium are reported as causes of infection in vertebrates, with A. kiliense being the most important {2085}. Agents of Acremonium hyalohyphomycoses include, in alphabetical order, A. alabamensis, A. falciforme, A. kiliense, A. potroni, A.

recifei, A. roseo-griseum and A. strictum {2121; 2092; 2091}. However, many case reports of infections only identify Acremonium species at the genus level. Clinical manifestations of these hyalohyphomycoses caused by Acremonium include arthritis, osteomyelitis, peritonitis, endocarditis, pneumonia, cerebritis and subcutaneous infection {814}. This pathogen has also been reported to cause locally invasive diseases such as mycetoma, paranasal sinus and haemodialysis access graft infections as well as posttraumatic keratitis {2199}. More details Acremonium is usually of low pathogenicity {2102}, although this fungus is being increasingly recognised as an opportunistic pathogen causing a variety of infections in immunocompromised patients (organ transplant recipients, myeloma or leukemia patients and those receiving corticoid therapy) {2167; 2085; 2126}. Invasive or disseminated infections are almost exclusively observed in immunocompromised patients {2114; 2089}; among neutropenic patients suffering from a disseminated infection, about half of afflicted patients died {2159}.

The lungs and the gastrointestinal tract are the apparent portals of entry in many patients {316}, although the spectrum of invasive disease is large, ranging from sinusitis, osteomyelitis, arthritis, peritonitis and pneumonia; invasion of vascular structures results in thrombomycosis, tissue infarction and necrosis {2114}. In immunocompetent individuals, it is possible to contract an Accremonium infection due to traumatic penetration of the mould {2102}; high prevalence of Accremonium species in soil can lead to superficial infections after traumatic inocalation {2131}. Funga kerentis inocalation {2131}. Funga kerentis infection of the distal extremities usually causing only mild discomfort, although a prolonged infection can lead to underlying bone destruction. Virulence factors No particular virulence factor has been reported and most species of Accremonium do not grow at 37°C. Accremonium sp. has rarely been reported. Notwithstanding this nominal occurrence, Acremonium species have become significant problems in the treatment of infections. Alter metica isolates have been compared for phenotypic and biochemical similarities. In an American study, Accremonium sp. has rarely been reported. Notwithstanding this nominal occurrence, Acremonium sp. has energing pathogens if on future nosocomial transmission of Acremonium sp. has energing pathogens {312}. Even and for future nosocomial transmission of Acremonium sp. has rarely been reported. Notwithstanding this nominal occurrence, Acremonium sp. has rarely been reported. Notwithstanding this nominal occurrence, Acremonium sp. has energing pathogens {312}. Even and for future nosocomial transmission of holecular studies have been compared for phenotypic and biochemical similarities. In an American study, Acremonium sp. has rarely been reported. Notwithstanding this nominal occurrence, Acremonium sp. has appresent so for future nosocomial transmission of the mould {212}. Even and for future nosocomial transmission of the dustal strue transmission of the mould susch



The mechanism of disseminated infection is thought to be secondary to increased host susceptibility with prior colonisation. A recent outbreak of Acremonium kiliense endophthalmitis was reported following cataract surgery in an ambulatory surgical center in which the responsible reservoir was determined to be a contaminated humidifier within the <u>heating</u>, ventilation, and air-conditioning system located directly above the operating suite {2174}. This occurrence was somewhat surprising since, historically, only Aspergillus spp.



and Zygomycetes have been implicated as major airborne fungal pathogens occurring in immunocompromised patients in health care settings, usually associated with hospital construction or renovation {4330}. Many cases of post transplantation Acremonium infections underscore the nosocomial threat posed by this fungus even if hospital transmission is not incriminated. Indeed, Acremonium has been involved in infections following transplantation of blood stem cells {2089}, bone marrow {2091} or kidney {2112; 2139}. Fatal infections by A. strictum have also been reported in young patients with leukemia {2092; 2171; 2126}. A case an acute septic arthritis after kidney transplantation due to Acremonium species was also reported {2112}.

Finally, very rare cases of neonates with disseminated Acremonium infection have been reported 1208}. Occupational diseases No occupational infections due to Acremonium sp. have been reported in the workplace. More details Type III hypersensitivity pneumonits due to Acremonium sp. have been mostly reported in the home environment and is not particularly known in occupational settings. Because of the prevalence of Acremonium in both cold and warm weakers are utipational exposure to Acremonium species are utipations. However, because the clinical process may be transported from cow barns to farmers' homes {2203}. It is worthwhile mentioning that Acremonium species are utipations for a doep not and a other considered have a nonducting primary cultures of deep setted species are utipation of a depositive cases of invasive hyalohyphomycoses are very seldom considered when conducting primary cultures of deep setted species caunot be distinguished by histopathological presentation of a deep setted in the workplace. Canobian and Acremonium species cannot be distinguished by histopathology the histological presentation of a deep setted in the distinguished by histopathology the histological presentation of a deep setted in the distinguished by histopathology the possibility of this aetiology in terms in the varies have been reported to complete the diagnosis {2156} and kerattis by especial presentation of a deep setted in the workplace. Accemonium species are utipation and exposure and phaloconida in tissue sections routinely stained with haematoxy in and eosin, Gomori methanamice alume site and/or perioduci acid-Schift stains; cultures are required to complete the diagnosis {2156}. In one instance, A. stricture was shown to cause a dense nodular lymphohistic systis infiltrate cutaneous infection [2156]. Acremonium mycetomas provide and and morphology that are very similar to there are setimate on the advance and an approximate guide in correctly identifying the organism. Fusarium and Acremonium species are utipate and

G. (1997). Fungal colonization of air filters from hospitals. Am Ind.Hyg.Assoc.J. 58[12], 900-904. 412. Larone, D H. (1987). Medically important fungi. A guide to identification. 2nd edition, -230 p.

New York - Amsterdam - London, Elsevier Science Publishing Co., Inc. 415. St-Germain, G and Summerbell, R. (1996). Champignons filamenteux d'intérêt médical. Caractéristiques et idenfication. -314 p. Belmont, Star Publishing Company. 431. Roberts, S. (2006). Mold...What is all about?, p. 1-p. 7. Mold-Help. 3-10-0060. 468. Scott, J M. (2001). Studies on indoor fungi. -291 p. University of Toronto. 587. Krause, M., Geer, W., Swenson, L., Fallah, P., and Robbins, C. (2006). Controlled study of mold growth and cleaning procedure on treated and untreated wet gypsum wallboard in an indoor environment. J Occup.Environ Hyg. 3[8], 435-441. 594. Claeson, A. S., Levin, J. O., Blomquist, G., and Sunesson, A. L. (2002). Volatile metabolites from microorganisms grown on humid building materials and synthetic media. J Environ Monit. 4[5], 667-672. 605. Gravesen, S., Nielsen, P. A., Iversen, R., and Nielsen, K. F. (1999). Microfungal contamination of damp buildings--examples of risk constructions and risk materials. Environ Health Perspect. 107 Suppl 3:505-8., 505-508. 725. Gravesen, S., Frisvad, J. C., and Samson, RA. (1994). Microfungi. 1st edition, -168 p. Copenhagen, Munksgaard. 813. EMLAB. (2007). Environmental Microbiology Laboratory, Inc. (EMLab): An index of some commonly encountered fungal genera. 814. Ellis, D. (2007). Mycology online.

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School of molecular & biomedical science. The University of Adelaide. 854. Lugauskas, A., Krikstaponis, A., and Sveistyte, L. (2004). Airborne fungi in industrial environments--potential agents of respiratory diseases. Ann Agric.Environ Med. 11[1], 19-25. 1056. Samson, RA, Hoekstra, ES, and Frisvad, JC. (2004). Introduction to food and airbone fungi. 7th, -389 p. Baarn, Centralalbureau voor Schimmellcultures, Institute of the Royal Netherlands Academy of Arts and Sciences. 1148. Fischer, G., Muller, T., Schwalbe, R., Ostrowski, R., and Dott, W. (2000). Species-specific profiles of mycotoxins produced in cultures and associated with conidia of airborne fungi derived from biowaste. Int J Hyg.Environ Health. 203[2], 105-116. 1202. Pieckova, E. and Jesenska, Z. (1999).

Microscopic fungi in dwellings and their health implications in humans. Ann Agric. Environ Med. 6[1], 1-11.

1346. Douwes, J. (2005). (1-->3)-Beta-D-glucans and respiratory health: a review of the scientific evidence. Indoor Air. 15[3], 160-169. 1407. Suda, T., Sato, A., Ida, M., Gemma, H., Hayakawa, H., and Chida, K. (1995). Hypersensitivity pneumonitis associated with home ultrasonic humidifiers. Chest. 107[3], 711-717. 1584. Basilico, Mde L., Chiericatti, C., Aringoli, E. E., Althaus, R.

L., and Basilico, I. C. (2007).

Influence of environmental factors on airborne fungi in houses of Santa Fe City, Argentina.

Sci Total Environ. 376[1-3], 143-150. 1733. Goncalves, A. B., Paterson, R. R., and Lima, N. (2006). Survey and significance of filamentous fungi from tap water. Int J Hyg Environ Health. 209[3], 257-264. 1824. Kozak, P. P., Gallup, J., Cummins, L. H., and Gillman, S.

A. (1979). Factors of importance in determining the prevalence of indoor molds. Ann Allergy. 43[2], 88-94. 1973. Menezes, E. A., Trindade, E.

C., Costa, M. M., Freire, C. C., Cavalcante, Mde S., and Cunha, F. A. (2004). Airborne fungi isolated from Fortaleza city, State of Ceara, Brazil. Rev Inst.Med Trop Sao Paulo. 46[3], 133-137. 2051. Ezeonu, I. M., Price, D. L., Simmons, R. B., Crow, S. A., and Ahearn, D. G. (1994). Fungal production of volatiles during growth on fiberglass. Appl Environ Microbiol. 60[11], 4172-4173. 2076. Claeson, A. S., Sandstrom, M., and Sunesson, A. L. (2007). Volatile organic compounds (VOCs) emitted from materials collected from buildings affected by microorganisms. J Environ Monit. 9[3], 240-245.

2085. Keynan, Y., Sprecher, H., and Weber, G. (2007). Acremonium vertebral osteomyelitis: molecular diagnosis and response to voriconazole. Clin Infect Dis. 45[1], e5-e6. 2089. Yamazaki, R., Mori, T., Aisa, Y., Nakazato, T., Mihara, A., Ikeda, Y., and Okamoto, S. (2006).

Systemic infection due to Acremonium after allogeneic peripheral blood stem cell transplantation. Intern Med. 45[16], 989-990.

2091. Miyakis, S., Velegraki, A., Delikou, S., Parcharidou, A., Papadakis, V., Kitra, V., Papadatos, I., and Polychronopoulou, S. (2006). Invasive Acremonium strictum infection in a bone marrow transplant recipient. Pediatr Infect Dis J. 25[3], 273-275. 2092. Foell, J. L., Fischer, M., Seibold, M., Borneff-Lipp, M., Wawer, A., Horneff, G., and Burdach, S. (2007). Lethal double infection with Acremonium strictum and Aspergillus fumigatus during induction chemotherapy in a child with ALL. Pediatr Blood Cancer. 49[6], 858-861. 2097. Thuret, G., Carricajo, A., Vautrin, A.

C., Raberin, H., Acquart, S., Garraud, O., Gain, P., and Aubert, G. (2005). Efficiency of blood culture bottles for the fungal sterility testing of corneal organ culture media. Br J Ophthalmol. 89[5], 586-590. 2099. Lepidi, H., Casalta, J. P., Fournier, P. E., Habib, G., Collart, F., and Raoult, D. (2005). Quantitative histological examination of mechanical heart valves.

Clin Infect Dis. 40[5], 655-661. 2102. Kan, S. F., Tsai, T. H., Hu, C. H., and Lee, W. R. (2004).

Cutaneous hyalohyphomycosis caused by Acremonium in an immunocompetent patient.

Br J Dermatol. 150[4], 789-790. 2104. Airaksinen, M., Pasanen, P., Kurnitski, J., and Seppanen, O. (2004).

Microbial contamination of indoor air due to leakages from crawl space: a field study. Indoor Air. 14[1], 55-64. 2105. Schinabeck, M. K. and Ghannoum, M. A. (2003). Human hyalohyphomycoses: a review of human infections due to Acremonium spp., Paecilomyces spp., Penicillium spp., and Scopulariopsis spp. J Chemother. 15 Suppl 2:5-15., 5-15. 2108. Yalaz, M., Hilmioglu, S., Metin, D., Akisu, M., Nart, D., Cetin, H., Ozturk, C., Isik, E., and Kultursay, N. (2003). Fatal disseminated Acremonium strictum infection in a preterm newborn: a very rare cause of neonatal septicaemia. J Med Microbiol. 52[Pt 9], 835-837. 2109. Gadberry, M.

S., Denard, T. M., Spiers, D. E., and Piper, E. L. (2003). Effects of feeding ergovaline on lamb performance in a heat stress environment. J Anim Sci. 81[6], 1538-1545.

2111. Mori, H., Abe, F., Furukawa, S., Furukawa, S., Sakai, F., Hino, M., and Fujii, T. (2003). FR235222, a fungal metabolite, is a novel immunosuppressant that inhibits mammalian histone deacetylase (HDAC) II. Biological activities in animal models.

J Antibiot. (Tokyo). 56[2], 80-86. 2112. Beaudreuil, S., Buchler, M., Al, Najjar A., Bastides, F., Francois, M., Duong, T. H., Nivet, H., Richard-Lenoble, D., and Lebranchu, Y. (2003). Acute septic arthritis after kidney transplantation due to Acremonium. Nephrol.Dial.Transplant. 18[4], 850-851. 2114. Fleming, R. V., Walsh, T. J., and Anaissie, E. J. (2002). Emerging and less common fungal pathogens. Infect Dis Clin North Am. 16[4], 915-vii. 2116. Bibashi, E., Kokolina, E., Sigler, L., Sofianou, D., Tsakiris, D., Visvardis, G., Papadimitriou, M., and Memmos, D. (2002). Three cases of uncommon fungal peritonitis in patients undergoing peritoneal dialysis. Perit.Dial.Int. 22[4], 523-525. 2121. Anadolu, R., Hilmioglu, S., Oskay, T., Boyvat, A., Peksari, Y., and Gurgey, E. (2001). Indolent Acremonium strictum infection in an immunocompetent patient. Int J Dermatol. 40[7], 451-453. 2123. Walsh, T. J. and Groll, A. H. (1999). Emerging fungal pathogens: evolving challenges to immunocompromised patients for the twenty-first century. Transpl.Infect Dis. 1[4], 247-261. 2124. Parnell, P. and Wilcox, M. H. (2001).

Myobacterium chelonae and Acremonium species isolated from endoscope autodisinfector rinse water despite daily treatment with chlorine dioxide. J Hosp Infect. 48[2], 152-154. 2126. Warris, A., Wesenberg, F., Gaustad, P., Verweij, P. E., and Abrahamsen, T. G. (2000). Acremonium strictum fungaemia in a paediatric patient with acute leukaemia. Scand.J Infect Dis. 32[4], 442-444. 2131. Rodriguez-Ares, T., De, Rojas Silva, V, Ferreiros, M. P., Becerra, E. P., Tome, C. C., and Sanchez-Salorio, M. (2000). Acremonium keratitis in a patient with herpetic neurotrophic corneal disease. Acta Ophthalmol.Scand. 78[1], 107-109. 2133. Laurent, D., Guella, G., Roquebert, M. F., Farinole, F., Mancini, I., and Pietra, F. (2000).

Cytotoxins, mycotoxins and drugs from a new deuteromycete, Acremonium neo-caledoniae, from the southwestern lagoon of New Caledonia. Planta Med. 66[1], 63-66. 2134. Tosti, A., Piraccini, B. M., and Lorenzi, S. (2000). Onychomycosis caused by nondermatophytic molds: clinical features and response to treatment of 59 cases. J Am Acad Dermatol. 42[2 Pt 1], 217-224. 2139. Breton, P., Germaud, P., Morin, O., Audouin, A. F., Milpied, N., and Harousseau, J. L. (1998). [Rare pulmonary mycoses in patients with hematologic diseases]. Rev Pneumol.Clin.

54[5], 253-257. 2152. Grunwald, M. H., Cagnano, M., Mosovich, M., and Halevy, S. (1998). Cutaneous infection due to acremonium. J Eur Acad Dermatol. Venereol. 10[1], 58-61. 2153. Weissgold, D. J., Orlin, S. E., Sulewski, M. E., Frayer, W. C., and Eagle, R. C., Jr. (1998). Delayed-onset fungal keratitis after endophthalmitis. Ophthalmology. 105[2], 258-262. 2156. Liu, K., Howell, D.

N., Perfect, J. R., and Schell, W. A. (1998). Morphologic criteria for the preliminary identification of Fusarium, Paecilomyces, and Acremonium species by histopathology. Am J Clin Pathol. 109[1], 45-54. 2159. Guarro, J., Gams, W., Pujol, I., and Gene, J. (1997). Acremonium species: new emerging fungal opportunists--in vitro antifungal susceptibilities and review. Clin Infect Dis. 25[5], 1222-1229. 2167. Fridkin, S. K. and Jarvis, W. R. (1996). Epidemiology of nosocomial fungal infections. Clin Microbiol Rev. 9[4], 499-511. 2171. Schell, W. A. and Perfect, J. R. (1996). Fatal, disseminated Acremonium strictum infection in a neutropenic host. J Clin Microbiol. 34[5], 1333-1336. 2172. Perfect, J. R. and Schell, W. A. (1996). The new fungal opportunists are coming. Clin Infect Dis. 22 Suppl 2:S112-8., S112-S118. 2174. Fridkin, S. K., Kremer, F. B., Bland, L. A., Padhye, A., McNeil, M. M., and Jarvis, W. R. (1996). Acremonium kiliense endophthalmitis that occurred after cataract extraction in an ambulatory surgical center and was traced to an environmental reservoir. Clin Infect Dis. 22[2], 222-227. 2185. Cross, D. L., Redmond, L. M., and Strickland, J.

R. (1995).

Equine fescue toxicosis: signs and solutions. J Anim Sci. 73[3], 899-908. 2189. Porter, J. K. (1995). Analysis of endophyte toxins: fescue and other grasses toxic to livestock. J Anim Sci. 73[3], 871-880. 2192. Plumlee, K. H.

and Galey, F. D. (1994). Neurotoxic mycotoxins: a review of fungal toxins that cause neurological disease in large animals. J Vet.Intern Med. 8[1], 49-54. 2193. Ismail, M. A. (1994). Mycoflora inhabiting water closet environments. Mycoses. 37[1-2], 53-57. 2197. Jeffrey, W. R., Hernandez, J. E., Zarraga, A. L., Oley, G. E., and Kitchen, L. W. (1993). Disseminated infection due to Acremonium species in a patient with Addison's disease. Clin Infect Dis. 16[1], 170. 2199. Fincher, R. M., Fisher, J. F., Lovell, R. D., Newman, C. L., Espinel-Ingroff, A., and Shadomy, H. J. (1991). Infection due to the fungus Acremonium (cephalosporium). Medicine (Baltimore). 70[6], 398-409. 2203. Pasanen, A. L., Kalliokoski, P., Pasanen, P., Salmi, T., and Tossavainen, A. (1989). Fungi carried from farmers' work into farm homes.

Am Ind Hyg Assoc J. 50[12], 631-633. 2204. Alexopoulos, C., Mims, C., and Blackwell, M. (1996). Introductory mycology. -868. New York, John Wiley & Sons Inc. 2261. Patterson, R., Fink, J. N., Roberts, M., Kelly, J. F., and Sommers, H. M. (1978). Antibody activity in sera of patients with humidifier disease: studies of the water supply as a source of antigens. J Allergy Clin Immunol. 62[2], 103-108. 2342. Burge, H. A. (1985). Fungus allergens.

Clin Rev Allergy. 3[3], 319-329. 2809. Fischer, G., Schwalbe, R., Moller, M., Ostrowski, R., and Dott, W. (1999). Species-specific production of microbial volatile organic compounds (MVOC) by airborne fungi from a compost facility. Chemosphere. 39[5], 795-810. 2954. Velcovsky, H. G. and Graubner, M. (1981). [Allergic alveolitis following inhalation of mould spores from pot plant earth (author's transl)].

Dtsch.Med Wochenschr. 106[4], 115-120. 2968. Fiedler, K., Schutz, E., and Geh, S. (2001). Detection of microbial volatile organic compounds (MVOCs) produced by moulds on various materials. Int J Hyg Environ Health. 204[2-3], 111-121. 3095. Foundation for Allergy Research in Europe. (1984). Atlas of moulds in Europe causing respiratory allergy. Knud Wilken-Jensen et Suzanne Gravesen. -110. Danemark, ASK Publising. 3182. Bryden, W.

L. (2007). Mycotoxins in the food chain: human health implications. Asia.Pac.J Clin.Nutr. 16 Suppl 1:95-101., 95-101. 3184. Heberer, T., Lahrssen-Wiederholt, M., Schauzu, M., Braeunig, J., Goetz, M., Niemann, L., Gundert-Remy, U., Luch, A., Appel, B., Banasiak, U., Bol, G. F., Lampen, A., Wittkowski, R., and Hensel, A. (2007). Zero tolerances in food and animal feed -- are there any scientific alternatives? A European point of view on an international controversy. Toxicol.Lett. 175[1-3], 118-135. 3185. Kendra, D. F. and Dyer, R. B. (2007). Opportunities for biotechnology and policy regarding mycotoxin issues in international trade. Int.J Food.Microbiol. %20;119[1-2], 147-151. 3257. Brown, T.

P., Rottinghaus, G. E., and Williams, M. E. (1992). Fumonisin mycotoxicosis in broilers: performance and pathology. Avian. Dis. 36[2], 450-454. 3283. Larone, D H. (2002). Medically important fungi. A guide to identification. 4th edition, -409 p. New York - Amsterdam - London, Elsevier Science Publishing Co., Inc. 3284. Hollister-Stier (SPDR) (2002). All a statistical and the statisti

Laboratories. (2009). Allergenic extracts : Molds. Hollister-Stier Laboratories . 3285. Federal Drug Administration (FDA). (2008). Biological products deviation reporting (BPDR).

Non-blood product codes. 3-29-2009. 3730. Pharmacia Diagnostics AB. (2009). Allergy & autoimmunity. Diagnostics product catalogue 2009. internet , 1-48. Pharmacia. 3839. Yman, L. (1992). Molds & Yeasts: Allergen related documents.

ImmunoCAP InVitroSight , 1-3.

Pharmacia & Upjohn Diagnostics AB, Uppsala, Sweden. 3842. Kendrick, B. and Murase, G. (2003). Anamorph-teleomorph dabase. CBS. Centraalbureau voor Schimmelcultures. 2009. 3883. Wiszniewska, M., Walusiak-Skorupa, J., Pannenko, I., Draniak, M., and Palczynski, C. (2009). Occupational exposure and sensitization to fungi among museum workers. Occup.Med (Lond.). 59[4], 237-242. 3971. Robert, V., Stegehuis, G., and Stalpers, J. (2005). The MycoBank engine and related databases.

International Mycological Association .

International Mycological Association. 9-9-2009. 3972. Beguin, H. and Nolard, N. (1996). Prevalence of fungi in carpeted floor environment: analysis of dust samples from living-rooms, bedrooms, offices and school classrooms.

Aerobiologia 12, 113-120. 4080. Collier, L. (1998). Topley & Wilson 's Microbiology and Microbial Infections. Collier, L, Balows, A., and Sussman, M.

9th edition[6 vols]. Edward Arnold. 4274. Hunter, C. A., Grant, C., Flannigan, B., and Bravery, A. F. (1988). Mould in buildings: the air spora of domestic dwellings. International Biodeterioration 24[2], 81-101. 4286. Lockey, R. F. and Ledford, D. K. (2008).

Allergens and allergen immunotherapy. 4, -550. Informa Healthcare. 4305. lewis, R. A. (1998). Lewis' Dictionnary of Toxicology. -1127. Boca Raton , Fla, CRC Press. 4320. Escamilla Garcia, Beatriz. (1997). Étiologie comparative des maladies respiratoires d'une population d'enfants à Montréal et à Vancouver : regard sur les moisissures de l'air intérieur. Université de Montréal. 4321. Adelman, D. C., Casale, T. B., and Corren, J. (2002). Manual of allergy and immunology. 4, -544. 4322. Patterson, R., Fink, J. N., Miles, W. B., Basich, J.

E., Schleuter, D. B., Tinkelman, D. G., and Roberts, M. (1981). Hypersensitivity lung disease presumptively due to Cephalosporium in homes contaminated by sewage flooding or by humidifier water. J Allergy.Clin.Immunol. 68[2], 128-132. 4323. Hemken, R. W., Jackson, J. A., Jr., and Boling, J. A. (1984). Toxic factors in tall fescue. J Anim.Sci. 58[4], 1011-1016. 4324. Joost, R. E. (1995). Acremonium in fescue and ryegrass: boon or bane? A review. J Anim.Sci. 73[3], 881-888. 4325. Benkhelil, A., Grancher, D., Giraud, N., Bezille, P., and Bony, S. (2004). Intoxication par des toxines de champignons endophytes chez des taureaux reproducteurs. Revue Méd.Vét. 156[5], 243-247. 4326. Galey, F. D., Tracy, M. L., Craigmill, A. L., Barr, B. C., Markegard, G., Peterson, R., and O'Connor, M. (1991). Staggers induced by consumption of perennial ryegrass in cattle and sheep from northern California. J Am.Vet.Med Assoc. 199[4], 466-470. 4327. Strickland, J. R., Oliver, J. W., and Cross, D. L. (1993). Fescue toxicosis and its impact on animal agriculture. Vet.Hum.Toxicol. 35[5], 454-464. 4328. Garcia, M., Burt, J., Reilly, R., Kaul, R., Campbell, R., and Conly, J. M. (1994). Contamination of multi-dose vials of technetium-99Msestamibi with Acremonium kiliense.

Abstracts of the Association for Practitioners for Infection Control. 4329. Brown, N. M., Blundell, E. L., Chown, S. R., Warnock, D. W., Hill, J. A., and Slade, R. R. (1992). Acremonium infection in a neutropenic patient. J Infect. 25[1], 73-76. 4330. Eickhoff, T. C. (1994). Airborne nosocomial infection: a contemporary perspective. Infect. Control.Hosp.Epidemiol. 15[10], 663-672. 4331. Read, R. W., Chuck, R. S., Rao, N. A., and Smith, R. E. (2000). Traumatic Acremonium atrogriseum keratitis following laser-assisted in situ keratomileusis. Arch.Ophthalmol. 118[3], 418-421. 4332. Hay, R. J. and Mackenzie, D. W. (1982). The histopathological features of pale grain eumycetoma. Trans.R.Soc.Trop.Med Hyg. 76[6], 839-844. 4333. Popp, W., Braun, O., Zwick, H., Rauscher, H., Ritschka, L., and Flicker, M. (1988). Detection of antigen-specific antibodies on lung tissue in a patient with hypersensitivity pneumonitis. Virchows.Arch.A.Pathol.Anat.Histopathol. 413[3], 223-226. 4334. Bernstein, I. L., Li, J. T., Bernstein, D. I., Hamilton, R., Spector, S. L., Tan, R., Sicherer, S., Golden, D. B., Khan, D. A., Nicklas, R. A., Portnoy, J. M., Blessing-Moore, J., Cox, L., Lang, D. M., Oppenheimer, J., Randolph, C. C., Schuller, D. E., Ti (2008). Allergy diagnostic testing: an updated practice parameter. Ann.Allergy.Asthma.Immunol. 100[3 Suppl 3], S1-148. 4335. Martinez, Ordaz, V, Rincon Castaneda, C. B., Lopez, Campos C., and Velasco, Rodriguez, V. (1997). [Cutaneous hypersensitivity in patients with bronchial asthma in La Comarca Lagunera]. Rev.Alerg.Mex. 44[6], 142-145. 4336. Kruglov, A. G., Andersson, M. A., Mikkola, R., Roivainen, M., Kredics, L., Saris, N. E., and Salkinoja-Salonen, M. S. (2009). Novel Mycotoxin from Acremonium exuviarum Is a Powerful Inhibitor of the Mitochondrial Respiratory Chain Complex III. Chem.Res Toxicol.