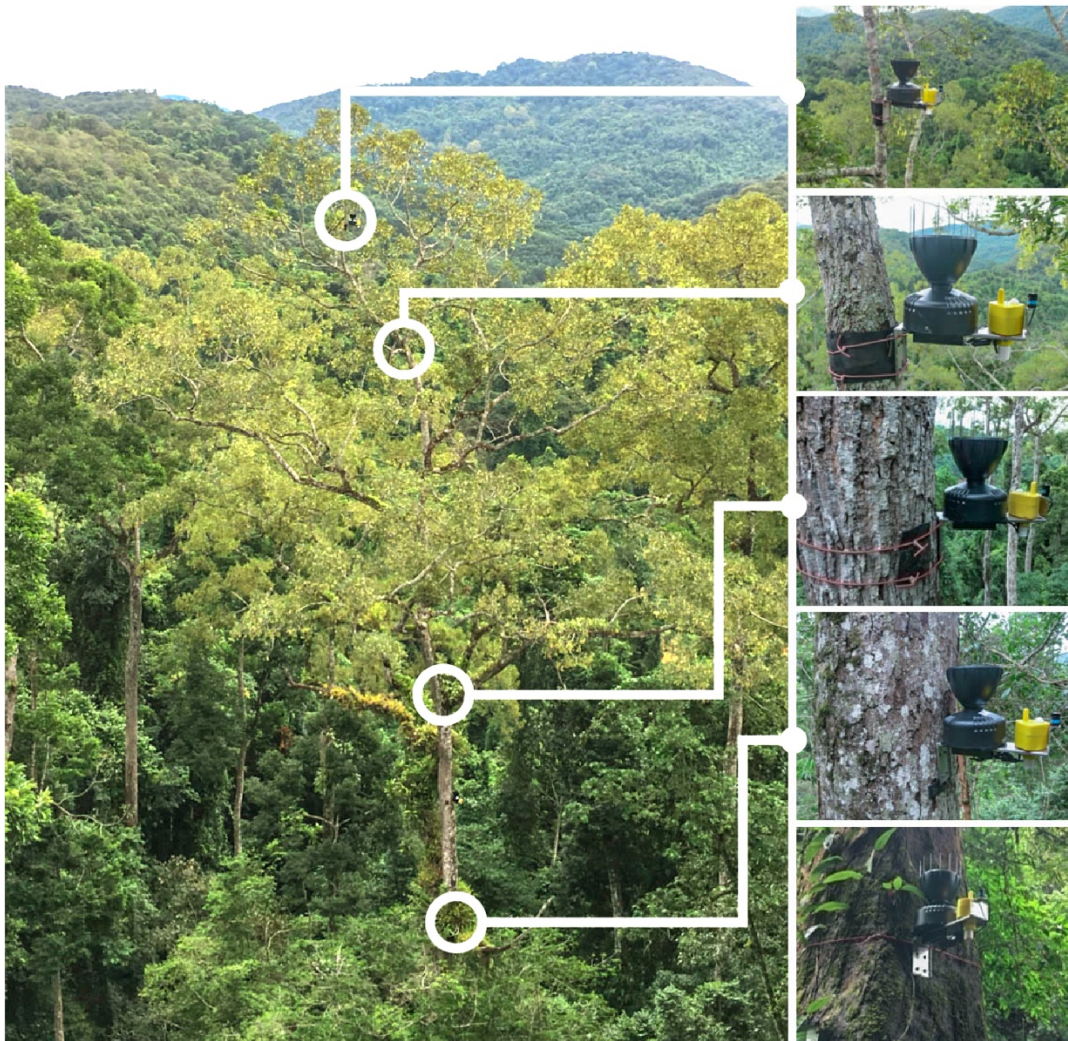
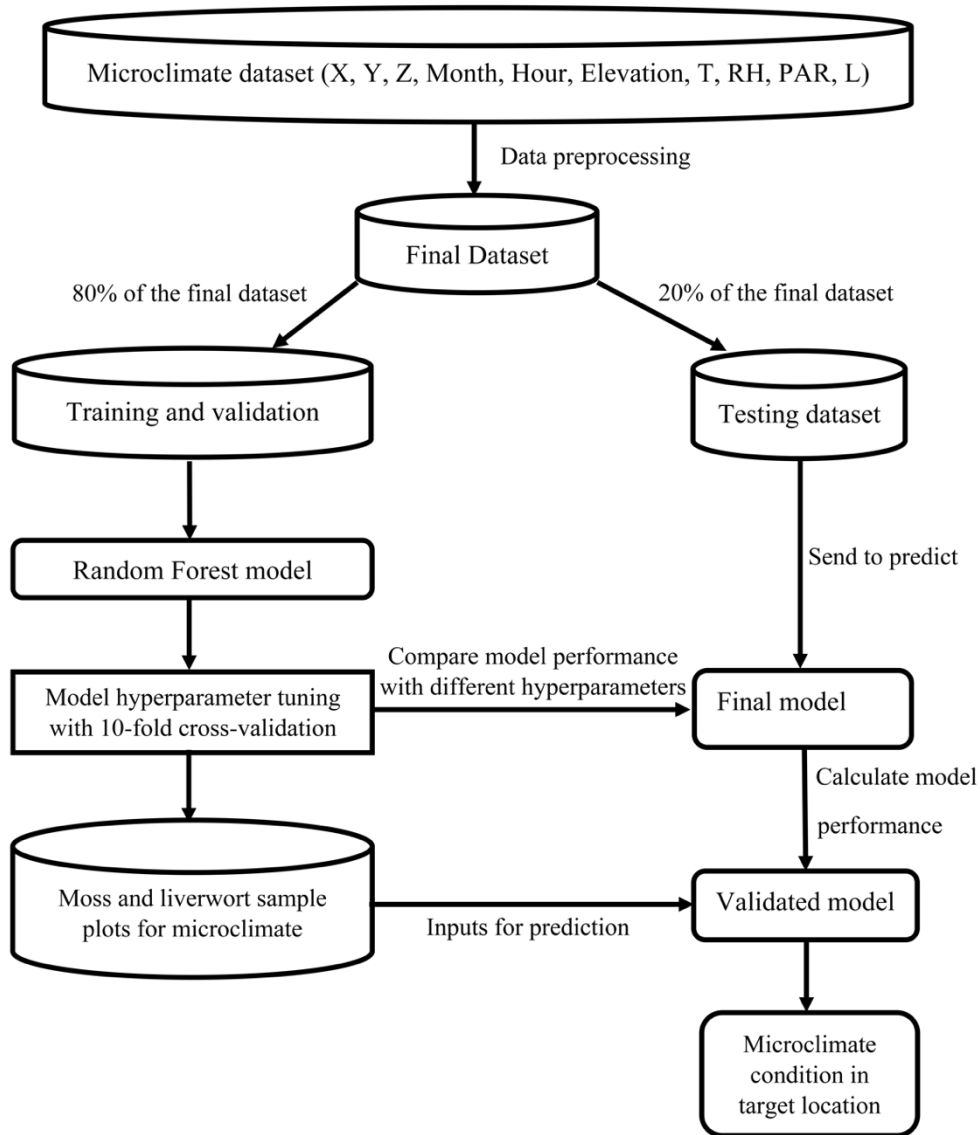


# 1 Supporting information



2

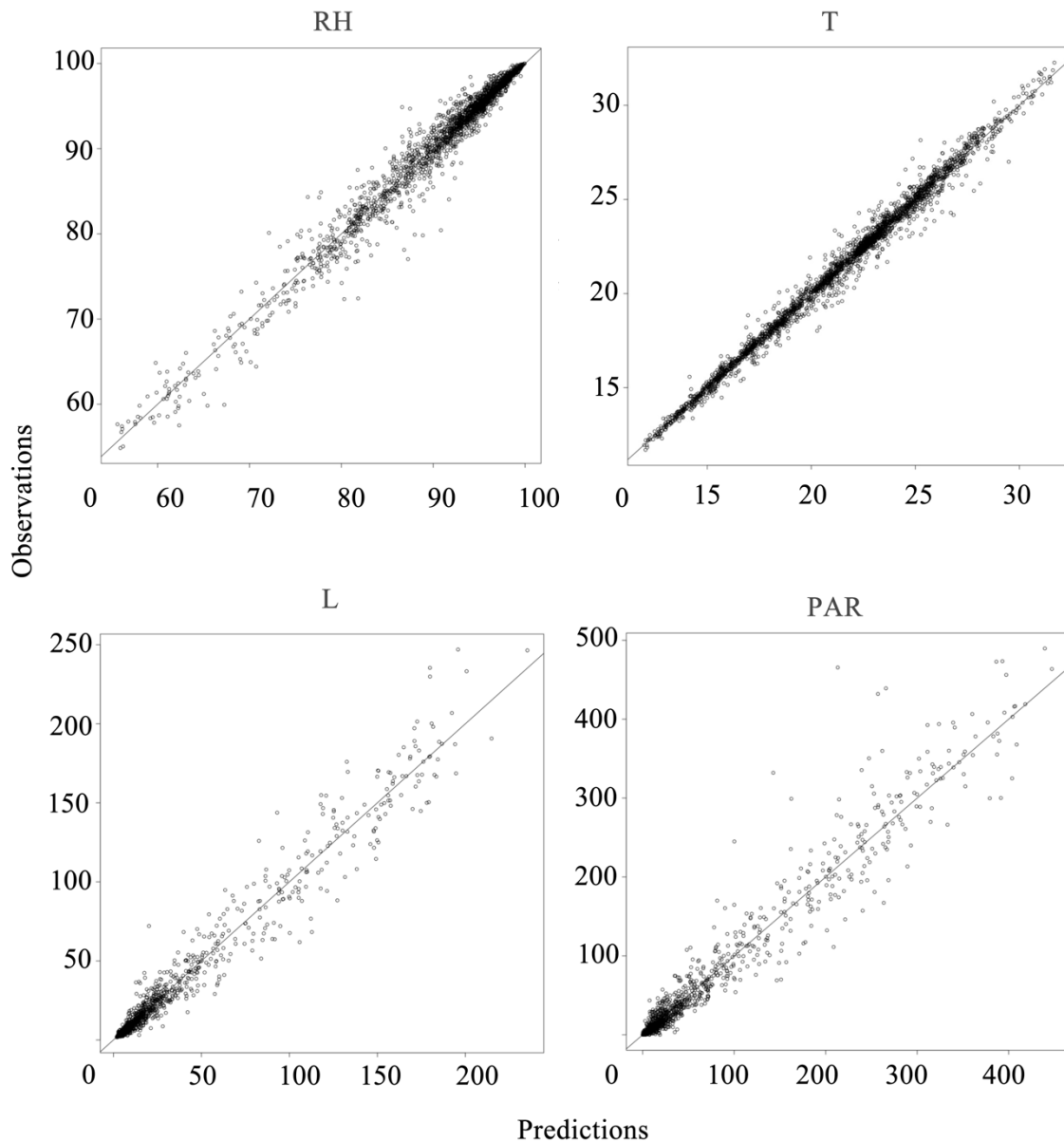
3 FIGURE S1 Data loggers used for recording light and microclimate conditions at the  
4 Xishuangbanna canopy crane facility, Yunnan, SW China. Temperature, relative humidity,  
5 photosynthetically active radiation and light intensity were recorded by 54 dataloggers on 12  
6 individual trees at five height zones (1: tree base, 2: middle trunk, 3: inner canopy, 4: middle  
7 canopy, 5: outer canopy, modified from Johansson, 1974) every hour during 30 months  
8 between 2017 and 2019.



9

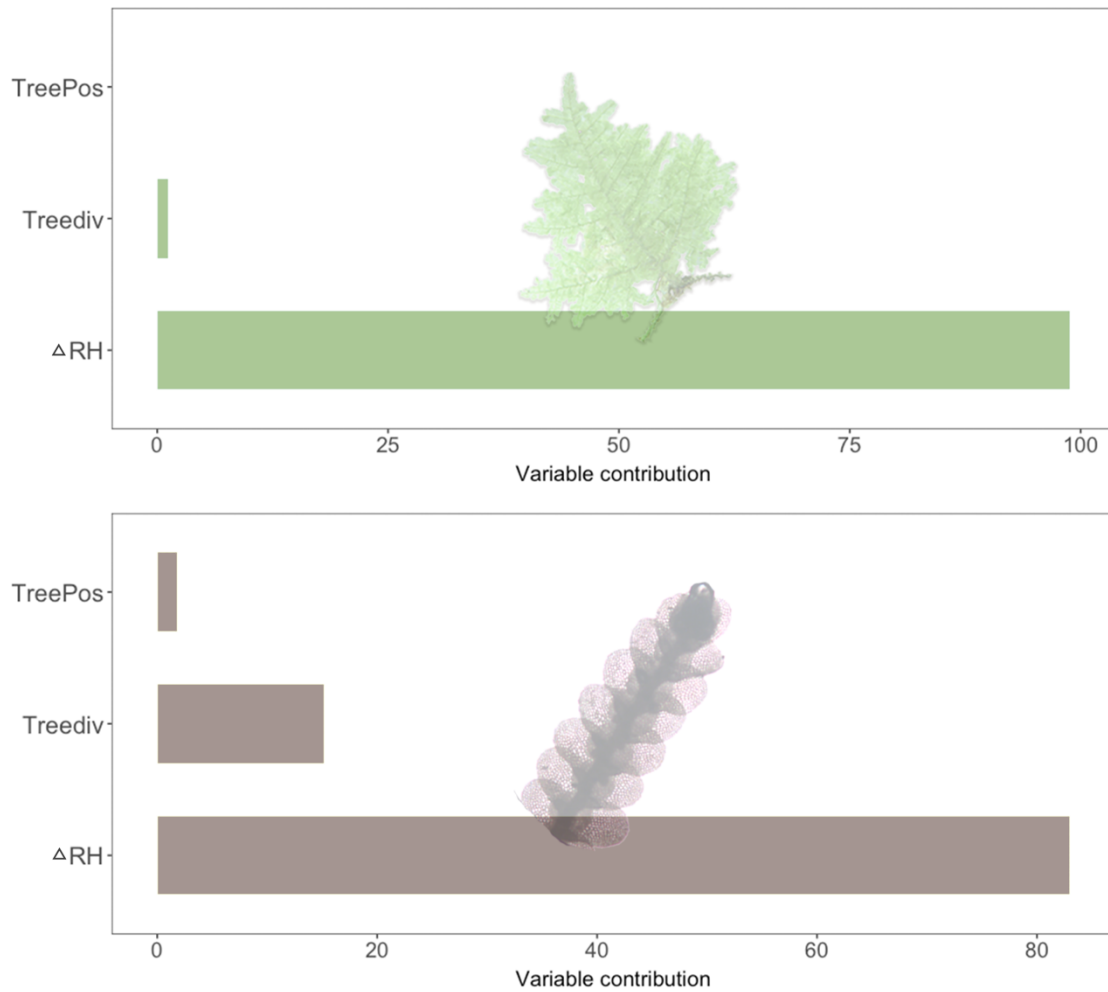
10 FIGURE S2 Flow chart of the protocol employed to model microclimatic variation using

11 Random Forest.



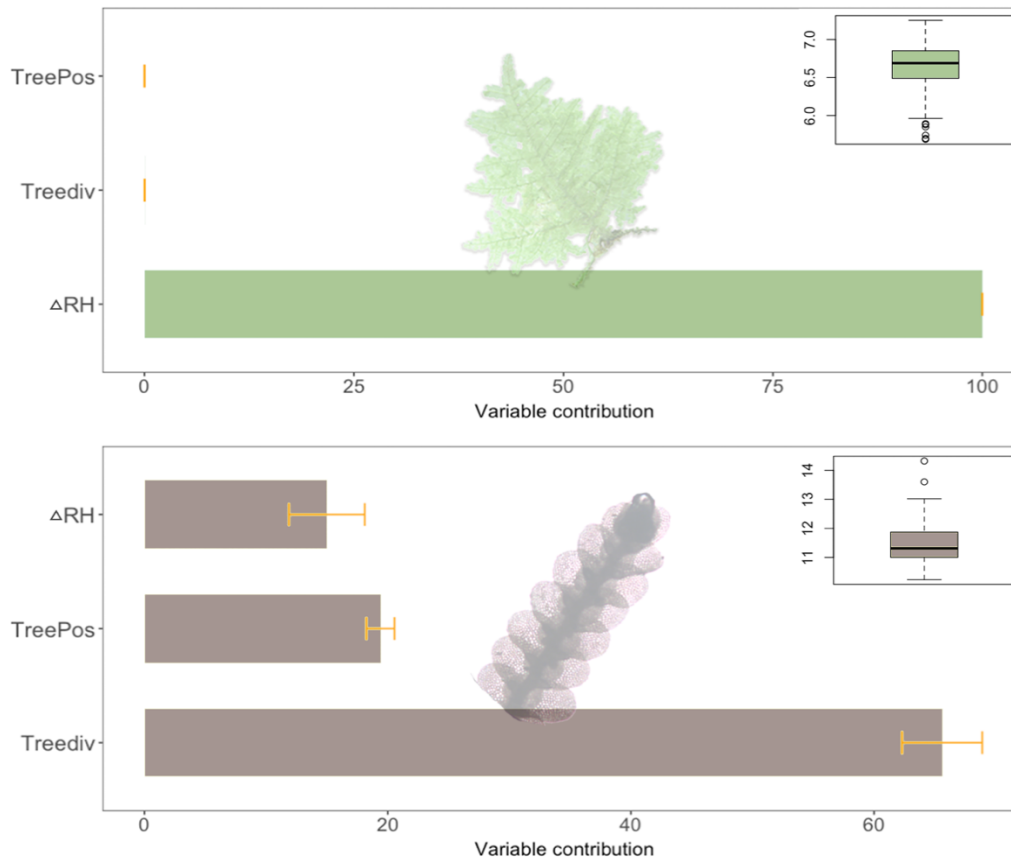
12

13 FIGURE S3 Observed vs predicted values of temperature (T), relative humidity (RH),  
 14 photosynthetically active radiation (PAR), and light intensity (L) from Random Forest  
 15 models in a 1.44 ha lowland dipterocarp forest at the Xishuangbanna canopy crane facility,  
 16 Yunnan, SW China. Hourly variation in L, PAR, RH, and T, recorded by 54 dataloggers on  
 17 12 trees during 30 months between 2017 and 2019, was modelled by Random Forest using  
 18 height on the tree, topography (X-Y coordinates and elevation), hour and month of the record  
 19 as predictors.



20

21 FIGURE S4 Importance (in % explained deviance) of the factors used to describe the vertical  
 22 variation in species turnover among epiphytic moss (green) and liverwort (brown)  
 23 communities by Generalized Dissimilarity Modelling. The percentage values illustrate the  
 24 contribution of the variables remaining in the model after backward selection. ΔRH:  
 25 difference in relative humidity among plots; TreePos: Position of each individual tree in the  
 26 x-y space (distance to a reference point); Treediv: variable indicating whether two plots being  
 27 compared are both located on the trunk or in the canopy, or in different parts of the tree.



28

29 FIGURE S5 Importance (in % explained deviance) of the factors used to describe the vertical

30 variation in phylogenetic turnover among epiphytic moss (green) and liverwort (brown)

31 communities by Generalized Dissimilarity Modelling. The main panel illustrates the

32 contribution (in %) of the variables remaining in the model after backward selection. The

33 upper right panels summarize how the percentage deviance explained by the GDM varies

34 across 100 trees with randomly resolved relationships among congeneric species using box-

35 plots (showing the 1st and 3rd quartiles (upper and lower bounds), 2nd quartile (center),

36 average (red dots), 1.5\* interquartile range (whiskers) and minima-maxima beyond the

37 whiskers).  $\Delta$ RH: difference in relative humidity among plots; TreePos: Position of each

38 individual tree in the x-y space (distance to a reference point); Treediv: variable indicating

39 whether two plots being compared are both located on the trunk or in the canopy, or in

40 different parts of the tree.

41

42 **Tables**

43 Table S1 Distribution of the dataloggers used to model microclimatic. 54 dataloggers were  
 44 installed on 12 trees, for which the ID number, species identity, elevation, and coordinates  
 45 (X-Y) are provided. Z: height of a datalogger on the tree.

ID	Host tree	Elevation	X	Y	Z
					2
					6.1
TD43-002	<i>Garcinia cowa</i>	658.65	64.9	48	8.66
					11.5
					15
<hr/>					
					2
					5.8
TD43-078	<i>Garcinia cowa</i>	660.28	77.8	52.8	10.47
					12.7
					16
<hr/>					
					2
					5.7
TD23-025	<i>Garcinia cowa</i>	651.27	22.2	47	9
					10.9
					13.4
<hr/>					
					2
					30.55
TD32-021	<i>Parashorea chinensis</i>	672.51	44	22.5	40.4
					54.3
					58.7
<hr/>					
					2
					30.6
TD53-111	<i>Parashorea chinensis</i>	666.21	89.7	54.6	41.3
					49.88

					57.8
					2
					37.2
TD46-023	<i>Parashorea chinensis</i>	693.95	68	105.7	50.05
					56.16
					61.34
TD43-066	<i>Pometia pinnata</i>	655.69	79	59.6	2
					2
					12.65
TD36-013	<i>Pometia pinnata</i>	682.75	43.5	103.2	19.66
					23.58
					27.54
					2
					9.8
TD65-029	<i>Pometia pinnata</i>	675.5	100.5	88	16.48
					20.73
					24.3
					2
					15.4
TD44-036	<i>Sloanea tomentosa</i>	668.41	61.5	77.6	28.83
					31.83
					2
					15.86
TD53-034	<i>Sloanea tomentosa</i>	660.28	84	45.3	23.4
					29.35
					2
					20.56
TD52-035	<i>Sloanea tomentosa</i>	683.46	86.8	28.7	32.57
					38.9

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47



48 Table S2 Moss genera not included in the phylogeny of Laenen et al. (2014) and re-assigned  
 49 to their closest genus based on phylogenetic evidence.

Genus missing in Laenen et al. 2014	Re-assigned genus	References
<i>Circulifolium</i>	<i>Himantocladium</i>	Olsson et al., 2016
<i>Caduciella</i>	<i>Himantocladium</i>	Olsson et al., 2016
<i>Haplohymenium</i>	<i>Herpetineuron</i>	Zhang et al., 2003
<i>Leucophanes</i>	<i>Syrrhopodon</i>	Pereira et al., 2019
<i>Macromitrium</i>	<i>Schlotheimia</i>	Li et al., 2013
<i>Nerkeropsis</i>	<i>Himantocladium</i>	Olsson et al., 2016
<i>Cyrto-hypnum</i>	<i>Thuidium</i>	Huttunen et al., 2012
<i>Entodontopsis</i>	<i>Pilosium</i>	Buck et al., 2000
<i>Pelekium</i>	<i>Thuidium</i>	Huttunen et al., 2012
<i>Calymperes</i>	<i>Syrrhopodon</i>	Pereira et al., 2019
<i>Aerobryidium</i>	<i>Meteoriopsis</i>	Huttunen et al., 2004
<i>Racopilum</i>	<i>Braithwaitea</i>	Liu et al., 2019

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 51  
 52  
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 54

55 Table S3 Setting and performance of microclimatic models in a 1.44 ha lowland dipterocarp  
 56 forest at the Xishuangbanna canopy crane facility, Yunnan, SW China. Variation in  
 57 temperature (T), relative humidity (RH), photosynthetically active radiation (PAR), and light  
 58 intensity (L) was predicted from Random Forest models calibrated from hourly records  
 59 between 2017 and 2019 of those variables by dataloggers installed on 12 trees. RMSE: Root  
 60 Mean Square Error; mtry: number of variables available for splitting at each tree node; ntree:  
 61 number of trees to grow.  
 62

Microclimatic variables	R <sup>2</sup>	RMSE	Mtry	Ntree
T	0.99	0.41	4	600
RH	0.97	1.43	6	100
L	0.97	6.06	5	1000
PAR	0.96	14.83	6	100

63

64

65 Table S4 Mosses and liverworts recorded on 42 *Parashorea chinensis* in a 1.44 ha lowland  
 66 dipterocarp forest at the Xishuangbanna canopy crane facility, Yunnan, SW China.

Mosses (50)	Liverworts (52)
<i>Aerobryidium aureo-nitens</i> (Schwaegr.) Broth.	<i>Acrolejeunea recurvata</i> Gradst.
<i>Aerobryopsis cochlearifolia</i> Dixon	<i>Caudalejeunea reniloba</i> (Gottsche) Steph.
<i>Brotherella nictans</i> (Mitt.) Broth.	<i>Cheilolejeunea eximia</i> P.C.Chen & P.C.Wu
<i>Caduciella mariei</i> (Besch) Enroth	<i>Cheilolejeunea intertexta</i> (Lindenb.) Steph.
<i>Calymperes erosum</i> Müll. Hal.	<i>Cheilolejeunea obtusilobula</i> (S.Hatt.) S.Hatt.
<i>Calypothecium phyllogonoides</i> Nog. & X.J.Li	<i>Cheilolejeunea ventricosa</i> (Schiffn. ex P.Syd.) X.L.He
<i>Calypothecium wightii</i> (Mitt.) Fleisch.	<i>Cheilolejeunea vittata</i> (Steph. ex G. Hoffm.) R.M.Schust. & Kachroo
<i>Circulifolium microdendron</i> (Mont.) M.Fleisch.	<i>Cheilolejeunea xanthocarpa</i> (Lehm. & Lindenb.) Malombe
<i>Claopodium prionophyllum</i> (Müll. Hal.) Broth.	<i>Cololejeunea appressa</i> (A.Evans) Benedix
<i>Cryphaea obovatocarpa</i> S.Okamura	<i>Cololejeunea descicens</i> Steph.
<i>Cyrto-hypnum gratum</i> (P.Beauv.) W.R.Buck & H.Crum	<i>Cololejeunea dinghuiana</i> R.L.Zhu & Y.F.Wang
<i>Entodontopsis wightii</i> (Mitt.) W.R.Buck & R.R.Ireland	<i>Cololejeunea indocinica</i> Tixier
<i>Erythrodontium julaceum</i> (Hook. ex Schwägr.) Paris	<i>Cololejeunea inflata</i> Steph.
<i>Fissidens hollianus</i> Dozy & Molk.	<i>Cololejeunea lanciloba</i> Steph.
<i>Fissidens linearis</i> var. <i>obscurirete</i> (Broth. & Par.) Stone	<i>Cololejeunea latilobula</i> (Herzog) Tixier
<i>Floribundaria pseudofloribunda</i> M.Fleisch.	<i>Cololejeunea peraffinis</i> (Schiffn.) Schiffn.
<i>Gollania tereticaulis</i> (Besch.) Broth.	<i>Cololejeunea planissima</i> (Mitt.) Abeyw.
<i>Groutiella tomentosa</i> (Hornsch.) Wijk & Margad	<i>Cololejeunea platyneura</i> (Spruce) A.Evans
<i>Haplohymenium triste</i> (Ces.) Kindb.	<i>Cololejeunea raduliloba</i> Steph.
<i>Homaliadelphus targionianus</i> (Mitt.) Dixon & P. de la Varde	<i>Cololejeunea subfloccosa</i> Mizut.
<i>Hypopterygium</i> sp.	<i>Cololejeunea tenella</i> Benedix
<i>Leucobryum chlorophyllosum</i> Müll. Hal.	<i>Cololejeunea trichomanis</i> Gottsche ex Steph.
<i>Leucophanes octoblepharioides</i> Brid.	<i>Frullania ericoides</i> (Nees ex Mart.) Mont.
<i>Mesonodon flavescens</i> (Hook.) W.R.Buck.	<i>Frullania monocera</i> (Taylor) Gottsche, Lindenb. & Nees
<i>Meteoriopsis reclinata</i> (Müll. Hal.) M.Fleisch.	<i>Frullania orientalis</i> S.Lac.
<i>Meteoriopsis</i> sp.	<i>Frullania polyptera</i> Taylor

*Meteorium polytrichum* Dozy & Molk.  
*Meteorium subpolytrichum* (Besch.) Broth.  
*Nerkeropsis crinita* (Griff.) M.Fleisch.  
*Pelekium bifarium* (Bosch et Lac.) Fleisch.  
*Pinnatella ambigua* (Bosch. & Lac.) Fleisch.  
*Pylaisiadelphina yokohamae* (Broth.)  
*Racopilum orthocarpum* Wilson ex Mitt.  
*Rhynchostegiella japonica* Dixon & Thér.  
*Rhynchostegium fauriei* Card.  
*Rhynchostegium* sp.1  
*Rhynchostegium* sp.2  
*Sematophyllum phoeniceum* (Müll. Hal.) M.Fleisch.  
*Sematophyllum subhumile* (Müll. Hal.) M.Fleisch.  
*Sematophyllum subpinnatum* (Brid.) E.Britton  
*Symphyodon perrottetii* Mont.  
*Syrhopodon parasiticus* (Brid.) Besch.  
*Taxiphyllum cuspidifolium* (Cardot) Z.Iwats.  
*Taxithelium oblongifolium* (Sull. & Lesq.) Z.Iwats.  
 Sp.1  
 Sp.2  
 Sp.3  
 Sp.4  
 Sp.5  
 Sp.6  
*Frullania riojaneirensis* (Raddi)  
 Spruce  
*Heteroscyphus argutus* (Schiffn.)  
 Schiffn.  
*Lejeunea anisophylla* Mont.  
*Lejeunea cocoes* Mitt.  
*Lejeunea flava* (Sw.) Ness  
*Lejeunea pallidevirens* S.Hatt.  
*Lejeunea sordida* (Nees) Nees  
*Lejeunea tuberculosa* Steph.  
*Leptolejeunea dapitana* Steph.  
*Leptolejeunea subacuta* Steph. ex  
 A.Evans  
*Lopholejeunea subfusca* (Nees)  
 Schiffn.  
*Lopholejeunea sikkimensis* Steph.  
*Metzgeria conjugata* Lindb.  
*Plagiochila fordiana* Steph.  
*Plagiochila furcifolia* Mitt.  
*Plagiochila gracilis* Lindenb. &  
 Gottsche  
*Plagiochila parviramifera* Inoue  
*Plagiochila trabeculata* Steph.  
*Plagiochila wightii* Nees ex Lindenb.  
*Porella* sp.  
*Ptychanthus striatus* (Lehm. &  
 Lindenb.) Nees  
*Radula retroflexa* Taylor  
*Spruceanthus planiusculus* (Mitt.)  
 X.Q.Shi, R.L.Zhu & Gradst.  
*Mastigolejeunea humilis* (Gott.)  
 Schiffn.  
*Thysananthus repletus* (Taylor)  
 Sukkharak & Gradst.  
*Tuzibeanthus chinensis* (Steph.)  
 Mizut.

68 Table S5 Generalized Dissimilarity Modeling of the horizontal variation (among trees within the same height zone) in turnover ( $\beta_{sim}$ ),  
69 nestedness ( $\beta_{sne}$ ) and phylogenetic turnover ( $\pi_{st}$ ) among epiphytic moss and liverwort communities. The table indicates, for each of the six  
70 height zones (Z1–Z6), the percent deviance explained by the models and the the relative contribution (in %) of the variables remaining in the  
71 models after backward selection. GeoDist: geographic distance among trees;  $\Delta DBH$ : difference in DBH among trees;  $\Delta Elev$ : difference in  
72 elevation among trees;  $\Delta T$ ,  $\Delta RH$ ,  $\Delta L$ ,  $\Delta PAR$ : differences in temperature, relative humidity, light intensity and PAR among plots. For  
73 phylogenetic turnover, the percent values indicate that different models were selected across the 100 phylogenetic trees and indicate the  
74 proportion of trees supporting a given model. At height zone 4, model selection varied among the 100 randomly resolved phylogenetic trees: in  
75 14% of the trees, selected models included GeoDist,  $\Delta DBH$  and  $\Delta T$ ; 35% of the models included GeoDist and  $\Delta T$ ; and the remaining models  
76 were not significant. ‘-’ indicates that no significant model was obtained for the considered height zone. Results for moss  $\beta_{sne}$  and liverwort  $\pi_{st}$   
77 are not displayed, because no significant model for moss  $\beta_{sne}$  was found at any height zone, and liverwort  $\pi_{st}$  is phylogenetic overdispersion  
78 ( $\pi_{st} < 0$ ).

Groups	Beta diversity	Variables	Z1		Z2		Z3		Z4		Z5		Z6	
			Percent Deviance Explained	Variable contribution	Percent Deviance Explained	Variable Contribution	Percent Deviance Explained	Variable contribution	Percent Deviance Explained	Variable contribution	Percent Deviance Explained	Variable contribution	Percent Deviance Explained	Variable contribution
Mosses	$\beta_{sim}$	GeoDist		1.8		-		0.3		-				1
		$\Delta DBH$	5.90%	68.8		-	7.50%	98.7		-			44.50%	99
		$\Delta Elev$		-		-		-		-				-



$\Delta RH$	-	-	-	-	-	-	-
$\Delta L$	-	-	-	-	-	-	-
$\Delta PAR$	-	-	-	-	-	-	-

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80 Table S6 P-values of community-level average  $\pi_{st}$  in epiphytic moss and liverwort  
81 communities in a lowland dipterocarp forest (Xishuangbanna, Yunnan, SW China) per DBH  
82 class (small, medium, large) and height zone (Z1 – Z6). Significant values are highlighted in  
83 bold. P-values of the significance test are given in parentheses, with ‘<’ and ‘>’ indicating  
84 whether the observed value is lower or higher than expected by chance.

DBH class	Height zone	Mosses $\pi_{st}$ (p-value)	Liverworts $\pi_{st}$ (p-value)
Small	1	0.16 (> 0.000)	-0.05 (< 0.22)
	2	<b>-0.07</b> (< 0.03)	<b>-0.07</b> (< 0.04)
	3	0.05 (0.84)	-0.01 (> 0.43)
	4	-	0.01 (0.57)
	5	-	0.01(0.77)
	6	-	-0.05 (0.19)
Medium	1	0.10 (1)	0.12 (0.92)
	2	<b>0.23</b> (> 0.03)	0.00 (0.44)
	3	-0.02 (0.31)	0.03 (0.76)
	4	0.04 (0.77)	-0.04 (0.2)
	5	0.00 (0.74)	0.02 (0.76)
	6	-	0.03 (0.77)
Large	1	0.04 (0.97)	-0.46 (< 0.07)
	2	0.01 (0.74)	-0.16 (< 0.07)
	3	-0.01 (0.29)	-0.02 (0.33)
	4	-0.05 (0.15)	-0.07 (0.38)
	5	-0.11 (< 0.12)	0.16 (0.91)
	6	<b>-0.12</b> (< 0.03)	-0.02 (0.33)

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