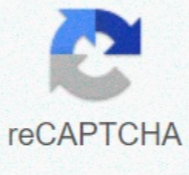




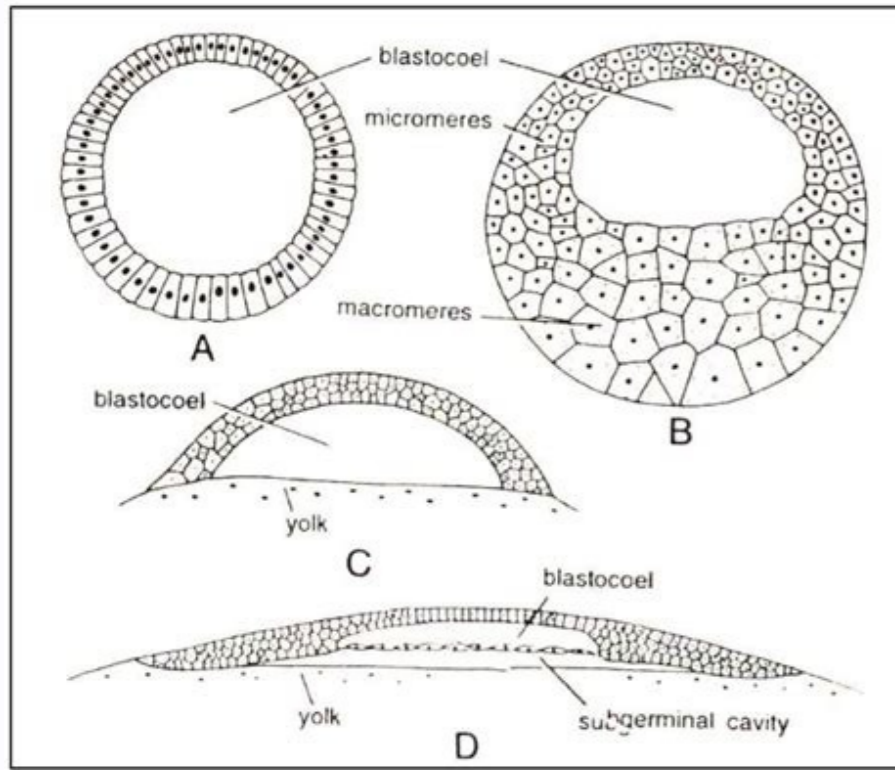
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## Blastulation in mammals pdf

If you're seeing this message, it means we're having trouble loading external resources on our website. If you're behind a web filter, please make sure that the domains \*.kastatic.org and \*.kasandbox.org are unblocked. Sphere of cells formed during early embryonic development in animals BlastulationBlastulation: from 1. morula to 2. blastulaDetailsDays4PrecursorMorulaGives rise toGastrulaIdentifiersMeSHD036703Anatomical terminology[edit on Wikidata] A. Morula and B. cross section of a blastula displaying the blastocoel and blastoderm of early animal embryonic development Blastulation is the stage in early animal embryonic development that produces the blastula. In mammalian development the blastula develops into the blastocyst with a differentiated inner cell mass and an outer trophoblast. The blastula (from Greek βλαστός (blastos meaning sprout)) is a hollow sphere of cells known as blastomeres surrounding an inner fluid-filled cavity called the blastocoel.[1][2] Embryonic development begins with a sperm fertilizing an egg cell to become a zygote, which undergoes many cleavages to develop into a ball of cells called a morula. Only when the blastocoel is formed does the early embryo become a blastula. The blastula precedes the formation of the gastrula in which the germ layers of the embryo form.[3] A common feature of a vertebrate blastula is that it consists of a layer of blastomeres, known as the blastoderm, which surrounds the blastocoel.[4][5] In mammals, the blastocyst contains an embryoblast (or inner cell mass) that will eventually give rise to the definitive structures of the fetus, and a trophoblast which goes on to form the extra-embryonic tissues.[3][6] During blastulation, a significant amount of activity occurs within the early embryo to establish cell polarity, cell specification, axis formation, and to regulate gene expression.[7] In many animals, such as *Drosophila* and *Xenopus*, the mid blastula transition (MBT) is a crucial step in development during which the maternal mRNA is degraded and control over development is passed to the embryo.[8] Many of the interactions between blastomeres are dependent on cadherin expression, particularly E-cadherin in mammals and EP-cadherin in amphibians.[7] The study of the blastula, and of cell specification has many implications in stem cell research, and assisted reproductive technology.[6] In *Xenopus*, blastomeres behave as pluripotent stem cells which can migrate down several pathways, depending on cell signaling.[9] By manipulating the cell signals during the blastula stage of development, various tissues can be formed. This potential can be instrumental in regenerative medicine for disease and injury cases. In vitro fertilisation involves the transfer of an embryo into a uterus for implantation.[10] Development The blastula stage of early embryo development begins with the appearance of the blastocoel. The origin of the blastocoel in *Xenopus* has been shown to be from the first cleavage furrow, which is widened and sealed with tight junctions to create a cavity.[11] In many organisms the development of the embryo up to this point and for the early part of the blastula stage is controlled by maternal mRNA, so called because it was produced in the egg prior to fertilization and is therefore exclusively from the mother.[12][13] Midblastula transition In many organisms including *Xenopus* and *Drosophila*, the midblastula transition usually occurs after a particular number of cell divisions for a given species, and is defined by the ending of the synchronous cell division cycles of the early blastula development, and the lengthening of the cell cycles by the addition of the G1 and G2 phases. Prior to this transition, cleavage occurs with only the synthesis and mitosis phases of the cell cycle.[13] The addition of the two growth phases into the cell cycle allows for the cells to increase in size, as up to this point the blastomeres undergo reductive divisions in which the overall size of the embryo does not increase, but more cells are created. This transition begins the growth in size of the organism.[3] The mid-blastula transition is also characterized by a marked increase in transcription of new, non-maternal mRNA transcribed from the genome of the organism. Large amounts of the maternal mRNA are destroyed at this point, either by proteins such as SMAUG in *Drosophila*[14] or by microRNA.[15] These two processes shift the control of the embryo from the maternal mRNA to the nuclei. Structure A blastula (blastocyst in mammals), is a sphere of cells surrounding a fluid-filled cavity called the blastocoel. The blastocoel contains amino acids, proteins, growth factors, sugars, ions and other components which are necessary for cellular differentiation. The blastocoel also allows blastomeres to move during the process of gastrulation.[16] In *Xenopus* embryos, the blastula is composed of three different regions. The animal cap forms the roof of the blastocoel and goes on primarily to form ectodermal derivatives. The equatorial or marginal zone, which composes the walls of the blastocoel differentiate primarily into mesodermal tissue. The vegetal mass is composed of the blastocoel floor and primarily develops into endodermal tissue.[7] In the mammalian blastocyst there are three lineages that give rise to later tissue development. The epiblast gives rise to the fetus itself while the trophoblast develops into part of the placenta and the primitive endoderm becomes the yolk sac.[6] In the mouse embryo, blastocoel formation begins at the 32-cell stage.



Diagrammatic comparison of blastula of an echinoderm (A), a frog (B), a fish (C) and a bird (D).

During this process, water enters the embryo, aided by an osmotic gradient which is the result of sodium-potassium pumps that produce a high sodium gradient on the basolateral side of the trophoblast. This movement of water is facilitated by aquaporins. A seal is created by tight junctions of the epithelial cells that line the blastocoel.[6] Cellular adhesion Main article: Cell adhesion Tight junctions are very important in embryo development. In the blastula, these cadherin mediated cell interactions are essential to development of epithelium which are most important to paracellular transport, maintenance of cell polarity and the creation of a permeability seal to regulate blastocoel formation. These tight junctions arise after the polarity of epithelial cells is established which sets the foundation for further development and specification.

Within the blastula, inner blastomeres are generally non-polar while epithelial cells demonstrate polarity.[16] Mammalian embryos undergo compaction around the 8-cell stage where E-cadherins as well as alpha and beta catenins are expressed. This process makes a ball of embryonic cells which are capable of interacting, rather than a group of diffuse and undifferentiated cells. E-cadherin adhesion defines the apico-basal axis in the developing embryo and turns the embryo from an indistinct ball of cells to a more polarized phenotype which sets the stage for further development into a fully formed blastocyst.[16] *Xenopus* membrane polarity is established with the first cell cleavage. Amphibian EP-cadherin and XB/U cadherin perform a similar role as E-cadherin in mammals establishing blastomere polarity and solidifying cell-cell interactions which are crucial for further development.[16] Clinical implications Fertilization technologies Experiments with implantation in mice show that hormonal induction, superovulation and artificial insemination successfully produce preimplantation mouse embryos. In the mice, ninety percent of the females were induced by mechanical stimulation to undergo pregnancy and implant at least one embryo.[17] These results prove to be encouraging because they provide a basis for potential implantation in other mammalian species, such as humans. Stem cells Blastula-stage cells can behave as pluripotent stem cells in many species. Pluripotent stem cells are the starting point to produce organ specific cells that can potentially aid in repair and prevention of injury and degeneration. Combining the expression of transcription factors and locational positioning of the blastula cells can lead to the development of induced functional organs and tissues. Pluripotent *Xenopus* cells, when used in an in vivo strategy, were able to form into functional retinas. By transplanting them to the eye field on the neural plate, and by inducing several mis-expressions of transcription factors, the cells were committed to the retinal lineage and could guide vision based behavior in the *Xenopus*.<sup>[18]</sup> See also Polarity in embryogenesis Diploblasty Triploblasty References ^ "Blastulation". web-books.com. ^ "Blastula". Encyclopædia Britannica. 2013. ^ a b c Gilbert, Scott (2010). *Developmental Biology* 9th Ed + Devbio Laboratory Vade Mecum3. Sinauer Associates Inc.

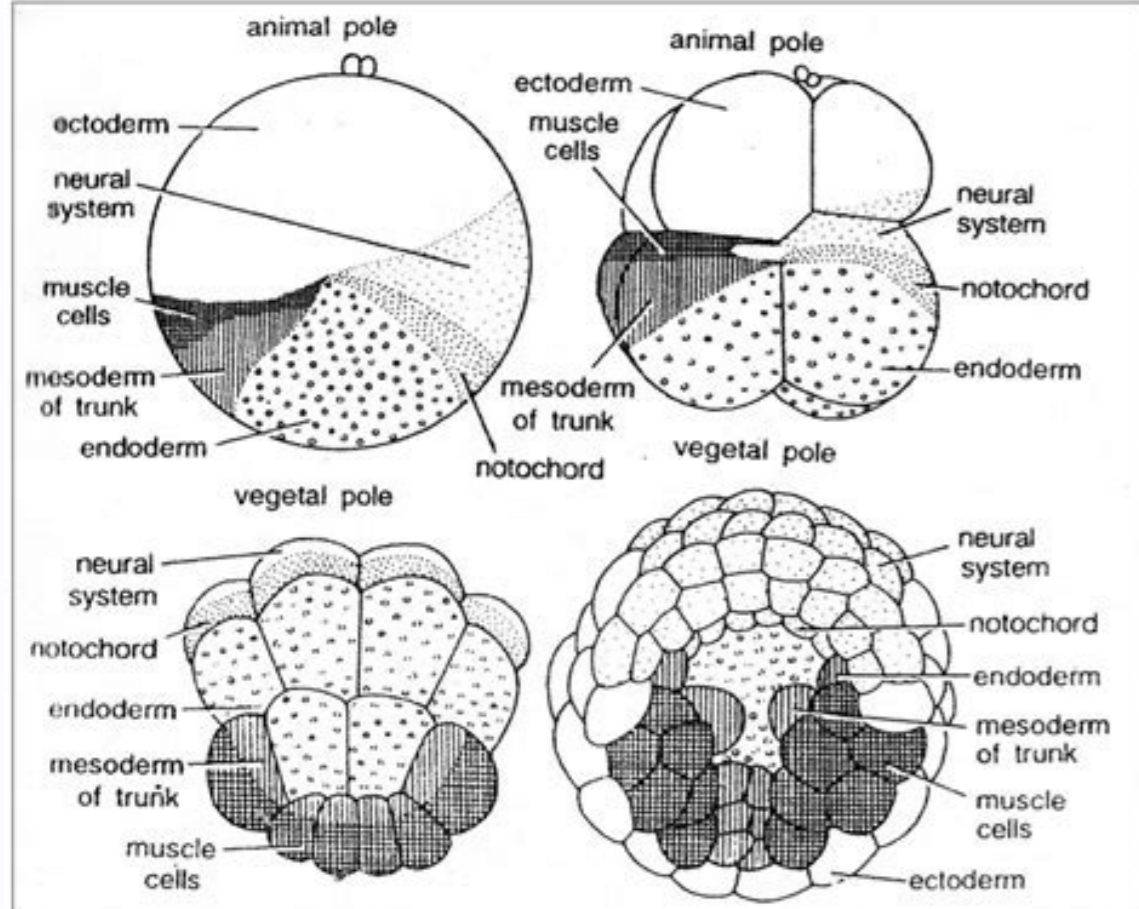
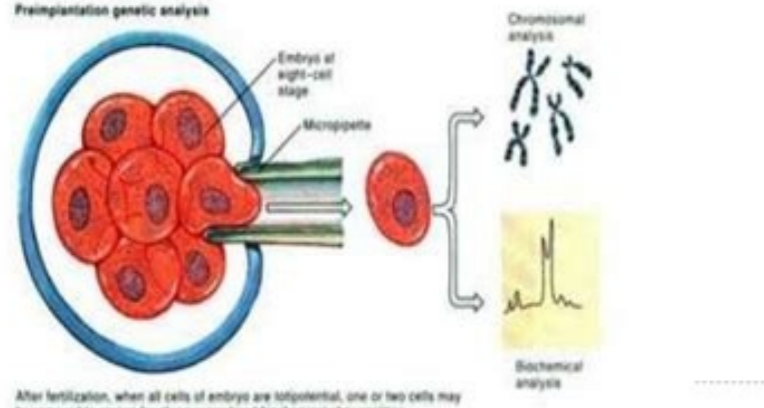


Fig. 3 : Cleavage of an ascidian egg showing bilateral cleavage.

pp. 243–247, 161. ISBN 978-0-87893-558-1.[permanent dead link] ^ Lombardi, Julian (1998). "Embryogenesis". *Comparative vertebrate reproduction*. Springer. p. 226. ISBN 978-0-7923-8336-9. ^ Forgács & Newman, 2005: p.

### PREIMPLANTATION GENETIC DIAGNOSIS

- Can be carried out 3 to 5 days after IVF of the oocyte.
- One or two cells (blastomeres) are removed from the embryo known to be at risk for a single gene defect



27 ^ a b c Cockburn, Katie; Rossant, Janet (1 April 2010). "Making the blastocyst: lessons from the mouse". *Journal of Clinical Investigation*. 120 (4): 995–1003. doi:10.1172/JCI41229. PMC 2846056. PMID 20364097.

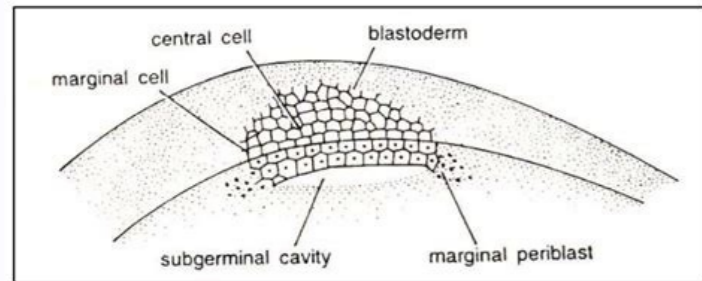
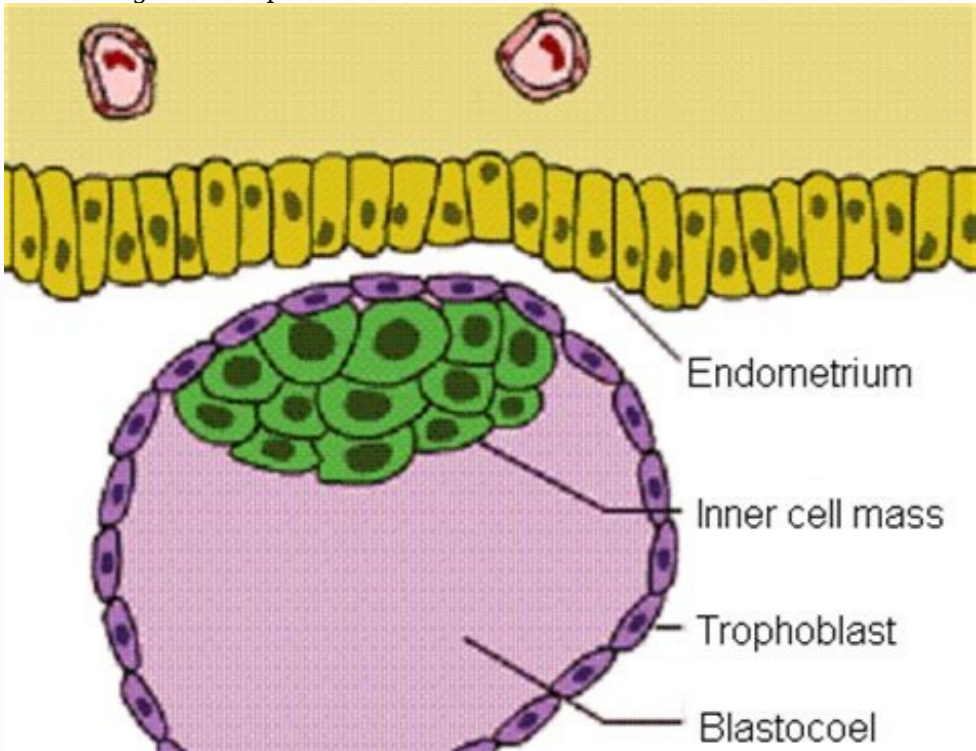


Fig. : Blastula stage of the chick embryo. The blastoderm (blastula) is shown cut transversely.

^ a b c Heasman, J (November 1997). "Patterning the *Xenopus* blastula".



Development. 124 (21): 4179–91. doi:10.1242/dev.124.21.4179. PMID 9334267.

^ Tadros, Wael; Lipshitz, Howard D. (1 March 2004). "Setting the stage for development: mRNA translation and stability during oocyte maturation and egg activation in *Drosophila*". *Developmental Dynamics*. 232 (3): 593–608. doi:10.1002/dvdy.20297. PMID 15704150. ^ Gourdon, John B.; Standley, Henrietta J. (December 2002). "Uncommitted *Xenopus* blastula cells can be directed to uniform muscle gene expression by gradient interpretation and a community effect". *The International Journal of Developmental Biology* (Cambridge, UK). 46 (8): 993–8. PMID 12533022. ^ Toth, Attila. "Treatment: Addressing the Causes of Infertility in Men and Women". Macleod Laboratory. Retrieved 22 March 2013. ^ Kalt, MR (August 1971). "The relationship between cleavage and blastocoel formation in *Xenopus laevis*. I. Light microscopic observations". *Journal of Embryology and Experimental Morphology*. 26 (1): 37–49. PMID 5565077. ^ Tadros, W; Lipshitz, HD (March 2005). "Setting the stage for development: mRNA translation and stability during oocyte maturation and egg activation in *Drosophila*". *Developmental Dynamics*. 232 (3): 593–608. doi:10.1002/dvdy.20297. PMID 15704150. ^ a b Etkin, LD (1988). "Regulation of the mid-blastula transition in amphibians". *Developmental Biology*. 5: 209–25. doi:10.1007/978-1-4615-6817-9\_7. ISBN 978-1-4615-6819-3. PMID 3077975. ^ Tadros, W; Westwood, JT; Lipshitz, HD (June 2007). "The mother-to-child transition". *Developmental Cell*. 12 (6): 847–9. doi:10.1016/j.devcel.2007.05.009. PMID 17543857. ^ Weigel, D; Izaurralde, E (24 March 2006). "A tiny helper lightens the maternal load". *Cell*. 124 (6): 1117–8. doi:10.1016/j.cell.2006.03.005. PMID 16564001. ^ a b c d Fleming, Tom P.; Papenbrock, Tom; Pesenko, Irina; Hausen, Peter; Sheth, Bhavanti (1 August 2000). "Assembly of tight junctions during early vertebrate development". *Seminars in Cell & Developmental Biology*. 11 (4): 291–299. doi:10.1006/scdb.2000.0179. PMID 10966863. ^ Watson, J.G. (October 1977). "Collection and Transfer of Preimplantation Mouse Embryos". *Biology of Reproduction*. 17 (3): 453–8. doi:10.1095/biolreprod17.3.453. PMID 901897. ^ Viczian, Andrea S.; Solessio, Eduardo C.; Lyou, Yung; Zuber, Michael E. (August 2009). "Generation of Functional Eyes from Pluripotent Cells". *PLOS Biology*. 7 (8): e1000174. doi:10.1371/journal.pbio.1000174. PMC 2716519. PMID 19688031. Bibliography Forgács, G. & Newman, Stuart A. (2005). "Cleavage and blastula formation". *Biological physics of the developing embryo*. Cambridge University Press. ISBN 978-0-521-78337-8. Cullen, K.E. (2009). "embryology and early animal development". *Encyclopedia of life science*, Volume 2. Infobase. ISBN 978-0-8160-7008-4. McGeady, Thomas A., ed. (2006). "Gastrulation". *Veterinary embryology*. Wiley-Blackwell. ISBN 978-1-4051-1147-8. Retrieved from "01:38Short Video: Sea Urchin Gastrulation Pearson 01:49Short Video: Sea Urchin Fertilization and Cleavage Pearson 21:22 05:41Neurulation (IB Biology) Alex Lee 01:59Fertilization Jason Amores Sumpter 13:09 08:06Embryology - Neurulation Armando Hasudungan 45:05Chapter 36 Animal Development Edward Kerschens 13:09 08:06Embryology - Neurulation Armando Hasudungan 11:35 00:53Embryology Animated - the First Three Weeks About Medicine 03:26SEA URCHIN DEVELOPMENT Kevin Maghinang 05:00Embryology | Fertilization, Cleavage, Blastulation Ninja Nerd 00:53Embryology Animated - the First Three Weeks About Medicine 03:26SEA URCHIN DEVELOPMENT Kevin Maghinang 00:47Sexual and Asexual Reproduction Explained Science Sauc 03:18General Embryology Review in 20 minutes Medical Animations 03:26SEA URCHIN DEVELOPMENT Kevin Maghinang 00:53Embryology | Fertilization, Cleavage, Blastulation Ninja Nerd 07:18Cleavage and Blastulation Jason Amores Sumpter 04:22Implantation and Gastrulation Jason Amores Sumpter 06:55Organogenesis and Neurulation Jason Amores Sumpter 13:09 08:06Embryology - Neurulation Armando Hasudungan 00:53Embryology Animated - the First Three Weeks About Medicine 03:26SEA URCHIN DEVELOPMENT Kevin Maghinang 05:00Embryology | Fertilization, Cleavage, Blastulation Ninja Nerd 00:53Embryology Animated - the First Three Weeks About Medicine