

# Benthic Marine Algae on Japanese Tsunami Marine Debris

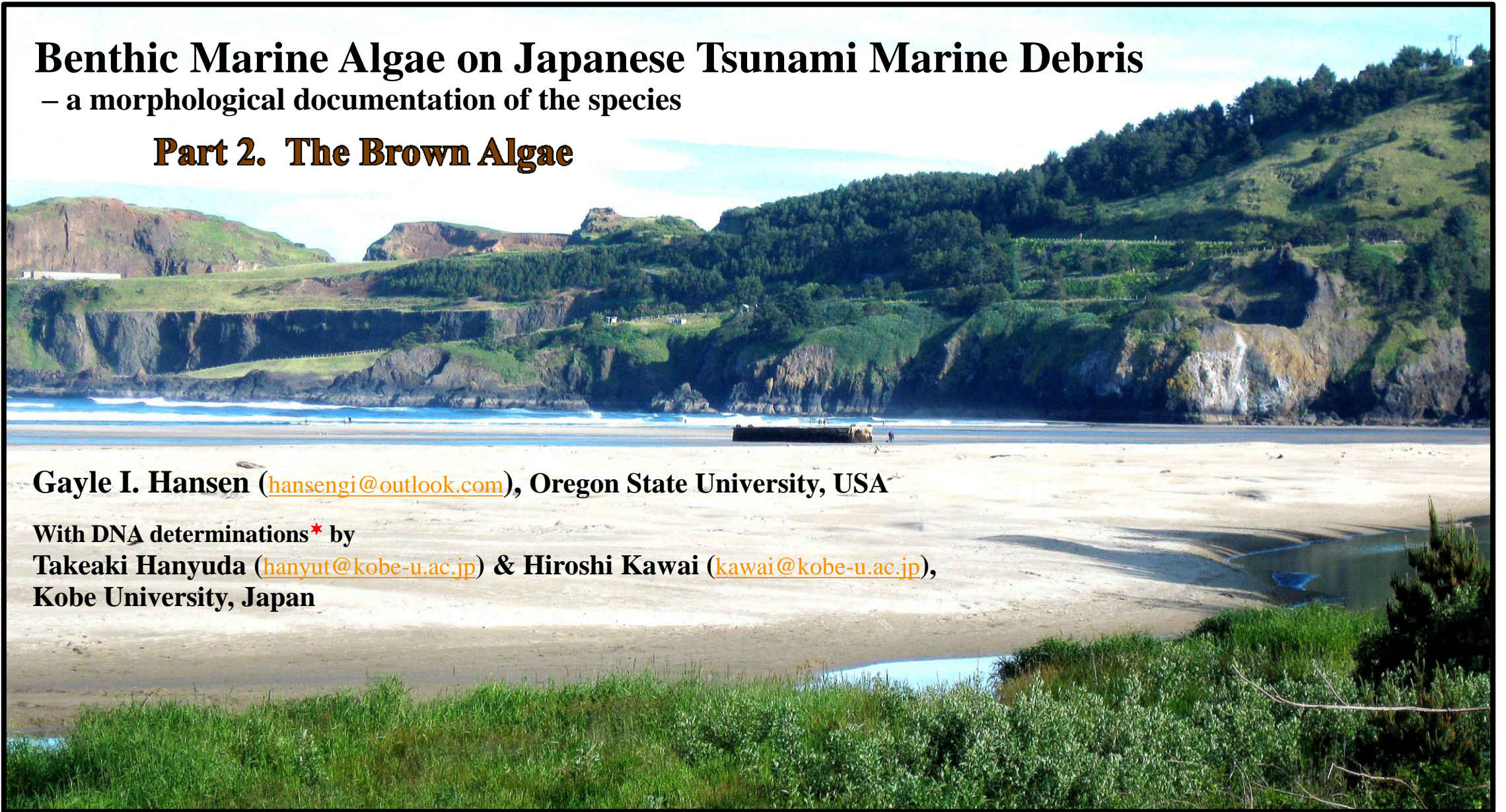
– a morphological documentation of the species

## Part 2. The Brown Algae

Gayle I. Hansen ([hansengi@outlook.com](mailto:hansengi@outlook.com)), Oregon State University, USA

With DNA determinations\* by

Takeaki Hanyuda ([hanyut@kobe-u.ac.jp](mailto:hanyut@kobe-u.ac.jp)) & Hiroshi Kawai ([kawai@kobe-u.ac.jp](mailto:kawai@kobe-u.ac.jp)),  
Kobe University, Japan





**Copyright:** 2017, CC BY-NC (attribution, non-commercial use). For photographs, please credit G.I. Hansen or those noted on the slides.

**Printing:** For better pdf printing, please reduce to letter (11" x 8.5") size, landscape orientation.

**Citations to be used for this series:**

- Hansen, G.I., Hanyuda, T. & Kawai, H. (2017). Benthic marine algae on Japanese tsunami marine debris – a morphological documentation of the species. Part 1. The tsunami event, the project overview, and the red algae. OSU Scholars Archive, Corvallis, pp. 1-50. <http://dx.doi.org/10.5399/osu/1110>
- Hansen, G.I., Hanyuda, T. & Kawai, H. (2017). Benthic marine algae on Japanese tsunami marine debris – a morphological documentation of the species. Part 2. The brown algae. OSU Scholars Archive, Corvallis, pp. 1-61. <http://dx.doi.org/10.5399/osu/1111>
- Hansen, G.I., Hanyuda, T. & Kawai, H. (2017). Benthic marine algae on Japanese tsunami marine debris – a morphological documentation of the species. Part 3. The green algae and cyanobacteria. OSU Scholars Archive, Corvallis, pp. 1-43. <http://dx.doi.org/10.5399/osu/1112>

**Other publications supported:** The Scholars Archive presentations above provide photographic documentation for the species included in the following publications. The poster is a pictorial overview of some of the larger debris algae made for teaching.

- Hansen, G.I., Hanyuda, T. & Kawai, H. (In Review). The invasion threat of benthic marine algae arriving on Japanese tsunami marine debris in Oregon and Washington, USA.
- Hanyuda, T., Hansen, G.I. & Kawai, H. (2017, In Press). Genetic identification of macroalgal species on Japanese tsunami marine debris and genetic comparisons with their wild populations. Marine Pollution Bulletin. <https://doi.org/10.1016/j.marpolbul.2017.06.053>
- Hansen, G.I. (2013). Some Marine Algae on Tsunami Debris. OSU Scholars Archive, Corvallis, a poster. <http://ir.library.oregonstate.edu/concern/defaults/ns064b84v>



Seaweeds on the Tohoku coast near Misawa, Japan, including *Saccharina japonica* and *Undaria pinnatifida*.

# Codes, Definitions & Abbreviations + Acknowledgements & Contents

## Special codes provided in each description:

- Definitions of terms and abbreviations (not provided below) are given in Part 1 of this series.
- ★ = Species that have been sequenced. We also list: the genes sequenced (p. 4), the debris item numbers (Appendix 1), and the collection numbers (with the descriptions).
- # = Identification was assisted by a monographic expert(s). Their names are provided on the species page.

**Approximate identifications:** The names for the Japanese Tsunami Marine Debris (JTMD) algae and cyanobacteria shown on the following pages are derived from: morphological accounts on the species, personal observations, and the genetic sequences (when available). Although the sequences and morphology often precisely match known species, situations can occur where either the sequences or the morphology vary slightly from the known observations and absolute identifications are impossible to determine. For these samples, I use the following qualifying terms to indicate approximate identifications: **sensu X** = an identification according to scientist X; **cf.** = refer to (the most probable species identification); **cpx.** = a clade or group of closely related species that includes the unnamed isolate. The term **cpx.** includes both: (1) **morphological variants** = species with identical sequences that have different morphology, and (2) **sequence variants** = species that are morphologically correct with the literature whose sequences do not match exactly those for the same species deposited in GenBank. These variant types are noted in the text.

**Longevity:** Life span data is from the literature and observations in the NEP. If it is estimated, a “~” precedes the most likely type (annual, perennial, ephemeral).

**Distributions:** See part 1 for all sources. Global distributions follow [www.AlgaeBase.org](http://www.AlgaeBase.org) (accessed July 2017). **Distribution Codes:** **G (Globally widespread)** = species that appear to be naturally widespread globally, occurring in different oceans and on multiple continents; **A (Asian-only)** = species occurring only in Asia, from Russia to the Philippines; **A+ (Asian+)** = Asian species that have also been exported globally by human activities; **NP-P (North Pacific-P)** = species limited primarily to both the NE and NW Pacific but with some occurrences in Alaska and the S. Pacific.

**Distribution Abbreviations:** Afr = Africa; AK = Alaska; A-Arc = Antarctic; Arc = Arctic; Aus = Australia; BC = British Columbia; Bra = Brazil; C = China; Car = Caribbean; ENA = Eastern North America; EUR = Europe including the British Isles; EUR-Arc = Europe and the European Arctic; HA = Hawaii; IO = Indian Ocean (including Indonesia); J = Japan; K = Korea; Med = Mediterranean; MX = Mexico; NEP = Northeast Pacific; NZ = New Zealand; OR = Oregon; Phil = Philippines; R = Eastern Russia; SA = South America (both coasts); Viet = Vietnam; WA = Washington. For brevity, we have excluded some island groups and Arctic areas. For more thorough distribution coverage, see the continually updated [www.algaebase.org](http://www.algaebase.org).

**Acknowledgements:** Financial support for this study was provided by Oregon Sea Grant, the Ministry of the Environment of Japan through the North Pacific Marine Science Organization (PICES) and personal savings. Collection assistance for the debris algae was generously provided by John Chapman, Russ Lewis, Nancy Treneman, Jessica Miller, Thomas Murphy and the state and volunteer agencies in Washington and Oregon responsible for debris removal. Jim Carlton kept the debris item database and provided his BF item numbers to all JTMD researchers. Cynthia Trowbridge gave valuable comments on the overall project. Judy Mullen (OSU libraries) provided essential and often obscure literature for the study. The US-EPA provided laboratory space for the Oregon part of the project. Additional credits are due to Mike Guiry and AlgaeBase.org for global distributions, reference information, and nomenclature advice – and also to the following taxonomic experts for helpful advice and identification assistance with the brown algae: S.M. Boo, S.G.A. Driasma, S. Loiseaux-de Goër, G.T. Kraft, D.G. Müller, A.F. Peters, W.F. Prud’homme van Reine, and J.R. Sears.

**Contents:** The Brown Algae – A Checklist of the Species on JTMD, The Species Descriptions, References, and Appendices 1-3.

# The Brown Algae

## A Checklist of the Species on JTMD and their global and NEP distributions

### KEY:

Pg = page number

JTMD = Japanese Tsunami Marine Debris

Global = general global occurrence

A = Asian only

A+ = Asian but also introduced by human activities globally

NP-P = Northwest and Northeast Pacific, some with Alaska and S. Pacific occurrences

G = globally widespread, including species with rare global occurrences

NEP = Northeast Pacific occurrence (Washington to Mexico)

y = species occurring in the NEP

y-s = yes but only California and/or Mexico

n = species not known in the NEP

DNA = genes sequenced or expert assistance

\* Gene codes: (1) *cox1*, (2) *cox3*, (3) ITS rDNA, (4) *rbcL*, (5) *psbC*

\* = further study required

# = monographic expert assistance

## The Species Descriptions

Please use page number or ^F to call up the individual species.



Pg	Brown Algae on JTMD	Global	NEP	DNA
5	<i>Alaria crassifolia</i> Kjellman in Kjellman et Petersen	A	n	(4)
6	<i>Analipus japonicus</i> (Harvey) M.J. Wynne	NP-P	y	(2)
7	<i>Costaria costata</i> (C.Agardh) De A. Saunders	NP-P	y	(2)
8	<i>Desmarestia japonica</i> H. Kawai et al. in Yang et al.	A	n	(1, 3)
9	<i>Desmarestia viridis</i> (O.F. Müller) J.V. Lamouroux	G	y	
10	<b>The <i>Ectocarpus</i> and <i>Kuckuckia</i> conundrum</b>			
11	<i>Ectocarpus acutus</i> Setchell et N.L. Gardner	G	y	(2)*
12-13	<i>Ectocarpus arctus</i> Kützing	G	n	
14	<i>Ectocarpus crouaniorum</i> Thuret in Le Jolis	G	n	(2, 4)
15	<i>Ectocarpus</i> cf. <i>penicillatus</i> (C. Agardh) Kjellman	G	n	(2)*
16	<i>Ectocarpus siliculosus</i> var. <i>pygmaeus</i> (J. Areschoug) T. Gallardo cpx.	G	y	(2)*
17-18	<i>Feldmannia irregularis</i> (Kützing) Hamel	G	y	(2)*
19-20	<i>Feldmannia mitchelliae</i> (Harvey) H.-S. Kim	G	y	(1, 2, 4)
21	<i>Hecatonema</i> cf. <i>streblonematoides</i> (Setchell et N.L.Gardner) Loiseaux	NP-P	y	#
22	<i>Hincksia granulosa</i> P.C. Silva in Silva, Meñez et Moe	G	y	
23-24	<i>Hincksia ovata</i> (Kjellman) P.C. Silva in Silva, Meñez et Moe	G	y	
25	<i>Hincksia sandriana</i> (Zanardini) P.C. Silva in Silva, Meñez et Moe	G	y	
26-28	<i>Kuckuckia</i> cf. <i>spinosa</i> (Kützing) Kornmann cpx.	G	n	(2)*
29	<i>Mutimo cylindricus</i> (Okamura) H. Kawai et T. Kitayama	A+	y-s	(2)
30-31	<i>Petalonia fascia</i> (O.F. Müller) Kuntze cpx.	G	y	(1, 2, 4)
32	<i>Petalonia zosterifolia</i> (Reinke) Kuntze	G	n	(1, 2, 3, 4)
33	<i>Petroderma maculiforme</i> (Wollny) Kuckuck	G	y	(4)
34	<i>Protectocarpus speciosus</i> (Børgesen) Kornmann in Kuckuck	G	n	#
35	<i>Pseudolithoderma</i> cf. <i>paradoxum</i> Sears et Wilce	G	n	#
36-37	<i>Punctaria latifolia</i> Greville cpx.	G	y	(1, 2, 4)*
38	<i>Saccharina japonica</i> (Areschoug) Lane, Mayes, Druehl et Saunders	A+	n	(2, 3)
39	<i>Scytosiphon gracilis</i> Kogame	A+	y-s	(1, 2, 4)
40-41	<i>Scytosiphon lomentaria</i> (Lyngbye) Link cpx.	G	y	(2)
42	<i>Sphacelaria rigidula</i> Kützing	G	y	(4, 5) #
43	<i>Sphacelaria solitaria</i> (Pringsheim) Kylin	G	n	
44-45	<i>Undaria pinnatifida</i> (Harvey) Suringar	A+	y-s	(2)
46-48	Unknowns			
49-56	References.			
57-58	Appendix 1. Tsunami Debris Items.			
59	Appendix 2. <i>Ectocarpus</i> – <i>Kuckuckia</i> <i>cox-3</i>			
60-61	Appendix 3. The JTMD <i>Ectocarpus</i> and <i>Kuckuckia</i> samples sequenced.			

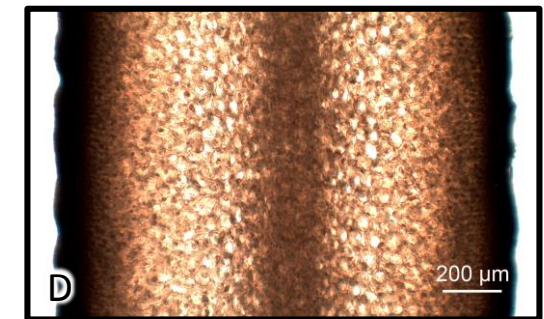
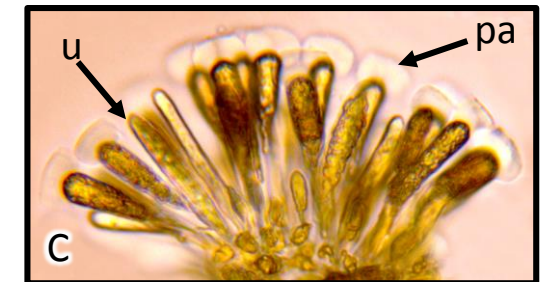


***Alaria crassifolia*★ – A – Asia only (NE Japan and Eastern Russia). Only on the Long Beach (Seaview) debris pipe (Jan). Fertile. Perennial and heteromorphic.**

- Ribbon-shaped blades with a stiff stipe and narrow midrib (~1 cm in diameter).
- Thalli were small on debris (up to 0.5 m tall) but are much larger in Japan (up to 2 m tall).
- Long narrow sporophylls develop bilaterally in dense clusters at the top of the rachis and stipe, are unusually thick when fertile and leave persistent scars when they drop off. Winged sporophyll tips as shown in Miyabe (1902) were not present.

- Reproductive soral areas cover all but the sporophyll tips and contain typical paraphyses with flared tips protecting the developing unangia.
- Frequent in the low intertidal and shallow subtidal in Japan where it is considered a pest in the aquaculture beds of *Saccharina*. Young blades are used for food.
- Kjellman & Petersen (1885), Miyabe (1902, pl. 22).

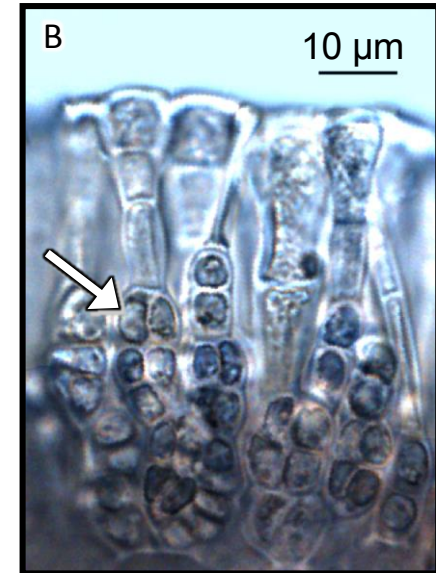
- A. Thalli from debris showing persistent sporophyll scars (arrow).
  - B. Thallus collected along the Tōhoku coast of Japan.
  - C. Squash of fertile sorus showing unangia (u) and paraphyses with flared tips (pa).
  - D. Section of thickened sporophyll.
- ★ SV-55 (BF-293).



***Analipus japonicus*★ – NP-P** – Asia (R, J, K) and AK-CA. Abundant on 2 debris items (May, Jun).  
 Fertile. Pseudo-perennial (the crust overwinters) and isomorphic. **With epiphytic *Pyropia yezoensis*.**



- Erect flattened to tubular axes reaching 10-20 cm. in height, ~1 mm in diameter, and arising from a perennial thick canaliculate crust.
- Axes are covered by abundant short unbranched laterals to 1 cm in length and often epiphytised (A).
- On debris, including the Agate Beach Dock, the species bore intercalary, often biseriate, plurangia (B, arrow).
- Although the same morphological species occurs on both coasts of the N. Pacific, studies of Nelson (1980) have shown that populations in the NE Pacific bear almost entirely plurangia while those in Asia bear both plurangia and unangia.
- Comparative sequencing has indicated that the NE and NW Pacific populations are closely related, but that 6 base pairs in the *cox3* gene do separate them (Hanyuda *et al.*, 2017). The debris specimen was the Japanese haplotype.
- Abbott & Hollenberg (1976, fig. 146); Nelson (1982a & b), Tokuda *et al.* (1994), Okamura (1908, pl. XXXV, as *Chordaria abietina*).
- ★ AB-5778 (BF-1). Not sequenced, Nye (BF-59).



Section through fertile branch with intercalary plurangia (arrow).



***Costaria costata***★ – NP-P – Asia (J, K), AK-CA. Only on the Agate Beach derelict dock (Jun).  
 “5-mid-ribbed kelp”. A sequence variant of NEP populations. Sterile. Heteromorphic and usually annual.

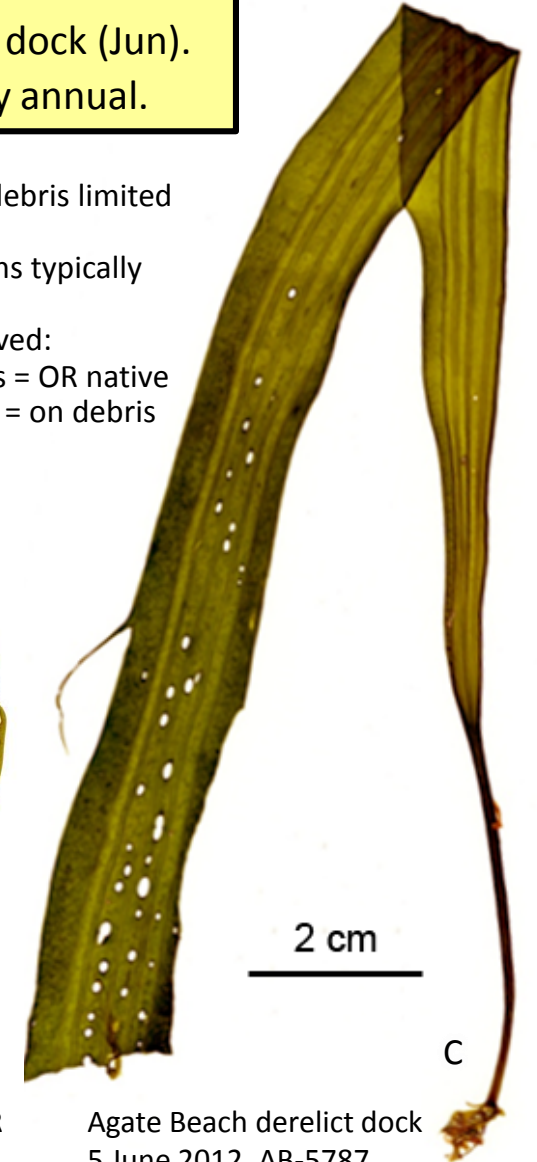


*Costaria* bed on the Tōhoku coast of Japan

- “Searsucker” kelp reaching 2 m long in Japan (on debris limited limited to 1 m) with a large digitate holdfast (A).
- With 5 midribs, 3 on top and 2 below and bullations typically forming between the ribs.
- The 2 forms of Miyabe & Nagai (1940) were observed:  
 B – f. *latifolia*, described as from sheltered habitats = OR native  
 C – f. *cuneata*, described as from exposed habitats = on debris
- The debris thalli had long stipes, acute bases, and long narrow blades, 2-5 cm wide (C).
- Oregon thalli had broader, more ovate blades with with rounder bases (B).
- Postels & Ruprecht (1840, tab. XXIV), Okamura (1925, pl. CCXXVI), Nagai (1940).
- ★ AB-5787 + pressing (BF-1) = sequence variant of OR & NEP material.  
 (Sequence variants occur on both coasts, but we are not yet certain how they relate to morphology.)



North Boardman, OR (native), 2 July 2000

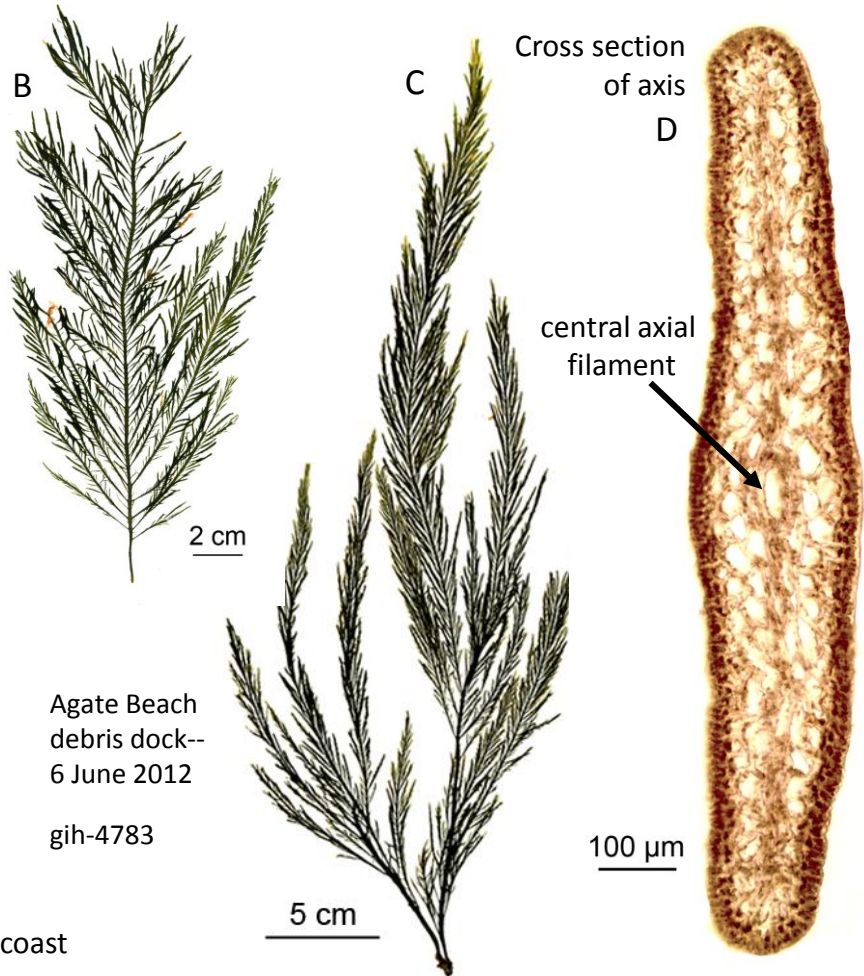


Agate Beach derelict dock 5 June 2012, AB-5787





***Desmarestia japonica*★ – A – Asia only (J).** On the Agate Beach derelict dock (Jun). Sterile. Annual, heteromorphous with a filamentous monoecious gametophyte.



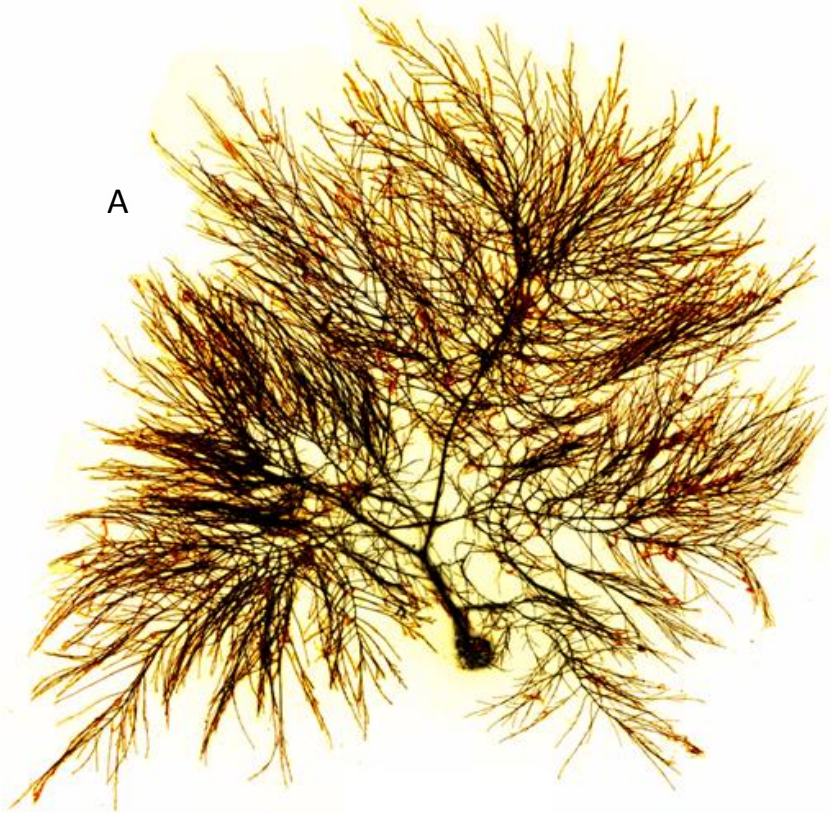
Agate Beach debris dock--  
6 June 2012  
gih-4783

*Desmarestia japonica* on the Tohoku coast

- Erect, highly branched dark brown thalli up to 1 (2) m long in Japan (A).
- Narrow, opposite branches, 2-6 mm in diameter, with a central midrib (B-C).
- Terminal and marginal trichothallic filaments that initiate growth were not apparent in the JTMD material.
- Axes and branches contain a large central axial filament surrounded by corticating filaments that make up the pseudo-parenchymatous structure (D).
- Vacuoles contain sulfuric acid.
- Similar morphologically to widespread *D. ligulata*, but molecular sequences, chromosomes, and subtle culture features indicate that this is a separate species unique to Japan.
- Yang *et al.* (2014), Peters & Müller (1986).
- ★ AB-5A (BF-1); gih-5783 pressings at left.

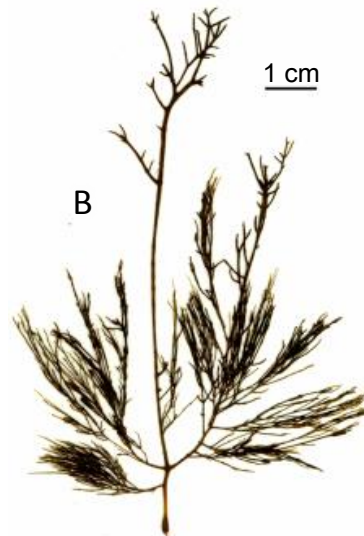


***Desmarestia viridis* – G (Globally widespread) –** Asia (R, J, C, K), Arc, AK-WA, CA-MX, SA, A-Arc, ENA, EUR, Medit. Only on the Agate Beach derelict dock (Jun). Sterile. Annual and heteromorphic.

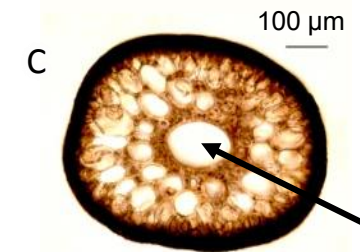


Possibly requires a fully saline sheltered habitat, explaining its rarity in Oregon.

- “Stringy acid weed” – a sulfuric acid containing *Desmarestia*.
- **On the “100 worst invaders in the Mediterranean” list** of Streftaris & Zenetos (2006).
- A highly branched thallus typically reaching 0.5 m in length in the NE Pacific (A, B).
- Trichothallic growth and a large central axial filament characterize the genus (C).
- Terete to slightly flattened axes, 1-2 mm broad, typically with primarily opposite, delicate and highly divided lateral branches.
- Abbott & Hollenberg (1976, fig. 187), Fletcher (1987, figs. 85, 86), Boo *et al.* (2010), Verlaque *et al.* (2015), Peters *et al.* (1997).
- AB-11 (BF-1), but the DNA extraction failed.



Eroded specimen from the Agate Beach debris dock. (BF-1)



Cross section of axis revealing large central axial filament (arrow).

## The *Ectocarpus* and *Kuckuckia* Conundrum

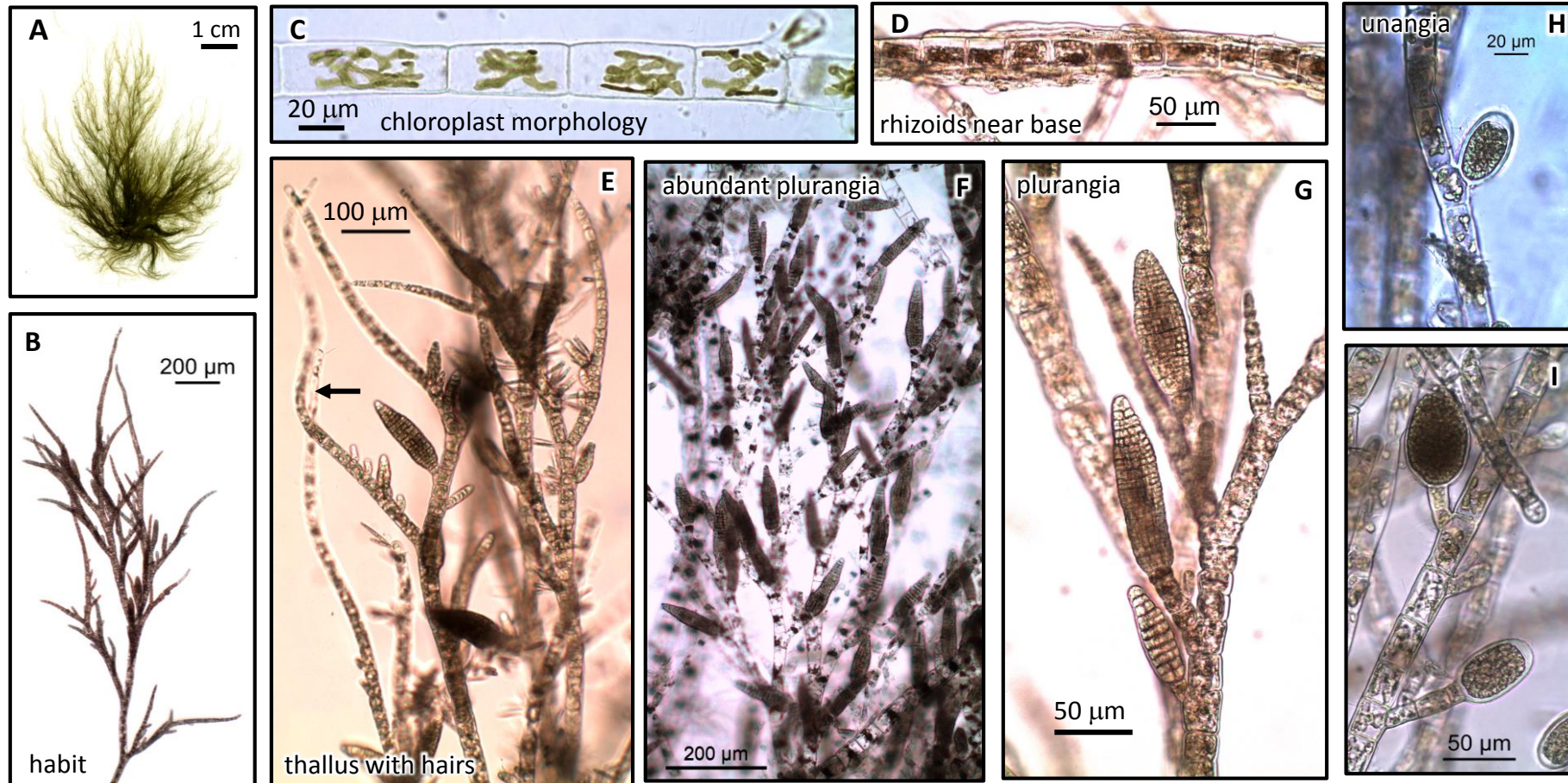
- The species in *Ectocarpus* and *Kuckuckia* were not only the most abundant on debris but also the most difficult to identify. Due to their morphological similarity, frequent deteriorated state and problematic genetic characterization, many of the final identifications in this group of species are approximate.
- Some of the problems encountered with the identifications:
  - Only partially disintegrated material without visible chloroplasts (important in generic identification) was present for some species.
  - The literature descriptions occasionally conflict. Over the years, different authors have applied slightly different features to identify the species and some have provided incorrect synonymies. These errors are then passed along by others in identification keys, etc. For this study, we provide references for the species identifications and try to use the following traditional features (when available) for our morphological identifications: chloroplast structure; shape, size and location of plurangia and unangia; filament diameter, cell width to length ratio, and shape of the filament apex; rhizoid presence; and the occurrence and abundance of hairs (differentiating phaeophycean hairs and pseudohairs when possible).
  - Field collections and separation of species. In field collected material, species are often intermixed, and they must be separated under a microscope for sequencing. For species that intertwine and attach to one another, separation is difficult. Although specific genes can be targeted for some species in these mixed samples, this is not always successful – so contamination might account for some of our DNA discrepancies.
  - Sequencing data could not be obtained from some samples due to their small size or to extraction difficulties.
  - Possible sequence errors in GenBank. The sequences deposited in GenBank occasionally come from misidentified species. For easily confused species, it is important that sequences (preferably multiple gene sequences) are obtained and deposited in GenBank from holotype (or lectotype) material so that truly accurate identifications are possible. Although research is underway in this area, it has happened for very few species in the *Ectocarpus* complex. We are looking forward to having access to the sequences of all of the holotypes (or lectotypes) in this group and also to additional culturing research on the species that will help us determine the stable morphological features useful in identification.
- Determining *Ectocarpus* and *Kuckuckia* species names for this project. Hanyuda *et al.* (2017) have produced a phylogenetic tree of the *Ectocarpus* complex, including the JTMD samples. JTMD species are in 5 of the clades, but names could be provided for only 2 of these: *Ectocarpus crowaniorum* and *Kuckuckia spinosa*. The morphological features of the JTMD samples grouped in these 2 known clades and in the 3 unnamed clades have been used, along with the morphological literature, to provide tentative names for the species. See Appendix 2 and 3 for this *Ectocarpus* and *Kuckuckia* tree and for the JTMD specimens used in this study.
- It is our hope that the final naming of the species in this abundant and complex group will be facilitated by our JTMD studies, including both the sequences provided by our molecular study (Hanyuda *et al.*, 2017) and by the photographic documentation in this account.



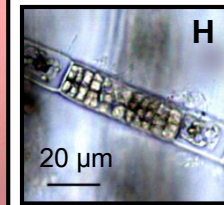
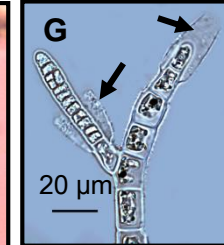
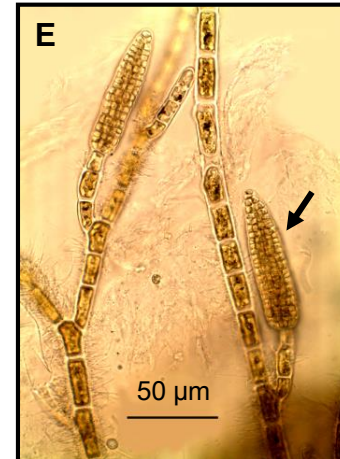
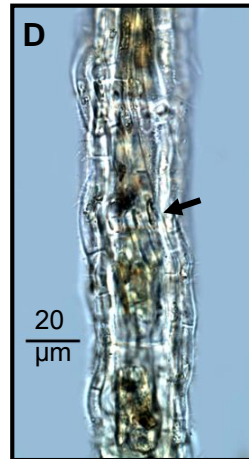
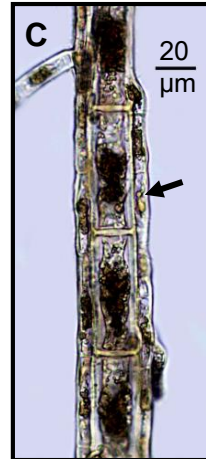
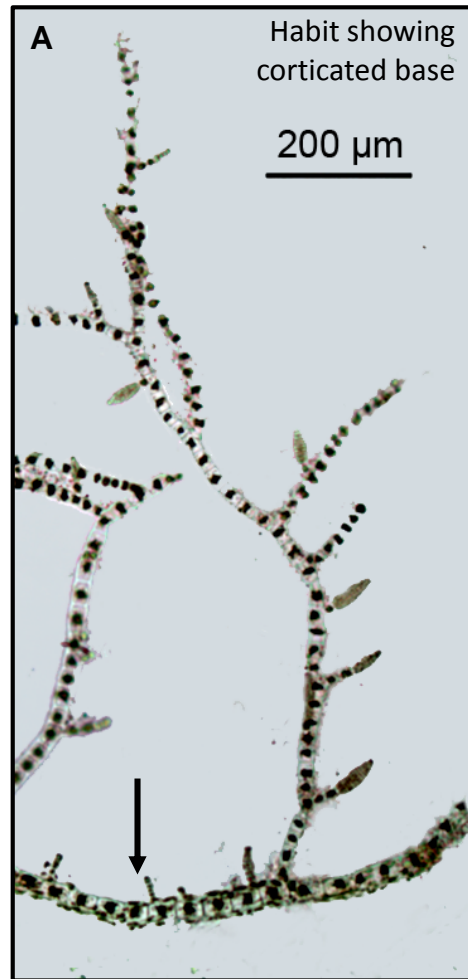
***Ectocarpus acutus*<sup>★</sup> – G (Globally widespread) — Asia (C, K), BC-MX, SA, Afr. On 12 debris items (Jan-May).**

Reproductive (plurangia, unangia). Spring ephemeral, isomorphic. ★ Wal-1 (BF-196); RE-686, 690, 699 (BF-533); Bev-113 (BF-288). Not sequenced: GB (BF-23), MP (BF-36). All are in Hanyuda clade *E. sp. 2* (see Appendix 2 and 3). Kim & Lee (1992a, figs. 9, 11 A-D as *penicillatus*), Kim (2010, fig. 2), Setchell & Gardner (1922, figs. 36-39).

Thalli are 4-5 cm tall with alternate branching and secund branches near tips, laterals that are often slightly reflexed, pointed apices, and occasional hairs (A, B, E). Chloroplasts are ribbon-shaped (C), and rhizoids envelope the axis near the base (D). Plurangia are cylindroconical with rounded tips (E-G). Unangia are ellipsoidal (H, I). Both are stipitate or sessile and often occur near the base. Diameters: axes (16-40  $\mu\text{m}$ ), rhizoids (8  $\mu\text{m}$ ), plurangia (16-22 x 70-122  $\mu\text{m}$ ), unangia (20-40 x 38-60  $\mu\text{m}$ ).



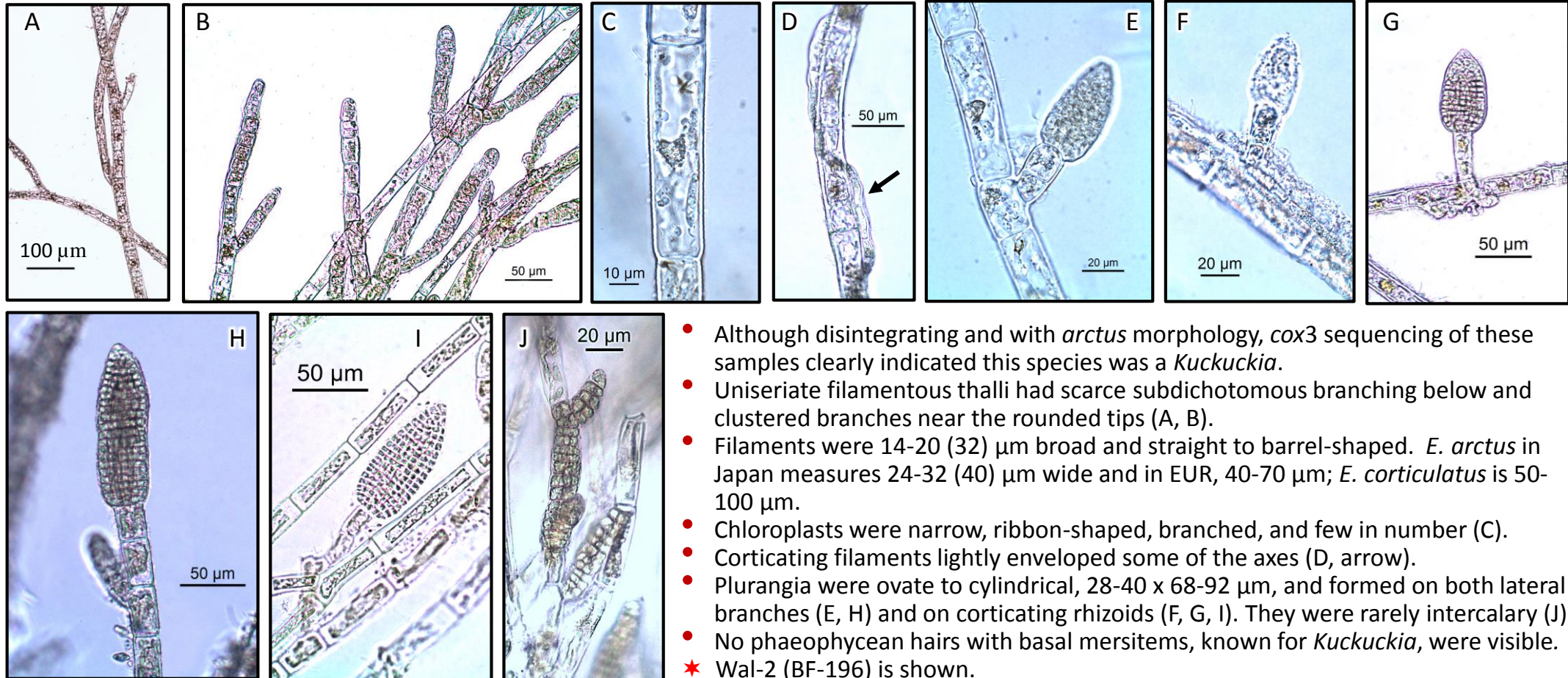
***Ectocarpus arctus* 1** (including *E. siliculosus* var. *arctus*) — **G** (Globally widespread) — Asia (R, J, K), ENA, EUR, Med). On 6 debris items (Feb, Apr-Jun, Oct). Reproductive (plurangia). Ephemeral. Similar species: *E. corticulatus* (a larger species).



- Uniseriate branching thallus, up to 2-3 cm tall, with multiple erect axes arising from a rhizomatous base (A, arrow).
- Branching is sub-dichotomous, alternate or unilateral.
- Main axial cells are 20-32 (40)  $\mu\text{m}$  in diameter with cells 1-3 times as long as broad, bearing a few ribbon-shaped chloroplasts (B). (EUR = 40-70  $\mu\text{m}$ ). Branch tips occasionally extended into hair-like extensions with rounded tips, 12  $\mu\text{m}$  in diameter.
- Rhizoids, 6-8  $\mu\text{m}$  broad, develop on the axes and often heavily corticate the lower branches and rhizome (C, D, arrows).
- Both normal branches and rhizoids form stalked or terminal plurangia (arrows) that are typically ellipsoidal with narrowed tips and ranging from 12-16 x 55-66  $\mu\text{m}$  in size (E, F).
- Empty plurangial walls were evident and occurred at the base of newly regenerating plurangia (G, arrows).
- Intercalary plurangia occurred but were very rare (H).
- Cormaci *et al.* (2012, pl. 70, figs. 1-4), Cardinal (1964, fig. 7), Lee (2008, p. 74), Yoshida (1998, pl. 2-1, figs. B, F). This species does not appear to be a synonym of *E. acutus* (that lacks rhizoidal plurangia) as proposed by Kim (2010, p. 19).
- Not sequenced: AB (BF-1), SBT (BF-234), CBD (BF-130), SVB 480 (BF-402).
- Although *E. arctus* is similar to *E. corticulatus* in having plurangia borne on both axial and rhizoidal filaments, the axial filaments of *arctus* are narrower than the 50-130  $\mu\text{m}$  wide filaments of *corticulatus*, a more robust species.



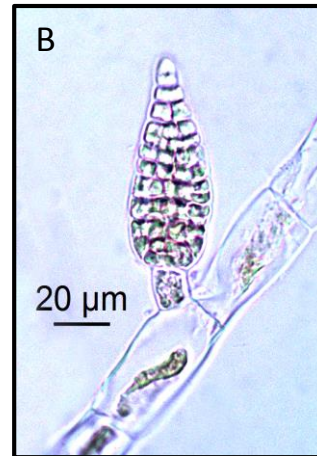
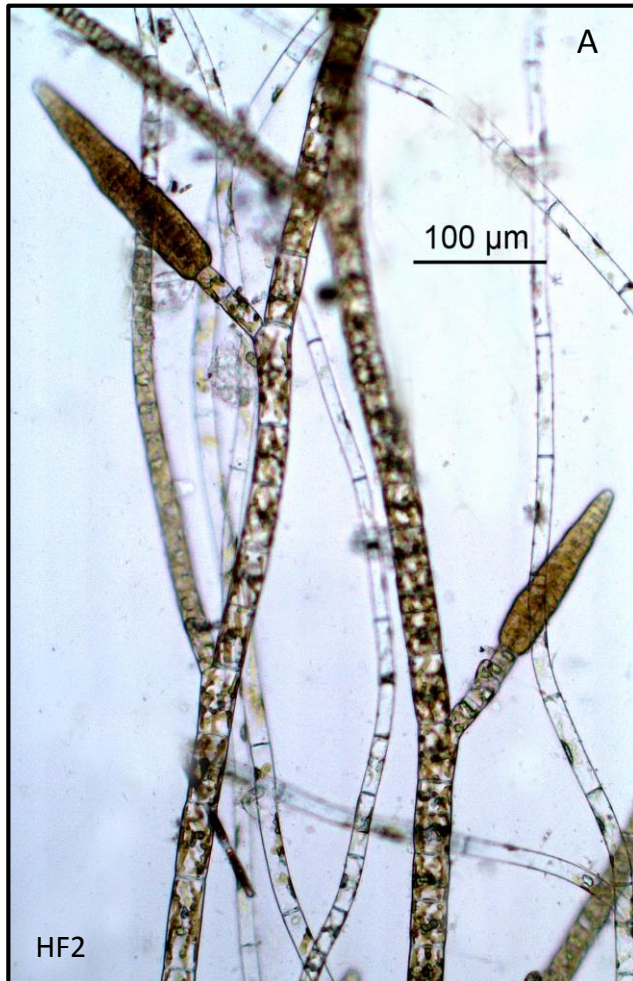
***Ectocarpus arctus* 2<sup>★</sup>** – but *cox3* sequencing indicated the *Kuckuckia* clade of Hanyuda. <sup>★</sup>Wal-2 (BF-196) was partially disintegrated, but it had the clear diagnostic characteristics of *E. arctus*: narrow filaments and small plurangia born on both normal and enveloping rhizoidal filaments. The sample appeared pure, but it sequenced as a *Kuckuckia spinosa*. There are several possibilities for this: (1) the genus may be highly variable morphologically, (2) the *cox3* gene may not separate the *Kuckuckia* clade well, (3) sample contamination. Note that the synonymy of *Ectocarpus spinosa* Kuetzing (1843) (= *Kuckuckia*) with *Ectocarpus arctus* Kuetzing (1843) was suggested by Ardissonne (1886) but was later rejected by Kuckuck (1958, p. 174).



- Although disintegrating and with *arctus* morphology, *cox3* sequencing of these samples clearly indicated this species was a *Kuckuckia*.
- Uniseriate filamentous thalli had scarce subdichotomous branching below and clustered branches near the rounded tips (A, B).
- Filaments were 14-20 (32)  $\mu\text{m}$  broad and straight to barrel-shaped. *E. arctus* in Japan measures 24-32 (40)  $\mu\text{m}$  wide and in EUR, 40-70  $\mu\text{m}$ ; *E. corticulatus* is 50-100  $\mu\text{m}$ .
- Chloroplasts were narrow, ribbon-shaped, branched, and few in number (C).
- Corticating filaments lightly enveloped some of the axes (D, arrow).
- Plurangia were ovate to cylindrical, 28-40 x 68-92  $\mu\text{m}$ , and formed on both lateral branches (E, H) and on corticating rhizoids (F, G, I). They were rarely intercalary (J).
- No phaeophycean hairs with basal mersiterns, known for *Kuckuckia*, were visible.
- ★ Wal-2 (BF-196) is shown.

***Ectocarpus crouaniorum***★ – G (Globally widespread) — Asia (K), EUR, Med, SA (Ch), but probably overlooked elsewhere.

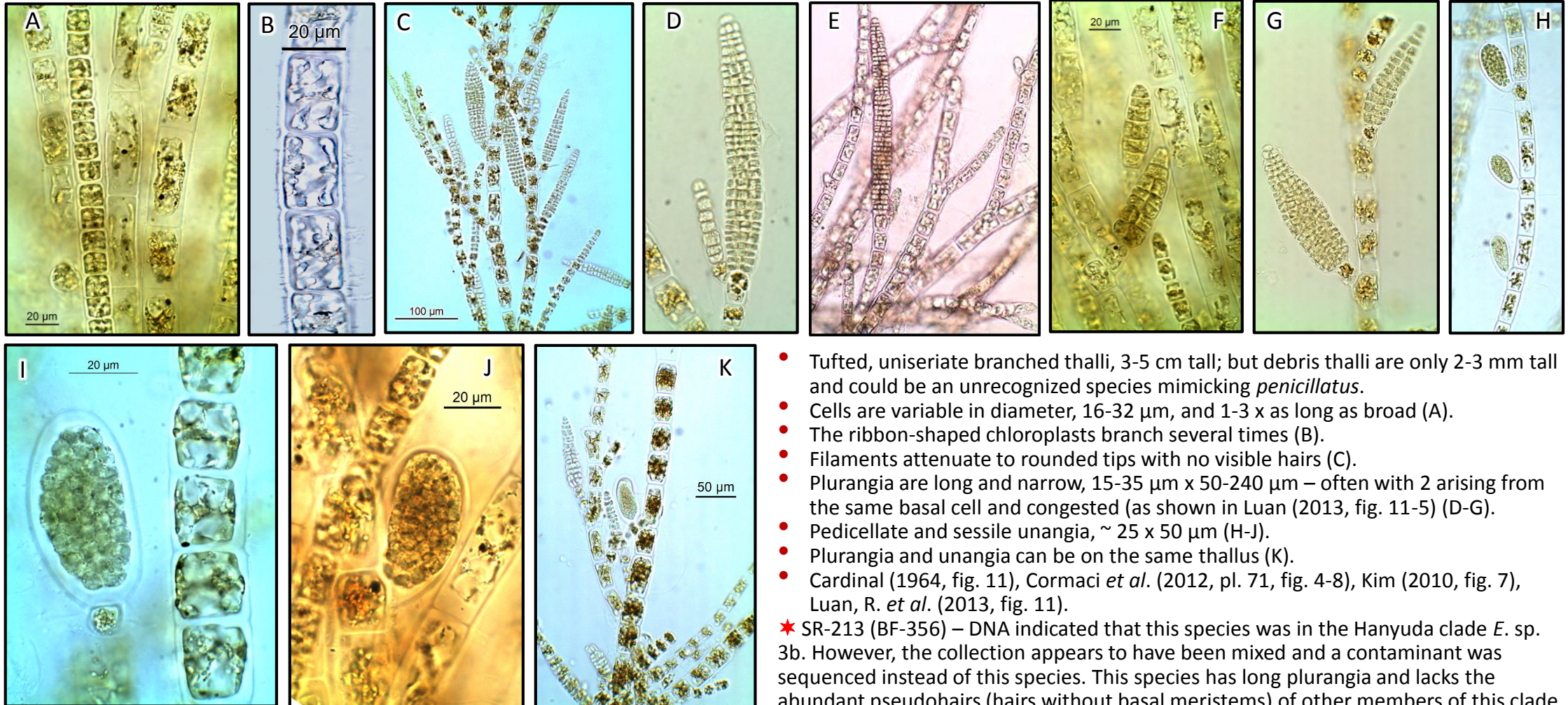
On 5 debris items (Jan-Apr). Ephemeral, isomorphic. Reproductive (plurangia and unangia). ★ NC-5 (BF-208). Not sequenced: HF2-650 (BF-526), Fal-932 (BF-652).



- Unilateral, alternate or irregularly branched filaments often with wide-angle branching that can reach 1-5 cm. in height (A). Rhizoids (not seen) occur near the thallus base.
- Axes were 30-40 μm in diameter near the base, 18-24 μm above and continue to narrow extending into long hyaline hairs, 11 μm in diameter.
- Cells were typically 1.5-2 times as long as broad and contained vertical ribbon-shaped plastids.
- Plurangia were terminal on long laterals (A), diagnostic for the species, and on shorter pedicels (B,C) or occasionally sessile in our material; 30-50 x 80-150 μm. Plurangia also occasionally had narrowed conical apices, a feature only shown in Cardinal (1964, fig. 8) and Cormaci *et al.* (2012, pl. 67, fig. 4).
- Unangia (E) were sessile and 35 x 50 μm in size (small for the species).
- Cardinal (1964, fig. 8 as *confervoides* var. *crouanii*); Cormaci *et al.* (2012, pl.67, figs. 1-4); Hamel (1939, fig. 2f), Kuckuck (1964, p. 12/228) and Peters *et al.* (2010, figs. 5-10).
- Similar species – drawings of *E. parvus*: Saunders (1898, pl. XXII, fig. 1., *E. siliculosus parvus* var. nov.), Norris (2010, fig. 89).



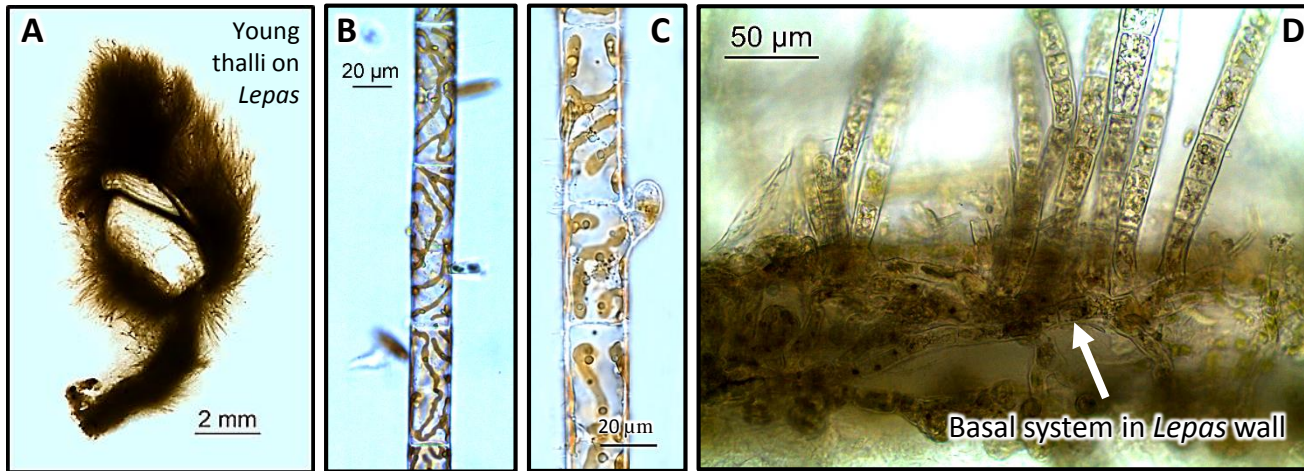
***Ectocarpus cf. penicillatus***★ – G (Globally widespread) — Asia (J, C, K) , SA, ENA, EUR, Med, A-Arc. On 6 debris items as hemispherical epiphytic tufts on *Scytosiphon* (Mar, May-Jul, Oct). Fertile. Isomorphic, ~ephemeral. Characterized by slender plurangia up to 240  $\mu\text{m}$  long and cylindrical or tapering to a point. Very problematic with mixed descriptions in the literature.



- Tufted, uniseriate branched thalli, 3-5 cm tall; but debris thalli are only 2-3 mm tall and could be an unrecognized species mimicking *penicillatus*.
- Cells are variable in diameter, 16-32  $\mu\text{m}$ , and 1-3 x as long as broad (A).
- The ribbon-shaped chloroplasts branch several times (B).
- Filaments attenuate to rounded tips with no visible hairs (C).
- Plurangia are long and narrow, 15-35  $\mu\text{m}$  x 50-240  $\mu\text{m}$  – often with 2 arising from the same basal cell and congested (as shown in Luan (2013, fig. 11-5) (D-G).
- Pedicellate and sessile unangia, ~ 25 x 50  $\mu\text{m}$  (H-J).
- Plurangia and unangia can be on the same thallus (K).
- Cardinal (1964, fig. 11), Cormaci *et al.* (2012, pl. 71, fig. 4-8), Kim (2010, fig. 7), Luan, R. *et al.* (2013, fig. 11).
- ★ SR-213 (BF-356) – DNA indicated that this species was in the Hanyuda clade *E. sp.* 3b. However, the collection appears to have been mixed and a contaminant was sequenced instead of this species. This species has long plurangia and lacks the abundant pseudohairs (hairs without basal meristems) of other members of this clade.

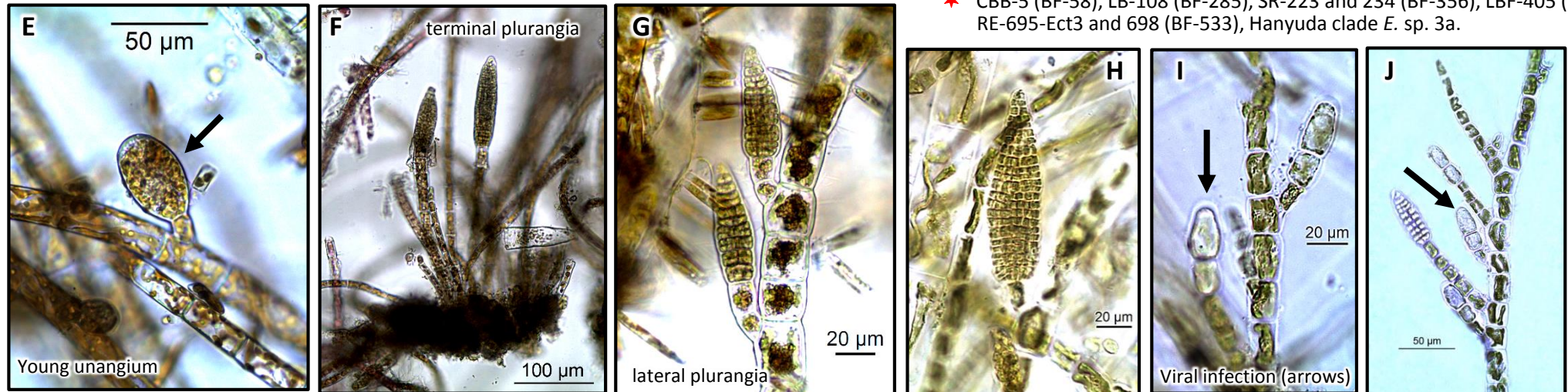


***Ectocarpus siliculosus* var. *pygmaeus* cpx.** \* – **G** (Globally widespread) — Asia (J, K, C, with synonyms listed below), AK-MX, SA (Br), ENA, EUR, Med. On 22 debris items (Jan-Jul). On *Lepas anatifera*, the pelagic goose neck barnacle, and on other algae. Reproductive (plurangia and unangia). Ephemeral and Isomorphic.



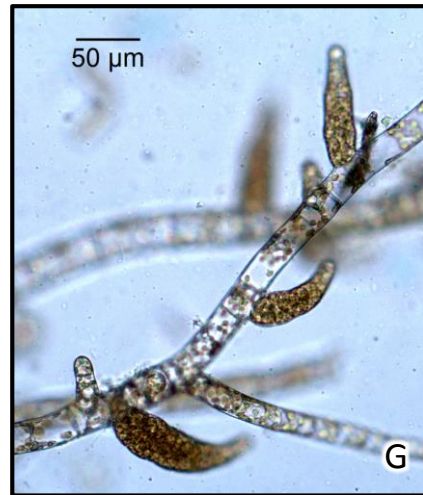
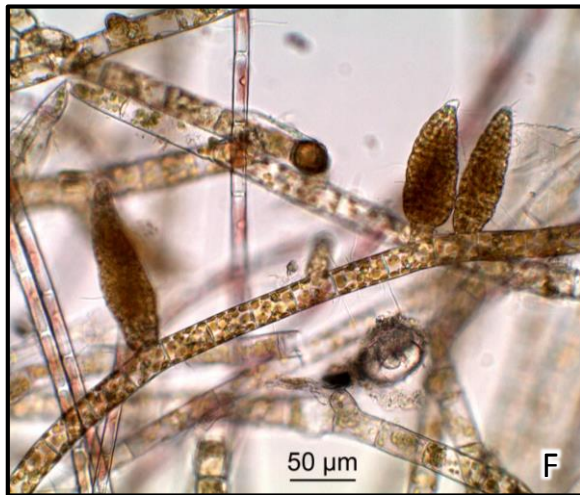
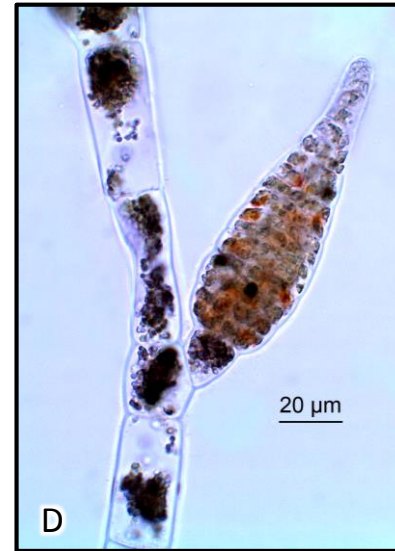
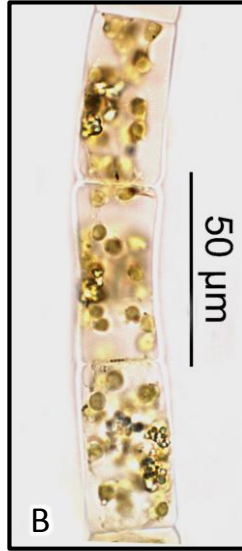
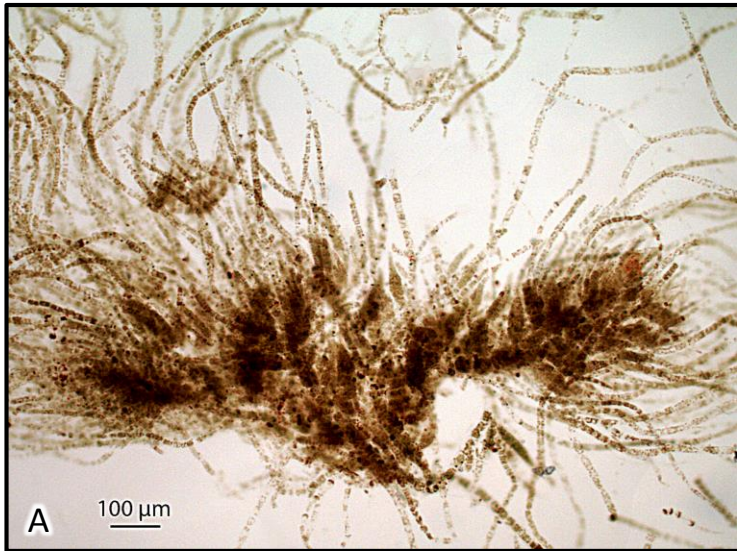
- Irregularly branched filaments, ~0.5-1.5 (3.0) cm tall on debris, epiphytic on and often penetrating the walls of *Lepas* and also larger algae (A).
- Cells are 12-25 μm in diameter, 1:1-1:3, and have variable band-shaped chloroplasts (B, C).
- The branched basal filaments penetrate into the host wall (D).
- Pseudohairs (without noticeable meristems) were present in larger thalli.
- Young ovoid unangia (38x44 μm) were present (E).
- Plurangia were terminal on younger filaments but often lateral in on mature filaments – cylindro-conical and 20-30 x 50-100 μm (F, G, H).
- Virally infected plurangia were found in one collection (I, J, arrows).
- Saunders (1898, pl. XV, figs. 5-9), Cormaci *et al.* (2012, pl. 72, figs. 1-3), Yamada & Tanaka (1944, fig.3, as *E. yezoensis*), Ohta (1973, fig.6), Setchell & Gardner (1922, pl. 48, fig 32, as *commensalis*), Cardinal (1964, fig. 10).
- Apparent synonyms: *E. commensalis* (non *parvus*), *E. confervoides* var. *pygmaeus*, *E. dimorphus*, *E. yezoensis*, *Kuckuckia kylinii* of Kim (2010, fig. 11A-C).

\* CBB-5 (BF-58), LB-108 (BF-285), SR-223 and 234 (BF-356), LBF-405 (BF-462), RE-695-Ect3 and 698 (BF-533), Hanyuda clade *E. sp.* 3a.





***Feldmannia irregularis* 1<sup>★</sup> – G (Globally widespread) –** Asia (R, J, C, K, Phil), Aus, NZ, SA, AK-CA, ENA, Car, Afr, EUR, Med, Arc. In turf on 3 derelict boats (May, Mar). Fertile (plurangia, unangia). Ephemeral and isomorphic.

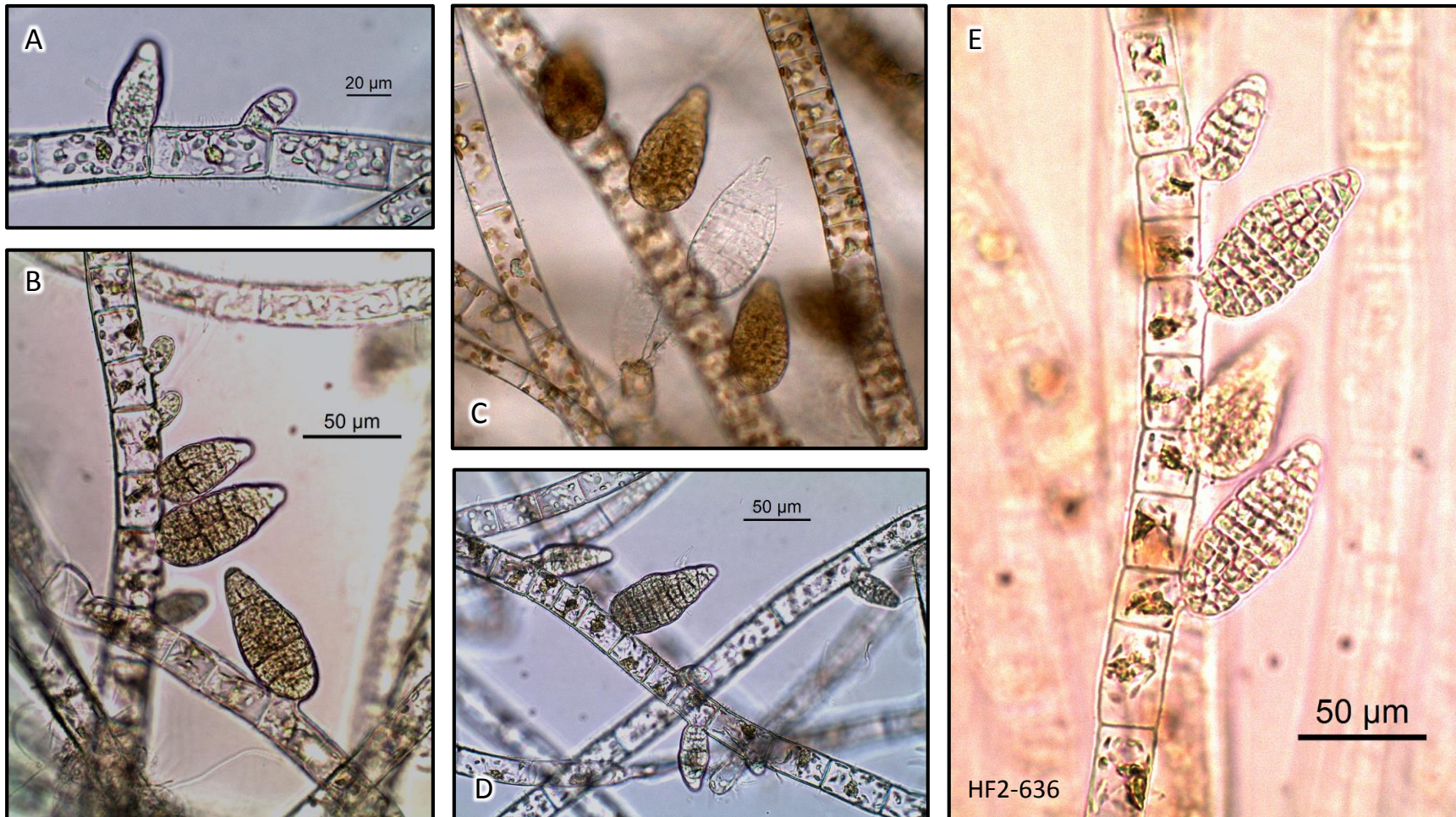


- Tufted filamentous brown alga occurring in the turf of derelict boats (A).
- Thalli 2-5 mm tall with filaments up to 24 μm wide bearing discoidal chloroplasts (B).
- Typically branched 1-3 times near the base with long hair-like filaments extending upward.
- Elongate-conical plurangia, curved or straight, form near the thallus base and occasionally in the upper branches, typically below the growth zones (C, D, F, G).
- Plurangia: sessile or rarely pedicellate, 20-40 μm x 90-150 μm (D).
- Unangia: sessile, globose and up to 50 μm in diameter (E).
- Kim & Lee (1994, fig. 1A-B, fig. 2A-C), Cardinal (1964, fig. 29), Sauvageau (1933, fig. 24-26), Stegenga & Mol (1983).
- Similar species: *Acinetospora filamentosa* (Noda) Yaegashi *et al.* (2015, fig. 2), a species originally described with ellipsoidal (not globose) unangia (Noda, 1970, fig. 3, 1980, fig. 33). Our species is morphologically similar to Yaegashi's images, but Hanyuda's sequencing indicates that our species is true *F. irregularis*.
- ★ SVB-404 (BF-402). Not sequenced: HF2 (BF-526), NC (BF-208).



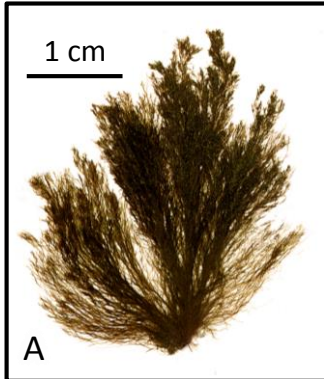
## ***Feldmannia irregularis* 2, the arabicus form – HF2-636 (Mar)**

Branching uniseriate filaments with discoidal chloroplasts – intermixed with normal *Feldmannia irregularis*. Axes are 20-24  $\mu\text{m}$  in diameter; sessile plurangia reach 24 x 68  $\mu\text{m}$  with slightly narrowed tips (A-E). Identical to drawings of Kuckuck (1963, fig. 6) for this species and illustrated by Kim & Lee (1994, fig. 4D-E).

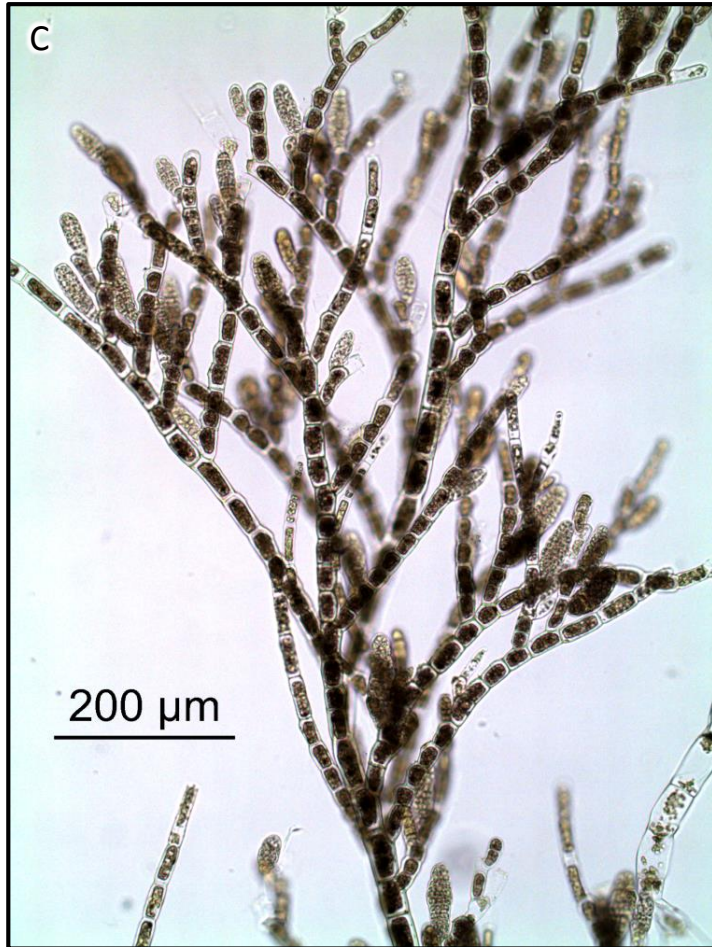




***Feldmannia mitchelliae***★ – **G** (Globally widespread) — Asia (J,C,K, Phil), IO, Aus, NZ, SA, CA-MX, ENA, Car, Afr, EUR, Med. Abundant on 24 debris items (all debris months). Fertile – some thalli with both plurangia and unangia. Ephemeral and isomorphic.



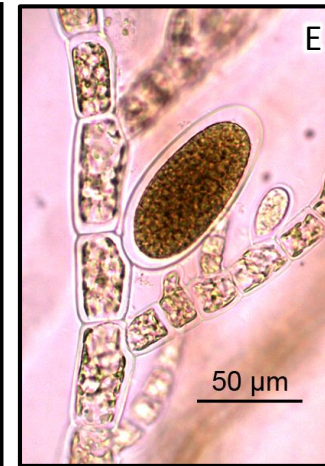
Often bi-lobed chloroplasts



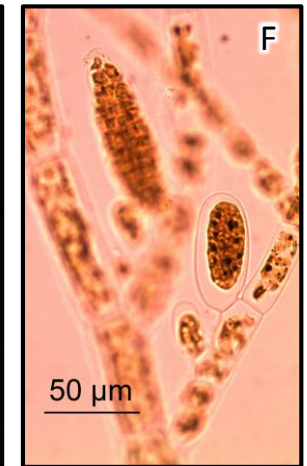
Upper branches with some unilateral branching



Arrangement of plurangia



A mature unangium



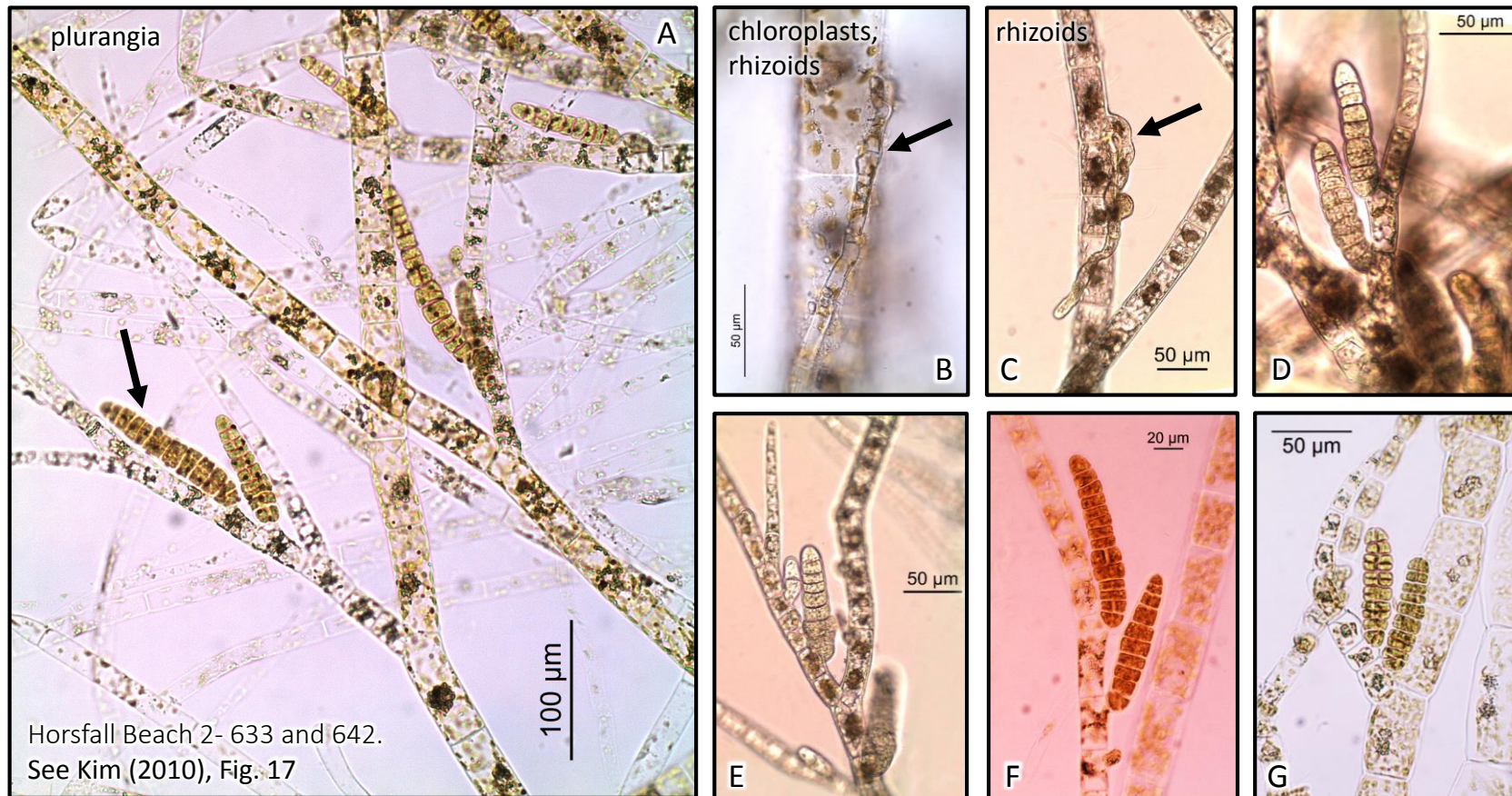
Both types

- Densely tufted to 3 (4) cm tall (A).
- Filaments tapering, 20-40 μm in diameter often with short-celled meristematic branch bases.
- Chloroplasts vary from discoidal to bilobed to slightly elongate (B).
- Sessile reproductive structures develop near the top of the thallus (C).
- Plurangia, 35-40 x 40-140 μm, with large locules (D).
- Unangia are ovoid, 15-20 μm x 40-45 μm (E) – some thalli have both unangia and plurangia (F).
- Kim & Lee (1992b, fig. 3), Kim (2010, figs. 21-22), Cardinal (1964, fig. 23).
- ★ MC-8 (BF-3), GB-1 (BF-23), HF-5 (BF-29), SR-201 (BF-356), SixR-724 (BF-538).



***Feldmannia mitchelliae* 2 – the *indica* form** ★ – True *F. indica* is also globally widespread: known in Asia (J, C, K, Phil), IO, Aus, NZ, SA, CA-MX, ENA, Car, Afr. This form was on 2 of the 24 debris items. Reproductive (plurangia) and ephemeral.

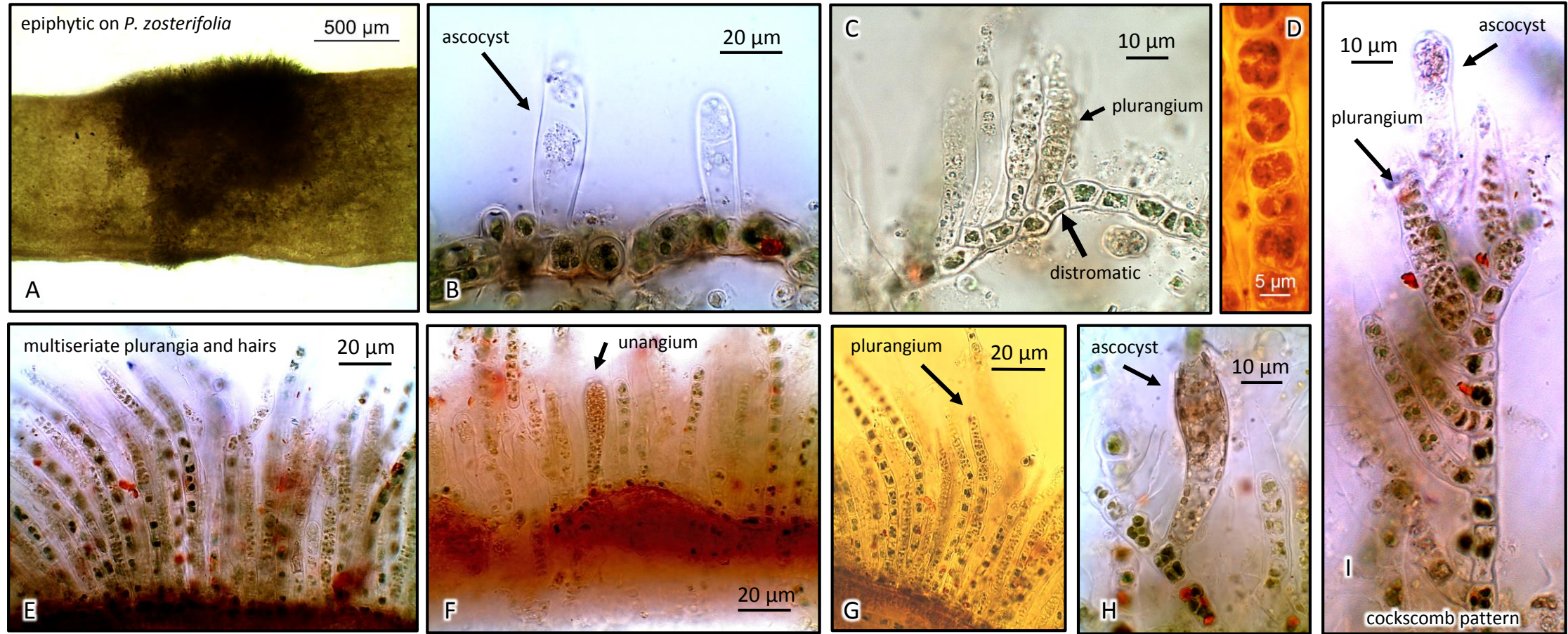
The “*indica*” form is characterized by discoidal chloroplasts (B) and sessile, long, narrow plurangia (A, D-G). Our form developed rare rhizoids (B, C, arrows). This isolate was found to be a morphological variant of *F. mitchelliae* since DNA sequences could not separate the two. See Kim (2010, fig.17) for *F. indica* in Korea. ★ HF2-633 (BF-526), RE-691 (BF-533); not sequenced HF2-642 (BF-526) (all Mar).





***Hecatonema cf. streblonematoides*<sup>#</sup> – NP-P –** Asia (J), AK-MX, SA (Ch). Only on the Seaview boat (May), SVB (BF-402). Not sequenced. Reproductive (multiseriate plurangia and sessile unangia). Ephemeral (year around in CA) and heteromorphic.

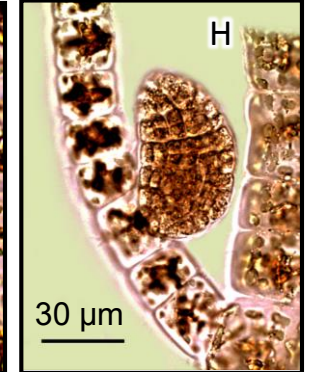
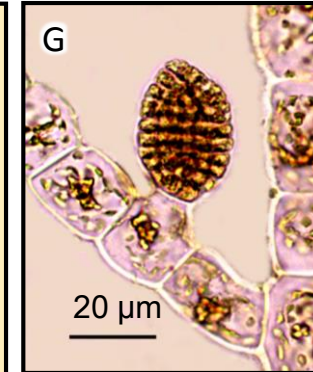
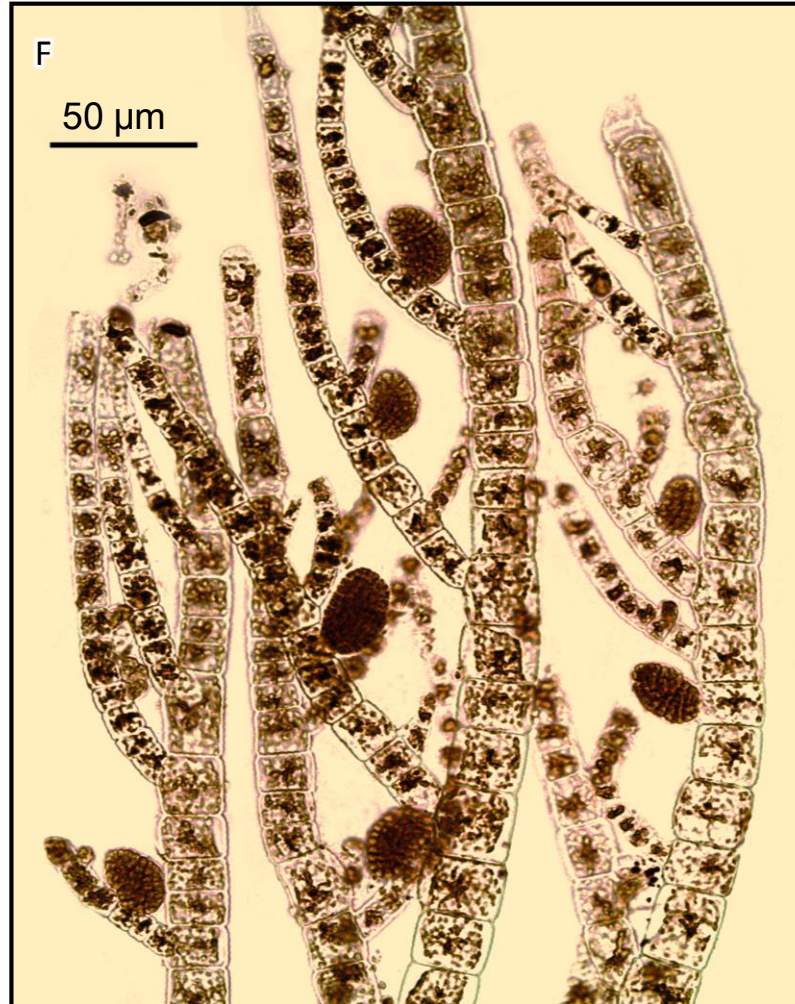
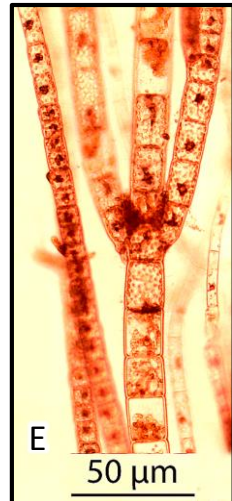
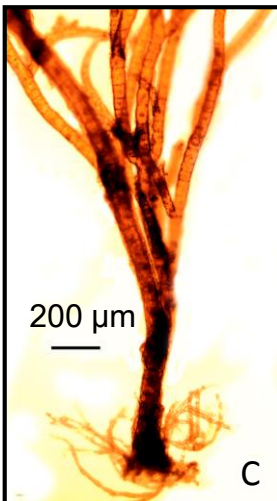
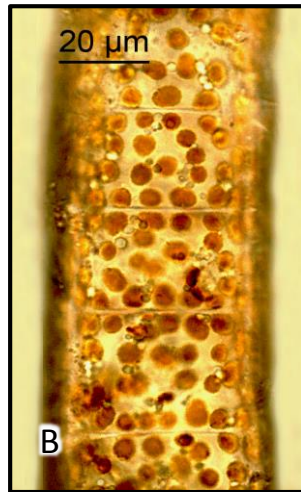
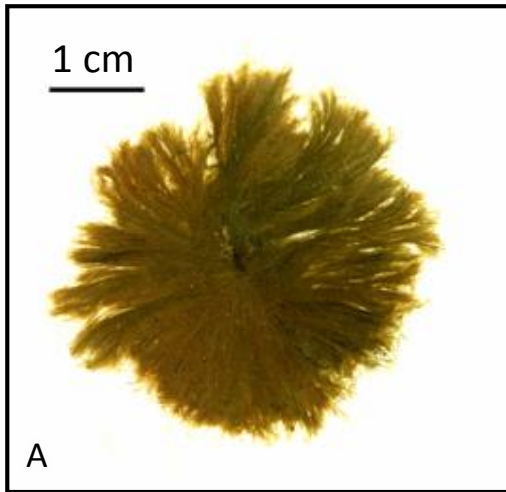
Forming 1-10 mm cushions on *Petalonia zosterifolia* (A). Partially distromatic basal layer (arrow) producing upright filaments, uni-multiseriate plurangia, unangia and ascocysts (B, C, E-H). Older uprights occasionally produce a cockscomb unilateral arrangements of plurangia and ascocysts (I). With discoidal chloroplasts (D).



<sup>#</sup> = Loiseaux-de Goër (4/21/2017 pers. com) confirmed the morphological identity of this species from pictures but also stated that sequencing and comparison with the type would be critical for absolute identification. See Loiseaux (1970), fig. 2.



***Hincksia granulosa* – G (Globally widespread) –** Asia (J, C, K), Aus, SA, AK-MX, ENA, Afr, EUR, Med.  
 On 7 debris items (Jan-Jun). Reproductive (only plurangia were observed). Ephemeral (Mar-Dec in EUR – Cardinal, 1964).



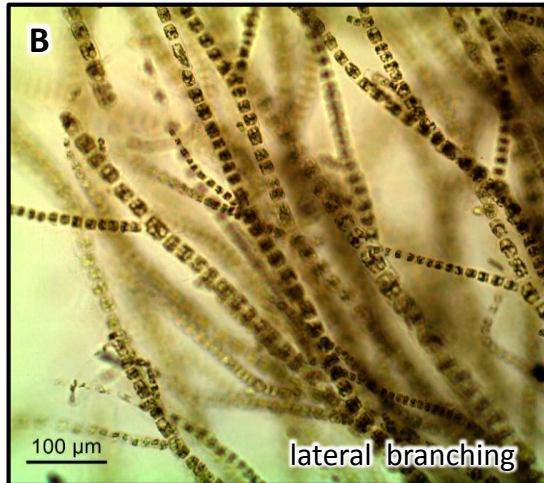
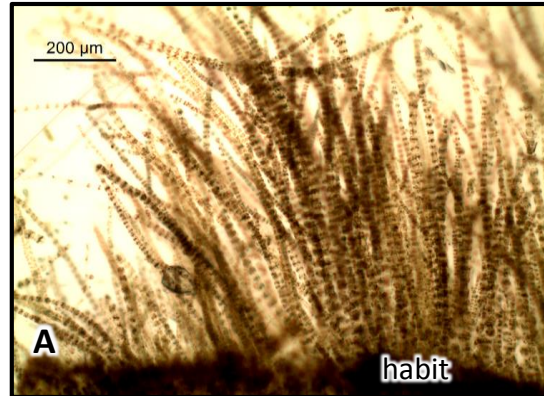
- A. Densely tufted branched filaments reaching up to 7 (25) cm, but on debris only 3-4 cm tall.
- B. Short cells with discoidal chloroplasts.
- C. Rhizoidal thallus base.
- D. Main axis with heavy cortication near the thallus base.
- E. Characteristic opposite branching in the main axis.
- F. Upper part of reproductive thallus with unilateral branching and sessile plurangia.
- G. & H. Straight and curved sessile plurangia.

Womersley (1987, fig. 13 A-D), Kim (2010, fig. 25), Kornmann & Sahling (1977, pl. 55), Cardinal (1964, fig. 19-20).

Examples not sequenced: AB (BF-1), SV (BF-293), SBT (BF-234), CBB (BF58), Nye (BF-59), Oys (BF-331), SixR (B-538).



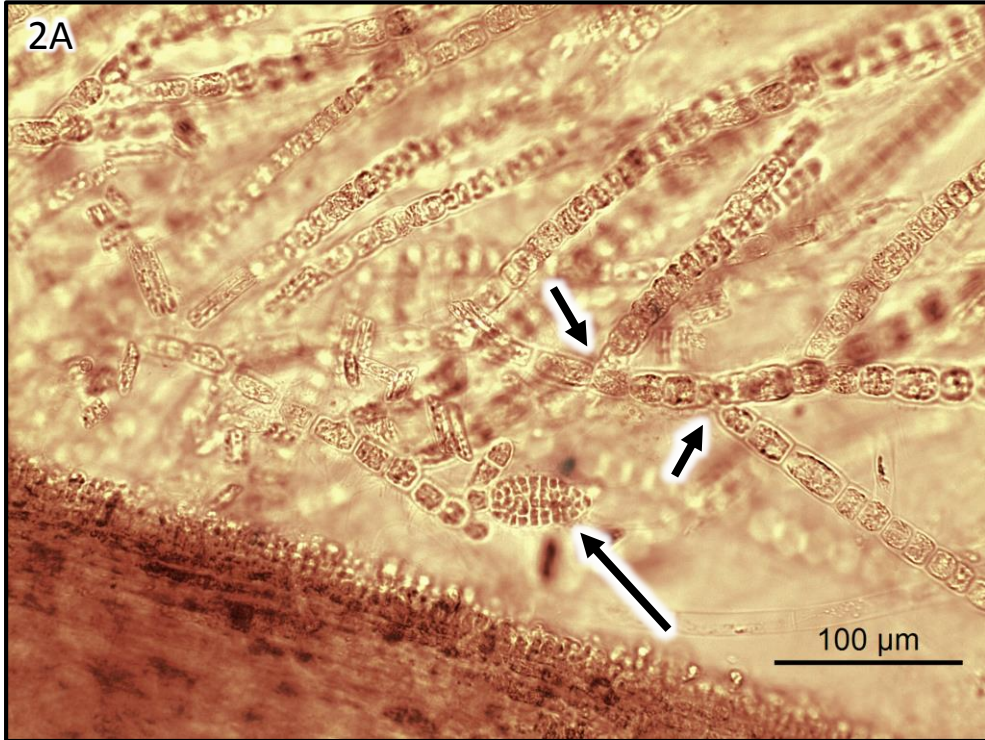
***Hincksia ovata* 1 – G (Widespread)** — Asia (R, J, K, C), Aus, SA (Arg, Ch), AK-WA, ENA, EUR, Med, IO. Epiphytic on *Scytosiphon* on 1 debris item (Apr). Cardinal indicates Mar-Sep in EUR. Fertile (plurangia and unangia). ~Ephemeral. Kim & Boo (2010) say thalli are isomorphic. Guiry (2017) indicates uniphasic direct development occurs. This sample has the spiral-alternate branching form of Kim & Lee (1992b, fig. 6c) and the *Giffordia intermedia*-form of Cardinal (1964, fig. 25), both without opposite plurangia that are often considered diagnostic for the species.



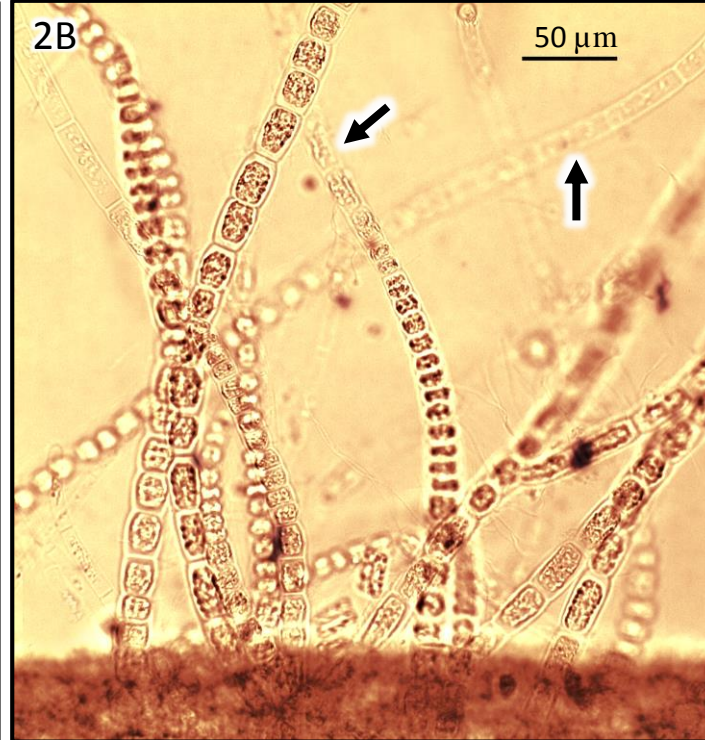
- Hemispherical tufts arising from a filamentous base with erect branches not having a clear main axis (A).
- Colonies 2-3 mm tall, 5 mm wide.
- Chloroplasts appear discoidal but are indistinct (see 2C).
- Filaments are 15-30 μm in diameter, often constricted at the cross walls, and have frequent alternate to irregular lateral branches that attenuate to rounded tips (B, 2A).
- Hairs are present in young thalli (2B).
- Growth zones occasionally occur near the base of the laterals (C).
- Unangia are ovate, 25x 50 μm, and pedicellate or sessile (D).
- Plurangia are ovate-conical, small (20-35 x 44-55 μm), and sessile or pedicellate (E, F, G).
- Cardinal (1964, fig. 24 as *G. intermedia*, fig. 26 as *G. ovata*), Cormaci *et al.* (2012, pl. 25, figs. 1-3), Kim (2010, fig. 28), Kim & Lee (1992b, fig. 6), Womersley (1987, fig. 14A-C).
- Not sequenced: SR-220 (BF-356), pressing available. DNA failed.



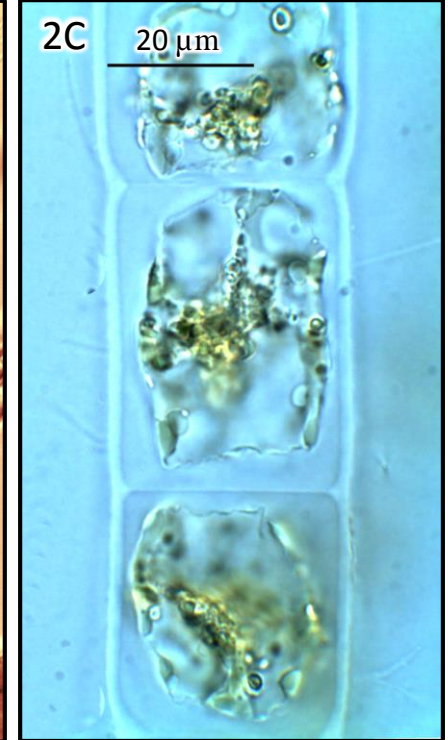
***Hincksia ovata* 2** – additional pictures. All specimens from SR-220 (Apr) sample.



Young tuft with plurangium (long arrow) and alternate branching (short arrows)



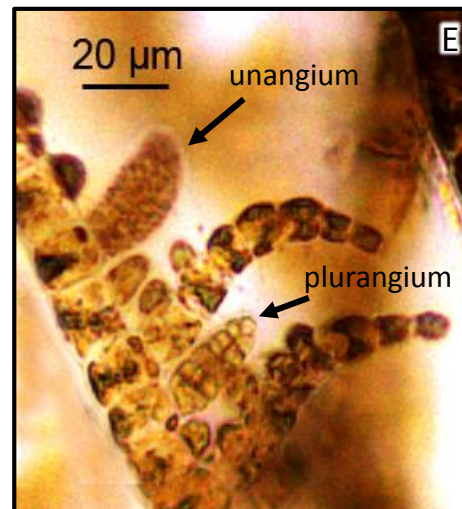
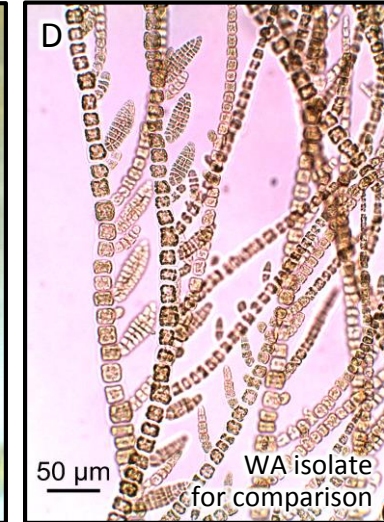
Young filaments with some bead-like cells and the production of hyaline hairs (arrows)



Plasmolyzed filament with indistinct discoidal chloroplasts



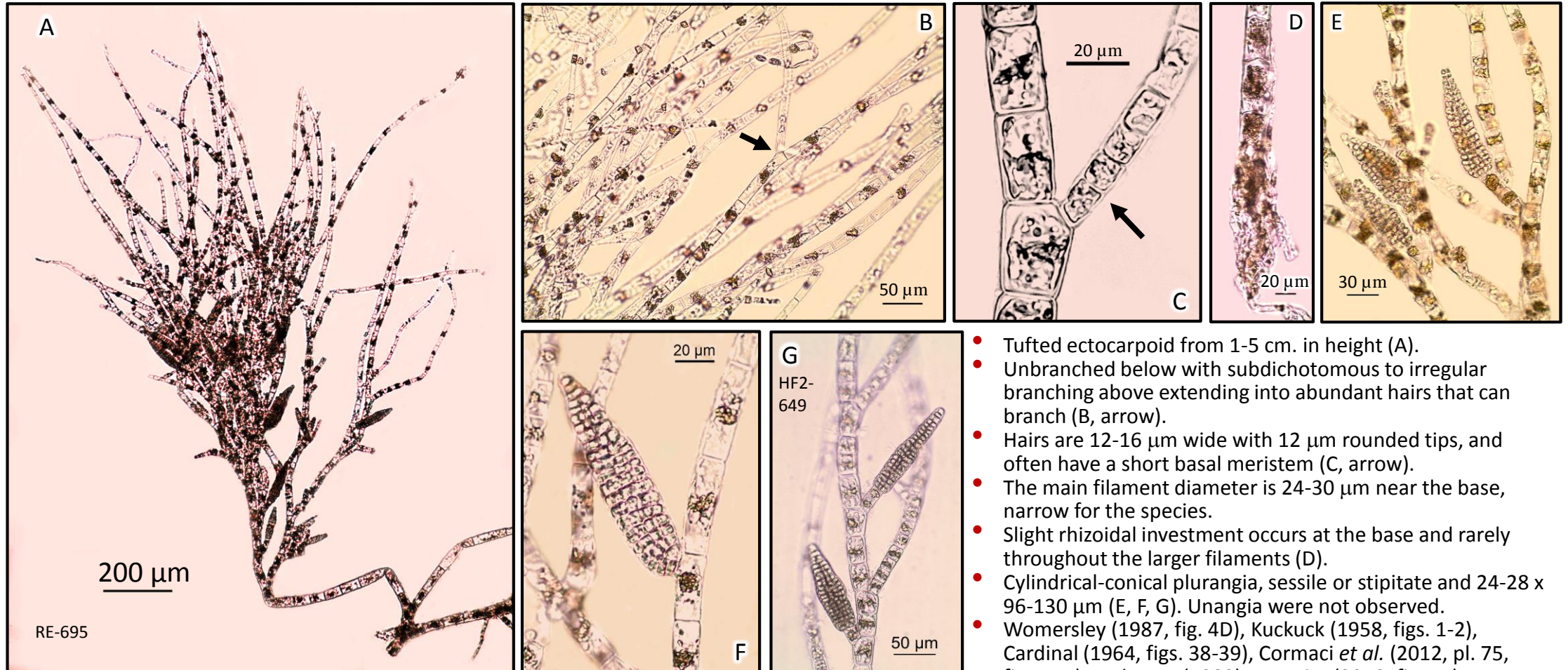
***Hincksia sandriana* – G (Globally widespread) –** Asia (J, C, K), Aus, BC-Mx, Aus, IO, SA, ENA, Afr, EUR, Med.  
 On 3 debris items (Feb, May, Jun). Mar-Dec in EUR (Cardinal, 1964). Reproductive with both plurangia and unangia. Ephemeral.



- Tufted uniseriate filaments with unilateral to irregular branching often intermixed with other filamentous brown algae.
- Lateral branches on debris thalli recurved (A), unusual for the species.
- Cells short, typically 15-25 μm in diameter, with discoidal chloroplasts.
- Up to 2 cm tall on debris but up to 10 cm tall in Korea.
- Sessile plurangia and unilateral branches often in a consecutive series (B, C). Empty plurangia were observed (C, arrow with ep).
- One ovoid sessile unangium was observed intermixed with a series of plurangia and branches (F). Note – not spherical as in *H. hincksiae*.
- Genetic studies (Kim *et al.*, 2010) have shown that *H. sandriana* is actually a complex of species.
- Kim & Lee (1992b, fig. 4), Womersley (1987, fig. 12 A-D), Yoshida (1998, pl. 2-1A), Cardinal (1964, fig. 18)
- Examples not sequenced: AB (BF-1), HF-1 (BF-28), CBB (BF-58).
- Similar species: *Hincksia hincksiae* and *Hincksia hincksiae* var. *californica*, but with shorter and cone-shaped plurangia.



***Kuckuckia cf. spinosa* cpx. 1<sup>★</sup> – G (Globally widespread) – Asia (C, K), Aus, SA (Arg, Br), ENA, EUR, Med. In turf or on *Lepas anatifera*. Fertile (plurangia). Isomorphic. On 9 debris items (Jan-May) but variable morphology. Sequenced from 3 items. This form, with abundant hyaline hairs, was on 2 debris items. ★ RE-695-Ect4 (BF-533). Not sequenced (G): HF2-649 (BF-462). Similar DNA to: *E. arctus*. ~ Ephemeral.**

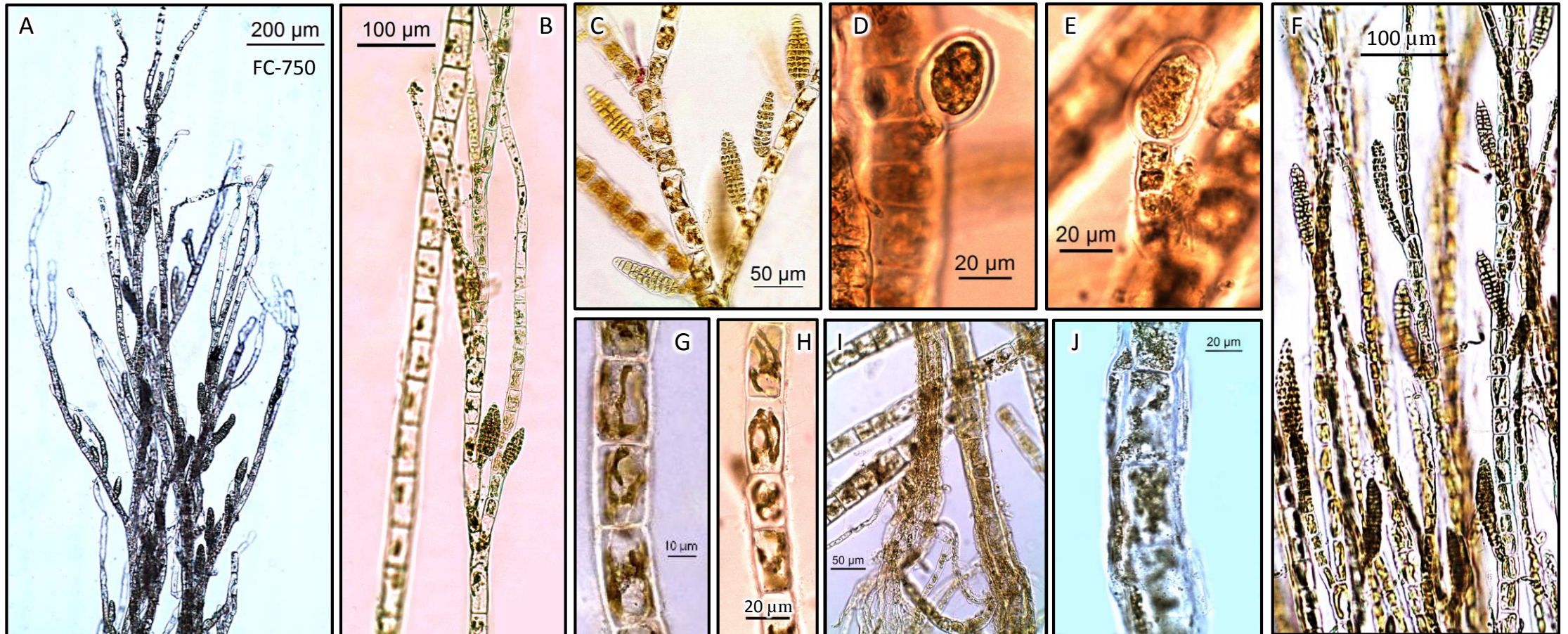


- Tufted ectocarpoid from 1-5 cm. in height (A).
- Unbranched below with subdichotomous to irregular branching above extending into abundant hairs that can branch (B, arrow).
- Hairs are 12-16 µm wide with 12 µm rounded tips, and often have a short basal meristem (C, arrow).
- The main filament diameter is 24-30 µm near the base, narrow for the species.
- Slight rhizoidal investment occurs at the base and rarely throughout the larger filaments (D).
- Cylindrical-conical plurangia, sessile or stipitate and 24-28 x 96-130 µm (E, F, G). Unangia were not observed.
- Womersley (1987, fig. 4D), Kuckuck (1958, figs. 1-2), Cardinal (1964, figs. 38-39), Cormaci *et al.* (2012, pl. 75, figs. 1-4), Pedersen (1989), non Kim (2010, fig. 11).



***Kuckuckia cf. spinosa* cpx. 2<sup>★</sup>** – A sequence variant occurring in Hanyuda Clade E-sp. 3b. Sequenced from 2 debris items (4 samples) (Jan, Jul). Fertile (plurangia and unangia). This form has the typical characteristics of *Kuckuckia*, but with less extensive hairs than in Form 1 and a growth zone that is rarely present. ★FC-750, 752 and 753 (BF-652), LBF-135 (BF 462), often on *Lepas anatifera*.

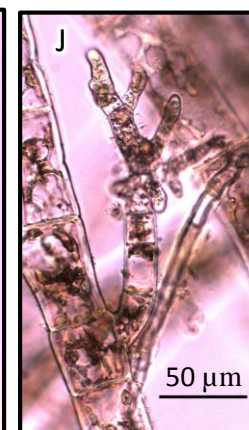
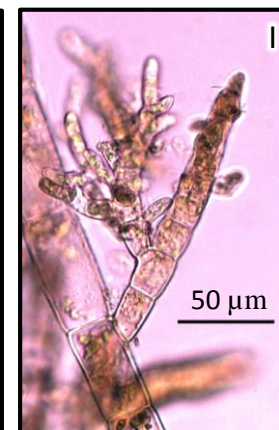
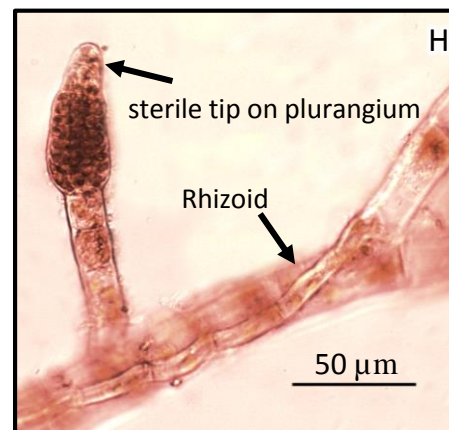
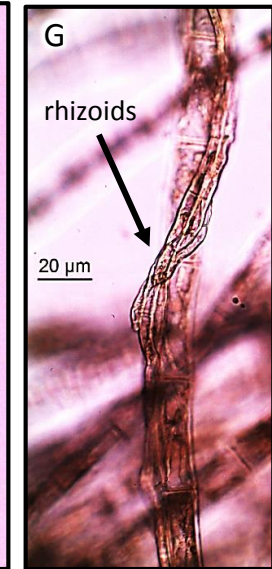
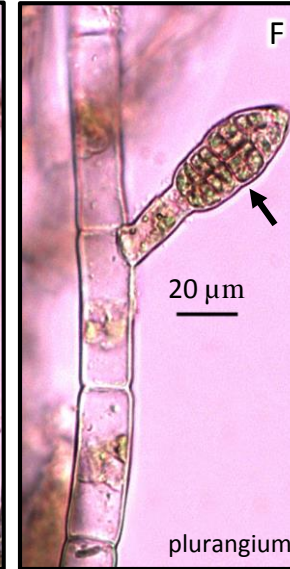
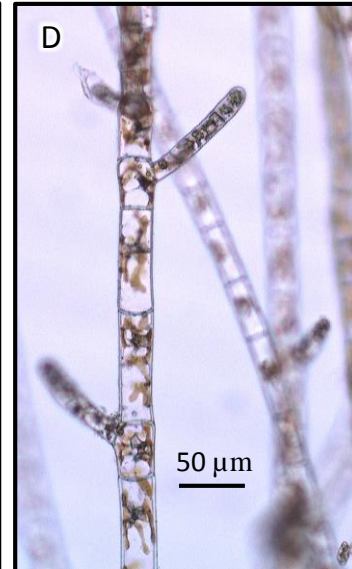
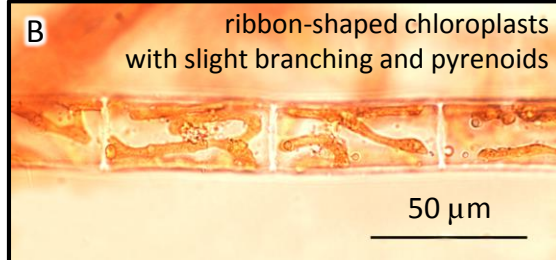
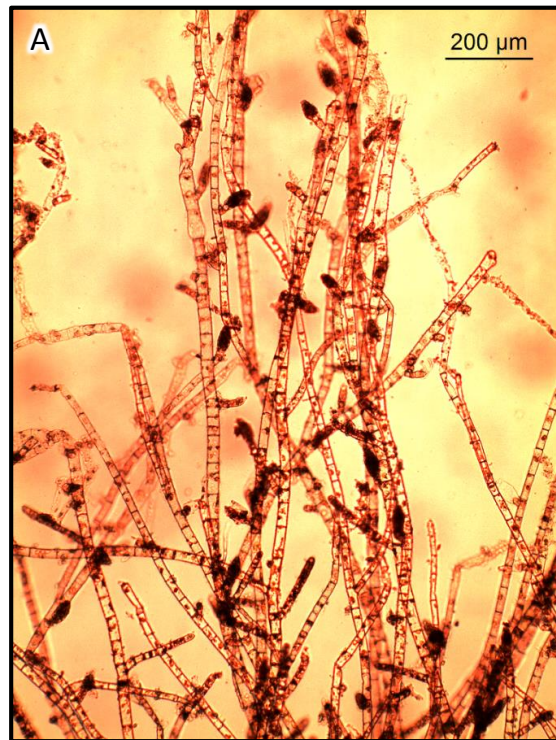
Branched filaments 24-30  $\mu\text{m}$  wide with cells having a 1:1 to 1:2 dimension (A, B); hairs were abundant and 12-15  $\mu\text{m}$  wide at filament tips but with a limited or absent basal meristem; a rhizoidal base and holdfast were present (I, J), sessile and pedicellate regenerative plurangia 20x80 (160  $\mu\text{m}$ )  $\mu\text{m}$  (C, F); sessile and pedicellate unangia = 40x26  $\mu\text{m}$  (D, E). Cells contained a few longitudinal to spiral ribbon-shaped chloroplasts (G, H), similar to true *Kuckuckia*.





***Kuckuckia* cf. *spinosa* cpx. 3<sup>★</sup>** – Sequences indicate *Kuckuckia*. <sup>★</sup>SixR-719 (BF-538) (Apr). Not sequenced: SixR-725.

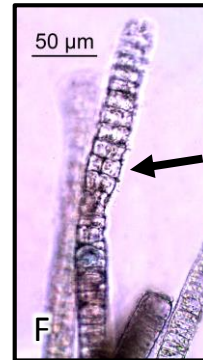
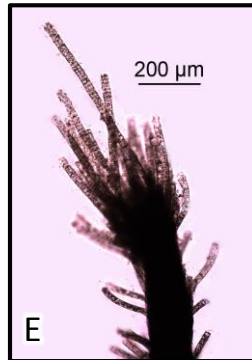
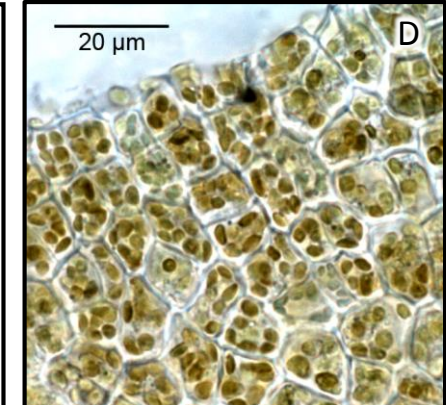
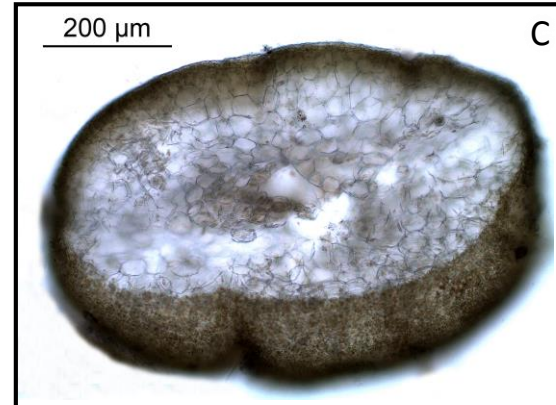
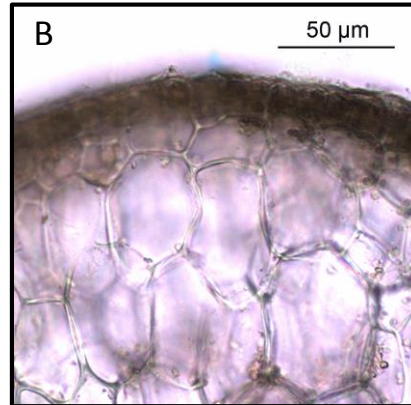
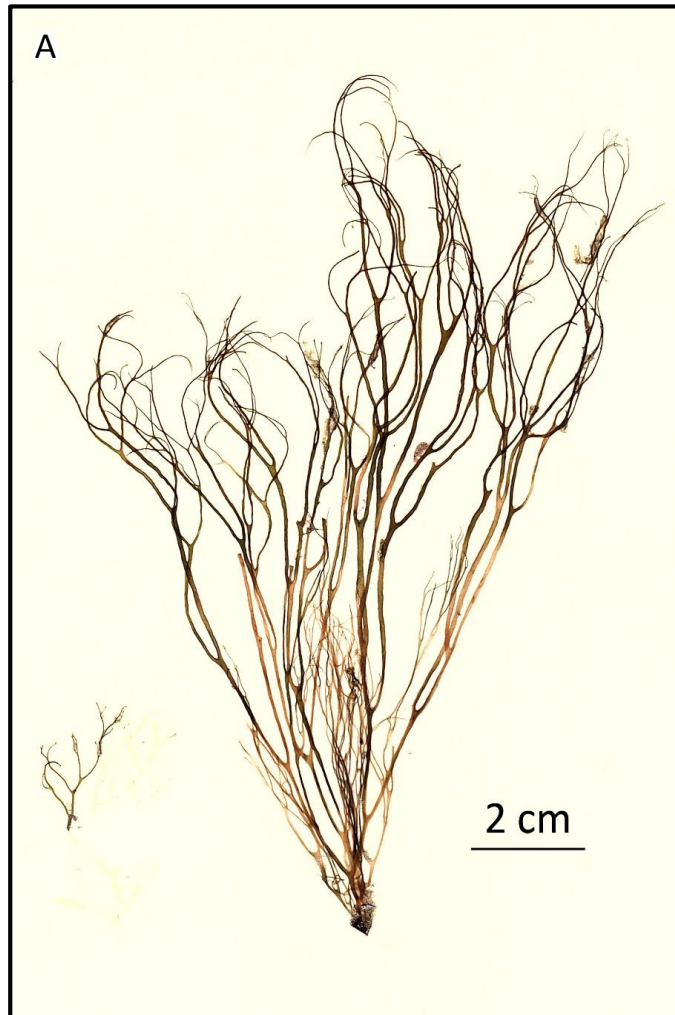
These samples have narrow filaments (like *arctus*), crampon-like laterals and small plurangia. Both also have unusual short highly branched laterals that may be the result of a virus infection (#D. Mueller). Drawings of *E. spinosa* by Kützing (1855, pl. 49) show hairs, small plurangia and also crampon-like laterals (but with more pointed tips).



- Uniseriate branched thalli (A).
- Ribbon-shaped chloroplasts (B-C).
- Filament diameter, 18-24 μm (B-C).
- Short lateral branches (C, D) similar to crampons in *Acinetospora*.
- Small plurangia, ~20-25 x 50-60 μm (E, F, H). Sterile tips may occur (arrow).
- Rhizoids that lightly envelop the axes, 10 μm in diameter (G, H).
- Rare highly branched laterals (I, J), unknown in either *arctus* or *spinosa*.
- Abundant hairs as occur in *Kuckuckia spinosa* were not observed.



***Mutimo cylindricus***★ – A+ – Asia (J, K, Phil) + CA-MX. On 1 debris item (Apr): Sixes River derelict boat. CA (1973) and MX (1990) introductions are a different haplotype than the debris species. Sterile. Annual and heteromorphic.



- An erect gametophyte alternates with an encrusting *Aglaozonia*-like sporophyte lacking rhizoids.
- Erect terete (solid to hollow), dichotomously branched thalli (A) are parenchymatous in structure (B, C) and reach 8-15 cm in height.
- Cortical cells have multiple discoidal chloroplasts without pyrenoids (D).
- Trichothallic filaments occur at the tips and sides of the branches (E).
- Dioecious: Male and female plurangia form in warty filamentous sori on the surface of the erect gametophytes (not seen).
- Introductions into California and Mexico are reported in Miller *et al.* (2011) but were shown to be a different haplotype from the debris species by Hanyuda *et al.* (2017).

- *Mutimo* was recently separated from *Cutleria* due primarily to genetics. Morphologically, it also differs due to the cylindrical shape of the branches and the ability of the trichothallic filaments to become multiseriate. Note the initial formation of the multiseriate state (F, arrow).
- Okamura (1900-1902, pl. XXVIII), Kogishi *et al.* (2010), Kawai *et al.* (2012).
- ★ SixR-716 (BF-538).



***Petalonia fascia* cpx. ★ – G (Globally widespread) –** Asia (R, J, C, K), Aus-NZ, SA (Ch), AK-MX, ENA, Afr, EUR, Med. On 24 debris items (Jan-May). Fertile (plurangia). Annual & heteromorphic (with a *Stragularia crustose* sporophyte). Variable both genetically and morphologically. ★ 4 of 8 *cox3* haplotypes known in Japan were found on debris (none of these occur in the NEP) – 3 types are illustrated.

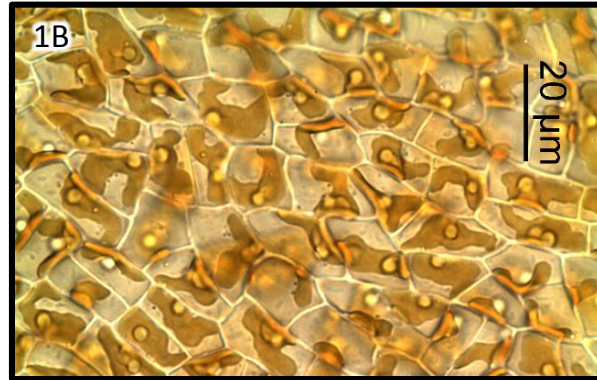
**Type 1 = H-6**

Sterile surface cells with single chloroplast and pyrenoid

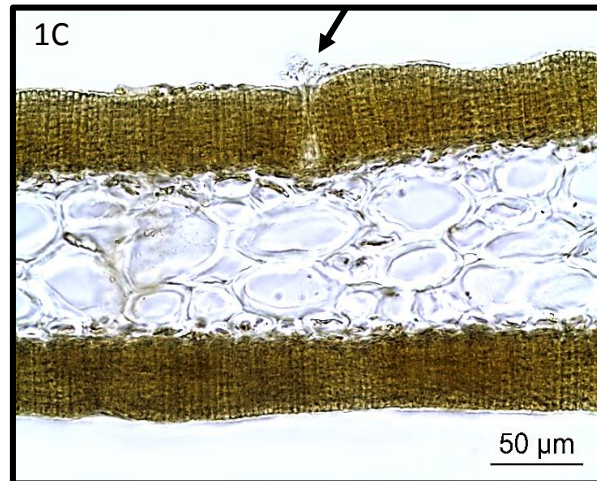


1A

Sterile thalli from Ponsler debris boat, MP-12 (BF-36, Hanyuda H-6)



1B



1C

Cross-section of fertile blade. Note surface of plurangia and clustered paraphyses (arrow)



1D

Fertile. Note banding. Seal Rock boat – SR-215 (BF-356, Hanyuda H-6)

- Type 1, described here, is dominant on debris.
- Lanceolate blades with cuneate bases arising from a small discoidal holdfast (1A, 1D).
- Most thalli have acute or pointed apices (1A).
- Blades are up to 40 cm tall, 1-3 cm wide and 100–250 μm thick.
- Surface cells are angular and contain a single chloroplast with a single pyrenoid (1B).
- Subcortical cells are also angular (see 1E, p. 31).
- Fertile gametophytes have banded sori (1D) containing densely packed uniseriate plurangia that may be up to 15 cells long (1C).
- Sori may contain clustered paraphyses. (1C, arrow).
- Fletcher (1987, figs. 61A, 62), Kogame (1997), Okamura (1900-1902, pl. X).
- ★ 17 samples sequenced – examples: MP-12 (BF-36, H-6), SR-215 (BF 356, H-6), SR-205 (BF-356, H-8), HF2-630 (BF-526, H-5), RE-662 (BF-533, H-2), SixR-727 (BF-538, H-2). Haplotypes (H-xx) are from Hanyuda *et al.* (2017).
- Japanese (including debris) and NEP haplotypes differ by numerous base pairs in the *cox3* gene, and seem to be separate species (Hanyuda *et al.*, 2017).
- Morphological variation appears to coincide with some of the haplotypes – see Types 2 and 3 (p. 31).

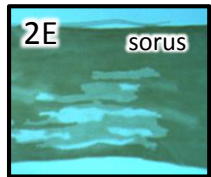


2 cm

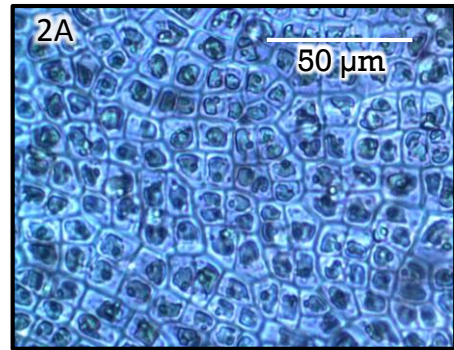


**Type 3.**  
3A. Seal Rock debris boat (BF-356)

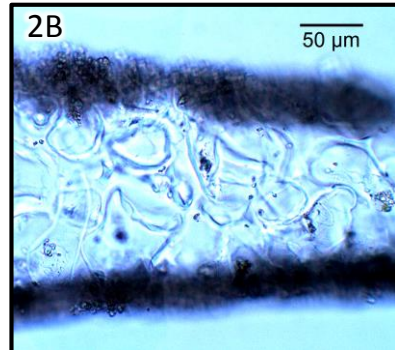
***Petalonia fascia* cpx. — Types 2★ and 3★ —** These morphological types are typically larger and have less acute tips than in Type 1 thalli (pg. 30, 1A and 1D). Differences also occur in the sori and subcortical layers. **Further study is required.**



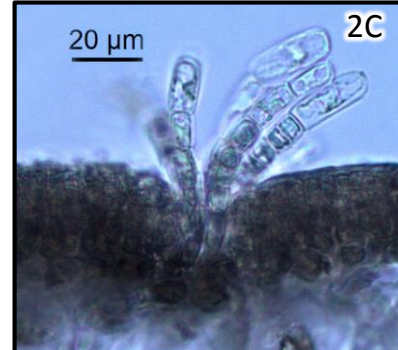
**Type 2.** ★RE-662 (BF-533, Hanyuda H-2): Blades without acute apices (not shown); blade anatomy is below:



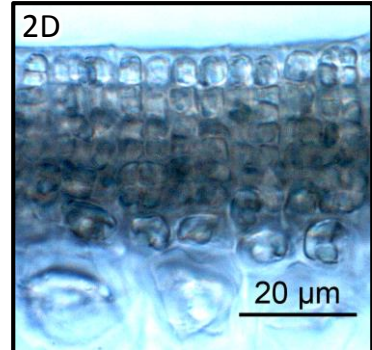
Square to angular surface cells.



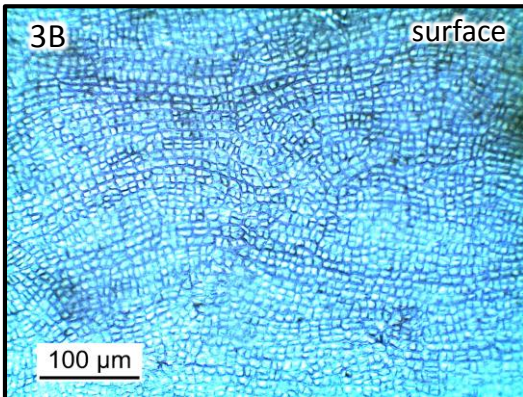
Irregular to hyphal-shaped medullary cells.



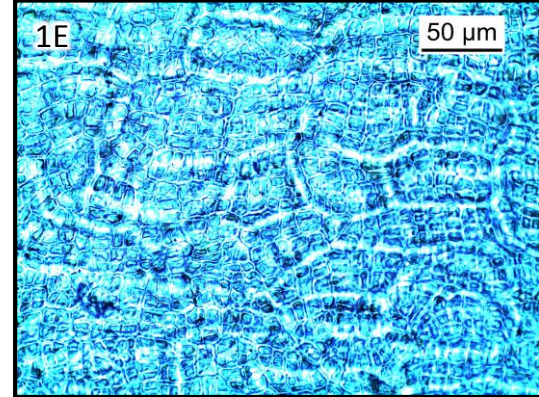
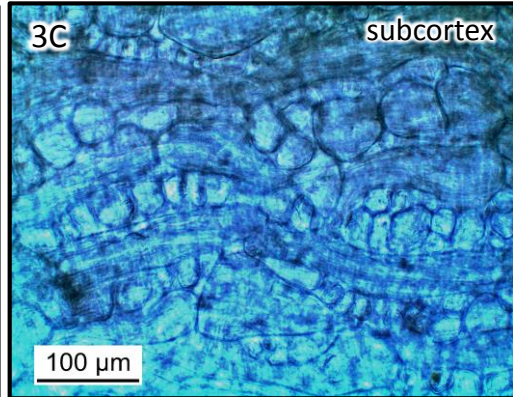
Unusually large tip cells on some surface hair clusters.



Plurangia – spore release pattern is in 2E above.



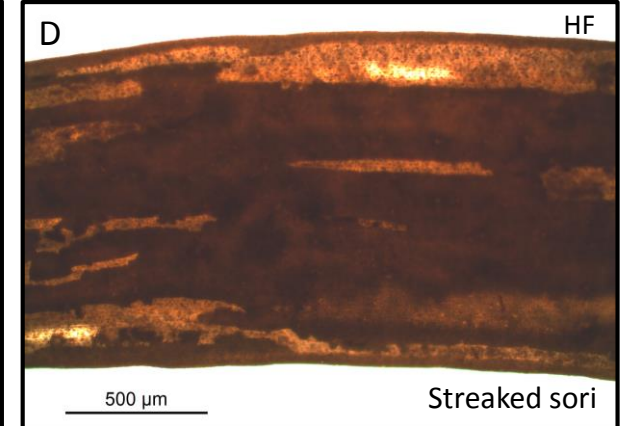
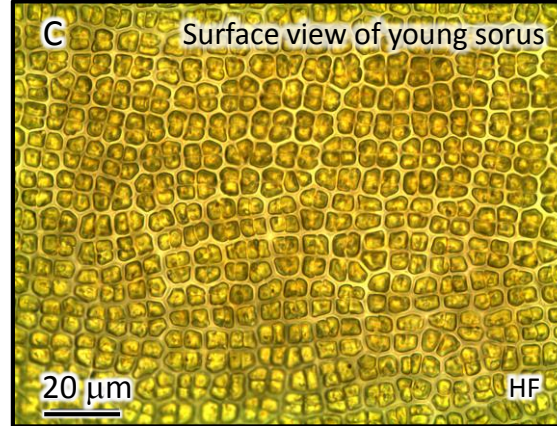
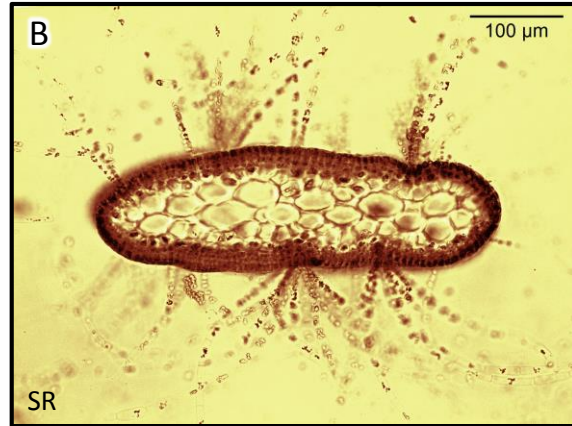
**Type 3.** ★SR-205 (BF-356, Hanyuda H-8) and the large specimen at left (3A). Mature thalli are larger than other *Petalonia* and have less pointed and often slightly curved apices (3A). Surface view (3B) and optical section of subcortex (3C) revealing the undulating pattern of the cell filaments and the cell size variation.



**Type 1.** ★MP-12 (BF-36, H-6, pg. 30). Optical section of the subcortex of Type 1 thalli showing angular cells.



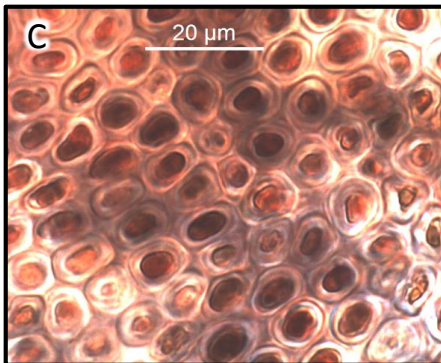
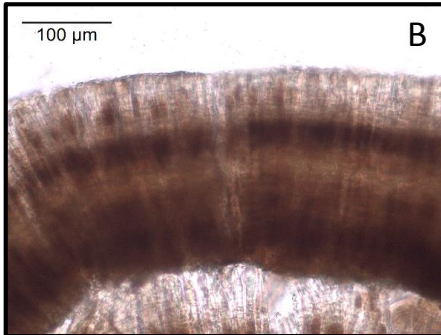
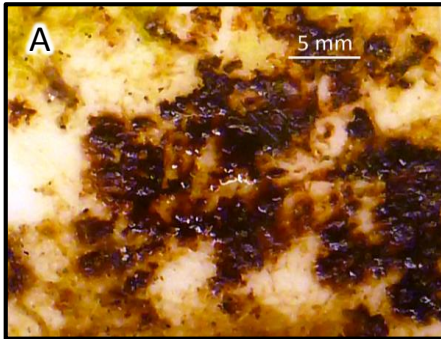
***Petalonia zosterifolia***★ – **G (Widespread)** — Asia (R, J, C, K), Arc, ENA, EUR, Med. On 12 debris items (Jan-May).  
 Reproductive. Annual and heteromorphic with direct development from a microscopic filamentous *Compsomena saxicolum* sporophyte.



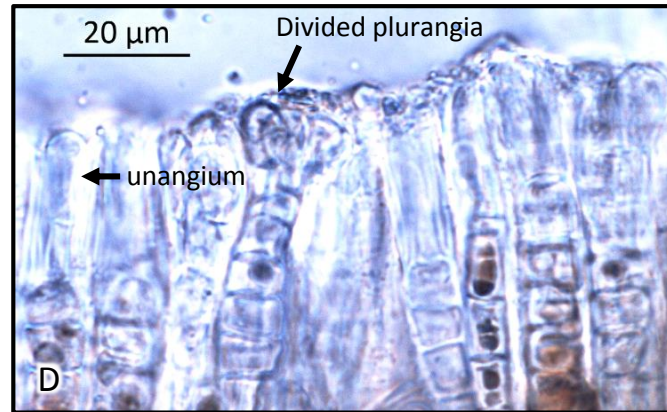
- Erect blades reach 10-20 cm in height and are flattened, 0.5-2 (8) mm in diameter, 60-120 μm thick – and are usually solid (A, B).
- The medulla contains broad elongate filaments, subglobose in cross section.
- A narrow cortex of 1-3 subglobose cells is present and, in surface view, the cells appear to be in longitudinal rows (C).
- Chloroplasts as for genus -- single and parietal with a single pyrenoid (E, chloroplasts shown in a juvenile terete phase).
- Basal holdfast containing rhizoidal filaments (F).
- Streaked sori develop on mature blades (C, D) and contain tightly packed erect uniseriate plurangia, 14-40 μm in diameter, each containing from 6-12 locules (not shown).
- In our material, hair clusters frequently occurred on sterile and fertile thalli (B), although these are reported as rare in Japan.
- Cho *et al.* (2002), Fletcher (1987), Kogame & Kawai (1993).
- ★ Examples sequenced: HF1-8 (BF-28); CBB-7 (BF-58), LC-2 (BF-173), SR-216 (BF-356), LBD-367 (BF-397), RE-701 (BF-533).



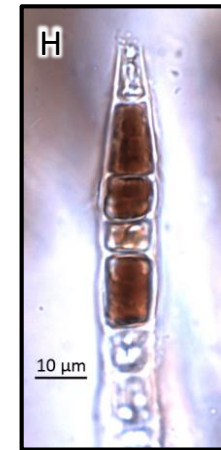
***Petroderma maculiforme*★ – G (Globally widespread) — Asia (R, J), Arc, A-Arc, SA (Argentina, AK (Hansen), WA, MX, ENA, EUR. On the Quail Street JTMD plastic carboy (Mar). Disintegrating with eroded unangia and plurangia. Perennial and isomorphic.**



- Congested thick 1 cm patches on plastic; soft to scrape off (A-C).
- Single-celled basal layer with no visible rhizoids (E).
- Erect filaments are 8-10 μm broad with rare branching.
- Single parietal plate-like chloroplasts were not visible.
- Eroded terminal unangia & divided plurangia were visible (D).



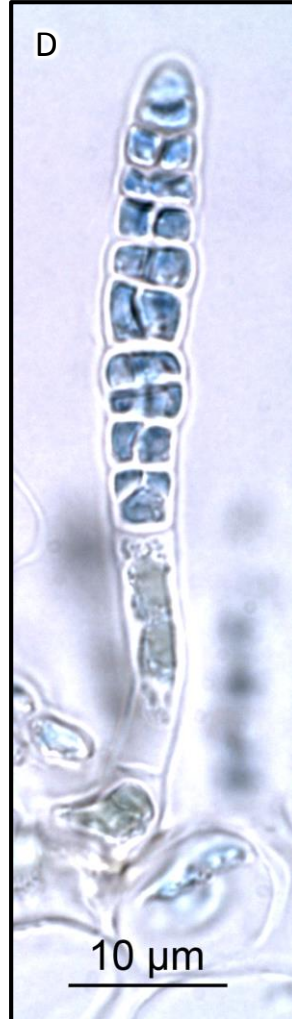
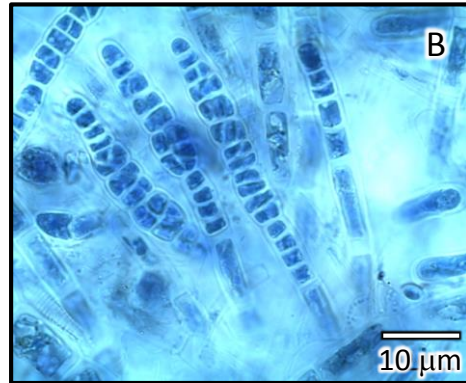
- Unangia tips were 10-12x30 μm, but rarely 10x50 μm.
- Eroded cells often contained dark-colored storage material (F-H).
- Wilce, *et al.* (1970, figs. 1 & 2), Waern (1949, pl. 1).
- ★ Quail Street carboy – QS-620 (BF-656).



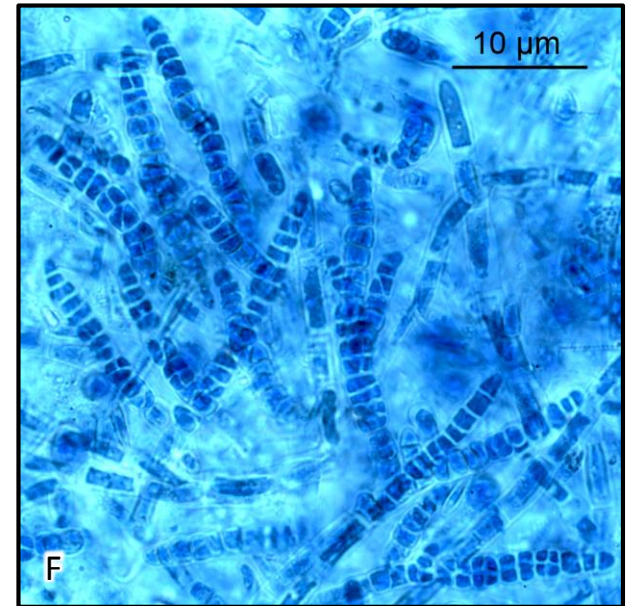
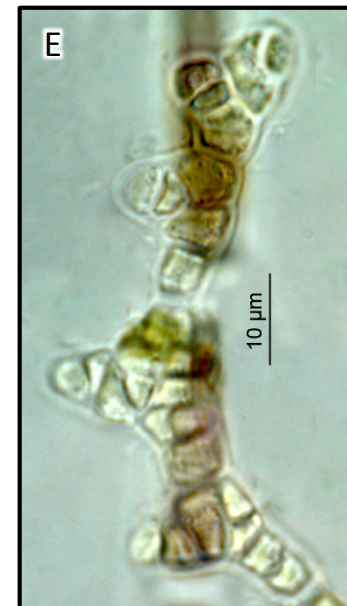


# *Protectocarpus speciosus* – G (Globally widespread) — Asia (J, K), IO, ENA, SA, EUR, Med, Canaries.

Epiphytic in the turf on *Lepas anatifera* and also on some algae. On 5 debris items (Feb-Jun). Fertile. ~Ephemeral and isomorphic.

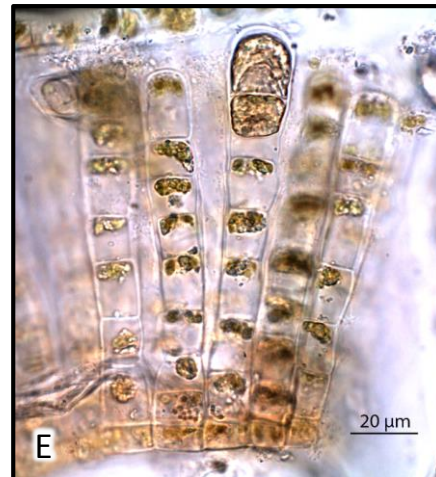
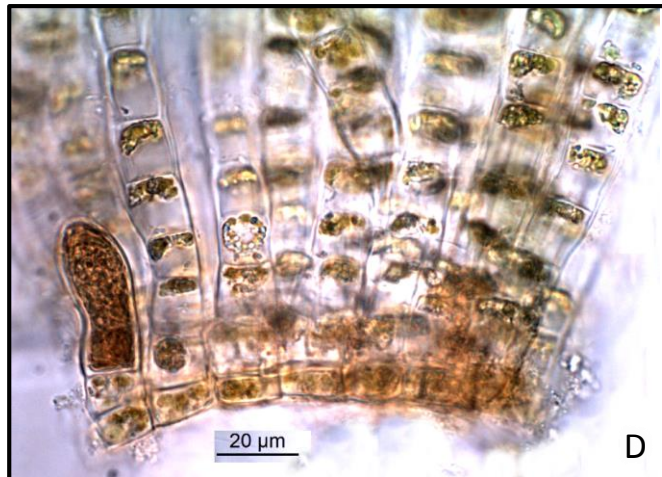
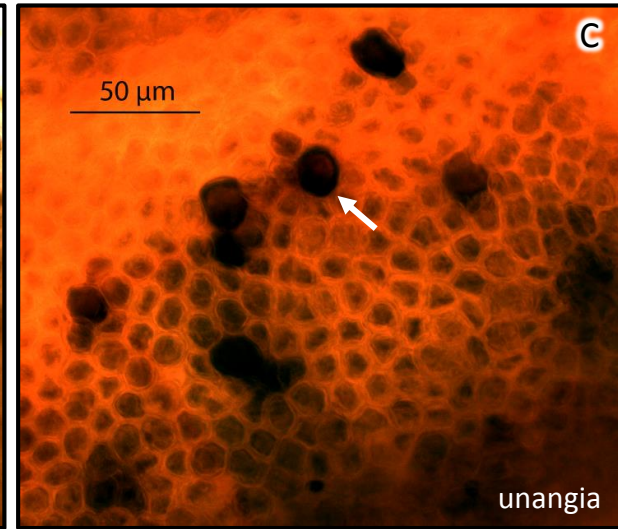
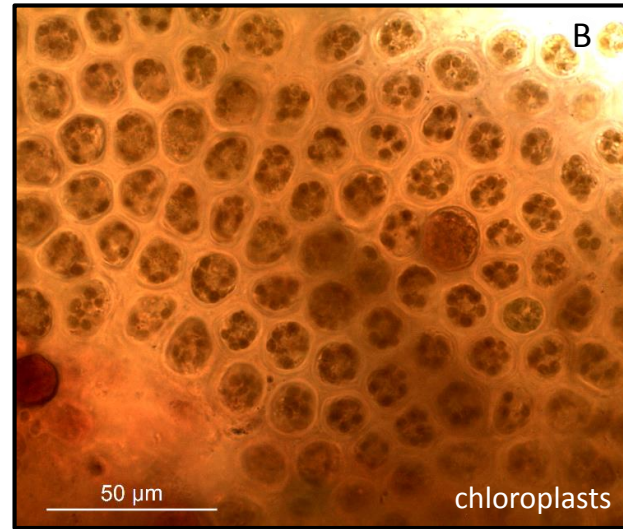
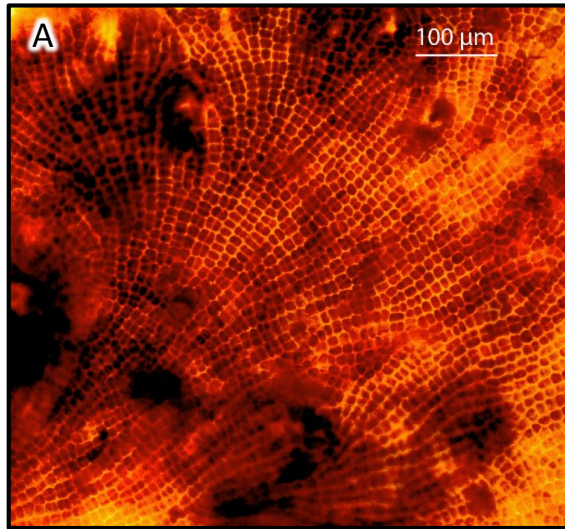


- A branched filamentous and monostromatic basal layer that forms upright, occasionally branched uniseriate filaments 5-10 μm in diameter and only 1-3 mm in height (A).
- Plurangia are uni to multiseriate on upright filaments -- when branching occurs the sporangia are continuous between the uprights and branches (B-F). A cockscomb pattern was occasionally observed (not shown). Unangia were not observed.
- Chloroplasts were not well-preserved in the debris material.
- Kuckuck (1958, p. 66/126, pl. 4), Tanaka (1986), Fletcher (2987, figs. 15-16), Kornmann & Sahling (1977, pl. 158).
- Not sequenced: AB (B-1), HF (BF-28), MP (BF-36), LC (BF-173), SVB (BF-402).



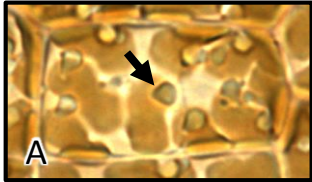


***Pseudolithoderma cf. paradoxum* – G** (rare global occurrences) — Asia (J? and debris), ENA (Massachusetts). On 1 debris item (Apr) – the fiberglass from the Sixes River Boat – SixR-730 (BF-538). Reproductive (unangia). Perennial and isomorphic.

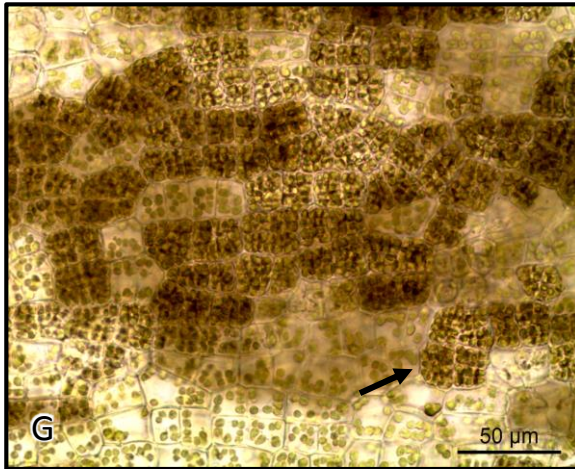
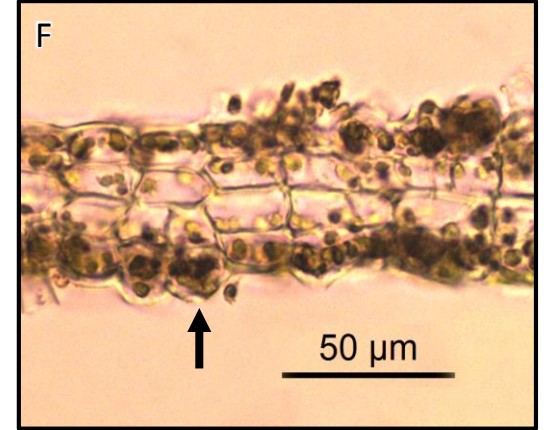
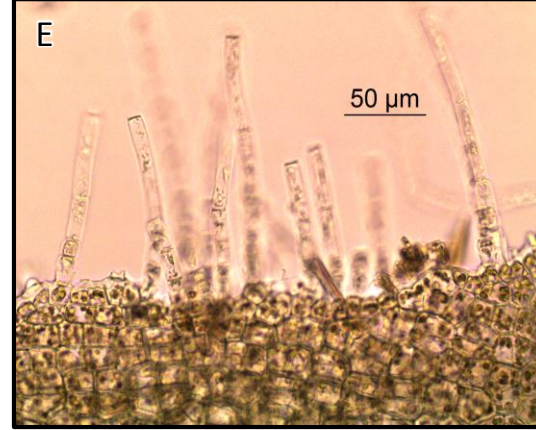
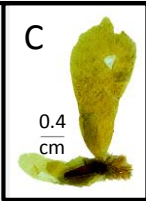


- A thin patchy crust on fiberglass with no visible thick gelatinous cuticle.
- Surface cells appeared as swirled radiating rows and contained no visible hairs (A).
- Cells had 6-9 discoidal plastids without pyrenoids (B).
- The basal layer was single-celled, ~ 8 μm in height, and bore tightly appressed upright unbranched filaments of uniform diameter (14-16 μm) that were most often of similar length, making the cells appear cuboidal (D, E).
- Unangia were scattered across the surface in sori (C), developed terminally on the upright filaments, and were 15-18 μm in diameter and cylindrical (D, E).
- *P. subextensum*, also reported from Japan, has cylindrical unangia and filament diameters similar to this species. However, it is also has a thick cuticle, hairs, and 3-4 chloroplasts/cell. Although hairs in many species are known to change with the environment, chloroplast number would not. So, I use *P. paradoxum* Sears et Wilce and suspect that this species has been overlooked in Japan.
- *P. paradoxum*: Sears & Wilce (1973). *P. subextensum*: Waern (1949, pl. III), Tanaka & Chihara (1981), Yoshida (1998, pl. 2-2, figs. F and G).





***Punctaria latifolia* cpx. 1<sup>★</sup> – G (Globally widespread) –** Asia (R, J, C, K), Aus, NZ, SA, AK, ENA, Afr, EUR, Med. A common epiphyte on debris on 14 JTMD items (Feb-Jun, Oct). Fertile with only plurangia. Annual. Heteromorphic with direct-development from a microthallus.

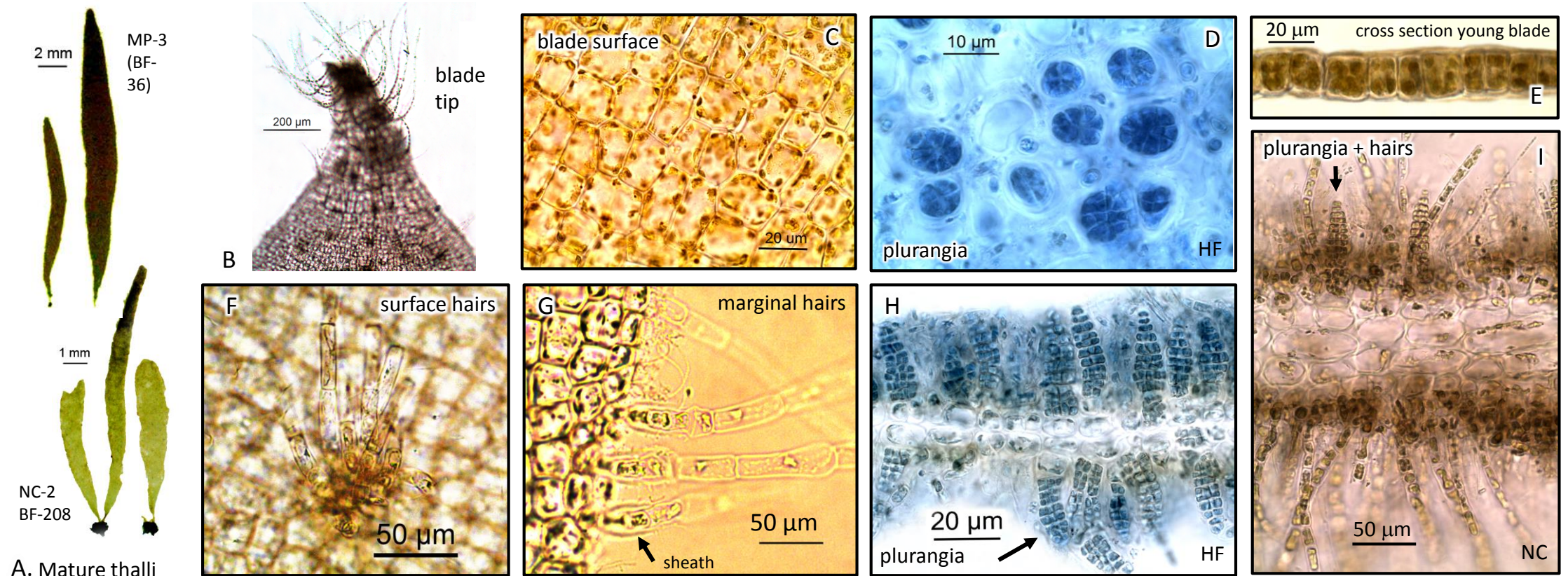


- Thalli were ovate to obovate and often split on debris (C).
- Blades on debris were small compared to other areas – up to only 3 cm tall. Shown here epiphytic on *Scytosiphon lomentaria* (B, arrow).
- Surface cells quadrate to rectangular with multiple discoidal plastids (D) and stalked pyrenoids (A, arrow).
- Hairs common on the margins and occasional on the surface (E).
- Thalli on debris were 4-6 cells thick (F).
- Plurangia (arrows) are ovate to conical in cross section (F) – and form in extensive patches on the blade surface (G).
- Fletcher (1987, figs. 52A, 53), Womersley (1987, figs. 114 A, 115 C-D), Okamura (1916).
- ★ Examples sequenced: SR-218 (BF-356), Bev-110 (BF-288), RE-703 (BF-533). Not sequenced SR-245 (BF-356).



***Punctaria latifolia* cpx. 2 – the *flaccida* form** ★ – Asian only (R, J, K). On 4 of the 14 JTMD *latifolia* items. Merged with *P. latifolia* for this study. Fertile with elongate plurangia extending outward 3-4 x the size of the outer cortical cells, similar to illustrations of true *P. flaccida* by Noda (1979, pl. 3, fig. 4 #2-3) and Nagai (1940, pl. II, figs. 12-13; pl. III, fig. 7). Annual – direct development from a microthallus.

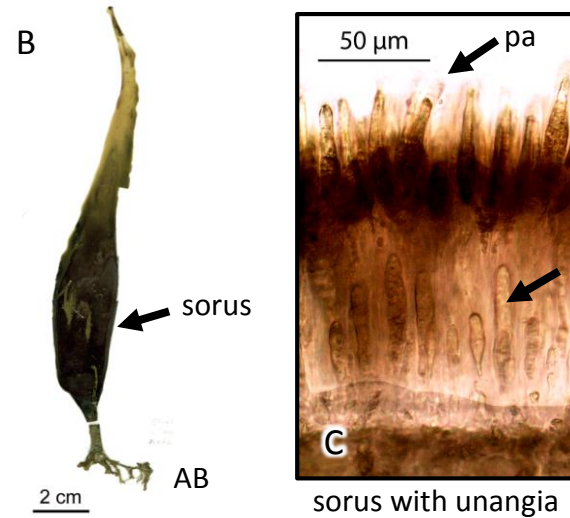
Thalli were lanceolate to oblanceolate often with a 1.5:10 ratio of width to length and reaching 5 cm (A). Blades are 1-4 (6) cells thick (E, I) with cortical cells slightly smaller than medullary cells. Sheathed hairs form in clusters on surface and solo along the margins (B, F, G). Our material was only plurangial and bore tall cylindroconical plurangia (D, H, I). DNA sequences could not differentiate this small form from *P. latifolia*. This similarity also occurs between *P. tenuissima* (not known in Japan) and *P. latifolia* (Parente *et al.*, 2010). Both smaller forms have protrudent plurangia. ★ Sequenced: MP-3 (BF-36), NC-2 (BF-208). Not sequenced: SJC (BF-39) and HF1 (BF-28).



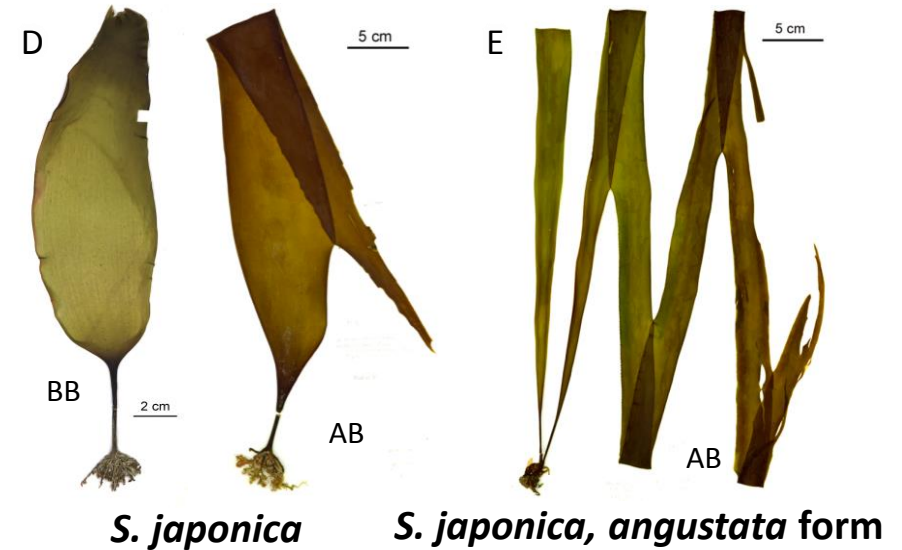
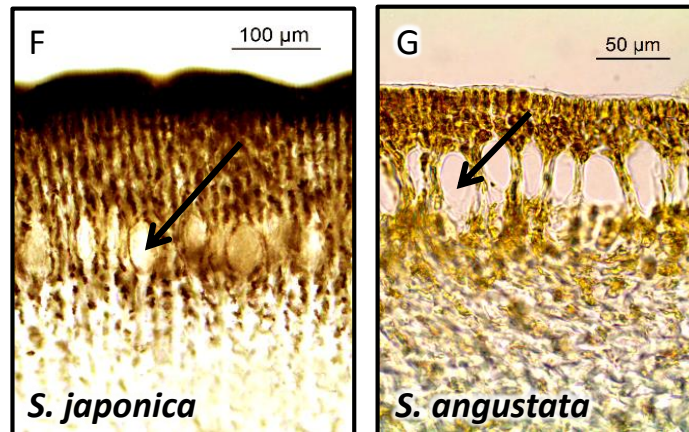


***Saccharina japonica*★/*angustata*★ – A+ – Asia (R, J, C, K) + France (*S. japonica*). “Kombu”, edible kelp.**

On 3 debris items (Mar, Jun), including the Agate Beach dock. Fertile. Biennial and heteromorphic. ★ 2 nearly identical haplotypes.



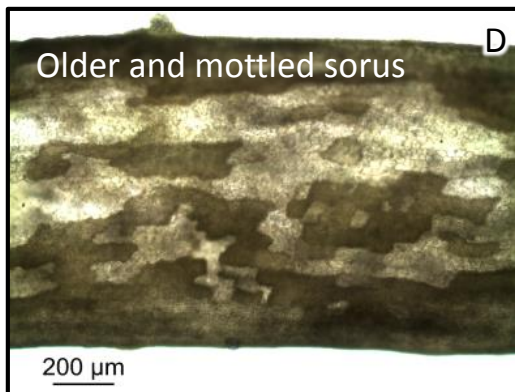
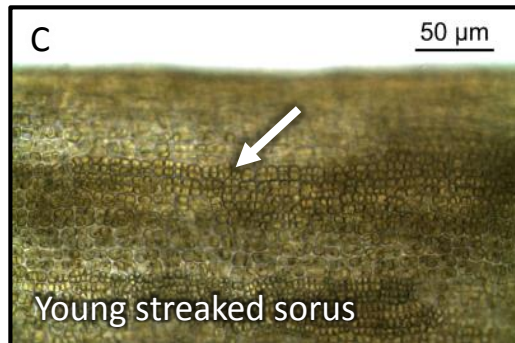
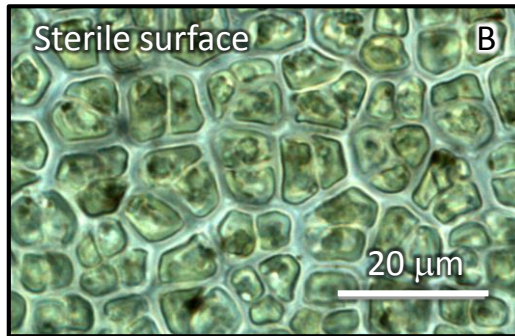
Stipe cross sections – arrows are mucilage ducts



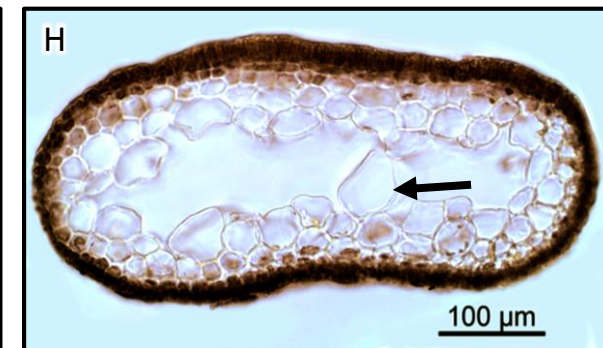
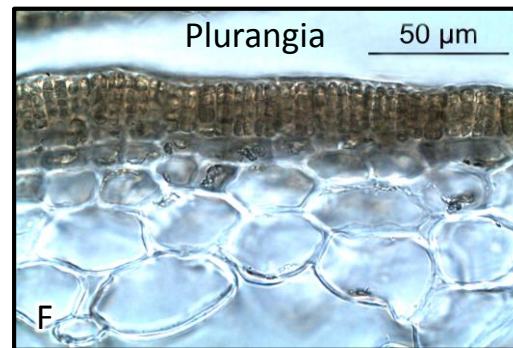
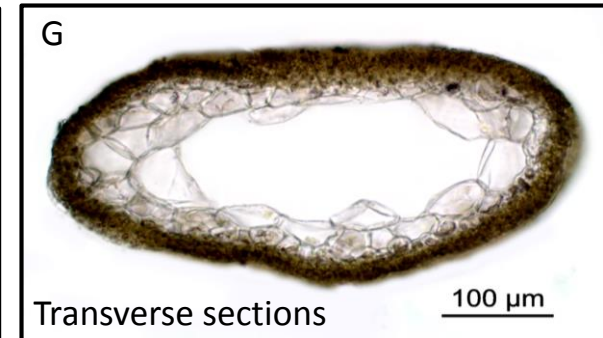
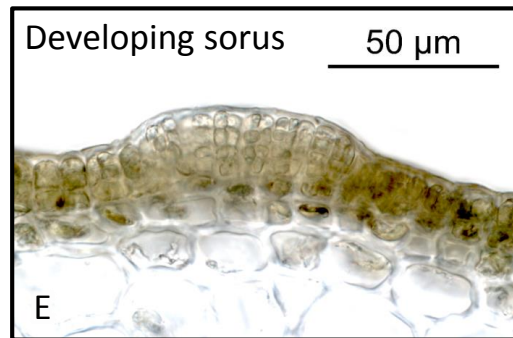
- Both are 1-2(4) m long flat to bullate simple blades with short round stipes (A, B, D, E).
- Reproductive sori form on the lower part of the blade (B, arrow).
- A section of the sorus (C) reveals plurangia (arrow) developing intermixed with typical protective paraphyses (arrow, pa).
- Morphologically, the 2 species differ in:
  - Blade and fascia width (D, E).
  - Mucilage duct location in the stipe (F, G, cross sections).
- Molecularly, the 2 are considered genetic variants of the same species.
- Introduced in 1976 into Thau Lagoon, France, the species has not spread or become invasive.
- Miyabe (1902, pls. I and IX), Okamura, (1925, pl. CCXXI), Okamura & Uyeda (1925), Nagai (1940), Yotsukura *et al.* (2006, 2010); Verlaque *et al.* (2015).
- ★ AB-6, 7, 8, 9 (all BF-1), BB-1 (BF-2). Not sequenced: Fish-1 (BF-40).



***Scytosiphon gracilis*\*** – A+ – Asia (J, K) + MX, Chile. On 10 debris items (Jan-May).  
 Similar to Oregon's *Scytosiphon dotyi* in lacking ascocysts. Fertile (plurangia). Annual and heteromorphic.

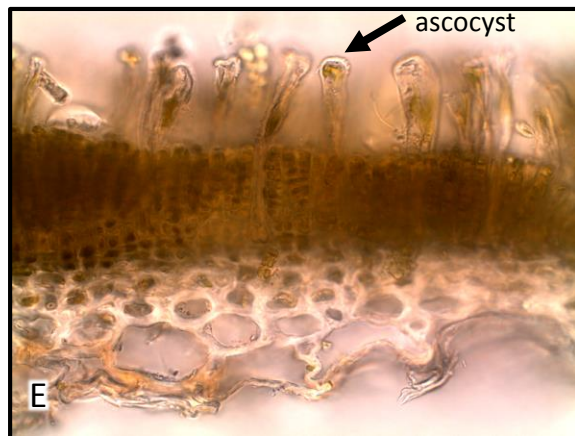
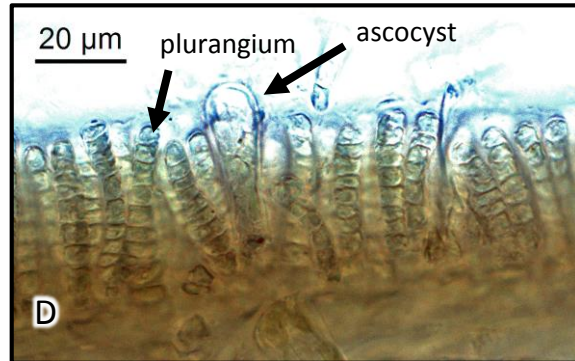
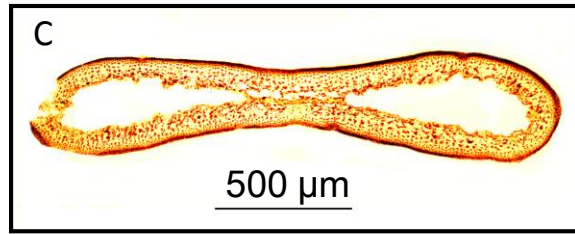


- Clustered, oval to compressed unbranched and unconstricted tubes (A)
- To 17 cm long x 0.4 – 4.0 mm wide.
- Hollow or occasionally with struts (G, H, arrow).
- The cortex is smooth with no ascocysts (B).
- The medulla under each cortex consists of 2-3 (4) large subglobose cells (G, H).
- Fertile tubes develop initially streaked sori consisting of vertical rows of coherent uniseriate plurangia, about 6-12 cells in length (C, E, F) .
- Older fertile thalli are mottled in appearance after gamete release (D).
- Kogami (1998) describes a microscopic *Compsonema*-like sporophyte.
- \* M-5 (BF-8). Not sequenced: GB (BF-23), SBT (BF-234), MP (BF-36), Bev (BF-288).

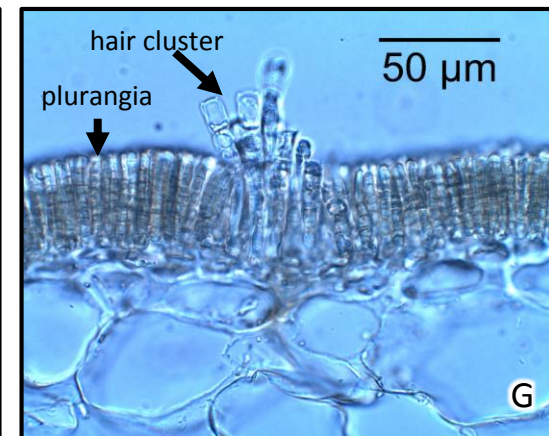
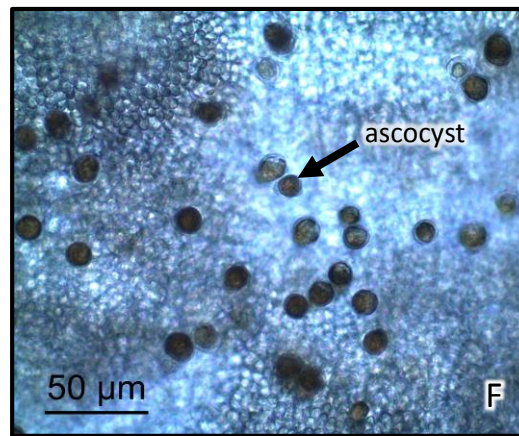




***Scytosiphon lomentaria* cpx. 1<sup>★</sup> – G (Globally widespread) –** Asia (R, J, C, K, Viet), IO, Aus, NZ, SA, AK-MX, ENA, EUR, Med, Afr. On 12 debris items (Jan-Jun). Reproductive (plurangia). Annual and heteromorphic.



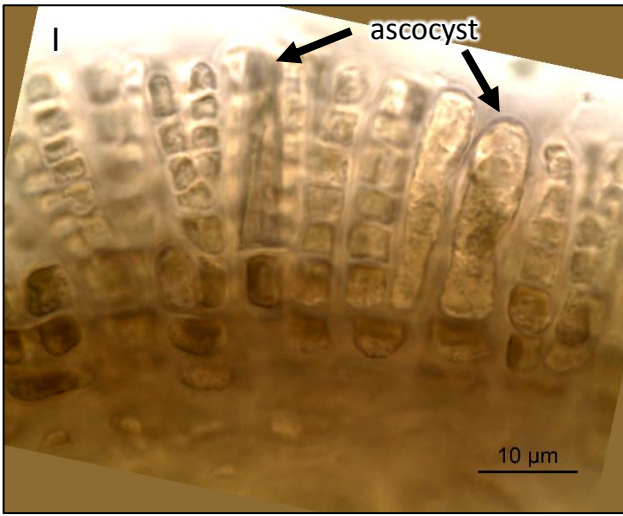
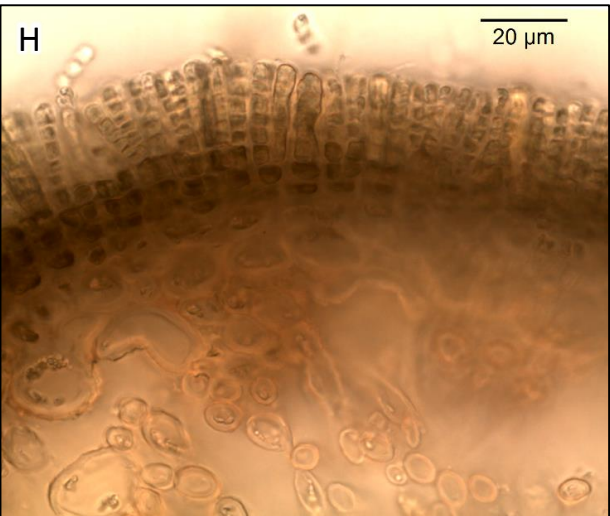
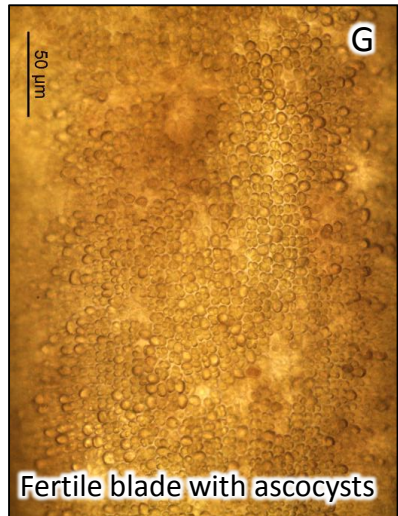
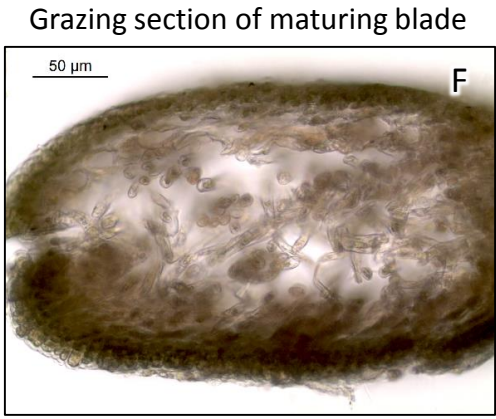
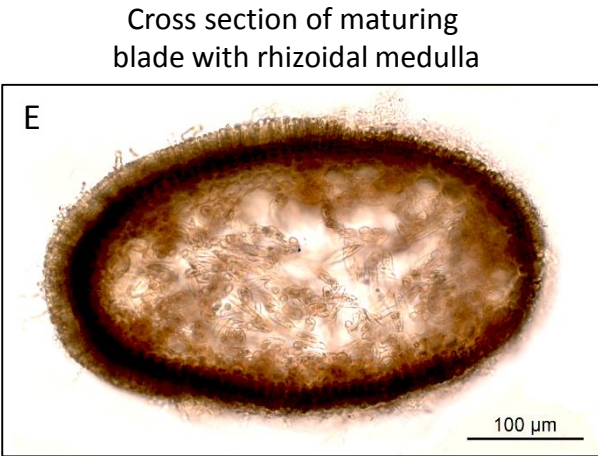
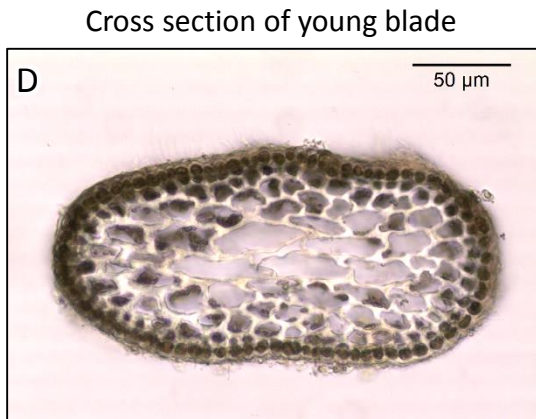
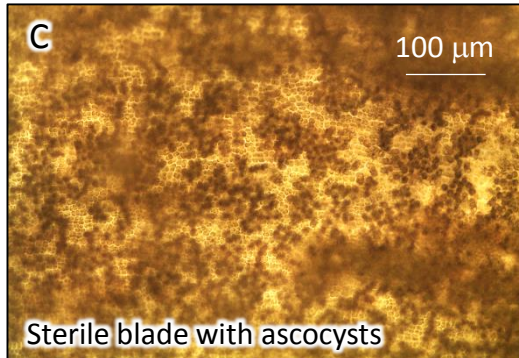
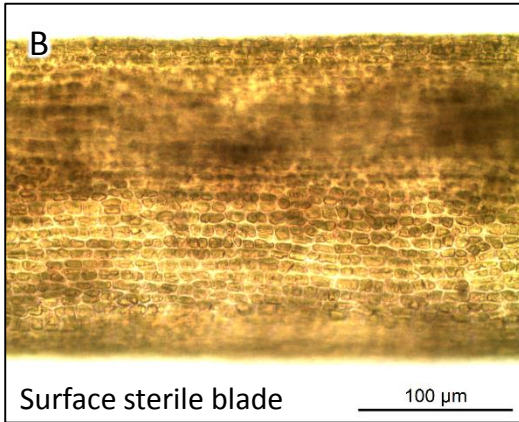
- Clustered tubular unbranched thalli to 30 or more cm tall (A).
- Each hollow tube is up to 5 mm in diameter and is often crimped periodically like a chain of sausages (A, B, C).
- The erect tubes narrow toward their tips and may be slightly flattened.
- Uniseriate plurangia coat the surface of fertile thalli dotted by ascocysts (arrows) (D, F) and occasional hair clusters (G).
- Ascocysts (arrows), generally submersed in the plurangia, are erumpent both before the plurangia fully develop and after they are released (E).
- Kogame (1998) describes a *Microsporangium*-like sporophyte.
- Tatewaki (1966), Clayton (1978), Fletcher (1987, figs. 67b, 69); Kogame (1998, figs. 52-65).
- Sequence variants are known on both Pacific coasts and in Europe (Cho *et al.*, 2007). 2 variants on JTMD are illustrated (see the following slide).
- ★ Examples sequenced: SR-212 (BF-356), SVB-382 (BF-401), LBF-409 (BF-462), RE-663 (BF-533), (Clade B of Hanyuda *et al.*, 2017).







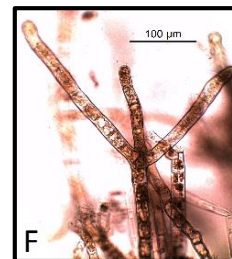
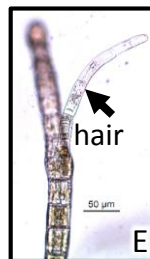
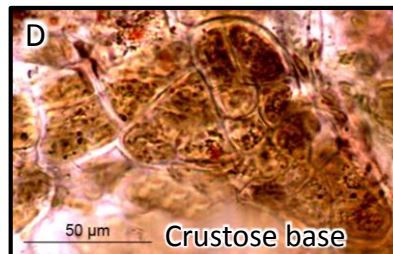
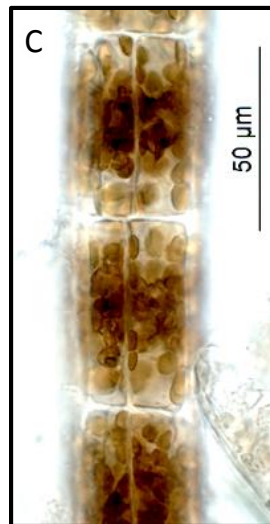
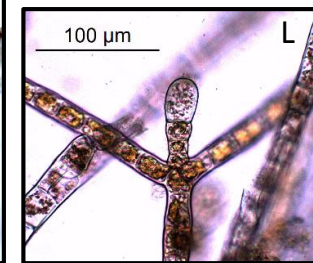
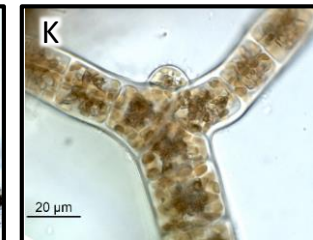
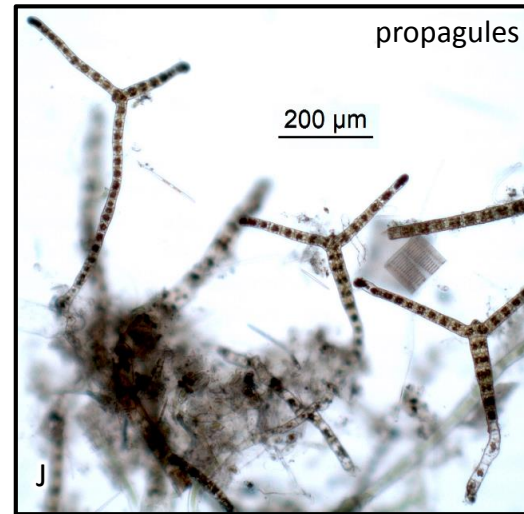
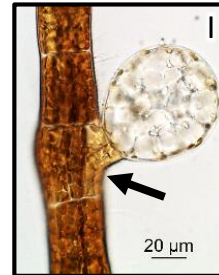
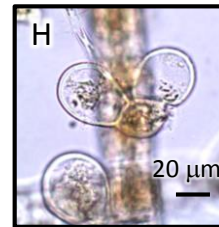
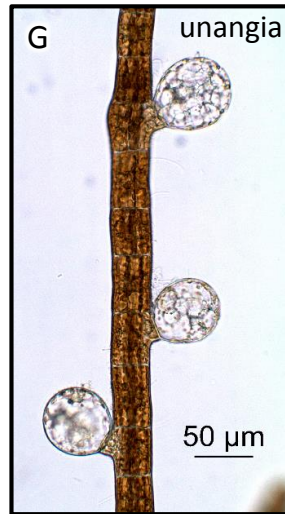
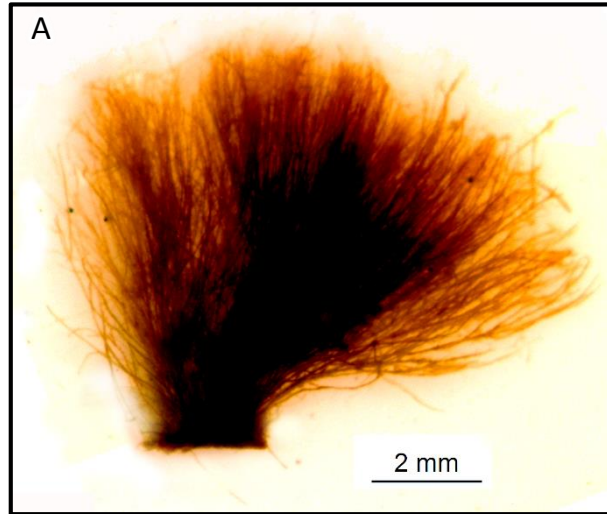
***Scytosiphon lomentaria cpx. 2\**** – Unusual small form from North Cove JTMD (May). Note rhizoidal medulla and ascocysts. Blades were terete but flatten slightly at tips.  
 ★ NC-1 (BF-208) -- DNA sequence variant in clade A of Hanyuda *et al.* (2017).



Sections of fertile blades with ascocysts and unbranched and branched uniseriate plurangia



***Sphacelaria rigidula* \*# – G (Globally widespread) –** Asia (R, J, C, K, Viet, Phil), IO, Aus-NZ, SA, AK-MX, ENA, Car, Afr., EUR, Med. On 8 debris items (Mar-May). Reproductive (propagules and unangia). Perennial and isomorphic.

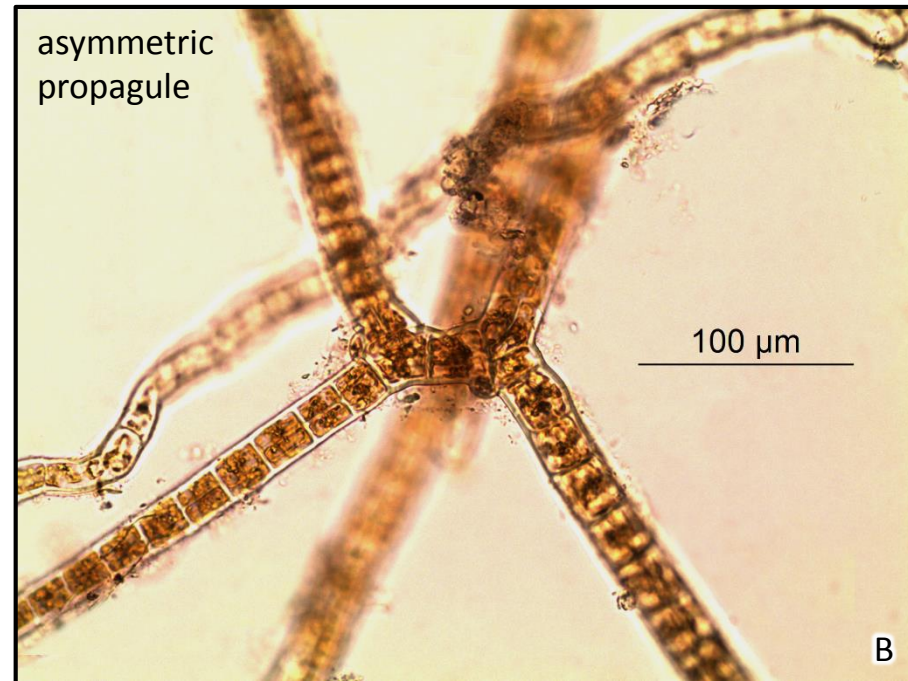
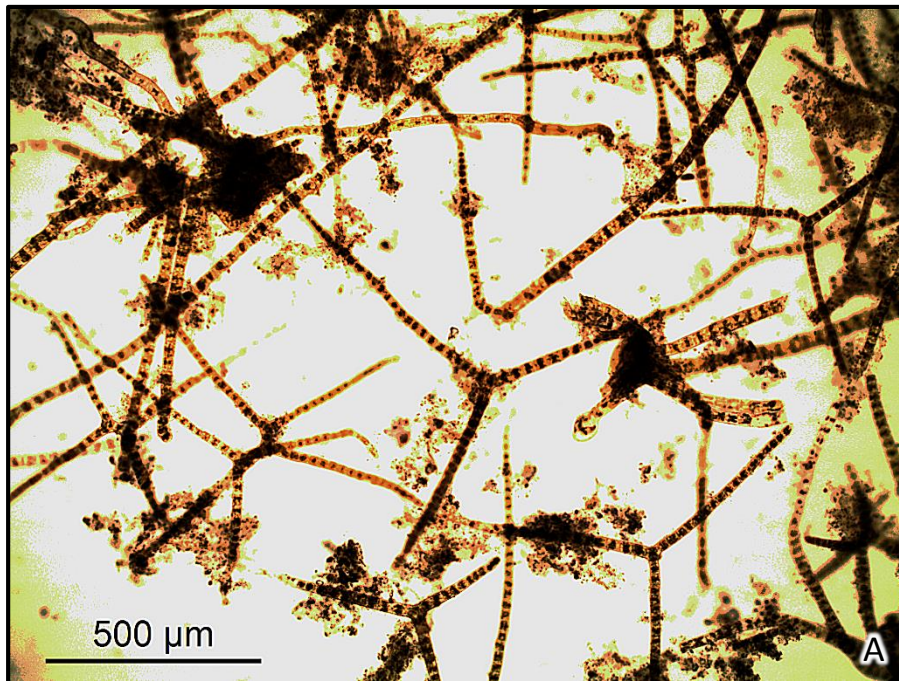


- Extensive turf with an encrusting base on hard substrata (rope, Styrofoam, boat surfaces) – about 1 cm tall (A, D).
- Typical large apical cell, 2-4 vertical cells in view on the filaments, rare hairs are produced (arrow) (B, C, E).
- Propagules bifurcate without narrowing of arms or base (C, K).
- Propagule apical cell visible (K) and may germinate – occasionally with unangium production (F, L).
- Unangia spherical, sessile or stipitate – 52 μm diameter; occasionally 2/stalk cell. Also note split (arrow) in stalk cell of unangium above (G, H, I).
- Macro-plurangia were not observed.
- Kitayama (1994, figs. 22-29), Driasma *et al.* (1998, 2010), Prud'homme van Reine (1982, figs. 508-553), Keum, *et al.* (2001b).
- # Experts consulted: Driasma, Prud'homme van Reine, Kitayama, and Kawai.
- \* Examples sequenced: Wal-3 (B-196), NC-3 (BF-208), RE-679 (BF-533).



***Sphacelaria solitaria* – G (Widespread) –** Asia (J, K, C, Viet), NZ, EUR. Only on the Mosquito Creek dock (Jan), MC (BF-8). Abundant propagules. Perennial and isomorphic.

- Irregularly branched filaments up to 1.8 cm in height and 35-60  $\mu\text{m}$  in diameter.
- Each tier of the filaments bear 2-5 longitudinal walls in surface view.
- Plants were found in the turf on the dock and were epilithic or epiphytic.
- The asymmetric bi-furcate propagules that characterize this species were present.
- Kitayama (1994, figs. 40-45), Keum *et al.* (2001a, figs. 1-17), Kylin (1947, non fig. 24D).

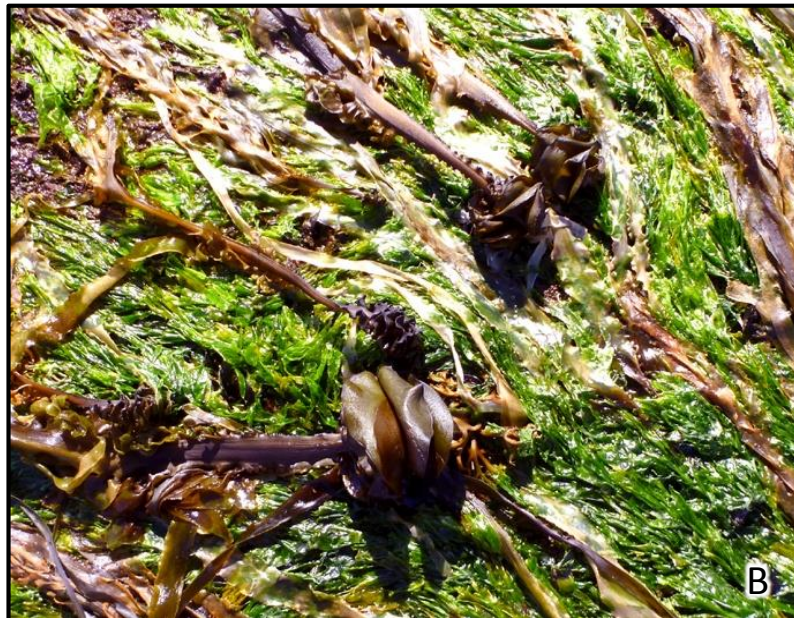




***Undaria pinnatifida***★ – A+ – Asia (R, C, J, K), Aus, NZ, SA (Argentina), CA-MX, EUR, Med. Only on the Agate Beach Dock (Jun). Abundant and fertile on debris. Annual and heteromorphic with a microscopic gametophyte.

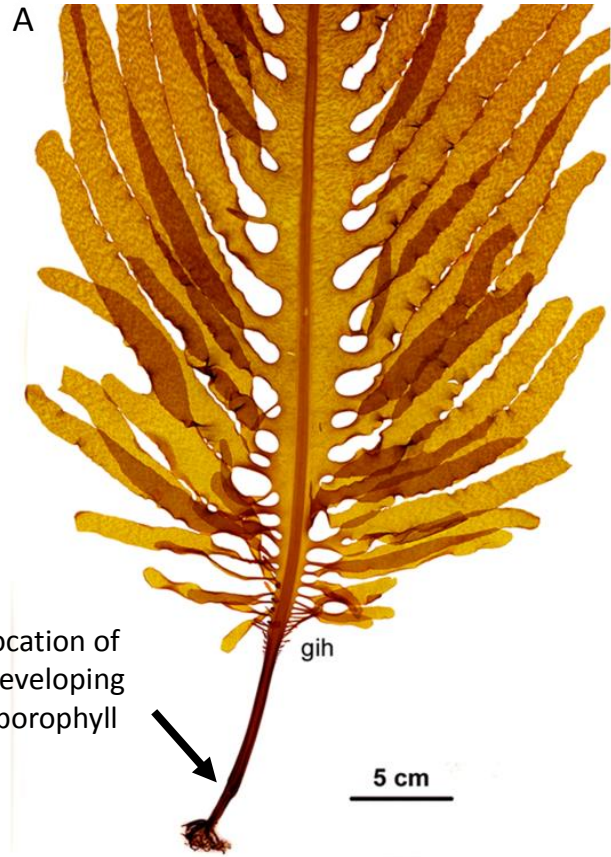


- “Wakame”, a food staple in Japan. Wild harvested and cultivated in Asia.
- Sporophytic blades on debris were highly lobed with a central midrib and reached 1+ m tall (A).
- Sporophylls form as a ruffled ring at the base of the stipe but are only fertile when the ruffle expands and becomes mature (B, C). See part 2.
- Thalli can regenerate and reproduce up to 3x/year as documented for NZ.
- Highly invasive listings: “**100 world’s worst invasive alien species**”, “**100 worst invaders in the Mediterranean**”, and 3rd among the “**most invasive algal species in Europe**”.
- First recorded in CA in 2000 (the CA-MX haplotype differs from the haplotype on JTMD).
- Okamura (1915, 1926, pl. CCXXVI), Saito (1972), Uwai *et al.* (2006).
- ★ **Sequenced:** AB-5790a (BF-1).



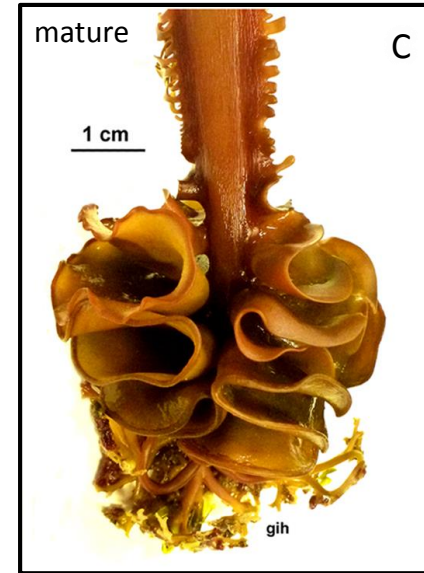
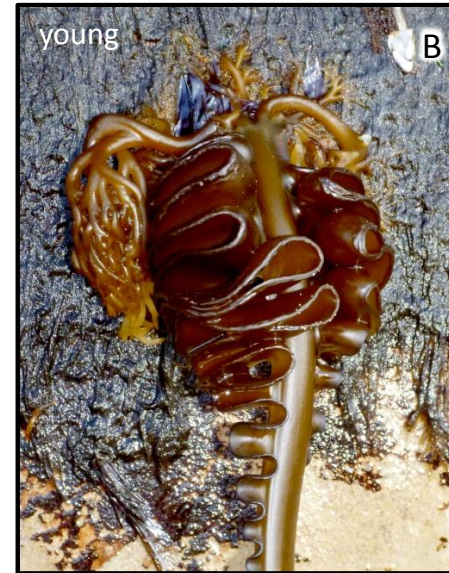


# Undaria 2 – Reproductive development



Sterile blade

Fertile sporophyll develops at the base of the stipe and is actively reproductive only after the ruffles mature.

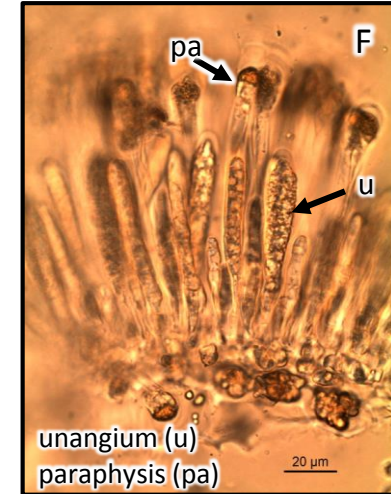
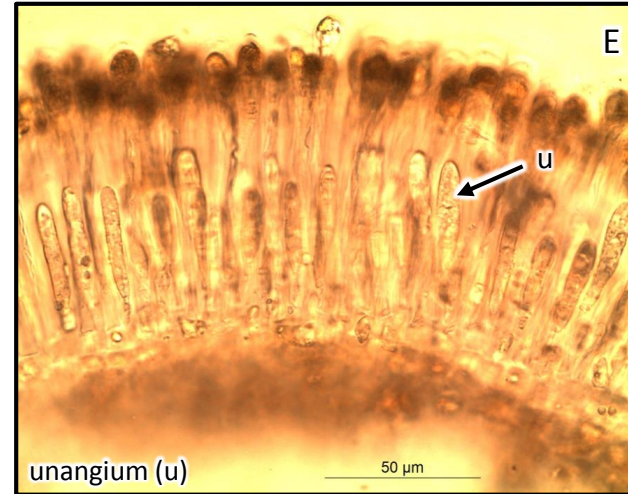


## Cross sections of:

(B, D) young sporophyll

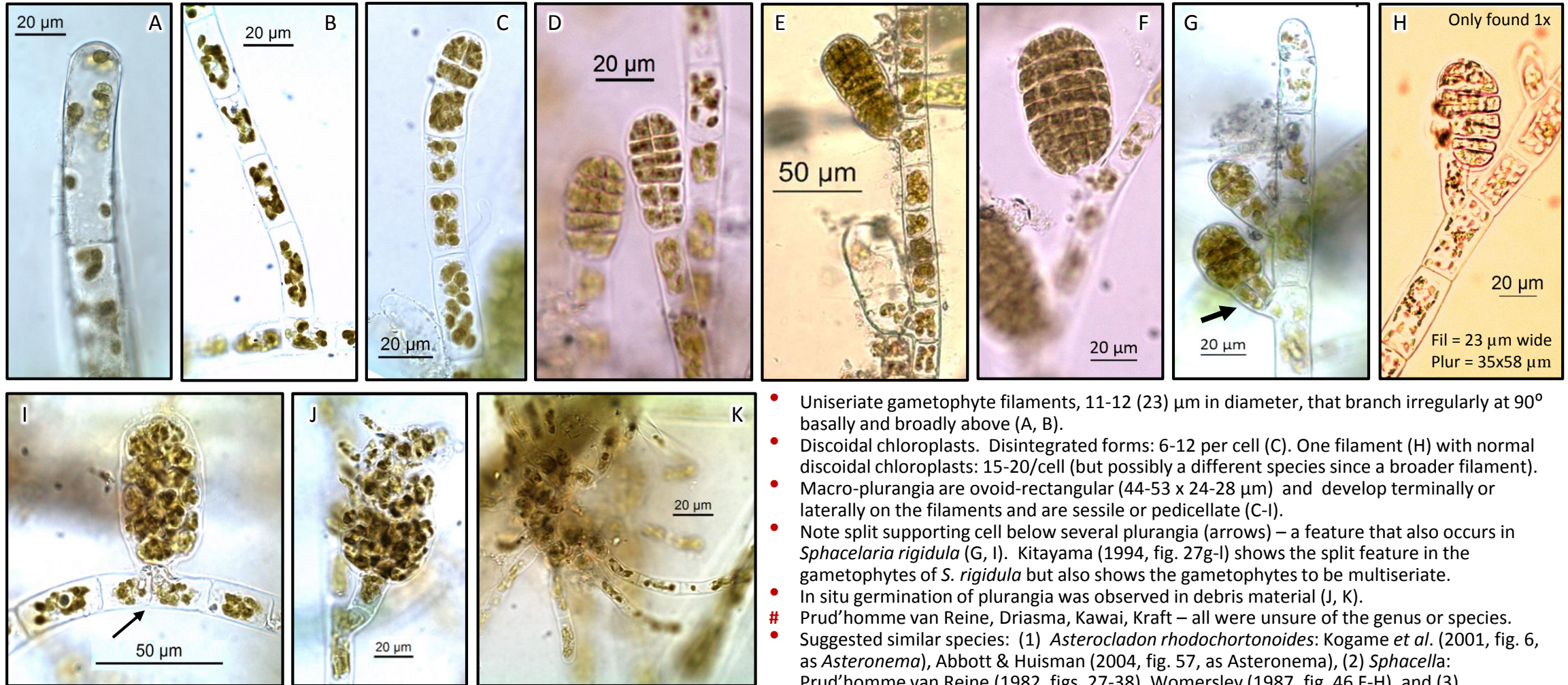
(C, E) maturing sporophyll

(C, F) mature sporophyll





**Unknown 1#** – Mixed with *Sphacelaria rigidula* in HF2-634 (BF-526), DNA failed. Chloroplasts appear plasmolyzed and abnormal (except H). Possibly an unusual gametophyte of *Sphacelaria rigidula* with only uniseriate filaments. Other suggestions by experts: *Sphacella* n. sp, *Asterocladon* sp., *Feldmannia* sp.

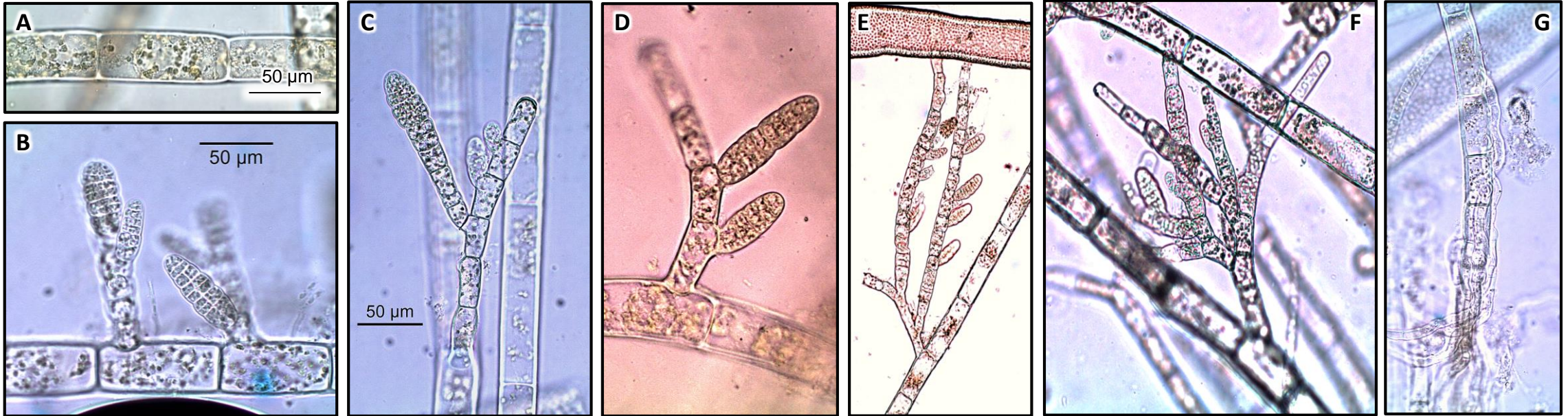


- Uniseriate gametophyte filaments, 11-12 (23)  $\mu\text{m}$  in diameter, that branch irregularly at  $90^\circ$  basally and broadly above (A, B).
- Discoidal chloroplasts. Disintegrated forms: 6-12 per cell (C). One filament (H) with normal discoidal chloroplasts: 15-20/cell (but possibly a different species since a broader filament).
- Macro-plurangia are ovoid-rectangular (44-53 x 24-28  $\mu\text{m}$ ) and develop terminally or laterally on the filaments and are sessile or pedicellate (C-I).
- Note split supporting cell below several plurangia (arrows) – a feature that also occurs in *Sphacelaria rigidula* (G, I). Kitayama (1994, fig. 27g-l) shows the split feature in the gametophytes of *S. rigidula* but also shows the gametophytes to be multiserial.
- In situ germination of plurangia was observed in debris material (J, K).
- # Prud'homme van Reine, Driasma, Kawai, Kraft – all were unsure of the genus or species.
- Suggested similar species: (1) *Asterocladon rhodochortonoides*: Kogame *et al.* (2001, fig. 6, as *Asteronema*), Abbott & Huisman (2004, fig. 57, as *Asteronema*), (2) *Sphacella*: Prud'homme van Reine (1982, figs. 27-38), Womersley (1987, fig. 46 F-H), and (3) *Feldmannia eilersii* cpx.: Kraft (2009, fig. 8).



**Unknown 2 – possibly *Feldmannia paradoxa* var. *caespitula* cpx.** (including *F. simplex* and *F. socialis*) – **G**

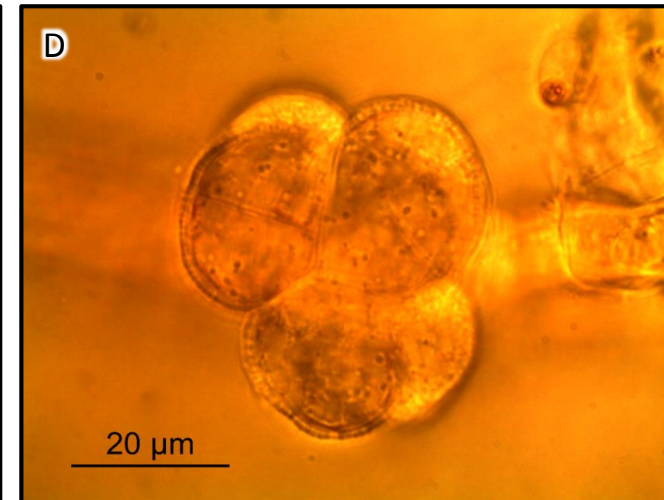
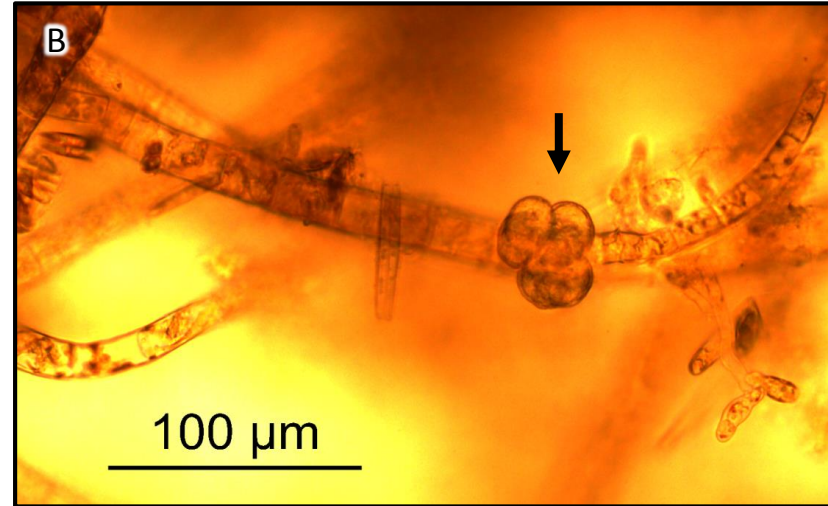
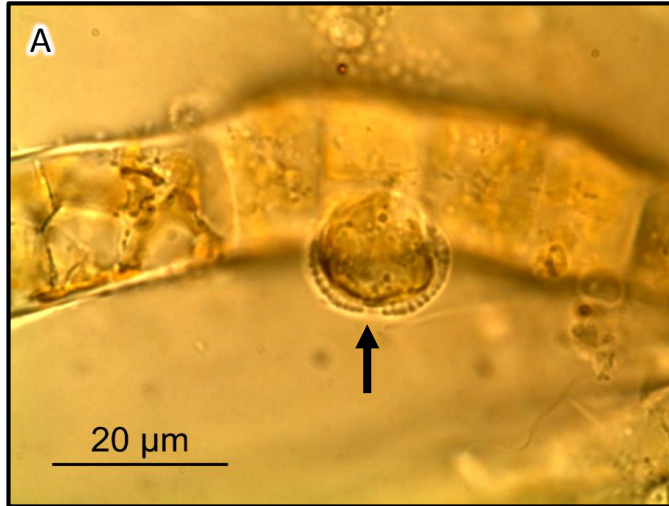
**(Widespread)** – Asia (J, K, C, Pak), AK-MX, ENA, SA, EUR, Med. Fragments only found on one JTMD site: CBD (BF-130). DNA not obtained.



- Small irregularly branching uniseriate filaments with main axes usually straight sided – fragments occurred intermixed with other algae in the Clatsop Beach Dock collection. Preserved – so no DNA.
- The pieces were too fragmented to be certain they were a single species, and therefore this sample was excluded from the main list.
- Chloroplasts appeared discoidal, although the material was partially disintegrated (A, B).
- Branches were 15-30 µm in diameter, varying from ~30 µm in the main axes to 10-15 µm in the often branched laterals (A-C).
- Small, sessile and stipitate, cylindrical plurangia with rounded apices were born only on the lateral branches and not on the main axes (B-F).
- Rhizoidal filaments occurred at the thallus base (G).
- Cormaci *et al.* (2012, pl. 21, figs. 4-6), Cardinal (1964, fig. 30. as *F. simplex*), Kim & Lee (1994, table 1 – similar to *Feldmannia paradoxa* var. *caespitula*).



**Unknown 3** – A wart-like infection of *Feldmannia mitchelliae*.





## References for the Brown Algae 1

- Abbott, I.A., & Hollenberg, G.J. (1976). Marine Algae of California. Stanford University Press, Stanford, pp. 1-827.
- Abbott, I.A. & Huisman, J.A. (2004). Marine Green and Brown Algae of the Hawaiian Islands. Bishop Museum Bulletin in Botany 4: [i-xi], 1-259.
- Ardisson, F. (1886). Phycologia mediterranea. P. II, Varese.
- Boo, S.-M., Lee, W.J., Hwang, I.K., Keum, Y.-S., Oak, J.H., & Cho, G.Y. (2010). Algal Flora of Korea. Volume 2, Number 2. Heterokontophyta: Phaeophyceae: Ishigeales, Dictyotales, Desmarestiales, Sphacelariales, Cutleriales, Ralfisales, Laminariales. Marine Brown Algae II. National Institute of Biological Resources, Incheon, pp. 1-203.
- Carlton, J.T., Chapman, J.W., Geller, J.B., Miller, J.A., Carlton, D.A., McCuller, M.I., Treneman, N.C., Steves, B.P. and Ruiz, G.M. (2017). Tsunami-driven rafting: Transoceanic species dispersal and implications for marine biogeography. Science 357: 1402–1406.
- Cardinal, A. (1964). Étude sur les Éctocarpacées de la Manche. Beihfte zur Nova Hedwigia 15: 1-86 + 41 figs.
- Cho, G.Y., Kogame, K., Kawai, H. & Boo, S.M. (2007). Genetic diversity of *Scytosiphon lomentaria* (Scytosiphonaceae, Phaeophyceae) from the Pacific and Europe based on RuBisCO large subunit and spacer, and ITS nrDNA sequences. Phycologia 46(6): 657-665.
- Cho, G.Y., Yang, E.C., Lee, S.H. & Boo, S.M. (2002). First description of *Petalonia zosterifolia* and *Scytosiphon gracilis* (Scytosiphonaceae, Phaeophyceae) from Korea with special reference to nrDNA ITS sequence comparisons. Algae 17(3): 135-144.
- Clayton, M.N. (1978). Morphological variation and life history in cylindrical forms of *Scytosiphon lomentaria* (Scytosiphoniaceae, Phaeophyta) from southern Australia. Marine Biology 47: 349-357.
- Cormaci, M., Furnari, G., Catra, M., Alongi, G. & Giaccone, G. (2012). Flora marina bentonica del Mediterraneo: Phaeophyceae. Bollettino dell'Accademia Gioenia 45: 1-508.
- DeCew, T. (1997). Guide to the Seaweeds of British Columbia, Washington, Oregon, and Northern California (In Part). Center for Phycological Documentation, University Herbarium, University of California, Berkeley. <http://ucjeps.berkeley.edu/guide/dq-toc.html>
- Driasma, S.G.A., Keum, Y.-S., Prud'homme van Reine, W.F., & Lokhorst, G. M. (1998). The species of *Sphacelaria* (Sphacelariales Phaeophyceae) in China with a description of a new species. Botanica Marina 41:181-190.
- Driasma, S.G.A., Prud'homme van Reine, W.F., & Kawai, H. (2010). A revised classification of the Sphacelariales (Phaeophyceae) inferred from a psbC and rbcL based phylogeny. European J. Phycology 45(3): 308-326.



## References for the Brown Algae 2

- Fletcher, R.L. (1987). Seaweeds of the British Isles. Vol. 3. Fucophyceae (Phaeophyceae). Part 1. British Museum (Natural History), London, pp. [i]-x, [1]-359.
- Gabrielson, P.W., Lindstrom, S.C. & O'Kelly, C.J. (2012). Keys to the Seaweeds and Seagrasses of Southeast Alaska, British Columbia, Washington, and Oregon. Phycological Contribution No. 8. Island Blue/Printorium Bookworks, Victoria, pp. [i]-iv, 1-192.
- Guiry, M.D. & Guiry, G.M. (searched 2017). AlgaeBase. World-wide electronic publication, National University of Ireland, Galway. <http://www.algaebase.org>
- Hamel, G. (1931-1939). Phéophycées de France. Published by the Author, Paris, pp. [i]-xlvi, 1-431 + 10 pls.
- Hanyuda, T., Hansen, G.I. & Kawai, H. (2017, In Press). Genetic identification of macroalgal species on Japanese tsunami marine debris and genetic comparisons with their wild populations. Marine Pollution Bulletin. <https://doi.org/10.1016/j.marpolbul.2017.06.053>
- Kawai, H., Kogishi, K., Hanyuda, T., and Kitayama, T. (2012). Taxonomic revision of the genus *Cutleria* proposing a new genus *Mutimo* to accommodate *M. cylindricus* (Cutleriaceae, Phaeophyceae). Phycological Research 60: 241–248.
- Keum, Y.-S., Oak, J.H., Prud'homme van Reine, W.F. & Lee, I.K. (2001a). Two species of *Sphacelaria* (Sphacelariales, Phaeophyceae), *S. solitaria* (Pringsheim) Kylin and *S. recurva* sp. nov. from Korea. Botanica Marina 44: 267-275.
- Keum, Y.-S., Oak, J.H., Draisma, S.G.A., Prud'homme van Reine, W.F. & Lee, I.K. (2001b). Taxonomic reappraisal of *Sphacelaria rigidula* and *S. fusca* (Sphacelariales, Phaeophyceae) based on morphology and molecular data with special reference to *S. didichotoma*. Algae 20: 1–13.
- Kim, H.-S. (2010). Ectocarpaceae, Acinetopsoraceae, Chordariaceae. In: Algal flora of Korea. Volume 2, Number 1. Heterokontophyta: Phaeophyceae: Ectocarpales. Marine Brown Algae I. (Kim, H.-S. & Boo, S.-M. Eds). National Institute of Biological Resources, Incheon, pp. [3]-137.
- Kim, H.-S. & Lee, I.K. (1992a). Morphotaxonomic studies on the Korean Ectocarpaceae (Phaeophyta). I. Genus *Ectocarpus* Lyngbye. Korean J. of Phycology 7(2): 225-242.
- Kim, H.-S. & Lee, I.K. (1992b). Morphotaxonomic studies on the Korean Ectocarpaceae (Phaeophyta). II. Genus *Hincksia* J.E. Gray. Korean J. of Phycology 7(2): 243-256.
- Kim, H.-S. & Lee, I.K. (1994). Morphotaxonomic studies on the Korean Ectocarpaceae (Phaeophyta). III. Genus *Feldmannia* Hamel, specially referred to morphogenesis and phylogenetic relationship among related genera. Korean J. of Phycology 9(2): 153-168.
- Kitayama, T. (1994). A taxonomic study of Japanese *Sphacelaria* (Sphacelariales, Phaeophyceae). Bulletin of the National Science Museum, Tokyo, Ser. B, 20 (2-3 ): 37-141.



## References for the Brown Algae 3

- Kjellman, F.R. (1883). The algae of the Arctic Sea. A survey of the species, together with an exposition of the general characters and the development of the flora. Kongliga Svenska Vetenskaps-Akademiens Handlingar 20(5): 1-351, 31 plates.
- Kjellman, F.R. & Petersen, J.V. (1885). Om Japans Laminariaceer. Vega-expeditionens Vetenskapliga Iakttagelser, Stockholm 4: 255-280, 1 table.
- Kogame, K. (1997). Sexual reproduction and life-history of *Petalonia fascia* (Scytosiphonales, Phaeophyceae). Phycologia 36: 389-394.
- Kogame, K. (1998). A taxonomic study of Japanese *Scytosiphon* (Scytosiphonales, Phaeophyceae), including two new species. Phycological Research 46: 39-51.
- Kogame, K. & Kawai, H. (1993). Morphology and life history of *Petalonia zosterifolia* (Reinke) O. Kuntze (Scytosiphonales, Phaeophyceae) from Japan. Japanese J. of Phycology 41: 29-37.
- Kogame, K., Uwai, S. & Kawaguchi, S. (2001). First record of *Asteronema rhodochortonoides* (Phaeophyceae) for the Pacific Ocean. Phycological Research 49: 281-284.
- Kogishi, K., Kitayama, T., Miller, K.A., Hanyuda, T. & Kawai, H. (2010). Phylogeography of *Cutleria cylindrica* (Cutleriales, Phaeophyceae) in northeastern Asia, and the identity of an introduced population in California (Note). J. of Phycology 46(3): 553-558.
- Kornmann, P. & Sahling, P.-H. (1977). Meeresalgen von Helgoland. Benthische Grün-, Braun und Rotalgen. Helgoländer Wissenschaftliche Meeresuntersuchungen 29: 1-289.
- Kraft, G.T. (2009). Algae of Australia. Marine Benthic Algae of Lord Howe Island and the Southern Great Barrier Reef, 2. Brown Algae. Australian Biological Resources Study and CSIRO Publishing, Canberra & Melbourne, pp. [i-vi], 1-364.
- Kuckuck, P. (1958). Ectocarpaceen-Studien V. *Kuckuckia*, *Feldmannia*. Helgoländer Wissenschaftliche Meeresuntersuchungen 6: 171-192.
- Kuckuck, P. (1963). Ectocarpaceen-Studien VIII. Einige Arten aus warmen Meeren. Helgoländer Wissenschaftliche Meeresuntersuchungen 9: 361-382.
- Kuckuck, P. (1964). Ectocarpaceen-Studien IX. Untersuchung von Herbarmaterial. Helgoländer Wissenschaftliche Meeresuntersuchungen : 1-23.
- Kützing, F.T. (1843). Phycologia generalis oder Anatomie, Physiologie und Systemkunde der Tange. Mit 80 farbig gedruckten Tafeln, gezeichnet und gravirt vom Verfasser. F. A. Brockhaus, Leipzig, pp. [part 1]: [i]-xxxii, [1]-142, [part 2:] 143-458, 1, err., pls. 1-80.
- Kützing, F.T. (1855). Tabulae phycologicae; oder, Abbildungen der Tange. Vol. V. Gedruckt auf kosten des Verfassers (in commission bei W. Köhne), Nordhausen, pp. i-ii, 1-30, 100 pls.



## References for the Brown Algae 4

- Kylin, H. (1947). Die Phaeophyceen der schwedischen Westküste. Acta Universitatis Lundensis 43(4): 1-99 + 18 pls.
- Lee, Y. (2008). Marine Algae of Jeju, Academy Publication, Seoul, [i]-xvi, 1-477 + map.
- Loiseaux, S. (1970). Notes on several Myrionemataceae from California using culture studies. J. of Phycology 6: 248-260.
- Luan, R., Ding, L., Lu, B. & Tseng, C.K. (2013). Flora Algarum Marinarum Sinicarum, Tomus III, Phaeophyta, No. I(1) Ectocarpales, Ralfsiales, Sphaceariales, and Dictyotales. Science Press, Beijing, pp. [i]-xxii, 1-195, pls. I-XXII.
- Mathieson, A.C. & Dawes, C.J. (2017). Seaweeds of the Northwest Atlantic. University of Massachusetts Press, Amherst and Boston, pp. [i-x], 1-798.
- Miller, K.A., Aguilar-Rosas, L.E., & Pedroche, F.F. (2011). A review of non-native seaweeds from California, USA, and Baja California, Mexico. Hidrobiológica 21 (3): 365-379.
- Miyabe, K. (1902). On the Laminariaceae of Hokkaido. J. of the Sapporo Agricultural College 1: 1-50, pls. I-XXIX.
- Montecinos, A.E., Couceiro, L., Peters, A.F., Desrut, A., Valero, M. & Guillemin, M.L. (2017). Species delimitation and phylogeographic analyses in the *Ectocarpus* subgroup *siliculosi* (Ectocarpales, Phaeophyceae). *Journal of Phycology* 53(1): 17-31.
- Nagai, M. (1940). Marine algae of the Kurile Islands, I. J. of the Faculty of Agriculture, Hokkaido Imperial University 46: 1-137.
- Nelson, W.A. (1982a). Development, anatomy and reproduction of *Analipus japonicus* (Harv.) Wynne (Phaeophyta, Heterochordariaceae). *Botanica Marina* 25: 357-369.
- Nelson, W.A. (1982b). A critical review of the Ralfsiales, Ralfsiaceae and the taxonomic position of *Analipus japonicus* (Harv.) Wynne (Phaeophyta). *British Phycological J.* 17 (3): 311-320.
- Noda, M. (1970). Some marine algae collected on the coast of Iwagasaki, Prov. Echigo facing the Japan Sea. Science Reports of Niigata University, Series D (Biology) 7: 27-35.
- Noda, M. (1972). Some marine algae collected on the coast of Kashiwazaki Province facing the Japan Sea (1). Science Reports of Niigata University Series D (Biology) 9: 1-15.
- Noda, M. (1979). The species of Phaeophyta from Sado Island in the Japan Sea. Science reports of Niigata University, Series D. Biology, 6: 1-64.



## References for the Brown Algae 5

- Noda, M. (1980). On the species of *Ectocarpus* Lyngbye from Echigo Province (Niigata Prefecture) facing the Japan Sea. *Seibutsu* 3: 1-25.
- Norris, J.N. (2010). Marine Algae of the Northern Gulf of California: Chlorophyta and Phaeophyceae. *Smithsonian Contributions to Botany* 94: 1-276.
- Ohta, T. (1973). Some new and rare marine algae from Tsugaru straits between Honshu and Hokkaido. *Scientific Reports of Niigata University, Series D (Biology)* 10: 11-28.
- Okamura, K. (1900-1902). Illustrations of the Marine Algae of Japan. Vol 1 (I-VI). Keigyosha & Co., Tokyo, pp. 1-93, plates I-XXX.
- Okamura, K. (1907-1909). Icones of Japanese Algae. Vol. I. The Author, Tokyo, pp. 1-258, pls. I-L + Index.
- Okamura, K. (1915). *Undaria* and its species. *Botanical Magazine, Tokyo* 29:266-278.
- Okamura, K. (1916). Icones of Japanese algae. Vol. IV. The Author, Tokyo, pp. 1-40, pls. CLI-CLX.
- Okamura, K. (1923-1928). Icones of Japanese Algae. Vol V. The Author, Tokyo, pp. 1-203, pls. CCI-CCL + Index.
- Okamura, K. & Uyeda, S. (1925). On *Laminaria angustata* Kjellm. and *L. longissimi* Miyabe. *J. of the Imperial Fisheries Institute* XXI (2): 20-25 + pl. 1.
- Pankow, H. (1971). Algenflora der Ostsee I. Benthos (Blau-, Grün, Braun- und Rotalgen). *Gustav Fischer Verlag, Stuttgart*, pp. 1-419.
- Parente, M.I., Fletcher, R.L., Neto, A.I., Tittley, I., Sousa, A.F., Draisma, S. & Gabriel, D. (2010). Life history and morphological studies of *Punctaria tenuissima* (Chordariaceae, Phaeophyceae), a new record for the Azores. *Botanica Marina* 53(3): 223-231.
- Pedersen, P.M. (1989). Studies on *Kuckuckia spinosa* (Fucophyceae, Sorocarpaceae): life history, temperature gradient experiments, and synonymy. *Nordic J. of Botany* 9 (4): 443-447.
- Pedroche, P.F., Silva, P.C., Aguilar Rosas, L.E., Dreckmann, K.M. & Aguilar Rosas, R. (2008). Catálogo de las algas bentónicas del Pacífico de México II. Phaeophycota. Universidad Autónoma Metropolitana and University of California Berkeley, Mexicali & Berkeley, pp. [i-viii], i-vi, 15-146.
- Peters, A.F. & Müller, D.G. (1986). Life-history studies—a new approach to the taxonomy of ligulate species of *Desmarestia* (Phaeophyceae) from the Pacific coast of Canada. *Canadian J. of Botany* 64: 2192-2196.
- Peters, A.F., Couceiro, L., Tsiamis, K., Kuepper, F.C., & Valero, M. (2015). Barcoding of cryptic stages of marine brown algae isolated from incubated substratum reveals high diversity in Acinetosporaceae (Ectocarpales, Phaeophyceae). *Cryptogamie Algologie* 36: 3–29.



## References for the Brown Algae 6

- Peters, A.F., van Oppen, M.J.H., Wiencke, C., Stam, W.T. & Olsen, J.L. (1997). Phylogeny and historical ecology of the Desmarestiaceae (Phaeophyceae) support a southern hemisphere origin. *J. of Phycology* 33: 294-309.
- Peters, A.F., Van Wijk, S.J., Scornet, D., Kawai, H., Schroeder, D.S., Cock, J.M. & Boo, S.M. (2010). Reinstatement of *Ectocarpus crouaniorum* Thuret in Le Jolis as a third common species of *Ectocarpus* (Ectocarpales, Phaeophyceae) in Western Europe, and its phenology at Roscoff, Brittany. *Phycological Research* 58(3): 157-170.
- Postels, A. & Ruprecht, F. (1840). *Illustrationes algarum* in itinere circum orbem jussu imperatoris Nicolai I. Atque auspiciis navarchi Friderici Lütke annis 1826, 1827, 1828 et 1829 celoce Seniavin exsecuto in Oceano pacifico, inprimis septemtrionale ad littora rossica asiatico-americana collectarum. Typis Eduardi Pratz, Petropoli [St. Petersburg], pp. [i-iv], 1-24 with index + 1 + 40 pls. [In Latin]
- Prud'homme van Reine, W.F. (1982). A taxonomic revision of the European Sphacelariaceae (Sphacelariales, Phaeophyceae). *Leiden Botanical Series* 6: [i-x], 1-293.
- Russell, G. (1966). The genus *Ectocarpus* in Britain. I. The attached forms. *J. of the Marine Biological Association of the United Kingdom* 46 (2): 267-294.
- Saito, Y. (1972). On the effects of environmental factors on morphological characteristics of *Undaria pinnatifida* and the breeding of hybrids in the genus *Undaria*. In Abbott, I.A. & Kurogi, M. (Eds.), *Contributions to the Systematics of Benthic Marine Algae of the North Pacific*. Japanese Society of Phycology, Kobe, pp. 117-133.
- Saunders, De A. (1898). *Phycological memoirs*. Proceedings of the California Academy of Sciences. Series 3, Botany 1: 147-168, pls. XII-XXXII.
- Sauvageau, C. (1933). Sur quelques Algues phéosporées de Guéthary (Basses-Pyrénées). *Bulletin de la Station biologique d'Arcachon* 30: 1-128.
- Schneider, C.W. & Searles, R.B. (1991). *Seaweeds of the Southeastern United States, Cape Hatteras to Cape Canaveral*. Duke University Press, Durham, pp. [i-xiv] 1-553.
- Sears, J.R. & Wilce, R.T. (1973). Sublittoral benthic marine algae of southern Cape Cod and adjacent islands: *Pseudolithoderma paradoxum* sp. nov. (Ralfsiaceae, Ectocarpales). *Phycologia* 12: 75-82.
- Setchell, W.A. & Gardner, N.L. (1922). *Phycological Contributions VI. New species of Ectocarpus*. University of California Publications in Botany 7 (11): 403-427.
- Setchell, W.A. & Gardner, N.L. (1925). *The marine algae of the Pacific coast of North America. Part III. Melanophyceae*. University of California Publications in Botany 8: 383-898.



## References for the Brown Algae 7

- Stache-Crain, B., Müller, D.G. & Goff, L.J. (1997). Molecular systematics of *Ectocarpus* and *Kuckuckia* (Ectocarpales, Phaeophyceae) inferred from phylogenetic analysis of nuclear- and plastid-encoded DNA sequences. *J. of Phycology* 33: 152-168.
- Stegenga, H. & Mol, I. (1983). Some marine brown algae new or rare to the Netherlands. *Acta botanica Neerlandica* 32 (3): 153-162.
- Streftaris, N. and Zenetos, A. (2006). Alien marine species in the Mediterranean – the 100 “worst Invasives” and their impact. *Mediterranean Marine Science* 7 (1): 87-118.
- Tanaka, J. (1986). The taxonomy of *Protectocarpus speciosus* (Borgesen) Kornmann (Myrionemataceae, Phaeophyceae). *Japanese J. Phycology* 34: 287-292.
- Tanaka, J. & Chihara, M. (1981). Taxonomic study of Japanese crustose brown algae. (6) *Pseudolithoderma* (Lithodermataceae, Ralfsiales). *J. of Japanese Botany* 56: 376-381.
- Tatewaki, M. (1966). Formation of a crustaceous sporophyte with unilocular sporangia in *Scytosiphon lomentaria*. *Phycologia* 6 (1) 62-66.
- Taylor, W.R. (1957). Marine algae of the northeastern coast of North America. The University of Michigan Press, Ann Arbor, pp. [i]-vii, [1]-509.
- Tokida, J. (1954). The marine algae of southern Saghalien. *Memoirs of the Faculty of Fisheries, Hokkaido University* 2: 1-264.
- Tokuda, H., Kawashima, S., Ohno, M., and Ogawa, H. (1994). A Photographic Guide to the Seaweeds of Japan. Midori Shobo Co., Ltd., Tokyo, pp. 1-193.
- Uwai, S., Nelson, W. Neill, K., Wang, W. D., Aguilar-Rosa, L. E., Boo, S. M., Kitayama, T., and Kawai, H. (2006). Genetic diversity in *Undaria pinnatifida* (Laminariales, Phaeophyceae) deduced from mitochondria genes – origins and succession of introduced populations. *Phycologia* 45 (6): 687-695.
- Verlaque, M. (2001). Checklist of the macroalgae of Thau Lagoon. *Oceanologica Acta* 24 (1): 29-49.
- Verlaque, M., Ruitton, S., Mineur, F. & Boudouresque, C.-F. (2015). CIESM Atlas of Exotic Species of the Mediterranean. Macrophytes. CIESM Publishers, Monaco, pp. 1-362.
- Waern, M. (1949). Remarks on some Swedish *Lithoderma*. *Svensk Botanisk Tidskrift* 43: 633-670 + 3 pls.
- Waern, M. (1952). Rocky-shore algae in the Öregrund Archipelago. *Acta Phytogeographica Suecica* 30: [i-xvi], 1-298.
- Wilce, R.R., Webber, E.E., and Sears, J.R. (1970). *Petroderma* and *Porterinema* in the New World. *Marine Biology* 5 (2): 119-135.
- Womersley, H.B.S. (1987). The Marine Benthic Flora of Southern Australia. Part II. South Australian Government Printing Division, Adelaide, pp. 1-484.



## References for the Brown Algae 8

- Yaegashi, K., Yamagishi, Y., Uwai, S., Abe, T., Santiañez, W.J.E. & Kogame, K. (2015). Two species of the genus *Acinetospora* (Ectocarpales, Phaeophyceae) from Japan: *A. filamentosa* comb. nov. and *A. asiatica* sp. nov. *Botanica Marina* 58(5): 331-343.
- Yamada, Y. and Tanaka, T. (1944). Marine algae in the vicinity of Akkesi Marine Biological Station. *Scientific Papers of the Institute of Algological Research, Hokkaido University* 3: 47-77.
- Yang, E.C., Peters, A.F., Kawai, H., Stern, R., Hanyuda, T., Bárbara, I., Müller, D.G., Strittmatter, M., Prud'homme van Reine, W.F. & Küpper, F.C. (2014). Ligulate *Desmarestia* (Desmarestiales, Phaeophyceae) revisited: *D. japonica* sp. nov. and *D. dudresnayi* differ from *D. ligulata*. *J. of Phycology* 50(1): 149-166.
- Yoshida, T. (1998). *Marine Algae of Japan*. Uchida Rokakuho Publishing Co., Ltd., Tokyo, pp. [1-2], 1-25, 1-1222.
- Yoshida, T., Suzuki, M. & Yoshinaga, K. (2015). Checklist of marine algae of Japan (Revised in 2015). *Japanese J. of Phycology* 63: 129-189.
- Yotsukura, N., Kawai, T., Kawashima, W., Ebata, H. & Ichimura, T. (2006). Nucleotide sequence diversity of the 5S rDNA spacer in the simple blade kelp genera *Laminaria*, *Cymanthaere* and *Kjellmaniella* (Laminariales, Phaeophyceae) from northern Japan. *Phycological Research* 54: 269-279.
- Yotsukura, N., Shimizu, T., Katayama, T. & Druehl, L.D. (2010). Mitochondrial DNA sequence variation of four *Saccharina* species (Laminariales, Phaeophyceae) growing in Japan. *J. of Applied Phycology* 22: 243-251.

## Appendix 1 – Japanese Debris Items

Japanese Tsunami Marine Debris (JTMD) items collected for the algal project, including their BF-numbers, state, site name, collection number abbreviations, collection date and year, and item type. All collections were made between Mosquito Creek, WA, and Sixes River, OR. Key: Abbrev.= collecting number abbreviation, BF # = biofouling number of Carlton *et al.* (2017, Table S1), OR = Oregon, WA = Washington.

BF #	State	Site Name	Abbrev.	Collection		Item
				Date	Year	
BF-1	OR	Agate Beach	AB	5-Jun	2012	dock
BF-2	WA	Ilwaco, Benson Beach	BB	15-Jun	2012	boat
BF-8	WA	Mosquito Creek	MC	5-Jan	2013	dock
BF-23	OR	Gleneden Beach, Salishan	GB	6-Feb	2013	boat
BF-28	OR	Horsfall Beach	HF1	21-Feb	2013	boat
BF-36	OR	Florence, Muriel Ponsler Park	MP	14-Mar	2013	boat
BF-39	OR	Cannon Beach, S Jockey Cap	SJC	22-Mar	2013	boat
BF-40	WA	Long Beach (fish boat)	Fish	22-Mar	2013	boat
BF-50	OR	Coos Bay North Spit	CBS	22-Apr	2013	boat
BF-58	OR	Clatsop Beach	CBB	30-May	2013	boat
BF-59/61	OR	Nye Beach	Nye	30-May	2013	post & beam
BF-108	OR	Cape Arago, Lighthouse Beach	CA	11-Jul	2013	post & beam
BF-130	OR	Clatsop Beach	CBD	9-Oct	2013	dock, pontoon
BF-134	WA	Twin Harbors State Park	TH	17-Jan	2014	boat
BF-135	OR	Yachats	Yac	18-Feb	2014	boat
BF-160	OR	Tillamook Bay spit	TBT	26-Apr	2014	tree
BF-171	OR	Tillamook Bay spit	TB	25-Apr	2014	post & beam
BF-173	OR	South Beach, Lost Creek	LC	27-Apr	2014	buoy
BF-188	OR	Cape Lookout Beach	CL	3-May	2014	boat



## Appendix 1 (continued) – Japanese Debris Items

BF #	State	Site Name	Abbrev.	Collection		Item
				Date	Year	
BF-196	OR	Waldport	Wal	12-May	2014	boat
BF-208	OR	Cape Arago, North Cove	NC	19-May	2014	boat
BF-223/224	WA	Long Beach, Ilwaco	Ilw2	29-May	2014	boats 2
BF-227/228	WA	Long Beach	LB2	5-Jun	2014	boats 2
BF-234	OR	South Beach	SBT	9-Feb	2013	tank
BF-235	WA	Long Beach	LBT	1-Mar	2013	tire
BF-277	OR	Seal Rock	SRT	30-Nov	2014	tote
BF-285	WA	Long Beach	LB	4-Jan	2015	boat fragment
BF-288	OR	Beverly Beach	Bev	20-Jan	2015	tote, pallet
BF-293	WA	Long Beach, Seaview	SV	28-Jan	2013	pipe
BF-331	WA	Oysterville	Oys	14-Mar	2014	boat
BF-356	OR	Seal Rock, in ocean	SR	10-Apr	2015	boat
BF-397	WA	Long Beach	LBD	1-May	2015	dock, pontoon
BF-402	WA	Long Beach, Seaview	SVB	12-May	2015	boat
BF-461	OR	Manzanita	Man	2-Mar	2015	tote, basket
BF-462	WA	Long Beach	LBF	4-Jan	2015	float
BF-500	WA	Long Beach	LBT	16-Feb	2016	tote
BF-526	OR	Horsfall Beach 2	HF2	22-Mar	2016	boat
BF-533	OR	Roads End	RE	28-Mar	2016	boat
BF-538	OR	Sixes River mouth	SixR	16-Apr	2016	boat
BF-545	OR	Umqua River mouth	Ump	26-Mar	2016	Boat
BF-652	OR	Falcon Cove beach	Fal	26-Jul	2016	Boat
BF-656	OR	Quail Street	QS	26-Mar	2016	carboy

## Appendix 2 – *Ectocarpus-Kuckuckia cox3* tree.

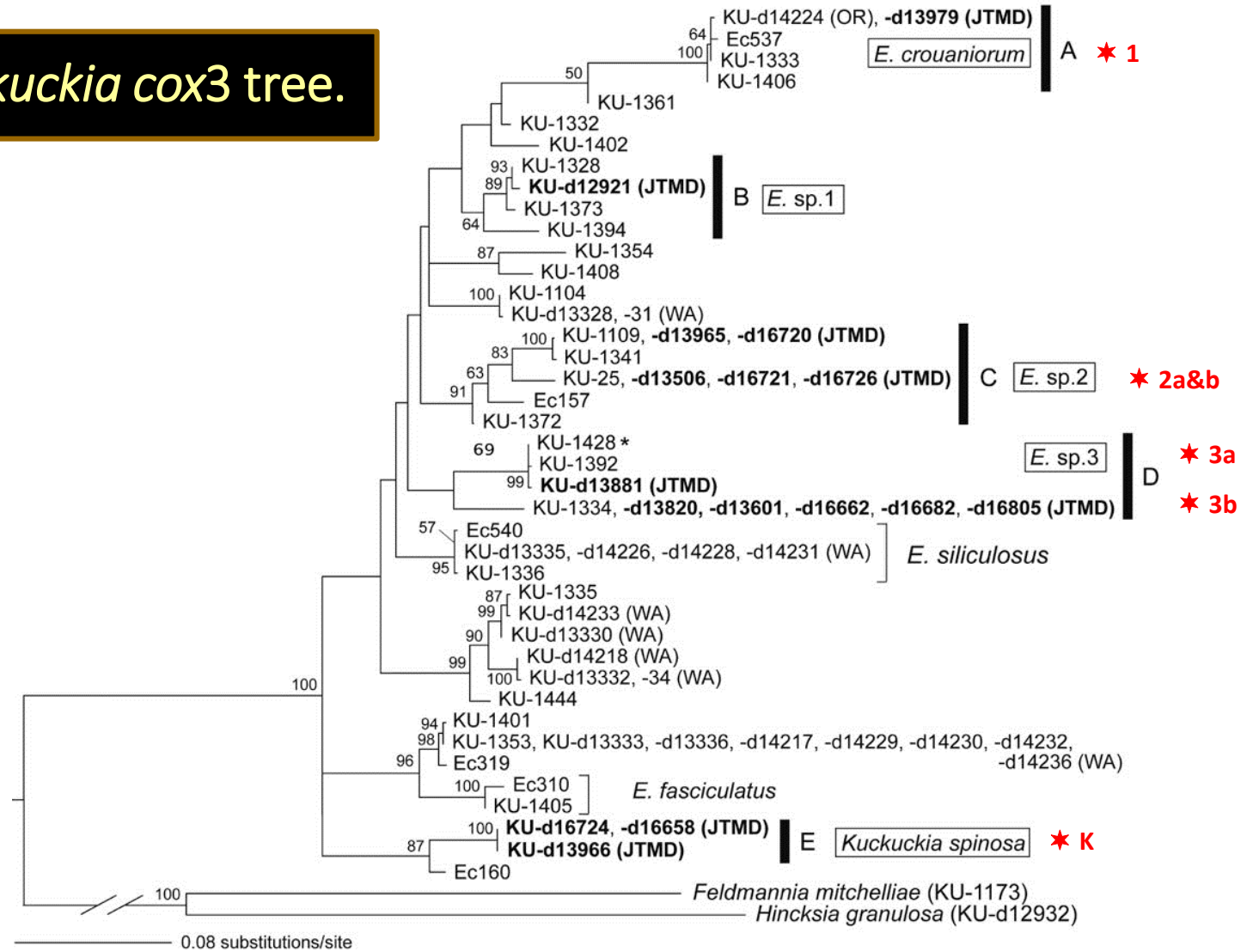
The phylogenetic tree at right is Fig. S1. from Hanyuda *et al.* (2017), and it is the molecular basis of the JTMD *Ectocarpus* and *Kuckuckia* identifications in this presentation.

### “Fig. S1. Molecular phylogeny of *Ectocarpus* spp. and *Kuckuckia* spp.” (modified)

“Maximum likelihood tree based on the mitochondrial *cox3* gene sequences (665 bp). Numbers at nodes indicate bootstrap values in maximum likelihood analysis. Only bootstrap values >50% are shown. JTMD isolates are shown in bold.”

★ = the clades examined in the morphological study

Appendix 3 is a guide to the KU-sequence numbers (Kobe University) shown for the JTMD species in this tree.



\*KU-d12934 (JTMD), -d12936 (JTMD), -d13501 (JTMD), -d13575 (WA), -d13576 (WA), -d13829 (JTMD), -d13837 (JTMD), -d16723 (JTMD), and -d16725 (JTMD) had the identical sequence with KU-1428.



## Appendix 3. The JTMD *Ectocarpus* and *Kuckuckia* samples sequenced.

The following tables are excerpts from Table S1 of Hanyuda *et al.* (2017). The sequence references are from: Peters *et al.* (2010-*cox3*, 2015-*cox1*), Kogishi *et al.* (2010-*cox3*). Accession numbers are listed in S4 of the 2017 paper.

Genetic identification	Clade in Appendix 2, GIH approximate identification	Apparent morphological variation	Kobe University sequence and silica gel numbers	BF Number	Location, item	Gene Sequenced
<b>REFERENCE SPECIMENS</b>						
<i>E. crouaniorum</i>	(A) <i>E. crouaniorum</i>		KU-1333		Portdeha, Aran Islands, Ireland	<i>cox3</i>
<i>E. sp. 1</i>	(B) <i>E. sp-1</i> , near <i>crouaniorum</i>		KU-1373		Tierra del Fuego, Argentina	<i>cox3</i>
<i>E. sp. 2</i>	(C) <i>E. sp-2a</i> , ~ <i>acutus</i>		KU-1109		Akkeshi, Hokkaido, Japan	<i>cox3</i>
<i>E. sp. 2</i>	(C) <i>E. sp-2b</i> , ~ <i>acutus</i>		KU-25		Awaji I., Hyogo Pref., Japan	<i>cox3</i>
<i>E. sp. 3</i>	(D) <i>E. sp-3a</i> , ~ <i>pygmaeus</i>		KU-1428		Roscoff, France	<i>cox3</i>
<i>E. sp. 3</i>	(D) <i>E. sp-3b</i> , sp-with-hairs	<i>K. spinosa</i>	KU-1334		Helgoland, Germany	<i>cox3</i>
<i>Kuckuckia spinosa</i>	(E) <i>Kuckuckia spinosa</i>	<i>E. arctus</i>	Ec-160		Arica, Chile	<i>cox3</i>

## Appendix 3 (continued)

Genetic identification	Clade in Appendix 2, GIH approximate identification	GIH Sample numbers	Kobe Univ. sequence and silica gel numbers	BF Number	Location, item	Gene Sequenced
<b>JAPANESE TSUNAMI MARINE DEBRIS SAMPLES</b>						
<i>Ectocarpus crouaniorum</i>	<i>E. crouaniorum</i>	NC-5	KU-d13979	BF-208	Cape Arago, OR, boat	<i>rbcL, cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-1, near crouaniorum</i>	SV-15-1	KU-d12921	BF-293	Long Beach, WA, pipe	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-2a, ~acutus</i>	Wal-1	KU-d13965	BF-196	Waldport, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-2a, ~acutus</i>	RE-686	KU-d16720	BF-533	Roads End, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-2b, ~acutus</i>	Bev-113	KU-d13506	BF-288	Beverly Beach, OR, pallet	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-2b, ~acutus</i>	RE-690	KU-d16721	BF-533	Roads End, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp. 2b, ~acutus</i>	RE-699	KU-d16726	BF-533	Roads End, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3a, ~pygmaeus</i>	CB-5	KU-d12934	BF-58	Clatsop Beach, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3a, ~pygmaeus</i>	LB-108+	KU-d13501, 75, 76	BF-285	Long Beach, WA, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3a, ~pygmaeus</i>	SR-223	KU-d13829	BF-356	Seal Rock, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3a, ~pygmaeus</i>	SR-234	KU-d13837	BF-356	Seal Rock, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3a, ~pygmaeus</i>	RE-695-Ect3	KU-d16723	BF-533	Roads End, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3a, ~pygmaeus</i>	RE-698	KU-d16725	BF-533	Roads End, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3a, ~pygmaeus</i>	SVB-383	KU-d13861	BF-402	Seaview, WA, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3a, ~pygmaeus</i>	LBF-405	KU-d13881	BF-462	Long Beach, WA, float	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3b, ~penicillatus?</i>	SR-213	KU-d13820	BF-356	Seal Rock, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3b, sp-with-hairs</i>	LBF-135	KU-d13601	BF-462	Long Beach, WA	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3b, sp-with-hairs</i>	Fal-750	KU-d16662	BF-652	Falcon Cove Beach, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3b, sp-with-hairs</i>	Fal-752	KU-d16682	BF-652	Falcon Cove Beach, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>E. sp-3b, sp-with-hairs</i>	Fal-753	KU-d16805	BF-652	Falcon Cove Beach, OR, boat	<i>cox3</i>
<i>Ectocarpus</i> sp.	<i>Kuckuckia</i>	RE-695-Ect4	KU-d16724	BF-533	Roads End, OR, boat	<i>cox3</i>
<i>Kuckuckia spinosa</i>	<i>Kuckuckia, with highly branched laterals</i>	SixR-719	KU-d16658	BF-538	Sixes River mouth, OR, boat	<i>cox3</i>
<i>Kuckuckia spinosa</i>	<i>E. arctus</i>	Wal-2	KU-d13966	BF-196	Waldport, OR, boat	<i>cox1, cox3</i>