

The Naturalis collections of pelagic Gastropoda (Mollusca)

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INTRODUCTION

Pelagic (or holoplanktic, see Emiliani, 1991) Gastropoda form a heterogeneous group of marine molluscs belonging to several remote systematic units that strongly differ in anatomical structure, but have an entirely oceanic pelagic life cycle in common. After dying, the shells sink to the seafloor where they are incidentally preserved. The best known group among them are the pteropods (Order Pteropoda), belonging to the subclass Heterobranchia. The second important group is commonly referred to as heteropods, but these are officially classified as Pterotracheoidea, a superfamily of the Order Littorinimorpha, in the subclass Caenogastropoda (Bouchet et al., 2017). The Pterotracheoidea and Pteropoda both include shelled, partially shelled and unshelled species. A few species in the Order Nudibranchia also live a holoplanktic life but a shell is absent, apart from little known larval shells that are

'Van onschatbare waarde ...'

200 years of natural history collections in Naturalis

We present a history of the various collections of pelagic, or holoplanktic Mollusca, available in the Naturalis Biodiversity Center, Leiden The Netherlands. Both for the fossil and present-day collections numbers of available lots, stratigraphic and geographic origin and availability of type specimens are documented. We discuss four issues related to these collections: 1) preservation of pyritised fossils, 2) the backgrounds of the Maltese fossil collection material, 3) a list of all 505 taxa considered valid, 216 of which are currently living species (many of which also have a fossil record) and 4) diagnosis of Vaginellinae subfamily nov.

shed during metamorphosis. Epitoniidae belonging to subclass Caenogastropoda are also included herein, although they are not really holoplanktic but, living in the air-water interface as adults, they are considered neuston.

Shells of pelagic gastropods consist of aragonite, a metastable morph of calcium carbonate. The calcareous shells can be found as fossils, but because of the instability of aragonite and very thin nature of the shells they are frequently poorly preserved or completely dissolved. Internal moulds remain recognisable and identifiable in most cases though. Pterotracheoidea, and particularly the Bellerophoniidae records extend as far back as the Triassic (Teichert & Nützel, 2015; Wall-Palmer et al., 2016; Pieroni & Nützel, 2020). Only few and usually doubtful records of pre-Cenozoic Pteropoda exist (reviewed in Janssen & Peijnenburg, 2017). After the Paleocene-Eocene boundary pteropods show a rapid evolution, developing numerous species (Janssen et al., 2016). Both groups of pelagic gastropods form an important component of the present-day oceanic zooplankton (Bednaršek et al., 2012; Wall-Palmer et al., 2016; Burridge et al., 2017; Buitenhuis et al., 2019).

Currently, holoplanktic molluscs, and especially the shelled pteropods, play an important role in the study of the effects of ocean acidification. In aragonite-undersaturated waters their shells dissolve which makes them potentially valuable indicator species. An extensive number of studies on the topic were performed during the last decennium (reviewed in Manno et al., 2017). This increased interest has simultaneously led to new studies concerning their systematics, biogeography and biostratigraphy, greatly amplifying the existing classical literature of the two last centuries. Modern techniques, such as scanning electron microscopy (SEM), CT-scanning, 3D-printing and especially DNA sequencing have opened up new research opportunities that have led, and will lead in the future, to a better understanding of these particular animal groups and their interrelationships. Two examples are the recent work of Peijnenburg et al. (2020) and Wall-Palmer et al. (2020) which explored the evolution of pteropods and shelled heteropods respectively, using a combination of genetic information and the ages of key fossils. Apart from shedding new light on the phylogenetic relationships in the respective groups, these studies also estimated divergence times. The results were surprisingly similar for the two groups of pelagic gastropods. Both the pteropods and the atlantid heteropods were found to be much older than previously thought, with both groups originating in the Early Cretaceous.

Currently only few universities and research institutes maintain animal systematics as a major topic. However, most of the current studies on pelagic molluscs require correct identifications at species level and therefore collections of material, both dry and preserved in buffered ethanol or formaldehyde solutions, are of utmost importance. It is for

that reason that we present in this paper a survey and quantification of the important collections of these groups represented in the Naturalis Biodiversity Center, comprising both fossil and present-day material in a wide variety and counting up to approximately 25,000 lots, which makes it one of the larger, if not the largest, collection of its kind.

The sections below have been written by groups of authors that are abbreviated as AWJ = Arie W. Janssen; BVDB = Bram van der Bijl; DWP = Deborah Wall-Palmer; JG = Jeroen Goud; KTCAP = Katja T.C.A. Peijnenburg; RP = Ronald Pouwer; FPW = Frank P. Wesselingh.

THE NATURALIS MOLLUSC COLLECTIONS IN GENERAL (AWJ, RP & BVDB)

Naturalis Biodiversity Center in Leiden (The Netherlands) has a complex history. It started in 1990 with the merging of two museums, the 'Rijksmuseum van Natuurlijke Historie' (RMNH) and the 'Rijksmuseum van Geologie en Mineralogie' (RGM), forming the Nationaal Natuurhistorisch Museum (NNM). In 1998 a new museum building was opened and the museum adopted the name Nationaal Natuurhistorisch Museum Naturalis, using Naturalis as the common name. In 2010 another major step was made with the merging of Naturalis with the Zoologisch Museum Amsterdam (ZMA) and the Nationaal Herbarium Nederland, forming the present-day Naturalis Biodiversity Center. The collections of recent molluscs of RMNH and ZMA were merged but the fossil molluscs of RGM were maintained as a separate collection. The two collections are differently organised: the RMNH/ZMA collection is basically stored in systematic order, whereas the RGM collection is arranged according to age and locality. Separation of fossil holoplanktic molluscs as a special, systematically arranged unit, was started in the 1980s by isolating species from the main collection of fossil molluscs.



Fig. 1. Naturalis mollusc collections in steel drawer cabinets.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Bartonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots'
Family Atlantidae																				
<i>Atlanta ariejansseni</i> Wall-Palmer et al., 2016	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	7
<i>Atlanta brunnea</i> Gray, 1850	72	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	107
<i>Atlanta californiensis</i> Seapy & Richter, 1993	3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-
<i>Atlanta diamesa</i> Woodring, 1928	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Atlanta echinogyra</i> Richter, 1972	19	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	+	4
<i>Atlanta fragilis</i> Richter, 1993	2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-
<i>Atlanta frontieri</i> Richter, 1993	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-
<i>Atlanta gaudichaudi</i> Gray, 1850	5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	20
<i>Atlanta gibbosa</i> Souleyet, 1852	2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	5
<i>Atlanta helicinoidea</i> Gray, 1850	39	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	76
<i>Atlanta inclinata</i> Gray, 1850	28	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	56
<i>Atlanta inflata</i> Gray, 1850	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	10
<i>Atlanta lesueurii</i> Gray, 1850	41	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	+	-	+	24
<i>Atlanta lingayanensis</i> Janssen, 2007	12	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Atlanta meteori</i> Richter, 1972	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-
<i>Atlanta oligogyra</i> Tesch, 1906	31	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	+	+	17
<i>Atlanta peronii</i> Lesueur, 1817	60	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	+	+	217
<i>Atlanta plana</i> Richter, 1972	40	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	-	+	3
<i>Atlanta richteri</i> Janssen, 2007	16	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Atlanta rosea</i> Gray, 1850	2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-
<i>Atlanta seapyi</i> Janssen, 2007	8	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Atlanta selvagensis</i> De Vera & Seapy, 2006	42	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	52
<i>Atlanta tokiokai</i> van der Spoel & Troost, 1972	18	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	+	26
<i>Atlanta turriculata</i> d'Orbigny, 1836 ²	22	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	24
<i>Atlanta vanderspoeli</i> Wall-Palmer, Hegmann & Peijnenburg, 2019²	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Atlanta</i> sp.	229	-	-	-	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	50
<i>Atlantidea rotundata</i> (Gabb, 1873)	93	-	-	-	-	-	-	-	+	+	-	-	-	-	-	+	-	+	+	9
<i>Mioatlanta soluta</i> Di Geronimo, 1974	3	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Oxygyrus inflatus</i> Benson, 1835	71	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	+	+	114
<i>Protatlanta kbiraensis</i> Janssen, 2012	22	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-
<i>Protatlanta souleyeti</i> (Smith, 1888)	81	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	-	+	+	97
<i>Protatlanta</i> sp.	12	-	-	-	-	-	-	-	-	+	-	-	-	-	+	-	-	-	-	-
Atlantidae sp.	3	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
Family Pterotracheidae																				
<i>Firoloida desmarestia</i> Lesueur, 1817	52	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	18
<i>Firoloida</i> sp.	4	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-

Table 1. Listing of all holoplanktic mollusc taxa and numbers of lots in the RGM and RMNH/ZMA collections as registered in the two databases, with an indication of their approximate stratigraphic age. Included are 450 taxa (36 of them in open nomenclature), 372 of which are represented in the RGM collection, and 162 in the RMNH/ZMA collection.

In the collection are various taxa that were introduced after 1960 at infrasubspecific rank. Such taxa are basically unavailable under ICZN art. 45.6.3. In Table 1 their number of lots is added to the currently valid species name. Several of these, however, were subsequently validated before 1985 under ICZN art. 45.6.4.1. Availability of such taxon names has to be ascertained individually in each case.

Names printed in **bold** mean that their primary types are in the Naturalis collection. (table continued on pp. 218-228).

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Bartonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots'
<i>Pterotrachea coronata</i> Forsskål in Niebuhr, 1775	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	224
<i>Pterotrachea hippocampus</i> Philippi, 1836	8	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	18
<i>Pterotrachea scutata</i> Gegenbaur, 1855	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	3
<i>Pterotrachea</i> / <i>Pterotracheidae</i> sp. (div. ?)	51	-	+	?	?	-	-	-	-	-	-	-	-	-	-	-	-	+	+	1
Family Carinariidae																				
<i>Cardiapoda placenta</i> (Lesson, 1830)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	18
<i>Carinaria cristata</i> (Linné, 1767)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	12
<i>Carinaria cithara</i> Benson, 1835	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	24
<i>Carinaria galea</i> Benson, 1835	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	3
<i>Carinaria lamarcki</i> Péron & Lesueur, 1810	86	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	+	+	+	28
<i>Carinaria maempeli</i> Janssen, 2012	4	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Carinaria rutschi</i> Robba, 1972	1	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Carinaria</i> sp.	45	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	+	+	2
<i>Pterosoma planum</i> Lesson, 1827	19	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	+	1
<i>Striocarinaria hugardi</i> (Pictet, 1855)	4	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-
<i>Carinariidae</i> sp.	36	-	-	-	-	-	-	-	-	+	+	-	-	-	+	-	-	-	+	-
Family Epitoniidae																				
<i>Janthina exigua</i> Lamarck, 1816	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	59
<i>Janthina globosa</i> Swainson, 1822	3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	+	116
<i>Janthina janthina</i> (Linnaeus, 1758)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	279
<i>Janthina pallida</i> Thompson, 1840	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	36
<i>Janthina umbilicata</i> d'Orbigny, 1841	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	22
<i>Janthina</i> / <i>Epitoniidae</i> sp.	17	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	14
<i>Recluzia lutea</i> (Bennett, 1840)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	14
Family Heliconoididae																				
<i>Heliconoides atypicus</i> (Laws, 1944)	6	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides auriformis</i> (Curry, 1982)	4	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides bartonensis</i> (Curry, 1965)	13	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides curryi</i> (Janssen, 1990)	6	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides daguini</i> Janssen, 2010	2	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides dilatata</i> (von Koenen, 1892)	11	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides ferax</i> (Laws, 1944)	2	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides hodgkinsoni</i> Garvie, 2020	3	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides hospes</i> (Rolle, 1862)	78	-	-	-	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides inflatus</i> (d'Orbigny, 1834)	421	-	-	-	-	-	-	-	-	+	+	+	+	+	+	+	+	+	+	349
<i>Heliconoides lillebaeltensis</i> Janssen, 2007	21	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides linneensis</i> Janssen, 2008	8	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides lunatus</i> (Janssen, 1990)	3	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides mercinensis</i> (Watelet & Lefèvre, 1885)	174	+	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides merlei</i> Janssen, 2010	3	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides mermuysi</i> Janssen, 2010	4	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-

Table 1, continued.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Barthian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots'
<i>Heliconoides nemoris</i> (Curry, 1965)	3	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides nikkieae</i> Janssen, 2017	31	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides nitens</i> (Lea, 1833)	44	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides paula</i> (Curry, 1982)	3	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides planus</i> (Tembrock, 1964)	1	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides pyrenaicus</i> Janssen, 2010	2	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides sondaari</i> Janssen, 2007	20	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Heliconoides stenzeli</i> (Garvie, 1992)	9	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides tatei</i> (Janssen, 1990)	2	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides taylori</i> (Curry, 1965)	33	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides tertiarus</i> (Tate, 1887)	90	-	-	-	-	-	+	+	+	+	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides texanus</i> (Garvie & Hodgkinson, 1992)	8	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides vanderweideni</i> Janssen, 2004	24	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides vonhachti</i> Janssen, 2012	14	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-
<i>Heliconoides wardijaensis</i> Janssen, 2004	3	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
Family Limacinidae																				
<i>Altaspiratella bearnensis</i> (Curry, 1982)	41	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Altaspiratella choctavensis</i> (Aldrich, 1887)	2	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Altaspiratella elongatoidea</i> (Aldrich, 1887)	36	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Altaspiratella gracilens</i> Hodgkinson, 1992	3	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Altaspiratella multispira</i> (Curry, 1882)	13	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Altaspiratella tavianii</i> Janssen, 2013	4	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Currylimacina asperita</i> Garvie, 2020	1	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Currylimacina cossmanni</i> (Curry, 1982)	19	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina acutimarginata</i> (Korobkov, 1966)	9	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina adornata</i> Hodgkinson, 1992	4	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina aegis</i> Hodgkinson, 1992	37	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina andrussowi</i> (Kittl, 1886)	27	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Limacina aryanaensis</i> Janssen, 2013	3	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina asiatica</i> Janssen, 2011	9	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina atlanta</i> (Mörch, 1874)	224	-	-	-	-	-	-	-	-	-	-	+	+	+	+	-	-	-	-	-
<i>Limacina bulimoides</i> (d'Orbigny, 1834)	119	-	-	-	-	-	-	-	+	-	-	-	-	-	+	+	-	+	+	177
<i>Limacina canadaensis</i> Hodgkinson, 1992	6	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina conica</i> (von Koenen, 1892)	5	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina convolutes</i> Hodgkinson, 1992	2	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina dzheroiensis</i> Janssen, 2011	10	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina erasmiana</i> Janssen, 2010	22	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina ernstkitkli</i> Janssen, 2012	34	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina gormani</i> (Curry, 1982)	15	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina gramensis</i> (Rasmussen, 1968)	95	-	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-
<i>Limacina guersi</i> Janssen, 2010	23	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina helicina antarctica</i> Woodward, 1854	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	39
<i>Limacina helicina helicina</i> (Phipps, 1774)	2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	6

Table 1, continued.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Bartonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots'	
<i>Limacina helicina rangii</i> (d'Orbigny, 1834)	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	
<i>Limacina helicina pacifica</i> Dall, 1871	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Limacina ingridae</i> Janssen, 1989	173	-	-	-	-	-	-	-	-	-	?	+	-	-	-	-	-	-	-	-	-
<i>Limacina irisae</i> Janssen, 1989	48	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Limacina karasawai</i> Ando, 2011	11	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina lesueurii</i> (d'Orbigny, 1836)	49	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	130
<i>Limacina lotschi</i> (Tembrock, 1989)	26	-	?	+	?	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina mariae</i> Janssen, 1989	35	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina novacaesarea</i> Janssen & Sessa, 2016	6	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina perforata</i> Janssen, 2013	2	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina pygmaea</i> (Lamarck, 1805)	42	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina retroversa retroversa</i> (Fleming, 1823)	60	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	50
<i>Limacina retroversa australis</i> (Eydoux & Souleyet, 1840)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	3
<i>Limacina robusta</i> (Eames, 1952)	10	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina smithvillensis</i> Hodgkinson, 1992	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina tanzaniaensis</i> Janssen, 2017	4	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina tarchanensis</i> (Kittl, 1886)	1	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Heliconoides taylori</i> (Curry, 1965)	33	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina timi</i> Janssen, 2017	20	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina trochiformis</i> (d'Orbigny, 1834)	84	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	9
<i>Limacina tutelina</i> (Curry, 1965)	15	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina ujiharai</i> Shibata, 1983	1	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina umbilicata</i> (Bornemann, 1855)	73	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina valvatina</i> (Reuss, 1867)	626	-	-	-	-	-	-	+	+	+	+	-	-	-	-	-	-	-	-	-	-
<i>Limacina vegrandis</i> Janssen, 2010	19	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina wechesensis</i> Hodgkinson, 1992	6	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina wilhelminae</i> Janssen, 1989	57	-	-	-	-	-	-	-	-	-	-	+	+	+	+	-	-	-	-	-	-
<i>Limacina yazdii</i> Janssen, 2013	4	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Limacina</i> sp.	131	-	+	+	+	+	+	+	+	+	+	+	+	-	+	+	-	-	-	-	57
<i>Striolimacina andaensis</i> Janssen, 2007	15	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-
<i>Striolimacina imitans</i> (Collins, 1934)	8	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-
Limacinidae sp.	62	-	+	+	-	+	+	+	+	+	-	-	-	-	+	+	-	-	-	+	-
Family Thieleidae																					
<i>Thielea helicoides</i> (Jeffreys, 1877)	4	-	-	-	-	-	-	-	-	-	-	-	+	-	-	+	-	-	+	-	10
Family Creseidae																					
<i>Boasia chierchiai</i> (Boas, 1886) s.lat.	64	-	-	-	-	-	-	-	-	+	-	-	-	-	-	+	-	+	+	-	2
<i>Bovicornu eocenense</i> Meyer, 1886	19	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Bovicornu gracile</i> Meyer, 1887	1	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Bowdenathea jamaicensis</i> Collins, 1934	18	-	-	-	-	-	-	-	-	-	-	-	+	-	+	+	-	-	-	-	-
<i>Bowdenathea miocenica</i> Janssen, 2004	14	-	-	-	-	-	+	?	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Bucanoides basiannulata</i> Hodgkinson, 1992	1	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Bucanoides divaricata</i> Hodgkinson, 1992	2	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Bucanoides tenuis</i> Hodgkinson, 1992	7	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

Table 1, continued.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Bartonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots'
<i>Camptocerotops americanus</i> Garvie, 1992	8	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Camptocerotops priscus</i> (Godwin-Austen, 1882)	92	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Cheilospicata cedrus</i> Garvie, 2020	2	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Cheilospicata repanda</i> Hodgkinson & Garvie, 1992	5	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Creseis acicula</i> (Rang, 1828)	111	-	-	-	-	-	-	-	-	-	-	-	-	+	+	+	+	+	+	156
<i>Creseis antoni</i> Janssen, 2010	3	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Creseis berthae</i> Janssen, 1989	19	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Creseis conica</i> Eschscholtz, 1829	80	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	32
<i>Creseis corpulenta</i> (Meyer, 1887)	3	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Creseis curta</i> Janssen, 2012	22	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Creseis cylindrica</i> Hodgkinson, 1992	10	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Creseis roesti</i> Janssen, 2010	28	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-
<i>Creseis simplex</i> (Meyer, 1886)	15	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Creseis spina</i> (Reuss, 1867)	120	-	-	-	+	+	+	+	+	+	+	-	-	-	+	-	-	-	-	-
<i>Creseis tugurii</i> Janssen, 2010	5	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Creseis virgula</i> (Rang, 1828)	34	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	99
<i>Creseis</i> sp.	40	-	-	-	-	-	+	+	+	+	-	-	-	-	+	-	-	-	+	14
<i>Euchilotheca elegans</i> Harris, 1894	55	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Euchilotheca ganensis</i> Curry, 1982	8	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Euchilotheca succincta</i> (Defrance, 1828)	14	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Euchilotheca</i> sp.	2	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Loxobidens aduncus</i> Hodgkinson, 1992	1	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Styliola schembriorum</i> Janssen, 2012	21	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-
<i>Styliola subula</i> (Quoy & Gaimard, 1827)	423	-	-	-	-	-	+	+	+	+	+	+	+	+	+	+	+	+	+	264
<i>Styliola</i> sp.	1	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Thecopsella fischeri</i> Cossmann, 1888	2	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Tibiella annulata</i> Garvie, 1992	2	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Tibiella marshi</i> Meyer, 1884	4	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Tibiella reflexa</i> Hodgkinson, 1992	1	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Tibiella texana</i> Collins, 1934	1	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Tibiella watupuruensis</i> Janssen, 2013	4	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Creseidae sp.	16	-	+	+	-	-	+	-	-	-	+	-	-	-	-	-	-	-	-	-
Family Hyalocylidae																				
<i>Hyalocylis marginata</i> Janssen, 2007	22	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Hyalocylis striata</i> (Rang, 1828)	79	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	60
<i>Praehyalocylis cincta</i> (von Koenen, 1892)	5	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Praehyalocylis maxima</i> (Ludwig, 1864)	32	-	-	-	+	+	+	?	-	-	-	-	-	-	-	-	-	-	-	-
<i>Praehyalocylis</i> sp.	4	-	-	-	-	-	+	+	-	+	+	-	-	-	-	-	-	-	-	-
Family Praeacuvierinidae																				
<i>Praeacuvierina lura</i> (Hodgkinson, 1992)	1	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Texacuvierina guta</i> (Hodgkinson, 1992)	1	-	-	?	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Texacuvierina hodgkinsoni</i> Janssen, 2013	3	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

Table 1, continued.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Barntonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots'
Family Cuvierinidae																				
<i>Cuvierina astesana</i> (Rang, 1829)	36	-	-	-	-	-	-	-	-	-	-	-	-	?	+	+	-	-	-	-
<i>Cuvierina atlantica</i> Bé, MacClintock & Currie, 1972	3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	140
<i>Cuvierina cancapae</i> Janssen, 2005	3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	25
<i>Cuvierina columnella</i> (Rang, 1827)	5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	30
<i>Cuvierina curryi</i> Janssen, 2005	19	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-
<i>Cuvierina grandis</i> d'Alessandro & Robba, 1981	19	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-
<i>Cuvierina inflata</i> (Bellardi, 1873)	14	-	-	-	-	-	-	-	-	-	-	-	+	+	-	+	-	-	-	-
<i>Cuvierina intermedia</i> (Bellardi, 1873)	13	-	-	-	-	-	-	-	-	-	+	+	+	-	+	-	-	-	-	-
<i>Cuvierina jagti</i> Janssen, 1995	5	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-
<i>Cuvierina ludbrooki</i> (Caprotti, 1962)	5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Cuvierina pacifica</i> Janssen 2005	9	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Cuvierina paronai</i> Checchia-Rispoli, 1921	122	-	-	-	-	-	-	-	-	-	+	+	+	-	-	-	-	-	-	-
<i>Cuvierina torpedo</i> (Marshall, 1918)	1	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Cuvierina tsudai</i> BurrIDGE et al., 2016	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	6
<i>Cuvierina urceolaris</i> (Mörch, 1850)	29	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	+	14
<i>Cuvierina</i> sp.	20	-	-	-	-	-	-	-	+	-	-	-	+	-	-	-	-	+	-	-
<i>Ireneia calandrellii</i> (Michelotti, 1847)	27	-	-	-	-	-	-	-	+	+	+	+	+	-	-	-	-	-	-	-
<i>Ireneia gracilis</i> Janssen, 2005	22	-	-	-	-	-	-	-	+	+	+	+	+	-	-	-	-	-	-	-
<i>Ireneia marqueti</i> Janssen, 1995	5	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-
<i>Ireneia nieulandei</i> Janssen, 1995	1	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Ireneia striatocarinata</i> Piehl, 2007	4	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Ireneia tenuistriata</i> (Semper, 1861)	47	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Ireneia testudinaria</i> (Michelotti, 1847)	3	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Johnjagtia baharensis</i> Janssen, 2012	7	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Johnjagtia moulinsi</i> (Benoist, 1873)	11	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Spoelia torquayensis</i> Janssen, 1990	59	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
Family Cliidae																				
<i>Clio aichinoi</i> Checchia-Rispoli, 1921	30	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Clio antarctica</i> Dall, 1908	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	232
<i>Clio bellardii</i> Audenino, 1899	48	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Clio berglundorum</i> Squires, 1989	7	-	-	-	-	-	+	?	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio blinkae</i> Janssen, 1989	53	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio braidensis</i> (Bellardi, 1873)	24	-	-	-	-	-	-	-	-	-	-	-	+	+	+	-	-	-	-	-
<i>Clio caralitana</i> Robba & Spano, 1978	38	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Clio carinata</i> Audenino, 1899	15	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Clio chadumica</i> Korobkov, 1966	2	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio chaptalii</i> Gray, 1850	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	3
<i>Clio coebana</i> Robba, 1972	3	-	-	-	-	-	+	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Clio convexa</i> (Boas, 1886) s.lat.	61	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	-	+	+	5
<i>Clio cuspidata</i> (Bosc, 1801)	85	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	+	+	89
<i>Clio deflexa</i> von Koenen, 1882	1	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Clio distefanoi</i> Checchia-Rispoli, 1921	94	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-

Table 1, continued.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Barntonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots'
<i>Clio fallauxi</i> (Kittl, 1886)	2	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Clio gargarica</i> Sirna, 1968	2	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Clio ghawdextensis</i> Janssen, 2004	25	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio giulioi</i> Janssen, 1995	17	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-
<i>Clio guidottii</i> Simonelli, 1896	11	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-
<i>Clio hataii</i> (Noda, 1972)	2	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-
<i>Clio itoigawai</i> Shibata, 1983	1	-	-	-	-	-	-	-	?	?	-	-	-	-	-	-	-	-	-	-
<i>Clio jacobae</i> Janssen, 1989	2	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio lozoueti</i> Janssen, 2010	3	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio lucai</i> Janssen, 2000	10	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-
<i>Clio merijni</i> Janssen, 2012	4	-	-	-	-	-	+	?	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio multicostata</i> (Bellardi, 1873)	3	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Clio nielseni</i> Janssen, 1990	7	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio oblonga</i> Rampal, 1996	7	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	-
<i>Clio pauli</i> Janssen, 1989	2	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Clio pedemontana</i> (Mayer, 1868)	87	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-
<i>Clio piatkowski</i> van der Spoel, Schalk & Bleeker, 1992	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	384
<i>Clio polita</i> Pelseneer, 1888	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	3
<i>Clio pulcherrima</i> (Mayer, 1868)	109	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Clio pyramidata angusta</i> (Boas, 1886)	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-
<i>Clio pyramidata pyramidata</i> Linné, 1767³	267	-	-	-	-	-	-	-	-	?	-	+	+	+	+	+	+	+	+	455
<i>Clio pyramidata tyrrhenica</i> Janssen, 2012	10	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	?	-
<i>Clio pyramidata</i> Linné, 1767 s.lat.	10	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2312
<i>Clio recurva</i> (Children, 1823)	3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	33
<i>Clio riccioli</i> (Calandrelli, 1844)	3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Clio "robbaei</i> Buonaiuto, 1979" (unpublished name)	2	-	-	-	?	?	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio saccoi</i> Checchia-Rispoli, 1921	43	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-
<i>Clio scheelei</i> (Munthe, 1888)	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-
<i>Clio sinuosa</i> (Bellardi, 1873)	4	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Clio sturarii</i> Robba, 1977	2	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Clio triplicata</i> Audenino, 1899	27	-	-	-	-	-	-	+	+	+	-	-	-	-	-	-	-	-	-	-
<i>Clio vasconiensis</i> Janssen, 2010	12	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio vilis</i> Janssen, 2012	4	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Clio yatsuoensis</i> Shibata, 1983	1	-	-	-	-	-	-	-	?	?	-	-	-	-	-	-	-	-	-	-
<i>Clio</i> sp.	92	-	-	-	-	-	+	-	+	+	+	+	+	-	-	+	-	-	-	11
Family Cavoliniidae																				
Subfamily Cavoliniinae																				
<i>Cavolinia baniensis</i> Janssen, 2007	6	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Cavolinia bituminata</i> Beets, 1953	2	-	-	-	-	-	-	-	-	-	-	?	?	-	-	-	-	-	-	-
<i>Cavolinia cookei</i> Simonelli, 1895	71	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Cavolinia floridana</i> Collins, 1934	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Cavolinia gatti</i> Janssen, 2012	7	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Cavolinia gibbosa</i> (d'Orbigny, 1834) s.lat.	8	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	83

Table 1, continued.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Bartonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots'
<i>Cavolinia gibbosa gibboides</i> Rampal, 2002	13	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-
<i>Cavolinia globulosa</i> (Gray, 1850)	5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	157
<i>Cavolinia grandis</i> (Bellardi, 1873)	16	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-
<i>Cavolinia gypсорum</i> (Bellardi, 1873)	22	-	-	-	-	-	-	-	-	+	?	+	+	-	-	-	-	-	-	-
<i>Cavolinia inflexa imitans</i> (Pfeffer, 1880)	19	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-
<i>Cavolinia inflexa inflexa</i> (Lesueur, 1813)	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-
<i>Cavolinia inflexa kakegawaensis</i> Shibata 1984	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	?	-	-	-
<i>Cavolinia inflexa labiata</i> (d'Orbigny, 1834)	3	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-
<i>Cavolinia inflexa</i> (Lesueur, 1813) s.lat.	22	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	275
<i>Cavolinia landaui</i> Janssen, 2004	20	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Cavolinia marginata hyugaensis</i> Ujihara, 1996	17	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Cavolinia marginata limatula</i> Beets, 1943	1	-	-	-	-	-	-	-	-	-	-	?	?	-	-	-	-	-	-	-
<i>Cavolinia marginata</i> (Bronn, 1862) s.lat.	13	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-
<i>Cavolinia marginata pliomediterranea</i> Janssen, 2004	6	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Cavolinia marginata vendryesiana</i> (Guppy, 1873)	5	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Cavolinia mexicana</i> (Collins, 1934)	5	-	-	-	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-
<i>Cavolinia microbesitas</i> Janssen, 2012	7	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Cavolinia perparvula</i> Janssen, 2007	11	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Cavolinia pycna</i> Jung, 1971	38	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Cavolinia shibatai</i> Janssen, 2007	24	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Cavolinia tridentata</i> (Forsskål in Niebuhr, 1775) s.lat.	49	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	+	+	160
<i>Cavolinia uncinata</i> (d'Orbigny, 1835) s.lat.	21	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	146
<i>Cavolinia ventricosa</i> (Guppy, 1882)	10	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Cavolinia yamabensis</i> Shibata, 1983	1	-	-	-	-	-	-	-	?	?	-	-	-	-	-	-	-	-	-	-
<i>Cavolinia zamboninii</i> Checchia-Rispoli, 1921	17	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Cavolinia</i> sp.	118	-	-	-	-	-	-	-	-	+	+	-	-	+	+	+	+	-	+	49
<i>Diacavolinia angulosa</i> (Gray, 1850)	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	10
<i>Diacavolinia aspina</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Diacavolinia atlantica</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Diacavolinia bandaensis</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Diacavolinia bicornis</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Diacavolinia constricta</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Diacavolinia deblainvillei</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	2
<i>Diacavolinia deshayesi</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Diacavolinia elegans</i> van der Spoel et al., 1993	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Diacavolinia flexipes</i> van der Spoel et al., 1993	15	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	14
<i>Diacavolinia grayi</i> van der Spoel et al., 1993	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Diacavolinia limbata</i> (d'Orbigny, 1836)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Diacavolinia longirostris</i> (de Blainville, 1821) s. lat.	18	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	348
<i>Diacavolinia ovalis</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Diacavolinia pacifica</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Diacavolinia pristina</i> Janssen, 2007	6	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Diacavolinia robusta</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Diacavolinia souleyeti</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1

Table 1, continued.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Bartonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots ¹	
<i>Diacavolinia strangulata</i> (Deshayes, 1823)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1	
<i>Diacavolinia striata</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Diacavolinia triangulata</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Diacavolinia v. vanutrechtii</i> van der Spoel et al., 1993	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Diacavolinia</i> sp.	6	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	1
<i>Gamopleura maxwelli</i> Grebneff & Janssen, 2011	1	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Gamopleura melitensis</i> Janssen, 1995	60	-	-	-	-	-	-	+	?	?	-	-	-	-	-	-	-	-	-	-	-
<i>Gamopleura pilula</i> Janssen, 2012	3	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Gamopleura taurinensis</i> (Michelotti, 1847)	4	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
Subfamily Diacriinae																					
<i>Diacria digitata</i> (Guppy, 1882)	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-
<i>Diacria italica</i> Grecchi, 1982 s.lat.	21	-	-	-	-	-	-	-	-	-	-	-	-	-	-	?	+	-	-	-	-
<i>Diacria major</i> (Boas, 1886)	2	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	16
<i>Diacria mbaensis</i> Ladd, 1934	7	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Diacria microstriata</i> Janssen, 2007	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Diacria paeninsula</i> Janssen, 2007	18	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Diacria rampalae</i> Dupont, 1979 emend.	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	4
<i>Diacria rubecula</i> Bontes & van der Spoel, 1998	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Diacria sangiorgii</i> Scarsella, 1934	47	-	-	-	-	-	-	-	-	-	?	-	+	-	-	-	-	-	-	-	-
<i>Diacria trispinosa</i> (de Blainville, 1821)	166	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	+	+	+	+	552
<i>Diacria trispinosa africana</i> van der Spoel, 1974	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Diacria</i> sp.	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	+	16
<i>Diacrolinia aquensis</i> (Grateloup, 1827)	7	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Diacrolinia aurita</i> (Bellardi, 1873)	212	-	-	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-
<i>Diacrolinia elioi</i> Janssen, 1995	54	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-
<i>Diacrolinia interrupta</i> (Bellardi, 1873)	32	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Diacrolinia larandaensis</i> Janssen, 1999	3	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Diacrolinia orbignyi</i> (Rang, 1827)	6	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Diacrolinia pumilionis</i> Janssen, 2012	2	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-
<i>Diacrolinia</i> sp.	118	-	-	-	-	-	-	+	+	+	+	-	-	-	-	-	-	-	-	-	-
<i>Telodiacria costata</i> (Pfeffer, 1879)	6	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	8
<i>Telodiacria danae</i> (van Leyen & van der Spoel, 1982)	11	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	99
<i>Telodiacria erythra</i> (van Leyen & van der Spoel, 1982)	31	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Telodiacria philippinensis</i> (Janssen, 2007)	34	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-
<i>Telodiacria quadridentata</i> (de Blainville, 1821) s. lat.	16	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	42
<i>Telodiacria schmidti</i> (van Leyen & van der Spoel, 1982)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	15
Subfamily Vaginellinae Janssen subfam. nov.⁵																					
<i>Edithinella bonaviai</i> Janssen, 2004	14	-	-	-	-	-	-	+	?	-	-	-	-	-	-	-	-	-	-	-	-
<i>Edithinella caribbeana</i> (Collins, 1934)	17	-	-	-	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-
<i>Edithinella curva</i> Janssen, 1998	9	-	-	-	-	-	-	-	-	-	?	+	-	-	-	-	-	-	-	-	-
<i>Edithinella doliarius</i> Janssen, 2006	17	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Edithinella katoi</i> (Shibata, 1983)	2	-	-	-	-	-	-	-	-	?	-	-	-	-	-	-	-	-	-	-	-
<i>Edithinella varanica</i> (Sirna, 1968)	32	-	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-

Table 1, continued.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Barntonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots'
<i>Edithinella</i> sp.	1	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Vaginella acutissima</i> Audenino, 1899	130	-	-	-	-	-	-	-	-	?	+	-	-	-	-	-	-	-	-	-
<i>Vaginella austriaca</i> Kittl, 1886	417	-	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-
<i>Vaginella basitruncata</i> Janssen, 2005	2	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Vaginella bicarinata</i> Tate, 1887	2	-	-	-	-	-	-	+	?	-	-	-	-	-	-	-	-	-	-	-
<i>Vaginella chattica</i> R. Janssen, 1979	69	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Vaginella chipolana</i> Dall, 1893	1	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
<i>Vaginella depressa</i> Daudin, 1800	145	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-
<i>Vaginella gaasensis</i> Janssen, 2010	1	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Vaginella gibbosa</i> Audenino, 1899	97	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Vaginella lapugyensis</i> Kittl, 1886	168	-	-	-	-	-	-	-	-	?	+	+	-	-	-	-	-	-	-	-
<i>Vaginella sannicola</i> Janssen, 1990	46	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Vaginella tricuspudata</i> Zorn & Janssen, 1993	68	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Vaginella venezuelana</i> Collins, 1934	2	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Vaginella victoriae</i> Janssen, 1990	7	-	-	-	-	-	-	-	-	-	+	-	+	-	-	-	-	-	-	-
<i>Vaginella</i> sp.	105	-	-	-	-	-	-	-	+	+	+	+	+	-	-	-	-	-	-	-
Superfamily Cavolinioidea s.lat.																				
Cavoliniidae/Cavolinioidea sp.	45	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	+	+	162
Family Sphaerocinidae																				
<i>Hameconia edmundi</i> Janssen, 2008	4	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Sphaerocina convolvula</i> Janssen, 2007	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	-	-	-	-
<i>Sphaerocina formai</i> (Audenino, 1899)	26	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-
<i>Sphaerocina</i> sp.	5	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-
Family Cymbuliidae																				
<i>Corolla calceola</i> (Verrill, 1880)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Corolla cupula</i> Rampal, 1996	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Corolla intermedia</i> (Tesch, 1903)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	6
<i>Corolla ovata</i> (Quoy & Gaimard, 1833)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	4
<i>Corolla spectabilis</i> Dall, 1871	1	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Corolla</i> sp.	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Cymbulia parvidentata</i> Pelseneer, 1888	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Cymbulia</i> sp.	138	-	-	?	-	-	-	-	-	+	+	-	-	-	-	+	-	-	+	57
<i>Cymbulia peronii</i> de Blainville, 1818	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1183
<i>Cymbulia sibogae</i> Tesch, 1903	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	4
<i>Gleba cordata</i> Forsskål in Niebuhr, 1776	64	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	5
<i>Gleba</i> sp.	5	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-
Family Desmopteridae																				
<i>Desmopterus papilio</i> Chun, 1889	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	3
Family Peraclidae																				
<i>Peracle amberae</i> Janssen, 2012	4	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	-	-	-
<i>Peracle bispinosa</i> (Pelseneer, 1888)	15	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	-	+	+	20
<i>Peracle charlotteae</i> Janssen & Little, 2010	3	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-

Table 1, continued.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Barntonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots'
<i>Peracle diversa</i> (Monterosato, 1875)	21	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	23
<i>Peracle elata</i> (Seguenza, 1875)	18	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	-	-	+	17
<i>Peracle grebneffi</i> Janssen, 2012	17	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Peracle lata</i> (Krach, 1979)	1	-	-	-	-	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-
<i>Peracle moluccensis</i> (Tesch, 1903)	4	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	14
<i>Peracle reticulata</i> (d'Orbigny, 1834)	87	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	53
<i>Peracle rissoides</i> Tesch, 1903	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Peracle valdiviae</i> (Meisenheimer, 1905)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Peracle</i> sp.	58	-	-	-	-	-	+	-	+	+	-	-	-	-	+	-	-	-	+	-
Family Pneumodermatidae																				
<i>Pneumoderma degraaffi</i> van der Spoel & Pafort-van Iersel, 1982	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Pneumoderma heronense</i> Newman & van der Spoel, 1989	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Pneumoderma mediterraneum</i> (van Beneden, 1838)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Pneumoderma peroni</i> (Cuvier, 1817)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Pneumoderma violaceum</i> d'Orbigny, 1835	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	25
<i>Pneumodermopsis ciliata</i> (Gegenbaur, 1855)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	9
<i>Pneumodermopsis michaelsarsi</i> (Bonnevie, 1913)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	23
<i>Pneumodermopsis teschi</i> van der Spoel, 1973	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Spongiobranchaea australis</i> d'Orbigny, 1836	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	237
Family Notobranchaeidae																				
<i>Notobranchaea bleekerae</i> van der Spoel & Pafort-van Iersel, 1985	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Notobranchaea inopinata</i> Pelseeneer 1887	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Notobranchaea macdonaldi</i> Pelseeneer, 1886	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	7
<i>Notobranchaea tetrabranchiata</i> Bonnevie, 1913	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	3
Family Cliopsidae																				
<i>Cliopsis krohni</i> Troschel, 1854	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	3
<i>Pruvotella danae</i> Pruvot-Fol, 1942	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
Family Clionidae																				
<i>Clione limacina antarctica</i> Smith, 1902	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	6
<i>Clione limacina limacina</i> (Phipps, 1774)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	565
<i>Clione? ignota</i> Janssen, 2012	24	-	-	-	-	-	?	+	+	+	+	-	-	-	-	-	-	-	-	-
<i>Clione? imdinaensis</i> Janssen, 2012	56	-	-	-	-	-	?	+	+	+	?	-	-	-	-	-	-	-	-	-
<i>Clione? phosphorita</i> Janssen, 2012	11	-	-	-	-	-	?	+	+	+	-	-	-	-	-	-	-	-	-	-
<i>Clione? tripartita</i> Janssen, 2012	6	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-
<i>Clione? tumidula</i> Janssen, 2012	13	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	-	-	-	-
<i>Clione?</i> sp.	143	-	-	-	-	-	-	-	+	-	-	-	-	-	+	-	+	+	-	-
<i>Fowlerina punctata</i> (Tesch, 1903)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Paedoclione doliiformis</i> Danforth, 1907	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Paraclione flavescens</i> (Gegenbaur, 1855)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
<i>Paraclione pelseeneeri</i> Tesch, 1903	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	4

Table 1, continued.

	RGM lots	Pre-Eocene	Ypresian	Lutetian	Bartonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	Pleistocene	Holocene	RMNH lots ¹
<i>Thliptodon antarcticus</i> Meisenheimer, 1906	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Thliptodon diaphanus</i> (Meisenheimer, 1902)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	4
<i>Thliptodon gegenbauri</i> Boas, 1886	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
<i>Thliptodon schmidti</i> Pruvot-Fol, 1942	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
Family Hydromylidae																				
<i>Hydromyles globulosus</i> (Rang, 1825)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	122
Family Glaucidae																				
<i>Glaucus atlanticus</i> Forster, 1777	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	1
Family Phylliroidae																				
<i>Phylliroe bucephala</i> Lamarck, 1816	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	4
Family Fionidae																				
<i>Fiona pinnata</i> (Eschscholtz, 1831)	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	2
Incerti ordinis																				
Aff. <i>Cheilospicata</i> sensu Garvie, in Garvie et al., 2020	2	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Aff. <i>Creseis</i> sensu Garvie, in Garvie et al., 2020	6	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Aff. <i>Currylimacina</i> sensu Garvie, in Garvie et al., 2020	2	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Aff. <i>Praehyalocylis</i> sensu Garvie, in Garvie et al., 2020	2	+	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
Beaked larva sensu van der Spoel & Newman, 1990	12	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	+	+	-
Veliger of benthic gastropod?	96	-	-	-	-	-	-	-	-	+	+	-	-	-	-	-	-	+	+	-

¹ No data signifies that either no samples are present in the RMNH collection or no sample data are present in the RMNH registry at the time of writing.

² Samples listed as *Atlanta turriculata* in both RGM and RMNH collections require reidentification and may contain *A. vanderspoeli*.

³ Reidentification of samples indicated in Table 11 as *Clio pyramidata lanceolata* is required; they are here grouped within *Clio pyramidata pyramidata*.

⁴ For diagnosis see Appendix 4.

Table 1, continued.

The RMNH/ZMA collection consists of two separate parts, a dry collection and the alcohol collection. Both the dry RMNH/ZMA collection as the RGM collection are stored in steel drawer cabinets (Fig. 1).

To make the material in both collections more easily available and accessible for further research a list is given here of all available identified species, with the number of lots present and their approximate age (Table 1). However, especially the RMNH/ZMA alcohol collection is incompletely registered. Although all lots received a RMNH or ZMA collection number, the data as present on the accompanying labels are only for a small part entered into the central Naturalis collection database. This undesirable situation requires a long period of careful work to be completed. At

the moment the fossil samples are registered in a separate database. Ultimately this will be incorporated into the central Naturalis database. Part of the RMNH/ZMA holoplanktic species is accessible through the ‘Naturalis bioportal’: <https://bioportal.naturalis.nl>.

There is a considerable overlap in the two collections. The RMNH/ZMA dry material includes numerous specimens collected from ocean bottom sediment samples, but many similar specimens are also in the RGM collection, dated as Holocene. Depending on the method used such samples may locally include material of Pleistocene age, as was several times the case in box cores from the Mediterranean.

THE NATURALIS (RGM) COLLECTION OF FOSSIL HOLOPLANKTIC MOLLUSCA (AWJ, RP & FPW)

History

In the early 1980's Unesco launched the International Geological Correlation Programme (IGCP). In one of the subprojects, IGCP 124: 'The Northwest European Tertiary Basin', AWJ participated for the molluscan discipline, together with a number of international colleagues, on the subject of mollusc-based stratigraphic correlation possibilities in western European countries. After some time researchers concluded that the application of (mainly benthic) mollusc assemblages was quite useful for local correlations, but offered severe difficulties over longer distances, as such assemblages depend strongly on facies. Finally it was the late Dr Chris King (U.K.), who first applied the barely known holoplanktic molluscs for correlations (King, 1981) which appeared to offer better possibilities, especially over longer distances, more or less comparable to other pelagic organisms, such as planktic foraminifera and calcareous nannoplankton. Ultimately this led to a first pteropod-based biostratigraphical zonation for the Cenozoic of the North Sea Basin, published in the final IGCP 124 report (Janssen & King, 1988). This early work triggered a lasting interest in planktic mollusc biostratigraphy and systematics and it developed into AWJ's main research topic for many years, the more so after his retirement in 1997.

Isolating fossil pelagic gastropods from existing collections in the Leiden museum led to a first concentration of this fossil group in a separate collection that during the years was extended by targeted field work and support of colleague researchers and institutes. The research included extensive studies of type specimens and historical collections housed in institutions in London, Paris, Bordeaux, Copenhagen, Berlin, Warsaw, Basel, Vienna, Turin, Milan,

Catania and Cagliari. The present worldwide collection of fossil holoplanktic molluscs comprises about 1200 lots, including a large number of primary and secondary type specimens and numerous illustrated specimens.

Preservation and storage

Specimens from unconsolidated rocks (sand or clay) have been isolated from the sediment by simple washing procedures. For sandy sediments usually a single treatment in water over a 0.2 or 0.3 mm sieve is sufficient to obtain a clean residue. For more clayey samples the procedure has to be repeated, soaking a residue in hot water after careful drying. Preservation in many types of clayey sediments is often in the form of internal pyritic moulds or specimens are filled with pyrite. For such samples application of hydrogenic peroxide (H_2O_2) to speed up disaggregation must be strongly discouraged, as it will damage or even completely destroy pyritic specimens. The tiny and fragile pelagic mollusc shells are easily damaged, so care has to be taken, also because frequently such specimens form only a minor part of residues that usually are dominated by benthic mollusc fragments that by their larger size and weight will rapidly damage heteropod and pteropod shells.

Incidentally, especially in fine-sandy sediments, smaller fossils including pteropods can be collected dry from the sediment contents of larger gastropods by careful tapping the shells. Samples obtained this way can be sieved without washing procedures. Specimens can be picked from dried and sieved residues using a 10 or 20 times magnification under a binocular microscope. Pelagic molluscs in shell preservation usually do not need further preservation and can be stored, after careful drying, in glass vials or slides. Pyritic specimens need special treatment for long term storage as pyrite will disintegrate under the influence of normal air humidity (Appendix 1). Pteropods (and similar fossils) in limestones or limestone concretions, mostly can-



Figs 2-3. Naturalis (RGM) fossil pelagic molluscs, stored in slides or in silicone oil glass vials (standing upright), in systematical order.



Fig. 4. Naturalis (RGM) fossil pelagic molluscs with smaller specimens in turn-top slides or glass vials and larger items in collection boxes, in systematical order; a red sticker indicates holotype, a green one paratype(s), specimens illustrated in publications are marked with a yellow sticker.

not be separated from the rock that has to be reduced in size for storage. If more specimens or species are represented on a single rock sample careful registration by cross-referencing has to be performed to be able to trace specimens.

In some Oligocene and Miocene limestones in the Mediterranean Basin (e.g. southern Italy, Malta, Sicily) specimens are preserved as internal phosphoritic moulds. These

can be concentrated from the rock using formic or acetic acid, as described extensively in Janssen (2012). After careful washing in water to remove acid remnants such specimens do not need further preservation anticipating long term storage. Specimens in limonitic internal mould preservation (such as in some Maltese or Sicilian Miocene clays) sometimes need treatment with an artificial resin to obtain sufficient solidity. Because of their very thin shell walls, pteropod specimens in internal mould preservation remain identifiable in most cases, although external ornamentation will be lost.

In the Naturalis collection fossil pelagic molluscs (Tables 2-3) are stored, either in turntop slides, glass vials (mainly 45 × 15/16 or 45 × 25 mm), or as rock samples, as much as possible in systematic order (Figs 2-4). Documentation is done by a registration number in the vial or on the slide, and adding a label with locality and further basic data (age, formation, collector, donator, number, date of collecting) and additionally one (or more) identification labels, stored per lot in standard PVC collection boxes (55 × 45 mm, or a multiple thereof). All data are assembled in a Microsoft Access database with RGM registration numbers, and cross-referencing in cases where a single lot contains more than one species. Primary type specimens are indicated with a red sticker (also on the drawer), secondary types have a green sticker and specimens illustrated in publications have a yellow one.

Smaller specimens that were used for illustration by scanning electron microscopy (SEM) cannot easily be

	No of lots	Atlantic	Mediterranean	Caribbean	Red Sea	Indian Ocean	Indonesia	Pacific
Atlantidae	527	20	257	20	150	24	6	50
Pterotracheidae	89	-	73	-	11	1	-	4
Carinariidae	91	2	69	1	4	9	1	5
Epitoniidae	19	1	8	-	7	1	-	2
Heliconoididae	45	7	5	3	19	1	3	7
Limacinidae	345	23	240	22	31	3	6	20
Thieleidae	2	2	-	-	-	-	-	-
Creseidae ¹	282	17	123	24	71	1	12	34
Hyalocylidae	66	2	36	3	16	4	1	4
Cuvierinidae	37	6	10	-	-	5	1	15
Cliidae	254	25	172	8	19	5	6	19
Cavoliniidae	369	32	175	32	72	16	11	31
Peraclidae	164	14	112	11	15	3	1	8
Cymbuliidae	149	-	123	-	17	1	-	8
Clionidae	128	2	107	-	14	2	-	3
Incerti ordinis ²	62	-	49	-	11	1	-	1

¹ Includes also the genus *Styliola*.

² Specimens provisionally included, but might be larvae of benthic species

Table 2. Quaternary (Pleistocene-Holocene) lots in the Naturalis (RGM) collection of fossil holoplanktic Mollusca and their origin per region.

	EOCENE					OLIGO-CENE		MIOCENE						PLIOCENE			Totals
	pre-Eocene ¹	Ypresian	Lutetian	Bartonian	Priabonian	Rupelian	Chattian	Aquitainian	Burdigalian	Langhian	Serravallian	Tortonian	Messinian	Zanclean	Piacenzian	Gelasian	
Belgium	-	30	3	-	-	118	-	-	-	26	-	-	6	3	4	-	190
Czech Republic	-	-	-	-	-	-	-	-	-	2	-	-	-	-	-	-	2
Denmark	4	6	29	-	-	4	38	11	50	9	-	-	-	-	-	151	
England	-	75	9	20	-	-	-	-	2	-	-	-	-	-	-	95	
Germany	-	9	-	-	-	29	126	39	275	582	146	19	1	-	-	1.226	
France	-	109	22	3	-	14	114	21	167	5	8	-	-	123	-	586	
Netherlands	-	263				90	7	-	-	69	12	14	-	-	19	-	469
Poland	-	-	-	-	-	1	-	-	-	43	-	-	-	-	-	44	
Portugal	-	-	-	-	-	-	-	-	-	-	2	-	-	-	-	2	
Portugal (Azores)	-	-	-	-	-	-	-	-	-	-	-	-	-	48	-	48	
Spain	-	-	-	-	-	-	-	-	-	-	1	9	4	-	68	82	
Ukraine	-	-	-	-	-	-	-	-	-	5	-	-	-	-	-	5	
Cyprus	-	-	-	-	-	-	-	-	-	312	-	273	-	-	-	585	
Egypt	-	4	6	1	-	-	-	-	1	6	-	-	-	-	-	17	
Italy	-	-	-	-	-	2	1	40	40	1.452	80	80	7	187	44	1.951	
Malta	-	-	-	-	-	-	115	-	471	1.092	126	103	15	-	-	1.922	
Turkey	-	-	-	-	-	-	-	-	-	27	21	-	-	-	-	48	
Iran	-	28		-	-	-	-	-	-	-	-	-	-	-	-	28	
Kazakhstan	-	14	-	-	3	-	-	-	-	-	-	-	-	-	-	17	
Uzbekistan	-	119		-	-	-	-	-	-	-	-	-	-	-	-	119	
Nigeria	-	-	-	3	-	-	-	-	-	-	-	-	-	-	-	2	
Tanzania	-	-	-	-	56	45	-	-	-	-	-	-	-	-	-	101	
Australia	-	3				21	-	-	-	78	-	-	-	-	-	-	102
Fiji	-	-	-	-	-	-	-	-	-	-	-	-	-	-	61	61	
Indonesia	-	-	-	-	-	-	-	-	-	14	-	-	-	-	5	19	
Japan	-	-	-	-	-	-	-	-	-	18	-	-	-	-	3	21	
New Zealand	-	-	-	-	-	-	1	-	-	11	-	-	-	-	1	13	
Philippines	-	-	-	-	-	-	-	-	-	-	-	-	-	-	707	707	
Canada	-	-	5	-	-	-	-	-	-	-	-	-	-	-	-	4	
Dominican Republic	-	-	-	-	-	-	-	-	-	-	1	-	-	-	-	1	
Jamaica	-	-	-	-	-	-	-	-	-	-	-	-	-	-	51	51	
Trinidad & Tobago	-	-	-	-	-	-	-	-	74	-	-	23	-	-	-	97	
U.S.A.	13?	164	159	34	-	41	-	7	12	1	-	-	-	-	-	395	
Chili	-	-	-	-	-	-	-	-	-	-	-	1	-	-	-	1	
Venezuela	-	1				-	-	-	-	-	-	-	-	-	-	-	1

¹ Includes Maastrichtian and Paleocene lots.

Table 3. Available number of Pliocene and older lots of holoplanktic Mollusca, specified per country and approximate age. In cases where correlation of local stages with the European subdivision is not ascertained or unknown numbers of lots are given per larger interval.

removed from the SEM-stub without damage and consequently a number of SEM-stubs form part of the collection. In a large number of Mediterranean core and bottom samples numerous smaller specimens are stored together in so-called 'composite samples', in order to save space in the

collection. Such specimens are registered under the same registration number, with added a, b, c etc. These composite samples frequently also contain larval bivalve shells (prodissoconchs) or larval specimens (protoconchs) of benthic gastropods from the same sample.

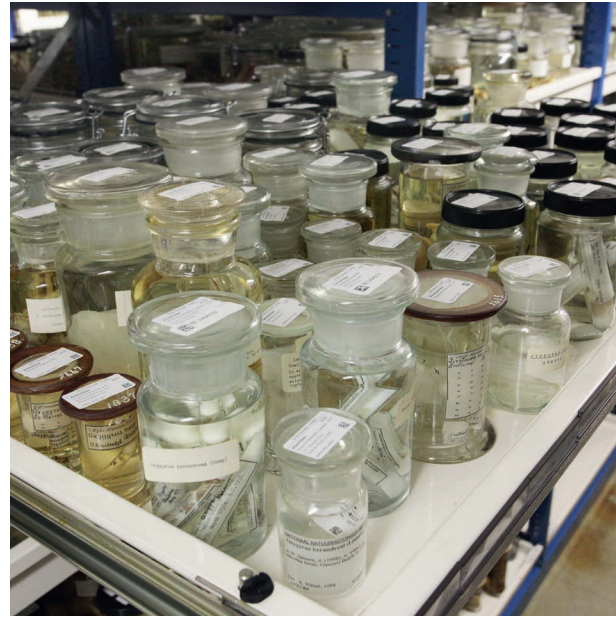
	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Atlantidae									
<i>Atlanta affinis</i> ¹⁰	6	-	-	-	-	-	-	6	-
<i>Atlanta ariejansseni</i> ²	7	7 ¹	-	-	-	-	-	-	-
<i>Atlanta brunnea</i>	102	9	85	3	-	2	1	-	2
<i>Atlanta echinogyra</i>	4	-	1	-	-	-	-	3	-
<i>Atlanta fusca</i> ³	5	-	4	-	-	1	-	-	-
<i>Atlanta gaudichaudi</i>	20	1	1	-	-	6	-	12	-
<i>Atlanta gibbosa</i>	5	1	-	-	-	-	-	4	-
<i>Atlanta helicinoidea</i>	76	1	59	10	-	2	-	3	1
<i>Atlanta inclinata</i>	50	3	33	-	-	9	-	3	2
<i>Atlanta inflata</i>	10	2	1	-	-	2 ⁴	2	3	-
<i>Atlanta keraudrenii</i>	1 ⁵	-	1	-	-	-	-	-	-
<i>Atlanta lesueurii</i>	24	3	3	8	-	1	2	5	2
<i>Atlanta oligogyra</i>	17	1	-	-	-	-	-	14	2
<i>Atlanta peronii</i>	216	24	130	20	-	18	2	20	2
<i>Atlanta plana</i>	3	-	-	-	-	-	-	1	2
<i>Atlanta selvagensis</i>	52	1	49	-	-	-	2 ⁶	-	-
<i>Atlanta tokiokai</i>	26	-	13	-	-	5	-	8	-
<i>Atlanta turriculata</i>	24	1	1	-	-	-	-	20	1
<i>Atlanta vanderspoeli</i>	1	-	-	-	-	-	-	-	1
<i>Atlanta</i> sp.	50	45	3	-	-	2	-	-	-
<i>Oxygyrus inflatus</i>	58	-	56	-	-	-	-	2	-
<i>Oxygyrus keraudrenii</i> ⁷	51	18	11	15	-	5	1	-	1
<i>Oxygyrus rangii</i> ⁸	5	1	-	1	-	-	3	-	-
<i>Protatlanta souleyeti</i> ⁹	97	4	81	7	-	5	-	-	-
Totals	909	122	532	64	-	58	13	104	16

¹ Not yet registered lots, data only available from the label.² Data not in register, but available from original publication (Wall-Palmer et al., 2016; all specimens from the Southern Subtropical Convergence Zone of the Atlantic and Indo-Pacific oceans).³ Junior synonym of *Atlanta brunnea*.⁴ Needs reidentification, *A. inflata* is not known to occur in the Atlantic or Caribbean (Janssen & Seapy, 2009).⁵ Neotype (Janssen, 2012b), synonym of *Atlanta peronii*.⁶ Needs reidentification, *A. selvagensis* is not known to occur in the Indian Ocean.⁷ Incorrect identification of *Oxygyrus inflatus*.⁸ Synonym of *Oxygyrus inflatus*.⁹ Needs reidentification: may include *Protatlanta sculpta* Issel, 1911 (Wall-Palmer et al., 2016).¹⁰ Junior synonym of *Atlanta inclinata*.**Table 4.** RMNH collection of Recent Atlantidae; number of lots specified per region. Taxon names in bold indicate that the primary type material is in the RMNH collection.

THE RECENT COLLECTION (BVDB & JG)

The study of pelagic Mollusca has always been a niche within the field of malacology. The number of Dutch scientists, who have been or still are active in this niche, is not very large, but as a result of their various publications their influence has been or still is quite substantial. The first Dutch researcher, who actively studied pelagic molluscs

was Johan Jacob Tesch (1877-1954). Tesch was a biologist at the Dutch Institute for Fishery Investigations, where he worked on population structures of commercially important North Sea fish. He graduated 11-5-1906 cum laude at Utrecht University (promotor A.A.W. Hubrecht) with a thesis on Pterotracheid pelagic molluscs obtained during the famous Dutch Siboga Expedition in the Indo-Pacific. From August 1907 to March 1908 and from 1915 to 1918 he



Figs 5-6. The Naturalis (RMNH) present-day mollusc collection; left the dry collection (glass vials or boxes in systematical order, right the alcohol collection.

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Carinariidae									
<i>Cardiapoda placenta</i>	15	-	1	-	-	-	-	11	3
<i>Cardiapoda sublaevis</i>²	3	-	-	-	-	-	-	3	-
<i>Cardiapoda trachydermon</i>³	2	-	-	-	-	-	-	2	-
<i>Carinaria cithara</i>	4	-	-	-	-	-	4	-	-
<i>Carinaria cristata</i>	12	6	-	-	-	-	-	6	-
<i>Carinaria cymbium</i> ⁴	1	-	-	1	-	-	-	-	-
<i>Carinaria galea</i>	3	-	-	-	-	-	1	-	2
<i>Carinaria lamarcki</i>	25	4	7	9	-	1	-	4	-
<i>Carinaria macrorhynchus</i>⁵	1	-	-	-	-	-	-	1	-
<i>Carinaria mediterranea</i> ⁴	19	8	-	11	-	-	-	-	-
<i>Carinaria</i> sp.	2	1	1	-	-	-	-	-	-
<i>Pterosoma planum</i>	1	1	-	-	-	-	-	-	-
Totals	88	20	9	21	-	1	5	27	5

¹ Not yet registered lots, data only available from the label.

² Synonym of *Cardiapoda placenta*, according to van der Spoel (1976: 157).

³ Synonym of *Carinaria lamarcki*, according to van der Spoel (1976: 152).

⁴ Synonym of *Carinaria cithara*, according to van der Spoel (1976: 155).

⁵ Synonym of *Carinaria lamarcki*, according to the WoRMS website (accessed February 2020).

Table 5. RMNH collection of Recent Carinariidae; number of lots specified per region. Taxon names in bold indicate that the primary type material is in the RMNH collection.

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Pterotracheidae									
<i>Firoloida desmarestia</i>	18	8	4	5	-	-	1	-	-
<i>Pterotrachea challengerii</i> ³	6	-	-	-	-	-	-	6	-
<i>Pterotrachea coronata</i>	215	210	-	1	-	-	-	4	-
<i>Pterotrachea mutabilis</i> ²	6	-	-	-	-	-	-	6	-
<i>Pterotrachea hippocampus</i>	10	1	-	3	-	-	-	6	-
<i>Pterotrachea intermedia</i> ³	3	-	-	-	-	-	-	3	-
<i>Pterotrachea microptera</i> ⁴	7	-	-	-	-	-	-	7	-
<i>Pterotrachea minuta</i>	2	-	-	-	1	-	-	1	-
<i>Pterotrachea mutica</i> ²	1	-	-	1	-	-	-	-	-
<i>Pterotrachea orthophthalmus</i> ²	1	-	-	-	-	-	-	1	-
<i>Pterotrachea scutata</i>	3	-	2	-	-	-	-	-	1
<i>Pterotrachea xenoptera</i> ¹	1	-	-	-	-	-	-	1	-
Totals	273	219	6	10	1	-	1	35	1

¹ Not yet registered lots, data only available from the label.

² Synonym of *Pterotrachea hippocampus*, according to van der Spoel (1976: 161-162).

³ Synonym of *Pterotrachea coronata*, according to van der Spoel (1976: 160).

⁴ Possibly a synonym of *Pterotrachea minuta*, according to van der Spoel (1976: 163).

Table 6. RMNH collection of Recent Pterotracheidae; number of lots specified per region. Taxon names in bold indicate that the primary type material is in the RMNH collection.

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Epitoniidae									
<i>Janthina bicolor</i> ²	1	-	-	1	-	-	-	-	-
<i>Janthina capreolata</i> ³	3	-	-	-	-	-	-	-	3
<i>Janthina exigua</i>	56	20	29	2	-	1	3	-	1
<i>Janthina fragilis</i> ²	3	1	-	-	2	-	-	-	-
<i>Janthina globosa</i>	111	45	4	3	-	37	6	8	8
<i>Janthina janthina</i>	274	153	23	7	2	47	15	2	22
<i>Janthina pallida</i>	36	21	8	2	-	4	-	-	1
<i>Janthina prolongata</i> ⁴	5	-	-	-	-	2	-	-	3
<i>Janthina rotundata</i> ⁵	1	-	1	-	-	-	-	-	-
<i>Janthina umbilicata</i>	22	7	1	2	-	9	2	-	1
<i>Recluzia jehennei</i> ⁶	3	1	-	-	-	-	1	1	-
<i>Janthina</i> sp.	10	5	2	-	3	-	3	-	-
<i>Recluzia rollandiana</i> ⁶	11	4	-	-	-	4	1	-	2
Epitoniidae indet.	4	1	1	-	-	-	1	-	1
Totals	540	258	69	17	7	104	32	11	42

¹ Not yet registered lots, data only available from the label.

² Synonym of *Janthina janthina*, according to Beu (2017: 186).

³ Synonym of *Janthina exigua*, according to Beu (2017: 191).

⁴ Synonym of *Janthina globosa*, according to Beu (2017: 181).

⁵ Synonym of *Janthina janthina*, according to WoRMS.

⁶ Synonym of *Recluzia lutea* (Bennett, 1840), according to Beu (2017: 203)

Table 7. RMNH collection of Recent Epitoniidae; number of lots specified per region.

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Heliconoididae									
<i>Heliconoides inflatus</i>	349	58	214	39	-	30	-	8	-
Family Limacinidae									
<i>Limacina balea</i> ²	7	7	-	-	-	-	-	-	-
<i>Limacina bulimoides</i>	177	24	127	5	-	14	-	7	-
<i>Limacina helicina antarctica</i>	39	28	11	-	-	-	-	-	-
<i>Limacina helicina helicina</i>	6	4	2	-	-	-	-	-	-
<i>Limacina helicina pacifica</i>	1	-	-	-	-	-	-	-	1
<i>Limacina lesueurii</i>	130	4	121	-	-	3	1	1	-
<i>Limacina retroversa</i>	43	15	19	8 ³	-	-	1 ⁴	-	-
<i>Limacina retroversa australis</i>	3	-	3	-	-	-	-	-	-
<i>Limacina trochiformis</i>	9	3	3	1	-	1	1	-	-
<i>Limacina</i> sp.	57	17	14	25	-	1	-	-	-
Family Thieleidae									
<i>Thielea helicoides</i>	10	5	5	-	-	-	-	-	-
Totals	831	165	519	78	-	49	3	16	1

¹ Not yet registered lots, data only available from the label.² Usually considered a forma of *L. retroversa*.³ Most probably Quaternary fossils (from sediment samples).⁴ Needs reidentification; *L. retroversa* is not known to occur in the Indian Ocean.**Table 8.** RMNH collection of Recent Limacinacea; number of lots specified per region. Taxon names in bold indicate that the primary type material is in the RMNH collection.

was employed at the RMNH. Apart from his fishery studies he published between 1904 and 1950 twelve papers on pelagic molluscs from several expeditions, among which the Dana expeditions, the material of which is housed in Copenhagen. Two of his publications (1908, 1909) are dedicated to material in the Rijksmuseum van Natuurlijke Historie (RMNH), Leiden. The Siboga material originally was in the Zoological Museum, Amsterdam, but currently in the Naturalis museum.

The second Dutch biologist, who actively studied pelagic Mollusca is Siebrecht van der Spoel. Apart from his PhD-thesis (van der Spoel, 1967) he wrote another substantial work on pelagic Mollusca (van der Spoel, 1976). Van der Spoel was professor at the University of Amsterdam, organised various expeditions on the Atlantic Ocean and published numerous papers on pelagic Mollusca, alone or with co-workers or students. Part of the type-material of the various new taxa described in these papers is nowadays incorporated in the Naturalis collection.

Next to his work on fossil pelagic Mollusca Arie W. Janssen also worked on present-day pelagic Mollusca. He performed extensive identification- and registration-work, mostly on substantial ocean bottom sediment samples collected during the various RMNH expeditions on the Atlantic and in the Caribbean (such as the CANCAP, Mauritania or Saba Bank expeditions). He published various papers on present-day pelagic Mollusca, alone or with co-workers. The type-material of the various newly described taxa is incorporated in the Naturalis collections. In the past years a new Naturalis research group under the leadership of Dr. K.T.C.A. (Katja) Peijnenburg on the topic of development and evolution of pelagic Mollusca emerged (see next chapter).

The way of storing the present-day collections is shown in Figures 5-6. A survey and quantification of the RMNH pelagic mollusc collection is presented in Tables 4-17.

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Creseidae									
<i>Creseis</i> sp.	14	12	-	-	-	2	-	-	-
<i>Creseis acicula</i>	123	69	2	15	1	31	4	1	-
<i>Creseis clava</i> ²	33	1	16	-	-	14	-	1	1
<i>Creseis conica</i>	32	-	23	-	-	4	-	5	-
<i>Creseis virgula</i>	99	29	32	13	1	19	1	2	2
<i>Creseis virgula constricta</i> ³	2	1	1	-	-	-	-	-	-
<i>Styliola subula</i>	264	55	117	73	-	18	1	-	-
Subtotals	567	167	191	102	2	88	6	9	3
Family Hyalocylidae									
<i>Hyalocylis striata</i>	60	17	18	18	1	2	1	2	1
Totals	627	184	209	119	3	90	7	11	4

¹ Not yet registered lots, data only available from the label.

³ Usually considered a forma of *Boasia chierchia* (Boas, 1886) s.lat..

² Synonym of *Creseis acicula*.

Table 9. RMNH collection of Recent Creseidae and Hyalocylidae; number of lots specified per region. Taxon names in bold indicate that the primary type material is in the RMNH collection.

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Cuvierinidae									
<i>Cuvierina atlantica</i>	140	38	84	-	-	18	-	-	-
<i>Cuvierina cancapae</i>	25	-	22	-	-	3	-	-	-
<i>Cuvierina columnella</i>	30	15	3 ²	1 ²	-	2 ²	1	5	3
<i>Cuvierina pacifica</i>	1	-	-	-	-	-	-	-	1
<i>Cuvierina tsudai</i>	6	6	-	-	-	-	-	-	-
<i>Cuvierina urceolaris</i>	14	6	-	-	-	-	1	7	-
Totals	216	65	109	1	-	23	2	12	4

¹ Not yet registered lots, data only available from the label.

² Needs reidentification, *C. columnella* does not occur in the Atlantic, the Mediterranean or the Caribbean.

Table 10. RMNH collection of Recent Cuvierinidae; number of lots specified per region. Taxon names in bold indicate that the primary type material is in the RMNH collection.

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Cliidae									
<i>Clio antarctica</i>	232	230	-	-	-	-	-	-	2
<i>Clio chaptalii</i>	3	2	-	-	-	-	-	-	1
<i>Clio convexa</i>	5	2	-	3 ²	-	-	-	-	-
<i>Clio cuspidata</i>	89	46	12	28	-	3	-	-	-
<i>Clio piatkowskii</i>	384	383	1 ³	-	-	-	-	-	-
<i>Clio polita</i>	3	3	-	-	-	-	-	-	-
<i>Clio pyramidata</i> ⁴ s.lat.	1602	1512	13	65	2	9	-	-	1
<i>Clio pyramidata excisa</i>	6	-	-	6	-	-	-	-	-
<i>Clio pyramidata lanceolata</i>	340	193	125	3	-	14	-	5	-
<i>Clio pyramidata pyramidata</i>	115	115	-	-	-	-	-	-	-
<i>Clio recurva</i>	33	27	6	-	-	-	-	-	-
<i>Clio sulcata</i>	704	694	9	-	-	-	1	-	-
<i>Clio</i> sp.	11	6	2	3	-	-	-	-	-
Totals	3527	3213	168	108	2	26	1	5	4

¹ Not yet registered lots, data only available from the label.

³ Holotype (from Weddell Sea, Antarctic Ocean).

² Needs reidentification, *Clio convexa* is not known to occur in the Mediterranean.

⁴ The *C. pyramidata*-group needs reidentification.

Table 11. RMNH collection of Recent Cliidae; number of lots specified per region. Taxon names in bold indicate that the primary type material is in the RMNH collection.

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Cavoliniidae									
<i>Cavoliniidae</i> non det. ²	162	162	-	-	-	-	-	-	-
<i>Cavolinia gibbosa</i> s.lat.	83	28	31	10	-	7	2	5	-
<i>Cavolinia globulosa</i>	157	150	-	-	-	-	2	5	-
<i>Cavolinia inflexa</i> s.lat.	275	40	173	35	-	25	-	1	1
<i>Cavolinia tridentata</i> s.lat.	153	36	64	9	-	34	2	6	2
<i>Cavolinia tridentata atlantica</i>	1 ⁴	-	1	-	-	-	-	-	-
<i>Cavolinia tridentata bermudensis</i>	1 ⁴	-	1	-	-	-	-	-	-
<i>Cavolinia tridentata dakarensis</i>	1 ⁴	-	1	-	-	-	-	-	-
<i>Cavolinia tridentata danae</i>	1 ⁴	-	-	-	-	-	1	-	-
<i>Cavolinia tridentata teschi</i>	2	-	-	-	-	-	-	2	-
<i>Cavolinia tridentata tridentata</i>	1	-	-	-	-	1	-	-	-
<i>Cavolinia uncinata</i>	144	40	42	1	2	34	1	23	1
<i>Cavolinia uncinata pulsatapusilla</i>	1 ⁴	-	-	-	1	-	-	-	-
<i>Cavolinia uncinata pulsatoides</i>	1 ⁴	-	-	-	1	-	-	-	-
<i>Cavolinia</i> sp.	49	48	-	-	-	-	-	1	-

Table 12. RMNH collection of Recent Cavoliniidae; number of lots specified per region. Taxon names in bold indicate that the primary type material is in the RMNH collection (continued on p. 238).

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
<i>Diacavolinia angulosa</i>	10	2	-	-	-	-	-	8	-
<i>Diacavolinia aspina</i>	1	-	-	-	-	-	1	-	-
<i>Diacavolinia atlantica</i>	2	-	2	-	-	-	-	-	-
<i>Diacavolinia bandaensis</i>	2	-	-	-	-	-	-	2	-
<i>Diacavolinia bicornis</i>	1	-	-	-	-	-	1	-	-
<i>Diacavolinia constricta</i>	1	-	1	-	-	-	-	-	-
<i>Diacavolinia deblainvillei</i>	2	-	2	-	-	-	-	-	-
<i>Diacavolinia deshayesi</i>	1	-	1 ⁵	-	-	-	-	-	-
<i>Diacavolinia elegans</i>	1	-	-	-	-	-	-	1 ⁵	-
<i>Diacavolinia flexipes</i>	16	2	-	-	10	-	-	2	2
<i>Diacavolinia grayi</i>	2	-	-	-	-	-	-	2	-
<i>Diacavolinia limbata</i>	2	2	-	-	-	-	-	-	-
<i>Diacavolinia longirostris africana</i>	2	-	2	-	-	-	-	-	-
<i>Diacavolinia longirostris angulata</i> ³	1	-	-	-	-	-	-	1	-
<i>Diacavolinia longirostris flexipes</i> ³	1	-	-	-	1	-	-	-	-
<i>Diacavolinia longirostris longirostris</i>	4	-	-	-	-	4	-	-	-
<i>Diacavolinia longirostris</i> s. lat. ⁶	361	100	112	-	3	131	-	14	1
<i>Diacavolinia ovalis</i>	1	-	1	-	-	-	-	-	-
<i>Diacavolinia pacifica</i>	2	-	-	-	-	-	-	2	-
<i>Diacavolinia robusta</i>	1	-	1	-	-	-	-	-	-
<i>Diacavolinia souleyeti</i>	1	-	-	-	-	-	1 ⁵	-	-
<i>Diacavolinia strangulata</i>	1	1	-	-	-	-	-	-	-
<i>Diacavolinia striata</i>	1	-	-	-	-	-	1 ⁵	-	-
<i>Diacavolinia triangulata</i>	2	-	-	-	-	-	-	-	2
<i>Diacavolinia vanutrechti vanutrechti</i>	1	-	-	-	-	-	-	-	1 ⁵
<i>Diacavolinia</i> sp.	1	1	-	-	-	-	-	-	-
<i>Diacria atlantica</i>	2	-	-	2	-	-	-	-	-
<i>Diacria major</i>	16	-	15	-	-	-	-	1	-
<i>Diacria rampalae</i>	4	3	-	-	-	-	-	-	1 ⁵
<i>Diacria rubecula</i>	2	-	2	-	-	-	-	-	-
<i>Diacria trispinosa trispinosa</i>	552	341	149	14	-	42	-	5	1
<i>Diacria trispinosa africana</i>	1	-	1 ⁵	-	-	-	-	-	-
<i>Diacria</i> sp.	16	15	-	-	-	1	-	-	-
<i>Telodiacria costata</i>	8	-	-	-	-	-	-	8	-
<i>Telodiacria danae</i>	99	1	96	-	-	-	-	2	-
<i>Telodiacria quadridentata</i>	36	14	3	1	5	12	1	-	-
<i>Telodiacria quadridentata erythra crassa</i>	1	-	-	-	1 ⁵	-	-	-	-
<i>Telodiacria quadridentata erythra erythra</i>	1	-	-	-	1 ⁵	-	-	-	-
<i>Telodiacria quadridentata quadridentata</i>	6	-	4	-	-	2	-	-	-
<i>Telodiacria quadridentata quadridentata</i> f. <i>schmidti</i>	15	13	-	-	-	-	-	-	2 ⁵
Totals	2212	999	705	72	25	293	13	91	14

¹ Not yet registered lots, data only available from the label.² Includes Creseidae s.lat.³ Probably includes more taxa⁴ Paratype (taxon unavailable)⁵ Paratype(s).⁶ Needs reidentification, may include more taxa.

Table 12, continued.

Name	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Peraclidae									
<i>Peracle apicifulva</i> ²	5	1	-	4	-	-	-	-	-
<i>Peracle bispinosa</i>	20	4	16	-	-	-	-	-	-
<i>Peracle diversa</i>	18	4	6	8	-	-	-	-	-
<i>Peracle elata</i>	14	2	12	-	-	-	-	-	-
<i>Peracle moluccensis</i>	14	2	-	-	-	-	-	12	-
<i>Peracle reticulata</i>	51	5	33	13	-	-	-	-	-
<i>Peracle reticulata minor</i> ³	2	-	-	-	-	-	-	2	-
<i>Peracle rissoides</i> ⁴	1	-	-	-	-	-	-	1	-
<i>Peracle triacantha</i> ⁵	3	1	2	-	-	-	-	-	-
<i>Peracle valdiviae</i>	1	1	-	-	-	-	-	-	-
Subtotals	129	20	69	25	-	-	-	15	-
Family Cymbuliidae									
<i>Corolla calceola</i>	1	-	-	-	-	-	-	-	-
<i>Corolla cupula</i>	1	-	1	-	-	-	-	-	-
<i>Corolla intermedia</i>	6	-	-	-	-	-	-	6	-
<i>Corolla ovata</i>	4	2	-	-	-	-	-	2	-
<i>Corolla spectabilis</i>	1	-	-	-	-	-	-	-	1
<i>Corolla</i> sp.	2	2	-	-	-	-	-	-	-
<i>Cymbulia parvidentata</i>	1	1	-	-	-	-	-	-	-
<i>Cymbulia peronii</i>	1182	1176	-	7	-	-	-	-	-
<i>Cymbulia peronii minor</i>	1	-	1	-	-	-	-	-	-
<i>Cymbulia sibogae</i>	4	-	-	-	-	-	-	4	-
<i>Cymbulia</i> sp.	57	57	-	-	-	-	-	-	-
<i>Gleba cordata</i>	5	3	-	2	-	-	-	-	-
Subtotals	1265	1241	2	9	-	-	-	12	1
Family Desmopteridae									
<i>Desmopterus papilio</i>	3	3	-	-	-	-	-	-	-
Totals	1397	1264	71	34	-	-	-	27	1

¹ Not yet registered lots, data only available from the label.⁴ Synonym of *Peracle reticulata*, according to van der Spoel (1976: 30).² Synonym of *Peracle diversa*.⁵ Synonym of *Peracle elata*.³ Synonym of *Peracle reticulata*, according to Tesch (1913: 73).**Table 13.** RMNH collection of Recent Pseudothecosomata; number of lots specified per region. Taxon names in bold indicate that the primary type material is in the RMNH collection.

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Pneumodermatidae									
<i>Pneumoderma atlanticum</i> ²	2	2	-	-	-	-	-	-	-
<i>Pneumoderma degraaffi</i>	1	-	-	-	-	-	-	-	1
<i>Pneumoderma heronensis</i>	1	-	-	-	-	-	-	-	1
<i>Pneumoderma mediterraneum</i>	2	1	-	1	-	-	-	-	-
<i>Pneumoderma violaceum</i>	19	17	1	1	-	-	-	-	-
<i>Pneumoderma violaceum souleyeti</i> ²	1	1	-	-	-	-	-	-	-
<i>Pneumoderma heterocotylum</i>	1	-	-	-	-	-	-	1	-
<i>Pneumoderma pygmaeum</i> ²	3	-	-	-	-	-	-	3	-
<i>Pneumoderma teschi</i>	1	1	-	-	-	-	-	-	-
<i>Pneumodermopsis ciliata</i>	9	8	-	1	-	-	-	-	-
<i>Pneumodermopsis michaelsarsi</i>	2	1	1	-	-	-	-	-	-
<i>Spongiobranchaea australis</i>	237	236	-	-	-	-	1	-	-
Subtotals	279	267	2	3	-	-	1	4	2
Family Notobranchaeidae									
<i>Notobranchaea bleekerae</i>	1	-	1	-	-	-	-	-	-
<i>Notobranchaea inopinata</i>	1	1	-	-	-	-	-	-	-
<i>Notobranchaea macdonaldi</i>	7	7	-	-	-	-	-	-	-
<i>Notobranchaea tetrabranchiata</i>	3	3	-	-	-	-	-	-	-
Subtotals	12	11	1	-	-	-	-	-	-
Family Cliopsidae									
<i>Cliopsis krohni</i>	2	1	-	1	-	-	-	-	-
<i>Cliopsis microcephalus</i> ³	1	1	-	-	-	-	-	-	-
<i>Pruvotella danae</i>	1	1	-	-	-	-	-	-	-
Subtotals	4	3	-	1	-	-	-	-	-
Family Clionidae									
<i>Clione gracilis</i> ⁴	3	1	2	-	-	-	-	-	-
<i>Clione limacina antarctica</i>	6	5	1	-	-	-	-	-	-
<i>Clione limacina meridionalis</i>	1	-	1	-	-	-	-	-	-
<i>Clione limacina</i> s.lat.	561	551	6	-	-	-	-	-	4
<i>Fowlerina punctata</i>	1	-	-	-	-	-	-	1	-
<i>Paedoclione doliiformis</i>	1	-	-	1 ⁵	-	-	-	-	-
<i>Paraclione flavescens</i>	2	2	-	-	-	-	-	-	-
<i>Paraclione pelseneeri</i>	4	-	-	-	-	-	-	4	-
<i>Thliptodon antarcticus</i>	1	1	-	-	-	-	-	-	-
<i>Thliptodon diaphanus</i>	4	3	-	-	-	-	-	1	-
<i>Thliptodon gegenbauri</i>	1	1	-	-	-	-	-	-	-
<i>Thliptodon schmidti</i>	1	1	-	-	-	-	-	-	-
Subtotals	586	565	10	1	-	-	-	6	4

Table 14. RMNH collection of Recent Gymnosomata; number of lots specified per region. Taxon names in bold indicate that the primary type material is in the RMNH collection (continued on p. 241).

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Hydromylidae									
<i>Hydromyles globulosus</i>	122	110	-	-	-	-	-	12	-
Family Laginiopsidae (not represented)	-	-	-	-	-	-	-	-	-
Totals for Gymnosomata	1003	956	13	5	-	-	1	22	6

¹ Not yet registered lots, data only available from the label.

² Synonym of *Pneomoderma violaceum*.

³ Synonym of *Cliopsis krohni* 'morph' *modesta*, according to van der Spoel (1976: 94).

⁴ Forma of *Clione limacina*, according to van der Spoel (1967: 114).

⁵ Needs reidentification, *Paedoclione doliiformis* is not known to occur in the Mediterranean. The record is for a shell, but Clionidae larval shells are not yet identifiable. Currently a correct identification would be 'Clionidae sp.'.

Table 14, continued.

	Number of lots	No data ¹	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Family Phylliroidae									
<i>Phylliroe bucephala</i>	4	-	1	3	-	-	-	-	-
Family Fionidae									
<i>Fiona pinnata</i>	2	-	-	2	-	-	-	-	-
Family Glaucidae									
<i>Glaucus atlanticus</i>	1	-	-	-	-	-	1	-	-
Totals	7	-	1	5	-	-	1	-	-

¹ Not yet registered lots, data only available from the label.

Table 15. RMNH collection of Recent pelagic Nudibranchia; number of lots specified per region.

	Number of lots	No data	Atlantic Ocean	Mediterranean	Red Sea	Caribbean	Indian Ocean	Indonesia	Pacific
Pterotracheoidea									
Atlantidae	909	122	532	64	-	58	13	104	16
Pterotracheidae	273	219	6	10	1	-	1	35	1
Carinariidae	88	20	9	21	-	1	5	27	5
Epitonioidae									
Epitoniidae	540	258	69	17	7	104	32	11	42
Limacinoidea									
Heliconoididae	349	58	214	39	-	30	-	8	-
Limacinidae	472	102	300	39	-	19	3	8	1
Thieleidae	10	5	5	-	-	-	-	-	-
Cavolinioidae									
Creseidae	567	167	191	101	2	88	6	9	3
Hyalocylidae	60	17	18	18	1	2	1	2	1
Cuvierinidae	216	65	109	1	-	23	2	12	4
Cliidae	3527	3213	168	108	2	26	1	5	4
Cavoliniidae	2212	999	705	72	25	293	13	91	14
Cymbulioidea									
Peraclidae	129	20	69	25	-	-	-	15	-
Cymbuliidae	1265	1241	2	9	-	-	-	12	1
Desmopteridae	3	3	-	-	-	-	-	-	-
Clionoidea									
Pneumodermatidae	279	267	2	3	-	-	1	4	2
Notobranchaeidae	12	11	1	-	-	-	-	-	-
Cliopsidae	4	3	-	1	-	-	-	-	-
Clionidae	586	565	10	1	-	-	-	6	4
Hydromyloidea									
Hydromylidae	122	110	-	-	-	-	-	12	-
Laginiopsidae	-	-	-	-	-	-	-	-	-
Nudibranchia									
Glaucidae	1	-	-	-	-	-	1	-	-
Phylliroidae	4	-	1	3	-	-	-	-	-
Fionidae	2	-	-	2	-	-	-	-	-
Totals	11.630	7.518	2.411	534	38	644	79	360	98

Table 16. Recapitulation of Recent pelagic Mollusca in the RMNH collection; numbers of lots specified per region.

	shells	whole/soft ¹	not specified		shells	whole/soft ¹	not specified
Atlantidae	806	102	-	Cymbuliidae	16	1249	-
Carinariidae	44	46	-	Desmopteridae	-	3	-
Pterotracheidae	16	257	-	Pneumodermatidae	1	278	-
Epitoniidae	515	24	-	Notobranchaeidae	-	12	-
Limacinidae	418	54	-	Cliopsidae	-	3	1
Thieleidae	5	5	-	Clionidae	3	583	-
Creseidae	436	22	62	Hydromylidae	-	122	-
Hyalocylidae	47	13	-	Laginiopsidae	-	-	-
Cuvierinidae	145	70	1	Phylliroidae	-	4	-
Cliidae	322	3150	55	Fionidae	-	7	-
Cavoliniidae	1652	547	6	Glaucidae	-	1	-
Peraclidae	128	-	1	Totals	4.910	6.591	126

¹ Whole organism or soft parts.

Table 17. Recapitulation of Recent pelagic Mollusca in the RMNH collection (left column predominantly dry collection, middle column alcohol collection, right column not specified in the register).

THE NATURALIS PLANKTON DIVERSITY AND EVOLUTION RESEARCH GROUP (KTCAP & AWJ)

In recent years there has been renewed interest in pelagic molluscs worldwide and at Naturalis, which resulted in a new research group, established in 2020, named ‘Plankton Diversity and Evolution’ and led by Dr. Katja Peijnenburg. The work on pelagic gastropods in this group has focussed on establishing species boundaries, biogeography and evolution, employing a combination of phylogenetic, morphological and genomic approaches. For this purpose, new collections were obtained during large oceanographic research expeditions mainly focussing on the Atlantic Ocean. These samplings were realised using different types of plankton nets such as Bongo nets (Fig. 7) or the Rectangular Midwater Trawl (Fig. 8), usually at night. Sorted samples of pelagic gastropods are available for three basin-scale transects (~45° North to ~45° South from 2012, 2014 and 2017) sampled in collaboration with the Atlantic Meridional Transect programme (<https://www.amt-uk.org>). In addition, samples of pelagic gastropods from several locations in the Indian and Pacific Oceans are available. These samples are mainly preserved in 96% ethanol and stored at -20°C to be suitable for molecular as well as morphological analyses. Samples that are used in publications have appropriate registration codes and data are accessible in public databases such as Barcode Of Life Data System (BOLD: <https://www.boldsystems.org>) and GenBank. In addition, analysed specimens have museum registration numbers,



Fig. 8. Preparing the Rectangular Midwater Trawl (RMTFF) for sampling of zooplankton during Atlantic Meridional Transect 22 in 2012.

DNA vouchers are stored at -80°C and type specimens of newly described species have been added to the collection.

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Fig. 7. Sampling of zooplankton with a Bongo net during Atlantic Meridional Transect 27 in 2017.

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APPENDIX 1.

STORAGE OF PYRITISED SPECIMENS (AWJ & RP)

Introduction

Long term preservation of pyritised fossils has always been a big problem in natural history collections, and there is extensive literature on the subject (see Howie, 1977a-b, 1978, 1979, 1992; Waller, 1987; Clark, 2003; Larkin, 2011; Cavallari et al., 2014, and references therein). Pyrite (FeS_2) is an unstable mineral that easily disintegrates when in contact with the atmosphere. Pyrite may replace fossils entirely, cover fossils, or fill cavities within. Many plant fossils have been found, in which tissues were completely replaced by pyrite, still preserving very fine details. Larger bones are sometimes filled or covered with pyrite. In the case of fossil molluscs usually the shell is filled with pyrite, but frequently the shell is dissolved, and only the internal pyrite mould remains. Especially the microcline type of pyrite is easily oxidised, but also other types of crystallisation are attacked.

Several treatments have been proposed. The minimal approach is to ‘do nothing, it’s no use’. Usual techniques involve covering the fossils with a natural or artificial resin, or another substance to prevent contact with air. Also pyritised specimens can be stored in various kinds of liquid (glycerine, paraffin oil, kerosine, even antifreeze, and silicone oil), treated with ammonium gas or closed-chamber treatment with ethanolamine thioglycollate (which required advanced laboratory facilities). For storage, specimens should be kept below a relative humidity of circa 45% (preferably 35% but certainly less than 60%) (Howie, 1992) and/or low oxygen levels (which needs micro- or macro-climate controlled storage). Another, but rather laborious method for the preservation of pyrite fossils is enclosing the

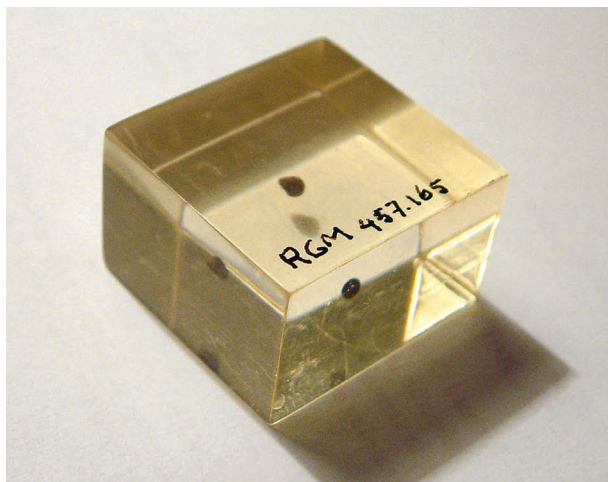
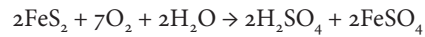


Fig. 9. RGM.457165, pyritic pteropod embedded in a block of artificial resin (donated by the late Karl Gürs).

specimens in blocks of artificial resin (Fig. 9). This method was applied in the Karl Gürs collection (see Gürs & Gürs, 1981). Air bubbles are difficult to avoid when embedding the specimens. The procedure, including polishing the blocks, is very timeconsuming.

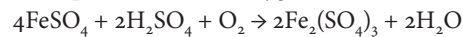
Pyrite disintegration

Cadée (1961) formulated the deterioration of pyrite as follows:



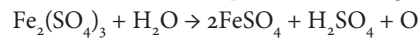
(pyrite + oxygen + water \rightarrow sulphuric acid + ferro sulphate)

In cases where the water soluble ferro sulphate is not removed (such as in collections) the ferro sulphate reacts with sulphuric acid and oxygen to form ferri sulphate:



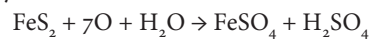
(ferro sulphate + sulphuric acid + oxygen \rightarrow ferri sulphate + water)

after which the ferri sulphate oxidises again:



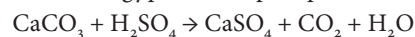
(ferri sulphate + water \rightarrow ferro sulphate + sulphuric acid + oxygen in statu nascendi)

Oxygen in statu nascendi of course strongly oxidises the pyrite:



(pyrite + oxygen s.n. + water \rightarrow ferro sulphate + sulphuric acid)

and the whole cycle starts all over again. If calcium carbonate is available (shells) it will be attacked by the sulphuric acid, and gypsum will precipitate:



(calcium carbonate + sulphuric acid \rightarrow gypsum + carbon dioxide + water)

Similar reactions were proposed by Buurman (1970), who also noted the occurrence of the minerals goethite and jarosite in mollusc samples with oxidised pyrite.

Preservation methods used in the RGM collection

In the 1970s and early 1980s preventing pyrite disintegration in molluscs housed in the RGM collections was attempted by immersion of fossils in a solvent of acetone + any kind of ‘plastic’ glue (so-called ‘placeton’) or, applying a diluted mix of acetone and ‘glyptal’, a synthetic resin used for preserving fossil mammals bones. The general idea was to seal off the pyrite from air to prevent oxidation. Both methods seemed to be helpful initially, but in the long run it was clear that further disintegration of the pyrite could not be stopped in that way. Even carefully coated specimens seemed to be attacked from inside. Humidity is supposed to affect pyritic fossils most. Similar results were described by Neal Clark (2003), who wrote: ‘We have had several carefully conserved specimens explode spectacularly due to pyrite “rot” building up under the protective skin of solvents.’



Fig. 10. Storage of pyritised pteropod samples in silicone oil, in upright glass vial, in polystyrene container, with labels separated, in standard RGM collection box.

Contact in 1986 with Mr John Cooper and Dr Peter Whybrow, of the (at that time) British Museum (Natural History), in London, led to the introduction of a strongly advised and entirely different way of preserving smaller pyritised fossils, viz. permanent immersion of specimens in silicone oil of a specifically prescribed type and extracting all air from the sample by using a vacuum pump. The type of silicone oil recommended was on sale from British Drug Houses (BDH) and indicated as DC 550 (viscosity 200/350 centistokes). Nowadays this type of silicone oil is widely available through many distributors. The inert silicone liquid is supposed to fill all fissures in the fossils and prevent air to come in contact. New oxygen or water from the air cannot reach the fossils in this way. To remove silicone oil from fossils for subsequent study many hydrocarbon solvents can be used, the least harmful and easiest available being white gas / solvent naphtha and white spirit / turpentine. This preservation method seemed so reliable and from the British Museum we experienced such favourable results that, after some tests, it was decided to transfer all Oligocene mollusc lots containing pyrite into silicone oil. For the purpose of smaller fossils special glass containers are used (with a plastic clip cap) that have to be stored upright, separate from labels, as the high surface tension of the silicone makes horizontal storage of the vials virtually impossible (Fig. 10). If stored horizontally, a thin layer of oil will creep onto the external surface of the vials as the plastic caps do not close them sufficiently. A small label with registration number is kept in the vials, together with the fossils. Lots in silicone oil are stored together with 'dry' lots, in drawer cabinets.

Ever since this system was initiated, over a thousand lots in silicone oil have been added to the holoplanktic mollusc collection. More than half of these lots (541) were obtained from the Landesamt für Natur und Umwelt Schleswig-Hol-

stein, at Flintbek, Germany, through the mediation of the late Dr Karl Gürs, to which institute we 'guaranteed' the long-lasting preservation of the specimens in silicone oil. This procedure was finished in 1997, but afterwards additionally obtained specimens were added. Also a large collection, received on loan from the Instytut Nauk Geologicznych PAN, Zakład Geologii Dynamicznej, at Kraków, Poland, was returned after publication (Janssen & Zorn, 1993), predominantly stored in silicone oil. The present state of this collection is not known.

Early 2003, when placing newly obtained material into the collection, it was discovered that some of the smaller labels bearing the registration numbers, inside the silicone vials, showed brown staining (Fig. 11).



Fig. 11. Label attacked by pyrite disintegration, kept in vial with silicone oil.

Closer inspection, under a binocular microscope, revealed that in such samples, unexpectedly, pyrite specimens had severely disintegrated. It was found that quite a number of samples showed similar characteristics and an inventory of the damage seemed necessary. Apparently disintegration of pyrite, that practically always has started already before the specimens are picked, identified, registered and stored, had continued in spite of immersion in silicone oil. Frequently such silicone storage (especially in the case of borehole samples) takes place several to even many years after the moment of collecting the material, giving enough time to start the destructive process. The initial idea, that immersion in silicone oil would stop disintegration of pyrite evidently appeared to be wrong. An inventory of the state of the collection was executed in August 2003, by inspecting all silicone lots, estimating the amount of damage in four grades (see also Fig. 12):

- 1 bad – all specimens are to be considered lost;
- 2 mainly bad – one or few specimens may still be recognisable;
- 3 mainly good – most specimens still recognisable, but sample clearly attacked;
- 4 good – all specimens recognisable, but some disintegration may be visible.

Brown colouring of the label with the registration number occurred only in those samples in which disintegration was bad or mainly bad. In some larger lots even the polyethylene cap of the vials showed brown discoloration.

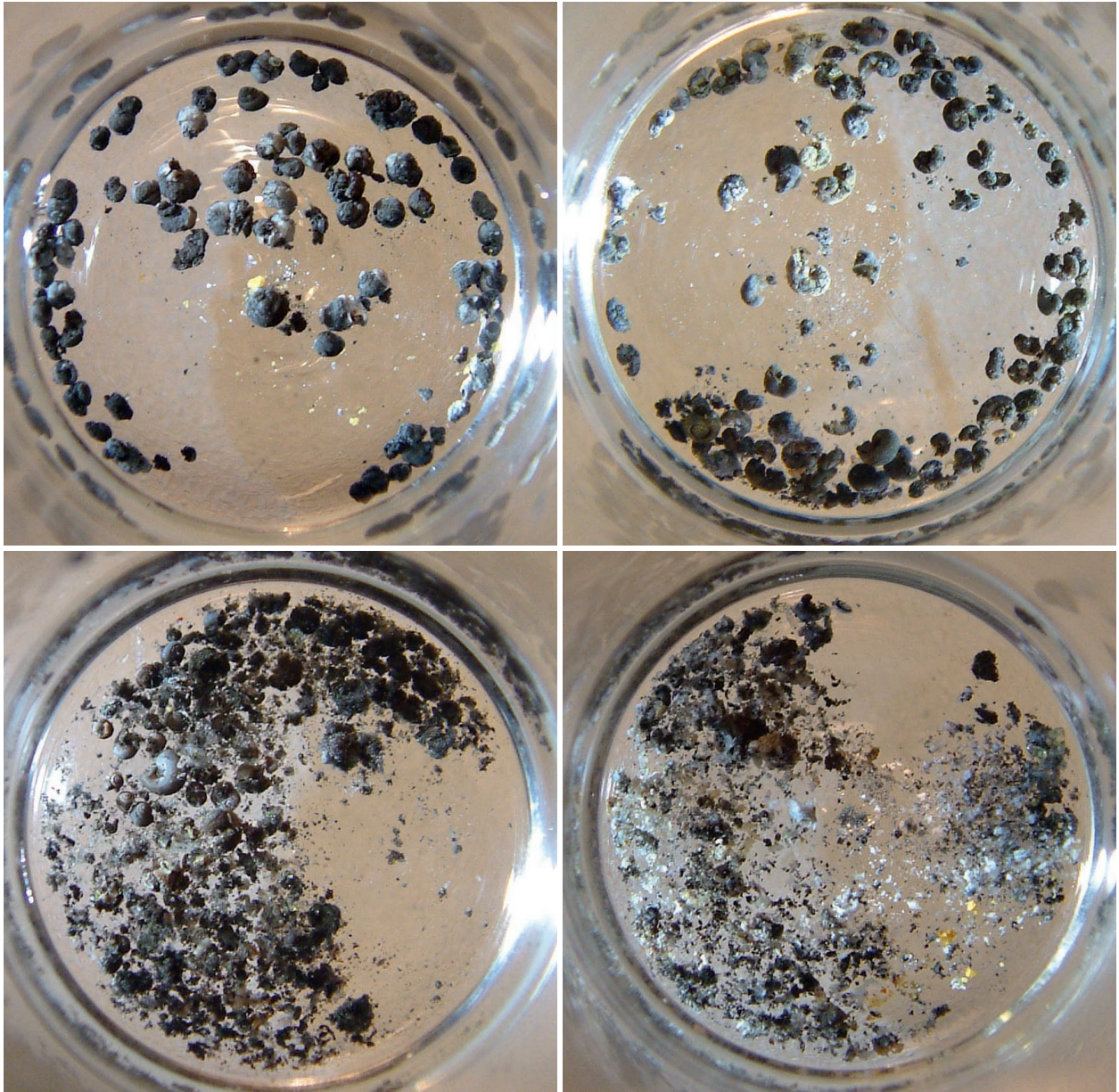


Fig. 12. Pyritised limacinid pteropods preserved in silicone oil, from upper left to lower right in good, mainly good, mainly bad and bad condition, respectively. *Limacina valvatina* (RGM.457059), indicated 'good', *Limacina miorostralis* (RGM.396519), indicated 'mainly good', *Limacina valvatina* (RGM.395976), indicated 'mainly bad', *Limacina atlanta* (RGM.397017), indicated 'bad'. Diameter of glass vial 18 mm.

Quantification of the damage

A total of 1.067 lots of holoplanktic molluscs in silicone oil were inspected, and the state of each sample was noted according to the four grades described above. The results were:

bad	186 lots	= 17.4%
mainly bad	114 lots	= 10.7%
mainly good	77 lots	= 7.2%
good	690 lots	= 64.7%

which means that more than a quarter of the samples were in bad condition. The 300 lots that have to be considered lost,

or mainly lost, unfortunately include several holotypes, and quite a number of paratypes and/or illustrated specimens.

Holotypes of the species *Limacina ingridae*, *L. irisaie*, *L. jessyae* and *Clio blinkae* are lost. Four other type specimens, however, are still in good condition: *Limacina mariae*, *L. wilhelminae*, *Creseis berthae* and *Clio jacobae*. All these species were introduced in Janssen (1989) and immersed in the silicone oil at the same time. For the first mentioned group of types a number of (paratype) lots are still in good condition. If the database with the results of the inventory

is sorted by locality it becomes immediately clear, that the condition of the specimens usually is not identical for samples of the same origin. Even in series from the same borehole, from which samples may be supposed to be collected and immersed more or less simultaneously, the condition may vary considerably. Some examples are given in Table 18.

The boreholes Karlum and Reinfeld are the only ones in which all samples are still in good condition. Most probably those series were studied and curated shortly after the boreholes were made. Most samples collected after 1997 (ever since the collection was housed in Malta, see Appendix 2) have not been treated with a vacuum pump, as such equipment was not available there. However, the series of samples from Kruikebeke was curated in Leiden (1988), where a vacuum pump was available, and more than half of the lots were in bad or mainly bad condition in 2003. According to F.P. Wesselingh, who was curator of the collection in 2003, especially thin-walled shells, such as *Thyasira* spp., kept in the silicone collection of fossil benthic molluscs housed in Leiden (mainly Oligocene of the North Sea Basin) suffered from disintegration. The damage, however, is less dramatic. Early 2020, when preparing the present paper, it was intended to execute another check of silicone lots, to find out if the damage has progressed since the first check in 2003. However, the COVID-19 crisis made work in the collection impossible. Consequently that task has to be postponed to a later date.

Future curation

According to the literature on pyrite preservation the best method to keep pyrite samples in good condition is housing them in environments with a low (c. 45%) relative humidity (RH) and low oxygen. The store rooms of Naturalis have a RH of 50%. For the majority of specimens, however, silicone immersion still seems to be a good method, provided that samples are curated as soon as possible after collecting. A certain amount of loss has to be accepted. A good suggestion might be to immerse specimens immediately when they are isolated from their original sediment samples, and not wait for identification and further curation. Possible application of silica gel, as in use for some products in the pharmaceutical industry, was suggested by the late Dr Karl Gürs as a solution for dry storage. An inquiry about these possibilities at ‘Fisher Group Ltd. Airconditioning’ at Glasgow (<https://www.fishergroup.co.uk/>) sent together with a description of the situation, resulted in the following answer (Mr Philip Handley, email dated August 18, 2003).

‘From the paper you attached it seems that the major problem is not so much with moisture but with oxidation causing deterioration. Using a dessiccant will have no effect on this at all and, in fact, having too dry an atmosphere could be detrimental as your samples may have “waters of crystallisation” as part of the mineral content. Dehydration could

lead to the disintegration of the crystals. I would recommend the use of silicon oil to act as a barrier to air or moisture. If you must have a dessiccant and oxygen free atmosphere then I would recommend the use of silica gel and an inert replacement atmosphere such as nitrogen.’

Clean pyrite does not have ‘waters of crystallisation’ (contrary to its weathering products goethite and jarosite), so after all application of silica gel might be helpful. In the latter case application of silica gel powder or grains, separated from the dry fossils by some cotton wool in a glass vial with tight screw cap might represent a long lasting solution. Creating a microclimate with not only low RH but also low oxygen levels will give the best results for dry storage. This can be achieved by storing the samples in a sealed bag or tightly closed box, using both a dessiccant like silicagel and an oxygen scavenger (Larkin, 2011).

Condition	Good	Mainly good	Mainly bad	Bad
Bargfeld (borehole, 28 lots)	12	-	2	14
Bönningstedt (borehole, 74 lots)	60	8	3	3
Bovenau (borehole, 31 lots)	14	4	4	9
Glinde (borehole, 15 lots)	4	2	-	9
Gross Pampau (borehole, 57 lots)	35	7	11	4
IJsselmuider (borehole, 54 lots)	39	3	-	12
Kaltenkirchen (borehole (97 lots)	41	10	21	25
Karlum (boreholes, 57 lots)	57	-	-	-
Kruikebeke (claypit, 33 lots)	13	2	8	10
Lübtheen (borehole, 73 lots)	69	2	2	-
Opende (borehole, 46 lots)	16	1	8	21
Reinfeld (borehole, 47 lots)	47	-	-	-
Rösing (borehole, 30 lots)	20	5	3	2
Springhirsch (borehole, 22 lots)	9	6	4	3
Viöl (borehole, 23 lots)	22	1	-	-

Table 18. Quantification of preservation condition of pyritised pteropod lots from various borehole- or outcrop locations.

APPENDIX 2. HISTORY OF THE RGM MALTA COLLECTION (AWJ)

In 1981 D'Alessandro & Robba published their study of pteropods from Miocene deposits in Puglia, southern Italy, which covered two main areas of research, namely Gargano and Salento. Material from the Gargano localities, such as Sannicola Varano, was available in the Leiden RGM collection. Specimens from the Salento localities (Melpignano and Cursi), however, were not yet represented and in 1992 the first author (AWJ) decided to attempt some collecting there during a trip in southern Italy. Several localities in the area were sampled successfully and rich material was obtained. Puzzling in that material was the occurrence of the species *Cavolinia cookei* Simonelli, 1895 (Fig. 13), according to D'Alessandro & Robba (1981: 659) originally described from the Early Miocene (Aquitanian) Lower Globigerina Limestone of Malta. Surprisingly, however, the same species was also recorded from much younger (Langhian) rocks of northern Italy, as described by Robba (1971). The assemblages from Salento were dated as 'reworked from deposits of Late Langhian to Early Serravallian age', which made an Aquitanian age of the *C. cookei* type material unexplainable. Did these type specimens indeed originate from the Lower Globigerina Limestones in Malta? These deliberations led to a visit to the Maltese Archipelago during the same 1992 trip.

In Malta contact was made with the at that time curator of the Maltese National Museum of Natural History in Mdina and the Ghar Dalam Cave and Museum in Birzebbugia, Dr George Zammit Maempel (Fig. 14). Explaining the problem



Fig. 13. Specimens of *Cavolinia cookei* Simonelli, 1895 in phosphoric internal mould preservation, from the Maltese Upper Globigerina Limestone Member (Langhian); specimen size approximately 6 mm.



Fig. 14. Dr George Zammit Maempel, during a visit to AWJ's working room at Xewkija, Gozo, August 2004.

of the origin of the pteropod *Cavolinia cookei* resulted in his immediate and enthusiastic cooperation and he demonstrated several outcrops on the main island of Malta. And indeed, the species was rather easily traced in several localities that were all located in the Upper Globigerina Limestone that is of a Langhian age. Together with Dr Zammit Maempel the information given in the publication of Simonelli (1895) was analysed. That author gave detailed stratigraphical data, inclusive of a composite section of Maltese rocks, supplied by the collector of the specimens, J.H. Cooke. Dr Zammit Maempel, being very well acquainted with Maltese stratigraphy, soon found out that the various layers composing Simonelli's section were upside down and that the lower layers of Simonelli that had yielded the *C. cookei* in reality represented the upper ones and so it was clear that the *C. cookei* type material was not of Aquitanian, but rather of Langhian age, agreeing with the occurrence in northern Italy (and also those in Gargano).

During the initial field work in Malta it was immediately clear how rich in pteropods various outcrops on the island are. Only the species *C. cookei* was documented from Malta, as well as some records of *Vaginella* sp. in older literature. In some of the outcrops pteropods occur by the thousands of specimens, in a wide range of species diversity. Collecting of this material was the second favourable result of this trip and it was followed by a second visit later in the same year and several ones in the following years, resulting in

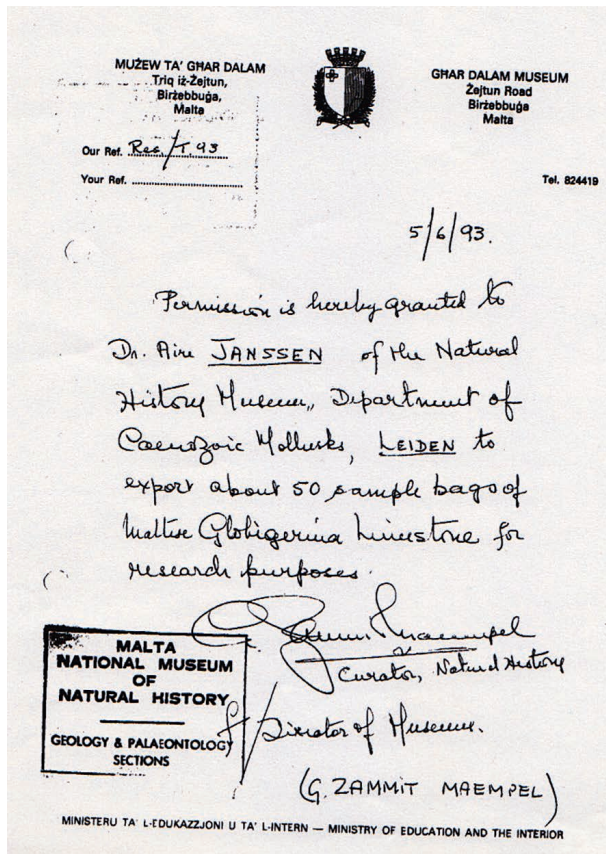


Fig. 15. Example of export permit of Maltese palaeontological samples to The Netherlands, issued by Dr G. Zammit Maempel, June 1993.

an extensive pteropod collection. All collected material was exported to the Netherlands on the basis of export permits issued by Dr Zammit Maempel (see Fig. 15 for an example).

These frequent visits led to the decision of the author's emigration from the Netherlands to Malta after retirement from Naturalis in 1997. Residence was found in the village of Xewkija on the second Maltese island, Gozo, where also numerous interesting outcrops are accessible. The complete RGM collection of fossil holoplanktic molluscs, then existing of about 8.000 lots, was permitted on long-term loan to Malta and was subsequently housed in the author's premises in Xewkija (Fig. 16). Dr Zammit Maempel issued another permit stating that the collection was the property of the Leiden museum and that re-exportation from Malta was permitted when no longer needed.

A first paper on Maltese geology, including the description of a very common but yet unnamed pteropod species (*Gamopleura melitensis*¹ Janssen, 1995), was published in

¹ This cavoliniid species is very common in part of the Globigerina Limestone of Malta and it is as far as known the only fossil pteropod with a local vernacular name. The common presence of this 'qanneb' indicates bad quality of building stone (Zammit Maempel, 1982: 11).

Facies (Rehfeld & Janssen, 1995). Several further new species were introduced in 2004, in the local periodical 'The Central Mediterranean Naturalist' (Janssen, 2004). Ultimately the study of the Maltese holoplanktic molluscs led to the publication of a voluminous monograph (> 400 pp) with the description of 85 species (20 of which new to science) and a biozonation based on the pteropods of the Maltese Late Oligocene and Miocene rocks, augmented with non-Maltese data to be valid for the entire Mediterranean Basin (Janssen, 2012). A collection of several hundred lots of Maltese 'microfossils', among which many pteropod species, was composed already in 2003 and donated to the National Natural History Museum in Mdina, Malta.

Problems concerning the formal status of the Maltese collection material started in 2007, when it was found that the Maltese government, already in 2002, had issued a 'Maltese Heritage Act' (<https://wipolex.wipo.int/en/text/201428>) covering the protection of Maltese cultural heritage objects, inclusive of palaeontological ones. Ultimately this situation led to long deliberations, but in 2017 it appeared to be possible to return the collection to the Netherlands on the basis of the originally issued permits. Currently the Maltese material is integrated in the main fossil pelagic mollusc collection where it will be accessible for any researcher.



Fig. 16. The RGM fossil pelagic mollusc collection in the house of AWJ, ca 2002. The cardboard boxes on top of the drawers contain the hundreds of RMNH lots of present-day specimens for identification and registration.

APPENDIX 3.
SYSTEMATIC LIST OF VALID PELAGIC MOLLUSC SPECIES, FOSSIL AND RECENT (AWJ, KTCAP, DWP)

Here we present a list of all species of pelagic (non-bottom-dwelling, cephalopods excluded) Mollusca we know and that we consider to be valid, with a rough indication of stratigraphic range (stages, E = Early, M = Middle, L = Late) and geographical distribution. The list is composed for the Pteropoda on the basis of the (simplified) table S4 of Peijnenburg et al. (2020) with minor additions and corrections

and including the new family- and genus-names published since Rampal (2019). Pteropoda higher systematics are as proposed by Peijnenburg et al. (2020). The Pterotracheoidea and Nudibranchia parts of the list are based on our own observation, completed with data from the existing literature and several websites. The Epitoniidae are from Beu (2017). The list includes 505 taxa, 216 of which are currently living species, many of which also have a fossil record. Additionally, we include a list of Mesozoic taxa for which currently a pelagic way of life has been convincingly argued for (see Nützel et al., 2016).

Class Gastropoda

Subclass Caenogastropoda

Order Littorinimorpha

Superfamily Pterotracheoidea

Family Atlantidae

<i>Atlanta arenularia</i> Gougerot & Braillon, 1965	Bartonian	France
<i>Atlanta ariejansseni</i> Wall-Palmer et al., 2016	present-day	circumglobal southern temperate
<i>Atlanta brunnea</i> Gray, 1850	Eemian - present-day	circumglobal tropic/subtropic
<i>Atlanta californiensis</i> Seapy & Richter, 1993	present-day	Pacific, California Current, northern temperate
<i>Atlanta cordiformis</i> Gabb, 1873	L Miocene ?	Dominican Republic
<i>Atlanta diamesa</i> Woodring, 1928	Piacenzian	Jamaica
<i>Atlanta echinogyra</i> Richter, 1972	Piacenzian? - present-day	Indopacific tropic/subtropic
<i>Atlanta fragilis</i> Richter, 1993	present-day	circumglobal tropic/subtropic
<i>Atlanta frontieri</i> Richter, 1993	present-day	Indopacific tropic/subtropic
<i>Atlanta gaudichaudi</i> Gray, 1850	Piacenzian - present-day	circumglobal tropic/subtropic
<i>Atlanta gibbosa</i> Souleyet, 1852	present-day	circumglobal tropic/subtropic
<i>Atlanta helicinoidea</i> Gray, 1850	Calabrian - present-day	circumglobal tropic/subtropic
<i>Atlanta inclinata</i> Gray, 1850	Gelasian - present-day	circumglobal tropic
<i>Atlanta inflata</i> Gray, 1850	Gelasian - present-day	Indopacific tropic/subtropic
<i>Atlanta lesueurii</i> Gray, 1850	Piacenzian - present-day	circumglobal tropic
<i>Atlanta lingayanensis</i> Janssen, 2007	Piacenzian	Philippines
<i>Atlanta meteori</i> Richter, 1972	present-day	circumglobal tropic/subtropic
<i>Atlanta oligogyra</i> Tesch, 1906	Piacenzian - present-day	circumglobal tropic/subtropic
<i>Atlanta peronii</i> Lesueur, 1817	Piacenzian - present-day	circumglobal tropic/subtropic
<i>Atlanta plana</i> Richter, 1972	Messinian - present-day	Indopacific tropic/subtropic
<i>Atlanta richteri</i> Janssen, 2007	Piacenzian	Philippines
<i>Atlanta rosea</i> Gray, 1850	present-day	circumglobal tropic/subtropic
<i>Atlanta seapyi</i> Janssen, 2007	Piacenzian	Philippines
<i>Atlanta selvagensis</i> De Vera & Seapy, 2006	present-day	Atlantic tropic/subtropic
<i>Atlanta tokiokai</i> van der Spoel & Troost, 1972	Piacenzian - present-day	circumglobal tropic/subtropic
<i>Atlanta turriculata</i> d'Orbigny, 1836	Gelasian - present-day	Indopacific tropic/subtropic
<i>Atlanta vanderspoeli</i> Wall-Palmer et al., 2019	present-day	Pacific southern tropic/subtropic
<i>Atlantidea rotundata</i> (Gabb, 1873)	Langhian - Piacenzian	Caribbean, Fiji
<i>Mioatlanta soluta</i> Di Geronimo, 1974	Langhian	Italy
<i>Oxygyrus inflatus</i> Benson, 1835	Eemian-present-day	circumglobal tropic/subtropic
<i>Protatlanta kbiraensis</i> Janssen, 2012	Langhian	Malta
<i>Protatlanta sculpta</i> Issel, 1911	Piacenzian - present-day	Atlantic tropic/subtropic; fossils from Indian Ocean, Philippines
<i>Protatlanta souleyeti</i> (Smith, 1888)	Piacenzian - present-day	circumglobal tropic/subtropic

Family Pterotracheidae

<i>Firolloida desmarestia</i> Lesueur, 1817	Eemian - present-day	circumglobal tropic/subtropic
<i>Pterotrachea coronata</i> Forsskål in Niebuhr, 1775	present-day	tropic/subtropic Atlantic, Pacific?
<i>Pterotrachea hippocampus</i> Philippi, 1836	present-day	no data
<i>Pterotrachea keraudrenii</i> Gray, 1850	present-day	no data
<i>Pterotrachea scutata</i> Gegenbaur, 1855	present-day	no data

Family Carinariidae

<i>Cardiapoda placenta</i> (Lesson, 1830)	present-day	Indonesia (New Guinea)
<i>Cardiapoda richardi</i> Vayssière, 1904	present-day	Atlantic, Pacific
<i>Carinaria cithara</i> Benson, 1835	present-day	equatorial Indo-Pacific
<i>Carinaria cristata</i> (Linné, 1767)	present-day	Indo-Pacific
<i>Carinaria galea</i> Benson, 1835	present-day	Indo-Pacific
<i>Carinaria japonica</i> Okutani, 1955	present-day	NE Pacific
<i>Carinaria lamarcki</i> Péron & Lesueur, 1810	present-day	Atlantic-Pacific tropic/subtropic
<i>Carinaria maempeli</i> Janssen, 2012	Langhian	Mediterranean
<i>Carinaria mirabilis</i> Cossmann, 1902	Lutetian	France
<i>Carinaria pseudorugosa</i> Vayssière, 1904	present-day	N Atlantic
<i>Carinaria rutschi</i> Robba, 1972	Langhian	Mediterranean
<i>Pterosoma planum</i> Lesson, 1827	present-day	Indo-Pacific
<i>Striocarinaria hugardi</i> (Pictet, 1855)	Burdigalian	Mediterranean

Order ?**Superfamily Epitonioidea****Family Epitoniidae**

<i>Janthina chavani</i> (Ludbrook, 1978)	L Piacenzian - Calabrian	Atlantic, Pacific
<i>Janthina exigua</i> Lamarck, 1816	Holocene	circumglobal tropic/subtropic
<i>Janthina globosa</i> Swainson, 1822	Piacenzian - present-day	circumglobal tropic/subtropic
<i>Janthina janthina</i> (Linnaeus, 1758)	present-day	circumglobal tropic/subtropic
<i>Janthina krejci</i> Beu, 2017	Zanclean	Azores
<i>Janthina pallida</i> Thomson, 1840	present-day	(almost) circumglobal, tropic/subtrop
<i>Janthina typica</i> (Bronn, 1861)	Messinian - L Piacenzian	Atlantic, Pacific
<i>Janthina umbilicata</i> d'Orbigny, 1841	present-day	circumglobal tropic/subtropic
<i>Recluzia johnii</i> (Holten, 1802)	present-day	tropical Indo-West Pacific, Red Sea
<i>Recluzia lutea</i> (Bennett, 1840)	present-day	circumglobal tropic/subtropic

Subclass Heterobranchia**Infraclass Euthyneura****Subterclass Tectipleura****Order Pteropoda****Suborder Thecosomata****Subterorder Euthecosomata****Superfamily Limacinoidea****Family Heliconoididae**

<i>Heliconoides atypicus</i> (Laws, 1944)	Chattian - Aquitanian	New Zealand
<i>Heliconoides auriformis</i> (Curry, 1982)	ML Ypresian	SW France
<i>Heliconoides bartonensis</i> (Curry, 1965)	L Lutetian - Bartonian	W Europe, USA
<i>Heliconoides curryi</i> (Janssen, 1990)	Chattian	S Australia
<i>Heliconoides daguini</i> Janssen, 2010	ML Ypresian-Lutetian	SW France
<i>Heliconoides dilatata</i> (von Koenen, 1892)	Priabonian - E Rupelian	E Germany, Ukraine
<i>Heliconoides ferax</i> (Laws, 1944)	Burdigalian	New Zealand
<i>Heliconoides hodgkinsoni</i> Garvie, 2020	L Lutetian	U.S.A.
<i>Heliconoides hospes</i> (Rolle, 1862)	Rupelian - Chattian	North Sea Basin, Aquitaine Basin
<i>Heliconoides inflatus</i> (d'Orbigny, 1834)	Chattian - present-day	Circumglobal, tropic/subtropic
<i>Heliconoides lillebaeltensis</i> Janssen, 2007	L Lutetian	Denmark

<i>Heliconoides linneensis</i> Janssen, 2008	Chattian	Aquitaine Basin
<i>Heliconoides lunatus</i> (Janssen, 1990)	Chattian	Australia
<i>Heliconoides mercinensis</i> (Watelet & Lef., 1885)	Thanetian - Ypresian	W Europe, USA, Iran, Uzbekistan
<i>Heliconoides merlei</i> Janssen, 2010	ML Ypresian	SW France
<i>Heliconoides mermuysi</i> Janssen, 2010	Burdigalian	Aquitaine Basin
<i>Heliconoides nemoris</i> (Curry, 1965)	L Lutetian - Priabonian	North Sea Basin, Aquitaine Basin
<i>Heliconoides nikkieae</i> Janssen, 2017	Priabonian - Rupelian	Tanzania
<i>Heliconoides nitens</i> (Lea, 1833)	Lutetian - Bartonian	W Europe, U.S.A., Japan, Nigeria
<i>Heliconoides paula</i> (Curry, 1982)	ML Ypresian	Aquitaine Basin
<i>Heliconoides planus</i> (Tembrock, 1964)	Rupelian	Poland
<i>Heliconoides pyrenaicus</i> Janssen, 2010	ML Ypresian	Aquitaine Basin
<i>Heliconoides sondaari</i> Janssen, 2007	Piacenzian	Philippines, Fiji
<i>Heliconoides stenzeli</i> (Garvie, 1992)	Lutetian	USA
<i>Heliconoides tatei</i> (Janssen, 1990)	Langhian - Serravallian	Australia
<i>Heliconoides taylori</i> (Curry, 1965)	Ypresian	W Europe, USA?
<i>Heliconoides tertiaris</i> (Tate, 1887)	Langhian - Serravallian	Australia, Mediterranean
<i>Heliconoides texanus</i> (Garvie & Hodgkinson, 1992)	Lutetian	USA
<i>Heliconoides vanderweideni</i> Janssen, 2004	Chattian	Mediterranean
<i>Heliconoides vonhachti</i> Janssen, 2012	Zanclean	Mediterranean
<i>Heliconoides wardijaensis</i> Janssen, 2004	Chattian	Mediterranean
Family Limacinidae		
<i>Altaspiratella bearnensis</i> (Curry, 1982)	M-L Ypresian - Lutetian	Europe, USA, Tanzania, New Zealand?
<i>Altaspiratella choctavensis</i> (Aldrich, 1887)	L Ypresian	USA
<i>Altaspiratella elongatoidea</i> (Aldrich, 1887)	L Ypresian	USA
<i>Altaspiratella gracilens</i> Hodgkinson, 1992	L Ypresian - L Lutetian ?	USA, Iran
<i>Altaspiratella labiata</i> (Hodgkinson, 1992)	Bartonian	USA
<i>Altaspiratella multispira</i> (Curry, 1982)	M-L Ypresian	SW France, USA ?
<i>Altaspiratella tavianii</i> Janssen, 2013	L Ypresian - L Lutetian	Iran
<i>Currylimacina cossmanni</i> (Curry, 1982)	ML Ypresian - Lutetian	W Europe, USA
<i>Currylimacina asperita</i> Garvie, 2020	L Lutetian	USA
<i>Limacina acutimarginata</i> (Korobkov, 1966)	Rupelian	Russia, W Europe
<i>Limacina adornata</i> Hodgkinson, 1992	Bartonian	USA
<i>Limacina aegis</i> Hodgkinson, 1992	Ypresian	USA
<i>Limacina andrussowi</i> (Kittl, 1886)	Langhian	Mediterranean
<i>Limacina antarctica</i> Woodward, 1854	present-day	Antarctic
<i>Limacina aryanaensis</i> Janssen, 2013	L Ypresian - E Lutetian	Iran
<i>Limacina asiatica</i> Janssen, 2011	L Ypresian - E Lutetian	Uzbekistan
<i>Limacina atlanta</i> (Mörch, 1874)	Messinian - Piacenzian	W Europe, Mediterranean
<i>Limacina bulimoides</i> (d'Orbigny, 1834)	Burdigalian - present-day	circumglobal tropics/subtropics
<i>Limacina canadaensis</i> Hodgkinson, 1992	Bartonian - ? Rupelian	USA, Canada, Japan
<i>Limacina conica</i> (von Koenen, 1892)	Priabonian - Rupelian	North Sea Basin
<i>Limacina convolutus</i> Hodgkinson, 1992	Lutetian	Texas, U.S.A.
<i>Limacina davidi</i> Hodgkinson, 1992	E Ypresian	Canada
<i>Limacina dzheroiensis</i> Janssen, 2011	L Ypresian - E Lutetian	Uzbekistan
<i>Limacina erasmiana</i> Janssen, 2010	Ypresian	Kazakhstan, Uzbekistan, North Sea Basin
<i>Limacina ernstkittli</i> Janssen, 2012	Rupelian - Chattian	North Sea Basin, Malta
<i>Limacina exceptispira</i> (Korobkov, 1966)	Rupelian	Paratethys
<i>Limacina gormani</i> (Curry, 1982)	M-L Ypresian	Aquitaine Basin, North Sea Basin
<i>Limacina gramensis</i> (Rasmussen, 1968)	Langhian ? - Tortonian	North Sea Basin, Poland, Australia
<i>Limacina guersi</i> Janssen, 2010	Ypresian	North Sea Basin
<i>Limacina heatherae</i> Hodgkinson 1992	Eocene ?	U.S.A.
<i>Limacina helicina helicina</i> (Phipps, 1774)	present-day	Arctic
<i>Limacina helicina ochotensis</i> Shkoldina, 1999	present-day	NW Pacific

<i>Limacina helicina pacifica</i> Dall, 1871	present-day	N Pacific
<i>Limacina helicina rangii</i> (d'Orbigny, 1835)	present-day	Antarctic
<i>Limacina helikos</i> Hodgkinson, 1992	Ypresian	N America
<i>Limacina ingridae</i> Janssen, 1989	Serravallian	North Sea Basin
<i>Limacina irisae</i> Janssen, 1989	Serravallian	North Sea Basin
<i>Limacina karasawai</i> Ando, 2011	Priabonian - L Rupelian	Japan, USA
<i>Limacina konkensis</i> (Zhizhchenko, 1937)	Miocene	Paratethys
<i>Limacina lesueurii</i> (d'Orbigny, 1836)	Saalian - present-day	circumglobal tropic/subtropic
<i>Limacina lotschi</i> (Tembrock, 1989)	Priabonian - E Rupelian	North Sea Basin
<i>Limacina mariaae</i> Janssen, 1989	Priabonian - E Rupelian	North Sea Basin
<i>Limacina minima</i> (Zhizhchenko, 1937)	? Miocene	Paratethys
<i>Limacina novacaesarea</i> Janssen & Sessa, 2016	Ypresian	NE USA
<i>Limacina nucleata</i> (Zhizhchenko, 1934)	Langhian ?	Prearal, Russia
<i>Limacina parvabrazensis</i> Garvie & Janssen, 2020	L Lutetian	USA
<i>Limacina perforata</i> Janssen, 2013	L Ypresian - E Lutetian	Iran
<i>Limacina planorbella</i> (Korobkov, 1966)	Rupelian	Paratethys
<i>Limacina pseudopygmaea</i> Garvie & Janssen, 2019	L Lutetian	USA
<i>Limacina pseudoumbilicata</i> (Korobkov, 1966)	Rupelian	Paratethys
<i>Limacina pygmaea</i> (Lamarck, 1805)	ML Ypresian - Lutetian	W Europe
<i>Limacina retroversa retroversa</i> (Fleming, 1823)	Saalian - present-day	N Atlantic, Mediterranean
<i>Limacina retroversa australis</i> (Eydoux & Souleyet, 1840)	present-day	Australia
<i>Limacina robusta</i> (Eames, 1952)	Priabonian	India, Tanzania
<i>Limacina smithvillensis</i> Hodgkinson, 1992	Lutetian	USA
<i>Limacina subtarchanensis</i> (Zhizhchenko, 1936)	Miocene	Paratethys (Crimea)
<i>Limacina tanzaniaensis</i> Janssen, 2017	Priabonian	Tanzania
<i>Limacina tarchanensis</i> (Kittl, 1886)	Langhian	Ukraine, Paratethys
<i>Limacina texanopsis</i> Garvie, 2020	Lutetian	USA
<i>Limacina timi</i> Janssen, 2017	Priabonian - Rupelian	Tanzania
<i>Limacina trochiformis</i> (d'Orbigny, 1834)	Saalian - present-day	tropics/subtropics
<i>Limacina tschokrakensis</i> (Zhizhchenko, 1934)	Langhian ?	Prearal, Russia
<i>Limacina tutelina</i> (Curry, 1965)	Ypresian	North Sea Basin, USA
<i>Limacina ujiharai</i> Shibata, 1983	Miocene	Japan
<i>Limacina umbilicata</i> (Bornemann, 1855)	Rupelian	North Sea Basin, USA?
<i>Limacina valvatina</i> (Reuss, 1867)	Chattian - Langhian	Europe, ?Australia
<i>Limacina variospirata</i> (Korobkov, 1966)	Rupelian	Paratethys
<i>Limacina vegrandis</i> Janssen, 2010	ML Ypresian - Lutetian	W Europe
<i>Limacina voluta</i> Hodgkinson, 1992	ML ? Eocene	Canada
<i>Limacina wechesensis</i> Hodgkinson, 1992	Lutetian	USA
<i>Limacina wilhelminae</i> Janssen, 1989	Tortonian - Messinian	North Sea Basin, Spain
<i>Limacina yasdii</i> Janssen in Janssen et al., 2013	L Ypresian - E Lutetian	Iran
<i>Striolimacina andaensis</i> Janssen, 2007	Piacenzian	Philippines, Fiji
<i>Striolimacina imitans</i> (Collins, 1934)	Zanclean	USA, Mediterranean
Family Thieleidae		
<i>Thielea helicoides</i> (Jeffreys, 1877)	Tortonian - present-day	Mediterranean, Atlantic
Superfamily Cavolinioidea		
Family Creseidae		
<i>Boasia chierchiaie chierchiaie</i> (Boas, 1886)	present-day	tropic/subtropic, Indo-Pacific, patchy
<i>Boasia chierchiaie constricta</i> (Chen & Bé, 1964)	Langhian - present-day	Australia, Philippines, tropic/subtropic circumglobal
<i>Bovicornu eocenense</i> Meyer, 1886	Rupelian	USA
<i>Bovicornu gracile</i> Meyer, 1887	L Bartonian - Priabonian	USA
<i>Bowdenathea jamaicensis</i> Collins, 1934	Messinian - Piacenzian	Caribbean, Italy
<i>Bowdenathea miocenica</i> Janssen, 2004	Chattian	Malta

<i>Bucanoides basiannulata</i> Hodgkinson, 1992	Lutetian	USA
<i>Bucanoides divaricata</i> Hodgkinson, 1992	Bartonian	USA
<i>Bucanoides tenuis</i> Hodgkinson, 1992	Bartonian	USA
<i>Camptoceratops americanus</i> Garvie, 1992	Lutetian	USA
<i>Camptoceratops priscus</i> (Godwin-Austen, 1882)	ML Ypresian	W. Europe, USA
<i>Cheilospicata cedrus</i> Garvie, 2020	Lutetian	USA
<i>Cheilospicata repanda</i> Hodgkinson & Garvie, 1992	Bartonian	USA
<i>Creseis acicula</i> (Rang, 1828)	Zanclean - present-day	tropic/subtropic, circumglobal
<i>Creseis antoni</i> Janssen, 2010	Rupelian	SW France, Canada?
<i>Creseis berthae</i> Janssen, 1989 (Annelida?)	Rupelian	North Sea Basin
<i>Creseis conica</i> Eschscholtz, 1829	Eemian - present-day	tropic/subtropic, circumglobal
<i>Creseis corpulenta</i> (Meyer, 1887)	Bartonian, Priabonian	USA, SW France
<i>Creseis curta</i> Janssen, 2012	Langhian	Malta
<i>Creseis cylindrica</i> Hodgkinson, 1992	Bartonian	USA
<i>Creseis roesti</i> Janssen, 2010	L Aquitanian	Aquitaine Basin
<i>Creseis simplex</i> (Meyer, 1886)	M Lutetian - Priabonian	W Europe, USA
<i>Creseis spina</i> (Reuss, 1867)	Bartonian - Zanclean	W-S Europe, USA
<i>Creseis tugurii</i> Janssen, 2010	Burdigalian	Aquitaine Basin
<i>Creseis virgula</i> (Rang, 1828)	present-day	tropic/subtropic, circumglobal?
<i>Euchilotheca elegans</i> Harris, 1894	Ypresian - Lutetian	W Europe, Uzbekistan
<i>Euchilotheca ganensis</i> Curry, 1982	ML Ypresian	SW Europe
<i>Euchilotheca succincta</i> (Defrance, 1828)	Lutetian - ? Bartonian	W Europe, USA
<i>Loxibidens aduncus</i> Hodgkinson, 1992	Lutetian	USA
<i>Styliola schembriorum</i> Janssen, 2012	Langhian	Mediterranean
<i>Styliola subula</i> (Quoy & Gaimard, 1827)	Chattian - present-day	tropic/subtropic circumglobal
<i>Thecopsella fischeri</i> Cossmann, 1888	Lutetian	W Europe
<i>Tibiella annulata</i> Garvie, 1992	Lutetian	USA
<i>Tibiella marshi</i> Meyer, 1884	Bartonian	USA
<i>Tibiella reflexa</i> Hodgkinson, 1992	Bartonian	USA
<i>Tibiella texana</i> Collins, 1934	Lutetian	USA
<i>Tibiella watapuruensis</i> Janssen, 2013	E Bartonian	Indonesia
Family Hyalocylidae		
<i>Hyalocylis marginata</i> Janssen, 2007	Piacenzian	Philippines, Fiji?
<i>Hyalocylis striata</i> (Rang, 1828)	? Miocene - present-day	tropics, subtropics circumglobal
<i>Praehyalocylis maxima</i> (Ludwig, 1864)	Priabonian - Rupelian	North Sea Basin, France (Paris, Mayence, Aquitane)
Family Praecuvierinidae		
<i>Praecuvierina lura</i> (Hodgkinson, 1992)	Lutetian	USA, SW France
<i>Texacuvierina gutta</i> (Hodgkinson, 1992)	Bartonian	USA
<i>Texacuvierina hodgkinsoni</i> Janssen, 2013	L Ypresian - L Lutetian	Uzbekistan
Family Cuvierinidae		
<i>Cuvierina astesana</i> (Rang, 1829)	Zanclean	Mediterranean, Mexico?, Dominican Republic?
<i>Cuvierina atlantica</i> Bé et al, 1972	present-day	Atlantic, Caribbean
<i>Cuvierina cancapae</i> Janssen, 2005	present-day	C Atlantic, Caribbean
<i>Cuvierina columnella</i> (Rang, 1827)	present-day	Indo-Pacific Ocean
<i>Cuvierina curryi</i> Janssen, 2005	Langhian	Mediterranean
<i>Cuvierina grandis</i> D'Alessandro & Robba, 1981	Serravallian	Mediterranean
<i>Cuvierina inflata</i> (Bellardi, 1873)	Tortonian	Mediterranean
<i>Cuvierina intermedia</i> (Bellardi, 1873)	Serravallian	Portugal, Mediterranean, Fiji, ?Japan
<i>Cuvierina jagti</i> Janssen, 1995	Tortonian	Mediterranean
<i>Cuvierina ludbrookii</i> (Caprotti, 1962)	Piacenzian	Mediterranean
<i>Cuvierina pacifica</i> Janssen, 2005	present-day	Pacific Ocean
<i>Cuvierina paronai</i> Checchia-Rispoli, 1921	Langhian	Mediterranean, C Paratethys
<i>Cuvierina torpedo</i> (Marshall, 1918)	Aquitanian	New Zealand

<i>Cuvierina tsudai</i> BurrIDGE et al., 2016	present-day	Pacific Ocean
<i>Cuvierina urceolaris</i> (Mörch, 1850)	Piacenzian - present-day	tropic/subtropic Indo-Pacific
<i>Ireneia calandrellii</i> (Michelotti, 1847)	L Burdigalian	Mediterranean, New Zealand, Chili
<i>Ireneia gracilis</i> Janssen, 2005	Burdigalian - Langhian	Mediterranean
<i>Ireneia marqueti</i> Janssen, 1995	Tortonian - Messinian ?	North Sea Basin
<i>Ireneia nieulandei</i> Janssen, 1995	Burdigalian	Aquitaine Basin, New Zealand, Patagonia?
<i>Ireneia tenuistriata</i> (Semper, 1861)	Chattian	North Sea Basin
<i>Ireneia testudinaria</i> (Michelotti, 1847)	L Burdigalian	Mediterranean
<i>Johnjagtia baharensis</i> Janssen, 2012	Chattian - Aquitanian	Malta
<i>Johnjagtia moulinsi</i> (Benoist, 1873)	Burdigalian	Aquitaine Basin
<i>Spoelia torquayensis</i> Janssen, 1990	Chattian - Aquitanian	Australia, Mediterranean, Aquitaine Basin
Family Cliidae		
<i>Clio aichinoi</i> Checchia-Rispoli, 1921	Langhian	Italy
<i>Clio andreae</i> (Boas, 1886)	present-day	Atlantic, Pacific
<i>Clio antarctica</i> Dall, 1908	present-day	Antarctic
<i>Clio bellardii</i> Audenino, 1899	Langhian	Mediterranean, North Sea Basin?
<i>Clio berglundorum</i> Squires, 1989	Oligocene (Rupelian ?)	USA
<i>Clio bittneri</i> (Kittl, 1886)	Langhian	Paratethys
<i>Clio blinkae</i> Janssen, 1989	Rupelian	North Sea Basin
<i>Clio braidensis</i> (Bellardi, 1873)	Zanclean	Mediterranean
<i>Clio calix</i> (Bellardi, 1873)	Aquitanian ?	Italy
<i>Clio caralitana</i> Robba & Spano, 1978	Langhian	Sardinia
<i>Clio carinata</i> Audenino, 1899	Burdigalian	Mediterranean
<i>Clio chadumica</i> Korobkov, 1966	Rupelian	Russia, USA
<i>Clio chaptalii</i> Gray, 1850	present-day	tropics, subtropics circumglobal
<i>Clio coebana</i> Robba, 1972	Chattian - Aquitanian	Mediterranean
<i>Clio collina</i> Janssen & Zorn, 2001	Burdigalian - Langhian	Italy
<i>Clio convexa convexa</i> (Boas, 1886)	Piacenzian - present-day	Indo-Pacific
<i>Clio convexa cyphosa</i> Rampal, 2002	present-day	Red Sea
<i>Clio cuspidata</i> (Bosc, 1801)	Piacenzian - present-day	tropics, subtropics circumglobal
<i>Clio deflexa</i> von Koenen, 1882	Aquitanian	North Sea Basin
<i>Clio distefanoi</i> Checchia-Rispoli, 1921	Langhian	Mediterranean
<i>Clio fallauxi</i> (Kittl, 1886)	Langhian	C Paratethys
<i>Clio gailae</i> Goedert & Janssen, 2020	Chattian	USA
<i>Clio gargarica</i> Sirna, 1968	Langhian	Mediterranean
<i>Clio ghawdexensis</i> Janssen, 2004	Chattian	Malta
<i>Clio giulioi</i> Janssen, 1995	Tortonian	Mediterranean
<i>Clio goedertorum</i> Squires, 1989	E Miocene	USA
<i>Clio guidottii</i> Simonelli, 1896	Zanclean	Mediterranean
<i>Clio hataii</i> (Noda, 1972)	Zanclean	Japan, Philippines?
<i>Clio itoigawai</i> Shibata, 1983	E ? Miocene	Japan
<i>Clio jacobae</i> Janssen, 1989	Rupelian	North Sea Basin
<i>Clio lavayssei</i> Rutsch, 1934	Langhian ?	Trinidad
<i>Clio lozoueti</i> Janssen, 2010	Chattian	Aquitaine Basin
<i>Clio lucai</i> Janssen, 2000	Calabrian	Italy
<i>Clio merijni</i> Janssen, 2012	Chattian	Malta
<i>Clio multcostata</i> (Bellardi, 1873)	Serravallian	Mediterranean
<i>Clio nielsenii</i> Janssen, 1990	Chattian	North Sea Basin, SW France
<i>Clio nuda</i> Korobkov, 1966	L Eocene	Paratethys
<i>Clio oblonga</i> Rampal, 1996	L Pleistocene	Mediterranean
<i>Clio ortheziana</i> (Benoist, 1889)	Serravallian	Aquitaine Basin
<i>Clio pauli</i> Janssen, 1989	Langhian	North Sea Basin
<i>Clio pedemontana</i> (Mayer, 1868)	Aquitanian ?- Langhian	Mediterranean

<i>Clio piatkowskii</i> van der Spoel et al., 1992	present-day	Antarctic
<i>Clio polita</i> Pelseneer, 1888	present-day	Atlantic
<i>Clio pulcherrima</i> (Mayer, 1868)	Langhian	Mediterranean, Caribbean?
<i>Clio pyramidata angusta</i> (Boas, 1886)	present-day	N Atlantic
<i>Clio pyramidata pyramidata</i> Linné, 1767	Tortonian - present-day	tropical/subtropical circumglobal
<i>Clio pyramidata tyrrhenica</i> Janssen, 2012	L Pleistocene	Mediterranean
<i>Clio recurva</i> (Children, 1823)	present-day	tropical/subtropical circumglobal
<i>Clio ricciolii</i> (Calandrelli, 1844)	Piacenzian	Italy
<i>Clio saccoi</i> Checchia-Rispoli, 1921	Langhian	Mediterranean
<i>Clio scheelei</i> (Munthe, 1888)	present-day	South Africa (Cape Horn)
<i>Clio sinuosa</i> (Bellardi, 1873)	Serravallian	Italy
<i>Clio sturanii</i> Robba, 1977	Serravallian	Italy
<i>Clio sulcosa</i> (Bellardi, 1873)	Burdigalian - Langhian	Italy
<i>Clio superba</i> Fuchs, 1902	Oligocene ?	?Czech Republic, ?USA
<i>Clio triplicata</i> Audenino, 1899	Aquitanian - Langhian	Mediterranean, New Zealand
<i>Clio vasconiensis</i> Janssen, 2010	Chattian	Aquitaine Basin
<i>Clio vilis</i> Janssen, 2012	Chattian	Malta
<i>Clio yatsuoensis</i> Shibata, 1983	Miocene	Japan
Family Cavoliniidae		
Subfamily Cavoliniinae		
<i>Cavolinia baniensis</i> Janssen, 2007	Piacenzian	Philippines
<i>Cavolinia bituminata</i> Beets, 1953	Tortonian - Messinian	Indonesia
<i>Cavolinia cookei</i> Simonelli, 1895	Langhian	Mediterranean
<i>Cavolinia floridana</i> Collins, 1934	M-L Pliocene	USA, Mediterranean, ? Japan
<i>Cavolinia gatti</i> Janssen, 2012	Serravallian - Tortonian	Mediterranean
<i>Cavolinia gibbosa flava</i> (d'Orbigny, 1834)	present-day	Atlantic, Mediterranean, ?Pacific
<i>Cavolinia gibbosa gibboides</i> Rampal, 2002	present-day	E Mediterranean
<i>Cavolinia gibbosa gibbosa</i> (d'Orbigny, 1834)	present-day	Pacific, ?Atlantic
<i>Cavolinia gibbosa plana</i> (Meisenheimer, 1905)	present-day	Pacific
<i>Cavolinia globulosa</i> (Gray, 1850)	present-day	tropics/subtropics circumglobal
<i>Cavolinia grandis</i> (Bellardi, 1873)	Zanclean	Mediterranean, New Zealand
<i>Cavolinia gypsorum</i> (Bellardi, 1873)	Tortonian - Messinian	Mediterranean, Fiji
<i>Cavolinia inflexa imitans</i> (Pfeffer, 1880)	present-day	tropical/subtropical, circumglobal
<i>Cavolinia inflexa inflexa</i> (Lesueur, 1813)	present-day	tropical/subtropical, mainly Atlantic
<i>Cavolinia inflexa kakegawaensis</i> Shibata, 1984	Piacenzian ?	Japan
<i>Cavolinia inflexa labiata</i> (d'Orbigny, 1834)	present-day	Indo-Pacific
<i>Cavolinia inflexa robusta</i> Rampal, 2002	present-day	Mediterranean
<i>Cavolinia landaui</i> Janssen, 2004	Piacenzian	Spain, Philippines
<i>Cavolinia longicostata</i> Rampal, 2002	present-day	NW Mediterranean
<i>Cavolinia marginata hyugaensis</i> Ujihara, 1996	Zanclean	Japan, Philippines
<i>Cavolinia marginata limatula</i> Beets, 1943	Tortonian/Messinian	Indonesia
<i>Cavolinia marginata marginata</i> (Bronn, 1862)	Zanclean	Azores
<i>Cavolinia marginata pliomediterranea</i> Janssen, 2004	Piacenzian	Spain
<i>Cavolinia marginata vendryesiana</i> (Guppy, 1873)	Piacenzian	Jamaica
<i>Cavolinia mexicana</i> (Collins, 1934)	Tortonian ? - Zanclean	Caribbean, Japan, Fiji, Indonesia
<i>Cavolinia microbesitas</i> Janssen, 2012	Langhian - L Burdigalian	Italy, Malta
<i>Cavolinia pachysoma</i> Rampal, 2002	present-day	NW Mediterranean
<i>Cavolinia perparvula</i> Janssen, 2007	Piacenzian	Philippines
<i>Cavolinia pycna</i> Jung, 1971	Langhian	Caribbean, Mediterranean
<i>Cavolinia shibatai</i> Janssen, 2007	Piacenzian	Spain, Philippines
<i>Cavolinia tridentata</i> (Niebuhr, 1775)	Piacenzian - present-day	tropical/subtropical circumglobal
<i>Cavolinia uncinata</i> (d'Orbigny, 1835)	present-day	tropical/subtropical circumglobal
<i>Cavolinia ventricosa</i> (Guppy, 1882)	Piacenzian	Caribbean, Philippines

<i>Cavolinia yamabensis</i> Shibata, 1983	Aquitanian - Burdigalian	Japan
<i>Cavolinia zamboninii</i> Checchia-Rispoli, 1921	Langhian	Mediterranean
<i>Diacavolinia angulata</i> (Souleyet, 1852)	present-day	tropics, Indian Ocean
<i>Diacavolinia angulosa</i> (Gray, 1850)	present-day	Indian Ocean
<i>Diacavolinia aspina</i> van der Spoel et al., 1993	present-day	tropical Atlantic
<i>Diacavolinia atlantica</i> van der Spoel et al., 1993	present-day	Indonesia
<i>Diacavolinia bandaensis</i> van der Spoel et al., 1993	present-day	Indonesia
<i>Diacavolinia bicornis</i> van der Spoel et al., 1993	present-day	tropical W Atlantic
<i>Diacavolinia constricta</i> van der Spoel et al., 1993	present-day	tropical Atlantic
<i>Diacavolinia deblainvillei</i> van der Spoel et al., 1993	present-day	tropical W Atlantic
<i>Diacavolinia deshayesi</i> van der Spoel et al., 1993	present-day	tropical/subtropical W Atlantic
<i>Diacavolinia elegans</i> van der Spoel et al., 1993	present-day	N Atlantic, C Pacific
<i>Diacavolinia flexipes</i> van der Spoel et al., 1993	present-day	tropical Atlantic
<i>Diacavolinia grayi</i> van der Spoel et al., 1993	present-day	tropical Atlantic
<i>Diacavolinia limbata</i> (d'Orbigny, 1836)	present-day	tropical Atlantic
<i>Diacavolinia longirostris</i> (de Blainville, 1821)	present-day	tropics, Atlantic, Indonesia
<i>Diacavolinia mcgowani</i> van der Spoel et al., 1993	present-day	Caribbean
<i>Diacavolinia ovalis</i> van der Spoel et al., 1993	present-day	N Atlantic
<i>Diacavolinia pacifica</i> van der Spoel et al., 1993	present-day	Indo-Pacific
<i>Diacavolinia pristina</i> Janssen, 2007	Piacenzian	Philippines
<i>Diacavolinia robusta</i> van der Spoel et al., 1993	present-day	central W Atlantic
<i>Diacavolinia souleyeti</i> van der Spoel et al., 1993	present-day	Indian Ocean
<i>Diacavolinia strangulata</i> (Deshayes, 1823)	present-day	central W Atlantic
<i>Diacavolinia striata</i> van der Spoel et al., 1993	present-day	Indian Ocean
<i>Diacavolinia triangulata</i> van der Spoel et al., 1993	present-day	Indian Ocean
<i>Diacavolinia vanutrechtii</i> van der Spoel et al., 1993	present-day	Pacific
<i>Gamopleura maxwelli</i> Grebneff & Janssen, 2011	Chattian	New Zealand
<i>Gamopleura melitensis</i> Janssen, 1995	Chattian - ?Burdigalian	Malta
<i>Gamopleura pilula</i> Janssen, 2012	Burdigalian	Malta
<i>Gamopleura taurinensis</i> (Michelotti, 1847)	Burdigalian?	Mediterranean, Aquitaine Basin
Subfamily Diacriinae		
<i>Diacria digitata</i> (Guppy, 1882)	Piacenzian	Jamaica
<i>Diacria italica fissicostata</i> Janssen, 2007	Piacenzian	Philippines, Italy
<i>Diacria italica italica</i> Grecchi, 1982	Piacenzian	Mediterranean
<i>Diacria major</i> (Boas, 1886)	present-day	Atlantic, Pacific
<i>Diacria mbaensis</i> Ladd, 1934	Tortonian - Messinian	Fiji, Indonesia
<i>Diacria microstriata</i> Janssen, 2007	Piacenzian	Philippines, Fiji
<i>Diacria paeninsula</i> Janssen, 2007	Piacenzian	Philippines
<i>Diacria piccola</i> Bleeker & van der Spoel, 1988	present-day	Philippines
<i>Diacria rampalae</i> Dupont, 1979	present-day	Caribbean
<i>Diacria sangiorgii</i> Scarsella, 1934	Tortonian	Mediterranean
<i>Diacria trispinosa</i> (de Blainville, 1821)	Messinian - present-day	Circumglobal tropic/subtropic
<i>Diacrolinia aquensis</i> (Grateloup, 1827)	Burdigalian	France
<i>Diacrolinia aurita</i> (Bellardi, 1873)	Langhian	N. Sea Basin, C. Paratethys, Mediterranean
<i>Diacrolinia cluzaudi</i> Janssen, 2010	L Aquitanian	Aquitaine Basin
<i>Diacrolinia elioi</i> Janssen, 1995	Tortonian	Mediterranean
<i>Diacrolinia interrupta</i> (Bellardi, 1873)	Burdigalian	Mediterranean
<i>Diacrolinia larandaensis</i> Janssen, 1999	Serravallian	Turkey, N. Italy
<i>Diacrolinia orbignyi</i> (Rang, 1827)	Burdigalian	Aquitaine Basin
<i>Diacrolinia pumilionis</i> Janssen, 2012	Langhian	Malta
<i>Diacrolinia revoluta</i> (Bellardi, 1873)	Burdigalian	Mediterranean
<i>Telodiacria costata</i> (Pfeffer, 1879)	present-day	Pacific
<i>Telodiacria danae</i> (van Leyen & van der Spoel, 1982)	present-day	circum tropical/subtropical

<i>Telodiacria erythra</i> (van Leyen & van der Spoel, 1982)	present-day	Red Sea
<i>Telodiacria philippinensis</i> (Janssen, 2007)	Piacenzian	Philippines, Fiji
<i>Telodiacria quadridentata</i> (de Blainville, 1821)	present-day	C Atlantic
<i>Telodiacria schmidti</i> (van Leyen & v.d. Spoel, 1982)	present-day	C + S Pacific
Subfamily Vaginellinae n. subfam.		
<i>Edithinella bonaviai</i> Janssen, 2004	Langhian	Malta
<i>Edithinella caribbeana</i> (Collins, 1934)	L Serravallian	Panama, Mediterranean, Aquitaine Basin
<i>Edithinella curva</i> Janssen, 1998	Langhian	Malta
<i>Edithinella doliarius</i> Janssen, 2006	Langhian	Malta
<i>Edithinella katoi</i> (Shibata, 1983)	Miocene	Japan
<i>Edithinella varanica</i> (Sirna, 1968)	Langhian	Mediterranean, Aquitaine Basin
<i>Vaginella acutissima</i> Audenino, 1899	L Langhian	Mediterranean
<i>Vaginella austriaca</i> Kittl, 1886	L Burdigalian - Langhian	Paratethys, North Sea Basin, Aquitaine Basin
<i>Vaginella basitruncata</i> Janssen, 2005	Chattian	North Sea Basin
<i>Vaginella bicarinata</i> Tate, 1887	Langhian	Australia
<i>Vaginella chattica</i> R. Janssen, 1979	Chattian	North Sea Basin, Aquitaine Basin, Mediterranean
<i>Vaginella chipolana</i> Dall, 1893	Burdigalian ?	Caribbean
<i>Vaginella depressa</i> Daudin, 1800	Aquitanian	Europe, Caribbean, Japan, Australia, New Zealand
<i>Vaginella floridana</i> Collins, 1934	Aquitanian/Burdigalian	USA
<i>Vaginella gaasensis</i> Janssen, 2010	Rupelian	France
<i>Vaginella gibbosa</i> Audenino, 1899	Langhian	Mediterranean
<i>Vaginella lapugyensis</i> Kittl, 1886	Langhian	W Europe, C. Paratethys, Mediterranean, Australia?
<i>Vaginella sannicola</i> Janssen, 1990	Langhian	Mediterranean
<i>Vaginella tricuspudata</i> Zorn & Janssen, 1993	Chattian	Aquitaine Basin, C. Paratethys, Mediterranean
<i>Vaginella venezuelana</i> Collins, 1934	Chattian - Aquitanian	Malta, Venezuela
<i>Vaginella victoriae</i> Janssen, 1990	Burdigalian	Australia, Aquitaine Basin, Mediterranean
Family Sphaerocinidae		
<i>Hameconia edmundi</i> Janssen, 2008	Chattian	Aquitaine Basin
<i>Sphaerocina convolvula</i> Janssen, 2007	Piacenzian	Philippines, Fiji
<i>Sphaerocina formai</i> (Audenino, 1899)	Langhian	Mediterranean
Suborder Pseudothecosomata		
Family Cymbuliidae		
<i>Corolla calceola</i> (Verrill, 1880)	present-day	Atlantic, W Pacific?
<i>Corolla cupula</i> Rampal, 1996	present-day	S Atlantic
<i>Corolla intermedia</i> (Tesch, 1903)	present-day	subtropical Atlantic, E Indian Ocean
<i>Corolla ovata</i> (Quoy & Gaimard, 1833)	present-day	tropical Atlantic, Pacific
<i>Corolla spectabilis</i> Dall, 1871	present-day	tropics, W Pacific, E Atlantic?
<i>Cymbulia parvidentata</i> Pelseneer, 1888	present-day	circumglobal
<i>Cymbulia peronii</i> de Blainville, 1818	present-day	Atlantic
<i>Cymbulia sibogae</i> Tesch, 1903	present-day	tropical Atlantic/Indo-Pacific
<i>Cymbulia tricavernosa</i> Zhang 1964	present-day	China
<i>Gleba chrysosticta</i> (Troschel, 1854)	present-day	S Atlantic, E Indian Ocean
<i>Gleba cordata</i> Forsskål in Niebuhr, 1776	Eemian - present-day	global, tropic/subtropic
Family Desmopteridae		
<i>Desmopterus gardineri</i> Tesch, 1910	present-day	Indian Ocean
<i>Desmopterus pacificus</i> Essenberg, 1919	present-day	NE Pacific
<i>Desmopterus papilio</i> Chun, 1889	present-day	tropics, subtropics circumglobal
Family Peraclidae		
<i>Peraclе amberae</i> Janssen 2012	Chattian	Malta
<i>Peraclе bispinosa</i> (Pelseneer, 1888)	Piacenzian - present-day	Atlantic
<i>Peraclе charlotteae</i> Janssen & Little, 2010	Langhian	Cyprus
<i>Peraclе depressa</i> Meisenheimer, 1906	present-day	Atlantic
<i>Peraclе diversa</i> (Monterosato, 1875)	present-day	Atlantic, Mediterranean

<i>Peracle elata</i> (Seguenza, 1875)	present-day	Atlantic
<i>Peracle grebneffi</i> Janssen, 2012	Langhian	Mediterranean
<i>Peracle lata</i> (Krach, 1981)	Langhian	Poland
<i>Peracle michaelisarsii</i> (Bonnevie, 1913)	present-day	N Atlantic
<i>Peracle moluccensis</i> (Tesch, 1903)	present-day	tropical Indo-Pacific
<i>Peracle philiporum</i> (Gilmer, 1990)	present-day	Bahamas
<i>Peracle reticulata</i> (d'Orbigny, 1834)	Zanclean - present-day	tropics, subtropics circumglobal
<i>Peracle rissoides</i> Tesch, 1903	present-day	Indonesia
<i>Peracle valdiviae</i> (Meisenheimer, 1905)	present-day	Indian Ocean
Suborder Gymnosomata		
Superfamily Clionoidea		
Family Cliopsidae		
<i>Cliopsis krohni</i> Troschel, 1854	present-day	tropics, subtropical circumglobal
<i>Pruvotella danae</i> Pruvot-Fol, 1942	present-day	N Atlantic, S Indo-Pacific
<i>Pruvotella pellucida</i> (Quoy & Gaimard, 1832)	present-day	Indian Ocean, Indonesia
Family Clionidae		
<i>Cephalobranchia bonnevii</i> Massy, 1917	present-day	N Atlantic
<i>Cephalobranchia macrochaeta</i> Bonnevie, 1913	present-day	N Atlantic, trop. Indo-Pacific
<i>Clione elegantissima</i> Dall, 1871	present-day	NW Pacific
<i>Clione limacina antarctica</i> Smith, 1902	present-day	Antarctic
<i>Clione limacina limacina</i> (Phipps, 1774)	present-day	northern oceans
<i>Clione okhotensis</i> Yamazaki & Kumahara, 2017	present-day	Okhotsk Sea, Japan
<i>Clione? ignota</i> Janssen, 2012	Chattian - Langhian	Mediterranean
<i>Clione? imdinaensis</i> Janssen, 2012	Chattian - Langhian	Mediterranean
<i>Clione? phosphorita</i> Janssen, 2012	Chattian - Langhian	Mediterranean
<i>Clione? tripartita</i> Janssen, 2012	Langhian	Mediterranean
<i>Clione? tumidula</i> Janssen, 2012	Chattian - Langhian	Mediterranean
<i>Fowlerina punctata</i> (Tesch, 1903)	present-day	central N Atlantic, Indonesia
<i>Fowlerina zetesios</i> Pelseneer, 1906	present-day	NE Atlantic
<i>Massya longecirrata</i> (Massy, 1917)	present-day	NE Atlantic
<i>Paedoclione doliiformis</i> Danforth, 1907	present-day	NW Atlantic
<i>Paraclyone flavescens</i> (Gegenbaur, 1855)	present-day	C Mediterranean
<i>Paraclyone longicaudata</i> (Souleyet, 1852)	present-day	Tropical/subtropical Atlantic, Indo-West Pacific
<i>Paraclyone pelseneeri</i> Tesch, 1903	present-day	Indonesia
<i>Thalassopterus zancleus</i> Kwietniewski, 1910	present-day	Mediterranean, W Indian Ocean
<i>Thliptodon akatukai</i> Tokioka, 1950	present-day	Japan
<i>Thliptodon antarcticus</i> Meisenheimer, 1906	present-day	Antarctic
<i>Thliptodon diaphanus</i> (Meisenheimer, 1902)	present-day	Tropical/subtropical Atlantic + Pacific
<i>Thliptodon gegenbauri</i> Boas, 1886	present-day	S Atlantic, SE Pacific
<i>Thliptodon schmidti</i> Pruvot-Fol, 1942	present-day	Indonesia
Family Notobranchaeidae		
<i>Notobranchaea bleekerae</i> van der Spoel & Pafort, 1985	present-day	N Atlantic, N Indian Ocean
<i>Notobranchaea grandis</i> Pruvot-Fol, 1942	present-day	Indian Ocean, central E Pacific
<i>Notobranchaea hjorti</i> Bonnevie, 1913	present-day	N Atlantic
<i>Notobranchaea inopinata</i> Pelseneer, 1887	present-day	W Pacific
<i>Notobranchaea longicollis</i> (Bonnevie, 1913)	present-day	N Atlantic
<i>Notobranchaea macdonaldi</i> Pelseneer, 1886	present-day	tropics, subtropics circumglobal
<i>Notobranchaea tetrabranchiata</i> (Bonnevie, 1913)	present-day	Atlantic, SW Pacific
<i>Notobranchaea valdiviae</i> Meisenheimer, 1905	present-day	Indian Ocean
Family Pneumodermatidae		
<i>Abranchaea chinensis</i> Zhang, 1964	present-day	China Sea
<i>Platybrachium antarcticum</i> Minichev, 1976	present-day	Antarctic Ocean
<i>Pneumoderma degraaffi</i> van der Spoel & Pafort, 1982	present-day	N Atlantic

<i>Pneumoderma heronense</i> Newman & v.d. Spoel, 1989	present-day	Australia, Heron Reef
<i>Pneumoderma mediterraneum</i> (Van Beneden, 1838)	present-day	Central Atlantic, central Indo-Pacific
<i>Pneumoderma meisenheimeri</i> Pruvot-Fol, 1926	present-day	Indonesia
<i>Pneumoderma pacificum</i> (Dall, 1871)	present-day	tropics/subtropics, Indo-Pacific
<i>Pneumoderma peroni</i> (Cuvier, 1817)	present-day	N Atlantic, tropical W Pacific
<i>Pneumoderma violaceum</i> d'Orbigny, 1835	present-day	tropics/subtropics, Atlantic, Pacific?
<i>Pneumodermopsis brachialis</i> Minichev, 1976	present-day	Antarctic Ocean
<i>Pneumodermopsis canephora</i> Pruvot-Fol, 1924	present-day	tropical/subtropical E Atlantic
<i>Pneumodermopsis ciliata</i> (Gegenbaur, 1855)	present-day	N Atlantic, Indo-Pacific
<i>Pneumodermopsis macrochira</i> Meisenheimer, 1905	present-day	circumglobal
<i>Pneumodermopsis macrocotyla</i> Zhang, 1964	present-day	NW Pacific
<i>Pneumodermopsis michaelsarsi</i> Bonnevie, 1913	present-day	N Atlantic
<i>Pneumodermopsis minuta</i> (Pelseener, 1887)	present-day	N Pacific
<i>Pneumodermopsis paucidens</i> (Boas, 1886)	present-day	N Atlantic, scattered southern oceans
<i>Pneumodermopsis polycotyla</i> (Boas, 1886)	present-day	NE Atlantic
<i>Pneumodermopsis pupula</i> Pruvot-Fol, 1926	present-day	NE Atlantic
<i>Pneumodermopsis simplex</i> (Boas, 1886)	present-day	Mediterranean
<i>Pneumodermopsis spoeli</i> Newman & Greenwood, 1988	present-day	no data
<i>Pneumodermopsis teschi</i> van der Spoel, 1973	present-day	NE Atlantic
<i>Schizobranchium polycotylum</i> Meisenheimer, 1903	present-day	all oceans
<i>Spongiobranchaea australis</i> d'Orbigny 1836	present-day	Antarctic oceans
<i>Spongiobranchaea intermedia</i> Pruvot-Fol, 1926	present-day	scattered all oceans
Superfamily Hydromyloidea		
Family Hydromylidae		
<i>Hydromyles globulosus</i> (Rang, 1825)	present-day	?Atlantic, ?Indo-Pacific
Family Laginiopsidae		
<i>Laginiopsis triloba</i> Pruvot-Fol, 1922	present-day	Indo-Pacific
Subterclass Ringipleura		
Order Nudibranchia		
Suborder Cladobranchia		
Superfamily Aeolidioidea		
Family Glaucidae		
<i>Glaucilla bennettiae</i> (Churchill et al., 2014)	present-day	S Pacific subtropical gyre system
<i>Glaucilla mcfarlanei</i> (Churchill et al., 2014)	present-day	N Pacific subtropical gyre system
<i>Glaucilla marginata</i> Reinhardt & Bergh, 1864	present-day	Indo-Pacific tropic/subtropic
<i>Glaucilla thompsoni</i> (Churchill et al., 2014)	present-day	N Pacific subtropical gyre system
<i>Glaucus atlanticus</i> Forster, 1777	present-day	Atlantic, Pacific tropic/subtropic
Superfamily (unassigned)		
Family Phylliroidea		
<i>Cephalopyge trematoides</i> Chun, 1889	present-day	circumglobal tropic/subtropic
<i>Phylliroe bucephala</i> Lamarck, 1816	present-day	circumglobal
<i>Phylliroe lichtensteinii</i> Eschscholtz, 1825	present-day	circumglobal tropic/subtropic
Superfamily Fionoidea		
Family Fionidae		
<i>Fiona pinnata</i> (Eschscholtz, 1831)	present-day	circumglobal
Mesozoic gastropods considered to be pelagic species		
Family Bellerophonidae		
<i>Bellerophina minuta</i> (Sowerby, 1814)	Albian	UK
<i>Frebaldia fluitans</i> Nützel & Schneider, 2016	Jurassic	Canada
<i>Frebaldia carinii</i> Pieroni & Nützel, 2020	Triassic	Italy

Family Carinariidae

Brunonia annulata (Yokoyama, 1890) Cretaceous Japan, Tethys ?

Family Pterotracheidae

Pterotrachea liassica Bandel & Hemleben, 1987 Jurassic Germany

Family Coelodiscidae

Coelodiscus minutus (von Zieten, 1830) Jurassic Germany

Coelodiscus fluegeli Bandel & Hemleben, 1987 Jurassic Germany

Tatediscus aratus (Tate, 1870) Jurassic UK, Germany, Argentina

Family ?

Costasphaera franconica Gründel & Nützel, 2015 Jurassic Germany

Globorilusopsis arcuatus Maubeuge, 1998 Jurassic Luxembourg

Globorilusopsis simoni Maubeuge, 1994 Jurassic Luxembourg

Globorilusopsis simoniiformis Maubeuge, 1998 Jurassic Luxembourg

Globorilusopsis turbinatus Maubeuge, 1998 Jurassic Luxembourg

Globorilusopsis erectusiformis Maubeuge, 1998 Jurassic Luxembourg

Globorilusopsis compactus Maubeuge, 1998 Jurassic Luxembourg

Globorilusopsis obesus Maubeuge, 1998 Jurassic Luxembourg

Globorilusopsis baculatus Maubeuge, 1998 Jurassic Luxembourg

Globorilusopsis gracilis Maubeuge, 1994 Jurassic Luxembourg

Globorilusopsis elegans Maubeuge, 1997 Jurassic Luxembourg

Globorilusopsis resurgens Maubeuge, 1998 Jurassic Luxembourg

Globorilusopsis inguis Maubeuge, 1998 Jurassic Luxembourg

Globorilusopsis regressum Maubeuge, 1998 Jurassic Luxembourg

Simonicerias commotum Maubeuge, 1998 Jurassic Luxembourg

Simonicerias concinnaticium Maubeuge, 1998 Jurassic Luxembourg

Simonicerias cornu Maubeuge, 1998 Jurassic Luxembourg

Simonicerias cornuaammoni Maubeuge, 1998 Jurassic Luxembourg

Simonicerias curvatum Maubeuge, 1998 Jurassic Luxembourg

Simonicerias delsatei Maubeuge, 1998 Jurassic Luxembourg

Simonicerias erectum Maubeuge, 1998 Jurassic Luxembourg

Simonicerias incurvatum Maubeuge, 1998 Jurassic Luxembourg

Simonicerias mirum Maubeuge, 1998 Jurassic Luxembourg

Simonicerias orbiculatum Maubeuge, 1998 Jurassic Luxembourg

Simonicerias spiratissimum Maubeuge, 1998 Jurassic Luxembourg

Simonicerias stompi Maubeuge, 1998 Jurassic Luxembourg

APPENDIX 4.

Diagnosis for Vaginellinae Janssen subfam. nov.

(AWJ) (Fig. 17)

Contrary to Cavoliniinae and Diacriinae the shell is not globose but more or less elongately triangular, dorsal and ventral shell parts equally convex, separated by lateral carinae at least in the apical part of the shell, no lateral slits; protoconch straight or obliquely positioned with a single subapical swelling, persisting (although usually broken) in

most, but shed in two species (*V. basitruncata*, *V. floridana*) and then with a basal septum; dorsal apertural margin somewhat higher than ventral one, simple, slightly flaring, with weak radial folds or with cusps. Shell with post-larval metamorphosis during which the flexible early teleoconch with a wide apical angle and small dorso-ventral diameter changes into the adult shell shape with a smaller apical angle and a wider dorso-ventral diameter, a process similar as seen in other Cavoliniidae.

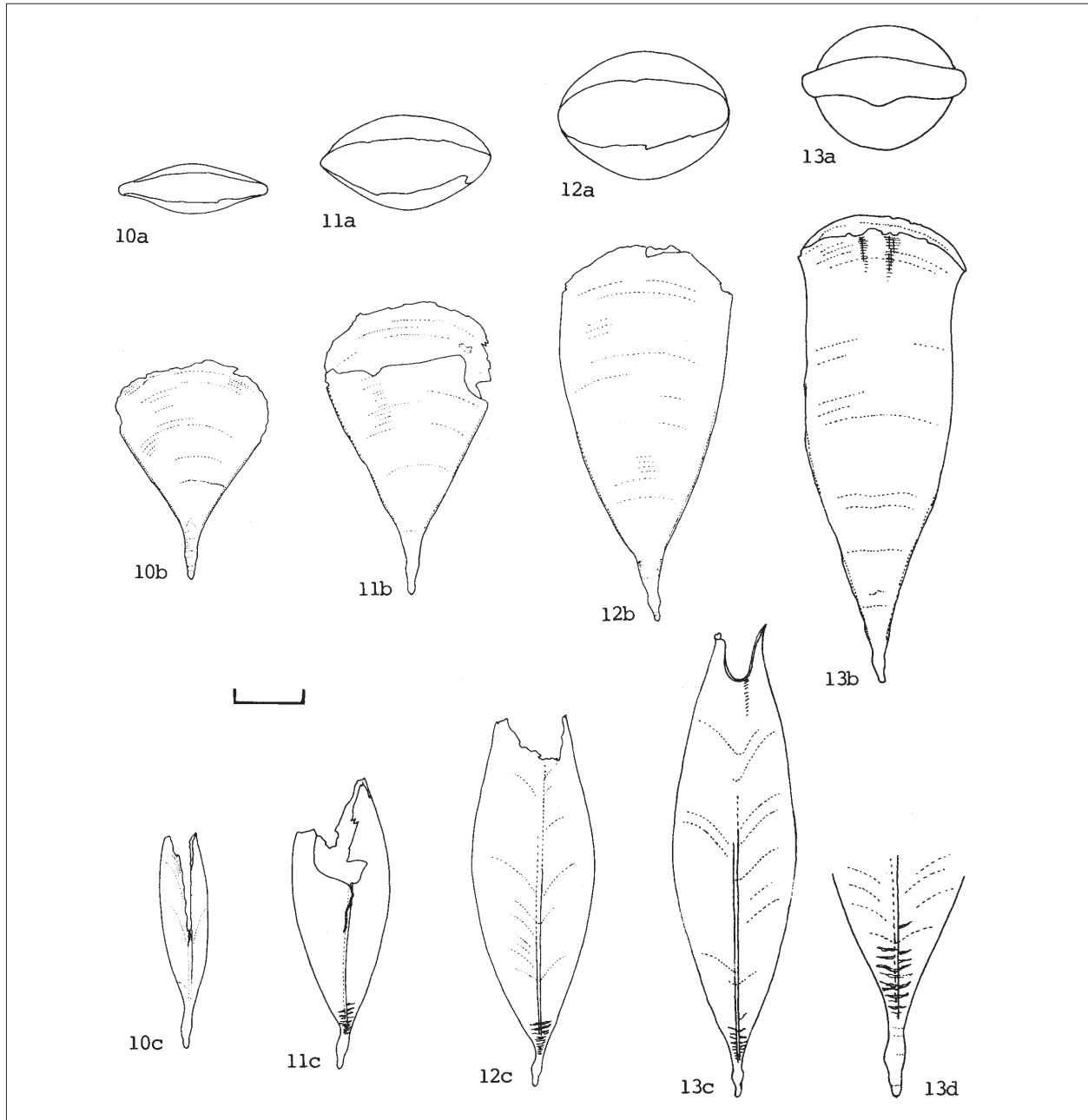


Fig. 17. Ontogenetic development of *Vaginella depressa* Daudin, 1800. Specimens from the Miocene (Burdigalian, Falun de Léognan) of Pas-de-Barreau, Martillac (Gironde, France); from Janssen, 1985: 202-203, figs 10-13. 10-12: successive stadia of juvenile shell, RGM.227565-7. 13: adult specimen, RGM.227568; a – apertural, b – ventral, c – left lateral views; 13d – protoconch, left lateral view. Bar = 1 mm, 13d magnified from 13c.