

Article

Ameliorative Effects of *Daphnopsis costaricensis* Extract against Oxazolone-Induced Atopic Dermatitis-Like Lesions in BALB/c Mice

Yunji Bae ^{1,†}, Taeyoung Kim ^{1,†}, Nojune Park ^{2,†}, Sangho Choi ³, Dongkeun Yi ³, Silvia Soto ⁴, Nelson Zamora ⁴, Sunam Kim ^{2,*} and Minhye Yang ^{1,*}

¹ College of Pharmacy, Pusan National University, Busan 46241, Korea

² Natural Products Research Center, Korea Institute of Science and Technology, Gangneung 25451, Korea

³ International Biological Material Research Center, Korea Research Institute of Bioscience and Biotechnology, 111 Gwahak-ro, Yuseong-gu, Daejeon 34141, Korea

⁴ Bioprospecting Research Unit, National Biodiversity Institute, Heredia 22-3100, Costa Rica

* Correspondence: snkim@kist.re.kr (S.K.); mhyang@pusan.ac.kr (M.Y.); Tel.: +82-33-650-3419 (S.K.); +82-51-513-6754 (M.Y.)

† These authors contributed equally to this work.

Abstract: The genus *Daphnopsis* has been traditionally used as a purgative, diuretic, stimulant, and psoriasis treatment. In this study, the anti-AD (atopic dermatitis) activities of the *Daphnopsis costaricensis* EtOH extract (DCE) were investigated in an oxazolone (OX)-induced mouse model of AD, and the anti-inflammatory effects of its active compounds were confirmed in PI-sensitized or IgE/DNP-BSA-sensitized RBL-2H3 cells. DCE improved the symptoms of OX-induced inflammatory dermatitis (swelling, erythema, and increased ear thickening) in OX-induced BALB/c mice ears and reduced epidermal thickness and mast cell infiltration. Eleven flavonoid compounds were isolated from DCE, and two compounds (7,8-dimethoxyflavone and 7,2'-dimethoxyflavone) significantly inhibited IL-4 overexpression in PI-induced RBL-2H3 cells and mast cell degranulation in IgE + DNP-BSA-induced RBL-2H3 cells. Our study indicates that DCE and two compounds (7,8-dimethoxyflavone and 7,2'-dimethoxyflavone) might effectively improve inflammatory and atopic skin symptoms.

Keywords: *Daphnopsis costaricensis*; atopic dermatitis; anti-inflammatory; 7,8-dimethoxyflavone; 7,2'-dimethoxyflavone; interleukin 4; β -hexosaminidase



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1. Introduction

Atopic dermatitis (AD) is a chronic skin disease caused by multiple factors. Although its main pathogenesis has not been clarified, it is known to be associated with genetic, immunological, environmental, and psychological factors [1]. Characteristic symptoms of AD are lichenification, dry skin, pruritus, eczema, and erythema [1]. In particular, itching is the major diagnostic criterion of AD, and can have severe impacts on physical and emotional life [2]. Immunologically, activation of serum immunoglobulin E (IgE) production due to Th2 cytokine (IL-4 and IL-13) overexpression is the main cause of skin lesion formation [3] because it mediates inflammation by inducing the release of histamine and prostaglandin E from various inflammatory factors and mast cells [4,5]. In addition, it also damages the skin barrier, and, thus, facilitates the entry of inflammatory mediators into skin and repeats a vicious cycle of inflammation [6]. Currently, topical corticosteroids, topical calcineurin inhibitors, and antihistamines along with moisturizers are used to suppress these immune reactions in AD [7,8]. In addition, the monoclonal antibody dupilumab, which targets IL-4R α and inhibits the biological actions of both IL-4 and IL-13, has recently been approved to treat adult patients with AD [9]. However, long-term treatment raises safety concerns of potential side effects (skin atrophy, increased percutaneous absorption, burning, and lethargy) [10,11].

Researchers are focusing on natural products to reduce the risks of conventional AD treatments [12], and various natural extracts have been reported to have the potential to treat AD [13,14]. Those described to date usually contain phenolics, such as flavonoids and coumarins, which are considered strong inhibitors of inflammatory and allergic reactions [15]. In particular, they potently inhibit the activities of inflammatory mediators such as IL-4, IgE, and proinflammatory cytokines in inflammatory animal models and cells [16,17]. Furthermore, many studies have demonstrated that natural products are safer and more effective compared to conventional chemical products [12,18]. Thus, natural products offer attractive alternatives for the treatment of AD.

Daphnopsis costaricensis is a small shrub of the family Thymelaeaceae indigenous to Costa Rica [19], and members of this genus have been used as diuretics, radical purgatives, and to treat psoriasis [20]. Previous phytochemical studies have identified secondary metabolites, including flavonoids, terpenoids, and terpenes, in this genus [20,21], but no such study has been conducted on *D. costaricensis*. Therefore, in the present study, to propose a potential natural anti-AD and anti-inflammatory substance, we investigated the effects of *D. costaricensis* EtOH extract (DCE) on AD-like lesions in an oxazolone (OX)-induced mouse model. In addition, we isolated and identified eleven compounds and examined their anti-inflammatory and anti-allergic activities on PI or IgE + DNP-BSA-induced RBL-2H3 cells.

2. Materials and Methods

2.1. Plant Material

The aerial parts of *Daphnopsis costaricensis* (Thymelaeaceae) were collected in the Osa Conservation Area, Golfo Dulce Forest Reserve, Costa Rica, in July 2013 and identified by Nelson Zamora (National Institute of Biodiversity, INBio). Voucher specimens (KRIB 0051617) are preserved at the International Biological Material Research Center (IBMRC) at the Korea Research Institute of Bioscience and Biotechnology, Daejeon, Republic of Korea.

2.2. General Experimental Procedures

JEOL 400 MHz (JEOL, Tokyo, Japan), Bruker 500 MHz (Bruker, Billerica, MA, USA), and Agilent Technologies 600 MHz instruments (Santa Clara, CA, USA) were used to obtain ^1H , ^{13}C , HMQC and HMBC NMR spectra. Sephadex LH-20 (25–100 μm ; Pharmacia, Stockholm, Sweden) and silica gel (230–400 mesh; Merck, Darmstadt, Germany) were used to perform column chromatography, and Merck precoated silica gel 60 F₂₅₄ Art. 5715 (Merck, Germany) plates were used to perform thin-layer chromatography (TLC). HR-ESI mass spectra were obtained using an Agilent Technologies 6530 Accurate-Mass Q-TOF LC/MS. Quantitative analysis was performed using a Shimadzu HPLC system (Tokyo, Japan) equipped with an SPD-20A UV/VIS detector, two LC-20AT pumps, and a CBM-20A HPLC system controller.

2.3. Extraction and Isolation

Air-dried aerial parts of *D. costaricensis* (4.4 kg) were subjected to three cycles of reflux extraction (95% EtOH (44 L) for 90 min at 30 °C and then held for 12 h at room temperature (RT)), filtered, and filtrates were combined and evaporated in vacuo at 40 °C to give DCE (225.3 g). The DCE obtained was suspended in distilled water (2 L) and partition extracted sequentially versus 4 L of *n*-hexane, ethyl acetate (EtOAc), and *n*-butanol (*n*-BuOH) to obtain 15.9 g, 13.2 g, and 32.3 g, respectively, of the corresponding extracts.

The EtOAc extract (13.2 g) was loaded to gradient silica gel column chromatography using Hexane:EtOAc (5:1 → 100% MeOH) as eluent to obtain 9 fractions (DCE1~DCE9). Fraction DCE4 (45 mg) was fractionated into three subfractions (DCE4-1~DCE4-3) using Sephadex LH-20 with MeOH. Subfraction DCE4-2 was subjected to RP HPLC (Watchers 120 ODS-BP, S-10 μm , 150 × 10 mm) at a flow rate of 2 mL/min using a UV detector (365 nm) and MeOH:H₂O (80:20) as eluant to yield 6 subfractions (DCE4-2-1~DCE4-2-6), which afforded compound **1** (0.7 mg, t_{R} 15 min), and compound **2** (0.8 mg, t_{R} 20 min).

Subfraction DCE4-2-5 was loaded to gradient silica gel column chromatography using Hexane:EtOAc (5:1 → 100% EtOAc) and afforded compounds **3** (1.9 mg) and **4** (1 mg). Fraction DCE6 (83.6 mg) was loaded to gradient silica gel column chromatography using Hexane:EtOAc (5:1 → 100% MeOH) to yield 11 fractions (DCE6-1~DCE6-11). Compound **5** (1.6 mg, t_R 40 min) and compound **6** (1.8 mg, t_R 55 min) were afforded from subfraction DCE6-7 (18 mg) by RP HPLC (Watchers 120 ODS-BP, S-10 μ m, 150 × 10 mm) using a flow rate of 2 mL/min, a 365 nm UV detector, and isocratic elution with MeOH:H₂O (60:40) as eluent. Fraction DCE8 (577.1 mg) was fractionated into 12 fractions (DCE8-1~DCE8-12) using gradient silica gel column chromatography with Hexane:EtOAc (3:1 → 100% MeOH). Subfraction DCE8-7 (103.3 mg) was subjected to Sephadex LH-20 using MeOH to afford compound **7** (1.6 mg). Subfraction DCE8-9 (93.4 mg) was loaded to gradient silica gel column chromatography using Hexane:EtOAc (7:1 → 100% MeOH) to yield 8 fractions (DCE8-9-1~DCE8-9-8). DCE8-9-4 (16.1 mg) was conducted to RP HPLC (Watchers 120 ODS-BP, S-10 μ m, 150 × 10 mm) at a flow rate of 2 mL/min using a UV detector 330 nm by isocratic elution with 0.1% formic acid in ACN:0.1% formic acid in H₂O (42:58) as eluant to afford compounds **8** (0.6 mg, t_R 40 min), **9** (1.5 mg, t_R 52 min), and **10** (0.8 mg, t_R 69 min). Subfraction DCE8-9-5 (23 mg) was conducted to RP HPLC (Watchers 120 ODS-BP, S-10 μ m, 150 × 10 mm) using a flow rate of 2 mL/min, a 330 nm UV detector by isocratic elution with 0.1% formic acid in ACN:0.1% formic acid in H₂O (36:64) to afford compound **11** (2.5 mg, t_R 31 min).

2.4. Animals

Female BALB/c mice of six-week-old were purchased from Orient Bio, Inc. (Seongnam, Korea), housed in a controlled environment (23 ± 3 °C and 55 ± 5% RH under a 12 h light/dark cycle), and allowed standard laboratory food and water ad libitum. All experimental procedures complied with the Guide for the Care and Use of Laboratory Animals issued by the National Institutes of Health (NIH publication No. 85-23, revised 2011) and were approved beforehand by the Institutional Animal Care and Use Committee of KIST (Certification No. KIST-2016-011).

2.5. Oxazolone-Induced Atopic Dermatitis BALB/c Mice

BALB/c mice were sensitized by applying 20 μ L of 1% oxazolone in a mixture of acetone and olive oil (4:1) to ear surfaces once daily for 7 days. Subsequently, AD was induced by applying 20 μ L of 0.1% oxazolone for 3 weeks every 2 days (the Oxazolone-group). During the AD induction period, mice in the DCE and Dexa groups were administered 1% DCE or 0.1% dexamethasone twice daily. Mice were divided into four groups ($n = 4$), that is, the OX, DCE, Dexa, and control (CON) groups. Animals in the CON group were administered distilled water instead of oxazolone throughout the sensitization and induction periods. On the final application day (day 28), mice were sacrificed, and samples were collected.

2.6. Histological Examination

Ear skins from BALB/c mice were fixed in 10% formaldehyde (Sigma) for 24 h, embedded in paraffin wax, serially sectioned at 4 μ m, stained with hematoxylin and eosin (H&E) or toluidine blue for general morphology and mast cell infiltration. Epidermal thickness was measured using HKBasic software (KOPTIC, Seoul, Korea), and the number of mast cells infiltrating into the dermal layer was counted by randomly selecting three sections in toluidine blue-stained tissue. Histopathological changes (×200 magnification) were evaluated using the ProgRes[®] CapturePro application software (JENOPTIK laser, Jena, Germany).

2.7. Cell Culture

RBL-2H3 (a rat basophilic leukemia cell line) cells were purchased from the American Type Culture Collection (ATCC, Rockville, MD, USA). RBL-2H3 cells were maintained

in 150 cm² cell culture dish with DMEM (Dulbecco's modified essential medium, HyClone, Logan, UT, USA) containing 10% FBS (fetal bovine serum), 100 U/mL penicillin, and 100 µg/mL streptomycin (HyClone) at 37 °C in a humidified atmosphere containing 5% CO₂.

2.8. Measurement of IL-4 mRNA Expression

RBL-2H3 cells were injected with DMSO or compounds isolated from DCE (10 µM) for 1 h, and inflammation was triggered by adding PI (PMA (phorbol 12-myristate 13-acetate, Sigma-Aldrich, St. Louis, MO, USA) at 50 ng/mL and ionomycin at 1 µM (Sigma-Aldrich), which induced states comparable to AD [22]. The control group was injected with DMSO without PI. After treatment for 20 h, total mRNA was harvested to synthesize cDNA, and IL-4 mRNA levels were calculated by quantitative real-time PCR (qPCR). Total RNA extraction was accomplished with the RNeasy mini kit (Qiagen, Valencia, CA, USA), and cDNA synthesis was conducted with the RevertAid First Strand cDNA Synthesis Kit (Thermo Fisher, Waltham, MA, USA). qPCR was performed using the QuantStudio™ 6 Pro Real-Time PCR System (Applied Biosystems, Foster City, MA, USA) and Fast SYBR® Green Master Mix (Applied Biosystems). qPCR analysis was repeated twice and desinged by duplication per sample. Expression levels of cytokines in exposed cells were compared to those in control cells at predetermined time points using the comparative cycle threshold (Ct) method. The sequences of the primers used were IL-4 forward: 5'-ACC TTG CTG TCA CCC TGT TC-3'; IL-4 reverse: 5'-TTG TGA GCG TGG ACTCAT TC-3'; β-actin forward: 5'-TCA TCA CCA TCG GCA ACG-3', β-actin reverse: 5'-TTC CT GAT GTC CAC GTC GC-3'. mRNA expressions were normalized versus β-actin.

2.9. Measurement of β-Hexosaminidase Release

RBL-2H3 cells were seeded in 24-well plates, cultured overnight, sensitized with 100 ng/mL IgE for 4 h at 37 °C in 5% CO₂. The IgE-sensitized cells were incubated with each of the eleven compounds (10 µM) isolated from DCE for 1 h, followed by 30 min incubation with 10 µg/mL DNP-BSA to stimulate degranulation. To assess β-hexosaminidase activity, the culture medium was supplemented with 10 mM poly-N-acetyl glucosamine in 0.1 M sodium citrate buffer (pH 4.5) in 96-well plates and incubated for 1 h at 37 °C. Absorbances at 405 nm were read using a microplate reader (Tecan Infinite M1000 Microplate Reader, Männedorf, Switzerland) [23].

2.10. Statistical Analysis

In vitro data are shown as means ± standard deviations (SDs) and in vivo data as mean ± standard errors of means (SEMs) (*n* = 4). Statistical analysis was performed using one-way analysis of variance (ANOVA). *p* values of < 0.05 were considered statistically significant.

3. Results

3.1. Effects of *D. costaricensis* EtOH Extract (DCE) on Oxazolone-Induced AD-Like Lesions

To verify the anti-AD effect of DCE on mouse ear skin, BALB/c mice were administered OX for four weeks. OX-induced mice showed AD-like skin symptoms, which included increases in the ear and epidermal thickness, tumefaction, erythema, and dry skin, but it was improved in the DCE group (Figure 1A). On the final application day (day 28), the animals were sacrificed and ear thickness was determined. Ear thickness on the last day of the experiment was thicker in the OX group (0.32 mm) than in the CON group (0.19 mm) (Figure 1B). However, DCE significantly reduced ear thickness (to 0.24 mm) versus the OX group. The Dexa group (0.16 mm), used as positive control, also showed reduced ear thickness versus the OX group.

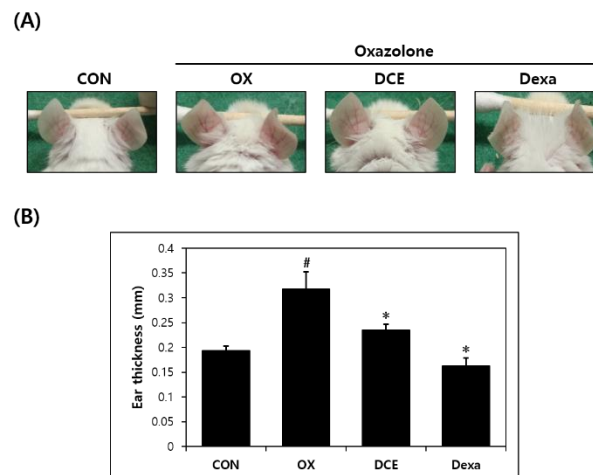


Figure 1. Effect *D. costaricensis* EtOH extract (DCE) on OX-induced AD-like lesions in the BALB/c mouse model. (A) Clinical features of AD-like symptoms. (B) Ear thickness. CON group: vehicle controls, OX group: oxazolone-treated controls, DCE group: oxazolone plus 1% DCE-treated, and Dexa group: oxazolone plus 0.1% dexamethasone-treated. Data values are presented as means \pm SEMs. [#] $p < 0.05$ vs. the CON group, ^{*} $p < 0.05$ vs. the OX group.

3.2. Histopathologic Effects of DCE on Oxazolone-Induced AD-Like Lesions

To examine the anti-AD effect of DCE on histopathological features, the epidermal tissues of mouse ears were stained with H&E or toluidine blue to determine epidermal thicknesses and degrees of mast cell infiltration. Microscopic examination revealed epidermal thickness was thinner in the DCE group than in the OX group (Figure 2A). Mast cell counts were also lower in the DCE group (Figure 3A). Oxazolone treatment increased epidermal thickness and mast cell infiltration by 2- and 4.5-fold, respectively, versus the CON group (Figures 2B and 3B). However, in the DCE group, epidermal thickness was reduced by 22%, and mast cell infiltration was significantly reduced by 45% versus the OX group. In the Dexa group, the epidermal thickness and mast cell infiltration were reduced by 65% and 63%, respectively, versus the OX group.

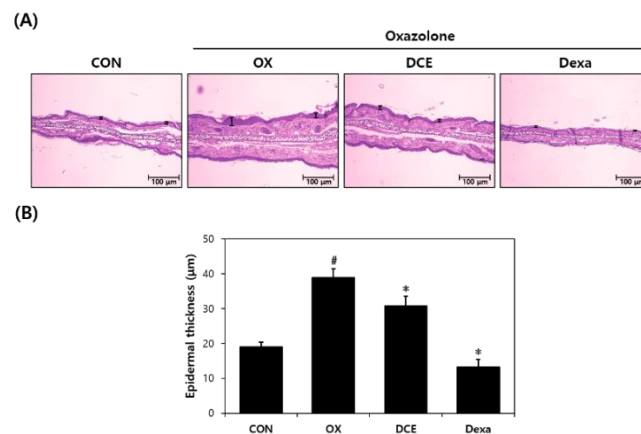


Figure 2. Effect of *D. costaricensis* EtOH extract (DCE) on H&E findings and epidermal thickness. (A) H&E staining results. (B) Epidermal thicknesses of mouse ears. CON group: vehicle controls, OX group: oxazolone-treated controls, DCE group: oxazolone plus 1% DCE-treated, and Dexa group: oxazolone plus 0.1% dexamethasone-treated. Data values are expressed as the means \pm SEMs. [#] $p < 0.05$ vs. the CON group, ^{*} $p < 0.05$ vs. the OX group.

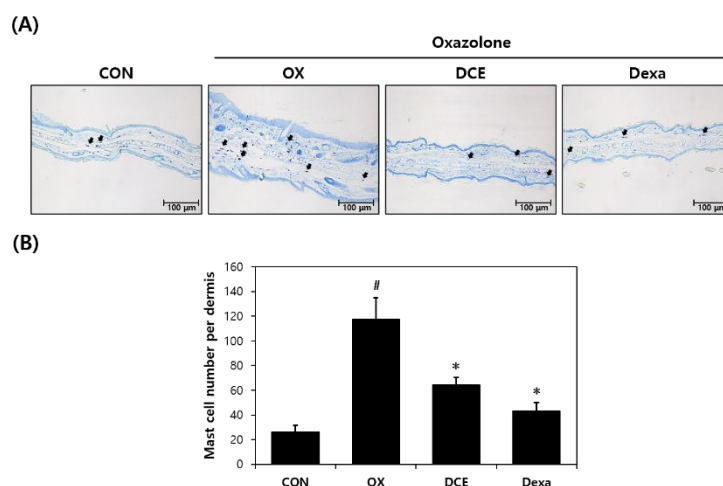
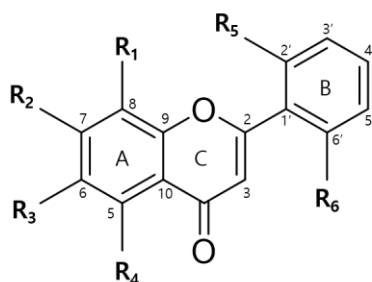


Figure 3. Effect of *D. costaricensis* EtOH extract (DCE) on toluidine blue staining findings and mast cell numbers in dermal tissues. (A) Toluidine blue staining results. (B) Mast cell numbers in dermal tissue. CON group: vehicle controls, OX group: oxazolone-treated controls, DCE group: oxazolone plus 1% DCE-treated, and Dexa group: oxazolone plus 0.1% dexamethasone-treated. Data values are expressed as means \pm SEMs. # $p < 0.05$ vs. the CON group, * $p < 0.05$ vs. the OX group.

3.3. Isolation of Compounds from DCE

Figure 4 provides details of the eleven compounds isolated from DCE as identified by 1D and 2D NMR and HR-MS data, and comparisons with the literature. The isolated compounds were identified as follows: 5,2'-dimethoxyflavone (1) [24], 6-methoxyflavone (2) [24], 6'-methoxyflavone (3) [25], 7-methoxyflavone (4) [24], 7,8-dimethoxyflavone (5) [26], 7,2'-dimethoxyflavone (6) [27], 7-hydroxyflavone (7) [28], 6,7-dimethoxyflavone (8) [29], 7,8,6'-trimethoxyflavone (9) [30], 5,7,8,2'-tetramethoxyflavone (10) [31], and 7-hydroxy-6'-methoxyflavone (11) [32].



- 1 $R_1 = H, R_2 = H, R_3 = H, R_4 = OCH_3, R_5 = OCH_3, R_6 = H$
- 2 $R_1 = H, R_2 = H, R_3 = OCH_3, R_4 = H, R_5 = H, R_6 = H$
- 3 $R_1 = H, R_2 = H, R_3 = H, R_4 = H, R_5 = H, R_6 = OCH_3$
- 4 $R_1 = H, R_2 = OCH_3, R_3 = H, R_4 = H, R_5 = H, R_6 = H$
- 5 $R_1 = OCH_3, R_2 = OCH_3, R_3 = H, R_4 = H, R_5 = H, R_6 = H$
- 6 $R_1 = H, R_2 = OCH_3, R_3 = H, R_4 = H, R_5 = OCH_3, R_6 = H$
- 7 $R_1 = H, R_2 = OH, R_3 = H, R_4 = H, R_5 = H, R_6 = H$
- 8 $R_1 = H, R_2 = OCH_3, R_3 = OCH_3, R_4 = H, R_5 = H, R_6 = H$
- 9 $R_1 = OCH_3, R_2 = OCH_3, R_3 = H, R_4 = H, R_5 = H, R_6 = OCH_3$
- 10 $R_1 = OCH_3, R_2 = OCH_3, R_3 = H, R_4 = OCH_3, R_5 = OCH_3, R_6 = H$
- 11 $R_1 = H, R_2 = OH, R_3 = H, R_4 = H, R_5 = H, R_6 = OCH_3$

Figure 4. Chemical structures of the compounds isolated from DCE.

The active compound **5** exhibits a peak at m/z 283.0974 $[M+H]^+$ in HREISMS, corresponding to its molecular formula as $C_{17}H_{14}O_4$. The 1H NMR spectrum of **5** indicated five aromatic protons of B-ring at δ_H 8.08 (m, 2H, H-2', H-6'), 7.61 (m, 3H, H-3', H-4', H-5'), and *ortho*-coupling protons of A-ring at δ_H 7.79 (d, $J = 8.9$ Hz, 1H, H-5), 7.29 (d, $J = 8.9$ Hz,

1H, H-6). The observation of a sharp singlet at δ_H 6.98 (s, 1H) with the above data and a carbonyl group at δ_C 176.6 (C-4) showed that compound **5** was a flavone. In addition, the positions of two methoxy groups were determined by correlations between δ_H 3.96 (d, 6H, OCH₃-7, OCH₃-8) and δ_C 156.5 (C-7), 136.4 (C-8) in the HMBC spectrum. Based on this spectral data, the structure of **5** was identified as 7,8-dimethoxyflavone.

The active compound **6** has a molecular formula C₁₇H₁₄O₄ as a peak at m/z 283.0968 [M+H]⁺ in HREISMS. The ¹H NMR spectrum of **6** identified 1', 2'-substituted aromatic protons of B-ring at δ_H 7.94 (m, 1H, H-6'), 7.58 (td, $J = 8.49, 7.4, 1.74$ Hz, 1H, H-4'), 7.26 (m, 1H, H-3'), 7.16 (td, $J = 7.4, 7.2, 2.4$ Hz, 1H, H-5'), and *ortho*-coupling protons [δ_H 7.94 (d, $J = 8.0$ Hz, 1H, H-5), 7.07 (dd, $J = 8.8, 2.3$ Hz, 1H, H-6)] and a singlet [δ_H 7.26 (s, 1H, H-8)] of A-ring. Moreover, a singlet at δ_H 6.86 (s, 1H, H-3) and a carbonyl group at δ_C 176.4 (C-4) proved that compound **6** was a flavone. The positions of two methoxy groups were determined by correlations between δ_H 3.93 (s, 3H, OCH₃-2') and δ_C 157.5 (C-2'), and δ_H 3.91 (s, 3H, OCH₃-7) and δ_C 163.8 (C-7), respectively, in the HMBC spectrum. On the basis of these spectral data, the structure of **6** was determined as 7,2'-dimethoxyflavone.

3.4. Compounds Isolated from DCE and Their Effects on RBL-2H3 Cells

The anti-inflammatory and anti-allergic effects of the eleven compounds isolated from DCE were assessed using IL-4 levels and β -hexosaminidase degranulation. PI-stimulation significantly increased the gene expression of IL-4 in RBL-2H3 cells. Pretreatment with 7,8-dimethoxyflavone (**5**), 7,2'-dimethoxyflavone (**6**), or 6,7-dimethoxyflavone (**8**) inhibited PI-induced increases in IL-4 levels by 46%, 35%, and 35%, respectively (Figure 5A). Furthermore, DNP-BSA-stimulation induced the release of β -hexosaminidase in IgE-sensitized RBL-2H3 cells, and 6'-methoxyflavone (**3**), 7-methoxyflavone (**4**), 7,8-dimethoxyflavone (**5**), or 7,2'-dimethoxyflavone (**6**) pretreatment inhibited β -hexosaminidase release by 17%, 16%, 27%, and 30%, respectively, versus IgE + DNP-BSA-stimulated RBL-2H3 cells (Figure 5B).

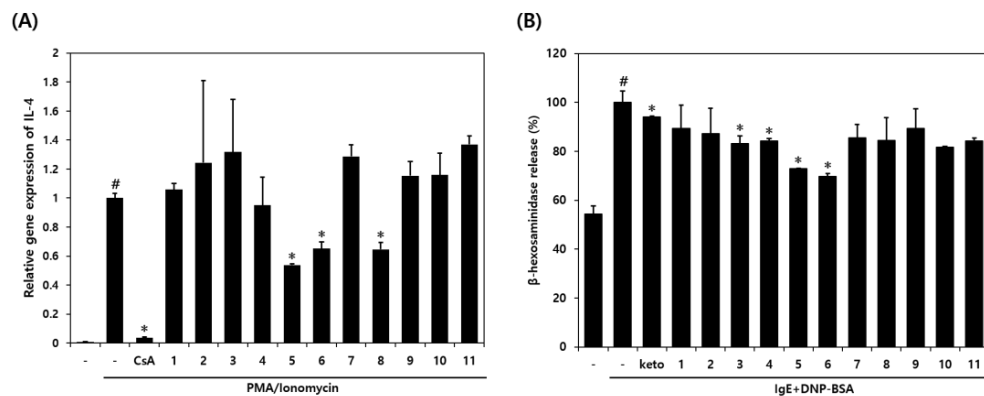


Figure 5. Anti-inflammatory and anti-allergic effects of compounds isolated from DCE on IL-4 levels and β -hexosaminidase release. (A) Expressions of IL-4. (B) β -hexosaminidase release. CsA: PI plus cyclosporin A-treated group, keto: IgE + DNP-BSA plus ketotifen-treated group. Data values are presented as the means \pm SDs. # $p < 0.05$ vs. vehicle controls, * $p < 0.05$ vs. the PI or IgE + DNP-BSA-stimulated cells.

4. Discussion

Flavonoids are extensively produced by plants and present in edible plants, fruits, legumes, and tea [33]. The number of studies performed on plant flavonoids is increasing as they have been shown to have beneficial protective effects on human health without any side effects [34]. Structurally, plant-derived flavonoids have a C6-C3-C6 carbon skeleton with different substitution patterns, as demonstrated by luteolin, apigenin, diosmetin, and quercetin [34,35]. Plant flavonoids have been shown to act as anti-inflammatory agents in epidemiologic, clinical, and animal studies [35,36]. In particular, they have been shown to regulate various inflammation-related enzyme systems and transcription factors [37], and some have been reported to have affirmative therapeutic effects on chronic inflammatory

skin diseases (atopic dermatitis, urticaria, and psoriasis) [37,38]. Accordingly, the discovery of novel flavonoids is attracting attention as a means of identifying prospective candidate anti-inflammatory drugs.

The Thymelaeaceae family has a broad range of biological activities, which include anti-inflammatory, anti-cancer, and antibacterial activities, and has been used to treat human diseases for centuries [20,21]. Plant extracts of the Thymelaeaceae family commonly contain highly diverse phenolics and possess potent anti-inflammatory properties [21]. Representatively, *Daphne* species inhibited acute and chronic inflammation in an inflammation-induced mouse model [39,40]. *Wikstroemia* species ameliorated AD-like skin lesions and decreased serum IL-4 and IgE levels in mice [41], and the genus *Daphnopsis* has been used in diuretic, laxative, and psoriasis remedies [20]. Although bioactive compounds, including flavonoids and terpenoids, with anti-inflammatory activities have been reported in the genus *Daphnopsis*, studies on the treatment of inflammatory diseases in this genus are still rare. For this reason, we undertook to characterize the anti-AD effect of DCE in an AD-induced mouse model and to examine the anti-allergic and anti-inflammatory effects of bioactive compounds isolated from DCE on inflammatory mediator degranulation in vitro.

AD is a representative allergic inflammatory skin disease. Patients with allergic inflammatory skin diseases typically exhibit symptoms such as keratinization, swelling, and erythema [42]. These inflammatory hypersensitivity reactions increase mast cell infiltration in the epidermis and thicken the epidermis [43]. Anti-AD examination of DCE via a representative oxazolone-induced BALB/c mouse model has been performed, and mice with repeated application of oxazolone exhibited erythema and edematous dermatitis along with immunological inflammatory response [44]. As a result, DCE improved AD-like lesions, severe scratching behavior, and ear thickness induced by oxazolone treatment. In the histological study, H&E and toluidine blue staining showed DCE significantly reduced mast cell infiltration and epidermal thickening. These results show that DCE has the potential to suppress allergic inflammatory symptoms in skin.

Inflammatory hypersensitivity reactions promote the differentiation of T cells and the release of various inflammatory cytokines, including IL-4, by Th2 cells [43]. Excessive IL-4 secretion degranulates immune cells, exacerbates epidermal barrier dysfunction, and causes itching [6,45]. For this reason, many authors considered IL-4 inhibitors key markers for the development of anti-inflammatory and allergy treatments [46]. Therefore, we examined the expression of mRNA IL-4 and degranulation of mast cells in PI or IgE/DNP-BSA pretreated RBL-2H3 cells subsequently treated with each of the eleven isolated flavonoids from DCE. RBL-2H3 cells have commonly been reported to be a mucosal mast cell line, and extensively used to study IgE-Fc ϵ RI interactions [47,48]. We found that 7,8-dimethoxyflavone (5), 7,2'-dimethoxyflavone (6), and 6,7-dimethoxyflavone (8) significantly decreased the expression of IL-4 mRNA. In addition, 6'-methoxyflavone (3), 7-methoxyflavone (4), 7,8-dimethoxyflavone (5), and 7,2'-dimethoxyflavone (6) showed strong inhibitory effects in the β -hexosaminidase release assay which is considered as a biomarker of mast cell degranulation. These observations support previous studies that methoxylation of the flavone A-ring 7-position enhances the anti-inflammatory activity and that multiple substituents have little effect on anti-inflammatory activity [49]. In addition, our results support earlier studies that hydroxylation of the flavone A-ring-7 position attenuates the anti-inflammatory activity, while hydroxyl groups on the flavone B-ring increase their inhibitory action [50]. Our results suggest that DCE containing flavonoids with anti-allergic and anti-inflammatory effects might be useful for the development of prophylactic and therapeutic agents for AD.

5. Conclusions

In summary, this study shows that DCE ameliorated AD-like pathology by decreasing ear epidermal thicknesses and mast cell infiltration in an oxazolone-induced BALB/c mouse model. Eleven active compounds were isolated from DCE, and 7,8-dimethoxyflavone and 7,2'-dimethoxyflavone were found to inhibit IL-4 overproduction and mast cell degranu-

lation in vitro. Accordingly, our results provide that DCE has potential use as a natural treatment for AD and chronic skin disease.

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References

1. Leung, D.Y.M. Pathogenesis of atopic dermatitis. *J. Allergy Clin. Immunol.* **1999**, *104*, 99–108. [[CrossRef](#)]
2. Koblenzer, C. Itching and the atopic skin. *J. Allergy Clin. Immunol.* **1999**, *104*, 109–113. [[CrossRef](#)]
3. Deo, S.S.; Mistry, K.J.; Kakade, A.M.; Niphadkar, P.V. Role played by Th2 type cytokines in IgE mediated allergy and asthma. *Lung India* **2010**, *27*, 66–71. [[CrossRef](#)] [[PubMed](#)]
4. Leung, D.Y.M. Immunopathology of atopic dermatitis. *Springer Semin. Immunopathol.* **1992**, *13*, 427–440. [[CrossRef](#)] [[PubMed](#)]
5. Branco, A.C.C.C.; Yoshikawa, F.S.Y.; Pietrobon, A.J.; Sato, M.N. Role of Histamine in Modulating the Immune Response and Inflammation. *Mediat. Inflamm.* **2018**, *2018*, 952407. [[CrossRef](#)] [[PubMed](#)]
6. Furue, M. Regulation of Skin Barrier Function via Competition between AHR Axis versus IL-13/IL-4-JAK-STAT6/STAT3 Axis: Pathogenic and Therapeutic Implications in Atopic Dermatitis. *J. Clin. Med.* **2020**, *9*, 3741. [[CrossRef](#)]
7. Giam, Y.C.; Hebert, A.A.; Dizon, M.V.; Van Bever, H.; Tiongco-Recto, M.; Kim, K.-H.; Soebono, H.; Munasir, Z.; Diana, I.A.; Luk, D.C.K. A review on the role of moisturizers for atopic dermatitis. *Asia Pac. Allergy* **2016**, *6*, 120–128. [[CrossRef](#)]
8. Simpson, E.L. Atopic dermatitis: A review of topical treatment options. *Curr. Med. Res. Opin.* **2010**, *26*, 633–640. [[CrossRef](#)]
9. Albader, S.S.; Alharbi, A.A.; Alenezi, R.F.; Alsaif, F.M. Dupilumab side effect in a patient with atopic dermatitis: A case report study. *Biologics* **2019**, *13*, 79–82. [[CrossRef](#)]
10. Kim, H.S.; Cho, S.H. Treatment for atopic dermatitis. *J. Korean Med. Assoc.* **2014**, *57*, 226–233. [[CrossRef](#)]
11. Berger, T.G.; Duvic, M.; Van Voorhees, A.S.; Frieden, I.J. The use of topical calcineurin inhibitors in dermatology: Safety concerns. Report of the American Academy of Dermatology Association Task Force. *J. Am. Acad. Dermatol.* **2006**, *54*, 818–823. [[CrossRef](#)] [[PubMed](#)]
12. Wu, S.; Pang, Y.; He, Y.; Zhang, X.; Peng, L.; Guo, J.; Zeng, J. A comprehensive review of natural products against atopic dermatitis: Flavonoids, alkaloids, terpenes, glycosides and other compounds. *Biomed. Pharmacother.* **2021**, *140*, 111741. [[CrossRef](#)] [[PubMed](#)]
13. Dohil, M.A. Natural ingredients in atopic dermatitis and other inflammatory skin disease. *J. Drugs Dermatol.* **2013**, *12*, 128–132.
14. Dawid-Pač, R. Medicinal plants used in treatment of inflammatory skin diseases. *Postep. Dermatol. Alergol.* **2013**, *30*, 170–177. [[CrossRef](#)]
15. Ambriz-Pérez, D.L.; Leyva-López, N.; Gutierrez-Grijalva, E.P.; Heredia, J.B. Phenolic compounds: Natural alternative in inflammation treatment. A Review. *Cogent Food Agric.* **2016**, *2*, 1131412. [[CrossRef](#)]
16. Shi, X.; Niu, L.; Zhao, L.; Wang, B.; Jin, Y.; Li, X. The anti-allergic activity of flavonoids extracted from *Citri Reticulatae* Pericarpium. *J. Food Process. Preserv.* **2018**, *42*, 13588. [[CrossRef](#)]
17. Li, Y.-J.; Guo, Y.; Yang, Q.; Weng, X.-G.; Yang, L.; Wang, Y.-J.; Chen, Y.; Zhang, D.; Li, Q.; Liu, X.-C.; et al. Flavonoids casticin and chrysosplenol D from *Artemisia annua* L. inhibit inflammation in vitro and in vivo. *Toxicol. Appl. Pharmacol.* **2015**, *286*, 151–158. [[CrossRef](#)]
18. Ullah, A.; Munir, S.; Badshah, S.L.; Khan, N.; Ghani, L.; Poulson, B.G.; Emwas, A.-H.; Jaremko, M. Important Flavonoids and Their Role as a Therapeutic Agent. *Molecules* **2020**, *25*, 5243. [[CrossRef](#)]
19. *Daphnopsis Cotaricensis* Barringer & Grayum | International Plant Names Index. Available online: <https://www.ipni.org/n/275663-2> (accessed on 8 October 1984).
20. Blaskó, G.; Xun, L.; Cordell, G.A. Studies in the Thymelaeaceae. V. 2'-Hydroxyflavone from *Daphnopsis Sellowiana*: Isolation and Synthesis. *J. Nat. Prod.* **1988**, *51*, 60–65. [[CrossRef](#)]

21. Borris, R.P.; Blaskó, G.; Cordell, G.A. Ethnopharmacologic and Phytochemical Studies of the Thymelaeaceae. *J. Ethnopharmacol.* **1988**, *24*, 41–91. [[CrossRef](#)]
22. Jin, M.; Yoon, S.J.; Pyo, M.Y. Down-regulation of T helper 2-associated cytokine expression and selective transcription factors by fisetin. *J. Appl. Biol. Chem.* **2011**, *54*, 949–958. [[CrossRef](#)]
23. Lee, E.-J.; Yu, M.-H.; Garcia, C.V.; Jhee, K.-H.; Yang, S.-A. Inhibitory effect of *Zizania latifolia* chloroform fraction on allergy-related mediator production in RBL-2H3 cells. *Food Sci. Biotechnol.* **2017**, *26*, 481–487. [[CrossRef](#)] [[PubMed](#)]
24. Lee, S.H.; Moon, B.H.; Park, Y.H.; Lee, E.J.; Hong, S.W.; Lim, Y.H. Methyl Substitution Effects on ¹H and ¹³C NMR Data of Methoxyflavones. *Bull. Korean Chem. Soc.* **2008**, *29*, 1793–1796. [[CrossRef](#)]
25. McKendall, M.; Smith, T.; Anh, K.; Ellis, J.; McGee, T.; Foroozesh, M.; Zhu, N.; Stevens, C.L.K. Methoxyflavone Inhibitors of Cytochrome P450. *J. Chem. Crystallogr.* **2008**, *38*, 231–237. [[CrossRef](#)]
26. Herath, W.; Mikell, J.R.; Khan, I.A. Microbial metabolism. Part 10: Metabolites of 7,8-dimethoxyflavone and 5-methoxyflavone. *Nat. Prod. Res.* **2009**, *23*, 1231–1239. [[CrossRef](#)] [[PubMed](#)]
27. Lee, J.-I.; Son, H.-S.; Park, H. An efficient synthesis of flavones from 2-hydroxybenzoic acids. *Bull. Korean Chem. Soc.* **2004**, *25*, 1945–1947. [[CrossRef](#)]
28. Aksnes, D.W.; Standnes, A.; Andersen, Ø.M. Complete Assignment of the ¹H and ¹³C NMR Spectra of Flavone and Its A-Ring Hydroxyl Derivatives. *Magn. Reson. Chem.* **1996**, *34*, 820–823. [[CrossRef](#)]
29. Yoon, H.; Eom, S.L.; Hyun, J.Y.; Jo, G.H.; Hwang, D.S.; Lee, S.H.; Yong, Y.J.; Park, J.C.; Lee, Y.H.; Lim, Y.H. ¹H and ¹³C NMR Data on Hydroxy/Methoxy Flavonoids and the Effects of Substituents on Chemical Shifts. *Bull. Korean Chem. Soc.* **2011**, *32*, 2101–2104. [[CrossRef](#)]
30. Kim, Y.S.; Keyser, S.G.L.; Schneekloth, J.S. Synthesis of 2',3',4'-trihydroxyflavone (2-D08), an inhibitor of protein sumoylation. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 1094–1097. [[CrossRef](#)]
31. Bhardwaj, D.K.; Gupta, A.K.; Chand, R.; Jain, K. Syntheses of *Andrographis paniculata* flavones. *Curr. Sci.* **1981**, *50*, 750–753.
32. Laget, M.; De Méo, M.; Wallet, J.C.; Gaydou, E.M.; Guiraud, H.; Duménil, G. Antimutagenic Activities of 24 Synthetic Flavones with the *Salmonella* Microsomal Assay. *Arch. Pharm. Res.* **1995**, *18*, 415–422. [[CrossRef](#)]
33. Panche, A.N.; Diwan, A.D.; Chandra, S.R. Flavonoids: An overview. *J. Nutr. Sci.* **2016**, *5*, 47. [[CrossRef](#)] [[PubMed](#)]
34. Karak, P. Biological activities of flavonoids: An overview. *Int. J. Pharm. Sci. Res.* **2019**, *10*, 1567–1574.
35. Singh, M.; Kaur, M.; Silakari, O. Flavones: An important scaffold for medicinal chemistry. *Eur. J. Med. Chem.* **2014**, *84*, 206–239. [[CrossRef](#)]
36. Lee, D.; Park, J.; Choi, J.; Jang, H.; Seol, J. Anti-inflammatory effects of natural flavonoid diosmetin in IL-4 and LPS-induced macrophage activation and atopic dermatitis model. *Int. Immunopharmacol.* **2020**, *89*, 107046. [[CrossRef](#)]
37. Maleki, S.J.; Crespo, J.F.; Cabanillas, B. Anti-inflammatory effects of flavonoids. *Food Chem.* **2019**, *299*, 125124. [[CrossRef](#)] [[PubMed](#)]
38. Gębka, N.; Adamczyk, J.; Gębka-Kępińska, B.; Mizgala-Izworska, E. The role of flavonoids in prevention and treatment of selected skin diseases. *J. Pre Clin. Clin. Res.* **2022**, *16*, 99–107. [[CrossRef](#)]
39. Moshiaşvili, G.; Tabatadze, N.; Mshvildadze, V. The genus *Daphne*: A review of its traditional uses, phytochemistry and pharmacology. *Fitoterapia* **2020**, *143*, 104540. [[CrossRef](#)]
40. Yeşilada, E.; Taninaka, H.; Takaishi, Y.; Honda, G.; Sezik, E.; Momota, H.; Ohmoto, Y.; Taki, T. In vitro inhibitory effects of ssp. on inflammatory cytokines and activity-guided isolation of active constituents. *Cytokine* **2001**, *13*, 359–364. [[CrossRef](#)]
41. Jegal, J.; Park, N.-J.; Kim, T.-Y.; Choi, S.; Lee, S.W.; Hang, J.; Kim, S.-N.; Yang, M.H. Effect of Topically Applied *Wikstroemia Dolichantha* Diels on the Development of Atopic Dermatitis-Like Skin Symptoms in Mice. *Nutrients* **2019**, *11*, 914. [[CrossRef](#)]
42. Guttman-Yassky, E.; Nograles, K.E.; Krueger, J.G. Contrasting pathogenesis of atopic dermatitis and psoriasis—Part I: Clinical and pathologic concepts. *J. Allergy Clin. Immunol.* **2011**, *127*, 1110–1118. [[CrossRef](#)] [[PubMed](#)]
43. Hong, S.-W.; Kim, M.-R.; Lee, E.-Y.; Kim, J.H.; Kim, Y.-S.; Jeon, S.G.; Yang, J.-M.; Lee, B.-J.; Pyun, B.-Y.; Gho, Y.S.; et al. Extracellular vesicles derived from *Staphylococcus aureus* induce atopic dermatitis-like skin inflammation. *Allergy* **2011**, *66*, 351–359. [[CrossRef](#)] [[PubMed](#)]
44. Jin, H.; He, R.; Oyoshi, M.; Geha, R.S. Animal models of atopic dermatitis. *J. Investig. Dermatol.* **2009**, *129*, 31–40. [[CrossRef](#)]
45. Fujii, M. Current Understanding of Pathophysiological Mechanisms of Atopic Dermatitis: Interactions among Skin Barrier Dysfunction, Immune Abnormalities and Pruritus. *Biol. Pharm. Bull.* **2020**, *43*, 12–19. [[CrossRef](#)] [[PubMed](#)]
46. Tanaka, T.; Takahashi, R. Flavonoids and asthma. *Nutrients* **2013**, *5*, 2128–2143. [[CrossRef](#)]
47. Kinet, J.-P.; Blank, U.; Brini, A.; Jouvin, M.-H.; Küster, H.; Mejan, O.; Ra, C. The high-affinity receptor for immunoglobulin E: A target for therapy of allergic diseases. *Int. Arch. Allergy Appl. Imm.* **1991**, *94*, 51–55. [[CrossRef](#)]
48. Oliver, J.M.; Seagrave, J.; Stump, R.F.; Pfeiffer, J.R.; Deanin, G.G. Signal transduction and cellular response in RBL-2H3 mast cells. *Chem. Immunol. Allergy* **1988**, *42*, 185–245.
49. Wang, X.; Cao, Y.; Chen, S.; Lin, J.; Bian, J.; Huang, D. Anti-inflammation Activity of Flavones and Their Structure–Activity Relationship. *J. Agric. Food Chem.* **2021**, *69*, 7285–7302. [[CrossRef](#)]
50. Matsuda, H.; Morikawa, T.; Ando, S.; Toguchida, I.; Yosikawa, M. Structural requirements of flavonoids for nitric oxide production inhibitory activity and mechanism of action. *Bioorg. Med. Chem.* **2003**, *11*, 1995–2000. [[CrossRef](#)]