

A new species of *Triplophysa* (Cypriniformes, Nemacheilidae) from Weihe River in Gansu Province, China

DEAR EDITOR,

A new species of Tibetan loach, *Triplophysa weiheensis* sp. nov., is described from the Weihe River in Gansu Province, China, based on morphological and molecular analyses. The new species can be distinguished from all known congeners by a unique combination of the following characters: scaleless; snout abruptly sloping downward, anterior to anterior nostril; lower jaw crescentic, not sharp; body without obvious mottling; lateral line interrupted on posterior trunk at pelvic-fin distal extremity; caudal-peduncle length 2.0–2.7 times its depth; branched rays of pectoral fin 10–11; branched rays of pelvic fin 5–6; inner gill rakers on 1st gill arch 14–16; vertebrae 4+34–36; intestine with 6–7 loops, length ca. 1.8 times SL ($n=3$); bony capsule of air bladder small and thin; posterior chamber of air bladder absent.

Species of the genus *Triplophysa* Rendahl 1933 are the most common fish among the nemacheilids found in the Qinghai-Tibet Plateau (QTP) (Wu & Wu, 1992; Zhu, 1989). These fish are found in almost all water bodies in the region, with new species of *Triplophysa* still being reported (Huang et al., 2019; Liu et al., 2017; Wu et al., 2018; Yang et al., 2016). As a result, a total of 147 valid species of *Triplophysa* have been recorded to date (Froese & Pauly, 2019).

Weihe River is a tributary of the Yellow River and originates from the southern part of Gansu Province (Figure 1A). Previous studies have reported that southern Gansu is a hotspot area for *Triplophysa*, with an extensive distribution of species across the river systems (Feng et al., 2017a, 2017b, 2019b). Until now, 17 species of *Triplophysa* have been reported from the Yellow River system (Chen et al., 1987; Ding, 1994; Feng et al., 2017a; Wu & Wu, 1992; Zhu, 1989), five of which have been recorded from Weihe River

(Supplementary Table S1). Following an investigation of *Triplophysa* species from Weihe River (Figure 1A), 15 specimens superficially resembling *Triplophysa stoliczkae* Steindachner 1866 (Supplementary Figure S1) were collected and are described herein as a new species based on morphological and molecular analyses.

After euthanization (see Supplementary Methods), the left ventral fin of some specimens was removed and preserved in 95% ethanol for DNA extraction. Voucher specimens were labeled and stored in 70% ethanol. Specimens were deposited in the collection of the Northwest Institute of Plateau Biology (NWIPB), Chinese Academy of Sciences, Xining, Qinghai, China. Morphological measurements and counts followed Kottelat (1990) and Prokofiev (2007). Additional measurements are described in the Supplementary Methods. Measurements were taken with digital calipers to the nearest 0.1 mm. Previous research has reported that *T. stoliczkae* is a striking case of morphological convergence and consists of distinct lineages that are not close relatives (Feng et al., 2019a). As there is no formal taxonomic revision for *T. stoliczkae*, we treated it as a morphological species in this study but considered its different genetic lineages in phylogeny. These lineages, which were initially mistaken as *T. stoliczkae*, exhibit very similar morphology. Thus, they represent a known morphological unit in the genus *Triplophysa*. We specifically measured 61 *T. stoliczkae* specimens collected from various water systems and used principal component analysis (PCA) (Supplementary Table S2) to visualize morphological differences between *T. stoliczkae* and the new species. Furthermore, we employed a Micro CT (Quantum GX2, PerkinElmer Corporation, USA) to build a skeletal model of the new species.

DNA extraction and complete *cyt b* gene (1140 bp) amplification were carried out, as detailed in the

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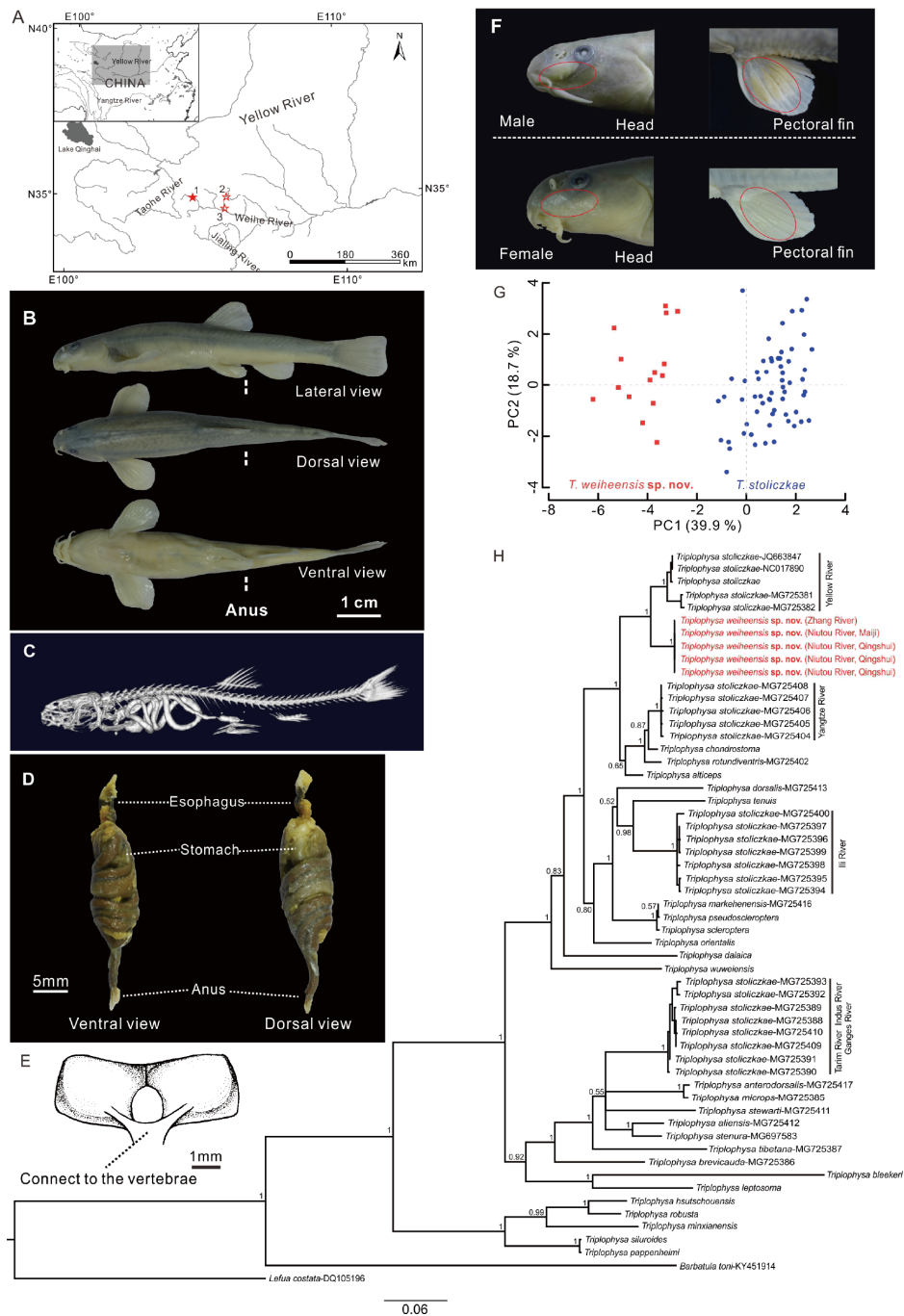


Figure 1 Collection site, morphological characters and phylogenetic position of *Triplophysa weiheensis* sp. nov.

A: Map showing sampling sites of *Triplophysa weiheensis* sp. nov. (star symbol, solid star for type locality). 1: Zhang River, at Chenjiamao Village, Zhangxian County; 2: Niutou River, at Hongbao Town, Qingshui County; 3: Niutou River, at Maiji District, Tianshui City. B: Lateral view, dorsal view, and ventral view of *Triplophysa weiheensis* sp. nov., NWIPB 1505189, holotype, 65.8 mm SL. C: Micro CT graph of skeletal system of *Triplophysa weiheensis* sp. nov. NWIPB 1505183, paratype, 64.1 mm SL. D: Structure of alimentary canal, *Triplophysa weiheensis* sp. nov., NWIPB 1505185, paratype, 65.5 mm SL. E: Dorsal view of bony capsule of air bladder, *Triplophysa weiheensis* sp. nov., NWIPB 1505185, paratype, 65.5 mm SL. F: Characters of sexual dimorphism of *Triplophysa weiheensis* sp. nov. Characters are highlighted in red circles. G: Morphological comparison between *Triplophysa weiheensis* sp. nov. and *T. stoliczkae* by PCA with 13 morphometric characters (Supplementary Table S2). H: Phylogenetic relationships of *Triplophysa* species based on *cyt b* gene sequences. Posterior probabilities (PP) obtained from Bayesian analyses are indicated on branches.

Supplementary Methods. Polymerase chain reaction (PCR) products were sequenced from both directions using an ABI PRISM 3700 sequencing system. Finally, 20 new *cyt b* sequences of *Triplophysa* were obtained, including five sequences from the new species (Supplementary Table S3). To assess the phylogenetic status of the new species, 37 additional *cyt b* sequences of *Triplophysa* and outgroups (*Barbatula toni* and *Lefua costata*) were retrieved from GenBank (Supplementary Table S3), as per previous studies (Feng et al., 2017b; He et al., 2006; Wang et al., 2016). Phylogenetic analysis was performed using Bayesian inference (BI) (Supplementary Methods). Additionally, Kimura's 2-parameter (K2P) genetic distances (Kimura, 1980) were estimated using MEGA 6.0 (Tamura et al., 2013).

Taxonomic account

Triplophysa weiheensis sp. nov. Feng, Zhang, Tong, Zhou et Zhao (Figure 1B–F; Table 1)

Holotype: NWIPB 1505189, 65.8 mm SL (standard length); Zhang River, a tributary of Weihe River, at Chenjiamo Village, Zhangxian County, Dingxi City, Gansu Province, China (N34°48', E104°31'; elevation 1 768 m a.s.l.), collected by Chenguang Feng, Chao Tong, and Kai Zhao on 12 May 2015.

Paratypes: NWIPB 1505181–183, 3 ex. (examined specimens), 64.1–77.1 mm SL; 1505185–188, 4 ex., 59.7–68.0 mm SL, same collection information as holotype. NWIPB 1505922, 53.6 mm SL, Niutou River, a tributary of Weihe River, at Maiji District, Tianshui City, Gansu Province, China (N34°34', E105°57'; elevation 1 360 m a.s.l.), on 21 May 2015. NWIPB 1505985–988, 4 ex., 56.4–65.9 mm SL; 1505990, 48.3 mm SL; 1505992, 60.5 mm SL, Niutou River, a tributary of Weihe River, at Hongbao Town, Qingshui County, Tianshui City, Gansu Province, China (N34°42', E106°1'; elevation 1 434 m a.s.l.), on 22 May 2015.

Etymology: The specific epithet *weiheensis* is derived from Weihe River (渭河 in Chinese, type locality) with the Latin suffix -ensis.

Table 1 Morphometric data of *Triplophysa weiheensis* sp. nov.

Morphometric character	Holotype	Paratype (Range n=14)	Mean	SD
Standard length (mm)	65.8	48.3–77.1		
Percentage of standard length (%)				
Body depth	18.1	16.2–19.2	18.2	0.9
Body width	16.5	14.1–17.6	16.0	1.0
Head length	17.9	16.3–21.5	19.6	1.7
Dorsal-fin length	16.9	14.1–20.6	17.7	1.9
Pelvic-fin length	14.2	12.2–14.7	13.6	0.7
Pectoral-fin length	18.2	15.1–20.5	17.3	1.8
Anal-fin length	15.2	13.1–17.1	15.0	1.2
Caudal-fin length	21.1	16.4–22.4	19.3	1.6
Predorsal length	49.5	47.9–55.1	50.9	2.2
Preanus length	68.5	61.9–71.0	67.3	2.6
Preanal length	72.2	63.5–73.4	69.8	2.5
Prepelvic length	56.7	51.1–58.4	54.6	2.4
Prepectoral length	19.7	16.9–21.9	19.8	1.3
Caudal-peduncle length	23.1	19.4–24.3	22.0	1.3
Caudal-peduncle depth	10.3	8.7–10.7	9.7	0.6
Pectoral-pelvic distance	32.5	31.6–38.8	35.2	2.4
Pectoral-anal distance	48.4	46.1–54.9	50.3	2.9
Pelvic-anal distance	15.5	12.7–18.1	15.3	1.7
Head length (mm)	11.8	10.0–14.0		
Percentage of head length (%)				
Head depth	67.4	59.2–72.2	63.5	4.4
Head width	81.9	68.1–91.3	79.7	8.0
Snout length	35.6	31.2–38.8	35.0	2.3
Eye length	19.4	16.6–22.9	18.7	1.7
Interorbital width	34.7	28.6–43.0	36.2	4.0
Postorbital length	50.6	41.4–53.0	48.4	3.1
Inner rostral barbel length	28.9	15.1–28.1	21.6	3.6
Outer rostral barbel length	27.0	16.7–27.5	22.9	2.9
Maxillary barbel length	31.4	18.7–28.9	23.4	3.2

Diagnosis: *Triplophysa weiheensis* sp. nov. can be distinguished from all known congeners by a combination of the following characters: (1) scaleless; (2) snout abruptly bending down before anterior nostril; (3) snout length shorter than postorbital length; (4) lower jaw crescentic, not sharp; (5) body without obvious mottling; (6) lateral line interrupted on posterior trunk behind vertical line of pelvic-fin distal extremity; (7) caudal-peduncle length 2.0–2.7 times its depth; (8) caudal fin slightly emarginate; (9) branched rays of pectoral fin 10–11; (10) pelvic-fin insertion behind vertical line through dorsal-fin origin, distal fin tip attaining anal-fin origin when adpressed, branched rays 5–6; (11) inner gill rakers on 1st gill arch 14–16; (12) vertebrae 4+34–36; (13) intestine with 6–7 loops, length ca. 1.8 times SL; (14) posterior chamber of air bladder absent, bony capsule of air bladder small and thin.

Description: Morphometric and meristic data are given in Table 1 and Supplementary Table S4, respectively.

Body thick, cylindrical. Dorsal profile of body arch-like (Figure 1B). Maximum depth of body slightly greater than maximum width, occurring between pectoral and dorsal fins. Caudal peduncle laterally compressed, depth nearly uniform toward caudal-fin base, length longer than head length (HL, 101.0–133.7% of HL). Head width greater than depth. Cheeks slightly inflated, V-shaped outline in ventral view. Snout obtuse, sloping downward anterior to anterior nostril (Figure 1B, C). Snout length shorter than postorbital length. Anterior and posterior nostrils close together. Valves around anterior nostrils, but not around posterior. Eyes small, dorsolaterally in head. Interorbital space wide (28.6–43.0% of HL, 155.6–239.0% of eye length). Mouth inferior (Supplementary Figure S2). Lips thick and well-developed with furrows; lower lip continuous with shallow median incision. Lower jaw crescentic, uncovered by lower lip. Three pairs of barbels thick, moderately short; inner rostral barbels almost reaching corner of mouth; outer rostral barbels horizontally reaching posterior nostril; maxillary barbels horizontally reaching mid-point of eyes.

Fins short (Figure 1B). Dorsal fin rounded distally, originating anterior to pelvic-fin origin; dorsal-fin origin near midway between tip of snout and caudal-fin base or slightly nearer to caudal-fin base. Pectoral fin short, not reaching halfway point to pelvic-fin base. Pelvic fin reaching past anus, distal fin tip attaining anal-fin origin when adpressed. Anal fin rounded distally, just posterior to anus. Caudal fin slightly emarginate.

Skin scaleless. Lateral line tapering, interrupted on posterior trunk at pelvic-fin distal extremity; few lateral line pores close to caudal-fin base. Stomach U-shaped, expanded. Intestine long, with 6–7 loops (Figure 1D and Supplementary Figure S3; one hidden loop surrounded by 5–6 loops). Bony capsule of air bladder small and thin, closed laterally; posterior chamber of air bladder absent (Figure 1E).

Sexual dimorphism: In mature males, unbranched and four outer branched pectoral-fin rays thickened, covered with breeding tubercles on dorsal surface. Small breeding tubercles also present on both sides of head in liber

apophyses extending from anterior lower margin of orbit to base of outer rostral barbel. These characters do not occur in females (Figure 1F).

Color pattern: In life: silver-gray base in individuals. In 70% EtOH: ground color of body brown dorsally and laterally, becoming yellowish ventrally. Fin membranes hyaline and slightly gray, with dark-brown faint spots on both sides of dorsal and caudal fins and upper side of pectoral fin. Body without obvious mottling. Peritoneum silvery with scattered dark melanophores becoming dense at vertebral column.

Ecology: Specimens were collected from flowing streams with gravel or sandy substrates at 1 360–1 768 m a.s.l. (Supplementary Figure S4). River water became muddy at about 0.5 m deep. Periphytic algae, sand, and insect larvae (e.g., chironomids) were found in the stomach of specimens. Other species collected with *Triplophysa weiheensis* sp. nov. included *T. dalaica* Kessler, *T. minxianensis* Wang and Zhu, *Pseudorasbora parva* Temminck and Schlegel, and *Gobio huanghensis* Lo, Yao et Chen.

Distribution: *Triplophysa weiheensis* sp. nov. is known only from the upper reaches of the Weihe River (Figure 1A).

Comparisons: Intestine and posterior chamber of air bladder are two important characters in the morphological taxonomy of the genus *Triplophysa* (He et al., 2006; Wu and Wu, 1992; Zhu, 1989). *Triplophysa weiheensis* sp. nov. differs from most *Triplophysa* species based on the combination of intestine screw shaped and posterior chamber of air bladder absent (Figure 1D, E), and can be classified into the same category as *T. stoliczkae* Steindachner, *T. tanggulaensis* Zhu, *T. crassilabris* Ding, *T. alticeps* Herzenstein, *T. cakaensis* Cao and Zhu, *T. chondrostoma* Herzenstein, *T. stenura* Herzenstein, *T. rotundiventris* Wu and Chen, *T. nujiangensis* Chen et al., and *T. daochengensis* Wu et al. (Chen et al., 2004; Ding, 1994; Wu & Wu, 1992; Wu et al., 2016; Zhu, 1989). Among these, the new species is most similar to *T. stoliczkae*.

Triplophysa weiheensis sp. nov. can be distinguished from *T. stoliczkae* by a combination of the following characters: lower jaw crescentic, not sharp (vs. spade-like, sharp); lateral line incomplete (vs. complete); vertebrae 4+34–36 (vs. 4+38–41); intestine ca. 1.8 times as long as SL (vs. 1.0–1.3 times); body without obvious mottling (vs. with mottling). The PCA results also indicated that the new species differs from *T. stoliczkae* in its whole morphology (Figure 1G) and the two species can be clearly separated from each other by PC1. Five morphometric measurements, including head depth/head length, interorbital width/head length, body depth/standard length, caudal peduncle depth/standard length, and postorbital length/head length, were highly correlated with and substantially contributed to PC1 (Supplementary Figure S5A, B). The five morphometric measurements were greater in the new species than in *T. stoliczkae* (Supplementary Figure S5C), suggesting a comparatively stubby body for the new species relative to *T. stoliczkae*.

Additionally, *Triplophysa weiheensis* sp. nov. can be distinguished from *T. tanggulaensis* by the following

characters: pelvic fin i, 5–6 (vs. i, 7–9); inner gill rakers on 1st gill arch 14–16 (vs. 10–13); vertebrae: 4+34–36 (vs. 4+37–38); intestine with 6–7 loops (vs. 3–4 loops). The new species can be distinguished from *T. crassilabris* by the following characters: pectoral fin i, 10–11 (vs. i, 8–9); pelvic fin i, 5–6 (vs. i, 7–8); inner gill rakers on 1st gill arch 14–16 (vs. 8–9); intestine with 6–7 loops (vs. 4–5 loops). The new species can be distinguished from *T. alticeps* by the following characters: intestine with 6–7 loops (vs. 3–4 loops); bony capsule of air bladder small and thin (vs. large and inflated). The new species can be distinguished from *T. cakaensis* by the following characters: snout abruptly sloping downward anterior to nostril (vs. snout gently sloping downward); lateral line interrupted on posterior trunk at pelvic fin distal extremity (vs. ending above pectoral fin); body without obvious mottling (vs. with mottling). The new species can be distinguished from *T. chondrostoma* by the following characters: lower jaw crescentic, not sharp (vs. spade-like, sharp); bony capsule of air bladder small and thin (vs. large and inflated); caudal-peduncle length 2.0–2.7 times its depth (vs. 3.2–3.8 times); body without obvious mottling (vs. with distinct mottling). The new species can be distinguished from *T. stenura* by the following characters: caudal peduncle depth nearly uniform towards caudal-fin base (vs. tapered); lateral line interrupted (vs. complete); intestine ca. 1.8 times as long as SL (vs. 1.0–1.3 times). The new species can be distinguished from *T. rotundiventris* by the following characters: caudal-peduncle length 2.0–2.7 times its depth (vs. 3.0–3.6 times); intestine ca. 1.8 times as long as SL (vs. 2.0–2.5 times); body without obvious mottling (vs. with mottling). The new species can be distinguished from *T. nujiangensa* by the following characters: vertebrae: 4+34–36 (vs. 4+38–39); intestine with 6–7 loops (vs. 3 loops); pelvic fin reaching past anus (vs. not reaching). The new species can be distinguished from *T. daochengensis* by the following characters: intestine with 6–7 loops (vs. 3 loops); pelvic fin reaching past anus (vs. not reaching); caudal fin slightly emarginate (vs. deeply emarginate).

Molecular analysis: Our results were in accordance with previous study, which suggested that *T. stoliczkae* consists of distinct lineages that are not close relatives (Feng et al., 2019a; Figure 1H). Phylogenetic analysis recovered the monophyly of *Triplophysa weiheensis* **sp. nov.** with strong support and showed that it was close to the *T. stoliczkae* population from the Yellow River system. The K2P genetic distance between *Triplophysa weiheensis* **sp. nov.** and *T. stoliczkae* was 3.7%, which is larger than that between several pairs of recognized species (Supplementary Table S5). These analyses suggest that *Triplophysa weiheensis* **sp. nov.** is a separately evolving lineage and genetic differences from its sister lineage in the genus *Triplophysa* may have reached species-level differentiation. Morphologically, this distinct phylogenetic lineage differed from all described *Triplophysa* species. Thus, based on an integrative taxonomic approach (Wu et al., 2019), the specimens collected from Weihe River are designated as a distinct species.

NOMENCLATURE ACTS REGISTRATION

The electronic version of this article in portable document format will represent a published work according to the International Commission on Zoological Nomenclature (ICZN), and hence the new names contained in the electronic version are effectively published under that Code from the electronic edition alone (see Articles 8.5–8.6 of the Code). This published work and the nomenclature acts it contains have been registered in ZooBank, the online registration system for the ICZN. The ZooBank LSIDs (Life Science Identifiers) can be resolved and the associated information can be viewed through any standard web browser by appending the LSID to the prefix <http://zoobank.org/>.

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SCIENTIFIC FIELD SURVEY PERMISSION INFORMATION

The field surveys in the Weihe River area in Gansu Province were approved by the Department of Fisheries of Gansu Province, China.

SUPPLEMENTARY DATA

Supplementary data to this article can be found online.

COMPETING INTERESTS

The authors declare that they have no competing interests.

AUTHORS' CONTRIBUTIONS

K.Z. and C.G.F. conceived and designed the study. K.Z., C.G.F., and C.T. collected specimens in the field. C.G.F., Y.Z., B.Z.Z., X.H.L., Y.T.T., and W.Z.S. performed the experiments and analyzed the data. C.G.F., Y.Z., C.T., and K.Z. prepared the manuscript. All authors read and approved the final version of the manuscript.

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REFERENCES

- Chen JX, Xu TQ, Fang SM, Song SL, Wang XT. 1987. Fishes in Qinling Mountain Area. Beijing: Science Press, 19–29. (in Chinese)
- Chen XY, Cui GH, Yang JX. 2004. A new fish species of genus *Triplophysa* (Balitoridae) from Nu Jiang, Yunnan, China. *Zoological Research*, **25**(6): 504–509. (in Chinese)
- Ding RH. 1994. The Fishes of Sichuan, China. Chengdu: Sichuan Publishing House of Science and Technology, 63–94. (in Chinese)
- Feng CG, Tang YT, Liu SJ, Tian F, Zhang CF, Zhao K. 2019a. Multiple convergent events created a nominal widespread species: *Triplophysa stoliczkae* (Steindachner, 1866) (Cobitoidea: Nemacheilidae). *BMC Evolutionary Biology*, **19**(1): 177.
- Feng CG, Tong C, Zhang RY, Li GG, Wanghe KY, Tang YT, Zhang CF, Zhao K. 2017a. Biodiversity and distribution patterns of *Triplophysa* species in the northeastern margin of the Tibetan Plateau. *Biodiversity Science*, **25**(1): 53–61. (in Chinese)
- Feng CG, Wu YJ, Tian F, Tong C, Tang YT, Zhang RY, Li GG, Zhao K. 2017b. Elevational diversity gradients of Tibetan loaches: the relative roles of ecological and evolutionary processes. *Ecology and Evolution*, **7**(23): 9970–9977.
- Feng CG, Zhou WW, Tang YT, Gao Y, Chen JM, Tong C, Liu SJ, Wanghe KY, Zhao K. 2019b. Molecular systematics of the *Triplophysa robusta* (Cobitoidea) complex: extensive gene flow in a depauperate lineage. *Molecular Phylogenetics and Evolution*, **132**: 275–283.
- Froese R, Pauly D. 2019. FishBase. World Wide Web electronic publication. version (08/2019). www.fishbase.org.
- He DK, Chen YX, Chen YF. 2006. Molecular phylogeny and biogeography of the genus *Triplophysa* (Osteichthyes: Nemacheilinae) in the Tibetan Plateau inferred from cytochrome b DNA sequences. *Progress in Natural Science*, **16**(11): 1395–1404. (in Chinese)
- Huang TF, Zhang PL, Huang XL, Wu T, Gong XY, Zhang YX, Peng QZ, Liu ZX. 2019. A new cave-dwelling blind loach, *Triplophysa erythraea* sp. nov. (Cypriniformes: Nemacheilidae), from Hunan Province, China. *Zoological Research*, **40**(4): 331–336.
- Kimura M. 1980. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, **16**(2): 111–120.
- Kottelat M. 1990. Indochinese Nemacheilines: A Revision of Nemacheiline Loaches (Pisces: Cypriniformes) of Thailand, Burma, Laos, Cambodia, and Southern Viet Nam. Munich: Verlag Dr. Friedrich Pfeil, 262.
- Liu SW, Pan XF, Yang JX, Chen XY. 2017. A new cave-dwelling loach, *Triplophysa xichouensis* sp. nov. (Teleostei Nemacheilidae) from Yunnan, China. *Journal of Fish Biology*, **90**(3): 834–846.
- Prokofiev AM. 2007. Materials towards the revision of the genus *Triplophysa* Rendahl, 1933 (Cobitoidea: Balitoridae: Nemacheilinae): a revision of nominal taxa of Herzenstein (1888) described within the species “*Nemachilus*” *stoliczkae* and “*N.*” *dorsonotatus*, with the description of the new species *T. scapanognatha* sp. nova. *Journal of Ichthyology*, **47**(1): 1–20.
- Tamura K, Stecher G, Peterson D, Filipiński A, Kumar S. 2013. MEGA6: molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, **30**(12): 2725–2729.
- Wang Y, Shen YJ, Feng CG, Zhao K, Song ZB, Zhang YP, Yang LD, He SP. 2016. Mitogenomic perspectives on the origin of Tibetan loaches and their adaptation to high altitude. *Scientific Reports*, **6**(1): 29690.
- Wu TJ, Wei ML, Lan JH, Du LN. 2018. *Triplophysa anshuiensis*, a new species of blind loach from the Xijiang River, China (Teleostei, Nemacheilidae). *ZooKeys*, **744**: 67–77.
- Wu YF, Wu CZ. 1992. The Fishes of the Qinghai-Xizang Plateau. Chengdu: Sichuan Publishing House of Science & Technology. (in Chinese)
- Wu YH, Suwannapoom C, Poyarkov Jr NA, Paawangkhant P, Xu K, Jin JQ, Murphy RW, Che J. 2019. A new species of the genus *Xenophys* (Anura: Megophryidae) from northern Thailand. *Zoological Research*, **40**(6): 564–574.
- Wu YY, Sun ZY, Guo YS. 2016. A new species of the genus *Triplophysa* (Cypriniformes: Nemacheilidae), *Triplophysa daochengensis*, from Sichuan Province, China. *Zoological Research*, **37**(5): 290–295.
- Yang HF, Li WX, Chen ZM. 2016. A new cave species of the Genus *Triplophysa* from Yunnan, China. *Zoological Research*, **37**(5): 296–300.
- Zhu SQ. 1989. The Loaches of the Subfamily Nemacheilinae in China (Cypriniformes: Cobitidae). Nanjing: Jiangsu Science and Technology Publishing House. (in Chinese)

Supplementary Materials

Supplementary Methods

Euthanization and ethics statement

After preliminary identification, collected specimens were euthanized as per Feng et al. (2019). They were placed in a dry ice box for rapid hypothermic anesthesia within about 20 s. Then, they were preserved in 70% ethanol for laboratory works. All animal experiments for this project were approved by the Ethics Committee of the Northwest Institute of Plateau Biology, Chinese Academy of Sciences [NWIPB201503018].

Additional measurements

Measurements and counts follow that of Kottelat (1990) and Prokofiev (2007). Additional measurements are the following: postorbital length is measured from the posterior margin of the orbit to the posterior end of the operculum; pectoral-pelvic distance is measured from the pectoral-fin origin to the pelvic-fin origin; the number of gill rakers was counted on the inner side of the first arch.

Amplification, sequencing and phylogenetic analysis

Total genomic DNA was extracted from fins using the standard 3-step phenol-chloroform method (Sambrook et al., 1989). The complete *cyt b* gene (1 140 bp) was amplified with the universal primer pairs L14724 (5'-GACTTGAAAAACCACCGTTG-3') and H15915 (5'-CTCCGATCTCCGGATTACAAGAC-3') (Xiao et al., 2001) in total reaction volumes of 35 μ L, containing approximately 100 ng of template DNA, 0.7 μ L of each primer (10 μ mol/L), 3.5 μ L 10 \times reaction buffer, 3 μ L dNTPs (2.5 mmol/L each), and 1.0 U TaKaRa rTaq (TaKaRa Corp., Dalian, China). The PCR cycling profile was at 94 $^{\circ}$ C for 5 min, 35 cycles of at 94 $^{\circ}$ C for 30 sec, at 52 $^{\circ}$ C for 30 sec and at 72 $^{\circ}$ C for 1 min 30 sec, followed by 72 $^{\circ}$ C for 10 min. The PCR products were sequenced from both directions using an ABI PRISM 3700 sequencing system.

The presence of *cyt b* pseudogene was checked by scanning for stop codons or indels in MEGA v6.0 (Tamura et al., 2013). Phylogenetic analysis was performed using the Bayesian inference (BI) implemented in MRBAYES 3.2.0 (Ronquist et al., 2012). The best model of DNA substitution (GTR+I+G) was identified by jModelTest 0.1.1 (Guindon and Gascuel, 2003; Posada, 2008) based on Bayesian Information Criterion (BIC) (Schwarz, 1978; Luo et al., 2010). Markov Chain Monte Carlo (MCMC) was set with one cold chain and three heated chains. Samples of trees and the parameters were drawn every 100 steps from a total of 1 000 000 MCMC generations, with the first 25% samples discarded as burn-in.

REFERENCES

- Feng CG, Tang YT, Liu SJ, Tian F, Zhang CF, Zhao K. 2019. Multiple convergent events created a nominal widespread species: *Triplophysa stoliczkae* (Steindachner, 1866) (Cobitoidea: Nemacheilidae). *BMC Evolutionary Biology*, **19**(1): 177.
- Guindon S, Gascuel O. 2003. A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Systematic Biology*, **52**, 696–704.
- Kottelat M. 1990. Indochinese nemacheilines: A revision of nemacheiline loaches (Pisces: Cypriniformes) of Thailand, Burma, Laos, Cambodia and southern Viet Nam. Verlag Dr Friedrich Pfeil, Pub., München, 262 pp.
- Luo A, Qiao H, Zhang Y, Shi W, Ho SY, Xu W, Zhang A, Zhu C. 2010. Performance of criteria for selecting evolutionary models in phylogenetics: a comprehensive study based on simulated datasets. *BMC Evolutionary Biology*, **10**, 1.
- Posada D. 2008. jModelTest: phylogenetic model averaging. *Molecular Biology and Evolution*, **25**, 1253–1256.
- Prokofiev A. 2007. Materials towards the revision of the genus *Triplophysa* Rendahl, 1933 (Cobitoidea: Balitoridae: Nemacheilinae): A revision of nominal taxa of Herzenstein (1888) described within the species “*Nemachilus*” *stoliczkae* and “*N.*” *dorsonotatus*, with the description of the new species *T. scapanognatha* sp. nova. *Journal of Ichthyology*, **47**, 1–20.
- Ronquist F, Teslenko M, Van Der Mark P, Ayres DL, Darling A, H Hna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology*, **61**, 539–542.
- Sambrook J, Fritsch EF, Maniatis T. 1989. Molecular cloning, New York, Cold spring harbor laboratory press.
- Schwarz G. 1978. Estimating the dimension of a model. *The Annals of Statistics*, **6**, 461–464.
- Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. 2013. MEGA6: molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, mst197.
- Xiao W, Zhang Y, Liu H. 2001. Molecular systematics of Xenocyprinae (Teleostei: Cyprinidae): taxonomy, biogeography, and coevolution of a special group restricted in East Asia. *Molecular Phylogenetics and Evolution*, **18**, 163–173.

Comparative materials

Triplophysa stoliczkae.

(The Indus river system): NWIPB 1407013–018, 6 ex., 63.7–85.2 mm SL, middle reach of Changchuan River, Rutog County, Ngari Prefecture, Tibet Autonomous Region, China; NWIPB 74–(280, 282, 284, 285, 289, 292, 294, 295), 1106009, 9 ex., 50.7–78.1 mm SL, a tributary of Lake Bangong, Rutog Town, Rutog County, Ngari Prefecture, Tibet Autonomous Region, China; NWIPB 74–(297, 312, 318), 3 ex., 67.0–80.6 mm SL, glang-chen gtsang-po, Zanda Zong, Ngari Prefecture, Tibet Autonomous Region, China.

(The Yellow river system): NWIPB 1505471–480, 10 ex., 44.51–72.69 mm SL, a tributary of Taohe River, Sigou Town, Minxian County, Gansu province, China; NWIPB 1205321, 1205322, 2 ex., 88.7–95.0 mm SL, Datong River, Haomen Town, Menyuan County, Haixi Mongolian and Tibetan Autonomous Prefecture, Qinghai Province, China; NWIPB 1707001, 1 ex., 85.1 mm SL, Huangshui River, Xihai Town, Haiyan County, Haixi Mongolian and Tibetan Autonomous Prefecture, Qinghai Province, China.

(The Ganges river system): NWIPB 1106001, 1106002, 2 ex., 104.9–112.2 mm SL, a tributary of Manasarovar Lake, Burang County, Ngari Prefecture, Tibet Autonomous Region, China; NWIPB 74–(257, 269, 267), 3 ex., 58.2–82.6 mm SL, Kongque River, Burang County, Ngari Prefecture, Tibet Autonomous Region, China.

(The Heihe river system): NWIPB 1205128, 1205134, 2 ex., 85.9–94.0 mm SL, Babao River, Zhamashi Town, Qilian County, Haixi Mongolian and Tibetan Autonomous Prefecture, Qinghai Province, China; NWIPB 1205095-097, 1205099-102, 7 ex., 65.6–101.2 mm SL, Heihe River, Longshou Town, Zhangye City, Gansu Province, China.

(The Tarim river system): NWIPB 1107083, 1 ex., 97.5 mm SL, Yarkand River, Yecheng County, Kashgar Prefecture, Xinjiang Uygur Autonomous Region, China; NWIPB 1107084, 1 ex., 73.9 mm SL, Qaraqash River, Pishan County, Kashgar Prefecture, Xinjiang Uygur Autonomous Region, China.

(The Ili river system): NWIPB 1305044, 1 ex., 98.8 mm SL, Kashi River, Nilka County, Ili Prefecture, Xinjiang Uygur Autonomous Region, China; NWIPB 1305111, 1305113-115, 4 ex., 68.8–91.0 mm SL, Kunes River, Xinyuan County, Ili Prefecture, Xinjiang Uygur Autonomous Region, China; NWIPB 1305046-048, 1305052, 1305056, 1305060, 6 ex., 56.0–102.5 mm SL, Jinghe Forest Farm, Jinghe County, Bortala Mongol Autonomous Prefecture, Xinjiang Uygur Autonomous Region, China; NWIPB 1305131, 1305141, 1305142, 3 ex., 64.0–75.4 mm SL, Tekes River, Tekes County, Ili Prefecture, Xinjiang Uygur Autonomous Region, China.

Triplophysa chondrostoma. Topotype. NWIPB 1006052–055, 4 ex., 74.4–89.2 mm SL, Caidam River, Nuomuhong Town, Dulan County, Haixi Mongolian and Tibetan Autonomous Prefecture, Qinghai Province, China.

Triplophysa tanggulaensis. NWIPB 75-061, holotype, 1 ex., 62.2 mm SL, 75-(051–054, 058, 060, 064), paratypes, 7ex., 61.3–73.2 mm SL, a hot spring between Yanshiping and T'ang-ku-la Pass, T'ang-ku-la Town, Geermu city, Haixi Mongolian and Tibetan Autonomous Prefecture, Qinghai Province, China.

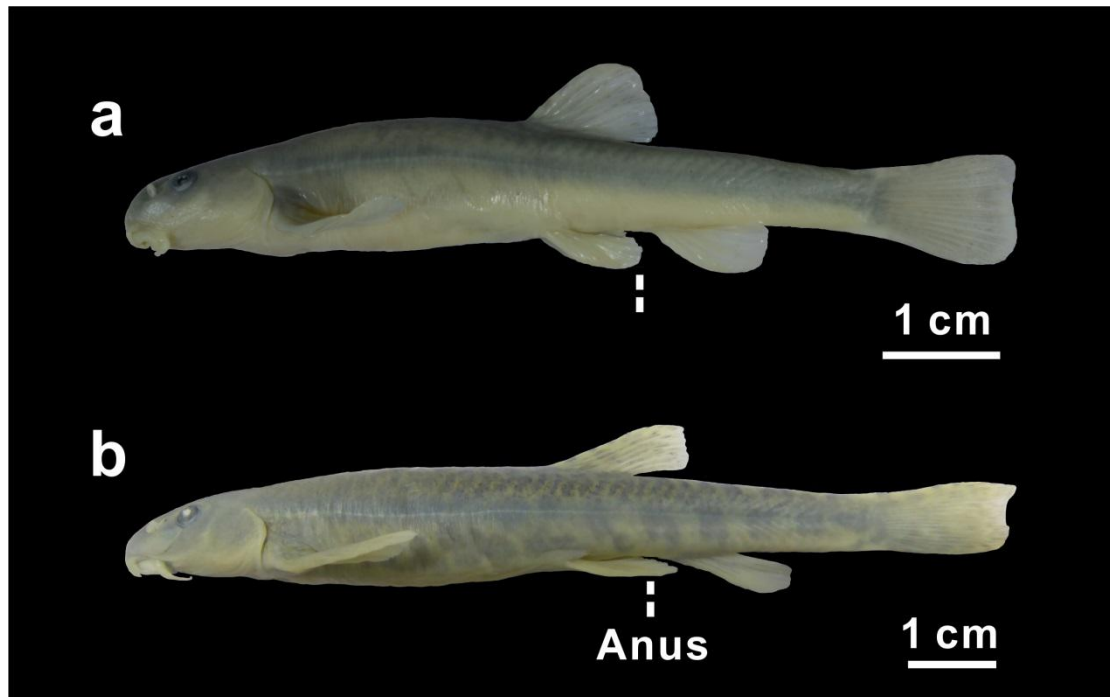
Triplophysa alticeps. Topotype. NWIPB 1206001–004, 4 ex., 45.6–60.1 mm SL, Lake Qinghai, in the Haiyan County, Tibetan Autonomous Prefecture of Haibei, Qinghai province, China; NWIPB 1310001–002, 2 ex., 74.35–86.54 mm SL, Huangshui River, at Haiyan County, Tibetan Autonomous Prefecture of Haibei, Qinghai province, China.

Triplophysa stenura. Topotype. NWIPB 1108062–1108065, 4 ex., 63.1–76.8 mm SL, Zhaqu River, Nangqian County, Yushu Tibetan Autonomous Prefecture, Qinghai province, China; NWIPB 1106110–1106112, 1106114–1106115, 1106117–1106119, 8 ex., 51.7–101.3 mm SL, headstream of Brahmaputra River, Zhongba County, Xigazê City, Tibet Autonomous Region, China.

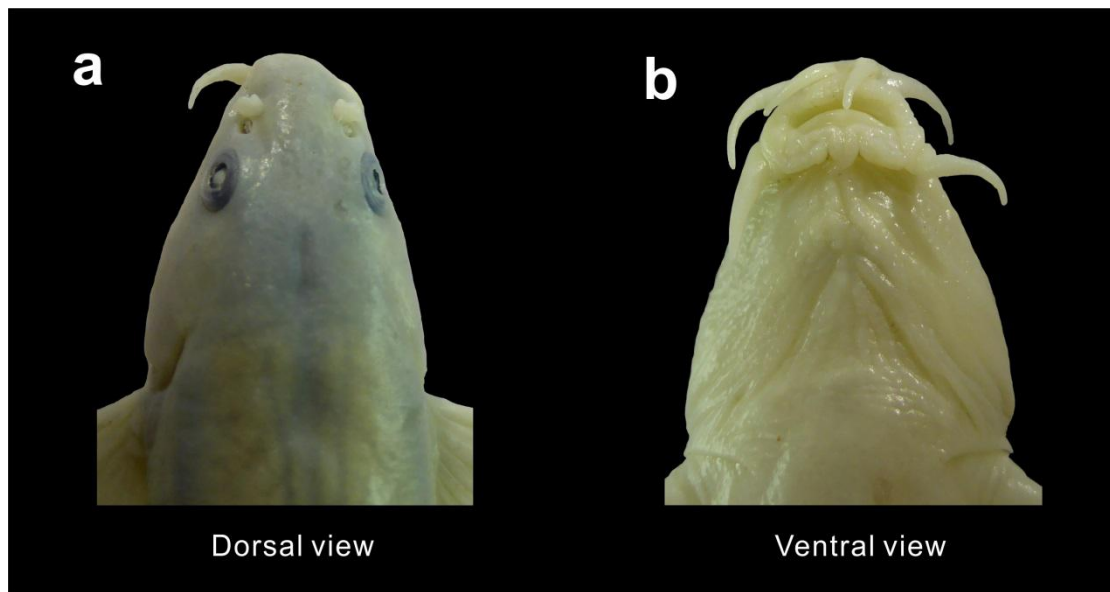
Triplophysa nujiangensa. Topotype. IHB 00915579, 00915605, 00915826, 3 ex., 65.7–77.3 mm SL, Pi River, Liuku Town, Lushui County, Nujiang of the Lisu Autonomous Prefecture, Yunnan Province, China.

Triplophysa rotundiventris. NWIPB 1107006, 1 ex., 70.1 mm SL, Naqu River, Naqu County, Naqu City, Tibet Autonomous Region, China.

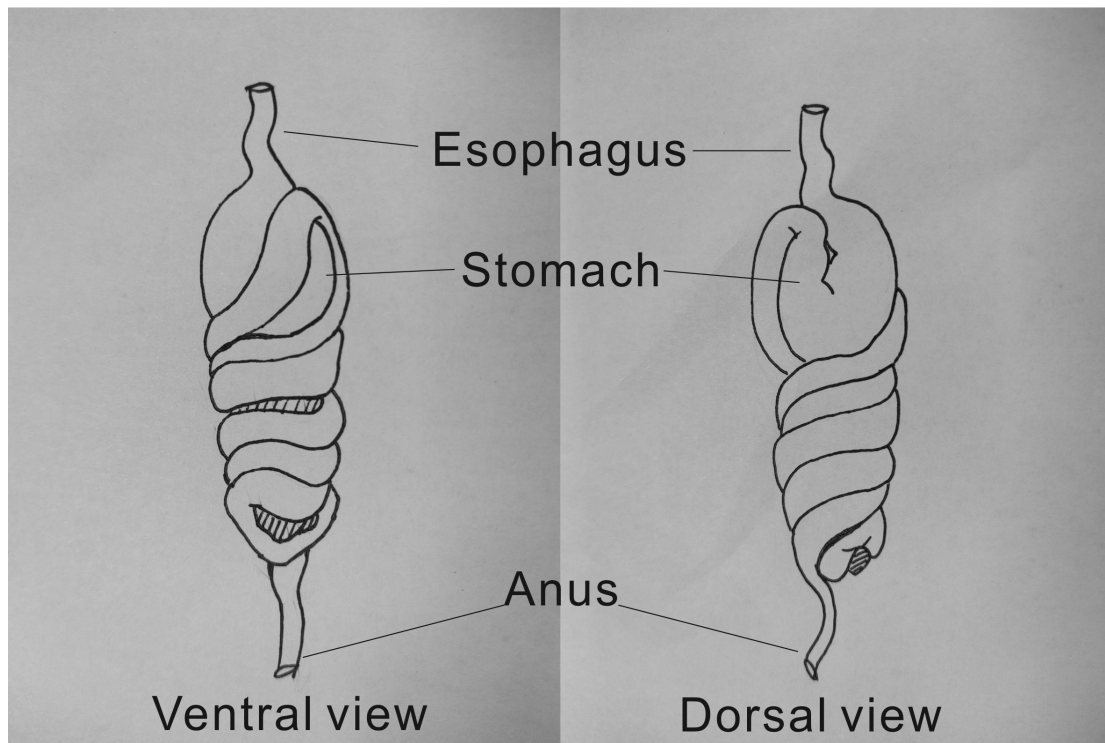
Supplementary Figures



Supplementary Figure S1 Lateral view of *Triplophysa weiheensis* sp. nov., NWIPB 1505189, holotype, 65.8 mm SL(**a**) and *T. stoliczkae* NWIPB 1707001, 85.1 mm SL, from the Yellow River system(**b**).



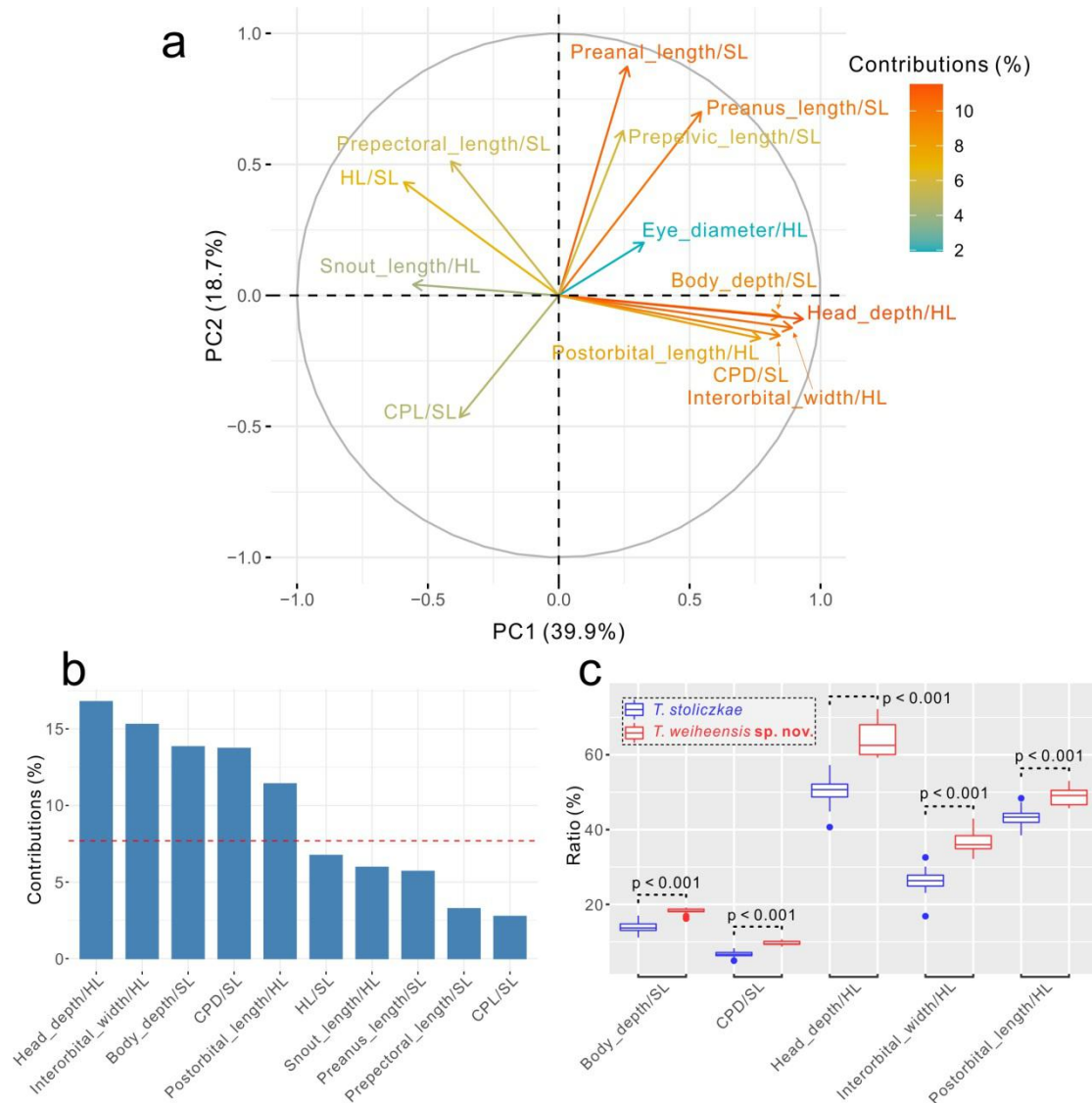
Supplementary Figure S2 *Triplophysa weiheensis* sp. nov., NWIPB 1505189, holotype, 65.8 mm SL. (a) dorsal and (b) ventral view of head.



Supplementary Figure S3 Hand drawing of alimentary canal, corresponding to the photograph of Figure 1d. The shaded area indicates the fat attached to the intestine. (Drawing by Yu Zhang).



Supplementary Figure S4 Habitat of *Triplophysa weiheensis* sp. nov.. **(a)** Type locality: Zhang River, at Chenjiamo Village, Zhangxian County; **(b)** Niutou River, at Hongbao Town, Qingshui County. (photo by Kai Zhao).



Supplementary Figure S5 Visualize principal component analysis (PCA) results for 13 variables of morphometric characters (Table S2). **(a)** Variable correlation plot. Variables on the correlation plot are colored according to their contribution values. **(b)** The contribution of variables to PC1. The red dashed line on the graph indicates the expected average contribution. Only the top 10 contributing variables are shown. **(c)** Boxplot of morphometric measurements of five high contributing variables to PC1. The p values indicate significance from the *t*-test analysis. Abbreviation: CPD, caudal peduncle depth; CPL, caudal peduncle length; HL, head length; SL, standard length; *T.*, *Triplophysa*.

Supplementary Tables

Supplementary Table S1 List of *Triplophysa* species that have been reported in the Yellow River system

Species name	Authors and year	Whether in the Weihe river
<i>Triplophysa minxianensis</i>	Wang and Zhu 1979	Yes
<i>Triplophysa sellaefer</i>	Nichols 1925	Yes
<i>Triplophysa shaanxiensis</i>	Chen in Chen et al 1987	Yes
<i>Triplophysa dalaica</i>	Kessler 1876	Yes
<i>Triplophysa stoliczkae</i>	Steindachner 1866	Yes
<i>Triplophysa robusta</i>	Kessler 1876	
<i>Triplophysa orientalis</i>	Herzenstein 1888	
<i>Triplophysa obscura</i>	Wang in Chen et al 1987	
<i>Triplophysa scleroptera</i>	Herzenstein 1888	
<i>Triplophysa pseudoscleroptera</i>	Zhu and Wu 1981	
<i>Triplophysa pappenheimi</i>	Fang 1935	
<i>Triplophysa siluroides</i>	Herzenstein 1888	
<i>Triplophysa leptosoma</i>	Herzenstein 1888	
<i>Triplophysa brevicauda</i>	Herzenstein 1888	
<i>Triplophysa longianguis</i>	Wu and Wu 1984	
<i>Triplophysa crassilabris</i>	Ding 1994	
<i>Triplophysa alticeps</i>	Herzenstein 1888	

Supplementary Table S2 13 morphometric characters used in the PCA analysis

Including body_depth/standard_length (SL), head_length (HL)/SL, preanus_length/SL, preanal_length/SL, prepelvic_length/SL, prepectoral_length/SL, caudal peduncle length (CPL)/SL, caudal peduncle depth (CPD)/SL, head_depth/HL, snout_length/HL, eye_diameter/HL, interorbital_width/HL, and postorbital_length/HL.

Supplementary Table S2 was listed as a separate csv file, because it's too big.

Supplementary Table S3 Mitochondrial cytochrome *b* (cyt *b*) sequence samples of *Triplophysa* species and outgroups used in the present study

Species	Voucher ID	Accession number	Sample site	Origin
<i>Triplophysa hsutschouensis</i>	NWIPB 0710002	KX373852	Beidahe river, Sunan, Gansu	This study
<i>Triplophysa stoliczkae</i>	NWIPB 1550000	KX373851	Daxiahe river, Linxia, Gansu	This study
<i>Triplophysa wuweiensis</i>	NWIPB 1205606	KX373838	Shiyanghe river, Wuwei, Gansu	This study
<i>Triplophysa tenuis</i>	NWIPB 1250174	KX373841	Heihe (Hexi drainage), Zhangye, Gansu	This study
<i>Triplophysa leptosoma</i>	NWIPB 1250353	KX373839	Datonghe river, Menyuan, Qinghai	This study
<i>Triplophysa pseudoscleroptera</i>	NWIPB 1505061	KX373844	Taohe river, Lintao, Gansu	This study
<i>Triplophysa minxianensis</i>	NWIPB 1505925	KX373849	Weihe river, Qingshui, Gansu	This study
<i>Triplophysa dalaica</i>	NWIPB 1205219	KX373845	Weihe river, Wushan, Gansu	This study
<i>Triplophysa robusta</i>	NWIPB 1505995	KX373850	Daxiahe river, Linxia, Gansu	This study
<i>Triplophysa pappenheimi</i>	NWIPB 1410049	KX373843	Taohe river, Luqu, Gansu	This study
<i>Triplophysa siluroides</i>	NWIPB 1410007	KX373842	Yellow river, Maduo, Qinghai	This study
<i>Triplophysa scleroptera</i>	NWIPB 1250405	KX373840	Yellow river, Guide, Qinghai	This study
<i>Triplophysa orientalis</i>	NWIPB 1505576	KX373846	Heihe (Yellow river), ruoergai, Sichuan	This study
<i>Triplophysa alticeps</i>	NWIPB 1108012	KX373837	Lake Qinghai, Qinghai	This study
<i>Triplophysa bleekeri</i>	NWIPB 1505622	KX373847	Baishui river, Wenxian, Gansu	This study
<i>Triplophysa weiheensis</i> sp. nov.	NWIPB 1505992	KX373834	Niutou river, Qingshui, Gansu	This study
<i>Triplophysa weiheensis</i> sp. nov.	NWIPB 1505181	KY781400	Zhang River, Zhangxian, Gansu	This study
<i>Triplophysa weiheensis</i> sp. nov.	NWIPB 1505922	KY781401	Niutou River, Maiji, Gansu	This study
<i>Triplophysa weiheensis</i> sp. nov.	NWIPB 1505985	KY781402	Niutou river, Qingshui, Gansu	This study
<i>Triplophysa weiheensis</i> sp. nov.	NWIPB 1505986	KY781403	Niutou river, Qingshui, Gansu	This study
<i>Triplophysa chondrostoma</i>	NA	KT213589	Caidam	Wang et al., 2016
<i>Triplophysa stoliczkae</i>	NA	JQ663847	Yellow river	Li et al., 2013
<i>Triplophysa stoliczkae</i>	NA	NC017890	Yellow river	Li et al., 2013

<i>Triplophysa stoliczkae</i>	NWPU 1208003	MG725381	Huangshui river, Huangyuan, Qinghai	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1208004	MG725382	Huangshui river, Huangyuan, Qinghai	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1108034	MG725408	Zhaqu river, Chindu, Qinghai	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1307052	MG725407	Zhaqu river, Chindu, Qinghai	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1307016	MG725406	Zhaqu river, Chindu, Qinghai	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1307007	MG725405	Zhaqu river, Chindu, Qinghai	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1307005	MG725404	Zhaqu river, Chindu, Qinghai	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1305148	MG725400	Kunes river, Xinyuan, Xinjiang	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1305127	MG725397	Tekes river, Tekes, Xinjiang	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1305125	MG725396	Tekes river, Tekes, Xinjiang	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1305111	MG725399	Kunes river, Xinyuan, Xinjiang	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1305065	MG725398	Kashi river, Nilka, Xinjiang	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1305045	MG725395	Kashi river, Nilka, Xinjiang	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1305043	MG725394	Kashi river, Nilka, Xinjiang	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1106008	MG725393	Kongque river, Purang, Tibet	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1106007	MG725392	Kongque river, Purang, Tibet	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1007084	MG725389	Qaraqash river, Pishan, Xinjiang	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1007083	MG725388	Yarkand river, Yecheng, Xinjiang	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1407014	MG725410	Changchuan river, Rutog, Tibet	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1407013	MG725409	Changchuan river, Rutog, Tibet	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1106002	MG725391	Lake Manasarovar, Purang, Tibet	Feng et al., 2019
<i>Triplophysa stoliczkae</i>	NWPU 1106001	MG725390	Lake Manasarovar, Purang, Tibet	Feng et al., 2019
<i>Triplophysa rotundiventris</i>	NWPU 1107006	MG725402	Naqu river, Amdo Zong, Tibet	Feng et al., 2019
<i>Triplophysa dorsalis</i>	NWPU 1305232	MG725413	Tekes river, Tekes, Xinjiang	Feng et al., 2019
<i>Triplophysa markehenensis</i>	NWPU 0907001	MG725416	Dadu river, Danba, Sichuan	Feng et al., 2019
<i>Triplophysa anterodorsalis</i>	NWPU 1506001	MG725417	Jinsha river, Gonjo, Tibet	Feng et al., 2019

<i>Triplophysa microps</i>	NWPU 1307038	MG725385	Lancang river, Zadoi, Qinghai	Feng et al., 2019
<i>Triplophysa stewarti</i>	NWPU 1107007	MG725411	Lake Duoqing, Kangmar, Tibet	Feng et al., 2019
<i>Triplophysa aliensis</i>	NWPU 1106031	MG725412	Lake Manasarovar, Purang, Tibet	Feng et al., 2019
<i>Triplophysa stenura</i>	NWPU 1108064	MG725583	Lancang, Nangqian, Qinghai	Feng et al., 2019
<i>Triplophysa tibetana</i>	NWPU 1160067	MG725387	Lake Manasarovar, Purang, Tibet	Feng et al., 2019
<i>Triplophysa brevicauda</i>	NWPU 1106029	MG725386	Pengqu river, Nyalam, Tibet	Feng et al., 2019
<i>Barbatula toni</i>	NA	KY451914	NA	GenBank, unpublished
<i>Lefua costata</i>	NA	DQ105196	NA	Tang et al., 2006

Supplementary Table S4 Meristic counts of *Triplophysa weiheensis* sp. nov. and *T. stoliczkae*

Characters	<i>T. weiheensis</i> sp. nov.		<i>T. stoliczkae</i> ^a (n=278)	<i>T. stoliczkae</i> ^b (n=261)	<i>T. stoliczkae</i> ^c (n=13)
	Holotype	Paratypes (n=14)			
Dorsal fin	iii, 7	iii, 6–7	iv, 6–9	iii~iv, 6–9	iv, 7–8
Pelvic fin	i, 6	i, 5–6	i, 6–8	i, 6–8	i, 6–8
Pectoral fin	i, 10	i, 10–11	i, 10–12	i, 10–12	i, 10–11
Anal fin	iii, 5	iii, 5	iii, 5	iii, 5	iii, 5
Caudal fin	16	15–16	16	13~17	16
Gill rakers	15	14–16	9–24	13–23	15–18
Vertebrae	4+34	4+34–36	4+38–41	4+36–41 (n = 95)	4+38–39 (n = 8)

^aData from Wu and Wu (1992);

^bData from Zhu (1989);

^cMeasurement data of *T. stoliczkae* from the Yellow River system (See the Comparative material for specimen information).

Supplementary Table S5. K2P genetic distances based on mitochondrial Cyt *b* sequences between species of genus *Triplophysa*. Genetic distance between *Triplophysa weiheensis* **sp. nov.** and *T. stoliczkae* (from the Yellow River system) was colored red, others less than this value were colored orange.

Supplementary Table S5 was listed as a separate excel file, because it's too big.

REFERENCES

- Feng CG, Tang YT, Liu SJ, Tian F, Zhang CF, Zhao K. 2019. Multiple convergent events created a nominal widespread species: *Triplophysa stoliczkae* (Steindachner, 1866) (Cobitoidea: Nemacheilidae). *BMC Evolutionary Biology*, **19**(1), 177.
- Li JX, Si SJ, Guo R, Wang Y, Song ZB. 2013. Complete mitochondrial genome of the stone loach, *Triplophysa stoliczkae* (Teleostei: Cypriniformes: Balitoridae). *Mitochondrial DNA*, **24**, 8–10.
- Tang QY, Liu HZ, Mayden R, Xiong BX. 2006. Comparison of evolutionary rates in the mitochondrial DNA cytochrome *b* gene and control region and their implications for phylogeny of the Cobitoidea (Teleostei: Cypriniformes). *Molecular Phylogenetics and Evolution*, **39**, 347–357.
- Wang Y, Shen YJ, Feng CG, Zhao K, Song ZB, Zhang YP, Yang LD, He SP. 2016. Mitogenomic perspectives on the origin of Tibetan loaches and their adaptation to high altitude. *Scientific Reports*, **6**, 1–10.
- Wu YF, Wu CZ. 1992. *The fishes of the Qinghai-Xizang plateau*, Sichuan Publishing House of Science & Technology.
- Zhu SQ. 1989. *The loaches of the subfamily Nemacheilinae in China (Cypriniformes: Cobitidae)*. Jiangsu Science and Technology Publishing House.