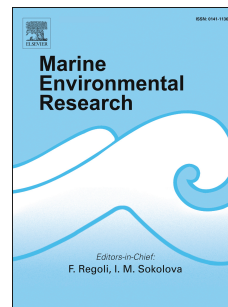


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Marine fouling invasions in ports of Patagonia (Argentina) with implications for legislation and monitoring programs

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1  
2 **Marine fouling invasions in ports of Patagonia (Argentina) with implications**  
3 **for legislation and monitoring programs**  
4

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ACCEPTED MANUSCRIPT

36 **Abstract**

37 Ports are a key factor in the understanding and solving of most problems  
38 associated with marine invasive species across regional and global scales. Yet  
39 many regions with active ports remain understudied. The aim of this work was to  
40 (a) identify and quantify the marine fouling organisms in all Patagonian ports of  
41 Argentina classifying them as native, exotic or cryptogenic species through a rapid  
42 assessment survey and experimental studies, (b) survey the environmental and  
43 anthropogenic variables of these ports and (c) analyze and discuss these results in  
44 the light of the South America context for the study of marine invasive species,  
45 legislation and commerce. We found 247 fouling species, including 17 introduced,  
46 one of which is a new record for the region, and other 15 species currently  
47 considered cryptogenic species that will need further attention to clarify their status.  
48 The analysis of mobile and sessile taxa, together with the environmental variables  
49 measured in this study and the port movement, allow us to discuss individual ports'  
50 vulnerability to future introductions. This is the first large scale study performed for  
51 this region on this topic, and it will help in developing monitoring programs and  
52 early detection plans to minimize new species introductions along the marine  
53 coastline of southern South America.

54

55 **Keywords:** Marine exotic species; Fouling; Ports; Southwestern Atlantic

56

## 57 1. Introduction

58 The introduction of invasive species is recognized as one of the top five  
59 threats to native biodiversity (Sala et al., 2000). An overwhelming number of  
60 species are transported worldwide every day by several means, and our  
61 understanding of their evolutionary history constantly reveals unexpected  
62 complexities (e.g. Geller, 1999; Fortune et al., 2008). Since ocean shipping is  
63 considered the most important vector for transporting and introducing species into  
64 new areas outside their native ranges (Ruiz and Carlton, 2003; Drake and Lodge,  
65 2007), the monitoring of ports and harbors helps us to predict the vulnerability of  
66 local harbors and to develop regional management policies (Bishop and Hutchings,  
67 2011). Indeed, harbors' vulnerability is extremely difficult to predict due to the  
68 complexity presented by variables such as propagule pressure (Johnston et al.,  
69 2009), resource availability (Olyarnik et al., 2009), diversity of resident species and  
70 environmental conditions of the receptive habitat (Byers, 2002). Within this context,  
71 it is clear necessity to create accurate baseline information about these  
72 environmental conditions (Bishop and Hutchings, 2011; Mead et al., 2011).

73 Port areas concentrate a variety of artificial structures that support many  
74 different organisms (Glasby, 1999; Connell, 2001), and it is known that artificial and  
75 natural habitats are not equally colonized by fouling species (Connell, 2001). In  
76 fact, man-made structures seem to favor the recruitment and survival of fouling  
77 exotic species even when the richness of native species is relatively high (Glasby  
78 et al., 2007). Indeed, man-made habitats might even act as corridors enhancing  
79 the spreading of exotic marine species, as shown by Bulleri and Airoidi (2005) for  
80 the invasive *Codium fragile* subsp. *tomentosoides*. Considering that the 90% of the  
81 global trade is carried by sea, our understanding of global marine invasion ecology  
82 is strongly related to the effort we dedicate to study port areas.

83 The Southwestern Atlantic (SWA) is currently placing a considerable effort to  
84 compile all the records of marine exotic and cryptogenic species (e.g., Orensanz et  
85 al., 2002; Scarabino, 2006; Schwindt, 2008). However, the lack of tradition in  
86 integrating coastal ecology and the regional maritime history hampers our ability to  
87 understand biological invasion patterns in this region (Bortolus and Schwindt,

88 2007). The earliest fouling studies in warm temperate Argentinean ports date from  
89 the 1960's (Bastida, 1971; Valentinuzzi de Santos, 1971), and since then, most  
90 cold temperate ports within this region have never been intensively surveyed and  
91 their biodiversity remains largely unknown. Argentina has the second longest  
92 shoreline of the SWA, after Brazil. However, in contrast with the heavily populated  
93 and industrialized coast of Brazil, Argentina has only ten major marine ports along  
94 a mostly exposed shoreline with a few marinas associated with recreational  
95 activities (Boltovskoy, 2008). Thus, the aim of this work was (a) to identify and  
96 quantify the marine fouling organisms in all Patagonian ports of Argentina by  
97 conducting a Rapid Assessment Survey (hereafter RAS) and experimental studies,  
98 and classifying them as native, exotic or cryptogenic species (b) to survey/describe  
99 the environmental and anthropogenic variables of these ports and (c) to analyze  
100 and discuss these results in the light of the South America context on marine  
101 invasion ecology, legislation and commerce. This is the first large scale study  
102 performed for this region on this topic, and it will help in developing monitoring  
103 programs and early detection plans to minimize new species introductions along  
104 the marine coastline of southern South America.

105

## 106 **2. Materials and Methods**

107

### 108 *2.1. Fouling sampling*

109 Of the ten main marine ports of Argentina, we surveyed six, all of them  
110 situated in the Patagonian region from 40°S to 54°S : San Antonio Este (SAE),  
111 Puerto Madryn (PM), Puerto Deseado (PD), Punta Quilla (PQ), Río Gallegos (RG)  
112 and Ushuaia (U, Fig. 1). At each port, a RAS (qualitative fouling sampling) was  
113 conducted in spring 2005 on the subtidal zone (i.e. just under the intertidal zone  
114 but never exposed to the air) by scuba diving and scraping the surface of different  
115 pilings (n = 3 to 5 samples per port, 25 x 25 cm each). Samples were collected by  
116 expert scientific divers, bagged separately, labeled, fixed in formalin (4%) and then  
117 preserved in ethanol (70%) excepting for the algae, which were kept in formalin.  
118 Later, samples were sorted and identified to the lowest possible taxonomic level

119 following the recommendations by Bortolus (2008; 2012a, b). Although most  
120 authors of this work have expertise in different taxa, we had the collaboration of  
121 several other expert taxonomists in order to cover most of the taxa found (see  
122 Acknowledgement section and Appendix A). Vouchers of the collected taxa were  
123 deposited in the Centro Nacional Patagónico (CENPAT) Invertebrate Collection.  
124 Planktonic and soft-bottom organisms were out of the scope of this study.

125 To identify the total biodiversity at each port, we complemented the RAS  
126 (qualitative sampling) with a survey with fouling plates (quantitative sampling).  
127 These plates ( $n = 15$  per port, 20 x 20 cm each, one plate per piling) were vertically  
128 deployed at each port along the subtidal zone, at 1.5 m below the average low tide,  
129 during 18-22 months. All plates were made of fiberglass homogeneously scratched  
130 to increase the roughness. Plates were deployed between October/November  
131 2005 (spring) and collected between June/July 2007 (Winter). At the end of this  
132 period all plates were placed separately in plastic bags and transported in coolers  
133 at  $\sim 5$  °C to the laboratory for processing. In the laboratory each plate was  
134 photographed, and the percentage cover of sessile species and the abundance of  
135 mobile species, were recorded. Then, all the organisms were removed from the  
136 plates, fixed and preserved following Hewitt and Martin (2001). All organisms  
137 collected were identified to the lowest taxonomic level possible and deposited in  
138 the Invertebrate Collection of the CENPAT. Organisms were classified as native,  
139 cryptogenic or exotic following Chapman and Carlton (1991). We noted if a species  
140 represented the first record for the region (FR), or if it was never previously  
141 mentioned in the regional literature as exotic or cryptogenic species (NM), and also  
142 those found outside their known regional geographic range (RE, range extension).

143

## 144 *2.2. Port characterization*

145 To assess differences and similarities among ports and to discuss the  
146 potential vulnerability of every port to marine invasive species, we developed a  
147 database with nine environmental variables based on field sampling and literature  
148 surveys (following Clarke et al., 2004, Table B.1 of Appendix B). The main  
149 variables considered were: 1) sea surface water temperature, 2) air temperature,

150 3) tidal amplitude, 4) wind speed, 5) surface salinity, 6) rainfall, 7) port depth, 8)  
151 type of port and 9) the environmental impact of the city. For the first seven  
152 variables we estimated their maximum, minimum and average values. The  
153 resultant matrix was composed by 26 different variables (see Appendix B for  
154 details). These variables were selected because they were identified influencing on  
155 the survivorship of intertidal and shallow subtidal organisms in the port  
156 environment, according to the studies carried on by the Globallast Programme (see  
157 for example Clarke et al. 2004 for the Port of Sepetiba, Brazil). In addition to these  
158 variables, we added wind speed because of its strong influence across the coastal  
159 area of Patagonia (Prohaska, 1970). The categorization of the environmental  
160 impact of the city was developed by Esteves (2007) considering coastal  
161 geography, the oceanographic and fluvial conditions, the pollution, and the  
162 eutrophication level recorded at each port (see Appendix B for details). In addition,  
163 to compare the port activity within the study area, we analyzed the average port  
164 movement (in tons) between 1998 and 2008 (Consejo Portuario Argentino, 2011  
165 and the references therein) and the average number of ship entries reported for the  
166 same period for all the ports excepting PD, PQ (both 1998-2005) and RG (2000-  
167 2005). The port movement was obtained from the statistics reported at each port  
168 and it represents the total cargo movement of domestic and international ships.  
169 The shipping entries represent the total number of vessels (domestic and  
170 international) entering at each port. Since ballast water discharge reports are not  
171 mandatories in Argentinean waters, this information was not available to analyze in  
172 this study (for detailed discussion see Boltovskoy et al., 2011).

173

### 174 2.3. *Data analysis*

175 To explain the relationships between environmental variables and the  
176 composition of the total biological assemblages among ports, two canonical  
177 correspondence analyses (CCA) were performed independently for mobile and  
178 sessile taxa using the package Vegan (Oksanen, 2011) in the R computing  
179 environment. Previously, a correlation matrix of the 26 environmental variables was  
180 studied to detect problems of multicollinearity (see Table B. 2 of Appendix B). The



181 final analysis of CCA was performed using the following seven variables which  
182 represented the main environmental characteristics of the ports that we studied:  
183 average annual surface water temperature, average tidal amplitude, average  
184 annual wind speed, salinity, average monthly rainfall, port's depth and type (see  
185 Table B. 3 of Appendix B for details). In addition, we used the one-way ANOVA to  
186 evaluate the null hypothesis of no differences in port movement (in tons) among  
187 ports (Zar, 1999). Another one-way ANOVA was used to evaluate the null  
188 hypothesis of no difference in taxonomic richness of the plates (mobile plus sessile  
189 taxa together) among ports (Zar, 1999). Levene and Kolgomorov-Smirnov tests  
190 were used to evaluate the homoscedasticity and normality of the data respectively.  
191 Data were square-root or log transformed to comply with the ANOVA assumptions.  
192 Finally, *a posteriori* Tukey tests were used to identify differences among means  
193 (Zar, 1999).

194

### 195 3. Results

196 A total of 247 fouling taxa and three associated fish species (Appendix A)  
197 were found; most organisms (77%) were recorded in the qualitative samples during  
198 the RAS, and most species (87%) were native. Overall, we found 17 exotic species  
199 (six macroalgae, five crustaceans, one bryozoan and five ascidians, Table 1) and  
200 15 cryptogenic species (four macroalgae, four hydrozoans, two polychaetes, two  
201 crustaceans, one bryozoan and two ascidians, Table 1). The use of plates allowed  
202 us to detect several species unrecorded during the RAS (Appendix A), including  
203 five cryptogenic species (the macroalgae *Bangia fuscopurpurea*, *Blidingia*  
204 *marginata*, *Dictyota dichotoma* and *Ectocarpus siliculosus*, and the ascidian  
205 *Cnemidocarpa robinsoni*) and five exotic species (the macroalgae *Anotrichium*  
206 *furcellatum*, the bryozoan *Bugula stolonifera* and the ascidians *Ciona intestinalis*,  
207 *Diplosoma listerianum* and *Molgula manhattensis*, Table 1).

208 The port of SAE showed the highest number of exotic and cryptogenic  
209 species with a total of 20, followed by PD with 12 species (Table 1). Our record for  
210 the colonial ascidian *Diplosoma listerianum* is the first for Argentinean waters,  
211 being observed in SAE (on 12 of 15 plates) and less abundantly in PD (on two

212 plates). We re-categorized as exotic two species previously known as native (the  
213 amphipods *Jassa marmorata* and *Crassikorophium bonnellii*), and four other  
214 species we re-categorized as cryptogenic (the hydrozoans *Amphisbetia operculata*,  
215 *Obelia bidentata* and *Halecium delicatulum*, Table 1). Finally, we detected a  
216 southward range extension for two known exotic species, the amphipod  
217 *Monocorophium insidiosum* and the ascidian *Molgula manhattensis*, found in U and  
218 PD, respectively. Nearly 50% of the surface mean cover on plates detected at SAE  
219 and PD were exotic or cryptogenic species (Fig. 2), while this percentage in the  
220 other ports was less than 13% (Fig. 2).

221 Mobile taxa were represented by turbellarians, polychaetes, brachyurans,  
222 carideans, isopods, amphipods, pycnogonids, gastropods, polyplacophorans,  
223 echinoderms and fishes (see Appendix A for complete species list). The first two  
224 CCA axes explained 90.9% (CCA1: 76.8% and CCA2: 14.1%) of the total variance  
225 in the analysis of mobile taxa (Fig. 3A). The ports of U, RG and PD were grouped  
226 showing similar taxa, mainly polychaetes, while PQ, SAE and PM differed their  
227 mobile taxa (Fig. 3A). Polychaetes, and particularly isopods, were abundant in PD.  
228 The port of SAE was the richest in terms of the mobile fauna. The carideans were  
229 present only in this port and the amphipods, mollusks, brachyurans and  
230 echinoderms showed their highest abundances there (Figs. 3A). Mobile fauna was  
231 almost absent in PQ. Ports were also separated by their environmental variables  
232 (Fig. 3A). The cold temperate ports of U, RG, PQ and PD were spread along the  
233 positive values of the first axis, while the warm temperate ports of SAE and PM  
234 were spread along the negative values also of the first axis. The ports of U, SAE  
235 and PM are situated in natural bays which were separated from PQ, PD and RG,  
236 located in estuarine areas. Salinity and temperature were high in SAE and PM and  
237 low in PQ and U. Rainfall was highest in U (Fig. 3A).

238 The cover values obtained from the plates for sessile taxa reached the  
239 maximum 100% in three ports (SAE, PD and U), ranging from 23% in RG to 72%  
240 in the remaining ports. The first two CCA axes explained 67.1% (CCA1: 37.8% and  
241 CCA2: 29.3%) of the total variance in the analysis of sessile taxa (Fig. 3B). Each  
242 port showed distinctive taxa composition, with the ascidians as the only taxonomic

243 group common to all ports. This taxon showed the highest average cover (85%) in  
244 PD, with eight species (three exotics and two cryptogenics, Table 1). Bryozoans,  
245 polychaetes and ascidians were the dominant faunal components in the ports of  
246 PD and U (Fig. 3B). The colonization by macroalgae registered on the plates was  
247 extremely low in most ports, excepting in PQ where they were dominant (average  
248 cover = 39%, Fig. 3B). Anthozoans were dominant in PM and abundant in SAE. In  
249 the latter the dominant taxon were the hydrozoans, mostly due to the presence of  
250 the cryptogenic *Ectopleura crocea* (average cover = 53%). The port of RG was  
251 very poor in terms of cover of sessile taxa, showing the lowest average cover  
252 (22.7%) compared to the other ports. Bivalves *Mytilus* spp. were the dominant  
253 taxon (17.3%). Environmental variables separated the ports in a similar way as the  
254 mobile taxa (Fig. 3B). The warm temperate ports of SAE and PM were also  
255 grouped by the high air and water temperatures, depth and salinity. The cold  
256 temperate ports (U, PD and PQ) were spread along the positive values of the  
257 second axis, except for RG which was closer to SAE and PM due to the high tidal  
258 amplitude. Rainfall was particularly high in U, and wind speed was highest in PQ.

259 The average port movement for the 1998-2008 period we analyzed showed  
260 significant differences among compared ports (square-root transformed data,  $F =$   
261  $123.4$ ,  $MS_{error} = 8941$ ,  $MS_{effect} = 1103881$ ,  $df_{error} = 60$ ,  $df_{effect} = 5$ ,  $p < 0.05$ , Fig. 4),  
262 with PM being the more active port with nearly 50% of the total movement, and  
263 significantly different from the others (Post-hoc Tukey test  $p < 0.05$ , Fig. 4). The U  
264 port was not significantly different from SAE or PD ( $p > 0.05$ ). Finally, the ports RG  
265 and PQ showed the lowest values in port movement (less than 5%,  $p < 0.05$ , Fig.  
266 4). These results were also accompanied by the average number of ship entries  
267 per port, excepting PD, in which the large number of ships showed a strong  
268 contrast with its port movement (Fig. 4).

269 Total taxonomic richness (considering both mobile and sessile taxa) was  
270 significantly different among the compared ports (square-root transformed data,  $F =$   
271  $78.9$ ,  $MS_{error} = 0.22$ ,  $MS_{effect} = 17.5$ ,  $df_{error} = 84$ ,  $df_{effect} = 84$ ,  $p < 0.05$ , Fig. 5A),  
272 showing the highest values for the plates deployed in SAE compared to the other  
273 ports (Post-hoc Tukey test  $p < 0.05$ , Fig. 5A). Also, the taxonomic richness was

274 higher in PD than in PM ( $p < 0.05$ ), but neither of them was found significantly  
275 different than U ( $p > 0.05$ ). The ports of PQ and RG showed the lowest taxonomic  
276 richness, with no significant differences between them ( $p > 0.05$ ). Finally, although  
277 the highest taxonomic richness was in SAE, the port of RG showed the highest  
278 percentage of exotic and cryptogenic species in relation to the total number of taxa  
279 found at that port (25%) mostly due to the high percentage of cryptogenic species  
280 (15.6%, Fig. 5B). In second place was SAE with 21.7% due to the high number of  
281 exotic species ( $n = 14$ ), which was the 15.2 % of the total number of the species  
282 observed (Fig. 5B).

283

## 284 4. Discussion

285

### 286 4.1. Assessment of marine exotic species and the port's environments

287 We detected a relatively large number of new records of exotic and  
288 cryptogenic species in addition to those reported in the literature for the ports we  
289 studied (see Orensanz et al., 2002; Schwindt, 2008). Some of them refer to  
290 species that had been previously misidentified as native, and which after reviewing  
291 the literature and museum collections, we re-classified them more properly as  
292 exotic or cryptogenic species. Our results include the third exotic colonial ascidian  
293 reported to have been introduced to Patagonia (*Diplosoma listerianum*, Table 1)  
294 after the styelid *Botryllus schlosseri*, collected for the first time in 1962 (Amor,  
295 1964), and *Lissoclinum fragile*, detected for the first time in 2004 (Rico et al., 2012)  
296 and which we recorded in SAE. *Diplosoma listerianum* and *L. fragile* are currently  
297 spread throughout the Western Pacific, South Pacific, and Indian Ocean; the  
298 Caribbean, Brazil, and West Africa (Rocha and Kremer, 2005; Carlton and  
299 Eldredge, 2009). Although *D. listerianum* is considered native to Europe (e.g.  
300 Monniot et al., 2001), its broad global distribution makes it difficult to determine a  
301 precise native area (Carlton and Eldredge, 2009) hence the need for DNA data.  
302 Ascidians are considered good indicators of anthropogenic transport over long  
303 distances because they have short lifespan and lecithotrophic larvae and,  
304 consequently, natural long distance dispersal is highly unlikely for these animals

305 (Lambert and Lambert, 1998). Moreover, since the larval stage is so short, the  
306 primary mode of anthropogenic transport of ascidians is likely to be hull fouling,  
307 which suggests that once introduced into a new region, local dispersal via domestic  
308 shipping is highly probable as a fouling species. This is particularly important for  
309 the Patagonian region, where a large proportion of the port entries are attributable  
310 to domestic shipping (Boltovskoy et al., 2011). We actually expect these tunicate  
311 species to disperse by shipping to other ports along the region in the near future,  
312 eventually reaching the Uruguayan coast in the North. In support to this we have  
313 recently found specimens of *D. listerianum* in PM (March 2012; Schwindt and  
314 Tatián, unpubl. data).

315 Most the ascidians found in PM were exotic species. Of the three exotic  
316 species found in this port, *Ascidella aspersa* is considered as pioneer organism on  
317 artificial substrates (Collins et al., 2002; Schwindt et al., 2013). In Argentina, forty  
318 years after the introduction of *Ascidella* (Tatián et al., 2010) studies showed that  
319 this species is not only one of the first species settling on fouling plates, but also  
320 that it quickly becomes a pest, overgrowing other exotic species like the invasive  
321 *Ciona intestinalis* (Schwindt et al., 2013). Among the eight ascidian species found  
322 in SAE, six of them (75%), are exotics or cryptogenics. Although *Diplosoma*  
323 *listerianum* is a new invader, this species showed the highest cover among all the  
324 ascidians we found growing on plates, and together with other exotic fouling  
325 species, were dominant over the native sessile species in this port. These species  
326 are well known because they can recruit rapidly and dominate the substrate and  
327 resist adverse conditions such as pollution from sewage, land runoff, heavy metals  
328 and periods of low salinity. Also, they show a high physiological flexibility that  
329 facilitates their success in all kind of ports and aquaculture facilities (Lambert and  
330 Lambert, 2003). Thus, the presence of new invader species like *A. aspersa*,  
331 *Molgula manhattensis* and *D. listerianum* could change dramatically the  
332 composition of the fouling communities in a short period.

333 The richness of the fouling species is not homogeneous across the ports of  
334 Patagonia, as each port was characterized by different taxonomic groups. It is  
335 noteworthy that the port of SAE showed not only the highest number of sessile and

336 mobile taxa (dominated by hydroids and amphipods respectively), but it also  
337 showed the highest number of exotic and cryptogenic species among the ports that  
338 we studied. Although the maritime activity of SAE (i.e. number of ship entries and  
339 port movement) was not the highest among the ports compared, it is still a major  
340 regional node for exporting goods, comparable to PD and U (Boltovskoy et al.,  
341 2011). In fact, these are the only ports almost exclusively receiving vessels laden  
342 with ballast water, and therefore the propagule pressure is expected to be higher  
343 there than in the other ports. Concordantly to this, we have found that SAE and PD  
344 ports have the highest number of exotic and cryptogenic species (20 and 12  
345 respectively) among all ports studied, suggesting that a closer surveillance is  
346 needed there.

347 Although port movement was similar in U and SAE, which are both export-  
348 oriented ports (Boltovskoy et al., 2011), the number of exotic and cryptogenic  
349 species found in U was among the lowest recorded ( $n = 4$ ). Only PQ had the same  
350 low number of exotic and cryptogenic species, being this port one of the least  
351 active in the region. On the other hand, we found that RG doubles the number of  
352 exotic and cryptogenic species of PQ port, which is very similar to RG in terms of  
353 regional shipping activity (scarce in both) and low taxonomic richness. The  
354 proportion of exotic and cryptogenic species in relation to the native biodiversity we  
355 found in RG is one of the highest among the ports studied. Considering that none  
356 of the non native species found in these ports were new arrivals, and that the port  
357 movement is relatively low there, it was expected that PQ and RG have a low  
358 vulnerability to new introductions. Since the sampling effort and level of expertise  
359 were the same in all ports, these unexpected results strongly support the  
360 hypothesis about the existence of environmental and biological variables able to  
361 modulate the propagule pressure for a given site, especially in the port of U  
362 (Boltovskoy et al., 2011).

363 The port of PM doubles the average number of ship entries of U and almost  
364 three times that of SAE. Although taxonomic richness of PM was lower than in SAE  
365 and comparable to U, the percentage of exotic and cryptogenic species found in  
366 this port was one of the highest within the ports studied. This is a striking finding

367 since PM is not one the ports receiving important discharges of ballast water  
368 (Boltovskoy et al., 2011). This port is situated within a natural bay with signs of  
369 contamination by heavy metals and/or eutrophication (Gil et al., 1999; Diaz et al.,  
370 2002) It was through this port that the macroalga *Undaria pinnatifida* was  
371 introduced and nowadays is one of the most aggressive marine invasive species in  
372 Southern South America, affecting the abundance and richness of native  
373 organisms (Casas et al., 2004; Irigoyen et al., 2011). Therefore, the results of this  
374 study suggest that more data about the shipping activity are needed to better  
375 understand bioinvasions and the vulnerability of this port to new introductions.

376

#### 377 4.2. The South American context of marine invasive species

378 While scientists have surveyed ports and coastal areas worldwide, cross-  
379 regional comparisons are still difficult to perform due to the implementation of  
380 different methods used and the often contrasting environmental conditions.  
381 Nevertheless, more efforts should be emphatically directed to coordinate  
382 international research teams to address cross-regional comparisons. In South  
383 America, other rapid assessment surveys of marine exotic species were performed  
384 in specific sites of Brazil (Ignacio et al., 2010; Marques et al., 2013), but exhaustive  
385 examinations of marine exotic and cryptogenic species were compiled only in  
386 Argentina, Uruguay (41 and 50 respectively, Orensanz et al., 2002; updated in  
387 Schwindt, 2008), Chile (51 and 47 respectively, Castilla and Neill, 2009) and  
388 Venezuela (22 and 67 respectively, Pérez et al., 2007). National reports and/or  
389 specific case-study publications were completed in Colombia (Gracia et al., 2011)  
390 and Brazil (e.g. Souza and Silva, 2004; Ferreira et al., 2009; Lopes, 2009;  
391 Farrapeira et al., 2011). The number of marine and brackish water exotic species  
392 reported in countries of South America is low if they are compared with other  
393 countries as Italy (89, Occhipinti-Ambrogi et al., 2011), South Africa (86, Mead et  
394 al., 2011), Britain (90, Minchin et al., 2013), Israel (296, Galil, 2007) and Germany  
395 (85, Gollasch and Nehring, 2006), among others. The scarce reports and  
396 compilations plus the intense maritime traffic of some South American countries  
397 (see below) calls the attention to the need of increase the surveys and monitoring

398 programs in and around ports and ports of South America. A step forward to  
399 achieve an international cross-regional collaboration is given by the Convention for  
400 the Control and Management of Ship's Ballast Water and Sediments, signed in  
401 2004 by 74 States. However, the only country in South America that ratified the  
402 Convention was Brazil (IMO, 2014).

403 Every protocol to survey marine invasive species has weaknesses and  
404 strengths (reviewed in Campbell et al., 2007) and they are strongly dependent of  
405 the budget and the availability of taxonomists. In spite of this, it is clear that the  
406 profuse maritime commercial activity linking South American countries must be  
407 complemented with effective sampling protocols to detect invasive species  
408 (Campbell et al., 2007; Bishop and Hutchings, 2011). To achieve this goal, it is  
409 critical to identify the major potential routes of introduction. For instance, the United  
410 States of America and China represent together the major import/export partners of  
411 South American countries (nearly 50% of the maritime relationships for Venezuela;  
412 The World Factbook, 2013-14). However, the countries facing the Pacific coast of  
413 South America, have more commercial relationships with USA, China and other  
414 countries of the Pacific like Japan and South Korea (ranging between 41 and 52%  
415 of exports and imports) than among them, being the intraregional commerce of  
416 imports and exports lower than 8% (The World Factbook, 2013-14). On the other  
417 hand, along the Atlantic coast of South America, Brazil, Argentina and Uruguay  
418 have more commercial interchange among them than with the countries situated  
419 on the Pacific coast. For these last group, Brazil is the major import/export partner  
420 (ranging between 16 and 27%, The World Factbook, 2013-14) and its commercial  
421 activity is so important that in 2011 the 19.1% of the total containership occurred in  
422 Latin America and the Caribbean region was operated through Brazilian ports  
423 (Sánchez, 2012). Moreover, Santos harbor (23° 58'S, 46° 17'W) is one of the 20  
424 most important harbors of the world with maritime activity, only compared to  
425 Panama (Kaluza et al., 2010). Thus, Brazil appears to be a major stepping stone in  
426 the region for marine invasions problem, that would likely contribute with their own  
427 biota (native and non native) to the rest of its commercial partners in South  
428 America.



429

**430 5. Conclusions**

431

432 Scientists' ability to predict the vulnerability of a given habitat or community to  
433 invasions is largely hampered by the multiple variables involved (Byers, 2002;  
434 Johnston et al., 2009; Olyarnik et al., 2009). However, it is by performing the  
435 analysis of global patterns that scientists will be able to provide the best support to  
436 managers and decision-makers. The expedient and extensive rapid assessment  
437 survey we present in this study, complemented by quantitative sampling of fouling  
438 plates and an extensive compilation of significant environmental variables, provide  
439 the first large-scale information baseline of bioinvasion analysis along the Southern  
440 South American ports. We expect that these results will assist managers to design  
441 more optimal monitoring programs and will speed up the development of  
442 appropriate legislation for preventing further bioinvasions.

443

**444 Conflict of interest**

445 Authors declare that they do not have any conflict of interest.

446

**447 Author contribution**

448 ES conceived the ideas; ES, AB, GL lead the field work; ES, MPR, MED, MMM,  
449 VS, MCS performed lab work; ES, JLG, MPR, MT, JMO, GA, MED, BD, GG, CL,  
450 MLP identified the taxa; ES, JLG analyzed the data; ES, JLG and AB led the  
451 writing. All authors have approved the final article.

452

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472

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715 **Figure Legends**

716 **Fig. 1.** Studied marine ports of Argentinean Patagonia: San Antonio Este (SAE),  
717 Puerto Madryn (PM), Puerto Deseado (PD), Punta Quilla (PQ), Río Gallegos (RG)  
718 and Ushuaia (U).

719 **Fig. 2.** Mean cover (in percentage, + SE) of exotic and cryptogenic species found  
720 on fouling plates at each port. Abbreviations are the same as in Fig. 1.

721 **Fig. 3.** Canonical correspondence analysis triplot showing the ordination of ports  
722 (SAE, PM, PD, PQ, RG and U, see abbreviations in Fig. 1), environmental  
723 variables (Te: temperature, Ti: tidal amplitude, Wi: wind speed, S: salinity, Ra:  
724 rainfall, De: depth and PT: Port type), mobile taxa (A, Is: isopods, Po: polychaetes,  
725 Py: pycnogonids, Ba: brachyurans, Ca: carideans, Am: amphipods, Mo: mollusks,  
726 Ec: echinoderms, Tu: turbellarians) and sessile taxa (B, Ma: macroalgae, Ci:  
727 cirripedia, Br: bryozoans, Mo: mollusks, Pr: porifera, Po: polychaetes, As:  
728 ascidians, An: anthozoans, Hy: hydrozoans).

729 **Fig. 4.** Average port movement + SD (bars, left y axis) between 1998 and 2008  
730 and average number of ship entries (diamonds, right y axis) reported for the same  
731 period for all the ports except for PD and PQ (1998-2005) and RG (2000-2005).  
732 Abbreviations of the ports are the same as in Fig. 1. Same letters indicate not  
733 statistically significant differences.

734 **Fig. 5.** Average taxonomic richness (A) and percentage of exotic and cryptogenic  
735 species (B) at each port. Same letters mean not statistically significant differences.  
736 Abbreviations are the same as in Fig. 1.

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738 **APPENDIX**

739 **APPENDIX A.** Organisms found during the qualitative and quantitative sampling in  
740 all ports.

741 **APPENDIX B.** Environmental variables studied at each port (Table B.1) and  
742 Spearman rank order correlation matrix (Tables B.2 and B.3).

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745 **Table 1.** Exotic and cryptogenic species recorded in Patagonian ports (SAE: San Antonio  
 746 Este, PM: Puerto Madryn, PD: Puerto Deseado, PQ: Punta Quilla, RG: Rio Gallegos, U:  
 747 Ushuaia) and their status (exotic, cryptogenic). TS: species that need taxonomic study,  
 748 FR: species that represents the first record for the Patagonian coast, NM: species that  
 749 were never mentioned in the SWA literature as exotic or cryptogenic, RE: exotic species  
 750 that extended the distribution range according to the earliest reports in the region, **P**: taxa  
 751 found only in fouling plates but not in the qualitative sampling, S: taxa found only during  
 752 the qualitative sampling but not on the plates, B: taxa found on plates and during the  
 753 qualitative sampling. Next to each taxon between brackets is the Phylum to which belongs  
 754 each taxon. R: Rodophyta, Cl: Chlorophyta, O: Ochrophyta, Cn: Cnidaria, An: Annelida,  
 755 Ar: Arthropoda, M: Mollusca, B: Bryozoa, Ch: Chordata,

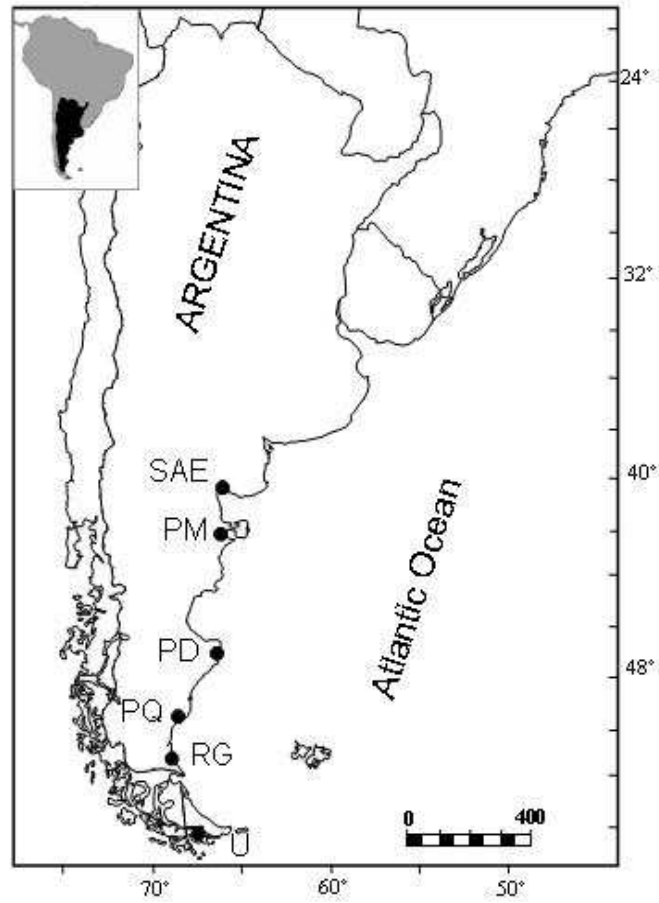
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Species	Ports						Comments
	SAE	PM	PD	PQ	RG	U	
<b>EXOTICS</b>							
<i>Anotrichium furcellatum</i> (R)	P						Observed in Argentina since 1984 (Boraso de Zaixso and Akselman, 2005)
<i>Lomentaria clavellosa</i> (R)	S	S					Native to Europe (Mathieson et al., 2008)
<i>Neosiphonia harveyi</i> (R)	S	S					Previously described as <i>Polysiphonia argentinica</i> in 1872 (Taylor, 1939)
<i>Rosenvingiella polyrhiza</i> (Cl)					S		First collected in 1972 (Boraso de Zaixso, 2002)
<i>Cutleria multifida</i> (O)	S						First reported in Argentina around 1965 (Asensi, 1971)
<i>Undaria pinnatifida</i> (O)		S					See Orensanz et al. (2002)
<i>Balanus glandula</i> (Ar)	S	B	S				First collected in 1974 (Spivak and L'Hoste, 1976)
<i>Monocorophium insidiosum</i> (Ar)	S				S	S	RE. First collected in 1968 in fouling communities (López Gappa et al., 2006)
<i>Monocorophium acherusicum</i> (Ar)	B		S		S		First collected in Argentina in 1961 (USNM # 127701)
<i>Crassikorophium bonellii</i> (Ar)			S			S	NM. The species was barely observed since 1892. A recent taxonomic study confirmed its presence and suggested its native area (Alonso, 2012)
<i>Jassa marmorata</i>	S						NM. Eastern North Atlantic

(Ar)					origin (Mead et al., 2011). Observed in Argentina and Uruguay since 1968 (Alonso de Pina, 2005) From 38° to 40°S strongly associated to port areas (López Gappa, 2000) See text (Tatián et al., 2010)
<i>Bugula stolonifera</i> (B)	P				
<i>Asciidiella aspersa</i> (Ch)	B	B	B		TS. More detailed studies are needed for this region (see Caputi et al., 2007). Regional records of <i>Ciona robusta</i> belong to <i>C. intestinalis</i> (Hoshino and Nishikawa, 1985)
<i>Ciona intestinalis</i> (Ch)	P	P			FR. Origin unknown First observed in 2004 (Rico et al., 2012). Origin unknown
<i>Diplosoma listerianum</i> (Ch)	P		P		RE. Strongly associated to port areas (Orensanz et al., 2002; Rico et al., 2012)
<i>Lissoclinum fragile</i> (Ch)	B				
<i>Molgula manhattensis</i> (Ch)	P		P		
<b>CRYPTOGENICS</b>					
<i>Bangia fuscopurpurea</i> (R)			P	S	TS. Observed in Argentina since 1969 (Mendoza, 1970). This species might be species complex (Guiry and Guiry, 2012)
<i>Blidingia marginata</i> (Cl)				P	TS. Idem to <i>B. fuscopurpurea</i>
<i>Dictyota dichotoma</i> (O)	S		P		TS. This species requires a global taxonomic revision
<i>Ectocarpus siliculosus</i> (O)	P				TS. Wide distribution in NE Atlantic
<i>Ectopleura crocea</i> (Cn)	B			B	See Imazu et al. (2014)
<i>Obelia bidentata</i> (Cn)				B	NM. Found only in port areas (Genzano et al., 2009). Origin unknown.
<i>Amphisbetia operculata</i> (Cn)		S	S	B	NM. Introduced in Australia (Hewitt et al., 2004)
<i>Halecium delicatulum</i> (Cn)				B	NM. Introduced in Australia (Hewitt et al., 2004)
<i>Boccardia polybranchia</i> (An)				S	TS. This species might be a species complex
<i>Syllis gracilis</i> (An)		S			See Orensanz et al. (2002)

<i>Amphibalanus improvisus</i> (Ar)	B						See Orensanz et al. (2002)
<i>Caprella equilibra</i> (Ar)	B						Strongly associated to port areas (López Gappa et al., 2006)
<i>Conopeum reticulum</i> (Ar)			S				Scattered records from 38° to 47°S (López Gappa, 2000)
<i>Cnemidocarpa robinsoni</i> (Co)	B	B	B			P	TS. Highly similar to <i>Asterocarpa humilis</i> reported as introduced in continental Chile (Clarke and Castilla, 2000)
<i>Corella eumyota</i> (Co)			B				Records in Argentina are scarce and reported for first time in the SWA in 1938 (Ärnback-Christie-Linde, 1938)
<b>Total Number of Exotic Species</b>	<b>14</b>	<b>6</b>	<b>6</b>	<b>0</b>	<b>3</b>	<b>2</b>	
<b>Total Number of Cryptogenic Species</b>	<b>6</b>	<b>1</b>	<b>6</b>	<b>4</b>	<b>4</b>	<b>2</b>	

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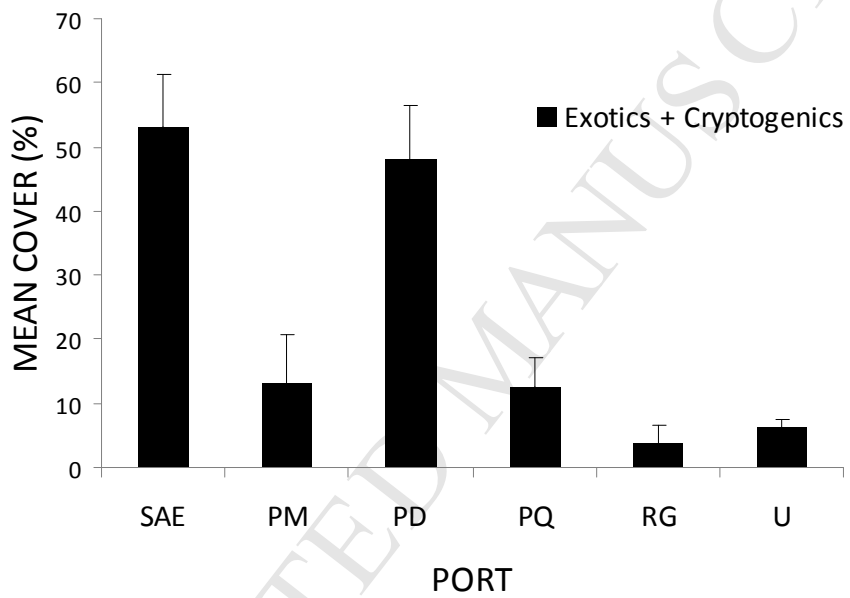
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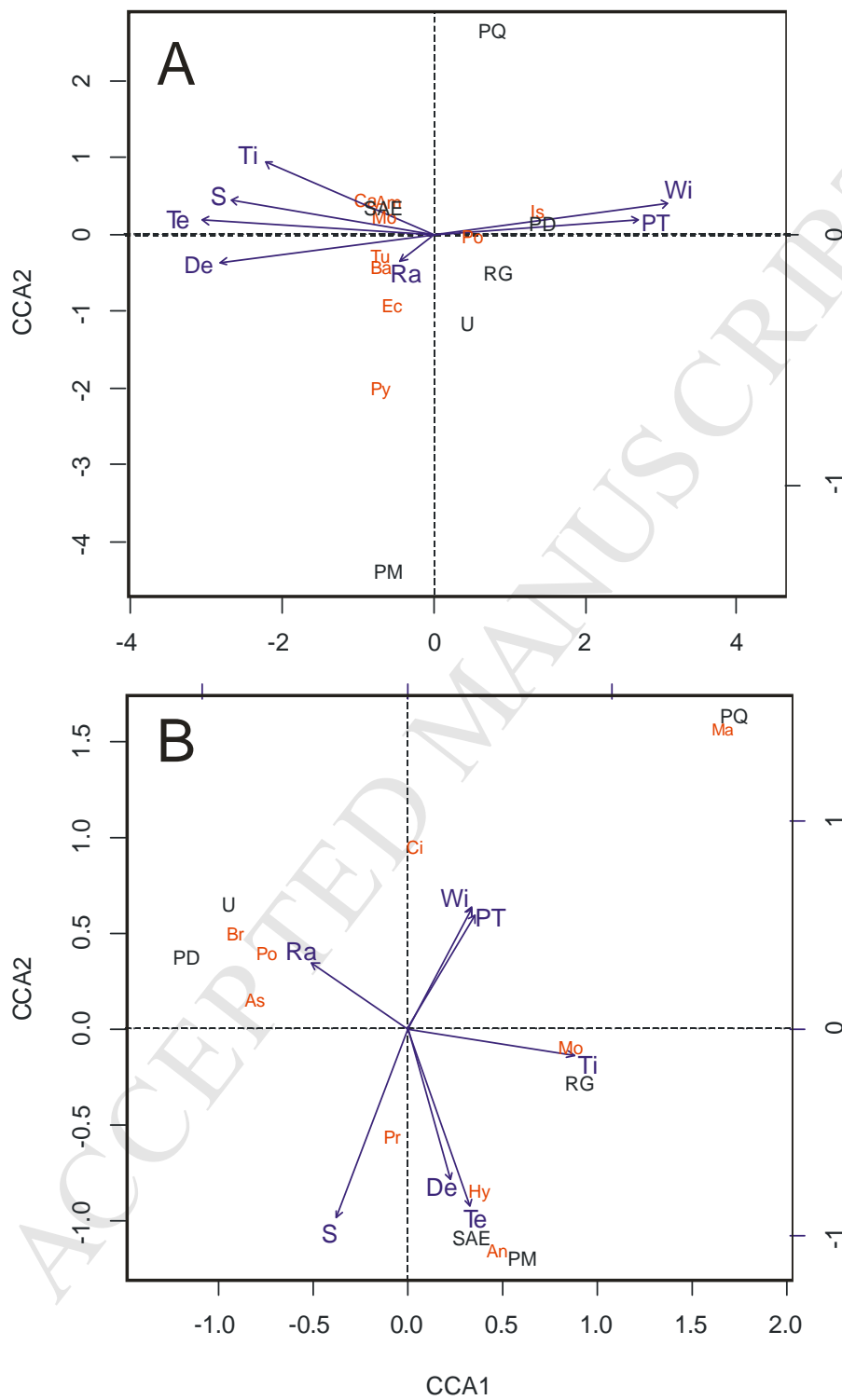
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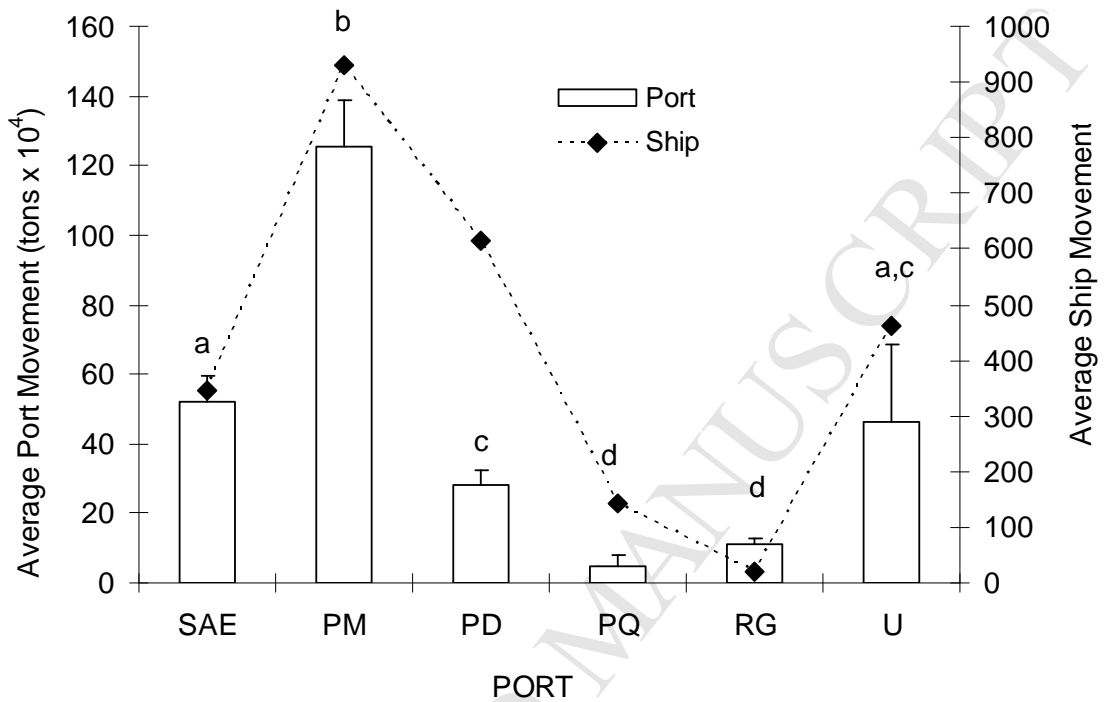
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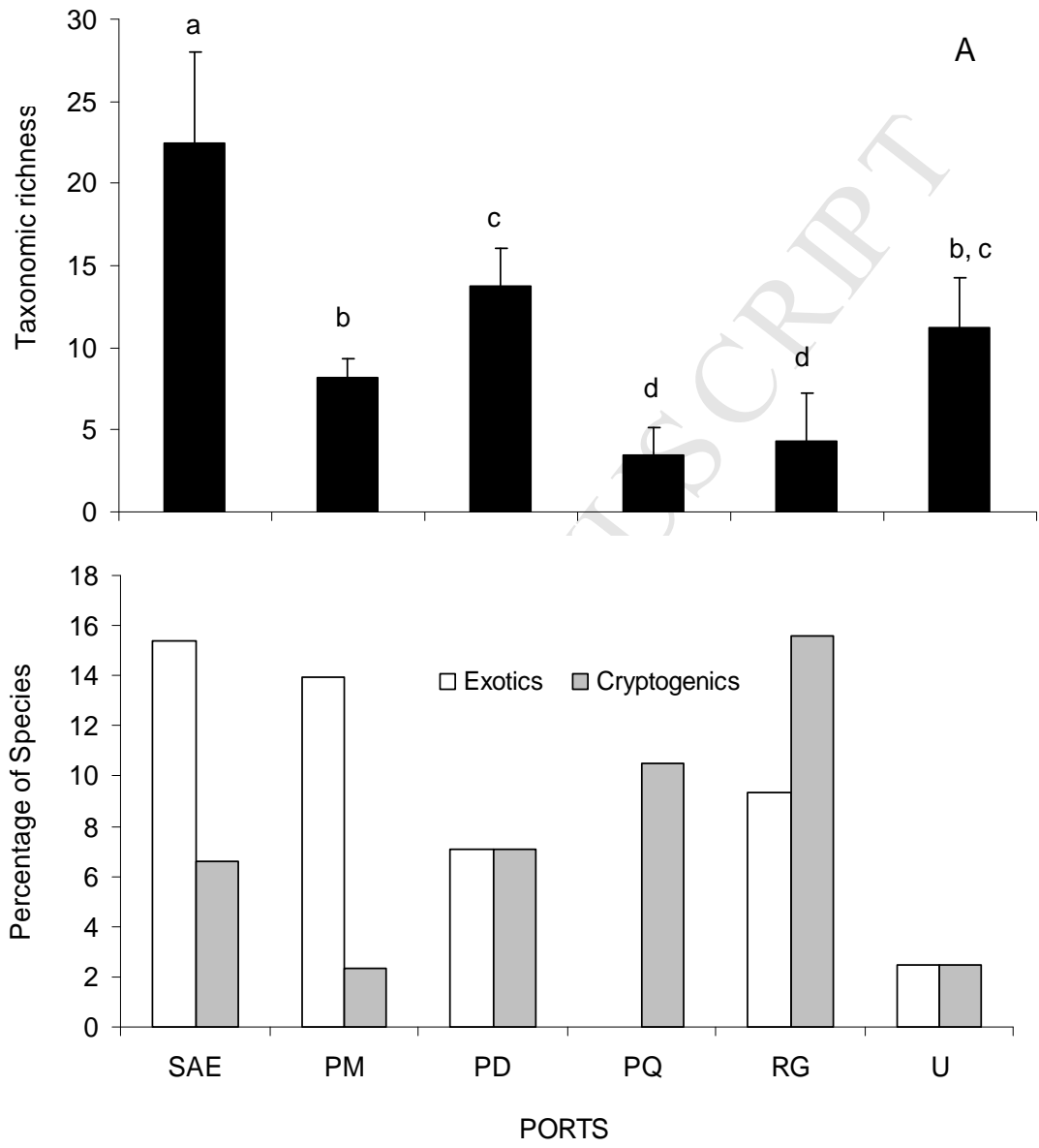


Fig. 5. Schwindt et al.

## **Marine fouling invasions in ports of Patagonia (Argentina) with implications for legislation and monitoring programs**

Evangelina Schwindt, Juan López Gappa, María Paula Raffo, Marcos Tatián, Alejandro Bortolus, José María Orensanz, Gloria Alonso, María Emilia Diez, Brenda Doti, Gabriel Genzano, Cristian Lager, Gustavo Lovrich, María Luz Piriz, María Martha Mendez, Verónica Savoya, María Cruz Sueiro

### **Highlights**

1. Marine native, exotic and cryptogenic species along major ports of Argentina are reported.
2. The port with the highest specific richness showed the highest number of exotic species.
3. A new marine exotic species is reported for Argentinean waters.
4. Taxa composition, environmental variables and port movement were different at each port.
5. Port's vulnerability to future introductions is discussed.

## Appendix A

**Table A.1.** Taxa identified at each Patagonian port (SAE: San Antonio Este, PM: Puerto Madryn, PD: Puerto Deseado, PQ: Punta Quilla, RG: Río Gallegos, U: Ushuaia) with the name of the taxonomic specialist responsible for its identification. Taxa are separated by Phylum and between brackets are the credits for the taxonomic identifications. P: taxa found only in fouling plates but not in the qualitative sampling, S: taxa found only during the qualitative sampling but not on the plates, B: taxa found on plates and during the qualitative sampling. Name of the Institutions abbreviated: UNLP: Universidad Nacional de La Plata (Argentina), UNPSJB: Universidad Nacional de la Patagonia San Juan Bosco (Argentina), UNMDP: Universidad Nacional de Mar del Plata (Argentina), UBA: Universidad de Buenos Aires (Argentina), DINARA: Dirección Nacional de Recursos Acuáticos (Uruguay), CENPAT: Centro Nacional Patagónico (Argentina), ECOSUR: El Colegio de la Frontera Sur (México).

Major taxonomic group	Ports					
	SAE	PM	PD	PQ	RG	U
<b>Phylum Rhodophyta</b> (ML Piriz)						
<i>Acanthococcus antarcticus</i> J.D. Hooker & Harvey			S	S		
<i>Acrochaetium</i> sp.	P					
<i>Anotrichium furcellatum</i> (J. Agardh) Baldock	P					
<i>Antithamnion</i> sp.	S		S			
<i>Aphanocladia robusta</i> Pujals	S					
<i>Ballia callitricha</i> (C. Agardh) Kützing			S	B		
<i>Bangia fuscopurpurea</i> (Dillwyn) Lyngbye				P	S	
<i>Callithamnion gaudichaudii</i> C. Agardh			P	S		
<i>Callophyllis</i> sp.				S		
<i>Ceramium tenuicorne</i> (Kützing) Waern	B			S		
<i>Ceramium virgatum</i> Roth	P		S		S	
<i>Chondria macrocarpa</i> Harvey				S		
<i>Cladodonta lyallii</i> (J.D. Hooker & Harvey) Skottsberg						P
Corallinaceae		S				
<i>Falklandiella harveyi</i> (J.D. Hooker) Kylin				S		
<i>Delesseria macloviana</i> Skottsberg						B
Delesseriaceae						B
<i>Erythrotrichia carnea</i> (Dillwyn) J. Agardh				P		
<i>Griffithsia antarctica</i> J.D. Hooker & Harvey						S
<i>Heterosiphonia merenia</i> Falkenberg			S	S		
<i>Hymenena falklandica</i> Kylin						S
<i>Hymenena laciniata</i> (J.D. Hooker & Harvey) Kylin			S			P
<i>Hymenena</i> sp.				S		
<i>Lomentaria clavellosa</i> (Lightfoot ex Turner) Gaillon	S	S				
<i>Lophurella hookeriana</i> (J. Agardh) Falkenberg				S		
<i>Medeiothamnion flaccidum</i> (J.D. Hooker & Harvey)			B			
Brauner						
<i>Neosiphonia harveyi</i> (Bailey) M.-S.Kim, H.-G.Choi, Guiry & G.W.Saunders	S	S				
<i>Phycodrys quercifolia</i> (Bory de Saint-Vincent)			S			S
Skottsberg						
<i>Picconiella pectinata</i> (J.D. Hooker & Harvey) De Toni						P
fil.						
<i>Picconiella plumosa</i> (Kylin) J. De Toni						S
<i>Plocamium secundatum</i> (Kützing) Kützing				S		
<i>Polysiphonia abscissa</i> J.D. Hooker & Harvey	S					

<i>Polysiphonia</i> spp.	P					
<i>Pseudolaingia larsenii</i> (Skottsberg) Levring						P
<i>Pterothamnion plumula</i> (J. Ellis) Nägeli				S		
<i>Pyropia columbina</i> (Montagne) W.A.Nelson				B	S	
Rhabdoniaceae	B					
<i>Rhodymenia corallina</i> (Bory de Saint-Vincent)						S
Greville						
Rhodymeniaceae						S
<i>Streblocladia camptoclada</i> (Montagne) Falkenberg	S					
<i>Streblocladia corymbifera</i> (C. Agardh) Kylin 1938	S					
<b>Phylum Chlorophyta</b> (ML Piriz)						
<i>Blidingia marginata</i> (J. Agardh) P. Dangeard						P
<i>Chaetomorpha aerea</i> (Dillwyn) Kützing				S		
<i>Cladophora falklandica</i> (J.D. Hooker & Harvey) J.D. Hooker & Harvey				P		
<i>Derbesia furcata</i> Ricker						P
<i>Derbesia</i> sp.						S
<i>Prasiola stipitata</i> Suhr ex Jessen						S
<i>Rhizoclonium</i> sp.				P		
<i>Rosenvingiella polyrhiza</i> (Rosenvinge) Silva						S
<i>Ulothrix flacca</i> (Dillwyn) Thuret					P	
<i>Ulothrix subflaccida</i> Wille				S		
<i>Ulva intestinalis</i> Linnaeus				S		
<i>Ulva lactuca</i> Linnaeus				S		S
<b>Phylum Ochrophyta</b> (ML Piriz)						
<i>Cladostephus spongiosus</i> (Hudson) C. Agardh						S
<i>Cutleria multifida</i> (Turner) Greville	S					
<i>Dictyota dichotoma</i> (Hudson) Lamouroux	B			P		
<i>Ectocarpus siliculosus</i> (Dillwyn) Lyngbye 1819	P					
<i>Macrocystis pyrifera</i> (Linnaeus) C. Agardh						B
<i>Microzonia velutina</i> (Harvey) J. Agardh						S
<i>Stypocaulon funiculare</i> (Montagne) Kützing						S
<i>Undaria pinnatifida</i> (Harvey) Suringar				S		
<b>Phylum Porifera</b> (J López Gappa)						
<i>Amphilectus</i> sp.						B
<i>Amphimedon</i> sp.	P					
<i>Cliona</i> sp.	B	S				
<i>Halichondria</i> sp.		S				
<i>Haliclona</i> sp.	P			B	S	B
<i>Mycale</i> sp.						S
<i>Spongia</i> sp.				B		
<i>Sycon</i> sp.	B					
<b>Phylum Cnidaria</b> (Actiniaria: E Schwindt and MP Raffo, Hydrozoa: G Genzano and MP Raffo)						
<i>Corynactis</i> sp.	B	S				
<i>Anthothoe chilensis</i> (Lesson, 1830)	P	B				
<i>Metridium senile lobatum</i> (Carlgren, 1899)		B				
<i>Antholoba achates</i> (Drayton in Dana, 1846)		B	B			
<i>Ectopleura crocea</i> (Agassiz, 1862)	B				B	
<i>Obelia geniculata</i> (Linnaeus, 1758)		S			S	
<i>Obelia bidentata</i> Clark, 1875						B
<i>Amphisbetia operculata</i> (Linnaeus, 1758)				S	S	B
<i>Symplectoscyphus milneanus</i> (d'Orbigny, 1846)					B	

<i>Sertularella polyzonias</i> (Linnaeus, 1758)					S	B
<i>Nemertesia</i> sp.					B	
<i>Lafoea dumosa</i> (Fleming, 1820)						B
<i>Halecium delicatulum</i> Coughtrey, 1876						B
<i>Eudendrium ramosum</i> (Linnaeus, 1758)						B
<b>Phylum Platyhelminthes</b> (F Brusa, UNLP)						
<i>Phrikoceros mopsus</i> (Marcus, 1952)	B					
<i>Thysanozoon</i> sp.	P					
SO. Acotylea			B			S
<b>Phylum Nemertea</b>	S	S				S
<b>Phylum Sipuncula</b>	S		S			
<b>Phylum Annelida</b> (S Salazar Vallejo (ECOSUR), ME Diez, JM Orensanz)						
F. Chaetopteridae						P
F. Chrysopetalidae	P					
F. Cirratulidae	P	S	B		S	B
F. Eunicidae	P	B				
<i>Eunice</i> cf. <i>argentinensis</i>		S	S			
<i>Marphysa</i> cf. <i>aenea</i>			S			
F. Flabelligeridae	S					
<i>Pherusa</i> sp.	S					
<i>Pherusa gymnopapillata</i> Hartmann-Schröder, 1965			P			
F. Lumbrineridae	B		P		P	S
F. Nereididae		S			P	B
<i>Perinereis</i> sp.			S			
<i>Platynereis australis</i> (Schmarda, 1861)				P		S
<i>Phylo</i> sp.			S			
<i>Arabella acuta</i> (Kinberg, 1865)			P			
<i>Halosydna patagonica</i> Kinberg, 1857			S	S	B	
<i>Halosydnella australis</i> (Kinberg, 1855)	S					
<i>Harmothoe</i> sp.	S					
<i>Harmothoe exanthema</i> (Grube, 1858)		S	P			
<i>Harmothoe madrynensis</i> Barnich, Orensanz & Fiege 2012	S	S	B			
<i>Harmothoe magellanica</i> (McIntosh, 1885)				S		
<i>Hermadion magalhaensis</i> Kinberg, 1855						S
<i>Lepidasthenia</i> cf. <i>esbelta</i>	S					
<i>Neopolynoe antarctica</i> (Kinberg, 1858)						S
F. Phyllodocidae			S			
<i>Eumida</i> sp.			P		P	
<i>Eteone</i> sp.	S					
F. Sabellidae	B		B			
<i>Parasabella</i> sp.	B		S			S
<i>Notaulax</i> sp.			S			
F. Serpulidae	P		P			
<i>Hydroides plateni</i> (Kinberg, 1867)		B				
SF. Spirorbinae		S	P		P	B
<i>Boccardia polybranchia</i> (Haswell, 1885)					S	
F. Syllidae	B	S	B		B	
<i>Syllis</i> sp.			S			
<i>Syllis gracilis</i> Grube, 1840			S			
F. Terebellidae	B	B	B		B	B
<i>Thelepus</i> sp.			S	S		S

<b>Phylum Arthropoda</b> (Caridea: E Gómez Simes, UNPSJB, Brachyura: MP Raffo, Cirripedia: E Schwindt, Pycnogonida: R Elias, UNMdP, Amphipoda: G Alonso, Isopoda: B. Doti)						
<i>Betaeus lilianae</i> Boschi, 1966	B					
<i>Nauticaris magellanica</i> (A. Milne Edwards, 1891)			S			S
<i>Rochinia gracilipes</i> A. Milne Edwards, 1875	B					
<i>Pelia rotunda</i> A. Milne Edwards, 1875	S					
<i>Libinia spinosa</i> H. Milne Edwards, 1834	S					
<i>Halicarcinus planatus</i> (Fabricius, 1775)	B	S	B	S	B	S
<i>Pilumnus reticulatus</i> (Stimpson, 1860)	B	S				
<i>Pachycheles chubutensis</i> Boschi, 1963	B	B				
<i>Eurypodius</i> sp.						S
<i>Austromegabalanus psittacus</i> (Molina, 1782)	S		S			S
<i>Balanus glandula</i> Darwin, 1854	S	B	S			
<i>Amphibalanus improvisus</i> (Darwin, 1854)	B					
<i>Balanus laevis</i> Brugière, 1789						B
<i>Elminius kingii</i> Gray, 1831						
<i>Anoplodactylus petiolatus</i> (Krøyer, 1844)	B					
<i>Achelia assimilis</i> (Haswell, 1885)			S			S
<i>Pycnogonum</i> spp.			S			
<i>Monocorophium insidiosum</i> (Crawford, 1937)	S				S	S
<i>Monocorophium acherusicum</i> (Costa, 1853)	B		S		S	
<i>Corophium</i> s.l.						S
<i>Crassikorophium bonnellii</i> (Milne Edwards, 1830)			S			S
<i>Caprella equilibra</i> Say, 1818	B					
<i>Caprella</i> sp. 1	B					
<i>Caprella</i> sp. 2	P					
<i>Stenothoe</i> sp.	B					
<i>Probolisca</i> sp.	S					
<i>Dulichella</i> sp.	S					
<i>Leucothoe</i> sp.	S					
cf. <i>Polycheria</i> sp.	S					
<i>Jassa marmorata</i> Holmes, 1905	S					
<i>Jassa</i> sp.	P					
<i>Erikus</i> sp.	B		P			
<i>Ampithoe</i> sp.	P					
<i>Austroregia huxleyana</i> (Bate, 1862)		S				
<i>Ultimachelium barnardi</i> (Alonso de Pina, 1993)			S			
<i>Liljeborgia octodentata</i> Schellenberg, 1931			S			
<i>Paramoera</i> sp.				S		P
<i>Atyloella dentata</i> K.H. Barnard, 1932						S
cf. <i>Lembos</i> sp.	B					
<i>Cymodoce</i> cf. <i>bentonica</i>		S				
<i>Exosphaeroma lanceolatum</i> (White, 1843)			B			S
<i>Exosphaeroma studeri</i> Vanhöffen, 1914			B	P	S	
<i>Ischyromene eatoni</i> (Miers, 1875)			S			
<i>lais pubescens</i> (Dana, 1852)			B			
<b>Phylum Mollusca</b> (Bivalvia: D Zelaya (UBA) and E. Schwindt, Gastropoda: D Zelaya and F Scarabino, DINARA, Polyplacophora: MP Raffo and D Zelaya)						
<i>Aulacomya atra</i> (Molina, 1782)		B	S	S	B	B
<i>Brachidontes purpuratus</i> (Lamarck, 1819)				S		



<i>Musculus viator</i> (d'Orbigny, 1846)	B						
<i>Mytilus</i> spp.		S			B	B	B
<i>Hiatella meridionalis</i> (d'Orbigny, 1846)							B
<i>Hiatella</i> sp.							S
<i>Entodesma patagonicum</i> (d'Orbigny, 1846)	S						
<i>Ostrea puelchana</i> d'Orbigny, 1842	B						
<i>Ostrea stentina</i> Payraudeau, 1826	S						
<i>Sphenia hatcheri</i> Pilsbry, 1899							B
<i>Bostrycapulus odites</i> Collin, 2005	B						
<i>Crepidatella</i> cf. <i>dilatata</i>				S			
<i>Crepidatella dilatata</i> (Lamarck, 1822)		B					
<i>Crepidula</i> sp.	B						
<i>Fissurella oriens</i> Sowerby, 1835							S
<i>Fissurella picta</i> (Gmelin, 1791)							S
<i>Fissurella radiosa radiosa</i> Lesson, 1831			S				
<i>Fissurellidea patagonica</i> (Strebel, 1907)				S			
<i>Margarella violacea</i> (King & Broderip, 1832)				S			
<i>Tegula patagonica</i> (d'Orbigny, 1835)			B				
<i>Costoanachis sertulariarum</i> (d'Orbigny, 1839)	P						
<i>Parvanachis paessleri</i> (Strebel, 1905)	P						
<i>Lachesis</i> (?) <i>euthrioides</i> Melvill & Standen, 1898				P			
<i>Pareuthria plumbea</i> (Philippi, 1844)				S			S
<i>Photinastoma taeniata</i> (Wood, 1828)				B			
<i>Trophon geversianus</i> (Pallas, 1774)					S	B	S
<i>Xymenopsis muriciformis</i> (King, 1832)							S
<i>Acteon biplicatus</i> (Strebel, 1908)				S	P		
<i>Odostomia</i> sp.							S
<i>Spurilla</i> sp.	P						
<i>Berghia rissodominguezi</i> Muniaín & Ortea, 1999		S					
<i>Callochiton puniceus</i> (Couthouy MS, Gould, 1846)							P
<i>Chaetopleura isabellei</i> (d'Orbigny, 1841)	S						
<i>Plaxiphora aurata</i> (Spalowsky, 1795)							S
<b>Phylum Entoprocta</b> (J López Gappa)							
<i>Pedicellina</i> sp.							B
<b>Phylum Bryozoa</b> (J López Gappa)							
<i>Alcyonidium australe</i> d'Hondt & Moyano, 1979						S	B
<i>Alcyonidium</i> sp.						S	
<i>Beania costata</i> (Busk, 1876)				S			
<i>Beania magellanica</i> (Busk, 1852)				B			B
<i>Bugula stolonifera</i> Ryland, 1960	P						
<i>Cellaria malvinensis</i> (Busk, 1852)				B			
<i>Celleporella hyalina</i> s.l.				S			
<i>Chaperiopsis galeata</i> (Busk, 1854).							B
<i>Conopeum reticulum</i> (Linnaeus, 1767)				S			
<i>Electra</i> sp.						S	S
<i>Fenestrulina</i> sp.	S	S	B				
<i>Membranipora isabelleana</i> (d'Orbigny, 1847)		S					
<i>Menipea patagonica</i> Busk, 1852		S					B
<i>Tricellaria aculeata</i> (d'Orbigny, 1847)			B				
<i>Disporella</i> sp.							S
<i>Metroperiella galeata</i> (Busk, 1854)							P
<i>Smittoidea</i> sp.	S						
<b>Phylum Echinodermata</b> (Asteroidea: T Rubilar,							

Ophiuroidea: M Brögger, UBA, Echinoidea: MP Raffo)						
<i>Allostichaster capensis</i> (Perrier, 1875)			S			
<i>Anasterias antarctica</i> (Lütken, 1857)			S			
<i>Diplodontias singularis</i> (Müller & Troschel, 1843)						S
<i>Ophiactis asperula</i> (Philippi, 1858)						B
<i>Amphipholis squamata</i> (Delle Chiaje, 1828)	P					
<i>Ophioplocus januarii</i> (Lütken, 1856)	P					
<i>Arbacia dufresnii</i> (Blainville, 1825)			B			
<i>Pseudechinus magellanicus</i> (Philippi, 1857)						B
<b>PHYLUM CHORDATA</b> (Ascidacea: M Tatián, C Lager, Osteichtheys: A Gosztanyi, CENPAT)						
<i>Aplidium meridianum</i> (Sluiter, 1906)						S
<i>Aplidium variabile</i> (Herdman, 1886)			S			B
<i>Ascidella aspersa</i> (Müller, 1776)	B	B	B			
<i>Cnemidocarpa robinsoni</i> Hartmeyer, 1916	B	B	B			P
<i>Polyzoa opuntia</i> Lesson, 1830			B			P
<i>Styela paessleri</i> (Michaelson, 1898)			S			P
<i>Ciona intestinalis</i> (Linnaeus, 1767)	P	P				
<i>Corella eumyota</i> Traustedt, 1882			B			
<i>Diplosoma listerianum</i> (Milne-Edwards, 1841)	P		P			
<i>Lissoclinum fragile</i> (Van Name, 1902)	B					
<i>Eudistoma platense</i> Van Name, 1945	P					
<i>Molgula manhattensis</i> (De Kay, 1843)	P		P			
<i>Paramolgula gregaria</i> (Lesson, 1830)	S		B	B	B	B
<i>Pyura legumen</i> (Lesson, 1830)						S
<i>Sycozoa gaimardi</i> (Herdman, 1886)						P
<i>Sycozoa sigillinoides</i> Lesson, 1830			B			
<i>Patagonotothen squamiceps</i> (Peters, 1877)						S
<i>Patagonotothen sima</i> (Richardson, 1845)						S
<i>Patagonotothen cornucola</i> (Richardson, 1844)						S
<b>Total number of taxa observed</b>	<b>92</b>	<b>43</b>	<b>85</b>	<b>38</b>	<b>32</b>	<b>80</b>

**APPENDIX B.****Table B.1.** List of variables studied at each port and the source of the information used.

<b>Main Variable</b>	<b>Specific Variable</b>	<b>Period recorded and Source</b>
Sea Surface Water Temperature (°C)	Annual mean (WTAM), maximum mean (WTMaxM), minimum mean (WTMinM), maximum at the hottest time of the summer season (WTMaxHS), mean during summer season (WTMS), minimum at the coldest time of the winter season (WTMinCW), mean during winter season (WTMW)	Servicio de Hidrografía Naval, Argentina (historical data from permanent oceanographic stations at the ports). For the port of San Antonio Este and Punta Quilla data were obtained from AVHRR Pathfinder, NOAA-NASA (period 1993-2003)
Air Temperature (°C)	Annual mean (ATAM), annual maximum mean (ATAMaxM), annual minimum mean (ATAMinM), mean of the maximum in summer season (ATMaxMS), mean of the minimum in winter season (ATMinMW)	Servicio Meteorológico Nacional 1981, 1986 (period 1961-1980). Data from Puerto Madryn obtained from Laboratorio de Datos CENPAT-CONICET (1982-2002)
Tidal Amplitude (m)	Mean (TAM), maximum in spring tides (TAMaxS), minimum in neap tides (TAMinN), mean with spring tides (TAMS), mean with neap tides (TAMN)	Charts of the Servicio de Hidrografía Naval, Argentina
Wind Speed (km/h)	Annual mean (WSAM)	Published data of the Servicio Meteorológico Nacional 1981, 1986 (period 1961-1980). Data from

		Puerto Madryn obtained from Laboratorio de Datos CENPAT-CONICET (1982-2002)
Superficial Salinity	Annual mean (SAM)	Tapella et al. (2002 for Ushuaia), Piola 2007, field surveys were performed in Punta Quilla, Río Gallegos and Puerto Deseado
Rainfall (mm)	Mean monthly (RMM), total Annual (RTA), total in the port's driest 6 months season (RTD), total in the port's wettest 6 months season (RTW)	Published data of the Servicio Meteorológico Nacional 1981, 1986 (period 1961-1980). Data from Puerto Madryn obtained from Laboratorio de Datos CENPAT-CONICET (1982-2002)
Port Depth (m)	Mean (De)	Consejo Portuario Argentino (2011)
Environmental impact of the city	This variable was categorized in high, medium and low considering the coastal geography, the oceanographic and fluvial conditions, the ecosystem disturbance, the pollution, and the eutrophication recorded at each port area (EIC)	Esteves (2007)
Port Type	Classification was based following Clarke et al (2004) in natural bay, breakwater port, tidal creek, and estuary (HT)	Hydrographic charts, Consejo Portuario Argentino (2011)

**Table B.2.** Spearman rank order correlation matrix for the Sea Surface Water Temperature (1: WTAM, 2: WTMaxM, 3: WTMinM, 4: WTMaxHS, 5: WTMS, 6: WTMinCW, 7: WTMW), Air Temperature (8: ATAM, 9: ATAMaxM, 10: ATAMinM, 11: ATMaxMS, 12: ATMinMW), Tidal Amplitude (13: TAM, 14: TAMaxS, 15: TAMinN, 16: TAMS, 17: TAMN), Wind Speed (18: WSAM), Superficial Salinity (19: SAM), Rainfall (20: RMM, 21: RTA, 22: RTD, 23: RTW), Depth (24: De), Environmental Impact of the City (25: EIC) and Port Type (26: HT). Abbreviations are the same as in Table 1. Values in italics within the grey cells show the significant results ( $p < 0.05$ ).

	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	
1	<b>0.98</b>	<b>0.99</b>	0.76	<b>0.98</b>	<b>0.96</b>	<b>0.97</b>	<b>0.99</b>	<b>0.97</b>	<b>0.97</b>	<b>0.93</b>	0.75	0.09	0	0.47	0.03	-0.17	-0.42	0.68	-0.46	-0.47	-0.38	-0.39	<b>0.84</b>	0.49	-0.57	
2		<b>0.95</b>	0.81	<b>0.93</b>	<b>0.93</b>	<b>0.98</b>	<b>0.94</b>	<b>0.91</b>	<b>0.91</b>	<b>0.9</b>	0.66	-0.04	-0.12	0.45	-0.1	-0.29	-0.58	0.7	-0.29	-0.32	-0.21	-0.23	<b>0.87</b>	0.63	-0.67	
3			0.77	<b>0.99</b>	<b>0.95</b>	<b>0.93</b>	<b>1</b>	<b>0.99</b>	<b>0.97</b>	<b>0.95</b>	0.75	0.19	0.11	0.43	0.13	-0.05	-0.3	0.67	-0.58	-0.59	-0.51	-0.51	0.78	0.39	-0.45	
4				0.8	0.6	0.68	0.73	0.69	0.61	<b>0.86</b>	0.17	-0.11	-0.16	0.26	-0.14	-0.24	-0.59	<b>0.81</b>	-0.28	-0.31	-0.2	-0.13	0.59	0.7	-0.43	
5					<b>0.92</b>	<b>0.89</b>	<b>0.98</b>	<b>0.98</b>	<b>0.95</b>	<b>0.97</b>	0.69	0.21	0.13	0.48	0.16	-0.02	-0.32	0.73	-0.59	-0.57	-0.51	-0.49	0.71	0.43	-0.4	
6						<b>0.95</b>	<b>0.95</b>	<b>0.96</b>	<b>0.95</b>	<b>0.81</b>	<b>0.87</b>	0.26	0.17	0.34	0.19	-0.02	-0.3	0.47	-0.48	-0.52	-0.42	-0.49	0.8	0.32	-0.48	
7							<b>0.94</b>	<b>0.92</b>	<b>0.93</b>	<b>0.83</b>	0.78	-0.05	-0.13	0.43	-0.11	-0.32	-0.51	0.57	-0.3	-0.33	-0.22	-0.26	<b>0.93</b>	0.52	-0.71	
8								<b>0.99</b>	<b>0.98</b>	<b>0.93</b>	0.79	0.16	0.08	0.46	0.1	-0.08	-0.28	0.65	-0.58	-0.57	-0.5	-0.5	0.81	0.37	-0.48	
9									<b>0.97</b>	<b>0.91</b>	<b>0.82</b>	0.26	0.18	0.4	0.21	0.02	-0.2	0.59	-0.64	-0.65	-0.58	-0.59	0.76	0.29	-0.39	
10										<b>0.9</b>	<b>0.84</b>	0.17	0.08	0.57	0.11	-0.09	-0.26	0.63	-0.54	-0.51	-0.46	-0.47	0.78	0.35	-0.49	
11											0.53	0.09	0.02	0.57	0.04	-0.11	-0.39	<b>0.86</b>	-0.52	-0.47	-0.43	-0.37	0.66	0.54	-0.41	
12												0.33	0.26	0.28	0.28	-0.08	0.07	0.11	-0.54	-0.56	-0.51	-0.61	0.7	-0.09	-0.34	
13													<b>1</b>	-0.26	<b>1</b>	<b>0.96</b>	0.63	-0.22	-0.7	-0.71	-0.77	-0.81	-0.32	-0.59	0.7	
14														-0.31	<b>1</b>	<b>0.98</b>	0.67	-0.27	-0.67	-0.68	-0.75	-0.78	-0.4	-0.64	0.76	
15																-0.29	-0.39	-0.37	0.74	-0.01	0.18	0.08	0.16	0.33	0.5	-0.45
16																	<b>0.97</b>	0.66	-0.25	-0.69	-0.7	-0.76	-0.8	-0.38	-0.63	0.75
17																		0.75	-0.33	-0.63	-0.64	-0.72	-0.73	-0.55	-0.71	<b>0.88</b>
18																			-0.57	-0.56	-0.53	-0.63	-0.63	-0.54	<b>-0.97</b>	0.81
19																				-0.18	-0.07	-0.08	0.04	0.43	0.75	-0.43
20																					<b>0.97</b>	<b>0.99</b>	<b>0.96</b>	-0.12	0.42	-0.39

21		<b>0.98</b>	<b>0.98</b>	-0.18	0.43	-0.36
22			<b>0.98</b>	-0.03	0.51	-0.48
23				-0.09	0.55	-0.44
24					0.52	<b>-0.85</b>
25						-0.75

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**Table B.3.** Spearman rank order correlation matrix reduced for the environmental variables of the ports being Te: temperature, Ti: tidal amplitude, Wi: wind speed, S: salinity, Ra: rainfall, De: depth and PT: port type. Significant results are shown within the grey cells ( $p < 0.05$ ).

Parameters	Ti	Wi	S	Ra	De	PT
<b>Te</b>	0.09	-0.42	0.68	-0.46	0.84	-0.57
<b>Ti</b>		0.63	-0.22	-0.7	-0.32	0.7
<b>Wi</b>			-0.57	-0.56	-0.54	0.81
<b>S</b>				-0.18	0.43	-0.43
<b>Ra</b>					-0.12	-0.39
<b>De</b>						-0.85

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