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Application of Nanoparticles to Control *Cuscuta* spp. in Horticultural Orchards: A Short Communication



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THE DODDER plants are well known as holo-parasitic stem weeds and belong *Cuscuta* genius (family Convolvulaceae), including about 200 species distributed worldwide. *Cuscuta* spp. have the ability to cause a serious damage to several agricultural and horticultural crops, which estimated to be than 300 million farmers in the world, causing severe losses in the yield of many crops. The main problem of dodder plants represents in penetrating host plant tissues by the haustoria of *Cuscuta* and uptake everything from the host plants including nutrients, water, macromolecules (i.e., proteins and mRNAs), and even pathogens through vascular connections. As parasitic weeds, several applied materials have been used to control the *Cuscuta* plants like nutrients (e.g., Ca, B, K, N, and Si) and herbicides, but the nanoparticles or nanomaterials are still need concern. This is a call for submission articles including original papers, reviews, notes on the expected role of nanoparticles like nano silica on control *Cuscuta* spp. in horticultural orchards.

Keywords: Nanoparticles, Nano-herbicides, Horticultural crops, Nano-nutrition, Nano silica.

1. Cuscuta sp. as a parasitic plant

As a parasitic plant, *Cuscuta* sp. can impact on the host plants and destroy their productivities by uptake the water, nutrients, proteins, metabolites, mRNAs, and pathogens like viruses (Albert et al. 2008; Zhou et al. 2021). A wide range of different crops can be infested by *Cuscuta* spp. including vegetables (e.g., asparagus, carrot, cucumber, garlic, onion, pepper, and potato), ornamental plants (e.g., chrysanthemum, dahlia, geranium, petunia, and periwinkle), and fruit trees (citrus, pomegranate, cranberry, coffee, etc.), causing a reduction in productivity and even death (Zaroug et al. 2014; Flores-Sánchez and Garza-Ortiz 2019). The genius *Cuscuta* includes more than 200 plant species (Table 1) such as *C. campestris*, *C. epithymum*, *C. europaea*, *C. gronovii*, *C. indecora*, *C. pentagona*, *C. planiflora*, *C. reflexa* and *C. suaveolens* (Wright et al. 2011, 2012; Riviere et al. 2013; Ali et al. 2017; Alamgeer et al. 2017; Aistova and Bezborodov 2017; Abu-Lafi et al. 2018; Al-Sultany et al. 2018; Flores-Sánchez and Garza-Ortiz 2019; Zhang et al. 2020; Al-Gburi 2021).

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| Cuscuta spp. | References |
|---------------------------|--|
| Cuscuta africana | Riviere et al. (2013); Ho and Costea (2018) |
| Cuscuta appendiculata | Wright et al. (2011) |
| Cuscuta approximata | Ho and Costea (2018) |
| Cuscuta attenuate | Mavlonov et al. (2008) |
| Cuscuta australis | Liu et al. (2020); Zhou et al. (2021) |
| Cuscuta campestris | Zaroug et al. (2014); Behdarvandi et al. (2015); Ho and Costea (2018); Wang et al. (2020); Al-Gburi (2021) |
| Cuscuta cephalanthi | McNeal et al. (2007b); Ho and Costea (2018) |
| Cuscuta chinensis | Marambe et al. (2002); Costea et al. (2011); Pan et al. (2013); Al-Sultany et al. (2018) |
| Cuscuta chapalana | Ho and Costea (2018) |
| Cuscuta compacta | McNeal et al. (2007b) |
| Cuscuta costaricensis | Banerjee and Stefanović (2019) |
| Cuscuta cotijana | Ho and Costea (2018) |
| Cuscuta epilinum | Costea and Tardif (2006); McNeal et al. (2007a) |
| Cuscuta epithymum | Costea and Tardif (2006) |
| Cuscuta europaea | McNeal et al. (2007a); Lukacova et al. (2019) |
| Cuscuta exaltata | McNeal et al. (2007a, b) |
| Cuscuta gronovii | Heide-Jorgensen (2008); Behdarvandi et al. (2015); Ho and Costea (2018) |
| Cuscuta indecora | McNeal et al. (2007a) |
| Cuscuta japonica | Ho and Costea (2018) |
| Cuscuta jepsonii | Costea and Stefanović (2009) |
| Cuscuta lupuliformis | McNeal et al. (2007a) |
| Cuscuta monogyna | Ho and Costea (2018) |
| Cuscuta nitida | McNeal et al. (2007a, 2009); Ho and Costea (2018) |
| Cuscuta obtusiflora | McNeal et al. (2007a, b) |
| Cuscuta odorata | Berg et al. (2003) |
| Cuscuta odontolepis | Write et al. (2012) |
| Cuscuta paitana | Riviere et al. (2013) |
| <i>Cuscuta palaestina</i> | Abu-Lafi et al. (2018) |
| Cuscuta pedicellata | Lyshede (1985, 1989) |
| Cuscuta pentagona | Costea et al. (2006b); Costea et al. (2015) |
| Cuscuta platyloba | Ahmad et al. (2017) |
| C.uscuta planiflora | Ho and Costea (2018) |
| Cuscuta polyanthemos | Wright et al. (2012) |
| Cuscuta purpurata | McNeal et al. (2007a); Ho and Costea (2018) |
| Cuscuta reflexa | Albert et al. (2010); Patel et al. (2012); Ali et al. (2017); Thomas et al. (2017) |
| Cuscuta rostrata | McNeal et al. (2007a) |
| Cuscuta rugosiceps | Costea et al. (2008) |
| Cuscuta tinctoria | Ahmad et al. (2017) |
| Cuscuta umbellate | Ho and Costea (2018) |
| Cuscuta umbrosa | Costea and Tardif (2006); Costea et al. (2006a) |
| Cuscuta veatchii | McNeal et al. (2007a) |
| Cuscuta warneri | Ho and Costea (2018) |

 TABLE 1. List of some plant species of genus "Cuscuta" as reported by several workers

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Cuscuta spp. cause significant reduction in the yield of some infested crops as reported in many studies such as the reduction of yield of tomato (Lycopersicum esculentum) and carrot (Daucus carota), which infested by C. pentagona to 50-75% and by 70-90%; respectively (Lanini and Kogan 2005). This reduction in yield differs from crop to other and depending on the species of Cuscuta as reported for the following crops Allium cepa (28-58%), Capsicum frutescens (60-65%), Cicer arietinum (86%), Lens culinaris (87%), Medicago sativa (60–70%), and Vigna mungo (31-34%) by dodder invasion (Mishra 2009, Zaroug et al. 2014). Many plant viruses could be transferred by the *Cuscuta* spp. as reported by Sastry et al. (2019) such as tobacco rattle virus by Cuscuta campestris in Bulgaria (Dikova 2006), tomato yellow leaf curl virus by Cuscuta spp. in Spain and Cyprus (Papayiannis et al. 2011), Arabis mosaic virus by Cuscuta spp. in Iran (Ghotbi and Shahraeen 2005), tomato yellow ring virus by Cuscuta spp. in Iran (Ghotbi et al. 2005). Figs. 1 and 2 shows the death impact of Cuscuta spp. on growth some horticultural plants.



Fig. 1. The infection by *Cuscuta europaea* on the ornamental plant of *Chrysanthemum morifolium* could present in the following stages: (A) normal plant, (B) first stage of infection, (C) sever plants injury, and (D) plants died after infection



Fig. 2. An overview for the infection on the ornamental plant of *Vitex trifolia* by *Cuscuta europaea* (B) compared to normal plant (A)

2. Nano-control of Cuscuta europaea infection

Several approaches have been applied to control these parasitic plants including herbicides (Angeles Alvarez et al. 2016), different nutrients like K (potassium sulphate), Ca (Ca compounds), B (boric acid), N (urea), and Si (Al-Gburi 2021). Cuscuta as weeds can control using the mechanical methods (removing, pruning, or deep-ploughing), chemical control using herbicides, and biological control (like bioherbicides, microbial herbicides, and genetically modified plants expressing herbicidal substances (Westwood et al. 2018). Some insects could be used in the biological control of dodders like the genus Smicronyx including Coleoptera and Curculionidae (Aistova and Bezborodov 2017; Flores-Sánchez and Garza-Ortiz 2019). Some recent reports confirmed the using of nanotechnology for delivery of natural therapeutic substances like Cuscuta chinensis, which its extract is rich in lignin and flavonoid with antioxidative and hepatoprotective effects (Saka and Chella 2021). Nanotechnology can overcome the poor solubility and permeability, low stability of the pharmacological agents by using the nanosuspension (Saka and Chella 2021). On the other hand, some nanomaterials like nano silica may be has the proper impact as nano-herbicide in control the Cuscuta. Concerning the using of nanoparticles or nanomaterials in Cuscuta control is important approach that needs more concerns. This call by EBSS journal for submission articles including original articles, reviews or comments on this issue.

Ethics approval and consent to participate

This article does not contain any studies with human participants or animals performed by any of the authors.

Consent for publication

All authors declare their consent for publication.

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The MS has been written, revised and agreed by all authors

Conflicts of Interest

The author declares no conflict of interest.

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