

SERPENTINE GEOECOLOGY OF EASTERN NORTH AMERICA: A REVIEW

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ABSTRACT. Serpentine outcrops are model habitats for geoeological studies. While much attention has been paid to serpentine outcrops worldwide, the literature on eastern North American serpentine and associated biota is scant. This review examines the available literature, published and unpublished, on geoeological studies conducted on serpentine in eastern North America, from Newfoundland through Québec and New England south to Alabama. Most serpentine outcrops in the region have been mapped, but there have been few intensive mineralogical and pedological investigations. The limited soil analyses available suggest elevated levels of heavy metals such as Ni, near-neutral pH values, and Ca:Mg ratios < 1 , characteristic of serpentine soils worldwide. Botanical studies to date have largely focused on floristic surveys and the influence of fire exclusion and grazing on indigenous vegetation. To date, 751 taxa of vascular plants belonging to 92 families have been reported from serpentine outcrops in the region. Two taxa, *Agalinis acuta* and *Schwalbea americana*, are federally endangered in the United States while many others are listed as rare, endangered, or imperiled in one or more states or provinces. Globally, six species, *Adiantum viridimontanum*, *Minuartia marcescens*, *Pycnanthemum torrei*, *S. americana*, *Scirpus longii*, and *Symphyotrichum depauperatum* are listed as imperiled (G2) while one species, *Agalinis acuta*, is listed as critically imperiled (G1). *Cerastium velutinum* var. *villosissimum* is the only recognized serpentine endemic plant for eastern North America while *Adiantum viridimontanum*, *Aspidotis densa*, *M. marcescens*, and *S. depauperatum* are largely restricted to the substrate. Based on current distributions, we propose that *A. viridimontanum* and *M. marcescens* be considered endemic to serpentine substrates in eastern North America. Studies on cryptogams list 165 species of lichens and 146 species of bryophytes for the region. None of the species found appear to be restricted to the substrate. Compared to other regions of the world, ecophysiological and evolutionary investigations are scant. Biosystematic investigations are restricted to the taxa *Adiantum aleuticum*, *C. velutinum* var.

villosissimum, and *S. depauperatum*. Plant-soil relations, especially the capacity to hyperaccumulate metals such as Ni and the ecological consequences of metal accumulation, are also under explored. One report from eastern Canada lists *Arenaria humifusa*, *M. marcescens*, *Packera paupercula*, and *Solidago hispida* as hyperaccumulating Ni although the findings have yet to be confirmed by subsequent investigations. Overall, serpentine geoecology in eastern North America remains largely unexplored.

Key Words: Ca:Mg, edaphic endemism, geobotany, heavy metal-hyperaccumulation, nickel, rare plants, serpentine soil, serpentine endemism, ultramafic ecology

Within a given climatic regime, geology plays a central role in the distribution and ecology of plant species and their associated biota (Jenny 1941, 1980). The most significant causes of localized or unusual plant distributions are discontinuities in geology and edaphics—geodaphics (Kruckeberg 1986). Extreme edaphic conditions as often seen on limestone (Lloyd and Mitchell 1973; Lousley 1950; Shimizu 1962, 1963), gypsum (Turner and Powell 1979), dolomite (Kruckeberg 2002; Lloyd and Mitchell 1973), granite (Ornduff 1986; Walters and Wyatt 1982; Wyatt and Fowler 1977), guano deposits (Gillham 1956; Ornduff 1965; Vasey 1985), vernal pools (Holland and Jain 1977, 1981), salt marshes (Flowers et al. 1986), and even mine tailings (Antonovics et al. 1971; Shaw 1990a), provide ideal settings for examining the role of the edaphic factor in the distribution and ecology of plants and their associated biota.

Serpentine outcrops have long provided model habitats for the study of geobotany worldwide (Brooks 1987; Kruckeberg 2002; Roberts and Proctor 1992). The word *serpentine* is applied in a general sense to describe soils rich in iron magnesium silicates derived from a range of ultramafic rocks (Coleman and Jove 1992; Wyllie 1979a). Serpentine more accurately refers to a group of hydrous magnesium phyllosilicate minerals, including antigorite, chrysotile and lizardite, in hydrothermally altered ultramafic rocks (Brooks 1987; Kruckeberg 1984). Nevertheless, researchers worldwide, us included, use the term *serpentine* loosely to describe rocks, soils, vegetation, and other biota associated with ultramafic outcrops.

Soils derived from serpentine outcrops provide harsh conditions for plant growth (Brady et al. 2005). These soils generally have a near-neutral pH, are high in metals such as Ni, Co, and Cr, and low in many essential nutrients such as P, K, and Mo (Kruckeberg 1984; Walker 2001). Although serpentine soils have often been considered

to be poor in N (Kruckeberg 2002), this generally applies only to serpentine barrens with little or no vegetation (Alexander et al. 2007). Calcium:magnesium ratios are generally < 1 , which are unfavorable for plant growth (Bradshaw 2005; Brady et al. 2005; Skinner 2005). Although physical features of serpentine soils can vary considerably from site to site (Alexander et al. 2007) and within a site (Rajakaruna and Bohm 1999), serpentine outcrops are often found in open, steep landscapes with soils that are generally shallow and rocky with a reduced capacity for moisture retention (Kruckeberg 2002). See Alexander et al. (2007) for a broader and more balanced perspective on available-water capacity in serpentine soils across more diverse landscapes. Given the extreme nature of these soils, their biota is often uniquely adapted and frequently restricted to such habitats.

Serpentine outcrops worldwide are known to harbor high numbers of endemic plant species with rates of endemism generally increasing toward the equator (Alexander et al. 2007; Brooks 1987; Kruckeberg 2002). Of the 1410 species endemic to California, 176 (12.5%) are restricted to serpentine (Hickman 1993). This is a remarkably high number given only 670 taxa are associated with serpentine soils in California, a substrate covering less than 1.5% of the state (Safford et al. 2005). Thus, it is no surprise that serpentine floras are well studied in California and other parts of western North America (Alexander et al. 2007; Harrison and Viers 2007; Kruckeberg 1984, 1992; Safford et al. 2005), not only for their taxonomic value, but also for their usefulness in testing ecological (Harrison, Davies, Grace, Safford, and Viers 2006; Harrison, Davies, Safford, and Viers 2006; Harrison, Safford, Grace, Viers, and Davies 2006; Safford and Harrison 2004) and evolutionary scenarios (Baldwin 2005; Bradshaw 2005; Patterson and Givnish 2004; Rajakaruna and Whitton 2004; Rajakaruna et al. 2003; Wright and Stanton 2007; Wright et al. 2006). The tropical islands of New Caledonia and Cuba also provide remarkable cases of serpentine endemism (Boyd et al. 2004; Brooks 1987; Kruckeberg 2002). In New Caledonia, 3178 taxa, approximately 50% of the native flora, are endemic to serpentine soils (Jaffré 1992). In Cuba, 920 species, approximately one-third of the taxa endemic to Cuba, have developed solely on serpentine soils (Borhidi 1992). Similar restrictions and notable floristic associations are also found in serpentine areas of the Mediterranean, Africa, Australia-New Zealand, and Asia (Baker et al. 1992; Balkwill 2001; Boyd et al. 2004; Brooks 1987; Chiarucci and Baker 2007; Jaffré et al. 1997).

Little attention has been paid to biota on serpentine outcrops in eastern North America despite its patchy occurrence along the Appalachian orogen, spreading south from Newfoundland and Québec to New Brunswick, Maine, Vermont, New York, Pennsylvania, New Jersey, Delaware, Maryland, Virginia, North Carolina, South Carolina, Georgia, and Alabama (Figure 1A, B). Although these outcrops have attracted some attention, resulting in unpublished theses (Carter 1979; Sirois 1984), conference proceedings (Roberts 1992), book chapters (Brooks 1987; Sirois and Grandtner 1992; Tyndall and Hull 1999), and numerous papers in regional journals, no extensive review has appeared to date for the region. An account of the natural history of serpentine outcrops in eastern North America is found in a book by Dann (1988). Similarly, Reed (1986) has provided an extensive treatment of the floras of the serpentine outcrops in eastern North America.

This review is the first attempt to highlight the geocology of serpentine outcrops of eastern North America, focusing on all unpublished and published reports we have been able to locate from the early 1900s to present. It is our hope that this review will generate renewed interest in the under-explored serpentine outcrops of the region and lead to much-needed geobotanical and geoecological investigations and conservation efforts.

GEOLOGY AND SOILS OF SERPENTINE OUTCROPS OF EASTERN NORTH AMERICA

The sharply demarcated boundaries of vegetation commonly found with geologic boundaries between serpentine and other substrates are nothing less than striking (Figure 2). This remarkable biological phenomenon seen at a local geographic scale is controlled primarily by geology and is a fitting reminder that the “plant world exists by geologic consent” (Kruckeberg 2002, p. ix).

Ultramafic rocks are chemically similar to the mantle of Earth (MacGregor 1979). The chemistry of the mantle is drastically different from that of the continental crust, the mantle being primarily composed of Mg-silicates whereas Ca is more abundant than Mg in the crust (Alexander et al. 2007). Relatively few plants are adapted to soils derived from rocks of the mantle (Brooks 1987); however, upon adaptation they often become specialized and restricted to such soils (Kruckeberg 1986; Rajakaruna and Boyd 2008; Rajakaruna and Whitton 2004). Most of the serpentine found

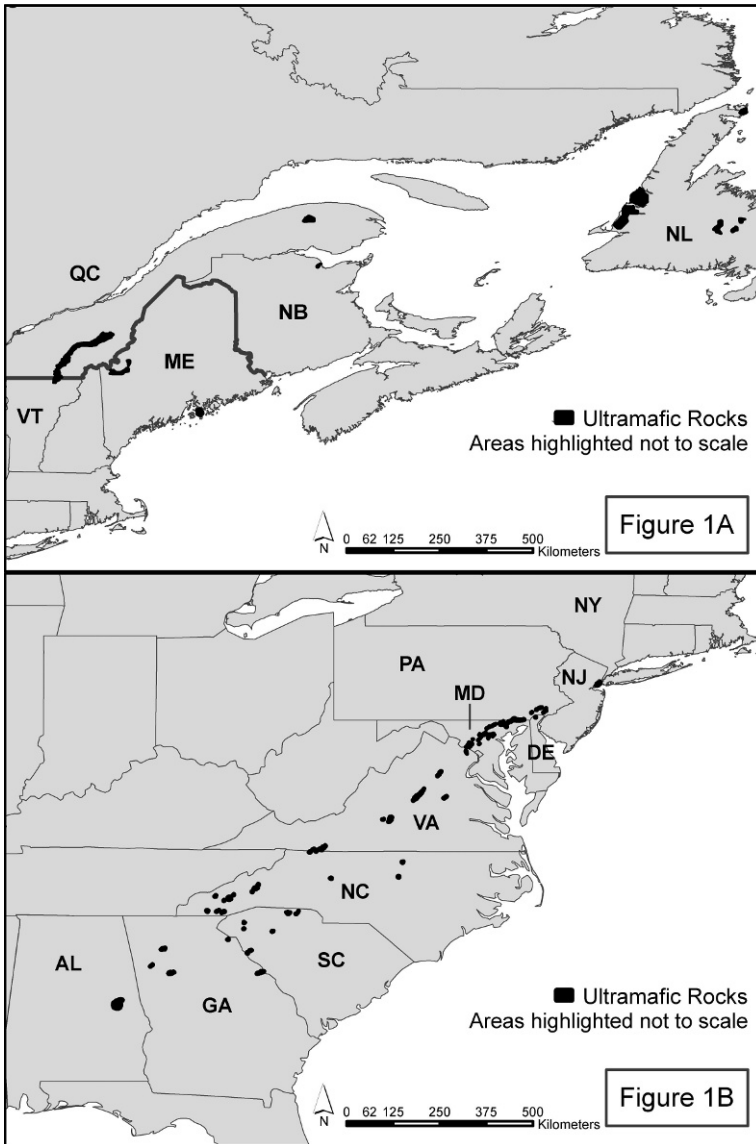


Figure 1. Locations of ultramafic rock exposures in the northern (A) and southern (B) Appalachian regions. Localities based on Larrabee (1966) and E. B. Alexander (unpubl. data). Credit: Jose Perez-Orozco and Apoorv Gehlot.



Figure 2. A sharply-demarcated floristic boundary under the control of an edaphic boundary at Jasper Ridge Biological Preserve, San Mateo County, California. The yellow-flowered *Lasthenia californica* (Asteraceae; in background) is restricted to serpentine soils. The boundary between *L. californica* and grasses is defined by a serpentine-sandstone transition. Credit: Bruce A. Bohm.

in eastern North America originated in the formation of the ocean floor. Mantle rocks are produced along ocean ridges where partial melting in the upper mantle produces reservoirs of gabbro melt, or magma, that feed basalt lava flows from ocean ridges. As fresh melt is added, lava spreads from the ridges where it cools to form rocks (Wyllie 1979b). The succession of rocks produced from this melting and cooling, from bottom to top, is peridotite (modified mantle), gabbro, and basalt—a sequence called ophiolite. Most ophiolite is subducted, sinking back into the mantle, but some is incorporated into continental crust (Coleman and Jove 1992; Wyllie 1979b).

The ultramafic outcrops of eastern North America have an interesting history that spans millions of years (Roberts 1996; Williams and Hatcher 1982, 1983; Williams and Talkington 1977). The fragmentation of the super-continent Rhodina, formed during the Proterozoic Era over 0.5 Ga ago, left an ocean off the coast of a large continental mass from which North America would eventually emerge. Volcanic deposits, ophiolites, and sediments that accumulated in this ocean were pushed against the margin of the precursor

of the North American continent several times during the Paleozoic Era, from about 500 to 250 Ma ago; each time some of the foreign materials were accreted onto the continent. Accreted peridotites have been altered to various degrees, with the most completely altered rocks becoming serpentinite, composed of the serpentine group of minerals (antigorite, chrysotile, and lizardite). Chemically, serpentinite is similar to peridotite (olivine and pyroxene minerals), except for the addition of water, which increases its volume by about one third. Because serpentinite is so chemically similar to peridotite, most plants do not discriminate between soils derived from these two rocks.

Paleozoic accreted terranes were added to the North American continent as far inland as the Brevard Zone in the Blue Ridge Mountains of the southern Appalachians and to the Baie Verte-Brompton (BV-B) line in the northern Appalachians (Williams and St. Julien 1982). Some foreign, or allochthonous, rocks (for example, the Hare Bay allochthon in Newfoundland) were pushed beyond the Brevard Zone and the BV-B line and deposited over rocks that had formed on the preaccretionary Proterozoic continent. The most well known of these is the Bay of Islands ophiolite in western Newfoundland. It is one of the most complete ophiolites exposed anywhere in the world (Coleman and Jove 1992). Ophiolites, with some ultramafic rocks in them, cover about 3% (318,000 ha) of Newfoundland's 106,000 km² (Roberts 1992), where they are more concentrated than in any other Appalachian area (Figure 1A). Besides in the ophiolites of western and northern Newfoundland and the Baltimore complex of the mid-Appalachian orogeny in Maryland and southeastern Pennsylvania, serpentine rocks are most concentrated along the BV-B line in the northern Appalachians, along the Brevard Zone from Alabama to North Carolina, and in the Albemarle-Nelson Soap-Stone Belt in Virginia, with small scattered serpentine exposures southeast of these lineaments and in the Blue Ridge Mountains. The ophiolite of Boil Mountain in western Maine (Caldwell 1998; Figure 1A) is a prominent exposure southeast of the BV-B line.

Unlike serpentine regions of western North America, serpentine areas of the northern Appalachian region south to Long Island were covered by ice during the Pleistocene glaciation (Dyke 2004). There may have been some unglaciated nunataks on Newfoundland and the Gaspé Peninsula. For some time after the Laurentide continental glacier had melted, icecaps persisted on Newfoundland and the Gaspé

Peninsula. Ice cover slowly disappeared from the Gaspé Peninsula between 10 and 13.5 ka ago (Dyke 2004). As such, the length of exposure to the forces of weathering and soil formation for these outcrops has been short in comparison to outcrops from unglaciated regions such as the Piedmont, south of the limit of glaciation in central New Jersey and eastern Pennsylvania.

Chemically, serpentine rocks differ from other rocks in having very high Mg and low Ca, K, and P levels. They have relatively high concentrations of elements 23 through 28 (V, Cr, Mn, Fe, Co, and Ni; Coles 1979). It is generally believed that the low ratio of exchangeable Ca:Mg found in soils derived from serpentine rock is the most limiting factor for plant growth, although high Ni levels in some serpentine soils may be toxic to many plants (Brady et al. 2005). Exchangeable Ca:Mg ratios are considerably >1 in most soils but are < 1 (mostly < 0.7) in serpentine soils and much lower in serpentine subsoils (Alexander et al. 2007; Brooks 1987).

The serpentine soils of the northern Appalachians are weakly developed, namely Entisols, Inceptisols, and Mollisols (Soil Survey Staff 1999), or Regosols, Brunisols, and Gleysols, respectively, in Canada (Roberts 1980, 1992; Sirois and Grandtner 1992). There are also several occurrences of poorly-drained serpentine organic soils in Canada termed Humisols (or Terric Cryosaprists in Soil Taxonomy; Soil Survey Staff 1999). Serpentine soils of the southern Appalachian orogen, beyond the limit of the Pleistocene glaciation, commonly have subsoil clay accumulations referred to as argillic horizons (Ogg and Smith 1993; Rabenhorst et al. 1982). These are Alfisols, or, where intensively leached, Ultisols. Physically the serpentine soils in the northern Appalachian region are similar to those of nonserpentine soils in the region, except that none of the serpentine soils have the bleached E horizons that form in Spodosols, which are more acidic than the serpentine soils. Although many serpentine soils south of the continental glacial limit have argillic horizons, they are lacking in serpentine soils north of there.

Serpentine soils cover a considerable extent of eastern Canada. Approximately 318,000 ha of ophiolite is found in Newfoundland (Roberts 1992), including 7700 ha with serpentine soils in the western part of the province (Roberts 1980). Québec's Mt. Albert region in the Gaspésie consists of at least 6400 ha of ultramafics (Sirois et al. 1988; L. Sirois, pers. comm.) while the Eastern Townships have approximately 250,000 ha (Brooks 1987), contributing to the majority of the serpentine in the province. Estimates by

E. B. Alexander from soil surveys in the Appalachian region of the United States suggest that there are approximately 6900 ha in Pennsylvania, 23,600 ha in Maryland, and at least 243 ha of serpentine soils in North Carolina. Although New Brunswick, Maine, Vermont, New York, New Jersey, Delaware, Virginia, South Carolina, Georgia, and Alabama also have small but appreciable amounts of serpentine (Brooks 1987; Reed 1986; Tyndall and Hull 1999), the total for the eastern United States is probably on the order of 40,000 ha. In comparison, California has 600,000 ha of serpentine (Safford et al. 2005), although estimates by E. B. Alexander based on soil surveys point to about half this area for both California and southwestern Oregon.

Serpentine is clearly more extensive in accreted terranes along the western coast of North America, and the greater variety of climates found there has contributed to a greater variety of soils (Alexander et al. 2007). This wide range of soils derived from ultramafic rocks has led to a remarkable array of plant species endemic to this region (Alexander et al. 2007; Kruckeberg 1984, 2002). The range of serpentine soils in eastern North America, from very cold Entisols and Histosols to cold Inceptisols, cool Alfisols, and warm Ultisols, has similarly given rise to a unique flora and associated biota worthy of appreciation, study, and conservation. Table 1 compares the pH, exchangeable Ca:Mg ratio, and Ni concentration of serpentine soils from Jasper Ridge Biological Preserve, San Mateo County, California (Rajakaruna and Bohm 1999) with those reported from serpentine soils in Newfoundland (Roberts 1992), Maine (Briscoe et al. 2009), New York (Parisio 1981), Maryland (Cumming and Kelly 2007), Pennsylvania (Miller and Cumming 2000), and Buck Creek, North Carolina (Mansberg and Wentworth 1984). While the geographically separated outcrops differ in their length of exposure to the forces of weathering and soil formation, soils collected 0–10 or 15 cm from the surface were surprisingly similar with respect to exchangeable Ca:Mg ratio (Table 1; all < 1, except in organic Histic epipedons in Newfoundland). Table 1 also shows that when compared to adjacent non-serpentine soils, the serpentine soils, regardless of geographical origin, had generally higher values of exchangeable Ni (8.5–116 $\mu\text{g g}^{-1}$ dry soil for serpentine vs. 0.13–0.4 $\mu\text{g g}^{-1}$ dry soil for nonserpentine) and pH (6.2–6.74 for serpentine vs. 4.2–5.25 for nonserpentine), and lower values of exchangeable Ca:Mg (0.025–0.56 for inorganic serpentine layers vs. 1.55–4.34 for nonserpentine).

Table 1. Soil surface chemical analyses for serpentine soil samples (S) collected from Jasper Ridge, CA (Rajakaruna and Bohm 1999); Soldiers Delight, MD (Cumming and Kelly 2007); Newfoundland, Canada (Roberts 1992); the Deer Isles, ME (Briscoe et al. 2009); Conowingo Barrens, PA (Miller and Cumming 2000); Staten Island, NY (Parisio 1981); and Buck Creek, NC (Mansberg and Wentworth 1984) along with values for adjacent nonserpentine soil samples (NS). Some of the Newfoundland soils have surface organic layers > 20 cm thick (Histic epipedons); these are listed separately from soils lacking thick surface organic layers (inorganic epipedons). Nickel was extracted by several different methods [DTPA, exchangeable (neutral ammonium acetate), Mehlich-3, and total dissolution with concentrated acids], some of which are not comparable. Values listed are means \pm standard errors.

Site	pH (soil water)	Exchangeable	Ni $\mu\text{g g}^{-1}$
		Ca:Mg	
Jasper Ridge, CA (S) (n = 123)	6.74 \pm 0.03	0.21 \pm 0.01	116.45 \pm 3.95 DTPA
Soldier's Delight, MD (S) (n = 3)	6.2 \pm 0.1	0.56 \pm 0.12	66.1 \pm 6.6 Mehlich-3
Newfoundland, Canada (S) (organic layer, n = 5)	6.1 \pm 0.6	1.39 \pm 0.69	21.9 \pm 8.1 exchangeable 842 \pm 294 total
Newfoundland, Canada (S) (inorganic layer, n = 6)	6.7 \pm 0.3	0.025 \pm 0.008	20.8 \pm 23.0 exchangeable 2949 \pm 512 total
Little Deer Isle, ME (S) (n = 18)	6.5 \pm 0.2	0.45 \pm 0.05	44.8 \pm 5.6 DTPA
Conowingo Barrens, PA (S) (n = 4)	6.6 \pm 0.1	0.17 \pm 0.01	21 \pm 6 Mehlich-3
Staten Island, NY (S) (n = 1)	6.9	0.17	8.5 exchangeable
Buck Creek, NC (S) (n = 11)	6.1 \pm 0.3	0.09 \pm 0.03	–
Deer Isle, ME (NS) (n = 15)	5.25 \pm 0.08	4.34 \pm 0.4	0.13 \pm 0.02 DTPA
Conowingo Barrens, PA (NS) (n = 3)	4.2 \pm 0.1	2 \pm 0.01	0.4 \pm 0.3 Mehlich-3
Buck Creek, NC (NS) (n = 5)	4.7 \pm 0.3	1.55 \pm 0.44	–

PLANT LIFE ON SERPENTINE OUTCROPS OF EASTERN NORTH AMERICA

Although few in comparison to other regions of the world, there are several descriptive and experimental studies highlighting aspects of the floristic associations and ecological relations of serpentine

outcrops in eastern North America. These studies include floristic surveys of lichens, bryophytes, algae, microbes, and vascular plants; ecophysiological studies focusing on plant-heavy metal relations; evolutionary studies focusing on cross-kingdom interactions and the role of edaphics in plant speciation; ecological studies concentrating on plant adaptation; and applied ecological studies exploring avenues for the remediation of previously mined serpentine habitats and the implications of fire, grazing, and land-use practices on the maintenance of native vegetation.

STUDIES ON LICHENS, BRYOPHYTES, ALGAE, MICROBES

The intimate and often inseparable relationship between cryptogams, such as lichens, and their substrates suggests a strong possibility of substrate effects for such species associated with extreme geodaphic habitats (Brodo 1974). Worldwide, however, there have been relatively few studies of lichens on serpentine soils (Favero-Longo et al. 2004), and only two published studies have examined lichens on serpentine soils in eastern North America (Harris et al. 2007; Sirois et al. 1988). Sirois et al. (1988) listed 202 lichen taxa associated with serpentine substrates on Mt. Albert, Gaspésian Provincial Park, Québec, Canada (Figure 1A); of these taxa, 36 were recorded for the first time in Québec, 16 were new to Canada, and 11 new to North America. The 11 species listed for the first time from North America include *Belonia russula* (Physciaceae), *Buellia dispersa* (as *B. tergestina*; Physciaceae), *Cladonia uliginosa* (as *C. stricta* var. *uliginosa*; Cladoniaceae), *Dactylospora urceolata* (Dactylosporaceae), *Endococcus propinquus* (Agyriaceae), *E. rugulosus* (Agyriaceae), *Miriquidica plumbeoatra* (as *Lecidea plumbeoatra*; Agyriaceae), *Lithographa tesserrata* (Agyriaceae), *Polyblastia melaspora* (Lecanoraceae), *Rinodina mniaroeiza* (Lecanoraceae), and *Scoliciosporum umbrinum* var. *compacta* (Lecanoraceae). They concluded that the ecological influences of serpentine on the lichens were, in many aspects, similar to those observed on vascular plants in the region (Rune 1954), where many taxa are largely restricted to areas with serpentinized rocks. A recent study by Harris et al. (2007) explored the lichen flora of a serpentine outcrop from Little Deer Isle, Hancock County, Maine, U.S.A. (Figure 1A). Sixty-three species were found, comprising 35 genera. Two species, *Buellia ocellata* (Physciaceae) and *Cladonia symphyrcarpia* (Cladoniaceae), were new reports for New England. Twenty species, including one genus, *Lobaria* (Lobariaceae),

were new reports for serpentine substrates worldwide (Favero-Longo et al. 2004). These studies suggest that there may be a serpentine substrate effect for lichens in eastern North America and that further study may reveal new species or interesting floristic associations even for outcrops that have only been exposed for less than 10,000 years since the retreat of the Pleistocene glaciers (Dyke 2004). Appendix 1 lists 165 lichen species recorded from serpentine outcrops in eastern North America as reported by Harris et al. (2007) and Sirois et al. (1988).

Little work has been undertaken for bryophytes growing on serpentine soils worldwide. The limited number of studies to date focus on British Columbia (Lewis et al. 2004), Québec (Belland 1987; Sirois 1984), and Newfoundland (Belland and Brassard 1988; Roberts 1992) in Canada; California (Sigal 1975), Maine (Briscoe et al. 2009), and Maryland (Shaw and Albright 1990) in the United States; and outside North America in the British Isles (Bates 1978), Cuba (Marín et al. 2004; Pócs 1988), Japan (Takaki 1968), and Poland (Samecka-Cymerman and Kempers 1994; Samecka-Cymerman et al. 2002). Sirois (1984) lists 115 species of bryophytes for serpentine substrates on Mt. Albert. Robinson (1966) described the moss *Bryum reedii* (Bryaceae) as a new species first collected from serpentine in Maryland (Anderson et al. 1990). It was subsequently found by C. F. Reed on a granite outcrop in Delaware (J. Spence, National Park Service, pers. comm.). Briscoe et al. (2009) compared the bryophyte floras of a serpentine and a granite outcrop from the Deer Isles, Hancock County, Maine, U.S.A. (Figure 1A) and examined tissue elemental concentrations for select species collected from both sites. Fifty-five species were found, 43 on serpentine and 26 on granite. Fourteen (25.5%) of these species were found at both sites. Out of the 43 species collected from serpentine soils 31 were previously reported to occur on such soils worldwide (Briscoe et al. 2009). Of the 43 species collected from serpentine on Little Deer Isle, 12 were shared with the 115 species collected from serpentinized areas of Mt. Albert (Sirois 1984). The tissue of mosses collected from the serpentine site had higher Mg, Ni, and Cr concentrations and lower Ca:Mg ratios than the tissue of those collected from the granite site. This trend is similar to that observed for vascular plants collected from serpentine soils worldwide (Brady et al. 2005; Brooks 1987). Appendix 2 lists 146 bryophyte species recorded from serpentine outcrops in eastern North America as reported by Belland and Brassard (1988), Briscoe et al. (2009), Roberts (1992), Robinson (1966), Shaw (1991), Shaw and Albright (1990), and Sirois (1984).

Unlike vascular plants, there are few records of serpentine endemism for cryptogamic species worldwide. Cryptogamic species appear to be broadly tolerant of substrate and show wide geographic distributions and range disjunctions that frequently span more than one continent (Schuster 1983). Thus, very few of these species are endemic to specific substrates, a pattern that has led some to argue that cryptogams such as bryophytes evolve more slowly or are genetically depauperate (Crum 1972), although recent molecular (Fernandez et al. 2006) and ecophysiological (Shaw 1990b) studies suggest otherwise. We were unable to locate examples of lichens endemic to serpentine soils; Favero-Longo et al. (2004) concluded that although several lichen species have been historically recorded as endemic to serpentine substrates, all such species have also been found on other non-mafic yet basic siliceous substrates. A similar scenario exists for bryophytes; of the 15 species of bryophytes listed as endemic to Cuba and occurring on serpentine, none are restricted to serpentine (Marín et al. 2004). One moss species, *Pseudoleskeella serpentiniensis* (Leskeaceae), however, is thought to be endemic to serpentine in western North America (Shevock 2003). Given the high percentage of serpentine endemism among vascular plants, the low percentage among cryptogams is intriguing and is a topic worthy of investigation.

Only one study has examined algal diversity on serpentine soils in eastern North America. Terlizza and Karlander (1979) described algae from serpentine soils at Soldiers Delight, Maryland (Figure 1B). The algae were found to be of the phylum Cyanophycota (as Cyanophyta; some of which fix nitrogen) and the divisions Chlorophyta and Chrysophyta. At these taxonomic levels, however, the soil algal composition was similar to that of non-serpentine soils. One study has also examined the microbe populations on an asbestos mine associated with a serpentine outcrop in southeastern Québec, Canada (Moore and Zimmermann 1977). Although the tailings were devoid of vegetation, they supported significant microbe populations, although less than in normal agricultural soil. A 12-year old tailing was found to contain 7.4×10^4 aerobic, heterotrophic bacteria; 6.9×10^3 actinomycetes; and 3.4×10^2 fungi per gram of tailings.

STUDIES ON VASCULAR PLANTS: FLORISTICS

Appendix 3 lists 751 taxa of vascular plants belonging to 92 families documented from serpentine outcrops in eastern North

America, noting those species listed as rare, endangered, or threatened in Canada or the United States and those with global protection status (Atlantic Canada Conservation Data Center 2007; Center for Plant Conservation 2007; Centre de données sur le patrimoine naturel du Québec 2007; NatureServe 2007; USDA, NRCS 2007). Serpentine outcrops of eastern North America harbor many taxa with localized distribution patterns, including many that are threatened in several states across the United States and imperiled in Newfoundland and Québec, Canada (Atlantic Canada Conservation Data Center 2007; Centre de données sur le patrimoine naturel du Québec 2007). Two taxa, *Agalinis acuta* and *Schwalbea americana* (Scrophulariaceae), found on serpentine outcrops in eastern North America (Hay et al. 1992; Tyndall and Hull 1999), are federally listed as endangered in the United States (Center for Plant Conservation 2007; NatureServe 2007). Globally, six species, *Adiantum viridimontanum* (Pteridaceae), *Minuartia marcescens* (Caryophyllaceae), *Pycnanthemum torrei* (Lamiaceae), *Schwalbea americana* (Scrophulariaceae), *Scirpus longii* (Cyperaceae), and *Symphyotrichum depauperatum* (Asteraceae) are listed as imperiled (G2), while *Agalinis acuta* is listed as critically imperiled (G1). *Cerastium velutinum* var. *villosissimum* (as *C. arvense* var. *villosissimum*; Caryophyllaceae) is the only recognized serpentine endemic plant for eastern North America (Gustafson et al. 2003; Morton 2004) while *Adiantum viridimontanum*, *Aspidotis densa* (Pteridaceae), *M. marcescens*, and *S. depauperatum* are largely restricted to the substrate (Brooks 1987; Roberts 1992; Tyndall and Hull 1999). *Symphyotrichum depauperatum*, once thought to be endemic to serpentine, was recently collected from mafic diabase glades in North Carolina (Gustafson and Latham 2005; Hart 1990; Levy and Wilbur 1990). Based on current distributions, we propose that *A. viridimontanum* and *M. marcescens* should be considered endemic to serpentine substrates from eastern North America. *Aspidotis densa*, a strong serpentine indicator in western North America, appears to be restricted to serpentine outcrops in eastern North America (Kruckeberg 2002). While this taxon is abundant in western North America and is sometimes found off serpentine outcrops there, it has not been found off serpentine in eastern North America. Even with the addition of these taxa, the number of serpentine-endemic species in eastern North America is in sharp contrast to the 176 species endemic to serpentine in California alone (Safford et al. 2005). Because many serpentinized areas in eastern

North America were under ice during the last glaciation, limiting the extent of soil development and length of plant colonization (Roberts 1992), it is likely that the plants associated with these young soils have not had adequate time to diverge and specialize as on similar soils in unglaciated lower latitudes of the world. Floristic surveys have been conducted on many of the prominent serpentine sites in eastern North America (Figure 1A, B), including the exhaustive surveys by Reed (1986) on many key serpentine sites across eastern North America. The serpentine barrens of Maryland are perhaps the most studied serpentine outcrops in the eastern United States (Tyndall 2005; Tyndall and Farr 1989, 1990; Tyndall and Hull 1999). Serpentine areas in Maryland, including the renowned Soldiers Delight site, host a number of rare species (Tyndall 1992a; Tyndall and Hull 1999): *Agalinis acuta*, *Carex hystericina* (Cyperaceae), *C. richardsonii* (Cyperaceae), *Desmodium obtusum* (as *D. rigidum*; Poaceae), *Dichantheium oligosanthos* var. *oligosanthos* (as *Panicum oligosanthos*; Poaceae), *Gentiana andrewsii* (Gentianaceae), *Gentianopsis crinita* (Gentianaceae), *Linum sulcatum* (Linaceae), *Panicum flexile* (Poaceae), *Pycnanthemum torrei* (Lamiaceae), *Sporobolus heterolepis* (Poaceae), *Symphyotrichum depauperatum* (as *Aster depauperatus*; Asteraceae), and *Talinum teretifolium* (Portulacaceae).

Serpentine areas of Delaware (Tyndall and Hull 1999), Georgia (Radford 1948), Maine (Carter 1979; N. Pope and N. Rajakaruna, unpubl. data), New York (Reed 1986), North Carolina (Mansberg and Wentworth 1984; Milton and Purdy 1988; Radford 1948), Pennsylvania (Latham 1993; Pennell 1910, 1912, 1930; Wherry 1963), and Vermont (Zika and Dann 1985) have also been exposed to restricted floristic explorations (see approximate locations of study sites in Figure 1A, B); however, much less is known about these floras than is known of the serpentine flora of Maryland.

Zika and Dann (1985) explored several serpentine outcrops in Vermont (Figure 1A) and found several rare and one possibly threatened plant species for the state. The rare species included *Adiantum aleuticum* (as *A. pedatum* var. *aleuticum*; Pteridaceae), *Agrostis borealis* (Poaceae), *Asplenium trichomanes-ramosum* (as *A. viride*; Aspleniaceae), *Carex scirpoidea* (Cyperaceae), *Dryopteris fragrans* (Dryopteridaceae), *Empetrum nigrum* (Ericaceae), *Minuartia marcescens* (as *Arenaria marcescens*; Caryophyllaceae), *Scirpus caespitosus* (Cyperaceae), *Thelypteris simulata* (Cyperaceae), *Vaccinium uliginosum* (Ericaceae), and *V. vitis-idaea* (Ericaceae). *Huper-*

zia selago (as *Lycopodium selago*; Lycopodiaceae) was recommended to be listed as threatened in Vermont (Zika and Dann 1985). They also found this taxon on non-serpentine substrates in two small alpine zones where it was threatened by heavy foot traffic. It was listed as rare throughout New England by Crow et al. (1981). Similarly, Crow et al. (1981) suggested *Adiantum aleuticum* (as *A. pedatum* var. *aleuticum*), *Asplenium trichomanes-ramosum* (as *A. viride*), *M. marcescens* (as *Arenaria marcescens*), and *Moehringia macrophylla* (as *Arenaria macrophylla*; Caryophyllaceae) be listed as threatened or endangered in New England. Currently, the serpentine-associated species state listed as threatened in Vermont are *A. trichomanes-ramosum*, *M. marcescens*, and *Adiantum viridimontanum* (Vermont Nongame and Natural Heritage Program 2005).

In a detailed floristic and phytogeographical analysis of several serpentine sites in Maine, Carter (1979) documented 250 taxa and concluded that although there was no overriding continuity in the composition of the serpentine flora in the state, the generally stunted vegetation reflected the presence of serpentine soils. The collection of *Oryzopsis asperifolia* (Poaceae) was a new record for Somerset County, while *Asplenium trichomanes-ramosum* (as *A. viride*), also collected from Somerset County, had been recorded from only one other site in the state. *Adiantum aleuticum* (as *A. pedatum* var. *aleuticum*) from Franklin County was listed as a new record for the state. The two fern species are currently state listed as S1 (highly rare statewide) with a global rarity status of G4 and G5, respectively (Maine Natural Areas Program 2005). Recent studies on serpentine on the Deer Isles, Maine (N. Pope and N. Rajakaruna, unpubl. data) also suggest interesting floristic associations, with known serpentine species such as *Asplenium trichomanes* and *Adiantum aleuticum* restricted to the Islands' serpentine substrates.

Adiantum aleuticum from the eastern North American serpentine sites has been exposed to rigorous biosystematic studies (Paris 1991; Paris and Windham 1988; Rugg 1922). The *A. aleuticum* complex includes *A. pedatum* subsp. *calderi* and *A. pedatum* var. *aleuticum* and is genetically divergent from the common eastern woodland maidenhair fern, *A. pedatum sensu stricto* (Paris 1991; Paris and Windham 1988). The allotetraploid derivative of *A. pedatum* and *A. aleuticum*, *A. viridimontanum*, is known only from a few serpentine outcrops in Vermont (Paris 1991) and is listed as threatened there (Vermont Nongame and Natural Heritage Program 2005). The parental taxa are found on several serpentine outcrops in

southeastern Canada and northeastern United States and are often considered rare where they are found (Carter 1979; Cody 1983; Paris 1991; Zika and Dann 1985).

Serpentine outcrops in eastern Canada have had a long history of botanical exploration (e.g., Fernald 1907, 1911, 1926, 1933). Outcrops in Québec (Bouchard et al. 1983; Legault and Blais 1968; Rune 1954; Sirois and Grandtner 1992) and Newfoundland (Bouchard et al. 1978, 1991; Damman 1965; Dearden 1977, 1979; Hay et al. 1992, 1994; Robertson and Roberts 1982) have been extensively botanized. As mentioned previously, *Aspidotis densa* (as *Cheilanthes siliquosa*) and *Minuartia marcescens* (as *Arenaria marcescens*), are listed as threatened in Québec (MDDEP 2007). A recent report by the Canadian Legal Information Institute (2008) also lists *Polystichum scopulinum* (Dryopteridaceae), *Salix chlorolepis* (Salicaceae), and *Solidago simplex* subsp. *simplex* var. *chlorolepis* (Asteraceae), all associated with serpentine on Mount Albert in the Gaspésie, as threatened in Québec. Several other rare species have been documented on serpentine in eastern Canada including *Danthonia intermedia* (Poaceae), *Eleocharis nitida* (Cyperaceae), *Festuca altaica* (Poaceae), *Salix arctica*, and three species of Caryophyllaceae: *Minuartia biflora*, *Sagina caespitosa*, and *S. saginoides*, all fairly recent additions to the flora of Newfoundland (Hay et al. 1994). Given their occurrence in mostly high altitude serpentine sites, including Table Mountain and the White Hill Mountains (Newfoundland) and Mt. Silver and Mt. Albert (Québec; Figure 1A), these plants are prone not only to the typical physiochemical stresses of serpentine soils but also to physical stresses such as drought, wind, snow, and cryoturbation (Roberts 1980, 1992).

ECOPHYSIOLOGICAL AND EVOLUTIONARY STUDIES

Serpentine habitats have long provided a model for ecophysiological and evolutionary studies (Brady et al. 2005; Roberts and Proctor 1992). While many long-term studies exist for other regions of the world, especially in California (Alexander et al. 2007; Kruckeberg 1984, 1992, 2002), few such rigorous studies have been conducted in eastern North America. The following is a summary of key studies focusing on ecological and evolutionary aspects of serpentine habitats, their plants, and associated biota in eastern North America.

Plant heavy-metal relations. Heavy metal accumulation in plants is an intriguing phenomenon and much work has been conducted to determine its physiological and genetic basis (Pollard et al. 2002) as well as the adaptive significance (Boyd 2004, 2007) of this unusual physiological process. Heavy metal hyperaccumulation can be found in plants growing on a range of metalliferous soils (Brooks 1998), and the hyperaccumulation of heavy metals, notably Ni, is a phenomenon commonly found in vascular plants from serpentine soils (Reeves 2003). For most metals, including Ni, hyperaccumulation is defined as the accumulation of metal to make up over 0.1% of dry leaf weight ($1000 \mu\text{g g}^{-1}$ dry leaf tissue), although the threshold level is over 1% for metals such as Zn and Mn. Only 320 Ni hyperaccumulators have been discovered worldwide, belonging to mostly the Brassicaceae and Euphorbiaceae (Reeves 2003). About two-thirds of the known Ni hyperaccumulators are found in the tropics, with the islands of New Caledonia and Cuba harboring the majority of such species. There are only two verified reports of Ni hyperaccumulators in the United States: *Thlaspi montanum* (Brassicaceae; Heath et al. 1997; Reeves et al. 1983) and *Streptanthus polygaloides* (Brassicaceae; Reeves et al. 1981), both restricted to western North America. *Thlaspi montanum* consists of three varieties (var. *montanum*, var. *siskiyouense*, and var. *californicum*) in western North America, all of which were found to hyperaccumulate Ni (Reeves et al. 1983). A third taxon, *Minuartia rubella* (as *Arenaria rubella*; Caryophyllaceae), has also been reported to hyperaccumulate Ni (Kruckeberg et al. 1993). Further investigation of this species from the original site, however, has cast doubt on the validity of the reported data (R. R. Reeves, pers. comm.). The only proposed hyperaccumulators of Ni from eastern North America—*A. humifusa* (Caryophyllaceae), *M. marcescens* (Caryophyllaceae), *Packera paupercula* (as *Senecio pauperculus*; Asteraceae), and *Solidago hispida* (Asteraceae)—occur in Newfoundland (Roberts 1992). These reports have yet to be verified (R. R. Reeves, pers. comm.), however, the Asteraceae and Caryophyllaceae are families with known Ni hyperaccumulators elsewhere (Brooks 1998). Brooks (1987) listed these four species as the only proposed Ni hyperaccumulators to be found in previously glaciated regions of the world.

Milton and Purdy (1988) sampled the foliage from several species of trees growing on serpentine soils in the Buck Creek and Webster-Addie districts in the Blue Ridge Mountains, North Carolina.

White oak (*Quercus alba*; Fagaceae) leaves accumulated the most Ni, about 400 to 700 $\mu\text{g g}^{-1}$ dry leaf tissue from five sites at Buck Creek, but $< 200 \mu\text{g g}^{-1}$ from sites at Webster-Addie. A recent study by Briscoe et al. (2009) demonstrated higher levels of Ni in mosses collected from serpentine soil compared to those collected from nonserpentine soil. *Polytrichum juniperinum* and *P. piliferum* (Polytrichaceae), found on and off of serpentine soil, contained 26.3 and 129 $\mu\text{g g}^{-1}$ Ni in dry leaf tissue, respectively, on serpentine, compared to < 1.5 and 3.69 $\mu\text{g g}^{-1}$ Ni in dry leaf tissue, respectively, from nonserpentine soils. *Weissia controversa* (Pottiaceae), a species known to inhabit metal-contaminated sites worldwide (Porley and Hodgetts 2005; Shaw et al. 1987), accumulated the highest levels of Ni (363 $\mu\text{g g}^{-1}$ Ni in dry leaf tissue) among all species sampled. Levels of metal accumulation have not been determined for the many serpentine taxa thus far listed for eastern North America (Appendix 1, 2, 3). Intense study should be directed at taxa with known accumulators, especially in the families Asteraceae, Brassicaceae, and Caryophyllaceae, known to harbor a disproportionately high number of species worldwide with the capacity to hyperaccumulate Ni and other metals (Brooks 1998; Reeves 2003).

Cross-kingdom interactions. Due to the harsh conditions ruling serpentine habitats, plants growing on serpentine and their associated biota—ranging from mutualistic organisms such as mycorrhizae, pollinators, and seed dispersers to antagonistic organisms such as pathogens and herbivores—show unique adaptations or biotic associations (Alexander et al. 2007; Kruckeberg 1984). Some animals are dependent on serpentine soils and their plants while many others spend at least some part of their lives on serpentine soils. Much of the work exploring serpentine plants and their cross-kingdom interactions has been conducted in western North America and other parts of the world (Boyd 2007). Notable studies include those on ants (Fisher 1997), butterflies (Gervais and Shapiro 1999; Harrison and Shapiro 1988), daddy long-leg spiders (Alexander et al. 2007), leaf beetles (Mesjasz-Przybyłowicz and Przybyłowicz 2001), and pocket gophers (Hobbs and Mooney 1995; Proctor and Whitten 1971).

In the only study of its kind from eastern North America, Wheeler (1988) found a beetle (*Diabrotica crista*; Chrysomelidae)—seldom found along the Atlantic coast but common farther west—

to be abundant on the Goat Hill and Nottingham barrens and present at Soldiers Delight. The main host-plant for the larvae is big bluestem (*Andropogon gerardii*; Poaceae) in the Midwest and assumed to be little bluestem (*Schizachyrium scoparium*; Poaceae) on serpentine prairies and savannas of the Baltimore complex. Although much work has been done elsewhere on insects and arthropods associated with plants that grow on serpentine soils, including their potential for metal accumulation (Boyd 2007; Boyd, Davis, Wall, and Balkwill 2006; Boyd, Wall, and Jaffré 2006), this topic is clearly under explored in eastern North America.

Microbe-soil relations of serpentine outcrops have been investigated by several groups worldwide (Balkwill 2001; Boyd et al. 2004). A number of researchers have examined mycorrhizae on serpentine soils in California (Hopkins 1987; Moser et al. 2005) and found distinct differences in taxa found on serpentine versus non-serpentine soils. In eastern North America, Panaccione et al. (2001) found a lower diversity of ectomycorrhizal fungi on serpentine plots at Soldiers Delight than on nearby non-serpentine soil. They collected *Cenococcum geophilum* (Ascomycota) isolates from *Pinus virginiana* (Pinaceae) seedlings in both serpentine and non-serpentine soils and found that the *C. geophilum* isolates from serpentine sites were genetically more similar to each other than to isolates from both local and distant non-serpentine sites. A study conducted in Virginia by Sheets et al. (2000) showed that the diversity of basidiocarps, mycorrhizas, and mycorrhizal inocula was lower on serpentine soil than on non-serpentine soil. Castelli and Casper (2003) demonstrated both inter- and intra-specific arbuscular mycorrhizal (AM) fungal variation among the dominant grass species in a serpentine community in Pennsylvania. Gustafson and Casper (2004) examined the impact of nutrient addition on AM fungal performance and expression of plant/fungal community feedback in three serpentine grasses found on the Goat Hill and Nottingham serpentine barrens in Chester County, Pennsylvania. Their study suggested implications for decoupling of plant/fungal community feedback by anthropogenic nutrient enrichment.

Serpentine grasslands often harbor plants with shallow root systems and brief life cycles and mycorrhizae appear to play an important role in plant nutrition and adaptation under such conditions (Hopkins 1987); improper management of such soils combined with atmospheric deposition of nutrients (Weiss 1999) may have unfavorable effects on plants and their microbe

communities. Thiet and Boerner (2007) examined the role of soil ectomycorrhizal (ECM) fungal inoculum in the invasion of *P. virginiana* at Soldiers Delight in Maryland. They suggested that ECM fungi facilitate rapid pine colonization from bordering mature pine forests and that current management practices should incorporate methods to kill or disrupt hyphal mats attached to mature pines to halt pine invasion to serpentine barrens. Cumming and Kelly (2007) investigated the effects of *P. virginiana* invasion at Soldiers Delight on soil properties, AM fungi, and native plant growth. They found drastic changes in soil pH (a drop from 6.2 to 4) and other changes in soil chemistry, AM fungal community structure, and plant growth, although varying in impact among serpentine grassland, savannah, and woodland habitats. This study has important implications for the management and restoration of serpentine habitats.

Plant ecology. Serpentine outcrops have long provided natural laboratories for exploring ecological theory (Alexander et al. 2007; Harrison, Davies, Grace, Safford, and Viers 2006; Harrison, Davies, Safford, and Viers 2006; Harrison, Safford, Grace, Viers, and Davies 2006) and evolutionary processes (Kruckeberg 2002; Rajakaruna 2004). Such studies in eastern North America are limited in number and scope.

Dearden (1979) examined factors influencing plant community location and composition on serpentine bedrock in western Newfoundland. He identified six plant community types and concluded that species composition was significantly correlated with available Ca, an element generally low in serpentine soils, and topography. The community types showing the greatest similarity to adjacent non-serpentine soils were found on soils with highest available Ca, lowest available Mg, and lowest total Ni concentrations.

It was once commonly thought that the availability of soil water was the main factor limiting plant growth on serpentine soils (Hughes et al. 2001; Proctor 1999). Hull and Wood (1984) examined plant water relations to determine if water availability was a limiting factor in the distribution of *Quercus* species on serpentine soils in Maryland. Although pre-dawn xylem water potentials were similar for the serpentine oaks (*Q. stellata*, *Q. marilandica*) and the non-serpentine oaks (*Q. alba*, *Q. velutina*) early in the growing season, by late summer they were higher for the serpentine species; however, the trend was not consistent for all species across the two substrates. Despite differences

in species responses to water availability, the authors concluded that water alone was not responsible for the distributional pattern of these species in Maryland.

Wood (1984) investigated plant-soil relationships for several elements known to vary significantly in concentration between serpentine and non-serpentine soils. There were no differences in heavy metal or Ca concentrations in the plants and soils tested; however, Mg concentrations differed significantly, suggesting an important role for Mg, especially in the distribution of oak species found on serpentine (*Quercus stellata* and *Q. marilandica*) and off serpentine (*Q. alba* and *Q. velutina*) in Maryland. The Ca:Mg ratio is now considered a major factor controlling plant growth and diversity on serpentine soils (Brady et al. 2005).

Hart (1980) examined the mechanisms by which serpentine-restricted taxa and their weedy congeners coexist on serpentine soils in southeastern Pennsylvania. He documented contrasting strategies for the congeneric pairs, with the weeds generally having higher potential growth rate, more mesic leaf structures, lower seedling mortality (on normal soils), lower Ca uptake, and earlier or more abundant seed production. He concluded that the most significant factors allowing weeds to persist on serpentine appear to be rapid growth when conditions are favorable and some reproduction early in the life cycle. The presence of early flowering times for several bodenvag species (Kruckeberg 1986) suggests that this is also true for certain weedy species on the serpentine outcrop on Little Deer Isle, Maine (N. Pope and N. Rajakaruna, unpubl. data). The serpentine-restricted congeners appear to allocate biomass not to rapid growth or early reproduction but to organs that enhance growth and survival during severe stress (Hart 1980). This study suggests that both weeds and their serpentine-restricted congeners can coexist when both moisture and nutrient availability are found within a particular range; when moisture and nutrients are found in abundance weeds grow faster and when low, serpentine-restricted taxa are favored. A similar trend was observed for edaphic races of *Lasthenia californica* (Asteraceae) in California (Rajakaruna and Bohm 1999).

Arabas (2000) examined the spatial and temporal patterns of disturbance and vegetation change in the Nottingham Barrens of Pennsylvania over the past 150 years. The study points to the importance of fire in maintaining the serpentine savannah conditions that support many rare and endemic serpentine taxa. Less frequent fires allow the savannahs and open woodlands to convert to closed

hardwood forests with immediate consequences on native plant diversity. Soil depth is also an important factor influencing rate and direction of succession; where soils are shallow, indigenous species have a competitive edge. This study, examining the relationships among fire frequency, vegetation, and soil depth of a serpentine barren, has important implications for land management.

Fire clearly plays a critical role in maintaining the vegetation of serpentine habitats in fire-prone regions (Harrison et al. 2003; Safford and Harrison 2004) and improper management of these sites can have dire consequences on plants uniquely adapted to grow there (Tyndall 1994). Tyndall and Hull (1999) provided a useful summary of pre- and post-settlement land-use history for both Maryland and Pennsylvania showing how fire suppression and livestock grazing have drastically altered the floristic composition of the serpentine barrens since the mid-1900s. In a study exploring succession patterns following fire on a serpentine barren in Pennsylvania, Miller (1981) suggested that the post-fire flora and succession patterns on serpentine are distinct from those on nonserpentine and that the dominant species on serpentine are well adapted to the occurrence of fires. In an exhaustive study of the role of fire on serpentine chaparral in California, Safford and Harrison (2004) reported that the effects of fire on less productive plant communities like serpentine chaparral may be longer lasting than the effects of fire on similar but more productive communities found off serpentine. All these studies point to the key role fires play in maintaining the diversity and ecology of serpentine habitats.

Only a handful of studies have examined the nature of ecotypic variation and divergence in response to serpentine soils in eastern North America despite many such studies elsewhere (Alexander et al. 2007; Brady et al. 2005). Ware and Pinion (1990) found little evidence of local adaptation to serpentine soils in serpentine populations of the bodenvag taxon, *Talinum teretifolium* (Portulacaceae; Kruckeberg 1984), found on granite from Virginia to Georgia, on serpentine in the northern portion of its range in Maryland and Pennsylvania, and on sandstone on the western and southern extreme of its range. Serpentine plants performed similarly on all substrates, showing no evidence of local adaptation to serpentine soils, but they developed chlorosis when grown on limestone soils. Despite slow growth, plants on serpentine soils remained healthy, persisting in the generally competition-free, shallow soils found there. Miller and Cumming (2000) examined the potential for ecotypic differentiation in *Pinus*

virginiana, an invasive species on serpentine barrens. They tested the effects of exchangeable Ca:Mg and Ni on growth, foliar pigment concentrations, and nutrient status of seedlings from serpentine and nonserpentine soils in Pennsylvania. They found that seedlings from trees currently growing on serpentine were no different in their response to Ca:Mg and Ni than those from off serpentine. This study implies that no ecotypic differentiation has occurred in this species with respect to key serpentine soil factors.

Habitat restoration and management studies. Serpentine outcrops have long been subjected to mining for the extraction of heavy metals such as Ni and Cr as well as minerals such as asbestos (Brooks 1998; Kruckeberg 1984, 2002). Moore and Zimmermann (1977) tested the revegetation potential of 23 species tolerant of asbestos tailings in Québec and found that the grasses (Poaceae) *Bromus inermis*, *Elymus junceus*, *Lolium perenne*, and *Poa pratensis* and the legumes (Fabaceae) *Medicago sativa*, *Melilotus alba*, and *Trifolium hybridum* could be successful at revegetating the site. The cost of revegetation and the low availability of seed from local populations were cited as obstacles to the revegetation effort.

Long-term suppression of fire and changes in grazing patterns have led to the rapid spread of *Pinus virginiana* and *Juniperus virginiana* (Cupressaceae), both fire-intolerant conifers, as well as *Frangula alnus* (as *Rhamnus frangula*; Rhamnaceae), which is rapidly replacing the native herbaceous plants restricted to the serpentine barrens of Maryland (Tyndall 1992a, b; Tyndall and Hull 1999). Native oaks, *Quercus stellata* and *Q. marilandica*, have also been affected by the fast-growing conifers, although *P. virginiana* succession appears to be inhibited to some extent by drought (Tyndall and Farr 1990). Barton and Wallenstein (1997) concluded that *P. virginiana* stands tend to increase soil depth (both mineral soil and litter depth), promoting suitable conditions for forests typical of non-serpentine sites in the region. Without immediate and proper management, all remaining serpentine barrens in Maryland are at risk of developing into conifer forests, representing a loss of habitat for the rare species living there (Knox 1984; Tyndall 1992b, 1994, 2005).

CRITICAL INFORMATION GAPS AND FUTURE DIRECTIONS

Serpentine outcrops provide unique opportunities for ecologists to explore the critical role of geology on biota worldwide. While

there appears to be a dearth of such studies in eastern North America relative to other parts of the world, our review points to a number of detailed studies examining the taxonomic and experimental aspects of biota on serpentine outcrops in the region. This descriptive work has pointed to several interesting taxa, floristic associations, cross-kingdom interactions, possible impacts of soils on the divergence of species, and implications for management of these unique habitats. However, there is still much work needed to reveal the geoecology and best management practices for serpentine outcrops in the region. While we report geoecological studies from Newfoundland, Québec, Maine, Vermont, New York, Delaware, Virginia, North Carolina, and Georgia, we failed to locate any published literature on serpentine geoecology from New Brunswick, New Jersey, South Carolina, and Alabama. We identify several areas where further research would significantly enhance the knowledge base for serpentine geoecology in eastern North America.

As is evident from the limited geographical knowledge, a more rigorous attempt at mapping the geologic extent of serpentine in eastern North America is greatly needed. Studies to characterize the mineralogy, pedology, and soil characteristics of serpentinized areas across eastern North America are also needed (R. G. Coleman, pers. comm.). While a few larger outcrops in Newfoundland (Dearden 1979; Roberts 1980, 1992), Québec (De Kimpe et al. 1973; Laurent 1975), Maryland (Rabenhorst and Foss 1981), and Pennsylvania (Miller and Cumming 2000) have received some attention (Brooks 1987; Tyndall and Hull 1999), smaller outcrops in New Brunswick, Maine, Vermont, New York, New Jersey, Delaware, Virginia, North Carolina, South Carolina, Georgia, and Alabama have gone largely unnoticed. Studies to date have shown that the soil chemical features, especially concentrations of heavy metals such as Ni and nutrient ratios such as exchangeable Ca:Mg, are comparable to those found in serpentine soils from California (Table 1) and other regions of the world (Alexander et al. 2007; Brooks 1987). A better understanding of the soils, especially from the rhizosphere of plants, would reveal the nature of soil-related stressors as well as specific tissue-ion relations for plants found on serpentine in the region.

Additional surveys should be conducted on floristics, including detailed studies of vascular plants, non-vascular plants, and lichens. While several studies have addressed the diversity of serpentine-

associated vascular plants of the region (Appendix 3), especially in Newfoundland, Québec, Maryland, Pennsylvania, and North Carolina, there are still many under-explored serpentine outcrops across eastern North America including in Maine, Vermont, New York, Delaware, and Virginia. We were unable to find any detailed floristic surveys for serpentine in New Jersey, Delaware, Virginia, North Carolina, South Carolina, Georgia, or Alabama other than those listed in Reed (1986). A better understanding of floristics can reveal taxa with site-associated variation, leading to experimental investigations on ecotypic or species-level divergence (Rajakaruna 2004). Currently, such biosystematic investigations exist for only three serpentine-associated taxa, *Adiantum aleuticum* (Paris and Windham 1988), *Cerastium arvense* var. *villosum* (Gustafson et al. 2003), and *Symphyotrichum depauperatum* (Gustafson and Latham 2005), compared to a plethora of such investigations from California (Alexander et al. 2007; Rice and Espeland 2007). *Aspidotis densa*, which shows an intriguing disjunct distribution on serpentine between western and eastern North America (Krukeberg 2002), and the two proposed narrow endemics, *Adiantum viridimontanum* and *Minuartia marcescens*, would be taxa worthy of investigation in this respect. To date, only a handful of investigations have focused on lichens in Québec and Maine (Appendix 1) and bryophytes in Québec, Maine, and Maryland (Appendix 2). This information is lacking for Newfoundland, New Brunswick, Vermont, New Jersey, New York, Delaware, Virginia, North Carolina, South Carolina, Georgia, and Alabama.

Studies of plant-metal relations are lacking compared to studies of this nature in other regions where outcrops occur (Alexander et al. 2007; Reeves 2003). The discovery of only a handful of species with the potential to accumulate high levels of Ni (Roberts 1992) may be due to insufficient study of soil-tissue relations for the region. Even the currently proposed Ni hyperaccumulators (Roberts 1992) have not been verified (R. R. Reeves, pers. comm.). Given that plant-available soil Ni levels in the region are comparable to levels found in areas such as California (Table 1), it is possible that additional taxa may be found with the capacity to accumulate this metal. Detailed studies of metal accumulation should be conducted on plants growing on outcrops across the region, as only three studies to date have examined aspects of heavy metal tolerance and accumulation—one in North Carolina (Milton and Purdy 1988), one in Maine (Briscoe et al. 2009), and Roberts'

study in Newfoundland. Intense study could reveal additional taxa with the unusual capacity to hyperaccumulate Ni and other metals, as well as reveal any unique relations plants may have with respect to other soil elements, particularly nutrients such as Ca and Mg. If such investigations do not reveal new hyperaccumulators or confirm the currently listed hyperaccumulators (Roberts 1992), they may provide the basis to explore the intriguing question as to why there are so few hyperaccumulators in North America and other temperate regions compared to tropical areas (Reeves 2003).

Studies of cross-kingdom interactions are lacking, especially those that examine herbivores and pathogens associated with serpentine plants (Boyd 2004). While many such studies exist for California and other tropical serpentine outcrops (Boyd 2007), the studies to date in eastern North America have focused on mycorrhizal associations on serpentine soil in only Maryland (Cumming and Kelly 2007; Panaccione et al. 2001; Thiet and Boerner 2007), Pennsylvania (Castelli and Casper 2003; Gustafson and Casper 2004), and Virginia (Sheets et al. 2000). Only one study from Pennsylvania and Maryland has examined a plant-insect association on serpentine in eastern North America (Wheeler 1988). Studies of cross-kingdom interactions do not exist for any of the other states or provinces. Investigations on the potential for metal transfer to higher trophic levels, key to examining how these metal-rich habitats could influence ecosystem health (Boyd, Davis, Wall, and Balkwill 2006; Boyd, Wall, and Jaffré 2006; Wall and Boyd 2002), do not exist for any state or province where serpentine occurs in eastern North America.

Ecological and evolutionary studies on serpentine systems are also needed at a foundational level. Serpentine outcrops provide model habitats to test ecological and evolutionary theory, as clearly documented by detailed geoecological studies in other parts of the world (Alexander et al. 2007; Baker et al. 1992; Boyd et al. 2004; Brooks 1987; Harrison and Viers 2007). In eastern North America many research possibilities still exist; research is needed on the ecology, physiology, and evolution of particular taxa associated with serpentine soils, including those taxa that appear to be endemic or largely restricted to the substrate. Studies on the evolutionary ecology of such taxa could reveal the nature of serpentine tolerance, direction and strength of selection imposed by serpentine factors, and factors and mechanisms responsible for divergence. Finally, studies focusing on soil remediation and site restoration are limited. Although Ni

mining has been considered largely economically unattractive in the eastern United States, extensive prospecting of Ni and other heavy metals has occurred in Virginia, North Carolina, Georgia, and Alabama (Worthington 1964), as well as in other parts of eastern North America (Cannon 1971; Wickland 1990). Serpentine outcrops that have undergone mining operations, such as those found in eastern Canada (Moore and Zimmermann 1977), can provide model habitats to investigate the potential for phytoremediation and phytomining using metal-accumulating serpentine plants. Both of these practices are quickly gaining recognition as environmentally friendly, low-cost technologies for the remediation of metal-contaminated sites worldwide (Angle and Linacre 2005; Boominathan et al. 2004; Brooks 1998; Pilon-Smits 2005). Plants associated with serpentine soils are not merely biological novelties suited for taxonomic, ecological, physiological, and evolutionary investigations; they also hold great potential as tools for the restoration of metal-contaminated sites around the world (Whiting et al. 2004).

While eastern North American serpentine outcrops provide a wealth of opportunities for geoecological investigations, ever-expanding agriculture, forestry, and mining activity, as well as fire suppression and urbanization, have drastically affected the biota of many eastern North American serpentinized areas. This is especially of concern as six of the serpentine taxa found in the region are globally imperiled (G2) and one taxon is listed as globally critically imperiled (G1). Recent years have seen the declaration of several preserves worldwide, set aside for their unique edaphic habitats and associated biota. Although spotty in their distribution and inadequate in number on a global scale, several preserves in the states of California, Oregon, and Washington, and in Cuba, Italy, New Caledonia, New Zealand, South Africa, and Sri Lanka have led the way in raising awareness of the immediate need for the conservation of these unique biotas (Kruckeberg 2002, 2004; Rajakaruna and Bohm 2002). There are several preserves in eastern North America, including the well-known Mt. Albert, Gaspésian Provincial Park, Québec, Canada (Kruckeberg 2004); Table Mountain, Gros Morne National Park, Newfoundland, Canada (Belland and Brassard 1988; Bouchard et al. 1986; Roberts 1992); State-Line Serpentine Barrens in Pennsylvania (Nature Conservancy 2007); Soldiers Delight Natural Environment Area in Maryland (Flanagan-Brown 2001; Soldiers Delight Conservation, Inc. 2007); and Pine Hill Preserve, Deer Isle,

Maine (Harris et al. 2007) set aside to preserve the unique biota associated with such sites. Preservation of such sites will assist in the conservation of rare and/or physiologically distinct species and provide avenues for much-needed long-term studies as well as opportunities to educate the general public of the role that extreme geologic settings play in maintaining and generating biotic diversity. It is our hope that this review will generate renewed interest in serpentine geoecology as a fruitful field with much promise for future research in eastern North America.

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APPENDIX 1

Lichen species reported from serpentine substrates in eastern North America based on studies by Harris et al. (2007; Maine, U.S.A. = ME) and Sirois et al. (1988; Québec, Canada = QC). Names standardized using Index Fungorum [Index Fungorum Partnership; website (<http://www.indexfungorum.org>); accessed 28 Jan 2008], Integrated Taxonomic Information System [website (<http://www.its.gov>); accessed 23 Jan 2008], and USDA, NRCS (2008). ¹Taxa that have undergone nomenclatural changes since this paper went to press; see Esslinger, 2008 for current taxonomy.

Species	Occurrence
<i>Acarospora fuscata</i> (Shrader) Arnold	ME
<i>Alectoria nigricans</i> (Ach.) Nyl.	QC
<i>A. ochroleuca</i> (Hoffm.) A. Massal.	QC
<i>Amygdalaria panaeola</i> (Ach.) Hertel & Brodo	QC
<i>Anaptychia palmulata</i> (Michx.) Vain.	ME
<i>Arctoparmelia centrifuga</i> (L.) Hale	QC
<i>A. incurva</i> (Pers.) Hale	QC
<i>Aspicilia cinerea</i> (L.) Körb.	ME; QC
<i>Bacidia sabuletorum</i> (Schreb.) Lettau ¹	QC
<i>Baeomyces carneus</i> Flörke	QC
<i>B. rufus</i> (Huds.) Rebert.	QC
<i>Bellemerea cinereorufescens</i> (Ach.) Clauzade & Cl. Roux	QC
<i>Belonia russula</i> Nyl.	QC
<i>Biatora vernalis</i> (L.) Fr.	QC
<i>Bryocaulon divergens</i> (Ach.) Kärnefelt	QC
<i>Bryoria nitidula</i> (Th.Fr.) Brodo & D. Hawksw.	QC
<i>Buellia dispersa</i> A. Massal.	QC
<i>B. leptocline</i> (Flot.) A. Massal.	QC
<i>B. ocellata</i> (Flot.) Körb.	ME
<i>B. papillata</i> (Sommerf.) Tuck.	QC
<i>Caloplaca ammiospila</i> (Wahlenb.) H. Olivier	QC
<i>C. holocarpa</i> (Ach.) A.E. Wade	QC
<i>C. lithophila</i> H. Magn.	ME
<i>C. microthallina</i> (Wedd.) Zahlbr.	ME
<i>C. scopularis</i> (Nyl.) Lettau	ME
<i>C. sinapisperma</i> (Lam. & DC.) Maheu & A. Gillet	QC
<i>C. tetraspora</i> (Nyl.) H. Olivier	QC
<i>Candelariella aurella</i> (Hoffm.) Zahlbr.	ME
<i>C. vitellina</i> (Hoffm.) Müll.Arg.	ME; QC
<i>Carbonea vorticosa</i> (Flörke) Hertel	QC
<i>Catillaria lenticularis</i> (Ach.) Th.Fr.	ME
<i>C. muscicola</i> Lyngé	QC
<i>Catolechia wahlenbergii</i> (Ach.) Körb.	QC
<i>Cetraria cucullata</i> (Bellardi) Ach. ¹	QC
<i>C. delisei</i> (Schaer.) Nyl. ¹	QC
<i>C. ericetorum</i> subsp. <i>ericetorum</i> Opiz	QC

Appendix 1. Continued.

Species	Occurrence
<i>C. hepatizon</i> (Ach.) Vain. ¹	QC
<i>C. islandica</i> (L.) Ach. subsp. <i>islandica</i>	QC
<i>C. islandica</i> subsp. <i>crispiformis</i> (Räsänen) Kärnefelt	QC
<i>C. laevigata</i> Rass.	QC
<i>C. nivalis</i> (L.) Ach. ¹	QC
<i>C. tilessi</i> Ach. ¹	QC
<i>Cladonia mitis</i> (Sandst.) W.L. Culb. ¹	QC
<i>C. rangiferina</i> (L.) Nyl. ¹	QC
<i>C. stellaris</i> (Opiz) Brodo ¹	QC
<i>Cladonia acuminata</i> (Ach.) Norrl.	ME; QC
<i>C. amaurocraea</i> (Flörke) Schaer.	QC
<i>C. boryi</i> Tuck.	ME
<i>C. cariosa</i> (Ach.) Spreng.	ME
<i>C. carneola</i> (Fr.) Fr.	QC
<i>C. cenotea</i> (Ach.) Schaer.	QC
<i>C. chlorophaea</i> (Sommerf.) Spreng.	QC
<i>C. coccifera</i> (L.) Willd.	QC
<i>C. coniocraea</i> (Flörke) Spreng.	QC
<i>C. crispata</i> (Ach.) Flot.	QC
<i>C. cristatella</i> Tuck.	ME; QC
<i>C. cyanipes</i> (Sommerf.) Nyl.	QC
<i>C. decorticata</i> (Flörke) Spreng.	QC
<i>C. deformis</i> (L.) Hoffm.	QC
<i>C. digitata</i> (L.) Hoffm.	QC
<i>C. ecmocyna</i> Leight. subsp. <i>ecmocyna</i>	QC
<i>C. furcata</i> (Huds.) Schrad.	QC
<i>C. glauca</i> Flörke	QC
<i>C. gracilis</i> (L.) Willd. subsp. <i>gracilis</i>	QC
<i>C. macilenta</i> Hoffm.	ME
<i>C. macilenta</i> var. <i>bacillaris</i> (Genth) Schaer.	QC
<i>C. macroceras</i> (Delise) Hav.	QC
<i>C. macrophylla</i> (Schaer.) Stenh.	QC
<i>C. maxima</i> (Asahina) Ahti	QC
<i>C. mitis</i> Sandst.	ME
<i>C. multiformis</i> G. Merr.	QC
<i>C. phyllophora</i> Hoffm.	QC
<i>C. pleurota</i> (Flörke) Schaer.	ME; QC
<i>C. polycarpoides</i> Nyl. ¹	ME
<i>C. pyxidata</i> (L.) Hoffm.	ME; QC
<i>C. rangiferina</i> (L.) F.H. Wigg.	ME
<i>C. rei</i> Schaer. ¹	ME
<i>C. scabriuscula</i> (Delise) Nyl.	QC
<i>C. squamosa</i> Hoffm.	ME; QC
<i>C. stricta</i> (Nyl.) Nyl. var. <i>uliginosa</i> Ahti	QC
<i>C. subulata</i> (L.) F.H. Wigg.	QC

Appendix 1. Continued.

Species	Occurrence
<i>C. sulphurina</i> (Michx.) Fr.	QC
<i>C. symphyrcarpia</i> (Flörke) Fr.	ME
<i>C. turgida</i> Hoffm.	ME; QC
<i>C. uncialis</i> (L.) F.H. Wigg.	ME; QC
<i>C. wainioi</i> Savicz	QC
<i>Coelocaulon aculeatum</i> (Schreb.) Link ¹	QC
<i>Collema subflaccidum</i> Degel.	ME
<i>Dactylospora urceolata</i> (Th.Fr.) Arnold	QC
<i>Dermatocarpon luridum</i> (With.) J.R. Laundon	QC
<i>D. miniatum</i> (L.) W. Mann	ME
<i>D. rivulorum</i> (Arnold) Dalla Torre & Sarnth.	QC
<i>Dibaeis baeomyces</i> (L. f.) Rambold & Hertel	ME
<i>Diploschistes scruposus</i> (Schreb.) Norman	QC
<i>Endococcus propinquus</i> (Körb.) D. Hawksw.	QC
<i>E. rugulosus</i> (Leight.) Nyl.	QC
<i>Ephebe lanata</i> (L.) Vain.	QC
<i>Flavoparmelia caperata</i> (L.) Hale	ME
<i>Fuscidea lowensis</i> (H. Magn.) R.A. Anderson & Hertel	QC
<i>Hypogymnia physodes</i> (L.) Nyl.	QC
<i>H. tubulosa</i> (Schaer.) Hav.	QC
<i>H. vittata</i> (Ach.) Parrique	QC
<i>Icmadophila ericetorum</i> (L.) Zahlbr.	QC
<i>Imshaugia aleurites</i> (Ach.) S.F. Meyer	QC
<i>Ionaspis odora</i> (Ach.) Th.Fr.	QC
<i>Lecanora argentea</i> Oksner & Volkova	ME
<i>L. dispersa</i> (Pers.) Sommerf.	ME
<i>L. epibryon</i> (Ach.) Ach.	QC
<i>L. hagenii</i> (Ach.) Ach.	QC
<i>L. placidensis</i> (H. Magn.) Knoph, Leuckert & Rambold	QC
<i>L. polytropa</i> (Hoffm.) Rabenh.	ME; QC
<i>Lecidea brunneofusca</i> H. Magn.	QC
<i>L. pycnocarpa</i> (Körb.) Ohlert	QC
<i>L. tessellata</i> Flörke	QC
<i>L. umbonata</i> (Hepp) Mudd	QC
<i>Lecidella carpathica</i> Körb.	QC
<i>L. euphorea</i> (Flörke) Hertel	QC
<i>L. stigmatea</i> (Ach.) Hertel & Leuckert	ME; QC
<i>L. wulfenii</i> (Hepp) Körb.	QC
<i>Lecidoma demissum</i> (Rutstr.) Gotth. Schneid. & Hertel	QC
<i>Lepraria caesioalba</i> (B. de Lesd.) J.R. Laundon	ME
<i>L. incana</i> (L.) Ach.	QC
<i>L. neglecta</i> (Nyl.) Erichsen	ME
<i>L. normandinoides</i> Lendemer & R.C. Harris	ME
<i>Leptogium cyanescens</i> (Rabenh.) Körb.	ME
<i>Lithographa tesserata</i> (DC.) Nyl.	QC

Appendix 1. Continued.

Species	Occurrence
<i>Lobaria pulmonaria</i> (L.) Hoffm.	ME
<i>Melanelia stygia</i> (L.) Essl.	QC
<i>Miriquidica leucophaea</i> (Rabenh.) Hertel & Rambold	QC
<i>M. plumbeoatra</i> (Vain.) A.J. Schwab & Rambold	QC
<i>Muellerella lichenicola</i> (Fr.) D. Hawksw.	QC
<i>Mycobilimbia berengeriana</i> (A. Massal.) Hafellner & V. Wirth	QC
<i>M. hypnorum</i> (Lib.) Kalb & Hafellner	QC
<i>Mycoblastus alpinus</i> (Fr.) Kernst.	QC
<i>M. sanguinarius</i> (L.) Norman	QC
<i>Nephroma parile</i> (Ach.) Ach.	ME
<i>Omphalina hudsoniana</i> (H.S. Jenn.) H.E. Bigelow ¹	QC
<i>Ophioparma lapponica</i> (Räsänen) Hafellner & R.W. Rogers	QC
<i>Pannaria rubiginosa</i> (Ach.) Bory	ME
<i>Parmelia saxatilis</i> (L.) Ach.	ME
<i>P. sulcata</i> Taylor	ME
<i>Parmotrema crinitum</i> (Ach.) M. Choisy	ME
<i>Peltigera didactyla</i> (With.) J.R. Laundon	ME
<i>P. rufescens</i> (Weiss) Humb.	ME
<i>Pertusaria amara</i> (Ach.) Nyl.	ME
<i>Phaeophyscia adiastrata</i> (Essl.) Essl.	ME
<i>P. rubropulchra</i> (Degel.) Essl.	ME
<i>P. sciastra</i> (Ach.) Moberg	ME
<i>Physcia caesia</i> (Hoffm.) Fűrnr.	ME
<i>Placynthiella icmalea</i> (Ach.) Coppins & P. James	ME
<i>Polyblastia melaspora</i> (Taylor) Zahlbr.	QC
<i>Porpidia subsimplex</i> (H. Magn.) Fryday	ME
<i>Psorula rufonigra</i> (Tuck.) Gotth. Schneid.	ME
<i>Rhizocarpon geminatum</i> Körb.	ME
<i>R. obscuratum</i> (Ach.) A. Massal.	ME
<i>Rinodina mmiaroeiza</i> (Nyl.) Arnold	QC
<i>Scoliosporum umbrinum</i> (Ach.) Arnold	ME
<i>S. umbrinum</i> var. <i>compacta</i> (Körb.) Vězda	QC
<i>Spilonema revertens</i> Nyl.	ME
<i>Stereocaulon glaucescens</i> Tuck.	ME
<i>Xanthoparmelia cumberlandia</i> (Gyeln.) Hale	ME
<i>X. plittii</i> (Gyeln.) Hale	ME
<i>Xanthoria elegans</i> (Link) Th.Fr.	ME
<i>X. parietina</i> (L.) Th.Fr.	ME

APPENDIX 2

List of mosses (Bryophyta) and liverworts (Marchantiophyta) recorded for serpentine soils in eastern North America. Occurrences based on Belland and Brassard (1988), and Roberts (1992) for Newfoundland, Canada (NL); Briscoe et al. (2009) for Maine, U.S.A. (ME); Sirois (1984) for Québec, Canada (QC); Shaw (1991), Shaw and Albright (1990), and Robinson (1966) for Maryland, U.S.A. (MD). Names standardized using Integrated Taxonomic Information System [website (<http://www.itis.gov>); accessed 28 Jan 2008].

Species	Occurrence
BRYOPHYTA	
<i>Amblystegium serpens</i> (Hedw.) Schimp. in BSG	ME
<i>Anastrophyllum minutum</i> (Schreb.) Schust.	QC
<i>A. saxicola</i> (Schrad.) Schust.	QC
<i>Andreaea rothii</i> var. <i>rothii</i> Web. & Mohr	QC
<i>Anomodon rostratus</i> (Hedw.) Schimp.	ME
<i>Aulacomnium androgynum</i> (Hedw.) Schwaegr.	ME
<i>A. palustre</i> (Hedw.) Schwaegr.	QC
<i>Bartramia pomiformis</i> Hedw.	ME
<i>Brachythecium calcareum</i> Kindb.	QC
<i>B. oedipodium</i> (Mitt.) Jaeg.	QC
<i>B. populeum</i> (Hedw.) Schimp. in BSG	QC
<i>B. reflexum</i> (Starke) Schimp. in BSG	QC
<i>B. rutabulum</i> (Hedw.) Schimp. in BSG	ME; QC
<i>B. velutinum</i> (Hedw.) Schimp. in BSG	ME; QC
<i>Bryum amblyodon</i> C. Muell.	QC
<i>B. argenteum</i> Hedw.	MD; QC
<i>B. knowltonii</i> Barnes	QC
<i>B. lisae</i> var. <i>cuspidatum</i> (Bruch & Schimp. in BSG) Marg.	QC
<i>B. pseudotriquetrum</i> (Hedw.) G. Gaertn., B. Mey. & Scherb.	QC
<i>B. reedii</i> Robins.	MD
<i>Callicladium haldanianum</i> (Grev.) Crum	ME
<i>Calliargon stramineum</i> (Brid.) Kindb.	QC
<i>Campylium chrysophyllum</i> (Brid.) J. Lange	ME
<i>C. hispidulum</i> (Brid.) Mitt.	QC
<i>C. stellatum</i> (Hedw.) C. Jens.	QC
<i>Catoscopium nigratum</i> (Hedw.) Brid.	NL
<i>Ceratodon purpureus</i> (Hedw.) Brid.	ME; QC
<i>Cirriphyllum piliferum</i> (Hedw.) Grout	QC
<i>Cynodontium alpestre</i> (Wahlenb.) Milde	QC
<i>Dicranum acutifolium</i> (Lindb. & Arnell.) Weinm.	QC
<i>D. bonjeanii</i> De Not.	QC
<i>D. elongatum</i> Schwaegr.	QC
<i>D. fragilifolium</i> Lindb.	QC
<i>D. fuscescens</i> Turner	QC
<i>D. majus</i> Sm.	QC

Appendix 2. Continued.

Species	Occurrence
<i>D. montanum</i> Hedw.	ME; QC
<i>D. ontariense</i> W.L. Peterson	QC
<i>D. polysetum</i> Sw.	ME; QC
<i>D. scoparium</i> Hedw.	ME; QC
<i>Funaria flavicans</i> Michx.	MD
<i>F. hygrometrica</i> Hedw.	ME
<i>Hedwigia ciliata</i> (Hedw.) P. Beauv.	ME
<i>Herzogiella striatella</i> (Brid.) Iwats.	ME
<i>Hylocomiastrum pyrenaicum</i> (Spruce) Fleisch.	QC
<i>H. umbratum</i> (Hedw.) Fleisch.	QC
<i>Hylocomium splendens</i> (Hedw.) Schimp. in BSG	NL; QC
<i>Hymenostylium recurvirostre</i> (Hedw.) Dixon	QC
<i>Hypnum mammillatum</i> (Brid.) Loeske	ME
<i>H. cupressiforme</i> Hedw.	ME
<i>H. imponens</i> Hedw.	ME
<i>H. pallescens</i> (Hedw.) P. Beauv.	ME; QC
<i>Isopterygiopsis pulchella</i> (Hedw.) Iwats.	QC
<i>Isothecium myosuroides</i> Brid.	ME
<i>Kiaeria glacialis</i> (Berggr.) I. Hagen	QC
<i>Lejeunea cavifolia</i> (Ehrh.) Lindb. emend. Buch	NL
<i>Leucobryum glaucum</i> (Hedw.) Angstr.	ME
<i>Limprichtia revolvens</i> (Sw.) Loeske	QC
<i>Loeskeobryum brevirostre</i> (Brid.) Fleisch.	QC
<i>Paludella squarrosa</i> (Hedw.) Brid.	QC
<i>Paraleucobryum longifolium</i> (Hedw.) Loeske	QC
<i>Philonotis fontana</i> (Hedw.) Brid.	QC
<i>Plagiothecium laetum</i> Schimp. in BSG	QC
<i>Platygyrium repens</i> (Brid.) Schimp. in BSG	ME
<i>Pleurozium schreberi</i> (Brid.) Mitt.	ME; NL; QC
<i>Pohlia cruda</i> (Hedw.) Lindb.	QC
<i>P. nutans</i> (Hedw.) Lindb.	QC
<i>P. sphagnicola</i> (Bruch & Schimp. in BSG) Lindb. & Arnell	QC
<i>Polytrichastrum alpinum</i> var. <i>alpinum</i> (Hedw.) G.L. Sm.	QC
<i>Polytrichum commune</i> Hedw.	QC
<i>P. formosum</i> Hedw.	QC
<i>P. juniperinum</i> Hedw.	ME; QC
<i>P. longisetum</i> Brid.	QC
<i>P. piliferum</i> Hedw.	ME
<i>P. strictum</i> Brid.	QC
<i>Pterigynandrum filiforme</i> Hedw.	ME
<i>Ptilium crista-castrensis</i> (Hedw.) De Not.	QC
<i>Pylaisiella polyantha</i> (Hedw.) Grout	QC
<i>Rhacomitrium heterostichum</i> (Hedw.) Brid.	QC
<i>R. lanuginosis</i> (Hedw.) Brid.	NL; QC
<i>Rhytidiadelphus squarrosus</i> (Hedw.) Warnst.	QC

Appendix 2. Continued.

Species	Occurrence
<i>R. triquetrus</i> (Hedw.) Warnst.	ME
<i>Rhytidium rugosum</i> (Hedw.) Kindb.	QC
<i>Sanionia uncinata</i> var. <i>uncinata</i> (Hedw.) Loeske	QC
<i>Schistidium apocarpum</i> (Hedw.) Bruch & Schimp. in BSG	QC
<i>Sphagnum angustifolium</i> (Russow) C. Jens.	QC
<i>S. capillifolium</i> (Ehrh.) Hedw.	QC
<i>S. centrale</i> C. Jens.	QC
<i>S. fuscum</i> (Schimp.) H. Klinggr.	QC
<i>S. girgensohnii</i> Russow	QC
<i>S. lindbergii</i> Schimp.	QC
<i>S. rubellum</i> Wils.	QC
<i>S. russowii</i> Warnst.	QC
<i>S. teres</i> (Schimp.) Ångstr.	QC
<i>S. warnstorffii</i> Russow	QC
<i>Splachnum sphaericum</i> Hedw.	QC
<i>Tetraphis pellucida</i> Hedw.	QC
<i>Tetraplodon angustatus</i> (Hedw.) Bruch & Schimp. in BSG	NL
<i>T. mnioides</i> (Hedw.) Bruch & Schimp. in BSG	QC
<i>Thuidium recognitum</i> (Hedw.) Lindb.	ME
<i>Tomenthypnum nitens</i> (Hedw.) Loeske	QC
<i>Uloa hutchinsiae</i> (Sm.) Hammar	ME
<i>Warnstorffia exannulata</i> var. <i>exannulata</i> (Schimp. in BSG) Loeske	QC
<i>Weissia controversa</i> Hedw.	ME; QC
MARCHANTIOPHYTA	
<i>Anastrophyllum minutum</i> (Schreb.) Schust.	ME
<i>Barbilophozia atlantica</i> (Kaal.) K. Mull.	QC
<i>B. attenuata</i> (Mart.) Loeske	QC
<i>B. barbata</i> (Schreb.) Loeske	ME
<i>B. floerkei</i> (Web. & Mohr) Loeske	QC
<i>B. hatcheri</i> (Evans) Loeske	QC
<i>B. kunzeana</i> (Huebener) Gams	QC
<i>B. lycopodioides</i> (Wallr.) Loeske	QC
<i>Blepharostoma trichophyllum</i> (L.) Dumort.	QC
<i>Calypogeia sphagnicola</i> (Arnell & J. Perss.) Warnst. & Loeske	QC
<i>Cephalozia connivens</i> (Dicks.) Lindb.	QC
<i>C. lunulifolia</i> (Dumort.) Dumort.	QC
<i>C. pleniceps</i> (Aust.) Lindb.	QC
<i>Cephaloziella hampeana</i> (Nees) Schiffn.	ME
<i>C. rubella</i> (Nees) Warnst.	QC
<i>Chandonanthus setiformis</i> (Ehrh.) Lindb.	QC
<i>Cladopodiella fluitans</i> (Nees) Joerg.	QC
<i>Cololejeunea biddlecomiae</i> (Aust.) Evans	ME
<i>Frullania tamarisci</i> subsp. <i>asagrayana</i> (Mont.) Hatt.	ME

Appendix 2. Continued.

Species	Occurrence
<i>Gymnocolea inflata</i> (Huds.) Dumort.	QC
<i>Harpanthus flotovianus</i> (Nees) Nees	QC
<i>Lejeunea lamacerina</i> (Steph.) Schiffn. subsp. <i>geminata</i> Schust.	ME
<i>L. cavifolia</i> (Ehrh.) Lindb. <i>emend.</i> Buch	ME
<i>Lophocolea heterophylla</i> (Schrad.) Dumort.	ME
<i>Lophozia alpestris</i> (Web.) Evans	QC
<i>L. ascendens</i> (Warnst.) Schust.	QC
<i>L. bicrenata</i> (Hoffm.) Dumort.	QC
<i>L. ventricosa</i> (Dicks.) Dumort.	QC
<i>Metzgeria conjugata</i> Lindb.	ME
<i>M. furcata</i> (L.) Dumort.	ME
<i>Mylia anomala</i> (Hook.) S. Gray	QC
<i>Odontoschisma elongatum</i> (Lindb.) A.W. Evans	QC
<i>O. denudatum</i> (Mart.) Dumort.	QC
<i>O. macounii</i> (Aust.) Underw.	QC
<i>Pellia endiviifolia</i> (Dicks.) Dumort.	QC
<i>Ptilidium ciliare</i> (L.) Nees	ME; QC
<i>P. pulcherrimum</i> (G. Web.) Hampe	QC
<i>Radula complanata</i> (L.) Dumort.	ME
<i>Scapania curta</i> (Mart.) Dumort.	QC
<i>S. irrigua</i> (Nees) Gottsche, Lindenb. & Nees	QC
<i>S. irrigua</i> subsp. <i>rufescens</i> (Loeske) Schust.	QC
<i>S. paludosa</i> (K. Mull.) K. Mull.	QC
<i>Tritomaria quinquedentata</i> (Huds.) Buch	QC

APPENDIX 3

Vascular plant species recorded from serpentine outcrops in eastern North America, followed by literature citations and protected status. Names standardized using Integrated Taxonomic Information System (<http://www.itis.gov>; accessed 23 Jan 2008), International Plant Names Index (<http://www.ipni.org>; accessed 23 Jan 2008), and USDA, NRCS (2008). Protected status data from USDA, NRCS (2008), Center for Plant Conservation (<http://www.centerforplantconservation.org>; accessed 28 Jan 2008), and NatureServe (<http://www.natureserve.org>; accessed 28 Jan 2008). Global (G1 = critically imperiled, G2 = imperiled); federal = USA; and U.S. state or Canadian province, both as postal abbreviations. U.S. locations are followed by rarity designations: (E) = endangered, (EV) = exploitably vulnerable, (H) = historical, (PREX) = probably extirpated, (PRX) = presumed extirpated, (PX) = possibly extirpated, (T) = threatened, (R) = rare, (S) = sensitive, (SC) = special concern, and (X) = extirpated. For Canadian locations, rarity designations are: (S1) = critically imperiled, (S2) = imperiled, (S3) = vulnerable, (S4) = apparently secure, (S5) = secure, (SX) = extinct, and (SH) = possibly extinct. Canadian protected status data for Newfoundland are from Atlantic Canada Conservation Data Center (<http://www.accdc.com/products/ranking.html>; accessed 24 Jan 2008) and, for Québec, from Centre de données sur le patrimoine naturel du Québec (<http://www.cdpnq.gouv.qc.ca/produits-en.htm>; accessed 23 Jan 2008).

Taxon	Citation	Global, Federal, State/Province Protected Status
ACERACEAE		
<i>Acer rubrum</i> L.	Brooks 1987; Carter 1979; Dearden 1979; Mansberg and Wentworth 1984; Miller 1981; Tyndall and Farr 1990; Wherry 1963; Zika and Dann 1985	–
<i>A. saccharum</i> Marshall	Brooks 1987; Carter 1979	–
<i>A. spicatum</i> Lam.	Brooks 1987; Carter 1979	KY (E)
ADOXACEAE		
<i>Viburnum acerifolium</i> L.	Miller 1981	–
<i>V. lantanoides</i> Michx.	Carter 1979	KY (E), NJ (E)
<i>V. nudum</i> var. <i>cassinoides</i> (L.) Torr. & A. Gray	Brooks 1987; Mansberg and Wentworth 1984; Zika and Dann 1985	IN (E)
<i>V. prunifolium</i> L.	Wherry 1963	CT (SC)
AGAVACEAE		
<i>Yucca constricta</i> Buckley	Maoui 1966	–
ANACARDIACEAE		
<i>Rhus copallina</i> L.	Miller 1981; Wherry 1963	–

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>R. glabra</i> L.	Tyndall and Hull 1999; Wherry 1963	QC (SH)
<i>R. hirta</i> L.	Miller 1981; Wherry 1963	—
<i>Robinia pseudoacacia</i> L.	Brooks 1987; Miller 1981	—
<i>Toxicodendron radicans</i> (L.) Kuntze subsp. <i>radicans</i>	Tyndall and Hull 1999; Wherry 1963	—
APIACEAE		
<i>Angelica venenosa</i> (Greenway) Fernald	Wherry 1963	CT (SC)
<i>Cicuta maculata</i> L.	Carter 1979; Wherry 1963	—
<i>Conioselinum chinense</i> Britton, Sterns & Poggenb.	Dearden 1979	IL (E), IN (E), MA (SC), NJ (E), NC (E), PA (E), WI (E)
<i>Heracleum maximum</i> Bartr.	Carter 1979	KY (E), TN (SC)
<i>Ligusticum scoticum</i> L. subsp. <i>hultenii</i> (Fernald) Calder & Roy L. Taylor	Carter 1979	—
<i>Sanicula marilandica</i> L.	Wherry 1963	WA (S)
<i>Thaspium trifoliatum</i> (L.) A. Gray	Mansberg and Wentworth 1984	MD (E)
<i>Zizia aptera</i> (A. Gray) Fernald	Wherry 1963	CT (E), IN (R), MI (T), RI (H)
<i>Z. aurea</i> Koch	Wherry 1963	—
APOCYNACEAE		
<i>Apocynum cannabinum</i> L.	Brooks 1987	—
<i>A. × floribundum</i> Greene	Wherry 1963	—
AQUIFOLIACEAE		
<i>Ilex ambigua</i> Elliott	Brooks 1987	—
<i>Nemopanthus mucronatus</i> (L.) Trel.	Brooks 1987; Dearden 1979	—
ARACEAE		
<i>Symplocarpus foetidus</i> (L.) W.P.C. Barton	Carter 1979	TN (E)
ARALIACEAE		
<i>Aralia nudicaulis</i> L.	Brooks 1987; Carter 1979	—

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
ARISTOLOCHIACEAE		
<i>Hexastylis arifolia</i> Small var. <i>ruthii</i> (Ashe) H.L. Blomq.	Mansberg and Wentworth 1984; Tyndall and Hull 1999	FL (T)
ASCLEPIADACEAE		
<i>Asclepias purpurascens</i> L.	Wherry 1963	CT (SC), MA (E), RI (H), TN (SC), WI (E)
<i>A. syriaca</i> L.	Wherry 1963	
<i>A. verticillata</i> L.	Brooks 1987; Tyndall 1992b, 1994, 1999; Wherry 1963	MA (T)
<i>A. viridiflora</i> Pursh	Brooks 1987; Tyndall 1992b, 1994; Wherry 1963	CT (SC), FL (E), NY (T)
ASPLENIACEAE		
<i>Asplenium platyneuron</i> (L.) Britton, Sterns & Poggenb.	Tyndall and Farr 1990; Wherry 1963	ME (SC), NY (EV); QC (S2)
<i>A. trichomanes</i> L.	Zika and Dann 1985	MN (T), NY (EV)
<i>A. trichomanes-ramosum</i> L.	Carter 1979; Zika and Dann 1985	ME (T), MI (T), NY (E), VT (T), WI (E)
ASTERACEAE		
<i>Achillea millefolium</i> L.	Carter 1979; Miller 1981; Tyndall and Farr 1990; Zika and Dann 1985	—
<i>A. millefolium</i> var. <i>borealis</i> (Bong.) Farw.	Pennell 1930	ME (SC)
<i>Ageratina aromatica</i> (L.) Spach var. <i>aromatica</i>	Tyndall and Farr 1990; Wherry 1963	MA (E)
<i>Ambrosia artemisiifolia</i> L.	Carter 1979; Wherry 1963	—
<i>Anaphalis margaritacea</i> (L.) Benth. & Hook. f.	Carter 1979	—
<i>Antennaria howellii</i> Greene subsp. <i>neodioica</i> (Greene) Bayer	Wherry 1963	—
<i>A. neglecta</i> Greene	Carter 1979; Wherry 1963	—
<i>A. plantaginifolia</i> (L.) Hook. <i>Artemisia campestris</i> (L.) subsp. <i>borealis</i> (Pall) H.M. Hall & Clem.	Carter 1979; Wherry 1963 Brooks 1987	MA (E), ME (PX), NY (E)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Aster umbellatus</i> Mill.	Carter 1979	—
<i>Cirsium discolor</i> (Willd.) Spreng.	Wherry 1963	—
<i>C. muticum</i> Michx.	Carter 1979; Wherry 1963	AR (T)
<i>C. pumilum</i> Spreng.	Wherry 1963	—
<i>C. vulgare</i> (Savi) Ten.	Miller 1981	—
<i>Erechtites hieracifolia</i> (L.) DC.	Tyndall 2005; Wherry 1963	—
<i>Erigeron strigosus</i> Willd.	Brooks 1987; Carter 1979	—
<i>Eupatorium maculatum</i> L.	Carter 1979	—
<i>E. perfoliatum</i> L.	Brooks 1987; Carter 1979; Wherry 1963	—
<i>E. purpureum</i> L.	Wherry 1963	—
<i>E. rotundifolium</i> L. var. <i>ovatum</i> (Bigelow) Torr.	Wherry 1963	NH (E), NY (E)
<i>Eurybia radula</i> (Aiton) G.L. Nesom	Brooks 1987	CT (E), KY (E), MD (E), NJ (E), NY (E)
<i>Euthamia graminifolia</i> (L.) Nutt. var. <i>graminifolia</i>	Carter 1979	—
<i>E. graminifolia</i> var. <i>nuttallii</i> (Greene) W. Stone	Wherry 1963	—
<i>Helianthus divaricatus</i> L.	Wherry 1963	QC (S3)
<i>H. giganteus</i> L.	Wherry 1963	IL (E)
<i>Heliopsis helianthoides</i> Sweet	Wherry 1963	—
<i>Hieracium aurantiacum</i> L.	Carter 1979	—
<i>H. caespitosum</i> Dumort.	Carter 1979	—
<i>H. canadense</i> Michx.	Carter 1979	—
<i>H. gronovii</i> L.	Wherry 1963	—
<i>H. pilosella</i> L.	Carter 1979	—
<i>H. piloselloides</i> Vill.	Carter 1979	—
<i>H. venosum</i> L.	Brooks 1987; Wherry 1963	ME (E)
<i>Krigia virginica</i> Willd.	Wherry 1963	IA (E), ME (PX), OH (T)
<i>Lactuca biennis</i> (Moench) Fernald	Wherry 1963	—
<i>L. canadensis</i> L.	Pennell 1930; Tyndall and Hull 1999	—
<i>Leucanthemum vulgare</i> Lam.	Carter 1979	—
<i>Liatris pilosa</i> Willd. var. <i>pilosa</i>	Brooks 1987	—
<i>L. spicata</i> Willd.	Wherry 1963	—

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Oclemena acuminata</i> (Michx.) Greene	Carter 1979	KY (T), OH (PRX)
<i>Omalotheca sylvatica</i> (L.) Sch.Bip. & F.W. Schultz	Brooks 1987	ME (SC), MI (T), NY (E), VT (E)
<i>Packera anonyma</i> (Wood) W.A. Weber & Á. Löve	Brooks 1987; Pennell 1930; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	PA (R)
<i>P. paupercula</i> (Michx.) Á. Löve & D. Löve	Brooks 1987; Pennell 1930; Roberts 1980, 1992	CT (E), NH (T), OH (T)
<i>P. plattensis</i> (Nutt.) W.A. Weber & Á. Löve	Mansberg and Wentworth 1984; Tyndall and Hull 1999	PA (X)
<i>Prenanthes alba</i> L.	Wherry 1963	KY (E)
<i>P. altissima</i> L.	Carter 1979	—
<i>P. nana</i> (Bigelow) DC.	Brooks 1987	ME (E), NY (E)
<i>P. serpentaria</i> Pursh	Wherry 1963	MA (E)
<i>P. trifoliata</i> (Cass.) Fernald	Carter 1979	OH (E)
<i>Pseudognaphalium</i> <i>obtusifolium</i> (L.) Hilliard & B.L. Burtt subsp. <i>obtusifolium</i>	Tyndall 2005; Wherry 1963	—
<i>Senecio sylvaticus</i> DC.	Carter 1979	—
<i>Sericocarpus asteroides</i> Britton, Sterns & Poggenb.	Brooks 1987; Wherry 1963	ME (E)
<i>Solidago bicolor</i> L.	Carter 1979; Wherry 1963	—
<i>S. caesia</i> L.	Wherry 1963	WI (E)
<i>S. caesia</i> var. <i>curtisii</i> (Torr. & A. Gray)	Mansberg and Wentworth 1984	—
<i>S. canadensis</i> L.	Brooks 1987; Carter 1979	—
<i>S. canadensis</i> var. <i>scabra</i> (Willd.) Torr. & A. Gray	Wherry 1963	—
<i>S. hispida</i> Willd.	Brooks 1987; Dearden 1979; Roberts 1992	MD (E, X)
<i>S. juncea</i> Aiton	Brooks 1987; Wherry 1963	—
<i>S. macrophylla</i> Pursh	Carter 1979	MA (T)
<i>S. multiradiata</i> Aiton	Brooks 1987	ME (T)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>S. nemoralis</i> Aiton	Carter 1979; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	—
<i>S. rugosa</i> Mill.	Carter 1979; Wherry 1963	—
<i>S. sempervirens</i> L.	Carter 1979	—
<i>S. simplex</i> subsp. <i>simplex</i> var. <i>chlorolepis</i> (Fernald) G.S. Ringius	Bouchard et al. 1983; Canadian Legal Information Institute 2008	QC (S1)
<i>S. simplex</i> Kunth var. <i>randii</i> (Porter) Kartesz & Gandhi	Brooks 1987	KY (SC), MA (E), TN (T)
<i>S. uliginosa</i> Nutt. var. <i>linoides</i> (Torr. & A. Gray) Fernald	Brooks 1987	NH (T)
<i>Sonchus arvensis</i> L.	Carter 1979	—
<i>Symphytotrichum cordifolium</i> (L.) G.L. Nesom	Wherry 1963	—
<i>S. depauperatum</i> (Fernald) G.L. Nesom	Brooks 1987; Gustafson and Latham 2005; Hart 1980; Miller 1981; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	G2; MD (E), PA (T)
<i>S. dumosum</i> (L.) G.L. Nesom var. <i>dumosum</i>	Wherry 1963	IA (E), OH (T)
<i>S. ericoides</i> (L.) G.L. Nesom var. <i>ericoides</i>	Brooks 1987	TN (T)
<i>S. foliaceum</i> (DC.) G.L. Nesom var. <i>foliaceum</i>	Brooks 1987	—
<i>S. laeve</i> (L.) Á. Löve & D. Löve var. <i>laeve</i>	Tyndall and Hull 1999; Wherry 1963	—
<i>S. laeve</i> var. <i>concinnum</i> (Willd.) G.L. Nesom	Tyndall and Hull 1999	MD (E, X), NY (E)
<i>S. lateriflorum</i> (L.) Á. Löve & D. Löve var. <i>lateriflorum</i>	Reed 1986	NY (E)
<i>S. novi-belgii</i> (L.) G.L. Nesom var. <i>novi-belgii</i>	Brooks 1987; Carter 1979	PA (T)
<i>S. novi-belgii</i> var. <i>villicaule</i> (A. Gray) Labrecque & Brouillet	Brooks 1987	QC (S1)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>S. patens</i> (Aiton) G.L. Nesom var. <i>patens</i>	Wherry 1963	ME (PX), NH (T)
<i>S. pilosum</i> (Willd.) G.L. Nesom var. <i>pilosum</i>	Hart 1980	
<i>S. pilosum</i> (A. Gray) G.L. Nesom var. <i>pringlei</i>	Wherry 1963	NY (T); QC (S1)
<i>S. puniceum</i> (L.) Á. Löve & D. Löve var. <i>puniceum</i>	Carter 1979; Wherry 1963	NJ (E), NY (E), PA (T)
<i>S. undulatum</i> (L.) G.L. Nesom	Mansberg and Wentworth 1984; Wherry 1963	–
<i>Taraxacum officinale</i> F.H. Wigg.	Carter 1979; Tyndall and Farr 1990	–
<i>Vernonia glauca</i> (L.) Willd.	Wherry 1963	NJ (E), PA (E)
<i>V. noveboracensis</i> (L.) Michx.	Brooks 1987; Wherry 1963	KY (SC), OH (PRX)
<i>Xanthium strumarium</i> L.	Tyndall and Farr 1990	–
BALSAMINACEAE		
<i>Impatiens capensis</i> Meerb.	Carter 1979; Wherry 1963	–
BERBERIDACEAE		
<i>Berberis vulgaris</i> L.	Carter 1979	–
<i>Mahonia trifoliolata</i> Fedde	Maoui 1966	–
BETULACEAE		
<i>Alnus incana</i> (L.) Moench subsp. <i>rugosa</i> (Du Roi) R.T. Clausen	Brooks 1987	IL (E)
<i>A. serrulata</i> (Aiton) Willd.	Wherry 1963	QC (S1)
<i>A. viridis</i> (Chaix) DC. subsp. <i>crispa</i> (Aiton) Turrill	Carter 1979; Dearden 1979	MA (T), PA (E), TN (SC)
<i>Betula alleghaniensis</i> Britton var. <i>alleghaniensis</i>	Brooks 1987; Mansberg and Wentworth 1984	IL (E)
<i>B. nana</i> L.	Brooks 1987; Dearden 1979	–
<i>B. papyrifera</i> Marshall	Brooks 1987; Carter 1979; Dearden 1979; Tyndall and Hull 1999; Zika and Dann 1985	–
<i>B. papyrifera</i> var. <i>cordifolia</i> (Regel) Fernald	Carter 1979	TN (E)
<i>B. pubescens</i> Ehrh. subsp. <i>borealis</i> (Spach) A. Löve & D. Löve	Zika and Dann 1985	–

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>B. pumila</i> L.	Brooks 1987; Dearden 1979; Roberts 1980	CT (SC), IA (T), MA (E), ME (SC), NH (E), NY (T), OH (T)
<i>Carpinus caroliniana</i> Walter subsp. <i>virginiana</i> (Marshall) Furlow	Wherry 1963	—
<i>Corylus americana</i> Walter	Tyndall and Hull 1999; Wherry 1963	QC (SH)
BRASSICACEAE		
<i>Arabis alpina</i> L.	Ryan 1988	—
<i>A. lyrata</i> L.	Brooks 1987; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	OH (T), VT (T)
<i>Cardamine bellidifolia</i> L.	Hay et al. 1992	ME (E), NH (E); NL (S1)
<i>C. diphylla</i> (Michx.) Alph. Wood	Carter 1979	QC (S4)
<i>C. pensylvanica</i> Willd.	Carter 1979	—
CAMPANULACEAE		
<i>Campanula rotundifolia</i> L.	Brooks 1987; Tyndall and Hull 1999; Zika and Dann 1985	NY (EV), OH (E)
<i>Lobelia inflata</i> L.	Brooks 1987	—
<i>L. spicata</i> Lam.	Tyndall and Farr 1990; Wherry 1963	—
<i>L. spicata</i> var. <i>scaposa</i> McVaugh	Brooks 1987	—
CAPRIFOLIACEAE		
<i>Diervilla lonicera</i> Mill.	Carter 1979	IN (R), TN (T)
<i>Linnaea borealis</i> L.	Dearden 1979	IA (T), IN (X), MD (E, X), NJ (E), OH (PRX), PA (T), RI (H), TN (PX, E)
<i>Lonicera canadensis</i> Marshall	Carter 1979	IN (X), MD (E), NJ (E), TN (SC)
<i>L. japonica</i> Thunb.	Miller 1981; Wherry 1963	—
<i>L. sempervirens</i> L.	Wherry 1963	ME (E)
<i>L. villosa</i> (Michx.) Schult.	Carter 1979; Dearden 1979	OH (PRX), PA (E)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>L. villosa</i> var. <i>calvescens</i> (Fernald & Wiegand) Fernald	Brooks 1987	—
<i>Sambucus nigra</i> L. subsp. <i>canadensis</i> (L.) R. Bolli	Wherry 1963	—
<i>S. racemosa</i> A. Gray var. <i>racemosa</i>	Carter 1979	IL (E), KY (E), RI (H)
CARYOPHYLLACEAE		
<i>Arenaria humifusa</i> Linden & Planch	Brooks 1987; Dearden 1979; Roberts 1980, 1992	—
<i>A. serpyllifolia</i> L.	Wherry 1963	—
<i>Cerastium alpinum</i> L.	Roberts 1980	—
<i>C. arvense</i> L.	Carter 1979; Zika and Dann 1985	—
<i>C. arvense</i> subsp. <i>strictum</i> (L.) Ugborogho	Hay et al. 1994	—
<i>C. arvense</i> var. <i>velutinum</i> (Raf.) Britton	Gustafson and Latham 2005; Hay et al. 1994	TN (E)
<i>C. arvense</i> var. <i>villosum</i> (Darl.) Hollick & Britton	Brooks 1987; Hart 1980; Pennell 1930; Ryan 1988; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	PA (E)
<i>C. beeringianum</i> Cham. & Schlecht.	Brooks 1987; Dearden 1979; Roberts 1980	—
<i>C. fontanum</i> Baumg. subsp. <i>vulgare</i> (Hartm.) Greuter & Burdet	Carter 1979; Hart 1980	—
<i>C. terrae-novae</i> Fernald & Wiegand	Brooks 1987; Dearden 1979; Hay et al. 1994; Tyndall and Hull 1999	—
<i>C. velutinum</i> Rafinesque var. <i>villosissimum</i> (Pennell) J.K. Morton [as <i>C. arvense</i> L. var. <i>villosissimum</i> Pennell]	Latham 1993; Gustafson et al. 2003; Morton 2004	PA (E)
<i>Dianthus armeria</i> L.	Brooks 1987	—
<i>Lychmis alpina</i> L.	Brooks 1987	—
<i>L. alpina</i> var. <i>americana</i> Fernald	Dearden 1979	—
<i>Minuartia biflora</i> (L.) Schinz & Thell.	Brooks 1987; Hay et al. 1994; Tyndall and Hull 1999	NL (S1)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>M. marcescens</i> ((Fernald) House	Brooks 1987; Dearden 1979; Roberts 1980, 1992; Sirois et al. 1988; Tyndall and Hull 1999; Zika and Dann 1985	G2; VT (T); NL (S2, S3), QC (S2)
<i>M. michauxii</i> (Fenzl) Farw. var. <i>michauxii</i>	Wherry 1963; Zika and Dann 1985	IN (R), NJ (E), RI (E)
<i>M. rubella</i> (Wahlenb.) Hiern.	Brooks 1987; Sirois et al. 1988	ME (T), VT (T)
<i>Moehringia macrophylla</i> (Hook.) Fenzl	Tyndall and Hull 1999; Zika and Dann 1985	CT (E), MA (E), MI (T), MN (T), WI (E); QC (S3)
<i>Sagina caespitosa</i> (J. Vahl) Lange	Tyndall and Hull 1999; Hay et al. 1994	NL (SH)
<i>S. nodosa</i> (L.) E. Mey.	Brooks 1987; Dearden 1979	MI (T)
<i>S. saginoides</i> (L.) Karst.	Tyndall and Hull 1999; Hay et al. 1994	NL (S1)
<i>Silene acaulis</i> L. Jacq.	Dearden 1979; Roberts 1980	ME (PX)
<i>S. acaulis</i> var. <i>exscapa</i> (All.) DC.	Brooks 1987	NH (T)
<i>S. stellata</i> (L.) W.T. Aiton	Wherry 1963	CT (SC), MI (T), RI (H)
<i>Spergularia rubra</i> J. Presl & C. Presl	Brooks 1987	–
CELASTRACEAE		
<i>Celastrus scandens</i> L.	Wherry 1963	NY (EV)
CHENOPODIACEAE		
<i>Atriplex prostrata</i> R. Br.	Carter 1979	–
<i>Suaeda maritima</i> (L.) Dumort.	Carter 1979	–
CISTACEAE		
<i>Helianthemum bicknellii</i> Fernald	Wherry 1963	KY (T), MD (E), OH (T), PA (E), TN (PX, E), VT (T)
<i>Lechea minor</i> L.	Wherry 1963	OH (T)
<i>L. pulchella</i> Raf. var. <i>pulchella</i>	Wherry 1963	MI (T), OH (T), TN (E)
<i>L. racemulosa</i> Michx.	Wherry 1963	IN (E), NY (R)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
CLUSIACEAE		
<i>Hypericum gentianoides</i> (L.) Britton, Sterns & Poggenb.	Brooks 1987; Miller 1981; Tyndall 1992b, 1994, 2005; Wherry 1963	IA (E)
<i>H. hypericoides</i> (L.) Crantz subsp. <i>hypericoides</i>	Wherry 1963	—
<i>H. perforatum</i> L.	Brooks 1987; Carter 1979	—
<i>H. punctatum</i> Lam.	Brooks 1987; Wherry 1963	—
COMMELINACEAE		
<i>Tradescantia virginiana</i> L.	Wherry 1963	—
CONVOLVULACEAE		
<i>Calystegia sepium</i> (L.) R. Br. subsp. <i>sepium</i>	Carter 1979	—
<i>C. spithamea</i> (L.) Pursh subsp. <i>spithamea</i>	Wherry 1963	NH (T)
CORNACEAE		
<i>Cornus alternifolia</i> L. f.	Wherry 1963	FL (E)
<i>C. canadensis</i> L.	Brooks 1987; Carter 1979; Dearden 1979	IA (T), IL (E), IN (E), MD (E), OH (T)
<i>C. florida</i> L.	Mansberg and Wentworth 1984	ME (E), NY (EV), VT (T)
CUPRESSACEAE		
<i>Juniperus communis</i> L.	Brooks 1987; Dearden 1979; Roberts 1980; Tyndall and Hull 1999; Zika and Dann 1985	IL (T), IN (R), MD (E, X), OH (E)
<i>J. communis</i> var. <i>depressa</i> Pursh	Brooks 1987	KY (T); NL (S4, S5)
<i>J. horizontalis</i> Moench	Brooks 1987; Dearden 1979; Roberts 1980	IA (T), IL (E), NH (E), NY (E), VT (T); NL (S5)
<i>J. virginiana</i> L.	Miller 1981; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	—
<i>Thuja occidentalis</i> L.	Carter 1979	CT (T), IL (T), IN (E), KY (T), MA (E), MD (T), NJ (E), TN (SC)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
CYPERACEAE		
<i>Bulbostylis capillaris</i> (Elliott) Fernald	Wherry 1963	—
<i>Carex annectens</i> E.P. Bicknell	Wherry 1963	—
<i>C. arctata</i> Hook.	Carter 1979	IN (E), NJ (E), OH (E)
<i>C. atlantica</i> L.H. Bailey subsp. <i>atlantica</i>	Wherry 1963	IN (T)
<i>C. atratiformis</i> Britton	Sirois et al. 1988	ME (SC), MI (T), NY (E), VT (T)
<i>C. bicknellii</i> Britton	Wherry 1963	ME (PX), NY (T), OH (T), PA (E)
<i>C. brunnescens</i> (Pers.) Poir. subsp. <i>sphaerostachya</i> (Tuck.) Kalela	Carter 1979	IL (E), IN (E), NJ (E), OH (T)
<i>C. bushii</i> Mack.	Wherry 1963	CT (SC), IN (E), MA (E), ME (PX), NJ (E), OH (E)
<i>C. buxbaumii</i> Wahlenb.	Brooks 1987; Dearden 1979	CT (E), KY (H), MD (T), NH (E), NY (T), PA (R), TN (SC), VT (E), WA (S)
<i>C. capitata</i> L. var. <i>arctogena</i> (Harry Sm.) Hiitonen	Sirois et al. 1988	NH (T)
<i>C. cephalophora</i> Willd.	Wherry 1963	QC (S2)
<i>C. communis</i> L.H. Bailey	Carter 1979	IL (T)
<i>C. conoidea</i> Willd.	Carter 1979; Sirosis et al. 1988	IN (E), MD (E), NC (T), OH (T)
<i>C. deflexa</i> Hornem.	Carter 1979	—
<i>C. echinata</i> Murray	Brooks 1987; Dearden 1979; Wherry 1963	IL (E), IN (E), OH (E)
<i>C. exilis</i> Dewey	Brooks 1987; Dearden 1979	CT (E), MD (E), NC (T), NH (T), WI (T)
<i>C. flaccosperma</i> Dewey var. <i>glaucoidea</i> (Olney) Kükenth.	Wherry 1963	—
<i>C. flava</i> L.	Sirois et al. 1988	IN (T), PA (T), WA (S)
<i>C. glaucoidea</i> Tuck.	Brooks 1987	MA (E)
<i>C. granularis</i> Willd.	Wherry 1963	NH (E)
<i>C. gynandra</i> Schwein.	Carter 1979	—
<i>C. hirsutella</i> Mack.	Wherry 1963	CT (SC), NH (E); QC (S2)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>C. hystericina</i> Willd.	Tyndall and Hull 1999; Wherry 1963	KY (H), MD (E), WA (S)
<i>C. interior</i> L.H. Bailey	Wherry 1963	–
<i>C. intumescens</i> Rudge	Carter 1979	IL (T)
<i>C. lachenalii</i> Schkuhr	Brooks 1987	–
<i>C. laxiculmis</i> Schwein.	Wherry 1963	ME (E), MN (T); QC (S1)
<i>C. laxiflora</i> Lam.	Carter 1979	–
<i>C. lenticularis</i> Michx. var. <i>lenticularis</i>	Sirois et al. 1988	MA (T), WI (T)
<i>C. limosa</i> L.	Dearden 1979	CT (E), IN (E), NJ (E), OH (E), PA (T), RI (H)
<i>C. lurida</i> Wahlenb.	Wherry 1963	–
<i>C. magellanica</i> Lam. subsp. <i>irrigua</i> (Wahlenb.) Hultén	Brooks 1987; Dearden 1979	WA (S)
<i>C. nigra</i> All.	Carter 1979	MI (E), NY (E)
<i>C. nigromarginata</i> Schwein.	Wherry 1963	CT (SC), IL (E), NY (E)
<i>C. normalis</i> Mack.	Wherry 1963	–
<i>C. novae-angliae</i> Schwein.	Carter 1979	CT (SC), MI (T)
<i>C. paleacea</i> Wahlenb.	Carter 1979	–
<i>C. pellita</i> Willd.	Wherry 1963	KY (E), TN (PX, E)
<i>C. pennsylvanica</i> Lam.	Wherry 1963	–
<i>C. pseudocyperus</i> L.	Carter 1979	CT (E), IN (E), NJ (E), OH (E), PA (E)
<i>C. retroflexa</i> Willd.	Wherry 1963	NH (T), NY (E)
<i>C. richardsonii</i> R. Br.	Sirois et al. 1988; Tyndall 1994; Tyndall and Hull 1999	IN (E), OH (PRX), PA (E), VT (E); QC (S1)
<i>C. rosea</i> Willd.	Wherry 1963	–
<i>C. scirpoidea</i> Michx.	Brooks 1987; Dearden 1979; Zika and Dann 1985	ME (T), MI (T), NH (T), NY (E), WA (S)
<i>C. scoparia</i> Schkuhr	Wherry 1963	–
<i>C. stipata</i> Willd.	Carter 1979; Wherry 1963	–
<i>C. straminea</i> Willd.	Wherry 1963	IN (T), KY (T), MI (E), NY (E)
<i>C. stricta</i> Lam.	Wherry 1963	–
<i>C. umbellata</i> Willd.	Carter 1979; Ryan 1988; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Wherry 1963	NH (E)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>C. vestita</i> Willd.	Wherry 1963	MD (E), ME (E), TN (PX, E)
<i>C. vulpinoidea</i> Michx.	Wherry 1963	—
<i>C. willdenowii</i> Schkuhr	Wherry 1963	CT (SC), IL (T), NY (T)
<i>Cyperus bipartitus</i> Torr.	Wherry 1963	WA (S)
<i>C. lupulinus</i> (Spreng.) Marcks subsp. <i>macilentus</i> (Fernald) Marcks	Wherry 1963	QC (S2)
<i>C. squarrosus</i> L.	Wherry 1963	ME (PX), NH (T), RI (E)
<i>C. strigosus</i> L.	Wherry 1963	—
<i>Dulichium arundinaceum</i> Britton	Wherry 1963	—
<i>Eleocharis erythropoda</i> Steud.	Wherry 1963	—
<i>E. melanocarpa</i> Torr.	Tyndall and Farr 1990	IN (T), MD (E), NJ (E), RI (E)
<i>E. nitida</i> Fernald	Hay et al. 1994; Tyndall and Hull 1999	MI (E), MN (T), WI (E); NL (S1)
<i>E. palustris</i> (L.) Roem. & Schult.	Wherry 1963	—
<i>E. tenuis</i> (Willd.) Schult.	Brooks 1987; Wherry 1963	NJ (E), NY (E), PA (E)
<i>Fimbristylis annua</i> (All.) Roem. & Schult.	Tyndall 1992b, 1994, 2005; Wherry 1963	PA (T)
<i>F. autumnalis</i> (L.) Roem. & Schult.	Brooks 1987	ME (T), VT (E); QC (S2)
<i>Rhynchospora capitellata</i> (Michx.) Vahl	Wherry 1963	QC (S2)
<i>Schoenoplectus maritimus</i> (L.) Lye	Carter 1979	CT (SC), IL (E), NJ (E), NY (E)
<i>S. tabernaemontani</i> (C.C. Gmel.) Palla	Wherry 1963	—
<i>Scirpus atrovirens</i> Muhl.	Wherry 1963	—
<i>S. caespitosus</i> (R. Br.) Poir.	Dearden 1979; Zika and Dann 1985	—
<i>S. cyperinus</i> (L.) Kunth.	Carter 1979; Wherry 1963	—
<i>S. hudsonianus</i> Fernald	Brooks 1987; Dearden 1979	—
<i>S. longii</i> Fernald	Hay et al. 1992	G2; CT (SC), MA (T), ME (T), NJ (E), RI (E)
<i>Scleria pauciflora</i> Willd.	Brooks 1987; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999	MA (E), MI (E), OH (T), PA (T)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>S. pauciflora</i> var. <i>caroliniana</i> (Willd.) Alph. Wood	Wherry 1963	CT (E), NY (E)
<i>S. triglomerata</i> Michx.	Wherry 1963	—
<i>Trichophorum caespitosum</i> (L.) Hartman [as <i>Scirpus</i> <i>caespitosus</i> (R. Br.) Poir. var. <i>callosus</i> Bigelow]	Brooks 1987	—
DENNSTAEDTIACEAE		
<i>Dennstaedtia punctilobula</i> (Michx.) T. Moore	Wherry 1963	IL (E), MI (PREX)
<i>Pteridium aquilinum</i> (L.) Kuhn	Zika and Dann 1985	—
<i>P. aquilinum</i> var. <i>latiusculum</i> (Desv.) A. Heller	Carter 1979; Wherry 1963	—
<i>P. aquilinum</i> var. <i>pubescens</i> Underw.	Brooks 1987	—
DIAPENSIACEAE		
<i>Diapensia lapponica</i> L.	Brooks 1987; Dearden 1979	ME (SC), NH (T), NY (T), VT (E)
DRYOPTERIDACEAE		
<i>Cystopteris fragilis</i> (L.) Bernh.	Carter 1979	NY (EV), OH (PRX)
<i>Dryopteris campyloptera</i> Clarkson	Carter 1979	CT (E), MD (E), NY (EV), PA (E)
<i>D. carthusiana</i> (Vill.) H.P. Fuchs	Carter 1979	AR (T), KY (SC), NY (EV), TN (T)
<i>D. fragrans</i> (L.) Schott	Zika and Dann 1985	ME (SC), NH (T), NY (E)
<i>D. intermedia</i> (Willd.) A. Gray	Carter 1979	IA (T), NY (EV)
<i>D. marginalis</i> (L.) A. Gray	Carter 1979; Wherry 1963	IA (T), MN (T), NY (EV)
<i>D. × triploidea</i> Wherry	Carter 1979	—
<i>Gymnocarpium dryopteris</i> (L.) Newman	Carter 1979	IL (E), IA (T), MD (E), NY (EV), OH (T)
<i>Matteuccia struthiopteris</i> (L.) Todaro	Carter 1979	IN (R), NY (EV)
<i>Onoclea sensibilis</i> L.	Carter 1979	—
<i>Polystichum acrostichoides</i> (Michx.) Schott	Tyndall and Farr 1990; Wherry 1963	MN (T), NY (EV)
<i>P. braunii</i> (Spenn.) Fée	Carter 1979; Rugg 1922	MA (E), MN (E), NY (EV), PA (E), WI (T)
<i>P. scopulinum</i> (D.C. Eaton) Maxon	Brooks 1987; Cody 1983	—

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Woodsia ilvensis</i> (L.) R. Br.	Carter 1979	IA (E), IL (E), MD (T), NY (EV), OH (PRX), RI (H)
EBENACEAE		
<i>Diospyros texana</i> Scheele	Maoui 1966	—
<i>D. virginiana</i> L.	Wherry 1963	CT (SC), NY (T)
EMPETRACEAE		
<i>Empetrum nigrum</i> L.	Dearden 1979; Zika and Dann 1985	MI (T), MN (E)
EQUISETACEAE		
<i>Equisetum arvense</i> L.	Carter 1979	—
<i>E. sylvaticum</i> L.	Carter 1979; Wherry 1963	IA (T), IL (E), MD (E)
ERICACEAE		
<i>Andromeda polifolia</i> L. var. <i>glaucophylla</i> (Link) DC.	Brooks 1987; Dearden 1979; Roberts 1980	CT (T), IN (R), NJ (E), OH (PRX), RI (E)
<i>Epigaea repens</i> L.	Brooks 1987; Dearden 1979; Wherry 1963; Zika and Dann 1985	FL (E), NY (EV)
<i>Gaultheria hispidula</i> Muhl.	Brooks 1987; Dearden 1979	CT (T), MD (E), NJ (E), OH (PRX), PA (R), WA (S)
<i>G. procumbens</i> L.	Carter 1979; Mansberg and Wentworth 1984; Zika and Dann 1985	IL (E)
<i>Gaylussacia baccata</i> (Wangenh.) K. Koch	Miller 1981; Tyndall and Hull 1999; Wherry 1963	IA (T)
<i>G. dumosa</i> Torr. & A. Gray	Brooks 1987; Dearden 1979	CT (T), NH (T), NY (E), PA (E), TN (T)
<i>Harrimanella hypnoides</i> (L.) Coville	Hay et al. 1994; Sirois et al. 1988	ME (T); NL (S2)
<i>Kalmia angustifolia</i> L.	Carter 1979; Zika and Dann 1985	NY (EV)
<i>K. latifolia</i> L.	Mansberg and Wentworth 1984; Tyndall and Hull 1999; Wherry 1963; Zika and Dann 1985	FL (T), ME (SC), NY (EV)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>K. polifolia</i> Wangenh.	Brooks 1987; Dearden 1979	NJ (E), NY (EV)
<i>Ledum groenlandicum</i> Oeder	Brooks 1987; Dearden 1979; Roberts 1980; Zika and Dann 1985	CT (T), OH (E), PA (R)
<i>Lyonia ligustrina</i> (L.) DC.	Wherry 1963	OH (PRX)
<i>L. mariana</i> (L.) D. Don	Wherry 1963	CT (SC), PA (E), RI (H)
<i>Oxydendrum arboreum</i> (L.) DC.	Brooks 1987	IN (T), MD (E)
<i>Phyllodoce caerulea</i> (L.) Bab.	Sirois et al. 1988	ME (T), NH (T)
<i>Rhododendron calendulaceum</i> (Michx.) Torr.	Mansberg and Wentworth 1984	OH (E), PA (X)
<i>R. canadense</i> (L.) Torr.	Brooks 1987; Carter 1979; Dearden 1979	NJ (E), NY (T)
<i>R. lapponicum</i> (L.) Wahlenb.	Brooks 1987; Dearden 1979; Sirois et al. 1988	ME (T), NY (E), WI (E)
<i>R. maximum</i> L.	Mansberg and Wentworth 1984	MA (T), ME (T), NY (EV), OH (T), VT (T)
<i>R. periclymenoides</i> (Michx.) Shinners	Miller 1981; Wherry 1963	NH (E), NY (EV), OH (T)
<i>R. viscosum</i> (L.) Torr.	Brooks 1987; Mansberg and Wentworth 1984; Wherry 1963	ME (E), NH (T), NY (EV)
<i>Vaccinium angustifolium</i> Aiton	Brooks 1987; Carter 1979; Dearden 1979; Wherry 1963	IA (T)
<i>V. caespitosum</i> Michx.	Carter 1979	MI (T), NY (E), WI (E)
<i>V. corymbosum</i> L.	Carter 1979; Wherry 1963	IL (E)
<i>V. fuscatum</i> Aiton	Wherry 1963	-
<i>V. macrocarpon</i> Aiton	Brooks 1987; Dearden 1979	IL (E), TN (T)
<i>V. myrtilloides</i> Michx.	Carter 1979; Zika and Dann 1985	CT (SC), IA (T), IN (E), OH (T), WA (S)
<i>V. oxycoccus</i> L.	Brooks 1987; Dearden 1979	IL (E), IN (T), MD (T), OH (T)
<i>V. pallidum</i> Aiton	Mansberg and Wentworth 1984; Miller 1981; Tyndall and Hull 1999; Wherry 1963	-

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>V. stamineum</i> L.	Mansberg and Wentworth 1984; Miller 1981; Tyndall 2005; Wherry 1963	VT (E)
<i>V. uliginosum</i> L.	Brooks 1987; Dearden 1979; Zika and Dann 1985	MI (T), MN (T), NY (R)
<i>V. vitis-idaea</i> L.	Brooks 1987; Dearden 1979; Zika and Dann 1985	MI (E)
EUPHORBIACEAE		
<i>Chamaesyce maculata</i> (L.) Small	Wherry 1963	—
<i>C. nutans</i> (Lag.) Small	Wherry 1963	—
<i>Euphorbia corollata</i> L.	Wherry 1963	—
FABACEAE		
<i>Acacia greggii</i> A. Gray	Maoui 1966	
<i>Amphicarpa bracteata</i> (L.) Fernald	Wherry 1963	NH (T)
<i>Baptisia tinctoria</i> (L.) R. Br.	Wherry 1963	KY (T), ME (E)
<i>Chamaecrista fasciculata</i> (Michx.) Greene var. <i>fasciculata</i>	Brooks 1987; Wherry 1963	—
<i>C. nictitans</i> Moench var. <i>nictitans</i>	Wherry 1963	NH (E)
<i>Crotalaria sagittalis</i> L.	Wherry 1963	NH (E), NY (E), VT (T)
<i>Desmodium ciliare</i> (Willd.) DC.	Wherry 1963	NY (T)
<i>D. marilandicum</i> (L.) DC.	Brooks 1987; Wherry 1963	NH (E)
<i>D. obtusum</i> (Willd.) DC.	Tyndall and Hull 1999; Wherry 1963	NY (E)
<i>D. paniculatum</i> (L.) DC.	Wherry 1963	QC (S1)
<i>D. perplexum</i> B.G. Schub.	Wherry 1963	—
<i>Lathyrus japonicus</i> Willd. var. <i>maritimus</i> (L.) Kartesz & Gandhi	Carter 1979	IL (E), IN (E), VT (T)
<i>L. palustris</i> L.	Carter 1979	KY (T), MD (E, X), PA (E), TN (SC), VT (T)
<i>Lespedeza capitata</i> Michx.	Wherry 1963	KY (SC)
<i>L. hirta</i> (L.) Hornem.	Wherry 1963	ME (PX), VT (T)
<i>L. procumbens</i> Michx.	Wherry 1963	MI (PREX), NH (E)
<i>L. repens</i> (L.) W. Bartram	Wherry 1963	CT (SC), NY (R)
<i>L. violacea</i> (L.) Pers.	Wherry 1963	NY (R), VT (T)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>L. virginica</i> (L.) Britton	Brooks 1987; Wherry 1963	NH (T), WI (T)
<i>L. × manniana</i> Mack. & Bush	Wherry 1963	–
<i>Strophostyles umbellata</i> (Willd.) Britton	Wherry 1963	NY (E)
<i>Stylosanthes biflora</i> Britton, Sterns & Poggenb.	Wherry 1963	PA (E)
<i>Tephrosia virginiana</i> (L.) Pers.	Wherry 1963	NH (E)
<i>Trifolium arvense</i> L.	Carter 1979	–
<i>T. campestre</i> Schreb.	Carter 1979	–
<i>T. pratense</i> L.	Carter 1979	–
<i>T. repens</i> L.	Carter 1979	–
FAGACEAE		
<i>Castanea dentata</i> (Marshall) Borkh.	Mansberg and Wentworth 1984; Wherry 1963	KY (E), ME (SC), MI (E), TN (SC)
<i>Quercus alba</i> L.	Brooks 1987; Mansberg and Wentworth 1984; Milton and Purdy 1988; Tyndall and Hull 1999; Wherry 1963	QC (S3)
<i>Q. coccinea</i> Wangenh.	Brooks 1987; Mansberg and Wentworth 1984	ME (E)
<i>Q. ilicifolia</i> Wangenh.	Wherry 1963	VT (E)
<i>Q. marilandica</i> Münchh.	Brooks 1987; Hull and Wood 1984; Miller 1981; Tyndall 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	–
<i>Q. palustris</i> Du Roi	Wherry 1963	
<i>Q. prinoides</i> Willd.	Wherry 1963	IN (E)
<i>Q. prinus</i> L.	Brooks 1987; Wherry 1963	IL (T), ME (T)
<i>Q. rubra</i> L.	Brooks 1987; Wherry 1963	–
<i>Q. stellata</i> Wangenh.	Brooks 1987; Hull and Wood 1984; Miller 1981; Tyndall 2005; Tyndall and Hull 1999; Wherry 1963	–

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Q. velutina</i> Lam.	Miller 1981; Tyndall and Hull 1999; Wherry 1963	—
FUMARIACEAE		
<i>Corydalis sempervirens</i> Pers.	Carter 1979	IA (T), IL (E), IN (E), KY (SC), TN (E)
<i>Dicentra canadensis</i> Walp.	Carter 1979	CT (T), ME (T), NH (T), NJ (E)
GENTIANACEAE		
<i>Gentiana andrewsii</i> Griseb.	Tyndall and Hull 1999	MA (T), MD (T), NH (T), NY (EV), RI (H), VT (T)
<i>G. villosa</i> L.	Wherry 1963	IN (E), MD (E), OH (E), PA (E)
<i>Gentianopsis crinita</i> (Froel.) Ma	Tyndall and Hull 1999; Wherry 1963	GA (T), MD (E), NC (SC, E), NH (T), NY (EV); QC (S1)
<i>Sabatia angularis</i> (L.) Pursh	Brooks 1987; Tyndall 1994; Wherry 1963	MI (T), NY (E)
GERANIACEAE		
<i>Geranium maculatum</i> L.	Wherry 1963	QC (SX)
GROSSULARIACEAE		
<i>Ribes cynosbati</i> L.	Carter 1979	—
<i>R. glandulosum</i> Ruiz & Pav.	Carter 1979	CT (E), NJ (E), OH (PRX)
<i>R. lacustre</i> (Pers.) Poir.	Carter 1979	CT (SC), MA (SC), PA (E)
<i>R. triste</i> Pall.	Carter 1979	CT (E), OH (E), PA (T)
HAMAMELIDACEAE		
<i>Hamamelis virginiana</i> L.	Mansberg and Wentworth 1984	—
IRIDACEAE		
<i>Hypoxis hirsuta</i> Coville	Wherry 1963	ME (PX), NH (T)
<i>Iris versicolor</i> L.	Carter 1979	—
<i>Sisyrinchium angustifolium</i> Mill.	Wherry 1963	—
<i>S. montanum</i> Greene var. <i>crebrum</i> Fernald	Carter 1979	IL (E), IN (E), NJ (E), OH (E), WA (S)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>S. mucronatum</i> Michx.	Brooks 1987; Mansberg and Wentworth 1984; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	MA (E), ME (SC), NY (E), OH (E)
JUGLANDACEAE		
<i>Carya glabra</i> (Mill.) Sweet	Brooks 1987	—
JUNCACEAE		
<i>Juncus acuminatus</i> Michx.	Wherry 1963	QC (S1)
<i>J. balticus</i> Willd.	Brooks 1987; Carter 1979	IN (R), MD (E, X), PA (T)
<i>J. biflorus</i> Elliott	Wherry 1963	NY (E), PA (T)
<i>J. brevicaudatus</i> (Engelm.) Fernald	Carter 1979	—
<i>J. dichotomus</i> Elliott	Brooks 1987; Wherry 1963	OH (E), PA (E)
<i>J. dudleyi</i> Wiegand	Brooks 1987	ME (SC)
<i>J. effusus</i> L.	Wherry 1963	—
<i>J. effusus</i> var. <i>conglomeratus</i> (L.) Engelm.	Carter 1979	—
<i>J. marginatus</i> Rostk.	Wherry 1963	—
<i>J. secundus</i> Poir.	Brooks 1987; Tyndall 1994; Tyndall and Farr 1990; Wherry 1963	ME (SC), IN (E), NH (E), OH (T), VT (E)
<i>J. tenuis</i> Willd.	Wherry 1963	—
<i>J. trifidus</i> L.	Brooks 1987; Dearden 1979	MD (E), NC (E), NY (T), TN (PX, E)
<i>Luzula acuminata</i> Raf.	Mansberg and Wentworth 1984	IL (E), IN (E), NJ (E)
<i>L. bulbosa</i> (Alph. Wood) Smyth & Smyth	Wherry 1963	OH (T), PA (E)
<i>L. campestris</i> (L.) DC.	Wherry 1963	—
<i>L. multiflora</i> (Ehrh.) Lej. subsp. <i>frigida</i> (Buchenau) Krecz.	Carter 1979	—
JUNCAGINACEAE		
<i>Triglochin palustre</i> L.	Carter 1979	IA (T), IL (T), IN (T), NY (T), PA (X), RI (H)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
LAMIACEAE		
<i>Cunila organoides</i> (L.) Britton	Brooks 1987; Wherry 1963	—
<i>Galeopsis tetrahit</i> L.	Carter 1979	—
<i>Lycopus virginicus</i> Michx.	Carter 1979	MI (T); QC (S2)
<i>Mentha arvensis</i> L.	Carter 1979	
<i>Monarda punctata</i> L.	Tyndall and Farr 1990	KY (H), OH (E), PA (E)
<i>Prunella vulgaris</i> L.	Brooks 1987; Mansberg and Wentworth 1984	—
<i>P. vulgaris</i> L. subsp. <i>lanceolata</i> (W. Bartram) Hultén	Carter 1979; Wherry 1963	—
<i>Pycnanthemum flexuosum</i> Britton, Sterns & Poggenb.	Brooks 1987; Miller 1981; Wherry 1963	—
<i>P. tenuifolium</i> Schrad.	Brooks 1987; Wherry 1963	—
<i>P. torrei</i> Benth.	Tyndall and Hull 1999	G2; CT (E), IL (E), MD (E), NH (E), NJ (E), NY (E), PA (E), TN (SC)
<i>Scutellaria elliptica</i> Epling	Mansberg and Wentworth 1984; Wherry 1963	—
<i>S. galericulata</i> L.	Carter 1979	—
<i>S. integrifolia</i> L.	Wherry 1963	CT (E), NY (E)
<i>S. lateriflora</i> L.	Carter 1979	
<i>S. parvula</i> Michx. var. <i>missouriensis</i> (Torr.) Goodman & C.A. Lawson	Wherry 1963	CT (E), MD (T), NJ (E)
<i>Trichostema dichotomum</i> L.	Wherry 1963	IN (R), MI (T); QC (SH)
Lauraceae		
<i>Lindera benzoin</i> Blume	Wherry 1963	ME (SC)
<i>Sassafras albidum</i> (Nutt.) Nees	Brooks 1987; Mansberg and Wentworth 1984; Miller 1981; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	ME (SC)
Liliaceae		
<i>Alertris farinosa</i> L.	Wherry 1963	ME (PX), NY (T), PA (E)
<i>Chamaelirium luteum</i> A. Gray	Wherry 1963	CT (E), IN (E), MA (E), NY (T)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Clintonia borealis</i> (Aiton) Raf.	Brooks 1987	IN (E), MD (T), OH (E), TN (SC)
<i>Erythronium americanum</i> Ker Gawl.	Carter 1979	–
<i>Hemerocallis fulva</i> L.	Carter 1979	–
<i>Lilium philadelphicum</i> L.	Wherry 1963	KY (T), MD (E, X), NM (E), NY (EV), OH (T), TN (E)
<i>Maianthemum canadense</i> Desf.	Brooks 1987; Dearden 1979	KY (T)
<i>M. racemosum</i> (L.) Link subsp. <i>racemosum</i>	Mansberg and Wentworth 1984; Wherry 1963	AZ (SR)
<i>Polygonatum biflorum</i> (Walter) Elliot var. <i>commutatum</i> (Schult. & Schult. f.) Morong	Wherry 1963	NH (E)
<i>Streptopus lanceolatus</i> (Aiton) Reveal var. <i>lanceolatus</i>	Carter 1979	KY (E)
<i>Trillium erectum</i> L.	Carter 1979	IL (E), NY (EV)
<i>Uvularia perfoliata</i> L.	Wherry 1963	IN (E), NH (E)
<i>U. puberula</i> Michx.	Mansberg and Wentworth 1984	NJ (E), NY (E), PA (R)
<i>Veratrum viride</i> Aiton	Carter 1979	–
LINACEAE		
<i>Linum floridanum</i> Trel.	Wherry 1963	MD (E, X)
<i>L. intercursum</i> E.P. Bicknell	Wherry 1963	CT (SC), IN (E), MA (SC), MD (T), NJ (E), NY (T), PA (E), RI (E)
<i>L. medium</i> (Planch.) Britton	Brooks 1987	NY (E)
<i>L. medium</i> var. <i>texanum</i> (Planch.) Fernald	Wherry 1963	MA (T), NY (T)
<i>L. sulcatum</i> Riddell	Tyndall 1992b, Tyndall 1994, Tyndall 2005; Tyndall and Hull 1999	CT (SC), IN (R), MD (E), NJ (E), NY (T), PA (E), RI (H)
<i>L. virginianum</i> L.	Wherry 1963	MI (T)
LYCOPODIACEAE		
<i>Huperzia lucidula</i> (Michx.) Trevis.	Carter 1979	NY (EV)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>H. selago</i> (L.) Schrank & Mart. var. <i>selago</i>	Tyndall and Hull 1999; Zika and Dann 1985	CT (SC), MA (E), ME (T), NY (E), PA (X),
<i>Lycopodium alpinum</i> L.	Hay et al. 1992	–
<i>L. clavatum</i> L.	Carter 1979	IA (E), IL (E), KY (E), NY (EV)
<i>L. digitatum</i> A. Braun	Wherry 1963	NY (EV)
<i>L. obscurum</i> L.	Carter 1979	IN (R), NY (EV)
LYTHRACEAE		
<i>Cuphea viscosissima</i> Jacq.	Wherry 1963	–
MAGNOLIACEAE		
<i>Liriodendron tulipifera</i> L.	Miller 1981	–
MONOTROPACEAE		
<i>Monotropa uniflora</i> L.	Carter 1979	–
MYRICACEAE		
<i>Comptonia peregrina</i> (L.) J.M. Coult.	Wherry 1963	IL (E), KY (E), OH (T), TN (E)
<i>Morella pensylvanica</i> (Mirb.) Kartesz	Carter 1979	NY (EV), OH (E)
<i>Myrica gale</i> L.	Brooks 1987; Carter 1979	NC (E), PA (T)
NAJADACEAE		
<i>Najas gracillima</i> Morong	Zika and Dann 1985	IN (E), KY (SC), MD (E, X), ME (SC), OH (E), PA (T)
NYSSACEAE		
<i>Nyssa sylvatica</i> Marshall	Brooks 1987; Mansberg and Wentworth 1984; Miller 1981; Wherry 1963	–
ONAGRACEAE		
<i>Circaea alpina</i> L.	Brooks 1987; Dearden 1979	IL (E), IN (X), KY (SC)
<i>Epilobium ciliatum</i> Raf. subsp. <i>glandulosum</i> (Lehm.) Hoch & P.H. Raven	Carter 1979	NY (E)
<i>Oenothera biennis</i> L.	Carter 1979	–
<i>O. fruticosa</i> L.	Miller 1981; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	CT (SC)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>O. fruticosa</i> L. subsp. <i>fruticosa</i>	Brooks 1987	—
<i>O. fruticosa</i> subsp. <i>glauca</i> (Michx.) Straley	Mansberg and Wentworth 1984	—
OPHIGLOSSACEAE		
<i>Botrychium dissectum</i> Spreng.	Wherry 1963	NY (EV)
<i>B. virginianum</i> (L.) Sw.	Carter 1979	NY (EV)
ORCHIDACEAE		
<i>Goodyera pubescens</i> (Willd.) R. Br.	Mansberg and Wentworth 1984; Wherry 1963	FL (E), NY (EV); QC (S2)
<i>G. tessellata</i> Lodd.	Carter 1979	MD (E, X), NJ (E), NY (EV), OH (PRX), PA (T)
<i>Liparis liliifolia</i> (L.) Lindl.	Wherry 1963	CT (E), MA (T), NY (E), RI (E), VT (T)
<i>L. loeselii</i> (L.) Rich.	Wherry 1963	AR (T), KY (T), NH (T), NY (EV), TN (E), WA (E)
<i>Malaxis unifolia</i> Michx.	Wherry 1963	CT (E), FL (E), IN (E), NH (T), NY (EV), RI (E)
<i>Platanthera dilatata</i> (Pursh) Beck var. <i>dilatata</i>	Carter 1979	CT (SC), IN (E), MA (T), NY (EV), PA (E)
<i>P. flava</i> (L.) Lindl. var. <i>flava</i>	Wherry 1963	IL (E), IN (E), NJ (E), TN (SC)
<i>Spiranthes lacera</i> (Raf.) Raf. var. <i>gracilis</i> (Bigelow) Luer	Wherry 1963	ME (PX)
<i>S. tuberosa</i> Raf.	Wherry 1963	CT (SC), FL (T), NY (EV), PA (X), RI (E)
OSMUNDACEAE		
<i>Osmunda cinnamomea</i> L.	Carter 1979	FL (CE), IA (E), NY (EV)
<i>O. claytoniana</i> L.	Carter 1979	AR (T), NY (EV)
<i>O. regalis</i> L.	Wherry 1963	FL (CE), IA (T), NY (EV)
OXALIDACEAE		
<i>Oxalis montana</i> Raf.	Carter 1979	OH (PRX)
<i>O. stricta</i> L.	Brooks 1987; Wherry 1963	—

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
PINACEAE		
<i>Abies balsamea</i> (L.) Mill.	Brooks 1987; Carter 1979; Tyndall and Hull 1999	CT (E)
<i>Larix laricina</i> (Du Roi) K. Koch	Brooks 1987; Dearden 1979; Roberts 1980	IL (T), MD (E); NL (S5)
<i>Picea glauca</i> (Moench) Voss	Brooks 1987; Carter 1979	–
<i>P. mariana</i> Britton, Sterns & Poggenb.	Brooks 1987; Carter 1979; Roberts 1980	NL (S5)
<i>P. rubens</i> Sarg.	Zika and Dann 1985	CT (SC), NJ (E)
<i>Pinus echinata</i> Mill.	Tyndall and Hull 1999	
<i>P. resinosa</i> Mill.	Brooks 1987; Carter 1979	CT (E), IL (E), NJ (E)
<i>P. rigida</i> Mill.	Brooks 1987; Mansberg and Wentworth 1984; Miller 1981; Tyndall and Hull 1999; Wherry 1963	QC (S1)
<i>P. strobus</i> L.	Brooks 1987; Carter 1979; Zika and Dann 1985	IN (R)
<i>P. virginiana</i> Mill.	Brooks 1987; Miller 1981; Tyndall 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	NY (E)
<i>Tsuga canadensis</i> Carrière	Brooks 1987; Mansberg and Wentworth 1984	–
PLANTAGINACEAE		
<i>Plantago maritima</i> L. var. <i>juncoides</i> (Lam.) A. Gray	Carter 1979	NY (T)
PLUMBAGINACEAE		
<i>Armeria maritima</i> (Mill.) Willd. subsp. <i>sibirica</i> (Boiss.) Nyman	Brooks 1987; Dearden 1979; Roberts 1980	–
<i>Limonium carolinianum</i> (Walter) Britton	Carter 1979	NY (EV)
POACEAE		
<i>Agrostis capillaris</i> L.	Carter 1979	–
<i>A. hyemalis</i> (Walter) Britton, Sterns & Poggenb.	Tyndall 2005; Wherry 1963	–
<i>A. mertensii</i> Trin.	Zika and Dann 1985	–
<i>A. perennans</i> (Walter) Tuck.	Brooks 1987; Carter 1979; Wherry 1963	–
<i>A. stolonifera</i> L.	Tyndall and Farr 1990	–

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Andropogon gerardii</i> Vitman	Mansberg and Wentworth 1984; Tyndall 1992b, 1994, 2005; Tyndall and Hull 1999; Wheeler 1988; Wherry 1963	–
<i>Anthoxanthum odoratum</i> L.	Miller 1981	–
<i>Aristida dichotoma</i> Michx.	Brooks 1987; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	MI (PREX)
<i>A. longispica</i> Poir.	Tyndall and Hull 1999; Wherry 1963	CT (SC), MI (T)
<i>A. oligantha</i> Michx.	Wherry 1963	–
<i>A. purpurascens</i> Poir.	Brooks 1987; Maoui 1966; Tyndall 1992b, 1994, 2005; Tyndall and Hull 1999; Wherry 1963	CT (SC), MA (T), PA (T)
<i>Bouteloua curtipendula</i> (Michx.) Torr.	Brooks 1987; Nixon and McMillan 1964; Wherry 1963	CT (E), KY (SC), MI (T), NJ (E), NY (E), PA (T)
<i>B. hirsuta</i> Lag.	Nixon and McMillan 1964	AR (E)
<i>B. rigidiseta</i> Hitchc.	Maoui 1966	AR (E)
<i>Bromus ciliatus</i> L. var. <i>ciliatus</i>	Carter 1979	MD (E, X)
<i>Calamagrostis canadensis</i> (Michx.) P. Beauv.	Carter 1979	–
<i>Cinna latifolia</i> Griseb.	Carter 1979	MD (T), NJ (E), OH (E)
<i>Danthonia intermedia</i> Vasey	Brooks 1987; Hay et al. 1994; Tyndall and Hull 1999	NL (S1,S2)
<i>D. spicata</i> (L.) Roem. & Schult.	Carter 1979; Mansberg and Wentworth 1984; Tyndall 2005; Tyndall and Farr 1990; Wherry 1963	–
<i>Deschampsia alpina</i> (L.) Roem. & Schult.	Brooks 1987	–
<i>D. caespitosa</i> (L.) P. Beauv.	Brooks 1987; Mansberg and Wentworth 1984; Roberts 1980; Wherry 1963; Zika and Dann 1985	CT (SC), IN (R), MA (E), MD (E), KY (E)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>D. flexuosa</i> (L.) Trin.	Carter 1979; Roberts 1980; Zika and Dann 1985	KY (T)
<i>Dichantherium acuminatum</i> (Sw.) Gould & C.A. Clark var. <i>fasciculatum</i> (Torr.) Freckmann	Carter 1979; Wherry 1963	IN (X), TN (E)
<i>D. acuminatum</i> var. <i>lindheimeri</i> (Nash) Gould & C.A. Clark	Wherry 1963	OH (E)
<i>D. boscii</i> (Poir.) Gould & C.A. Clark	Mansberg and Wentworth 1984; Wherry 1963	–
<i>D. clandestinum</i> (L.) Gould	Brooks 1987; Wherry 1963	–
<i>D. depauperatum</i> (Muhl.) Gould	Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	–
<i>D. dichotomum</i> (L.) Gould var. <i>dichotomum</i>	Brooks 1987; Mansberg and Wentworth 1984; Wherry 1963	FL (T), IL (E), IN (E), MA (E), OH (E), PA (E)
<i>D. linearifolium</i> (Scribn.) Gould	Brooks 1987; Wherry 1963	–
<i>D. meridionale</i> (Ashe) Freckmann	Wherry 1963	OH (T)
<i>D. oligoanthes</i> (Schult.) Gould var. <i>oligoanthes</i>	Tyndall and Hull 1999	NY (E)
<i>D. oligoanthes</i> var. <i>scribnerianum</i> (Nash) Gould	Wherry 1963	NY (E), PA (E)
<i>D. ovale</i> (Elliot) Gould & C.A. Clark var. <i>addisonii</i> (Nash) Gould & C.A. Clark	Tyndall 2005; Wherry 1963	IN (R), OH (E), PA (X)
<i>D. sphaerocarpon</i> (Elliot) Gould var. <i>sphaerocarpon</i>	Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	CT (SC), NH (E)
<i>D. sphaerocarpon</i> var. <i>isophyllum</i> (Scribn.) Gould & C.A. Clark	Wherry 1963	MI (E)
<i>D. villosissimum</i> (Nash) Freckmann var. <i>villosissimum</i>	Wherry 1963	MA (SC), OH (PRX)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Digitaria cognata</i> (Schult.) Pilg.	Maoui 1966	PA (T)
<i>D. filiformis</i> (L.) Koeler	Wherry 1963	MI (PREX), NY (T), OH (PRX)
<i>Elymus repens</i> (L.) Gould	Carter 1979	—
<i>E. trachycaulus</i> (Link) Gould subsp. <i>trachycaulus</i>	Brooks 1987; Mansberg and Wentworth 1984	MD (E, X)
<i>E. virginicus</i> L.	Carter 1979	—
<i>Eragrostis intermedia</i> Hitchc.	Maoui 1966	—
<i>E. pectinacea</i> (Michx.) Nees	Wherry 1963	—
<i>E. spectabilis</i> (Pursh) Steud.	Tyndall 2005; Tyndall and Farr 1990	—
<i>Festuca altaica</i> Trin.	Brooks 1987; Hay et al. 1994; Tyndall and Hull 1999	MI (T); NL (S2), QC (S2, S3)
<i>F. filiformis</i> Lam.	Carter 1979	—
<i>F. rubra</i> L.	Brooks 1987; Dearden 1979	ME (E), NH (E), 1979
<i>Glyceria melicaria</i> (Michx.) F.T. Hubb.	Carter 1979	—
<i>G. striata</i> (Lam.) Hitchc.	Carter 1979; Wherry 1963	—
<i>Hierochloe odorata</i> (L.) P. Beauv.	Carter 1979	MD (E), NC (E), PA (E)
<i>Hilaria belangeri</i> (Steud.) Nash	Nixon and McMillan 1964	—
<i>Leersia oryzoides</i> (L.) Sw.	Wherry 1963	—
<i>L. virginica</i> Willd.	Wherry 1963	NH (T), WI (T)
<i>Lolium pratense</i> (Huds.) Darbysh.	Carter 1979	—
<i>Muhlenbergia glomerata</i> Trin.	Brooks 1987; Mansberg and Wentworth 1984	WA (S)
<i>M. mexicana</i> Trin.	Wherry 1963	—
<i>M. sylvatica</i> Trin.	Wherry 1963	QC (S2)
<i>Nassella leucotricha</i> (Trin.) & Rupr.) R.W. Pohl	Maoui 1966	—
<i>Oryzopsis asperifolia</i> Michx.	Brooks 1987; Carter 1979	IN (E), MD (T), NJ (E), OH (E)
<i>Panicum anceps</i> Michx.	Wherry 1963	—
<i>P. capillare</i> L.	Wherry 1963	—
<i>P. dichotomiflorum</i> Michx.	Wherry 1963	—
<i>P. flexile</i> Scribn.	Tyndall and Hull 1999	MD (E), NJ (E), NY (T), VT (E); QC (S2)
<i>P. gattingeri</i> Nash	Wherry 1963	MA (SC)
<i>P. hallii</i> Vasey	Maoui 1966	—

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>P. philadelphicum</i> Trin.	Tyndall and Hull 1999; Wherry 1963	IA (T), MA (SC), NH (E), OH (T), PA (T); QC (S2)
<i>P. virgatum</i> L.	Nixon and McMillan 1964; Wherry 1963	QC (S1)
<i>Paspalum laeve</i> Michx.	Wherry 1963	CT (E), NY (E)
<i>P. setaceum</i> Michx.	Wherry 1963	CT (SC), NY (T)
<i>Phalaris arundinacea</i> L.	Carter 1979	–
<i>Piptochaetium avenaceum</i> (L.) Parodi	Tyndall and Hull 1999	IN (T), OH (PRX), PA (X)
<i>Poa compressa</i> L.	Tyndall 2005	–
<i>P. palustris</i> L.	Carter 1979	TN (E)
<i>P. pratensis</i> L.	Brooks 1987; Carter 1979; Wherry 1963	–
<i>P. saltuensis</i> Fernald & Weigand	Mansberg and Wentworth 1984	MA (E), MD (E), IL (E), KY (E), NJ (E), OH (E), PA (T), TN (SC)
<i>Puccinellia distans</i> (Jacq.) Parl.	Brooks 1987	–
<i>P. maritima</i> Parl.	Carter 1979	–
<i>P. tenella</i> A.E. Porsild subsp. <i>alaskana</i> (Scribn. & Merr.) Tzvelev	Carter 1979	CT (SC), NH (E)
<i>Schizachne purpurascens</i> Swallen	Carter 1979	CT (SC), IL (E), IN (E), KY (T), MD (E), NJ (E), OH (E)
<i>Schizachyrium scoparium</i> (Michx.) Nash	Flanagan-Brown 2001; Nixon and McMillan 1964; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999	OH (E), PA (R)
<i>S. scoparium</i> (Michx.) Nash var. <i>scoparium</i>	Brooks 1987; Mansberg and Wentworth 1984; Miller 1981; Wherry 1963	–
<i>Setaria faberi</i> Herrm.	Wherry 1963	–
<i>S. parviflora</i> (Poir.) Kerguelén	Wherry 1963	MA (SC), IN (E)
<i>S. punila</i> (Poir.) Roem. & Schult. subsp. <i>pallidifusca</i> (Schumach.) B.K. Simon	Carter 1979	–

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Sorghastrum nutans</i> (L.) Nash	Brooks 1987; Flanagan-Brown 2001; Nixon and McMillan 1964; Tyndall 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	ME (E); QC (S3)
<i>Spartina pectinata</i> Link	Carter 1979	WA (S)
<i>Sphenopholis obtusata</i> (Michx.) Scribn.	Brooks 1987; Tyndall and Farr 1990; Wherry 1963	ME (PX), NH (E), NY (E), OH (T), VT (E)
<i>Sporobolus cryptandrus</i> A. Gray	Maoui 1966	CT (E), NH (T), PA (R); QC (S2)
<i>S. heterolepis</i> A. Gray	Brooks 1987; Mansberg and Wentworth 1984; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	CT (E), KY (E), MD (E), NC (E), NY (T), OH (T), PA (E); QC (S2)
<i>S. vaginiflorus</i> (A. Gray) Alph. Wood	Wherry 1963	-
<i>Tridens flavus</i> Hitchc. var. <i>flavus</i>	Wherry 1963	-
<i>Vulpia octoflora</i> (Walter) Rydb. var. <i>glauca</i> (Nutt.) Fernald	Wherry 1963	NH (E)
POLEMONIACEAE		
<i>Phlox carolina</i> L.	Mansberg and Wentworth 1984	MD (E, X)
<i>P. subulata</i> L. subsp. <i>subulata</i>	Brooks 1987; Tyndall 2005; Wherry 1963	TN (T)
POLYGALACEAE		
<i>Polygala ambigua</i> Nutt.	Wherry 1963	-
<i>P. paucifolia</i> Willd.	Brooks 1987; Mansberg and Wentworth 1984	IN (E), KY (E), OH (E)
<i>P. sanguinea</i> L.	Wherry 1963	-
<i>P. senega</i> L.	Wherry 1963	CT (E), MD (T), ME (E), NJ (E); QC (S2)
<i>P. verticillata</i> L.	Brooks 1987	-
POLYGONACEAE		
<i>Polygonum aviculare</i> L.	Hart 1980	-
<i>P. cilinode</i> Michx.	Carter 1979	IN (E), OH (E), TN (T)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>P. cuspidatum</i> Siebold & Zucc.	Carter 1979	—
<i>P. persicaria</i> L.	Carter 1979	MA (SC)
<i>P. sagittatum</i> L.	Carter 1979	—
<i>P. tenue</i> Michx.	Brooks 1987; Hart 1980; Tyndall 1992b, 1994; Tyndall and Farr 1990; Tyndall and Hull 1999	ME (PX), NH (E), NY (R)
<i>Rumex acetosella</i> L.	Carter 1979	—
<i>R. crispus</i> L.	Carter 1979	—
POLYPODIACEAE		
<i>Athyrium filix-femina</i> (L.) Roth	Carter 1979; Wherry 1963	FL (T), NY (EV)
<i>Polypodium virginianum</i> L.	Carter 1979; Rugg 1922; Zika and Dann 1985	NY (EV)
PORTULACACEAE		
<i>Claytonia caroliniana</i> Michx.	Carter 1979	—
<i>Talinum teretifolium</i> Pursh	Brooks 1987; Tyndall 1992b, 1994; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	—
PRIMULACEAE		
<i>Androsace septentrionalis</i> L.	Brooks 1987; Dearden 1979	—
<i>Glaux maritima</i> L.	Carter 1979	MD (E, X), MN (E), NJ (E), RI (H)
<i>Lysimachia quadrifolia</i> L.	Mansberg and Wentworth 1984; Wherry 1963	NY (E), TN (SC)
<i>L. terrestris</i> Britton, Sterns & Poggenb.	Carter 1979	KY (E), TN (E)
<i>Primula mistassinica</i> Michx.	Dearden 1979	IL (E), ME (SC), NY (T), VT (T)
<i>P. stricta</i> Hornem.	Hay et al. 1994	NL (S1)
<i>Trientalis borealis</i> Raf.	Brooks 1987; Carter 1979; Dearden 1979; Zika and Dann 1985	GA (E), IL (T), KY (E), TN (T)
PTERIDACEAE		
<i>Adiantum aleuticum</i> (Rupr.) C.A. Paris	Paris 1991; Paris and Windham 1988; Tyndall and Hull 1999	ME (E); QC (S2)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>A. aleuticum</i> [as <i>A. pedatum</i> L. subsp. <i>calderi</i> Cody]	Cody 1983; Paris 1991; Paris and Windham 1988	–
<i>A. aleuticum</i> [as <i>A. pedatum</i> var. <i>aleuticum</i> Rupr.]	Brooks 1987; Carter 1979; Cody 1983; Dearden 1979; Paris 1991; Paris and Windham 1988; Rugg 1922; Tyndall and Hull 1999; Zika and Dann 1985	–
<i>A. pedatum</i> L.	Carter 1979; Wherry 1963	NY (EV); QC (S4)
<i>A. viridimontanum</i> C.A. Paris	Paris 1991; Tyndall and Hull 1999	G2; VT (T); QC (S3)
<i>Aspidotis densa</i> (Brack.) Lellinger	Brooks 1987; Tyndall and Hull 1999	QC (S1)
PYROLACEAE		
<i>Chimaphila maculata</i> (L.) Pursh	Mansberg and Wentworth 1984; Wherry 1963	IL (E), ME (E), NY (EV)
<i>C. umbellata</i> (L.) W. Bartram subsp. <i>cisatlantica</i> (S.F. Blake) Hultén	Carter 1979; Wherry 1963	IN (T)
<i>Pyrola americana</i> Sweet	Wherry 1963	IN (R), KY (H), TN (E)
<i>P. elliptica</i> Nutt.	Wherry 1963	–
RANUNCULACEAE		
<i>Actaea rubra</i> (Aiton) Willd.	Carter 1979	IN (R), OH (T)
<i>Coptis trifolia</i> Salisb.	Carter 1979	MD (E), WA (S)
<i>Ranunculus abortivus</i> L.	Carter 1979	–
<i>R. lapponicus</i> L.	Sirois et al. 1988	ME (T), MI (T), WI (E)
<i>R. pedatifidus</i> Sm. var. <i>affinis</i> (R. Br.) L.D. Benson	Hay et al. 1992	NL (S2)
<i>Thalictrum alpinum</i> L.	Brooks 1987; Dearden 1979	–
<i>T. macrostylum</i> Small & A. Heller	Brooks 1987; Mansberg and Wentworth 1984; Tyndall and Hull 1999	–
<i>T. pubescens</i> Pursh	Carter 1979	IN (T)
<i>T. revolutum</i> DC.	Hay et al. 1994; Wherry 1963	IA (E), RI (H); QC (S1)
<i>T. thalictroides</i> (L.) Eames & B. Boivin	Wherry 1963	FL (E), ME (PX), NH (T)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
RHAMNACEAE		
<i>Ceanothus americanus</i> L.	Ryan 1988; Wherry 1963	ME (T); QC (S2)
<i>Fragula alnus</i> Mill.	Tyndall and Hull 1999	—
ROSACEAE		
<i>Amelanchier arborea</i> (F. Michx.) Fernald	Brooks 1987; Carter 1979; Miller 1981; Mansberg and Wentworth 1984; Wherry 1963	—
<i>A. bartramiana</i> (Tausch) M. Roem.	Carter 1979	MA (T), PA (E)
<i>A. laevis</i> Wiegand	Wherry 1963	
<i>Dalibarda repens</i> L.	Carter 1979	CT (E), MI (T), NC (E), NJ (E), OH (T), RI (E)
<i>Fragaria virginiana</i> Mill.	Carter 1979	
<i>Malus coronaria</i> (L.) Mill. var. <i>coronaria</i>	Wherry 1963	NY (E)
<i>M. sylvestris</i> Mill.	Carter 1979	—
<i>Photinia melanocarpa</i> (Michx.) K.R. Robertson & J.B. Phipps	Wherry 1963	IA (E)
<i>Physocarpus opulifolius</i> (L.) Maxim.	Mansberg and Wentworth 1984	FL (E)
<i>Potentilla canadensis</i> L.	Brooks 1987; Mansberg and Wentworth 1984; Tyndall 1992b, Tyndall 1994, Tyndall 2005; Tyndall and Hull 1999; Wherry 1963	—
<i>P. fruticosa</i> L.	Brooks 1987	—
<i>P. simplex</i> Michx.	Carter 1979	—
<i>Prunus americana</i> Marshall	Wherry 1963	NH (T), VT (T)
<i>P. pensylvanica</i> L. f.	Brooks 1987; Carter 1979	IN (R)
<i>P. serotina</i> Ehrh.	Miller 1981	—
<i>P. virginiana</i> L.	Carter 1979	TN (SC)
<i>Rosa carolina</i> L.	Carter 1979; Wherry 1973	
<i>R. multiflora</i> Murray	Miller 1981	—
<i>R. virginiana</i> Mill.	Brooks 1987; Carter 1979	—
<i>Rubus allegheniensis</i> Porter	Miller 1981	—
<i>R. argutus</i> Link	Wherry 1963	—

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>R. cuneifolius</i> Pursh	Tyndall and Hull 1999	CT (SC), NH (E), NY (E), PA (E)
<i>R. flagellaris</i> Willd.	Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	IN (E); QC (S2)
<i>R. frondosus</i> Bigelow	Wherry 1963	—
<i>R. hispidus</i> L.	Carter 1979	—
<i>R. idaeus</i> subsp. <i>strigosus</i> (Michx.) Focke	Carter 1979	—
<i>R. occidentalis</i> L.	Miller 1981; Wherry 1963	—
<i>R. pensilvanicus</i> Poir.	Wherry 1963	—
<i>R. pubescens</i> Raf.	Brooks 1987; Carter 1979; Dearden 1979	IL (T)
<i>Sanguisorba canadensis</i> L.	Brooks 1987; Wherry 1963	GA (T), IL (E), IN (E), KY (E), MD (T), ME (T), MI (T), RI (E), TN (E)
<i>Sibbaldia procumbens</i> L.	Hay et al. 1992	NH (E); NL (S1)
<i>Sibbaldiopsis tridentata</i> (Aiton) Rydb.	Roberts 1980	CT (E), GA (E), IA (E), NJ (E), PA (E), RI (H), TN (SC)
<i>Sorbus americana</i> Marshall	Carter 1979	IL (E)
<i>S. decora</i> C.K. Schneid.	Carter 1979	IN (X), MA (E), OH (E), PA (E)
<i>S. groenlandica</i> (C.K. Schneid.) Á. Löve & D. Löve	Carter 1979	—
<i>Spiraea alba</i> Du Roi var. <i>latifolia</i> (Aiton) Dippel	Wherry 1963; Zika and Dann 1985	OH (PRX)
RUBIACEAE		
<i>Galium aparine</i> L.	Miller 1981	—
<i>G. asprellum</i> Michx.	Carter 1979; Wherry 1963	TN (SC)
<i>G. boreale</i> L.	Tyndall and Hull 1999; Wherry 1963	—
<i>G. pilosum</i> Aiton	Wherry 1963	NH (E)
<i>G. tinctorium</i> L.	Wherry 1963	—
<i>G. triflorum</i> Michx.	Carter 1979; Wherry 1963	—

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Houstonia caerulea</i> L.	Brooks 1987; Tyndall 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	—
<i>H. serpyllifolia</i> Michx.	Mansberg and Wentworth 1984	KY (E), PA (X)
<i>Mitchella repens</i> L.	Wherry 1963	IA (T)
SALICACEAE		
<i>Populus grandidentata</i> Michx.	Miller 1981; Wherry 1963	TN (SC)
<i>P. tremuloides</i> Michx.	Carter 1979; Miller 1981	—
<i>Salix arctica</i> Richardson	Brooks 1987; Dearden 1979; Hay et al. 1994; Roberts 1980; Tyndall and Hull 1999	NL (S2)
<i>S. arctophila</i> Cockerell	Hay et al. 1992	ME (E); NL (S2)
<i>S. argyrocarpa</i> Andersson	Hay et al. 1994	ME (E), NH (T); NL (S1)
<i>S. brachycarpa</i> Nutt.	Brooks 1987	—
<i>S. calcicola</i> Fernald & Wiegand	Hay et al. 1992	NL (S3)
<i>S. chlorolepis</i> Fernald	Bouchard et al. 1983; Canadian Legal Information Institute 2008	G1; QC (S1)
<i>S. cordata</i> Muhl.	Hay et al. 1992	IL (E), NY (E), WI (E); NL (S1)
<i>S. discolor</i> Muhl.	Carter 1979	KY (H)
<i>S. herbacea</i> L.	Hay et al. 1992, 1994	ME (T), NH (T), NY (E); NL (S2)
<i>S. humilis</i> Marshall var. <i>tristis</i> (Aiton) Griggs	Tyndall and Hull 1999; Wherry 1963	—
<i>S. pedunculata</i> Fernald	Hay et al. 1992	—
<i>S. reticulata</i> L.	Hay et al. 1992	—
<i>S. × wiegandii</i> Fernald	Hay et al. 1992	—
SANTALACEAE		
<i>Comandra umbellata</i> Nutt.	Wherry 1963	—
SAXIFRAGACEAE		
<i>Chrysosplenium americanum</i> Hook.	Carter 1979	IN (T), KY (E)
<i>Parnassia grandifolia</i> DC.	Brooks 1987	FL (E), KY (E), NC (T), TN (SC)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Saxifraga aizoides</i> L.	Brooks 1987; Dearden 1979	NY (T)
<i>S. oppositifolia</i> L.	Brooks 1987; Dearden 1979	NY (E)
<i>S. rivularis</i> L.	Hay et al. 1992	NH (E), WA (S); NL (S2)
<i>S. virginiensis</i> Michx.	Brooks 1987; Tyndall 1992b, 1994; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963; Zika and Dann 1985	IL (E), IN (R)
<i>Tiarella cordifolia</i> L.	Carter 1979	NJ (E), WI (E)
SCROPHULARIACEAE		
<i>Agalinis acuta</i> Pennell	Tyndall 1994; Tyndall and Hull 1999	G1; USA (E); CT (E), MA (E), MD (E), NY (E), RI (E)
<i>A. obtusifolia</i> Raf.	Brooks 1987	KY (E), MD (E)
<i>A. paupercula</i> (A. Gray) Britton var. <i>paupercula</i>	Wherry 1963	PA (E)
<i>A. paupercula</i> var. <i>borealis</i> Pennell	Wherry 1963	OH (E), NY (T)
<i>A. tenuifolia</i> (Vahl) Raf. var. <i>tenuifolia</i>	Wherry 1963	—
<i>Aureolaria flava</i> (L.) Farw.	Wherry 1963	—
<i>A. pedicularia</i> (L.) Raf.	Brooks 1987	IA (E), ME (SC), MN (T)
<i>A. pedicularia</i> var. <i>pedicularia</i>	Wherry 1963	OH (E)
<i>Castilleja coccinea</i> (L.) Spreng.	Brooks 1987; Mansberg and Wentworth 1984; Wherry 1963	CT (E), KY (E), MD (E), ME (PX), NY (E), RI (H)
<i>C. septentrionalis</i> Lindl.	Dearden 1979	ME (SC), MI (T), MN (E), NH (T), VT (T)
<i>Chelone glabra</i> L.	Wherry 1963	NY (EV)
<i>Euphrasia nemorosa</i> Pers.	Carter 1979	MI (T)
<i>Linaria vulgaris</i> Mill.	Brooks 1987	—
<i>Melampyrum lineare</i> Lam.	Brooks 1987; Carter 1979; Mansberg and Wentworth 1984	IN (R), OH (T)
<i>Pedicularis canadensis</i> L.	Mansberg and Wentworth 1984	—
<i>Rhinanthus minor</i> L. subsp. <i>minor</i>	Carter 1979	—

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>Schwalbea americana</i> L.	Hay et al. 1992	G2; USA (E); CT (SC), FL (E), GA (E), KY (H), MD (E, X), NC (E), NJ (E), TN (PX, E)
<i>Verbascum blattaria</i> L.	Brooks 1987	–
<i>V. thapsus</i> L.	Carter 1979	–
<i>Veronica americana</i> Benth.	Carter 1979	IL (E), IN (X), KY (H), TN (SC)
<i>Veronicastrum virginicum</i> Farw.	Wherry 1963	MA (T), NY (T), VT (E)
SELAGINELLACEAE		
<i>Selaginella rupestris</i> (L.) Spring	Carter 1979	IN (T), OH (PRX)
<i>S. selaginoides</i> (L.) Mart. & Schrank	Hay et al. 1992	ME (T), MN (E), WI (E); NL (S4, S5)
SMILACACEAE		
<i>Smilax glauca</i> Walter	Mansberg and Wentworth 1984; Miller 1981; Tyndall and Farr 1990; Wherry 1963	–
<i>S. herbacea</i> L.	Wherry 1963	–
<i>S. rotundifolia</i> L.	Hull and Wood 1984; Mansberg and Wentworth 1984; Miller 1981; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	–
SOLANACEAE		
<i>Solanum dulcamara</i> L.	Carter 1979	–
THELYPTERIDACEAE		
<i>Phegopteris connectilis</i> (Michx.) Watt	Carter 1979	IA (E), IL (E), NY (EV), TN (SC)
<i>P. hexagonoptera</i> (Michx.) Fée	Wherry 1963	ME (SC), MN (T), NY (EV); QC (S2)
<i>Thelypteris noveboracensis</i> (L.) Nieuwl.	Mansberg and Wentworth 1984; Wherry 1963	IL (E), NY (EV)

Appendix 3. Continued.

Taxon	Citation	Global, Federal, State/Province Protected Status
<i>T. simulata</i> (Davenport) Nieuwl.	Tyndall and Hull 1999; Zika and Dann 1985	MD (T), NC (T), NY (EV), TN (PX, E); QC (SH)
TYPHACEAE		
<i>Typha angustifolia</i> L.	Wherry 1963	—
<i>T. latifolia</i> L.	Wherry 1963	—
URTICACEAE		
<i>Boehmeria cylindrica</i> (L.) Sw.	Wherry 1963	—
VIOLACEAE		
<i>Viola blanda</i> var. <i>palustriformis</i> A. Gray	Carter 1979	IA (E), IL (E)
<i>V. conspersa</i> Rchb.	Wherry 1963	—
<i>V. macloskeyi</i> subsp. <i>pallens</i> (Ging) M.S. Baker	Carter 1979	—
<i>V. palmata</i> L.	Wherry 1963	ME (PX), NH (E)
<i>V. palustris</i> L.	Hay et al. 1994	ME (E), NH (T); NL (S2, S3)
<i>V. pedata</i> L.	Brooks 1987; Wherry 1963	NH (T), NY (EV), OH (T)
<i>V. sagittata</i> Aiton	Pennell 1930; Tyndall 1992b, 1994, 2005; Tyndall and Farr 1990; Tyndall and Hull 1999; Wherry 1963	—
<i>V. sagittata</i> var. <i>ovata</i> (Nutt.) Torr. & A. Gray	Brooks 1987; Pennell 1930	WI (E); QC (S1)
<i>V. sororia</i> Willd.	Carter 1979	—
VITACEAE		
<i>Vitis aestivalis</i> Michx.	Wherry 1963	ME (E)