

Review

Ethnobotanical, Phytochemical, and Pharmacological Properties of the Subfamily Nepetoideae (Lamiaceae) in Inflammatory Diseases

Nancy Ortiz-Mendoza ^{1,2} , Martha Juana Martínez-Gordillo ^{3,*} , Emmanuel Martínez-Ambriz ⁴ , Francisco Alberto Basurto-Peña ⁵, María Eva González-Trujano ⁶  and Eva Aguirre-Hernández ^{1,*}

- ¹ Laboratorio de Productos Naturales, Departamento de Ecología y Recursos Naturales, Facultad de Ciencias, Universidad Nacional Autónoma de México, Mexico City 04510, Mexico; nancy_om@ciencias.unam.mx
- ² Posgrado en Ciencias Biológicas, Unidad de Posgrado, Ciudad Universitaria Coyoacán, Edificio D, 1° Piso, Circuito de Posgrados, Mexico City 04510, Mexico
- ³ Departamento de Biología Comparada, Herbario de la Facultad de Ciencias, Universidad Nacional Autónoma de México, Mexico City 04510, Mexico
- ⁴ Instituto de Ecología, A.C., Red de Biodiversidad y Sistemática, Xalapa 91073, Veracruz, Mexico; emmanuel.martinez@inecol.mx
- ⁵ Jardín Botánico, Instituto de Biología, Universidad Nacional Autónoma de México, Mexico City 04510, Mexico; abasurto@ib.unam.mx
- ⁶ Laboratorio de Neurofarmacología de Productos Naturales, Dirección de Investigaciones en Neurociencias, Instituto Nacional de Psiquiatría Ramón de la Fuente Muñiz, Mexico City 14370, Mexico; evag@imp.edu.mx
- * Correspondence: mjmg@ciencias.unam.mx (M.J.M.-G.); eva_aguirre@ciencias.unam.mx (E.A.-H.)



Citation: Ortiz-Mendoza, N.; Martínez-Gordillo, M.J.; Martínez-Ambriz, E.; Basurto-Peña, F.A.; González-Trujano, M.E.; Aguirre-Hernández, E. Ethnobotanical, Phytochemical, and Pharmacological Properties of the Subfamily Nepetoideae (Lamiaceae) in Inflammatory Diseases. *Plants* **2023**, *12*, 3752. <https://doi.org/10.3390/plants12213752>

Academic Editor: Adriana Basile

Received: 8 September 2023

Revised: 15 October 2023

Accepted: 31 October 2023

Published: 2 November 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Nepetoideae is the most diverse subfamily of Lamiaceae, and some species are well known for their culinary and medicinal uses. In recent years, there has been growing interest in the therapeutic properties of the species of this group regarding inflammatory illnesses. This study aims to collect information on traditional uses through ethnobotanical, pharmacological, and phytochemical information of the subfamily Nepetoideae related to inflammatory diseases. UNAM electronic resources were used to obtain the information. The analysis of the most relevant literature was compiled and organised in tables. From this, about 106 species of the subfamily are traditionally recognised to alleviate chronic pain associated with inflammation. Pharmacological studies have been carried out *in vitro* and *in vivo* on approximately 308 species belonging to the genera *Salvia*, *Ocimum*, *Thymus*, *Mentha*, *Origanum*, *Lavandula*, and *Melissa*. Phytochemical and pharmacological evaluations have been performed and mostly prepared as essential oil or high polarity extracts, whose secondary metabolites are mainly of a phenolic nature. Other interesting and explored metabolites are diterpenes from the abietane, clerodane, and kaurane type; however, they have only been described in some species of the genera *Salvia* and *Isodon*. This review reveals that the Nepetoideae subfamily is an important source for therapeutics of the inflammatory process.

Keywords: inflammation; Lamiaceae; Nepetoideae; secondary metabolites

1. Introduction

Within the Angiosperms, the Lamiaceae is the sixth most diverse family worldwide. It is divided into 12 subfamilies, where Nepetoideae stands out due to its diversity. Based on molecular and morphological data, it is a monophyletic subfamily, with approximately 123 genera and 3685 species [1,2]. Morphologically, Nepetoideae is characterised by herbaceous, shrubby, or rarely arboreal individuals; they are generally aromatic as they contain a diversity of terpenoids and the presence of the well-known rosmarinic acid [3,4]. Many species belonging to genera of this subfamily are known for their medicinal and culinary uses as condiments, e.g., *Ocimum*, *Origanum*, *Thymus*, *Salvia*, *Melissa*, and *Lavandula*, among others [5–8]. Other genera of economic importance are *Perilla* and *Prunella*. *Perilla frutescens*

is used as a condiment in oriental cuisine and *Prunella vulgaris* is outstanding for its medicinal, culinary, and ornamental uses, respectively [9,10]. On the other hand, the need for alternative and complementary therapies to treat several health problems around the world is continuous. Relevant areas looking for relevant information on these issues include agriculture and biological sciences in the fields of biochemistry, genetics, and molecular biology, as well as in the fields of pharmacology, toxicology, and pharmaceuticals. Thus, certain countries, such as India and China, have contributed almost 25% of the publications in this research area [11,12]. An interesting topic is inflammation, which is a complex set of sequential tissue changes to eliminate the initial cause of cellular injury; it presents various signs, such as local redness, swelling, pain, heat, and loss of function [13,14]. In the fields of pharmacology, toxicology, and pharmaceuticals, therapies to treat inflammation are frequently mentioned, as the inflammatory process is implicated in a wide variety of physical and mental diseases that dominate current morbidity and mortality worldwide [15,16]. At the molecular level, several chemical substances are implicated in reducing or increasing inflammation. Frequently, certain cellular stimuli trigger inflammatory processes through the release of proinflammatory cytokines and chemokines (TNF, IL-1 β , IL-22, IL-17, IFN- γ , among others). Cytokines can activate endothelial cells and acute phase protein synthesis and recruit immune system cells that play a crucial role in phagocytosis and pathogen destruction. Once the cells of the immune system are activated, they release cytokines and stimulate the release of prostaglandins that mediate a series of signs and symptoms of this process [17]. Finally, for the closure of the inflammatory response, cytokines, such as IL-10, IL-37, and TGF- β , can largely suppress this mechanism and return to homeostasis; if the anti-inflammatory response is not very pronounced, it can lead to vulnerability [18,19]. Currently, glucocorticoids and non-steroidal anti-inflammatory drugs are the most commonly used therapies in the clinic to treat problems related to inflammation. These drugs provide pain relief to the patient. The main mechanism of action of these drugs is the inhibition of prostaglandins and other proteins released in the inflammatory process [20]. However, there is enough evidence demonstrating the risk of myocardial infarction, heart failure, kidney failure, and arterial hypertension as part of the common adverse effects [21,22]. Due to these inconveniences, research on medicinal plants is relevant because they are a large potential reservoir of active metabolites with fewer harmful effects. The genera and species of the subfamily Nepetoideae (Figure 1) are interesting and important for their useful properties. Therefore, this review aims to compile some of the medicinal uses attributed to species belonging to the subfamily Nepetoideae, as well as the pharmacological properties and phytochemical analysis to isolate bioactive metabolites responsible for the anti-inflammatory activity and mechanism of action, reinforcing them as a potential therapy to improve health.

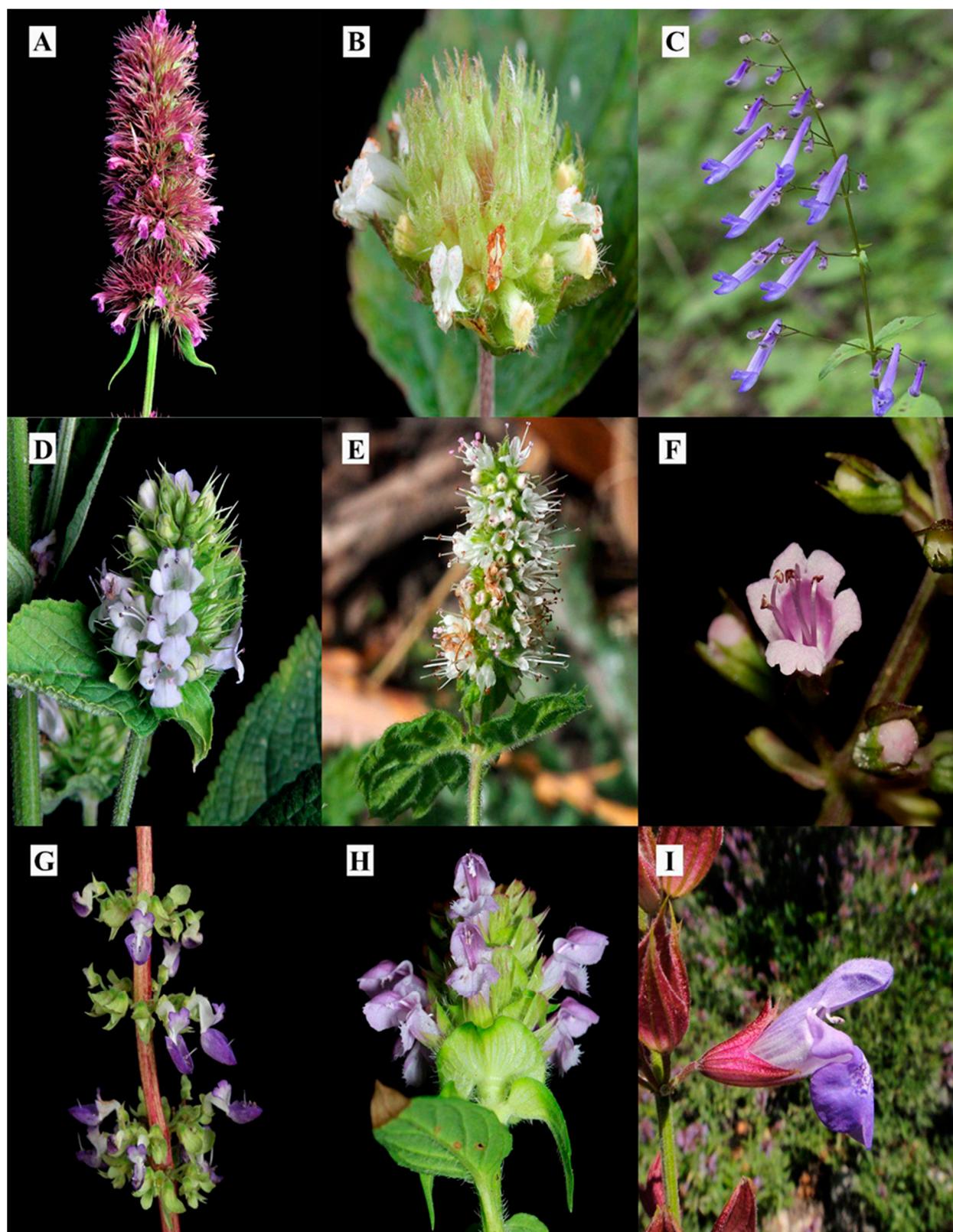


Figure 1. Selected species of the subfamily Nepetoideae (Lamiaceae) that have been studied for their anti-inflammatory properties. (A) *Agastache mexicana*, (B) *Hyptis atrorubens*, (C) *Isodon effusus*, (D) *Lepechinia caulescens*, (E) *Mentha spicata*, (F) *Ocimum carnosum*, (G) *Plectranthus scutellarioides*, (H) *Prunella vulgaris*, (I) *Salvia officinalis*. Photo credits: Canek Ledesma (A,D); Jonathan Amith (B,H); Toshihiro Nagata (C); Fred Melgert (E); Miriam Jiménez (F); Francisco Osbel (G); Christian Berg (I).

2. Results and Discussion

2.1. Ethnobotanical Information

According to the Encyclopedic Dictionary of Traditional Mexican Medicine, inflammation is a synonym for swelling; it is caused by blows, infections, rheumatism, local pain, heat, and redness, among others. In traditional medicine, inflammation is almost always understood as a sign or symptom present in various diseases and rarely as a condition [23]. Thus, the search for literature describing traditional uses related to the inflammatory process include 33 genera and 106 species of the Nepetoideae subfamily worldwide (Table S1). Regions where they are used include Chile, Brazil, Ecuador, Nicaragua, Panama, Costa Rica, Mexico, USA, and Canada in the American continent [24–31], Spain, Greece, Italy, Turkey, Algeria, Libya, Morocco, Mauritania, Tunisia, and Israel from the Mediterranean Sea, and China, India, Nepal, and Pakistan from the Himalayan region [32–40]. Other regions include the Philippines, Malaysia, and Thailand from Southeast Asia, and some islands, such as Monserrat, Samoa, and Madagascar [41–46] (Figure 2).

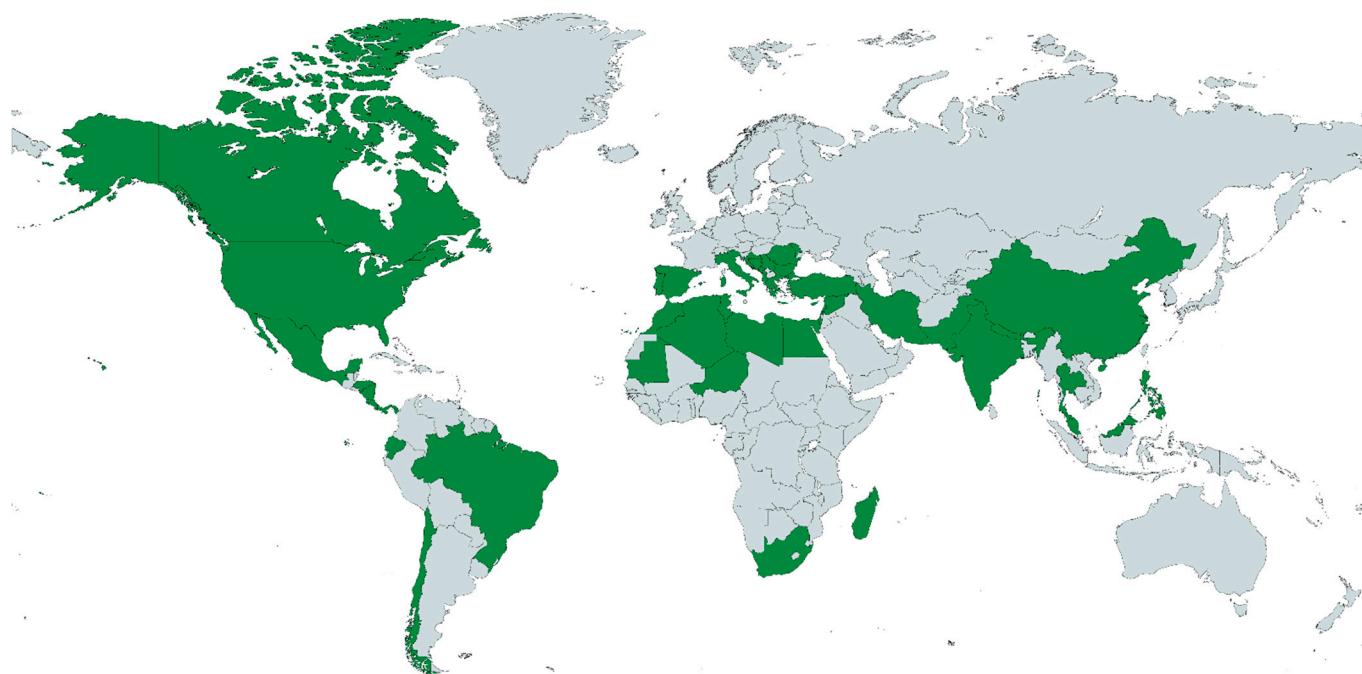


Figure 2. Countries included in reports of traditional uses related to the inflammatory process of species of the subfamily Nepetoideae are shown in green.

The genera with the most reports of traditional uses for inflammation-related conditions are *Mentha*, *Ocimum*, and *Salvia*. The most mentioned species was *Mentha longifolia* (L.) Hudson, *M. spicata* L., *Ocimum sanctum* L., and *Salvia rosmarinus* L. (Figure 3) (Table S1). *M. longifolia* and *M. spicata* are used for inflammation of the throat, gums, and eyes. *O. sanctum* is used in India to treat everything from constipated flu to chronic pain. Meanwhile, *S. rosmarinus* is used against neuritis, rheumatism, and uterine fibrosis [33,36,37,47]. The genera distributed in the American continent are *Agastache*, *Cunila*, *Hedeoma*, and *Hyptis*, all of which are used to treat various conditions, such as gastrointestinal pain and inflammation, gingivitis, blows, rheumatism, wounds, earaches, bones, and colic [23,48,49].

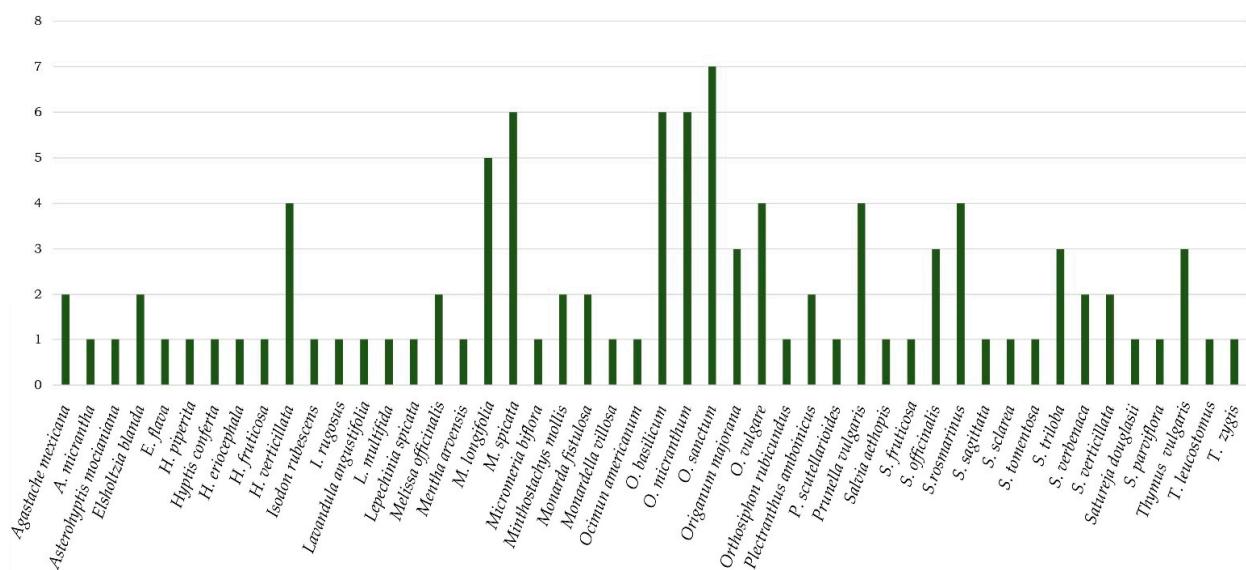


Figure 3. Number of mentions of some species belonging to the Nepetoideae subfamily for traditional uses associated with inflammatory processes.

Elsholtzia, *Isodon*, and *Orthosiphon* are widely used in Oriental medicine for skin conditions (e.g., wounds and psoriasis), the respiratory tract (tonsillitis and pharyngitis), and colic pain, respectively [50–52]. Some genera are part of the culinary culture of many countries, such as *Mentha*, *Ocimum*, *Origanum*, and *Thymus*, where they are used as condiments and for gastrointestinal ailments (diarrhea, dysentery, and colic), the respiratory system (asthma, colds, catarrh, and bronchitis), to reduce fever, to reduce swelling, and for some chronic issues, such as uterine fibrosis [32,53–55]. A highly cited genus is *Salvia*, known both in Mediterranean cuisine for its culinary use and in traditional medicine in different countries including Mexico, China, and India. Salvias are used to treat wounds, pain, infections, fever, rheumatism, uterine fibrosis, and burns, among others [31,34,52]. *Asterohyptis mociniana* (Benth.) Epling and *Isodon rubescens* (Hemsl.) H. Hara are used to treat gastroenteritis, whereas *Clinopodium brownei* Kuntze and *Plectranthus scutellarioides* Blume are used for swelling. In the cases of *Minthostachys mollis* (Kunth) Griseb. and *Monarda fistulosa* L., they have been reported for bronchitis, while *Calamintha acinos* Man. and *Lepechinia spicata* Willd. have been reported for lung problems [23,24,31,35,52,56–58]. All of these genera produce a considerable number of secondary metabolites, which alone or in synergy have beneficial biological properties for human health, making excellent functional foods, which could subsequently be developed as nutraceuticals [59].

2.2. Pharmacology

In general, it is known that polar preparations are the most commonly used medicinal plants in folk medicine, and they are included in the most representative studies explored in bioassays and phytochemical studies [60]. In the case of the Nepetoideae subfamily species, preparations commonly used in folk medicine are produced through infusion or decoction of the whole plant or other independent parts (Table S1). However, not only polar extractions but also non-polar extracts using different organic solvents and the essential oil have been reported to identify and isolate bioactive metabolites with anti-inflammatory activity. A total of 831 species of the Nepetoideae subfamily were obtained with at least one article indexed in any category in the Scopus database. From the 831 species, a total of 39 genera and 308 species were reported in 3124 articles, all of them associated with the anti-inflammatory activity (Table S2). In this regard, the genera with the most publications (100–1400) were *Salvia*, *Ocimum*, *Thymus*, *Mentha*, *Origanum*, *Lavandula*, and *Melissa* (Figure 4). It was followed by 17 genera with 10 to 100 articles (Figure 5), of which 12 belong to the Menthae tribe (*Prunella*, *Satureja*, *Zataria*, *Nepeta*, *Dracocephalum*,

Hyssopus, *Agastache*, *Glechoma*, *Ziziphora*, *Clinopodium*, *Monarda*, and *Lycopus*), 4 belong to the Ocimeae tribe (*Isodon*, *Orthosiphon*, *Plectranthus*, and *Hyptis*), and only 1 is from the Elsholtzieae tribe (*Elsholtzia*) (Figure 4).

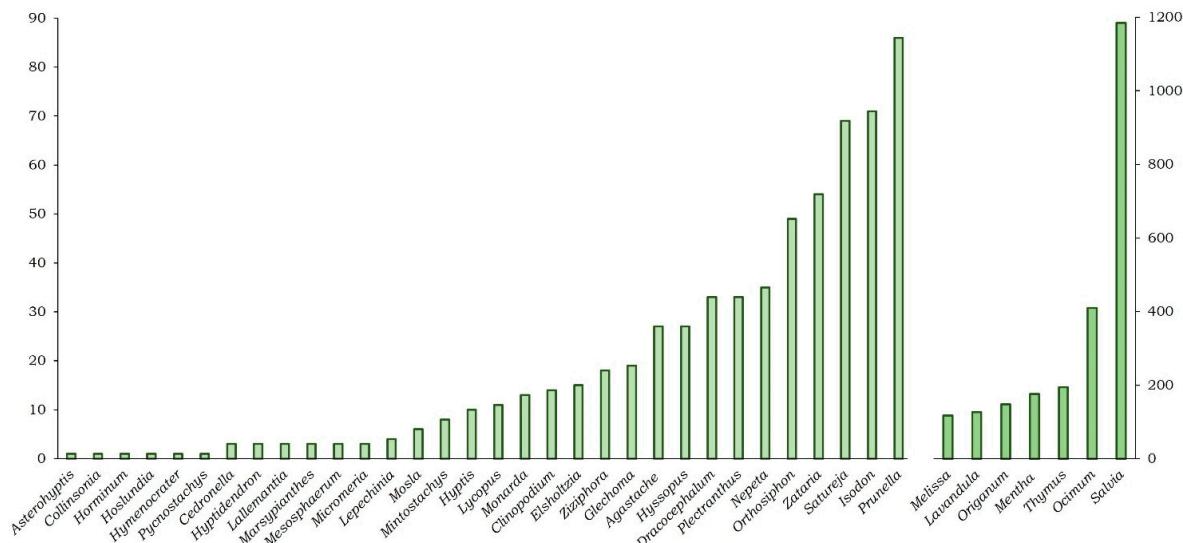


Figure 4. Number of articles related to anti-inflammatory activity indexed in Scopus for the genera of the subfamily Nepetoideae. The number of items for the genera *Melissa*, *Lavandula*, *Origanum*, *Mentha*, *Thymus*, *Ocimum*, and *Salvia* is shown on the right-hand axis, and on the left-hand axis are all of the remaining genera.



Figure 5. Number of articles related to anti-inflammatory activity indexed in Scopus in relation to the tribes of the subfamily Nepetoideae, including the genus for each tribe.

Within the genus *Salvia*, there are approximately 111 species reported in relation to the inflammatory process. The most studied species are *S. miltiorrhiza* Bunge and *S. officinalis* L. (Figure 6). For *S. miltiorrhiza*, its properties have been reported using the root extracts, where a number of isolated diterpenes called tanshinones were also identified as responsible for the effect on the reduction of proinflammatory cytokines in *in vitro*

assays [61]. The reduction of fibrosis in the liver, heart, lung, and kidney was reported in in vivo models, with improvement against allergies, asthma, and rhinitis using clinical tests [62,63]. Anti-inflammatory effects of *S. officinalis* have been described in in vivo tests prepared as organic extracts of different polarity and aqueous extracts, in which the main component was rosmarinic acid [64]. *S. dolomitica* Codd, *S. frigida* L., *S. nipponica* Miq., *S. petrophilla* G. X. Hu, E. D. Liu, and Yan Liu, *S. plebeia* R. Br., and *S. sclareoides* Brot. reduced the enzymatic activity of elastase and inflammatory mediator molecules, such as nitric oxide (NO), IL-4, IL-13, IL-5, TNF- α , cyclooxygenase (COX)-2, and prostaglandine PGE2 in RAW 264.7 macrophages induced with LPS in organic extracts of different polarities and aqueous extracts [65–70]. The polar extracts of *S. chudei* Batt. and Trab., *S. fruticosa* Mill., *S. leriiifolia* Benth., *S. macilenta* Boiss., *S. sclarea* L., *S. transsylvaniaica* (Schur ex Griseb. and Schenck) Schur, and *S. virgata* Ortega demonstrated anti-inflammatory activity in the carrageenan-induced edema model in rats at a dose range of 250–1500 mg/kg. Finally, the acetone extract and the methanol extract of *S. aegyptiaca* L. and *S. moorcroftiana* Wall. ex Benth., respectively, showed antipyretic effects in murine models of hyperthermia (Table S2) [71–78]. After the genus *Salvia*, the species *Ocimum tenuiflorum* L. (syn. *Ocimum sanctum* L.), *Thymus vulgaris* L., *Mentha spicata* L., and *Origanum vulgare* L. are the most studied of their respective genera (Figure 6). The organic and aqueous extracts of *O. tenuiflorum* showed positive effects on the epithelialization and a reduction in the induced edema in murine models. Likewise, phenolic compounds isolated from this species, including rosmarinic acid, decreased the concentration of COX-1 in in vitro models [79,80]. The aerial part of *T. vulgaris* increased the activity of antioxidant enzymes in models of renal and hepatic dysfunction in rabbits, and it also had an effect against cholestasis, chronic hepatitis, and liver fibrosis in clinic studies [81]. Both the organic and aqueous extracts of *M. spicata* decreased edema, granulomas, and mucositis induced in murine models [82]. The essential oil of *O. vulgare* produced significant effects on the proinflammatory cytokines and chemokines [83]. *Lavandula angustifolia* Mill. and *Melissa officinalis* L. have been also explored for their anti-inflammatory effects. Regarding the genus *Lavandula*, several species have been evaluated, except for *Melissa*, where only *M. officinalis* has been described. With respect to *L. angustifolia*, both the essential oil and the polyphenolic fraction reduced the levels of proinflammatory cytokines in murine models of induced edema, ischemia, chronic inflammatory pain, and sepsis [84–91]. Anti-inflammatory effects of *M. officinalis* were corroborated in a model of edema induced with carrageenan in a murine model, mainly from its ethanol and aqueous extract [92–95]. The genera explored for their anti-inflammatory effects that belong to the *Menthae* tribe are shown in Figure 5. In the case of *Prunella*, only *P. vulgaris* L. has been evaluated in this affection. The essential oil, polar extracts, as well as the isolated compounds from the inflorescences, such as the diterpene prunella diterpenol A and the phenolic compounds prunelanate A and prunelanate B, presented activity in the regulation of proinflammatory cytokines in in vitro tests [96–99]. Triterpenes, such as 2 α ,3 α ,23-trihydroxyursa-12,20(30)-dien-28-oic acid, β -amyrin, and eusapic acid, produced effects on histamine suppression in in vitro studies [100]. For the genera *Satureja*, *Zataria*, *Nepeta*, *Glechoma*, and *Lycopus*, few species have been studied in murine models using polar extracts of *Satureja montana* L. and *Nepeta dschuparensis* Bornm, which decreased IL-1 β levels in the model of traumatic brain injury and artery occlusion infarction, respectively. *Glechoma longituba* (Nakai) Kuprian attenuated proinflammatory gene expression in retinas exposed to bright light, *Lycopus lucidus* Turcz. ex Benth. inhibited histamine release in allergy models, and *Zataria multiflora* Boiss. improved levels of proinflammatory cytokines in asthma models [101–105]. From *Agastache mexicana* (Kunth) Lint and Epling and *Dracocephalum moldavica* L., which also belong to the *Mentheae* tribe, the glycosylated flavonoid tiliianin has been isolated, where a reduction in the mRNA expression of proinflammatory cytokines was observed in in vivo and in vitro models [106,107]. Likewise, some little-known diterpenes have been isolated from *D. moldavica*, such as dracocephalumoids A-E, uncinatone, trichotomone F, and caryopterisoid C, which suppressed TNF- α , IL-1 β , and NO in RAW 264.7 macrophages induced with LPS [108]. Finally, the aqueous extracts of

Hyssopus officinalis L. and *Ziziphora clinopodioides* Lam. were used for the synthesis of Zn and Fe nanoparticles, demonstrating their anti-inflammatory effect in murine models of carrageenan-induced edema and hemolytic anemia, respectively [109,110]. Diterpenoid-type compounds isolated from the aqueous extracts of species of the Ocimeae tribe have presented anti-inflammatory activity in both in vitro and in vivo models, such as the case of orthosiphon A and B obtained from *Orthosiphon stamineus* Benth., parvifloron D from *Plectranthus ecklonii* Benth., as well as suaveolol and methyl suveolate from *Hyptis suaveolens* (L.) Poit. [111–113]. Regarding the genus *Isodon*, such as *I. adenanthus* (Diels) Kudô, *I. enanderianus* (Hand.-Mazz.) H.W. Li, *I. eriocalyx* (Dunn) Kudô, *I. henryi* (Hemsl.) Kudô, *I. leucophyllus* (Dunn) Kudô, *I. rugosiformis* (Hand.-Mazz.) H. Hara, *I. scoparius* C.Y. Wu and H.W. Li H. Hara, and *I. rubescens* (Hemsl.) H. Hara, a significant amount of kaurane-type diterpenes have been isolated as responsible for the activity on proinflammatory cytokines both in RAW 264.7 macrophages induced with LPS as well as in a variety of models, such as in murine infections of encephalomyelitis, prostatitis, peritonitis, gouty arthritis, and type II diabetes [114–120]. In the case of *I. ternifolius* (D.Don) Kudô, the lignans ternifoliuslignans A, B, C, D, and E and the glycosylated phenylethanoid 3-carboxy-6,7-dihydroxy-1-(3',4'-dihydroxyphenyl)-naphthalene were characterized to suppress activity of PGE2 and TNF- α in macrophages induced with LPS [121]. *I. eriocalyx*, from which endophytic fungus *Phomopsis sp.* was first isolated, allowed the isolation of the phomopchalasins A, B, and C with NO inhibitory activity in in vitro assays [122]. Species with few studies reported in the literature were *Cedronella canariensis* (L.) Webb and Berthel., *Hoslundia opposita* Vahl, and *Micromeria biflora* (Buch. Ham. ex D.Don) Benth., prepared as chloroform extracts of flowers, root, and aerial part, respectively, which demonstrated anti-inflammatory effects in the murine model of edema induced with carrageenan [123–125]. Polar extracts of *Asterohyptis stellulata* (Benth.) Epling promoted skin regeneration in CD-1 mice. *Micromeria croatica* (Pers.) reduced the expression of proinflammatory cytokines in models of liver injury, and *Mosla chinensis* Maxim. and *M. scabra* attenuated levels of inflammatory mediators in models of ulcerative colitis [126–129]. From the petroleum ether extracts of *Horminum pyrenaicum* L., abietane-type diterpenes were isolated that suppressed the activity in immunometabolic pathways related to inflammatory processes in murine models [130] (See Table 1). All of this information together demonstrates that different classes of natural compounds are investigated for their anti-inflammatory potential properties in the species of the Nepetoideae subfamily. The chemical structure diversity found in natural products has served as an attractive approach in searching for relevant anti-inflammatory drugs, as it can be possible from this subfamily. Thus, the chemical structure diversity is not a factor in producing similar biological activity; however, the bioactivity can be improved by modifying the structure [131,132]. Due to this, it is important to notice that natural chemical compounds show a wide spectrum of activities and interaction in several molecular targets responsible for the anti-inflammatory activity of these species because it can be more than one at the same time. As an example, several compounds possessing antioxidant properties can prevent and/or reduce oxidative stress, which is relevant in inflammation and neurodegenerative diseases. The activity–structure relationship of several common polyphenols in plants, such as gallic acid reported in some species of the Nepetoideae subfamily, have demonstrated that the higher the number of phenolic hydroxyl groups, the stronger the antioxidant activity that regulates inflammatory mechanisms and pathways [132]. The interaction of some flavonoids in several targets at the same time, such as quercetin derivatives, has been reported using antagonists of inhibitory receptors, such as endogenous opioids, and those of serotonergic and/or dopaminergic neurotransmission involved in neurodegenerative diseases have been explored by using predictive molecular docking, too, in order to support their important role at peripheral and central levels [131]. Some natural products from a terpenoid nature, such as sclareol, which are also found in species of this subfamily, produced their anti-inflammatory activity by inhibiting not only NO production but also the expression of iNOS and COX-2 proteins, as well as in the MAPK signaling pathway [63]. Meanwhile, tanshinone II activity has been supported by regulating the

CCNA2-CDK2 complex and AURKA/PLK1 pathways [133]. Further structure activity relationship studies are encouraged for metabolites found in the Nepetoideae subfamily species, as in other plants, to identify not only the chemical compounds but also their mechanisms of action involved in their potential biological activities as anti-inflammatory therapy to improve health.

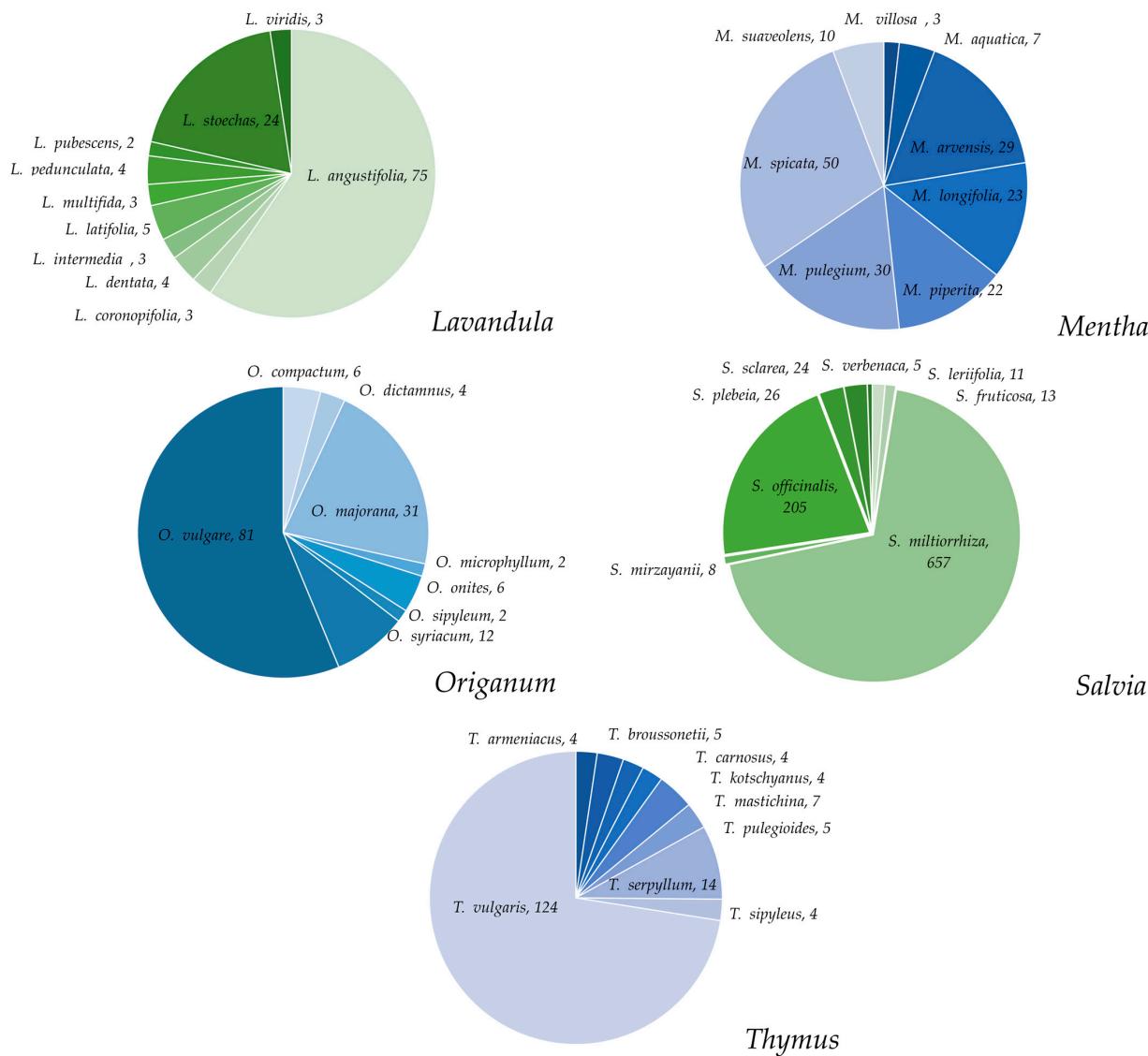


Figure 6. Species of the genera *Lavandula*, *Mentha*, *Origanum*, *Salvia*, and *Thymus* with the highest number of publications related to anti-inflammatory processes.

Table 1. Bioactive metabolites with anti-inflammatory properties isolated from species of the Nepetoideae (Lamiaceae) subfamily.

Species	Secondary Metabolites with Anti-Inflammatory Activity	References
<i>Agastache mexicana</i> Linton and Epling	Ursolic acid and limonene	[134,135]
<i>Clinopodium polycephalum</i> (Vaniot) C.Y. Wu and S.J. Hsuan	Saturol I; 3 β -22, 25-dihydroxy-tirucalla-7, 23-dieno; maslinic acid; 2 α , 3 α -dihydroxyolean-12-en-28-oic acid; hederagenin; 2 α , 3 α -dihydroxyursolic acid; alphitolic acid; rosmarinic acid; and hesperidin	[136]

Table 1. Cont.

Species	Secondary Metabolites with Anti-Inflammatory Activity	References
<i>Coleus scutellarioides</i> (L.) Benth.	Quercetin	[137,138]
<i>Dracocephalum heterophyllum</i> Benth.	Rosmarinate, luteolin, and diosmetin	[139,140]
<i>Dracocephalum kotschy</i> Boiss.	Apigenin	[141]
<i>Dracocephalum moldavica</i> L.	Tilianin, dracocephalumoid A, uncinatone, trichotomone F, and caryopterisoid C	[107,108]
<i>Dracocephalum palmatum</i> Stephan ex Willd.	Cosmosin, cymaroside, and eriodictyol	[142–144]
<i>Dracocephalum rupestre</i> Hance	Eriodictyol	[145]
<i>Elsholtzia ciliata</i> (Thunb.) Hyl.	Luteolin, caffeic acid, vitexin, pedalin, luteolin-7-O-β-D-glucopyranoside, apigenin-5-O-β-D-glucopyranoside, apigenin-7-O-β-D-glucopyranoside, chrysoeriol-7-O-β-D-glucopyranoside, 7,3'-methoxy luteolin-6-O-β-D-glucopyranoside, 5,6,4'-trihydroxy-7,3'-dimethoxyflavone, 5-hydroxy-6,7-dimethoxyflavone, 4-(E)-caffeooyl-L-threonic acid, 4-O-(E)-p-coumaroyl-L-threonic acid, and α-linolenic acid	[146–150]
<i>Elsholtzia rugulosa</i> Hemsl.	Rugulolide A, nepetoidin B, methyl rosmarinate, and syringaresinol	[151]
<i>Glechoma longituba</i> (Nakai) Kuprian.	Apigenin-7-diglucoronide	[105]
<i>Isodon adenanthus</i> (Diels) Kudô	Adenanitin	[117]
<i>Isodon amethystoides</i> (Benth.) H. Hara	Glauccalyxin A	[152]
<i>Isodon excisus</i> (Maxim.) Kudô	Inflexinol and inflexin	[153,154]
<i>Isodon henryi</i> (Hemsl.) Kudô	Rabdotoernin A and lasiodonin	[155]
<i>Isodon japonicus</i> (Burm. f.) H. Hara	Kamebanin, kamebakaurin, oridonin, kamebakaurin, effusinan C, and isodojaponin D	[156–159]
<i>Isodon melisooides</i> (Benth.) H.W. Li	Melissoidesin	[160,161]
<i>Isodon scoparius</i> C.Y. Wu and H.W. Li	Scopariusol L	[115]
<i>Isodon serra</i> (Maxim.) Kudô	Oridonin, serrin F, 14β-hydroxyrabdoestin A, serrins H-I, enanderianin N, megathyrin B, enmein, isoserrin A-I, and nodosin	[162–166]
<i>Isodon sculponeatus</i> (Vaniot) Kudô	Sculponeatin J, sculponin T, sculponeatin C, and sculponins Y	[167,168]
<i>Isodon ternifolius</i> (D.Don) Kudô	Ternifoliolignane A–D and 3-carboxy-6,7-dihydroxy-1-(3',4'-dihydroxyphenyl)-naftalene	[121]
<i>Isodon rubescens</i> (Hemsl.) H. Hara	Oridonin	[169]
<i>Isodon wikstroemioides</i> (Hand.-Mazz.) H. Hara	Iswikstroemins A–D, G, H, J, K, and macrocalyxin B	[170,171]
<i>Mentha cordifolia</i> Opiz ex Fresen.	Menthol	[172]
<i>Mentha longifolia</i> (L.) Huds.	Longifolin A and eucalyptol	[173,174]
<i>Nepeta cataria</i> Benth.	Lamiuside A and verbascoside	[175]
<i>Ocimum kilimandscharicum</i> Baker ex Gürke	Camphor and mixture of 1,8 cineole/limonene 1:1	[176]

Table 1. Cont.

Species	Secondary Metabolites with Anti-Inflammatory Activity	References
<i>Perilla frutescens</i> (L.) Britton	Ursolic acid, corosolic acid, 3-epicorosolic acid, pomolic acid, tormentic acid, hiptadienic acid, oleanolic acid, augustic acid, 3-epimaslinic acid, luteolin, monogalactosildiacilgliceroles, and rosmarinic acid	[177–183]
<i>Plectranthus ecklonii</i> Benth.	Parvifloron D, mixture of β-sitosterol, and stigmasterol (1:1)	[112]
<i>Plectranthus ornatus</i> Codd.	(11R,13E)-11-acetoxyhalima-5,13-dien-15-oic acid; 1α,6β-diacetoxyl-8α,13R-epoxy-14-labden-11-one; 1,6-di-O-acetylforskolin; 1,6-di-O-acetyl-9-deoxyforskolin, 1,6-di-O-acetylforskolin, and forskolin	[184]
<i>Prunella vulgaris</i> L.	2α, 3α, 23-trihydroxyursa-12,20(30)-dien-28-oic acid; β-amyrin, eusapic acid, 2α, 3α-dihydroxyursolic acid, prunelanate A, and pruneladiterpenol A	[96,97,99,100]
<i>Salvia digitaloides</i> Diels	Salviatalin A	[185]
<i>Salvia lachnostachys</i> Benth.	Fruticulin A	[186]
<i>Salvia miltiorrhiza</i> Bunge	Tanshinona IIA	[61]
<i>Salvia mirzayanii</i> Rech. f. and Esfand.	Teuclatriol	[187,188]
<i>Salvia petrophilla</i> G. X. Hu, E. D. Liu and Yan Liu	Petrofins A-E	[70]
<i>Salvia plebeia</i> R. Br.	8-epi-eudebeiolida C, salviplenoide A, luteoloside, nepitrin, homoplantagenin, luteolin, nepetin, hispidulin, and eupatorin	[189–192]
<i>Salvia rosmarinus</i> (L.) J.B. Walker, B.T. Drew and J.G. González (Syn.)	Carnosic acid	[193]

2.3. Phytochemistry

Chemical secondary metabolites have been identified and isolated using dissolvents of different polarity through several techniques of extraction [194]. Therefore, phytochemical techniques have allowed for the determination of the main components of some species of the subfamily Nepetoideae. This section generally describes some of the compounds commonly identified in the subfamily (mono- and tri-terpenoids, phenolic acids, and flavonoids). It also mentions the case of clerodane and kaurano-type diterpenes, which have different biological activities (see Section 2.2) and have only been isolated in certain genera, such as *Salvia* and *Isodon*. Table S3 lists the species studied phytochemically and the secondary metabolites isolated from them. Monoterpenes have been the most identified, such as thymol (C1) and carvacrol (C2), which are important components of the essential oils of *Collinsonia*, *Monarda*, *Ocimum*, *Origanum*, *Satureja*, *Thymbra*, *Thymus*, and *Zataria* [83,101,195–200]. On the contrary, there are exclusive molecules of certain genera, as exemplified by *Mentha*, where menthol (C3), menthone (C4), and eucalyptol (C5) predominate. In the case of *Lavandula* and *Melissa*, linalool and nerol have been identified as the most important constituents, respectively [94,201,202]. Different phenolic acids have been reported throughout the subfamily, such as caffeic acid, ferulic acid, gallic acid, chlorogenic acid, sinapic acid, and a high presence of rosmarinic acid (C6) [203–208]. Similarly, the presence of flavonoids and their glycosylated derivatives, such as quercetin, luteolin, and naringenin, have been very common [126,198,200–202]. In contrast, the presence of the flavonoid tiliianin (C7) has been reported only in *Agastache* and *Dracocephalum* (Figure 7) [104,105]. It is important to notice that diterpenes are the chemical group less explored throughout the subfamily. The species of *Dracocephalum taliense* Forrest and *Horminum pyrenaicum* L. included the abietanes sugiol, ferruginol, cryptojaponol, and tarol isolated from roots. *Dracocephalum* A, B, C, and D (C8–C11), and orthosiphonol A and B (C12–C13) were purified from *Dracocephalum moldavica* L. and *Orthosiphon stamineus*

Benth. [108,111,209]. Other genera, such as *Salvia* and *Isodon*, stand out for the great diversity of diterpenes that have been isolated and identified in several of their species. In the case of *Salvia*, terpenes of abietane, clerodane, labdane, and pimarane-type structures have been described. Some examples are tanshinone IIA (C14) and sclareol (C15) isolated from *S. miltiorrhiza* and *S. sclarea*, respectively [61–71]. In the case of the genus *Isodon*, compounds with a Kaurano-type skeleton predominated, and some common cases among the species were adenanthine (C16), eriocalyxin B (C17), and oridonine (C18) [114,117,169]. Finally, the presence of triterpenes, such as oleanolic acid (19), ursolic acid (20), stigmasterol (21), and their derivatives, was very common in the subfamily Nepetoideae (Figure 7) [210–212]. Chemical structures of the representative bioactive metabolites are included in Figure 7.

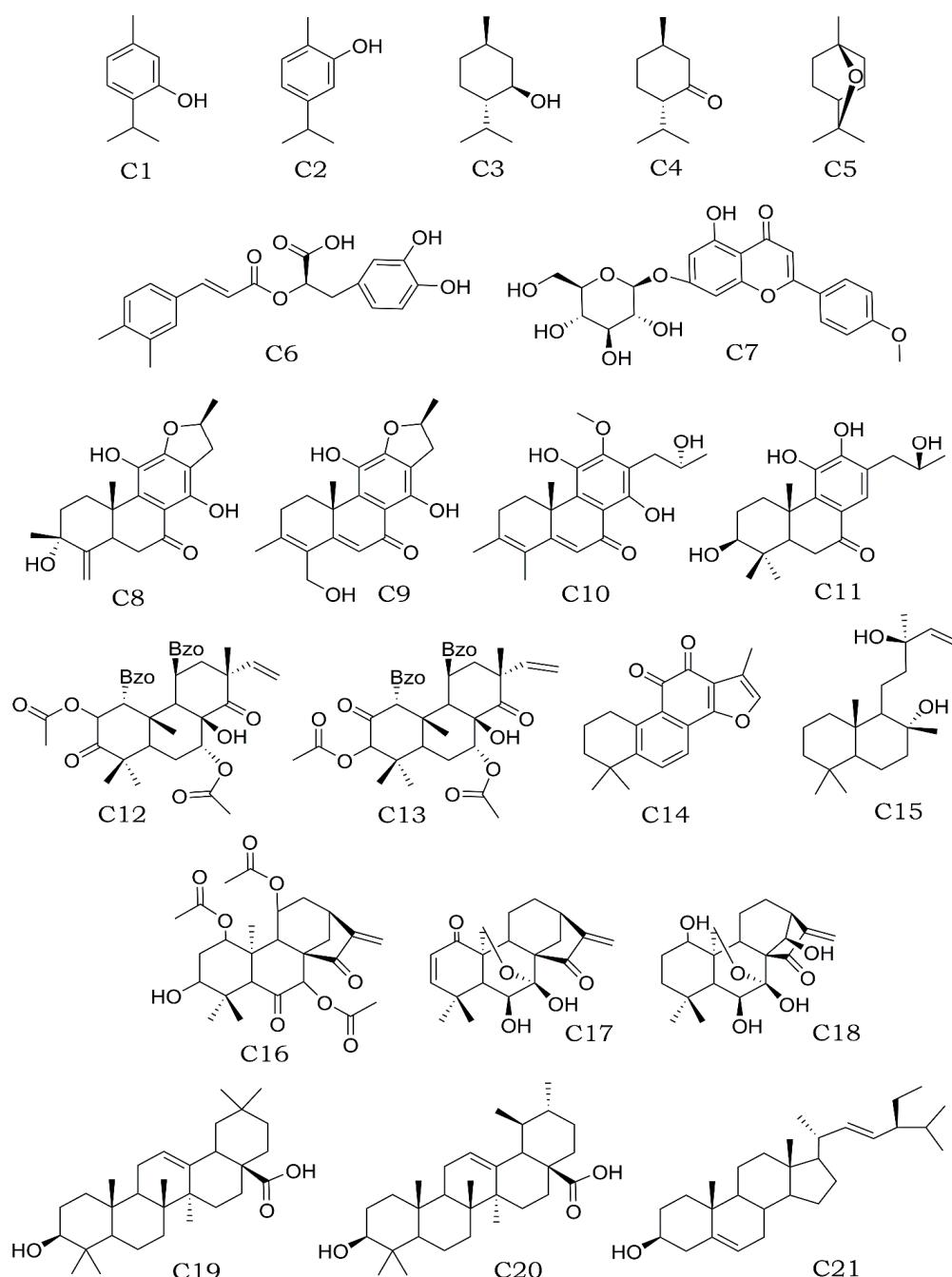


Figure 7. Representative non-polar (C1–C5 and C19–C21) and polar (C6–C18) secondary metabolites with anti-inflammatory activity isolated from species of the subfamily Nepetoideae. Thymol (C1),

carvacrol (C2), menthol (C3), menthone (C4), eucalyptol (C5), rosmarinic acid (C6), tiliatin (C7), dracocephalmoids A to D (C8–C11), orthosiphon A and B (C12–C13), tanshinone IIA (C14), sclareol (C15), adenanthine (C16), eriocalyxin B (C17), oridonine (C18), oleanolic acid (19), ursolic acid (20), and stigmasterol (21).

3. Materials and Methods

The species selected for this review, without including Mexican sage (previously reported in ref. [213]), were obtained from the Taxonomy Browser of the National Center for Biotechnology Information [214]. Thus, traditional uses were explored using the Economic Botany database and the books available at the Institute of Biology, UNAM. Then, pharmacological and phytochemistry properties were searched using the Scopus database. The literature research covers a period from the past to December 2022. The optimization of this search was made by developing a script based on the Python programming language through its interface 3.11.1 ([python.org](https://www.python.org)) and using the Application Programming Interface (API) provided by Scopus (dev.elsevier.com). For the development of the script, the following elements were used: (1) list of species belonging to the Nepetoideae subfamily, (2) SCOPUS website to access its database, (3) access credentials to the database (API-Key), (4) parameters, search algorithm, and information processing, and (5) printing of the results. The search strategy consisted of two phases. In the first, the presence/absence of information on each species of the subfamily Nepetoideae was explored in all of the indexed fields (All fields) in the Scopus database. In the second phase, the species with the presence of information obtained in the first phase were used, and the search was carried out with the following parameters: “Genus species” AND Anti-inflammatory OR Antiinflammatory (e.g., “*Zataria multiflora*” AND anti-inflammatory OR antiinflammatory, delimited to the indexed fields “Article title-Abstract-Keywords.” The articles of the species with information more related to anti-inflammatory processes and their phytochemistry were downloaded and analyzed. The results of this search were organized in tables and graphs, as above.

4. Conclusions

In this review, a total of 831 species of the subfamily Nepetoideae were obtained with at least one article indexed in any category based on a search in the Scopus database. From the 831 species, a total of 39 genera and 308 species were reported in 3124 articles, all of them associated with anti-inflammatory activity. Thus, traditional uses of the Nepetoideae subfamily species are reinforced by botanical, pharmacological, and phytochemical information described in the literature related to inflammatory illnesses and compiled in this review. The results revealed that this subfamily plays an essential role as another important source of potential anti-inflammatory drugs from natural products of different chemical natures not only at the central level, in part because of their neuroprotective activity, but also in peripheral systems. The most common extracts prepared not only for phytochemical but also for pharmacological investigation in this review were those obtained from a polar nature. Chemical secondary metabolites have been identified and isolated using dissolvents of different polarities through several techniques of extraction. These results together support medicinal use by preparing infusions or decoctions from the whole plants or from different parts of them.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/plants12213752/s1>, Table S1: Traditional uses of species from the subfamily Nepetoideae (Lamiaceae) related to the inflammatory process, Table S2: Pharmacological evaluation of the extracts related to inflammatory processes in species of the Nepetoideae (Lamiaceae) subfamily, Table S3: Phytochemistry related to inflammatory processes in species of the Nepetoideae (Lamiaceae) subfamily.

Author Contributions: Conceptualization, N.O.-M. and M.J.M.-G.; Methodology, N.O.-M. and M.J.M.-G.; Software, Formal analysis, Data curation, and Validation, N.O.-M., M.J.M.-G., F.A.B.-P., E.M.-A. and E.A.-H.; Writing—original draft preparation, N.O.-M.; Writing—review and editing, N.O.-M., M.J.M.-G., F.A.B.-P., E.M.-A., M.E.G.-T. and E.A.-H. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by grants provided by the UNAM-PAPIIT (No. IN221221). Additional funding to cover publication fees required by the National System of Researchers (SNI-CONAHCYT) was secured through grants provided to E.A.-H. (52151) and M.J.M.-G. (55788).

Acknowledgments: This paper is part of the requirement for obtaining a doctoral degree at the Posgrado en Ciencias Biológicas, UNAM of N.O.-M. The project also had the following CONAHCYT graduate scholarship number: 793655. This work was supported by UNAM-PAPIIT (No. IN221221). The authors thank Julio Javier Meraz Martínez for automating the information search through the development of a script based on the Python programming language and the usage of APIs provided by Scopus.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Gul, S.; Ahmad, M.; Zafar, M.; Bahadur, S.; Sultana, S.; Begum, N.; Shah, S.N.; Zaman, W.; Ullah, F.; Ayaz, A.; et al. Taxonomic study of subfamily *Nepetoideae* (Lamiaceae) by polynomorphological approach. *Microsc. Res. Tech.* **2019**, *82*, 1021–1031. [[CrossRef](#)]
- da Silva-Monteiro, F.K.; de Melo, J.I.M. Flora of Paraíba, Brazil: Subfamily *Nepetoideae* (Lamiaceae). *Rodriguesia* **2020**, *71*, 1–22. [[CrossRef](#)]
- Harley, R.M.S.; Atkins, A.L.; Budantsev, P.D.; Cantino, B.J.; Conn, R.; Grayer, M.M.; Harley, R.; De Kok, T.; Krestovskaja, R.; Morales, A.J.; et al. Labiateae. In *Labiatae. The Families and Genera of Vascular Plants VII. Flowering Plants Dicotyledons: Lamiales (Except Acanthaceae including Avicenniaceae)*; Kubitzki, J.K.Y., Kadereit, W., Eds.; Springer: Berlin/Heidelberg, Germany, 2004; pp. 167–275.
- Omar, S.H. Biophenols: Impacts and prospects in anti-Alzheimer drug discovery. In *Discovery and Development of Neuroprotective Agents from Natural Products*; Brahmachari, G., Ed.; Elsevier: Amsterdam, The Netherlands, 2018; pp. 103–148. [[CrossRef](#)]
- Burk, L.; O’Brien, B.; Charron, J.M.; Sherman, K.J.; Bullock, M.L. Psychic/intuitive diagnosis: Two case reports and commentary; experiences with aromatherapy in the elderly; use of *Lavandula latifolia* as an expectorant; characteristics and complaints of patients seeking therapy. *J. Altern. Complement. Med.* **1997**, *3*, 209–212. [[CrossRef](#)]
- Bornowski, N.; Hamilton, J.P.; Liao, P.; Wood, J.C.; Dudareva, N.; Buell, C.R. Genome sequencing of four culinary herbs reveals terpenoid genes underlying chemodiversity in the Nepetoideae. *DNA Res.* **2020**, *27*, 1–12. [[CrossRef](#)]
- El Yaagoubi, M.; Mechqoq, H.; el Hamdaoui, A.; Jrv Mukku, V.; el Mousadik, A.; Msanda, F.; el Aouad, N. A review on Moroccan *Thymus* species: Traditional uses, essential oils chemical composition and biological effects. *J. Ethnopharmacol.* **2021**, *278*, 114205. [[CrossRef](#)]
- Draginic, N.; Jakovljevic, V.; Andjic, M.; Jeremic, J.; Srejovic, I.; Rankovic, M.; Tomovic, M.; Nikolic Turnic, T.; Svistunov, A.; Bolevich, S.; et al. *Melissa officinalis* L. as a nutritional strategy for cardioprotection. *Front. Physiol.* **2021**, *12*, 1778. [[CrossRef](#)]
- Li, P.; Qi, Z.C.; Liu, L.X.; Ohi-Toma, T.; Lee, J.; Hsieh, T.H.; Fu, C.X.; Cameron, K.M.; Qiu, Y.X. Molecular phylogenetics and biogeography of the mint tribe Elsholtzieae (Nepetoideae, Lamiaceae), with an emphasis on its diversification in East Asia. *Sci. Rep.* **2017**, *7*, 2057. [[CrossRef](#)]
- Pan, J.; Wang, H.; Chen, Y. *Prunella vulgaris* L.—A review of its ethnopharmacology, phytochemistry, quality control and pharmacological effects. *Front. Pharmacol.* **2022**, *13*, 3171. [[CrossRef](#)]
- Elsevier, B.V. Analyze Search Results: Lamiaceae 1980–2023. Available online: www.scopus.com (accessed on 15 January 2023).
- Youn, B.Y.; Moon, S.; Mok, K.; Cheon, C.; Ko, Y.; Park, S.; Jang, B.H.; Shin, Y.C.; Ko, S.G. Use of traditional, complementary and alternative medicine in nine countries: A cross-sectional multinational survey. *Complement. Ther. Med.* **2022**, *71*, 102889. [[CrossRef](#)]
- Liu, C.H.; Abrams, N.D.; Carrick, D.M.; Chander, P.; Dwyer, J.; Hamlet, M.R.J.; Macchiarini, F.; Prabhudas, M.; Shen, G.L.; Tandon, P.; et al. Biomarkers of chronic inflammation in disease development and prevention: Challenges and opportunities. *Nat. Immunol.* **2017**, *18*, 1175–1180. [[CrossRef](#)]
- What Is an Inflammation? Institute for Quality and Efficiency in Health Care (IQWiG): Cologne, Germany, 2006. Available online: www.ncbi.nlm.nih.gov/books/NBK279298 (accessed on 12 December 2022).
- Slavich, G.M. Understanding inflammation, its regulation, and relevance for health: A top scientific and public priority. *Brain Behav. Immun.* **2015**, *45*, 13–14. [[CrossRef](#)]
- Bennett, J.M.; Reeves, G.; Billman, G.E.; Sturmberg, J.P. Inflammation—nature’s way to efficiently respond to all types of challenges: Implications for understanding and managing “the epidemic” of chronic diseases. *Front. Med.* **2018**, *5*, 316. [[CrossRef](#)]
- Netea, M.G.; Balkwill, F.; Chonchol, M.; Cominelli, F.; Donath, M.Y.; Giamarellos-Bourboulis, E.J.; Golenbock, D.; Gresnigt, M.S.; Heneka, M.T.; Hoffman, H.M.; et al. A guiding map for inflammation. *Nat. Immunol.* **2017**, *18*, 826–831. [[CrossRef](#)]

18. Ouyang, W.; Rutz, S.; Crellin, N.K.; Valdez, P.A.; Hymowitz, S.G. Regulation and functions of the IL-10 family of cytokines in inflammation and disease. *Annu. Rev. Immunol.* **2011**, *29*, 71–109. [CrossRef]
19. Dinarello, C.A.; Nold-Petry, C.; Nold, M.; Fujita, M.; Li, S.; Kim, S.; Bufler, P. Suppression of innate inflammation and immunity by interleukin-37. *Eur. J. Immunol.* **2016**, *46*, 1067–1081. [CrossRef]
20. Nunes, C.d.R.; Barreto Arantes, M.; Menezes de Faria Pereira, S.; Leandro da Cruz, L.; de Souza Passos, M.; Pereira de Moraes, L.; Vieira, I.J.C.; Barros de Oliveira, D. Plants as sources of anti-inflammatory agents. *Molecules* **2020**, *25*, 3726. [CrossRef]
21. Harirforoosh, S.; Asghar, W.; Jamali, F. Adverse effects of nonsteroidal antiinflammatory drugs: An update of gastrointestinal, cardiovascular and renal complications. *J. Pharm. Pharm. Sci.* **2013**, *16*, 821–847. [CrossRef]
22. Furman, D.; Campisi, J.; Verdin, E.; Carrera-Bastos, P.; Targ, S.; Franceschi, C.; Ferrucci, L.; Gilroy, D.W.; Fasano, A.; Miller, G.W.; et al. Chronic inflammation in the etiology of disease across the lifespan. *Nat. Med.* **2019**, *25*, 1822–1832. [CrossRef]
23. Argueta, A.; Cano, L.; Rodarte, M.; Gallardo, C. *Atlas de las Plantas de la Medicina Tradicional Mexicana*; Instituto Nacional Indigenista: Mexico City, Mexico, 1994; 1786p.
24. Core, E.L. Ethnobotany of the southern Appalachian aborigines. *Econ. Bot.* **1966**, *21*, 198–214. [CrossRef]
25. Turner, N.; Bell, M. The ethnobotany of the coast Salish Indians of Vancouver island. *Econ. Bot.* **1971**, *25*, 63–99. [CrossRef]
26. Aldunate, C.; Armesto, J.J.; Castro, V.; Villagran, C. Ethnobotany of pre-altiplanic community in the Andes of Northern Chile. *Econ. Bot.* **1983**, *37*, 120–135. [CrossRef]
27. Hazlett, D.L. Ethnobotanical observations from Cabecar and Guaymi settlements in Central America. *Econ. Bot.* **1986**, *40*, 339–352. [CrossRef]
28. Bye, R.A. Medicinal plants of the Sierra Madre: Comparative study of Tarahumara and Mexican market plants. *Econ. Bot.* **1986**, *40*, 103–124. [CrossRef]
29. Barret, B. Medicinal plants of Nicaragua’s Atlantic coast. *Econ. Bot.* **1994**, *48*, 8–20. [CrossRef]
30. Voeks, R.A. Tropical forest healers and habitat preference. *Econ. Bot.* **1996**, *50*, 381–400. [CrossRef]
31. Rios, M.; Koziol, M.; Borgtoft, H.; Granda, G. *Plantas Útiles del Ecuador*; Abya-Yala: Quito, Ecuador, 2007; p. 652.
32. Fujita, T.; Sezik, E.; Tabata, M.; Yesilada, G.; Takeda, Y.; Tanaka, T.; Takaishi, Y. Traditional medicine in Turkey VII. Folk medicine in middle and west black sea regions. *Econ. Bot.* **1995**, *49*, 406–422. [CrossRef]
33. Martínez-Lirola, M.J.; Gonzalez-Tejero, M.R.; Molero-Mesa, J. Ethnobotanical resources in the province of Almería, Spain: Campos de Níjar. *Econ. Bot.* **1996**, *50*, 40–56. [CrossRef]
34. Rivera, D.; Obón, C.; Cano, F. The botany, history and traditional uses of three-lobed sage (*Salvia fruticosa* Miller) (Labiatae). *Econ. Bot.* **1994**, *48*, 190–195. [CrossRef]
35. Brussell, D. A medicinal plant collection fromMontserrat, west indies. *Econ. Bot.* **2004**, *58*, s203–s220. [CrossRef]
36. Dagar, H.S.; Dagar, J.C. Plant folk medicines among the Nicobarese of Katchal Island, India. *Econ. Bot.* **1990**, *45*, 114–119. [CrossRef]
37. Bhattarai, N.K. Medical ethnobotany in the Karnali zone, Nepal. *Econ. Bot.* **1992**, *46*, 257–261. [CrossRef]
38. Huai, H.Y.; Pei, S.J. Plants used medicinally by folk healers of the Lahu people from the autonomous county of Jinping Miao, Yao, and Dai in southwest China. *Econ. Bot.* **2004**, *58*, 265s–273s. [CrossRef]
39. Ajaib, M.; Ishtiaq, M.; Bhatti, K.H.; Hussain, I.; Maqbool, M.; Hussain, T.; Mushtaq, W.; Ghani, A.; Azeem, M.; Khan, S.M.R.; et al. Inventorization of traditional ethnobotanical uses of wild plants of Dawarian and Ratti Gali areas of District Neelum, Azad Jammu and Kashmir Pakistan. *PLoS ONE* **2021**, *16*, e0255010. [CrossRef]
40. Lahoulou, R.A.; Samba, N.; Soeiro, P.; Alves, G.; Gonçalves, A.C.; Silva, L.R.; Silvestre, S.; Rodilla, J.; Ismael, M.I. *Thymus hirtus* Willd. ssp. *algeriensis* Boiss. and Reut: A Comprehensive Review on Phytochemistry, Bioactivities, and Health-Enhancing Effects. *Foods* **2022**, *11*, 3195. [CrossRef]
41. Anderson, E.F. Ethnobotany of hill tribes of northern Thailand. I. Medicinal plants of Akha, I. *Econ. Bot.* **1986**, *40*, 38–53. [CrossRef]
42. Cox Bodner, C.; Gereau, R.E. A contribution to Bontoc ethnobotany. *Econ. Bot.* **1988**, *42*, 307–369. [CrossRef]
43. Christensen, H. *Ethnobotany of the Iban & the Kelabit*; Oko-Tryk Press: Copenhagen, Denmark, 2002; 384p.
44. Uhe, G. Medicinal plants of Samoa a preliminary survey of the use of plants for medicinal purposes in the Samoan Islands. *Econ. Bot.* **1974**, *28*, 1–30. [CrossRef]
45. Quansah, N. Ethnomedicine in the Maroantsetra region of Madagascar. *Econ. Bot.* **1988**, *42*, 370–375. [CrossRef]
46. Brussell, D. Medicinal plants of Mt. Pelion, Greece. *Econ. Bot.* **2004**, *58*, 176s–202s. [CrossRef]
47. Weckerle, C.; Huber, F.; Yongping, Y.; Weibang, S. Plant knowledge of the Shuhi in the Hengduan mountains, southwest China. *Econ. Bot.* **2006**, *60*, 3–23. [CrossRef]
48. Gispert Cruells, M.; Rodríguez González, H. *Los Coras: Plantas Alimentarias y Medicinales de su Ambiente Natural*; CONACULTA Culturas Populares: Mexico City, Mexico, 1988; 128p.
49. Monroy-Ortiz, C.; Monroy, R. *Las plantas, Compañeras De Siempre: La Experiencia En Morelos*; Universidad Autónoma del Estado de Morelos: Cuernavaca, Mexico, 2006; 184p, ISBN 9701816145.
50. Manandhar, N.P. Medicinal plant-lore of Tamang tribe of Kabhrepalanchok district, Nepal. *Econ. Bot.* **1991**, *45*, 58–71. [CrossRef]
51. Pal, D.C.; Jain, S.K. Notes on Lodha medicine in Midnapur district, West Bengal, India. *Econ. Bot.* **1989**, *43*, 464–470. [CrossRef]
52. Chen, X.; Dai, X.; Liu, Y.; He, X.; Gong, G. *Isodon rubescens* (Hemsl.) Hara.; A comprehensive review in traditional uses, phytochemistry, and pharmacological activities. *Front. Pharmacol.* **2022**, *13*, 766581. [CrossRef] [PubMed]

53. Balick, M.J.; Kronenberg, F.; Ososki, A.L.; Fugh-Berman, A.; Roble, M.; Lohr, P.; Atha Balick, D.; Reiff, M. Medicinal plants used by Latino healers for women's health conditions in New York city. *Econ. Bot.* **2000**, *54*, 344–357. [CrossRef]
54. Dolores, B.M.; Croce, H.M.; Cerda, L.M. *Plantas Útiles de la Región Semiárida de Aguascalientes*; Universidad Autónoma de Aguascalientes: Aguascalientes, Aguascalientes, 2003.
55. Pérez-Nicolás, M.; Vibrans, H.; Romero-Manzanares, A.; Saynes-Vásquez, A.; Luna-Cavazos, M.; Flores-Cruz, M.; Lira-Saade, R. Patterns of Knowledge and Use of Medicinal Plants in Santiago Camotlán, Oaxaca, México. *Econ. Bot.* **2017**, *71*, 209–223. [CrossRef]
56. Breedlove, D.; Laughlin, R. *The Flowering of Man: A Tzotzil Botany of Zinacantán, Volume I*; Smithsonian Contributions to Anthropology, Number 35; Smithsonian Institution Press: Washington, DC, USA, 1993.
57. Isidro, V.M. *Etnobotánica de los Zoques de Tuxtla Gutiérrez, Chiapas*; Instituto de Historia Natural: Chiapas, México, 1997; p. 59.
58. De La Cruz-Jiménez, L.; Guzmán-Lucio, M.; Viveros-Valdez, E. Traditional medicinal plants used for the treatment of gastrointestinal diseases in Chiapas, México. *World Appl. Sci. J.* **2014**, *31*, 508–515. [CrossRef]
59. Carović-Stanko, K.; Petek, M.; Grdiša, M.; Pintar, J.; Bedeković, D.; Herak, M. Medicinal plants of the family Lamiaceae as functional foods—A Review. *J. Food Sci.* **2016**, *5*, 377–390. [CrossRef]
60. Abubakar, A.R.; Haque, M. Preparation of medicinal plants: Basic extraction and fractionation procedures for experimental purposes. *J. Pharm. Bioallied Sci.* **2020**, *12*, 1–10. [CrossRef]
61. Feng, J.; Liu, L.; Yao, F.; Zhou, D.; He, Y.; Wang, J. The protective effect of tanshinone IIA on endothelial cells: A generalist among clinical therapeutics. *Expert. Rev. Clin. Pharmacol.* **2021**, *14*, 239–248. [CrossRef]
62. Mahalakshmi, B.; Huang, C.Y.; da Lee, S.; Maurya, N.; Kiefer, R.; Bharath Kumar, V. Review of Danshen: From its metabolism to possible mechanisms of its biological activities. *J. Funct. Foods* **2021**, *85*, 104613. [CrossRef]
63. Yang, N.; Zou, C.; Luo, W.; Xu, D.; Wang, M.; Wang, Y.; Wu, G.; Shan, P.; Liang, G. Scclareol attenuates angiotensin II -induced cardiac remodeling and inflammation via inhibiting MAPK signaling. *Phytother. Res.* **2022**, *37*, 578–591. [CrossRef] [PubMed]
64. Miraj, S.; Kiani, S. A review study of therapeutic effects of *Salvia officinalis* L. *Pharm. Lett.* **2016**, *8*, 299–303. Available online: www.researchgate.net/publication/305147951 (accessed on 20 January 2023).
65. Kamatou, G.P.P.; Van Zyl, R.L.; Van Vuuren, S.F.; Viljoen, A.M.; Figueiredo, A.C.; Barroso, J.G.; Tilney, P.M. Chemical composition, leaf trichome types and biological activities of the essential oils of four related *Salvia* species indigenous to southern Africa. *J. Essent. Oil Res.* **2006**, *18*, 72–79. [CrossRef]
66. Chan, H.H.; Hwang, T.L.; Su, C.R.; Reddy, M.V.B.; Wu, T.S. Anti-inflammatory, anticholinesterase and antioxidative constituents from the roots and the leaves of *Salvia nipponica* Miq. var. *formosana*. *Phytomedicine* **2011**, *18*, 148–150. [CrossRef]
67. Jang, H.H.; Cho, S.Y.; Kim, M.J.; Kim, J.B.; Lee, S.H.; Lee, M.Y.; Lee, Y.M. Anti-inflammatory effects of *Salvia plebeia* R. Br. extract in vitro and in an ovalbumin-induced mouse model. *Biol. Res.* **2016**, *49*, 41. [CrossRef]
68. Batista, D.; Falé, P.L.; Serralheiro, M.L.; Araújo, M.E.; Dias, C.; Branco, I.; Grossi, C.; Coelho, J.; Palavra, A.; Madeira, P.J.A.; et al. Phytochemical characterization and biological evaluation of the aqueous and supercritical fluid extracts from *Salvia sclareaoides* Brot. *Open Chem.* **2017**, *15*, 82–91. [CrossRef]
69. Hassan, Z.Y.; Hassan, T.Y.; Abu-Raghif, A.R. Evaluation the Effectiveness of Phenolic Compound of *Salvia frigida* on Induced Atopic Dermatitis in Experimental Mice. *Iraqi J. Pharm. Sci.* **2022**, *31*, 154–166. [CrossRef]
70. Zou, Z.Q.; Ning, D.S.; Liu, Y.; Fu, Y.X.; Li, L.C.; Pan, Z.H. Five new germacrane sesquiterpenes with anti-inflammatory activity from *Salvia petrophila*. *Phytochem. Lett.* **2022**, *47*, 111–114. [CrossRef]
71. Moretti, M.D.L.; Peana, A.T.; Satta, M. A study on anti-inflammatory and peripheral analgesic action of *Salvia sclarea* oil and its main components. *J. Essent. Oil Res.* **1997**, *9*, 199–204. [CrossRef]
72. Hosseinzadeh, H.; Yavary, M. Anti-inflammatory effect of *Salvia leriifolia* Benth. leaf extract in mice and rats. *Pharm. Pharmacol. Lett.* **1999**, *9*, 60–61.
73. Maklad, Y.A.; Aboutabl, E.A.; El-Sherei, M.M.; Meselhy, K.M. Bioactivity studies of *Salvia transsylvanica* (Schur. ex Griseb.) grown in Egypt. *Phytother. Res.* **1999**, *13*, 147–150. [CrossRef]
74. Al-Yousuf, M.H.; Bashir, A.K.; Ali, B.H.; Tanira, M.O.M.; Blunden, G. Some effects of *Salvia aegyptiaca* L. on the central nervous system in mice. *J. Ethnopharmacol.* **2002**, *81*, 121–127. [CrossRef] [PubMed]
75. Boukhary, R.; Raafat, K.; Ghoneim, A.I.; Aboul-Ela, M.; El-Lakany, A. Anti-inflammatory and antioxidant activities of *Salvia fruticosa*: An HPLC determination of phenolic contents. *Evid.-Based Complement. Altern. Med.* **2016**, *2016*, 7178105. [CrossRef] [PubMed]
76. Hussain, L.; Sajid Hamid Akash, M.; Imran Qadir, M.; Pharm Sci, P.J.; Iqbal, R.; Irfan, M.; Imran Qadir, M. Analgesic, anti-inflammatory, and antipyretic activity of *Salvia moorcroftiana*. *Pak. J. Pharm. Sci.* **2017**, *30*, 7134. Available online: https://www.researchgate.net/publication/316007134_Analgesic_anti-inflammatory_and_antipyretic_activity_of_Salvia_moorcroftiana (accessed on 25 June 2023).
77. Semaoui, R.; Ouafi, S.; Machado, S.; Barros, L.; Ferreira, I.C.F.R.; Oliveira, M.B.P.P. Infusion of aerial parts of *Salvia chudaei* Batt. & Trab. from Algeria: Chemical, toxicological and bioactivities characterization. *J. Ethnopharmacol.* **2021**, *280*, 114455. [CrossRef]
78. Taheri, S.; Khalifeh, S.; Shajee, H.; Ashabi, G. Dietary uptake of *Salvia macilenta* extract improves Nrf2 antioxidant signaling pathway and diminishes inflammation and apoptosis in amyloid beta-induced rats. *Mol. Biol. Rep.* **2021**, *48*, 7667–7676. [CrossRef]
79. Pattanayak, P.; Behera, P.; Das, D.; Panda, S. *Ocimum sanctum* L. A reservoir plant for therapeutic applications: An overview. *Pharmacogn. Rev.* **2010**, *4*, 95–105. [CrossRef]

80. Singh, D.; Chaudhuri, P.K. A review on phytochemical and pharmacological properties of Holy basil (*Ocimum sanctum* L.). *Ind. Crops Prod.* **2018**, *118*, 367–382. [[CrossRef](#)]
81. Bacalbasa, N.; Balescu, I.; Stoica, C.; Pop, L.; Varlas, V.; Martac, C.; Voichitoiu, A.; Gaspar, B. The anti-inflammatory, anti-infectious and anti-cancerous effects of *Thymus vulgaris*. *Rom. Med. J.* **2022**, *17*, 21–23. [[CrossRef](#)]
82. el Meniyi, N.; Mrabti, H.N.; el Omari, N.; Bakili, A.E.; Bakrim, S.; Mekkaoui, M.; Balahbib, A.; Amiri-Ardekani, E.; Ullah, R.; Alqahtani, A.S.; et al. Medicinal Uses, Phytochemistry, Pharmacology, and Toxicology of *Mentha spicata*. *Evid.-Based Complement. Altern. Med.* **2022**, *2022*, 7990508. [[CrossRef](#)]
83. Sharifi-Rad, M.; Berkay Yilmaz, Y.; Antika, G.; Salehi, B.; Tumer, T.B.; Kulandaismy Venil, C.; Das, G.; Patra, J.K.; Karazhan, N.; Akram, M.; et al. Phytochemical constituents, biological activities, and health-promoting effects of the genus *Origanum*. *Phytother. Res.* **2021**, *35*, 95–121. [[CrossRef](#)]
84. Hajhashemi, V.; Ghannadi, A.; Sharif, B. Anti-inflammatory and analgesic properties of the leaf extracts and essential oil of *Lavandula angustifolia* Mill. *J. Ethnopharmacol.* **2003**, *89*, 67–71. [[CrossRef](#)] [[PubMed](#)]
85. Zhao, J.; Xu, F.; Huang, H.; Ji, T.; Li, C.; Tan, W.; Chen, Y.; Ma, L. Evaluation on bioactivities of total flavonoids from *Lavandula angustifolia*. *Pak. J. Pharm. Sci.* **2015**, *28*, 1245–1251. [[PubMed](#)]
86. Giovannini, D.; Gismondi, A.; Basso, A.; Canuti, L.; Braglia, R.; Canini, A.; Mariani, F.; Cappelli, G. *Lavandula angustifolia* mill. Essential oil exerts antibacterial and anti-inflammatory effects in macrophage mediated immune response to *staphylococcus aureus*. *Immunol. Investig.* **2016**, *45*, 11–28. [[CrossRef](#)] [[PubMed](#)]
87. Georgiev, Y.N.; Paulsen, B.S.; Kiyohara, H.; Ciz, M.; Ognyanov, M.H.; Vasicek, O.; Rise, F.; Denev, P.N.; Yamada, H.; Lojek, A.; et al. The common lavender (*Lavandula angustifolia* Mill.) pectic polysaccharides modulate phagocytic leukocytes and intestinal Peyer’s patch cells. *Carbohydr. Polym.* **2017**, *174*, 948–959. [[CrossRef](#)]
88. Cardia, G.F.E.; Silva-Filho, S.E.; Silva, E.L.; Uchida, N.S.; Cavalcante, H.A.O.; Cassarotti, L.L.; Salvadego, V.E.C.; Spironello, R.A.; Bersani-Amado, C.A.; Cuman, R.K.N. Effect of lavender (*Lavandula angustifolia*) essential oil on acute inflammatory response. *Evid. Based Complementary Altern. Med.* **2018**, *2018*, 1413940. [[CrossRef](#)]
89. Souris, F.; Rakhshan, K.; Erfani, S.; Azizi, Y.; Maleki, S.; Aboutaleb, N. Natural lavender oil (*Lavandula angustifolia*) exerts cardioprotective effects against myocardial infarction by targeting inflammation and oxidative stress. *Inflammopharmacology* **2019**, *27*, 799–807. [[CrossRef](#)]
90. Chen, X.; Zhang, L.; Qian, C.; Du, Z.; Xu, P.; Xiang, Z. Chemical compositions of essential oil extracted from *Lavandula angustifolia* and its prevention of TPA-induced inflammation. *Microchem. J.* **2020**, *153*, 104458. [[CrossRef](#)]
91. Xie, Q.; Wang, Y.; Zou, G.L. Protective effects of lavender oil on sepsis-induced acute lung injury via regulation of the NF-κB pathway. *Pharm. Biol.* **2022**, *60*, 968–978. [[CrossRef](#)]
92. Guginski, G.; Luiz, A.P.; Silva, M.D.; Massaro, M.; Martins, D.F.; Chaves, J.; Mattos, R.W.; Silveira, D.; Ferreira, V.M.M.; Calixto, J.B.; et al. Mechanisms involved in the antinociception caused by ethanolic extract obtained from the leaves of *Melissa officinalis* (lemon balm) in mice. *Pharmacol. Biochem. Behav.* **2009**, *93*, 10–16. [[CrossRef](#)]
93. Müzell, D.; Lunardelli, A.; Leite, C.E.; Medeiros Fagundes, R.; Saciura, V.C.; Reichel, C.L.; Rodrigues De Oliveira, J.; Vieira Astarita, L. Nephroprotective and anti-inflammatory effects of aqueous extract of *Melissa officinalis* L. on acetaminophen-induced and pleurisy-induced lesions in rats. *Braz. Arch. Biol. Technol.* **2013**, *56*, 383–392. [[CrossRef](#)]
94. Bounihi, A.; Hajjaj, G.; Alnamer, R.; Cherrah, Y.; Zellou, A. In vivo potential anti-inflammatory activity of *Melissa officinalis* L. essential oil. *Adv. Pharmacol. Sci.* **2013**, *2013*, 101759. [[CrossRef](#)] [[PubMed](#)]
95. Draganic, N.; Andjic, M.; Jeremic, J.; Zivkovic, V.; Kocovic, A.; Tomovic, M.; Bozin, B.; Kladar, N.; Bolevich, S.; Jakovljevic, V.; et al. Anti-inflammatory and antioxidant effects of *Melissa officinalis* extracts: A comparative study. *Iran. J. Pharm. Res.* **2022**, *21*, 126561. [[CrossRef](#)]
96. Hwang, Y.J.; Lee, E.J.; Kim, H.R.; Hwang, K.A. NF-κB-targeted anti-inflammatory activity of *Prunella vulgaris* var. *lilacina* in macrophages RAW 264.7. *Int. J. Mol. Sci.* **2013**, *14*, 21489–21503. [[CrossRef](#)] [[PubMed](#)]
97. Ryu, S.Y.; Oak, M.-H.; Yoon, S.-K.; Cho, D.-I.; Yoo, G.-S.; Kim, T.-S.; Kim, K.-M.; Thieme, G.; Stuttgart, V.; York, N. Anti-allergic and anti-inflammatory triterpenes from the herb of *Prunella vulgaris*. *Planta Med.* **2000**, *66*, 358. [[CrossRef](#)]
98. Tang, Y.Q.N.; Deng, J.; Li, L.; Yan, J.; Lin, L.M.; Li, Y.M.; Lin, Y.; Xia, B.H. Essential oil from *Prunella vulgaris* L. as a valuable source of bioactive constituents: In vitro anti-bacterial, anti-viral, immunoregulatory, anti-inflammatory, and chemical profiles. *S. Afr. J. Bot.* **2022**, *151*, 614–627. [[CrossRef](#)]
99. Zheng, X.Q.; Song, L.X.; Qiu, H.; Yang, Y.B.; Han, Z.Z.; Wang, Z.T.; Gu, L.H. Novel phenolic and diterpenoid compounds isolated from the fruit spikes of *Prunella vulgaris* L. and their anti-inflammatory activities. *Phytochem. Lett.* **2022**, *49*, 60–64. [[CrossRef](#)]
100. Choia, H.; Kim, T.; Kim, S.-H.; Kim, J. Anti-allergic inflammatory triterpenoids isolated from the spikes of *Prunella vulgaris*. *Nat. Prod. Commun.* **2016**, *11*, 2–31. [[CrossRef](#)]
101. Khazdair, M.R.; Ghorani, V.; Alavinezhad, A.; Boskabady, M.H. Pharmacological effects of *Zataria multiflora* Boiss L. and its constituents focus on their anti-inflammatory, antioxidant, and immunomodulatory effects. *Fundam. Clin. Pharmacol.* **2018**, *32*, 26–50. [[CrossRef](#)]
102. Milijasevic, B.; Steinbach, M.; Mikov, M.; Raskovic, A.; Capo, I.; Zivkovic, J.; Borisev, I.; Canji, J.; Teofilovic, B.; Vujcic, M.; et al. Impact of winter savory extract (*Satureja montana* L.) on biochemical parameters in serum and oxidative status of liver with application of the principal component analysis in extraction solvent selection. *Eur. Rev. Med. Pharmacol. Sci.* **2022**, *26*, 4721–4734.

103. Shin, T.Y.; Kim, S.H.; Suk, K.; Ha, J.H.; Kim, I.K.; Lee, M.G.; Jun, C.D.; Kim, S.Y.; Lim, J.P.; Eun, J.S.; et al. Anti-allergic effects of *Lycopus lucidus* on mast cell-mediated allergy model. *Toxicol. Appl. Pharmacol.* **2005**, *209*, 255–262. [CrossRef] [PubMed]
104. Mousavi Nia, A.; Pari Kalantarpour, T.; Basiri, M.; Vafaei, F.; Asadi-Shekaari, M.; Eslami, A.; Darvish Zadeh, F. *Nepeta Dschuparensis* Bornm extract moderates COX-2 and IL-1 β proteins in a rat model of cerebral ischemia. *Iran. J. Med. Sci.* **2017**, *42*, 179–186.
105. Bian, M.; Zhang, Y.; Du, X.; Xu, J.; Cui, J.; Gu, J.; Zhu, W.; Zhang, T.; Chen, Y. Apigenin-7-diglucuronide protects retinas against bright light-induced photoreceptor degeneration through the inhibition of retinal oxidative stress and inflammation. *Brain Res.* **2017**, *1663*, 141–150. [CrossRef] [PubMed]
106. García-Díaz, J.A.; Navarrete-Vázquez, G.; García-Jiménez, S.; Hidalgo-Figueroa, S.; Almanza-Pérez, J.C.; Alarcón-Aguilar, F.J.; Gómez-Zamudio, J.; Cruz, M.; Ibarra-Barajas, M.; Estrada-Soto, S. Antidiabetic, antihyperlipidemic and anti-inflammatory effects of tilianin in streptozotocin-nicotinamide diabetic rats. *Biomed. Pharmacother.* **2016**, *83*, 667–675. [CrossRef] [PubMed]
107. Shen, W.; Anwaier, G.; Cao, Y.; Lian, G.; Chen, C.; Liu, S.; Tuerdi, N.; Qi, R. Atheroprotective Mechanisms of Tilianin by Inhibiting Inflammation Through Down-Regulating NF- κ B Pathway and Foam Cells Formation. *Front. Physiol.* **2019**, *10*, 825. [CrossRef]
108. Nie, L.; Li, R.; Huang, J.; Wang, L.; Ma, M.; Huang, C.; Wu, T.; Yan, R.; Hu, X. Abietane diterpenoids from *Dracocephalum moldavica* L. and their anti-inflammatory activities in vitro. *Phytochemistry* **2021**, *184*, 112680. [CrossRef]
109. Chen, S.; Fang, A.; Zhong, Y.; Tang, J. *Ziziphora clinopodioides* Lam leaf aqueous extract mediated novel green synthesis of iron nanoparticles and its anti-hemolytic anemia potential: A chemobiological study. *Arab. J. Chem.* **2022**, *15*, 103561. [CrossRef]
110. Mohammad, G.; Tabrizi, M.; Ardalan, T.; Yadaman, S.; Safavi, E. Green synthesis of zinc oxide nanoparticles and evaluation of anti-angiogenesis, anti-inflammatory and cytotoxicity properties. *J. Biosci.* **2019**, *44*, 30.
111. Singh, M.K.; Gidwani, B.; Gupta, A.; Dhongade, H.; Kaur, C.D.; Kashyap, P.P.; Tripathi, D.K. A review of the medicinal plants of genus *Orthosiphon* (Lamiaceae). *Int. J. Biol. Chem.* **2015**, *9*, 318–331. [CrossRef]
112. Andrade, J.M.; Custódio, L.; Romagnoli, A.; Reis, C.P.; Rodrigues, M.J.; Garcia, C.; Petruccioli, E.; Goletti, D.; Faustino, C.; Fimia, G.M.; et al. Antitubercular and anti-inflammatory properties screening of natural products from *Plectranthus* species. *Future Med. Chem.* **2018**, *10*, 1677–1691. [CrossRef]
113. Almeida-Bezerra, J.W.; Rodrigues, F.C.; Lima Bezerra, J.J.; Vieira Pinheiro, A.A.; Almeida De Menezes, S.; Tavares, A.B.; Costa, A.R.; Augusta De Sousa Fernandes, P.; Bezerra Da Silva, V.; Martins Da Costa, J.G.; et al. Traditional Uses, Phytochemistry, and Bioactivities of *Mesosphaerum suaveolens* (L.) Kuntze. *Evid.-Based Complement. Altern. Med.* **2022**, *10*, 3829180. [CrossRef] [PubMed]
114. Leung, C.H.; Grill, S.P.; Lam, W.; Gao, W.; Sun, H.D.; Cheng, Y.C. Eriocalyxin B inhibits nuclear factor- κ B activation by interfering with the binding of both p65 and p50 to the response element in a noncompetitive manner. *Mol. Pharmacol.* **2006**, *70*, 1946–1955. [CrossRef] [PubMed]
115. Jiang, H.; Li, X.; Sun, H.; Zhang, H.; Puno, P. Scopariusols L-T, nine new ent-kaurane diterpenoids isolated from *Isodon scoparius*. *Chin. J. Nat. Med.* **2018**, *16*, 456–464. [CrossRef]
116. Zhang, Y.Y.; Jiang, H.Y.; Liu, M.; Hu, K.; Wang, W.G.; Du, X.; Li, X.N.; Pu, J.X.; Sun, H.D. Bioactive ent-kaurane diterpenoids from *Isodon rubescens*. *Phytochemistry* **2017**, *143*, 199–207. [CrossRef] [PubMed]
117. Yin, Q.-Q.; Liu, C.-X.; Wu, Y.-L.; Wu, S.-F.; Wang, Y.; Zhang, X.; Hu, X.-J.; Pu, J.-X.; Lu, Y.; Zhou, H.-C.; et al. Preventive and Therapeutic Effects of Adenanthin on Experimental Autoimmune Encephalomyelitis by Inhibiting NF- κ B Signaling. *J. Immunol.* **2013**, *191*, 2115–2125. [CrossRef]
118. Lu, Y.; Chen, B.; Song, J.H.; Zhen, T.; Wang, B.Y.; Li, X.; Liu, P.; Yang, X.; Zhang, Q.L.; Xi, X.D.; et al. Eriocalyxin B ameliorates experimental autoimmune encephalomyelitis by suppressing Th1 and Th17 cells. *Proc. Natl. Acad. Sci. USA* **2013**, *110*, 2258–2263. [CrossRef]
119. Zhang, L.G.; Yu, Z.Q.; Yang, C.; Chen, J.; Zhan, C.S.; Chen, X.G.; Zhang, L.; Hao, Z.Y.; Liang, C.Z. Effect of Eriocalyxin B on prostatic inflammation and pelvic pain in a mouse model of experimental autoimmune prostatitis. *Prostate* **2020**, *80*, 1394–1404. [CrossRef]
120. He, H.; Jiang, H.; Chen, Y.; Ye, J.; Wang, A.; Wang, C.; Liu, Q.; Liang, G.; Deng, X.; Jiang, W.; et al. Oridonin is a covalent NLRP3 inhibitor with strong anti-inflammasome activity. *Nat. Commun.* **2018**, *9*, 2550. [CrossRef]
121. Zhang, Y.; Wang, K.; Chen, H.; He, R.; Cai, R.; Li, J.; Zhou, D.; Liu, W.; Huang, X.; Yang, R.; et al. Anti-inflammatory lignans and phenylethanoid glycosides from the root of *Isodon ternifolius* (D.Don) Kudô. *Phytochemistry* **2018**, *153*, 36–47. [CrossRef]
122. Yan, B.C.; Wang, W.G.; Hu, D.B.; Sun, X.; Kong, L.M.; Li, X.N.; Du, X.; Luo, S.H.; Liu, Y.; Li, Y.; et al. Phomopchalasins A and B, two cytochalasans with polycyclic-fused skeletons from the endophytic fungus *Phomopsis* sp. *Org. Lett.* **2016**, *18*, 1108–1111. [CrossRef]
123. López-García, R.E.; Rabanal, R.M.; Darias, V.; Martín-Herrera, D.; Carreiras, M.C.; Rodríguez, B. A preliminary study of *Cedronella canariensis* (L.) var. *canariensis* extracts for antiinflammatory and analgesic activity in rats and mice. *Phytther. Res.* **1991**, *5*, 273–275. [CrossRef]
124. Olajide, O.A.; Oladiran, O.O.; Awe, S.O.; Makinde, J.M. Pharmacological evaluation of *Hoslundia opposita* extract in rodents. *Phytther. Res.* **1998**, *12*, 364–366. [CrossRef]
125. Aljohani, A.S.M.; Alhumaydhi, F.; Rauf, A.; Hamad, E.; Rashid, U. In Vivo and In Vitro biological evaluation and molecular docking studies of compounds isolated from *Micromeria biflora* (Buch. Ham. ex D.Don) Benth. *Molecules* **2022**, *27*, 3377. [CrossRef]
126. Vladimir-Knežević, S.; Cvijanović, O.; Blažeković, B.; Kindl, M.; Štefan, M.B.; Domitrović, R. Hepatoprotective effects of *Micromeria croatica* ethanolic extract against CCl₄-induced liver injury in mice. *BMC Complement. Altern. Med.* **2015**, *15*, 1–12. [CrossRef]

127. Wang, X.; Cheng, K.; Liu, Z.; Sun, Y.; Zhou, L.; Xu, M.; Dai, X.; Xiong, Y.; Zhang, H. Bioactive constituents of *Mosla chinensis*-cv. Jiangxiangru ameliorate inflammation through MAPK signaling pathways and modify intestinal microbiota in DSS-induced colitis mice. *Phytomedicine* **2021**, *93*, 153804. [[CrossRef](#)] [[PubMed](#)]
128. Álvarez-Santos, N.; Estrella-Parra, E.A.; Benítez-Flores, J.d.C.; Serrano-Parrales, R.; Villamar-Duque, T.E.; Santiago-Santiago, M.A.; González-Valle, M.D.R.; Avila-Acevedo, J.G.; García-Bores, A.M. *Asterohyptis stellulata*: Phytochemistry and wound healing activity. *Food Biosci.* **2022**, *50*, 102150. [[CrossRef](#)]
129. Chen, J.; Wang, J.B.; Yu, C.H.; Chen, L.Q.; Xu, P.; Yu, W.Y. Total flavonoids of *Mosla scabra* leaves attenuates lipopolysaccharide-induced acute lung injury via down-regulation of inflammatory signaling in mice. *J. Ethnopharmacol.* **2013**, *148*, 835–841. [[CrossRef](#)]
130. Becker, K.; Schwaiger, S.; Waltenberger, B.; Fuchs, D.; Pezzei, C.K.; Schennach, H.; Stuppner, H.; Gostner, J.M. Immunomodulatory effects of diterpene quinone derivatives from the roots of *Horminum pyrenaicum* in human PBMC. *Oxid. Med. Cell. Longev.* **2018**, *2018*, 2980295. [[CrossRef](#)]
131. Moreno-Pérez, G.F.; González-Trujano, M.E.; Hernandez-Leon, A.; Valle-Dorado, M.G.; Valdés-Cruz, A.; Alvarado-Vásquez, N.; Aguirre-Hernández, E.; Salgado-Ceballos, H.; Pellicer, F. Antihyperalgesic and Antiallodynic Effects of Amarisolide A and *Salvia amarissima* Ortega in Experimental Fibromyalgia-Type Pain. *Metabolites* **2023**, *13*, 59. [[CrossRef](#)]
132. Chen, H.; Li, Y.; Wang, J.; Zheng, T.; Wu, C.; Cui, M.; Feng, Y.; Ye, H.; Dong, Z.; Dang, Y. Plant Polyphenols Attenuate DSS-induced Ulcerative Colitis in Mice via Antioxidation, Anti-inflammation and Microbiota Regulation. *Int. J. Mol. Sci.* **2023**, *24*, 10828. [[CrossRef](#)]
133. Li, Z.; Zhang, Y.; Zhou, Y.; Wang, F.; Yin, C.; Ding, L.; Zhang, S. Tanshinone IIA suppresses the progression of lung adenocarcinoma through regulating CCNA2-CDK2 complex and AURKA/PLK1 pathway. *Sci. Rep.* **2021**, *11*, 23681. [[CrossRef](#)] [[PubMed](#)]
134. Verano, J.; González-Trujano, M.E.; Déciga-Campos, M.; Ventura-Martínez, R.; Pellicer, F. Ursolic acid from *Agastache mexicana* aerial parts produces antinociceptive activity involving TRPV1 receptors, cGMP and a serotonergic synergism. *Pharmacol. Biochem. Behav.* **2013**, *110*, 255–264. [[CrossRef](#)] [[PubMed](#)]
135. González-Ramírez, A.; González-Trujano, M.E.; Pellicer, F.; López-Muñoz, F.J. Anti-nociceptive and anti-inflammatory activities of the *Agastache mexicana* extracts by using several experimental models in rodents. *J. Ethnopharmacol.* **2012**, *142*, 700–705. [[CrossRef](#)]
136. Liu, Y.; Song, H.; Xu, J.; Bi, G.; Meng, D. Anti-inflammatory abietanes diterpenes and triterpenoids isolated from *Clinopodium polyccephalum*. *Fitoterapia* **2022**, *161*, 105244. [[CrossRef](#)] [[PubMed](#)]
137. Mustarichie, R.; Moektiwardojo, M.; Apriani Dewi, W. Isolation, identification, and characteristic of essential oil of Iller (*Plectranthus scutellarioides* (L.) R. Br leaves. *J. Pharm. Sci. Res.* **2017**, *9*, 2218–2223.
138. Mustarichie, R.; Ramdhani, D.; Mekar Saptarini, N. The anti-inflammatory tablet formulation of Coleus (*Plectranthus scutellarioides*) leaves extract using Kollicoat® protect coating. *Int. J. Appl. Pharm.* **2022**, *14*, 159–162. [[CrossRef](#)]
139. Shi, Q.Q.; Dang, J.; Wen, H.X.; Yuan, X.; Tao, Y.D.; Wang, Q.L. Anti-hepatitis, antioxidant activities and bioactive compounds of *Dracocephalum heterophyllum* extracts. *Bot. Stud.* **2016**, *57*, 16. [[CrossRef](#)]
140. Bian, J.; Wang, K.; Wang, Q.; Wang, P.; Wang, T.; Shi, W.; Ruan, Q. *Dracocephalum heterophyllum* (DH) exhibits potent anti-proliferative effects on autoreactive CD4+ T cells and ameliorates the development of experimental autoimmune uveitis. *Front. Immunol.* **2020**, *11*, 575669. [[CrossRef](#)]
141. Minaiyan, M.; Sadraei, H.; Asghari, G.; Khanabadi, M.; Minaiyan, M. Anti-inflammatory effect of apigenin and hydroalcoholic extract of *Dracocephalum kotschy*i on acetic acid-induced colitis in rats. *Res. Pharm. Sci.* **2017**, *12*, 322–329. [[CrossRef](#)]
142. Olennikov, D.N.; Chirikova, N.K.; Okhlopkova, Z.M.; Zulfugarov, I.S. Chemical composition and antioxidant activity of Tánara Ótó (*Dracocephalum palmatum* Stephan), a medicinal plant used by the North-Yakutian nomads. *Molecules* **2013**, *18*, 14105–14121. [[CrossRef](#)]
143. Andreyeva, A.A.; Sivtseva, V.V.; Pashayeva, A.T.; Chirikova, N.D.; Okhlopkova, Z.M.; Kim, S.W.; Huseynova, I.M.; Zulfugarov, I.S. Dragonhead as a model plant for biotechnology of biologically active compounds. *Azerbaijan Pharm. Pharmacother. J.* **2020**, *1*, 31–35.
144. Chirikova, N.K.; Nokhsorov, V.V.; Nikolaev, W.M. Ethnopharmacological research of plant resources of central and eastern Yakutia. *IOP Conf. Ser. Earth Environ. Sci.* **2021**, *670*, 012010. [[CrossRef](#)]
145. Zhu, C.-S.; Liu, K.; Wang, J.-L.; Li, J.-F.; Liu, M.-F.; Hao, N.; Lin, Y.-X.; Xiao, Z.-F. Antioxidant activities and hepatoprotective potential of *Dracocephalum rupestre* Hance extract against CCl4-induced hepatotoxicity in Kunming mice. *J. Food Biochem.* **2018**, *42*, e12484. [[CrossRef](#)]
146. Zhang, Q.; Porto, N.M.; Guilhon, C.C.; Fernandes, P.D.; Boylan, F. Pharmacognostic study on *Elsholtzia ciliata* (Thunb.) Hyl.: Anatomy, phytochemistry and pharmacological activities. *Pharmaceutics* **2021**, *14*, 1152. [[CrossRef](#)] [[PubMed](#)]
147. Nguyen, D.T.X.; Tran, H.; Schwaiger, S.; Stuppner, H.; Marzocco, S. Effect of non-volatile constituents of *Elsholtzia ciliata* (Thunb.) Hyl. from southern Vietnam on reactive oxygen species and nitric oxide release in macrophages. *Chem. Biodivers.* **2021**, *18*, e2000577. [[CrossRef](#)] [[PubMed](#)]
148. Pudziuveliyte, L.; Liaudanskas, M.; Jekabsone, A.; Sadauskienė, I.; Bernatoniene, J. *Elsholtzia ciliata* (Thunb.) Hyl. extracts from different plant parts: Phenolic composition, antioxidant, and anti-inflammatory activities. *Molecules* **2020**, *25*, 1153. [[CrossRef](#)] [[PubMed](#)]
149. Kim, H.-H.; Yoo, J.-S.; Lee, H.-S.; Shin, T.-Y.; Kim, S.-H. *Elsholtzia ciliata* inhibits mast cell-mediated allergic inflammation: Role of calcium, p38 mitogen-activated protein kinase and nuclear factor-kB. *Exp. Biol. Med.* **2011**, *236*, 1070–1077. [[CrossRef](#)]

150. Kim, T.-W.; Kim, Y.-J.; Seo, C.-S.; Lee, M.-Y.; Jung, J.-Y. *Elsholtzia ciliata* (Thunb.) Hylander attenuates renal inflammation and interstitial fibrosis via regulation of TGF- β and Smad3 expression on unilateral ureteral obstruction rat model. *Phytomedicine* **2016**, *23*, 331–339. [CrossRef]
151. Yang, F.; Pu, H.-Y.; Yaseen, A.; Chen, B.; Li, F.; Gu, Y.-C.; Shen, X.-F.; Wang, M.-K.; Guo, D.-L.; Wang, L. Terpenoid and phenolic derivatives from the aerial parts of *Elsholtzia rugulosa* and their anti-inflammatory activity. *Phytochemistry* **2021**, *181*, 112543. [CrossRef]
152. Xiang, Z.; Wu, X.; Liu, X.; Jin, Y. Glaucocalyxin A: A review. *Nat. Prod. Res.* **2014**, *28*, 2221–2236. [CrossRef]
153. Lee, J.W.; Lee, M.S.; Kim, T.H.; Lee, H.J.; Hong, S.S.; Noh, Y.H.; Hwang, B.Y.; Ro, J.S.; Hong, J.T. Inhibitory effect of Inflexinol on nitric oxide generation and iNOS expression via inhibition of NF- κ B activation. *Mediat. Inflamm.* **2007**, *2007*, 93148. [CrossRef] [PubMed]
154. Ko, H.M.; Koppula, S.; Kim, B.-W.; Kim, I.S.; Hwang, B.Y.; Suk, K.; Park, E.J.; Choi, D.-K. Inflexin attenuates proinflammatory responses and nuclear factor- κ B activation in LPS-treated microglia. *Eur. J. Pharmacol.* **2010**, *633*, 98–106. [CrossRef] [PubMed]
155. Cheng, E.; Chi, J.; Li, Y.X.; Zhang, W.J.; Huang, N.; Wang, Z.M.; Dai, L.P.; Xu, E.P. Diverse ent-kaurane diterpenoids from *Isodon henryi*. *Tetrahedron Lett.* **2022**, *108*, 154119. [CrossRef]
156. Hwang, B.; Lee, J.; Koo, T.; Kim, H.; Hong, Y.; Ro, J.; Lee, K.; Lee, J. Kaurane diterpenes from *Isodon japonicus* inhibit nitric oxide and prostaglandin E2 production and NF- κ B activation in LPS-stimulated macrophage RAW264.7 cells. *Planta Med.* **2001**, *67*, 406–410. [CrossRef]
157. Kim, B.W.; Koppula, S.; Kim, I.S.; Lim, H.W.; Hong, S.M.; Han, S.D.; Hwang, B.Y.; Choi, D.K. Anti-neuroinflammatory activity of kamebakaurin from *Isodon japonicus* via inhibition of c-Jun NH₂-terminal kinase and p38 mitogen-activated protein kinase pathway in activated microglial cells. *J. Pharmacol. Sci.* **2011**, *116*, 296–308. [CrossRef]
158. Kim, J.Y.; Kim, H.S.; Kim, Y.J.; Lee, H.K.; Kim, J.S.; Kang, J.S.; Hong, J.T.; Kim, Y.; Hwang, B.Y.; Han, S.B. Effusain C inhibits inflammatory responses via blocking NF- κ B and MAPK signaling in monocytes. *Int. Immunopharmacol.* **2013**, *15*, 84–88. [CrossRef]
159. Ikoma, K.; Takahama, M.; Kimishima, A.; Pan, Y.; Taura, M.; Nakayama, A.; Arai, M.; Takemura, N.; Saitoh, T. Oridonin suppresses particulate-induced NLRP3-independent IL-1 α release to prevent crystallopathy in the lung. *Int. Immunol.* **2022**, *34*, 493–504. [CrossRef]
160. Yu, Z.-Y.; Liang, Y.-G.; Xiao, H.; Shan, Y.-J.; Dong, B.; Huang, R.; Fu, Y.-L.; Zhao, Z.-H.; Liu, Z.-Y.; Zhao, Q.-S.; et al. Melisoidesin G, a diterpenoid purified from *Isodon melisoides*, induces leukemic-cell apoptosis through induction of redox imbalance and exhibits synergy with other anticancer agents. *Int. J. Cancer* **2007**, *121*, 2084–2094. [CrossRef]
161. Kumar, A.; Singh, S.; Kumar, A.; Umrao Bawankule, D.; Tandon, S.; Kumar, A.; Swaroop Verma, R.; Saikia, D. Chemical composition, bactericidal kinetics, mechanism of action, and anti-inflammatory activity of *Isodon melisoides* (Benth.) H. Hara essential oil. *Nat. Prod. Res.* **2021**, *35*, 690–695. [CrossRef]
162. Zhang, Y.; Liu, J.; Jia, W.; Zhao, A.; Zhang, T.L.; Liu, Y.; Jia, J.; Zhao, W.; Li, A. Distinct immunosuppressive effect by *Isodon serra* extracts. *Int. Immunopharmacol.* **2005**, *5*, 1957–1965. [CrossRef]
163. Li, J.; Du, J.; Sun, L.; Liu, J.; Quan, Z. Anti-Inflammatory function of nodosin via inhibition of IL-2. *Am. J. Chinese Med.* **2010**, *38*, 127–142. [CrossRef] [PubMed]
164. Zhou, L.; Sun, L.; Wu, H.; Zhang, L.; Chen, M.; Liu, J.; Zhong, R. Oridonin ameliorates lupus-like symptoms of MRLlpr/lpr mice by inhibition of B-cell activating factor (BAFF). *Eur. J. Pharmacol.* **2013**, *715*, 230–237. [CrossRef] [PubMed]
165. Wan, J.; Liu, M.; Jiang, H.Y.; Yang, J.; Du, X.; Li, X.N.; Wang, W.G.; Li, Y.; Pu, J.X.; Sun, H.D. Bioactive ent-kaurane diterpenoids from *Isodon serra*. *Phytochem.* **2016**, *130*, 244–251. [CrossRef] [PubMed]
166. Xing, H.; An, L.; Song, Z.; Li, S.; Wang, H.; Wang, C.; Zhang, J.; Tuerhong, M.; Abudukeremu, M.; Li, D.; et al. Anti-Inflammatory ent-kaurane diterpenoids from *Isodon serra*. *J. Nat. Prod.* **2020**, *83*, 2844–2853. [CrossRef] [PubMed]
167. Jiang, H.-Y.; Wang, W.-G.; Zhou, M.; Wu, H.-Y.; Zhan, R.; Du, X.; Pu, J.-X.; Sun, H.-D. 6,7-seco-ent-Kaurane diterpenoids from *Isodon sculponeatus* and their bioactivity. *Chin. Chem. Lett.* **2014**, *25*, 541–544. [CrossRef]
168. Jiang, H.-Y.; Wang, W.-G.; Zhou, M.; Wu, H.-Y.; Zhan, R.; Li, X.-N.; Du, X.; Li, Y.; Pu, J.-X.; Sun, H.-D. Diterpenoids from *Isodon sculponeatus*. *Fitoterapia* **2014**, *93*, 142–149. [CrossRef]
169. Jia, T.; Cai, M.; Ma, X.; Li, M.; Qiao, J.; Chen, T. Oridonin inhibits IL-1 β -induced inflammation in human osteoarthritis chondrocytes by activating PPAR- γ . *Int. Immunopharmacol.* **2019**, *69*, 382–388. [CrossRef]
170. Wu, H.-Y.; Wang, W.-G.; Du, X.; Yang, J.; Pu, J.-X.; Sun, H.-D. Six new cytotoxic and anti-inflammatory 11,20-epoxy-ent-kaurane diterpenoids from *Isodon wikstroemioides*. *Chin. J. Nat. Med.* **2015**, *13*, 383–389.
171. Wu, H.-Y.; Wang, W.-G.; Jiang, H.-Y.; Du, X.; Li, X.-N.; Pu, J.-X.; Sun, H.-D. Cytotoxic and anti-inflammatory ent-kaurane diterpenoids from *Isodon wikstroemioides*. *Fitoterapia* **2014**, *98*, 192–198. [CrossRef]
172. Bayat, M.; Kalantar, K.; Amirghofran, Z. Inhibition of interferon- γ production and T-bet expression by menthol treatment of human peripheral blood mononuclear cells. *Immunopharmacol. Immunotoxicol.* **2019**, *41*, 267–276. [CrossRef]
173. Murad, H.A.S.; Abdallah, H.M.; Ali, S.S. *Mentha longifolia* protects against acetic-acid induced colitis in rats. *J. Ethnopharmacol.* **2016**, *190*, 354–361. [CrossRef] [PubMed]
174. Wang, M.; Wang, Y.; Shi, L.; Lu, Y.; Zhu, H.; Liang, L.; Sun, Z.L. A new phenylpropanoid-substituted flavan-3-ol from aerial part of *Mentha longifolia*. *Chem. Nat. Compd.* **2022**, *58*, 237–239. [CrossRef]

175. Prescott, T.A.K.; Veitch, N.C.; Simmonds, M.S.J. Direct inhibition of calcineurin by caffeoyl phenylethanoid glycosides from *Teucrium chamaedrys* and *Nepeta cataria*. *J. Ethnopharmacol.* **2011**, *137*, 1306–1310. [CrossRef] [PubMed]
176. de Lima, V.T.; Vieira, M.C.; Kassuya, C.A.L.; Cardoso, C.A.L.; Alves, J.M.; Foglio, M.A.; De Carvalho, J.E.; Formagio, A.S.N. Chemical composition and free radical-scavenging, anticancer and anti-inflammatory activities of the essential oil from *Ocimum kilimandscharicum*. *Phytomedicine* **2014**, *21*, 1298–1302. [CrossRef] [PubMed]
177. Banno, N.; Akihisa, T.; Tokuda, H.; Yasukawa, K.; Higashihara, H.; Ukiya, M.; Watanabe, K.; Kimura, Y.; Hasegawa, J.I.; Nishino, H. Triterpene acids from the leaves of *Perilla frutescens* and their anti-inflammatory and antitumor-promoting effects. *Biosci. Biotechnol. Biochem.* **2004**, *68*, 85–90. [CrossRef]
178. Huang, B.P.; Lin, C.H.; Chen, Y.C.; Kao, S.H. Anti-inflammatory effects of *Perilla frutescens* leaf extract on lipopolysaccharide stimulated RAW264.7 cells. *Mol. Med. Rep.* **2014**, *10*, 1077–1083. [CrossRef]
179. Ueda, H.; Yamazaki, C.; Yamazaki, M. Luteolin as an anti-inflammatory and anti-allergic constituent of *Perilla frutescens*. *Biol. Pharm. Bull.* **2002**, *25*, 1197–1202. [CrossRef]
180. Zuo, J.; Zhang, T.H.; Xiong, L.; Huang, L.; Peng, C.; Zhou, Q.M.; Dai, O. Two pairs of 7,7'-Cyclolignan enantiomers with anti-Inflammatory activities from *Perilla frutescens*. *Molecules* **2022**, *27*, 6102. [CrossRef]
181. Liu, D.; Wang, Y.; Lu, Z.; Lv, F.; Bie, X.; Zhao, H. Separation, characterization, and anti-inflammatory activities of galactoglycerolipids from *Perilla frutescens* (L.) Britton. *Nat. Prod. Res.* **2022**, *37*, 3610–3615. [CrossRef]
182. Kangwan, N.; Pintha, K.; Khanaree, C.; Kongkarnka, S.; Chewonarin, T.; Suttajit, M. Anti-inflammatory effect of *Perilla frutescens* seed oil rich in omega-3 fatty acid on dextran sodium sulfate-induced colitis in mice. *Res. Pharm. Sci.* **2021**, *16*, 464–473. [CrossRef]
183. Zhao, J.; Xu, L.; Jin, D.; Xin, Y.; Tian, L.; Wang, T.; Zhao, D.; Wang, Z.; Wang, J. Rosmarinic acid and related dietary supplements: Potential applications in the prevention and treatment of Cancer. *Biomolecules* **2022**, *12*, 1410. [CrossRef] [PubMed]
184. Sitarek, P.; Kowalczyk, T.; Synowiec, E.; Merecz-Sadowska, A.; Bangay, G.; Princiotto, S.; Sliwinski, T.; Rijo, P. An evaluation of the novel biological properties of diterpenes isolated from *Plectranthus ornatus* Codd. In vitro and in silico. *Cells* **2022**, *11*, 3243. [CrossRef] [PubMed]
185. Wu, S.J.; Chan, H.H.; Hwang, T.L.; Qian, K.; Morris-Natschke, S.; Lee, K.H.; Tian-Shung Wu, T.S. Salviatalin A and salvitrijudin A, two diterpenes with novel skeletons from roots of *Salvia digitaloides* and anti-inflammatory evaluation. *Tetrahedron Lett.* **2010**, *51*, 4287–4290. [CrossRef]
186. Piccinelli, A.C.; Figueiredo de Santana Aquino, D.; Morato, P.N.; Kuraoka-Oliveira, A.M.; Strapasson, R.L.; Dos Santos, E.P.; Stefanello, M.É.; Oliveira, R.J.; Kassuya, C.A. Anti-inflammatory and antihyperalgesic activities of ethanolic extract and fruticulin A from *Salvia lachnostachys* leaves in mice. *Evid.-Based Complement. Altern. Med.* **2014**, *2014*, 835914. [CrossRef]
187. Zarshenas, M.M.; Krenn, L. Phytochemical and pharmacological aspects of *Salvia mirzayanii* Rech. f. & Esfand. *Evid.-Based Complement. Altern. Med.* **2015**, *20*, 65–72. [CrossRef]
188. Ziae, A.; Hoppstädter, J.; Kiemer, A.K.; Ramezani, M.; Amirghofran, Z.; Diesel, B. Inhibitory effects of teuclatriol, a sesquiterpene from *Salvia mirzayanii*, on nuclear factor- κ B activation and expression of inflammatory mediators. *J. Ethnopharmacol.* **2015**, *160*, 94–100. [CrossRef]
189. Zou, Y.H.; Zhao, L.; Xu, Y.K.; Bao, J.M.; Liu, X.; Zhang, J.S.; Li, W.; Ahmed, A.; Yin, S.; Tang, G.H. Anti-inflammatory sesquiterpenoids from the Traditional Chinese Medicine *Salvia plebeia*: Regulates pro-inflammatory mediators through inhibition of NF- κ B and Erk1/2 signaling pathways in LPS-induced Raw 264.7 cells. *J. Ethnopharmacol.* **2018**, *210*, 95–106. [CrossRef]
190. Jeong, H.; Sung, M.; Kim, Y.; Ham, H.; Choi, Y.; Lee, J. Anti-inflammatory activity of *Salvia plebeia* R. Br. leaf through heme oxygenase-1 induction in LPS-stimulated RAW264.7 macrophages. *J. Korean Soc. Food Sci. Nutr.* **2012**, *41*, 888–894. [CrossRef]
191. Akram, M.; Syed, A.S.; Kim, K.A.; Lee, J.S.; Chang, S.Y.; Kim, C.Y.; Bae, O.N. Heme oxygenase 1-mediated novel anti-inflammatory activities of *Salvia plebeia* and its active components. *J. Ethnopharmacol.* **2015**, *174*, 322–330. [CrossRef]
192. Jang, H.H.; Lee, S.; Lee, S.J.; Lim, H.J.; Jung, K.; Kim, Y.H.; Lee, S.W.; Rho, M.C. Anti-inflammatory activity of eudesmane-type sesquiterpenoids from *Salvia plebeia*. *J. Nat. Prod.* **2017**, *80*, 2666–2676. [CrossRef]
193. Borges, R.S.; Ortiz, B.L.S.; Pereira, A.C.M.; Keita, H.; Carvalho, J.C.T. *Rosmarinus officinalis* essential oil: A review of its phytochemistry, anti-inflammatory activity, and mechanisms of action involved. *J. Ethnopharmacol.* **2019**, *229*, 29–45. [CrossRef] [PubMed]
194. Zhang, Q.W.; Lin, L.G.; Ye, W.C. Techniques for extraction and isolation of natural products: A comprehensive review. *Chin. Med.* **2018**, *13*, 20. [CrossRef] [PubMed]
195. Albano, S.M.; Miguel, M.G. Biological activities of extracts of plants grown in Portugal. *Ind. Crops Prod.* **2011**, *33*, 338–343. [CrossRef]
196. Abbasloo, E.; Dehghan, F.; Khaksari, M.; Najafipour, H.; Vahidi, R.; Dabiri, S.; Sepehri, G.; Asadikaram, G. The anti-inflammatory properties of *Satureja khuzistanica* Jamzad essential oil attenuate the effects of traumatic brain injuries in rats. *Sci. Rep.* **2016**, *6*, 31866. [CrossRef]
197. Salehi, B.; Mishra, A.P.; Shukla, I.; Sharifi-Rad, M.; Contreras, M.d.M.; Segura-Carretero, A.; Fathi, H.; Nasrabadi, N.N.; Kobarfard, F.; Sharifi-Rad, J. Thymol, thyme, and other plant sources: Health and potential uses. *Phytother. Res.* **2018**, *32*, 1688–1706. [CrossRef]
198. Mahoomodally, M.F.; Devi Dursun, P.; Venugopala, K.N. *Collinsonia canadensis* L. In *Naturally Occurring Chemicals against Alzheimer's Disease*; Elsevier: Amsterdam, The Netherlands, 2021; pp. 373–377. [CrossRef]

199. Aminian, A.R.; Mohebbati, R.; Boskabady, M.H. The Effect of *Ocimum basilicum* L. and its main ingredients on respiratory disorders: An experimental, preclinical, and clinical review. *Front. Pharmacol.* **2022**, *12*, 805391. [CrossRef]
200. Pandur, E.; Micalizzi, G.; Mondello, L.; Horváth, A.; Sipos, K.; Horváth, G. Antioxidant and anti-Inflammatory effects of Thyme (*Thymus vulgaris* L.) essential oils prepared at different plant phenophases on *Pseudomonas aeruginosa* LPS-Activated THP-1 macrophages. *Antioxidants* **2022**, *11*, 1330. [CrossRef]
201. Sun, Z.; Wang, H.; Wang, J.; Zhou, L.; Yang, P. Chemical composition and anti-inflammatory, cytotoxic and antioxidant activities of essential oil from leaves of *Mentha piperita* grown in China. *PLoS ONE* **2014**, *9*, e114767. [CrossRef]
202. Pandur, E.; Balatinácz, A.; Micalizzi, G.; Mondello, L.; Horváth, A.; Sipos, K.; Horváth, G. Anti-inflammatory effect of lavender (*Lavandula angustifolia* Mill.) essential oil prepared during different plant phenophases on THP-1 macrophages. *BMC Complement. Med.* **2021**, *21*, 287. [CrossRef]
203. Kim, K.M.; Kim, S.Y.; Mony, T.J.; Bae, H.J.; Han, S.D.; Lee, E.S.; Choi, S.H.; Hong, S.H.; Lee, S.D.; Park, S.J. *Dracocephalum moldavica* ethanol extract suppresses LPS-induced inflammatory responses through inhibition of the JNK/ERK/NF- κ B signaling pathway and IL-6 production in raw 264.7 macrophages and in endotoxic-treated mice. *Nutrients* **2021**, *13*, 12. [CrossRef]
204. Wang, Y.Y.; Lin, S.Y.; Chen, W.Y.; Liao, S.L.; Wu, C.C.; Pan, P.H.; Chou, S.T.; Chen, C.J. *Glechoma hederacea* extracts attenuate cholestatic liver injury in a bile duct-ligated rat model. *J. Ethnopharmacol.* **2017**, *204*, 58–66. [CrossRef] [PubMed]
205. Tubon, I.; Bernardini, C.; Antognoni, F.; Mandrioli, R.; Potente, G.; Bertocchi, M.; Vaca, G.; Zannoni, A.; Salaroli, R.; Forni, M. *Clinopodium tomentosum* (Kunth) govaerts leaf extract influences in vitro cell proliferation and angiogenesis on primary cultures of porcine aortic endothelial cells. *Oxid. Med. Cell. Longev.* **2020**, *2020*, 298461. [CrossRef] [PubMed]
206. Sheychenko, O.P.; Sheychenko, V.I.; Goryainov, S.V.; Zvezdina, E.V.; Kurmanova, E.N.; Ferubko, E.V.; Uyutova, E.V.; Potanina, O.G.; Fadi, K. Chemical composition and biological activity of the dry extract “Rosmatin” from the herb of *Dracocephalum moldavica* L. *Khimiya Rastitel'nogo Syr'ya* **2021**, *3*, 253–264. [CrossRef]
207. Zotsenko, L.; Kyslychenko, V.; Kalko, K.; Drogovoz, S. The study of phenolic composition and acute toxicity, anti-inflammatory and analgesic effects of dry extracts of some *Elsholtzia* genus (Lamiaceae) species. *Archives* **2021**, *2*, 637–649.
208. Mićović, T.; Katanić Stanković, J.S.; Bauer, R.; Nöst, X.; Marković, Z.; Milenović, D.; Jakovljević, V.; Tomović, M.; Bradić, J.; Stešević, D.; et al. In vitro, in vivo and in silico evaluation of the anti-inflammatory potential of *Hyssopus officinalis* L. subsp. *aristatus* (Godr.) Nyman (Lamiaceae). *J. Ethnopharmacol.* **2022**, *293*, 115201. [CrossRef]
209. Deng, Y.; Hua, J.; Wang, W.; Zhan, Z.; Wang, A.; Luo, S. Cytotoxic terpenoids from the roots of *Dracocephalum taliense*. *Molecules* **2018**, *23*, 57. [CrossRef]
210. Feng, X.; Wang, X.; Liu, Y.; Di, X. Linarin inhibits the acetylcholinesterase activity in-vitro and ex-vivo. *Iran. J. Pharm. Res.* **2015**, *14*, 949–954.
211. Yang, Z.; Qi, J.; Ping, D.; Sun, X.; Tao, Y.; Liu, C.; Peng, Y. *Salvia miltiorrhiza* in thorax and abdominal organ fibrosis: A review of its pharmacology. *Front. Pharmacol.* **2022**, *13*, 9604. [CrossRef]
212. Vergara-Martínez, V.M.; Estrada-Soto, S.E.; Valencia-Díaz, S.; García-Sosa, K.; Peña-Rodríguez, L.M.; de Jesús Arellano-García, J.; Perea-Arango, I. Methyl jasmonate enhances ursolic, oleanolic and rosmarinic acid production and sucrose induced biomass accumulation, in hairy roots of *Lepechinia caulescens*. *Peer J.* **2021**, *9*, 11279. [CrossRef]
213. Ortiz-Mendoza, N.; Aguirre-Hernández, E.; Fragoso-Martínez, I.; González-Trujano, M.E.; Basurto-Peña, F.A.; Martínez-Gordillo, M.J. A review on the ethnopharmacology and phytochemistry of the neotropical sages (*Salvia* subgenus *Calosphace*; Lamiaceae) emphasizing Mexican species. *Front. Pharmacol.* **2022**, *19*, 867892. [CrossRef]
214. Schoch, C.L. NCBI Taxonomy: A Comprehensive Update on Curation, Resources, and Tools. Database (Oxford). 2020. Available online: <https://www.ncbi.nlm.nih.gov/taxonomy> (accessed on 12 December 2022).

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.