UNIVERSITY OF CALIFORNIA, DAVIS Public Strawberry breeding program

New genomic tools for studying cultivated and wild octoploid strawberry



Roadmap Forward...

UC Davis Strawberry Breeding Program





Current Focus:

- Germplasm knowledge Octoploid genomics tools
- Building foundation for genomic breeding

Future Goals:

- Increased fruit quality
- Genomic prediction
 - Leveraging untapped diversity to improve flavor

UC Davis Strawberry Breeding Program

1290

81100

790

500

The last 30 years...

 Successful focus on yield and marketable
 quality for California field production Day neutral breeding has increased potential

important trait for growers

- Virtually no introduction of new g
- program Limited use of genomic tools and
- breeding process



A short history of strawberry genomic breeding resources

2011 - Fragaria vesca genome assembly (Shulaev et al) diploid progenitor genome, translational system for non-climacteric fruit genetic research

Wide variety of published marker genotyping strategies – SSRs, target capture, DArtSeq, ddRADseq

- 2014 First pass at an octoploid assembly (Hirakawa et al) highly fragmented
- 2015 Affymetrix iStraw90 Axiom array (Bassil et al) diploid-segregating markers iStraw35 spin-off, high quality subset



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2016-present - push for a high-quality octoploid genome

collaboration at UC Davis, Michigan State

multiple other groups

Challenges for Octoploids

Total subgenomic ambiguity, fails to account for inter-species divergence and gene content across subgenomes

Low quality





PacBio Assembly – Finding the Right **Parameters**

Primary goal for assembly: Zero subgenome collapsing with haploid-level subgenomes

How can we estimate and avoid subgenome collapses during the assembly process?

Approach 1. How many contigs stack up against the F. vesca genome?

and 2. How many contine align to concentral single conv. C



proach 2. How many contigs	aligh to conserved single-copy Fragana vesca genes?	
F. vesca chromosome F. vesca SCG PB contigs		

Building Resources for Molecular Breeding



PacBio Assembly - Finding the Right **Parameters**

Key Parameters:

read overlap – aligner thresholds for error-correction, pre-assembly of longreads



relaxed = 2500, 2000

length cutoff – minimum length of input reads for pre-assembly, final assembly

inclusion of alternate contigs

Strict-Overlap Relaxed Overlap Strategy: Prioritize haplotype separation to minimize intersubgenome collapses, address within-subgenome haplotype separation downstream (goal=haploid)





Edger et al. (2019) Origin and Evolution of the Octoploid Strawberry Genome

P. Edge^{nb3}, Thomas J. Poorten⁷, Robert VamBuren¹⁴, Michael A. Hardigan⁷, Marri Coll, R. McKain⁸, Ronald D. Smith, Scott Terezi, Andree D. L. Netson, Chron Jam Wal⁸, Elist Kein A. Bird⁸, Nan E. Yocca⁸, Nathan Pumplin⁶, Shujun Ou¹⁴, Gill Ben-Ze⁸, Artal Brod Thomas Swale¹, Lij Shiuel, Charlotte B. Acharya⁷, Giom S. Cole⁶, Jeffrey P. Moverk, H. Ning Jiang¹⁸, Enc Lyons⁸, Michael Freeling¹⁹, Joshua R. Puzey¹, and Steven J. Knapp⁻¹

- Childs¹, Ving Jiangh², Eric Ljoner, Michael Freeling¹, Johna R. Puzey¹, and Serien J. Krappi¹-Bogentreic of Horizonian. Methops State University, East Landing, MU, GA. 5. Ecology, Teolutionay Biology and Behavior, Michigan State University, East Landing, MU, GA. 5. Dapameter GHan States and State University, East Landing, MU, GA. 1. Part Relineare Instands, Michigan State University, East Landing, MU, GA. 5. School, T. States, Stat

PacBio Assembly – Dovetail HiC Scaffolding

Dovetail scaffolding of Falcon assembly improved





NRGene and PacBio Genome – Assembly Metrics romosome-Scale Contiguity in Both Assemblie® 28 Pseude ed Gen ne Lengths Close to licted G me Size Pace 1 805,488,70 806,643, PacBio + 306.592. NRGe

nificantly Longer Contigs & Fewe Gaps in the PACBIO Assembly

Signific

New Challenge: Reducing Assemblies to Haploid Level without Collapsing Subgenomes

NRGene and PacBio Genome -Genes and Repetitive Elements





Constructing a Haploid Assembly





California Beach Strawberry Genome

Fragaria chiloensis spp. lucida (Del Norte)

- Native California beach strawberry
- Characteristics
- High salt tolerance
 Small soft fruit with unique aroma
- Pathogen resistance
- · Genomic and breeding data collected
 - 70X PacBio SEQUEL data → <u>NEW assembly</u>
 160X PE 2x250 Illumina data
 - Mapping population (Camarosa x Del Norte)

 - 200 progeny
 Illumina sequenced
 - Phenotypesd for multiple traits



WGS-based Octoploid Mapping

- F. chiloensis mapping with Illumina data:
- 28 chromosome-scale linkage groups
 >2 million genome+map anchored markers
- >2 million genome+map anchored markets
 High-confidence chromosome scaffolding
- mapping multiple traits of interest





Challenge for subgenome-specific alignment?



 145 octoploids at present
 Illumina sequence (PE 150nt)
 Used to identify genomic variants for SNP array
 Magned to NPGene

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•	High	rates	of s	subg	enom



F. chiloensis genetic map shows high concordance with F. x ananassa genome





Octoploid Sequence Diversity Panel

Assessing genetic diversity of the octoploid genome

Selection Targets:

- 1) Capture UC Davis germplasm diversity
- Capture alleles from founders, high pedigree contribution heirlooms
- Capture progenitor species diversity





Wild vs. Domesticated Population Divergence

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					Position (Mh)			



Increased Likelihood of Subgenome Specific Probe Hybridization (HomRO >= 0.30)



78.5% of Polymorphic SNPs in Screening Panel

Development of Marker Array Panel



Overview: Strawberry SNP Selection

- PHASE 1: Identify and select candidate marker probes basedon variant calling in sequenced cotpolidpanel oppulated 850K screening array Selection priority in/around genes (600K) genome coverage (250K) alled elversity in domest (cat dgermplasm low probe interference
- ✓ PHASE 2: Test probes on Affymetrix screening array
 - 446,644 AxiomQC-passing polymorphic markers
 78.5% polymorphichi-resolution (PHR)
 12,624 passing IStraw35 markers (>10K anchored)
- PHASE 3: Select markers to populate production array
 49K probe capacity
 Numerous criteria: technical quality, diversity, coverage

- Marker Analysis Areas of Interest Genetic Diversity Heterozygosity, allelic diversity across cultivated sub-populations
- Mapping quality
 2 mapping populations on screening array (n=48)
- Major QTL regions represented
- Candidate genes
 Fruit texture and quality
 Volatile biosynthesis
 Hormone-mediated fruit development
 Flowering and maturity
 R genes
- Physical genome coverage

Genetic Diversity Across Programs



Marker Genome Distribution





Early Array Applications

Genetic Mapping (49K array)



Camarosa (maternal) map F. x ananassa ~11k QC-passing het x hom SNPs

~1.8k QC-passing het x het SNPs

Del Norte (paternal) map *F. chiloensis* (wild) ~2.1k QC-passing hom x het SNPs

Octoploid Germplasm Diversity



Conversion to Single Marker Assays for Fine Mapping and Breeding

Fw1 Fusarium Resistance

850K array GWAS identified Fvb2-4 locus
Subgenome-specific KASP assays flanking Fw1



Nicolas Cobo







Strawberry Maturity Locus





Strawberry Maturity Locus



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UCDAVIS DEPARTMENT of PLANT SCIENC College of Agricultural and Environmental Scien)		

Strawberry Maturity Locus





Future work: • Conversion of 850K panel markers to KASP assays • Multiple segregating day neutral populations for fine mapping (2019) THANKS FOR LISTENING!





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