FLOW CYTOMETRY PRINCIPLES AND PRACTICE

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Aims and Objectives

- Principles of flow cytometry
 - Preparation
 - Steps involved
 - Interpretation of flow cytometry plots
 - Use of panels
- Cases

Introduction

- Identifies cell types present in cell suspension
- Fluorescent-labelled antibodies specific for lineage or maturation markers
- 'marker' molecules: usually on cell surface
- Possible to permeabilize cells to allow antibody penetration to detect intracellular antigens e.g. TdT and Myeloperoxidase

Preparation of cells

- Blood and bone marrow red cells lysed (e.g. ammonium chloride).
- White cells recovered by washing and centrifugation
- Aliquots of the cell suspension are incubated with different combinations of fluorescentlyconjugated antibodies
- CSF, ascitic fluid, pleural fluid or a cell suspension made by 'mashing' solid tumour samples can be analyzed in a similar way.

Step 1: Add a fluorescently-labelled antibody specific for the cell marker



Different Fluorochromes



Different emission spectra for different fluorochromes means that multiple cell surface markers may be interrogated.

Step 2: Separating out subpopulations of cells for identification



Three parameters are recorded

- Forward scatter this reflects cell size
- Side scatter reflects granularity
- Fluorescence signal no. of fluorochromes/cell

Using CD45/SCC to 'gate' different sorts of cells in blood



• CD45: common leucocyte antigen

 protein tyrosine phosphatase that is specifically expressed in haemopoietic cells.

- Its variable positivity allows separation of blood cells.
- It is negative on plasma cells, RBCs and some ALL

Choosing a panel of antibodies

Inspect morphology + clinical picture, and choose a panel:



Acute leukaemia panel

Lineage CD79a – B cell receptor component CD3 – T cell receptor component MPO – myeloperoxidase

CD117 – c-kit (most AML, few ALL) CD34 – expressed on stem cells CD19 – pan-B cell marker CD10 – expressed on ALL CD7 – T cell marker HLADR CD33 – myeloid marker TdT – all ALL, 25% AML Lysozyme – monocytic marker Chronic lymphoproliferative disease panel



Lineage

CD3 – T cell receptor component CD19 – pan-B cell marker CD79b – B cell receptor component Clonality of B cells ? Kappa/lambda light chains

CD20 –B cells; not early blasts/plasma cells CD23 – B-CLL FMC7 – conformational variant of CD20 CD5 – T cell marker, also expressed in B-CLL CD45 – pan-leucocyte marker CD10 – T/B ALL and germinal centre marker If T cell population: extended T cell panel (e.g. CD2, CD7, TCR, CD4, CD8, CD16, CD56, CD57...)

Analysis of Acute Leukaemia: Identify Lineage:

Does the case express:-

1. MPO? myeloid

2. CD79a? B cell

3. Cyt CD3 OR sCD3 ? T cell

4. TdT ? Primitive; often ALL



40 year old Female

Mucous bleeding and easy bruising

Hb 9 Plt 12 WCC 15

Deranged clotting

NORMAL

CD45

800

Side scatter







Case 1 Acute promyelocytic Leukaemia (APML)

Characteristic markers:

CD117 usually positive may be weak CD33 positive MPO strongly positive HLADR negative CD 9 positive

12 year old boy Lethargy and night sweats Hb 7 WCC 70 Plts 50







Case 2 B-cell precursor lymphoblastic leukaemia/lymphoma

Characteristic markers:

CD117 negative CD19 positive CD10 positive CD79a positive TdT positive MPO negative CytCD3 negative

Chronic lymphoproliferative disorders



Mantle cell lymphoma and Chronic lymphocytic leukaemia are the 2 major B cell chronic lymphoproliferative diseases that express CD5 and CD19.

Practicalities: how to perform the chronic lymphoproliferative panel



к FITC	FMC7 <i>FITC</i>	CD79b <i>FITC</i>	CD38 FITC	CD5 FITC
λ <i>ΡΕ</i>	CD23 PE	CD20 PCP	IgM PE	CD10 <i>PE</i>
CD19 PCP	CD19 PCP	CD200 <i>PE</i>	CD19 PCP	CD19 PCP
CD5 <i>APC</i>				
CD45 APCCY7				



65 year old male, just retired Lethargic, abdominal fullness Hb 80, WCC 30, platelets 90





CD19

Case 3 Mantle Cell Lymphoma

Characteristic markers:

CD19 and CD5 positive CD23 negative CD79b positive FMC7 positive High level of slg Strong kappa or lambda expression



Characteristic markers:

CD19 and CD5 positive CD23 positive CD79b negative FMC7 negative Low level of slg – may be difficult to detect clonality

B-cell CLL Scoring system



92% of CLL score 4 or 5, 6% score 3 and 2% score 1-2.

Moreau et al, 1997; Okaly et al, Journal of Clinical and Diagnostic Research, 2013.

CD19 positive

High levels of slg so CD79b positive

Obvious clonality

High levels of CD20 - FMC7 is positive

CD10 is positive in FL

Flow cannot separate cases of LPL vs MZL



70 year old male Atypical infection Pancytopenia Early satiety WCC 12 Hb 70 Platelets 45







CD103





Case 5 : Hairy cell Leukaemia

Characteristic markers:

CD19 and CD5 negative CD79b positive CD20 strongly positive FMC7 positive CD11c, CD103 and CD25 positive

Diagnosis	CD5	CD10	CD19	CD20	CD23	CD79b	FMC-7	CD25	CD11c	CD103
CLL/SLL	+	-	+	+(w)	+	-	-	-/+	+/-	-
Mantle cell Iymphoma	+	-	+	+	-	+	+	-	-	-
Follicular Lymphoma	-	+	+	+	-/+	+/-	+/-	-	-	-
Marginal zone Iymphoma	-	-	+	+	-	+/-	+/-	-/+	+	-
Hairy cell leukemia	-	-	+	+	-	+/-	+/-	+/-	+	+





55 year old female History of Rheumatoid arthritis RhF+ve Hb 130, Plts 225, WCC 11 Neutropenia Neut 0.7





Case 6: Large Granular Lymphocytosis

Characteristic markers:

CD 2 positive CD3 positive CD4 negative CD8 positive Often CD16, CD57 positive TCR A/B or D/G expression