2	Psoroma spinuliferum (Pannariaceae), a new corticolous lichen species
3	from Alaska with two different types of cephalodia
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13	ABSTRACT. The species <i>Psoroma spinuliferum</i> is described as new to science here. It is only known from the
14	holotype on a <i>Picea sitchensis</i> trunk near a sea-shore in southern, coastal Alaska. The species is distinct in having short,
15	brittle, spinule-like hairs on both apothecium margins, thalline squamules and on pulvinate to coarsely coralloid
16	cephalodia with emerald-coloured Nostoc photobionts. These spinules are unique within Pannariaceae as they are
17	developed on both the chlorobiont and on one of two cyanobionts, but it is uncertain whether they can act as vegetative
18	propagules. The presence of two types of cephalodia is also unique within Pannariaceae. The second type consists of
19	glabrous, small-foliose, geotropically arranged cephalodia, containing a Nostoc strain with cells of an intense ultramarine
20	color, when observed after long storage. The species also has shorter ascospores than <i>Psoroma paleaceum</i> , another hairy
21	species. The hair types of <i>Psoroma hypnorum</i> and <i>P. paleaceum</i> are here by contrast referred to as tomentum and scales,
22	respectively.
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24	KEY WORDS. Taxonomy, biodiversity, North America, <i>Nostoc</i> , photobionts, vegetative propagules.
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27	The genus <i>Psoroma</i> , as defined by its type species <i>P. hypnorum</i> (Vahl) S. F. Gray
28	(Jørgensen 1978), is a <i>Pannariaceae</i> genus, mostly tripartite, consisting of chlorobiont squamules

and cephalodia, interconnected by an inconspicuous hypothallus. The genus has traditionally been interpreted to comprise practically all tripartite species within *Pannariaceae*. However, all foliose species and several squamulose species have now been transferred to other genera, see e.g. Elvebakk et al. (2016). On the other hand, Ekman et al. (2014) transferred six bipartite species of *Pannaria* and *Santessoniella* to *Psoroma*. The genus has apothecia with well-developed thalline excipuli, and asci with IKI+ amyloid, tube-like internal structures. Its color is dominated by dark brown to greyish melanins. TLC-detectable substances are absent, except for pannaric acid and substances related to porphyrilic acid in the *Psoroma tenue* group (Henssen & Renner 1981) and in *P. asperellum* Nyl. and *P. multifidum* P. M. Jørg. (Jørgensen 2004a). Another exception is pannarin in *P. aphthosum* Vain., a surprising occurrence. In *Pannariaceae* this substance in restricted to the genus *Pannaria* and to other even more distantly related genera (Ekman et al. 2014).

Like most genera within *Pannariaceae*, *Psoroma* has its center of biodiversity in the Southern Hemisphere. Øvstedal & Smith (2001) included six species from Antarctica, a number which has now increased to 10, in addition to another four species occurring in subantarctic areas (Park et al. 2018). All these species, except the saxicolous *P. saccharatum* (Scutari & Calvelo 1995; Olech 2004), share the terricolous to muscicolous habit of *P. hypnorum*. However, some corticolous species occur in austral forests, such as *P. aphthosum* in South America (Vainio 1899), and *P. asperellum* Nyl., *P. coralloideum* Nyl., *P. geminatum* P. M. Jørg. and *P. multifidum* P. M. Jørg. in New Zealand (Galloway 2007), partly also in Australia. The species *P. filicicola* P. M. Jørg. & Sipman described from Papua New Guinea by Jørgensen & Sipman (2006) is also corticolous, and confined to stems of tree ferns. In addition, there are some austral, corticolous and squamulose species presently positioned within *Psoroma*, but with deviating chemistry, indicating the need for further studies and revised generic affiliations.

All the four *Psoroma* species known to be bipolar are present in North America. *Psoroma* cinnamomeum Malme could rather be referred to as an amphi-tropical species, as its only locality in the Northern Hemisphere is on one of the Channel Islands in California (Jørgensen 2001). In addition to the widespread *P. hypnorum*, *Psoroma tenue* var. *boreale* Henssen and *P. paleaceum* (Fr.) Timdal

& Tønsberg may both have been overlooked in North America as very few localities have been published so far (Jørgensen 2004b; 2005), whereas ongoing herbarium revisions by the first author indicate that they are regionally common. During field work in Alaska, the second author collected a strange *Psoroma* growing on a *Picea* trunk. On closer inspection, this collection turned out to be very different from all known species in this genus. Except for *P. paleaceum*, it is the only known species with erect hairs in *Psoroma* s. str. The species is also distinct in other characters. The aim of the present paper is to describe this new species, still only known from its holotype collection.

MATERIALS AND METHODS

Herbarium material for this study is housed at BG. Material of the species has not been found during the first author's extensive studies of the Pannariaceae collections in B, BM, C, CANB, O, S, SGO, UPS, W, and WIS. Some reference samples of other species were also studied for comparison. In microscope sections, iodine reactions were tested by adding IKI to mounts pretreated with KOH (Orange et al. 2001). Perispore structures were studied in water mounts and restricted to spores liberated from the asci. Ascospore morphology was studied in detail by drawing detailed sketches of ascospores, and copies of all original drawings have been included with the sample. In an attempt to reproduce true colors of the different parts of the lichen when moist, a painting was made by the first author, after comparisons with colors of moistened fragments of the lichen, 17 years after its collection. Thin-layer chromatography of acetone extracts followed standardized procedures and used solvents A and C (Culberson 1972; Orange et al. 2001). Nomenclature of ascospore structures follows Nordin (1997).

RESULTS

Psoroma spinuliferum Elvebakk & Tønsberg, sp. nov.

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Differs from Psoroma paleaceum in having a cover of small, erect spinules on the sides of the thalline excipuli and on many squamules and on one type of cephalodia, by spinules being circular in cross-section and at bases concolorous with the adjacent parts of the thallus, by having smaller apothecia with shorter ascospores with low verrucae and without apical extensions, and by the occurrence of two different types of cephalodia.

TYPE: USA. ALASKA: City and Borough of Yakutat, Yakutat Foreland, S of village Yakutat, Cannon Beach, 59°29.6'N, 139°43.6'W, alt. 0–10 m, corticolous on trunk of *Picea sitchensis* at upper edge of beach, 29 May 2001, *T. Tønsberg* 29882 (BG-L-70447; holotype).

Description. Thallus squamulose, corticolous, 4–5 cm wide, tripartite. Chlorobiont squamules c. 200 μm thick, pulvinate, entire to very weakly lobate, 0.1–0.3 (–0.5) mm wide, appressed to the substratum, and attached by pale rhizohyphae to the hypothallus. Upper surface pale to ochraceous brown, weakly glossy, commonly, but not uniformly, with numerous erect and brittle, 30–60 μm tall spinules, at bases concolorous with the adjacent thallus, isodiametrical, tapering from a 10–15 μm wide base, initially with long hyphae, later with paraplectenchymatic tissue extending from the cortex, with inclusion of chlorobiont cells in the central lowermost parts of large spinules. Upper cortex 15–25 μm thick, sclerenchymatic, hyaline and paraplectenchymatic; lumina elongate, 2–5 × 2–8 μm; walls 2–4 μm thick. Chlorobiont layer c. 60 μm thick, of cf. Myrmecia cells, globose to irregularly globose, 8–17 μm diam. Medulla 70–100 μm thick; lower cortex absent.

107 Prothallus/hypothallus whitish, forming a weak, but distinct byssoid network.

Cephalodia common, and of two types. The most striking form developes from tiny granules into 0.2–0.8 mm wide, mini-foliose, suberect and geotropically arranged, weakly scalariform, glabrous cephalodia, divided into 0.1 mm broad lobules, and positioned directly on the hypothallus,

in some cases with visible contact with the chlorobiont squamules; lobes c. $100~\mu m$ thick, with both upper and lower cortex layers $10{\text -}15~\mu m$ thick, paraplectenchymatic with lumina $2{\text -}3\times3{\text -}6~\mu m$, walls $2{\text -}3~\mu m$, uppermost part appearing sclerenchymatic. *Cyanobiont Nostoc*, small-celled, blue to ultramarine blue, $3{\text -}4\times3{\text -}6~\mu m$. The other cephalodium type is pulvinate to coarsely coralloid, $0.1{\text -}0.3~m m$ wide, directly attached to the hypothallus, or associated or embedded within the chlorobiont squamules, with spinules like on the chlorobiont squamules, and with cyanobiont cells extending into basal, central parts of the largest spinules; *Nostoc* cells deep sea-green or emerald green to turquoise, $4{\text -}7\times5{\text -}8~\mu m$. Both *Nostoc* types are arranged in $10{\text -}30~\mu m$ large glomeruli or goniocysts without chain structures, with glomeruli filling out most of the medullary layer.

Apothecia common, substipitate, 0.7–1.3 mm wide; disc orange-brown, flat, becoming distinctly convex; thalline excipulum partly lacking, partly with scattered, very low and appressed squamules, densely covered with spinules of the same type as on chlorobiont squamules. Epithecium c. 15 μm thick, sclerenchymatic, pale brown. Hymenium c. 100 μm thick, colourless, but strongly IKI+ blue. Asci clavate, 15×70 –80 μm, with 8 ascospores and with cap to tube-like IKI + blue, apical structures. Proper ascospores hyaline, non-septate, short-ellipsoid, 12.5– 16×8.5 –10.5 μm. Perispores of the same shape, 13– 17×9 –11.5 μm, low-verrucose, no apical extensions seen. Parafyses septate, simple to sparingly branched, c. 2.5 μm thick, apices slightly swollen. Hypothecium light brown, 40–50 μm thick, IKI negative.

Pycnidia not seen.

Chemistry: brownish melanins present, but no TLC-detectable components found.

Habitat ecology: only known from relatively smooth bark of a Picea sitchensis trunk at upper edge of a sandy sea-shore beach.

Distribution: U.S.A., Alaska; only known from the type collection.

Etymology: 'Carrying spinules', referring to the spinules on apothecia, squamules and cephalodia.

Two collections representing rare occurrences of P. hypnorum and P. paleaceum growing on smooth bark were studied in detail for comparison and are cited below. The hair type of P. paleaceum was studied on several collections in addition. These hairs were found to be white when dry, $100-500~\mu m$ long, $20-80~\mu m$ wide at base, unistratose to thin and flattened, transparent when moist, and composed of very long, septate hyphae, connected laterally (Fig. 3A). It is proposed here that they should be referred to as long narrow scales, rather than hairs. Fig. 3C shows the hair types of P. hypnorum, being composed of c. $3~\mu m$ wide hyphae, weak, and often branched and with scattered septae, forming a c. $20~\mu m$ high tomentum.

Additional specimens examined: *Psoroma hypnorum* (Vahl) S.F. Gray: USA. ALASKA: Klondike, U.S.A., Alaska, NNW of Skagway, Klondike Gold Rush National Historical Park, along Chilkoot Trail, N59 39.930 W135 15.912 (NAD27 Alaska), corticolous at base of *Tsuga* snag, 27 July 2008, *T. Tønsberg 38976* (BG). *Psoroma paleaceum* (Nyl.) Timdal & Tønsberg: AUSTRALIA. TASMANIA: Cradle Mountain-Lake St. Clair National Park, S end of Lake St. Clair, 2 km SW of Cynthia Bay along path to Mt. Rufus. 42°7.078'S, 146°8.534'E, 975 m, on basis of an *Eucalyptus* with smooth bark, 22 April 2008, *A. Elvebakk 08:196* (TROM).

DISCUSSION

The new species represents a striking discovery, as it is the first accepted Northern Hemisphere species of a genus otherwise strongly concentrated to austral areas. It also grows on relatively smooth bark, an uncommon habitat among *Psoroma* lichens. In the absence of molecular data, the question therefore arises, does the holotype instead merely represent a habitat modification of a widespread species such as *P. hypnorum* or *P. paleaceum*?

Most collections of both these species look very different from *P. spinuliferum*. However, in two rare cases, where *P. hypnorum* (*Tønsberg 38976*) and *P. paleaceum* (*Elvebakk 08:196*) had been collected from smooth bark, the specimens have an overall similarity to the new species. The

squamules are smaller and paler than in collections from terricolous/muscicolous habitats, apothecia are also more orange-brown and smaller, although significantly larger than those of *P. spinuliferum*, and are flat to weakly convex. These modifications may be a response to a habitat exposed to relatively low light intensities. A closer examination of the collection of the corticolous *P. hypnorum* specimen, which has been published by Spribille et al. (2010), immediately reveals its distinctive squamulose thallus margins, different from both *P. paleaceum* and *P. spinulosum*. The two smoothbark reference collections of *P. hypnorum* and *P. paleaceum* have significantly larger apothecia than *P. spinuliferum*, and also larger ascospores. In *P. spinuliferum* the proper spores are in the range 11–16 μm, in both *P. hypnorum* and *P. paleaceum* they are 16–20 μm. In addition, the two latter have very distinct nodulose apical extensions of the perispores, lacking in *spinuliferum*. Thus the total perispore lengths are 12–17 μm (*P. spinuliferum*), 18–21 μm (*P. hypnorum*) and 19–23 μm (*P. paleaceum*). The spore widths are quite similar in these specimens, although the two latter have higher verrucae.

The hair types of *P. hypnorum* var. *hypnorum* and *P. paleaceum* (as *P. hypnorum* var. *paleaceum*) were illustrated by Jørgensen (1978). The former was shown to have 'short, irregular hairs', the latter 'long, straight hairs' which are '100–300 µm long', 'situated at the uppermost parts', 'on the outer part of the apothecia', 'with somewhat variable density' (Jørgensen 1978). Jørgensen & Kristinsson (2003) stated that two hairy taxa were present in Europe, *P. hypnorum* var. *paleaceum* with glabrous thalline squamules, and *P. hirsutulum* Nyl. ex Crombie differing e.g. by hairy thalli. However, these two taxa were united by Timdal & Tønsberg (2006) as *P. paleaceum*, and our studies confirm that the same type of hairs are frequently also found also on the thalline squamules and on cephalodia, similar to the situation in *P. spinuliferum*. The hair types are very different in these three species and they are referred to here as *scales* in *P. paleaceum*, *spinules* in *P. spinuliferum*, and *tomentum* in *P. hypnorum* (Fig. 3). The tomentum of *P. hypnorum* is very distinct, forming a low mat of branched hyphae, bent towards all directions when seen in the microscope, more erected upwards when dry. *P. paleaceum* scales are long and flat and consistently white when dry. Large spinules of

P. spinuliferum, on the other hand, have a circular cross-section and take on the color of the mycobiont in lower parts, where the photobiont is present.

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The spinules are numerous, stiff and brittle, and break off easily when old. Theoretically, they can act as vegetative propagules also when alive, as well-developed spinules contain both the mycobiont and one of the photobionts. This would then be the first Pannariaceae species which has similar vegetative propagules on both the chlorobiont and on one among two cyanobionts. On the former they are scattered, on the latter they dominate on every cephalodium of the pulvinate to coralloid type, whereas they are totally absent from the small-foliose cephalodia with ultramarine *Nostoc* cells.

The photobiont diversity in *Pannariaceae* is diverse. The family is basically cyanobacterial, however, 30 % of the genera also include tripartite species or green-algal species or are entirely composed of tripartites (Jørgensen & Zhurbenko 2002; Ekman et al. 2014; Elvebakk et al. 2016). Among the latter, there is a high diversity in cephalodium morphology, in several cases with adaptations to vegetative dispersal (James & Henssen 1976; Jørgensen & Wedin 1999; Jørgensen 2004a). There are also several cases where cyanobiont diversity results in morphological diversity. The generitype of the tropical genus Lepidocollema deviates significantly from the other species of the genus by being homoiomerous from having a distinctly chain-celled *Nostoc* strain (Ekman et al 2014). Elvebakk (2016) indicated that the tropical species Lepidocollema polyphyllinum (P. M. Jørg.) P. M. Jørg. has two different cyanobionts, one forming the 'normal' thallus, the other forming cephalodiumlike lobe systems. Recently, the homoiomerous cyanbacterial genus *Kroswia* was shown to be included in Fuscopannaria, a surprising result as the homoiomerous thalli of the former look very different from the heteromerous thalli of the latter (Magain & Sérusiaux 2015). However, P. spinuliferum is the first example of a Pannariaceae lichen, where two clearly anatomically different cyanobionts lead to the formation of two morphologically different types of cephalodia. The two types are obviously attached to a common hypothallus.

Figure 2 illustrates *Psoroma spinuliferum* as a very colourful lichen. The chlorobiont squamules have been given a fresher green colour corresponding to those of other fresh *Psoroma*

species, the colour is less intense in the 17-year old specimen. The cephalodia are illustrated with exact colours, the scalariform cephalodia are truly ultramarine blue, the other type intensely emerald green, less intensive on the outside, although the colours have probably altered and become more intensive after storage. Still, the first author has not observed such intensity in these colours of cyanobionts in numerous herbarium specimens of tripartite Pannariaceae species studied from the Southern Hemisphere. When studied by microscope, cyanobiont diversity appears to be very large in Pannariaceae, a fact which has also been documented genetically, e.g. by Elvebakk et al. (2008) and Magain, & Sérusiaux (2014). However, without striking cyanobiont colors, the two known *P. spinuliferum* specimens known so far are inconspicuous and not easily discovered, and the species may not be common. Only one of the large *Psoroma* collections in North America has so far been studied by the first author, and *P. spinuliferum* was lacking from 218 examined WIS specimens originally determined as *Psoroma hypnorum*, many of these collected in Alaska.

The species grew on a trunk on the leeward side of a narrow forest belt just above a sandy sea-shore beach. Thus, the two holotype specimens were probably exposed to occasional sea-spray, and were obviously in a shaded position and therefore less exposed to drought there than in neighboring habitats.

To conclude, *P. spinuliferum* is basically different from *P. paleaceum* and *P. hypnorum* in its hair types and ascospores. It is also unique in its bicephalodiate cyanobiont structure. Pale and convex apothecia and small, pale squamules are characters shared by rare smooth-bark occurrences of *P. hypnorum* and *P. paleaceum*, and one would therefore predict future collections of *P. spinuliferum* from open habitats to be more robust, without modifications from habitats with low light intensities. Hopefully, the particular cyanobiont structure will be confirmed by studies of future collections, however, a given lichen species might also occur as associated with other photobionts. It should also be added that the genus *Psoroma* is still very insufficiently known world-wide, and a rich material under study by the first author and co-workers will also reveal additional novelties for both North and South America.

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Figure 1. The holotype of *Psoroma spinuliferum*; A) the entire collection, scale bar = 10 mm; B) close-up, scale bar = 5 mm.



Figure 2. A painted presentation of *Psoroma spinuliferum* with colors matching moistened fragments of the holotype, scale bar = 1 mm.

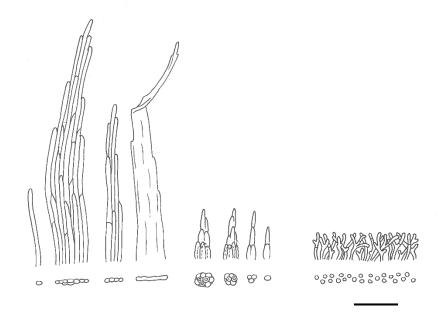


Figure 3. Hair types in *Psoroma*. A) *P. paleaceum* scales (left), B) *P. spinuliferum* spinules (center) and C) *P. hypnorum* tomentum (right), scale bar = $30 \, \mu m$. The illustrations are based on microscoped samples, except one folded *P. paleaceum* scale, drawn from its dry state.

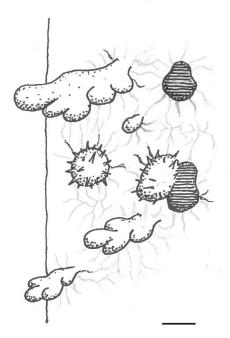


Figure 4. Minifoliose cephalodia with ultramarine Nostoc cells, spinulose cephalodia with emerald green Nostoc, and chlorobiont squamules (hatched) interconnected by hypothalline hyphae, scale bar = 0.1 mm