


Article

Target Site Resistance to Acetolactate Synthase Inhibitors in *Diplotaxis erucooides* and *Erucaria hispanica*—Mechanism of Resistance and Response to Alternative Herbicides

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Received: 8 February 2020; Accepted: 25 March 2020; Published: 29 March 2020



Abstract: *Diplotaxis erucooides* and *Erucaria hispanica* are common weeds of the Mediterranean region; they infest various habitats including cultivated fields and roadsides. In several fields across Israel, farmers have reported on poor control of *D. erucooides* and *E. hispanica* plants using acetolactate synthase (ALS) inhibitors. Greenhouse experiments were conducted to determine the effect of various ALS inhibitors on plants from two potentially resistant *D. erucooides* and *E. hispanica* populations. Additionally, alternative management strategies using auxinic herbicides were studied. Plants from both populations exhibited resistance to all tested ALS inhibitors, up to 20-fold the label field rate, as compared with ALS sensitive populations of *D. erucooides* and *E. hispanica*. Sequencing of the ALS gene revealed Trp574 to Leu substitution in ALS-resistant *D. erucooides* plants, whereas a Pro197 to Ser substitution was detected in ALS-resistant *E. hispanica* plants. Although high levels of resistance were observed in individuals from both putative resistant populations, sensitive individuals were also detected, suggesting the evolution of resistance in these two populations is still in progress. Auxinic herbicides, 2,4-D, and mecoprop-P, provided excellent control of plants from both ALS-resistant populations. This study documents and confirms the first case of evolution of resistance to ALS inhibitors in *D. erucooides* and *E. hispanica* populations.

Keywords: alternative management; auxinic herbicides; herbicide resistance; Spanish pink mustard; white wall-rocket

1. Introduction

Diplotaxis erucooides (white wall-rocket) and *Erucaria hispanica* (Spanish pink mustard) are common weeds of the Brassicaceae family spread across the Mediterranean region [1]. These species are frequently found as native weeds mostly in Portugal, Spain, Italy, Cyprus, Greece, Turkey, and Israel; recently, they had also invaded as alien species other countries, such as Romania, Norway, Switzerland, and Slovakia [2]. High germination rate and reproductive success may contribute to the high abundance of these two species [3,4]. Furthermore, both species have high seed fecundity with reproduction mechanisms that prevent self-pollination [5,6]. These species can germinate in several flushes in a season, enabling them to better compete with crop seedlings, dominate the entire field, and cause high yield losses (B Rubin, personal communication). The fact that these weed species are exogamous (i.e., they rely mainly on outcross-pollination) may enhance the spread of beneficial adaptive traits, especially if they are inherited as a single dominant gene, such as in the case of target-site resistance [7].

Acetolactate synthase (ALS) inhibiting herbicides (group B, HRAC/WSSA) are the main mode of action (MOA) used for selective weed control in crops [8,9]. With more than 50 known active ingredients, this MOA can be used almost in every crop; thus, herbicide rotations are often neglected [10]. Five different chemical groups are known as ALS inhibitors: sulfonyleurea (SU), imidazolinone (IMI), triazolopyrimidine (TP), pyrimidiny-lthiobenzoate (PTB), and sulfonyl-aminocarbonyl-triazolinone (SCT) [10].

The evolution of resistance to photosystem II (PSII) inhibiting herbicides (group C) has resulted in a dramatic increase in the use of ALS inhibitors in agricultural and non-agricultural areas [11]. This over-use across all types of habitats resulted in a fast increase in the number of cases of evolved resistance and even lead to the evolution of multiple-herbicide resistance species [12]. Mechanisms of resistance to ALS inhibitors in weeds typically involve an altered target site [13]. Different mutations and substitutions may result in various levels of resistance to ALS inhibitors [14]. Generally, the evolution of an herbicide resistance trait is associated with a fitness cost to the individuals showing the trait when the selective factor (herbicide) is removed from the environment, thus the frequency of such mutations is presumed to be very low [15]. However, in most cases, target-site resistance to ALS inhibitors does not carry a fitness penalty [16]. Resistance to ALS inhibitors can also be endowed by non-target site mechanisms as reviewed by Délye (2013). This type of ALS-resistance was reported in *Sinapis arvensis* L. [17] and *Papaver rhoeas* [18], as well as in plants showing multiple-resistance with other modes of action, such as glyphosate and ACCase-inhibitors [19].

In Israeli dry-land farming, wheat (*Triticum aestivum*) and pea (*Pisum sativum*) are the main crops. Two common three-year crop rotations are traditionally in practice: (i) the main rotation is wheat/wheat/fallow, where SU, TP, or SCT herbicides are mainly used; and (ii) the minor one is wheat/peas/wheat where IMI herbicides are used in peas, and SU, PTB, or SCT are commonly used in wheat.

Recently, farmers have reported on poor control of *D. erucoides* and *E. hispanica* plants using ALS-inhibitors. We suggest that the intensive use of ALS inhibitors in both rotations resulted in the evolution of ALS-resistant individuals. The aim of this study was to (1) confirm ALS resistance in two potentially resistant *D. erucoides* and *E. hispanica* populations and (2) to elucidate the mechanism of resistance. Moreover, (3) we tested the use of auxinic herbicides (Group O) as an alternative management practice, in preventing further buildup of ALS-resistant weed populations.

2. Materials and Methods

2.1. Plant Material

Seeds of two potentially resistant populations of *D. erucoides* (DER) and *E. hispanica* (EHR) were collected at Kibbutz Dvir (31°24'45" N 34°49'30" E) and Kibbutz Beeri (31°25'26" N 34°29'28" E), respectively. Samples were collected from fields (20–30 hectares) where farmers reported on weed control failure using ALS inhibitors. Seeds of sensitive populations of *D. erucoides* (DES) and *E. hispanica* (EHS) were collected from an uncultivated area where no herbicides have been used at Kibbutz Re'em (31°23'08" N 34°27'34" E). To ensure appropriate representation of field population, mature seedpods from 30 to 40 randomly selected plants in each population were collected and pooled. Collected seeds were air-dried and stored at 4 °C until used. Seeds from each population were germinated in flats filled with commercial potting media (Tuff, Marom Golan, Israel), including Osmocote®(The Scotts Company, Marysville, OH, USA) slow release fertilizer. Seedlings of *D. erucoides* and *E. hispanica* populations at the fully developed cotyledons stage were transplanted into pots, 7 by 7 by 6 cm (one plant per pot), filled with the same potting media. Pots were kept in a greenhouse (22/16 °C, day/night), at natural light conditions, and watered daily.

2.2. Plant Responses to ALS Herbicides

At the stage of three to four true leaves, plants were treated with 0, 1/32, 1/16, 1/8, 1/4 1/2, 1, 2, or 4 times the label field rate of each ALS inhibiting herbicides using a chain-driven sprayer delivering 300 L ha⁻¹, with a flat-fan 8001E nozzle (TeeJet®, Spraying Systems Co., Wheaton, IL, USA). Different ALS inhibitors (commercial formulations) and their label field rate are detailed in Table 1. The level of resistance in both DER and EHR populations was determined in a separate experiment when plants from all populations were treated with high rates of 10 and 20 times the label field rate of each herbicide as previously described. All experiments were arranged in a completely randomized-factorial design, with three to five replicates for each treatment, and repeated three times. No significant treatment by experimental run were observed; therefore, the data obtained from the repeated experiments for each herbicide were pooled. Shoot fresh weight (FW) and survival rate (alive or dead) were recorded 21 days after treatment (DAT).

Table 1. Herbicides used in this study. ALS = acetolactate synthase; MOA = mode of action; SU = sulfonylureas; IMI = imidazolinones; TP = triazolopyrimidines; SCT = sulfonylaminocarbonyltriazolinones.

MOA	Common Name	ALS Chemical Family	Trade Name	Label Field Rate (g ai ha ⁻¹)	Manufacturer
ALS Inhibitors	Tribenuron-methyl	SU	Express®	15	DuPont
	Imazamox	IMI	Pulsar®	24	BASF
	Florasulam	TP	Darbuka®	4	ADAMA-Agan
	Propoxycarbazone-sodium	SCT	Olympus®	45.5	Bayer
Auxinic Herbicides	2,4-D		Albar Super®	670	ADAMA-Makhteshim
	Mecoprop-P		Duplosan®	1200	Nufarm

2.3. Plant Responses to Auxinic Herbicides

At the stage of three to four true leaves, plants from both DER and EHR populations were treated with two different auxinic herbicides, as detailed in Table 1. Each of the auxinic herbicides was applied at the following rates: 1/4, 1/2, 1, 2, or 4 times the label field rate as described above. The experiment was arranged in a completely randomized-factorial design, with three to five replicates of each treatment and was repeated three times. No significant treatment by experimental run were observed; therefore, the data obtained from the repeated experiments for each herbicide were pooled. Shoot FW and survival rate (alive or dead) were recorded 21 DAT.

2.4. DNA Extraction and Molecular Studies

Leaf tissue (3 cm²) was excised from individual plants of all four populations (DER, EHR, DES, and EHS). Each sample was placed separately in a microtube. DNA was extracted using the Puregene DNA isolation kit (Gentra Systems, Minneapolis, MN, USA) according to the manufacturer's instructions and diluted 10-fold before further use. Specific primers were used to identify the ALS gene and locate common point mutations previously shown to endow resistance to ALS inhibitors. Primers were designed according to the known sequences of the ALS genes from *Arabidopsis thaliana* (X51514) and *Lolium rigidum* (DQ184640.1) (Table 2). Sequence analyses and alignment were performed using the Bioedit software [20]. The obtained sequences were compared to the known sequence of the ALS gene of *A. thaliana* (X51514).

Table 2. Primers used for PCR and sequencing reactions.

Primers	Sequence (5'-3')	Position	Product	Reference
			Size (bp)	
ALS-A F ALS-A R	GCTGATATCCTCGTCAAGC GAATCGGAAGCTGTTGA	122, 197, 205	490	<i>Arabidopsis thaliana</i> (X51514) and <i>Lolium rigidum</i> (DQ184640.1)
ALS-B F ALS-B R	CGCTGTTGATAAGGCTGACC ACAAGTATGGCCCAGGAGTC	376, 377, 574	800	
ALS-C F ALS-C R	AAGTACTGGTGTCTGGGCAAC GGCAACACATGTTCTGGTG	574, 653, 654	500	

F-forward, and R-reverse primers. ALS-A, B and C—first, second and third fragment of the ALS gene sequence.

2.5. Statistical Analyses

Data of FW were analyzed using ANOVA in JMP (ver. 13) statistical package (SAS Institute Inc., Cary, NC, USA) as the percentage of FW reduction compared to the untreated control. Means were compared using Student's *t*-test ($\alpha = 0.05$). Dose-response curve experiments were conducted to evaluate the effect of different herbicides on the average shoot FW and survival rate of plants from each population. For each treatment, data is presented and plotted as a percentage of untreated control. Data were fit to a nonlinear sigmoidal logistic three-parameter model [21] as described in Equation (1):

$$Y = \frac{a}{1 + \left(\frac{x}{X_0}\right)^b} \quad (1)$$

In this model, if $b > 0$, then a describes the upper limit of Y ; X_0 describes the herbicide rate causing 50% shoot FW reduction (ED_{50}) or 50% mortality (LD_{50}), and b describes the slope of the curve. For each species, the resistance index (RI) was calculated as the R: S (resistant: susceptible) ratio.

3. Results and Discussion

3.1. Response to ALS Inhibitors

Although they are very common in the Mediterranean region, apart from this case, herbicide resistance has never before been reported for both *D. eruroides* or *E. hispanica* [9]. Plants from the DER population exhibited resistance in response to imazamox, propoxycarbazone-sodium, and tribenuron-methyl (RI > 10). EHR plants, were highly resistant to all tested ALS inhibitors except imazamox (Table 3; Figures 1 and 2). A wide range of herbicide responses among DER and EHR populations were recorded, including highly resistant, as well as highly sensitive individuals were found in both populations. This was mainly pronounced in imazamox-treated DER plants and tribenuron-methyl treated EHR plants, as increasing rates did not show the same response as some of the lower rates did (Figure 1b). No significant reduction in FW and survival percentage of EHR plants was recorded in response to tribenuron-methyl; thus, the ED_{50} and LD_{50} values have exceeded the highest applied rate (60 g ha^{-1}), which eventually resulted in failure to fit a correct model to describe this treatment (Figure 2a).

Table 3. Effects of post-emergence treatments on the shoot fresh weight (FW) of plants from *D. erucoides* (DES) and *E. hispanica* (EHS) ALS-sensitive and -resistant populations (DES, *D. erucoides* (DER), EHS, and *E. hispanica* (her), respectively). Herbicides were applied at 1 and 4 times the label field rate (X).

Pop'	Shoot FW (% of untreated control)															
	Imazamox ^a				Propoxycarbazone-Sodium ^b				Tribenuron-methyl ^c				Florasulam ^d			
	X	4X	ED ₅₀	RI	X	4X	ED ₅₀	RI	X	4X	ED ₅₀	RI	X	4X	ED ₅₀	RI
DER	21 ± 40	16 ± 31	23.7	14	24 ± 10 *	17 ± 7 *	14.5	72.5	47 ± 19 *	25 ± 17	3.4	85	20 ± 24	31 ± 31	1	5
DES	0.2 ± 0.1	0.2 ± 0.1	1.7		1.7 ± 0.3	0.9 ± 0.4	0.2		1.5 ± 1	2.6 ± 0.7	0.04		0.6 ± 0.9	0.16 ± 0.1	0.2	
EHR	51 ± 12 **	22 ± 31	28.5	7	55 ± 33 *	31 ± 27 *	55.9	69.8	128 ± 35 **	57 ± 64	>60	>60	106 ± 31 **	47 ± 55	14.4	24
EHS	2 ± 1.3	0.2 ± 0.2	4.1		0.9 ± 1.1	0.5 ± 0.3	0.8		1.3 ± 0.3	0.1 ± 0.1	0.4		2.6 ± 2.7	0.2 ± 0.1	0.6	

ED₅₀ value represent herbicide rate reducing plant growth by 50% were extracted from the dose response curves. RI was calculated as the ratio of the ED₅₀ value of the resistant population compared with the ED₅₀ of the sensitive one. Each comparison was between a resistant and sensitive population from the same species under the same treatment. * or ** asterisks indicate on significant differences between treatments, as determined using Student *t* test ($P \leq 0.05$ and 0.01) ($n = 12$). Label field rate for each herbicide (X): ^a 24 g ai ha⁻¹, ^b 45.5 g ai ha⁻¹, ^c 15 g ai ha⁻¹, and ^d 4 g ai ha⁻¹.

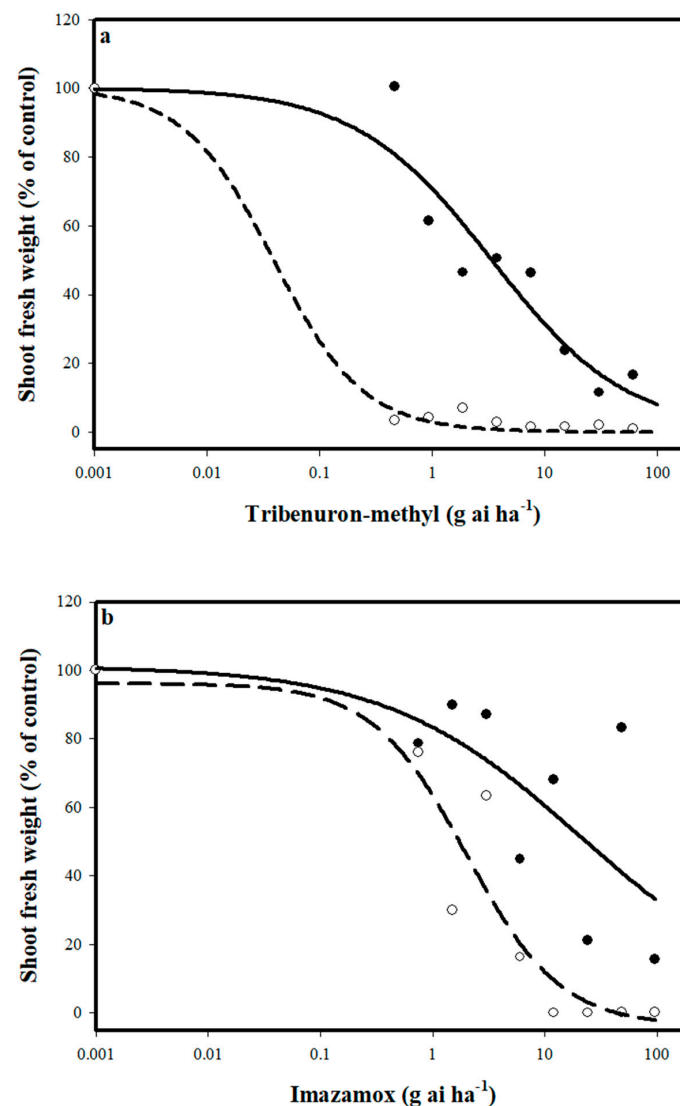


Figure 1. Effects of tribenuron-methyl (a) and imazamox (b) applied post-emergence at increasing rates on the shoot fresh weight of S (open circles) and R (closed circles) populations of *Diplotaxis erucoides*. Data was collected 21 days after treatment.

The fact that both resistant populations (DER and EHR) exhibited high level of variability in the response to ALS inhibitors may indicate that these populations are still in transition, comprising a mixture of resistant and sensitive individuals. The data concerning survival rates show that even though shoot fresh weight was not significantly different from sensitive populations (DES and EHS), more than 25% of plants from both resistant populations survived the highest rate (4X) of all four ALS inhibitors (Table 4). When treated with the highest rate of tribenuron-methyl, DER showed a higher survival rate than EHR; however, LD₅₀ values of both populations exceeded the highest rate (>60). In contrast, EHR had higher survival rates following treatment with imazamox, as presented also by higher LD₅₀ values (36.67 vs. 19.42; Table 4). When both sets of data (shoot FW and survival rate) are viewed side by side, it appears that the DER and EHR populations are at a transitional stage and have not yet developed homogenic response to the tested ALS inhibitors. Using high herbicide rates, 10 and 20 times the label field rate, we have detected high number of survivors in both species alongside susceptible individuals (data not shown), reinforcing our hypothesis that the evolution of resistance to ALS inhibitors in these two populations is still in progress. Previous studies on *D. erucoides* or *E. hispanica* mainly deal with their ecological traits and adaptation ability to thrive in different

environments [3,4,6,22]. These highly adaptive traits emphasize the risk in the evolution of herbicide resistance of these two species.

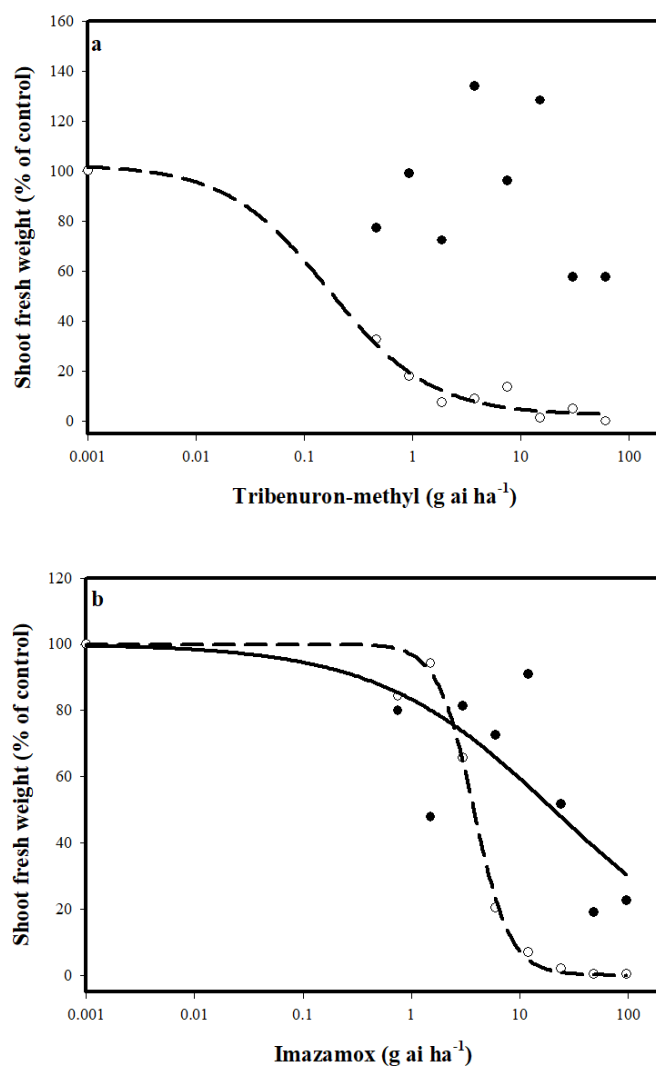


Figure 2. Effects of tribenuron-methyl (a) and imazamox (b) applied post-emergence at increasing rates on the shoot fresh weights of S (open circles) and R (closed circles) *Erucaria hispanica* populations. Data was collected 21 days after treatment.

Table 4. Effects of post-emergence treatments on the survival rate of plants from *D. eruroides* and *E. hispanica* ALS-sensitive and -resistant populations (DES, DER, EHS, and EHR, respectively). Herbicides were applied at 1 and 4 times the label field rate (X).

9	Survival (%)															
	Imazamox ^a				Propoxycarbazone-sodium ^b				Tribenuron-methyl ^c				Florasulam ^d			
	X	4X	LD ₅₀	RI	X	4X	LD ₅₀	RI	X	4X	LD ₅₀	RI	X	4X	LD ₅₀	RI
DER	50	25	70.30	19.42	50	50	69.93	>182	100	75	>60	>60	50	75	132.17	>16
DES	0	0	3.62	19.42	0	0	0.002	>182	0	0	0.01	>60	0	0	1.02	>16
EHR	100	33	93.82	36.67	100	50	50.00	>182	100	50	62.45	>60	100	75	>16	>16
EHS	0	0	2.85	36.67	0	0	0.002	>182	0	0	0.70	>60	0	0	0.69	>16

LD₅₀ value represent herbicide rate reducing survival rate by 50% were extracted from the dose response curves. RI was calculated as the ratio of the LD₅₀ value of the resistant population compared with the LD₅₀ of the sensitive one. Label field rate for each herbicide: ^a 24 g ai ha⁻¹, ^b 45.5 g ai ha⁻¹, ^c 15 g ai ha⁻¹, and ^d 4 g ai ha⁻¹.

3.2. Mechanism of Resistance to ALS Inhibitors

Several different *ALS* gene point mutations have been previously identified to endow resistance to ALS inhibitors [14]. Specific point mutation may alter plant response to ALS inhibitors, as will different amino acid substitutions at the same point mutation [23].

Alignment of sequences from DER plants revealed a Trp574 to Leu substitution in the *ALS* gene sequence (Figure 3a). In EHR plants, a different substitution, Pro197 to Ser was detected (Figure 3b). To better understand mutations rate and their nature (homozygous and heterozygous) in both DER and EHR populations, random individuals from each population were sequenced. In the DER population, the homozygous (R/R) form of the Trp574 to Leu substitution was found in four out of 15 individuals, two of these individuals exhibited heterozygosity (R/S) and the rest were homozygous for the ALS-sensitive allele (S/S). Out of 12 individual plants from the EHR population, six were found to be homozygous (R/R) for the Pro197 to Ser substitution, four exhibited heterozygosity (R/S), and two were homozygous for the ALS-sensitive allele (S/S). Differences in allelic frequency between both resistant populations can also be correlated with the variation in response to ALS inhibitors. In most studies, mutations in Trp574 confers high level of resistance to all ALS inhibitors [24–27], while mutations in position Pro197 are mainly associated with high level of resistance to SU herbicides [18,28,29]. The fact that resistance to IMI and SCT herbicides was higher in DER, while resistance to SU was higher in EHR can be associated with the different target site mutations detected in the *ALS* gene sequence of plants from both populations. It can be suggested that the observed differences in resistance levels and the abundance of mutations are characteristic of a weed population in a transitional stage toward homogenic resistance.

a	570 580
<i>Arabidopsis thaliana</i> X51514.1
<i>Diplotaxis erucoides</i> (DES)	LLLNQHLGMVMQWEDRFYKANRAHT
<i>Diplotaxis erucoides</i> (DER)L.....
<i>Diplotaxis erucoides</i> (DER)X.....
b	190 200
<i>Arabidopsis thaliana</i> X51514.1
<i>Erucaria hispanica</i> (EHS)	DALLDSVPLVAITGQVPRRMIGTDA
<i>Erucaria hispanica</i> (EHR)	..M.....
<i>Erucaria hispanica</i> (EHR)	..M.....S.....
<i>Erucaria hispanica</i> (EHR)	..M.....X.....

Figure 3. Alignment of partial *ALS* gene sequence of plants from S and R populations of *Diplotaxis erucoides* and *Erucaria hispanica* (DES, DER, EHS, and EHR, respectively). Amino acid substitutions of Trp574 to Leu in DER plants (a) and Pro197 to Ser in EHR plants (b). Positions refer to the known *ALS* gene sequence of *A. thaliana* (X51514).

3.3. Alternative Management Using Auxinic Herbicides

Several chemicals that confer auxinic activity are used as herbicides [30]. Epinasty, shoot reorientation, curly leaf, and early senescence have been documented as plant response to auxinic herbicide activity [31,32]. When treated with auxinic herbicides, 2,4-D, or mecoprop-P, ALS-resistant *D. erucoides* and *E. hispanica* plants were severely and rapidly damaged, and no survivors were detected at any tested rate. Even at the rate of 1/4X, none of the treated plants survived (Figure 4). Auxinic herbicides may be used to control ALS-resistant individuals to contain and reduce the problem we currently face, preventing further buildup of resistance, such as in the case of *Raphanus raphanistrum* [33] in Australia, *Kochia scoparia* [34] in Western Canada, and *Amaranthus tuberculatus* [35] in the USA.

To reduce the probability of evolution of resistance to auxinic herbicides, proper crop rotation, as well as other components of integrated weed management, should be applied.

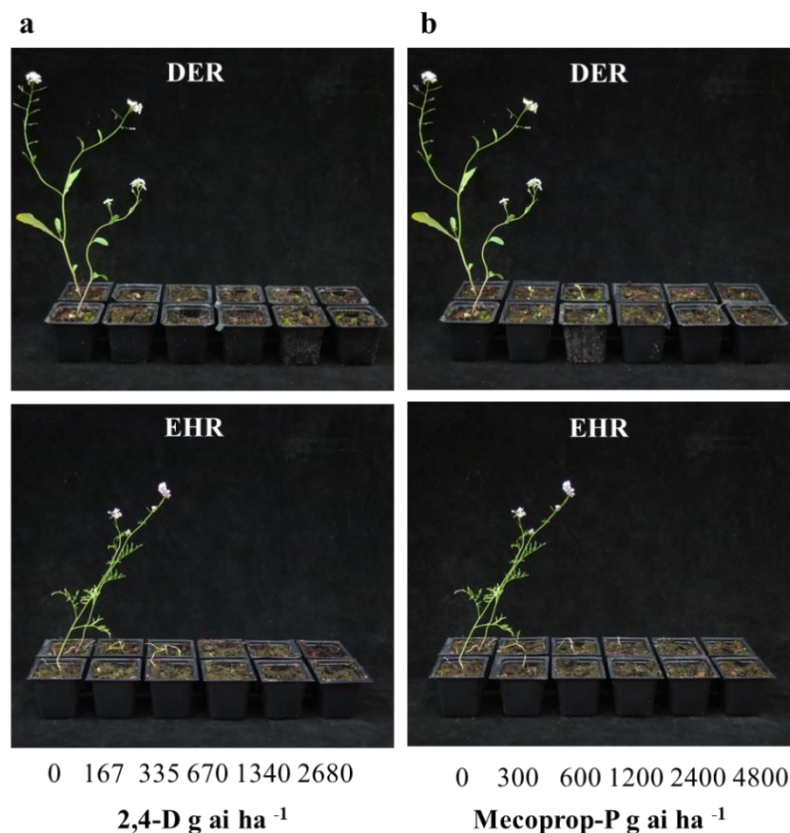


Figure 4. Effect of increasing rates of two auxinic herbicides on survival and shoot fresh weight of plants from *D. erucoides* (DER) and *E. hispanica* (EHR) ALS-resistant populations. Recommended rate for Mecoprop-P (a) and 2,4-D (b) are 670 and 1200 g ai ha⁻¹, respectively.

As was shown before, the over-use of one herbicide can eventually lead to resistance to several other herbicides from the same MOA [36,37]. According to the records of herbicide applications received from the farmers, extensive and prolonged use of tribenuron-methyl and iodosulfuron was documented in fields where DER and EHR populations were found. The evolution of ALS-resistant populations of *D. erucoides* and *E. hispanica* in pea and wheat fields may be associated with the repeated use of ALS inhibitors.

4. Conclusions

Populations of *D. erucoides* and *E. hispanica* collected from pea and wheat fields were found to be resistant to ALS inhibitors. In both populations, resistance was found to be associated with target site mutations in the *ALS* gene sequence. Auxinic herbicides provided excellent control of plants from both ALS-resistant populations. According to the record of herbicide applications received from the farmers, ALS inhibitors have been extensively used in these fields, which in turn may explain the evolution of resistance to ALS inhibitors. Different resistance levels may suggest these populations are in a transitional stage toward homogenic resistance. This report should increase the awareness and alert farmers and researchers in the region and beyond in order to proactively challenge the evolution of herbicide resistance in *D. erucoides* and *E. hispanica* in arable crops.

Author Contributions: Conceptualization, M.M. and B.R.; Data curation, M.M. and O.G.; Formal analysis, M.M. and O.G.; Methodology, M.M., M.S. and B.R.; Visualization, M.M.; Writing—original draft, M.M. and B.R.;

Writing–review & editing, M.M., O.G., M.S. and B.R. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: The authors would like to thank Tom Kliper and Ziv Kleinman for their technical assistance. First (M.M.) and second (O.G.) authors are the recipient of scholarships from the Teomim Foundation, the Natan Yaffe Foundation and the Zion Cohen Foundation.

Conflicts of Interest: The authors declare no conflict of interest.

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