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Pathogenesis and causes of spontaneous primary ovarian insufficiency (premature ovarian failure)

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INTRODUCTION

Primary hypogonadism in women is defined as ovarian failure accompanied by high serum folliclestimulating hormone (FSH) concentrations. Premature ovarian failure (POF), now referred to as primary ovarian insufficiency (POI), is defined as primary hypogonadism in a woman under the age of 40 years.

POI is characterized by loss of oocytes, lack of folliculogenesis and ovarian estrogen production, and infertility. Transient or partial resumption of ovarian activity has been documented in over 50 percent of women with POI based upon hormonal measurements, pelvic ultrasonography, or conception [1,2].

There are several known causes of ovarian failure, including chromosomal defects like Turner syndrome and fragile X syndrome premutation carriers, exposure to radiation and certain drugs, and autoimmune disease. The list of mutations that can cause ovarian failure has increased rapidly, as discoveries from the human genome project and next-generation sequencing increase our understanding of the factors involved in ovarian development. Unfortunately, the etiology remains unknown in approximately 75 to 90 percent of cases [<u>3</u>].

The pathogenesis and causes of POI will be reviewed here (<u>table 1</u>); its evaluation and treatment are discussed separately. (See <u>"Clinical manifestations and diagnosis of spontaneous primary</u> ovarian insufficiency (premature ovarian failure)" and <u>"Management of spontaneous primary</u> ovarian insufficiency (premature ovarian failure)".)

PATHOGENESIS

Normal ovarian development is dependent on a carefully timed cascade of events; as a result, there are multiple potential etiologies for ovarian dysfunction. Pathophysiologically, these defects can be divided into two major categories:

- Accelerated follicle depletion
- Decreased steroid production without oocyte loss

Other theoretical causes of ovarian failure, including absent germ cell migration or defective ovarian development, have not yet been described, although some genes appear to play a role in normal ovarian development.

ACCELERATED FOLLICLE DEPLETION

The genetic, autoimmune, and environmental or toxic causes of primary ovarian insufficiency (POI), formerly referred to as premature ovarian failure (POF), often result in accelerated follicle depletion. Complete follicular depletion indicates that there are no remaining primordial follicles in the ovary. For most women, this process is usually complete by approximately age 51 years (the average age of menopause). However, the rate of follicle depletion may be accelerated by a number of genetic disorders and ovarian toxins, resulting in early menopause.

Genetic causes

X chromosome disorders

Turner syndrome — The lack of a second X chromosome (Turner syndrome) is the most common chromosomal defect in humans, occurring in up to 1.5 percent of conceptions, 10 percent of spontaneous abortions, and 1 of 2500 live births [4,5]. It is also one of the most common causes of POI.

The ovaries in Turner syndrome characteristically consist of small amounts of connective tissue and no follicles or only a few atretic follicles ("streak gonads"). The initial reported cases had extreme forms of this phenotype with absent pubertal development, primary amenorrhea, and multiple morphologic defects [6]; however, the degree of ovarian dysfunction and the extent of the defects are variable. While most affected women have no pubertal development and primary amenorrhea, some develop normally and then have secondary amenorrhea, and some have no morphologic defects and normal stature.

Early histologic studies of the ovaries of eight XO fetuses are the basis of the understanding of the gonadal failure in Turner syndrome [$\underline{7}$]. These ovaries contained apparently normal numbers of primordial germ cells up to at least 12 weeks of gestation, but at later gestational ages, the numbers of germ cells were decreased and connective tissue was increased, as compared with age-matched normal fetuses. These results indicate that the ovarian failure is more likely caused by accelerated atresia than by abnormal germ cell formation. The diagnosis of Turner syndrome is

established by karyotypic analysis. This topic is reviewed in more detail separately. (See <u>"Clinical manifestations and diagnosis of Turner syndrome"</u>, section on 'Pathogenesis'.)

Other X chromosome deletions/translocations — In addition to X monosomy and deletions in the short arm of the X chromosome causing Turner syndrome, X chromosome deletions, inversions, and duplications and balanced X chromosome-to-autosome translocations are the most common causes of POI. Small deletions and breakpoints in translocated X chromosomes lead to the identification of an X chromosome region critical for ovarian development and function: Xq13 to Xq26 with the exception of Xq22 [8]. Unfortunately, mapping the precise location of these breakpoints has not elucidated genes involved in ovarian function, because a gene was not interrupted or the interrupted gene had no link to ovarian function. The exception is diaphanous (*DIAPH2*), which was identified at a breakpoint in an X chromosome-to-autosome translocation in a woman with secondary amenorrhea [9]. Mutations in the Drosophila melanogaster diaphanous homolog of *DIAPH2* disrupt oogenesis and spermatogenesis, supporting a role for *DIAPH2* in ovarian function [9].

Fragile X syndrome premutation carriers — There is a strong relationship between age at menopause, including POI, and premutations (defined below) for the fragile X syndrome (the *FMR1* gene) [10,11]. Fragile X syndrome is an X-linked form of intellectual disability that is one of the most common causes of mental retardation worldwide [12].

The genetics of the syndrome are complex. Affected subjects have more than 200 CGG repeats in the 5' untranslated region of the *FMR1* gene; as a result, a full mutation refers to an allele with more than 200 repeats. Alleles with less than 40 CGG are normal, repeat lengths 40 to 55 are intermediate or "gray zone," and repeat lengths of 55 to 200 are relatively unstable and called "premutations." Premutations can expand to a full mutation when transmitted by females with a rate dependent upon the number of repeats. In males, the premutation may stay the same, expand to less than a full mutation, or even regress when transmitted. Of note, the term premutation is defined based on the ability of the repeat to expand to a full mutation in one generation. Some studies suggest that the "gray zone" or intermediate repeat alleles, which may expand to a full mutation over two or more generations, may also be associated with POI [13,14].

Carriers of the *FMR1* gene premutation who have repeat lengths of 55 to 200 were originally thought to be unaffected, as they do not have the severe neurodevelopmental problems associated with the full mutation. However, in addition to POI, premutation carriers, particularly males, can present with fragile X-associated tremor-ataxia syndrome (FXTAS) or mild cognitive and behavioral deficits on the spectrum of those seen in fragile X syndrome [<u>15-17</u>]. (See <u>"The spinocerebellar ataxias", section on 'Fragile X-associated tremor/ataxia syndrome'</u>.)

The percentage of premutation carriers having menopause before the age of 40 years ranges from 12 to 28 percent [<u>11</u>]. The number of CGG repeats within the premutation range appears to influence the timing of menopause [<u>18</u>].

The mechanism by which a premutation causes ovarian failure is not known. Subjects with premutations appear to have increased *FMR1* mRNA expression and decreased but not absent FMR1 protein [<u>19,20</u>]. In comparison, full mutations in mentally retarded subjects are associated with methylation of the *FMR1* gene and lack of transcription and protein production.

There is no association between POI and full *FMR1* mutations (see <u>"Fragile X syndrome: Clinical features and diagnosis in children and adolescents", section on 'Pathogenesis</u>). Therefore, since premutation carriers have a normal amount of protein, the increased levels of mRNA are postulated to cause a toxic effect [21].

Although early menopause occurs in less than one-third of patients with premutations, the remainder exhibit hormonal changes of early ovarian aging while still experiencing regular menstrual cycles. Women who carry the premutation and have children or relatives affected by fragile X syndrome have an earlier menopause compared with control women, and the age of menopause is dependent on the premutation repeat length [22]. This was also illustrated in a study of 11 affected women ages 24 to 41 years and 22 age-matched controls who underwent daily blood samples for gonadotropin, sex steroid, and inhibin levels with the following results [23]:

- Follicular phase length was significantly shorter in the carriers compared with controls (12.9 versus 14.5 days).
- Serum estradiol concentrations were not different between the groups, but mean follicular phase follicle-stimulating hormone (FSH) concentrations were higher in the premutation carriers (21.9 versus 11.2 international units/L).
- Inhibin B concentrations in the follicular phase and inhibin A and progesterone concentrations in the luteal phase were lower in the premutation carriers compared with controls, changes consistent with ovarian aging.

A separate issue is the frequency of fragile X premutations in women with sporadic or familial POI, which has been addressed in several studies. The prevalence of *FMR1* premutations ranges from 2 to 5 percent in women with sporadic POI and from 12 to 14 percent in women with POI with at least one family member who also had menopause before age 40 years [24-27]. The frequency of fragile X premutations in normal women is estimated to be approximately 1 in 259 [28]. (See "Prenatal screening and diagnosis for fragile X syndrome", section on 'Candidates for screening'.)

Thus, the American College of Obstetricians and Gynecologists (ACOG) now recommends that women with POI or an elevated FSH level before age 40 years without known cause be screened for *FMR1* premutations [29].

Although women with POI have relative infertility, many are still actively trying to conceive when the diagnosis of POI is made. Discovering that she may be a carrier for a mental retardation allele (since premutations are unstable) may help these women resolve their decision to become 2/9/2020

pregnant with their own oocytes. In addition, identification of carrier status is likely to prompt screening of sisters and cousins who are still fertile, and the discovery of fragile X carrier status may exclude a sister as a potential oocyte donor.

Somatic chromosomal defects

Galactosemia — POI is common in women with galactosemia. Because the hypogonadism cannot be eliminated by early metabolic control, it is thought to be caused by the toxic effects of galactose metabolites [30], rather than abnormalities in the carbohydrate composition of gonadotropins that reduce their bioactivity [31]. (See <u>"Galactosemia: Clinical features and diagnosis"</u>.)

Even subtle defects in galactose metabolism may have an impact on ovarian function. In a study of 104 normal postmenopausal women, 15 had decreased galactose-1-phosphate uridyl transferase activity and an abnormal transferase allele by gel electrophoresis; these women had a lower mean age at menopause (45 versus 49 years) [<u>32</u>].

In contrast, in a second report of 58 pre- and postmenopausal women who were heterozygous for the classical galactosemia mutation, there were no differences in serum markers of ovarian aging (FSH, inhibin B, anti-müllerian hormone) or antral follicle count on ultrasound in the 42 premenopausal carriers when compared with a cohort of proven fertile women. In addition, the mean age at menopause in the postmenopausal carriers was 49.7 years, no different from that seen in the general population [<u>33</u>].

Genes associated with 46,XX gonadal dysgenesis — A number of new genes have been discovered using whole exome sequencing in consanguineous families [<u>34-38</u>]. These mutations, which appear to be rare, cause primary amenorrhea associated with POI (XX gonadal dysgenesis). The majority of the mutations identified in these consanguineous families in the absence of other syndromic features have been found in genes important for meiosis, homologous recombination, and DNA damage and repair [<u>34-38</u>]. However, some of the genes involved overlap with regions of the genome that are associated with age of menopause in the general population [<u>35,39</u>]. These include:

- STAG3 A homozygous, recessive base pair deletion in STAG3 [34], a meiosis-specific subunit of the cohesion ring, which is responsible for proper synapsing of sister chromatids. The oocytes in mice with STAG3 deletions are arrested at the prophase I stage, unable to form complete meiosis and resulting in oocyte loss.
- SYCE1, MCM8, MCM9, SPIDR, and PSMC3IP.
- BRCA2 Compound, heterozygous protein truncating mutations in BRCA2, resulting in reduced protein levels, were identified in sisters with XX gonadal dysgenesis, microcephaly,

and early-onset leukemia in one sister [<u>40</u>]. The mutation results in impaired double-strand DNA damage repair.

 ESR2 – A loss-of-function heterozygous missense mutation in the estrogen receptor beta gene (ESR2) was identified in a 16-year-old female with XX gonadal dysgenesis and osteoporosis [41]. The mutation (in a co-factor binding domain) disrupted estradiol signaling and demonstrated a dominant negative effect, suggesting a critical role for ESR2 in ovarian development. (See <u>"Molecular biology and physiology of estrogen action", section on</u> <u>'Estrogen receptors'</u>.)

Other ovarian genes

- FIGLA FIGLA is an oocyte-specific transcription factor that has been demonstrated to carry heterozygous variants affecting gene function in a candidate gene study in 100 women of Han Chinese ethnicity [42].
- NOBOX Heterozygous mutations in NOBOX, the homeodomain binding to NOBOX DNAbinding element (NBE), have been demonstrated to cause dominant, negative disruption of DNA binding and are implicated in POI [43].
- Additional genes have been identified in sporadic or familial cases of POI. A rare heterozygous nonsense mutation in the eukaryotic translation initiation factor 4E nuclear import factor 1 (*eIF4ENIF1*) has been identified in a family with nine women in three consecutive generations who developed menopause at approximately age 30 years, suggesting dominant inheritance [44]. This gene may play a role in meiosis [45].

BMP15 — A mutation in bone morphogenic protein-15 (*BMP15*), a regulator of ovulation and folliculogenesis in animal models, and a member of the TGF family of growth factors that includes activin and inhibin, was initially identified in two women with POI [46]. In functional assays, the mutant *BMP15* was processed abnormally, associated with reduced granulosa cell growth, and antagonized the stimulatory effect of wild-type *BMP15* on granulosa cell proliferation. In a follow-up study of 166 patients with POI who underwent genetic screening, seven (4.2 percent) were found to have one of three heterozygous mutations in the *BMP15* gene. These mutations were not found in control groups of 95 women who had natural menopause after age 50 and 116 subjects from the general population [47].

Other rare syndromic defects — Several neurologic, cancer, and other syndromic disorders present with POI, pointing to the pleiotropic effects of the genes involved. They include:

Mutations in the *FOXL2* gene, which are associated with blepharophimosis/ptosis/epicanthus inversus syndrome (BPES) [<u>48-50</u>]. BPES type I, but not type II, is associated with POI [<u>51</u>]. In addition, mutations in *FOXL2* have been described in patients with isolated POI [<u>51</u>].

- Ataxia telangiectasia presents clinically with unstable gait, progressive motor degeneration, dilated blood vessels (telangiectasias), and germ cell failure. Mutations in ataxia telangiectasia mutated (ATM), a protein kinase that plays a role in the cellular response to genomic damage, are responsible. Mouse models suggest that ATM monitors the normal progression of meiosis [52].
- Bloom syndrome (BLM) is characterized by growth deficiency, predisposition to malignancy, chromosome instability, and POI [53]. Bloom syndrome results from mutations in RecQ protein-like-3 on chromosome 15, a DNA helicase that plays a role in the response to genomic damage.
- White matter abnormalities on magnetic resonance imaging (MRI) associated with neurologic deficits and ovarian failure are due to mutations in the eukaryotic translation initiation factor 2B (*EIF2B*) [54]. Mutations in three separate *EIF2B* factors have been identified: *EIF2B2* (14q), *EIF2B4* (2p), and *EIF2B5* (3q).
- Women with progressive external ophthalmoplegia, a mitochondrial disorder, have an increased risk of POI. Recent reports demonstrated segregation of a mutation in DNA polymerase gamma (*POLG*) with POI that occurs in this disorder [55].
- Perrault syndrome is a disorder of sensorineural deafness and various additional neurologic manifestations [56]. Women with Perrault syndrome have primary amenorrhea caused by POI. A number of gene defects causing Perrault syndrome affect mitochondrial function.
- FSH receptor mutations. (See 'FSH receptor mutations' below.)

Autoimmune ovarian failure — Autoimmunity was first postulated as a cause of POI when it was noted that some women with adrenal insufficiency also had ovarian insufficiency. These women include many with the type I and type II syndromes of polyglandular autoimmune failure, which are associated with autoantibodies to multiple endocrine and other organs (<u>table 2</u>). A possible association between myasthenia gravis and POI has also been reported [<u>57,58</u>].

There is strong histologic evidence that POI, when it occurs in association with adrenal autoimmunity, is a specific entity mediated by an intense lymphocytic infiltration of thecal cells [59]. In this disorder, primordial follicles are spared, although there is likely spillover of the inflammatory infiltrate with time. Theoretically, ovarian function might be restored if a safe and effective immunosuppressive regimen could be developed and used early in the course.

In one series, 4 percent of women with 46,XX POI had lymphocytic autoimmune oophoritis and positive antiadrenal antibodies. The frequency of autoimmune oophoritis among women with ovarian failure without coexisting adrenal failure is not known. (See <u>"Clinical features and diagnosis of autoimmune primary ovarian insufficiency (premature ovarian failure)"</u>.)

Ovarian toxins — Ovarian toxins associated with POI include chemotherapy and radiation therapy. Viral infections have been suspected but not established.

Chemotherapy/radiation therapy — Chemotherapeutic drugs and radiation therapy are the most common causes of toxin-induced ovarian failure (<u>table 3</u>). (See <u>"Ovarian failure due to</u> <u>anticancer drugs and radiation"</u>.)

Viruses — Viruses such as mumps have been associated with orchitis and testicular failure in men and have therefore been presumed to cause oophoritis and ovarian failure in women. However, acute oophoritis is much more difficult to diagnose in women than is orchitis in men, and most of the evidence that viral infections can cause primary hypogonadism in women is circumstantial [60]. Although viral oophoritis has been experimentally induced in cows [61], few cases of viral infection of the ovaries have been documented histologically in women.

A report described a woman with widespread cytomegalovirus (CMV) infection who had histologic evidence of CMV oophoritis at autopsy; however, she had had no clinical evidence of hypogonadism [62,63]. While some women with primary hypogonadism can recall a preceding viral illness, a cause-and-effect relationship has not been established.

Other — Other toxins may have more subtle effects on ovarian function. As an example, cigarette smokers reach menopause approximately two years earlier than nonsmokers [<u>64,65</u>]. Endogenous factors that correlate with age at menopause include weight and socioeconomic status, which increase it, and parity, which decreases it [<u>64,66</u>].

PRIMARY HYPOGONADISM WITHOUT FOLLICLE DEPLETION

Some disorders that are not associated with follicular depletion may present clinically as primary hypogonadism. These disorders are genetic disorders of estradiol precursor production or aromatase function, resulting in decreased estradiol and absence of normal follicle-stimulating hormone (FSH) negative feedback.

There are two major mechanisms of primary hypogonadism without follicle depletion: endogenous mediators of gonadotropin receptor activation and steroidogenic enzyme defects that prevent estradiol production. Secretion of biologically inactive gonadotropins, a cause of secondary hypogonadism, presents in a similar fashion to primary ovarian insufficiency (POI) in that serum immunoreactive gonadotropin concentrations are high. (See <u>"Evaluation and management of secondary amenorrhea"</u> and <u>"Evaluation and management of primary amenorrhea"</u>.)

Intraovarian modulators — Many substances that may serve as paracrine regulators of ovarian responsiveness have been identified. Abnormal ovarian production of any of these modulators could cause primary hypogonadism, either directly or by decreasing cellular responses to

gonadotropins. However, only polymorphisms in the alpha subunit of inhibin have been implicated in ovarian failure [<u>67</u>].

Steroidogenic enzyme defects — Some of the uncommon causes of congenital adrenal hyperplasia are due to genetic defects in the enzymes involved in androstenedione and estradiol biosynthesis in addition to cortisol biosynthesis. These defects result in low estrogen and, therefore, high serum FSH concentrations. Although these disorders are not associated with follicular depletion, patients present with hypergonadotropic hypogonadism. (See <u>"Uncommon congenital adrenal hyperplasias", section on 'Lipoid congenital adrenal hyperplasia'</u>.)

The reported enzyme defects include mutations in:

- Steroidogenic acute regulatory enzyme (StAR), the enzyme that moves cholesterol from the outer to the inner mitochondrial membrane [68]. Patients with congenital lipoid hyperplasia caused by StAR mutations typically have severe adrenal insufficiency very soon after birth, although they occasionally present later in infancy. In patients treated with steroid replacement early, there is absence of pubertal development [69].
- Patients with CYP17 deficiency usually present at about the time of expected puberty because of hypertension, hypokalemia, and hypogonadism. Female (46,XX) patients have primary amenorrhea and absent secondary sexual characteristics.
- Mutations in the aromatase gene also cause absence of estradiol secretion and primary amenorrhea [70]. However, the concomitant deficiency of placental aromatase results in androgenization of the mother and fetus due to the failure to aromatize fetal androgens during pregnancy. (See <u>"Adrenal hyperandrogenism"</u>.)
- Mutations of *NR5A1* (also called steroidogenic factor 1) were first described in four families, causing 46,XY disorders of sex development without adrenal insufficiency and 46,XX primary ovarian failure; the mutation was also identified in 2 of 25 women (8 percent) with spontaneous ovarian failure, but in no controls (n = 700) [71]. However, in a second study of 358 women with POI, the overall frequency of *NR5A1* gene mutation rate was only 1.4 percent, suggesting that this is a rare cause of POI [72]. Similar results were seen in a second study [73].

FSH receptor mutations — While the ovaries of all women with POI are unresponsive to FSH and luteinizing hormone (LH), specific receptor defects had not been identified until not long ago. Mutations of the gene for the FSH (or LH) receptor that result in production of receptors that either do not bind FSH or cannot transduce the signal normally initiated by the binding of the hormone to the receptor would be expected to cause primary hypogonadism. As noted above, a mutation of the FSH-receptor gene that results in production of receptors that bind FSH poorly was identified in some young women with POI [74-76]. The mutation resulted in a single amino acid substitution in the extracellular domain of the FSH receptor that prevents FSH binding. Thus, FSH does not

stimulate aromatization of precursors to estradiol, and the women are estrogen deficient. Whether this mutation affects oocyte number and viability remains to be determined. Its effect on male fertility is variable.

A mutation in the LH receptor gene has also been reported [77]. This defect was identified in males who had hypogonadism, and an affected female in the family had secondary amenorrhea with high serum LH but normal FSH concentrations.

Gs alpha subunit gene mutations — In addition to defects in gonadotropin receptors, defects have been described in the guanine nucleotide regulatory protein of adenylate cyclase (G-protein) that is linked to the receptors and via which activated receptors stimulate adenylate cyclase activity. As expected from the multiplicity of receptors activated by the same G-proteins, including parathyroid hormone, FSH, LH, and thyrotropin receptors, affected subjects may have pseudohypoparathyroidism, abnormal gonadal function, and hypothyroidism, among other problems [78-80]. (See "Etiology of hypocalcemia in infants and children", section on 'End-organ resistance to PTH (pseudohypoparathyroidism)'.)

SOCIETY GUIDELINE LINKS

Links to society and government-sponsored guidelines from selected countries and regions around the world are provided separately. (See <u>"Society guideline links: Primary ovarian insufficiency"</u>.)

INFORMATION FOR PATIENTS

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 Basics topics (see <u>"Patient education: Early menopause (primary ovarian insufficiency) (The</u> <u>Basics)</u>") Beyond the Basics topics (see <u>"Patient education: Early menopause (primary ovarian</u> insufficiency) (Beyond the Basics)")

SUMMARY

Although many cases of premature ovarian failure (POF), now known as primary ovarian insufficiency (POI), remain idiopathic, the number of known causes and genetic factors is growing rapidly. However, for 75 to 90 percent of patients, the cause of their ovarian failure remains unexplained. The more common known causes of POI include:

- Turner syndrome, the lack of a second X chromosome (45,XO), is one of the most common causes of POI. Although their ovaries have a normal complement of primordial germ cells early in gestation, a process of accelerated follicular depletion follows. (See <u>'Turner syndrome'</u> above.)
- Fragile X syndrome Women who are carriers of the *FMR1* gene premutation are at increased risk for POI. The prevalence of *FMR1* premutations is estimated to range from 0 to 3.3 percent in women with sporadic POI and from 12 to 16 percent in women with POI with at least one family member who also had menopause before age 40 years. (See <u>'Fragile X syndrome premutation carriers'</u> above.)
- Somatic and X chromosome gene defects associated with POI include mutations in FOXL2, eIF4ENIF1, STAG3, NR5A1, BMP15, follicle-stimulating hormone (FSH) receptor, Gs alpha, and steroidogenic enzymes. However, the majority of the known genetic factors are extremely rare and do not warrant widespread genetic screening outside a research setting. (See <u>'Somatic chromosomal defects'</u> above.)
- Rare mutations in genes have also been identified that are important for meiosis, homologous recombination, and DNA damage and repair. However, some of the genes involved overlap with regions of the genome that are associated with age of menopause in the general population. (See <u>'Genes associated with 46,XX gonadal dysgenesis'</u> above.)
- Autoimmune ovarian failure Approximately 4 percent of women with 46,XX POI have lymphocytic autoimmune oophoritis and positive antiadrenal antibodies. The frequency of autoimmune oophoritis among women with ovarian failure without coexisting adrenal failure is not known. (See <u>'Autoimmune ovarian failure</u>' above.)
- Toxins Chemotherapeutic drugs and radiation therapy are the most common causes of toxin-induced ovarian failure (<u>table 3</u>). (See <u>'Ovarian toxins'</u> above.)

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REFERENCES

- <u>Nelson LM, Anasti JN, Kimzey LM, et al. Development of luteinized graafian follicles in</u> patients with karyotypically normal spontaneous premature ovarian failure. J Clin Endocrinol <u>Metab 1994; 79:1470.</u>
- <u>Taylor AE, Adams JM, Mulder JE, et al. A randomized, controlled trial of estradiol</u> replacement therapy in women with hypergonadotropic amenorrhea. J Clin Endocrinol <u>Metab 1996; 81:3615.</u>
- 3. Nelson LM. Clinical practice. Primary ovarian insufficiency. N Engl J Med 2009; 360:606.
- 4. <u>Jacobs PA. The incidence and etiology of sex chromosome abnormalities in man. Birth</u> <u>Defects Orig Artic Ser 1979; 15:3.</u>
- Hook EB, Warburton D. The distribution of chromosomal genotypes associated with Turner's syndrome: livebirth prevalence rates and evidence for diminished fetal mortality and severity in genotypes associated with structural X abnormalities or mosaicism. Hum Genet 1983; 64:24.
- 6. <u>Albright F, Smith PH, Fraser R. A syndrome characterized by primary ovarian insufficiency</u> and decreased stature: Report of 11 cases with a digression on hormonal control of axillary and pubic hair. Am J Med Sci 1942; 204:625.
- 7. <u>Singh RP, Carr DH. The anatomy and histology of XO human embryos and fetuses. Anat</u> <u>Rec 1966; 155:369.</u>
- Schlessinger D, Herrera L, Crisponi L, et al. Genes and translocations involved in POF. Am J Med Genet 2002; 111:328.
- Bione S, Sala C, Manzini C, et al. A human homologue of the Drosophila melanogaster diaphanous gene is disrupted in a patient with premature ovarian failure: evidence for conserved function in oogenesis and implications for human sterility. Am J Hum Genet 1998; 62:533.
- 10. <u>Fryns JP. The female and the fragile X. A study of 144 obligate female carriers. Am J Med</u> <u>Genet 1986; 23:157.</u>
- 11. <u>Allingham-Hawkins DJ, Babul-Hirji R, Chitayat D, et al. Fragile X premutation is a significant</u> risk factor for premature ovarian failure: the International Collaborative POF in Fragile X study--preliminary data. Am J Med Genet 1999; 83:322.

- 12. <u>de Vries BB, Halley DJ, Oostra BA, Niermeijer MF. The fragile X syndrome. J Med Genet</u> <u>1998; 35:579.</u>
- 13. <u>Bodega B, Bione S, Dalprà L, et al. Influence of intermediate and uninterrupted FMR1 CGG</u> <u>expansions in premature ovarian failure manifestation. Hum Reprod 2006; 21:952.</u>
- Bretherick KL, Fluker MR, Robinson WP. FMR1 repeat sizes in the gray zone and high end of the normal range are associated with premature ovarian failure. Hum Genet 2005; <u>117:376.</u>
- 15. <u>Hagerman RJ, Leavitt BR, Farzin F, et al. Fragile-X-associated tremor/ataxia syndrome</u> (FXTAS) in females with the FMR1 premutation. Am J Hum Genet 2004; 74:1051.
- 16. <u>Greco CM, Hagerman RJ, Tassone F, et al. Neuronal intranuclear inclusions in a new</u> <u>cerebellar tremor/ataxia syndrome among fragile X carriers. Brain 2002; 125:1760.</u>
- 17. Jacquemont S, Hagerman RJ, Leehey M, et al. Fragile X premutation tremor/ataxia syndrome: molecular, clinical, and neuroimaging correlates. Am J Hum Genet 2003; 72:869.
- 18. <u>Sullivan AK, Marcus M, Epstein MP, et al. Association of FMR1 repeat size with ovarian</u> <u>dysfunction. Hum Reprod 2005; 20:402.</u>
- Kenneson A, Zhang F, Hagedorn CH, Warren ST. Reduced FMRP and increased FMR1 transcription is proportionally associated with CGG repeat number in intermediate-length and premutation carriers. Hum Mol Genet 2001; 10:1449.
- 20. <u>Tassone F, Hagerman RJ, Taylor AK, et al. Clinical involvement and protein expression in</u> <u>individuals with the FMR1 premutation. Am J Med Genet 2000; 91:144.</u>
- 21. <u>Tassone F, Hagerman RJ, Taylor AK, et al. Elevated levels of FMR1 mRNA in carrier males:</u> <u>a new mechanism of involvement in the fragile-X syndrome. Am J Hum Genet 2000; 66:6.</u>
- 22. <u>Mailick MR, Hong J, Greenberg J, et al. Curvilinear association of CGG repeats and age at</u> <u>menopause in women with FMR1 premutation expansions. Am J Med Genet B</u> <u>Neuropsychiatr Genet 2014; 165B:705.</u>
- 23. <u>Welt CK, Smith PC, Taylor AE. Evidence of early ovarian aging in fragile X premutation</u> <u>carriers. J Clin Endocrinol Metab 2004; 89:4569.</u>
- 24. <u>Gersak K, Meden-Vrtovec H, Peterlin B. Fragile X premutation in women with sporadic</u> premature ovarian failure in Slovenia. Hum Reprod 2003; 18:1637.
- 25. <u>Marozzi A, Vegetti W, Manfredini E, et al. Association between idiopathic premature ovarian</u> <u>failure and fragile X premutation. Hum Reprod 2000; 15:197.</u>

- 26. <u>Uzielli ML, Guarducci S, Lapi E, et al. Premature ovarian failure (POF) and fragile X</u> premutation females: from POF to to fragile X carrier identification, from fragile X carrier <u>diagnosis to POF association data. Am J Med Genet 1999; 84:300.</u>
- 27. <u>Sherman SL. Premature ovarian failure in the fragile X syndrome. Am J Med Genet 2000;</u> 97:189.
- 28. Rousseau F, Rouillard P, Morel ML, et al. Prevalence of carriers of premutation-size alleles of the FMRI gene--and implications for the population genetics of the fragile X syndrome. Am J Hum Genet 1995; 57:1006.
- 29. American College of Obstetricians and Gynecologists Committee on Genetics. ACOG committee opinion. No. 338: Screening for fragile X syndrome. Obstet Gynecol 2006; 107:1483.
- 30. <u>Chen YT, Mattison DR, Feigenbaum L, et al. Reduction in oocyte number following prenatal</u> <u>exposure to a diet high in galactose. Science 1981; 214:1145.</u>
- 31. <u>Kaufman FR, Kogut MD, Donnell GN, et al. Hypergonadotropic hypogonadism in female</u> patients with galactosemia. N Engl J Med 1981; 304:994.
- 32. <u>Cramer DW, Harlow BL, Barbieri RL, Ng WG. Galactose-1-phosphate uridyl transferase</u> activity associated with age at menopause and reproductive history. Fertil Steril 1989; 51:609.
- 33. <u>Knauff EA, Richardus R, Eijkemans MJ, et al. Heterozygosity for the classical galactosemia</u> <u>mutation does not affect ovarian reserve and menopausal age. Reprod Sci 2007; 14:780.</u>
- 34. <u>Caburet S, Arboleda VA, Llano E, et al. Mutant cohesin in premature ovarian failure. N Engl</u> J Med 2014; 370:943.
- 35. <u>AlAsiri S, Basit S, Wood-Trageser MA, et al. Exome sequencing reveals MCM8 mutation</u> <u>underlies ovarian failure and chromosomal instability. J Clin Invest 2015; 125:258.</u>
- 36. <u>de Vries L, Behar DM, Smirin-Yosef P, et al. Exome sequencing reveals SYCE1 mutation</u> <u>associated with autosomal recessive primary ovarian insufficiency. J Clin Endocrinol Metab</u> <u>2014; 99:E2129.</u>
- 37. <u>Wood-Trageser MA, Gurbuz F, Yatsenko SA, et al. MCM9 mutations are associated with</u> <u>ovarian failure, short stature, and chromosomal instability. Am J Hum Genet 2014; 95:754.</u>
- Smirin-Yosef P, Zuckerman-Levin N, Tzur S, et al. A Biallelic Mutation in the Homologous Recombination Repair Gene SPIDR Is Associated With Human Gonadal Dysgenesis. J Clin Endocrinol Metab 2017; 102:681.

- 39. Day FR, Ruth KS, Thompson DJ, et al. Large-scale genomic analyses link reproductive aging to hypothalamic signaling, breast cancer susceptibility and BRCA1-mediated DNA repair. Nat Genet 2015; 47:1294.
- 40. <u>Weinberg-Shukron A, Rachmiel M, Renbaum P, et al. Essential Role of BRCA2 in Ovarian</u> <u>Development and Function. N Engl J Med 2018; 379:1042.</u>
- Lang-Muritano M, Sproll P, Wyss S, et al. Early-Onset Complete Ovarian Failure and Lack of Puberty in a Woman With Mutated Estrogen Receptor β (ESR2). J Clin Endocrinol Metab 2018; 103:3748.
- 42. <u>Zhao H, Chen ZJ, Qin Y, et al. Transcription factor FIGLA is mutated in patients with</u> premature ovarian failure. Am J Hum Genet 2008; 82:1342.
- 43. <u>Qin Y, Choi Y, Zhao H, et al. NOBOX homeobox mutation causes premature ovarian failure.</u> <u>Am J Hum Genet 2007; 81:576.</u>
- 44. <u>Kasippillai T, MacArthur DG, Kirby A, et al. Mutations in eIF4ENIF1 are associated with</u> primary ovarian insufficiency. J Clin Endocrinol Metab 2013; 98:E1534.
- 45. <u>Pfender S, Kuznetsov V, Pasternak M, et al. Live imaging RNAi screen reveals genes</u> essential for meiosis in mammalian oocytes. Nature 2015; 524:239.
- 46. <u>Di Pasquale E, Beck-Peccoz P, Persani L. Hypergonadotropic ovarian failure associated with</u> <u>an inherited mutation of human bone morphogenetic protein-15 (BMP15) gene. Am J Hum</u> <u>Genet 2004; 75:106.</u>
- 47. <u>Di Pasquale E, Rossetti R, Marozzi A, et al. Identification of new variants of human BMP15</u> gene in a large cohort of women with premature ovarian failure. J Clin Endocrinol Metab 2006; 91:1976.
- 48. <u>Crisponi L, Deiana M, Loi A, et al. The putative forkhead transcription factor FOXL2 is</u> <u>mutated in blepharophimosis/ptosis/epicanthus inversus syndrome. Nat Genet 2001; 27:159.</u>
- 49. Raile K, Stobbe H, Tröbs RB, et al. A new heterozygous mutation of the FOXL2 gene is associated with a large ovarian cyst and ovarian dysfunction in an adolescent girl with blepharophimosis/ptosis/epicanthus inversus syndrome. Eur J Endocrinol 2005; 153:353.
- 50. <u>Settas N, Anapliotou M, Kanavakis E, et al. A novel FOXL2 gene mutation and BMP15</u> variants in a woman with primary ovarian insufficiency and blepharophimosis-ptosisepicanthus inversus syndrome. Menopause 2015; 22:1264.
- 51. <u>Beysen D, De Paepe A, De Baere E. FOXL2 mutations and genomic rearrangements in</u> <u>BPES. Hum Mutat 2009; 30:158.</u>

- 52. <u>Barlow C, Liyanage M, Moens PB, et al. Atm deficiency results in severe meiotic disruption</u> <u>as early as leptonema of prophase I. Development 1998; 125:4007.</u>
- 53. Ellis NA, German J. Molecular genetics of Bloom's syndrome. Hum Mol Genet 1996; 5 Spec No:1457.
- 54. Fogli A, Rodriguez D, Eymard-Pierre E, et al. Ovarian failure related to eukaryotic initiation factor 2B mutations. Am J Hum Genet 2003; 72:1544.
- 55. Pagnamenta AT, Taanman JW, Wilson CJ, et al. Dominant inheritance of premature ovarian failure associated with mutant mitochondrial DNA polymerase gamma. Hum Reprod 2006; 21:2467.
- 56. Jenkinson EM, Clayton-Smith J, Mehta S, et al. Perrault syndrome: further evidence for genetic heterogeneity. J Neurol 2012; 259:974.
- 57. Li Y, Xiao B, Xiao L, et al. Myasthenia gravis accompanied by premature ovarian failure and aggravation by estrogen. Intern Med 2010; 49:611.
- 58. <u>Ryan MM, Jones HR Jr. Myasthenia gravis and premature ovarian failure. Muscle Nerve</u> 2004; 30:231.
- 59. <u>Hoek A, Schoemaker J, Drexhage HA. Premature ovarian failure and ovarian autoimmunity.</u> <u>Endocr Rev 1997; 18:107.</u>
- 60. <u>Morrison JC, Givens JR, Wiser WL, Fish SA. Mumps oophoritis: a cause of premature</u> <u>menopause. Fertil Steril 1975; 26:655.</u>
- 61. <u>Smith PC, Nusbaum KE, Kwapien RP, et al. Necrotic oophoritis in heifers vaccinated</u> <u>intravenously with infectious bovine rhinotracheitis virus vaccine during estrus. Am J Vet Res</u> <u>1990; 51:969.</u>
- 62. <u>Williams DJ, Connor P, Ironside JW. Pre-menopausal cytomegalovirus oophoritis.</u> <u>Histopathology 1990; 16:405.</u>
- 63. <u>Subietas A, Deppisch LM, Astarloa J. Cytomegalovirus oophoritis: ovarian cortical necrosis.</u> <u>Hum Pathol 1977; 8:285.</u>
- 64. <u>Willett W, Stampfer MJ, Bain C, et al. Cigarette smoking, relative weight, and menopause.</u> <u>Am J Epidemiol 1983; 117:651.</u>
- 65. <u>McKinlay SM, Bifano NL, McKinlay JB. Smoking and age at menopause in women. Ann</u> <u>Intern Med 1985; 103:350.</u>

- 66. <u>Stanford JL, Hartge P, Brinton LA, et al. Factors influencing the age at natural menopause. J</u> <u>Chronic Dis 1987; 40:995.</u>
- 67. <u>Shelling AN, Burton KA, Chand AL, et al. Inhibin: a candidate gene for premature ovarian</u> <u>failure. Hum Reprod 2000; 15:2644.</u>
- 68. <u>Lin D, Sugawara T, Strauss JF 3rd, et al. Role of steroidogenic acute regulatory protein in</u> adrenal and gonadal steroidogenesis. Science 1995; 267:1828.
- 69. <u>Mallin SR. Congenital adrenal hyperplasia secondary to 17-hydroxylase deficiency. Two</u> sisters with amenorrhea, hypokalemia, hypertension, and cystic ovaries. Ann Intern Med <u>1969; 70:69.</u>
- 70. <u>Ito Y, Fisher CR, Conte FA, et al. Molecular basis of aromatase deficiency in an adult female</u> with sexual infantilism and polycystic ovaries. Proc Natl Acad Sci U S A 1993; 90:11673.
- 71. Lourenço D, Brauner R, Lin L, et al. Mutations in NR5A1 associated with ovarian insufficiency. N Engl J Med 2009; 360:1200.
- 72. Janse F, de With LM, Duran KJ, et al. Limited contribution of NR5A1 (SF-1) mutations in women with primary ovarian insufficiency (POI). Fertil Steril 2012; 97:141.
- 73. <u>Voican A, Bachelot A, Bouligand J, et al. NR5A1 (SF-1) mutations are not a major cause of primary ovarian insufficiency. J Clin Endocrinol Metab 2013; 98:E1017.</u>
- 74. Aittomäki K. The genetics of XX gonadal dysgenesis. Am J Hum Genet 1994; 54:844.
- 75. <u>Aittomäki K, Lucena JL, Pakarinen P, et al. Mutation in the follicle-stimulating hormone</u> receptor gene causes hereditary hypergonadotropic ovarian failure. Cell 1995; 82:959.
- 76. <u>Nakamura Y, Maekawa R, Yamagata Y, et al. A novel mutation in exon8 of the follicle-</u> <u>stimulating hormone receptor in a woman with primary amenorrhea. Gynecol Endocrinol</u> <u>2008; 24:708.</u>
- 77. Latronico AC, Anasti J, Arnhold IJ, et al. Brief report: testicular and ovarian resistance to luteinizing hormone caused by inactivating mutations of the luteinizing hormone-receptor gene. N Engl J Med 1996; 334:507.
- 78. <u>Levine MA, Downs RW Jr, Moses AM, et al. Resistance to multiple hormones in patients with</u> <u>pseudohypoparathyroidism. Association with deficient activity of guanine nucleotide</u> <u>regulatory protein. Am J Med 1983; 74:545.</u>
- 79. <u>Shapiro MS, Bernheim J, Gutman A, et al. Multiple abnormalities of anterior pituitary</u> <u>hormone secretion in association with pseudohypoparathyroidism. J Clin Endocrinol Metab</u> <u>1980; 51:483.</u>

80. <u>Faull CM, Welbury RR, Paul B, Kendall-Taylor P. Pseudohypoparathyroidism: its phenotypic</u> variability and associated disorders in a large family. Q J Med 1991; 78:251.

Topic 7435 Version 24.0

GRAPHICS

Major causes of primary ovarian insufficiency (premature ovarian failure) in women

Accelerated follicular atresia		
Genetic defects		
Turner syndrome		
Fragile X premutations		
X chromosome deletions and translocations		
Galactosemia		
Ovarian toxins		
Chemotherapeutic drugs (especially alkylating agents)		
Radiation		
Mumps or cytomegalovirus infection		
Autoimmune injury		
Isolated or part of polyglandular autoimmune syndromes		
Abnormal follicular stimulation		
Intraovarian modulators		
BMP15		
Steroidogenic enzyme defects		
CYP17 deficiency, StAR mutation		
Aromatase gene mutations		
Gonadotropin receptor function		
FSH receptor mutations		
Gs alpha subunit gene mutations		

FSH: follicle-stimulating hormone.

Graphic 74029 Version 4.0

Types of endocrine and nonendocrine autoimmune syndromes associated with adrenal insufficiency

Disorder	Prevalence (percent)	
Polyglandular autoimmune syndrome type I		
Endocrine		
Hypoparathyroidism	89	
Chronic mucocutaneous candidiasis	75	
Adrenal insufficiency	60	
Primary hypogonadism	45	
Hypothyroidism	12	
Type 1 diabetes mellitus	1	
Hypopituitarism	<1	
Diabetes insipidus	<1	
Nonendocrine		
Malabsorption syndromes	25	
Alopecia totalis or areata	20	
Pernicious anemia	16	
Chronic active hepatitis	9	
Vitiligo	4	
Polyglandular autoimmune syndrome type II		
Endocrine		
Adrenal insufficiency	100	
Autoimmune thyroid disease	70	
Type 1 diabetes mellitus	50	
Primary hypogonadism	5 to 50	
Diabetes insipidus	<1	
Nonendocrine		
Vitiligo	4	
Alopecia, pernicious anemia, myasthenia gravis, immune thrombocytopenia purpura, Sjogren's syndrome, rheumatoid arthritis	≤1	

Data from:

1. Leshin M. Polyglandular autoimmune syndromes. Am J Med Sci 1985; 290:77.

2. Neufeld M, Maclaren NK, Blizzard RM. Two types of autoimmune Addison's disease associated with different polyglandular autoimmune (PGA) syndromes. Medicine (Baltimore) 1981; 60:355.

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Chemotherapy-associated ovarian toxicity

Drug	Class (action)	
Definitely associated with ovarian damage		
Nitrogen mustard	Mechlorethamine (alkylating agent)	
L-phenylalanine mustard	Mechlorethamine (alkylating agent)	
Chlorambucil	Chloroethylamine (alkylating agent)	
Cyclophosphamide	Chloroethylamine (alkylating agent)	
Melphalan	Mechlorethamine (alkylating agent)	
Busulfan	Alkylalkane sulfonate (alkylating agent)	
Procarbazine	Substituted hydrazine	
Dacarbazine	Alkylating agent	
Probably associated with ovarian damage		
Vinblastine	Vinca alkaloid	
Cytosine arabinoside (Ara-C)	Antimetabolite	
Cis-platinum	Heavy metal	
Carmustine	Nitrosourea (alkylating agent)	
Lomustine	Nitrosourea (alkylating agent)	
VP-16 (etoposide)	Podophyllotoxin	
Imatinib	Tyrosine kinase inhibitor	
Low probability of ovarian damage		
Methotrexate	Antimetabolite	
Fluorouracil (5-FU)	Antimetabolite	
6-mercaptopurine	Antimetabolite	
Vincristine	Vinca alkaloid	
Mitomycin	Antibiotic (alkylating agent)	
Unknown		
VM-26	Podophyllotoxin	
Daunorubicin	Anthracycline	
Bleomycin	Peptide	
Vindesine	Vinca alkaloid	
Doxorubicin	Anthracycline	

Graphic 58142 Version 13.0

Contributor Disclosures

Corrine K Welt, MD Consultant/Advisory Board (Spouse): Medtronic [Cardiac stents for catheterization]. **Robert L Barbieri, MD** Nothing to disclose **William F Crowley, Jr, MD** Equity Ownership/Stock Options: Dare Bioscience [Endocrinology]. Consultant/Advisory Boards: Dare Bioscience [Endocrinology]. **Kathryn A Martin, MD** Nothing to disclose

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