Revision of the Australian genus Lawrencia Hook. (Malvaceae: Malveae)

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Abstract

Lander, N. S. Revision of the Australian genus Lawrencia Hook. (Malvaceae: Malveae). Nuytsia 5 (2): 201-271 (1984). Lawrencia Hook. (including species formerly placed in the genus Selenothamnus Melville) is revised. The genus contains 12 species; four of these are described for the first time: L. buchananensis from north-eastern Queensland, L. viridi-grisea from Western Australia and the Northern Territory, and L. chrysoderma and L. cinerea which are both endemic to Western Australia. A new infrageneric classification is presented and its taxa described. Each species is fully described; its distribution, habit and flowering period are indicated where appropriate; its conservation status is assessed. Several species are illustrated. A key to infrageneric taxa and species of Lawrencia is provided. The comparative morphology of the genus is discussed and related to that of the tribe Malveae in general and the Plagianthus alliance in particular. It is suggested that diocey in the genus has evolved in response to pollinator-mediated selection. The dispersal mechanisms of species of Lawrencia are discussed. It is hypothesized that the genus had a littoral origin where its initial diversification occurred followed by dispersal to the margins of inland salt lakes, further diversification and colonization of the arid zone. Plagianthus monoicus Helms ex Ewart is excluded from the Malvaceae and synonymized with Ricinocarpos velutinus F. Muell. (Euphorbiaceae).

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Introduction

The taxa considered in this study are Australian members of the so-called *Plagianthus* alliance in the tribe Malveae of the family Malvaceae (Bates 1968). In the last definitive treatment of this group, Melville (1966) distributed its species amongst six genera. Perhaps because of the limited material at his disposal, Melville's generic descriptions are rather brief and provide an insufficient basis for more than a superficial comparison of morphological

features or for proper circumscription of his taxa. Species descriptions were not presented by Melville. The lack of effective keys together with the presence in Australia of several hitherto unrecognized species has led to much confusion in herbarium collections.

The present revision is based on a reassessment of morphological characters of species previously consigned to the genera Lawrencia and Selenothamnus. Despite considerable variation in habit, trichome type, sexuality and fruit and seed morphology, it is considered that the division between Lawrencia and Selenothamnus can no longer be upheld and that the 12 species concerned are best placed in a single genus. Although clearly delimitable groups emerge, these do not correspond to previous generic concepts, nor does the evidence seem strong enough to support their recognition at generic level. A possible exception is L. berthae, a species so much at variance morphologically with those remaining in the genus that a strong case could be made for its segregation. This has been deferred pending further investigation of the genera of the Plagianthus alliance.

Taxonomic History

The genus Lawrencia was erected by Hooker (1840) to accommodate a single species, L. spicata, specimens of which had been sent to him from Tasmania by Ronald Gunn. Hooker (1842) described a second species, L. glomerata, sent to him from Western Australia by James Drummond. A third species, L. squamata, was added to the genus by Miquel (1845).

Bentham (1862) united under *Plagianthus* J. R. & G. Forst. (1776) non Jaub. & Spach (1845) species variously placed until that date in *Asterotrichion* Klotzsch, *Blepharanthemum* Klotzsch, *Halothamnus* F. Muell., *Hoheria* A. Cunn, *Lawrencia* and *Plagianthus*. This action was followed by subsequent authors, including Bentham in Bentham & Hooker (1862), Bentham (1863), Baker (1892), Schumann (1890), Dalla Torre & Harms (1900-7), Lemeé (1934) and Burbidge (1963). Several new species of *Plagianthus* sensu lato were described during this period.

In January of 1909, O. Stapf exhibited specimens and lantern slides of *Plagianthus helmsii* to members of the Linnean Society of London pointing out that it appeared more natural to him to treat that species together with *P. microphyllus* and *P. squamatus* as members of a distinct genus for which F. Mueller's name *Halothamnus*, orginally applied to *P. microphyllus*, would have to stand (Anon. 1909). Amongst those participating in the ensuing discussion were E. G. Baker and T. A. Sprague.

Sprague and Summerhayes (1926) examined the New Zealand representatives of *Plagianthus* sensu lato which they assigned to two genera, *Plagianthus* sensu stricto and *Hoheria*, noting in passing that the Australian species of *Lawrencia* and *Halothamnus* should be excluded from *Plagianthus*.

In the late 1930s, E. G. Baker prepared the draft for a paper which described three putative new species of *Lawrencia* and provided a partial key to the genus. From his draft it is clear that Baker intended to reinstate the genus *Lawrencia*. It is worthy of note that *Plagianthus berthae* and *P. helmsii* were excluded from consideration, although the significance of this is not explained. Copies of his unpublished manuscript are held at BM and PERTH. Baker annotated specimens held at BM, K and PERTH.

In a more comprehensive study of the *Plagianthus* complex, Melville (1966) divided its species amongst six genera. In particular, species placed by Bentham (1862) in *Plagianthus* section *Lawrencia* were now allocated to two separate genera, namely *Lawrencia* sensu stricto and *Selenothamnus*, founded, as Stapf had originally suggested, on Mueller's

Halothamnus, first described in 1862. These two genera were distinguished by Melville on the basis of differences in habit, vestiture and the nature of the thickening of the mericarp walls. Melville's classification was adopted by Hutchinson (1967), Baines (1981) and Mitchell (1983). In his review of generic relationships in the tribe Malveae of the Malvaceae, Bates (1968) declined to comment on Melville's realignment of the *Plagianthus* complex, although he accepted it as one of his informal sub-tribal alliances, the *Plagianthus* alliance.

Barker (1981) drew attention to the overlapping of Melville's floral diagnostic characters and questioned the separation of *Selenothamnus* and *Lawrencia*.

Materials and Methods

The taxonomic decisions made in this paper were based upon a study of the gross morphology of all available herbarium specimens examined at or loaned by the major Australian and relevant British herbaria, and on collection and field observation of species present in Western Australia. The methods and procedures adopted were those summarized by Leenhouts (1968). Measurements listed are based upon the total variation observed in the herbarium specimens cited at the end this paper.

The total geographic range of Lawrencia illustrated in Map I and the distribution of all the species in Maps 3-8 respectively were compiled by recording their occurrence in one degree squares superimposed on a Bonnes Equal Area Projection of Australia or on a Lambert Zenithal Equal Area Projection of Western Australia where a species is confined to that state. In the taxonomic treatment that follows, the distribution of each species is indicated by reference to the maps and to the geographical sub-divisions generally adopted by botanists in each state, namely Anon. (1972) for Queensland, Anon. (1979) for South Australia, Beard (1980) for Western Australia, Beauglehole (1980) for Victoria, Chippendale (1971) for the Northern Territory, and Pickard & Dlugaj (1981) for New South Wales. There is no comparable treatment of Tasmania.

A complete list of herbarium specimens examined and annotated in the course of this study is provided in abbreviated form at the conclusion of this paper. The more detailed lists in the following taxonomic treatment are intended primarily to permit the location of populations at sites throughout the range of each species. With the exception of *Lawrencia viridi-grisea*, all available specimens of species newly described here are cited in full. Localities given in single inverted commas are pastoral stations. In the absence of a collector's number the date of collection is cited where available; herbarium register or sheet numbers are cited only in otherwise ambiguous cases.

I regret to have to report that all the specimens, including types, from both the National Herbarium of Victoria (MEL) and the Tasmanian Herbarium (HO) cited in this paper were destroyed when the truck returning loan material to their home states left the road and caught fire. Fortunately, all the type specimens involved are represented either by isotypes or syntypes elsewhere. In the text, recently distributed duplicates not involved in this accident are indicated by an asterisk. A comprehensive list of all the destroyed specimens with localities, collectors' names and dates of gathering, together with an indication of available duplicates may be found in Lander (in press).

Comparative Morphology

This section is concerned with comparative aspects of morphology rather than attempting to provide an exhaustive account of the morphology of *Lawrencia*. Emphasis is placed on the range of variation of significant morphological patterns common to all members of the genus. The morphological traits considered here provide a basis for the ensuing classification of *Lawrencia* and for the general discussion of their taxonomic and evolutionary significance which follows the formal taxonomic presentation.

Habit and duration. The branching pattern of Lawrencia is monopodial. In the seedling stage all species produce a single actively growing main stem. The main axis of the young plant normally soon loses its dominance and several to many primary branches, often basal, achieve the same stature. Secondary branching is particularly frequent as the inflorescences are produced and during the second year.

Six species are suffrutescent: Lawrencia diffusa is diminutive and prostrate, L. repens is decumbent, L. densiflora (Figure 6A), L. cinerea, L. glomerata (Figure 4) and L. spicata (Figure 3) are conspicuous, erect sub-shrubs. The herbaceous stems of these species become soft-woody with age. Occasionally, such plants die back or are burnt or grazed back to a ligneous branched caudex at ground level from which new growth is initiated.

The remaining six species are small shrubs which may be weakly ascending little-branched plants, such as Lawrencia buchananensis and L. viridi-grisea (Figure 5), or divaricate shrubs, such as L. berthae (Figure 2A) and L. squamata. In L. helmsii and, to a lesser extent, in L. chrysoderma (Figure 7A) marked contracton of densely ramulose secondary stems gives the plants their distinctive appearance. Shrubby species normally produce new growth from erect woody stems.

Although various species of *Lawrencia* have been reported on labels of some herbarium specimens to be annual, it appears that all species are perennial, the few instances of annual duration being attributable to the plants flowering in their first year. All of the taxa observed in the field appear to reach reproductive maturity in their first year with adequate rainfall.

Thorns. All but one species of Lawrencia are unarmed. Specimens of L. squamata often have prominent thorns. These are tapered, depauperate branches.

Vestiture. Variation in trichome types and their distribution is invaluable in recognizing taxa at all levels in Lawrencia. In this genus, as in many others in the tribe Malveae, the usual multiradiate stellate hair of the Malvaceae has apparently been modified to produce a wide range of forms which grade imperceptibly into one another.

The commonest hair type encountered is represented by the haplomorphic (multiangulate), multicellular stellate hairs found in all species with the exception of the glabrous Lawrencia spicata and glabrous plants of L. diffusa and L. glomerata. Haplomorphic stellate hairs (Figure 1A) are the predominant hair type found in L. berthae, L. cinerea, L. densiflora, L. diffusa, L. glomerata and L. repens in which they occur on stems, stipules, leaves and the abaxial surfaces of calyces, invariably in conjunction with bifurcate and simple hairs into which they merge.

Haplomorphic stellate hairs are found on the abaxial surfaces of petals in Lawrencia cinerea, L. densiflora and L. glomerata. They are present to a lesser extent on all other pubescent species of Lawrencia, although nearly lacking in some.

On the stems, petioles and peduncles of Lawrencia berthae, haplomorphic stellate hairs are often distinctly raised on wart-like mounds of epidermal and possibly sub-epidermal tissue thus forming tufted stellate hairs (Figure 1D), a phenomenon observed in no other species of Lawrencia. Haplomorphic stellate hairs are also found in L. berthae at the junction of the style branches and on the surface of the pericarp.

Actinomorphic (flattened radiate), multicellular stellate hairs with more or less basally coalescent rays on a multicellular base or stalk (Figure 1E) are the predominant hair type found in *Lawrencia viridi-grisea* and in many populations of *L. squamata*. In these species, such hairs are found on stems, stipules, leaves and the abaxial surfaces of calyces, occurring

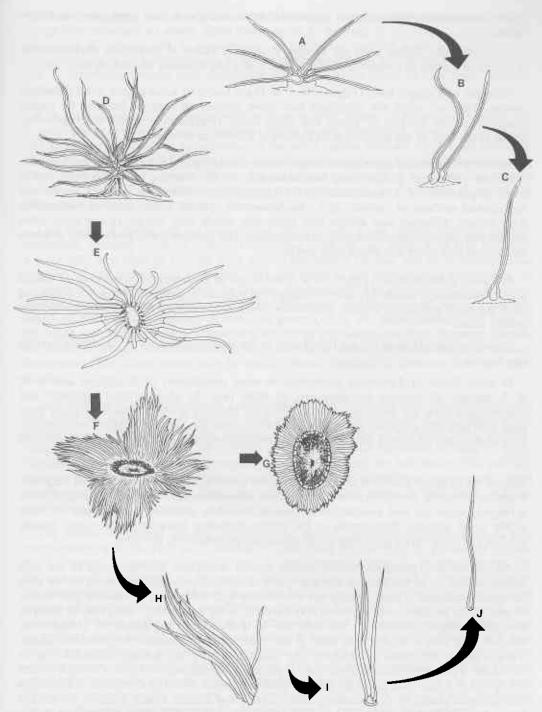


Figure 1. Elaboration and reduction of stellate hairs in Lawrencia. A — Unspecialized haplomorphic stellate hair of L. densiflora. B — Reduction: birfurcate hair of L. densiflora. C — Simple hair of L. densiflora. D — Multiplication of rays: haplomorphic stellate hair with raised base L. berthae. E — Plantation and reorientation: actinomorphic stellate hairs of L. viridi-grisea. F — Coalescence of rays: fimbriate peltate scale of L. chrysoderma. G — Entire peltate scale of L. helmsii. H — Reduction and reorientation: palmate scale of L. chrysoderma. I — Palmate scale of L. chrysoderma. J — simple hair of L. chrysoderma.

in conjunction with haplomorphic stellate, bifurcate and simple hair types into which they merge.

Actinomorphic stellate hairs are present to a lesser extent in *Lawrencia buchananensis* in which they occur in conjunction with peltate scales and bifurcate and simple hair types.

Bifurcate and simple hairs (Figure 1B, \mathbb{C} & J) are found to some degree in all pubescent species. They are often the dominant hair types present on stipules, but can be readily observed on the margins of leaves and calyx lobes. Intermeshing simple hairs are often found at the base of the petals of male flowers of *Lawrencia berthae*.

Minutely fimbriate or entire multicellular peltate scales (Figure 1F & G) are the predominant trichome type found in Lawrencia buchananensis, L. chrysoderma and L. helmsii, and in many populations of L. squamata. Such trichomes may cover stems, stipules, leaves and the abaxial surfaces of calyces. In L. buchananensis, peltate scales occur in conjunction with stellate, bifurcate and simple hair types into which they merge. In the three other species with peltate scales, these occur in conjunction with palmate scales, and stellate, bifurcate and simple hair types into which they merge.

Bract-like palmate scales (Figure 1H & I) are found on Lawrencia chrysoderma, L. helmsii and L. squamata in which they often dominate the vestiture of stems and branches, extending onto the basal portion of stipules and leaves where they can clearly be seen to merge into peltate scales.

Glandular hairs (Figure 2I) are found only in the sepaline nectaries of Lawrencia berthae (see below).

In some forms of Lawrencia glomerata, in most populations of L. diffusa, and in all of L. spicata, all vestiture has been lost. In other taxa the above-ground vegetative and reproductive parts are variously clothed. Thus the vestiture gives the plants or plant parts their characteristic textures: smooth and glabrous, scabrous, velvety or shiny and armour-like. Most of the trichomes are colourless, vitreous and rigid at maturity; none can penetrate skin.

Leaves. As in many genera of the Malvaceae, the leaves of Lawrencia are spirally disposed, stipulate, petiolate, subsessile or sessile, and have actinodromous venation. Although spirally arranged, reduction and contraction of lateral branches, observed to a greater or lesser extent in all species, often results in the leaves appearing fasciculate, sometimes densely so. The leaf form varies according to the taxon and placement on the stem.

All species bear persistent, paired lateral stipules which are shortly adnate to the base of the petiole, or to the base of the leaf blade of sessile leaves, and appressed to the stem or slightly reflexed. These stipules are translucent and colourless, or opaque and brown or pale green, membranous, succulent and leaf-like, or dry and spongy. They may be filiform, triangular, elliptic, obovate or sub-orbicular in shape. They are minute or conspicuous, 0.4-10.0 mm long x 0.2-3.6 mm wide at the base, and symmetrical or somewhat falcate. Their margins are entire or more or less ciliate and they are apically acuminate, acute or obtuse. Their vestiture resembles that of the stem and leaves, but there is often a greater frequency of simple and bifurcate hairs as is noted above. With the exception of *L. spicata* and glabrous plants of *L. glomerata*, both abaxial and adaxial stipule surfaces are usually hairy, but the adaxial surface is typically less densely hairy and often glabrous or nearly so. Stipules may be rather variable in any one species.

By virtue of their presence or absence, length, cross-sectional outline and vestiture the petioles of Lawrencia are of considerable diagnostic value. The leaves may be distinctly

petiolate throughout as in L. diffusa and L. repens, or else distinctly petiolate basal leaves merge into subsessile or sessile floral leaves as in L. berthae, L. cinerea, L. densiflora, L. glomerata, L. spicata and L. viridi-grisea, or the leaves may be more or less subsessile throughout as in L. buchananensis or sessile throughout as in L. chrysoderma, L. helmsii and L. squamata. Petioles may exceed the leaf blade by as much as four times, notably in L. spicata.

The petioles are basically triquetrous in cross-section. Those of Lawrencia berthae and L. viridi-grisea are triquetrous throughout. Whilst mostly triquetrous, those of L. densiflora and L. glomerata are often canaliculate; in the latter species they are often quite flattened. Those of L. diffusa and L. repens are canaliculate throughout, becoming flattened only towards the base. In L. spicata the petioles are either triquetrous or else somewhat swollen and hence trigonal. The latter tendency is most pronounced in L. cinerea in which the petioles are semiterete throughout.

Leaf blades of Lawrencia are symmetrical and variously linear, elliptic, cuneate, deltoid, spathulate, obovate, ovate or orbicular in outline, with the size range 1-70(115) mm long x 0.7-35(60) mm wide in size. In most species the blades are flat, but in L. chrysoderma (Figure 7B), L. cinerea and L. helmsii they are more or less conduplicate throughout. In L. squamata the leaf blades may be flat or conduplicate. The blade-bases are variously cuneate, acute, obtuse, truncate or cordate. The blade margins may be entire, shallowly lobed or more or less coarsely dentate. In general, they are entire towards the base of the blade with shallow lobing or toothing becoming prominent and more frequent towards the apex. A notable exception to this is L. cinerea in which the blade margin is dentate throughout. The blade apices may be acute, obtuse, rounded or truncate; they are often lobed.

Leaf venation is actinodromous in *Lawrencia*. The mid-vein is the most strongly developed and there are 1-3 additional pairs of veins radiating from the base of the blade. These primary veins extend towards the margins, often terminating in marginal lobes or teeth. Secondary veins branch from the primary veins and may also terminate in lobes or teeth. Smaller reticulate veins occur throughout the remainder of the leaf blade. The primary veins are sometimes completely obscure, sometimes indistinct, sometimes conspicuously raised on the abaxial surface of the blade and slightly impressed above in both fresh and dry specimens.

The leaves of several species of Lawrencia exhibit heteroblastic development. This phenomenon was observed in L. cinerea, L. densiflora (Figure 6A), L. glomerata (Figure 4), L. repens, L. spicata (Figure 3) and L. viridi-grisea (Figure 5). In these species the leaves vary in their degree of petiolation and their blade size and shape corresponding to their placement on the plant. Mature vegetative leaves produced on the primary stems during periods of most active growth are distinctly petiolate, the blades are at their largest, and lobing or toothing of the margins is at its maximum extent. In comparison, leaves produced on primary stems later in the season or on lateral branches have shorter petioles and their blades are smaller and narrower with marginal lobing or toothing reduced in prominence and frequency, particularly towards the base. The final leaves produced with the flowers are usually subsessile or sessile and their blades are by far the smallest on the plants with lobing or toothing highly reduced, often confined to the apical region. These tendencies are most dramatically expressed in L. spicata (Figure 3), most subtly in L. repens.

Heteroblastic leaf-development was not observed in the remaining species in all of which the leaves are more or less uniform throughout in any one plant. The leaf blades of all but one of the species without developmental heterophylly lack marginal lobes or teeth,

the only exception being certain populations of *L. squamata* in which they are 3-5-dentate towards the apex.

Inflorescences. Like other genera in the tribe Malveae, inflorescence patterns in Lawrencia are rather plastic and hence easily modified according to the age and vigour of the individuals. Thus, inflorescences often exhibit striking differences in expression when compared at different stages of development, even on a single plant.

With the exception of a single species, the flowers of Lawrencia are subsessile or sessile, solitary, born in the axils of leaves and their associated stipules. Flowers produced later in the season are often congested towards the stem apices. In many species reduction of floral leaves and of the internodes produce terminal and axillary sub-spicate inflorescences. Further reduction of axillary sub-spicate inflorescences located in the upper part of the plant results in the congested terminal spicate inflorescence observed in fully mature individuals of L. densiflora, L. glomerata and L. spicata (Figure 3), as indeed the names of these species suggest. In all of these species each flower is subtended by a single leaf.

One species is exceptional: the flowers of *Lawrencia berthae* are long-pedicellate. The first flowers are borne near the base of the plant and are usually solitary in the leaf-axils, but later in the season they are borne in axillary fascicles of 2-5 subtended by a single leaf (Figure 2A).

Flowers. An epicalyx is lacking in all species of Lawrencia. Peculiarities of calyx, corolla, androecium and style morphology are discussed below.

It is worth noting at the outset that three stages of sexual differentiation are found in Lawrencia:

- 1. Hermaphroditism: flowers of all plants bisexual. This condition is found in L. buchananensis, L. diffusa, L. repens, L. spicata (Figure 3) and L. viridi-grisea.
- 2. Polygamodioecy: flowers of male plants with larger spreading calyx and corolla and a sterile vestigial gynoecium; flowers of female plants with smaller erect calyx and corolla, with or without sterile anthers; flowers of bisexual plants with erect calyx and corolla, as large as those of males or almost so, with fertile anthers and gynoecium. This condition is found only in *L. glomerata*.
- 3. Complete dioecy: flowers of male plants with larger spreading calyx and corolla, either lacking gynoecium or with a sterile vestigial gynoecium; flowers of female plants with smaller erect calyx and corolla, with or without sterile anthers. This condition is found in *L. berthae* (Figure 2B & C), *L. chrysoderma* (Figure 7C, D & F), *L. cinerea*, *L. densiflora* (Figure 6B, D & E), *L. helmsii* and *L. squamata*.

All bisexual flowers of *Lawrencia* are protandrous. The style branches are closely appressed in early flower and are exserted from the filament tube. Anther dehiscence occurs well before the stigmas separate and become receptive. As the flower begins to wilt the style branches reflex into the anther cluster: it is thus theoretically possible for self-pollination to occur at this stage.

Calyx. The persistent calyx in Lawrencia is composed of five sepals that are plicate and valvate in bud and connate for part of their length. At anthesis the basal united portion is turbinate or broadly to very broadly obconic; the lobes are flat, erect, narrowly to broadly triangular, and obtuse, acute or acuminate, and they lack accessory teeth.

The calyces range from 1.5-4.8 mm in length from pedicel to lobe apex. Species may sometimes be distinguished on the basis of calyx length alone. For example, *Lawrencia viridi-grisea* has calyces 3.8-5.3 mm long and *L. buchananensis* has calyces 7.0-10.0 mm long. Since the range in calyx length in most species overlaps either in whole or part that in other species, size alone is of limited use diagnostically.

The degree of fusion of the sepals varies widely between species, ranging from 1/4-9/10 their length. The extent of overlap is so great that this character also is of little value diagnostically. However, it is usually possible to separate *Lawrencia densiflora*, with its sepals connate 1/2-4/5 their length, from *L. repens*, with its sepals connate 1/4-1/2 their length.

In dioecious species there is a significant difference in both the calyx length and the degree of connation of the sepals between male and female flowers. The calyces of male flowers are generally longer than those of female flowers and tend to be fused for a greater proportion of their length. This syndrome is seen at its most extreme in *Lawrencia cinerea* in which male calyces are 5.0-7.5 mm long with sepals connate for 1/2-3/4 their length, whereas female calyces are 3.0-5.0 mm long with sepals connate for only 1/10-1/4 their length.

Externally, calyces may be glabrous as in Lawrencia spicata and some forms of L. glomerata and L. diffusa, but usually they display the full species-specific range of hair types. Internally, calyces are glabrous with the exception of female calyces of L. spicata which are sometimes densely stellate-hairy and both male and female calyces of L. berthae which have a few scattered apical hairs. L. berthae is also exceptional in possessing tufts of multicellular papillae on the basal adaxial surfaces of male calyces which appear to be glandular and are interpreted here as nectaries (Figure 21 & J).

The venation of the sepals is actinodromous and reticulate. It may be obscure or distinct with 3-5 primary sub-parallel veins.

Corolla. The corolla of Lawrencia consists of five antisepalous petals which are adnate to the filament tube above and connate below to form a delicate, membranous, transparent ovary cup. As the fruit ripens the connate region of the petals dehisces along its sutures to release the spent corolla which remains loosely attached merely by means of the style passing through the staminal tube. At anthesis the corolla is campanulate or more or less rotate.

Measured from base to lobe apex the corolla length ranges from 2-12 mm. Although there is a mode characteristic of each species, the degree of overlap is such that it is not usually possible to distinguish between species on the basis of this measurement alone. However, *Lawrencia buchananensis*, with a corolla length of 6-8 mm, can easily be separated from *L. chrysoderma* (3.0-5.0 mm), *L. repens* (3.0-5.0 mm) and *L. squamata* (1.7-5.6 mm).

The degree of fusion of the petals varies widely between species. Petals may be free almost to their base or connate for as much as 4/5 their length. This character overlaps too often to be of much value diagnostically, although each species has its characteristic mode.

Corolla lobes of *Lawrencia* are imbricate in bud. At anthesis they may be obovate, ovate or elliptic, and spreading and flat or more or less erect and carinate. In width (measured at the widest point) corolla lobes range from 0.6-7.0 mm. Despite considerable overlap between species this measurement is of somewhat more use diagnostically than corolla length or the degree of fusion of petals. In particular, it serves to distinguish *L. berthae* (2.7-7.0 mm) from *L. buchananensis* (2.2-2.5 mm), *L. cinerea* (0.9-2.5 mm), *L. chrysoderma*

(0.8-2.0 mm), L. diffusa (1.2-1.5 mm), L. helmsii (0.5-2.0 mm), L. repens (1.1-1.8 mm) and L. viridi-grisea (1.6-2.6 mm).

Auricles, basal wing-like outgrowths at the base of the petals of many species of Malvaceae, are entirely lacking in *Lawrencia*.

The apices of corolla lobes are acute, obtuse or truncate and may be entire, emarginate or irregularly dentate. The last feature was observed only in *Lawrencia squamata* and *L. chrysoderma*.

The corolla lobes are usually glabrous on both surfaces. There are several notable exceptions to this: corolla lobes of *Lawrencia berthae* sometimes have scattered, intermeshing simple hairs in the basal area on both abaxial and adaxial surfaces; those of *L. cinerea* are stellate-hairy along the mid-veins on the abaxial surface, weakly so in male fowers, strongly in females; those of *L. densiflora* are often sparsely stellate-hairy abaxially in male flowers, densely in females; those of *L. glomerata* can be stellate-hairy along the abaxial mid-vein.

In dioecious species there are striking differences between male and female flowers in corolla length, the degree of fusion of the petals, corolla lobe width, shape and orientation, and, in species with stellate-hairy petals, in the extent of their vestiture. The corollas of male flowers are generally larger than those of female flowers; their petals, which are almost free or only slightly fused, are flat and spreading at anthesis and, if stellate-hairy, then sparsely so. Corollas of female flowers are smaller by comparison; their petals, which are fused to a far greater extent, tend to be carinate and erect at anthesis and, if stellate-hairy, then densely so.

In colour, the corollas are white, cream-coloured, pale yellow or pale green, and may be tinted red or purple. Corolla colour is often quite variable within a single species.

The petals are membranous and elaborately vascularized with actinodromous reticulate veins which anastamose before reaching the margins. They thus resemble to a remarkable degree the wings of certain insects.

One species, Lawrencia berthae, is notable for its sweetly scented flowers, a character detected in no other species in the genus.

Androecium. The androecium of Lawrencia is composed of monadelphous stamens which arise from five groups of primordia (Heel 1966). This five-ranked symmetry is usually not evident at anthesis when it is obscured by the densely packed nature of the anther cluster. Only in L. glomerata is the apex of the filament tube ever distinctly five-lobed. In male and bisexual flowers of Lawrencia, the filaments are united into a tube 0.4-7.0 mm long and are free at their apices for an additional 0.2-2.0 mm. In female flowers, the filaments are united for 0.1-2.6 mm and, if they have anthers (sterile), are free at their apices for an additional 0.1-0.9 mm. Within these two measurements in each case there is a continuum. The filament tube is identical in colour to the corolla and is glabrous in all species with the single exception of L. berthae in which scattered simple hairs are often present.

Often the free portions of filaments are paired. At an extreme, this results in two anthers terminating a single filament, a condition frequently observed in *Lawrencia glomerata* and *L. repens*.

The number of anthers in each flower varies considerably between taxa. The lowest number of anthers encountered is 5, found regularly in flowers of *L. diffusa* and sometimes encountered in specimens of *L. glomerata*, *L. repens* and *L. helmsii*. The highest number observed is

in *L. berthae* which has 10-30 anthers in each flower. Some species have a constant number of anthers in each male or bisexual flower, namely *L. densiflora* (20), *L. diffusa* (5) and possibly *L. buchananensis* (20). The remaining species have a variable number of anthers in each flower. Anthers are normally white but in *L. berthae* they are very occasionally faintly pink.

In three of the dioecious species, namely Lawrencia chrysoderma, L. helmsii and L. squamata, anthers are present in female flowers, but are smaller in size than in male flowers and lack pollen. In the first two of the above-mentioned species, anthers of female flowers are fewer than in male flowers. In the remaining dioecious species, female flowers lack anthers altogether.

Variation in characters of the androecium of *Lawrencia* can be useful in distinguishing species. Nonetheless, there is a continuum in this variation between the various species in the genus. One species stands apart from the others, namely *L. berthae* with its often simple-hairy filament tube and generally freer filaments.

Styles. As in all genera of the Malvaceae, the style of Lawrencia is tubular at the base where it is continuous with the columella; above, it is divided into distinct branches. In Lawrencia, the total length of the style varies from 0.4-10.0 mm in bisexual and female flowers. Each species is modally characterized by its style length, and this character can occasionally be of assistance in distinguishing between closely related species, notably L. chrysoderma, with styles 2.0-7.0 mm long, and L. helmsii, with styles 0.4-3.2 mm long.

The presence or absence of a gynoecium in male flowers is often diagnostic in Lawrencia. Of the dioecious species, L. chrysoderma, L. cinerea, L. helmsii and L. squamata may lack gynoecia in male flowers. Conversely, they are always present in male flowers of L. berthae, L. glomerata and L. densiflora. When a gynoecium is present in male flowers the styles are much reduced, ranging from 1.2-4.0 mm in length.

Branching of the styles may occur from the base itself to as far as 7/10 of the distance above the apex of the columella. In Lawrencia diffusa, L. glomerata and L. viridi-grisea, branching always occurs from the base: thus, the presence of branches which are only shortly united or united at some considerable distance from the style base is a useful diagnostic aid. In all species the number of branches equals that of the carpels, that is 2-11. This is, perhaps, the most useful character provided by the styles and is treated further in the consideration of fruit below.

At anthesis the style branches are more or less flattened and divergent. They are most usually filiform, but in four species they are often clavate, namely in Lawrencia berthae, L. chrysoderma, L. helmsii and L. squamata. The frequently broadly clavate styles of L. chrysoderma (Figure 7E) are often apically lobed, a most useful character in distinguishing that species from L. helmsii. In width the style branches vary from 0.1-1.0 mm. In colour they may be white, yellow, pink, purple or brown. The dark purple stigmas of L. squamata are unique in the genus.

In all but one species of *Lawrencia* the style is completely glabrous externally. That of *L. berthae* is conspicuously stellate-hairy at the junction of its branches.

Fruit. The mature ovary of Lawrencia is a complex oblate or ovoid schizocarp. It consists of a single whorl of mature carpels or mericarps supported by a central columella, subtended by a membranous carpocrater (see below) and enclosed by the peristent calyx. The number of mericarps always corresponds to the number of style branches. The pedicel, calyx and columella do not abscise from the inflorescence axis. In most cases the withered corolla persists for a while.

The variable fruit morphology encountered in Lawrencia is usefully summarized as follows:

- A. Fruit with indehiscent mericarps; pericarp rugose or reticulate.
 - 1. Mericarps 9-11, all fertile; pericarp with apical and distal walls stellate-hairy, lateral walls coriaceous and rugose or obscurely reticulate. This condition is found in *L. berthae*. See Figure 2D, G & E.
 - 2. Mericarps 5, all fertile; pericarp glabrous, lateral and distal-basal walls membranous, becoming distinctly reticulate with fenestrate or alveolate areolae. This condition is found in *L. buchananensis*, *L. glomerata*, *L. spicata* and *L. viridi-grisea*. See Figures 3 (4, 6 & 7) and 4 (4 & 5).
- B. Fruit with dehiscent mericarps; pericarps unornamented.
 - 3. All mericarps more or less well-developed.
 - (a) Mericarps 5, all fertile. This condition is found in L. diffusa and L. repens.
 - (b) Mericarps 2-5, only 1 fertile. This condition is found in *L. densiflora* (mericarps 2-5) and *L. cinerea* (mericarps 2-3). See Figure 6F & G.
 - 4. Sterile mericarps undeveloped and obliterated by developing fertile mericarps.
 - (c) Mericarps 2-5, only 1 or 2 fertile. This condition is found in *L. squamata* (mericarps 2-3, 1 or 2 fertile), *L. chrysoderma* (mericarps 2-3, only 1 fertile) and *L. helmsii* (mericarps 2, only 1 fertile). See Figure 7F & G.

The mericarp is traditionally the most important structure used to determine taxa in the tribe Malveae and keys to the taxa are generally dependent on its character. In *Lawrencia* the number of mericarps, their fertility, development, dehiscence, shape and size, and the vestiture, colour, texture and ornamentation of the pericarp are of great diagnostic value.

In number the mericarps of *Lawrencia* vary from 2 to 11 in each fruit. For a particular species this number may be constant or else of limited range.

In a given species of *Lawrencia* all the mericarps may be fertile, or else only one or two mericarps in each fruit produce mature seed. Mericarps containing sterile, aborted or unfertilized seed continue to develop more or less normally in all but three species: in these they regularly fail to develop and are crushed and obliterated by the developing fertile mericarps. In the latter case each developing mericarp assumes the swollen irregularly globular shape of the seed it contains.

In those fruit in which all mericarps are well developed these are shaped roughly like a wedge from an inverted pudding basin. There are four surfaces to each mericarp: two lateral walls, an apical wall, and a continuous distal-basal wall. There is a single, nearly straight proximal margin and two curved lateral margins.

Individual mericarps are either dehiscent or indehiscent. Indehiscent mericarps separate regularly at maturity as their pericarp becomes light, dry and stiff. In one species, *Lawrencia berthae*, the pericarp is black, conspicuously hairy on its apical and distal-basal walls and coriaceous and rugose or obscurely reticulate on its lateral walls. In other species with indehiscent mericarps the pericarp is always glabrous, white or tan, thin and delicate, becoming translucent at maturity with a pronounced reticulum. The areolae enclosed by the thickenings

of the reticulum may be of firm texture, but more often are thin and translucent (fenestrate), and in some cases no tissue is evident and the areolae are quite open (alveolate). In species with indehiscent mericarps the lateral margins of the pericarp are often projected slightly to form narrow wings which may connect across the apical-distal region to give the distal wall a distinctive shield-like appearance. The apical region of the pericarp projects somewhat to form a short, acute cusp.

When indehiscent mericarps separate from the columella along the proximal margin an opening forms in the apical proximal region where the funicle is attached to the columella. Thus the seed is no longer connected to the mericarp. This opening is far too small to allow the seed to escape.

In species with dehiscent mericarps, dehiscence takes place by the operation of two mechanisms, namely complete or partial opening of the suture along the proximal edge of the pericarp coupled with disintegration or weakening and rupture of the basal region of the pericarp. These species have a pericarp which is smooth and glabrous and becomes translucent or transparent at maturity.

All mericarps in *Lawrencia* are uniovular. No trace of an endoglossum was observed in any species. An endoglossum is an internal tongue-like appendage dividing the mericarp partially or completely into two cells, present in many Malvaceae.

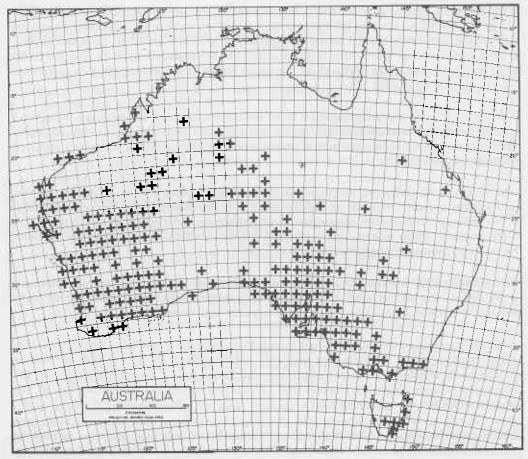
The columella is a generally persistent cylindrical, clavate or peltate structure at the centre of the calyx to which the mericarps and style are attached (Figure 2F). Near its base is an outgrowth which is chartaceous centrally and membranous marginally known as the carpocrater, a term coined by Hochreutiner (1920). The carpels are completely free of this latter structure but are subtended by it.

Seed. Lawrencia seeds are basically reniform in lateral view with their proximal apical lobe projected into a short blunt or acute cusp. They have a thick, firm testa that is completely glabrous. The outer layer of the seed coat may produce a thin waxy layer giving the seed a glaucous appearance. This layer is often lacking or worn away by the time the seed is shed. The colour of the seed coat varies from grey, to light brown, chestnut brown or black. See Figures 2H, 4(6 & 7), 6H & 7G.

In species with fruit in which several mericarps are fertile, seed shape is determined by the shape of the enclosing pericarp and hence is triquetrous. In species in which the pericarp ruptures early and only one or two seeds in each fruit are fertile these seeds become swollen and irregularly globular as they mature. Seeds range in size from 0.6-3.3 mm high x 0.4-2.5 mm wide. Seed size overlaps to such an extent that it is of virtually no use in distinguishing between taxa.

A single seed is produced in each mericarp to which it is attached apically by its funicle. The seed is almost completely filled by the embryo which is apically pendulous with its radicle uppermost. Only a thin layer of endosperm surrounds the embryo. See Figure 3(8 & 9).

In all but three species the seed surface is smooth and featureless. In *Lawrencia densiflora* the entire seed surface is conspicuously rugose (Figure 6H); in *L. repens* only the distal surface is rugose. The lateral seed surfaces of both these species are prominently angular, characteristic also of *L. diffusa* which is otherwise featureless.



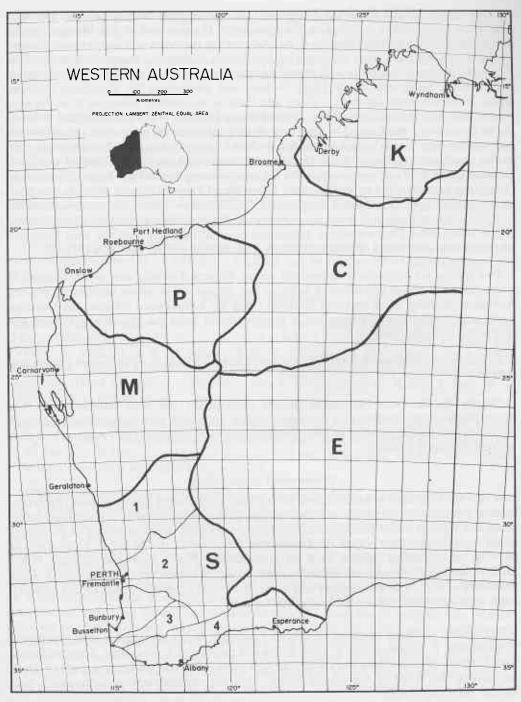
Map 1. Distribution of the genus Lawrencia.

Phytogeography

Lawrencia is endemic to Australia, extending from 18° S latitude southwards (Map 1). It occurs in all States but principally in Western Australia where 11 of the 12 species are to be found, six of which are endemic to that State. Its species commonly inhabit the margins of salt lakes, saline depressions and estuaries.

For convenience, in the following taxonomic treatment species distributions are indicated by reference to the sub-divisions indicated above which are generally adopted by botanists in each State. However, it has been found more meaningful in Western Australia to summarize these distributions in terms of the drainage systems they occupy. Mulcahy and Bettenay (1972) recognize six drainage divisions within Western Australia. Their classification is refined by Bettenay and Mulcahy (1972) and further elucidated by Beard (1973). These divisions and the lake systems within the South West Division are indicated in Map 2. Lawrencia occurs in all drainage divisions.

Two species are peripheral to the Great Plateau. Lawrencia cinerea is confined to the Great Sandy Desert and Little Sandy Desert in the Canning Division (Map 6). L. viridigrisea occurs along the coastline of the Monger System of the South West Division, and of the Murchison, Pilbara and Canning Divisions; in the Canning Division it extends inland to the Great Sandy Desert, thence to the Northern Territory (Map 4).



Map 2. Drainage Divisions in Western Australia:
C. Canning Division
K. Kimberl
E. Eucla Division
M. Murchi

- K. Kimberley Division M. Murchison Division
- P. Pilbara Division
- S. South West Division

- Drainage Systems in the South West Division:
 1. Monger System
 2. Avon System

- 3. Blackwood System
- 4. South Coast System

After Mulcahy & Bettenay (1972), Bettenay & Mulcahy (1972) and Beard (1973).

Seven species inhabit the Great Plateau and the coastal plain fringing its eastern margin. Lawrencia chrysoderma is found in the Murchison Division and in the Monger System of the South West Division (Map 8). L. berthae inhabits the Avon and South Coast Systems of the South West Division, although its distribution is disjunct, reappearing in the Eyre Peninsula of South Australia and extending into western Victoria (Map 3). L. densiflora occurs in the north west of the Eucla Division and throughout the Murchison Division, where it extends onto the coastal plain; and there is an isolated collection from between Nickol Bay and the De Grey River in the Pilbara Division (Map 6). L. diffusa is found in the far west of the Eucla Division as well as in the eastern part of the Avon System and in the South Coast System of the South West Division (Map 6).L. helmsii is restricted to the south-eastern corner of the Murchison Division and the western half of the Eucla Division (Map 8). L. repens occupies the far south-western area of the Eucla Division and the eastern part of the Avon System of the South West Division (Map 6). With the exception of an isolated specimen from Loongana on the Nullarbor Plain, L. spicata is confined to river banks, swamps and estuaries on the Swan Coastal Plain and South Coast Systems of the South West Division with a disjunction in its distribution until the Eyre Peninsula whence it skirts the coasts of South Australia, Victoria and eastern Tasmania (Map 4).

Two species are particularly widespread within Western Australia and in other mainland states, too. Lawrencia glomerata is more or less ubiquitous in saline habitats in Western Australia occurring in all drainage divisions (Map 5). L. squamata occupies the Canning, Eucla and South West Divisions but is notably absent from the Murchison, Pilbara and Kimberley Divisions (Map 7).

Lawrencia buchananensis is known only from Lake Buchanan in north-eastern Queensland (Map 4).

Despite the fact that the species occupy substantially the same habitats, that their geographical ranges overlap and that often several are found growing together or in close proximity, no natural hybrids or intermediates have been observed: this suggests that they are reproductively isolated.

Taxonomic Considerations

In this work, taxa have been defined on the basis of correlated morphological characters. Such a procedure is invariably arbitrary in that taxa may be chosen to encompass larger or smaller units of variation. In particular, I have attempted to define species of *Lawrencia* in the light of character variation documented for other of the Malveae, particularly in recently monographed genera such as *Anisodontea Presl*, *Kearnemalvastrum Bates*, *Malacothamnus E*. Greene and related groups (Bates 1967, 1969) and *Malvastrum A*. Gray (Hill 1982 a, b, c). This has resulted in a somewhat broad definition of the species in which many localized variations are not recognized formally.

Taxonomic problems certainly remain in Lawrencia below the species level, notably in L. glomerata, L. squamata and L. viridi-grisea and possibly in L. densiflora. Their resolution would require extensive field work supplemented by experimental procedures, both of which are beyond the scope of the present study. Thus the establishment of infraspecific taxa has not been undertaken.

Cytogenetic evidence has not been considered. It is likely that in the wider context of the presently developing classification of the Malveae (Bates 1968, 1969, Bates & Blanchard, 1970, Bates pers comm.) this line of investigation will prove important in elucidating the relationships of the *Plagianthus* alliance to other alliances in the tribe.

Within the sequence of species examined here are major discontinuities in the presence of sepaline nectaries, the presence of a floral scent, the vestiture of the filament tube, style branches and pericarp, and the number of carpels in each fruit. Further, these discontinuities are distinctly correlated and hence have prompted recognition of the new subgenus *Panifex* pending further study of the *Plagianthus* alliance (see Key to Infrageneric Taxa and Species below).

In the subgenus Lawrencia there is minor variation in many characters, in some of which discontinuities have been the basis for generic recognition elsewhere in the tribe Malveae. This variation is correlated sufficiently to allow the recognition of several infrageneric taxa serving to emphasize the heterogeneity in these taxonomically important characters as well as providing a useful means of grouping species within the genus (see Key to Infrageneric Taxa and Species below). Species of the series Selenothamnus have been segregated as the genus Selenothamnus by Melville (1966): I believe his approach to be inconsistent with the morphological evidence presented above and discussed in more general terms below.

The infrageneric classification offered here is phylogenetically based only insofar as the morphological similarities on which it is founded may be interpreted to suggest propinquity of descent.

Taxonomic Treatment

Lawrencia Hook., Hooker's Icon. Pl. 3: t. 261 & 262 (1840); Melville, Kew Bull. 20: 514 (1966); Hutchinson, Gen. Fl. Pl. Dicot. 2: 52 (1967); J. A. Baines, Austral. Pl. Gen. 338-9 (1981; A. Mitchell, in B. D. Morley & H. R. Toelken, Fl. Pl. Australia 126-9 (1983). Wrenciala A. Gray, U.S. Explor. Exped., Bot. 1: 181 (1854), nom. inval. Plagianthus section Lawrencia (Hook.) Benth., J. Linn. Soc., Bot. 6: 97 (1862); E. G. Baker, J. Bot. 30: 72 (1892). Type: L. spicata Hook.

Halothamnus F. Muell., Pl. Victoria 1: 158-9 (1862) non Jaub. & Spach, Ill. Pl. Orient. 2: 50, t. 136 (1845). Selenothamnus Melville, Kew Bull. 20: 514-5 (1966); Hutchinson, Gen. Fl. Pl. Dicot. 2: 53 (1967); J. A. Baines, Austral. Pl. Gen. 338-9 (1981). Type: H. microphyllus F. Muell. [Lawrencia squamata Nees ex Miq.].

Prostrate, decumbent or erect, perennial sub-shrubs or shrubs to 180 cm tall, monopodial and simple, little-branched and divaricate or else densely ramulose; dioecious, polygamodioecious or hermaphroditic. Above-ground vegetative and reproductive parts glabrous or variously hairy with multicellular stellate trichomes often modified to form haplomorphic stellate, actinomorphic stellate, bifurcate or simple hairs and peltate and palmate scales; texture smooth, scabrous, velvety or shiny and armour-like. Stems sometimes with a spongy periderm producing dry fibrous or flaky bark. Leaves alternate, often fasciculate on much-reduced lateral branches, subsessile, sessile or distinctly petiolate, stipulate, often heteroblastic with large subsessile or distinctly petiolate basal leaves merging into smaller. subsessile or sessile stem and floral leaves; stipules persistent, free from petiole when present, shortly adnate to the petiole-base or to the blade-base of sessile leaves, minute or conspicuous, variously filiform, linear, triangular, ovate, elliptic, obovate or suborbicular, 0.4-10.0 mm long x 0.2-3.6 mm wide at the base, colourless, pale green, or brown, delicate and membranous, more or less succulent and leaf-like or dry and spongy, margin entire or more or less ciliate. apex variously acuminate, acute or obtuse; petiole lacking, minute or up to 400 mm long, slender or stout, basically triquetrous but often canaliculate, almost flat, trigonal or semiterete; blade variously linear, elliptic, cuneate, deltoid, spathulate, obovate, ovate or orbicular, 1-70(115) mm long x 0.7-35(60) mm wide, flattened or more or less conduplicate, sometimes falcate, sometimes swollen and succulent, venation actinodromous, obscure or more or less conspicuously 1-7 veined with one pronounced mid-vein, veins distinctly raised below and

somewhat depressed above, base variously cuneate, acute, obtuse, truncate or cordate, margin entire, coarsely dentate or obtusely lobed throughout or towards the apex only, apex variously acuminate, acute, obtuse, rounded or truncate, often lobed. Flowers pedicellate to subsessile or sessile, 2-5 in axillary fascicles subtended by a single leaf, or solitary and axillary, often in densely leafy clusters on reduced lateral branches or at stem apices to form congested spicate inflorescences, unisexual or bisexual, more or less sexually dimorphic in dioecious species, usually odourless, rarely sweetly scented. Pedicels lacking, very short and stout, or slender and up to 4.5 cm long. Epicalyx lacking. Calyx turbinate, 1.0-12.0 mm long at anthesis, green, sepals 5, connate 1/10-9/10 their length, often strongly plicate in bud, adaxially glabrous, rarely densely stellate-hairy or with a few simple apical hairs; lobes erect at anthesis, triangular, 1.0-4.8 mm wide at the base; accessory teeth lacking; apices variously obtuse, acute or acuminate; nectaries usually lacking, rarely with basal tufts of minute, nectariferous, multicellular papillae on adaxial surfaces; venation reticulate, obscure or distinct with 3-5 primary subparallel veins to each lobe. Corolla campanulate, broadly campanulate or more or less rotate at anthesis, exceeding or subequal to the calyx, 2-12 mm long, membranous, white, yellow or tinged green, red or purple, entirely glabrous or else abaxially stellate- or simple-hairy, rarely adaxially hairy, adnate to the base of the filament tube to form an ovary cup; petals 5, almost free to their bases or connate up to 4/5 their length, glabrous or else abaxially simple- or stellate-hairy, rarely adaxially hairy; auricles lacking; lobes imbricate in bud, erect or spreading at anthesis, obovate, ovate or elliptic, 0.6-7.0 mm wide, flat or more or less carinate; apices variously acute or obtuse, and entire, emarginate or irregularly dentate. Androecium of male and bisexual flowers with 5-30 unranked stamens; filament tube shorter than the petals and matching them in colour, 0.4-7.0 mm long, glabrous or rarely subglabrous with scattered simple hairs; free portion of filaments 0.2-2.0 mm long; anthers dorsifixed, terminal, white, extrorse; androecium of female flowers more or less reduced with anthers lacking or smaller in size, fewer in number and sterile. Gynoecium of female and bisexual flowers with a single style branching from its base or from up to 7/10 its length from the base into 2-11 branches equal in number to the carpels; style 0.4-10.0 mm long, glabrous or stellate-hairy at the junction of the branches; branches divergent at anthesis, filiform, linear or clavate, rarely broadly clavate and lobed, 0.1-1.0 mm wide, variously white, yellow, pink or brown; stigmas introrsely decurrent on the style branches which are papillate over their whole length or on the upper portion only; carpels in a single discoid whorl attached by their proximal margins to a central, apically expanded columella, subtended by a thin, membranous carpocrater; ovules solitary in each carpel, pendulous, attached to the columella above the carpel notch; gynoecium of male flowers more or less reduced with carpels lacking or fewer in number, smaller in size and sterile. Fruit an oblate or ovoid schizocarp, 1.4-4.7 mm high x 0.8-6.2 mm in diameter, exposed or enclosed by the calyx, readily or tardily shed from the shrunken columella and calvx at maturity; mericarps 2-11, completely to partially dehiscent or indehiscent, free from one another, laterally compressed and wedge-shaped or, if sterile, then often crushed and obliterated by the developing mericarp(s); pericarp coriaceous, chartaceous or membranous, black, tan, white or translucent, becoming rugose, more or less reticulate or smooth and featureless at maturity, glabrous, rarely stellate-hairy, with lateral margins sometimes projected slightly to form narrow wings which may connect apicallydistally to form a distinctive shield, and the apical region often projected to form a short, acute cusp; endoglossum lacking. Seeds solitary in each mericarp, attached apically to the columella by a funicle, triquetrous and basically reniform in lateral view or irregularly globular, 0.6-3.3 mm high x 0.4-2.5 mm in radial width, variously grey, brown or black, glabrous, sometimes glaucous with a layer of wax, smooth or more or less rugose, sometimes with angular lateral faces, with only a thin layer of endosperm surrounding the embryo. Columella filamentous, slender or stout at maturity, 1.2-4.5 mm high x <0.1-2.0 mm in diameter. A genus of 12 species confined to Australia.

The name *Lawrencia* honours Robert William Lawrence (1807-1833), a young settler and botanical collector in the northern part of Tasmania who, until his untimely death at the age of 26, communicated specimens to Sir William Hooker (Hooker 1855, Sharr 1978).

Gray (1854 loc. cit.) pointed out that Lawrencia Hook. is antedated by Laurencia Lemouroux in Ann. Mus. Hist. Nat. Paris 20: 130 (1813), a red seaweed. Gray suggested the anagram Wrenciala to replace Lawrencia. There is no basis for such a substitution: the names Lawrencia and Laurencia are not orthographic variants; they have not been a source of persistent error and are most unlikely to become so. This matter is discussed fully by Melville (1966 loc. cit.).

Key to infrageneric taxa and species

- 1. Prostrate, decumbent or erect sub-shrubs, or erect shrubs; flowers subsessile or sessile, axillary, always solitary, often in densely leafy clusters; sepals adaxially glabrous; petals never with basal hairs; style branches glabrous at their junction; mericarps 2-5 in each fruit; pericarps glabrous. (SUBGENUS LAWRENCIA.)
 - 2. Erect sub-shrubs or shrubs; female flowers of dioecious species lacking anthers altogether (in *L. glomerata*, a polygamodioecious species, sterile anthers may be present); mericarps indehiscent, 5 in each fruit, all fertile; pericarps laterally reticulate with fenestrate or alveolate areolae; seed triquetrous, smooth. (SECTION LAWRENCIA.)
 - 3. Short-lived, suffrutescent, simple perennial; vestiture stellate-hairy or glabrous.
 - 4. Plant strongly rank-smelling; to 180 cm tall with one or several erect stout stems diverging from a conspicuous basal rosette, slender secondary stems contracted to form dense terminal flowering spikes; glabrous throughout; hermaphroditic.
 - 4. Plant not strongly smelling; to 100 cm tall without obvious rosette, secondary stems not contracted to form dense flowering spikes; vestiture stellate-hairy or glabrous; flowers solitary, often in densely leafy clusters; polygamodioecious.
 - 3. Small, woody, divaricate, perennial shrub; vestiture stellate-hairy or densely squamulose with peltate scales.
 - 5. Vestiture stellate-hairy; petiolate basal leaves merging into subsessile or sessile floral leaves, blades with 1-5 prominent veins, margin becoming dentate or lobed towards apex; calyx 3.8-5.3 mm long.................4. L. viridi-grisea
 - 5. Vestiture densely squamulose with peltate scales; all leaves sessile, blades with a single prominent mid-vein, margin entire throughout; calyx 7-10 mm long.

 5. L. buchananensis
 - 2. Prostrate, decumbent or erect sub-shrubs, or erect shrubs; female flowers of dioecious species with sterile anthers present; mericarps dehiscent, 2-5 in each fruit, all fertile or only 1-2 fertile; pericarps unornamented; seed triquetrous or swollen and irregularly globular, rugose or smooth. (SECTION SELENOTHAMNUS.)
 - 6. Erect, decumbent or prostrate sub-shrubs; dioecious or hermaphroditic; stems without spongy periderm; leaves distinctly petiolate throughout or petiolate basal leaves merging into subsessile or sessile stem and floral leaves, stellate-hairy or glabrous; female flowers completely lacking anthers; mericarps all well-developed; seed triquetrous, surface smooth or rugose. (Series Halophyton.)

- 7. Plant erect; vestiture sparse to dense; leaves conduplicate or flat, distinctly petiolate basal leaves merging into subsessile or sessile stem and floral leaves; dioecious; petals glabrous or sparsely to densely stellate-hairy; fruit with 2-5 mericarps a single one of which is fertile.
 - 8. Plant to 130 cm tall; vestiture shortly stellate-hairy giving plant an ashengrey appearance; leaves strongly conduplicate along prominent midvein; petals glabrous; fruit with 2-3 mericarps; seed smooth.........6. *L. cinerea*
 - 8. Plant to 60 cm tall; vestiture sparsely to densely loosely stellate-hairy, plant green in appearance; leaves flat with several more or less prominent veins; petals stellate-hairy on abaxial surface, rarely glabrous; fruit with 2-5 mericarps; seed conspicuously rugose on all surfaces. 7. L. densiflora
- 7. Plant decumbent or prostrate; vestiture sparse to dense or glabrous; leaves flat, distinctly petiolate throughout; hermaphroditic; petals glabrous; fruit with 5 mericarps all of which are fertile.
 - 9. Plant decumbent, to 15 cm tall x 40 cm wide; vestiture densely stellatehairy; anthers 5-10; seeds conspicuously rugose on abaxial surface.
- 6. Erect shrubs; dioecious; stems with spongy periderm producing dry fibrous or flaky bark; leaves sessile throughout, stellate-hairy or with armour-like covering of peltate scales; dioecious; female flowers with or without sterile anthers; sterile mericarp crushed and obliterated by developing fertile mericarp(s); seed swollen and irregularly globular, smooth. (Series Selenothamnus.)

 - 10. Vestiture of young branches densely pannose with palmate hairs, dense covering of peltate scales always present on leaves and calyces; thorns lacking; secondary branches densely leafy and much contracted; leaf margin entire throughout; only 1 mericarp ever fertile in each fruit.
 - 11. Secondary stems openly branched or loosely contracted; leaves 4.0-12.0 mm long; corollas of female and male flowers indistinguishable; anthers 10-20 in male flowers, 5-15 in female flowers; pistils 2-3; style of female flowers 2.0-7.0 mm long, branches often briefly lobed towards the apices.

 11. L. chrysoderma
 - 11. Secondary stems much contracted giving plant a characteristic cactus-like appearance; leaves 1.5-7.2 mm long; corollas of female flowers c. ¾ length of corollas of male flowers; anthers 5-16 in male flowers, 0-8 in female flowers; pistils 2; style of female flowers 0.4-3.2 mm long, branches always entire.

......12. L. helmsii

Subgenus PANIFEX

Lawrencia subgenus Panifex Lander, subgen. nov.

Frutices erecti, inermes; indumentum e pilis stellatis compositum; flores manifeste pedicellati, 1-5-ni in cymas axillaris dispositi; sepala ventrali basi papillosa; styli rami ad conjunctionem manifeste stellato-pilosi; mericarpia indehiscentia, per fructum 9-11-na, omnia fertilia; pericarpium apicaliter et distaliter pilosum, lateraliter coriaceum et rugosum vel obscure reticulatum; semina triquetra laevia.

Typus: L. berthae (F. Muell.) Melville

Erect shrubs; without thorns; vestiture stellate-hairy; flowers distinctly pedicellate, in axillary cymes of 1-5, sweetly scented; sepals of male flowers with basal tufts of minute, nectariferous papillae on adaxial surface; petals of male fowers often with intermeshing basal hairs; filament tubes often sparsely simple-hairy; style branches conspicuously stellate-hairy at their junction; mericarps indehiscent, 9-11 in each fruit, all fertile; pericarps apically and distally stellate-hairy, laterally coriaceous and rugose or obscurely reticulate; seed triquetrous, smooth.

The name of this subgenus honours the British botanist Edmund Gilbert Baker (1864-1949) a student of the Malvaceae (Stafleu & Cowan 1976) and of *Lawrencia* in particular (Baker unpublished).

1. Lawrencia berthae (F. Muell.) Melville, Kew Bull. 20: 514 (1966); Beard, Descrip. Cat. Western Austral. Pl. 84 (1970); J. H. Willis, Handb. Pl. Victoria 2: 381 (1973); J. Green, Census Vasc. Pl. Western Australia 67 (1981); Jessop, List, Vasc. Pl. South Australia 62 (1983). Plagianthus berthae F. Muell., Fragm. Phytogr. Austral. 5: 103 (1866); E. G. Baker, J. Bot. 30: 72 (1892); C. A. Gardner, Enum. Pl. Austral. Occid. 78 (1931); J. M. Black, Fl. South Australia, edn 2, 3: 557 & t. 719 (1952); Blackall & Grieve, How to Know Western Austral. Wildfl. 2. 346 (1956). Sida berthae F. Muell., Fragm. Phytogr. Austral. 5: 103-104 (1866), nom. inval. Type: Gardners River, Western Australia, s.dat., G. Maxwell s.n. (holo: K; possible iso: MEL, M non vidi).

Shrub to 80 cm tall, spreading with slender branches, dioecious. Vestiture of stems, stipules, leaves, pedicels and abaxial surface of calyces sparsely to densely hairy with simple, bifurcate and haplomorphic stellate hairs. Leaves alternate, distinctly petiolate basal leaves merging into more or less subsessile floral leaves; stipules green and leaf-like, narrowly to broadly triangular or elliptic, 1.0-2.5 mm long x 0.5-0.8 mm wide, margin ciliate, apex acuminate, acute or obtuse; petiole slender, triquetrous, to 17(45) mm long; blade elliptic, ovate, obovate or deltoid, 5-20(45) mm long x 3-16(35) mm wide, flat, venation distinct with 3-5 conspicuous primary veins, base acute- or obtuse-cuneate, margin coarsely dentate or obtusely lobed. except towards the base, apex acute, obtuse or rounded. Flowers unisexual, sweet-scented, born on slender peduncles to 45 mm long, solitary or 2-5 in axillary fascicles. Calvx of male flower turbinate, green, 4.2-7.0 mm long; sepals connate 2/5-2/3 their length, each with a basal tuft of minute, nectariferous, multicellular papillae at the base; lobes triangular, 2.0-2.5 mm wide; apices acute. Corolla of male flower white, 6.5-12.0 mm long; petals almost free to their bases, connate for less than 1/8 their length, glabrous or with intermeshing simple hairs at base on both abaxial and adaxial surfaces; lobes spreading, obovate to ovate, 4.5-7.0 mm wide; apices obtuse, entire. Androecium of male flower with filament tube 1.0-4.0 mm long, glabrous or with scattered simple hairs; free portion of filaments 1.5-2.0 cm long; anthers 10-30, very rarely pink. Gynoecium of male flower lacking, or present but sterile and much reduced; style 2.0-4.0 mm long; branches 5-8, connate for ¼ their length, 0.2-0.5 mm wide, conspicuously stellate-hairy at their junction. Calyx of

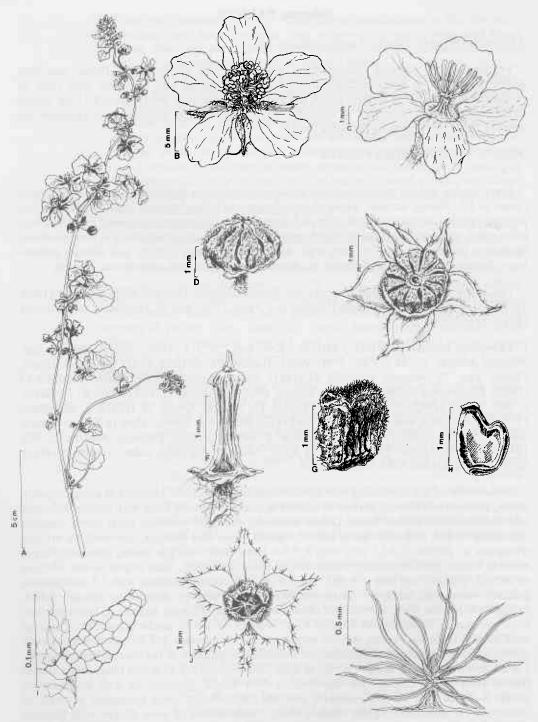


Figure 2. Lawrencia berthae. A — Male plant, habit. B — Male flower. C — Female flower. D — Mature fruit, lateral view. E — Mature fruit, apical view. F — Columella. G — Ripe mericarp, lateral-distal view. H — Seed, lateral view. I — Nectariferous papilla. J — Calyx, apical view showing vestigial nectaries. K — Trichome.

Drawn from George 5692 (male) and Helms, Sept. 1891 (female).

female flower turbinate, green, 4.0-5.0 mm long; sepals connate 1/3-1/2 of their length, adaxially glabrous, sometimes with a few simple hairs towards apices; lobes triangular, 1.4-2.5 mm wide; apices acute. Corolla of female flower white, 4.0-4.8 mm long; petals connate ½-¼ of their length, glabrous; lobes broadly imbricate, erect, ovate, 2.7-3.0 mm wide; apices obtuse, entire or emarginate. Androecium of female flower much reduced; filament tube 1.5-2.0 mm long; anthers lacking. Gynoecium of female flower with style 2.5-3.5 mm long; branches 9-11, free to base or connate for up to 2/5 their length, clavate, 0.2-0.5 mm wide, pale yellow, conspicuously stellate-hairy at their junction, introrsely stigmatose on upper 1/2-2/3. Fruit oblate, 2.0 mm high x 3.7-6.2 mm wide; mericarps 9-11; pericarp black, coriaceous, lateral walls rugose or obscurely reticulate, apical and distal surfaces densely stellate-hairy, indehiscent. Seed triquetrous, smooth, brown, 1.5 mm high x 1.3-1.7 mm in radial width. Columella stout, 1.2-1.5 mm high, 0.7-1.2 mm in diameter. Figure 2.

Flowering period. July to December.

Habitat. Occurs on clay soils in open woodland and shrubland adjoining shallow wide depressions, also on disturbed ground by roadsides.

Selected specimens from 45 collections examined. WESTERN AUSTRALIA: N of Borden, A.M. Ashby 1940 (AD, PERTH); Israelite Bay, 1893, Brooks s.n. (MEL); Bendering, C.A. Gardner 1993 (PERTH); Near Salmon Gums, Sept. 1935, C.A. Gardner s.n. (PERTH); Northam, 1901, J.H. Gregory s.n. (BM); E bank of Swan River, 1889, M. Heal s.n. (MEL).

SOUTH AUSTRALIA: Alawoona, c. 30 km S of Loxton, Sept. 1936, E.H. Ising s.n. (AD); Hambidge Flora and Fauna Reserve, Eyre Peninsula, D.E. Symon 4300 (AD, CANB); Ardrossan, Yorke Peninsula, Oct. 1879, O. Tepper s.n. (AD).

VICTORIA: Beside Ouyen Highway, c. 5 km E of Murrayville, M.G. Corrick 6239 (AD,

MEL); Mildura, Oct. 1932, W.J. Zimmer s.n. (MEL).

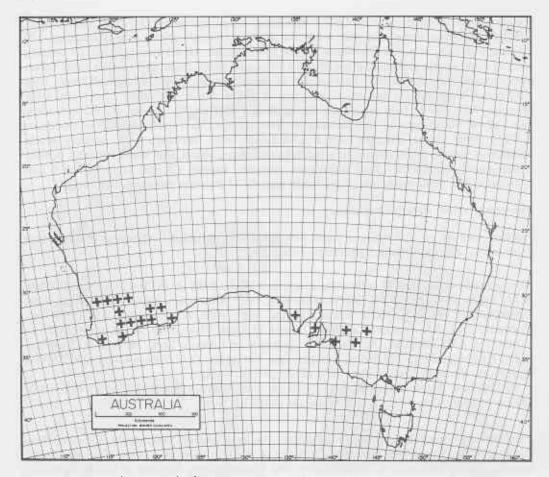
Distribution. Found in the Avon, Coolgardie, Darling (Warren Subdistrict), Eyre and Roe Districts of the South-West Botanical Province of Western Australia; in the Eyre Peninsula, Murray, Yorke Peninsula, Southern Lofty, and South Eastern Regions of South Australia; and in the Mallee Study Area of Victoria. See Map 3.

Etymology. The specific epithet honours Bertha Cosson, daughter of the botanist and famous traveller in Algeria (Mueller loc. cit.).

Notes. It is, perhaps, significant that in the draft of his paper concerning Lawrencia, Baker (unpublished) excluded Plagianthus berthae from consideration. I have chosen to follow Melville (1966) in retaining this species in Lawrencia. The problem remains that L. berthae has no close relatives in the genus and most of the character states that separate it from other species are viewed as primitive (see Key to Infrageneric Taxa and Species above and Discussion below). Obviously, a good case could be made for its placement in a monotypic genus: this should, in my opinion, await further investigation of the Plagianthus alliance.

The geographical range of Lawrencia berthae overlaps with those of L. glomerata, L. repens, L. spicata and L. squamata. L. berthae has been found growing in close proximity to these other species.

Despite its widespread and disjunct distribution, there is no morphological-geographical variation discernible within *Lawrencia berthae*.



Map 3. Distribution of Lawrencia berthae.

Subgenus LAWRENCIA

Lawrencia Hook. subgenus Lawrencia

Plagianthus section Lawrencia (Hook.) Benth., J. Linn. Soc., Bot. 6: 123 (1862); E. G. Baker, J. Bot. 30: 72-3 (1892). Sida section Lawrencia (Hook.) F. Muell., Pl. Victoria 1: 162-3 (1862). Lectotype (here designated): Pl. spicata (Hook.) Benth. [L. spicata Hook.].

Prostrate, decumbent or erect sub-shrubs, or erect shrubs; with or without thorns; vestiture stellate-hairy or squamulose with peltate scales; flowers subsessile or sessile, axillary, solitary, often crowded in leafy fascicles, odourless; sepals adaxially glabrous; petals never with intermeshing basal hairs; filament tubes glabrous; style branches glabrous at their junction; mericarps indehiscent or dehiscent, 2-5 in each fruit, all fertile or only 1-2 fertile; pericarps glabrous, laterally reticulate with fenestrate or alveolate areolae or unornamented; seed triquetrous or swollen and irregulary globular, smooth or rugose.

SECTION LAWRENCIA

Lawrencia section Lawrencia

Erect sub-shrubs or shrubs; without thorns; female flowers of dioecious species lacking anthers altogether (in *L. glomerata*, a polygamodioecious species, sterile anthers may be present); styles filiform; mericarps indehiscent, 5 in each fruit, all fertile; pericarps laterally reticulate with fenestrate or alveolate areolae; seed triquetrous, smooth.

This section comprises two apparently related species pairs. Young plants of Lawrencia spicata bear a remarkable resemblance to plants of the glabrous type form of L. glomerata which is confined to the Irwin and Austin Botanical Districts of Western Australia, well to the north of the present-day range of the former. These two species are, of course, readily distinguishable in their adult phases by features of odour, habit, leaf shape, inflorescence and sexuality.

Lawrencia viridi-grisea and L. buchananensis are superficially similar in that both are hermaphroditic shrubs. However, they are always easily distinguishable by features of vestiture, leaf shape, venation and development, and dimensions of floral parts.

Lawrencia glomerata, L. spicata and L. viridi-grisea are extremely widely distributed across Australia; L. buchananensis is confined to a single, isolated population in north-eastern Queensland (Maps 4 & 5).

2. Lawrencia spicata Hook., Hooker's Icon.. Pl. 3: t. 262 (1840); Hooker, J. Bot. (Hooker) 2: 413 (1840); J. D. Hooker, Fl. Tasman. 1: 48 (1855); Melville, Kew Bull. 20: 514 (1966); J. Hutchinson, Evol. & Phyl. Fl. Pl. t. 229 (1969); Beard, Descr. Cat. Western Austral. Pl. 84 (1970); J. H. Willis, Handb. Pl. Victoria 2: 382 (1973); W. M. Curtis, Stud. Fl. Tasmania Ed. 2, 1: 88 (1975); J. Green, Census Vasc. Pl. Western Australia 67 (1981); Jacobs & Pickard, Pl. New South Wales 144 (1981); Jessop, List. Vasc. Pl. South Australia 62 (1983). Sida lawrencia F. Muell., Pl. Victoria 1: 162 (1862). Plagianthus spicatus (Hook.) Benth., J. Linn. Soc., Bot. 6: 103 (1862); Bentham, Fl. Austral. 1: 189-190 (1863); E. G. Baker, J. Bot. 30: 72 (1892); C. Moore, Census Pl. New South Wales 6 (1884); C. Moore, Handb. Fl. New South Wales: 57 (1893); Rodway, Tasman. Fl. 16 (1903); Maiden & Betche, Census New South Wales Pl. 136 (1916); C. A. Gardner, Enum. Pl. Austral. Occid. 78 (1931); J. M. Black, Fl. South Australia, edn 2, 3: 556 & t. 716 (1952); Hj. Eichler, Suppl. Black's Fl. South Australia: 221 (1965); Blackall & Grieve, How to know Western Austral. Wildfl. 2: 344 (1966). Neotype (here designated): Hooker's Icon. Pl. 3: t. 261 (1840).

Sub-shrub to 180 cm tall, ascending, with one to several stout stems diverging from a basal rosette and slender secondary stems contracted to form dense leafy spikes, hermaphroditic. Vestiture lacking, plant glabrous throughout. Leaves alternate, distinctly petiolate basal leaves merging into subsessile or sessile floral leaves; stipules green and leaf-like, narrowly triangular, 2-11 mm long x 0.3 mm wide, margin entire, apex narrowly acuminate; petiole stout, trigonal or triquetrous, to 40 cm long; blade of basal leaves elliptic, narrowly elliptic or narrowly ovate, often falcate, as large as 11.5 cm long x 6.0 cm wide but generally less than 7.0 cm long x 3.0 cm wide, blade of floral leaves narrowly ovate, narrowly elliptic or narrowly obovate, as small as 4.0 mm long x 0.5 mm wide, blades flat, venation distinct with 1-7 primary veins, base obtuse- to acute-cuneate, margin of basal leaves coarsely dentate throughout becoming dentate only towards apex or else entire throughout in floral leaves, apex of basal leaves broadly obtuse to acute, becoming narrowly acute in floral leaves. Flowers bisexual, axillary, sessile or subsessile, solitary, often crowded in leafy fascicles. Calyx turbinate, green, 5.5-10.0 mm long; sepals connate 1/3-2/3 their

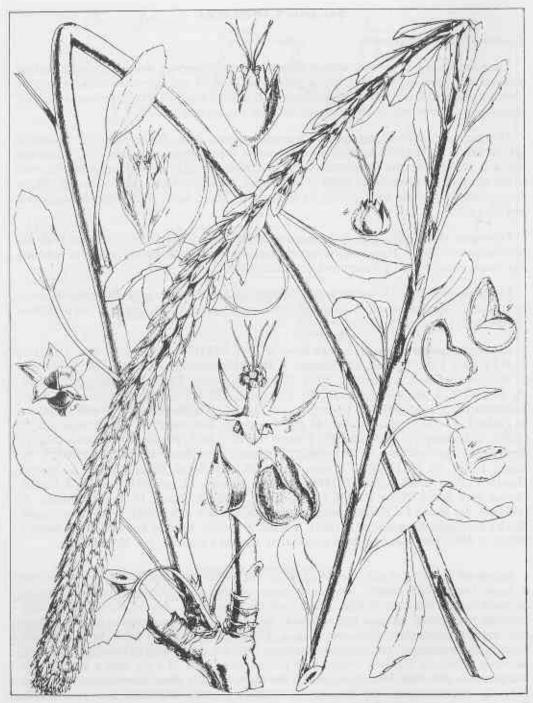


Figure 3. Lawrencia spicata. 1 — Flower and subtending leaf. 2 — Flower. 3 — Corolla, staminal column and style branches. 4 — Gynoecium. 5 — Calyx and columella. 6 — Ripe mericarp. 7 — Mericarp opened to reveal seed. 8 — Seed opened to reveal embryo surrounded by endosperm. 9 — Seed opened to reveal embryo itself. 10 — Embryo.

Neotype (see text).

Reproduced from Hooker, Hook., Ic. Pl. 3: t 262 (1840).

length; lobes broadly triangular, 2.5-4.8 mm wide; apices acute or acuminate. Corolla white to pale yellow, 4.0-8.0 mm long; petals almost free to their bases or connate to 2/5 their length, erect or somewhat spreading, ovate, 1.8-3.0 mm wide; apices obtuse, usually emarginate, sometimes entire. Androecium with filament tube 1.8-3.7 mm long, glabrous; free portion of filaments 0.2-0.8 mm long; anthers 10-20, usually 20. Gynoecium with style 4.5-8.0 mm long; branches 5, free to base or united for up to 2/5 their length, filiform, 0.1-0.5 mm wide, pink to dark brown, introrsely stigmatose, often on upper 4/5 only. Fruit somewhat oblate to ovoid, 2.2-4.0 mm high x 1.6-4.4 mm in diameter; mericarps 5; pericarp white or tan, membranous, becoming translucent and reticulate on lateral and distal-basal surfaces, margin prolonged to form narrow distal-lateral wings often connected across apical distal region to give the distal wall a distinctive shield-like appearance. Seed triquetrous, brown, smooth, 1.6-2.7 mm high x 1.0-2.0 mm wide. Columella slender, 2.5-3.8 mm high x 0.2-0.4 mm in diameter. Figure 3.

Flowering period. October to February.

Habitat. Occurs in alluvial soils of temperate coastal salt-marshes, estuaries and river banks, and in peaty swamps, occasionally colonizing clay soils along roadsides.

Selected specimens from 130 collections examined. NEW SOUTH WALES: Sydney, s.dat., B. Bynoe s.n. (K) a doubtful record.

SOUTH AUSTRALIA: Bay 10 (Port Lincoln), s.dat. R. Brown 5110 (BM, CANB, K, MEL); mouth of Inman River, c. 65 km S of Adelaide, Jan. 1926, J.B. Cleland s.n. (AD); E of Reserve, Hundred of Murlong, c. 120 km N of Port Lincoln, Dec. 1959, J.B. Cleland s.n. (AD); 15 km S of Streaky Bay, c. 90 km SE of Ceduna, T. Dennis 152 (AD); Hundred of Messent, c. 190 km SSE of Adelaide, T.R.N. Lothian 1255 (AD); Holdfast Bay, 1850, F. Mueller s.n. (MEL); Lake Albert, c. 10 km SW of Meningie, c. 110 km SE of Adelaide, L.D. Williams 4102 (AD).

TASMANIA: Flinders Island, 1956 Anon. (HO 12972); Port Arthur, 1892, J. Bufton s.n. (MEL); Bellaire, Old Canal, H.F. Comber 2024 (K); Ralphs Bay Canal, towards Sandford, Feb. 1966, W.M. Curtis s.n. (HO); Koonya, at mouth of Newmans Creek, Hj. Eichler 16859 (AD); Clarke Island, 1894, E. Maclaine s.n. (MEL); Double Creek, on main road between Orford and Triabunna, Feb. 1976, J.W. Parham s.n. (HO); Coles Bay, April 1930, L. Rodway s.n. (HO, K).

VICTORIA: Melbourne, Feb. 1946, L.C. Bartels s.n. (NSW); smaller island near Goat Island, Mallacoota Inlet National Park, A.C. Beauglehole 31285 (MEL); Rigby Island, S of Kalimna, P.K. Gullan 384 (MEL); Polkemmet, near river, Wimmera, F.M. Reader 8 (MEL); Point Lonsdale, J. Tilden 797 (BM, K); Lake Tyers, April 1977, J. Turner s.n. (MEL).

WESTERN AUSTRALJA: Claremont, C. Andrews 72 (BM, K, PERTH); Mandurah, Feb. 1928, A.J. Hall s.n. (PERTH); 22.6 km E of Ravensthorpe on road to Esperance, N.S. Lander 1166 (BH, K, MEL, PERTH); Loongana, J. Lowry 43 (PERTH); near Jurien Bay, Jan. 1963, F. Lullfitz s.n. (PERTH); Neendaling, W of Lake Grace, Nov. 1978, P. Stevenson s.n. (PERTH).

Distribution. Found in the Avon, Darling (Drummond Subdistrict), Eyre and Roe Districts of the South-West Botanical Province of Western Australia; in the Eyre Peninsula, Northern Lofty, Southern Lofty, and Kangaroo Island Regions of South Australia; in the South West Coranamite, Melbourne, Southern Gippsland, Gippsland Lakes and Eastern Gippsland Study Areas of Victoria; on the east coast of Tasmania and islands of Bass Strait; and possibly near Sydney on the Central Coast of New South Wales. See Map 5.

Etymology. The specific epithet draws attention to the dense spicate inflorescence characteristic of this species.

Notes. In his protologue, W. J. Hooker (1840 loc. cit.) cites specimens sent to him by Ronald Gunn collected at "Port Arthur, Van Diemans Land; and at Port Fairy, South Australia, growing on the side of a salt-water inlet, where the ground was marshy". Neither these specimens nor any duplicates of them have been found in the course of the present study, nor are they cited by Bentham (1863 loc. cit.).

A specimen sent to J. D. Hooker, Gunn 746 (K), has a note in Gunn's hand appended: "Numerous specimens of this plant collected at various times are packed up separately. They are from Port Fairy, South Coast of New Holland, but the Lawrencia spicata occurs at Flinders Island and I doubt not upon our North Coast although I have not yet seen it. Remove Port Arthur from the habitats as I now hardly feel satisfied about having seen it from that place". Further, Hooker (1855 loc. cit.) notes that "Mr Gunn informs me that the Port Arthur Station formerly assigned to this plant by him is doubtful". Clearly, Gunn 746 was dispatched well after publication of Lawrencia spicata. It is just conceivable that it is a duplicate of Gunn's original collection from Port Fairy probably made between 1836 and 1838 (see Willis 1949). On the other hand it could as easily have been gathered at some later date on Flinders Island. In view of this uncertainty it seems best to designate the above neotype. Fortunately, L. spicata is sufficiently characteristic for there to be no mistaking it from the detailed illustration provided by W. J. Hooker which is reproduced here (Figure 3).

Mueller (1862 loc. cit.) published the name Sida lawrencia indicating that it was composed of Lawrencia spicata and L. glomerata distinguishing a "glabrous erect variety with simple spikes" and a "procumbent or ascendent velvet-downy branched variety with glomerate spikes". Collections cited by Mueller under this name are referred variously to L. glomerata, L. spicata and L. viridi-grisea in the present treatment.

A specimen of Lawrencia spicata from W. J. Hooker's herbarium and now held at K is annotated "Sydney Bynoe." Benjamin Bynoe was a surgeon with H.M.S. 'Beagle' on King's 1837-1843 expedition to survey the north-western coast of Australia. Bynoe made botanical collections on Depuch Island, the Abrolhos, the Victoria River, off Bass Strait and in New South Wales (Maiden 1906). His specimen of L. spicata was cited tentatively by J. D. Hooker (1855 loc. cit.) Subsequent authors recorded the occurrence of this species in New South Wales under Plagianthus spicatus, including C. Moore (1884 loc. cit., 1893 loc. cit.) and Maiden & Betche (1916 loc. cit.), although no further collections from that state are known. Jacobs & Pickard (1981 loc. cit.) consider its presence in New South Wales unsubstantiated. It is possible that Bynoe encountered L. spicata at the Victoria River or off Bass Strait where it is still to be found today.

With the notable exception of a single immature specimen collected at Loongana on the Nullarbor Plain, the distribution of *Lawrencia spicata* shows a pronounced disjunction between south-western and south-eastern Australia. It seems most likely that the Nullarbor specimen is of merely sporadic occurrence from seed dropped by trains, although future collecting from this region may demonstrate otherwise.

Despite its disjunct distribution, there is no morphological-geographical variation discernible in *Lawrencia spicata*.

The geographical range of Lawrencia spicata overlaps with those of L. berthae, L. diffusa, L. glomerata and L. repens. L. spicata has been found growing in close proximity to these other species.

3. Lawrencia glomerata Hook., Hooker's Icon. Pl. 5: t. 417 (1842); C. Moore, Handb. Fl. New South Wales 57 (1893); J. H. Willis, Handb. Pl. Victoria 2: 382 (1973); N. Beadle, Stud. Fl. N.E. New South Wales 1: 300 & t. 141 (1976); J. Green, Census Vasc. Pl. Western Australia 67 (1981); Jessop, Fl. Centr. Australia 211-2 (1981); Jacobs & Pickard, Pl. of New South Wales 144 (1981); G. M. Cunningham et al., Pl. W. New South Wales 491-2 (1982); Jessop, List Vasc. Pl. South Australia 62 (1983). Plagianthus glomeratus (Hook.) Benth., J. Linn. Soc., Bot. 6: 103 (1862); Bentham, Fl. Austral. 1: 190 (1863); Bailey, Queensland Fl. 1: 110 (1899); Maiden & Betche, Census New South Wales Pl. 136 (1916); C. A. Gardner, Enum. Pl. Austral. Occid. 78 (1931); J. M. Black, Fl. South Australia, edn 2, 3: 556-7 & t. 717 (1952); Blackall & Grieve, How to Know Western Austral. Wildfl. 2: 345 (1956); Hj. Eichler, Suppl. Black's Fl. South Australia: 221 (1965); Chippendale, Proc. Linn. Soc. New South Wales 96: 248 (1971). Type: Swan River, Western Australia, 1843, Drummond 55 (holo: K; iso: K, BM, MEL, PERTH).

Plagianthus spicatus var. pubescens Benth., Fl. Austral. 1: 189-190 (1863); E. G. Baker, J. Bot. 30: 72 (1892); J. M. Black, Fl. South Australia ed. 2, 3: 556 (1952); Hj. Eichler, Suppl. Black's Fl. South Australia: 221 (1965). Lectotype (here designated): Lake Hindmarsh, s. dat., F. Mueller s.n. (lecto: NSW; isolecto: BM, MEL 98684, 98686, 986887 pro pte). Lectoparatypes: Lake Weering (as "Lake Waringa"), 3 October 1860, J. Dallachy 237 (MEL 98687 pro pte); Lake Weering (as "Lake Waringa"), Victoria, 3 October 1860, J. Dallachy 238 (MEL); Murray Desert, 14 October 1835, F. Mueller s.n. (MEL); Trial Bay, s.dat., P.W. Warburton s.n. (MEL).

Sub-shrub to 100 cm tall, ascending, polygamodioecious. Vestiture of stems, stipules, leaves and adaxial surfaces of calyces glabrous to densely tomentose with simple, bifurcate and haplomorphic stellate hairs. Leaves alternate, distinctly petiolate basal leaves merging into subsessile or sessile floral leaves; stipules green and leaf-like, narrowly linear, obovate or broadly triangular, 1.3-10.0 mm long x 0.3-3.0 mm wide, margin often ciliate, apex acuminate, acute or broadly obtuse; petiole slender, broadly to shallowly triquetrous, often appearing canaliculate or almost flat, to 60 mm long; blades orbicular, broadly to narrowly obovate or broadly to narrowly elliptic, 5-50 mm long x 2-40 mm wide, flat, venation obscure or distinct with 3-7 conspicuous primary veins, base broadly obtuse- to narrowly acute-cuneate, margin coarsely dentate except towards the base of basal leaves, becoming dentate only towards the apex of floral leaves, apex broadly obtuse on basal leaves, becoming narrowly acute or 3-lobed on floral leaves. Flowers unisexual or bisexual, axillary, sessile, solitary, often crowded in leafy fascicles. Calyx turbinate, green, 2.5-12.0 mm long, sepals connate 1/4-9/10 their length; lobes broadly triangular, 1.5-4.6 mm wide; apices obtuse, acute or acuminate. Corolla white or green, 2.5-8.5 mm long; petals almost free to their bases or connate for up to 2/5 their length, glabrous or stellate-hairy along mid-vein; lobes flat and spreading in male flowers, erect and carinate in bisexual and female flowers, elliptic or ovate, 0.9-3.0 mm wide; apices obtuse, usually entire, sometimes emarginate. Androecium with filament tube 1.3-7.0 mm long, glabrous; free portion of filaments 0.2-1.0 mm long; anthers up to 25 in male and bisexual flowers, with or without sterile anthers in female flowers. Gynoecium with style 1.3-10.0 mm long; branches 5, free to base, filiform, 0.1-0.5 mm wide, salmon pink or yellow, introrsely stigmatose; gynoecium sterile and much reduced in male flowers. Fruit somewhat oblate to ovoid, 1.4-4.7 mm high x 0.8-5.0 mm in diameter; mericarps 5; pericarp white or tan, membranous, becoming translucent and reticulate on lateral and distal-basal surfaces, margin often prolonged to form narrow distallateral wings sometimes connected across apical-distal region to give the distal wall a distinctive shield-like appearance. Seed triquetrous, brown, smooth, 1.0-3.2 mm high x 0.5-2.5 mm in radial width. Columella slender, 1.6-4.5 mm high, c. 0.2 mm in diameter. Figure 4.

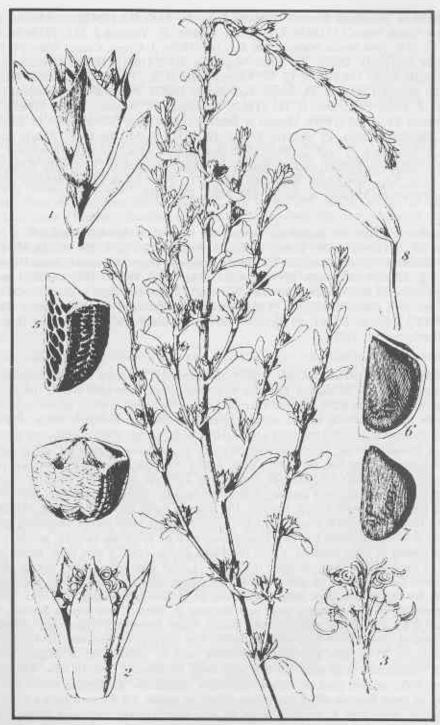


Figure 4. Lawrencia glomerata. 1. Flower, 2. — Corolla, 3 — Staminal column and style branches, 4 — Immature fruit, 5 — Ripe mericarp, 6 — Mericarp opened to reveal seed, 7 — Ripe seed, 8 — Lower leaf,

Drawn from Drummond 55 (holotype).

Reproduced from Hooker, Hook. Ic. Pl. 5: t. 417 (1842).

Flowering period. May to March.

Habitat. Occurs in subsaline sand or clay in succulent steppe, *Triodia* grassland, low shrubland and open woodland in and around estuaries, coastal inlets, inland playa lakes, saline depressions and watercourses or on gibber plains or gypsum or limestone ridges, occasionally colonizing sand along roadsides.

Selected specimens from 258 collections examined. NEW SOUTH WALES: Cobham Salt Lake, W. Bauerlen 256 (MEL); "Barrakee", near Bourke, May 1968, G.L. Jacob s.n. (NSW); West Valley, Acacia Vale, 12 km W of Broken Hill, J.W. Leigh 2047 (NSW); Nucha Lake (as "Lake Mucha"), near Mootwingee, P.L. Milthorpe 576 (AD); Mt Mulyah, c. 60 miles (96 km) NW of Louth, C.W.E. Moore 6367 (CANB); Waroo, near Bourke, Oct. 1936, K.A. Morris s.n. (BRI).

NORTHERN TERRITORY: Palm Valley, G. Chippendale 10612, (AD, CANB, MEL, NSW); "New Haven", Lake Bennet, C. Dunlop 2541 (AD); Stirling Swamp, P.K. Latz 5591 (AD); Lake Neale, J.R. Maconochie 1893 (AD, MEL, NT); Little Sandy Desert, A.S. Mitchell 995 (NT, PERTH); 1 mile (1.6 km) W of Central Mt Wedge, A.O. Nicholls 818

(AD, CANB, K, MEL, PERTH).

QUEENSLAND: Sources of the Thomson River, 1891, C.W. Birch s.n. (MEL); Gregory

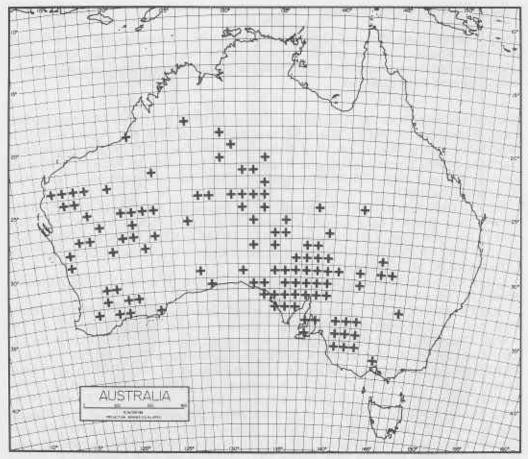
South, M. Olsen 717 (BRI).

SOUTH AUSTRALIA: 7 miles (11.2 km) E of Lyndhurst, J. Carrick 1802 (AD); Bight Well, near Head of Bight, Nov. 1955, J.B. Cleland s.n. (AD); Gawler Ranges, c. 6.5 km NW of Chilpuddie, along the Minnipa-Yardea road, Hj. Eichler 19550 (AD); 20 miles (32 km) W of Emu, N. Forde 479 (AD, K, MEL); near Arckarunga, E.N.S. Jackson 741 (AD, PERTH); Allandale Station, c. 20 km SE of Oodnadatta, R.H. Kuchel 659 (AD); "Purple Downs", c. 50 km NNE of Pimba, R. Swinbourne 180 (AD); Fenelon Island, c. 60 km SW of Ceduna, N.M. Wace 193 (AD); St Francis Island, c. 60 km SW of Ceduna, N.M. Wace 219 (AD, MEL); c. 5 km SE of Coward Springs, on gibber plain just E of Blanche Cup Springs, J.Z. Weber 5786 (AD, PERTH); Billeroo Creek area, c. 45 km ENE of Frome Downs Homestead, D.J.E. Whibley 3425 (AD); c. 18 km S of Mooneree Homestead, near W margin of Lake Gairdner, P.G. Wilson 602 (AD); c. 3 km S of Penong, P.G. Wilson 1588 (AD).

VICTORIA: Raak Plain, c. 46 km S of Mildura, M.D. Crisp 3417 (CBG, MEL); Wyperfeld National Park, 1959, J.O. Maroske s.n. (MEL); Junction of the Murray and Darling Rivers, 1887, J. Minchin s.n. (MEL); near Lake Hindmarsh, J.D.M. Pearson 2024 (MEL); 5.6 km W along road which runs W of the Sunset Tank-Merrinee road, P.S. Short 1185 (AD, CBG, MEL); Gypsum workings 4 miles (6.4 km) SW of Nowingl, Aug. 1955, J.H. Willis

s.n. (MEL).

WESTERN AUSTRALIA: c. 20 km from Mount Augustus Homestead on the Landor road, A.M. Ashby 3408 (AD); between "Barren Downs" and Muellers Range, s.dat., C.W. Birch s.n. (MEL); 6 miles (9.6 km) N of Salmon Gums, W.E. Blackall 1009 (PERTH); "Anna Plains", N.T. Burbidge 1433 (PERTH); 78 km ENE of Cosmo Newberry Mission, R.J. Chinnock 666 (AD); Calvary Gap, Kennedy Range, C.A. Gardner 6075 (PERTH); Lake Auld, A.S. George 9132 (MEL*, NSW, PERTH); upper Ashburton River, W.E.P. Giles 1876, W.E.P. Giles s.n. (MEL); Mt Murchison, 1876, W.E.P. Giles s.n. (MEL); Lake Christopher area, R.H. Kuchel 271 (AD); Lake Austin, c. 15 km S of Day Dawn on Great Northern Highway, N.S. Lander 1113 (BH, CANB, K, MEL*, NT, PERTH); N. end of salt lake extending from Coolimba to Leeman, N.S. Lander 1160, (BH, K, NSW, PERTH); Culham Inlet, Fitzgerald National Park, N.S. Lander 1168 (BH, BRI, NSW, NT, PERTH); Carnamah, Nov. 1906, A. Morrison s.n. (BM, BRI, PERTH); Eucla, 1877, T. Richards s.n. (MEL); 1 mile (1.6 km) SW of Southern Cross, on Great Eastern Highway, M.D. Tindale 130321 (NSW).



Map 4. Distribution of Lawrencia glomerta.

Distribution. Found in the Avon, Eyre, Irwin and Roe Districts of the South-West Botanical Province, in the Ashburton, Canning, Carnarvon, Coolgardie, Eucla, Giles, Helms and Keartland Districts of the Eremaean Botanical Province, and in the Dampier and Fitzgerald Districts of the Northern Botanical Province of Western Australia; in the western half of the Central Australia Pastoral District of the Northern Territory; in each and every Region of South Australia; in the Mallee, Melbourne, South West and Wimmera Study Areas of Victoria; in the Far Western Plains (North Subdistrict) and Western Plains (North and South Subdistricts) Botanical Divisions of New South Wales; and in the Gregory South Pastoral District of Queensland. See Map 4.

Etymology. The specific epithet draws attention to the clustered flowers usual in this species.

Notes. Bentham (1863 loc. cit.) clearly based his *Plagianthus spicatus* var. *pubescens* on the second of Mueller's unnamed varieties of *Sida lawrencia* (*Lawrencia spicata*) discussed above. All of the specimens designated here as lectotype and lectoparatypes were cited by Mueller (loc. cit.) under *Sida lawrencia* and have been examined and initialed by Bentham.

In the draft of a paper made in the late 1930s concerning *Lawrencia*, the British botanist E. G. Baker described and named two putative new species, which were segregates of *L. glomerata*. These putative new species were founded on differences observed amongst the

few specimens available to Baker in London supplemented by a small number from Western Australia selected for his examination by Western Australian Government Botanist Charles Gardner. Several specimens held at BM, K and PERTH have been annotated with two manuscript names by Baker which have never been published.

Lawrencia glomerata is a highly polymorphic taxon displaying very considerable plasticity in vegetative and floral characters. Compared on an individual basis specimens from different populations can be strikingly dissimilar. Throughout Central Australia, for instance, are populations of stellate-hairy plants with particularly large juvenile leaves; in the extreme eastern and western reaches of this species' geographical range are to be found populations of glabrous or subglabrous small-leaved forms (the type form, in fact). As dramatic as these and other examples may seem, the present study, involving the examination of all available herbarium specimens from throughout Australia, found them to be merely part of a continuum and that interpopulational variability in this species does not exhibit a morphological-geographical pattern clear enough to be readily exploited taxonomically. Although particular forms do predominate in particular areas of the range of the complex they do not do so to the exclusion of other forms; some forms also recur at different places within the overall range. L. glomerata is by far the most widespread species in the genus, but its populations are allopatric: under such conditions one might expect local fixation of character states.

Lawrencia glomerata is also remarkable for its polygamodioecy. Flowers of male plants generally have a larger spreading calyx and corolla and a sterile vestigial gynoecium. Flowers of female plants have a smaller erect calyx and corolla and either lack anthers or have sterile anthers which remain small and closed, producing no pollen. Morphologically, the flowers of hermaphrodite plants are intermediate between these two extremes, but they are as large as those of males or almost so. In view of the continuous morphological variation between them, a distinction between male and female flower-types has not been made in the above description.

The geographical range of *Lawrencia glomerata* overlaps with those of all other species in the genus with the exception of *L. buchananensis*. *L. glomerata* is often found growing in close proximity to these other species.

4. Lawrencia viridi-grisea Lander, sp.nov.

Lawrencia viridi-grisea est frutex parce ramosus hermaphroditus, pilis brevibus stellatis dense vestitus. Fructus e mericarpiis quinque indehiscentibus compositus, pericarpio facie laterali et distali-basali hyalino reticulato. Lawrencia viridi-grisea Lawrencia buchananensi similis, a qua indumento stellato nec peltato, foliis basalibus petiolatis nec sessilibus floribusque conspicue minoribus facile distinguitur.

Typus: At turnoff to Cardabia Homestead on road to Point Maud, Western Australia, October 1981, N.S. Lander 1143 (holo: PERTH; iso: BM, BR, K, NSW, NT).

Shrub to 100 cm tall, little-branched, weakly ascending, hermaphroditic. Vestiture of stems, stipules, leaves and adaxial surfaces of calyces densely tomentose with short simple, bifurcate and actinomorphic stellate hairs, the latter with more or less basally coalescent rays, giving the plant a distinctive green-grey appearance. Leaves alternate or clustered on reduced branches, distinctly petiolate basal leaves merging into subsessile floral leaves; stipules green and leaf-like, filiform or narrowly triangular, 1.5-3.0 mm long x 0.3-0.9 mm wide, margin ciliate, apex narrowly acuminate; petiole slender, triquetrous, to 50 mm long; blade narrowly linear, narrowly elliptic to elliptic and sometimes falcate, or obovate to broadly obovate,

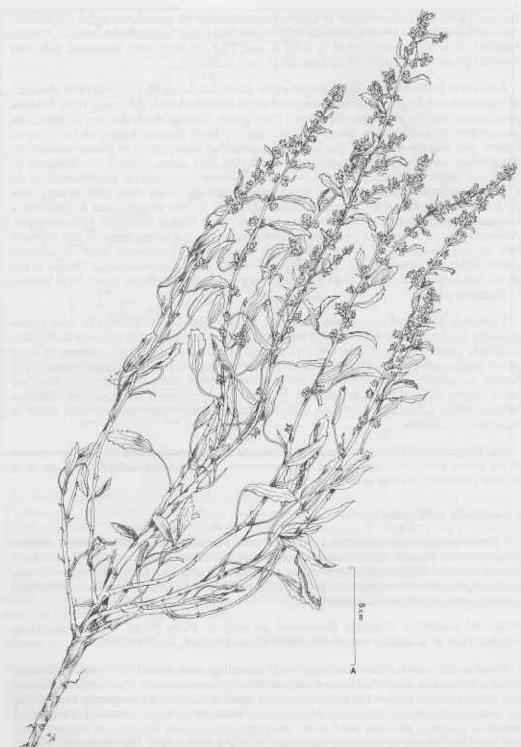


Figure 5. Lawrencia viridi-grisea.

Drawn from Lander 1149

6-48 mm long x 1-26 mm wide, flat, venation sometimes obscure, usually distinct with 1-5 conspicuous primary veins, base acute- to obtuse-cuneate, margin entire, usually becoming coarsely dentate towards the apex, sometimes merely 3-lobed, rarely entire throughout, apex acute, obtuse or truncate. Flowers bisexual, sessile, axillary, solitary, often crowded in leafy fascicles. Calyx turbinate, 3.8-5.3 mm long; sepals connate 1/2-2/3 their length; lobes triangular, 2.0-2.5 mm wide; apices broadly acute. Corolla white or pale yellow, 4.5-6.2 mm long; petals connate 1/10-2/5 their length, glabrous; lobes slightly carinate, ovate, 1.6-2.6 mm wide; apices emarginate. Androecium with filament tube 1.5-3.0 mm long, glabrous; free portion of filaments 0.5-1.0 mm long; anthers 10-20. Gynoecium with style 1.2-5.5 mm long; branches 5, free to base, filiform, white or pink, introrsely stigmatose on upper 2/3-3/4. Fruit oblate, 2.0-3.1 mm high x 3.0-4.0 mm in diameter; mericarps 5; pericarp white or tan, membranous, becoming translucent and reticulate on lateral and distal-basal surfaces, margin prolonged to form narrow distal-lateral wings, indehiscent. Seed triquetrous, brown, smooth, 0.9-2.5 mm high x 0.8-1.5 mm in radial width. Columella slender, 1.5-2.5 mm high x 0.2-0.3 mm in diameter. Figure 5.

Flowering period. Throughout the year.

Habitat. Occurs on subsaline sand and clay soils in succulent steppe in and around estuaries, coastal inlets, inland playa lakes and saline depressions. It is also prominent amongst *Spinifex* on the leeward side of coastal sand-dunes.

Selected specimens from 40 collections examined. NORTHERN TERRITORY: Lake Bennett, N.M. Henry 378 (MEL); S of "Mongrel Downs", P.K. Latz 6567a (NT); Lake Neale, J.R. Maconochie 1891 (AD, CANB, CBG, MEL, NT); 45 km SW of Mongrel Downs Homestead, S. Parker 280 (AD, K, M).

WESTERN AUSTRALIA: "Frazier Downs" (as "Fraser Downs"), Sept. 1964, *C. De Marci s.n. (PERTH); 18*° 16′ S, 122° 04′ E, 1879, A. *Forrest s.n.* (MEL); Dirk Hartog Island, Sept. 1972, *A.S. George s.n.* (PERTH); Hermite Island, *F.L. Hill* 416 (CANB, K); Claypan adjoining Lyndon River on Exmouth-Minilya road, *N.S. Lander* 1149 (MEL*, NT, PERTH); N end of salt lake extending from Coolimba to Leeman on coast E of Eneabba, *N.S. Lander* 1159 (PERTH); Little Sandy Desert, 23° 05′ S, 123° 22′ E, *A.S. Mitchell* 1010 (NT, PERTH); Lake Tobin, Canning Stock Route, *P. Smith* 53 (PERTH).

Distribution. Found in the Irwin District of the South-West Botanical Province, in the Canning, Carnarvon and Fortescue Districts of the Eremaean Botanical Province, and in the Dampier District of the Northern Botanical Province of Western Australia; and in the Central Australia Pastoral District of the Northern Territory. See Map 5.

Etymology. The specific epithet draws attention to the close, dense indumentum which gives this species its characteristic grey-green appearance.

Notes. Lawrencia viridi-grisea displays considerable morphological-geographical variation, particularly in its vestiture and its leaf architecture. In general, plants from coastal populations in Western Australia have long linear, narrowly elliptic or elliptic leaves, and the arms of the radiate stellate hairs that clothe them are only shortly coalescent basally; those from Central Australia tend to bear short obovate to broadly obovate leaves and are clothed in stellate hairs with strongly coalescent arms approaching the peltate scales of L. buchananensis in appearance. Although plants from the two extremes of this range of variation can appear very different indeed, there would appear to be no compelling argument for the taxonomic sub-division of this species. Broadly speaking, there is an east-west continuum between these morphological extremes, all other characters remaining constant.

Herbarium specimens of Lawrencia viridi-grisea have been relegated to L. glomerata in the past.

Lawrencia viridi-grisea is a conspicuous element of the hind-dune flora along the coastline of Western Australia from Leeman to Broome. Its geographical range overlaps with those of L. cinerea, L. densiflora, L. glomerata and L. squamata. L. viridi-grisea has been found growing in close proximity to these other species.

5. Lawrencia buchananensis Lander, sp. nov.

Lawrencia buchananensis est frutex hermaphroditus trichomatibus peltatis dense obsitus. Fructus e mericarpiis quinque indehiscentibus compositus, pericarpio facie laterali et distalibasali hyalino et reticulato. Haec species Lawrenciae viridi-grisea simillima, a qua facile indumento peltato nec stellato, foliis basalibus sessilibus nec petiolatis floribusque conspicue majoribus distinguende.

Typus: Lake Buchanan, Queensland, June 1976, J. Gasteen 85 (holo: BRI).

Shrub to 100 cm tall, ascending, hermaphroditic. Vestiture of stems, leaves and adaxial surface of calyces densely squamulose, predominantly with peltate scales, but also with scattered simple, bifurcate or actinomorphic stellate hairs, the latter with more or less basally coalescent rays. Leaves more or less subsessile throughout, alternate or clustered on reduced branches; stipules membranous, glabrous, linear, c. 2.5 mm long x 0.5 mm wide, margin ciliate, apex acuminate; blade narrowly ovate, obovate or cuneate, 10-30 mm long x 2-60 mm wide, flat, venation obscure except for a prominent mid-vein, base narrowly cuneate, canaliculate, margin entire, apex broadly acute to obtuse. Flowers bisexual, subsessile, axillary, solitary, often crowded in leafy fascicles. Calyx turbinate, green, 7.0-10.0 mm long; sepals connate 2/5-3/4 their length; lobes narrowly triangular, 2.0-2.5 mm wide; apices acute. Corolla white, 6.0-8.0 mm long; petals connate 1/4-1/3 their length, glabrous; lobes erect, ovate, 2.0-2.5 mm wide; apices emarginate. Androecium with filament tube 2.4-4.0 mm long, glabrous; free portion of filaments 0.6-1.0 mm long, anthers 20. Gynoecium with style 4.5-6.5 mm long; branches 5, free to base or connate for up to 1/3 their length, filiform, dark brown, introrsely stigmatose. Fruit oblate, 1.5-3.5 mm high x 2.5-4.2 mm in diameter; mericarps 5; pericarp tan, membranous, becoming translucent and reticulate on lateral and distalbasal surfaces, indehiscent. Seed triquetrous, smooth, brown, 2.1-2.4 mm high x 1.2-1.4 mm in radial width. Columella slender, 1.5-3.5 mm high x 0.2-0.3 mm in diameter.

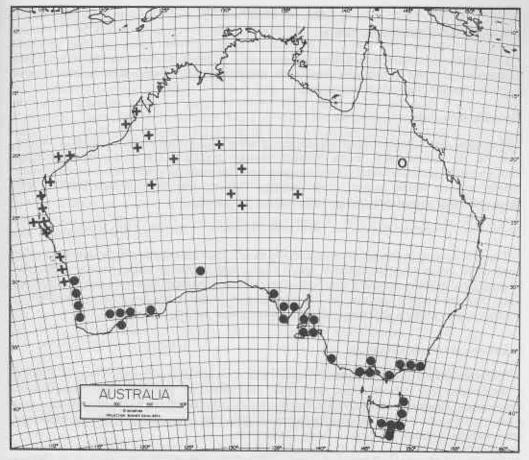
Flowering period. June.

Habitat. ". . . forming clumps in lower part of salt meadows adjacent to lake. Heavily browsed by cattle and roots sought by pigs. Withstands long immersion in salt water."

Conservation status. Lawrencia buchananensis is known only from a single population restricted to a highly specific habitat where both its above-ground and below-ground parts are heavily browsed by cattle and pigs respectively. Thus it must be considered a rare and vulnerable species according to the criteria established by Leigh et al. (1981).

Specimens examined. QUEENSLAND: Lake Buchanan, June 1976, J. Gasteen 85 (holo: BRI); Lake Buchanan, c. 130 miles (208 km) SW of Charters Towers, H.S. Laveny 52 (BRI).

Distribution. Found only at Lake Buchanan in the South Kennedy Pastoral District of Queensland. See Map 5.



Map 5. Distributions of Lawrencia buchananensis (O), L. viridi-grisea (+) and L. spicata. (1)

Etymology. The specific epithet draws attention to the fact that this taxon has been found only at Lake Buchanan in north-eastern Queensland.

Notes. Lawrencia buchananensis is land-locked and geographically isolated and has been found growing with no other species in the genus.

Section SELENOTHAMNUS

Lawrencia sectio Selenothamnus (Melville) Lander, stat. nov. Selenothamnus Melville, Kew Bull. 20: 514-5 (1966); Hutchinson, Gen. Fl. Pl. Dicot. 2: 53 (1967); Baines, Austral. Pl. Gen. 338-9 (1981). Type: S. squamatus (Nees ex Miq.) Melville [Lawrencia squamata Nees ex Miq.].

Prostrate, decumbent or erect sub-shrubs, or erect shrubs; with or without thorns; female flowers of dioecious species with sterile anthers present; styles filiform or clavate; mericarps dehiscent, 2-5 in each fruit, all fertile or only 1-2 fertile; pericarp unornamented; seed triquetrous or swollen and irregularly globular, rugose or smooth.

Etymology. The name probably alludes to the moon-like landscapes and gypseous soils preferred by species in this section.

Series HALOPHYTON

Lawrencia series Halophyton Lander, ser. nov.

Suffrutices prostrati decumbentes vel erecti, glabri vel stellato-pilosi, dioici vel hermaphroditi; flores feminei sine staminodiis; mericarpia omnia fructus bene evoluta; semina triquetra, laevia vel angularia vel rugosa.

Typus: L. diffusa (Benth.) Melville

Prostrate, decumbent or erect sub-shrubs; dioecious or hermaphroditic; without thorns; glabrous or stellate-hairy; stems without spongy periderm; leaves distinctly petiolate throughout or petiolate basal leaves merging into subsessile or sessile floral leaves; female flowers without sterile anthers; style branches filiform; mericarps in each fruit all well developed; seed triquetrous and smooth, angular or rugose.

Etymology. The name refers to the saline soils frequented by species in this series.

The four species in this series are extremely closely interrelated and display a considerable number of independent reductional series in characters such as habit, vestiture, leaf size and marginal incision, mericarp number, anther number, seed sterility and external sculpturing. They have been much confused in the past. Species in this series are confined to Western Australia and exhibit a striking north-south distribution pattern (Map 6).

6. Lawrencia cinerea Lander, sp. nov.

Suffrutex erectus dioicus indumento denso e pilis parvis stellatis composito cinereus. Fructus e mericarpiis 2-3 dehiscentibus, pericarpio membranaceo pellucido non ornamentato compositus. Species nova versimiliter *Lawrenciae densiflorae* affinis, a qua indumento denso e pilis parvis stellatis composito potius cinereo quam viridi, foliis conduplicatis nec applanatis seminibusque laevibus nec rugosis facile distinguitur.

Typus: Near Lake Auld, Western Australia, 22° 15′ S, 123° 45′ E, July 1967, A.S. George 9137 (holo: PERTH). This sheet includes male and female plants.

Sub-shrub to 130 cm tall, ascending, dioecious. Vestiture of young stems, stipules, leaves and adaxial surface of calyces densely cinereous-tomentose with small simple, bifurcate or haplomorphic stellate hairs. Leaves alternate or clustered on reduced branches, distinctly petiolate basal leaves merging into subsessile or sessile floral leaves; stipules green and leaflike, soon becoming brown and scarious, triangular to broadly ovate, 2.5-5.0 mm long x 1.8-3.0 mm wide, adaxial surface sometimes glabrous apart from a dense basal tuft of hairs, margin entire, apex acute to obtuse; petiole stout, semiterete, to 30 mm long; blade narrowly ovate to ovate, elliptic or obovate, 7-35 mm long x 2-23 mm wide, strongly conduplicate, venation indistinct except for a prominent thickened mid-vein, base cordate, truncate, obtuse or acute, margin coarsely dentate throughout, apex broadly obtuse to narrowly acute. Flowers unisexual, subsessile to sessile, axillary, solitary, often crowded in leafy fascicles. Calyx of male flower turbinate, 5.0-7.5 mm long; sepals connate ½-¾ their length; lobes triangular, 2.0-3.0 mm wide; apices acute. Corolla of male flower white becoming purple, 5.0-7.5 mm long; petals free almost to their bases or connate for up to 1/3 their length, weakly stellatehairy on abaxial surface along mid-vein, otherwise glabrous; lobes spreading, ovate, 2.0-2.5 mm wide; apices narrowly acute to acute; margin entire. Androecium of male flower with filament tube 3.1-4.0 mm long, glabrous; free portion of filaments 0.4-1.0 mm long; anthers 10-20. Gynoecium of male flower lacking. Calyx of female flower turbinate, 3.0-5.5 mm long; sepals connate 1/10-1/4 their length; lobes triangular, 1.3-2.0 mm wide; apices acute to obtuse. Corolla of female flower white becoming purple, 3.0-4.0 mm long; petals connate 1/3-5/8 their length, strongly stellate-hairy on abaxial surface along mid-vein, otherwise glabrous; lobes imbricate, erect, carinate, 0.9-1.2 mm wide; apices emarginate; margin entire. Androecium of female flower much reduced; filament tube 1.5-2.0 mm long; anthers lacking. Gynoecium of female flower with style 4.3-8.0 mm long; branches 2-3, free to base or connate for up to 1/7 their length, 0.1-0.2 mm wide, light brown, filiform, introrsely stigmatose, often on upper 2/3 only. Fruit ovoid, c. 2.2 mm high x 1.3-1.5 mm in diameter; mericarps 2-3; pericarp white or tan, membranous, becoming translucent, dehiscing to release naked seed a single one of which is fertile in each fruit. Seed triquetrous, smooth, brown, 1.8-2.2 mm high x 0.8-1.5 mm in radial width. Columella slender, 1.2-1.5 mm high x c. 0.1 mm in diameter.

Flowering period. July to October.

Habitat. Occurs on subsaline sandy clay or loam along ephemeral watercourses between sand dunes with 'Desert Oak' (Casuarina decaisneana F. Muell.), and in the Melaleuca zone adjacent to playa lakes.

Specimens examined. WESTERN AUSTRALIA: Well 30, Canning Stock Route, 22° 30′, 124° 08′, P. de Rebeira 415 (AD, PERTH); W of Lake Disappointment, 14 km N of Well 19, P. de Rebeira 422 (AD, PERTH); Near Lake Auld, A.S. George 9137 (holo: PERTH); Just W of Dragon Tree Soak, 19° 40′ S, 123° 21′ E, A.S. George 14812 (BH, NT, PERTH); Just W of Dragon Tree Soak, 19° 40′ S, 123° 21′ E, A.S. George 14813 (PERTH); s. loc., 1876, W.E.P. Giles s.n. (MEL); Well 30, Canning Stock Route, 22° 30′, 124° 08′, A.S. Mitchell 1023 (NT, PERTH).

Distribution. Found in the Canning and Keartland Districts of the Eremaean Botanical Province of Western Australia. See Map 6.

Etymology. The specific epithet draws attention to the dense vestiture of small stellate hairs which give this species its characteristic ashen-grey appearance.

Notes. The collection of Lawrencia cinerea made by W. Ernest P. Giles in 1876 is without locality details. Giles' Fifth Expedition of 1876 passed across the Little Sandy Desert (Giles 1889) which is in all likelihood the provenance of his specimen.

Herbarium specimens of Lawrencia cinerea have been referred to L. glomerata in the past.

The geographical range of Lawrencia cinerea overlaps with those of L. glomerata, L. squamata and L. viridi-grisea. L. cinerea has been found growing in close proximity to these other species.

7. Lawrencia densiflora (E. G. Baker) Melville, Kew Bull. 20: 514 (1966); J. Green, Census Vasc. Pl. Western Australia 67 (1981). *Plagianthus densiflorus* E. G. Baker, J. Bot. 30: 72 (1892). *Type:* Nickol Bay (as "Nicol Bay") and De Grey River, Western Australia, April-September 1863, *J. B. Ridley* s.n. (holo: K; iso: BM).

Plagianthus gardneri E. G. Baker ex Blackall & Grieve, How to Know Western Austral. Wildfl. 2: 346 (1956), nom. inval.

Sub-shrub to 60 cm tall, ascending, dioecious. Vestiture of stems, stipules, leaves and abaxial surface of calyces sparsely to densely hairy with simple, bifurcate and haplomorphic stellate hairs. Leaves alternate, distinctly petiolate basal leaves merging into subsessile or sessile floral leaves; stipules green and leaf-like, narrowly elliptic, often falcate, 2.5-4.0 mm long x 0.5-0.8 mm wide, margin ciliate, apex acute; petiole slender, triquetrous, to 70 mm

long: blade oboyate or broadly-cuneate in basal leaves, oboyate, ovate or narrowly cuneate in floral leaves, 4-25 mm long x 2-23 mm wide, flat, venation obscure or more or less distinct with 1-5 inconspicuous primary veins, base narrowly acute- to broadly obtuse-cuneate, margin coarsely dentate, apex acute obtuse or truncate. Flowers unisexual, sessile, solitary, often crowded in leafy fascicles arranged in dense spikes. Calyx of male flower turbinate, green, 3.0-6.5 mm long; sepals connate 2/5-4/5 their length, adaxially glabrous; lobes triangular, 1.8-3.5 mm wide; apices obtuse, acute or acuminate. Corolla of male flower membranous, white, 4.0-7.5 mm long; petals almost free to their bases or connate for up to 1/3 their length, glabrous or weakly stellate-hairy on abaxial surface; lobes 1.5-2.8 mm wide, spreading, ovate to obovate; apices obtuse, entire; margin entire or conspicuously ciliate. Androecium of male flower with filament tube 2.1-3.4 mm long, glabrous; free portion of filaments 0.9-1.5 mm long; anthers 20. Gynoecium of male flower lacking, or rarely present but sterile and much reduced; style filiform, 2.5-4.0 mm long; branches 3-5, 0.1-0.2 mm wide, free to bases or connate for up to 1/3 their length, dark brown. Calyx of female flower turbinate, green, 2.5-6.0 mm long; sepals connate ½-¾ their length, adaxially glabrous; lobes triangular, 1.5-4.5 mm wide; apices obtuse, acute or acuminate. Corolla of female flower white, 2.0-5.4 mm long; petals connate for 1/5-1/2 their length, glabrous to densely stellate-hairy on abaxial surfaces; lobes erect, carinate, ovate, 0.8-2.0 mm wide; apices obtuse to acute, entire; margin entire or conspicuously ciliate. Androecium of female flower much reduced with filament tube 0.5-2.6 mm long; anthers lacking. Gynoecium of female flower with style 2.0-7.0 mm long; branches 2-5, free to base of style or connate for up to 1/3 its length, filiform, 0.1-0.2 mm wide, brown, introrsely stigmatose. Fruit somewhat oblate to ovoid, 2.0-3.0 mm high x 2.0-3.4 mm in diameter; mericarps 2-5; pericarp white, membranous, becoming translucent, dehiscing to release naked seed a single one of which is fertile in each fruit. Seed triquetrous, angular on lateral surfaces, rugose on all surfaces, black, 1.3-2.2 mm high x 1.2-2.0 mm in radial width. Columella slender, 1.0-2.5 mm high x 0.1-0.3 mm in diameter. Figure 6.

Flowering period. July to October.

Habitat. Occurs on subsaline sand or clay soils in succulent steppe and low shrubland around playa lakes, saline depressions, dry watercourses and on limestone and gypsum ridges, often colonizing disturbed roadsides.

Selected specimens from 56 collections examined. WESTERN AUSTRALIA: c. 16 km SW of Earaheedy Homestead, R.J. Chinnock 941, (AD); between Mt Morgans & Laverton, C.A. Gardner 2441 (PERTH); 19 km NW of Cue on road to Weld Range, N.S. Lander 1125b (BH, BRI, CANB, MEL*, NSW, NT, PERTH); On Giralia road, 28 km E of Exmouth-Minilya road, N.S. Lander 1139, (NSW, PERTH); 8 km S of Wooramel Roadhouse, on North West Coastal Highway, N.S. Lander 1156, (BH, BM, BRI, CANB, K, NSW, PERTH); Nickol Bay (as "Nicol Bay") & De Grey River, April-Sept. 1863, J.B. Ridley s.n. (holo: K).

Distribution. Found in the Ashburton, Austin, Carnarvon, Fortescue and eastern margin of the Helms Districts of the Eremaean Botanical Province of Western Australia. See Map 6.

Etymology. The specific epithet draws attention to the dense inflorescences often observed in this species.

Notes. Several specimens held at BM, K and PERTH have been annotated as *Plagianthus gardneri* by E. G. Baker. However, the paper by Baker in which he set out to describe and name this putative new species (see notes under *Lawrencia glomeratus*) was never

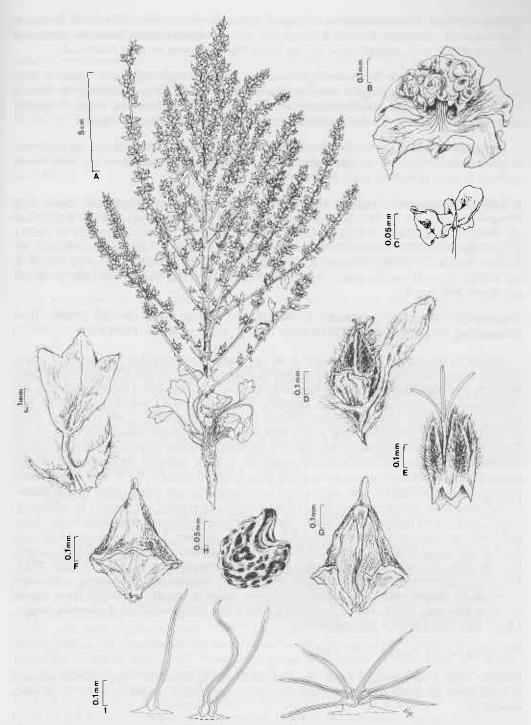


Figure 6. Lawrencia densiflora. A — Female plant, habit. B — Corolla of male flower and staminal tube. C — Anther after dehiscence. D — Female flower and subtending leaf. E — Female corolla and style branches after dehiscence of ovary cup. F — Immature fruit. G — Immature fruit with a single mericarp removed to show columella and membranous lateral mericarp walls. H — Ripe seed. I — Trichomes. J — Cauline leaf.

Drawn from Lander 1156.

published, hence the combination is invalid under the International Code of Botanical Nomenclature. Curiously, Blackall & Grieve (loc. cit.) adopted this invalid name for specimens of *P. densiflorus* (= *L. densiflora*) occurring in the region covered by their handbook.

Lawrencia densiflora is remarkably constant in its morphology over quite a wide geographical range. Although it varies somewhat in the extent of its branching, the density of its inflorescence and the density of its vestiture, nowhere does this variation appear geographically discrete or extensive enough to warrant sub-division of the species.

The geographical range of Lawrencia densiflora overlaps with those of L. chrysoderma, L. glomerata, L. helmsii, L. squamata and L. viridi-grisea. L. densiflora has been found growing in close proximity to these other species.

8. Lawrencia repens (S. Moore) Melville, Kew Bull. 20: 514 (1966); Beard, Descr. Cat. Western Austral. Pl. 84 (1970); Marchant & Keighery, Poorly Col. & Pres. Rare Vasc. Pl. Western Australia 61 (1979); J. Green, Census Vasc. Pl. Western Australia 67 (1981). Plagianthus repens S. Moore, J. Linn. Soc. 34: 179 (1899); C. A. Gardner, Enum. Pl. Austral. Occid. 78 (1931); Blackall & Grieve, How to Know Western Austral. Wildfl. 2: 345 (1956). Type: Gibraltar, near Coolgardie, Western Australia, September 1895, S. Moore s.n. (holo: BM; iso: K).

Plagianthus repens var. pentandra E. Pritzel, Bot. Jahrb. Syst. 35: 361 (1904). Type: Bullabulling, Western Australia, 29 October 1901, L. Diels 5204 (iso: PERTH).

Sub-shrub decumbent, to 15 cm tall x 40 cm wide, hermaphroditic. Vestiture of stems, stipules, leaves and abaxial surface of calvees moderately dense to densely hairy with simple, bifurcate and haplomorphic stellate hairs. Leaves alternate, petiolate; stipules colourless and membranous or green and leaf-like, narrowly triangular, 1.5-4.5 mm long x 0.3-0.6 mm wide, margin entire, apex acuminate; petiole slender, canaliculate, 2-45 mm long; blade cuneate or deltoid, 8-10 mm long x 3-12 mm wide, flat, venation obscure or more or less distinct with 1-3 inconspicuous primary veins, base obtuse-cuneate or truncate, margin coarsely dentate, apex obtuse. Flowers bisexual, subsessile, axillary, solitary, often crowded in leafy fascicles. Calvx turbinate, green, 4.0-6.0 mm long; sepals connate 3/5-4/5 their length; lobes broadly triangular, 2.0-3.5 mm wide; apices acuminate. Corolla white, often tinged yellow, 3.5-5.0 mm long; petals connate 1/5-1/2 their length, glabrous; lobes erect to spreading, ovate. 1.1-1.8 mm wide; apices obtuse. Androecium with filament tube 0.8-1.0 mm long, glabrous; free portion of filaments 0.5-1.2 mm long; anthers 5-10. Gynoecium with style 2.0-2.6 mm long; branches 5, free to their bases or connate for up to 1/5 their length, filiform, light to dark brown, introrsely stigmatose. Fruit oblate, 2.2-4.3 mm high x 3.0-4.0 mm in diameter; mericarps 5; pericarp white or tan, membranous, becoming translucent, dehiscing to release naked seed. Seed triquetrous, smooth, angular on lateral faces, rugose on distal face only, black, 1.5-2.2 mm high x 1.2-1.7 mm in lateral width. Columella slender, 1.8-2.5 mm high x 0.3-0.5 mm in diameter.

Flowering period. August to November.

Habitat. Occurs on subsaline sand, clay and loam soils in succulent steppe on the margins of small playa lakes and saline depressions; less commonly found on lateritic soil in *Eucalyptus* woodland.

Conservation status. On the basis of the single named specimen available to them at PERTH, Marchant & Keighery (1979) noted that Lawrencia repens was restricted to the Ravensthorpe area. Subsequent collecting and the determination of previously unidentified material at PERTH and from other herbaria in the course of the present study has shown this species

to have a much wider distribution. It can no longer be considered poorly collected or rare and would seem not to be endangered.

Selected specimens from 26 collections examined. WESTERN AUSTRALIA: 5 km NE of Norseman, c. 1 km N of Eyre Highway, A.C. Beauglehole 49335 (NT) (NT); Bullabulling, L. Diels 5204 (PERTH); Mt Moore, 1889, E. Merrall s.n. (MEL); near Lake Mason, June 1982, A.V. Milewski s.n. (PERTH); 19 km ENE of Norseman, K. Newbey 7537 (PERTH); 14 km NNW of Buningonia Spring, c. 35 km S of Zanthus, K. Newbey 7239 (PERTH).

Distribution. Found in the Avon District of the South-West Botanical Province, and in the Austin and Coolgardie Disticts of the Eremaean Botanical Province of Western Australia. See Map 6.

Etymology. The specific epithet draws attention to the prostrate, spreading (though not strictly repent) habit characteristic of this species.

Notes. Pritzel described his *Plagianthus repens* var. pentandra to accommodate specimens with 5 anthers. In the present study a continuum between androecia with 5 and 10 anthers was observed. Hence the variety has not been recognized.

The geographical range of Lawrencia repens overlaps with those of L. berthae, L. diffusa, L. glomerata, L. helmsii and L. squamata. L. repens has been observed growing in close proximity to these other species.

9. Lawrencia diffusa (Benth.) Melville, Kew Bull. 20: 514 (1966); Beard, Descr. Cat. Western Austral. Pl. 84 (1970); Marchant & Keighery, Poorly Col. & Pres. Rare Vasc. Pl. Western Australia 61 (1979); J. Green, Census Vasc. Pl. Western Australia 67 (1981). Plagianthus diffusus Benth., Fl. Austral. 1: 190 (1863); E. G. Baker, J. Bot. 30: 72 (1892); C. A. Gardner, Enum. Pl. Austral. Occid. 78 (1931). Lectotype (here designated): Swan River, Western Australia, s.dat., J. Drummond 104 (lecto & isolecto: K). Lectoparatypes: Swan River, J. Drummond 137 (K-L glomerata); Swan River, J. Drummond 246 (K, BM-L. glomerata); Swan River, J. Drummond 275 (K, BM-L. berthae).

Diminutive sub-shrub, prostrate, compact, to c. 15 cm wide, hermaphroditic. Vestiture of stems, stipules, leaves and abaxial surface of calyces moderately densely hairy or subglabrous with simple, bifurcate and haplomorphic stellate hairs, but usually glabrous. *Leaves* alternate, distinctly petiolate throughout; stipules colourless and membranous, narrowly ovate, ovate or orbicular, c. 3 mm long x 0.7-1.4 mm wide, margin ciliate, apex obtuse or broadly obtuse; petiole slender, canaliculate, 2-15 mm long; blade elliptic, ovate, obovate or deltoid, 2.0-8.5 mm long x 2.0-7.0 mm wide, somewhat succulent, flat, venation obscure, base acuteor obtuse-cuneate, margin coarsely dentate, apex acute, obtuse or rounded. Flowers bisexual, subsessile, axillary, solitary, often crowded in leafy fascicles. Calyx broadly turbinate, green, 3.5-5.5 mm long; sepals connate 1/4-1/2 their length, abaxially glabrous or with a few stellate hairs at the junction of the lobes; lobes broadly triangular, 2.0-3.0 mm wide; apices acuminate. Corolla white, 3.0-3.5 mm long; petals free almost to their bases or connate to 2/3 their length, glabrous; lobes erect to spreading, ovate, 1.2-1.5 mm wide; apices obtuse. Androecium with filament tube 0.7-1.0 mm long, glabrous; free portion of filaments 0.8-1.0 mm long; anthers 5. Gynoecium with style 1.8-2.5 mm long; branches 5, free to their bases, c. 0.1 mm wide, white or light brown, filiform, introrsely stigmatose. Fruit oblate, 1.5-2.0 mm high x 2.5-3.0 mm in diameter; mericarps 5; pericarp membranous, becoming translucent, dehiscing to release naked seed. Seed triquetrous, angular on lateral faces, black, 1.4-1.7 mm high x 0.8-1.3 mm in radial width. Columella slender, 1.5-2.0 mm high x 0.2-0.3 mm in diameter.

Flowering period. September to November.

Habitat. Occurs on subsaline clay soils in succulent steppe in and around playa lakes and saline depressions.

Conservation status. On the basis of the single named specimen available to them at PERTH, Marchant & Keighery (1979) noted that Lawrencia diffusa was restricted to the Fitzgerald River National Park. Leigh et al. (1981) categorized it as rare but not currently considered endangered or vulnerable. Subsequent collecting and the determination of previously unidentified material at PERTH and from other herbaria in the course of the present study has shown this species to be of much wider distribution. It can no longer be considered poorly collected or rare and would seem not to be endangered at present.

Selected specimens from 16 collections examined. WESTERN AUSTRALIA: c. 4.9 km N of Lake Cronin Crossroads, R.J. Chinnock 4136 (AD); near Woody Lake, NE of Esperance, June 1970, A.S. George s.n. (PERTH); 14 km S of Clear Streak Well, c. 70 km SE of Norseman, K. Newbey 7575a (PERTH); Hunts Well, Nov. 1891, R. Helms s.n. (AD, K, MEL); Yuinmery Homestead, near Youanmi Lake, Oct. 1981, A.V. Milewski s.n. (PERTH); Fitzgerald River National Park, c. 20 km N of Bremer Bay on E side of No. 2 Vermin Proof Fence, (P.G. Wilson 10163 PERTH); 6 km N of Borden, Sept. 1966, P.G. Wilson s.n. (PERTH).

Distribution. Found in the Eyre and Roe Districts of the South-West District and in the Austin and Coolgardie Districts of the Eremaean Botanical Province of Western Australia. See Map 6.

Etymology. The specific epithet refers to the prostrate, spreading habit characteristic of this species.

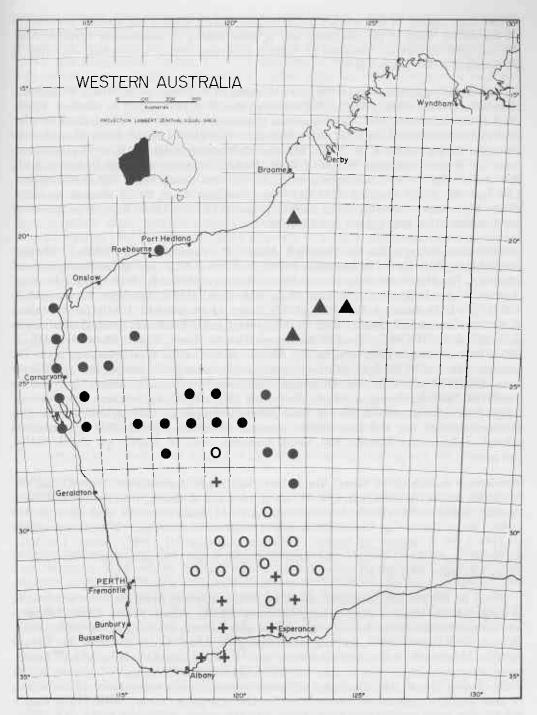
Notes. In his protologue of *Plagianthus diffusus*, Bentham (loc. cit.), listed four syntypes, namely Swan River, Drummond 104 (K), Drummond 137 (K), Drummond 246 (K, BM) and Drummond 275 (K, BM). These specimens comprise three distinct taxa which can easily be separated. Of them, Drummond 104 corresponds most nearly to the protologue and has been designated as the lectotype above. Thus the remaining collections become lectoparatypes. However, Drummond 137 & 246 are clearly specimens of *Lawrencia glomerata* Hook.; likewise, Drummond 275 is clearly a specimen of *L. berthae* (F. Muell.) Melville.

The geographical range of Lawrencia diffusa overlaps with those of L. berthae, L. glomerata, L. helmsii, L. repens and L. spicata. L. diffusa has been found growing in close proximity to each of these species.

Series SELENOTHAMNUS

Lawrencia series Selenothamnus (Melville) Lander, stat. nov. Selenothamnus Melville, Kew Bull. 20: 514-5 (1966); Hutchinson, Gen. Fl. Pl. Dicot. 2: 53 (1967); Baines, Austral. Pl. Gen 338-9 (1981). Type: S. squamatus (Nees ex Miq.) Melville [=Lawrencia squamata Nees ex Miq.].

Erect shrubs; dioecious; with or without thorns; vestiture stellate-hairy or squamulose with peltate scales, never glabrous; stems with spongy periderm producing dry fibrous or flaky bark; leaves sessile throughout; female flowers with or without sterile anthers; style branches filiform or clavate; sterile mericarps crushed and obliterated by developing fertile mericarps; seed swollen, irregularly globular, smooth.



Map 6. Distributions of Lawrencia cinerea (▲), L. densiflora (•) L. repens (O) and L. diffusa (+)

The three species in this section are extremely closely related and careful attention to all the details noted in the diagnostic key is often needed to differentiate between them. Two of these species are confined to Western Australia; one is widespread, occurring in all mainland states (Maps 7 & 8).

10. Lawrencia squamata Nees ex Miq., in Lehmann, Pl. L. Preiss. 1: 242 (1845). Plagianthus squamatus (Nees ex Miq.) Benth., J. Linn. Soc., Bot. 6: 103 (1862); C. A. Gardner, Enum. Pl. Austral. Occid. 78 (1931). Selenothamnus squamatus (Nees ex Miq.) Melville, Kew Bull. 20: 515 (1966); Cochrane et al, Fl. & Pl. Victoria t. 1677 (1968); Beard, Descr. Cat. Western Australia Pl. 84 (1970); J. H. Willis, Handb. Pl. Victoria 2: 38l (1973); J. Green, Census Vasc. Pl. Western Australia 67 (1981); W. R. Barker, in Jessop, Fl. Central Australia 212 & t. 246 (1981); G. M. Cunningham et al., Pl. W. New South Wales 492 (1982); Jessop, List Vasc. Pl. South Australia 62 (1983). Type: Southern River, Perth, Western Australia, Preiss 1231, Sept. 1841 (holo: LD non vidi; iso: MEL). Southern River is a branch of the Canning River, near present day Thornlie.

Halothamnus microphyllus (F. Muell.) F. Muell., Pl. Victoria 1: 159 (1862); F. Mueller, Second Gen. Rep. Gov. Parliamentary Papers — Votes & Proc. of the Legislative Council 1854-5 (A. No. 18): 10, sine descriptione. Plagianthus microphyllus F. Muell., Fragm. Phytogr. Austral. 1: 29 (1858) Benth., J. Linn. Soc., Bot. 6: 103 (1862); Bentham, Fl. Austral. 1: 190 (1863); E. G. Baker, J. Bot. 30:72 (1892); Bailey, Queensland Fl. 1: 110 (1899); Maiden & Betche, Census New South Wales Pl. 136 (1916); J. M. Black, Fl. South Australia, edn 2, 3: 557 & t. 718 (1952); Blackall & Grieve, How to Know Western Austral. Wildfl. 2: 345 (1956). Lectotype (here designated): Murray River, s.dat., F. Mueller, s.n. (lecto: K; isolecto: BM, Mel 98788 & 98789). Lectoparatypes: Spencers Gulf, October 1857, F. Mueller s.n. (MEL 98790, 98812 & 584132); Spencers Gulf near Port Pirie, October 1857, F. Mueller s.n. (MEL 584133); Murray, s.dat., F. Mueller s.n. (MEL 98791, K); Seaflats on the Murray, s.dat. F. Mueller s.n. (MEL 98787); Lake Albert Salt Springs, 1854, F. Mueller s.n. (K); Between Guichen Bay and Lake Albert, s.dat., F. Mueller s.n. (MEL 584131 — material affixed to sheet only); Guichen Bay, South Australia, s.dat., F. Mueller s.n. (MEL 584131 pro pte).

Plagianthus incanus J. M. Black, Trans. Roy. Soc. South Australia 49: 274 (1925); J. M. Black, Fl. South Australia, edn 2, 3: 557 (1952); Blackall & Grieve, How to Know Western Austral. Wildfl. 2: 345 (1956). Lawrencia incana (J. M. Black) Melville, Kew Bull. 20: 514 (1966); Beard, Descr. Cat. Western Austral. Pl.; J. Green, Census Vasc. Pl. Western Australia 67 (1981); W. R. Barker, in Jessop, Fl. Central Australia 211 (1981); Jessop, List Vasc. Pl. South Australia 62 (1983). Type: Gawler Range, South Australia, September 1912, S. White s.n. (holo: AD; iso: K).

Shrub to 100 cm tall, spreading, dioecious, with secondary branches often terminating in a rigid thorn. Vestiture of branches glabrous to densely tomentose with simple, bifurcate and actinomorphic stellate hairs, or squamulose with minutely fimbriate peltate scales, rarely pannose with palmate hairs, clothing a thin spongy periderm which produces grey fibrous bark on older stems; leaves and abaxial surfaces of calyces densely tomentose with simple, bifurcate and actinomorphic stellate hairs, or squamulose with minutely fimbriate peltate scales. Leaves densely fasciculate on short lateral branches, sessile throughout; stipules brown or pale green, membranous, narrowly to broadly triangular, falcate, 0.4-2.1 mm long x 0.2-2.3 mm wide, vestiture of abaxial and adaxial surfaces sparse to dense with peltate scales or palmate hairs, margin ciliate with stellate hairs, apex acuminate or acute; blade linear, narrowly elliptic, ovate, obovate, spathulate or suborbicular, flat or conduplicate, 1.6-19.0 mm long x 0.5-11.0 mm wide, abaxial and adaxial surfaces dense with either stellate hairs or with peltate scales, or with palmate scales at the base merging into peltate scales,

venation obscure, base narrowly to broadly cuneate, margin entire or coarsely 3-5 dentate, apex acuminate, acute, obtuse or truncate. Flowers unisexual, sessile, axillary, solitary, often crowded in leafy fascicles. Calyx of male flowers turbinate to broadly turbinate, green, 2.1-4.8 mm long; sepals connate 3/5-9/10 their length, vestiture on abaxial surface dense with stellate hairs or peltate scales, adaxial surface glabrous; lobes triangular, 1.0-2.5 mm wide; apices narrowly to broadly acuminate, acute or obtuse, frequently with a small dark mucro. Corolla of male flower white, yellow or reddish, often tinged violet in bud, 2.0-6.5 mm long; petals free almost to base or connate up to 1/2 their length, glabrous; lobes spreading, flat, narrowly to broadly elliptic, ovate or obovate, 0.8-2.5 mm wide; apices acute or obtuse to broadly obtuse, entire or emarginate, often faintly to coarsely irregularly dentate. Androecium of male flower with filament tube 0.4-3.6 mm long, glabrous; free portion of filaments 0.2-0.8 mm long; anthers 10-20. Gynoecium of male flower much reduced and sterile or lacking. Calyx of female flowers turbinate to broadly turbinate, green, 2.2-5.9 mm long; sepals connate 3/4-9/10 their length, vestiture on abaxial surface dense with stellate hairs or peltate scales, adaxial surfaces usually glabrous, rarely dense with stellate hairs; lobes triangular, 1.0-2.3 mm wide; apices narrowly to broadly acuminate, acute or obtuse, frequently with a small dark mucro. Corolla of female flower white or yellow or reddish, often tinged violet in bud, 2.3-6.9 mm long; petals connate 1/5-3/5 their length, glabrous; lobes erect to somewhat spreading, carinate, conduplicate or almost flat, narrowly to broadly elliptic, ovate or obovate, 0.5-3.0 mm wide; apices narrowly to broadly acute or obtuse, entire or emarginate, often faintly to coarsely irregularly dentate. Androecium of female flower with filament tube 0.1-1.8 mm long, glabrous; free portion of filaments 0.1-0.4 mm long; anthers 10-20, sterile. Gynoecium of female flower with style 1.8-6.1 mm long; branches 2-5, free to base of style or connate for up to 1/3 its length, filiform or clavate, entire, 0.1-0.7 mm wide, introrsely stigmatose. Fruit ovoid, 2.3-3.6 mm high x 2.3-3.8 mm in diameter; mericarps 2-5; pericarp tan, membranous becoming translucent, dehiscing to release naked seed a single one of which is fertile in each fruit, sterile mericarps crushed and almost obliterated. Seed swollen, irregularly globular, dark brown, smooth, 0.6-3.3 mm high x 0.4-2.5 mm in radial width. Columella filamentous, 2.8-3.8 mm high.

Flowering period. Mostly from August to January, sporadically throughout the year.

Habitat. Occurs on subsaline sand or calcareous red soils in succulent steppe, low shrubland or open woodland on the margins of inland playa lakes, saline flats and depressions, ephemeral watercourses, tidal flats and coastal inlets, sometimes amongst coastal limestone or granite debris, occasionally on gypsum or limestone ridges inland.

Selected specimens from 298 collections examined. NEW SOUTH WALES: "Sunnyside", near Balranald, B.M. Alchin 13 (NSW); Cobham Salt Lake, W. Bauerlen 254 (NSW); "Mt Mulyah", c. 80 km NW of Louth, C.W.E. Moore 6760 (CANB, NSW); 17 miles (27 km) W of Euston, J.H. Leigh W2O (NSW); Utah Lake, K. Paijmans 3319 (CANB).

NORTHERN TERRITORY: Mt Wedge, 22° 48′ S, 131° 44′ E, C.R. Dunlop 2452 (K, NSW); "Newhaven", 22° 51′ S, 131° 13′ E, P.K. Latz 2129 (AD); "Napperby", 22° 43′ S, 132° 23′ E, P.K. Latz 5946 (AD, NT); Dalhousie Springs, 26° 27′ S, 135° 28′ E, P.K. Latz 4797 (AD, BH, MEL, NT, PERTH); c. 1 mile (1.6 km) W of Central Mt Wedge, 22° 50′ S, 131° 50′ E, A.O. Nichols 808 (K, MEL).

QUEENSLAND: 5 km NE of South Glen Homestead, 28° 11′ S, 146° 33′ E, R.W.

Purdie 759 D (BRI).

SOUTH AUSTRALIA: c. 7 km SW of Copley, on Coply-Beltana road, *Hj. Eichler* 13001 (AD); Diamond Lake, between Beaufort and Whitwarta, *R. Hill* 1006 (AD); Walkers Flat, c. 93 km NE of Adelaide, *D.N. Kraehenbuehl* 918 (AD); c. 1.6 km W of Coober Pedy, *T.R.N. Lothian* 4346 (AD); c. 20 km W of Oldea, *T.R.N. Lothian* 5508 (AD); Wittons Bluff, Pt Noarlunga, *T.J. Smith* 439 (AD); Royal Park, c. 10 km NE of Adelaide, *T.J.*

Smith 1727 (AD); 9 miles (14.5 km) SW of Malinong Hall, M.C.R. Sharrard 1224 (AD); Dalhousie Springs, c. 80 miles (128 km) N of Oodnadatta in the vicinity of Spring Hut, D.E. Symon 3251 (CANB, K); Shell Beach, Innes National Park, J.Z. Weber 4229 (AD); Head of Bight, P.G. Wilson 1621 (AD); St Francis Island, c. 60 km SW of Ceduna, N.M. Wace 136 (AD).

VICTORIA: Raak Salt Plains, 9 miles (14.5 km) WNW of Hattah, A.C. Beauglehole 40583 (MEL); near Kiatta, R. Melville 988 (K, MEL); Wimmera, s.dat., J. Dallachy s.n.

(K); near Annuello, Sept. 1971, R. Wade s.n. (MEL).

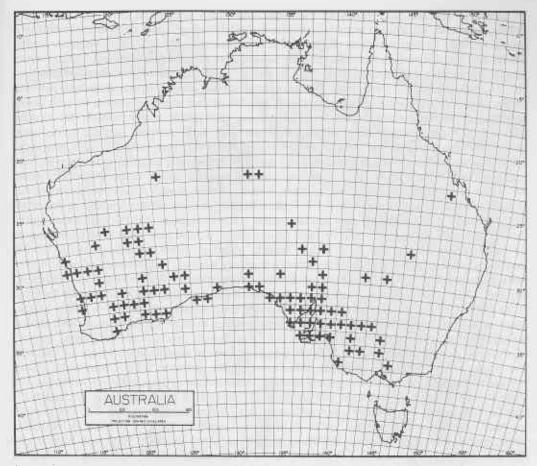
WESTERN AUSTRALIA: 10 miles (16 km) S of Leonora on road to Menzies, T.E.H. Aplin 230((PERTH); 17 miles (27 km) N of Melrose Homestead, J.S. Beard 6534 (PERTH); Israelite Bay, 1885, S.T.C. Brooks (as "Brooke") s.n. (MEL); 6.5 miles (10.5 km) W of Winchester, s.dat., C. Chapman s.n. (PERTH); Bern Pool, "Yelma", R.J. Chinnock 772 (AD); Cannington Swamp, Wattle Grove, R. Coveny 8258 (NSW, PERTH); c. 30 km NNE of Stokes' Inlet, Hj. Eichler 20322 (AD, PERTH); Lake Auld, A.S. George 9131 (BRI, MEL, NSW, NT, PERTH); 13 miles (20.5 km) from Chester Pass down South Stirlings Road, Dec. 1964, A.S. George s.n. (PERTH); 12 km NE of Norseman, K. Newbey 8095 (AD, PERTH); Rawlinna, P.G. Wilson 5798 (PERTH); 1 km W of Meckering in flood plain of Mortlock River, P.G. Wilson 6403 (AD, BH, BRI, MEL, NSW, NT, PERTH). N end of Lake Moore, P.G. Wilson 8635 (K, PERTH).

Distribution. Found in the Avon, Darling (Drummond Subdistrict), Eyre, Irwin and Roe Districts of the South-West Botanical Province, and in the Austin, Canning, Coolgardie, Eucla and Helms Districts of the Eremaean Botanical Province of Western Australia; in the western sector of the Central Australia Pastoral District of the Northern Territory; in the Eastern, Eyre Peninsula, Flinders Range, Gairdner-Torrens Basin, Lake Eyre Basin, Murray, Northern Lofty, Nullarbor, south-eastern Southern Lofty and Yorke Peninsula Regions of South Australia; in the Ballarat, Mallee, Murray Valley, South West and Wimmera Study Areas of Victoria; in the Far Western Plains (North and South Subdivisions) and Western Plains (North and South Subdivisions) Botanical Divisions of New South Wales; and in the Maranoa Pastoral District of Queensland. See Map 7.

Etymology. The specific epithet refers to the densely squamulose vestiture of peltate scales found on the stems, leaves and calyces of the type form of this species, giving the plant an ashen appearance.

Notes. Like Lawrencia glomerata (see above), L. squamata is a widespread and highly polymorphic species comprising many allopatric populations. Nonetheless, it is rather surprising that specimens formerly referred to L. incana and L. squamata should have been located in different genera by Melville (1966). Barker (1981) has drawn attention to this bewildering anomaly. To be sure, branching pattern, leaf shape and size, thorniness, and hair type are extremely variable, but examination of all available herbarium specimens in the course of this study reveals a continuum in each of these characters. Specimens with predominantly actinomorphic stellate hairs usually have longer, broader leaves than those with predominantly peltate scales, and they have a generally more northerly distribution. Plants at either extreme can look strikingly different, and in the past L. incana and L. squamata sensu stricto have been distinguished on this basis. However, in their essential characters of flowers, fruit and seed these and other forms are remarkably constant: in keeping with the principles expressed earlier in this paper, I am unwilling to maintain their separation, even at an infraspecific level.

Lawrencia squamata is the only species in the genus to possess thorns. These are not invariably present and their presence or absence seems not to be geographically correlated. R. F. Parsons (pers. comm.) has suggested that small-leaved, non-thorny plants and large-



Map 7. Distribution of Lawrencia squamata.

leaved, thorny plants of L. squamata may be edaphically segregated, with the former occupying sandy, gypseous soils and the latter inhabiting loamy, non-gypseous soils. Whilst this is evidently the case in New South Wales and Victoria, I have not observed such a pattern elsewhere, nor have other collectors commented on it.

The geographical range of Lawrencia squamata overlaps with those of all other species in the genus with the exception of L. buchananensis. L. squamata has been found growing in close proximity to these other species.

11. Lawrencia chrysoderma Lander, sp. nov.

Lawrencia chrysoderma est frutex dioicus ramis secundariis aliquantum contractis caules dense foliatos formantibus, indumento magna pro parte e pilis peltatis consistente. Fructus e mericarpiis 2-3 dehiscentibus compositus, pericarpio membranaceo translucente. Haec species Lawrenciae helmsii simillima, a qua ramificatione magis aperta, foliis majoribus, calycibus corollisque uniformibus nec sexualiter dimorphis distinguitur.

Typus: Billi Billi Claypans, W end of Weld Range, Western Australia, 26° 58′ S, 117° 33′ E, October 1969, R.A. Saffrey 816 (holo: PERTH). The holotype sheet includes both male and female plants.

Shrub to 100 cm tall, spreading, ramulose and much contracted to form dense leafy stems, dioecious. Vestiture of branches, yellowish, densely pannose with palmate scales clothing an exceedingly spongy periderm which produces conspicuous dry flaky bark on older stems; leaves and abaxial surface of calyces densely squamulose with fimbriate peltate scales. Leaves sessile throughout, densely fasciculate; stipules pale green, dry and spongy, narrowly triangular, 1.5-7.0 mm long x 0.5-2.3 mm wide, falcate, abaxial surface densely squamulose with palmate scales at the base merging into peltate scales, adaxial surface almost glabrous or with dense vestiture of palmate scales on the mid-rib merging into peltate scales towards the margin and apex, margin ciliate with simple, bifurcate and haplomorphic stellate hairs, apex acuminate; blade ovate, narrowly obovate or spathulate, 4-12 mm long x 1.7-3.0 mm wide, conduplicate, reflexed, abaxial and adaxial surfaces dense with palmate scales at the base merging into peltate scales, base narrowly cuneate, margin entire, apex acuminate, acute or obtuse. Flowers unisexual sessile, axillary, solitary. Calyx of male flowers turbinate, green, 3.4-4.5 mm long; sepals connate 3/5-4/5 their length, abaxial surface densely squamulose with palmate scales merging into peltate scales; lobes triangular, 1.0-1.5 mm wide; apices acute to broadly obtuse. Corolla of male flower white, 3.5-5.0 mm long; petals free almost to base or connate for up to 3/5 their length, glabrous; lobes spreading, ovate, 0.8-2.0 mm wide; apices acute, obtuse or truncate, irregularly dentate. Androecium of male flower with filament tube 1.7-2.5 mm long, glabrous; free portion of filaments 0.3-1.5 mm long; anthers 10-20. Gynoecium of male flower very much reduced and sterile, sometimes lacking. Calyx of female flower turbinate, green, 3.3-5.5 mm long; sepals connate 1/2-4/5 their length, abaxial surface densely squamulose with palmate scales at base merging into peltate scales, adaxial surface glabrous; lobes triangular, 0.8-2.0 mm wide apices acute to broadly obtuse. Corolla of female flowers white, 3.0-5.0 mm long; petals almost free to base or connate for up to 3/5 their length, glabrous; lobes erect, ovate, carinate, 0.8-2.0 mm wide; apices obtuse to truncate, entire or irregularly dentate. Androecium of female flower with filament tube 0.7-1.6 mm long, glabrous; free portion of filaments 0.1-0.9 mm long; anthers 5-15, sterile. Gynoecium of female flower with style 2.0-7.0 mm long; branches 2-3, free almost to base of style or connate for up to 7/10 of its length, filiform or clavate, 0.1-1.0 mm wide, apices often briefly-lobed, introrsely stigmatose. Fruit ovoid, 2.3-4.0 mm high x 1.8-2.5 mm in diameter; mericarps 2-3; pericarp white or tan, membranous, becoming translucent, dehiscing to release naked seed a single one of which is fertile in each fruit, sterile mericarps crushed and almost obliterated. Seed swollen, irregularly globular, smooth, brown, 1.5-2.6 mm high x 1,1-1.5 mm in radial width. Columella filamentous, c. 2.0 mm high. Figure

Flowering period. August to January.

Habitat. Occurs on subsaline sand or clay soils in lightly wooded succulent steppe on margins of playa lakes, particularly on gypsum ridges.

Specimens examined. WESTERN AUSTRALIA: 44 miles (70.4 km) NW of Cue on road to Mileura, T.E.H. Aplin 2531 (PERTH); "Modoonga", 26° 56′ S, 117° 36′ E, J.S. Beard 6612 (PERTH); Eneabba Flora Reserve, Oct. 1967, C. Chapman s.n. (BM, BRI, CANB, K, NSW, NT, PERTH); Salt lake SW of Winchester, J. Coleby-Williams 117 (PERTH); Salt lake SW of Winchester, J. Coleby-Williams 118 (PERTH); 9 km from Winchester along road to Eneabba, 0.5 km S of road, S end of Yarra Yarra Lakes, M.D. Crisp 5469 (CANB); "Barnong", A.W. Humphries P48 (PERTH); N of Weld Range, opposite Madoonga Homestead beside road from Cue, N.S. Lander 1122 (K, MEL*, PERTH); N of Weld Range opposite Madoonga Homestead beside road from Cue, N.S. Lander 1123 (CANB, PERTH); locality unknown (as "Puttingup"), s.dat, G. Maxwell s.n. (MEL); "Roderick", c. 150 km N of Yalgoo, A.A. Mitchell 968 (MEL*, PERTH); Billi Billi Claypans, W end of Weld Range, 26° 58′ S, 117° 33′ E, R. A. Saffrey 816 (holo: PERTH); "Barnong", April

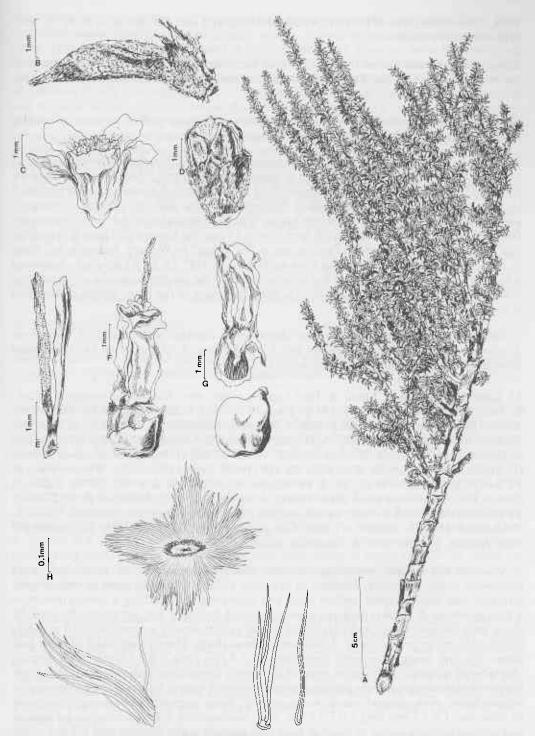


Figure 7. Lawrencia chrysoderma. A — Male plant, habit. B — Leaf. C — Corolla of male flower. D — Female flower. E — Style. F — Withered corolla and mature fruit in process of dehiscence. G — Withered corolla, spent fruit retaining crushed mericarp and sterile ovule, and released seed. H — Trichomes.

Drawn from Saffrey 816 (holotype).

1954, D.G. Wilcox s.n. (PERTH); N end of Mongers Lake, 28° 46′ S, 117° 22′ E, P.G. Wilson 8604 (PERTH).

Distribution. Found in the Irwin District of the South-West Botanical Province and in the Austin District of the Eremaean Botanical Province of Western Australia. See Map 8.

Etymology. The specific epithet draws attention to the dense yellow vestiture of stellate hairs on the young branches characteristic of this species.

Notes. A mixed collection of Lawrencia chrysoderma and L. squamata communicated to Ferdinand von Mueller by G. Maxwell is annotated "Puttingup." This locality is cited by Mueller (1875) under Plagianthus microphyllus [L. squamata]. A thorough search has failed to reveal that any place name exists in Western Australia with the spelling "Puttingup" and no aboriginal place name anywhere begins "Put..." (Nomenclature Advisory Committee, Office of the Surveyor General, Perth, pers. comm.) From the handwritten label it is possible that "Pullingup" was intended. There is not a "Pullingup" in Western Australia, but there is a Pulingup Spring near Pallinup river at 34° 18′ S, 118° 23′ E. Although L. squamata is to be found in that vicinity today it is far removed from the present range of L. chrysoderma: it would seem that the attribution of Maxwell's specimen of the latter species to this area is in error.

The geographical range of Lawrencia chrysoderma overlaps with those of L. densiflora, L. glomerata, L. helmsii, L. squamata and L. viridi-grisea. L. chrysoderma has been found growing in close proximity to all these species.

12. Lawrencia helmsii (F. Muell. & Tate) Lander, comb. nov. Plagianthus helmsii F. Muell. & Tate, Bot. Centralb. 55: 316 (1893); Trans. Roy. Soc. South Australia 16: 339 (1896); Anon., Proc. Linn. Soc. London 1908-9: 9 (1909); Morrison, J. Nat. Hist. & Sci. Soc. Western Australia 3: t.2 (1907); C. A. Gardner, Enum. Pl. Austral. Occid. 78 (1931); Blackall & Grieve, How to Know Western Austral. Wildfl. 2: 346 (1956). Selenothamnus helmsii (F. Muell. & Tate) Melville, Kew Bull. 20: 515 (1960); Beard, Descr. Cat. Western Austral. Pl. 84 (1970); Erickson et al., Fl. & Pl. Western Australia 154 & t. 487 (1973); J. Green, Census Vasc. Pl. Western Australia 67 (1981); A. Mitchell, in B. D. Morley & H. R. Toelken, Fl. Pl. Australia t. 79c & d (1983). Lectotype (here designated): Lake Lefroy, Western Australia, 7 November 1891, R. Helms s.n. (lecto: AD; isolecto: K, NSW), 981553261). Lectoparatype: Lake Annean, October 1893, S. Dixon s.n. (AD).

Shrub to 100 cm tall, ascending, ramulose and much contracted to form dense leafy cactus-like stems, dioecious. Vestiture of branches yellowish, densely pannose with simple, bifurcate and haplomorphic stellate hairs and palmate scales clothing a spongy periderm which produces dry flaky bark on mature exposed branches but normally obscured by dense leafy fascicles; leaves and abaxial surfaces of calyces densely squamulose with minutely fimbriate to entire peltate scales. Leaves sessile throughout, densely fasciculate; stipules pale green, dry and spongy, narrowly triangular, 1.2-3.5 mm long x 0.9-3.0 mm wide, falcate, chartaceous, abaxially glabrous or sparsely to densely squamulose with palmate scales, often becoming glabrous towards the margin, margin ciliate with simple, bifurcate and haplomorphic stellate hairs, apex narrowly acute to acuminate; blade narrowly obovate, elliptic, ovate or orbicular, 1.5-7.2 mm long x 0.7-2.2 mm wide, conduplicate, reflexed, vestiture on abaxial and adaxial surfaces sparse to dense at base with palmate scales merging into peltates scales on lamina, base narrowly cuneate, margin entire, apex acute to broadly obtuse. Flowers unisexual, sessile, axillary, solitary. Calyx of male flowers turbinate, 1.5-4.5 mm long, green; sepals connate at least 4/5 their length, abaxial surface densely squamulose with stellate

hairs merging into palmate and peltate scales; lobes shallow, broadly triangular, 1.0-1.3 mm wide; apices broadly acute to broadly obtuse. Corolla of male flower white, or light green, 2.0-5.6 mm long; petals connate for 1/5-3/5 their length, glabrous; lobes flat, spreading, obovate, 0.6-2.0 mm wide; apices acute, obtuse to broadly obtuse, entire or emarginate. Androecium of male flower with filament tube 0.5-2.7 mm long, glabrous; free portion of filaments 0.3-0.8 mm long; anthers 5-15, sterile. Gynoecium of male flower much reduced and sterile or lacking. Calyx of female flowers campanulate, 1.8-3.8 mm, long, green; sepals connate at least 7/10 their length, abaxial surface densely squamulose with stellate hairs merging into palmate or peltate scales; lobes triangular, 0.6-0.8 mm wide; apices acute to broadly obtuse. Corolla of female flowers white or light green, 1.7-3.0 mm long; petals connate for 1/3-4/5 their length, glabrous; lobes erect, flat or slightly carinate, ovate, 0.5-1.5 mm wide; apices broadly acute to broadly obtuse, entire or emarginate. Androecium of female flowers with filament tube 0.3-1.0 mm long, glabrous; free portion of filaments 0.2-0.3 mm long; anthers 0-8, sterile. Gynoecium of female flowers with style 0.4-3.2 mm long; branches 2, filiform or clavate, entire, free to base of style or connate for up to 1/3 its length, 0.1-0.4 mm wide, introrsely stigmatose. Fruit ovoid, 1.5-2.4 mm high x 1.6-2.0 mm in diameter; mericarps 2; pericarp tan, membranous, becoming translucent, dehiscing to release naked seed a single one of which is fertile in each fruit, sterile mericarps crushed and almost obliterated. Seed swollen irregularly globular, brown, smooth, 0.8-1.7 mm high x 1.0-1.4 mm in radial width. Columella filamentous, c. 1.5 mm high.

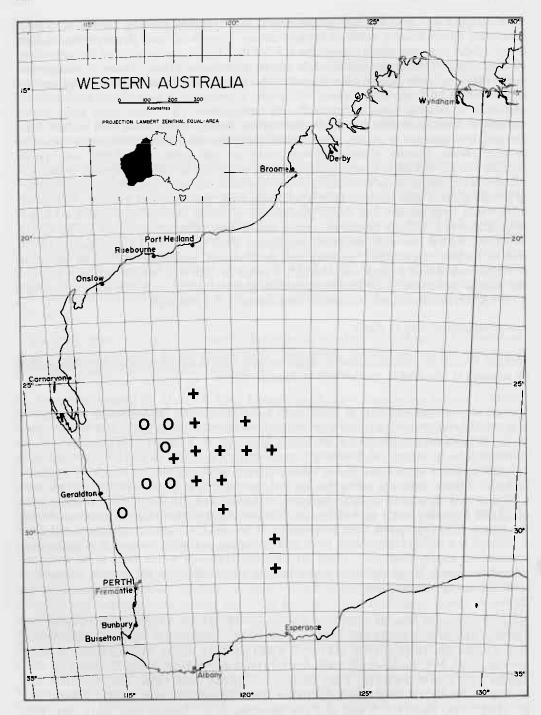
Flowering period. July to April.

Habitat. Occurs on subsaline sand or gypseous clay soils in lightly wooded succulent steppe on margins of playa lakes, particularly on gypsum ridges.

Conservation status. As Rye et al. (1980) note, Lawrencia helmsii (as Selenothamnus helmsii) is at present commercially exploited for use in dried flower arrangements, doubtless because of its extraordinary cactus-like appearance. Marchant & Keighery (1979) considered its occurrence to be in need of assessment and monitoring. Leigh et al. (1981) categorize it as a species with a range over 100 km but occurring in small populations which are restricted to highly specific habitats, vulnerable but not presently endangered although at risk over a long period through continued depletion. Burgman and Hopper (1982) record a total of 22,000 flowering stems picked for the Western Australian wildflower industry during the period 1980-1981 taken from populations at Mount Sir Samuel, Sandstone and Cue. L. helmsii is a highly conspicuous plant and its habitat is readily located. It is gregarious, proliferating readily from seed. From my own observation it would seem to be under no immediate threat, but populations could become vulnerable were the present intensity of bush harvesting to be sustained.

Selected specimens from 66 collections examined. WESTERN AUSTRALIA: 2 miles (3.2 km) S of Mount Sir Samuel, Oct. 1947, G.E. Brockway, s.n. (CANB, PERTH); Lake Austin, L.A. Craven 5033 (BRI, CANB, MEL, NT, PERTH); Lake Austin, on road from Daydawn to "Lakeside", N.S. Lander 1120 (MEL*, PERTH); Lake Annean, Great N. Highway, N.S. Lander 1128 (NSW, PERTH); Lake Barlee, P.G. Wilson 8866 (BRI, BM, CANB, CBG, K, MEL*, NSW, NT, PERTH); Lake Barlee, P.G. Wilson 8867 (BH, BRI, CANB, CBG, K, MEL*, NT, PERTH); S end of Lake Miranda, H.R. Toelken 6093 (AD, BH, BM, CANB, PERTH).

Distribution. Found in the Austin and Coolgardie Districts of the Eremaean Botanical Province of Western Australia. See Map 8.



Map 8. Distribution of Lawrencia chrysoderma (O) and L. helmsii (+).

Etymology. The specific epithet commemorates Robert Helms, naturalist on the Elder Exploring Expedition, 1891-1892, in the north-west of South Australia and across the Victoria Desert of Western Australia (Beard 1970, Eardley 1950).

Notes. The geographical range of Lawrencia helmsii overlaps with those of L. chrysoderma, L. densiflora, L. diffusa, L. glomerata, L. repens and L. squamata. L. helmsii has been found growing in close proximity to these other species.

A note on Plagianthus monoicus Helms ex Ewart

In the course of this study it was necessary to examine the types of *Plagianthus monoicus* Helms ex Ewart, a species overlooked by Melville (1966). The type collection, made at Lake Deborah in Western Australia by R. Helms, comprises specimens of *Ricinocarpos velutinus* F. Muell., a species in the Euphorbiaceae, hence the following synonymy:

Ricinocarpos velutinus F. Muell., Fragm. Phytogr. Austral. 9: 2 (1875). Type: upper reaches of the Irwin River, Western Australia, March 1874, J. Forrest s.n. (holo: MEL, non vidi).

Plagianthus monoicus Helms ex Ewart, in Ewart & Tovey, Proc. Roy. Soc. Victoria 32: 203 (1920), as "monoica"; C. A. Gardner, Enum. Pl. Austral. Occid 78 (1931), as "monoica"; Blackall & Grieve, How to Know Western Austral. Wildf. 2: 346 (1956), as "monoica". Type Lake Deborah, November 1891, R. Helms s.n. (lecto AD 96628004; isolecto; AD 97714338, MEL). The above lectotypification is necessitated by the destruction of the holotype from MEL (see note p. 203).

Discussion

A. Morphology

Habit. Melville (1966) considered species of the genus Lawrencia to be "perennial herbs often woody at the base" and species of the genus Selenothamnus to be shrubs. In the course of the present study the full spectrum of growth forms between these two extremes was observed. Whilst differences in habit and duration are certainly invaluable in the delimitation of species, in conjunction with vestiture and fruit characters they also contribute to the syndromes permitting the recognition of series in the section Selenothamnus. Bates (1968) suggests that in the tribe Malveae the shrubby habit is primitive, a trait shared by the family as a whole. All other members of the Plagianthus alliance are shrubs or trees

Thorns. The thorns of Lawrencia squamata are unique in the genus and would seem to be of rare occurrence in the tribe Malveae. Whilst otherwise unremarkable in themselves, it is worth noting that these thorns are not unlike those of Cratystylis subspinescens (F. Muell. & Tate) S. Moore, Chenopodium nitrariaceum (F. Muell.) F. Muell. ex Benth., Lycium australe F. Muell., Roycea spinescens C. A. Gardner and Scaevola spinescens R. Br., species which also inhabit the margins of salt lakes and ephemeral watercourses and which are often found growing alongside Lawrencia squamata. Presumably, such thorns provide protection against browsing by animals visiting these habitats.

Vestiture. The Malvaceae show a rich assortment of hair types: glandular hairs, simple and bifurcate hairs, and, in particular, stellate hairs which have developed into peltate and palmate scales.

Despite their great morphological diversity, these various forms of hair integrade imperceptibly into one another. In *Lawrencia*, this intergradation can often be observed on the same plant, a process seen most clearly in the vestiture of the stipules.

The primitive hair type in Lawrencia is taken to be the haplomorphic, multicellular stellate hair common to all but the glabrous species in the genus. Modification of this basic hair type has apparently proceeded in several directions (Figure 1). From the basic haplomorphic stellate hair (Figure 1A or D) can be derived the bifurcate hair (Figure 1B), simply by reduction in the number of rays; further reduction leads to a simple hair (Figure 1C). Planation and fusion of the rays of the basic hair type result in actinomorphic stellate hairs (Figure 1E) and, with complete fusion, peltate scales (1F & G). Subsequent reduction of the fused rays of peltate scales, coupled with a change in orientation of the hair, produces the unique bract-like palmate scales (Figure 1H & I) observed in the series Selenothamnus. Continuation of this process produces a simple hair (Figure 1J). Neither peltate nor palmate scales occur elsewhere in the Plagianthus alliance.

In Lawrencia, the protective role usually attributed to plant hairs is probably three-fold. Firstly, hairs effect water economy by restricting movement of air near stomata and thus reducing transpiration. Secondly, hairs may prevent tissue damage from high incident light by virtue of their great reflectivity and by providing an insulating layer. Thirdly, they may help to avoid damage from salt-spray by preventing droplets of sea-water or salty lakewater from reaching live tissues. Hairs on the flower buds and soft vegetative parts of Lawrencia may also discourage insect predation.

The intermeshing basal hairs on the petals of male flowers of *Lawrencia berthae* associated with the sepaline nectaries of that species are discussed below.

The bristle-like hairs on the persistent mericarps of Lawrencia berthae probably assist their dispersal (see below).

Leaves. Variation observed in the leaf-characters of Lawrencia falls well within the limits recorded for other members of the tribe Malveae (Hutchinson 1967, Bates 1968a).

Although stipules are usually present in the Malvaceae (Cronquist 1981) and in the tribe Malveae, throughout the *Plagianthus* alliance there is a trend towards stipule reduction and loss (Bates 1968): *Gynatrix* Alef., like *Lawrencia*, has conspicuous and persistent stipules; *Plagianthus* sensu stricto and *Hoheria* have stipules which are small and caducous; *Asterotrichion* is exstipulate (Melville 1968). The considerable variation observed in texture, vestiture and size of the stipules of *Lawrencia* is striking.

Melville (1966) and Bates (1968) have noted the trend towards palmati-pinnate venation in the genera of the *Plagianthus* alliance. In the present study venation in *Lawrencia* was observed to be actinodromous throughout, but with a marked tendency towards reduction of the lateral primary veins with only the mid-vein really distinct or with the venation quite obscure. Leaf blades in the intermediate category certainly have the superficial appearance of being pinnate: on closer inspection, faint lateral primary veins radiating from a central point towards the blade base are invariably revealed. Leaf venation is variable even within species and no overall pattern in *Lawrencia* was discerned. In other genera of the *Plagianthus* alliance reduction of the lateral primary veins has proceeded further than in *Lawrencia* and is coupled with the development of stronger secondary veins from the mid-rib giving a distinctly palmati-pinnate venation. This syndrome is most advanced in certain species of *Hoheria* which appear to have pinnate venation.

Heteroblastic leaf development such as that observed in *Lawrencia* is widespread in the tribe Malveae. Elsewhere in the *Plagianthus* alliance it has been reported in *Plagianthus* and *Hoheria* (Allan 1961, Melville 1966, Salmon 1980).

Inflorescences. Lawrencia itself does not display the wide range of variation in inflorescence characteristics found in other genera of the tribe Malveae. However, inflorescence structure within the *Plagianthus* alliance as a whole is highly variable and includes solitary axillary flowers, and axillary racemes, panicles, cymes and fascicles. Despite the complexity of their mature inflorescences both *Hoheria* and *Plagianthus* produce solitary axillary flowers early in their growing seasons.

The seemingly anomalous axillary fascicles of *Lawrencia berthae* probably represent a transitional stage in a reduction series leading from an ancestral cymose condition to the single axillary flowers which serve as basic inflorescence units in all the remaining species of *Lawrencia*.

Epicalyx. In Lawrencia, as in all other genera of the Plagianthus alliance, an epicalyx is entirely lacking. The presence or absence of this structure has been used to delimit subtribes and to circumscribe the alliances recognized by Bates (1968) in the tribe Malveae. Bates considers the absence of an epicalyx primitive in the tribe and notes that this condition is typical of members of the Abutilon alliance from which he postulates the Plagianthus alliance was derived.

Calyx. In its basic calyx morphology Lawrencia conforms to the pattern observed elsewhere in the Plagianthus alliance. Amongst the more generally useful characters in Lawrencia are the total calyx length and its ratio with lobe length. Variation in these and other features of the calyx manifests itself only in subtle overall differences which, when considered in conjunction with variation in the characters of other organs, assume importance in the characterization of each species.

Contrary to the general tendency observed by Bates (1969) in the tribe Malveae, in the Plagianthus alliance characters of the calvx are clearly of generic significance. Hitherto, they have not been given the attention they deserve. What is important here is that the calyx of Lawrencia is decidedly turbinate or obconic and never tubular, campanulate or cupular as reported in other genera of the Plagianthus alliance, it lacks the accessory teeth reported in one species of Hoheria, and its lobes remain erect at maturity enclosing the ripened fruit rather than becoming widely spreading or strongly reflexed as in species of Plagianthus and Hoheria (Allan 1961, Melville 1966, Curtis 1975, Moore & Irwin 1978). Further, tufts of multicellular, nectariferous papillae similar in appearance to those found on the adaxial surface at the base of the sepals of L. berthae are found elsewhere in the Plagianthus alliance only in Gynatrix, where they can be observed in both male and female flowers, and in Asterotrichion (Melville 1966). Although nectar-secreting hairs are found in this position in many genera of the tribe Malveae and indeed throughout the family, their presence in both L. berthae and Gynatrix is reported for the first time here. Intermeshing basal hairs on the corolla lobes of male flowers of L. berthae appear to be associated with the nectary of this species (see below).

Other genera in the *Plagianthus* alliance for which a perfume similar to that of *Lawrencia* berthae has been noted are *Hoheria* and *Plagianthus*.

Corolla. Despite its relative constancy in the Malvaceae, a few more striking variations of the corolla have been employed in generic definitions.

In Lawrencia the petals seem to correspond in form to that typical of the family, that is broadest above the middle and hence basically obovate. However, in many species, even in those with petals almost free to the base (usually male flowers of dioecious species) these are often merely ovate or elliptic. This condition seems to be widespread in the *Plagianthus* alliance.

The lack of auricles at the base of the petals in *Lawrencia* is a feature common to all members of the *Plagianthus* alliance, even in those genera with distinctly clawed petals, namely *Hoheria* and *Plagianthus*.

A greater range of colours is found in the corollas of Lawrencia than in other genera of the Plagianthus alliance. In particular, the red- or purple-tinted corollas found in several species of Lawrencia have not been reported in other genera of the alliance. Only Plagianthus shows much variation in the colour of its corolla which may be white, cream-coloured, yellow or green. The corollas of Asterotrichion, Gynatrix and Hoheria are white or cream-coloured throughout.

Several species of Lawrencia have simple-hairy or stellate-hairy corolla lobes. Although this has not previously been reported from any other genus in the Plagianthus alliance, the petals of Hoheria angustifolia Raoul are densely stellate-hairy, like those of certain specimens of L. densiflora. Many genera of the tribe Malveae have hairy petals, particularly in the region of the basal claw and auricles where they intermesh serving to protect the delicate nectariferous sepaline papillae below (Bates 1969, Hill 1982a). Protective hairs similar to these are found on the petals of male flowers of L. berthae, the only species in the genus possessing sepaline nectaries (see above). Such hairs are entirely lacking in Gynatrix, but in female flowers their function seems to have been assumed by the carpocrater which, although not previously reported, is fleshy and extended, forming a nectariferous collar like that described for Asterotrichion by Melville (1966). Curiously, although Hoheria apparently lacks nectaries of any kind, some species of this genus have petals which are simple-hairy in the region of the basal claw, also reported for the first time here.

Apical insertion of the corolla lobes on the ovary cup appears to be characteristic of all genera in the *Plagianthus* alliance. Also constant in the *Plagianthus* alliance is the tendency for female flowers of dioecious species to be carinate and erect, forming a campanulate rather than rotate corolla. This tendency has been reported in *Asterotrichion, Plagianthus* and *Gynatrix* (Curtis 1975, Moore & Irwin 1978, Salmon 1981).

As in Lawrencia, the corollas of Hoheria and Plagianthus are released from the maturing fruit by dehiscence of the basal sutures of the ovary cup. The remaining two genera of the Plagianthus alliance, Asterotrichion and Gynatrix, are unusual in that their corollas are regularly circumcissile (Melville 1966, Curtis 1975).

Androecium. Contrary to Heel's observation that in *Plagianthus* sensu lato only half-stamens leave the filament tube, forked stamens and stamens with closely paired (but never actually bilocular) anthers were commonly found in several species of *Lawrencia* in the present study.

Although the five-ranked symmetry of the filaments often evident in the apical lobing of the filament tube of *Lawrencia glomerata*, the frequently simple-hairy filament tube and occasionally pink anthers of *L. berthae* are exceptional in *Lawrencia*, similar variation in one or more of these same characters has been encountered in other genera of the *Plagianthus* alliance, namely *Hoheria* and *Plagianthus*.

Another number varies more widely in Lawrencia than in any other genus of the Plagianthus alliance. Obviously there has been a marked trend towards reduction of this figure in Lawrencia. The same trend may be observed in the Plagianthus alliance as a whole: only Lawrencia and Plagianthus have species with as few as 5 anthers in each flower, the other genera have a minimum of 10 and, like Lawrencia, a maximum of 30.

Styles. Style differences have been given much emphasis in the taxonomy of the tribe Malveae at sub-tribal and generic levels. Recently, Bates (1968, 1969) has questioned the extent of this emphasis, nonetheless agreeing that the greatest taxonomic value of style characteristics lies in the delimitation and grouping of genera. In particular, Bates (1969) has identified three basic stigma types. The first of these is elongate and introrsely decurrent on a filiform style branch. The second type is that capping, and decurrent on either edge of, a thickened, laterally compressed style branch. The third type includes a variety of essentially capitate forms. In general, the stigmas of *Lawrencia* conform to the first of these types, but the style branches of several species are decidedly thickened, broadened and, in one species, often apically lobed. As in other genera of the tribe Malveae, the exent of the introrsely decurrent stigmatic area on the branches varies: it may be complete or it may cover only the upper portions.

Bates (1968) has suggested that the styles of the early Malveae had large stigmatic areas decurrent on thickened style branches. In *Lawrencia* there seem to have been independant trends leading to loss of external vestiture, reduction in thickness of the style branches, increasing fusion of the style branches, and reduction in the extent of the stigmatic area. These same trends may be observed in other genera of the *Plagianthus* alliance. *Asterotrichion* has externally pubescent styles with longitudinally decurrent stigmatic areas on free, clavate branches; *Gynatrix* has externally glabrous or stellate hairy styles with longitudinally decurrent stigmatic areas on the upper portions of shortly united filiform, linear or sub-clavate branches; *Plagianthus* has externally glabrous styles with longitudinally decurrent stigmatic areas on almost completely united clavate branches; *Hoheria* has externally glabrous styles with slightly decurrent, obliquely capitate or capitate stigmatic areas on free or shortly united, clavate branches. (Allan 1961, Melville 1966, Curtis 1975).

The apical lobing observed on the style branches of Lawrencia chrysoderma has been recorded for no other taxon of the Plagianthus alliance.

Fruit. The rather variable fruit morphology of Lawrencia clearly involves modification of the basic schizocarp typical of the tribe Malveae. This is manifested in several trends: reduction in mericarp number and fertility and supression of the development of sterile mericarps, reduction in pericarp thickness, loss of vestiture, supression of dehiscence along the proximal suture and abaxial mid-vein, development of pericarp ornamentation, development of a zone of weakness in the basal region allowing dehiscence by gross rupture. Similar variability in fruit morphology and dehiscence is found in other genera in the tribe Malveae and cannot be used as a basis for generic recognition.

The laterally narrow-ridged or winged mericarps observed in several species of *Lawrencia* are quite unlike the abaxially broad-winged mericarps observed in *Hoheria* (see Figure 18B in Schumann 1890). Other genera of the *Plagianthus* alliance are not ridged or winged.

Elsewhere in the *Plagianthus* alliance the external vestiture of stellate hairs observed on the fruit of *Lawrencia berthae* is also found to a greater or lesser extent in *Asterotrichion* (Melville 1966, Curtis 1975), in *Hoheria* (reported for the first time here) and in *Plagianthus* (Allan 1961); the fruit of *Gynatrix* has scattered simple hairs (Burbidge & Gray 1970).

Like Lawrencia, the mericarps of other genera in the *Plagianthus* alliance are usually indehiscent or irregularly dehiscent at maturity. The sole exception to this is *Gynatrix* in which the mericarps split open regularly along abaxial and adaxial sutures (Melville 1966, Curtis 1975).

The trend towards reduction in carpel number is found in all genera of the *Plagianthus* alliance (Allan 1961, Melville 1968, Bates 1968), the trend towards reduction in seed fertility

is recorded only in Asterotrichion (Melville 1966, Curtis 1975) and Lawrencia but may be more widespread in the alliance (see Bates 1968).

Of the species considered in this paper known to him Melville (1966) relegated those he believed to have reticulate mericarps to the genus *Lawrencia* and those with unornamented mericarp walls to the genus *Selenothamnus*. Ornamentation of the pericarp probably had its origin in the venation still evident in certain pluriovulate members of the tribe Malveae. Such reticulation of the mericarp walls occurs in many seemingly unrelated or only distantly related taxa in the tribe Malveae and is generally thought to have evolved independently several times. It has not been recorded for other genera in the *Plagianthus* alliance.

On the basis of fruit morphology alone, Lawrencia berthae is clearly peripheral to other species in the genus by virtue of its 9-11 indehiscent mericarps which have a stellate-hairy, coriaceous pericarp with little or no reticulation. Further support for division of the genus Lawrencia is provided by the axillary fascicles of 1-5 conspicuously pedicellate unisexual flowers, the sepaline nectaries, and the stellate-hairy style bases found only in L. berthae. In the present treatment L. berthae is placed in the subgenus Panifex; the remaining species form the subgenus Lawrencia.

In the subgenus Lawrencia reticulation of the pericarp is always associated with indehiscent mericarps all of which are fertile. On the other hand, pericarps without reticulation are associated with dehiscent mericarps in which there is a strong tendency towards reduction in number and fertility. These two syndromes provide the basis for the segregation of the sections Lawrencia and Selenothamnus, respectively, in the present treatment. It should be noted that the section Selenothamnus recognized here is considerably broader in concept than Melville's genus Selenothamnus.

Within the section Selenothamnus two further syndromes involving fruit characters are apparent. Fruit in which all mericarps are well developed are associated with plants of suffrutescent habit which have a vestiture of stellate hairs and which are hermaphroditic or dioecious with female flowers completely lacking anthers. Fruit in which only fertile mericarps develop are associated with plants of a distinctly shrubby habit which have a vestiture of peltate trichomes or stellate hairs and which are dioecious with female flowers often possessing sterile anthers. These two syndromes have prompted the recognition of the series Halophyton and Selenothamnus, respectively.

Seed. Bates (1969) argues that in the tribe Malveae the uniovulate state is derived from the pluriovulate condition. All species of Lawrencia have uniovulate carpels with pendulous ovules as do all taxa in the Plagianthus alliance with the possible exception of P. divaricatus which rarely has 2 pendulous ovules in each locule, according to Melville (1966). The complete lack of any trace of an endoglossum in Lawrencia is a major barrier to speculation on whether its uniovulate condition has been arrived at by loss of the upper or lower ovules in the carpels of the presumed pluriovulate ancestor. The fact that the solitary ovule is pendulous may indicate the latter process, but this condition might conceivably have arisen by reorientation of the basal ascendent ovule following loss of the ovules above it.

Sculpturing such as that found on seeds of species in the series *Halophyton* has not been recorded for other genera in the *Plagianthus* alliance.

B. Sexuality

Melville (1966) noted in the *Plagianthus* alliance "a strong tendency towards dioecy which has nowhere reached the final stage of complete supression of one sex." In the present study of *Lawrencia*, hermaphroditism, polygamodioecy and complete dioecy were all

encountered, revealing a trend leading from hermaphroditism to complete dioecy. This trend is expressed morphologically by partial or total supression of the gynoecium in flowers of male plants complemented by sterility and partial or complete supression of the anthers in flowers of female plants.

Dioecy is infrequent in the Malvaceae (Cronquist 1981). Elsewhere in the tribe Malveae similar floral sexual dimorphy to that of *Lawrencia* is found in the *Plagianthus* alliance in *Asterotrichion, Gynatrix* and *Plagianthus* itself (Allan 1961, Melville 1966, Curtis 1975, Moore & Irwin 1978); it is also found in *Napaea* L. and *Kydia* Roxb., the latter a genus of uncertain affinities (Bates 1968, Fryxell 1979). The polygamodioecy observed in *L. glomerata* has also been recorded in both species of *Plagianthus* which, in addition, may sometimes be monoecious (Allan 1961, Melville 1966).

Evolution of dioecy directly from hermaphroditism or via gynodioecy, monoecy or heterostyly has been documented by Bawa (1980) who also allows of the theoretical possibility that dioecy could also be established via androdioecy. In *Lawrencia*, flowers of male plants often possess a vestigial gynoecium and flowers of female plants often possess sterile anthers, but neither gynodioecy nor androdioecy were observed.

It has long been assumed that true dioecy ensures the maintenance of considerable heterozygosity in the population, which is generally considered to be an evolutionary advantage. Although bisexual flowers in all hermaphroditic species of *Lawrencia* are protandrous, as the flowers wilt their style branches reflex into the anther cluster thus apparently increasing the likelihood of self-pollination at this late stage. It is not yet known to what extent self-compatibility and hermaphroditism are linked in *Lawrencia*. In recent years several alternative models have been propounded to explain the evolution of dioecy without invoking the genetic benefits of outcrossing as the selective force of most importance (Bawa 1980, Bawa & Beach 1981).

The sexual systems of plants are fundamentally linked to their pollination biology. Unlike those of most Malvaceae, the flowers of all but one species of Lawrencia produce no nectar, so pollen would seem to be the only reward offered visitors. As pollen is vital to the economy of many pollinating insects it may be advantageous for certain plants to produce pollen in excess of that needed for fertilization in order to enhance the activity and maintain the numbers of pollinators over a long time. Supplying the pollen in bisexual flowers may prejudice maternal success, in the sense that pollen and ovules utilize the same energy and nutritional resources and hence limit each other's production, or it may be too short-lived or result in overloading the plant with more fruit than it could normally mature. Such a need for the economical deployment of energy and nutritional resources available for developing embryos, seeds and fruits is indicated by the uniovulate mericarps found in all species of Lawrencia and by the strong tendency towards reduction in mericarp number and fertility characteristic of the subgenus Selenothamnus.

Since sexual selection must operate via pollinating agents, it is significant that small generalist bees, flies and other insects that are the predominant pollinators of dioecious species respond dramatically to changes in floral resources, particularly changes in flower number (Bawa 1980). An increase in flower number on hermaphrodite plants may lead to a disproportionate increase in male fitness because plants with larger floral displays may either attract more pollinators and thereby disperse more pollen, or be visited earlier in the day and thereby transmit more genes via pollen than via ovules. As a consequence, males would be established in an initially hermaphroditic population. Similarly, females may become established when individuals increase their fitness due to resources saved from reduced pollen production and dispersal (Bawa & Beach 1981).

In these complementary processes representing a response to pollinator-mediated selection of the kind hypothesized above lies a possible explanation for the evolution of dioecy in *Lawrencia* which may repay further investigation.

C. Dispersal

The single species in the subgenus *Panifex, Lawrencia berthae*, probably exploits hydrochory or epizoochory. Its persistent mericarps are densely stellate-hairy on their apical and distal surfaces. Such hairs may aid flotation by means of air-bubbles adhering to the seed coat in the manner described for species of *Salicornia* L. by Waisel (1972), or they may serve to affix the disseminule to an animal's coat.

The mericarps of all species in the subgenus Lawrencia section Lawrencia have persistent indehiscent pericarps which become light, dry and stiff at maturity. Their apical- and lateral-distal margins are often projected to form narrow wings, and there is a substantial air-space between the single seed contained in each mericarp and the pericarp. These bladder-like fruit would seem to be well adapted for both hydrochory and for amenochory (see Pijl 1982). The areolae which develop in the reticulate walls of species in this section may further aid hydrochory by trapping air-bubbles.

Species in section Selenothamnus appear to have no special aids to dispersal. On the contrary, the early disintegration of the pericarp and the release of naked seeds would seem to favour rapid germination at the parent site. Presumably, further dispersal occurs when seed is released by wind-action on the plant or tumbling of broken branches. The seeds of these species are small and triquetrous, or swollen and irregularly globular, and may be smooth, angular or rugose. The significance of the shallow pits or depressions observed in several species in this section is obscure. Ridley (1930) suggests that such ridges and other emanations found on the seeds of aquatic species may act as anchoring devices serving to fix them to their substrate, thus preventing them from being carried away by water currents. If such seeds do indeed readily adhere to mud then this may also aid their transport on the feet of waterfowl and grazing mammals (see Stebbins 1974). Conversely, such sculpturing may assist the flotation of seeds by means of trapped air bubbles thus favouring hydrochory rather than epizoochory (see Pijl 1982).

Endozoochory is unlikely in *Lawrencia*. The seeds are not particularly hard and would easily be destroyed by a mammal's teeth or a bird's gizzard. The persistent pericarp of species in the section *Lawrencia* would easily be broken in like fashion.

D. Phytogeography

In general, those species of Lawrencia with mericarps favouring epizoochory, amenochory or hydrochory are of wide distribution; those lacking specialized aids to dispersal, are of considerably less widespread occurrence. There are two notable exceptions to this. Despite its bladder-like fruit, L. buchananensis is restricted to a land-locked and geographically isolated salt-lake system in north eastern Queensland. Its taxonomic affinities would seem closest to L. viridi-grisea, a species extending from the western coast of Western Australia across the Great Sandy Desert into the Northern Territory. These two may well represent a vicarious species pair which have diverged morphlogically through long separation by unfavourable edaphic and climatic conditions in the intervening region. L. squamata, which releases large naked, smooth seeds, is widely distributed across southern mainland Australia.

Green (1964) mentions that several hundred species of flowering plants have marked disjunctions between south-western and south-eastern Australia; Beard (1969) records 280 such species. Lawrencia berthae is clearly a good example of a disjunctly distributed species.

So too is *L. spicata*, apart from a single immature specimen from Loongana, possibly of sporadic occurrence from seed dropped by trains. Green argues that such plant disjunctions are unlikely to be the result of a single separation of eastern and western Australia by, say, the Miocene innundation of the Nullarbor Plain or, alternatively, by late Pleistocene changes. He points out that such disjunctions may also be the result of long-distance dispersal, a view supported by Fryxell (1967). The lack of evolutionary divergence shown by each of these two bicentric species indicates a very recent achievement of their disjunctions; the impressive distance barrier separating their populations points to long-distance dispersal rather than migration. As discussed above, both species have mericarps apparently well adapted for dispersal. Since all *Lawrencia* species frequent saline conditions their seeds may well tolerate immersion in sea water. Thus the absence of *L. berthae* and *L. spicata* from the Nullarbor Plain and the coast of the Great Australian Bight may merely reflect the lack of suitable habitats in this region. It is, of course, conceivable that these disjunctions are ancient and the species highly stable, but in the absence of evidence supporting this view, the alternative is more likely.

Although there is danger in using present-day distributions to identify centres of origin, it does seem likely that the various infrageneric taxa of Lawrencia were segregated in and have undergone their principal evolution within Western Australia where the genus has its major centre of diversity today. In particular, there has been an obvious proliferation of taxa of the section Selenothamnus in the interior where it appears that the lake systems of Western Australia have furnished opportunities for speciation in Lawrencia. In this region it is easy to envisage the isolation of populations not only between divisions but between lake systems as well (Short 1981). Even today lakes within a system may be isolated for extended periods with water linking them only in exceptionally wet years (Beard 1973, Bettenay 1962, Chapman 1962). The very close morphological similarities between the various species of Lawrencia, together with their present distribution patterns, suggest that some have evolved in quite recent times, perhaps in response to the rapid and profound hydrologic oscillations evident over the last 400,000 years (Bowler 1981, 1982), causing isolation of individual lakes and therefore of populations within any one system.

There seems now to be general agreement that the arid zone has been colonized successfully by numerous biotypes selected from adjacent populations growing under favourable conditions rather than producing its own characteristic flora by means of internal evolutionary radiation (Barlow 1981, Beadle 1981, White, 1982). Burbidge (1960) suggested extensive colonization of the arid zone of Australia from populations which first appeared and diversified in strand habitats, a view which, until recently, has met with widespread acceptance (Beard 1976). In his review of studies on the phytogeography of arid Australia, Carolin (1982) finds scant evidence in support of this putative littoral connection: he cites the Salicornieae (Chenopodiaceae) and Atriplex (Chenopodiaceae) as possible illustrations and suggests that a consideration of taxa amongst the Caryophyllaceae may also vindicate such a link. Lawrencia, too, appears to be relevant in this context.

Lawrencia glomerata, a species with predominantly primitive characters within the genus, is widely distributed throughout the arid zone. Its considerable heterogeneity would seem to indicate active radiation and segregation in the arid environment. To a lesser extent the same is true of L. viridi-grisea. On the other hand, L. squamata, a taxon with many advanced characteristics, is also highly variable and widely dispersed. These examples may reflect radiation patterns of other taxa in the past.

The hypothesis that Lawrencia first appeared and diversified in strand habitats from whence taxa dispersed to colonize the margins of inland salt-lakes the subsequent expansion and contraction of which has provided a mechanism whereby secondary diversification and

colonization of the arid zone has occurred deserves further exploration. It seems likely that cytological evidence and cladistic analysis might prove most useful in such a study.

E. Specimens from Giles' Fifth Expedition

Willis (1981) notes that specimens collected by W. Ernest P. Giles during his Fifth Expedition of 1876 are presumed to have been lost. Thus the discovery of Giles' specimens of Lawrencia cinerea presumed to have been gathered in the Little Sandy Desert and of L. glomerata from the upper Ashburton River and Mt Murchison is significant: ironically, these specimens were amongst those accidentally destroyed (see note p. 203). It is possible that further botanical collections from Giles' last long traverse are to be found at The National Herbarium of Victoria (MEL).

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The abbreviations used below for titles of journals are those given by Lawrence et al. (1976) or are formed according to the principles codified therein.

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Exsiccatae

The following list is intended to serve as a record of all specimens of Lawrencia studied and annotated by the author together with their contributing herbaria and to facilitate the identification of duplicates of these not examined in the course of this revision. Collectors are listed alphabetically and the initials of all their given names are cited wherever possible. Numbers in parentheses refer to the corresponding numbered species in the text. Abbreviations for herbaria are those given in Holmgren et al. (1981). In the case of Kings Park and Botanic Garden, Perth, there is no abbreviation so KP is used informally here. In the absence of a collector's number the date of collection is cited where available; herbarium register or sheet numbers are cited only in otherwise ambiguous cases. All specimens from HO and MEL examined were destroyed in a road accident during their return to those herbaria. Specimens from MEL marked with an asterisk represent recently distributed duplicates which are still extant. A comprehensive account of the lost specimens may be found in Lander (in press).

Alchin, B.M. 13 (10-NSW). Alcock, C.R. 663 (3-AD), 722 (10-AD), 912 (3-AD), 1093 (1-AD), 1096 (2-AD), 1113 (1-AD), 1114 (3-AD), 1255 (2-AD), 2143 (3-AD), 2763 (3-AD), 2764 (10-AD), 4184 (10-AD), 4743 (10-AD), 4758 (2-AD). Alin, W. s.n., s.dat. (2-MEL). Allan, K.M. 89 (12-PERTH), 695 (10-PERTH). Allender, B.M. s.n., May 1966 (12-UWA). Amtsberg, H. s.n., Nov. 1073 (2-AD). Andrews, C. 72 (2-BM, K, PERTH), s.n., Sept. 1904 (10-PERTH), s.n., June 1927 (12-PERTH). Anonymous Oct. 1877 (4-MEL), Oct. 1877 (7-MEL 98655, 98657), —1897 (12-K), Nov. 1905 (10-MEL), May 1917 (2-AD), June 1917 (2-AD), Oct. 1918 (10-AD), Nov. 1936 (3-AD), Oct. 1949 (10-AD), Oct. 1950 (10-AD), May 1956 (10-AD), — 1956 (1-NSW), Oct. 1966 (3-AD), Nov. 1966 (10-AD), May 1967 (3-AD), s.dat. (2-HO), s.dat. (2-MEL 98747), s.dat. (2-MEL 98756), s.dat. (2-NSW), s.dat. (2-1UWA), s.dat. (3-AD), s.dat. (2-MEL), s.dat. (10-AD), s.dat. (2-NSW), s.dat. (2-UWA), s.dat. (3-AD), s.dat. (3-MEL), s.dat. (9-MEL 98726), s.dat. (10-AD), s.dat. (10-K), s.dat. (10-MEL 98794), s.dat. (10-MEL 98805), s.dat. (10-MEL 98807), s.dat. (10-MEL 98807) MEL 98809). Aplin, T.E.H. 1489 (10-PERTH), 2145 (10-PERTH), 2308 (10-PERTH), 2419 (7-PERTH), 2531 (11-PERTH), 2369 (12-PERTH). Archer, W.H. s.n., s.dat. (2-NSW). Armitsberg, H. s.n., Feb. 1973 (AD). Armstrong, (11-PERTH), 2369 (12-PERTH). Archer, W.H. S.N., S.dat. (2-INSW). Armisoerg, H. S.N., Feb. 1973 (AD). Armistong, S. S.N., July 1966 (12-PERTH). Ashby, A.M. 1204 (1-AD), 1940 (1-AD, PERTH), 2214 (10-AD, PERTH), 2969 (7-AD, PERTH), 3408 (3-AD), 3705 (12-AD, MEL). Atkin, Rev. S.N., Dec. 1896 (12-NSW). Backhouse, J. S.N., S.dat. (2-K). Bailey, E.T. 1-22 (12-PERTH). Banssley, D. 971 (10-CBG, NT). Barrels, L.C. S.N., Feb. 1946 (2-NSW, PERTH). Bates, R. 223 (3-AD). Batt, J.D. 64 (3-MEL), 98 (3-MEL), s.N., 1886 (3-MEL), Jan. 1887 (10-MEL), s.N., 1889 (10-MEL), s.N., 1890 (10-MEL. Bauerlen, W. 254 (10-MEL, NSW), 256 (3-MEL), s.N., Oct. 1887 (10-BRI). Beard, J.S. 6079 (3-NSW, PERTH), 6534 (10-PERTH), 656 (3-PERTH), 6612 (11-PERTH). Beauglehole, A. C. 989 (10-MEL), 2808 (3-NT), 29241 (3-AD), 10048 (3-NT), 12991 (3-NT), 13207 (10-NT, PERTH), 13244 (8-NT), 13361 (8-NT), 16088 (3-MEL), 19403 (2-AD), 21349 (2-MEL), 24451 (3-NT), 29357 (3-MEL), 29614 (12-NT), 29528 (7-NT), 31285 (2-MEL), 32165 (2-MEL), 33027 (2-MEL), 39124 (2-MEL), 39209 (3-MEL), 39210 (10-MEL), 39459 (2-MEL), 40390 (10-MEL), 40453 (10-MEL), 40583 (10-MEL), 40592 (3-MEL), 42985 (3-MEL), 42987 (3-MEL), 42993 (10-MEL), 45704 (4-NT), 46175 (3-NT), 48276 (3-NT), 49093 (12-NT, PERTH), (3-MEL), 42987 (3-MEL), 42993 (10-MEL), 45704 (4-NT), 46175 (3-NT), 48276 (3-NT), 49093 (12-NT, PERTH), 49335 (8-NT), 49434 (3-NT), 49486 (10-AD, NT) 50759 (3-NT), 50932 (3-NT), 55588 (10-MEL), 55731 (3-MEL), 55867 (3-MEL), 55955 (10-MEL), 59528 (7-NT). Bechervaise, J.M. s.n., Sept. 1947 (3-MEL; Bennett, E.M. 45 (2-PERTH). Berston, J. 7 (7-PERTH). Bettenay, E. 172 (10-PERTH). Birch, C.W. s.n., 1871 (3-MEL), s.n., s.dat. (3-MEL). Black, E.C. Oct. 1917 (3-AD), s.n., Oct. 1947 (3-AD), s.n., Oct. 1948 (3-AD), s.n., Oct. 1950 (3-AD). Black, R.A. s.n., April 1922 (2-MEL), s.n., Nov. 1936 (3-AD), Blackall, W.E. 390 (3-PERTH), 391 (10-PERTH), 534 (7-PERTH), 1009 (3-PERTH), 1228a (12-PERTH), 1232 (10-PERTH), 3495 (10-PERTH), 4375 (10-PERTH), s.n., Oct. 1931 (3-PERTH), s.n., Sept. 1939 (3-PERTH), s.n., Sept. 1940 (7-PERTH), s.n., s.dat. (1-PERTH). Blackwell, M. Y 310 (7-PERTH). Blaylock, B.J. 639 (10-AD), 1126 (10-AD), 1698 (10-AD). Blockley, J.V. 760 (7-KP). Boswell, P. 24 (8-PERTH). Bowen, H. 198 (3-K), 271 (3-K), 311 (10-K). Bridgewater, P. s.n., Oct. 1979 (PERTH). Brockway, G.E. s.n., Oct. 1947 (12-CANB, PERTH). Brooker, M.G. 3647 (3-PERTH). Brooks, S.T.C. (as Miss "Brooke", "Brookes") s.n., 1883 (3-MEL), s.n., 1884 (3-MEL), s.n., 1885 (10-MEL), s.n., 1893 (1-MEL). Brown, C.D. s.n., 1890 (7-MEL). Brown, R. 4998 (10-BM, (3-MEL), s.n., 1885 (10-MEL), s.n., 1893 (1-MEL). Brown, C.D. s.n., 1890 (7-MEL). Brown, R. 4998 (10-BM, K), 5103 (3-BM), 5110 (2-BM, CANB, K, MEL). Brown, Mrs T. s.n., Oct. 1907 (10-NSW). Browne, J.H. 125 (10-PERTH), 126 (10-PERTH). Brumby, Mrs s.n., 1937 (3-AD). Brummit, R. s.n., Aug. 1892 (10-AD). Buckley, R. 7135 (4-PERTH), 7136 (4-PERTH), 7137 (4-PERTH), 7147 (4-PERTH). Bufton, J. s.n., 1892 (2-MEL), s.n., 1893 (2-MEL). Burbidge, N.T. 263 (8-K), 1433 (3-PERTH), 2738 (10-CANB, PERTH), 4088 (10-CANB), 4610 (3-CANB), 4713 (3-PERTH), 4755 (12-BRI, CANB), 4756 (12-CANB, PERTH), s.n., Aug. 1938 (7-PERTH). Burgman, M.A. 2704 (7-PERTH). Butler, R. s.n., July 1965 (4-PERTH). Bynoe, B. s.n., s.dat. (2-K). Callen, R. s.n., 1969 (10-AD). Carr, G.W. 2396 (3-NT), 4498 (3-AD, NT), 1925 (4-NT). Carr, S.G.M. 512 (12-PERTH), 1925 (4-NT), 4498 (3-NT). Carrick, J. 1802 (3-AD), 3710 (10-AD). Carrol, E.J. s.n., Sept. 1965 (3-CBG, NSW). Caulfield, H.W. 146a (3-AD, BM, BRI, CANB, K, MEL, NSW, NT, PERTH). Chadwick, - 2317 (3-CBG, Caugheid, H.W. 140a (3-AD), BM, BRI, CANB, K, MEL, NSW, NI, FERTH). Chalwick, — 2517 (3-CBG, NSW). Chapman, C. s.n., Oct. 1967 (11-PERTH), s.n., s.dat. (10-PERTH). Chapman, D. s.n., Oct. 1967 (11-BM, BRI, K, NSW, NT, PERTH). Cheal, P.D.C. s.n., Dec. 1980 (10-MEL). Chimnock, R.J. 87 (10-AD), 772 (10-AD), 773 (10-AD), 336 (3-AD), 666 (3-AD), 941 (7-AD), 1495 (3-AD), 1505 (10-AD), 1506 (10-AD), 2617 (3-AD), 2930 (3-AD), 4006 (12-AD), 4007 (12-AD), 4132 (9-AD), 4157 (9-AD). Chippendale, G. 10611 (3-AD), 1505 (10-AD), 1 BRI, CANB, K, MEL, NSW, PERTH), 10612 (3-AD, CANB, MEL, NSW, NT), 6375 (3-AD), 10612 (3-AD, CANB, MEL, NSW). Chorney 1066 (3-AD, PERTH). Churchill, D. 202 (12-UWA). Clarke, E. de C. s.n., 1916 (3-BM, PERTH). Clarke, W. s.n., s.dat. (2-MEL). Cleland, J.B. s.n., Nov. 1913 (1-AD), s.n., Jan. 1924 (2-AD),

s.n., Nov. 1924 (2-AD), s.n., Jan. 1926 (2-AD), s.n., 5 Nov. 1926 (10-AD 96601963), s.n., 5 Nov. 1926 (10-AD 96805867), s.n., 7 Nov. 1926 (10-AD), s.n., 1 Nov. 1928 (10-AD), s.n., 27 Nov. 1928 (10-AD), s.n., June 1933 (3-NT), s.n., Aug. 1939 (3-AD), s.n., Nov. 1941 (3-AD), s.n., July 1943 (3-AD), s.n., Nov. 1950 (10-AD), s.n., Dec. 1953 (3-AD), s.n., Dec. 1953 (10-AD), s.n., Nov. 1955 (3-AD), s.n., Nov. 1956 (3-AD), s.n., Nov. 1959 (2-AD), s.n., July 1960 (3-PERTH), s.n., Feb. 1964 (3-AD), s.n., Feb. 1968 (10-AD), s.n., Nov. 1968 (3-AD), s.n., s.dat. (3-NT). Clement, E. s.n., s.dat. (4-K). Coleby-Williams, J. 117 (11-PERTH), 118 (11-PERTH). Comber, H.F. 2024 (2-K). Cooper, H.M. s.n., March 1941 (3-AD). Copley, B. 817 (3-AD), 1020 (10-AD), 1136 (2-AD), 1235 (10-AD), 1563 (3-AD), 1571 (3-AD), 1603 (3-AD), 2398 (10-AD), 2719 (10-AD), 3678 (3-AD), 3843 (3-AD), 4541 (10-AD). Cornwall, G.C. 129 (3-AD). Corrick, M.G. 5072 (3-MEL), 6239 (1-AD, MEL), 6376 (1-MEĹ), 6380 (1-MEĹ), 6656 (10-MEĹ). Coveny, R. 8258 (10-NSW, PERTH). 8411 (12-PERTH). Cranfield, R.J. 638 (8-PERTH), 1723 (7-PERTH), 1856 (7-PERTH), 2521 (4-PERTH), 2584 (4-PERTH), 2595 (4-PERTH), 4009 (10-PERTH). Craven, L.A. 5033 (12-BRI, MEL, NT, PERTH). Crawford, — 39 (10-MEL), 48 (10-MEL). Crisp, B.C. 576 (3-CANB). Crisp, M.D. 676 (3-AD, CANB), 1231 (10-MEL), 3295 (10-CBG, MEL), 3417 (3-CBG, MEL), 5469 (11-CANB), 5630 (10-CBG, NT). Crocker, R.L. s.n., July 1939 (3-AD), s.n., Aug. 1939 (3-AD), s.n., Aug. 1939 (10-AD), s.n., 1939 (10-AD), s.n., s.dat. (10-AD). Cronin, M. s.n., 1890 (3-MEL), s.n., 1893 (1-MEL). Cummings, D.J. 232 (10-CBG, NT). Cunningham, G.M. 3026 (3-NSW), 4776 (10-NSW). Curtis, W.M. s.n., Jan. 1944 (2-HO), s.n., Jan. 1946 (2-HO), s.n., Dec. 1946 (2-HO), s.n., April 1953 (2-HO), s.n., Feb. 1966 (2-HO). Czornij (as "Chorney"), K. 1066 (3-AD, BH, K, PERTH). Dalachi, — s.n., s.dat. (2-MEL, NSW, PERTH). Dallachy, J. 237 (3-MEL), 238 (3-MEL), s.n., s.dat. (10-K). D'Alton, St. E. 16 (3-MEL), 26 (3-NSW), s.n., Oct. 1899 (3-MEL). Davies, C.F. 283 (1-PERTH). De Marci, C. s.n., Scpt. 1964 (4-PERTH). Dennis, T. 152 (2-AD). de Rebeira, P. 415 (6-AD, PERTH), 422 (6-AD, PERTH), 433 (12-PERTH). Demarz, H. 4445 (3-KP, PERTH), 4610 (10-KP, PERTH), 4712 (7-AD, KP, PERTH), 4587 (3-KP, PERTH), 4690 (3-KP, PERTH), 4829 (4-PERTH), 4831 (7-KP, PERTH), 4894 (8-KP, PERTH), 5292 (10-KP), 5367 (3-KP, PERTH), 6015 (2-KP), 6310 (10-KP, PERTH), 7140 (7-KP, PERTH), 8241 (7-KP), 8995 (7-KP, PERTH), 9258 (2-PERTH), 9540 (4-PERTH). Deseglise, L. s.n., 1866 (3-BM). Diels, L. 5204 (8-PERTH). Dixon, S. s.n., Oct. 1893 (12-AD). Donner, N.N. 1785 (3-AD), 2457 (10-AD), 2458 (10-AD). Drummond, J. 6 (3-K), 24 (9-BM, K), 55 (3-BM, MEL, PERTH, K), 104 (9-K), 106 (10-BM, K, PERTH), 137 (3-K), 208 (10-BM, K, MEL, PERTH), 252 (10-BM, K, MEL), 246 (3-K), 275 (1-K), 302 (2-BM, K, MEL), s.n., 1848 (10-K), s.n., Jan. 1854 (2-MEL), s.n., s.dat. (1-MEL), s.n., s.dat. (2-K, MEL 98729, 98730 98754), s.n., s.dat. (4-MEL 98692), s.n., s.dat. (10-K), s.n., s.dat. (10-MEL 98771, 98772, 98781, 98785). Dunlop, C. 1810 (3-AD), 2452 (10-K, NSW), 2541 (3-AD, MEL, NT). Eckert, J.P. s.n., 1892 (10-MEL). Eichler, Hi. 12536 (3-AD), 12958 (3-AD), 13001 (10-AD), 1300 14186 (10-AD), 15732 (10-AD), 16859 (2-AD), 17629 (3-AD), 18745 (3-AD), 19417 (10-AD), 19433 (10-AD), 19434 (10-AD), 19445 (3-AD), 19550 (3-AD), 20317 (9-AD, PERTH), 20321 (10-AD, PERTH), 20322 (10-AD), Elkington, J. 330 (12-PERTH). Erroy, E.G. 3228 (7-NT), 3314 (12-NT). Everist, S.L. 9164 (7-BRI). Fagg, M. 376 (3-AD), Fairall, A.R. 1825 (12-KP). Fell, L. 678 (10-KP). Fillagan, I. 3 (2-MEL). Fitzgerald, W.V. s.n., 1893 (2-HO), s.n., Oct. 1898 (8-NSW, PERTH), s.n., Jan. 1903 (10-PERTH), s.n., Aug. 1903 (10-PERTH), s.n., A W.V. S.n., 1893 (2-HO), S.n., Oct. 1898 (8-NSW, PERTH), S.n., Jan. 1903 (10-PERTH), S.n., Aug. 1903 (10-NSW, PERTH), S.n., Sept. 1903 (7-PERTH), S.n., Nov. 1907 (10-NSW). Flounders, B. 8 (3-AD). Forde, N. 479 (3-AD, K, MEL), 1330 (10-CANB), 1355 (12-CANB). Forrest, A. S.n., 1879 (3 & 4-MEL). Forrest, J. S.n., June 1874 (3-MEL), s.n., 28 June 1874 (3-MEL), S.n., 30 June 1874 (3-MEL). S.n., 1882 (3-MEL). Fraser, C. 27 (2-BM), 126 (2-BM), 178 (2-K). French, C. S.n., April 1923 (3-MEL). Galbraith, J. 10 (10-AD). Gardner, C.A. 1993 (1-PERTH), 2094 (10-PERTH), 2128 (12-BM, PERTH), 2403 (7-PERTH), 2441 (7-BM, K, PERTH), 2444 (10-PERTH), 2545 (7-PERTH), 3264 (3-BM, PERTH), 3264A (3-K, PERTH), 6075 (3-PERTH), 6089 (7-PERTH), 2562 (3-PERTH), 3264 (3-PERTH), 7532 (10-PERTH), 7800 (3-PERTH), 7814 (12-PERTH), 8618 (10-PERTH), 13352 (7-PERTH), s.n., Aug. 1927 (12-PERTH), s.n., Aug. 1931 (7-PERTH), s.n., Sept. 1935 (1-PERTH). Gasteen, J. 85 (5-BRI). Gauba. E. s.n., Oct. 1942 (3-NSW, CANB). George, A.S. 915 (12-PERTH), 923 (12-PERTH), 952 (7-PERTH), 1482 (7-PERTH), 2290 (10-CANB), 2291 (10-CANB), 2948 (3-PERTH), 4214 (8-PERTH), 5692 (1-AD, BH, K, PERTH), 8111 (3-AĎ, BĤ, PERTH), 9131 (10-AD, PÉRTĤ), 9132 (3-MEL*, NSW, PÉRTĤ), 9137 (6-PÉRTH, AD), 9765 (3-BH, K. PERTH), 10207 (7-PERTH), 10214 (4-BH, K. MEL*, NSW, NT, PERTH), 11380 (NT, PERTH), 14783 (4-PERTH), 14812 (6-BH, NT, PERTH), 14813 (6-PERTH), s.n., Dec. 1964 (10-PERTH), s.n., June 1970 (9-PERTH), s.n., Sept. 1972 (4-PERTH). George, D.N. s.n., July 1931 (3-AD). Giles, W.E.P. s.n., 1872-4 (3-MEL), s.n., 1875 (10-MEL), s.n., 1876 (6-MEL), s.n., 1876 (3-MEL 98695), s.n., 1876 (3-MEL 98731), Gil. W. s.n., Feb. 1903 (2-NSW). Glennon, P. 109 (4-PERTH). Goodall, D.W. 2910 (10-PERTH), 3313 (7-PERTH, UWA). Goss, G.F. s.n., April 1955 (3-AD). Gratte, S. s.n., Aug. 1970 (3-AD), Green, A. s.n., Nov. 1924 (1-PERTH). Gregory, J.H. s.n., 1901 (1-BM). Gregory, Prof. s.n., May 1909 (12-MEL). Grieve, B.J. s.n., Oct. 1960 (10-UWA). Griffin, E.A. 3084 (4-PERTH). Griffith, H.H.D. s.n., Dec. 1907 (2-AD), s.n., Oct. 1927 (10-AD). Gullan, P.K. 384 (2-MEL). Gunn, R. 746 (2-K). Hall, A.J. s.n., Feb. 1928 (2-PERTH). Hannaford, S.A. 9 (10-NSW). Heal, M. s.n., 1889 (1-MEL). Helms, R. s.n., Sept. 1891 (1-MEL), s.n., Nov. 1891 (9-AD, K, MEL), s.n., Nov. 1891 (12-AD, K, MEL, NSW, PERTH), s.n., 1899 (12-K, PERTH). Henderson, W. 2556 (10-NSW), s.n., Oct. 1946 (10-NSW). Henry, N.M. 378 (4-MEL), 383 (3-AD, NT), 953 (4-AD). Henshall, T. s.n., Sept. 1966 (10-NSW). Herbert, D.A. s.n., July 1918 (12-PERTH). Heyligers, P.C. 79140 (10-CANB), 80136 (10-CANB). AD, CANB). Hill, F.L. 119 (4-BM), 124 (4-BM, K), 416 (4-CANB, K), 619 (4-K). Hill, R. 54 (3-AD), 229 (3-AD), 1006 (10-AD), 1173 (3-AD). Holst, N.O. s.n., April 1896 (12-MEL). Horbury, — s.n., Oct. 1937 (3-PERTH). Humphries, A.W. P48 (11-PERTH). Ince, W.H. s.n., June 1903 (12-K). Ising, E.H. s.n., Dec. 1922 (2-AD), s.n., Jan. 1925 (2-AD), s.n., Aug. 1925 (8-AD), s.n., Scpt. 1936 (1-AD), s.n., Jan. 1937 (10-AD), s.n., Jan. 1925 (2-AD), s.n., Jan. 1925 (2-AD), s.n., Jan. 1925 (3-AD), Nov. 1938 (10-AD 96935429 & 97416003), s.n., Sept. 1938 (3-AD), s.n., Oct. 1950 (3-AD), s.n., Sept. 1951 (3-AD), s.n., Aug. 1952 (3-AD), s.n., Aug. 1955 (3-AD), s.n., Sept. 1955 (3-AD), s.n., Oct. 1955 (3-AD), s.n., Oct. 1955 (3-AD), s.n., Dec. 1958 (10-AD), s.n., Sept. 1960 (1-AD). Jackson, E.N.S. 741 (3-AD, PERTH), 1136 (AD), 1835 (10-MEL), 2559 (10-AD). Jacob, G.L. s.n., May 1968 (3-NSW). Jacobs, S.W.L. 1114 (10-NSW). Johns, C.W. s.n., 1941 (10-AD). Johnson, U. 30 (10-NSW). Keighery, G.J. 1956 (10-KP, PERTH), 2966 (8-KP), 3883 (2-KP), 5543 (10-PERTH). Kenneally, K.F. 7351 (7-PERTH). Kenny, R. s.n., Feb. 1946 (2-UWA). Kingsmill, W. 485 (12-BRI, NSW). Kniep, N. s.n., 1937 (12-PERTH). Koch, M. 1167 (10-K, NSW). Kraehenbuehl, D.N. 495 (10-AD), 918 (10-AD). Kuchel, R.H. 271 (3-AD), 515 (10-AD), 659 (3-AD), 681 (10-AD) 716 (3-AD), 2147 (8-

AD), 3328 (10-AD). Lander, N.S. 1101 (10-BH, BM, BRI, MEL*, NSW, NT, PERTH), 1113 (3-BH, CANB, K, MEL*, NT, PERTH), 1116 (12-BH, K, PERTH), 1120 (12-MEL*, PERTH), 1122 (11-K, MEL*, NSW, PERTH), 1123 (11-BH, CANB, PERTH), 1125 (7-BH, BRI, K, MEL*, NT, PERTH), 1128 (12-NSW, PERTH), 1129 (7-CANB, MEL*, NSW, NT, PERTH), 1139 (7-NSW, PERTH), 1140 (7-PERTH), 1142 (7-MEL*, NSW, PERTH), 1143 (4-BM, BRI, K, NSW, NT, PERTH), 1145 (7-BH, BM, CANB, K, MEL*, NSW), 1149 (4-MEL*, NT, PERTH), 1150 (4-PERTH, NT), 1151 (7-PERTH), 1154 (7-BM, PERTH), 1155 (4-K, PERTH), 1156 (7-BH, PERTH), (150 (4-PERTH, N1), (151 (7-PERTH), 1154 (7-BM, PERTH), 1159 (4-P. 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Lothian, T.R.N. 698 (10-AD), 1101 (3-AD), 1122 (3-AD), 1255 (2-AD). 1407 (10-AD), 1937 (3-AD), 2634 (10-AD), 3929 (3-AD), 4346 (10-AD), 4900 (3-AD), 5508 (10-AD, PERTH). Lowry, J. 43 (2-PERTH). Lullfitz, F. 1496 (12-KP, PERTH). 1588 (12-KP), s.n., Jan. 1963 (2-PERTH). McDougall, V.F. 22(1-PERTH). McGillivray, D.J. 3374 (12-NSW, PERTH). Mackinnon, D.D. 12 (7-PERTH). Maclaine, E. s.n., 1894 (2-MEL). Maconochie, J.R. 1891 (4-AD, CANB, CBG, MEL, NT), 1893 (3-AD, MEL, NT). Maddox, M.D. s.n., Sept. 1924 (2-MEL). Malcom, C.V. s.n., Oct. 1960 (3-PERTH), s.n., Oct. 1960 (10-PERTH). Maroske, J.O. s.n., 1959 (3-MEL). Maxwell, G. s.n., s.dat. (1-K, MEL), s.n., s.dat. (1-MEL), s.n., s.dat. (1-MEL), s.n., s.dat. (1-MEL), Melville, R. 219 (10-K, MEL), 988 (10-K, MEL), 4011 (12-AD, BRI, K, MEL, NSW, PERTH), 4019 (12-K, PERTH). Merrall, E. s.n., 1889 (8-MEL), s.n., 1890 (8-MEL), s.n., 1890 (1-MEL), s.n., 1892 (1-MEL), s.n., 1889 (1-MEL). Milewski, A.V. s.n., Oct. 1981 (3-PERTH), s.n., Oct. 1981 (9-PERTH), s.n., June 1982 (8-PERTH). Milligan, J. s.n., Feb. 1844 (2-HO). Milthorpe, P.L. 576 (3-AD), 2440 (10-NSW). Minchin, J. s.n., Oct. 1887 (3-MEL). Mitchell, A.A., 45 (10-NT, PERTH), 517 (4-PERTH), 845 (3-PERTH), 968 (11-MEL*, PERTH), 969 (7-PERTH). Mitchell, A.S., 517 (4-PERTH), 995 (3-NT, PERTH), 1010 (4-NT, PERTH), 1023 (6-NT, PERTH), 1059 (10-NT, PERTH), 1247 (3-NT), 1259 (3-NT, PERTH), 1059 (10-NT, PERTH), 1247 (3-NT), 1259 (3-NT, PERTH), 1059 (10-NT, PERTH), 1247 (3-NT), 1259 (3-N NT). Moore, C. W. E. 5087 (3-CANB), 6367 (3-CANB), 6760 (10-CANB, NSW), 6761 (10-CANB, NSW). Moore, S. s.n., 1895 (8-BM, K). Morris, K.A. s.n., Oct. 1936 (3-BRI). Morrison, A. 11 (12-k), 15 (4-K), s.n., Feb. 1903 (2-BM, CANB, MEL), s.n., June 1903 (2-BM), s.n., Nov. 1906 (3-BM, BRI, PERTH), s.n., s.dat. (12-PERTH, KEW). Mueller, F. s.n., Dec. 1850 (2-MEL 98721, 98722, 98723, 98757), s.n., Nov. 1852 (2-MEL), s.n., Jan. 1854 (3-MEL), s.n., Oct. 1857 (10-98790, 98812 & 584132), s.n., Oct. 1857 (10-MEL 584133), s.n., Oct. 1867 (10-MEL), s.n., Oct. 1867 (10-MEL), s.n., 1880 (3-MEL), s.n., s.dat. (2-MEL 98725, 98755), s.n., s.dat. (2-MEL 98732), s.n., s.dat. (2-MEL 98747), s.n., s.dat. (2-K, MEL 98763), s.n., s.dat. (3-MEL 98683), s.n., s.dat. (3-BM, MEL 98684, 98686 & 98687 pro pte, NSW), s.n., s.dat. (3-MEL 98688), s.n., s.dat. (3-MEL 98710), s.n., s.dat. (3-MEL 98712), s.n., s.dat. (3-NSW), s.n., s.dat. (10-MEL 98775), s.n., s.dat. (10-MEL 98785), s.n., s.dat. (10-MEL 98787), s.n., s.dat. (10-MEL 98791, K), s.n., s.dat. (10-MEL 98812), s.n., s.dat. (10-MEL 584131), s.n., s.dat. (10-NSW). Muir, E.T. ACB 42987 (3-MEL), ACB 42994 (10-MEL), s.n., 1948 (3-MEL). Murray, B.J. s.n., Nov. 1927 (3-AD), Newbey, K. 407 (1-PERTH), 1463 (1-PERTH), 3084 (10-PERTH), 4247 (1-PERTH), 6879 (1-PERTH), 7239 (8-PERTH), 7521 (10-PERTH), 7537 (8-PERTH), 7575a (9-PERTH), 7658 (3-PERTH), 8095 (10-PERTH), 8304 (8-BH, NSW, PERTH), 8523 (8-PERTH), 8853 (8-PERTH), 9011 (8-PERTH), 9136 (10-PERTH), 9456 (3-PERTH), Nicholls, A.O. 808 (10-K, MEL), 818 (3-AD, CANB, K, MEL, PERTH, NT). Noble, J.C. 184 (3-AD), 622 (2-AD). Norton, J. s.n., Nov. 1936 (2-AD). O'Farrell, R. 16 (3-PERTH). Oldfield, A. s.n., s.dat. (4-K, MEL). Ollerenshaw, P. 148 (3-AD), 249 (3-CANB). Olsen, M. 717 (3-BRI). Orchard, A.E. 114 (10-AD), 1445 (9-AD), BH, PERTH). Osborn, T.G. B. s.n., Oct. 1918 (AD). Paijmans, K. 3166 (3-CANB), 3319 (10-CANB). Parham, J. W. s.n., Feb. 1976 (2-HO). Parker, S. 242 (3-AD), 280 (4-AD, K, MEL, NT). Parr-Smith, G. 1359 (10-PERTH). Payne, A. 163 (4-PERTH). Peake-Jones, K. s.n., Aug. 1952 (3-AD). Pearman, G. s.n., s.dat. (12-UWA). Pearson, J.D.M. 2024 (3-MEL). Perry, R.A. s.n., Oct. 1944 (10-CANB). Phillips, M.E. 49 (10-CBG), 237 (10-CBG), CBG 20817 (8-CANB), CBG 21616 (10-AD, CBG). Pickard, J. 2458 (3-NSW). Polak, J. (as "Pollack") s.n., 1882 (3-MEL). Preiss, L. 1231 (10-MEL), 2381 (2-MEL), 2387 (2-MEL). Probert, L. 356 (10-AD), 362 (10-AD), 418 (10-AD), 420 (10-AD), 422 (10-AD), 431 (10-AD). Purdie, R.W. 759 D (10-BRI). Quartermaine, E. s.n., Dec. 1934 (2-PERTH). Reader, F.M. 2 (3-MEL), 8 (2-MEL), s.n., March 1895 (3-BM), s.n., Oct. 1895 (10-MEL), s.n., Dec. 1895 (2-MEL), s.n., Nov. 1898 (3-MEL). Reader. J. 5 (10-MEL), 8 (10-MEL), s.n., March 1895 (3-MEL). MEL). Rees, J.D. s.n., s.dat. (10-AD). Richards, T. 4 (3-MEL), s.n., 1875 (10-MEL, 98810), s.n., 1875 (10-MEL, 98810), s.n., 1877 (3-MEL) 98709 & 98676), s.n., 1800 (3-MEL), s.n., 1800 (3-MEL), s.n., s.dat. (3-MEL). Ridley, J.B. s.n., April-Sept. 1863 (7-BM, K). Ritman, H. s.n., Feb. 1960 (2-MEL). Robertson, J.G. s.n., s.dat. (2-K). Robinson, A. s.n., Oct. 1976 (10-AD). Rodway, E. 2358 (2-K, NSW). Rodway, L. s.n., 1883 (2-HO), s.n., Jan. 1897 (2-HO), s.n., April 1930 (2-HO, K), s.n., s.dat. (2-HO 12978), s.n., s.dat. (2-NSW), Rogers, Mrs. R. s.n., Oct. 1915 (10-NSW), Royce, R.D. 5357 (10-PERTH), 5829 (10-PERTH), 6630 (7-PERTH), 6655 (3-PERTH), 9137 (2-PERTH), Saffrey, R.A. 657 (7-PERTH), 803 (12-PERTH), 816 (11-PERTH). Saunders, Mrs. s.n., Oct. 1971 (1-KP). Schomburgk, R.D. 82 (10-AD), K. PERTH), s.n., March 1872 (2-K), s.n., s.dat. (1-AD, BRI), s.n., s.dat. (2-AD), s.n., s.dat. (10-AD). Serventy, D.L. s.n., Feb. 1982 (2-PERTH). Sharrad, M.C.R. 1224 (10-AD). Short, P.S. 1185 (3-AD, CBG, MEL). Sims, E.B. s.n., Aug. 1969 (10-AD). Smith, G.G. s.n., Aug. 1958 (10-UWA). Smith, P. 53 (4-PERTH). Smith, R.V. 517 (12-MEL). Smith, T.J. 439 (10-AD), 580 (10-AD), 1096 (2-AD), 1109 (2-AD), 1225 (10-AD), 1485 (10-AD), 1727 (10-AD), 1796 (10-AD), 1860 (2-AD), 1864 (2-AD), 1871 (2-AD), 1900 (10-AD). Somerville, J. s.n., Dec. 1960 (2-HO). Southcott, R.V. s.n., Dec. 1960 (3-AD), Specht, 1912 (10-AD), 1913 (10-AD), 1913 (10-AD), 1913 (10-AD), 1914 (10-AD), 1915 (10-R.L. 24 (10-AD), 2227 (2-AD), s.n., Nov. 1959 (10-AD), Speck, N.H. 644 (3-CANB, PERTH), 683 (3-PERTH), 730 (3-CANB, PERTH), 744 (12-BRI, CANB, NSW, PERTH), 1346 (10-AD, BRI, K, MEL, NSW), 1435 (3-AD, CANB, MEL). Spooner, A.G. 1677 (10-AD), 2430 (3-AD), 3224 (2-AD), 6149 (10-AD). Stevenson, P. s.n., Nov. 1978 (2-PERTH). Stoward, F. s.n., Nov. 1914 (3-BM, NSW 146993 & 146994). Story, Dr. s.n., s.dat. (2-MEL). Streich, V. s.n., 1895 (12-MEL). Strid, A. 20150 (12-PERTH). Stuart, J.M. s.n., 1859 (3-K). Sutton, C.S. 1174 (10-K), 1175 (3-K), s.n., Oct. 1905 (2-MEL). Swinbourne, R. 94 (3-AD), 180 (3-AD). Symon, D.E. 2338 (3-AD, PERTH), 3205 (10-NSW), 3215 (10-CANB, K), 3268 (3-CANB, K), 4300 (1-AD, CANB), 4308 (3-CANB), 5460 (12-AD, PERTH), 8149 (3-K), 9945 (12-AD, PERTH). Tate, R. s.n., 1880 (3-AD), s.n., 1883 (2-AD), s.n., 1883 (3-AD), s.n., 1889 (3-AD), telford, I.R. 5859 (12-CANB). Tepper, J.G.O. 82 (1-MEL), 444 (1-MEL), 756 (2-MEL), 880 (2-MEL), 916 (3-MEL), 932 (2-MEL), 1209 (2-MEL), s.n., Oct. 1879 (1-AD), s.n., Jan. 1880 (1-MEL), s.n., s.dat. (1-MEL). Telkins, W.H. s.n., 1880 (3-MEL). Tilden, J.E. 797 (2-BM, K). Tindale, M.D. 130321 (3-NSW). Tindale, N.B. s.n., 8 Aug. 1935 (3-AD). 535 (3-AD). Toelken, H.R. 6093 (12-AD, BH, BM, CANB, PERTH), 6094 (3-AD, BH, PERTH), 6129 (10-AD). Trezise, P. 359 (3-AD), 370 (3-AD). Turner, H. s.n., Sept. 1960 (3-AD). Turner, J. s.n., April 1977 (2-MEL). Tyson, I. 18 (10-MEL), s.n., 1892 (7-MEL). Tikery, J.W. s.n., Oct. 1899 (10-NSW). Mace, N.M. 9 (10-AD), 255 (3-AD), 136 (10-AD), 146 (3-AD), 172 (10-AD), 193 (3-AD), 212 (10-AD), 219 (3-AD, MEL), 228 (10-AD), 255 (3-AD), 262 (10-AD), 270 (10-AD), 285 (3-AD). Wade, R. s.n., Sept. 1971 (10-MEL). Wakefield, N.A. 2243 (2-MEL). Walter, C. s.n., Oct. 1899 (3-NSW), s.n., Oct. 1899 (10-NSW), s.n., Oct. 1900 (10-BR), K), S.n., Feb. 1902 (2-NSW). Warburton, R.E. s.n., s.dat. (2-MEL), s.n., s.dat. (3-MEL). Warnes, K.B. 31 (3-AD), 34 (3-AD), 425 (10-AD), 426 (10-AD

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