





**Appendix E1**  
**Flora Species Records of the Northern Territory Herbarium Database and**  
**Environment Australia Listings of Potential Flora Presence Based on Potential Habitat Presence**  
**for the Area 12°09' to 12°15'S; and 136°40' to 136°50'E**

**Key to Conservation Status**

*Territory Parks and Wildlife Commission Act 2000*

LC – Least Concern

DD – Data Deficient

NE – Not Evaluated

*Environment Protection and Biodiversity Conservation Act 1999*

V - Vulnerable

Nomenclature for native flora follows Wheeler (1992), Wightman & Andrews (1989), Brooker & Kleinig (1994), Brock (2001), except where more recent taxonomic revisions are known to have been published (eg. *Checklist of Northern Territory Vascular Plant Species*<sup>1</sup> Northern Territory Herbarium, 2003), and/or where the Northern Territory recognises a different binomial name. Other texts used to assist in identification include, Yunupinu et al. (1995), Milson (2000), Hacker (1990), Sainty & Jacobs (1994), Stephens & Dowling (2002), Smith (2002), Auld & Medd (1999).

Taxon	Conservation Status	
	NT	Comm.
ACANTHACEAE Hypoestes floribunda R.Br. Ruellia tuberosa L.		
AIZOACEAE Trianthema portulacastrum L.		
AMARANTHACEAE Achyranthes aspera L. Alternanthera dentata (Moench) Stuchlik Amaranthus sp Gomphrena celosioides Mart. Ptilotus spicatus F.Muell. ex Benth.		
ANACARDIACEAE Buchanania obovata Engl.		
ANNONACEAE Cyathostemma glabrum (Span.) Jessup Miliusa traceyi Jessup		
APOCYNACEAE Alyxia spicata R.Br. Catharanthus roseus (L.) G.Don Wrightia saligna (R.Br.) F.Muell. ex Benth.		

<sup>1</sup> [http://www.nt.gov.au/ipe/pwcnt/index.cfm?attributes.fuseaction=open\\_page&page\\_id=1794](http://www.nt.gov.au/ipe/pwcnt/index.cfm?attributes.fuseaction=open_page&page_id=1794)

Taxon	Conservation Status	
	NT	Comm.
ARECACEAE Arenga australasica Livistona humilis R.Br. Livistona inermis R.Br.		V
ASCLEPIADACEAE Cryptostegia madagascariensis Bojer ex Decne. Cynanchum carnosum (R.Br.) Schltr. Gymnanthera oblonga (Burm.f.) P.S.Green Marsdenia geminata (R.Br.) P.I.Forst. Sarcostemma viminale (L.) R.Br. subsp. brunonianum Sarcostemma viminale (L.) R.Br. subsp. brunonianum Tylophora flexuosa R.Br.		
ASTERACEAE Blumea saxatilis Zoll. & Moritzi Emilia sonchifolia (L.) DC. Melanthera biflora (L.) Willd. Pleurocarpaea denticulata Benth. Tridax procumbens L. Vittadinia spechtii N.T.Burb.	DD	
BIXACEAE Cochlospermum gillivraei Benth.		
BORAGINACEAE Cordia subcordata Lam. Heliotropium ventricosum R.Br.		
BURSERACEAE Canarium australianum F.Muell.		
CAESALPINIACEAE Cassia sp Senna sp Senna alata (L.) Roxb. Senna occidentalis (L.) Link		
CAPPARACEAE Capparis quiniflora DC. Capparis sepiaria L.		
CARYOPHYLLACEAE Polycarpaea violacea (Mart.) Benth.		
CASUARINACEAE Casuarina equisetifolia J.R.Forst. & G.Forst.		
CELASTRACEAE		

Taxon	Conservation Status	
	NT	Comm.
Cassine melanocarpa (F.Muell.) Kuntze Denhamia obscura (A.Rich.) Meisn. ex Walp.		
<b>COMBRETACEAE</b> Lumnitzera racemosa Willd. Terminalia latipes Benth.		
<b>COMMELINACEAE</b> Cartonema parviflorum Hassk. Commelina ensifolia R.Br.		
<b>CONVOLVULACEAE</b> Ipomoea diversifolia R.Br. Ipomoea graminea R.Br. Ipomoea pes-caprae (L.) Sweet Ipomoea pes-caprae (L.) Sweet subsp. brasiliensis (L.) Ooststr. Ipomoea quamoclit L. Jacquemontia paniculata (Burm.f. ) Hallier f. Merremia dissecta (Jacq.) Hallier f. Merremia quinata (R.Br.) Ooststr.		
<b>CUCURBITACEAE</b> Coccinia grandis (L.) Voigt Cucumis melo L. Diplocyclos palmatus (L.) C.Jeffrey Zehneria mucronata (Blume) Miq.	DD	
<b>CYPERACEAE</b> Bulbostylis barbata (Rottb.) C.B.Clarke Cyperus brevifolius (Rottb.) Hassk. Cyperus compressus L. Cyperus involucratus Roth Cyperus iria L. Cyperus polystachyos Rottb. Cyperus rotundus L. Cyperus sphacelatus Rottb. Cyperus stoloniferus Retz. Fimbristylis ferruginea Vahl Fimbristylis littoralis Gaudich. Fimbristylis sericea R.Br. Fuirena ciliaris (L.) Roxb. Schoenoplectus litoralis (Schrad.) Palla	LC	
<b>DILLENIACEAE</b> Hibbertia brownii Benth. Hibbertia cistifolia R.Br. ex DC. Hibbertia dealbata (R.Br. ex DC.) Benth. Hibbertia lepidota R.Br. ex DC. Pachynema complanatum R.Br. ex DC.		

Taxon	Conservation Status	
	NT	Comm.
DIOSCOREACEAE Dioscorea bulbifera L.		
EBENACEAE Diospyros compacta (R.Br.) Kosterm. Diospyros cordifolia Roxb. Diospyros humilis (R.Br.) F.Muell. Diospyros maritima Blume		
ELAEOCARPACEAE Elaeocarpus arnhemicus F.Muell.		
ERIOCAULACEAE Eriocaulon australe R.Br.		
EUPHORBIACEAE Croton habrophyllus Airy Shaw Drypetes deplanchei (Brongn. & Griseb.) Merr. Euphorbia armstrongiana Boiss. ex DC. Euphorbia atoto G.Forst. Euphorbia cyathophora Murray Euphorbia heterophylla L. Euphorbia hirta L. Euphorbia mitchelliana Boiss. Flueggea virosa (Roxb. ex Willd.) Voigt Glochidion xerocarpum (O.Schwarz) Airy Shaw Jatropha gossypifolia L. Petalostigma pubescens Domin Petalostigma quadriloculare F.Muell. Phyllanthus sp Phyllanthus amarus K.Schum. & Thonn.		
FABACEAE Abrus precatorius L. Aphyllodium schindleri Pedley Canavalia papuana Merr. & L.M.Perry Clitoria ternatea L. Crotalaria brevis Domin Dalbergia sissoo Roxb. Daviesia reclinata A.Cunn. Desmodium scorpiurus (SW.) Desv. Desmodium tortuosum (Sw.) DC. Desmodium triflorum (L.) DC. Gompholobium subulatum Benth. Stylosanthes hamata (L.) Taub. Tephrosia sp Tephrosia laxa Domin Tephrosia leptoclada Benth.		

Taxon	Conservation Status	
	NT	Comm.
Tephrosia oligophylla Benth.	NE	
Tephrosia porrecta R.Br. ex Benth.		
Vigna adenantha (G.Mey.) Marechal	Y	
FLAGELLARIACEAE		
Flagellaria indica Willd.		
GOODENIACEAE		
Goodenia sp		
LAMIACEAE		
Ocimum tenuiflorum L. var. anisodorum (F.Muell.) Domin		
Plectranthus scutellarioides (L.) R.Br.		
LAURACEAE		
Cassytha filiformis L.		
Litsea glutinosa (Lour.) C.B.Rob.		
LEMNACEAE		
Spirodela polyrhiza (L.) Schleid.		
LENTIBULARIACEAE		
Utricularia sp		
LILIACEAE		
Crinum angustifolium R.Br.		
Curculigo ensifolia R.Br.		
Curculigo ensifolia R.Br. var. longifolia Benth.		
Thysanotus chinensis Benth.		
LOGANIACEAE		
Mitrasacme brachystemonea Domin		
Mitrasacme elata R.Br.		
Mitrasacme laevis Benth.		
Strychnos lucida R.Br.		
LORANTHACEAE		
Amyema miquelii (Lam. ex Miq.) Tiegh.		
Amyema villiflora (Domin) Barlow		
Decaisnina petiolata (Barlow) Barlow		
Decaisnina petiolata (Barlow) Barlow subsp. angustata Barlow		
Dendrophthoe glabrescens (Blakely) Barlow		
LYCOPODIACEAE		
Lycopodiella cernua (L.) Pic.-Serm.		
MALVACEAE		
Abelmoschus moschatus Medik.		
Abutilon indicum (L.) Sweet		
Gossypium hirsutum L.		

Taxon	Conservation Status	
	NT	Comm.
Hibiscus sp Sida acuta Burm.f.		
<b>MELIACEAE</b> Aglaia brownii Pannell		
<b>MENISPERMACEAE</b> Pachygone ovata (Poir.) Hook.f. ex Thomson Tinospora smilacina Benth.		
<b>MIMOSACEAE</b> Acacia drepanocarpa F.Muell. Acacia gonocarpa F.Muell. Acacia leptocarpa A.Cunn. ex Benth Acacia oncinocarpa Benth. Acacia plectocarpa A.Cunn. ex Benth. subsp. plectocarpa Acacia simsii A.Cunn. ex Benth. Acacia simsii A.Cunn. ex Benth. Acacia sublanata Benth. Acacia yirrkallensis Specht Leucaena leucocephala (Lam.) de Wit Neptunia gracilis Benth.		
<b>MORACEAE</b> Trophis scandens (Lour.) Hook. & Arn.		
<b>MYRISTICACEAE</b> Myristica insipida R.Br. Aegiceras corniculatum (L.) Blanco Ardisia humilis Vahl		
<b>MYRTACEAE</b> Calytrix exstipulata DC. Eucalyptus miniata A.Cunn. ex Schauer Melaleuca acacioides F.Muell. Melaleuca cajuputi Powell Melaleuca dealbata S.T.Blake Melaleuca nervosa (Lindl.) Cheel Osbornia octodonta F.Muell. Syzygium angophoroides (F.Muell.) B.Hyland Syzygium minutiflorum (F.Muell.) B.Hyland		
<b>NYCTAGINACEAE</b> Pisonia aculeata L. Nymphaea violacea Lehm.		
<b>OLEACEAE</b> Jasminum aemulum R.Br. Jasminum didymum G.Forst. Jasminum molle R.Br.		



Taxon	Conservation Status	
	NT	Comm.
ONAGRACEAE Ludwigia hyssopifolia (G.Don) Exell		
OPILIACEAE Cansjera leptostachya Benth.		
ORCHIDACEAE Dendrobium affine Steud. Habenaria ochroleuca R.Br. Liparis habenarina (F.Muell.) Benth.	DD	
PANDANACEAE Pandanus spiralis R.Br.		
PASSIFLORACEAE Passiflora foetida L.		
PHILYDRACEAE Philydrum lanuginosum Sol. ex Gaertn.		
PLUMBAGINACEAE Aegialitis annulata R.Br.		
POACEAE Alloteropsis semialata (R.Br.) Hitchc. Arundo donax L.		
RUBIACEAE Spermacoce remota Lam.		



A summary of the community types found in each of the dominant landforms in the survey area are summarised below. A detailed species list follows.

## Community Types

### 1. Marine Deposits

This is an association of several concise marine plant communities. They occur on quaternary marine deposits subject to periodic inundation by saline or brackish waters. Typically these comprise of saline marine flats dominated by halophytic chenopods with occasional mangroves through to closed mangrove forests (mangals). The distribution, abundance and form of these communities is related to inundation frequency, degree of salination, freshwater inputs and a variety of other ecological factors. The substrates are generally fine silts to sands.

#### 1a Mangroves

The two main mangrove areas are the deltaic tidal swamps of Macassar and Crawford Creeks. In addition, there is a discontinuous, narrow fringe of mangroves along the shore of Melville Bay. Mangrove forests range from closed forests to low open woodlands. Species present include *Ceriops tagal*, *Bruguiera exaristata*, *Excoecaria ovalis*, *Avicennia marina* white mangrove, *Lumnitzera racemosa* red flowered mangrove, *Osbornea octodonata*, *Scyphiphora hydrophylla*, *Aegiceras corniculatum* river mangrove and rarely, *Rhizophora stylosa* stilt root mangrove. Typically they form mosaic communities with saline swamps.

#### 1b Saline Flats

Typically saline flats and swamps form mosaic communities with mangroves. Saline swamps consist of *Acrostichum speciosum* mangrove fern and samphire flats that can contain such species as *Acrostichum speciosum* mangrove fern, *Tectornia* sp, *Halosarcia* spp. and *Sesuvium portulacastrum*. Plants of the landward mangrove fringe include *Melaleuca acacoides* small leaved paperbark and *Scyphiphora hydrophylacae*.

### 2. Quaternary Coastal Dune Vegetation

Vegetation within this zone occupies quaternary coastal dunes, sand plains, swales, sand plains and beaches. The origin of the substrate is primarily marine in origin comprising mainly of siliceous sands.

#### 2a. Foredune Vegetation

A few elongated sand dunes lie parallel to and only a few meters from the seashore. These are all fixed dunes with little apparent sign of sand movement. The supralittoral area is colonised by a few hardy species tolerant of the harsh exposed conditions of the foreshore being nutrient poor and exposed to salt laden sea breezes, storm surges and cyclonic winds. Species For the most part they are fixed with tussock or hummock grasses such as the tall *Sorghum plumosum* perennial sorghum, *Spinifex longifolius*, *Chrysopogon elongatus*, the creeping vines of *Ipomoea pes caprae*, the herbs *Gomphrena canescens* pink everlasting and *Euphorbia atoto*, the shrub *Guettarda speciosa*, and the tree *Casuarina equestifolia* casuarina which may form copses seaward of the dunes.

#### 2b. Coastal Monsoon Vine Thicket

Coastal Monsoon Vine Thicket (CMVT) are a common community scattered throughout the fringing dune ridges extending into the hind dune swales on consolidated beach sands. On the seaward edge these low (3-4 m), dense wind pruned thickets range from copses of several plants to larger extensive patches on the beach dune ridges and are more extensive on southerly facing beaches where they are more sheltered from prevailing winds. Typically

these communities are only 30-50 m wide and often have extensive convolutions along the seaward edge, arising from wind shear. They provide protection to the dunes from the strong on-shore winds that buffet the coast especially during the wet season. CMVT also forms a band between terrestrial vegetation and marine plant communities. Common species include *Aglaia brownii*, *Celtis philippensis* celtis, *Cordia subcordata* cordia, *Canarium australianum*, *Diospyros* spp., *Premna* spp., *Emmenosperma cunninghamii*, *Scaevola taccada* pipe tree, *Sterculia quadrifida* peanut tree, *Pouteria sericea* pouteria, *Strychnos lucida* Strychnine tree and *Thespesia populnoides*.

### **2c. Mixed Eucalypt Coastal Woodland**

Mixed Eucalypt Coastal Woodland is a eucalypt woodland on consolidated coastal sand masses. Typically it occupies dune swales but also occurs on sand masses landward of headlands. Soils are typically deep, coarse white sands with a grey organic sandy layer for the top 3-5 cm. It is likely that two communities are represented, one being found in dune swales, the second being found on sand sheets around rocky headlands. However owing to the reduced areal extent of the latter (as a result of the refinery) it is not possible to definitively separate the two. Both contain constituent CMVT species.

The canopy is dominated by *Corymbia polycarpa* long fruited bloodwood with *Corymbia polysciada*, *Corymbia ferruginea* and *Eucalyptus albens* also present in reduced numbers. The shrub layer is characterised by *Pandanus spirilis* fan pandanus, *Erythrophleum chlorostachys* ironwood, *Buchanania obovata* Munydjutj, *Brachychiton diversifolius* northern kurrajong, *Brachychiton paradoxus* red flowering kurrajong, *Livistonia humilis* fan palm, *Planchonia careya* cocky apple and wattles *Acacia* spp. Lower shrubs, herbs and groundcovers include *Ficus scobina* sandpaper fig, *Persoonia falcata* milky plum, *Cassytha filiformis* dodder, *Gomphrena canescens* pink everlasting and *Smilax australis* smilax. Grasses include *Heteropogon triticeus* balck spear grass, *Cymbopogon bombycinus* lemon scented grass, *Panicum mindanese*, *Themeda arguens* and *Imperata cylindrica*

## **3. Communities on Cainozoic Sand Deposits**

The dominant geology of the Gove Peninsula, is a Cainozoic laterite containing ferricrete and bauxite. Outcrops of Proterozoic Metamorphics and Proterozoic Granite also occur in the area. Within the study area, soils are typically undifferentiated cainozoic brown to red earthy sands, silts and clays often with lateritic gravels or outcrops. Outcropping of Proterozoic Metamorphics and Proterozoic Granite also occurs though in limited extent.

### **3a. Eucalyptus Communities**

#### **3ai Eucalyptus tetradonta Darwin stringybark Open Forest on Sands**

This forest occurs on deep sandy soils generally have little gravel content. In this community, *Eucalyptus tetradonta* Darwin Stringybark 12-16m in height is accompanied by a few tall *Eucalyptus polycarpa*, *Eucalyptus miniata* yellowjack. The shrub layer is characterised by *Brachychiton diversifolius* and *B. paradoxus*, *Buchanania obovata*, *Planchonia careya*, *Petalostigma pubescens* quinine tree and shorter shrubs such as *Livistonia humilis* fan palm, *Hibbertia dealbata* and *Pachynema dilatatum*. Vines, twinners and prostate herbs include *Grewia retusifolia*, *Cassytha filiformis* dodder, *Smilax australis* smilax and *Vigna lanceolata*. Gramineous flora is sparse and is dominated by *Heteropogon triticeus* black spear grass with *Chrysopogon fallax* spear grass, *Mnesithea rottboellioides*, *Aristida.browniana* and *Themeda arguens*.

### **3aii Eucalyptus tetradonta Darwin stringybark Open Forest on Lateritic Gravels**

This community occurs on pebbling lateritic surfaces to the south of the RDA. The soils are typically grey to yellow, gravelly sands. Structurally the canopy vegetation is similar to unit 3ai, if not slightly denser (greater foliage projection) but has a sparser shrub and groundcover. The canopy is dominated by *Eucalyptus tetradonta* to 16m. Associated species in the shrub layers are *Petalostigma quadriloculare*, *Livistona humilis*, *Acacia lamprocarpa* hickory wattle, *Buchanania obovata* the small shrub *Hibertia dealbata* and *Pachynema dilatatum*. Few grass species occur, *Heteropogon triticeus* being the dominant grass, with some *Themeda arguens*, *Aristida browniana* kerosene grass and *Ectrosia schultzei*.

### **3aiii Eucalyptus tetradonta Darwin stringybark Woodland**

This vegetation type occurs on the deep sandy soils derived from granite, similar to (3ai) but obviously of lower fertility and water holding capacity. In this community *Eucalyptus tetradonta* is dominant but the trees are more widely spaced than in the above communities. There are a few tall *Eucalyptus polycarpa* and *Eucalyptus alba* an understorey comprised of *Acacia lamprocarpa*, *Pandanus spirilis*, occasional *Livistona humilis*, *Grevillia pteridiifolia* fern leaf grevillea, *Acacia lamprocarpa* and the flowering shrub *Calythrix exstipulata*. Gramineous flora is sparsely distributed and comprises *Aristida browniana*, *Pseudopogonatherum contortum* and *Setaria* sp. These grasses are indicative of the drier soil regime.

## **3b. Melaleuca species Dominated Communities**

### **3bi Melaleuca viridiflora broad leaved paperbark/ Pandanus spirilis spring pandanus Open (Savanna) Woodland with a Grassy Understorey**

This community occupies the sand (sheet) plain at the seaward edge of the elevated lands occupied by *Eucalyptus tetradonta* Darwin stringybark communities and littoral vegetation types (marine plant communities). The soils appear to be kept moist for much of the year with the exception of the mid to late dry season. These soils are typically deep, coarse white sands with a grey organic sandy layer for the top 3-5 cm.

The community is dominated by *Melaleuca viridiflora* broad leaved paperbark interspaced with *Pandanus spirilis* spring pandanus, *Grevillea pteridiiflora*. *Corymbia polycarpa* are present especially around the edges of the savanna plain where it fringes *Eucalyptus tetradonta* communities. The bulk of the ground space is covered by dense short grasses and beneath these where the ground is kept moist *Drosera* sp. sundew are present. Grasses include taxa from the genera "Dimeria, Schizachrium, Chrysopogon, Mnesithea, and Alloeropsis. *Pandanus spirilis* becomes more abundant closer to localised ephemeral drainage lines and soaks.

### **3bii Melaleuca spp. / Pandanus spirilis spring pandanus Woodland to Open Forest**

Situated entirely within Vegetation Community 3bi, this vegetation occupies a drainage depression/soak system to the south east of the disposal area where the soil is more or less continually moist. These soils are typically deep, coarse clayey-sands with a grey organic sandy layer.

This community is dominated by the tall *Melaleuca dealbata* which are clustered along a semi permanent drainage line and fringed by *M. viridiflora*. *Corymbia polycarpa* is present within the canopy and *Pandanus spirilis* forms a dense layer. Other species present include *Eucalyptus* spp and species characteristic of monsoon vine thickets. The bulk of the ground space is covered by dense short grasses belonging to the genera *Dimeria* and *Schizachrium*. Tall grasses include *Chrysopogon elongatus* spear grass, *Mnesithea rottboellioides*, *Alloeropsis semialata* grass and tall reeds and herbs colonise the small gutters. *Lobelia dioica*, *Utricularia chrysantha*, *Ludwigia octovalvis*, *Cyperus* spp. and *Fimbristylis* sp. area common in the wettest areas.

#### 4. Disturbed Land

Disturbed areas comprise species that have colonised disturbed sites, have been planted for landscape amenity purposes (L), have been planted for revegetation (Rv), as well as regrowth (Re) which has naturally recolonised following disturbance and which is characterised by species from surrounding areas. Disturbed land also includes untended or tended land composed of native and exotic species (D). The proposed construction accommodation area at Nhulunbuy falls into the regrowth category as it was previously used as the construction area for the initial refinery construction. The vegetation surrounding the construction accommodation area comprises *Eucalyptus tetradonta* Open Forest and the regrowth in this area has affinities with this.

**Table E2**  
**Flora Species List**

Family	Species	Status	Common Name	Community Type											
				1a/1b	2a	2b	2c	3ai	3aii	3aiii	3bi	3bii	4		
													D	R	
Acanthaceae	<i>Acanthus ilicifolius</i>		Mangrove Holly	X											
Aizoaceae	<i>Sesuvium portulacastrum</i>			X											
Amaranthaceae	<i>Gomphrena canescens</i>				X		X	X	X	X				X	X
Anacardaceae	<i>Buchanania obovata</i>		Green plum					X							
	<i>Semecarpus australiensis</i>		Native Cashew									X			
Annonaceae	<i>Miliusa brahei</i>					X									
	<i>Polyalthia australis</i>					X									
Apocynaceae	<i>Alyxia spicata</i>					X									
	<i>Parsonisa velutina</i>							X							
	<i>Tabernaemontana orientalis</i>		Ervatamia			X									
Arecaccae	<i>Livistonia humilis</i>											X			
Asclepidaceae	<i>Calotropis gigantea</i>		Giant Rubber Bush											X	
	<i>Sarcostemma viminalis</i>		Caustic Vine		X										
Asteraceae	<i>Biddens</i> sp	*				X									
	<i>Conyza bonariensis</i>	*					X				X				
	<i>Emelia sonchifolia</i>		Emelia					X						X	
	<i>Tridax procumbens</i>		Tridax Daisy											X	
Avicennaceae	<i>Avicennia eucalyptifolia</i>			X										X	
Bixaceae	<i>Cochlospermum fraseri</i>		Yellow Kapok												
Boraginaceae	<i>Cordia subcordata</i>		Cordia			X									
Burseraceae	<i>Canarium australianum</i>		Canarium			X									
Caesalpinaceae	<i>Erythrophleum chlorostachys</i>		Ironwood								X				

Family	Species	Status	Common Name	Community Type											
				1a/1b	2a	2b	2c	3ai	3aii	3aiii	3bi	3bii	4		
													D	R	
	<i>Peltophorum pterocarpum</i>		Yellow Flame Tree			X									
	<i>Senna alata</i>	B	Candlebush					X	X					X	
	<i>Tamarindus indica</i>		Tamarind											X	
Campanulaceae	<i>Lobelia dioica</i>												X		
Capparaceae	<i>Capparis quiniflora</i>				X	X		X							
	<i>Capparis sepiaria</i>							X							
Caryophyllaceae	<i>Polycarpaea</i> sp							X							
Casuarinaceae	<i>Casuarina equestifolia</i>		Casuarina		X									X	X
Chenopodaceae	<i>Halosarcia</i> sp			X											
Combretaceae	<i>Lumnitzera racemosa</i>		Red Mangrove	X											
	<i>Terminalia carpentariae</i>		Billy Goat Plum					X			X			X	
	<i>Terminalia catappa</i>		Indian Almond			X									
	<i>Terminalia</i> sp						X								
Commelinaceae	<i>Commelina</i> sp.						X								
Convolvulaceae	<i>Evolvulus alsinoides</i>						X		X		X				
	<i>Ipomoea pes-capare</i>		Beach Morning Glory		X	X								X	
	<i>Ipomoea pes-tigridis</i>	*												X	
	<i>Merremia aegyptia</i>	*	White Convolvulus Creeper											X	
	<i>Merremia</i> sp							X	X		X	X			
Cyperaceae	<i>Cyperus exaltatus</i>						X						X		
	<i>Cyperus javanicus</i>													X	
	<i>Cyperus polystachos</i>												X	X	
	<i>Cyperus</i> sp 1						X						X		
	<i>Cyperus</i> sp 2						X								
	<i>Fimristylis</i> sp												X		
Dilleniaceae	<i>Hibbertia dealbata</i>							X	X		X				
	<i>Hibbertia lepidota</i>										X				
	<i>Hibbertia oblongata</i>						X	X	X						
	<i>Pachynema dilatatum</i>							X	X		X				
Dioscoreaceae	<i>Dioscorea transversa</i>		Long Yam			X									
Droseraceae	<i>Drosera petiolaris</i>										X				
Ebenaceae	<i>Diospyros humilis</i>					X									

Family	Species	Status	Common Name	Community Type										
				1a/1b	2a	2b	2c	3ai	3aii	3aiii	3bi	3bii	4	
													D	R
	<i>Diospyros maritima</i>													
Elaeocarpaceae	<i>Elaeocarpus arnhemensis</i>					X								
Euphorbiaceae	<i>Antidesma ghesaembilla</i>		Black Current			X								
	<i>Breynia cernua</i>					X								
	<i>Bridelia tomentosa</i>					X								
	<i>Drypetes deplanchei</i>					X						X		
	<i>Euphorbia atoto</i>				X									
	<i>Euphorbia cyanthophora</i>	*	Painted Spurge											X
	<i>Euphorbia peplus</i>	*	Petty Spurge											X
	<i>Euphorbia</i> sp 1	*												X
	<i>Euphorbia</i> sp 2	*												X
	<i>Excoecaria ovalis</i>			X										
	<i>Flueggea virosa</i>		White Currant			X								
	<i>Macaranga involuocrata</i>		Macaranga			X						X	X	
	<i>Mallotus nesophyllus</i>					X								
	<i>Omalanthus novo-guineensis</i>					X								
	<i>Petalostigma pubescens</i>		Quinine Tree				X	X		X	X			
	<i>Petalostigma quadriloculare</i>						X	X	X	X	X			
	<i>Phyllanthus</i> sp.										X	X		
Fabaceae	<i>Abrus precatorius</i>		Crab's Eye Vine			X								
	<i>Austrodolichos errabundus</i>		Yam				X		X					
	<i>Canavalia rosea</i>				X									
	<i>Crotalaria goreensis</i>	*	Gambia Pea			X		X	X					X
	<i>Desmodium</i> sp							X						
	<i>Glycine</i> sp.							X						
	<i>Jacksonia dilatata</i>						X	X	X	X	X			
	<i>Macroptilium atropupureum*</i>	*	Sirato											X
	<i>Sesbania cannabina</i>		Yellow Bush Pea											
	<i>Stylosanthes hamata</i>	*												X
	<i>Stylosanthes viscosa</i>	*					X			X				X
	<i>Vigna lanceolata</i> var. <i>filiformis</i>							X	X					X
	<i>Vigna vexillata</i>		Stringy Yam					X	X					
	<i>Zornia</i> sp								X					
Flagellariaceae	<i>Flagellaria indica</i>		Flagellaria			X								
Goodenaceae	<i>Calogyne pilosa</i>						X							
	<i>Scaveola taccada</i>		Pipe Tree		X	X								
Haemodoraceae	<i>Haemodorum brevicaule</i>		Red Root				X				X			
Hypoxidaceae	<i>Curculigo ensifolia</i>										X	X		
Iridaceae	<i>Patersonia macrantha</i>											X		



Family	Species	Status	Common Name	Community Type											
				1a/1b	2a	2b	2c	3ai	3aii	3aiii	3bi	3bii	4		
													D	R	
Lamiaceae	<i>Coleus scutellaroides</i>														
	<i>Hyptis suaveolens</i>	B	Hyptis			X	X	X	X		X		X		
Lauraceae	<i>Cassytha filiformis</i>		Dodder				X	X	X	X					
	<i>Litsea glutinosa</i>					X									
Lecythidaceae	<i>Planchonia careya</i>		Cocky Apple					X							
Lentibulariaceae	<i>Utricularia fulva</i>		Bladderwort								X	X			
Liliaceae	<i>Crinum augustifolium</i>		Onion Lily									X			
	<i>Protasparagus racemosus</i>		Native Asparagus			X			X	X					
Loganiaceae	<i>Strychnos lucida</i>		Strychnine Tree			X									
Lythraceae	<i>Pemphis acidula</i>						X							X	
Malvaceae	<i>Abutilon indicum</i>							X	X						
	<i>Hibiscus meraukensis</i>							X			X		X		
	<i>Hibiscus tiliaceus</i>		Beach Hibiscus		X	X							X	X	X
	<i>Sida acuta</i>	B											X		
	<i>Sida cordifolia</i>	B	Flannel Weed										X		
	<i>Thespesia populneoides</i>				X	X									
	<i>Urena lobata</i>						X				X				
Melastomataceae	<i>Osbeckia australiana</i>										X	X			
Meliaceae	<i>Aglaia brownii</i>					X									
	<i>Dysoxylon</i> sp					X									
Menispermaceae	<i>Tinospora smilacina</i>		Sanke Vine			X									
Mimosaceae	<i>Acacia auriculiformis</i>		Black Wattle				X	X	X		X				
	<i>Acacia holosericea</i>							X					X	X	
	<i>Acacia lamprocarpa [aulacocarpa]</i>		Hickory Wattle				X	X	X	X	X			X	X
	<i>Acacia leptocarpa</i>										X		X	X	
	<i>Acacia multisilique</i>							X					X	X	
	<i>Acacia simsii</i>												X	X	
	<i>Acacia sublanata</i>							X							
	<i>Acacia torulosa</i>												X	X	
	<i>Leucaena leucocephala</i>	*	Coffee Bush										X		
Moraceae	<i>Ficus benjamina</i>					X									
	<i>Ficus hispida</i>					X									
	<i>Ficus opposita</i>														X
	<i>Ficus scobina</i>		Sandpaper Fig					X							
	<i>Ficus virens</i>		Banyan										X		

Family	Species	Status	Common Name	Community Type											
				1a/1b	2a	2b	2c	3ai	3aii	3aiii	3bi	3bii	4		
													D	R	
Myrsinaceae	<i>Aegiceras corniculatum</i>		Club Mangrove	X											
	<i>Ardisia humilis</i>	*	Ardisia											X	
Myrtaceae	<i>Calytrix exstipulata</i>		Balarrwalarr					X		X					
	<i>Corymbia ferruginea</i>		Long Fruited Bloodwood				X	X	X					X	X
	<i>Corymbia polycarpa</i>						X	X						X	X
	<i>Corymbia polysiada</i>		small leaved paperbark												
	<i>Melaleuca acacoides</i>				X									X	X
	<i>Melaleuca cajuputi</i>													X	
	<i>Melaleuca dealbata</i>													X	
	<i>Melaleuca viridiflora</i>		Broad Leaved Paperbark								X	X			
	<i>Eucalyptus albens</i>						X			X				X	X
	<i>Eucalyptus miniata</i>		Yellowjack					X						X	X
	<i>Eucalyptus papuana</i>													X	X
	<i>Eucalyptus ptychocarpa</i>		Swamp Bloodwood					X							
	<i>Eucalyptus tetradonta</i>		Darwin Stringybark					X	X	X					X
	<i>Osbornia octodonta</i>			X											
	<i>Syzygium suborbiculare</i>		Red Bush Apple			X	X								
Oleaceae	<i>Jasminum didymum</i>	lc	Native Jasmin			X									
Ongaraceae	<i>Ludwigia octovalvis</i>	lc											X		
Orchidaceae	<i>Dendrobium</i> sp										X				
Pandanaceae	<i>Pandanus spirilis</i>		Spring Pandanus								X				
Passifloraceae	<i>Adenia heterophylla</i>					X									
	<i>Passiflora foetida</i>	*	Wild Passionfruit			X								X	
Philydraceae	<i>Philydrum lanuginosum</i>		Frogsmouth										X		
Poaceae	<i>Alloteropsis semialata</i>		Cockatoo Grass				X				X	X			
	<i>Aristida browniana</i>		Kerosene Grass					X	X	X					
	<i>Bambusa arnhemica</i>		Bamboo			X								X	
	<i>Cenchrus ciliaris</i>	*	Buffel Grass											X	
	<i>Chloris gayana</i>	*	Rhodes Grass											X	
	<i>Chrysopogon elongatus [Vetiveria elongata]</i>						X				X	X			
	<i>Chrysopogon fallax</i>		Spear Grass					X	X		X				
	<i>Cymbopogon bombycinus</i>		Lemon Grass					X	X	X					
	<i>Cynodon dactylon</i>													X	
	<i>Dimeria</i> sp										X	X			
	<i>Ectrosia leporina</i>							X			X	X	X		
	<i>Ectrosia schultzei</i>											X			
	<i>Eriachne obtusa</i>							X							

Family	Species	Status	Common Name	Community Type											
				1a/1b	2a	2b	2c	3ai	3aii	3aiii	3bi	3bii	4		
													D	R	
	<i>Eriachne schultzi</i>							X				X	X		
	<i>Heteropogon contortus</i>		Bunch Spear Grass				X								
	<i>Heteropogon triticeus</i>		Giant Spear Grass/Bush Sugar Cane					X						X	
	<i>Imperata cylindrica</i>		Blady Grass				X								
	<i>Mnesteia rottboellioides</i>										X	X			
	<i>Panicum sp</i>							X	X	X					
	<i>Panicum mindanese</i>													X	
	<i>Pennisetum polystachion</i>	B	Mission Grass											X	
	<i>Pseudopogonatherum contortum</i>						X	X	X	X	X	X			
	<i>Setaria sp</i>						X	X	X	X					
	<i>Schizachrium sp</i>										X	X			
	<i>Schizachrium fragile</i>										X	X	X		
	<i>Sorghum intrans</i>		Annual Sorghum											X	
	<i>Sorghum plumosum</i>		Perennial Sorghum		X	X	X	X	X	X				X	
	<i>Spinifex longifolius</i>		Coastal Spinifex		X	X									
	<i>Thaumastochloa major</i>				X	X								X	
	<i>Themeda arguens</i>							X							
	<i>Themeda quadrivalvis</i>	B	Grader Grass											X	
	<i>Vetiver zizanioides</i>	*	Vetiver Grass												X
Polygonaceae	<i>Antigonon leptopus</i>	*	Coral Vine											X	
	<i>Comesperma secundum</i>							X							
Proteaceae	<i>Banksia dentata</i>										X	X			
	<i>Grevillea formosa</i>														X
	<i>Grevillea heiosperma</i>														X
	<i>Grevillea pteroidifolia</i>		Fern Leaved Grevillea						X		X			X	
	<i>Hakea arborescens</i>		Hakea				X							X	
	<i>Persoonia falcata</i>		Milky Plum				X	X							
Pteridaceae	<i>Acrostichum speciosum</i>		Mangrove Fern	X											
Restionaceae	<i>Dapsilanthus spathaceus</i>												X		
Rhamnaceae	<i>Alphitonia excelsa</i>		Red Ash			X	X	X							
	<i>Emmenosperma cunninghamii</i>				X	X									
Rhizophoraceae	<i>Bruguiera exaristata</i>			X											
	<i>Ceriops tagal</i>			X											
	<i>Rhizophora stylosa</i>		Stilt Root Mangrove	X											
Rubiaceae	<i>Coeliospermum reticulatum</i>							X							
	<i>Guettarda speciosa</i>				X	X									
	<i>Pogonolobus reticulatus</i>							X							
	<i>Scyphiphora hydrophyllacea</i>			X											

Family	Species	Status	Common Name	Community Type											
				1a/1b	2a	2b	2c	3ai	3aii	3aiii	3bi	3bii	4		
													D	R	
Rutaceae	<i>Murraya paniculata var ovatifoliata</i>					X									
Santalaceae	<i>Exocarpos latifolius</i>		Native Cherry				X							X	
Sapindaceae	<i>Cupaniopsis anacarinoides</i>					X									
	<i>Distochostemon hispidulus</i>						X								
	<i>Dodonaea physocarpa</i>		Hop Bush											X	
	<i>Ganophyllum falcatum</i>					X									
Sapotaceae	<i>Pouteria sericea</i>		Pouteria				X								
Scrophulariaceae	<i>Buchnera linearis</i>							X	X		X				
Solanaceae	<i>Physlais minima</i>		Goosberry											X	
Sterculiaceae	<i>Brachychiton diversifolius</i>		Northern Kurrajong				X	X	X	X	X			X	X
	<i>Brachychiton megaphyllum</i>													X	X
	<i>Brachychiton paradoxus</i>		Red Flowering Kurrajong			X	X	X	X	X	X				
	<i>Helicteres cana</i>						X	X							
	<i>Sterculia quadrifida</i>		Peanut Tree						X					X	X
Tiliaceae	<i>Grewia breviflora</i>					X									
	<i>Grewia multiflora</i>					X									
	<i>Grewia retusifolia</i>		Emu Berry				X	X	X	X					
Typhaceae	<i>Typha dominigensis</i>		Bullrush										X		
Ulmaceae	<i>Celtis philippensis</i>		Celtis			X									
	<i>Trema tomentosa</i>														
Vitaceae	<i>Ampelocissus acetosa</i>		Wild Grape					X							
	<i>Cissus adnata</i>					X									
Verbenaceae	<i>Avicennia marina</i>		White Mangrove	X											
	<i>Premna acuminata</i>				X	X									
	<i>Premna serratifolia</i>				X	X									
	<i>Stachytarpheta australis</i>	B	Snakeweed										X		
	<i>Vitex glabrata</i>		Black Plum			X			X		X				
Zygophyllaceae	<i>Tribulus cistoides</i>	B	Bindi Eye		X										

**Status:** \* - exotic species/environmental weeds; N – native species not endemic to Gove Peninsula; Ic – lease concern (source: Territory Parks and Wildlife Commission Act 2000); A – weeds that must be eradicated, B – weeds that must be contained (source: Weeds Management Act 2001)

## Vegetation Communities

### *Communities on Quaternary Marine Deposits*

1a/1b – Marine Plant Communities

### *Communities in Quaternary Coastal Sand Deposits*

2a – Foredune Vegetation

2b – Coastal Monsoon Vine Thicket

2c – Mixed Eucalypt Coastal Woodland

### **Communities on Cainozoic Sand Plains**

3a *Eucalyptus* dominated communities

3ai – *Eucalyptus tetradonta* Darwin stringybark Open Forest on Sands

3aii – *Eucalyptus tetradonta* Darwin stringybark Open Forest on Lateritic Gravels

3aiii – *Eucalyptus tetradonta* Darwin stringybark Woodland

3b *Melaleuca* dominated communities

3bi – *Melaleuca viridiflora* broad leaved paperbark/*Pandanus spirillis* spring pandanus Open (Savanna) Woodland with a Grassy Understorey

3bii – *Melaleuca* spp./*Pandanus spirillis* spring pandanus Woodland to Open Forest

4 – Vegetation on Disturbed surfaces (various geologies and soils)

R – Revegetation

D – Disturbed (includes mapping units D, Re, L)



**Table E3**  
**Rirratjinu Ethnobotany of Species Recorded During Field Survey (aboriginal use and significance after Yunupinu et al, 1995)**

Family	Species Name	Rirratjinu Name	Rirratjinu Use
Acanthaceae	<i>Acanthus ilicifolius</i>	Banuminy (Dhuwa)	
Aizoaceae	<i>Sesuvium portulacastrum</i>	Birrkpirrknanin (Dhuwa)	
Amaranthaceae	<i>Gomphrena canescens</i>	Banbalarri (Dhuwa)	
Anacardaceae	<i>Buchanania obovata</i>	Munydjutj (Yirritja)	Green fruits eaten. Inner bark mixed with water as treatment for toothache. Fruit and seeds pounded to make an infant food.
	<i>Semecarpus australiensis</i>	Ganyawu (Yirritja)	Three to four weeks after the fruit has fallen it is collected and roasted on a fire; seeds are then eaten.
Annonaceae	<i>Miliusa brahei</i>	Gutjawutja (Dhuwa)	Fruit eaten when ripe.
	<i>Polyalthia australis</i>	Dhaman (Dhuwa)	Ripe fruit collected from ground and eaten
Apocynaceae	<i>Alyxia spicata</i>	Burrba (Dhuwa)	
Arecaccae	<i>Livistonia humilis</i>	Dhalpi (Yrritja)	The inner cabbage was eaten in the past but is not eaten now; emus and occasionally children eat the black fruit.
Asclepidaceae	<i>Sarcostemma viminalis</i>	Dhalkurrnanin (Dhuwa)	
Bixaceae	<i>Cochlospermum fraseri</i>	Djiranbulk (Dhuwa)	Root of young plants roasted and eaten.
Boraginaceae	<i>Cordia subcordata</i>	Buyama	Timber used for ornamental carving, seeds can be eaten.
Burseraceae	<i>Canarium australianum</i>	Barrata (Dhuwa)	Timber used for making ornamental carvings.
Caesalpiniaceae	<i>Erythrophleum chlorostachys</i>	Buwatji (Yirritja)	Adult plants are called Maypiny while the young plants are called Buwatji; the wood is considered dangerous as it is sharp and strong and can cause injuries to children if they step or fall on it; it is used to make clapsticks (Bilma) and boomerangs (Galiwali); leaves are crushed and washed over a person who is the victim of sorcery.
	<i>Senna alata</i>		Used to treat ringworms.
	<i>Tamarindus indica</i>	Djamban (Yirritja)	Fruit can be soaked in water and the liquid drunk to cure colds.
Capparaceae	<i>Capparis quiniflora</i>	Dirridirri (Dhuwa)	
Caryophyllaceae	<i>Polycarpaea sp</i>		
Casuarinaceae	<i>Casuarina equestifolia</i>	Mawurraki (Yirritja)	Timber used as firewood and digging sticks; good shade tree.
Combretaceae	<i>Lumnitzera racemosa</i>		Red flowers are sucked (especially by children) for nectar. Hard timber used to make clasp sticks.
	<i>Terminalia catappa</i>	Matpana (Yirritja)	The fruit are eaten when ripe; seed is eaten after the fruit is cracked open
	<i>Terminalia carpentariae</i>	Mamanbu (Dhuwa)	Green fruit eaten, gum cooked or eaten raw.
Convolvulaceae	<i>Ipomoea pes-capare</i>	Murukun (Dhuwa)	Tuberous roots eaten after roasting, leaves used to stop bleeding from cuts.
Dioscoreaceae	<i>Dioscorea transversa</i>	Manmuna (Dhuwa)	Tuber cooked in coals or in a ground oven; it has a taste and texture similar to sweet potato and is the most sought after yam.
Ebenaceae	<i>Diospyros humilis</i>	Burrpurr (Yirritja)	Orange fruit are eaten.
	<i>Diospyros maritima</i>	Yundidi (Yirritja)	The juice from the yellow fruit can cause a severe skin irritation and the whole plant is considered poisonous; the leaves and bark can be used as a

Appendix E.3  
*Rirratjinu Ethnobotany of Species  
Recorded During Field Survey  
(aboriginal use and significance after  
Yunupinu et al, 1995)*

Family	Species Name	Rirratjinu Name	Rirratjinu Use
			fish poison.
Elaeocarpaceae	<i>Elaeocarpus arnhemensis</i>	Yulumuru (Dhuwa)	
Euphorbiaceae	<i>Antidesma ghesaembilla</i>	Barrata (Dhuwa)	The fruit are eaten when ripe (purple); the fruit flesh stains the lips and tongue purple; they are sweet, very tasty and much sought after when in season (December to April).
	<i>Drypetes deplanchei</i>	Djilka (Dhuwa)	Fruit eaten when ripe. Leaves used to flavour seafood dishes.
	<i>Euphorbia atoto</i>	Birrpirknanin (Dhuwa)	
	<i>Flueggea virosa</i>	Bulun/Gumbu (Dhuwa)	The white globbular fruit are eaten when ripe; they are sweet and much sought after; fruiting signifies the end of the wet season and also that the parrot fish are ready to be hunted; small spears are made from the hard springy straight stems.
	<i>Mallotus nesophyllus</i>	Galurra (Dhuwa)	The fruits are eaten when pale and soft (ripe).
	<i>Omalanthus novo-guineensis</i>	Gudatpa (Dhuwa)	The stems are used to make light fishing spears.
	<i>Petalostigma pubescens</i>	Mayadilmatla (Dhuwa)	
Fabaceae	<i>Abrus precatorius</i>	Yirinanin	The red and black seeds are boiled to make them soft and then used to make necklaces. The name also refers to the necklace.
	<i>Austrodolichos errabundus</i>	Wanydjarrpu	A low twiner with a perennial tuber; the yam is roasted in the coals, then cleaned and softened by crushing gently.
	<i>Glycine tomentella</i>	Yuluk (Dhuwa)	The tubers are dug up, cleaned and coked in hot sand and ashes before eating.
	<i>Jacksonia dilatata</i>	Dhurrurrungitj	Smoke from the plant is used to stop lactation.
	<i>Vigna vexillata</i>	Yukuwa (Yirritja)	Tuber is eaten after roasting.
Flagellariaceae	<i>Flagellaria indica</i>	Darwirr (Yirritja)	Stems are split and made into ceremonial armbands and bracelets.
Goodenaceae	<i>Scaveola taccada</i>	Luniny (Yirritja)	Smoking pipes are made from straight stems.
Haemodoraceae	<i>Haemodorum brevicaule</i>	Yirrinanin	Small perennial tuber is dug up and used to produce a red/brown dye.
Hypoxidaceae	<i>Curculigo ensifolia</i>	Dhunguruk (Dhuwa)	The elongate fleshy tubers are dug up and roasted on coals.
Lauraceae	<i>Cassytha filiformis</i>	Burrnburrun (Yirritja)	Children eat the fruit when ripe; the vine is used as a string to wrap up meat when nothing better is available.
	<i>Litsea glutinosa</i>	Butjirinanin (Dhuwa)	The leaves are crushed into the hands and rubbed directly onto the chest to alleviate chest congestion
Lecythidaceae	<i>Planchonia careya</i>	Dhangi (Dhuwa)	Soft yellow inner fruit eaten when ripe; bark used as a fish poison.
Liliaceae	<i>Crinum augustifolium</i>	Warrkarr (Yirritja)	Annual herb with an onion like tuber. When flowers are produced stingrays are fat and ready to be hunted.
Lythraceae	<i>Pemphis acidula</i>	Dakul (Dhuwa)	Tree is used for yam digging, woomera hooks, and picks to extract oysters from shell.
Malvaceae	<i>Abutilon indicum</i>	Banbalarri (Dhuwa)	A small shrub with yellow flowers that when flowering signals that the terns are laying their eggs and it is a good time to collect them.



Appendix E.3  
*Rirratjinu Ethnobotany of Species  
Recorded During Field Survey  
(aboriginal use and significance after  
Yunupinu et al, 1995)*

Family	Species Name	Rirratjinu Name	Rirratjinu Use
	<i>Hibiscus meraukensis</i>	Malnany (Yirritja)	An annual herb with a perennial tuber that grows in open forests during wet season; tuber is eaten after roasting in the ashes.
	<i>Hibiscus tiliaceus</i>	Yal (Dhuwa)	Pale light timber is used for ornamental carving, straight stems used to make short fishing spears.
Meliaceae	<i>Thespesia populneoides</i>	Meli (Dhuwa)	Stems used for spears.
	<i>Aglaia brownii</i>	Djirrkawul (Dhuwa)	Stems are used to make fighting spears called Dhumadal/Wararri.
Menispermaceae	<i>Tinospora smilacina</i>	Burrpa (Dhuwa)	Leaves used as a curse (mixed with the urine or faeces or sweat of the person) or heated in a fire and waved at arms length to stop the wind from blowing.
Mimosaceae	<i>Acacia lamprocarpa [aulacocarpa]</i>	Dhurrtji (Dhuwa)	When the ancestral Djan'kwa arrived at Yalanbara he walked on the land carrying his digging sticks (Mawalan) and clap sticks (Bilma) made from Dhurrtji. When the re-enactment ceremony of this event is undertaken clap sticks and digging sticks made from Dhurrtji are used. Digging sticks and clap sticks for everyday use are occasionally made from Dhurrtji. The timber is also used to make woomera (Galpu) shafts and hooks.
	<i>Acacia holosericea</i>	Dhurrtji (Dhuwa)	As for <i>A. lamprocarpa</i> .
	<i>Acacia leptocarpa</i>	Dhurrtji (Dhuwa)	As for <i>A. lamprocarpa</i> .
	<i>Acacia torulosa</i>	Galayin (Dhuwa)	A shade tree and a 'calendar' plant. When in flower it indicates that the turtles are fat and ready to be hunted. The bark was used to make grass skirts which were worn during ceremonies. The flowers are called Mayatili. The gum from the stems and branches may be eaten. The timber is used to make Woomera (Galpu) shafts.
Moraceae	<i>Ficus scobina</i>	Muthi' (Dhuwa)	Fruits eaten when ripe, Inner bark used as a traement for vomiting and diarrhoea.
	<i>Ficus virens</i>	Rripipi (Dhuwa)	Fruits eaten when ripe; bark of the prop roots used for string baks, when men prepare for sacred ceremony they sit beneath it to sing.
Myrsinaceae	<i>Aegiceras corniculatum</i>	Bitjininy (Dhuwa)	Leaves have a salt coating and can be used to flavour meats.
Myrtaceae	<i>Melaleuca acacoides</i>	Gulun'kulun (Dhuwa)	Leaves are used as medicine.
	<i>Melaleuca leucadendron</i>	Dhulwu (Yirritja)	The inner bark was boiled and the liquid used to cure vomiting and diarrhoea; timber used to make canoes; bark used to wrap bread made from <i>Cycas armstrongii</i> (Nathu); water may be released from chopping into swellings on the trunk; bark can be used to make coolamons to carry food and other articles.
	<i>Melaleuca viridiflora</i>	Dindin (Dhuwa)	Young leaves are crushed in the hands and the scent inhaled to alleviate colds and congestion; alternatively the leaves are boiled and the vapour inhaled; flowering signals the best time to hunt turtles.
	<i>Corymbia ferruginea</i>	Badawili (Yirritja)	The honey from the bee-hives collected from this species is especially sweet.
	<i>Corymbia polycarpa</i>	Dhumulu (Dhuwa)	Grows in low lying areas; insect galls found in the branches are broken off the tree and opened up; the grubs and the white flesh are eaten raw; hollow stems and branches used to make didgeridus.

Appendix E.3  
*Rirratjinu Ethnobotany of Species  
Recorded During Field Survey  
(aboriginal use and significance after  
Yunupinu et al, 1995)*

Family	Species Name	Rirratjinu Name	Rirratjinu Use
Pandaceae	<i>Corymbia polysciada</i>	Gudirri (Yirritja)	<p>This plant is considered to be the grandmother of <i>Corymbia papuana</i> and <i>C. polysciada</i> which have the same name.</p> <p>The fruits of this tree are called Guyurru and the small black seeds eaten by children; hollow stems and branches used to make didgeridus.</p> <p>Considered to be the grandchild of <i>E. albens</i>.</p> <p>Hollow stems and branches used to make didgeridus.</p> <p>The bark is peeled off, flattened, dried and used for traditional paintings; sheets of bark may be used as roofing and wall material for shelters; new leaves are crushed and boiled and the liquid rubbed onto sores, children eat the seeds; ; hollow stems and branches used to make didgeridus (Yidaki); when in flower it is time to collect sugarbag.</p> <p>Fruit eaten when ripe.</p> <p>The young leaves are used to make string for bracelets, mats and dilly bags; soft flesh (cabbage) at base of leaf applied to open wounds as an antiseptic; the fruit are burnt in a pit and the stems of <i>Elaeocharis dulcis</i> is then placed over the fire to produce steam, a person lies on the steaming bed and this relieves back pain.</p>
	<i>Eucalyptus albens</i>	Gudirri (Yirritja)	
	<i>Eucalyptus miniata</i>	Gunurra (Yirritja)	
	<i>Eucalyptus papuana</i>	Gudirri (Yirritja)	
	<i>Eucalyptus ptychocarpa</i>	Dhumulu (Dhuwa)	
	<i>Eucalyptus tetradonta</i>	Gadayka (Dhuwa)	
	<i>Osbornia octodonta</i>	Manyarr (Dhuwa)	
	<i>Syzygium suborbiculare</i>	Larrani (Dhuwa)	
	<i>Pandanus spirilis</i>	Gunga (Dhuwa)	
	Passifloraceae	<i>Adenia heterophylla</i>	
Philydraceae	<i>Passiflora foetida</i>	Gana (Dhuwa)	pulp is eaten when ripe.
	<i>Philydrum lanuginosum</i>	Burrumburr (Dhuwa)	Stems and leaves soaked in warm or hot water and the liquid is consumed to treat diarrhoea.
Poaceae	<i>Alloteropsis semialata</i>	Bulmirri (Yirritja)	Basal stem is pulled out of the ground roughened to remove fibres and dirt and then used to dip honey out of sugarbag.
Polygonaceae	<i>Chrysopogon elongatus</i>	Bawu (Dhuwa)	Sometimes used to tie or wrap articles.
	<i>Chrysopogon fallax</i>	Ritharr (Yirritja)	When it shoots at the start of the wet season the new shoots are sharp and can stick in your feet.
	<i>Cymbopogon bombycinus</i>	Gawulurmanin (Dhuwa)	The leaves are boiled in water and cooled liquid is used as an external wash to treat vomiting and Diarrhoea.
	<i>Heteropogon triticeus</i>	Gaditjirri (Yirritja)	The stems of this grass are broken into shorter lengths and chewed to obtain a sweet juice; stems also used a toy spears by children.
	<i>Imperata cylindrica</i>	Ritharr (Yirritja)	Shoots in the early wet are sharp and can pierce the skin; leaf blades are sharp and can cut the skin
Proteaceae	<i>Comesperma secundum</i>	Nangungangu (Dhuwa)	<p>When this plant is in flower it signals the best time to collect sugarbag (Guku), as native bee-hives contain the most honey; early in the morning when the dew is on the ground, children collect the flowers and suck the sweet nectar from them; The hard dense timber is used to pierce the nasal septum; the young inflorescence spike is used to carry fire.</p>
	<i>Banksia dentata</i>	Gulpu (Yirritja)	

Appendix E.3  
*Rirratjinu Ethnobotany of Species  
Recorded During Field Survey  
(aboriginal use and significance after  
Yunupinu et al, 1995)*

Family	Species Name	Rirratjinu Name	Rirratjinu Use
Pteridaceae	<i>Persoonia falcata</i>	Dangapa (Dhuwa)	Fruit eaten when ripe.
	<i>Acrostichum speciosum</i>	Mayawarku (Yirritiya)	
Restionaceae	<i>Dapsilanthus spathaceus</i>	Muyukuya (Yirritija)	The stems are pulled up and laid on the ground and used as a clean layer to rest food on during preparation.
Rubiaceae	<i>Guettarda speciosa</i>	Gamarran (Yirritija)	Leaves are used to cook damper or used as plates
	<i>Pogonolobus reticulatus</i>		Roots are dug up and the bark chipped off and then added to water along with fibres to be dyed. Water is boiled to produce colours ranging from yellow to brown depending upon length of boiling.
	<i>Scyphiphora hydrophylacae</i>	Milinyarr (Dhuwa)	Production of fruit signals the kingfish are ready to hunt. Production of a swollen orange peduncle means that the box jellyfish is at its most potent and children are not allowed to swim.
Rhizophoraceae	<i>Rhizophora stylosa</i>	Walmu (Dhuwa)	Mangrove worms (Latjin) found in the trunks and dead branches.
Sapotaceae	<i>Pouteria sericea</i>	Wunapu (Dhuwa)	Fruit eaten when ripe.
Santalaceae	<i>Exocarpos latifolius</i>	Dakul'nanin (Dhuwa)	smoke repels mosquitos and sandflies.
Sapindaceae	<i>Ganophyllum falcatum</i>	Dilminyin (Yirritija)	The fruit are eaten when ripe (red) but too much makes the mouth sore and induces coughing.
Scrophulariaceae	<i>Buchnera linearis</i>		
Solanaceae	<i>Physalis minima</i>	Bodan	The small globular fruit inside the papery lantern like casing can be eaten.
Sterculiaceae	<i>Brachychiton diversifolius</i>	Nanunguwa (Yirritija)	bark used to make fibre for string bags and arm bands, seeds are eaten after the fruit placed on a fire to cook the seeds and remove irritant hairs.
	<i>Brachychiton paradoxus</i>	Dharrangulk (Dhuwa)	Tap root of young plant dug up, peeled and chewed for water. Seeds are roasted to remove irritant hairs or ground to make a paste for infants.
Tiliaceae	<i>Grewia retusifolia</i>	Murrjumun (Dhuwa)	The bark of the root is stripped, pounded and boiled. The resultant jelly like substance is used to draw and heal boils.
Typhaceae	<i>Typha dominigensis</i>	Gulwani (Yirritija)	Occasionally used to make small spears for children.
Ulmaceae	<i>Celtis philippensis</i>	Lilirtjin (Dhuwa)	Fruit eaten when ripe.
Vitaceae	<i>Ampelocissus acetosa</i>	Lingarr (Yirritija)	Sweet fruit eaten when ripe, but leaves a bitter aftertaste; seeds not eaten.
	<i>Cissus adnata</i>	Burpunanin (Dhuwa)	Leaved are pounded a little and waved in the air to make the wind stop blowing.
Verbenaceae	<i>Avicennia marina</i>	Manyarr (Dhuwa)	Timber is used medicinally. It is burnt and the ash soaked in salt water and applied to sores, boils, yinea and ringworm.
	<i>Premna serratifolia</i>	Duttji (Yirritija)	The straight stems are used as firesticks.
	<i>Vitex glabrata</i>	Wundan (Dhuwa)	Sweet fruit eaten.



**Appendix E4**  
**EFFECTS OF SULFUR DIOXIDE ON VEGETATION**  
**AT GOVE, NORTHERN TERRITORY**

A Report to URS Ltd  
D Doley  
Brisbane

August 2003

## Table of Contents

	Page
1 Introduction	1
2 Ambient conditions	1
2.1 The Gove environment	1
2.2 Estimated SO <sub>2</sub> concentrations at selected locations	1
2.3 Selecting a benchmark SO <sub>2</sub> concentration	2
3 Evidence of vegetation sensitivity to SO <sub>2</sub>	4
3.1 Short-term exposure	4
3.2 Long-term exposure	7
3.2.1 Experimental studies	7
3.2.2 Ecological studies	8
3.3 Future SO <sub>2</sub> emissions	8
4 Conclusions	9
References	9
Appendix 1	10

### List of Tables

Table 1. Summary of 1-h maximum ground level concentrations of SO <sub>2</sub> at selected locations at Gove, interpolated from data of PAE (2002)	2
Table 2. Plant species from southern Australia assigned to SO <sub>2</sub> sensitivity classes by O'Connor et al. (1974)	6

### List of Figures

Figure 1 Relationship between ambient guideline SO <sub>2</sub> concentration and averaging period (Data from NEPC 1998)	3
Figure 2 Relationships between median extent of injury in plants from seven classes of sensitivity to SO <sub>2</sub> fumigation and dose from data of O'Connor et al. (1975)	5

## 1 INTRODUCTION

Ambient air quality guidelines for SO<sub>2</sub> have been established with the objective of protecting human health, in particular the more sensitive groups within the population (NEPC, 1998). If these guidelines can be shown to adequately protect vegetation, then there is little need to consider additional secondary or informal guidelines for the protection of vegetation.

The effects of SO<sub>2</sub> on Australian plant species have not been studied extensively. Two principal types of investigation have been carried out; firstly the short-term exposure of plant seedlings to high concentrations of SO<sub>2</sub> (O'Connor et al., 1975), and longer-term studies on crop plants or small specimens of tree species (Murray et al., 1991). The results of these investigations cannot be reconciled directly, as both the concentrations and exposure times are very different. However, both types of study are relevant to an understanding of the effects of SO<sub>2</sub> on plants, as there are circumstances in which vegetation may be exposed for long periods to low concentrations, or to relatively high concentrations for shorter periods.

At Gove, most attention has been directed towards short-term ambient air quality (PAE, 2002), in which the maximum one-hour mean concentrations of SO<sub>2</sub> have been measured and modelled in the vicinity of the alumina refinery. Therefore, before considering vegetation responses, it is necessary to determine the most likely maximum ambient concentrations of SO<sub>2</sub> in locations where the establishment or retention of vegetation may be important.

## AMBIENT CONDITIONS

### 2.1 The Gove environment

Gove has a markedly seasonal rainfall environment, which results in vigorous plant growth during the wet season and a dying back of some species and quiescence in others during the dry season. Because vegetation is generally more susceptible to injury by SO<sub>2</sub> during periods of active growth, the exposure conditions during the wet season are likely to be of greater relevance to the occurrence of injury than are dry season conditions.

An air quality monitoring station at the alumina refinery provided records of SO<sub>2</sub> concentration that were analysed in 1-h intervals by PAE (2002). The maximum 1-h average concentration for the period reported was 2300 µg m<sup>-3</sup> (Table 11, PAE 2002).

### 2.2 Estimated SO<sub>2</sub> concentrations at selected locations

PAE (2002) reported dispersion modelling that allowed the prediction of the maximum 1-h SO<sub>2</sub> concentrations in the vicinity of the alumina refinery for a whole year, and the mean SO<sub>2</sub> concentrations for both wet and dry seasons, of 5 and 7 months' duration respectively. The interpolated maximum concentrations for locations near the refinery where there may be plant communities of interest are indicated in Table 1, which also indicates the maximum 1-h SO<sub>2</sub> concentration at the refinery site. It is relevant that for 1-h averaging periods, the maximum concentration permitted under the guidelines is approximately 570 µg m<sup>-3</sup>.

For the purpose of this review, it will be assumed that some plant species occurring on the Gove Peninsula will be susceptible to SO<sub>2</sub> injury at concentrations similar to those that affect SO<sub>2</sub> susceptible species elsewhere. That is, responses are likely to become evident only at concentrations that are substantially higher than those established in the Ambient Air Quality Guidelines (NEPC, 1998).

**Table 1**  
**Summary of maximum ground level concentrations of SO<sub>2</sub> (µg m<sup>-3</sup>) for wet and dry seasons at selected locations at Gove, interpolated from modelled data of PAE (2002).**

Assessment source	Averaging Time	SO <sub>2</sub> concentrations at specified locations			
		Refinery	Drimmie Peninsula	Nhulunby	West Gove Harbour
Fig 8. Dry season, model	7 months	200	1.5	<0.5	10
Fig 9. Wet season, model	5 months	200	30	3	17
Fig 10. Annual, model	1 hour	5000	1500	300	800
Fig 11. Annual, measured	1 hour	2300	na	na	na

Mean SO<sub>2</sub> concentrations modelled by PAR (2002) for wet and dry season conditions used different averaging times, and except for the refinery location the dry season maxima were lower than those for the wet season (Table 1). The maximum 1-hour modelled SO<sub>2</sub> concentration at the refinery was approximately twice the maximum measured value (Table 1), although PAE (2002) concluded that the measured value may have been an underestimate of the concentration due to the effects of buildings in the vicinity of the monitoring station.

The model predicts (cf. Fig 11, PAE 2002) that the upper 1 percentile of observations at the alumina refinery are associated with ambient SO<sub>2</sub> concentrations up to 10 times those actually recorded. Therefore, the predicted maximum ground level concentrations of SO<sub>2</sub> at sites around the refinery and indicated in Table 1 may be somewhat higher than those actually experienced on a seasonal basis. In addition, the predictions for seasonal maxima and the annual maximum use different averaging times, so that it is appropriate to examine the relationship between concentration and averaging time.

### 2.3 Selecting a benchmark ambient SO<sub>2</sub> concentration

Because air quality guidelines have been established for 1h, 24h and 1 year (NEPC 1998), the process of relating the duration of exposure at which plants respond to a given ambient SO<sub>2</sub> concentration to the guideline values for concentration and exposure duration is not straightforward. However, there is a close mathematical relationship between exposure durations and ambient guideline values (Figure 1), which can be described by

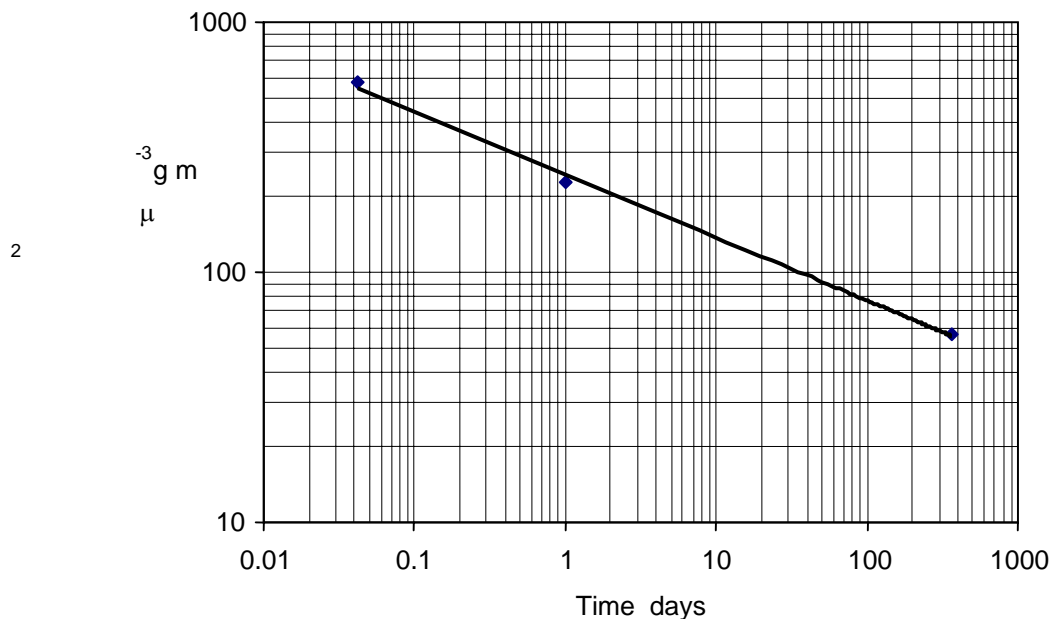
$$[\text{SO}_2] = 246.2 T^{0.252}$$

where  $T$  is averaging time in days.

Using the equation or the graphical representation (Figure 1), an interpolation may be used to assess the likelihood that a plant may be injured at an ambient concentration other than the published guideline values.

In the case of the seasonal mean SO<sub>2</sub> concentrations, the interpolated wet season guideline value would be approximately 70 µg SO<sub>2</sub> m<sup>-3</sup> and the interpolated dry season guideline value would be 65 µg SO<sub>2</sub> m<sup>-3</sup>. It is suggested that these values can be compared directly with the predicted seasonal average concentrations in Table 1.





**Figure 1**

**Relationship between ambient guideline SO<sub>2</sub> concentration and averaging period (Data from NEPC 1998). The regression equation is  $[SO_2] = 246.2 T^{-0.252}$  where  $T$  is averaging time in days.**

This approach may be challenged on the basis that the guideline SO<sub>2</sub> concentrations for each averaging time were not intended to conform to a mathematical relationship. However, if there is any consistency in the response of organisms to SO<sub>2</sub> exposure, there should be a continuous relationship between exposure duration and the concentration at which an effect becomes evident. On this basis, the use of procedures to permit interpolation between averaging times of 1 day and 1 year is defended.

### 3 EVIDENCE OF VEGETATION SENSITIVITY TO SO<sub>2</sub>

The evaluation of vegetation responses to pollutant exposure requires consideration of both the concentration of the pollutant and the duration of exposure. The majority of studies, summarised in Appendix 1, have been concerned with longer-term exposures, and have applied relatively low ambient concentrations. These conditions are appropriate where exposure is expected to be chronic, such as in regions where there are multiple sources. For single, geographically restricted sources, short-term exposures may be more important, particularly if there is not a very definite prevailing wind direction. Under these circumstances, exposure to a relatively high pollutant concentration for one hour may be followed by many hours of very low concentration. During the period of very low pollutant concentration, a substantial amount of tissue repair may occur, so that dysfunction occurring during and immediately after a fumigation event may be eliminated in time.

In the same way, the assessment of injury during a fumigation event, immediately after it, or at some later time may provide quite different indications of injury.

### 3.1 Short-term exposure

Short-term fumigation studies, with exposure durations of 4 or 6 hours were carried out by O'Connor et al. (1974) using SO<sub>2</sub> concentrations of 1, 2 or 3 ppm (1 ppm being equivalent to 2860 µg m<sup>-3</sup> under standard conditions). That is, the concentrations used were at least 4 times and up to 14 times the 1-hour guideline value of approximately 600 µg m<sup>-3</sup>. Assessments were made at the end of the fumigation period, although the authors did note the development of symptoms during fumigations in some sensitive species. Therefore, extrapolation of their sensitivity scales to lower exposures introduces considerable uncertainties, and must be treated with caution.

O'Connor et al. (1974) used a scale of injury to foliage to establish seven sensitivity groups. If the product of SO<sub>2</sub> concentration and duration exposure is described as the dose, then the lowest dose applied was approximately 17000 µg h m<sup>-3</sup> (2860 µg m<sup>-3</sup> for 6 hours). If the median plant responses in terms of leaf injury for each of the sensitivity classes are compared with the SO<sub>2</sub> dose (Figure 2), it is evident that the threshold for the appearance of injury in the most sensitive class (number 6) occurs at a dose of approximately 6000 µg h m<sup>-3</sup>. This extrapolation corresponds to the observation by O'Connor et al. (1975) that sensitive species first showed injury after 2 to 3 h of exposure to 1 ppm (2860 µg m<sup>-3</sup>) SO<sub>2</sub>.

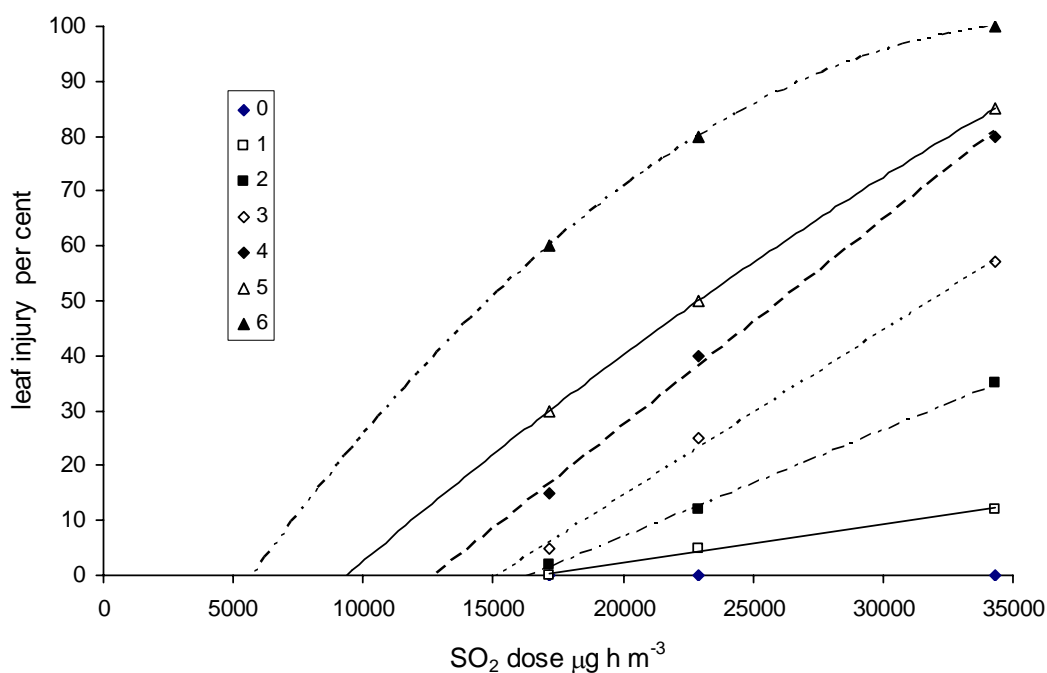


Figure 2

Relationships between the median extent of leaf injury in plants from seven classes of sensitivity to SO<sub>2</sub> fumigation at products of concentration (µg m<sup>-3</sup>) and duration (h) indicated. Class 0 is least sensitive and Class 6 is most sensitive. Calculated from data of O'Connor et al. (1974).

Experimental long-term fumigations of some important crop and tree species from southern Australia are reviewed in Appendix 1. Again, there is no information for species of tropical origin. In the absence of data to the contrary, it will be assumed that there will be a similar range in species sensitivities in the Northern Territory as there is in the species occurring in southern Australia; that is, there will be some species that are resistant to SO<sub>2</sub> exposure, and others that are susceptible.

A summary of experiments by O'Connor et al. (1974) is provided in Table 2. Most of these species are from southern Australia, but they may indicate the range of sensitivities that could be encountered in Arnhem Land.

The list of most sensitive species includes several *Acacia* and *Eucalyptus* species, and fewer representatives of *Banksia* and *Melaleuca*. The sensitive species of *Eucalyptus* are found in several subgenera, and within each subgenus there is generally a range of sensitivities. Therefore, it is not possible to extrapolate from this data set to indicate which of the many untested species may be particularly sensitive to SO<sub>2</sub>. There are almost equal numbers of species of both *Acacia* and *Eucalyptus* that are resistant to SO<sub>2</sub>, and all species of *Casuarina* investigated were very resistant to SO<sub>2</sub>. Therefore, it is very difficult to predict from taxonomic associations the likely sensitivity of species, or even genera, of plants to SO<sub>2</sub>. No direct information is available on the sensitivity of rainforest species to SO<sub>2</sub>. However, there is evidence that some rainforest and vine thicket species may be sensitive to gaseous fluoride (Doley, unpublished), and this sensitivity may also be expressed with respect to SO<sub>2</sub>.

In the absence of specific information, it needs to be assumed that some species in the Gove area will be very sensitive to SO<sub>2</sub>.

**Table 2**

**Plant species from southern Australia assigned to SO<sub>2</sub> sensitivity classes by O'Connor et al., (1974) on the basis of short-term (4 or 6 h), high-concentration fumigations of seedlings.**

Species	Common name	Sensitivity class
Most sensitive		
<i>Acacia verniciflua</i>	Varnish wattle	6
<i>Eucalyptus crenulata</i>	Silver gum	6
<i>Eucalyptus elata</i>	River peppermint	6
<i>Eucalyptus forrestiana</i>	Fuchsia gum	6
<i>Eucalyptus nicholii</i>	Willow-leaved peppermint	6
<i>Eucalyptus polyanthemus</i>	Red box	6
<i>Eucalyptus radiata</i>	Narrow leaved peppermint	6
<i>Eucalyptus regnans</i>	Mountain ash	6
Sensitive		
<i>Acacia parvissima</i>	Ovens wattle	5
<i>Acacia prominens</i>	Golden rain wattle	5
<i>Banksia collina</i>	Hill banksia	5
<i>Banksia marginata</i>	Silver banksia	5
<i>Eucalyptus aggregata</i>	Black gum	5
<i>Eucalyptus astringens</i>	Brown mallet	5
<i>Eucalyptus cosmophylla</i>	Bog gum	5
<i>Eucalyptus macrorhyncha</i>	Red stringybark	5
<i>Eucalyptus occidentalis</i>	Swamp yate	5
<i>Eucalyptus perriniana</i>	Spinning gum	5
<i>Eucalyptus risdonii</i>	Silver peppermint	5
<i>Eucalyptus saligna</i>	Sydney blue gum	5
<i>Eucalyptus viminalis</i>	Manna gum	5
<i>Leptospermum lanigerum</i>	Woolly tea-tree	5
<i>Melaleuca ericifolia</i>	Swamp paperbark	5
<i>Melaleuca heugellii</i>	Chenille honey-myrtle	5
<i>Melaleuca lanceolata</i>	Moonah	5
<i>Melaleuca squarrosa</i>	Scented paperbark	5

Species	Common name	Sensitivity class
<i>Melaleuca styphelioides</i>	Prickly-leaved paperbark	5

O'Connor et al. (1974) observed that the most plant species exhibited injury after exposure to 1 ppm (2860  $\mu\text{g m}^{-3}$ ) for two hours. This is equivalent to a dose of 5700  $\mu\text{g h m}^{-3}$ , or slightly more than the maximum 1-h concentration predicted to occur near the refinery. If all these assumptions hold, then the most sensitive plant species may be visibly injured by  $\text{SO}_2$  concentrations predicted to occur at least once during a year with the environmental conditions specified by PAE (2002).

Provided that the frequency of occurrence of the maximum concentration events is low, it is concluded that the extent of injury occurring in plants at the refinery site would be small, and may be confined to a small proportion of all the leaves on a plant.

### 3.2 Long-term exposure

Long-term exposure studies are more relevant to regional pollutant sources than to the relationship between point sources and their environments. However, the effects of point sources may be discernible after a period of time due to the recurrence of elevated ambient  $\text{SO}_2$  concentrations.

#### 3.2.1 Experimental studies

The experimental studies summarised in Appendix 1 show that the thresholds for significant reductions in plant growth attributes vary greatly between species and between plant attributes assessed. However, in crop and tree species ambient concentrations of more than about 200  $\mu\text{g m}^{-3}$  for exposure durations of more than 100 days were required in order to depress growth.

Plants that show an injury response after exposure to 100  $\mu\text{g SO}_2 \text{ m}^{-3}$  for 40 days should just be protected from injury by the air quality guideline (Figure 2). Plants that are affected only at higher  $\text{SO}_2$  concentrations greater than 100  $\mu\text{g SO}_2 \text{ m}^{-3}$  for 40 days should be protected from injury by the application of the guideline.

In the cases of the species surveyed in Appendix 1, injury was recorded in the most sensitive after exposure to 122  $\mu\text{g SO}_2 \text{ m}^{-3}$  for 120 days (*Eucalyptus marginata* in Table 5). The air quality guideline value appropriate to an averaging period of 100 days is approximately 80  $\mu\text{g SO}_2 \text{ m}^{-3}$ . Therefore, an ambient concentration 50% greater than the air quality guideline would be necessary in order to induce injury in this species.

From these considerations, it may be concluded that  $\text{SO}_2$  concentrations that meet the longer-term ambient air quality guidelines are very unlikely to result in detectable reductions in plant growth or in the appearance of visible injury to foliage.

Species occurring in the Gove area were not included in the studies reviewed in Appendix 1. However, if it can be assumed that species from tropical and temperate regions have a similar range of sensitivities to  $\text{SO}_2$ , it remains very unlikely that plant growth would be compromised by the patterns of exposure likely to occur in the Gove area.

#### 3.2.2 Ecological studies

A field survey was carried out at Gove in June 2003 as part of the Gove Refiner Expansion EIS. It revealed no evidence of visible injury to plant species near the present alumina refinery. This lack of injury indicates that, even in the areas predicted to be exposed to the highest 1-hour maximum and seasonal average concentrations (Table 1), there was no detectable effect of  $\text{SO}_2$  on the vegetation.

A survey of plant communities in the vicinity of Mt Isa, Queensland (Griffiths, 1998), is relevant to the present situation, except that the quantities and duration of emissions of SO<sub>2</sub> are much greater at Mt Isa than would be expected to occur at Gove. At Mt Isa, 50 years of very substantial SO<sub>2</sub> releases had resulted in changes in plant community composition and structure within 15 km of the major sources. This does not imply that an effect of similar dimension will occur at Gove, because of the marked differences in the quantities of SO<sub>2</sub> emitted at Gove and Mt Isa. However, there remains the possibility that operation of the alumina refinery for 50 years may result in some ecological changes in the immediate proximity of the works site. The nature of these changes is difficult to predict, but may be similar in principle to the effects reported for Mt Isa, including alteration of relative species abundance in plant communities and possibly growth rates of species. The effects of these changes in terms of the appearance of the vegetation in 50 years' time are even more difficult to predict, as vegetation appearance will be influenced by many factors other than ambient SO<sub>2</sub> concentration. However, it is considered that the contribution of SO<sub>2</sub> effects to changes in vegetation structure and condition at Gove are likely to be minor, if detectable at all.

At low concentrations in the environment, sulfur acts as an essential mineral element, and additions from the atmosphere may in some situations rectify soil deficiencies. There is usually a wide range of soil sulfur concentrations that are regarded as satisfactory, and the direct deposition of SO<sub>2</sub> on vegetation supplements this soil supply. Sulfur is incorporated rapidly into biological systems, and is recycled rather than accumulated in vegetation, so low rates of uptake from the air can occur without adverse effects appearing at a later time (Lauenroth and Preston, 1984). Consequently, small additions of sulfur from deposition of atmospheric SO<sub>2</sub> are not considered to be a material hazard to the Gove environment.

### **3.2 Future SO<sub>2</sub> emissions**

It is assumed that the majority of the SO<sub>2</sub> released into the Gove environment originates in the fuel used for steam generation. A proposal to replace bunker oil with natural gas as the fuel for the refinery processes and electricity generation would be expected to lead to a reduction in SO<sub>2</sub> emissions, even with an increase in fuel consumption. Such a reduction in SO<sub>2</sub> emissions would be expected to reduce the maximum and average ground level SO<sub>2</sub> concentrations, and reduce the possibility of adverse effects on vegetation.

### **3. Conclusions**

On the basis of recently measured ground level concentrations of SO<sub>2</sub> at the Gove refinery and associated predictions of 1-h maximum or seasonal average ground level concentrations in the surrounding area, it is concluded that injury is very unlikely to occur to plant species growing in the vicinity of the Gove alumina refinery.

No evidence of visible injury to vegetation has been obtained in recent surveys of the refinery site and surrounding area, although some of the plant species at Gove can be assumed to be sensitive to injury by SO<sub>2</sub>.

Long-term effects of SO<sub>2</sub> exposure are difficult to predict, but at low concentrations in the environment the sulfur is likely to act as a nutrient source rather than as a pollutant.

A change in fuel for the refinery is considered to be likely to reduce total SO<sub>2</sub> emissions, and to further reduce any likelihood of adverse effects on vegetation.

### **References**

Griffiths, AD (1998) 'Impact of sulphur dioxide emissions on savanna biodiversity at Mt Isa, Queensland' Supporting document to final report of Mount Isa Mines Limited Panel Assessment Study. CRC for Sustainable use of Tropical Savannas and CSIRO Division of Wildlife & Ecology, Winnellie, NT.

Lauenroth, WK, Preston, E.M. (1984) 'The Effects of SO<sub>2</sub> on a Grassland'. Ecological Studies 45. Springer Verlag, New York.

NEPC (1998). National Environment Protection Measure for Air. National Environment Protection Council, Canberra.

O'Connor, JA, Parberry, DG, Strauss, W (1974) The effects of phytotoxic gases on native Australian plant species: Part 1. Acute effects of sulphur dioxide. *Environmental Pollution* 7, 7-23.

PAE (2002) Summary Report: Work performed to date. Gove Refinery Expansion EIS. Air Quality. Pacific Air & Environment, Brisbane.

## **Appendix 1**

### **SULFUR DIOXIDE AND AUSTRALIAN VEGETATION**

D. Doley

Department of Botany, The University of Queensland, Qld 4072

This discussion represents part of a larger review prepared for submission to *Clean Air*. On publication, copyright to the material will be vested in the Clean Air Society of Australia and New Zealand, but remains with the author until that time.

#### **SUMMARY**

Ambient air quality standards are designed to protect identified organisms (e.g. humans) or they may be intended to protect the environment at large, including organisms that are very sensitive to the pollutant in question. Standards may be established using evidence of effects from other places, or they may be based upon research carried out specifically for that purpose. Comparatively little information is available on the responses of Australian plant species to experimental fumigations of sulfur dioxide, but those studies that have been conducted indicate that the National Environment Protection Measure for Air provides adequate protection.

#### **INTRODUCTION**

Historically, environmental standards were concerned first with human health, and subsequently with human well-being. As a result, there has been a dichotomy of regulation. Primary standards are designed to protect human health, and should be applicable throughout the world, assuming an absence of racial variation in human sensitivity to atmospheric pollutants. Secondary standards are designed to protect human well-being and the functioning of the large majority of organisms under the large majority of environmental conditions. Many of the species most sensitive to pollutants are plants. In the past, the plants considered to be most important to human society have been those that provide food, either directly or indirectly, and clothing or shelter. Now, human societies are now concerned with more general aspects of well-being, including the stability and diversity of the ecosystems in which they live (Tingey et al. 1990), and this raises to importance all organisms, whether or not they are of direct utility to humans. All these concerns require that ambient air quality is maintained at levels that permit a range of human activities to occur, and also protect values that may not be easy to assess in commercial terms. For the majority of air pollutants, including sulfur dioxide, environmental conditions that protect plants ensure the protection of other organisms, so that the well-being of plant species assumes a central role in environmental protection. Ultimately, a social decision must be made concerning the total level of impact humans should have on the environment. Air quality is one of many such impacts.

Air quality standards that are designed to protect some identified part of the environment are conveniently described as *receptor* standards. These standards require any activity to be conducted in such a way that the critical part of the receptor environment will not be affected adversely. Alternatively, standards may be applied to sources, as *emission* standards, which limit the amount of pollutant added to the environment. The contrast between the two types of standard is important. In receptor standards, the pollutant emitter is responsible for calculating the relationship between emission and receptor condition, whilst in emission standards the community accepts this responsibility through the regulating agency. The following discussion will focus on receptor standards, as these can be interpreted more directly in terms of plant response.

The simplest environmental situation, with no pollution, could not support technologically based, urban societies. Practical environmental standards accept that some species, and some members of any species are likely to be adversely affected by pollutants that are acceptable to a defined majority of humans.

#### **AUSTRALIAN STANDARDS**

Australia has applied a national air quality standard for SO<sub>2</sub> since 1981 (cf. Steer and Heiskanen 1993), which was refined in the National Environment Protection Measure for Air (Air NEPM) (NEPC 1998) (Table 1).

**Table 1**  
**Selected air quality standards for sulfur dioxide in Australia and New Zealand.**  
**Values are expressed as µg m<sup>-3</sup> at 0C and 101.3 kPa.**

Authority	Zone	Status	Averaging Time		
			1 Year	24 Hours	1 Hour
Victoria (1987)		detrimental		314	972
		acceptable		171	486
Tasmania (1974)			54	160	450
Western Australia (1992a)	industrial <sup>1</sup>	limit	80	365	1400
		standard	60	200	700
	buffer	limit	60	200	1000
		standard	50	150	500
	general	limit	60	200	700
		standard	50	125	350
Murray et al. (1991)		secondary	60	75 <sup>2</sup>	600
Queensland (1997)		goal	57	230	572
NEPC (1998)		goal	57	230	572
New Zealand (2002)	agricultural	guideline	30	120	350
	forest	guideline	20		
	lichen	guideline	10		

<sup>1</sup> Kwinana region; <sup>2</sup> recommended 4-h standard.

The Queensland Environmental Protection Policy (QEPP Air) (Queensland 1997) established goals for SO<sub>2</sub> concentrations that were based principally on human health effects, and a similar approach was adopted in the development of the National Environment Protection Measure for Air (Air NEPM) (NEPC 1998). The purpose of this measure was to provide uniformity of protection for all Australians, as the States, which have primary responsibility for air quality matters, had acted to control emissions in different ways.

For example, Western Australia (1992a) set regulations for the Kwinana industrial area near Perth, in which different ground level concentrations were permitted in three zone: the industrial area, the industrial buffer zone, and residential and general use areas (Table 1). This method of zoning and regulation is based on the recognition of *beneficial use*, namely "Any lawful human activity within the relevant portion of the environment which is conducive to the health, welfare, convenience, comfort or amenity of persons within the relevant portion of the environment is declared to be a beneficial use to be protected under this policy." (Western Australia 1992b). Beneficial uses may include industrial or agricultural activities, residential development, or nature conservation. Under this policy, for each specified area, the concentrations of atmospheric pollutants are regulated by *limits* "... which shall not be exceeded..." and *standards* "... which it is desirable not to exceed....".

Table 1 shows that, for short exposures, the limit values applied in Western Australia are double the standard values, but for one-year averaging times, the limits are one-quarter to one-third greater than the standard values. Also, the differences between industrial and general use zones decrease with increasing averaging time. The relationship implies that the effects of SO<sub>2</sub> are very time-dependent in short exposures (a change in exposure duration from 4 to 1 hour enabling the environment to tolerate an almost four-fold increase in SO<sub>2</sub> concentration. In contrast, between 4-hour and one-year averaging times, there is a three- to four-fold change in acceptable concentration.



In Victoria, the State Environmental Protection Policy (Victoria 1981) established *beneficial uses* (Clause 8(1)) which were to be protected, including: (a) life, health and well-being of humans; (b) life health and well-being of other forms of life, including animals and vegetation; (c) visibility; (d) useful life and aesthetic appearance of buildings, structures, properties and materials; and (e) aesthetic enjoyment and local amenity. These beneficial uses, "...except life, health and well-being of humans..." are to be protected if they are located outside an industrial buffer zone (Clause 8(2)). An acceptable level of a pollutant was defined as "...that concentration of an indicator at or below which all beneficial uses listed in Clause 8 are protected." A detrimental level of a pollutant was defined as "...that concentration of an indicator at or above which a substantial proportion of the exposed population may be adversely affected or significant changes are likely to be caused to some segments of the environment." Therefore, comparisons between Victoria and other jurisdictions should be made on the basis of secondary (environmental) rather than primary (human) criteria. It is noteworthy that the intention of the term *beneficial use* appears to be different in Victoria, where it does not explicitly mention lawful activities of human beings, from than in Western Australia, where such activities are mentioned specifically.

In New Zealand, an extensive review of air quality guidelines resulted in the adoption of short-term guidelines for the protection of human health, and annual guidelines for the protection of ecosystems (Ministry for Environment 2002). The long-term guidelines were adopted from those developed by the World Health Organization (2000) for the purpose of protection of ecosystems from chronic exposure to SO<sub>2</sub> under the humid, low temperature conditions that prevail in northern Europe, and may be replicated in the cool temperate climate of New Zealand.

## STUDIES ON AUSTRALIAN VEGETATION

Investigations into the effects of SO<sub>2</sub> on plant species have been used principally to determine whether the sensitivities Australian species are markedly different from those in other parts of the world. In view of the importance of secondary standards for SO<sub>2</sub>, the analysis presented to be here covers a surprisingly limited amount of work.

The first examinations of effects of SO<sub>2</sub> on Australian native plant species were conducted by O'Connor et al. (1974), using high concentrations and exposure times of about 3 hours. Whilst their results indicated orders of sensitivity to acute fumigation, they did not indicate the quantitative responses to more realistic, low concentration and long duration exposures. Their conclusions were that many of the native tree and shrub species tested, and particularly eucalypts, were sensitive to SO<sub>2</sub>, but less sensitive to ozone.

Field studies of the unintended effects of SO<sub>2</sub> on *Pinus radiata* were reported by Turner and Lambert (1980). On sulfur-deficient soils, SO<sub>2</sub> presumed to have originated from coal-fired power stations had resulted in significantly increased stem volume growth in trees. There were no estimates of ambient SO<sub>2</sub> concentrations available for this study. Perhaps surprisingly, there has not been a report to the converse, namely that SO<sub>2</sub> emissions have resulted in significantly depressed tree growth in the vicinity of Australian power stations.

A landmark series of fumigations, initiated by Murray (1984a) and summarised by Murray et al. (1991) has been conducted, largely in Western Australia, using open-top chambers at ambient temperatures and humidities, with the addition of regulated concentrations of SO<sub>2</sub> ranging from about 150 to 1000 µg m<sup>-3</sup>, generally for four hours in the middle of each day for several days per week. The majority of fumigations were applied for more than 100 days, and the responses of plants were assessed at the end of each experimental period. Attributes recorded included plant height and dry weight, and for cereal species, seed weight, seed number, ear number, and ear weight. The data were tested by analyses of variance in order to determine whether significant differences in attributes were associated with the fumigation treatments.

From these analyses, it is possible to identify concentrations of SO<sub>2</sub> in each experiment and for each species at which significant reductions in attributes below the control values were recorded. These summary data are reproduced in Table 2 for pasture legume species, Table 3 for cereal species, Table 4 for crop species, and Table 5 for tree species. The summaries consider only the direct effects of SO<sub>2</sub>, and not the interactions with either HF or NO<sub>2</sub>, which were examined in several experiments.

Analyses of variance showed that statistically significant reductions in plant attributes occurred in branch length of *Trifolium subterraneum* cv. Trikkala (subterranean clover) at 157 µg m<sup>-3</sup>, and in shoot dry weight and branch length in *Medicago polymorpha* cv. Santiago (burr medic) at 228 µg m<sup>-3</sup> (Table 2). These represented the most sensitive responses of all species studied. In other pasture species, dry weight reductions were observed at SO<sub>2</sub> concentrations above 700 µg m<sup>-3</sup>.

**Table 2**  
**The lowest concentrations of SO<sub>2</sub> causing significant reductions in growth related attributes of pasture species.**

Species	Ref	Exposure days	SO <sub>2</sub> concentration (µg m <sup>-3</sup> ) significantly reducing attribute			
			Shoot dry wt	Root dry wt	Branch length	Leaf area
<i>Lolium perenne</i> cv. Tetralite (perennial ryegrass)	1	48	>164	>164	na*	na
<i>Medicago sativa</i> cv. Siriver (lucerne)	4	108	750	na	750	na
<i>M. sativa</i> cv CUF101 (lucerne)	2	301	>215	>215	na	>215
<i>M. truncatula</i> cv Paraggio (barrel medic)	5	108	425	na	750	na
<i>M. polymorpha</i> cv. Santiago (burr medic)	4	149	<b>228</b>	na	228	228
<i>Trifolium repens</i> cv. Haifa (white clover)		149	730	na	730	na
<i>T. repens</i> cv Regal (white clover)	3	63	>215	>215	na	>215
<i>T. subterraneum</i> cv Woogenellup (subterranean clover)	1	48	>164	>164	na	na
<i>T. subterraneum</i> cv. Trikkala (subterranean clover)	4	108	750	na	750	na

\*na indicates no data available.

References: 1, Murray 1984b; 2, Murray 1985a; 3, Murray 1985b; 4, Murray et al. 1991; 5, Murray and Wilson 1991.

Amongst cereals, shoot dry weight was reduced significantly in *Triticum aestivum* cv Banks at 425 µg m<sup>-3</sup>, and in *X. triticosecale* cv. Currency at 228 µg m<sup>-3</sup> (Table 3). Reductions in growth-related attributes were uncommon in crop and pasture species at SO<sub>2</sub> concentrations about 150 µg m<sup>-3</sup>, and in the tree species tested, these effects were limited at 230 µg m<sup>-3</sup>.

**Table 3**  
**The lowest concentrations of SO<sub>2</sub> causing significant reductions in growth-related attributes of cereals.**

Species	Ref	Exposure days	SO <sub>2</sub> Concentration (µg m <sup>-3</sup> ) significantly reducing attribute		
			Total dry wt	Shoot dry wt	Total grain wt
<i>Avena sativa</i> cv Echidna (oats)	1	182	1015	730	1015
<i>Hordeum vulgare</i> cv. Clipper (barley)	2	90	na*	>267	>267
<i>H. vulgare</i> cv. Schooner (barley)	1	90	750	750	750
<i>Triticum aestivum</i> cv. Banks (wheat)	1	108	750	750	750
<i>T. aestivum</i> cv. Halberd (wheat)	2	90	na	<b>267</b>	>267
<i>X. triticosecale</i> cv. Currency (triticale)	1	182	1100	640	1100

\*na indicates no data available.

References: 1, Murray et al 1991; 2, Murray and Wilson 1988d.

Murray et al. (1991) concluded: "Clearly the concentration-response relationship is different for each species, especially *Pinus radiata* which was resistant to SO<sub>2</sub> exposure over the concentration range considered. Wheat and triticale showed a similar response to SO<sub>2</sub> with an initial stimulation in yield below about 0.2 ppm of SO<sub>2</sub> [572 µg m<sup>-3</sup>], whereas barley and oats showed a no-threshold response. Burr medic, barrel medic and white clover also showed no-threshold responses while sub- clover and lucerne were little affected until SO<sub>2</sub> concentrations reached 0.10 and 0.20 ppm [286 and 572 µg m<sup>-3</sup>] respectively. *Eucalyptus pilularis* showed a threshold around 0.15 ppm [430 µg m<sup>-3</sup>] but *E. microcorys* and *E. regnans* showed a steady decline in growth across the entire SO<sub>2</sub> concentration range considered."

Of the species that showed a "no-threshold response", the reduction in plant weight was statistically significant at 230 µg m<sup>-3</sup> for burr medic and white clover (Table 2) and triticale (Table 3), although in the latter case not at 400 µg m<sup>-3</sup>. Other species were recognized as having response thresholds well above 150 µg m<sup>-3</sup>. A very conservative interpretation of the analyses of variance would be that, for the species examined, the threshold for significant reductions in plant growth was in the vicinity of 150 µg m<sup>-3</sup>.

**Table 4**

**The lowest concentrations of SO<sub>2</sub> causing significant reductions in growth-related attributes of crop species.**

Species	Ref	Exposure days	SO <sub>2</sub> Concentration (µg m <sup>-3</sup> ) causing significant reduction in attribute		
			Total dry weight	Pod/cob number	Seed weight
<i>Arachis hypogea</i> cv. Virginia Bunch (peanut)	1	70	136		>282
<i>Cucumis sativus</i> cv. Green Gem Cucumber	2	21	250	na	na
<i>Glycine max</i> cv. Dragon (soybean)	1	91	>279	>279	>279
<i>Phaseolus vulgaris</i> cv. Gallaroy (navy bean)	1	51	<b>141</b>	>290	290
<i>Zea mays</i> cv. QK958 (maize)	1	70	na*	>282	na

\* na indicates no data available.

References: 1, Murray and Wilson 1990; 2, Dodd & Doley (1998)

**Table 5**

**The lowest concentrations of SO<sub>2</sub> causing significant reductions in selected attributes of Australian tree species under experimental conditions.**

Species	Ref	Exposure days	SO <sub>2</sub> concentration (µg m <sup>-3</sup> ) significantly reducing attribute				
			Total dry wt	Leaf dry wt	Stem dry wt	Stem diam	Height growth
<i>Araucaria cunninghamii</i> (hoop pine)	5	115	na*	na	na	>477	477
<i>Corymbia calophylla</i> (marri)	4	120	na	271	>271	na	na
<i>Eucalyptus crebra</i> (ironbark)	1	40	>217	>217	na	na	>217
<i>E. gomphocephala</i> (tuart)	4,6	126	>303	>303	>303	>303	>303
<i>E. marginata</i> (jarrah)	4	120	na	<b>122</b>	>271	na	na
<i>E. microcorys</i> (tallow-wood)	7	107	500	500	>950	350	>950
<i>E. moluccana</i> (gum-top box)	1	40	>217	>217	na	na	>217
<i>E. pilularis</i> (blackbutt)	7	107	500	500	500	500	>950

<i>E. regnans</i> (mountain ash)	7	107	500	>950	500	500	500
<i>E. rudis</i> (flooded gum)	3	119	na	>274	na	>274	>274
<i>E. tereticornis</i> (forest red gum)	2	90	>267	na	na	na	na
<i>Pinus radiata</i> (radiata pine)	7	107	>950	>950	950	>950	>950

\* na indicates no data available.

References: 1, Murray 1984a; 2, Murray and Wilson 1988c; 3, Clarke and Murray 1990; 4, Murray and Wilson 1989b; 5, Murray et al. 1990; 6, Fulford and Murray 1990; 7, Murray et al. 1991.

An alternative analysis of the data was provided by Murray et al. (1991), based upon linear regressions of relative growth or yield on SO<sub>2</sub> concentration, or (SO<sub>2</sub>-concentration) x (hours of exposure). The results of these analyses were very similar, suggesting that the effects of total exposure duration were limited. The concentration of SO<sub>2</sub> was concluded to be the best term for prediction of growth and yield responses. Daily duration of exposure, frequency of exposure, as well as total length of exposure appeared to be far less important in determining the nature of the dose-response relationship. Australian crop, forestry and native plant species were considered to all show similar overall dose-response curves for frequent exposure to SO<sub>2</sub>, leading Murray et al. (1991) to establish a single dose-response function for all Australian plant species. They selected a straight-line relationship of the form

$$y = 4.81 - 0.19x \quad (9)$$

where y is the percentage change in yield or growth and x is the SO<sub>2</sub> concentration in nL L<sup>-1</sup>. The correlation coefficient, r, was 0.79, so the regression explained 62% of the variation in yield.

As indicated by Doley and McCune (1993), the use of a linear regression implies that there is a constant relationship between SO<sub>2</sub> concentration and the responses of all species, that is, there is no threshold concentration for the appearance of an effect, and all species respond in the same manner. Also, a regression analysis based on a central statistic of a sample submerges the differences in species responses that were given prominence in the earlier discussion of their results by Murray et al. (1991). If it is desired to establish the air quality criteria on the response of the most sensitive species for which information is available, then it is inappropriate to amalgamate data for all species in a single regression that weights equally the data for sensitive and tolerant species. In addition, the qualifications regarding detoxification and loss of SO<sub>2</sub>, and the influence of irregular exposures are not incorporated in this prediction.

Where different growth parameters are analysed in the same regression, for example, shoot dry weight, total plant dry weight, and grain weight, it is assumed that the effects of SO<sub>2</sub> on these parameters, i.e. on the production and distribution of dry weight between different plant parts, are identical, and that the variation observed in the results is due to random variation within the sample. This assumption is generally acceptable, but because of different patterns of functioning between species, it is not entirely sustained by the data presented in the literature. Again, if the most sensitive response is to be used, it is inappropriate to combine data for all responses and to apply a central statistic and a response function for one of the most sensitive species should be used.

Murray et al. (1991) concluded that the effects of SO<sub>2</sub> were independent of exposure duration, between 4, 8 and 24 h d<sup>-1</sup>, or between 2.5 h d<sup>-1</sup>, 3 days per week and daily exposure. However, in developing their standard, they did incorporate a time function: "These three *Eucalyptus* species were also sensitive to SO<sub>2</sub> exposure showing significant growth depression once the SO<sub>2</sub> concentration reached 175 nL L<sup>-1</sup> [500 µg m<sup>-3</sup>, 4 h d<sup>-1</sup>]. As a 24-h average this is only 29 nL L<sup>-1</sup> [83 µg m<sup>-3</sup>]. This is equivalent to the lowest SO<sub>2</sub> concentration known to affect northern hemisphere tree species.." Also, "*E. microcorys* showed a trend towards growth depression at 50 nL L<sup>-1</sup> [143 µg m<sup>-3</sup>] for a 4-h/day, equivalent to a 24 hour average of only 8 nL L<sup>-1</sup> [23 µg m<sup>-3</sup>] daily."

If the only important characteristic of the SO<sub>2</sub> fumigation is the concentration during the period of exposure (the middle of the day), and the duration of exposure has no influence, then there should not be any difference between the SO<sub>2</sub> standards for different averaging times, provided all these averaging times include the middle of the day. If different averaging times should be associated with different ambient concentrations, evidence obtained from fumigations of 4 h d<sup>-1</sup> should not be applied to the determination of standards for continuous exposure, as reflected in a 24-hour average, unless sufficient direct evidence of the identity of effects has been obtained. The fact that statistically significant differences between continuous and intermittent fumigations have not been demonstrated is not sufficient evidence of identity of effects, as the experiments reviewed by Murray et al. (1991) were not designed in a manner that could adequately test the effect of exposure duration or the timing of exposure during the day on plant response to SO<sub>2</sub>.

An appropriate conclusion from the evidence of Murray et al. (1991) is that the SO<sub>2</sub> exposure regime causing significant leaf dry weight reduction in jarrah (*Eucalyptus marginata*) was associated with 122 µg m<sup>-3</sup>, applied for 4 h d<sup>-1</sup> during bright sunlight and 5 d w<sup>-1</sup> over a period of 120 days, and the highest experimental concentration not resulting in significant shoot dry weight reduction was 271 µg m<sup>-3</sup>. The value of 122 µg m<sup>-3</sup> is approximately half of the 24-h NEPC (1998) and Queensland goal value of 230 µg m<sup>-3</sup> and similar to the 24-h guideline value of 120 µg m<sup>-3</sup> adopted in New Zealand (2002).

## CONCLUSION

Experimental studies on Australian plant species have not revealed any pasture, crop or tree species that will be affected adversely by exposure to sulfur dioxide under regimes that comply with the goals established through the National Environment Protection Measure (1998). In view of the margins between the lowest SO<sub>2</sub> concentrations resulting in significant effects in the experimental studies and the ambient goal values, it is concluded that Australian plant species are not more sensitive to SO<sub>2</sub> than are species from the temperate Northern Hemisphere.

## REFERENCES

- Barker, I. and Barker, J. (eds) (1988). 'Clean Air Around the World: The Law and Practice of Air Pollution Control in 14 Countries in 5 Continents'. International Union of Air Pollution Prevention Associations. Brighton.
- Bennett, J.P. (1985). Regulatory uses of SO<sub>2</sub> effects data. In Winner, W.E., Mooney, H.A. and Goldstein, R.A. (eds) 'Sulfur Dioxide and Vegetation'. pp 23-36. Stanford University Press. Stanford.
- Canada (1989). Department of the Environment, Annexation to the Environmental Protection Act, Canada 1989. Canada Gazette Part 1, pp 3642-3645.
- Clarke, K. and Murray, F. (1990). Stimulatory effects of SO<sub>2</sub> on growth of *Eucalyptus rudis*. *New Phytologist*. 115, 633-637.
- Cochran, L.S., Peilke, R.A. and Kovács, E. (1992). Selected international receptor-based air quality standards. *Journal of the Air and Waste Management Association* 42, 1567-1572.
- Davieson, G., Murray, F. and Wilson, S. (1990). Effects of sulphur dioxide and hydrogen fluoride, singly and in combination, on growth and yield of wheat in open-top chambers. *Agricultural Ecosystems and Environment* 30, 317-325.
- Dodd, I.C. and Doley, D. (1998). Growth responses of cucumber seedlings to sulphur dioxide fumigation in a tropical environment. *Environmental and Experimental Botany* 39, 41-47.

Doley, D. and McCune, D.C. (1993). Ambient air quality standards for sulfur dioxide in Australia: I. Criteria and analysis. *Clean Air* 27, 122-132.

Environment Protection Authority of Victoria. (1994). 'State Environment Protection Policy (the Air Environment) Draft Recommendations on Class 1 Indicators'. Government of Victoria. pp 53.

Fulford, G. and Murray, F. (1990). Morphogenetic changes in *Eucalyptus gomphocephala* exposed to SO<sub>2</sub>. *Environmental and Experimental Botany*. 30, 343-347.

Heck, W.W., Taylor, O.C. and Tingey, D.T. (eds) (1988). 'Assessment of Crop Loss from Air Pollutants'. Elsevier Science Publishing, Barking.

Hogsett, W.E., Olszyk, D., Ormrod, D.P., Taylor, G.E. and Tingey, D.T. (1987a). 'Air Pollution Exposure Systems and Experimental Protocols: Volume 1: A Review and Evaluation of Performance' U.S. Environmental Protection Agency, Publication 600/3-87/037a, Corvallis, Oregon.

Hogsett, W.E., Olszyk, D., Ormrod, D.P., Taylor, G.E. and Tingey, D.T. (1987b). 'Air Pollution Exposure Systems and Experimental Protocols: Volume 2: Description of Facilities' U.S. Environmental Protection Agency, Publication 600/3-87/037b, Corvallis, Oregon.

Ministry for Environment (2002). Ambient Air Quality Guidelines. Air Quality Report No. 32. New Zealand Ministry for Environment, Wellington.

Murray, F. (1984a). Effects of sulfur dioxide on three *Eucalyptus* species. *Australian Journal of Botany*. 32, 139-145.

Murray, F. (1984b). Responses of subterranean clover and ryegrass to sulphur dioxide under field conditions. *Environmental Pollution (Series A)* 36, 239-249.

Murray, F. (1985a). Changes in growth and quality characteristics of lucerne (*Medicago sativa* L.) in response to sulphur dioxide exposure under field conditions. *Journal of Experimental Botany*. 36, 449-457.

Murray, F. (1985b). Some responses of ladino clover (*Trifolium repens* L. cv. Regal) to low concentrations of sulphur dioxide. *New Phytologist*. 100, 57-62.

Murray, F., Clarke, K and Wilson, S (1990). The response of *Araucaria cunninghamii* (hoop pine) to intermittent exposures of sulfur dioxide and nitrogen oxides. Report to Queensland Electricity Commission. Murdoch University. pp 26.

Murray, F., Clarke, K., Wilson, S. and Monk, R. (1991). 'Development of Australian Secondary Ambient Air Quality Criteria for Sulfur Dioxide and Nitrogen Oxides'. Report to the Energy Research and Development Corporation. Murdoch University. pp 169.

Murray, F. and Wilson, S. (1988a). 'Effects of SO<sub>2</sub>, HF and their Combination on Plants of Agricultural or Ecological Importance in Australia'. Final Report to the National Energy Research Development and Demonstration Council. Murdoch University, Perth.

Murray, F. and Wilson, S. (1988b). Effects of sulphur dioxide, hydrogen fluoride and their combination on three *Eucalyptus* species. *Environmental Pollution*. 52, 265-279.

Murray, F. and Wilson, S. (1988c). Joint action of sulfur dioxide and hydrogen fluoride on growth of *Eucalyptus tereticornis*. *Environmental and Experimental Botany*. 28, 343-349.

Murray, F. and Wilson, S. (1988d). The joint action of sulphur dioxide and hydrogen fluoride on quality of wheat and barley. *Environmental Pollution*. 55, 239-249.

Murray, F. and Wilson, S. (1989a). The relationship between sulphur dioxide concentration and yield of five crops in Australia. *Clean Air* 23, 51-58.

Murray, F. and Wilson, S. (1989b). Sulfur dioxide-induced growth changes in *Eucalyptus calophylla*. *European Journal of Forest Pathology*. 19, 193-199.

Murray, F. and Wilson, S. (1990). Yield responses of soybean, maize, peanut and navy bean exposed to SO<sub>2</sub>, HF and their combination. *Environmental and Experimental Botany*. 30, 215-223.

Murray, F. and Wilson, S. (1991). The effects of SO<sub>2</sub> on the final growth of *Medicago truncatula*. *Environmental and Experimental Botany*. 31, 319-325.

NAS (1978). 'Sulfur Oxides'. National Research Council, National Academy of Sciences, Washington DC.

NEPC (1998). National Environment Protection Measure for Air. National Environment Protection Council, Canberra.

O'Connor, J.A., Parberry, D.G. and Strauss, W. (1974). The effects of phytotoxic gases on native Australian plant species: Part 1. Acute effects of sulphur dioxide. *Environmental Pollution*. 7, 7-23.

Queensland Environmental Protection Agency (1997) Queensland Environmental Protection Policy (Air).

Rawlings, J.O. and Cure, W. (1985). The Weibull function as a dose-response model to describe ozone effects on crop yields. *Crop Science*. 25, 807-814.

Steer, K. and Heiskanen, L. (1993). 'Options for Australian Air Quality Goals for Oxides of Sulphur: Draft for Technical Review'. Commonwealth Department of Health, Housing and Community Services, Canberra. pp 166

Streeton, J.A. (1990). 'Air Pollution Health Effects and Air Quality Objectives in Victoria'. Report to Environment Protection Authority of Victoria. pp 97.

Tingey, D.T., Hogsett, W.E. and Henderson, S. (1990). Definition of adverse effects for the purpose of establishing secondary national ambient air quality standards. *Journal of Environmental Quality* 19, 635-639.

Turner, J. and Lambert, M.J. (1980). Sulphur nutrition of forests. In 'Atmospheric Sulphur Deposition - Environmental Impact and Health Effects' ed by D.S. Shriner, C.R. Richmond and S.E. Lindberg. pp 321- 333. Ann Arbor Science Publ.

USEPA (1987). Environmental Protection Agency Regulations on National Primary and Secondary Ambient Air Quality Standards. Environment Reporter

USEPA (1988). Proposed decision not to revise National Ambient Air Quality Standards for sulfur oxides (sulfur dioxide): proposed rule. Federal Register 53(8), 14926-14951.

Victoria (1981). State environmental protection policy (the air environment). Victoria Government Gazette 63, 2293-2305.

Victoria (1987). State environmental protection policy (the air environment), amendments. Victoria Government Gazette 120 3895-3902.

Western Australia (1992a). 'Environmental Protection (Kwinana) (Atmospheric Wastes) Regulations 1992'. Government Printer, Perth.

Western Australia (1992b). 'Environmental Protection (Kwinana) (Atmospheric Wastes) Policy 1992'. Government Printer, Perth.

World Health Organization (2000). Air Quality Guidelines for Europe 2<sup>nd</sup> Edition. WHO Regional Publications, European Series, No. 91. WHO, Geneva.





The appendix provides information on the fauna sampling methodology and further information on each of the sampling sites.

## **Sampling Methodology**

### ***Live Capture/Release Trapping***

Small mammals and reptiles were surveyed using live trapping methods, conducted under Parks and Wildlife Commission of the Northern Territory Permit Number 15641 and Animal Ethics Clearance No. A01013.

Live capture/release methods included aluminium type A Elliott box traps for small mammals, and pitfall traps with drift fences. Elliott traps were placed at 10 m intervals along transects at survey sites (20 Elliott traps per transect). Traps were baited with a standard bait mixture and were operated over a two night period at each trap site.

Pitfall trap systems incorporated PVC buckets approximately 40 cm deep with a screen drift fence (5 m long x 0.4 m high). Pitfall trapping was undertaken at 4 sites (2 traps per site) and traps were operated over 4 nights (32 pitfall trap nights altogether). Pitfall traps were cleared of captures in the morning and late afternoon.

### ***Bird Census***

Diurnal birds were sampled using a point census method and broad observational surveys. Birds were systematically sampled at each of the four main sample sites. Censuses were carried out in the early morning (in the first 3-4 hours after sunrise) and late afternoon (last 3-4 hours before sunset) due to variation in avian activity during the day. Ten census points were located at least 100 m apart along transects at each site. During each census, birds seen and/or heard were counted and their distance from the observer noted over a ten minute period. An effort was made to avoid counting the same bird twice. Species flying overhead were not included in census data but were noted for analysis of species presence.

Additional bird species records were compiled incidentally, during spotlight survey and from vocalisations. Nocturnal species were recorded during spotlight surveys and identified from characteristic calls.

### ***Microchiropteran Bat Call Detection & Trapping***

Microchiropteran bat echolocation calls were recorded using an ultrasonic bat call detector (Anabat II; Titley Electronics). Anabat detection was conducted from fixed points during spotlight survey. Echolocation call detection was undertaken at all survey sites for a period of 30 minutes in the period between dusk and 9:00 pm, and at other sites opportunistically, especially in areas of observed bat activity, such as near water bodies. Mist netting for small bat species (microchiropteran or small megachiropteran taxa) was undertaken at two sites.

### ***Active Searches***

Active diurnal searching for reptiles, amphibians and small mammals included scanning of trees and ground, removal of cover such as rocks and fallen logs, and peeling the bark from trees. Searches also focussed on locating and identifying tracks and traces. Large mammals were recorded when encountered during trapping, bird surveys and spotlight surveys, and along roads and tracks throughout the study area. Observations made of wildlife recorded outside of the main sampling sites were noted according to the habitat in which they were observed.

### Spotlight Survey

Spotlighting on foot using low-watt hand-held torches was undertaken at all sample sites and in other areas of representative habitat. Spotlighting from a slow moving vehicle was undertaken along the main tracks of the study area.

### Taxonomy and Nomenclature

Scientific nomenclature and common names used in this report generally follow Stanger et al., (1998) with the exception of recently published taxonomic revisions. Additional texts utilised for species identifications and ecological data include Cogger (2000) for reptiles, Slater et al., (1989) for birds, and Menkhorst & Knight (2001) for mammals.

### Survey Site Details

Site Number	F1	F2	F3	F4
<b>Date</b>	28/05/2003	29/05/2003	30/05/2003	31/05/2003
<b>Locality</b>	Adjacent to Yacht Club	Headland near Burnt Lime Processor	Freshwater Swamp, RDA area	RDA area
<b>Map Reference (UTM/Lats-Longs)</b>	83 685371E 86 51606N	53 683146E 86 52248N	53 689633E 86 50133N	53 688800E 86 49498N
<b>Habitat type of landscape</b>	Coastal Monsoon Vine Thicket	Mixed Woodland	Freshwater Swamp	Open Forest
<b>Vegetation Classification</b>	Low (4-5 m height) Closed Forest	Mid-height mixed woodland to 5 m growing on beach dunes	Mid-high (10-15 m) Melaleuca open forest	Tall (12 m+) Eucalyptus Open Forest
<b>Dominant plants</b>	<i>Sterculia quadrifolia</i> , <i>Strychnos lucida</i> , <i>Canarium australinum</i>	<i>Pandanus spiralis</i> ; <i>Casuarina cunninghamii</i> ; <i>Acacia</i> spp.	<i>Melaleuca dealbata</i> ; <i>Melaleuca cajuputi</i> ; <i>Pandanus spiralis</i>	<i>Eucalyptus tetradonta</i> ; <i>Acacia</i> spp; <i>Livistona humilis</i>
<b>Adjacent habitats</b>	Mangroves, Beach, Open Forest	Beach	Open Forest; Mangroves	None
<b>Topography</b>	Flat	Undulating	Flat	Flat
<b>Rocks</b>	<2%	<2%	<2%	<2
<b>Soil</b>	Sand	Sand	Sand	Lateritic gravel
<b>Ground Cover</b>	Leaf Litter 90%; Bare Ground 10%	Leaf Litter 90%; Bare Ground 10%	Leaf Litter 40%; grass 50%	Leaf litter 20%; Bare Ground 20%; Grass 40%
<b>Refuges (presence of absence)</b>	None	Tree hollows	Tree hollows	Tree hollows
<b>Distance to nearest water (permanent/ephemeral)</b>	Beach – 20 m	Beach – 10 m	Mangroves – 100 m	Beach – 300 m
<b>Last burnt</b>	>3 years	>3 years	1-3 years	>3 years
<b>Site Disturbance</b>	Evidence of feral buffaloes; camps; baech flotsam and other litter	Site is heavily disturbed by noise from the nearby plant operations. Also, all vegetation covered with alumina dust and tope 10 cm of soil impregnated with alumina dust	Buffaloes; area close to RDA ponds	Tracks used by local Aboriginals; Beach access, hunting and bark removal (for bark paintings)



Common Name	Scientific Name	Sample Sites				Other Observations		Mangroves/Beach
		F1 Vine Thicket	F2 Mixed Woodland	F3 Freshwater Swamp	F4 Open Forest	Sewage Ponds	Open Forest/ RDA area	
<b>Amphibians</b>								
Giant Frog	<i>Cyclorana australis</i>					x		
Marbled Frog	<i>Limnodynastes convexiusculus</i>			P				
Bicolor Tree Frog	<i>Litoria bicolor</i>						x	
Green Tree Frog	<i>Litoria caerulea</i>					x	x	
Rocket Frog	<i>Litoria nasuta</i>						x	
Roth's Tree Frog	<i>Litoria rothi</i>					x	x	
Desert Tree Frog	<i>Litoria rubella</i>						x	
<b>Reptiles</b>								
Estuarine Crocodile	<i>Crocodylus porosus</i>					x		x
Binoe's Gecko	<i>Heteronotia binoei</i>				P			
Asian House Gecko	<i>Hemidactylus frenatus</i>						x	
Callose-palmed Shinning-skink	<i>Cryptoblepharus plagiocephalus</i>	x	x				x	
Closed-litter Rainbow-skink	<i>Carlia longipes</i>	x						
Shaded-litter Rainbow-skink	<i>Carlia munda</i>	x	x	x	x		x	
Lowlands Plainbacked Ctenotus	<i>Ctenotus essingtonii</i>	x						
Plain Ctenotus	<i>Ctenotus inornatus</i>						x	
Northern Sand Monitor	<i>Varanus panoptes</i>						x	
Friiled Lizard	<i>Chlamydosaurus kingii</i>						x	
Two-lined Dragon	<i>Diporiphora cf. bilineata</i>	x						
Olive Python	<i>Liasis olivaceus</i>						x	
Western Brown Snake	<i>Pseudonaja nuchalis</i>			x				
<b>Birds</b>								
Orange-footed Scrubfowl	<i>Megapodius reinwardt</i>	x						
Radjah Shelduck	<i>Tadorna radjah</i>					x	x	x
Eastern Reef Egret	<i>Egretta sacra</i>							x

Common Name	Scientific Name	Sample Sites				Other Observations		Mangroves/Beach
		F1 Vine Thicket	F2 Mixed Woodland	F3 Freshwater Swamp	F4 Open Forest	Sewage Ponds	Open Forest/ RDA area	
Great Egret	<i>Ardea alba</i>							x
Intermediate Egret	<i>Ardea intermedia</i>			x			x	x
Australian White Ibis	<i>Threskiornis molucca</i>						x	x
Black-necked Stork	<i>Ephippiorhynchus asiaticus</i>			x			x	
Osprey	<i>Pandion haliaetus</i>				1		x	
Black Kite	<i>Milvus migrans</i>						x	
Whistling Kite	<i>Haliastur sphenurus</i>						x	x
Brahminy Kite	<i>Haliastur indus</i>					x		
White-bellied Sea-eagle	<i>Haliaeetus leucogaster</i>							x
Brown Goshawk	<i>Accipiter fasciatus</i>		2					
Brown Falcon	<i>Falco berigora</i>							x
Australian Hobby	<i>Falco longipennis</i>						x	
Terek Sandpiper	<i>Xenus cinereus</i>							x
Beach Stone-curlew	<i>Esacus neglectus</i>							x
Masked Lapwing	<i>Vanellus miles</i>					x		x
Silver Gull	<i>Larus novaehollandiae</i>							x
Crested Tern	<i>Sterna bergii</i>							x
Peaceful Dove	<i>Geopelia striata</i>	1		7	5	x	x	x
Bar-shouldered Dove	<i>Geopelia humeralis</i>	1	1	3	2		x	x
Red-tailed Black-Cockatoo	<i>Calyptorhynchus banksii</i>			1				
Little Corella	<i>Cacatua sanguinea</i>				1		x	x
Sulphur-crested Cockatoo	<i>Cacatua galerita</i>	1		2	6		x	
Red-collared Lorikeet	<i>Trichoglossus rubritorquis</i>			4	7	x	x	x
Varied Lorikeet	<i>Psitteuteles versicolor</i>			1	1			
Red-winged Parrot	<i>Aprosmictus erythropterus</i>			4	2		x	
Southern Boobook Owl	<i>Ninox novaeseelandiae</i>					x		
Tawny Frogmouth	<i>Podargus strigoides</i>						x	
Blue-winged Kookaburra	<i>Dacelo leachii</i>			x			x	
Forest Kingfisher	<i>Todiramphus macleayii</i>	1		1	1	x		

Common Name	Scientific Name	Sample Sites				Other Observations		Mangroves/Beach
		F1 Vine Thicket	F2 Mixed Woodland	F3 Freshwater Swamp	F4 Open Forest	Sewage Ponds	Open Forest/ RDA area	
Sacred Kingfisher	<i>Todiramphus sanctus</i>							x
Collared Kingfisher	<i>Todiramphus chloris</i>							x
Rainbow Bee-eater	<i>Merops ornatus</i>		2	3	1	x	x	x
Red-backed Fairy-wren	<i>Malurus melanocephalus</i>				1			
Mangrove Gerygone	<i>Gerygone levigaster</i>	1						x
Helmeted Friarbird	<i>Philemon buceroides</i>	1					x	x
Silver-crowned Friarbird	<i>Philemon argenticeps</i>			4			x	
Little Friarbird	<i>Philemon citreogularis</i>	1						
Blue-faced Honeyeater	<i>Entomyzon cyanotis</i>						x	
Yellow-throated Miner	<i>Manorina flavigula</i>				1			
White-gaped Honeyeater	<i>Lichenostomus unicolor</i>					x	x	
White-throated Honeyeater	<i>Melithreptus albogularis</i>			7	2			
Brown Honeyeater	<i>Lichmera indistincta</i>	1			3	x	x	x
Bar-breasted Honeyeater	<i>Ramsayornis fasciatus</i>			1				
Dusky Honeyeater	<i>Myzomela obscura</i>			8			x	x
Red-headed Honeyeater	<i>Myzomela erythrocephala</i>			2				x
Jacky Winter	<i>Microeca fascinans</i>					x		
Australian Magpie-lark	<i>Grallina cyanoleuca</i>						x	x
Northern Fantail	<i>Rhipidura rufiventris</i>			1				
Spangled Drongo	<i>Dicrurus bracteatus</i>			1				
Black-faced Cuckoo-shrike	<i>Coracina novaehollandiae</i>		1					
Varied Triller	<i>Lalage leucomela</i>		1					
Yellow Oriole	<i>Oriolus flavocinctus</i>	1						
Olive-backed Oriole	<i>Oriolus sagittatus</i>	1						
Black-faced Woodswallow	<i>Artamus cinereus</i>						x	
Grey Butcherbird	<i>Cracticus torquatus</i>				2			
Pied Butcherbird	<i>Cracticus nigrogularis</i>						x	
Torresian Crow	<i>Corvus orru</i>		2	6	9		x	
Great Bowerbird	<i>Chlamydera nuchalis</i>	5					x	

Common Name	Scientific Name	Sample Sites				Other Observations		Mangroves/Beach
		F1 Vine Thicket	F2 Mixed Woodland	F3 Freshwater Swamp	F4 Open Forest	Sewage Ponds	Open Forest/ RDA area	
Double-barred Finch	<i>Taeniopygia bichenovii</i>						x	
Tree Martin	<i>Hirundo nigricans</i>						x	
<b>Mammals</b>								
Northern Brown Bandicoot	<i>Isoodon macrourus</i>	T						
Agile Wallaby	<i>Macropus agilis</i>			T			x	
Black Flying-fox	<i>Pteropus alecto</i>					x	x	x
Little Red Flying-fox	<i>Pteropus scapulatus</i>					x	x	
Northern Blossom Bat	<i>Macroglossus minimus</i>						M	
Long-eared Bat	<i>Nyctophyllus sp.</i>			A		A		
Unidentified Bat	<i>Scotorepens/Chalinolobus sp</i>					A		
Northern Pipistrelle	<i>Pipistrellus westralis</i>					A		
Yellow-bellied Sheath-tail-bat	<i>Saccolaimus flaviventris</i>			A				
Grasslands Melomys	<i>Melomys burtoni</i>			E				
Water Buffalo	<i>Bubalus bubalis</i>	T	T	x		x	T	
Feral cat	<i>Felis catus</i>						x	
Dingo	<i>Canis lupus dingo</i>	x	T				x	x





Sample Site		F1	F1	F2	F2	F3	F3	F4	F4
Trap Type		Elliots	Pitfalls	Elliots	Pitfalls	Elliots	Pitfalls	Elliots	Pitfalls
Total Trappnights		80	8	80	8	80	8	80	8
<b>Common name</b>	<b>Scientific name</b>								
Marbled Frog	<i>Limnodynastes convexiusculus</i>						4		
Closed-litter Rainbow-skink	<i>Carlia longipes</i>		1		1				
Shaded-litter Rainbow-skink	<i>Carlia munda</i>								1
Callose-palmed Shinning-skink	<i>Cryptoblepharus plagiocephalus</i>				1				1
Lowlands Plainbacked Ctenotus	<i>Ctenotus essingtonii</i>								
Binoe's Gecko	<i>Heteronotia binoei</i>			1					2
Two-lined Dragon	<i>Diporiphora cf. bilineata</i>								1
Grasslands Melomys	<i>Melomys burtoni</i>					2			



Data Source		Current Survey	Brady (unpubl)	Gambold et al (1995)	URS 2003	NT Fauna Atlas 2003
<b>Amphibians</b>						
Bilingual Froglet	<i>Crinia bilingua</i>			x		
Marbled Frog	<i>Limnodynastes convexiusculus</i>	x		x		x
Ornate Burrowing Frog	<i>Limnodynastes ornatus</i>			x		
Floodplain Toadlet	<i>Uperoleia inundata</i>			x		
Giant Frog	<i>Cyclorana australis</i>	x		x		x
Bicolor Tree Frog	<i>Litoria bicolor</i>	x		x		x
Green Tree Frog	<i>Litoria caerulea</i>	x		x		x
Dwarf Rocket Frog	<i>Litoria dorsalis</i>			x		
Rocket Frog	<i>Litoria nasuta</i>	x		x		x
Roth's Tree Frog	<i>Litoria rothi</i>	x		x		x
Desert Tree Frog	<i>Litoria rubella</i>	x		x		x
Wotjulum Frog	<i>Litoria wotjulumensis</i>			x		x
Wood Frog	<i>Rana daemeli</i>			x		
<b>Reptiles</b>						
Estuarine Crocodile	<i>Crocodylus porosus</i>	x		x		x
Loggerhead Turtle	<i>Caretta caretta</i>			x		
Green Turtle	<i>Chelonia mydas</i>			x		x
Hawksbill Turtle	<i>Eretmochelys imbricata</i>			x		x
Olive Ridley	<i>Lepidochelys olivacea</i>			x		
Flatback Turtle	<i>Natator depressus</i>			x		
Leatherback Turtle	<i>Dermochelys coriacea</i>			x		
Northern Long-necked Turtle	<i>Chelodina rugosa</i>			x		
Crowned Gecko	<i>Diplodactylus stenodactylus</i>			x		
Northern Dtella	<i>Gehyra australis</i>			x		
Binoe's Gecko	<i>Heteronotia binoei</i>	x		x		
Marbled Velvet Gecko	<i>Oedura marmorata</i>			x		x
Zig Zag Gecko	<i>Oedura rhombifer</i>			x		
Asian House Gecko	<i>Hemidactylus frenatus</i>	x				
Beaked Gecko	<i>Rhynchoedura ornata</i>			x		
Rusty-topped Delma	<i>Delma borea</i>			x		
Excitable Delma	<i>Delma tincta</i>			x		x
Burton's Snake-lizard	<i>Lialis burtonis</i>			x		x
Black-headed Scaly-foot	<i>Pygopus nigriceps</i>			x		
Frilled Lizard	<i>Chlamydosaurus kingii</i>	x		x		x
Two-lined Dragon	<i>Diporiphora cf. bilineata</i>	x		x		
Gilbert's Lashtail	<i>Lophognathus gilberti</i>			x		
Swamplands Lashtail	<i>Lophognathus temporalis</i>			x		
Gould's Monitor	<i>Varanus gouldii</i>			x		x
Northern Sand Monitor	<i>Varanus panoptes</i>	x				
Mangrove Monitor	<i>Varanus indicus</i>			x		
Merten's Water Monitor	<i>Varanus mertensi</i>			x		x
Spotted Tree Monitor	<i>Varanus scalaris</i>			x		x
Black-headed Monitor	<i>Varanus tristis</i>			x		

Data Source		Current Survey	Brady (unpubl)	Gambold et al (1995)	URS 2003	NT Fauna Atlas 2003
Two-spined Rainbow-skink	<i>Carlia amax</i>			x		x
Closed-litter Rainbow-skink	<i>Carlia longipes</i>	x		x		x
Shaded-litter Rainbow-skink	<i>Carlia munda</i>	x		x		
Callose-palmed Shinning-skink	<i>Cryptoblepharus plagiocephalus</i>	x		x		x
Lowlands Plainbacked Ctenotus	<i>Ctenotus essingtonii</i>	x		x		x
Plain Ctenotus	<i>Ctenotus inornatus</i>	x		x		
Robust Ctenotus	<i>Ctenotus robustus</i>			x		
Straight-browed Ctenotus	<i>Ctenotus spaldingi</i>			x		
Scant-striped Ctenotus	<i>Ctenotus vertebralis</i>			x		
Cape York Mulch-skink	<i>Glaphromorphus crassicaudus</i>			x		
Douglas' Skink	<i>Glaphromorphus douglasi</i>			x		
Smooth-scaled Skink	<i>Glaphromorphus isolepis</i>			x		
Dark-tailed Skink	<i>Glaphromorphus nigrocaudis</i>			x		
Lesser Robust Fine-lined Slider	<i>Lerista karlschmidti</i>			x		
Single-toed Lerista	<i>Lerista stylis</i>			x		
Macfarlan's Skink	<i>Lygisaurus macfarlani</i>			x		
Storr's Snake-eyed Skink	<i>Morethia storri</i>			x		x
Northern Blue-tongue Skink	<i>Tiliqua scincoides</i>			x		x
Faint-striped Blind Snake	<i>Ramphotyphlops broomi</i>			x		
Northern Blind Snake	<i>Ramphotyphlops diversus</i>			x		
Brown-snouted Blind Snake	<i>Ramphotyphlops wiedii</i>			x		
Yirrkala Blind Snake	<i>Ramphotyphlops yirrikalae</i>			x		
Children's Python	<i>Liasis childreni</i>			x		x
Water Python	<i>Liasis fuscus</i>			x		x
Olive Python	<i>Liasis olivaceus</i>	x		x		
Carpet Python	<i>Morelia spilota</i>			x		
Little File Snake	<i>Acrochordus granulatus</i>			x		
Brown Tree-snake	<i>Boiga irregularis</i>			x		x
Bockadam	<i>Cerberus rhynchops</i>			x		
Common Tree Snake	<i>Dendrelaphis punctulatus</i>			x		x
Macleay's Water Snake	<i>Enhydryis polylepis</i>			x		
White-bellied Mangrove Snake	<i>Fordonia leucobalia</i>			x		
Slaty-grey Snake	<i>Stegnotus cucullatus</i>			x		
Common Keelback	<i>Tropidonophis mairii</i>			x		x
Northern Death Adder	<i>Acanthophis praelongus</i>			x		x
Black Whip Snake	<i>Demansia atra</i>			x		x
Papuan Whip Snake	<i>Demansia papuensis</i>			x		
Olive Whip Snake	<i>Demansia olivacea</i>			x		
Collared Whip Snake	<i>Demansia torquata</i>			x		
Moon Snake	<i>Furina ornata</i>			x		
Taipan	<i>Oxyuranus scutellatus</i>			x		x
King Brown Snake	<i>Pseudechis australis</i>			x		x
Western Brown Snake	<i>Pseudonaja nuchalis</i>	x		x		x
Northern Small-eyed Snake	<i>Rhinoplocephalus pallidiceps</i>			x		x
Little Spotted Snake	<i>Suta punctata</i>			x		
Northern Bandy Bandy	<i>Vermicella multifasciata</i>			x		

Data Source		Current Survey	Brady (unpubl)	Gambold et al (1995)	URS 2003	NT Fauna Atlas 2003
<b>Birds</b>						
Emu	<i>Dromaius novaehollandiae</i>			x		
Orange-footed Scrubfowl	<i>Megapodius reinwardt</i>	x		x		x
Brown Quail	<i>Coturnix ypsilophora</i>		x	x		
King Quail	<i>Coturnix chinensis</i>			x		
Magpie Goose	<i>Anseranas semipalmata</i>			x		x
Plumed Whistling-Duck	<i>Dendrocygna eytoni</i>			x		x
Wandering Whistling-Duck	<i>Dendrocygna arcuata</i>			x		x
Radjah Shelduck	<i>Tadorna radjah</i>	x		x		x
Green Pygmy-goose	<i>Nettapus pulchellus</i>			x		x
Pacific Black Duck	<i>Anas superciliosa</i>		x	x		x
Grey Teal	<i>Anas gracilis</i>			x		
Pink-eared Duck	<i>Malacorhynchus membranaceus</i>			x		
Australasian Grebe	<i>Tachybaptus novaehollandiae</i>			x		x
Masked Booby	<i>Sula dactylatra</i>			x		
Brown Booby	<i>Sula leucogaster</i>			x		
Darter	<i>Anhinga melanogaster</i>			x	x	x
Little Pied Cormorant	<i>Phalacrocorax melanoleucas</i>			x		x
Pied Cormorant	<i>Phalacrocorax varius</i>			x		x
Little Black Cormorant	<i>Phalacrocorax sulcirostris</i>			x		x
Australian Pelican	<i>Pelicanus conspicillatus</i>			x		x
Great Frigatebird	<i>Fregata minor</i>			x		
Lesser Frigatebird	<i>Fregeta ariel</i>			x		
White-faced Heron	<i>Egretta novaehollandiae</i>			x		
Little Egret	<i>Egretta garzetta</i>			x		x
Eastern Reef Egret	<i>Egretta sacra</i>	x		x	x	x
White-necked Heron	<i>Ardea pacifica</i>			x		x
Great-billed Heron	<i>Ardea sumatrana</i>					x
Pied Heron	<i>Ardea picata</i>		x	x	x	x
Great Egret	<i>Ardea alba</i>	x		x	x	x
Intermediate Egret	<i>Ardea intermedia</i>	x		x		x
Striated Heron	<i>Butorides striatus</i>			x	x	x
Nankeen Night Heron	<i>Nycticorax caledonicus</i>			x		x
Black Bittern	<i>Ixobrychus flavicollis</i>			x		x
Glossy Ibis	<i>Plegadis falcinellus</i>			x		x
Australian White Ibis	<i>Threskiornis molucca</i>	x		x	x	x
Straw-necked Ibis	<i>Threskiornis spinicollis</i>			x		x
Royal Spoonbill	<i>Platalea regia</i>			x		x
Black-necked Stork	<i>Ephippiorhynchus asiaticus</i>	x		x		x
Osprey	<i>Pandion haliaetus</i>	x	x	x	x	x
Pacific Baza	<i>Aviceda subcristata</i>			x		
Black-shouldered Kite	<i>Elanus axillaris</i>		x			x
Square-tailed Kite	<i>Lophoictinia isura</i>					x
Black-breasted Buzzard	<i>Hamirostra melanosternon</i>		x			
Black Kite	<i>Milvus migrans</i>	x	x	x		x
Whistling Kite	<i>Haliastur sphenurus</i>	x	x	x		x
Brahminy Kite	<i>Haliastur indus</i>	x	x	x	x	x

Data Source		Current Survey	Brady (unpubl)	Gambold et al (1995)	URS 2003	NT Fauna Atlas 2003
White-bellied Sea-eagle	<i>Haliaeetus leucogaster</i>	x	x	x		x
Spotted Harrier	<i>Circus assimilis</i>			x		
Swamp Harrier	<i>Circus approximans</i>		x			
Brown Goshawk	<i>Accipiter fasciatus</i>	x	x	x		x
Grey Goshawk	<i>Accipiter novaehollandiae</i>		x	x		x
Collared Sparrowhawk	<i>Accipiter cirrhocephalus</i>			x		x
Red Goshawk	<i>Erythrotriorchis radiatus</i>			x		x
Wedge-tailed Eagle	<i>Aquila audax</i>			x		x
Brown Falcon	<i>Falco berigora</i>	x	x	x		x
Australian Hobby	<i>Falco longipennis</i>	x		x		x
Peregrine Falcon	<i>Falco peregrinus</i>		x	x		x
Nankeen Kestrel	<i>Falco cenchroides</i>		x	x		x
Brolga	<i>Grus rubicunda</i>		x	x		x
Bush Hen	<i>Amauornis olivaceus</i>					x
Chestnut Rail	<i>Eulabeornis castaneiventris</i>			x		
Purple Swamphen	<i>Porphyrio porphyrio</i>			x		x
Eurasian Coot	<i>Fulica atra</i>			x		
Australian Bustard	<i>Ardeotis australis</i>		x	x		x
Red-backed Button-quail	<i>Turnix maculosa</i>		x			
Chestnut-backed Button-quail	<i>Turnix castanota</i>			x		
Black-tailed Godwit	<i>Limosa limosa</i>			x	x	
Bar-tailed Godwit	<i>Limosa lapponica</i>			x		
Little Curlew	<i>Numenius minutus</i>			x		x
Whimbrel	<i>Numenius phaeopus</i>			x	x	x
Eastern Curlew	<i>Numenius madagascariensis</i>			x	x	x
Marsh Sandpiper	<i>Tringa stagnatilis</i>			x		
Common Greenshank	<i>Tringa nebularia</i>			x	x	
Terek Sandpiper	<i>Xenus cinereus</i>	x		x		
Common Sandpiper	<i>Actitis hypoleucos</i>			x	x	x
Grey-tailed Tattler	<i>Heteroscelus brevipes</i>			x	x	
Ruddy Turnstone	<i>Arenaria interpres</i>			x		
Great Knot	<i>Calidris tenuirostris</i>			x		
Red Knot	<i>Calidris canutus</i>			x		
Red-necked Stint	<i>Calidris ruficollis</i>			x	x	x
Sharp-tailed Sandpiper	<i>Calidris acuminata</i>			x		
Curlew Sandpiper	<i>Calidris ferruginea</i>			x	x	x
Broad-billed Sandpiper	<i>Limicola falcinellus</i>					x
Comb-crested Jacana	<i>Irediparra galliacea</i>			x		x
Bush Stone-curlew	<i>Burhinus grallarius</i>			x		x
Beach Stone-curlew	<i>Esacus neglectus</i>	x		x		x
Pied Oystercatcher	<i>Haematopus longirostris</i>			x		x
Sooty Oystercatcher	<i>Haematopus fuliginosus</i>			x	x	
Black-winged Stilt	<i>Himantopus himantopus</i>			x	x	x
Pacific Golden Plover	<i>Pluvialis fulva</i>			x		
Grey Plover	<i>Pluvialis squatarola</i>			x		
Red-capped Plover	<i>Charadrius ruficapillus</i>			x	x	
Lesser Sand Plover	<i>Charadrius mongolus</i>			x		

Data Source		Current Survey	Brady (unpubl)	Gambold et al (1995)	URS 2003	NT Fauna Atlas 2003
Greater Sand Plover	<i>Charadrius leschenaultii</i>			x	x	
Black-fronted Dotterel	<i>Euseyornis melanops</i>			x		x
Red-kneed Dotterel	<i>Erythrogonys cinctus</i>					x
Masked Lapwing	<i>Vanellus miles</i>	x		x		
Australian Pratincole	<i>Stiltia isabella</i>			x		x
Kelp Gull	<i>Larus dominicanus</i>			x		
Silver Gull	<i>Larus novaehollandiae</i>	x		x	x	x
Gull-billed Tern	<i>Sterna nilotica</i>			x		x
Caspian Tern	<i>Sterna caspia</i>			x		x
Lesser Crested Tern	<i>Sterna bengalensis</i>			x		x
Crested Tern	<i>Sterna bergii</i>	x		x	x	x
Roseate Tern	<i>Sterna dougallii</i>			x	x	
Black-naped Tern	<i>Sterna sumatrana</i>			x	x	x
Common Tern	<i>Sterna hirundo</i>					x
Little Tern	<i>Sterna albifrons</i>			x	x	x
Sooty Tern	<i>Sterna fuscata</i>			x		
Whiskered Tern	<i>Chlidonias hybridus</i>			x	x	x
White-winged Black Tern	<i>Chlidonias leucopterus</i>			x		
Common Noddy	<i>Anous stolidus</i>			x		
Rock Dove	<i>Columba livia</i>			x		x
Emerald Dove	<i>Calcophaps indica</i>		x	x		x
Common Bronzewing	<i>Phaps chalcoptera</i>			x		x
Partridge Pigeon	<i>Geophaps smithii</i>			x		
Chestnut-quilled Rock-pigeon	<i>Petrophassa rufipennis</i>			x		
Diamond Dove	<i>Geopelia cuneata</i>			x		x
Peaceful Dove	<i>Geopelia striata</i>	x	x	x		x
Bar-shouldered Dove	<i>Geopelia humeralis</i>	x	x	x		x
Banded Fruit-dove	<i>Ptilinopus cinctus</i>			x		
Rose-crowned Fruit-dove	<i>Ptilinopus regina</i>			x		x
Pied Imperial-Pigeon	<i>Ducula bicolor</i>		x	x		x
Red-tailed Black-Cockatoo	<i>Calyptorhynchus banksii</i>	x	x	x		x
Galah	<i>Cacatua roseicapilla</i>					x
Little Corella	<i>Cacatua sanguinea</i>	x	x	x		x
Sulphur-crested Cockatoo	<i>Cacatua galerita</i>	x	x	x		x
Rainbow Lorikeet	<i>Trichoglossus haematodus</i>	x	x	x		x
Varied Lorikeet	<i>Psitteuteles versicolor</i>	x	x	x		x
Red-winged Parrot	<i>Aprosmictus erythropterus</i>	x	x	x		x
Northern Rosella	<i>Platycercus venustus</i>		x	x		x
Oriental Cuckoo	<i>Cuculus saturatus</i>			x		
Pallid Cuckoo	<i>Cuculus pallidus</i>					x
Brush Cuckoo	<i>Cacomantis variolosus</i>		x	x		x
Horsfield's Bronze-Cuckoo	<i>Chrysococcyx basalis</i>		x	x		
Little Bronze-Cuckoo	<i>Chrysococcyx minutillus</i>			x		
Common Koel	<i>Eudynamis scolopacea</i>		x	x		x
Pheasant Coucal	<i>Centropus phasianinus</i>		x	x		x
Rufous Owl	<i>Ninox rufa</i>			x		
Barking Owl	<i>Ninox connivens</i>			x		



Data Source		Current Survey	Brady (unpubl)	Gambold et al (1995)	URS 2003	NT Fauna Atlas 2003
Southern Boobook Owl	<i>Ninox novaeseelandiae</i>	x	x	x		x
Barn Owl	<i>Tyto alba</i>			x		
Grass Owl	<i>Tyto capensis</i>					x
Tawny Frogmouth	<i>Podargus strigoides</i>	x	x	x		x
Spotted Nightjar	<i>Eurostopodus argus</i>			x		x
Large-tailed Nightjar	<i>Caprimulgus macrurus</i>			x		x
Australian Owlet-nightjar	<i>Aegotheles cristatus</i>		x	x		x
White-throated Needletail	<i>Hirundapus caudacutus</i>			x		
Fork-tailed Swift	<i>Apus pacificus</i>			x		
Azure Kingfisher	<i>Alcedo azurea</i>			x		x
Blue-winged Kookaburra	<i>Dacelo leachii</i>	x	x	x		x
Forest Kingfisher	<i>Todiramphus macleayii</i>	x	x	x		x
Red-backed Kingfisher	<i>Todiramphus pyrrhopygia</i>			x		x
Sacred Kingfisher	<i>Todiramphus sanctus</i>	x	x	x	x	x
Collared Kingfisher	<i>Todiramphus chloris</i>	x		x		x
Rainbow Bee-eater	<i>Merops ornatus</i>	x	x	x	x	x
Dollarbird	<i>Eurystomus orientalis</i>		x	x		x
Rainbow Pitta	<i>Pitta iris</i>			x		
Variegated Fairy-wren	<i>Malurus lamberti</i>			x		
Red-backed Fairy-wren	<i>Malurus melanocephalus</i>	x	x	x		x
Striated Pardalote	<i>Pardalotus striatus</i>		x	x		x
Weebill	<i>Smicromis brevirostris</i>		x	x		
Mangrove Gerygone	<i>Gerygone levigaster</i>	x			x	x
Large-billed Gerygone	<i>Gerygone magnirostris</i>			x		
Green-backed Gerygone	<i>Gerygone chloronotus</i>			x		x
White-throated Gerygone	<i>Gerygone olivacea</i>		x	x		
Helmeted Friarbird	<i>Philemon buceroides</i>	x	x	x	x	x
Silver-crowned Friarbird	<i>Philemon argenticeps</i>	x	x	x	x	x
Little Friarbird	<i>Philemon citreogularis</i>	x	x	x		x
Blue-faced Honeyeater	<i>Entomyzon cyanotis</i>	x	x	x		x
Yellow-throated Miner	<i>Manorina flavigula</i>	x	x	x		
White-gaped Honeyeater	<i>Lichenostomus unicolor</i>	x	x	x		x
Yellow-tinted Honeyeater	<i>Lichenostomus flavescens</i>			x		
White-throated Honeyeater	<i>Melithreptus albogularis</i>	x	x	x	x	x
Brown Honeyeater	<i>Lichmera indistincta</i>	x	x	x	x	x
Bar-breasted Honeyeater	<i>Ramsayornis fasciatus</i>	x	x	x		x
Rufous-banded Honeyeater	<i>Conopophila albogularis</i>			x		x
Rufous-throated Honeyeater	<i>Conopophila rufogularis</i>			x		x
Banded Honeyeater	<i>Certhionyx pectoralis</i>			x		
Dusky Honeyeater	<i>Myzomela obscura</i>	x	x	x		x
Red-headed Honeyeater	<i>Myzomela erythrocephala</i>	x		x	x	x
Jacky Winter	<i>Microeca fascinans</i>	x		x		x
Lemon-bellied Flycatcher	<i>Microeca flavigaster</i>		x	x		x
Hooded Robin	<i>Melanodryas cucullata</i>					x
Grey-crowned Babbler	<i>Pomatostomas temporalis</i>			x		x
Varied Sittela	<i>Daphoenositta chrysoptera</i>			x		x
Grey Whistler	<i>Pachycephala simplex</i>			x		x

Data Source		Current Survey	Brady (unpubl)	Gambold et al (1995)	URS 2003	NT Fauna Atlas 2003
Rufous Whistler	<i>Pachycephala rufiventris</i>		x	x		x
White-breasted Whistler	<i>Pachycephala lanoides</i>				x	
Little Shrike-thrush	<i>Colluricincla megarhyncha</i>			x		
Grey Shrike-thrush	<i>Colluricincla harmonica</i>			x		
Broad-billed Flycatcher	<i>Myiagra ruficollis</i>			x		x
Leaden Flycatcher	<i>Myiagra rubecula</i>		x	x	x	x
Shining Flycatcher	<i>Myiagra alecto</i>			x	x	
Restless Flycatcher	<i>Myiagra inquieta</i>			x		x
Australian Magpie-lark	<i>Grallina cyanoleuca</i>	x	x	x		x
Rufous Fantail	<i>Rhipidura rufifrons</i>			x		
Grey Fantail	<i>Rhipidura fuliginosa</i>					x
Northern Fantail	<i>Rhipidura rufiventris</i>	x	x	x		x
Willie Wagtail	<i>Rhipidura leucophrys</i>			x		x
Spangled Drongo	<i>Dicrurus bracteatus</i>	x	x	x		x
Black-faced Cuckoo-shrike	<i>Coracina novaehollandiae</i>	x	x	x		x
White-bellied Cuckoo-shrike	<i>Coracina papuensis</i>		x	x	x	x
Cicadabird	<i>Coracina tenuirostris</i>			x		
White-winged Triller	<i>Lalage sueurii</i>		x	x		x
Varied Triller	<i>Lalage leucomela</i>	x	x	x	x	x
Yellow Oriole	<i>Oriolus flavocinctus</i>	x	x	x		x
Olive-backed Oriole	<i>Oriolus sagittatus</i>	x	x	x		x
Figbird	<i>Sphecotheres viridus</i>		x	x		x
White-breasted Woodswallow	<i>Artamus leucorhynchus</i>		x	x		x
Black-faced Woodswallow	<i>Artamus cinereus</i>	x	x	x		x
Grey Butcherbird	<i>Cracticus torquatus</i>	x	x	x		x
Pied Butcherbird	<i>Cracticus nigrogularis</i>	x	x	x		x
Torresian Crow	<i>Corvus orru</i>	x	x	x		x
Apostlebird	<i>Struthidea cinerea</i>					x
Great Bowerbird	<i>Chlamydera nuchalis</i>	x	x	x		x
Richard's Pipit	<i>Anthus novaeseelandiae</i>		x	x		
Red-throated Pipit	<i>Anthus cervinus</i>		x			
Double-barred Finch	<i>Taeniopygia bichenovii</i>	x	x	x		
Long-tailed Finch	<i>Poephila acuticauda</i>			x		x
Masked Finch	<i>Poephila personata</i>			x		x
Crimson Finch	<i>Neochmia phaeton</i>			x		x
Gouldian Finch	<i>Erythrura gouldiae</i>			x		
Mistletoebird	<i>Dicaeum hirundinaceum</i>		x	x		x
Tree Martin	<i>Hirundo nigricans</i>	x	x	x		x
Tawny Grassbird	<i>Megalurus timoriensis</i>			x		
Rufous Songlark	<i>Cincloramphus mathewsi</i>			x		
Golden-headed Cisticola	<i>Cisticola exilis</i>		x	x		x
Yellow White-eye	<i>Zosterops luteus</i>			x		
<b>Mammals</b>						
Irrawady Dolphin	<i>Orcaella brevirostris</i>					x
False Killer Whale	<i>Pseudorca crassidens</i>					x
Dugong	<i>Dugong dugon</i>					x

Data Source		Current Survey	Brady (unpubl)	Gambold et al (1995)	URS 2003	NT Fauna Atlas 2003
Short-beaked Echidna	<i>Tachyglossus aculeatus</i>			x		x
Fawn Antechinus	<i>Antechinus bellus</i>			x		
Brush-tailed Phascogale	<i>Phascogale tapoatafa</i>			x		
Northern Quoll	<i>Dasyurus hallucatus</i>			x		
Red-cheeked Dunnart	<i>Sminthopsis virginiae</i>			x		
Common Planigale	<i>Panigale maculata</i>			x		
Golden Bandicoot	<i>Isoodon auratus</i>			x		x
Northern Brown Bandicoot	<i>Isoodon macrourus</i>	x		x		
Common Brushtail Possum	<i>Trichosurus vulpecula</i>			x		
Sugar Glider	<i>Petaurus breviceps</i>			x		
Rock Ringtail Possum	<i>Pseudocheirus dahli</i>			x		
Short-eared Rock Wallaby	<i>Petrogale brachyotis</i>			x		
Agile Wallaby	<i>Macropus agilis</i>	x		x		x
Antilopine Wallaroo	<i>Macropus antilopinus</i>			x		x
Common Wallaroo	<i>Macropus robustus</i>			x		
Little Red Flying-fox	<i>Pteropus scapulatus</i>	x		x		x
Black Flying-fox	<i>Pteropus alecto</i>	x		x		
Northern Blossom Bat	<i>Macroglossus minimus</i>	x		x		
Ghost Bat	<i>Macroderma gigas</i>			x		
Dusky Horseshoe-bat	<i>Hipposideros ater</i>			x		
Orange Horseshoe-bat	<i>Rhinonicteris aurantius</i>			x		
Yellow-bellied Sheathtail-bat	<i>Saccolaimus flaviventris</i>	x		x		
Common Sheathtail-bat	<i>Taphozous georgianus</i>			x		x
Hoary Wattled Bat	<i>Chalinolobus nigrogriseus</i>	?		x		
Northern Brown Bat	<i>Eptesicus caurinus</i>			x		
Common Bentwing Bat	<i>Miniopterus shreibersii</i>			x		
Arnhem Land Long-eared Bat	<i>Nyctophilus arnhemensis</i>	?		x		
Northern Pipistrelle	<i>Pipistrellus westralis</i>	x				
Dusky Rat	<i>Rattus colletti</i>			x		
Black Rat	<i>Rattus rattus</i>			x		x
Pale Field rat	<i>Rattus tunneyi</i>			x		
Water Rat	<i>Hydromys chrysogaster</i>			x		
Brush-tailed Rabbit-rat	<i>Conilurus penicillatus</i>			x		
Black-footed Tree Rat	<i>Mesembriomys gouldii</i>			x		
Delicate Mouse	<i>Pseudomys delicatulus</i>			x		
House Mouse	<i>Mus musculus</i>			x		x
Northern Hopping Mouse	<i>Notomys aquilo</i>			x		
Grasslands Melomys	<i>Melomys burtoni</i>	x		x		x
Dingo	<i>Canis lupus dingo</i>	x		x		
Feral Cat	<i>Felis catus</i>	x		x		
Feral Horse	<i>Equus caballus</i>			x		
Feral Pig	<i>Sus scrofa</i>			x		
Water Buffalo	<i>Bubalus bubalis</i>	x		x		
European Cattle	<i>Bos taurus</i>			x		



E=Endangered; V=Vulnerable; nt-near threatened  
S=seabird; W-wader/shorebird; L=landbird

		TPWC Act 2000 <sup>1</sup>	EPBC Act <sup>2</sup> 1999	EPBC Act 1999 Migratory spp
<b>Amphibians</b>				
Wood Frog	<i>Rana daemeli</i>	V		
<b>Reptiles</b>				
Estuarine Crocodile	<i>Crocodylus porosus</i>			x
Loggerhead Turtle	<i>Caretta caretta</i>	E	E	x
Green Turtle	<i>Chelonia mydas</i>		V	x
Hawksbill Turtle	<i>Eretmochelys imbricata</i>		V	x
Olive Ridley	<i>Lepidochelys olivacea</i>		E	x
Flatback Turtle	<i>Natator depressus</i>		V	x
Leatherback Turtle	<i>Dermochelys coriacea</i>	V	V	x
<b>Birds</b>				
Emu	<i>Dromaius novaehollandiae</i>	nt		
Masked Booby	<i>Sula dactylatra</i>			S
Brown Booby	<i>Sula leucogaster</i>			S
Great Frigatebird	<i>Fregata minor</i>			S
Lesser Frigatebird	<i>Fregata ariel</i>			S
Eastern Reef Egret	<i>Egretta sacra</i>			W
Great Egret	<i>Ardea alba</i>			W
Glossy Ibis	<i>Plegadis falcinellus</i>			W
Osprey	<i>Pandion haliaetus</i>			L
White-bellied Sea-eagle	<i>Haliaeetus leucogaster</i>			L
Red Goshawk	<i>Erythrorhynchus radiatus</i>	V	V	
Bush Hen	<i>Amaurornis olivaceus</i>	nt		
Australian Bustard	<i>Ardeotis australis</i>	nt		
Black-tailed Godwit	<i>Limosa limosa</i>			W
Bar-tailed Godwit	<i>Limosa lapponica</i>			W
Little Curlew	<i>Numenius minutus</i>			W
Whimbrel	<i>Numenius phaeopus</i>			W
Eastern Curlew	<i>Numenius madagascariensis</i>			W
Marsh Sandpiper	<i>Tringa stagnatilis</i>			W
Common Greenshank	<i>Tringa nebularia</i>			W
Terek Sandpiper	<i>Xenus cinereus</i>			W
Common Sandpiper	<i>Actitis hypoleucos</i>			W
Grey-tailed Tattler	<i>Heteroscelus brevipes</i>			W
Ruddy Turnstone	<i>Arenaria interpres</i>			W
Great Knot	<i>Calidris tenuirostris</i>			W
Red Knot	<i>Calidris canutus</i>			W
Red-necked Stint	<i>Calidris ruficollis</i>			W
Sharp-tailed Sandpiper	<i>Calidris acuminata</i>			W
Curlew Sandpiper	<i>Calidris ferruginea</i>			W
Broad-billed Sandpiper	<i>Limicola falcinellus</i>			W
Bush Stone-curlew	<i>Burhinus grallarius</i>	nt		
Pacific Golden Plover	<i>Pluvialis fulva</i>			W

		TPWC Act 2000 <sup>1</sup>	EPBC Act <sup>2</sup> 1999	EPBC Act 1999 Migratory spp
Grey Plover	<i>Pluvialis squatarola</i>			W
Lesser Sand Plover	<i>Charadrius mongolus</i>			W
Greater Sand Plover	<i>Charadrius leschenaultii</i>			W
Caspian Tern	<i>Sterna caspia</i>			S
Lesser Crested Tern	<i>Sterna bengalensis</i>			S
Black-naped Tern	<i>Sterna sumatrana</i>			S
Common Tern	<i>Sterna hirundo</i>			S
Little Tern	<i>Sterna albifrons</i>			S
Common Noddy	<i>Anous stolidus</i>			S
Partridge Pigeon	<i>Geophaps smithii</i>	nt	V	
Banded Fruit-dove	<i>Ptilinopus cinctus</i>	nt		
Oriental Cuckoo	<i>Cuculus saturatus</i>			L
White-throated Needletail	<i>Hirundapus caudacutus</i>			L
Fork-tailed Swift	<i>Apus pacificus</i>			L
Rainbow Bee-eater	<i>Merops ornatus</i>			L
Leaden Flycatcher	<i>Myiagra rubecula</i>			L
Restless Flycatcher	<i>Myiagra inquieta</i>			L
Rufous Fantail	<i>Rhipidura rufifrons</i>			L
Gouldian Finch	<i>Erythrura gouldiae</i>	E	E	
<b>Mammals</b>				
Irrawady Dolphin	<i>Orcaella brevirostris</i>			x
Dugong	<i>Dugong dugon</i>	nt		x
Fawn Antechinus	<i>Antechinus bellus</i>	nt		
Northern Quoll	<i>Dasyurus hallucatus</i>	nt		
Golden Bandicoot	<i>Isodon auratus</i>	E	V	
Ghost Bat	<i>Macroderma gigas</i>	nt		
Orange Horseshoe-bat	<i>Rhinonicteris aurantius</i>	nt		
Pale Field rat	<i>Rattus tunneyi</i>	nt		
Black-footed Tree Rat	<i>Mesembriomys gouldii</i>	nt		
Northern Hopping Mouse	<i>Notomys aquilo</i>	V	V	
<b>Invertebrates</b>				
Gove Crow Butterfly	<i>Euploea alcatheae enastri</i>	E		

<sup>1</sup> Territory Parks and Wildlife Conservation Act 2000

<sup>2</sup> Environment Protection and Biodiversity Conservation Act 1999