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# New data on nrITS phylogeny of *Lotus* (Leguminosae, Loteae)

Galina V. Degtjareva, Tatiana E. Kramina, Dmitry D. Sokoloff, Tahir H. Samigullin, Graeme Sandral & Carmen M. Valiejo-Roman

Summary: Some new nrITS sequences of Lotus are produced and added to the data set analysed in DEGTJAREVA et al. (2006). Lotus burttii and L. filicaulis are revealed as members of the /Lotus corniculatus clade. Lotus conimbricensis is found to be sister to the entire /Lotus corniculatus clade; the /Lotus pedunculatus clade is more distantly related. The New Caledonian Lotus anfractuosus is closest among species sampled to two Australian endemics, though its molecular divergence is considerable. The NE African Lotus torulosus is close to some other red-flowered species from the same region; it does not group with any other Lotus species with dimorphic leaflets. In general, dimorphic vs. monomorphic leaflets is a quite homoplastic character in Lotus. Molecular divergence is weak within the /Pedrosia clade, where the morphological divergence is especially high. In contrast, molecular divergence is considerable but morphological differentiation is weak in the /Lotus corniculatus clade.

*Keywords:* Leguminosae, Loteae, *Lotus*, biogeography, nuclear ribosomal ITS sequences, morphology, phylogeny

The genus *Lotus* includes about 120–130 species native to Europe, Asia, Africa, Australia and some islands of Atlantic Ocean (many endemics in Macaronesia), Pacific Ocean (two endemic species in Ryukyu, Taiwan, New Caledonia, Vanuatu) and Socotra archipelago (two endemic species) in the Indian Ocean. All native New World species formerly placed in *Lotus* are now segregated in four (e.g. ARAMBARRI et al. 2005; SOKOLOFF & LOCK 2005; SOKOLOFF et al. 2007) or two (BROUILLET 2008) distinct genera. In the Old World, three monotypic segregate genera are accepted (*Kebirita, Podolotus* and *Pseudolotus*), while two commonly recognized genera (*Dorycnium* and *Tetragonolobus*) are placed in synonymy of *Lotus* (reviewed in DEGTJAREVA et al. 2006).

Molecular phylogenetic studies of *Lotus* were mainly concentrated on analyses of nrITS sequences (ALLAN & PORTER 2000; ALLAN et al. 2003, 2004; DEGTJAREVA et al. 2006). To-date, nrITS sequences of more than 80 *Lotus* species are published; they represent most traditionally recognized sections and species groups of the genus. This makes *Lotus* one of relatively well-explored legume genera with respect of nrITS phylogeny. Nevertheless, several critical species are not sampled so far, and phylogenetic placement of some other species is intriguing. Therefore, we produced new nrITS sequences for some *Lotus* species.

# Material and Methods

Most nrITS sequences used for phylogenetic analysis and all methods, respectively are the same as described in Degtjareva et al. (2006). Information on new sequences is shown in Table 1. In addition, a GenBank sequence AY294302 of *Lotus mascaënsis* (referred here as 2) is used (first published by Allan et al. 2004).

#### G. V. Degtjareva et al.

Species	Voucher	GenBank number
<i>Lotus anfractuosus</i> (Bak. f.) Kramina & D. D. Sokoloff	New Caledonia, Loyalty Is., Mare, 18.07.1951, <i>Baumann-Bodenheim</i> (Z)	FJ411111
Lotus arinagensis Bramwell	Canary Is., Gran Canaria, Punta Arinaga, 24.03.1996, <i>Royl 778</i> (B)	FJ411112
<i>Lotus burttii</i> Borsos	B-303 (inbred line, plants grown from seeds collected on 03.07.2006 by N. Sandal; the origin of the seeds for establishment of the inbred line: Kabul River, Peshawar, Pakistan)	FJ411113
Lotus conimbricensis Brot. (1)	Spain, 27.04.1966, <i>Segura Zubizarreta</i> <i>960</i> (Z)	FJ411114
Lotus conimbricensis Brot. (2)	Israel, 19.04.1957, Lorch & Grizi 644 (Z)	FJ411115
<i>Lotus filicaulis</i> Durieu	B37 (inbred line, plants grown from seeds collected on 18.09.2002 by N. Sandal; the origin of the seeds for establishment of the inbred line: Algeria, W. F. Grant's collection)	FJ411116
Lotus loweanus Webb & Berthel.	Madeira, Porto Santo, 08.03.1985, <i>Luck</i> <i>MD 33</i> (B)	FJ411117
Lotus mascaënsis Burchard (1)	Canary Is., Tenerife, Masca, 19.03.1985, <i>J. J. &amp; W. Greuter 286</i> (B)	FJ411118
<i>Lotus tetraphyllus</i> Murr.	Baleares, Mallorca, Torrent de Sa Coma de S'Arrom, 10.05.1986, <i>Orell 52</i> (B)	FJ411119
Lotus torulosus (Chiov.) Fiori	cultivated at the Botanical Garden of Moscow University, 2006	FJ411120

Table 1: Voucher information and GenBank accession numbers of new nrITS sequences.

## Results and Discussion

The length of the ITS region (ITS1, 5.8S and ITS2) ranges from 587 to 617 bp for the 108 accessions of the ingroup (i.e. members of the tribe Loteae) and two outgroup (i.e., *Robinia* and *Sesbania*) taxa studied. In the 10 new generated sequences of *Lotus*, the length of the ITS ranges from 588 to 617 bp. The alignment of 110 ITS sequences resulted in a matrix of 599 nucleotide positions after excluding 256 ambiguous positions. Of the remaining sites, 262 characters were parsimony-informative, 82 characters were parsimony-uninformative and 255 were constant.

Maximum parsimony analyses yielded 73000 shortest trees (1496 steps), with a consistency index of 0.4 and a retention index of 0.75. The strict consensus tree based on all shortest trees does not differ significantly in topology from the bootstrap consensus tree (Figs. 1, 2). The topology of the Bayesian tree (Fig. 3) is generally similar to those of the trees inferred from the maximum

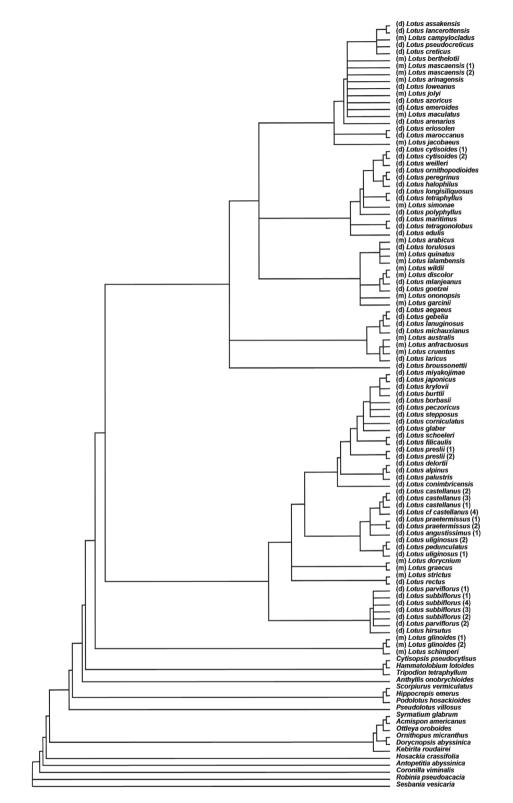


Figure 1: Strict consensus of most parsimonious trees with leaf morphology indicated for *Lotus* species: (m) = leaflets of a leaf are (almost) monomorphic, (d) = leaflets of a leaf are markedly dimorphic.

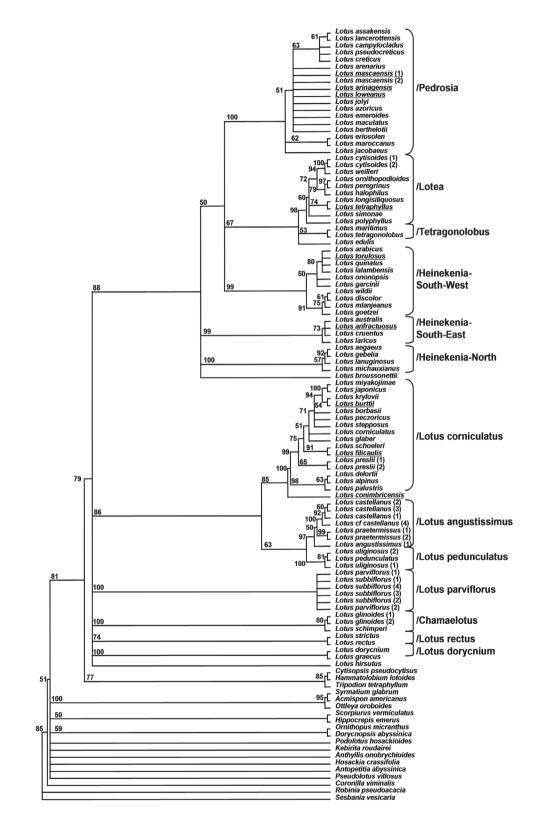


Figure 2: Majority-rule bootstrap consensus tree inferred from the maximum parsimony analysis. Numbers indicate bootstrap support of clades. New nrITS accessions are underlined. Clade names used in the text are outlined.

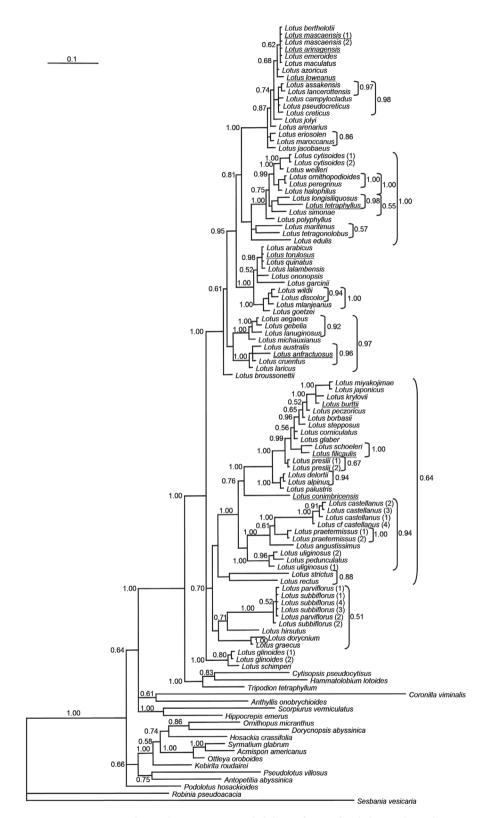


Figure 3: Bayesian tree. Numbers indicate posterior probabilities of particular clades. Nodes with posterior probabilities less than 0.5 are shown as unresolved. Scale = 0.1 substitution/site. New nrITS accessions are underlined.

parsimony analyses. Adding new sequences to the data set analyzed by DEGTJAREVA et al. (2006) does not bring about significant changes of the overall tree topology. The general pattern of phylogenetic relationships within *Lotus* is still the same as revealed by DEGTJAREVA et al. (2006). Therefore, we discuss below details of relationships within some clades.

/Lotus corniculatus clade. The clade comprising the tetraploid *Lotus corniculatus* and its closest predominantly diploid relatives (*L. palustris, L. alpinus, L. delortii, L. preslii, L. schoelleri, L. glaber, L. stepposus, L. peczoricus, L. borbasii, L. krylovii, L. japonicus, L. miyakojimae*) was found to be highly supported in the molecular phylogenetic analysis by DEGTJAREVA et al. (2006). This study added to this clade two more species, namely *L. burttii* and *L. filicaulis*. The chromosome number of both species is the same, 2n = 12 (Sz.-Borsos et al. 1972; CHENG & GRANT 1973).

*Lotus filicaulis* Dur. was described in 1846 from coastal sandy places in Oran and limestone slopes in Mascara (Mediterranean coast in northwestern Algeria). It is characterized by filiform stems and long peduncles, about ten times longer than subtending leaves (BRAND 1898). BRAND considered it as a variety of *Lotus corniculatus* (*L. corniculatus* L. var. *filicaulis* (Durieu) Brand).

*Lotus burttii* Borsos was described in 1972 on the basis of plants grown from seeds collected by Dr. B. L. Burtt (Royal Botanic Garden, Edinburgh) on the 3<sup>rd</sup> May 1958 from plants on the bank of Kabul River, Peshawar, West Pakistan (Sz.-BORSOS et al. 1972). Dr. Burtt identified those plants as *L. corniculatus* L. var. *minor* Bak. The plants of *L. burttii* are small, almost glabrous annual herbs with prostrate or ascending shoots, oblanceolate upper leaflets of a leaf and one- or, rarely, two-flowered umbels. The flowers are small, 7–9 mm long, with primrose to pale pink petals. They are also characterized by non-shattering behavior of pods (KAWAGUCHI et al. 2005).

Our observations on the plants of inbred lines of *L. burttii* B-303 (KAWAGUCHI et al. 2005) and *L. filicaulis* Dur. B-37 (GRANT et al. 1962) did not reveal deep morphological differences between them. Plants of *L. filicaulis* possessed slightly longer peduncles and leaflets than *L. burttii* plants, but the number of flowers in umbel was similar in two species. The petal colour seems to be a variable character in the *Lotus corniculatus* group. We observed pink petal colour more often in *L. burttii* than in *L. filicaulis*, but this character needs a more detailed investigation.

Very complicated relationships among diploid species of *Lotus corniculatus* complex were demonstrated in a series of experimental works carried out by W. F. Grant and collaborators. SOMAROO & GRANT (1971) reported that diploid and self-fertile species such as *L. burttii*, *L. alpinus*, *L. krylovii*, *L. filicaulis*, *L. japonicus*, and *L. schoelleri* could cross more or less with each other in artificial conditions and produce F1 hybrids. The success of crossing was counted as a percent of developed pods from the total number of pollinated flowers. Pollen fertility in hybrids was also analysed. It was shown that species with close geographical distribution crossed worse than species distributed distantly from each other. For example, crosses *L. schoelleri* × *L. filicaulis* had 14% success and pollen fertility was 17% or less. *L. filicaulis* could cross as a male parent with *L. burttii* with 50% success, pollen fertility in this hybrid was 32%. *L. krylovii* was crossed as a female parent with *L. filicaulis*, *L. schoelleri*, and *L. burttii* with success of 41%, 37%, and 10%, respectively. The most geographically distant species *L. japonicus* (as a female parent) produced hybrids with other diploid species more successfully, thus F1 hybrids between *L. japonicus* and

*L. filicaulis*, *L. schoelleri*, *L. burttii* and *L. krylovii* had 57%, 40%, 10–30% and 13% success, respectively (SOMAROO & GRANT 1971), which is negatively correlated with geographical distance between type localities of these species.

Both *L. filicaulis* and *L. burttii* were used as crossing partners to *L. japonicus* in experimental studies aimed to a construction of chromosomal and genetic linkage maps of the model legume *L. japonicus* (PEDROSA et al. 2002; SANDAL et al. 2002; KAWAGUCHI et al. 2005). The occurrence of inversions on chromosomes 1 and 3 between the closely related species *L. filicaulis* and *L. japonicus* was demonstrated, suggesting that these chromosomal rearrangements are early events in speciation of this group (PEDROSA et al. 2002). Besides, *L. filicaulis* has a very high level of DNA polymorphism by AFLP (amplified fragment length polymorphism) analysis (up to 49%) (SANDAL et al. 2002). The general chromosome morphology and genome size of *L. burttii* are similar to those of *L. japonicus*. The translocation event, involving chromosomes 1 and 2, which was observed in *L. miyakojimae*, is not present in *L. burttii*. However, the long arm of chromosome 1 of *L. burttii* was altered in comparison to that of *L. japonicus*. The close proximity between two BAC clones on the long arm of *L. burttii* chromosome 1 is similar to the position of the species and an inversion in the long arm of chromosome 1 might have occurred during the speciation of *L. japonicus* (KAWAGUCHI et al. 2005).

Thus, *L. burttii* and *L. filicaulis* are morphologically rather similar species showing small morphological differences, however, some chromosomal rearrangements exist between them. In phylogenetic trees presented in this paper, *L. filicaulis* is grouped with *L. schoelleri*, and *L. burttii* is placed in a subclade with *L. krylovii*, *L. japonicus* and *L. miyakojimae*. This grouping is well corresponding with geographical distribution of the species, i.e. closely distributed species are clustered together.

Generally, recognition of the highly supported (100% bootstrap support in the parsimony analysis) /Lotus corniculatus clade does not contradict traditional views and morphological data. All species that belong to this clade have been traditionally classified within the section Lotus. The section, in its traditional circumscription (e.g. BALL & CHRTKOVÁ-ŽERTOVÁ 1968; KRAMINA 1999a; see also DEGTJAREVA et al. 2006) comprises both annuals and perennials. Most members of the /Lotus corniculatus clade are perennials or plants that can be either annuals or perennials. In the classification of KRAMINA (1999b) other perennial members of the section Lotus are L. pedunculatus, L. uliginosus, and L. granadensis. Morphologically, these three species are closely related to each other and known as a *Lotus pedunculatus* group. Members of this group have subterranean shoots (stolons) with reduced scaly leaves. In contrast, members of the /Lotus corniculatus clade, with very rare exceptions, do not have subterranean shoots with reduced leaves. Species of *L. uliginosus* group occur in wet and swampy places, which is not typical to species of *L.* corniculatus group. L. uliginosus and related species are characterized by a series of morphological, cytological and biochemical distinctions from *L. corniculatus* group. They have hollow stems, long ciliate trichomes on calyx teeth, small usually one-coloured seeds. It was shown that two pairs of chromosomes of *L. pedunculatus* possessed satellites (ZANDSTRA & GRANT 1968). This species differs from L. corniculatus, L. krylovii and their relatives by phenolic compounds (ZANDSTRA & GRANT 1968) and isoenzymes of leaves and pollen grains (RAELSON & GRANT 1988, 1989). Molecular phylogenetic data show that the *Lotus pedunculatus* group (two of three species are studied so far – the /Lotus pedunculatus clade) is not sister to the /Lotus corniculatus clade; instead, the *Lotus pedunculatus* group appears to be sister to a clade of three annual members of the section *Lotus*, namely the /Lotus angustissimus clade (DEGTJAREVA et al. 2006; this study). This suggests a complex picture of evolution of life form in *Lotus*.

**The** *Lotus angustissimus* **complex and** *Lotus conimbricensis.* The *Lotus angustissimus* complex, as traditionally circumscribed, comprises a group of morphologically similar species of the section *Lotus*. The plants are mostly annual, sometimes biennial or rarely perennial. Morphological diagnostic characters of different species are overlapping, and their identification is sometimes problematic (KRAMINA 2006). Nevertheless, (mostly) annuals of the section *Lotus* do not form a clade on molecular trees. They form two clades that are not closely related: the /Lotus angustissimus clade and the /Lotus parviflorus clade. In the previous analysis (DEGTJAREVA et al. 2006), we used a GenBank accession of *L. conimbricensis*, and this species was found to be a member of the /Lotus parviflorus clade (together with *L. parviflorus* and *L. subbiflorus*). In the current analysis, we used two new sequences of *L. conimbricensis* based on different vouchers. Both new sequences were identical to each other but quite different from the GenBank sequence. In the current phylogeny, *L. conimbricensis* does not group with *L. parviflorus* and *L. subbiflorus*. Instead, it is sister to the /Lotus corniculatus clade.

Traditionally, *L. conimbricensis* was classified within the section *Erythrolotus*. As defined by BRAND (1898), the section *Erythrolotus* is characterized by red (to white) flowers. Members of the section *Lotus* have predominantly yellow flowers. KRAMINA & SOKOLOFF (2003) re-defined the section *Erythrolotus* as comprising only *L. conimbricensis* and placed other red-flowered species to the section *Heinekenia*. The presence of the basic chromosome number x = 6 (instead of x = 7) is an obvious synapomorphy for members of the section *Lotus* plus *L. conimbricensis*; besides, data on biogeography and hair morphology agree with a placement of *L. conimbricensis* among members of the section *Lotus* (see DEGTJAREVA et al. 2006).

Grouping of *L. conimbricensis* with *L. parviflorus* and *L. subbiflorus* agreed with the fact that all three species are annual plants (*L. subbiflorus* is annual to perennial). However, it was curious that *L. conimbricensis* has probably the greatest fruit length/width ratio in the genus *Lotus*, while the fruits of *L. parviflorus* and *L. subbiflorus* are, instead, short. *Lotus parviflorus* is one of few species with shortest fruits in the genus *Lotus*. The current tree topology is more congruent with fruit morphology. Interestingly, the /Lotus parviflorus clade shows weak association with *L. hirsutus*, a species formerly classified as a member of the genus *Dorycnium*. Other former members of the genus *Dorycnium* (*L. dorycnium*, *L. rectus*, *L. graecus*) are in an unresolved position close to the /Lotus parviflorus clade. One of the features used to characterize *Dorycnium* was fruit length: the fruits are generally short in *Dorycnium* and sometimes 1-seeded (*L. dorycnium*). SERINGE (1825) placed both *L. subbiflorus* and *L. parviflorus*, apparently on the basis of short fruits, in the genus *Dorycnium*, a view that was rejected by almost all subsequent authors. The current molecular data show that the SERINGE's view was not completely misleading.

**/Heinekenia clades**. Members of the section *Heinekenia*, as defined by KRAMINA & SOKOLOFF (2003) belong to three highly supported clades. Relationships between these clades and some other clades of *Lotus* are unresolved. As found by DEGTJAREVA et al. (2006) phylogenetic placement of membes of the section *Heinekenia* agrees with pattern of geographical distribution. For this

study, we added two more species (*L. anfractuosus* and *L. torulosus*), and their placement supports the earlier view.

The /Heinekenia-North clade includes *L. aegaeus, L. gebelia, L. lanuginosus, L. michauxianus.* These species occur mainly in the south-western part of the Irano-Turanian floristic region (sensu TAKHTAJAN 1978) and partly in the Arabian peninsula and in the Mediterranean region.

The /Heinekenia-South-East clade comprises *L. laricus* (along around the Persian Gulf to Pakistan), two Australian species (*L. australis* and *L. cruentus*) and the newly sequenced *L. anfractuosus* from New Caledonia. The morphologically close *L. taitungensis* from Japan and Taiwan may also belong to this clade, but no molecular data are available.

The members of the /Heinekenia-South-East clade are characterized by an unique deletion (Fig. 4). All members of this clade have very short ITS sequences (588 bp). Some other species of *Lotus*, such as *L. dorycnium*, *L. garcinii*, *L. graecus* also possess short ITS sequences (587–588 bp), but these species have other deletions, non homologous to the deletion in the /Heinekenia-South-East clade.

The /Heinekenia-South-West clade is most diverse and comprises the rest of the members of the section *Heinekenia* sequenced so far plus two members of the traditional section *Ononidium* (*L. ononopsis, L. garcinii*). The members of this clade occur in Africa, except its Mediterranean part, and in the western and southern part of the Arabian peninsula. Placement of *L. torulosus* (Djibouti, Ethiopia, Eritrea, Sudan) within this clade further supports importance of the biogeographic pattern found by DEGTJAREVA et al. (2006). On the basis of morphology, *L. torulosus* was thought to be relatively close to *L. laricus*, in particular because in both species basal leaflets of a leaf differ in shape from the terminal leaflets and have their maximum length in the basal part. In our phylogenetic trees *L. torulosus* groups with *L. arabicus, L. quinatus* and *L. lalambenis*. These three species (especially *L. quinatus*) are close to *L. torulosus* in terms of biogeography, but differ in leaf morphology (all five leaflets of about the same shape). In general, species of *Lotus* with dimorphic leaflets do not form a clade in our phylogeny. Rather, they are scattered and 'mixed' with species

#### ITS2

Lotus arabicus	CCAACGTCAAACGTCTAGTGCAATGTGGCGTGT
Lotus torulosus	CCAACGTCAAACGTCTAGTGCAATGTGGCGTGT
Lotus quinatus	CCAACGTCAAACGTCTAGTGCAATGTGGCGTGT
Lotus lalambensis	CCAACGTCAAACGTCTAGTGCAATGTGGCGTGT
Lotus ononopsis	CCAACGTCAAACGTTTAGTGCAATGTGGCGTGT
Lotus garcinii	CCAACGTCAAACGTCTAGTGCAATGTGGCGTGT
Lotus wildii	CCAACGTCAAACGTCTAGTGCAATGTGGCGTGT
Lotus discolor	CCAACGTCAAATGTCTAGTGCAATGTGGCGTGT
Lotus mlanjeanus	CCAACGTCAAACGTCTAGTGCAATGTGGCGTGT
Lotus goetzei	CCAACGTCAAACGTCTAGTGCAATGTGGCGTGT
Lotus australis	CCAATGCCAATATGGCGTGT
Lotus anfractuosus	CCGATGCCAATATGGCGTGT
Lotus cruentus	CCAATGCCAATATGGCGTGT
Lotus laricus	CCAATGCCAATATGGCGTGT
Lotus aegaeus	CCAACGCCAAACGCCTAGTGCAATGTGGCGTGT
Lotus gebelia	CCAACGCCAAACGCCTAGTGCAATGTGGCGTGT
Lotus lanuginosus	CCAACGCCAAATGCCTAGTGTAATGTGGCGTGT
Lotus michauxianus	CCAACGCCAAACGCCTAGTGCAATGTGGCGTGT

Figure 4: A portion of the alignment showing a deletion that characterized the /Heinekenia-South-East clade.

having monomorphic leaflets (Fig. 1). At the higher level, a tendency to have dimorphic leaflets is a very important feature of the genus *Lotus*. However, this character is quite homopolastic within *Lotus* and does not represent an unequivocal synapomorphy of the genus.

/Lotea clade. This clade was found by DEGTJAREVA et al. (2006) as including all members of the section Lotea (except L. creticus) plus L. simonae. The present analysis added to this clade L. tetraphyllus. Traditionally, L. tetraphyllus has been classified within a monospecific section Quadrifolium (BRAND 1898; BALL & CHRTKOVÁ-ŽERTOVÁ 1968; VALDÉS 2000). Our data suggest that L. tetraphyllus can be placed within the section Lotea. It seems that presence of four rather than five leaflets is the only significant morphological difference between sections Lotea and Quadrifolium. BRAND (1898) described the section Quadrifolium within the framework of his general ideas on classification of *Lotus*. Since the vast majority of *Lotus* species constantly possess five leaflets, BRAND (1898) considered this character as an important one and therefore segregated species of his subgenus *Edentolotus* with leaflet numbers other than five into two separate sections. All four species with three leaflets that were known at the time of BRAND (1898) formed the section Ononidium, while the only species with four leaflets (L. tetraphyllus) formed the section Quadrifolium. Molecular data showed that Lotus species with three leaflets do not form a natural monophyletic group. Two species constantly showing presence of three leaflets studied so far (Lotus ononopsis and L. simonae) do not form a monophyletic group (DEGTJAREVA et al. 2006). Besides, leaflet number varies in some Lotus species (3-5 in L. brunneri, L. jacobaeus, L. garcinii and L. quinatus, 4-5 in L. hebranicus, 6-9 in L. benoistii, 5-9 in members of the section *Rhyncholotus*); some *Lotus* species that normally possess five leaflets (*L. taitungensis* [= *L. pacificus*], L. corniculatus, L. weilleri, L. australis, L. cruentus) sporadically produce leaves with other leaflet numbers (Lassen 1986; Sokoloff 2003; Kramina & Sokoloff 2004; Sandral et al. 2006). Finally, leaves with five leaflets can be rarely found in L. tetraphyllus (VALDÉS 2000; KRAMINA pers. obs.). In summary, leaflet number cannot be effectively used as a criterion for sectional delimitation in Lotus. In the re-circumscribed section Lotea, most species have five, one species (L. tetraphyllus) has four, and one species (L. simonae) has three leaflets.

**/Pedrosia clade.** This clade comprises all members of sections *Pedrosia* and *Rhyncholotus* studied so far. Monophyly of this clade and its detailed phylogeny were discussed by ALLAN et al. (2004). Although *Pedrosia* and *Rhyncholotus* have been traditionally accepted as two subgenera or sections (see MONOD 1980 and SANDRAL et al. 2006, for discussion), molecular data show that *Rhyncholotus* is derived from the paraphyletic *Pedrosia*. The section *Rhyncholotus* is endemic to Canary Islands, while the section *Pedrosia* has main diversity in Canary Islands, Madeira, Cape Verde and Morocco (SANDRAL et al. 2006).

An important gap in molecular sampling of *Pedrosia* was the absence of data on *Lotus loweanus*, which is endemic to the Island of Porto Santo (Madeira). This is probably the most remarkable species of the section. It differs from the rest of the section by a combination of such features as very short one-flowered peduncles (flowers appear to be solitary in axils of foliage leaves), blackish-purple to pink flowers, hairy outer surface of the standard, fruits densely pubescent along whole surface of valves and usually with one or several very deep constrictions between seeds (SANDRAL et al. 2006). Our present phylogenetic data clearly confirm placement of *L. loweanus* in the /Pedrosia clade.

Two other new sequences of the section *Pedrosia* produced for this paper are those of *L. arinagensis* and L. mascaënsis. These two species have been analyzed by ALLAN et al. (2004). We sequenced them again because we were intrigued by the fact that they formed an unresolved position with species of the section Rhyncholotus (ALLAN et al. 2004). The section Rhyncholotus differs significantly from *Pedrosia* in floral morphology. The new sequence of *L. arinagensis* is identical to the sequence published by ALLAN et al. (2004). In contrast, the sequence of L. mascaënsis slightly differs from the previously published one. Figure 5 shows a portion of alignment of nrITS sequences of *L. mascaënsis* and its closest relatives of the /Pedrosia clade. For this figure, only those parts of the alignment are selected that possess parsimony informative substitutions. Sites with unique substitutions are not considered. In contrast to the previously published sequence (Lotus mascaënsis 2), our new sequence (Lotus mascaënsis 1) shows two polymorphic sites (Y = C/T). In the first site, where Lotus mascaënsis 1 is polymorphic, most examined members of the tribe Loteae have C, while T is much less common (though the presence of T is not unique for the /Pedrosia clade). In the second site, where Lotus mascaënsis 1 is polymorphic, most members of the tribe Loteae have G, while T is less common; the presence of C is recorded only from within the /Pedrosia clade and from a totally unrelated monotypic Kebirita.

The discovery of variation in nrITS sequences of *L. mascaënsis* is important because of a very limited distribution range of this species, which is restricted to just one valley (Valle de Masca) on Tenerife. Very few collections of this species have been made. Interestingly, within these few collections a morphological variation is also observed (leaf rachis present or absent). Presence/ absence of a leaf rachis is an important taxonomic character in the section *Pedrosia* (SANDRAL et al. 2006). Morphological and molecular variation may suggest possible hybrid origin of *L. mascaënsis* or an increased evolution rate in a small population. *L. mascaënsis* is an endangered species (BRAMWELL & BRAMWELL 2001). SANDRAL et al. (2006) suggested that the species might be even extinct in the wild.

Apart from incongruence between the two sequences of *Lotus mascaënsis*, Figure 5 shows absence of any differences in this portion of the alignment between species of the section *Rhyncholotus* (e.g. *L. maculatus, L. berthelotii*) and their closest relatives of the section *Pedrosia* (*L. arinagensis, L. mascaënsis, L. emeroides*). This shows that nrITS sequences are not enough informative for detailed phylogeny reconstruction within the /Pedrosia clade.

ITS 1

ITS 2

Lotus berthelotii	TTAGTTGTACTCTTTGCCCC-AACGCCAAATGCCTAGTGCTATGTGGTGGCTGGGGCGA
Lotus mascaensisl	TTAGTTGTACTCTTTGCCCC-AACGYCAAATGCCTAGTGCTATGTGGTGGYTGGGGCGA
Lotus mascaensis2	TTAGTTGTACTCTTTGCCCC-AACGCCAAATGCCTAGTGCTATGTGGTGGCTGGGGCGA
Lotus arinagensis	TTAGTTGTACTCTTTGCCCC-AACGCCAAATGCCTAGTGCTATGTGGTGGCTGGGGCGA
Lotus emeroides	TTAGTTGTACTCTTTGCCCC-AACGCCAAATGCCTAGTGCTATGTGGTGGCTGGGGCGA
Lotus maculatus	TTAGTTGTACTCTTTGCCCC-AACGCCAAATGCCTAGTGCTATGTGGTGGCTGGGGCGA
Lotus azoricus	TTAGTTGTACTCTTTGCCCC-AACGCCAAATGCCTAGTGCTATGTGGTGGTTGGGGCGA
Lotus loweanus	TTAGTTGTACTCTTTGCCCC-AACGTCAAATGCCTAGTGCTATGTGGTGGTTGGGGCGA
Lotus assakensis	TTAGTTGCACTCTTTGCCCCCAACATCAAATGCCTTGTGCTATGTGGTGGTTGGGGGCGA
Lotus lancerottensis	TTAGTTGCACTCTTTGCCCCCAACATCAAATGCCTTGTGCTATGTGGTGGTTGGGGGCGA
Lotus campylocladus	TTAGTTGYACTCTTTGCCCC-AACATCAAATSCCTAGTGCTATGTGGTGGTTGGGGCGA
Lotus pseudocreticus	TTAGTTGCACTCTTTGCCCC-AACATCAAATGCCTAGTGCTATGTGGTGGTTGGGGGCGA
Lotus creticus	TTAGTTGCACTCTTTGCCCC-AACATCAAATGCCTAGTGCTATGTGGTGGTGGGGGGGGA

Figure 5: A portion of the alignment showing differences between the two sequences of *Lotus mascaënsis* and sequences of its closest relatives of the /Pedrosia clade. See text for the details.

Unequal levels of molecular differentiation within different clades. Data on nrITS sequences show considerable differences between patterns of molecular variation within different clades. In particular, differences between /Pedrosia and /Lotus corniculatus clades are remarkable. Morphologically, the /Pedrosia clade is extremely diverse, with taxa differentiated in such substantive characters as corolla shape (as an adaptation to insect versus bird pollination), presence/absence of hairs on standard, presence/absence of leaf rachis, leaflet shape (all range of leaflet length/width ratio, also leaflets dimorphic vs. monomorphic), flower colour (all range from dark brown to pale yellow), growth form (annuals to shrubs), inflorescence architecture (including differences in plant axiality), umbel peduncle length, fruit shape, presence/absence of hairs on fruits (e.g. SANDRAL et al. 2006). Nevertheless, very little resolution is found in the nrITS trees within the /Pedrosia clade. No clade with bootstrap support more than 80% is found within the /Pedrosia clade. In contrast, species of the /Lotus corniculatus clade differ in such quantitative characters as leaflet shape (the range of variation here is much narrower than in /Pedrosia), flower size, calyx shape, pubescence of the calyx and vegetative parts of the plant, and some others (e.g. KRAMINA 1999a, b). In terms of nrITS sequences, differentiation within the /Lotus corniculatus clade is considerable, and several well-supported subclades can be identified. This is unexpected from a group with reportedly occurrence of limited or wide hybridization in wild populations (e.g. GAUTHIER et al. 1998; KRAMINA 2000).

Within the /Pedrosia clade, molecular differences between mainland and insular taxa (from the Azores, Madeira, Canary Is., and Cape Verde) are low (see also ALLAN et al. 2004). In contrast, *L. anfractuosus* from New Caledonia differs considerably in nrITS sequence from its closest relatives in Australia (*L. australis* and *L. cruentus*). This is clear because of a relatively long branch leading to *L. anfractuosus* in Figure 3.

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## References

- ALLAN G. J. & PORTER J. M. (2000): Tribal delimitation and phylogenetic relationships of Loteae and Coronilleae (Faboideae: Fabaceae) with special reference to *Lotus*: evidence from nuclear ribosomal ITS sequences. – Amer. J. Bot. 87: 1871–1881.
- ALLAN G. J., ZIMMER E. A., WAGNER W. L. & SOKOLOFF D. D. (2003): Molecular phylogenetic analyses of tribe Loteae (Leguminosae): implications for classification and biogeography. In: KLITGAARD B. B. & BRUNEAU A. [eds.]: Advances in Legume Systematics. Pt. 10, Higher Level Systematics: 371–393. Kew: Royal Botanic Gardens.
- ALLAN G. J., FRANCISCO-ORTEGA J., SANTOS-GUERRA A., BOERNER E. & ZIMMER E. A. (2004): Molecular phylogenetic evidence for the geographic origin and classification of Canary Island *Lotus* (Fabaceae: Loteae). – Molec. Phylog. Evol. **32**: 123–138.
- ARAMBARRI A. M., STENGLEIN S. A., COLARES M. N. & NOVOA M. C. (2005): Taxonomy of the New World species of *Lotus* (Leguminosae: Loteae). Austral. Journ. Bot. 53: 797–812.
- BALL P. W. & CHRTKOVÁ-ŽERTOVÁ A. (1968): Lotus L. In: TUTIN T. G., HEYWOOD V. H., BURGES N. A., MOORE D. M., VALENTINE S. M. & WEBB D. A. [eds.]: Flora Europaea. Vol. 2: 173–176. – Cambridge: Cambridge University Press.

- BRAMWELL D. & BRAMWELL Z. (2001): Wild Flowers of the Canary Islands. [2<sup>nd</sup> ed.] Madrid: Editorial Rueda.
- BRAND A. (1898): Monographie der Gattung Lotus. Bot. Jahrb. Syst. 25: 166–232.
- **BROUILLET L.** (2008): The taxonomy of north American *Loti* (Fabaceae: Loteae): new names in *Acmispon* and *Hosackia* J. Bot. Res. Inst. Texas 2: 387–394.
- CHENG R. I.-J. & GRANT W. F. (1973): Species relationships in the *Lotus corniculatus* group as determined by karyotype and cytophotometric analyses. Can. J. Genet. Cytol. **15**: 101–115.
- DEGTJAREVA G. V., KRAMINA T. E., SOKOLOFF D. D., SAMIGULLIN T. H., VALIEJO-ROMAN C. M. & ANTONOV A. S. (2006): Phylogeny of the genus *Lotus* (Leguminosae, Loteae): evidence from nrITS sequences and morphology Can. J. Bot. 84: 813–830.
- GAUTHIER P., LUMARET R. & BÉDÉCARRATS A. (1998): Genetic variation and gene flow in Alpine diploid and tetraploid populations of *Lotus (L. alpinus* (D.C.) Schleicher/*L. corniculatus* L.). II. Insights from RFLP of chloroplast DNA. – Heredity **80**: 694–701.
- GRANT W. F., BULLEN M. R. & DE NETTANCOURT D. (1962): The cytogenetics of *Lotus*. I. Embryocultured interspecific diploid hybrids closely related to *L. corniculatus* L. – Can. J. Genet. Cytol. 4: 105–128.
- KAWAGUCHI M., PEDROSA-HARAND A., YANO K., HAYASHI M., MUROOKA Y., SAITO K., NAGATA T., NAMAI K., NISHIDA H., SHIBATA D., SATO S., TABATA S., HAYASHI M., HARADA K., SANDAL N., STOUGAARD J., BACHMAIR A. & GRANT W. F. (2005): Lotus burttii takes a position of the third corner in the Lotus molecular genetics triangle. – DNA Res. 12: 69–77.
- KRAMINA T. E. (1999a): Taxonomic revision of *Lotus* L. section *Lotus* (Leguminosae) in the territory of European Russia and adjacent countries. PhD Thesis: Moscow State University. [In Russian]
- KRAMINA T. E. (1999b): A contribution to the taxonomic revision of the *Lotus corniculatus* complex (Leguminosae, Loteae) in the European part of the former USSR. – Syst. Geogr. Pl. 68: 265– 279.
- KRAMINA T. E. (2000): A new species of *Lotus* L. (Leguminosae) from *L. corniculatus* L. group Bull. Moscow Soc. Natur., Biol. Ser. 105: 35–40. [In Russian]
- KRAMINA T. E. (2006): A contribution to the taxonomic revision of the *Lotus angustissimus* complex (Leguminosae, Loteae). Wulfenia 13: 57–92.
- KRAMINA T. E. & SOKOLOFF D. D. (2003): On *Lotus* sect. *Erythrolotus* and related taxa (Leguminosae). – Bull. Moscow Soc. Naturalists, Biol. Ser. **108**(5): 59–62. [In Russian]
- Ккаміна Т. Е. & Sokoloff D. D. (2004): A taxonomic study of *Lotus australis* complex (Leguminosae), with special emphasis on plants from Pacific Ocean islands. Adansonia, ser. 3, **26**: 171–197.
- LASSEN P. (1986): Lotus benoistii. Willdenowia 16: 111.
- MONOD T. (1980): Contribution à l'étude des *Lotus* (Papilionaceae) ouest-sahariens et macaronésiens. - Adansonia, ser. 2, 19: 367-402.
- PEDROSA A., SANDAL N., STOUGAARD J., SCHWEIZER D. & BACHMAIR A. (2002): Chromosomal map of the model legume *Lotus japonicus*. Genetics 161(4): 1661–1672.
- RAELSON J. V. & GRANT W. F. (1988): Evaluation of hypotheses concerning the origin of *Lotus corniculatus* using isoenzyme data. Theor. Appl. Genet. **76**: 267–276.
- RAELSON J. V. & GRANT W. F. (1989): An isoenzyme study in the genus *Lotus* (Fabaceae). Experimental protocols and genetic basis of electrophoretic phenotype. Theor. Appl. Genet. 77: 595–607.
- SANDAL N., KRUSELL L., RADUTOIU S., OLBRYT M., PEDROSA A., STRACKE S., SATO S., KATO T., TABATA S., PARNISKE M., BACHMAIR A., KETELSEN T. & STOUGAARD J. (2002): A genetic linkage map of the model legume *Lotus japonicus* and strategies for fast mapping of new loci. Genetics 161(4): 1673–1683.

- SANDRAL G., REMIZOWA M. V. & SOKOLOFF D. D. (2006): A taxonomic survey of *Lotus* section *Pedrosia* (Leguminosae, Loteae) Wulfenia. 13: 97–192.
- SERINGE N. (1825): *Dorycnium, Lotus.* In: DE CANDOLLE A. P.: Prodromus Systematis Naturalis Regni Vegetabilis. P. 2: 208–215. Paris: Treufel et Würtz.
- Soкоloff D. D. (2003): Morphology and classification of the tribe Loteae DC. of the family Leguminosae. – Dr. Sci. Thesis: Moscow State University. [In Russian]
- Sokoloff D. D., Degtjareva G. V., ENDRESS P. K., REMIZOWA M. V., SAMIGULLIN T. H. & VALIEJO-ROMAN C. M. (2007): Inflorescence and early flower development in Loteae (Leguminosae) in a phylogenetic and taxonomic context. – Int. Journ. Plant Sci. **168**: 801–833.
- Sokoloff D. D. & Lock J. M. (2005): Tribe Loteae. In: Lewis G., Schrire B., Mackinder B. & Lock M. [eds.]: Legumes of the world: 455–465. Kew: Royal Botanic Gardens.
- Somaroo B. H. & GRANT W. F. (1971): Interspecific hybridization between diploid species of *Lotus* (Leguminosae). Genetica 42: 353–367.
- Sz.-Borsos O., Somaroo B. H. & Grant W. F. (1972): A new diploid species of *Lotus* (Leguminosae) in Pakistan. Can. J. Bot. **50**: 1865–1870.
- TAKHTAJAN A. L. (1978): The floristic regions of the world. Leningrad: Nauka. [In Russian]
- VALDÉS B. (2000): Lotus L. In: TALAVERA S. et al. [eds.]: Flora Iberica. Vol. 7: 776–812. Madrid: Real Jardin Botanico.
- ZANDSTRA I. I. & GRANT W. F. (1968): The biosystematics of the genus *Lotus* (Leguminosae) in Canada. I. Cytotaxonomy. – Can. J. Bot. **46**: 557–583.

Addresses of the authors:

Dr. Galina V. Degtjareva Botanical Garden of Moscow University Biological Faculty Moscow State University Moscow, 119991 Russia E-mail: degavi@genebee.msu.su

Dr. Tatiana E. Kramina Dr. Dmitry D. Sokoloff Department of Higher Plants Biological Faculty Moscow State University Moscow, 119991 Russia E-mail: kramina@herba.msu.ru sokoloff@dds.srcc.msu.su

Dr. Tahir H. Samigullin Dr. Carmen M. Valiejo-Roman Department of Evolutionary Biochemistry A. N. Belozersky Institute of Physicochemical Biology Moscow State University Moscow, 119991 Russia E-mail: valiejo@genebee.msu.su samigul@belozersky.msu.ru Mr. Graeme A. Sandral NSW Department of Primary Industries Wagga Wagga Agricultural Research Institute Private Mail Bag - Pine Gully Road Wagga Wagga NSW 2650 Australia

E-mail: gsandral@cyllene.uwa.edu.au graeme.sandral@dpi.nsw.gov.au