

Review Article

Medicinal Plants of *Solanum* Species: The Promising Sources of Phyto-Insecticidal Compounds

Kumarappan Chidambaram ¹, **Taha Alqahtani** ¹, **Yahia Alghazwani**¹, **Afaf Aldahish**¹, **Sivakumar Annadurai**², **Kumar Venkatesan**³, **Kavitha Dhandapani**⁴, **Ellappan Thilagam**⁵, **Krishnaraju Venkatesan** ¹, **Premalatha Paulsamy**⁶, **Rajalakshimi Vasudevan**¹ and **Geetha Kandasamy**⁷

¹Department of Pharmacology and Toxicology, College of Pharmacy, King Khalid University, Al-Qara, Abha 61421, Saudi Arabia

²Department of Pharmacognosy, College of Pharmacy, King Khalid University, Al-Qara, Abha, Saudi Arabia

³Department of Pharmaceutical Chemistry, College of Pharmacy, King Khalid University, A-Qara, Abha, Saudi Arabia

⁴Department of Biochemistry, Biotechnology and Bioinformatics, Avinashilingam Institute for Home Science and Higher Education for Women, Coimbatore 641043, Tamil Nadu, India

⁵Department of Pharmacognosy, JKKMMRF's Annai JKK Sampoorani Ammal College of Pharmacy, Namakkal 638183, Tamilnadu, Tamil Nadu 638183, India

⁶Faculty of Nursing, King Khalid University, Abha 61421, Saudi Arabia

⁷Department of Clinical Pharmacy, College of Pharmacy, King Khalid University, Abha 61421, Saudi Arabia

Correspondence should be addressed to Kumarappan Chidambaram; ctkumarrx@gmail.com

Received 22 November 2021; Revised 14 May 2022; Accepted 31 July 2022; Published 21 September 2022

Academic Editor: Mohammed Bourhia

Copyright © 2022 Kumarappan Chidambaram et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Several medicinal plants have the potential to be a promising alternative pharmacological therapy for a variety of human illnesses. Many insects, including mosquitoes, are important vectors of deadly pathogens and parasites, which in the world's growing human and animal populations can cause serious epidemics and pandemics. Medicinal plants continue to provide a large library of phytochemicals, which can be used to replace chemically synthesized insecticides, and utilization of herbal product-based insecticides is one of the best and safest alternatives for mosquito control. Identifying new effective phyto-derived insecticides is important to counter increasing insect resistance to synthetic compounds and provide a safer environment. *Solanum* genus (Solanaceae family or nightshades) comprises more than 2500 species, which are widely used as food and traditional medicine. All research publications on insecticidal properties of Solanaceae plants and their phytoconstituents against mosquitoes and other insects published up to July 2020 were systematically analyzed through PubMed/MEDLINE, Scopus, EBSCO, Europe PMC, and Google Scholar databases, with focus on species containing active phytoconstituents that are biodegradable and environmentally safe. The current state of knowledge on larvicidal plants of *Solanum* species, type of extracts, target insect species, type of effects, name of inhibiting bioactive compounds, and their lethal doses (LC₅₀ and LC₉₀) were reviewed in this study. These studies provide valuable information about the activity of various species of *Solanum* and their phytochemical diversity, as well as a roadmap for optimizing select compounds for botanical repellents against a variety of vectors that cause debilitating and life-threatening human diseases.

1. Introduction

Medicinal plants are traditionally used to treat numerous human infections, and their bioactive compounds have long

been important in therapeutic development, particularly in cancer and infectious diseases. Medicinal plant-derived natural products have garnered much interest in recent years as potential bioactive agents for insect vector control. Vector

control is threatened by the emergence of resistance to conventional synthetic insecticides in vectors, among which mosquitoes pose high threats to human and animal health and life, often leading to the transmission of serious diseases, such as dengue, Ebola, filariasis, and malaria, resulting in millions of deaths each year [1–4]. Because chemical control of mosquitoes has been linked to such detrimental outcomes as the development of insect resistance, it is urgently necessary to discover and develop reliable and environmentally sustainable alternatives to current synthetic chemical insecticides.

As an alternative to synthetic insecticides, plant-based insecticide preparations have the advantages of rapid biodegradability and low toxicity to humans and animals [5]. Several plants and their constituents, especially those in medicinal herbs, have been traditionally used as insecticides, due to being rich in various bioactive phytochemicals and providing potential sources of natural mosquito control agents [6–9]. Recently, attention has been given to preparations of mosquito-larvicidal compounds based on herbal origin to enhance insecticidal effects and reduce the probability of development of resistance by the target pest population [10, 11]. While several plants from different families have been reported with mosquito-larvicidal properties, only a few species show promising effects and could be developed into natural insecticidal agents [12].

The *Solanum* family of plants is a large genus within the Solanaceae family that contains up to 2,000 species ranging from food crops to medicinal herbs. The genus *Solanum* has received much interest in chemical and biological studies over the last 30 years. Several steroidal saponins, steroidal alkaloids, disaccharides, flavonoids, and phenols have been implicated in the biological activities [13]. The genus *Solanum* appears to have a lot of potential, although most of the species are unknown or have had little research on their chemical contents. Several reviews of the *Solanum* genus and their phytochemistry have been published. These compounds have been linked to various health-promoting activities in the fight against several noncommunicable diseases, which are the leading causes of death worldwide. Many species belonging to this genus present a huge range of pharmacological activities such as anticancer, hepatoprotective, antimalarial, anthelmintic, and other activities [14]. Plants in this family are recognized for having a wide spectrum of alkaloid compounds, some of which are therapeutically the most potent. Steroidal glycoalkaloids are the most common and important group of nitrogen-containing secondary metabolites identified in Solanaceae plants. More than 350 *Solanum* species have yielded more than 100 different forms of glycoalkaloids [15, 16]. Many medicinal plants belonging to the Solanaceae family are promising therapeutic candidates to develop as bioinsecticides against vector-borne human diseases such as malaria, leishmaniasis, and dengue fever due to the presence of different phytoconstituents. Various *Solanum* spp. provide a potential source of useful adulticidal drugs because of the presence of phytochemicals that can be used for the treatment of many diseases [17]. Thus, more scientific efforts should be made to identify and develop *Solanum*-based phyto-insecticides. Our

literature review revealed 19 *Solanum* medicinal plants used in all parts (leaves, roots, bark, and flowers). The goal of this review is to compile most of the scientific literature on mosquito-larvicidal and insecticidal investigations of *Solanum* plants and their active bioactive chemicals from various scientific sources, including the types of extracts examined, dosages, and effect on target organisms.

2. Source of Data

A comprehensive systematic review of the literature up to July 2020 on Solanaceae plants with larvicidal effects present in standard electronic databases, such as EBSCO, Europe PMC, Google Scholar, MEDLINE, PubMed, Scopus, and Web of Science, was conducted using various keywords (adulticidal, botanical, essential oil, insecticidal, larvicidal, repellency, Solanaceae, *Solanum*, and steroidal alkaloids). The search was restricted to publications having English titles. In addition, a manual search was performed to categorize related articles using references from the retrieved literature.

A total of 51 full-text original research articles published in peer-reviewed journals on Solanaceae plants were retrieved, and data were culled for larvicidal effects. Roles of larvicidal activities were assessed in Solanaceae plant solvent extracts, such as acetone, chloroform, ethyl acetate, hexane, and methanol from seventeen different medicinal plants. Other parts of these plants with significant larvicidal properties against various mosquito vectors were highlighted.

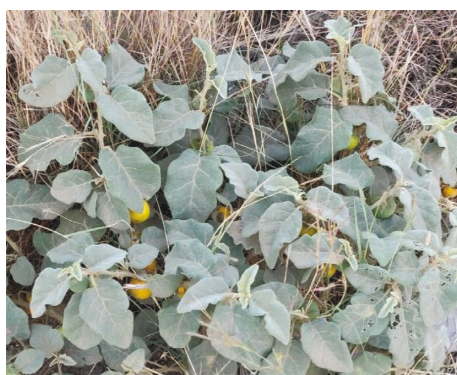
3. Solanaceae Family

Solanum L genus is the largest of the Solanaceae family or nightshades containing approximately 85–90 genera and 2,500–3000 species distributed in tropical and subtropical regions (Table 1) [12, 13]. Local names are given in various languages to describe a specific species for a particular local use. In Saudi Arabia, some species of Solanaceae are found primarily in the Asir Region and Jizan Region of Abha (Figure 1). A recent ethnobotanical study recorded three new collections of *Solanum* spp. in the southwest regions of Saudi Arabia [18].

3.1. Ethnopharmacological Use. Solanaceae is the most economically important family in the genus *Solanum* (Table 2). Solanaceae family offers a diversity of medicinal, culinary, and ornamental applications. The genus *Solanum* has attracted much interest in chemical and biological investigations over the last 30 years. Biologically important products for medicine and food include atropine, hyoscyne, solasodine, and withanolide [19–21]. Although rich in alkaloids of medical importance, Solanaceae plants contain alkaloids with toxicity to humans and animals, ranging from mild irritation to fatal outcomes [22–25]. In addition, *Solanaceae* spp. have potential importance as food supplements worldwide [22, 26]. *S. nigrum*, *S. xanthocarpum*, *S. tuberosum*, and *S. lycopersicum* are a few economically important species of the *Solanum* genus. Various species in

TABLE 1: Taxonomy of *Solanaceae* family.

Taxonomic placement	Scientific division
Kingdom	Plantae
Subkingdom	Tracheobionta
Infrakingdom	Streptophyta
Superdivision	Spermatophyta
Division	Magnoliophyta/Tracheophyta
Class	Magnoliopsida
Subclass	Asteridae
Superorder	Asteranae
Order	Solanales
Family	Solanaceae
Subfamily	Nicotianoideae
Genus	<i>Solanum</i> L
Common name	Nightshade



(a)



(b)

FIGURE 1: Leaves (a) and fruits (b) of *Solanum incanum*, Asir Region, Abha, Saudi Arabia.

this genus have completed various pharmacological research to verify and validate their ethnopharmacological usage. However, various reviews of the *Solanum* genus have been published, most of which focused on a single species [14, 27–30]. Table 3 summarizes the scientific literature and reveals a variety of ethnopharmacologically based traditional insect repellents derived from *Solanum* plants utilized by local ethnic communities in various countries to avoid mosquito bites.

Solanum genus has several species found in tropical and subtropical areas and is used in folk medicine and dietary supplements. Among them, *S. nigrum* has been considered ethnobotanically important due to its use in the traditional healthcare system to cure various ailments. The leaves and bitter berries with pungent have been traditionally used against severe ulcers, heart diseases, piles, dysentery, gastritis, and stomachache [27]. *S. sisymbriifolium*, known as “wild tomato,” is a traditional medicine used by indigenous people of Central and South America to treat veterinary and human diseases. Various parts of the wild tomato have been widely used to prevent and treat numerous diseases, including hypertension, diarrhea, and respiratory and urinary tract infections [31].

S. tuberosum is used in folk medicine to treat burns, constipation, hemorrhoids, corns, cough, tumors, scurvy,

and warts and to prevent wrinkles on the face [32]. *S. integrifolium* is native to Africa; its unripe fruits are eaten daily to check high blood pressure, inflammation, pain remedy to alleviate edema or cure stomach pain, lymphadenopathy, or sore armpits in indigenous medicine [34]. *S. villosum* is a traditionally important plant used in various systems of medicine for the treatment of leucorrhea, nappy rash, wounds, and cold sores, and as an ointment for sores and abscesses. A well-known traditional herb *S. xanthocarpum* is widely used in India to manage different ailments, including urolithiasis [35]. *S. trilobatum* is a widely used plant in the Indian indigenous systems of medicine. It is mainly used to treat respiratory diseases such as bronchial asthma [37]. *S. virginianum* L. has been used to manage fever, bronchial asthma, and cough for thousands of years [48].

In traditional medicine in Peru, *S. mammosum* is used to treat fungal infections and respiratory disorders via topical application. *S. incanum* is commonly found in Africa and is used as a folklore remedy for sore throat, angina, stomachache, colic, headache, wounds, pain relief in toothache, cure of snake bites, and sexually transmitted disease in wounds [49]. *S. elaeagnifolium* is called silverleaf nightshade and traditionally is used for the treatment of sore throats as an antiseptic agent, toothaches, and gastrointestinal disorders.

TABLE 2: Common and scientific names of *Solanum* spp.

Common name	Scientific name
Silverleaf nightshade	<i>Solanum elaeagnifolium</i>
Scarlet eggplant	<i>S. integrifolium</i>
Sticky nightshade	<i>S. sisymbriifolium</i>
Potato	<i>S. tuberosum</i>
Black nightshade	<i>S. nigrum</i>
Red nightshade	<i>S. villosum</i>
Yellow-fruit nightshade	<i>S. xanthocarpum</i>
Thai nightshade	<i>S. virginianum</i> L.
Indian ginseng (ashwagandha)	<i>S. trilobatum</i>
Nipplefruit nightshade	<i>Withania somnifera</i>
Garden tomato	<i>S. mammosum</i>
Jasmine nightshade	<i>S. lycopersicum</i>
Mullein nightshade	<i>S. laxum</i>
Jerusalem cherry	<i>S. verbascifolium</i>
Thorn apple	<i>S. pseudocapsicum</i>
Turkey berry	<i>S. incanum</i>
	<i>S. torvum</i>

TABLE 3: List of various phytochemicals and ethnopharmacological uses from *Solanum* plants.

Species name	Medicinal uses	Parts used	Phytochemicals	Country used	References
<i>S. sisymbriifolium</i>	Used as contraceptive febrifuge, to treat syphilis, hypertension, diarrhea, and respiratory and urinary tract infections, and as analgesics	Whole plants	Solanargine and β -solanargine, cuscohygrine, sisymbriifolin, neolignan	Paraguay, India, Brazil, Peru, and Argentina	[31]
<i>S. tuberosum</i>	Used to treat burns, constipation, hemorrhoids, corns, cough, tumors, scurvy, and warts, to prevent wrinkles on face, pain, acidity, and swollen gums, and to heal burns	Tubers, skins, raw juice	Solanidine, demissidine, α -chaconine, α -solanine, solavilline, solasdamine	Europe and South America	[32]
<i>S. nigrum</i>	Used to treat Liver disorders, diarrhea, inflammatory conditions, chronic skin ailments (psoriasis and ringworm), fever, hydrophobia, painful periods, and eye diseases	Whole plants	Steroidal alkaloids Steroidal saponins Glycoprotein	Kenya, China, India, and Pakistan	[27]
<i>S. villosum</i>	Used to treat leucorrhea, nappy rash, wounds, and cold sore	Whole plants	Solanidine, α -chaconine, (d) α -solanine	Africa, Central and South America, China, India, and Pakistan	[33]
<i>S. integrifolium</i>	Used to treat high blood pressure and edema or to cure stomach, lymphadenopathy, and inflammation, and as pain remedy to alleviate edema	Fruits	N-caffeoyl putrescine, 5-caffeoylquinic acid, and 3-acetyl-5-caffeoylquinic acid	South-East Asia, Brazil, Argentina, Uruguay, and Paraguay	[34]
<i>S. xanthocarpum</i>	Used to treat urolithiasis, respiratory disorders (expectorant, coughs, bronchial asthma, and chest pain), gonorrhoea, pest repellent, tympanitis, misperistalsis, piles, and dysuria	Whole plants	Saponins, solanacarpine, solanacarpidine, solancarpine, solasonine	South-East Asia including India, Malaysia, and tropical Australia	[35, 36]

TABLE 3: Continued.

Species name	Medicinal uses	Parts used	Phytochemicals	Country used	References
<i>S. trilobatum</i>	Used to treat cough and cold, respiratory disease (chronic bronchitis and tuberculosis), and male fertility, and to cure snake poison, dyspnea, anorexia, worm infestation, skin diseases, hemiplegia, edema, urinary calculi, amenorrhea, and urinary tract disorders.	Leaves	Sobatum, solasodine, solanine, tomatidine, diosgenin, soladunalinidine	China, Myanmar, Thailand, Vietnam, Sri Lanka, Peninsular Malaysia, and southern India	[37–39]
<i>S. virginianum</i>	Used to treat sore throats, chest pain and catarrh, stomach and respiratory complaints, fever, influenza, painful and difficult urination, bladder stones, and rheumatism	Whole plants	Arabinogalactan, chlorogenic and caffeic acid, khasianine, solasonine, solamargine, beta-solamargine, solanocarpine, and solanocarpidine	India, Sri Lanka, South-East Asia, Malaysia, tropical Australia, and Polynesia	[40]
<i>S. mammosum</i>	Used to treat fungal infections and respiratory disorders (asthma, cough, cold, and sinusitis), skin ulcer, scabies, furunculosis and rashes, insecticide, and rat poison Used to treat skin and cardiovascular diseases, cancer, burns, scalds and sunburn, rheumatism and severe headaches, filarial worm swellings, incipient leprosy spots, and toothache.	Leaves, fruits, and seeds	Indioside D, solamargine, tomatine, solasonine, diosgenin, solamargine and β -solamargine	Northern and South America, Caribbean islands, and Africa	[41]
<i>S. lycopersicum</i>	Used to treat sore throat, angina, stomachache, ear inflammation, snake bites, wounds, liver disorders, skin ailments (ringworm), warts, inflammatory conditions, painful periods, and fever	Fruits	Lycopene, zeaxanthin, esculoside A, beta-carotene	South and Central America	[42]
<i>S. elaeagnifolium</i>	To cure cold and infant, typhoid, pneumonia, sore throats, an antiseptic agent, toothaches, and gastrointestinal disorders	Whole plants	Solanine, solasonine, solasodine, kaempferol 8-C-beta-galactoside, β -solamarine, solanidine	Asia, Africa, Australia, and tropical and subtropical America	[43]
<i>S. incanum</i>	Used to treat sore throat, angina, stomachache, ear inflammation, snake bites, wounds, liver disorders, skin ailments (ringworm), warts, inflammatory conditions, painful periods, and fever	Whole plants	Khasianine, incanumine, solasodine, kaempferol, isoquercitrin, yamogenin	Africa, Middle East and Far East Asia, and Arabian Peninsula	[44]
<i>S. jasminoides</i> or <i>S. laxum</i>	Used to kill insects	Aerial parts	Steroidal glycosides—inunigroside A; steroidal sapogenol—jasminoside A, solasodine, laxumin A, laxumin B	Uruguay, Brazil, South America, Paraguay, Uruguay, and Argentina	[13]
<i>S. pseudocapsicum</i>	Used to treat boils and gonorrhoea, male tonic and abdominal pain, somnolence, and diabetes	Bark, fruit, leaves, and seeds	Solanocapsine, solacanine, solateinimine, O-methylsolanocapsine, episolacapsine, and isosolacapsine	India, Nepal, and the Philippines	[45]
<i>S. torvum</i>	Used to treat fever, wounds, tooth decay, reproductive problems, and arterial hypertension	Fruits and leaves	Chlorogenin, torvoside A-L, chlorogenone	Thailand, India, West Indies, and South America	[46]
<i>S. verbasicum</i>	Used to treat diarrhea, dysentery, eczema, edema, gout, headaches, ulcers, fever, hematuria, and toothache	Leaves and roots	Pentanone and γ -sitosterol	India and China	[47]

Phytochemical analysis of berry extracts *S. elaeagnifolium* revealed the presence of kaempferol 8-C- β -galactoside that possesses medicinal proprieties [50]. *S. verbascifolium* is used in Chinese folklore for diarrhea, dysentery, eczema, edema, gout, headaches, ulcers, fever, hematuria, and toothache [45]. Despite being a poisonous plant, *S. pseudocapsicum* is used in traditional medicine to treat boils and gonorrhoea and relieve abdominal pain, and as a male tonic [47]. *S. torvum* is another commonly used Solanaceae herb in traditional medicine. The plant extracts have been widely used to treat fever, wounds, tooth decay, reproductive problems, and arterial hypertension [46]. Thus, leaves, fruits, roots, and aerial parts of *Solanum* plants can benefit humans by enhancing their health when consumed as part of a daily diet, nutraceutical, or biopharmaceutical.

3.2. Phytopharmacology and Insecticidal Properties of Solanaceae spp. Medicinal Solanaceae plants have traditionally been used as insecticidal, anti-infectious, and antimicrobial agents [51, 52]. Table 4 shows the different types of test organisms, bioassays, and doses applied to investigate the mosquitocidal activity of crude plant extracts from the *Solanum* genus. Crude and chloroform-methanol extracts of *S. tuberosum* at very low concentrations are effective in mosquito control [54]. Volatile oils of *S. xanthocarpum* were effective as insect repellents, giving rise to >5 hours of protection against *Culex quinquefasciatus* without apparent dermal irritation to human skin [72]. Chloroform-methanol extract of *S. villosum* green berries was used as a biocontrol agent against *Aedes aegypti* [90]. *S. villosum* green berries had the greatest biocidal activity against *St. aegypti. aegypti*, and *Cx. quinquefasciatus* in chloroform and methanol extracts. As a result, crude extracts or protein fractions/isolated bioactive phytochemicals from *S. villosum* could be utilized as a possible biocontrol agent against these mosquitoes, especially because of its larvicidal impact [58, 59, 90]. *S. integrifolium* chitin-binding lectins (CBLs) inhibit *Spo-doptera frugiperda* (sf21) insect cell growth by binding to carbohydrates and depolarizing mitochondrial membrane potential [60].

Table 5 summarizes detailed investigations of the mosquito-larvicidal efficiency of various *Solanum* species. Some examples are highlighted (according to the author's viewpoint) as follows: *S. xanthocarpum* extracts show various larvicidal and pupicidal activity against Cx's first-to-fourth instars. *Cx. quinquefasciatus* fruit aqueous extract exhibits 100% killing after 48-hour exposure compared with its root extract [61, 62]. The previous study has reported that the fruit extract of *S. xanthocarpum* and copepod *Mesocyclops thermocyclopoides* could serve as a potential highest mortality rate against dengue vector *Ae. aegypti* [63]. This mosquitocidal efficiency may be caused by detrimental effects of the *S. xanthocarpum* active principle compounds (solanocarpine and solanocarpidine) on the mosquito larvae. Similarly, *S. xanthocarpum* fruit extracts had larvicidal action against *An. stephensi* and *Cx. quinquefasciatus*, as well as one culicine species, *Ae. aegypti*. The toxic concentrations of fruit extract against *An. culicifacies*, *An. stephensi*, and *Ae.*

aegypti were found to be 0.112 and 0.258%, 0.058 and 0.289%, and 0.052 and 0.218%, respectively, at the LC₅₀ and LC₉₀ levels. It was discovered that crude extracts have larvicidal capability due to their volatile oil content, implying that they could be used as an environmentally friendly, effective larvicidal in managing various vector-borne epidemics [66, 97]. Methanol leaf extract of *S. trilobatum* is effective against *Ae. aegypti*, *Cx. quinquefasciatus*, and *An. stephensi* pupae and larvae with an LC₅₀ value of 125, 128, and 117 ppm, respectively [73]. Chloroform: methanol (1:1 v/v) extract of *S. nigrum* mature leaves is toxic against Cx's early 3rd instar larvae of the *Cx. vishnui* group and *An. subpictus* [56].

The seed hexane extract of *S. trilobatum* exhibited (38%) acaricidal and insecticidal activities against the adult of *H. bispinosa* (Ixodidae) and hematophagous fly *H. maculata* Leach (Hippoboscidae). Therefore, this study provides the first report on the parasitic activities of plant extracts from southern India [75]. The leaf extract of *S. trilobatum* was found to have an oviposition deterrent effect, reducing egg-laying by *An. stephensi* by 18 to 99% and providing 70 to 120 minutes of mosquito bite protection skin repellent activities. *S. trilobatum* leaf extract had dose-dependent oviposition deterrent and skin repellent effects. Several solvent extracts of *S. trilobatum* were tested against the filarial vector *Cx. quinquefasciatus*; petroleum ether had the highest larvicidal activity, with LC₅₀ values of 203.87 and 165.04, respectively, after 24 and 48 hours, followed by acetone and chloroform extracts [96]. According to the findings, *S. trilobatum* leaf extract is an efficient oviposition preventive and cutaneous repellent against *A. stephensi* [72]. The crude extract of the leaves or fruits of *S. incanum* and *W. somnifera* has an equal effect on the *A. messinae* mortality (96% mortality). However, the percentage mortality of the termite was 100% with 135 μ g from the crude extract of *S. incanum* leaves. Based on findings, both crude extracts have the potential to be used as termite control agents in termite breeding areas in the field or infested homes [80].

The larvicidal efficacy of *S. torvum* was tested against *An. stephensi* and *Cx. quinquefasciatus*, with the results indicating that the leaf methanol extract of *S. torvum* had the highest LC₉₀, ranging from 70.38 to 210.68 ppm. As a result, isolated plant metabolites from *S. torvum* from southern India have the potential to be used as environmentally safe and long-lasting mosquito repellents [85]. The mosquito repellent effect of *S. lycopersicum esculentum* leaf hydro-ethanolic extract on the larvae of multiple mosquito species was tested at varied concentrations of 50, 100, 150, 200, and 250 ppm, with larva mortality seen within 24 hours. The hydro-ethanolic extract caused complete mortality in mosquitoes at 200 ppm in 18–19 hours, and the study found that *S. lycopersicum esculentum* may kill mosquitoes at a lower concentration [77]. The insecticidal effects of methanolic extracts of *S. elaeagnifolium* seeds were investigated further against *S. littoralis*, and 100% larval mortality was observed with the strongest growth inhibition (59.68%) compared to leaves [110]. Methanolic extracts from the leaves and seeds of *S. elaeagnifolium* also showed insecticidal efficacy against *P. operculella* and *T. castaneum*. Seed extract

TABLE 4: Different types of test organisms, bioassays, and doses used to study the mosquitocidal activity of crude plant extracts from *Solanum* genus.

Species name	Species tested	Types of bioassays	Dose	References
<i>S. sisymbriifolium</i>	<i>Anophelinae</i> (insects and larvae)	Biocidal assay	0.005–5 g/ml	[53]
<i>S. tuberosum</i>	<i>Cx. quinquefasciatus</i> and <i>An. stephensi</i>	Larvicidal assay	1.1–0.5% (AE) 1.2 25–75 ppm (CME)	[54]
<i>S. nigrum</i>	<i>Cx. quinquefasciatus</i> and <i>An. stephensi</i>	Mosquito-larvicidal assay	2.5, 5, and 10 ppm	[55]
	<i>Cx. quinquefasciatus</i>	Larvicidal bioassay	1–3%	[56]
	<i>Cx. quinquefasciatus</i>	Mosquitocidal assay	15, 20, and 25 mg/L	[57]
	<i>Cx. quinquefasciatus</i>		30, 50, and 100 ppm	
<i>S. villosum</i>	<i>An. subpictus</i>	Larvicidal assay	30, 50, 100, and 200 ppm	[58, 59]
	<i>S. aegypti</i>		0.1–0.5% and 15, 25, and 30 ppm	
<i>S. integrifolium</i>	<i>Spodoptera frugiperda</i>	Insecticidal assay	1 µg/mL	[60]
	<i>Cx. quinquefasciatus</i>	Mosquito-larvicidal and pupicidal assays	50–650 ppm	[61]
	<i>Culicine larvae</i>	Larvicidal assay	1–5 ml	[62]
	<i>Ae. aegypti</i>	Mosquitocidal assay	100, 150, 200, 250, and 300 ppm	[63]
	<i>Ae. aegypti</i>	Insecticidal	0.82 mg/ml	[64]
<i>S. xanthocarpum</i>	<i>Cx. quinquefasciatus</i>	Mosquito-larvicidal assay	62.5, 125, 250, 500, and 1000 mg/L	[65]
	<i>Cx. quinquefasciatus</i>	Larvicidal assay	7500–20 000 ppm	[66]
	<i>An. stephensi</i>	Larvicidal assay	1 : 1, 1 : 2, and 1 : 4%	[67–69]
	<i>Cx. quinquefasciatus</i>	Larvicidal assay	1 : 1, 1 : 2, and 1 : 4%	
	<i>An. stephensi</i>	Larvicidal assay	7500–20 000 ppm	[70]
	<i>Cx. Vishnui</i> and <i>L. acuminata</i>	Mosquito-larvicidal assay	75, 100, and 150 ppm	[71]
	<i>Cx. quinquefasciatus</i>	Mosquito-larvicidal assay	15, 20, and 25 ppm	
		Oviposition deterrent assay	0.01, 0.025, 0.05, 0.075, and 0.1%	[72]
	<i>An. stephensi</i>	Skin repellent assay	0.001, 0.005, 0.01, 0.015, and 0.02%	[73]
<i>S. trilobatum</i>	<i>Ae. aegypti</i> , <i>Cx. quinquefasciatus</i> , and <i>An. stephensi</i>	Larvicidal and pupicidal assays	50, 100, 150, 200, and 250 ppm	[74]
	<i>Ae. aegypti</i> and <i>Cx. quinquefasciatus</i>	Larvicidal assay	100, 200, 300, 400, and 500 ppm	[74]
	<i>Haemaphysalis bispinosa</i> and <i>Hippobosca maculata</i>	Acaricidal and insecticidal assays	46.88 to 3,000 ppm	[75]
Solanum Mammosum-Silver Nanoparticles (Sm-AgNPs)	<i>Ae. aegypti</i>	Larvicidal assay	1500, 3000, 4500, and 6000 ppm 0.05, 0.06, 0.07, and 0.08 ppm	[76]
<i>S. lycopersicum</i>	<i>Ae. aegypti</i> and <i>Cx. quinquefasciatus</i>	Larvicidal activity	50, 100, 150, 200, and 250 ppm	[77]
	<i>Fourth instar larvae</i>	Larvicidal assay		[78]
<i>S. elaeagnifolium</i>	<i>Tribolium castaneum</i> and <i>Phthorimaea operculella</i>	Insecticidal assay	2% extract (5 µl spray)	[79]
<i>S. incanum</i>	<i>A. messinae</i> and <i>M. najdensis</i>	Insecticidal assay	2.5–135 µg/ml	[80]
<i>S. laxum—laxumin A</i>	<i>Schizaphis graminum</i>	Insecticidal assay	50–500 µm	[81]
<i>S. laxum—luciamin</i>	<i>Schizaphis graminum</i>	Repellant assay	50–500 µm	[82]
<i>S. jasminoides</i>	<i>Phlebotomus papatasi</i> and <i>Bougainvillea glabra</i>	Insecticidal assay	—	[83]
<i>S. surattense</i>	<i>Callosobruchus chinensis</i>	Insecticidal assay	1, 2.5, 5, and 10%	[84]
<i>S. torvum</i>	<i>An. stephensi</i> and <i>Cx. quinquefasciatus</i>	Larvicidal bioassay	1.25 to 400 ppm	[85]
<i>S. verbasicum</i>	<i>Cx. quinquefasciatus</i>	Larvicidal activity	100, 300, 500, or 1000 ppm	[86]
<i>S. asperum</i>	<i>Biomphalaria glabrata</i>	Molluscicidal activity	10, 50, and 100 µg/ml	[87]
<i>Nicotiana glauca</i>	<i>Pieris rapae</i>	Larvicidal assay	0.7 mg to 2.8 mg/ml	[88]
<i>S. elaeagnifolium</i>	<i>Tribolium castaneum</i>	Repellent and antifeedant assay	200 µl/disc (2% extract)	[89]

TABLE 5: Insecticidal efficacy of Solanaceae plant extracts and their fraction/compound as adulticides.

Solanum spp.	Part used	Target insect species	Effect	Extract/compound fraction	Bioactive compound	LC ₅₀ , LC ₉₀	Reference
<i>S. sisymbriifolium</i>	F	<i>Anopheles funestus</i> and <i>Anopheles gambiae</i>	Insecticidal	Total alkaloid fraction	Solamargine β -solamarine	0.45–0.75 mg/ml	[53]
<i>S. tuberosum</i>	T	<i>An. stephensi</i> (malaria vector) <i>Culex quinquefasciatus</i> (filaria vector)	Mosquito-larvicidal	Aqueous, chloroform: methanol (1 : 1)	NI	1.18–1.30 mg/l	[54]
<i>S. nigrum</i>	B, L	<i>An. stephensi</i> (malaria vector) <i>Cx. quinquefasciatus</i> (filaria vector)	Mosquito-larvicidal	Silver nanoparticle (AgNP)	Alkaloids	1.26–2.44 ppm	[55]
<i>S. villosum</i>	B	<i>Aedes aegypti</i> (dengue vector)	Mosquito-larvicidal	Chloroform:methanol	Steroids	21.02 (3 rd instar) ppm	[90]
<i>S. integrifolium</i>	F	<i>Spodoptera frugiperda</i>	Insecticidal	Chitin-binding lectins (CBLs) and crude	Polysaccharide	1 μ g/ml	[60]
<i>S. xanthocarpum</i>	L	<i>Cx. quinquefasciatus</i> (filaria vector)	Mosquito-larvicidal and pupicidal	Crude ethanolic	NI	155.3–448.4 ppm, 687.1–1,141.6 ppm	[61]
<i>S. xanthocarpum</i>	F	<i>Ae. aegypti</i> (dengue vector)	Mosquito-larvicidal and pupicidal	Crude ethanolic	SolanocarpidineSolanocarpine	253.2, 435.2 ppm/79.5, 462.1 ppm	[63]
<i>S. xanthocarpum</i>	L	<i>Cx. quinquefasciatus</i> (filarial vector)	Mosquito repellent effect	Volatile oil	Volatile oil	8% repellency; 311 minutes of protection	[72]
<i>S. trilobatum</i>	L	<i>An. stephensi</i> (malaria vector)	Oviposition deterrent and skin repellent	Leaf extract	Volatile compounds	99.4% repellency; 123 minutes of protection	[72]
<i>S. xanthocarpum</i>	WP	<i>Cx. quinquefasciatus</i> (filaria vector)	Larvicidal and pupicidal	Chloroform fraction, crude	Quinine, terpenoids, and other compounds	227.9 ppm, 411.4 ppm	[91]
<i>S. xanthocarpum</i>	F, R	<i>Culicine larvae</i>	Larvicidal	Crude aqueous	AlkaloidsSaponins	~100% mortality	[62]
<i>S. trilobatum</i>	L	<i>Ae. aegypti</i> (dengue vector) <i>An. stephensi</i> (malaria vector) <i>Cx. quinquefasciatus</i> (filaria vector)	Adulticidal	Crude methanolic	NI	116.6–127.8 ppm	[73]
<i>S. nigrum</i>	L	<i>An. subpictus</i> <i>Cx. vishnui</i> group	Larvicidal	Chloroform:methanol (1 : 1 v/v)	Phytosteroids	3.68–5.64 mg/l, 24.74–44.33 mg/l	[56]
<i>S. villosum</i>	L	<i>Ae. aegypti</i> (dengue vector) <i>An. stephensi</i> (malaria vector) <i>Cx. quinquefasciatus</i> (filaria vector)	Mosquito-larvicidal	Leaf protein	Polypeptides	644.7–747.2 ppm, 1,882.4–2,220.0 ppm	[90]
<i>S. virginianum</i>	WP	<i>Ae. aegypti</i> (dengue vector)	Insecticidal	Methanolic	NI	0.82 mg/ml	[92]

TABLE 5: Continued.

<i>Solanum</i> spp.	Part used	Target insect species	Effect	Extract/compound fraction	Bioactive compound	LC ₅₀ , LC ₉₀	Reference
<i>S. nigrum</i>	B, L	<i>Ae. aegypti</i> (dengue vector)	Larvicidal	Crude	Eugenol (E)-6-hydroxy-4,6-dimethyl-3-heptene-2-one	Leaf: 9.8 ml/l, 26.4 ml/l Green berry: 51.4 ml/l, 459.8 ml/l Black berry: 9.9 ml/l, 56.1 ml/l	[93]
<i>S. nigrum</i>	L	<i>Cx. quinquefasciatus</i> (filaria vector)	Mosquito-larvicidal	Crude	Alkaloids	0.08%, 0.37%	[57]
<i>S. nigrum</i>	B	<i>Cx. quinquefasciatus</i> (filaria vector)	Mosquito-larvicidal	Crude, chloroform: methanol (1:1, v/v)	Aromatic amide compounds	61.5 mg/l, 297.0 mg/l SX: 48.4 mg/l, 218.2 mg/l SX: WS (1:1 v/v): 32.7 mg/l, 149.4 mg/l/SX: WS (1:2 v/v): 22.9 mg/l, 109.8 mg/l ISX: WS (1:3 v/v): 50.2 mg/l, 361.9 mg/l	[55]
<i>S. xanthocarpum</i> , <i>Withania somnifera</i>	F	<i>Ae. aegypti</i> (dengue vector) <i>An. stephensi</i> (malaria vector) <i>Cx. quinquefasciatus</i> (filaria vector)	Synergistic larvicidal	Crude aqueous	NI		[65]
<i>S. xanthocarpum</i>	WP	<i>Ae. aegypti</i> (dengue vector) <i>An. culicifacies</i> <i>An. stephensi</i> (malaria vector) <i>Cx. quinquefasciatus</i> (filaria vector)	Larvicidal	Methanolic	Edible oils	91.7–450.6 ppm, 379.0–1,881.0 ppm	[94]
<i>S. xanthocarpum</i>	NI	<i>Ae. aegypti</i> (dengue vector) <i>Cx. quinquefasciatus</i> (filaria vector)	Mosquito-larvicidal	Ethanollic	NI	788.10, 1288.91 mg/l, 573.20, 1066.93 mg/l	[95]
<i>S. mammosum</i>	NI	<i>Ae. aegypti</i> (dengue vector)	Larvicidal	Aqueous silver nanoparticles	Steroidal alkaloids	1,631.3 ppm, 4,756.2 ppm; 0.06 ppm, 0.08 ppm	[76]
<i>S. trilobatum</i>	L	<i>Ae. aegypti</i> (dengue vector) <i>Cx. quinquefasciatus</i> (filaria vector)	Mosquito-larvicidal	Acetone	Cyclodecanol and other compounds	189.5 ppm, 444.3 ppm 167.4 ppm, 371.8 ppm	[74]

TABLE 5: Continued.

<i>Solanum</i> spp.	Part used	Target insect species	Effect	Extract/compound fraction	Bioactive compound	LC ₅₀ , LC ₉₀	Reference
<i>S. trilobatum</i>	AP	<i>Cx. quinquefasciatus</i> (filarial vector)	Mosquito-larvicidal	Acetone, chloroform, petroleum ether	NI	Acetone: 186.4 mg/L, 366.5 mg/L Chloroform: 346.1 mg/L, 595.6 mg/L Petroleum ether: 165.0 mg/L, 293.5 mg/L	[96]
<i>S. xanthocarpum</i>	R	<i>An. stephensi</i> (malaria vector)	Larvicidal	Petroleum ether	NI	0.93 ppm, 8.48 ppm	[67]
<i>S. xanthocarpum</i>	R	<i>An. stephensi</i> (malaria vector)	Larvicidal	Petroleum ether with temephos (1:1)	NI	0.02 ppm, 0.09 ppm	[69]
<i>S. xanthocarpum</i>	F	<i>An. stephensi</i> (malaria vector) <i>Cx. quinquefasciatus</i> (filarial vector)	Larvicidal	Carbon tetrachloride/petroleum ether	NI	1.27 ppm, 59.45 ppm	[97]
<i>S. xanthocarpum</i>	R	<i>Cx. quinquefasciatus</i> (filaria vector)	Larvicidal	Petroleum ether	NI	38.48 ppm, 80.83 ppm	[66]
<i>S. xanthocarpum</i>	R	<i>Cx. quinquefasciatus</i> (filaria vector)	Larvicidal	Temephos/plant (1:1)	NI	0.01 ppm, 0.02 ppm	[67]
<i>S. xanthocarpum</i>	F, R	<i>Ae. aegypti</i> (dengue vector) <i>An. culicifacies</i> <i>An. stephensi</i> (malaria vector)	Larvicidal	Fruit, root	NI	0.05–1.16 ppm, 0.22–3.58 ppm	[98]
<i>S. villosum</i>	L	<i>Cx. quinquefasciatus</i> (filaria vector)	Larvicidal	Chloroform:methanol (1:1 v/v)	NI	39.19 ppm	[58]
<i>S. villosum</i>	L	<i>An. subpictus</i>	Larvicidal	Chloroform-methanol	Glycoalkaloids	23.47–30.63 ppm	[59]
<i>S. lycopersicum</i>	L	<i>Culex</i> and <i>Aedes</i> spp.	Larvicidal	Aqueous ethanolic	NI	100% mortality at 250 ppm	[77]
<i>S. nigrum</i>	L	<i>Ae. aegypti</i> (dengue vector) <i>An. culicifacies</i> <i>Cx. quinquefasciatus</i> (filaria vector)	Larvicidal	Crude aqueous	NI	0.027–0.032%, 0.027–0.212%	[99]
<i>S. elaeagnifolium</i>	F	<i>Blattella germanica</i>	Repellent effect	Ethanolic, hexane	NI	50 mg/ml	[100]
<i>S. elaeagnifolium</i>	L, B	<i>An. labranchiae</i>	Larvicidal effect	Aqueous, ethanolic	Glycoalkaloid extracts	LC ₉₀ (24 h) 209.8, 123.4 ppm	[78]
<i>S. elaeagnifolium</i>		<i>Fasciola hepatica</i> <i>Galba truncatula</i> Müll. <i>Amiatermes</i>	Molluscicidal activity	Total saponin fraction	β -solamarine	0.94 mg/L/14.67 mg/L	[101]
<i>S. incanum</i>	F, L	<i>messinae</i> <i>Microtermes majdensis</i>	Insecticidal	Crude hexane	β -chaconine, α -solanine	40% mortality at 67.5 μ g/ml	[80]

TABLE 5: Continued.

<i>Solanum</i> spp.	Part used	Target insect species	Effect	Extract/compound fraction	Bioactive compound	LC ₅₀ , LC ₉₀	Reference
<i>S. jasminoides</i>	WP	<i>Phlebotomus papatasi</i> (Leishmania vector)	Larvicidal and repellent effect	Branch	NI	Median survival = 8 days (confidence interval: 17.1–18.9)	[83]
<i>S. nigrum</i>	F	<i>Ae. aegypti</i> (dengue vector) <i>An. culicifacies</i> <i>AAn. culicifacies</i> <i>CAn. stephensi</i> (malaria vector) <i>Cx. quinquefasciatus</i> (filaria vector)	Mosquito-larvicidal	Aqueous, hexane	NI	<20 ppm, <100 ppm	[102]
<i>S. pseudocapsicum</i>	NI	<i>Helicoverpa armigera</i> <i>Spodoptera litura</i>	Antifeedant, insecticidal	Ethyl acetate (5%)	NI	Maximum insecticidal = 66.5–75.3%	[103]
<i>S. pseudocapsicum</i>	L, SD	<i>Agrotis ipsilon</i>	Antifeedant, insecticidal	Ethyl acetate (5%)	NI	Maximum insecticidal = 60.1%	[104]
<i>S. nigrum</i>	L, F	<i>Ae. caspius</i> <i>Cx. pipiens</i>	Larvicidal	70% ethanolic	NI	3.37 mg/l	[105, 106]
<i>S. surattense</i>	F, L, R, S	<i>Callosobruchus chinensis</i>	Pesticidal	Aqueous extract, aqueous suspension	NI	Reduction in oviposition = 2–5 eggs/pair	[84]
<i>S. surattense</i> and <i>S. trilobatum</i>	L	<i>Cx. quinquefasciatus</i> (filaria vector) <i>An. stephensi</i> (malaria vector) <i>Cx. quinquefasciatus</i> (filaria vector)	Insecticidal	Ethyl acetate, petroleum ether	NI	46.04 ppm	[107]
<i>S. torvum</i>	L	<i>quinquefasciatus</i> (filaria vector)	Larvicidal	Methanolic	NI	(LC ₉₀) 70.38–210.68 ppm	[85]
<i>S. trilobatum</i>	L, SD	<i>Hippobosca maculata</i>	Insecticidal	Hexane	NI	495.61–432.77 ppm, 1, 914.84–1,872.33 ppm	[75]
<i>S. verbasicum</i>	L	<i>Cx. quinquefasciatus</i> (filaria vector) <i>Leptinotarsa decemlineata</i> <i>Empoasca fabae</i>	Larvicidal	Various solvents	NI	100% mortality at 72 hours	[86]
<i>S. tuberosum</i>	T	<i>decemlineata</i> <i>Empoasca fabae</i>	Insecticidal	Crude sample	Chaconine	100% defoliation	[108]
<i>S. laxum</i>	WP	<i>Schizaphis graminum</i>	Insecticidal	Steroidal glycoalkaloid fraction	Laxumin A and B	4.3 μM and 6.1 μM	[81]
<i>S. laxum</i>	AP	<i>Schizaphis graminum</i>	Insecticidal	Ethanolic	Luciamin	70% mortality at 24 hours	[82]
<i>S. nigrum</i>	B	<i>Cx. quinquefasciatus</i>	Mosquito-larvicidal	Chloroform: methanol (1:1, v/v) solvent	NI	80% mortality at 72 hours	[57]
Solanum steroidal alkaloids and glycoalkaloids	SA	<i>Tribolium castaneum</i>	Larvicidal	Isolated compounds in diet	Steroidal glycoalkaloids	89–100% larvicidal effect	[109]

TABLE 5: Continued.

<i>Solanum</i> spp.	Part used	Target insect species	Effect	Extract/compound fraction	Bioactive compound	LC ₅₀ , LC ₉₀	Reference
<i>S. nigrum</i>	L	<i>Lymnaea acuminata</i> <i>Cx. vishnui</i> (vector of <i>Japanese encephalitis</i>)	Mosquito-larvicidal	Aqueous	Aliphatic amide compounds	LC ₅₀ 55.45 and 11.59 ppm, respectively at 72 h. 32-48% mortality at 72 hours of 4th larval instars	[70]
<i>S. nigrum</i>	L	<i>Cx. quinquefasciatus</i>	Mosquito-larvicidal	Ethyl acetate	Glucosisaustrin		[71]
<i>S. asperum</i>	L	<i>Biomphalaria glabrata</i>	Molluscicidal activity	Methanolic extract	Solanandaine	LC9044.1;17.3399. 772.063.6	[87]
<i>Nicotiana glauca</i>	L	<i>Pieris rapae</i>	Larvicidal effect	SolanandaineSolasonineSolamargine Total alkaloid anabasine	SolanandaineSolasonineSolamargine Anabasine	EC50-1.202 mg/larva 0.572 mg/larva	[88]
<i>S. elaeagnifolium</i>	S, L	<i>Tribolium castaneum</i>	Insecticidal (repellent assay)	Methanol (seed and leaves)	Glycoalkaloids	% repellent effect: Seeds: 94% after 2 hrsLeaves: 74% after 2 hrs	[89]

AP: aerial part; B: berry; F: fruit; FL: flower; L: leaf; LC₅₀: 50% lethal concentration; LC90: 90% lethal concentration; LC100: 100% lethal concentration; mg/l: milligrams per liter; mg/ml: milligrams per milliliter; ml/l: milliliter per liter; µg/ml: micrograms per milliliter; µM: micromoles; NI: not identified; ppm: parts per million; R: root; S: stem; SA: *Solanum* alkaloids; SD: seed; SX: *S. xanthocarpum*; T: tuber; v/v: volume/volume; WP: whole plant; WS: *W. somnifera*.

inhibited oviposition and egg hatching the most (95.9% and 98.6%, respectively), with an aphid mortality rate of 23.6% [79]. These findings suggested that numerous *Solanum* species might be used as plant-based mosquitocidal. They could be a valuable source for developing novel natural repellents as an alternative to chemical repellents in the future. The structure of phytochemicals with promising mosquitocidal and insecticidal effects from *Solanum* species is summarized in Figure 2.

3.3. Solanaceae spp. Phytochemicals with Insecticidal Properties. For decades, *Solanum* species have been widely used in healthcare systems as a source for various phytochemicals, including steroidal alkaloids. *Solanum* is distinguished by the presence of the steroidal alkaloid solasodine, which is a potential starting material for the manufacture of steroid hormones. Because of the wide spectrum of biological activities such as antibacterial, anti-inflammatory, antioxidant, and anticancer, *Solanum* alkaloids have been a topic of interest in pharmacological and therapeutical investigations. Because of metabolites such glycoalkaloids, some of the *Solanum* species are poisonous. Several pharmacologically important lead compounds are found in *Solanum* species, including steroidal alkaloids such as solasodine, solasonine, solamargine, and other medicinally important alkaloids; solasodine and its glycosylated derivatives, such as solamargine and solanine; and other chemicals with medicinal potential for developing new drugs against various human diseases.

S. xanthocarpum is an important source of many pharmacologically and medicinally useful alkaloids. Recent GC-MS analysis showed that several essential oils from leaves, fruits, roots, and stems of *S. xanthocarpum* were responsible for larvicidal activity [111]. Six important phytosteroids (1, 2-benzenedicarboxylic acid, dibutyl phthalate, phytol, lauric acid, 3,7,11,15-tetramethyl-2-hexadec, and 7-hexadecenal) with larvicidal activity against early 3rd instar larvae of the *Cx. vishnui* group were identified from mature leaves of *S. nigrum* using GC-MS [32]. A short polypeptide (15 amino acids) from mature leaves of *S. villosum* exhibited a moderate larvicidal effect. Further studies will be needed to determine its mode of action and appropriate formulations for field applications [90]. Eugenol and (E)-6-hydroxy-4,6-dimethyl-3-heptene-2-one in *S. nigrum* crude extract were proposed as being the main compounds responsible for mosquito-larvicidal activity [93]. The GC-MS analysis of acetone leaf extract of *S. trilobatum* revealed cyclodecanol (12.42%), β -sitosterol (10.25%) and 2-tetradecyclohexane (6.07%) as the major components and were possibly responsible for larvicidal activity against *Cx. quinquefasciatus* and *Ae. aegypti* [74].

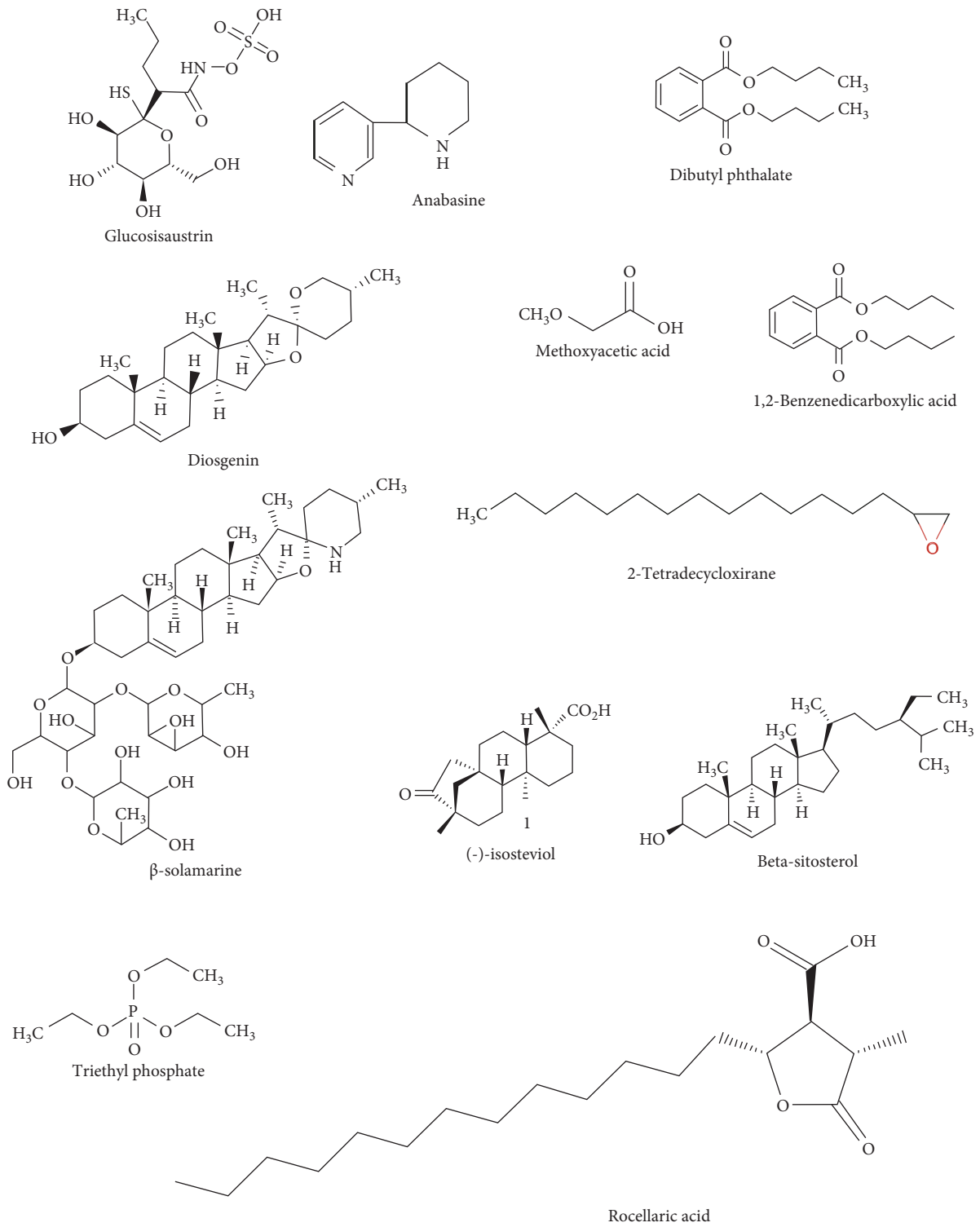
β -Solamarine isolated from the methanolic extract of seeds of *S. elaeagnifolium* was found to have molluscicidal activity against *G. truncatula* and *F. hepatica*. The median lethal concentration of β -solamarine in molluscicidal activity (LC₅₀) was of 0.49, and the study emphasizes that this glycoalkaloid may be used as molluscicides [101]. Another mosquitocidal investigation revealed that the leaf extract of

S. trilobatum possesses oviposition deterrent and skin repellent activity against *An. stephensi*. Both oviposition deterrent and skin repellent activity were dose-dependent [112]. Two major compounds of steroidal glycoalkaloids were isolated from the fraction C MeOH extract of *S. sisymbriifolium* and were identified as solamargine (1) and β -solamarine. The toxicity of fraction C is 10-fold higher than that of *Anophelinae* larvae. These two steroidal alkaloids were known to possess molluscicidal activity where they could be used as a molluscicide in the future [53].

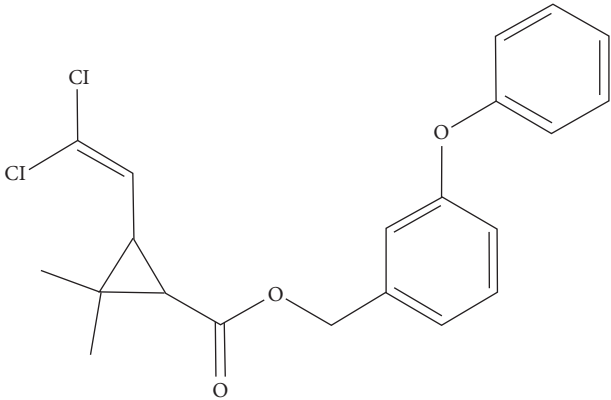
Luciamin, a spirostanol saponin, was isolated from the ethanolic extract of the aerial parts of *S. laxum* and was tested against the aphid *S. graminum* by incorporation in artificial diets. Luciamin showed a deterrent (toxic) activity against the insect and is the first spirostanol glycoside reported to have this activity. Luciamin's aphid repelling effect deserves further exploration to determine its biological and economic effects against viral vectors [81]. Two *Solanum* glycosides isolated from *S. laxum* were found to have insecticidal effects against *S. graminum* with LC₅₀ 4.3 μ M (laxumin A) and LC₅₀ 6.1 μ M (laxumin B), respectively [82]. The insecticidal effect of the isolated steroidal alkaloids fraction B from *S. sisymbriifolium* was investigated on *Anophelinae* larvae (*A. gambia*, *A. funestus*). Compared with other extracts and fractions, fraction B, which contains solamargine and β -solamarine, appears less hazardous to larvae. As a result, fraction B could potentially be employed as an insecticide in the future [53]. *Solanum* species steroidal alkaloids are unique in their pharmacological properties and are important lead molecules for drug development [113].

Glucosisaustrin, a glucosinolate group of bioactive compounds from *S. nigrum*, was responsible for larval mortality. Glucosinolate is a plant-derived secondary metabolite and hydrophilic, having potent mosquito-larvicidal properties against *Cx. quinquefasciatus* and found to be safe for the environment [87]. For several decades, *N. glauca* has been known for its content of the pyridine alkaloids, such as anabasine, nicotine, and nornicotine. In the *P. rapae* larval bioassay, the median effective concentrations of anabasine were 0.572 mg/larva. Despite this, the insecticidal activities of the *N. glauca* extract are likely related to anabasine, as several phytochemical experiments and bioassays have shown [88]. This review intended to collect all of the scientific data on mosquitocidal, insecticidal, and larvicidal investigations on medicinally important *Solanum* compounds including steroidal alkaloids. Because most of the studies are laboratory-based and do not meet clinical standards, this comprehensive review is expected to bolster investigators in furthering their research into this field, which could lead to the development of plant-based mosquito repellents with significant economic benefits.

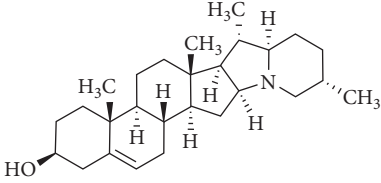
3.4. Solanum Plant-Mediated Nanoparticles. Plant extract-based silver nanoparticles have recently been developed to improve the control of mosquitoes without causing any significant harm to humans [114, 115]. According to the recent literature, silver nanoparticles (AgNPs) synthesized from aqueous extracts of *S. nigrum* and *S. mammosum* have



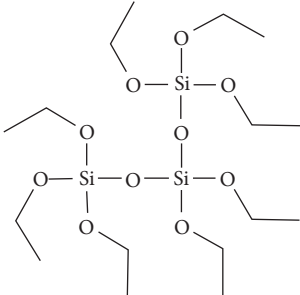
(a)
 FIGURE 2: Continued.



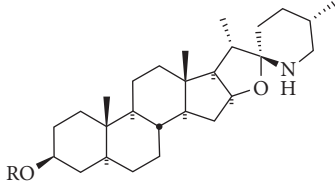
Permethrin



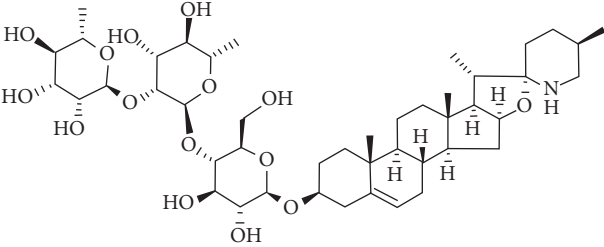
Solanidine



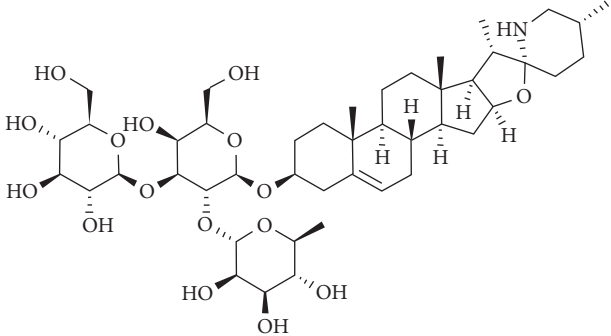
Silicic acid, diethyl bis (trimethylsilyl) ester



Tomatidine (TO) and Tomatine (TN). For TO, R=H, For TN, R=Lycotetraose

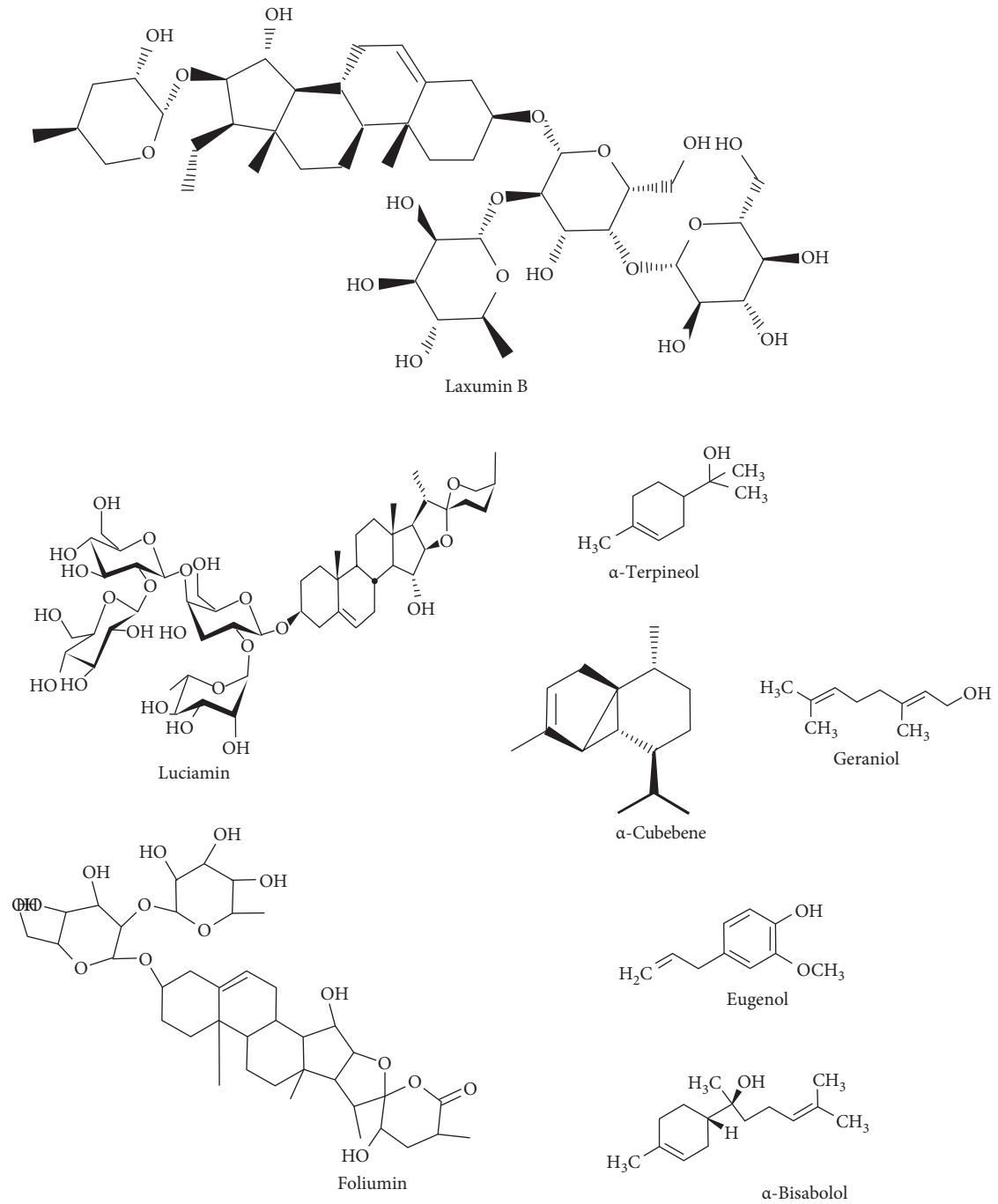


Solamargine



Solasonine

(b)
FIGURE 2: Continued.



(c)

FIGURE 2: Continued.

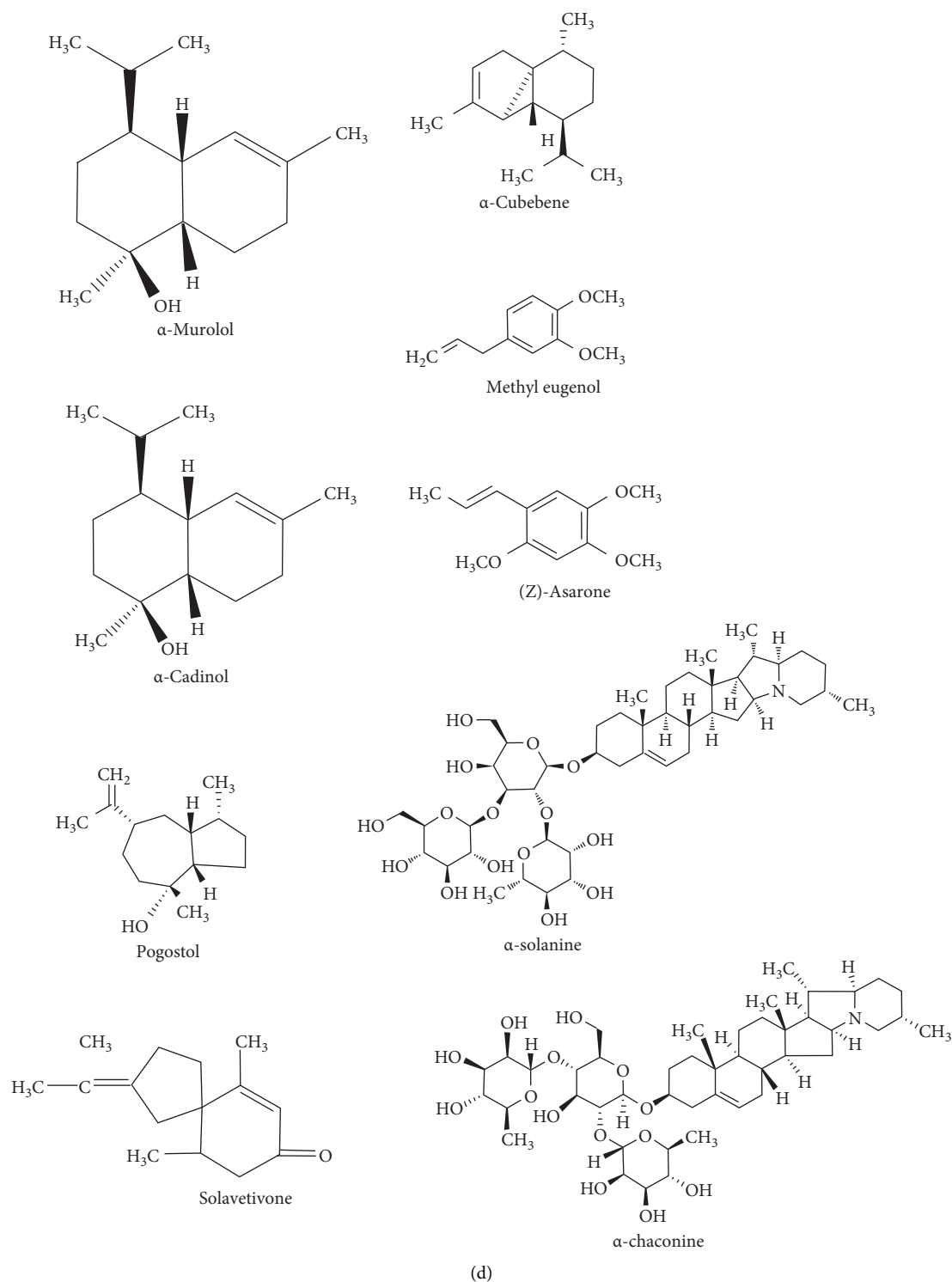


FIGURE 2: Chemical structures of insecticidal and mosquitocidal phytochemicals isolated from various *Solanum* species.

demonstrated adulticidal and insecticidal activity against *Ae. aegypti*, implying that AgNPs produced from plant extracts have higher levels of toxicity than the extracts alone [55, 76]. Thus, these herbal-based AgNPs hold great promise as potent larvicides, but their environmental impact requires further investigation for controlling target vector mosquitoes.

4. Conclusion

Mosquito control is an important public health policy in tropical areas. Mosquitoes constitute a part of natural biodiversity, and their total eradication is not necessary, but only the diseases they transmit need to be eradicated. Nevertheless, bites from disease-bearing insects can be

minimized and even avoided altogether. Inappropriate application of chemical insecticides in insect pest control can lead to insect resistance and pose environmental hazards and raises pest resistance to insecticides. Plants contain a variety of larvicidal secondary metabolites, and given their low environmental impact and minimal toxicity to humans, medicinal plants present a promising alternative to synthetic pesticides [116,117]. *Solanum* spp. constitute a large and diverse genus (~2,000 species) of flowering plants, which provide food sources, eggplant, potato and tomato, ornamental flowers and fruits, and herbal medications. *Solanum* species grow in various habitats and can be annuals and perennials, vines, subshrubs, shrubs, and small trees.

Solanum spp. contain a diversity of phytochemicals that can be culled for their insecticidal properties, particularly mosquito larvicide, adulticide, and repellent. Although crude extracts have higher insecticidal potency than pure components, probably due to synergy among their bioactive constituents, optimal utilization of crude extracts is limited by an inability to control their contents, which can vary depending on plant species, cultivation conditions, the season of harvest, and extraction methods and solvents used. Thus, an understanding of the mechanism of insecticidal activity of the major bioactive compounds can lead to consistent and optimal formulation and adjustment to match the target insect pests of interest.

The review highlights current knowledge of phytochemicals (glycoalkaloids, phytoosteroids, plant proteins, and volatile oils) reported as larvicides, adulticides, and repellents against a variety of insect pests, against vectors of important human diseases (dengue, filaria, and malaria). The exquisitely low larvicidal activity of silver nanoparticles formed from plant fruit and aqueous leaf extracts indicates the synergism between traditional medicinal herbal knowledge and modern (nano)technology. As a result, the usage of environmentally beneficial and cost-free plant-based products for insect/mosquito control is now unavoidable. Although the evaluation of phytochemicals is still in its early stages, with much more research needed to characterize promising agents and discover new ones, some of the findings presented in this review suggest that *Solanum* genus-based botanical phytochemicals should not be dismissed as a potential future alternative to synthetic insecticides. Hence, *Solanaceae* plants should be mined for their inexpensive, eco-friendly, safe, and effective alternatives to current chemical larvicides.

Data Availability

The data used to support the findings of the study can be obtained from the corresponding author upon request.

Conflicts of Interest

The authors declare no conflicts of interest.

Acknowledgments

The authors extend their appreciation to the Deanship of Scientific Research at King Khalid University for supporting through Large Groups (RGP.2/52/1443).

References

- [1] L. S. Tusting, J. Thwing, D. Sinclair et al., "Mosquito larval source management for controlling malaria," *Cochrane Database of Systematic Reviews*, vol. 8, Article ID CD008923, 2013.
- [2] L. H. V. Franklinos, K. E. Jones, D. W. Redding, and I. Abubakar, "The effect of global change on mosquito-borne disease," *The Lancet Infectious Diseases*, vol. 19, no. 9, pp. e302–312, 2019.
- [3] E. Massaro, D. Kondor, and C. Ratti, "Assessing the interplay between human mobility and mosquito borne diseases in urban environments," *Scientific Reports*, vol. 9, no. 1, Article ID 16911, 2019.
- [4] T. Iwamura, A. Guzman-Holst, and K. A. Murray, "Accelerating invasion potential of disease vector *Aedes aegypti* under climate change," *Nature Communications*, vol. 11, no. 1, p. 2130, 2020.
- [5] L. M. Turchen, L. Cosme-Junior, and R. N. C. Guedes, "Plant-derived insecticides under meta-analyses: status, biases, and knowledge gaps," *Insects*, vol. 11, no. 8, p. 532, 2020.
- [6] E. A. S. Shaalan, D. Canyon, M. W. F. Younes, H. Abdel-Wahab, and A. H. Mansour, "A review of botanical phytochemicals with mosquitocidal potential," *Environment International*, vol. 31, no. 8, pp. 1149–1166, 2005.
- [7] N. Kishore, B. B. Mishra, V. K. Tiwari, V. Tripathi, and N. Lall, "Natural products as leads to potential mosquitocides," *Phytochemistry Reviews*, vol. 13, no. 3, pp. 587–627, 2014.
- [8] A. T. H. Mossa, "Green pesticides: essential oils as biopesticides in insect-pest management," *Journal of Environmental Science and Technology*, vol. 9, no. 5, pp. 354–378, 2016.
- [9] A. Asadollahi, M. Khoobdel, A. Zahraei-Ramazani, S. Azarmi, and S. H. Mosawi, "Effectiveness of plant-based repellents against different *Anopheles* species: a systematic review," *Malaria Journal*, vol. 18, no. 1, p. 436, 2019.
- [10] R. Pavela, F. Maggi, R. Iannarelli, and G. Benelli, "Plant extracts for developing mosquito larvicides: from laboratory to the field, with insights on the modes of action," *Acta Tropica*, vol. 193, pp. 236–271, 2019.
- [11] M. Piplani, D. P. Bhagwat, G. Singhvi et al., "Plant-based larvicidal agents: an overview from 2000 to 2018," *Experimental Parasitology*, vol. 199, pp. 92–103, 2019.
- [12] D. Bekele, "Review on insecticidal and repellent activity of plant products for malaria mosquito control," *Biomedical Research and Reviews*, vol. 2, pp. 1–7, 2018.
- [13] J. S. Kaunda and Y. J. Zhang, "The genus *Solanum*: an ethnopharmacological, phytochemical and biological properties review," *Natural Products and Bioprospecting*, vol. 9, pp. 77–137, 2019.
- [14] C. A. Elizalde-Romero, L. A. Montoya-Inzunza, L. A. Contreras-Angulo, J. B. Heredia, and E. P. Gutierrez-Grijalva, "*Solanum* Fruits: phytochemicals, bio-accessibility and bioavailability, and their relationship with their health-promoting effects," *Frontiers in Nutrition*, vol. 8, Article ID 790582, 2021.
- [15] K. Jayakumar and K. Murugan, "Solanum alkaloids and their pharmaceutical roles: a review," *Journal of Analytical & Pharmaceutical Research*, vol. 3, no. 6, Article ID 00075, 2015.
- [16] K. Roshan, K. Mona, H. Rupali, and U. Milind, "A review of the phytochemistry and pharmacological activities of

- solanaceae family,” *International Journal of Pharmaceutical Chemistry Research*, vol. 2, pp. 13–19, 2020.
- [17] V. A. Royo, L. J. Matias, J. Rocha, E. Menezes, AdM. Junior, and D. de Oliveira, “Phytochemistry in medicinal species of solanum L. (*Solanaceae*),” *Pharmacognosy Research*, vol. 11, no. 1, pp. 47–50, 2019.
- [18] G. A. El-Shaboury, H. M. Al-Wadi, and A. Badr, “Biodiversity of some *Solanum* species from southwestern Saudi Arabia’s highlands,” *Botany Letters*, vol. 168, no. 2, pp. 246–255, 2021.
- [19] T. M. Silva, M. G. Carvalho, R. Braz-Filho, and M. F. Agra, “Occurrence of flavones, flavonols and its glycosides in species of the genus *Solanum* (*Solanaceae*),” *Quimica Nova*, vol. 26, pp. 517–522, 2003.
- [20] T. Atul and S. Ray, “Certain medicinal plants of *solanaceae* and their alkaloids screening,” *International Journal of Research in Medical Sciences*, vol. 2, pp. 4–6, 2014.
- [21] S. Chowanski, Z. Adamski, P. Marciniak et al., “A review of bioinsecticidal activity of *Solanaceae* alkaloids,” *Toxins*, vol. 8, no. 3, p. 60, 2016.
- [22] A. Dafni and Z. Yaniv, “*Solanaceae* as medicinal plants in Israel,” *Journal of Ethnopharmacology*, vol. 44, no. 1, pp. 11–18, 1994.
- [23] A. A. Awan and G. Murtaza, “Ethnobotanical uses of plants of family solanaceae muzaffarabad division azad Jammu and Kashmir, Pakistan-13100,” *International Journal of Pharmaceutical Science Invention*, vol. 2, pp. 5–11, 2013.
- [24] R. S. Meyer, B. D. Whitaker, D. P. Little et al., “Parallel reductions in phenolic constituents resulting from the domestication of eggplant,” *Phytochemistry*, vol. 115, pp. 194–206, 2015.
- [25] A. B. Pomilio, E. M. Falzoni, and A. A. Vitale, “Toxic chemical compounds of the Solanaceae,” *Natural Product Communications*, vol. 3, no. 4, Article ID 1934578X0800300, 2008.
- [26] M. M. Rigano, G. De Guzman, A. M. Walmsley, L. Frusciante, and A. Barone, “Production of pharmaceutical propeptides in *Solanaceae* food crops,” *International Journal of Molecular Sciences*, vol. 14, no. 2, pp. 2753–2773, 2013.
- [27] R. Jain, A. Sharma, S. Gupta, I. P. Sarethy, and R. Gabrani, “*Solanum nigrum*: current perspectives on therapeutic properties,” *Alternative Medicine Review: A Journal of Clinical Therapeutic*, vol. 16, no. 1, pp. 78–85, 2011.
- [28] S. E. Milner, N. P. Brunton, P. W. Jones, N. M. O’ Brien, S. G. Collins, and A. R. Maguire, “Bioactivities of glycoalkaloids and their aglycones from *Solanum* species,” *Journal of Agricultural and Food Chemistry*, vol. 59, no. 8, pp. 3454–3484, 2011.
- [29] M. Kumar, M. Tomar, D. J. Bhuyan et al., “Tomato (*Solanum lycopersicum* L.) seed: a review on bioactives and biomedical activities,” *Biomedicine & Pharmacotherapy*, vol. 142, Article ID 112018, 2021.
- [30] P. Shukla, K. Bajpai, S. Tripathi, S. Kumar, and G. K. Gautam, “A review on the taxonomy, ethnobotany, chemistry and pharmacology of solanum lycopersicum linn,” *International Journal of Chemistry and Pharmaceutical Sciences*, vol. 8, pp. 521–527, 2013.
- [31] G. K. More, “A review of the ethnopharmacology, phytochemistry and pharmacological relevance of the South African weed solanum sisymbriifolium Lam. (*Solanaceae*),” *Environment, Development and Sustainability*, vol. 21, no. 1, pp. 37–50, 2019.
- [32] M. E. Camire, S. Kubow, and D. J. Donnelly, “Potatoes and human health,” *Critical Reviews in Food Science and Nutrition*, vol. 49, no. 10, pp. 823–840, 2009.
- [33] H. Hammami and A. Ayadi, “Molluscicidal and antiparasitic activity of solanum nigrum villosum against *Galba truncatula* infected or uninfected with fasciola hepatica,” *Journal of Helminthology*, vol. 82, no. 3, pp. 235–239, 2008.
- [34] L. Wang, S. Y. Chiou, Y. T. Shen, F. T. Yen, H. Y. Ding, and M. J. Wu, “Anti-inflammatory effect and mechanism of the green fruit extract of *Solanum integrifolium* poir,” *BioMed Research International*, vol. 2014, Article ID 953873, 11 pages, 2014.
- [35] S. Govindan, S. Viswanathan, V. Vijayasekaran, and R. Alagappan, “A pilot study on the clinical efficacy of Solanum xanthocarpum and Solanum trilobatum in bronchial asthma,” *Journal of Ethnopharmacology*, vol. 66, no. 2, pp. 205–210, 1999.
- [36] S. Parmar, A. Gangwal, and N. Sheth, “*Solanum xanthocarpum* (yellow berried night shade): A review,” *Der Pharmacia Lettre*, vol. 2, pp. 373–383, 2010.
- [37] M. S. Ranjith, A. J. A. Ranjitsingh, S. G. Shankar et al., “*Solanum trilobatum* in the management of atopy: through inhibition of mast cell degranulation and moderation of release of interleukins,” *Pharmacognosy Research*, vol. 2, no. 1, pp. 10–14, 2010.
- [38] P. P. Balakrishnan, T. Ansari, T. A. M. Gani, S. Sreenath, and S. Kumaran, “A perspective on bioactive compounds from solanum trilobatum,” *Journal of Chemical and Pharmaceutical Research*, vol. 78, pp. 507–512, 2015.
- [39] J. Sahu, B. Rathi, S. Koul, and R. L. Khosa, “*Solanum trilobatum* (*Solanaceae*)—an overview,” *Journal of Natural Remedies*, vol. 13, pp. 76–80, 2013.
- [40] M. H. Rane, N. K. Sahu, S. S. Ajoankar, N. C. Teli, and D. R. Verma, “A holistic approach on review of solanum virginianum. L,” *Research and Reviews in Pharmacy and Pharmaceutical Sciences*, vol. 3, pp. 1–4, 2014.
- [41] B. Cabanillas, F. Chassagne, P. Vasquez-Ocmin et al., “Pharmacological validation of solanum mammosum L. as an anti-infective agent: role of solamargine,” *Journal of Ethnopharmacology*, vol. 280, Article ID 114473, 2021.
- [42] M. A. Buabeid, E. S. A. Arafa, T. Rani et al., “Effects of solanum lycopersicum L. (tomato) against isoniazid and rifampicin induced hepatotoxicity in wistar albino rats,” *Brazilian Journal of Biology*, vol. 84, Article ID e254552, 2022.
- [43] U. W. Hawas, G. M. Soliman, L. T. A. El-Kassem, A. R. H. Farrag, K. Mahmoud, and F. Leon, “A new flavonoid C-glycoside from solanum elaeagnifolium with hepatoprotective and curative activities against paracetamol-induced liver injury in mice,” *Zeitschrift für Naturforschung C*, vol. 68, pp. 0019–0028, 2013.
- [44] A. Al-Marby, C. Ejike, M. J. Nasim et al., “Nematicidal and antimicrobial activities of methanol extracts of seventeen plants, of importance in ethnopharmacology, obtained from the Arabian peninsula,” *Journal of Intercultural Ethnopharmacology*, vol. 5, no. 2, pp. 114–121, 2016.
- [45] S. Badami, S. A. Manohara Reddy, E. P. Kumar, P. Vijayan, and B. Suresh, “Antitumor activity of total alkaloid fraction of solanum pseudocapsicum leaves,” *Phytotherapy Research*, vol. 17, no. 9, pp. 1001–1004, 2003.
- [46] A. D. Agrawal, P. S. Bajpei, A. A. Patil, and S. R. Bavaskar, “*Solanum torvum* sw.-a phytopharmacological review,” *Der Pharmacia Lettre*, vol. 2, pp. 403–407, 2010.

- [47] L. X. Zhou and Y. Ding, "A cinnamide derivative from *Solanum verbascifolium* L.," *Journal of Asian Natural Products Research*, vol. 4, no. 3, pp. 185–187, 2002.
- [48] W. Raja, G. Nosalova, K. Ghosh, V. Sivova, S. Nosal, and B. Ray, "In-vivo antitussive activity of a pectic arabinogalactan isolated from *Solanum virginianum* L. in guinea pigs," *Journal of Ethnopharmacology*, vol. 156, pp. 41–46, 2014.
- [49] R. W. Bussmann, G. G. Gilbreath, J. Solio et al., "Plant use of the Maasai of sekenani valley, Maasai mara, Kenya," *Journal of Ethnobiology and Ethnomedicine*, vol. 2, no. 1, p. 22, 2006.
- [50] M. Houda, S. Derbre, A. Jedy et al., "Combined anti-ages and antioxidant activities of different solvent extracts of *Solanum elaeagnifolium* cav (*Solanaceae*) fruits during ripening and related to their phytochemical compositions," *EXCLI Journal*, vol. 13, pp. 1029–1042, 2014.
- [51] R. S. Meyer, M. Bamshad, D. Q. Fuller, and A. Litt, "Comparing medicinal uses of eggplant and related Solanaceae in China, India, and the Philippines suggests the independent development of uses, cultural diffusion, and recent species substitutions," *Economic Botany*, vol. 68, no. 2, pp. 137–152, 2014.
- [52] M. Afroz, S. Akter, A. Ahmed et al., "Ethnobotany and antimicrobial peptides from plants of the *Solanaceae* family: an update and future prospects," *Frontiers in Pharmacology*, vol. 11, p. 565, 2020.
- [53] J. J. M. Bagalwa, L. Voutquenne-nazabadioko, C. Sayagh, and A. S. Bashwira, "Evaluation of the biological activity of the molluscicidal fraction of *Solanum sisymbriifolium* against non-target organisms," *Fitoterapia*, vol. 81, no. 7, pp. 767–771, 2010.
- [54] S. Singha and G. Chandra, "Mosquito larvicidal activity of some common spices and vegetable waste on *Culex quinquefasciatus* and *Anopheles stephensi*," *Asian Pacific Journal of Tropical Medicine*, vol. 4, pp. 288–293, 2011.
- [55] A. Rawani, A. Ghosh, and G. Chandra, "Mosquito larvicidal and antimicrobial activity of synthesized nano-crystalline silver particles using leaves and green berry extract of *Solanum nigrum* L. (*Solanaceae*: solanales)," *Acta Tropica*, vol. 128, no. 3, pp. 613–622, 2013.
- [56] A. Rawani, A. S. Ray, A. Ghosh, M. Sakar, and G. Chandra, "Larvicidal activity of phytosteroid compounds from leaf extract of *Solanum nigrum* against *Culex vishnui* group and *Anopheles subpictus*," *BMC Research Notes*, vol. 10, no. 1, p. 135, 2017.
- [57] A. Rawani, A. Ghosh, and G. Chandra, "Mosquito larvicidal activities of *Solanum nigrum* L. leaf extract against *Culex quinquefasciatus* say," *Parasitology Research*, vol. 107, no. 5, pp. 1235–1240, 2010.
- [58] N. Chowdhury, I. Bhattacharjee, S. Laskar, and G. Chandra, "Efficacy of *Solanum villosum* mill. (*Solanaceae*: solanales) as a biocontrol agent against fourth instar larvae of *Culex quinquefasciatus* say," *Turkish Journal of Zoology*, vol. 31, pp. 365–370, 2007.
- [59] N. Chowdhury, S. K. Chatterjee, S. Laskar, and G. Chandra, "Larvicidal activity of *Solanum villosum* mill (*Solanaceae*: solanales) leaves to *Anopheles subpictus* grassi (diptera: culicidae) with effect on non-target *Chironomus circumdatus* kieffer (diptera: chironomidae)," *Journal of Pest Science*, vol. 82, no. 1, pp. 13–18, 2009.
- [60] C. S. Chen, C. Y. Chen, D. M. Ravinath, A. Bungahot, C. P. Cheng, and R. I. You, "Functional characterization of chitin-binding lectin from *Solanum integrifolium* containing anti-fungal and insecticidal activities," *BMC Plant Biology*, vol. 18, p. 1, 2018.
- [61] P. Mahesh Kumar, K. Murugan, K. Kovendan, J. Subramaniam, and D. Amaresan, "Mosquito larvicidal and pupicidal efficacy of *Solanum xanthocarpum* (Family: solanaceae) leaf extract and bacterial insecticide, *Bacillus thuringiensis*, against *Culex quinquefasciatus* say (diptera: culicidae)," *Parasitology Research*, vol. 110, no. 6, pp. 2541–2550, 2012.
- [62] I. A. Simon-Oke, O. J. Afolabi, and O. T. Ajayi, "Larvicidal activity of a perennial herb, *Solanum xanthocarpum* against the larvae of culicine species," *FUTA Journal of Research in Sciences*, vol. 1, pp. 152–162, 2015.
- [63] P. Mahesh Kumar, K. Murugan, K. Kovendan et al., "Mosquitocidal activity of *Solanum xanthocarpum* fruit extract and copepod *Mesocyclops thermocyclopoides* for the control of dengue vector *Aedes aegypti*," *Parasitology Research*, vol. 111, no. 2, pp. 609–618, 2012.
- [64] M. Govindarajan, "Larvicidal and repellent properties of some essential oils against *Culex tritaeniorhynchus* giles and *Anopheles subpictus* grassi (diptera: culicidae)," *Asian Pacific Journal of Tropical Medicine*, vol. 4, no. 2, pp. 106–111, 2011.
- [65] S. K. Bansal, K. V. Singh, and H. Sharma, "Synergistic efficacy of *Solanum xanthocarpum* and *Withania somnifera* on larvae of mosquito vector species," *Journal of Environmental Biology*, vol. 36, pp. 633–638, 2015.
- [66] L. Mohan, P. Sharma, and C. N. Srivastava, "Evaluation of *Solanum xanthocarpum* extract as a synergist for cypermethrin against larvae of the filarial vector *C. quinquefasciatus* (say)," *Entomological Research*, vol. 36, no. 4, pp. 220–225, 2006.
- [67] L. Mohan, P. Sharma, and C. N. Srivastava, "Comparative efficacy of *Solanum xanthocarpum* extracts alone and in combination with a synthetic pyrethroid, cypermethrin, against malaria vector, *Anopheles stephensi*," *Southeast Asian Journal of Tropical Medicine and Public Health*, vol. 38, no. 2, pp. 256–260, 2007.
- [68] L. Mohan, P. Sharma, and C. N. Srivastava, "Combination larvicidal action of *Solanum xanthocarpum* extract and certain synthetic insecticides against filarial vector, *Culex quinquefasciatus* (SAY)," *Southeast Asian Journal of Tropical Medicine and Public Health*, vol. 41, no. 2, pp. 311–319, 2010.
- [69] L. Mohan, P. Sharma Shrankhla, and C. N. Srivastava, "Relative toxicity of different combinations of temephos and fenthion with *Solanum xanthocarpum* extract against anopheline larvae," *Entomological Research*, vol. 43, no. 4, pp. 236–242, 2013.
- [70] A. Rawani, A. Ghosh, and G. Chandra, "Laboratory evaluation of molluscicidal & mosquito larvicidal activities of leaves of *Solanum nigrum* L.," *Indian Journal of Medical Research*, vol. 140, no. 2, pp. 285–295, 2014.
- [71] A. Rawani, A. Ghosh, S. Laskar, and G. Chandra, "Glucosinolate from leaf of *Solanum nigrum* L. (*Solanaceae*) as a new mosquito larvicide," *Parasitology Research*, vol. 113, no. 12, pp. 4423–4430, 2014.
- [72] S. Rajkumar and A. Jebanesan, "Oviposition deterrent and skin repellent activities of *Solanum trilobatum* leaf extract against the malarial vector *Anopheles stephensi*," *Journal of Insect Science*, vol. 5, no. 1, p. 15, 2005.
- [73] S. Premalatha, K. Elumalai, and A. Jeyasankar, "Mosquitocidal properties of *Solanum trilobatum* L. (*Solanaceae*) leaf extracts against three important human vector mosquitoes (diptera: culicidae)," *Asian Pacific Journal of Tropical Medicine*, vol. 6, no. 11, pp. 854–858, 2013.
- [74] A. Lalitha and S. Thangapandiyam, "Mosquito larvicidal efficacy of the acetone leaf extract of *Solanum trilobatum*

- against *Culex quinquefasciatus* and *Aedes aegypti*,” *Asian Journal of Pharmaceutical and Clinical Research*, vol. 11, no. 12, pp. 273–276, 2018.
- [75] A. A. Zahir, A. A. Rahuman, A. Bagavan et al., “Evaluation of botanical extracts against *Haemaphysalis bispinosa neumann* and *Hippobosca maculata leach*,” *Parasitology Research*, vol. 107, no. 3, pp. 585–592, 2010.
- [76] F. Pilaquinga, B. Morejon, D. Ganchala et al., “Green synthesis of silver nanoparticles using solanum mammosum L. (*Solanaceae*) fruit extract and their larvicidal activity against *Aedes aegypti* L. (diptera: culicidae),” *PLoS One*, vol. 14, no. 10, Article ID e0224109, 2019.
- [77] M. Maragatham and D. Joseph, “Larvicidal evaluation of aged and fresh leaf extract of solanum lycopersicum esculentum,” *International Journal of Mosquito Research*, vol. 6, pp. 57–60, 2019.
- [78] M. Markouk, K. Bekkouche, M. Larhsini, M. Bousaid, H. Lazrek, and M. Jana, “Evaluation of some moroccan medicinal plant extracts for larvicidal activity,” *Journal of Ethnopharmacology*, vol. 73, no. 1-2, pp. 293–297, 2000.
- [79] A. B. Hamouda, K. Zarrad, I. Chaieb, and A. Laarif, “Insecticidal effect of solanum elaeagnifolium extracts under laboratory conditions,” *Journal of Entomology and Zoology Studies*, vol. 3, pp. 187–190, 2015.
- [80] G. Elsayed, “Insecticidal effect of plant extracts on two termite species,” *Archives of Phytopathology and Plant Protection*, vol. 44, pp. 356–361, 2011.
- [81] S. Soule, C. Guntner, A. Vazquez, V. H. Argandona, F. Ferreira, and P. Moyna, “Effect of solanum glycosides on the aphid *Schizaphis graminum*,” *Journal of Chemical Ecology*, vol. 25, no. 2, pp. 369–374, 1999.
- [82] S. Soule, C. Guntner, A. Vazquez, V. V. Argandona, P. Moyna, and F. Ferreira, “An aphid repellent glycoside from solanum laxum,” *Phytochemistry*, vol. 55, no. 3, pp. 217–222, 2000.
- [83] Y. Schlein, R. L. Jacobson, and G. C. Muller, “Sand fly feeding on noxious plants: a potential method for the control of leishmaniasis,” *The American Journal of Tropical Medicine and Hygiene*, vol. 65, no. 4, pp. 300–303, 2001.
- [84] M. Srivastava and L. Gupta, “Effect of formulations of solanum surratense (family: solanaceae) an Indian desert plant on oviposition by the pulse beetle *Callosobruchus chinensis* linn,” *African Journal of Agricultural Research*, vol. 2, pp. 552–554, 2007.
- [85] C. Kamaraj, A. Abdul Rahman, A. Bagavan et al., “Larvicidal efficacy of medicinal plant extracts against *Anopheles stephensi* and *Culex quinquefasciatus* (Diptera: Culicidae),” *Tropical Biomedicine*, vol. 27, no. 2, pp. 211–219, 2010.
- [86] S. Arivoli, T. Narendran, and S. Ignacimuthu, “Larvicidal activity of some botanicals against *Culex quinquefasciatus* say,” *Journal of Advanced Zoology*, vol. 21, pp. 19–23, 2000.
- [87] T. M. S. Silva, C. A. Camara, K. R. L. Freire, T. Gd. Silva, M. F. Agra, and J. Bhattacharyya, “Steroidal glycoalkaloids and molluscicidal activity of solanum asperum rich fruits,” *Journal of the Brazilian Chemical Society*, vol. 19, no. 5, pp. 1048–1052, 2008.
- [88] M. Zammit, C. Shoemake, E. Attard, and L. M. Azzopardi, “The effects of anabasine and the alkaloid extract of *Nicotiana glauca* on Lepidopterous larvae,” *International Journal of Biology*, vol. 6, no. 3, pp. 46–53, 2014.
- [89] A. B. Hamouda, I. Chaieb, K. Zarrad, and A. Laarif, “Insecticidal activity of methanolic extract of silverleaf nightshade against *Tribolium castaneum*,” *International Journal of Entomological Research*, vol. 3, pp. 23–28, 2015.
- [90] N. Chowdhury, A. Ghosh, and G. Chandra, “Mosquito larvicidal activities of solanum villosum berry extract against the dengue vector *Stegomyia aegypti*,” *BMC Complementary and Alternative Medicine*, vol. 8, no. 1, p. 10, 2008.
- [91] K. Baskar, J. Ananthi, and S. Ignacimuthu, “Toxic effects of solanum xanthocarpum sch & wendle against *Helicoverpa armigera* (hub), *Culex quinquefasciatus* (say) and *Eisenia fetida* (savigny, 1826),” *Environmental Science and Pollution Research Science and Pollution Research International*, vol. 25, no. 3, pp. 2774–2782, 2018.
- [92] P. K. Tr, R. Hl, R. Mr, A. Hc, A. Gn, and K. Kn, “Antimicrobial, insecticidal, and antiradical activity of solanum virginianum L. (*Solanaceae*),” *Asian Journal of Pharmaceutical and Clinical Research*, vol. 10, no. 11, pp. 163–167, 2017.
- [93] M. Kumar Patel, A. Tiwari, and V. Laxmi Saxena, “Larvicidal activity of crude solanum nigrum leaf and berries extract against dengue vector-*Aedes aegypti*,” *International Journal of Current Research and Review*, vol. 10, no. 14, pp. 16–21, 2018.
- [94] S. K. Bansal, K. V. Singh, and M. R. K. Sherwani, “Evaluation of larvicidal efficacy of solanum xanthocarpum storage against vector mosquitoes in north-western Rajasthan,” *Journal of Environmental Biology*, vol. 30, no. 5, pp. 883–888, 2009.
- [95] T. Changbunjong, W. Wongwit, S. Leemingsawat, Y. Tongtokit, and V. Deesin, “Effect of crude extract of solanum xanthocarpum against snails and mosquito larvae,” *The Southeast Asian Journal of Tropical Medicine and Public Health*, vol. 41, no. 2, pp. 320–325, 2010.
- [96] M. Sakthivadivel, P. Gunasekaran, M. Sivakumar, S. Arul, S. Arivoli, and T. Samuel, “Evaluation of solanum trilobatum L. (*Solanaceae*) aerial extracts for mosquito larvicidal activity against the filarial vector *Culex quinquefasciatus* say (diptera: culicidae),” *Journal of Entomology and Zoology Studies*, vol. 2, pp. 102–106, 2014.
- [97] L. Mohan, P. Sharma, and C. N. Srivastava, “Evaluation of Solanum xanthocarpum extracts as mosquito larvicides,” *Journal of Environmental Biology*, vol. 26, no. 2, pp. 399–401, 2005.
- [98] K. V. Singh and S. K. Bansal, “Larvicidal properties of a perennial herb solanum xanthocarpum against vectors of malaria and dengue/DHF,” *Current Science*, vol. 84, pp. 749–751, 2003.
- [99] S. P. Singh, K. Raghavendra, R. K. Singh, and S. K. Subbarao, “Studies on larvicidal properties of leaf extract of solanum nigrum linn. (family: solanaceae),” *Current Science*, vol. 81, pp. 1529–1530, 2001.
- [100] C. S. Chopra, V. Benzi, R. Alzogaray, and A. A. Ferrero, “Repellent activity of hexanic and ethanolic extracts from fruits of solanum eleagnifolium (*Solanaceae*) against *Blattella germanica* (insecta, dictyoptera, blattellidae) adults,” *Latin American and Caribbean Bulletin of Medicinal and Aromatic Plants*, vol. 8, pp. 172–175, 2009.
- [101] F. Njeh, H. Feki, I. Koubaa et al., “Molluscicidal activity of solanum elaeagnifolium seeds against *Galba truncatula* intermediate host of *Fasciolahepatica*: identification of β -solamarine,” *Pharmaceutical Biology*, vol. 54, no. 4, pp. 726–731, 2016.
- [102] K. Raghavendra, S. P. Singh, S. K. Subbarao, and A. P. Dash, “Laboratory studies on mosquito larvicidal efficacy of aqueous & hexane extracts of dried fruit of solanum nigrum

- linn,” *Indian Journal of Medical Research*, vol. 130, no. 1, pp. 74–77, 2009.
- [103] A. Jeyasankar, S. Premalatha, and K. Elumalai, “Biological activities of solanum pseudocapsicum (solanaceae) against cotton bollworm, *Helicoverpa armigera* hubner and armyworm, *Spodoptera litura* fabricius (lepidoptera: noctuidae),” *Asian Pacific Journal of Tropical Biomedicine*, vol. 2, no. 12, pp. 981–986, 2012a.
- [104] A. Jeyasankar, S. Premalatha, and S. Jancy Rani, “Bio-efficacy of solanum pseudocapsicum L. (*Solanaceae*) against black cutworm, *Agrotis ipsilon* hufnagel (lepidoptera: noctuidae),” *Journal of Biological Sciences*, vol. 12, no. 3, pp. 174–179, 2012b.
- [105] A. H. Ahmed and R. M. Ramzy, “Seasonal variation in molluscicidal activity of *Solanum nigrum* L,” *Journal of the Egyptian Society of Parasitology* *Egyptian Society of Parasitology*, vol. 28, no. 3, pp. 621–629, 1998.
- [106] A. H. Ahmed, I. H. Kamal, and R. M. Ramzy, “Studies on the molluscicidal and larvicidal properties of *Solanum nigrum* L. leaves ethanol extract,” *Journal of the Egyptian Society of Parasitology* *Egyptian Society of Parasitology*, vol. 31, no. 3, pp. 843–852, 2001.
- [107] J. Muthukrishnan, E. Pushpalatha, and A. Kasthuribhai, “Biological effects of four plant extracts on *Culex quinquefasciatus* Say larval stages,” *International Journal of Tropical Insect Science*, vol. 17, no. 3-4, pp. 389–394, 2011.
- [108] J. J. Hlywka, G. R. Stephenson, M. K. Sears, and R. Y. Yada, “Effect of insect damage on glycoalkaloid content in potatoes (*Solanum tuberosum*),” *Journal of Agricultural and Food Chemistry*, vol. 42, no. 11, pp. 2545–2550, 1994.
- [109] M. Weissenberg, A. Levy, J. A. Svoboda, and I. Ishaaya, “The effect of some solanum steroidal alkaloids and glycoalkaloids on larvae of the red flour beetle, *Tribolium castaneum*, and the tobacco hornworm, *manduca sexta*,” *Phytochemistry*, vol. 47, no. 2, pp. 203–209, 1998.
- [110] A. B. Hamouda, K. Zarrad, I. Chaieb, and A. Laarif, “Antifeedant and insecticidal properties of solanum elaeagnifolium extracts on the african cotton leafworm,” *Azarian Journal of Agriculture*, vol. 2, pp. 71–74, 2015.
- [111] P. Satyal, S. Maharjan, and W. N. Setzer, “Volatile constituents from the leaves, fruits (berries), stems and roots of solanum xanthocarpum from Nepal,” *Natural Product Communications*, vol. 10, no. 2, Article ID 1934578X1501000, 2015.
- [112] S. Rajkumar and A. Jebanesan, “Repellency of volatile oils from moschosma polystachyum and solanum xanthocarpum against filarial vector *Culex quinquefasciatus* say,” *Tropical Biomedicine*, vol. 22, pp. 139–142, 2005.
- [113] J. Jerzykiewicz, “Alkaloids of solanaceae (nightshade plants),” *Postepy Biochemii*, vol. 53, no. 3, pp. 280–286, 2007.
- [114] G. Suganya, S. Karthi, and M. S. Shivakumar, “Larvicidal potential of silver nanoparticles synthesized from *Leucas aspera* leaf extracts against dengue vector *Aedes aegypti*,” *Parasitology Research*, vol. 113, no. 5, pp. 1673–1679, 2014.
- [115] K. Veerakumar, M. Govindarajan, and M. Rajeswary, “Green synthesis of silver nanoparticles using *Sida acuta* (*Malvaceae*) leaf extract against *Culex quinquefasciatus*, *Anopheles stephensi*, and *Aedes aegypti* (diptera: culicidae),” *Parasitology Research*, vol. 112, no. 12, pp. 4073–4085, 2013.
- [116] A. Ghosh, N. Chowdhury, and G. Chandra, “Plant extracts as potential mosquito larvicide: a review,” *Indian Journal of Medical Research*, vol. 135, no. 5, pp. 581–598, 2012.
- [117] M. B. Isman, “Plant essential oils for pest and disease management,” *Crop Protection*, vol. 19, pp. 603–608, 2000.