RESEARCH ARTICLE



A new genus and a new species in the subfamily Polyzosteriinae (Blattodea, Blattidae) from China

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Abstract

Laevifacies quadrialata gen. et sp. nov. is described from Hainan Province, China based on morphological data. COI data (DNA barcodes) is utilized to confirm the sexual dimorphism occurring in *Laevifacies quadrialata* gen. et sp. nov. *Melanozosteria nitida* Brunner von Wattenwyl, 1865, is reported from Guangxi Province, China. A key to the Chinese Polyzosteriinae is provided.

Keywords

Blattaria, cockroaches, Laevifacies, Melanozosteria, molecular identification, morphology

Introduction

Polyzosteriinae is a relatively species-abundant subfamily in the Blattidae. The subfamily is flightless (except Methanini), having lobiform vestigial tegmina or being totally apterous. Some species have pits or tubercles scattered on the pronotum (Rentz 2014), short tarsi and large pulvilli and arolia (Mackerras 1968b). Members of Polyzosteriinae were firstly mentioned by Burmeister (1838), with the establishment of genus *Polyzosteria* Burmeister, 1838. Tepper (1893) erected the subfamily Polyzosteriinae with two genera, *Polyzosteria* and *Platyzosteria* Brunner von Wattenwyl, 1865. The revisionary works of Mackerras (1965a, 1965b, 1965c, 1966a, 1966b, 1967a, 1967b, 1968a,

1968b) included 16 genera, e.g. *Polyzosteria* Burmeister, 1838; *Platyzosteria* Brunner von Wattenwyl, 1865; *Cosmozosteria, Melanozosteria, Zonioploca* Stål, 1874; *Methana* Stål, 1877; *Anamesia, Drymaplaneta, Leptozosteria, Pseudolampra* and *Temnelytra* Tepper, 1893; *Desmozosteria, Euzosteria* and *Scabina* Shelford, 1909; *Eppertia* Shaw, 1925 and *Megazosteria* Mackerras, 1966a. In the catalogue, Princis (1966) recorded *Eurycotis* Stål, 1874. McKittrick (1964) provided a detailed description of the genitalia and proventriculus of the Polyzosteriinae female and male for the first time. Mackerras (1965a) divided the Australian members into two tribes: Polyzosteriini and Methanini, compiling the most complete account of Polyzosteriinae to date. Grandcolas (1997) described 5 genera from New Caledonia. Up to now, Polyzosteriinae contains 22 genera and 305 species (Beccaloni 2014), most of which are distributed in Australia, Southeast Asia, America, and the Pacific Islands.

The genus Melanozosteria was established with Polyzosteria nitida Brunner von Wattenwyl, 1865 as type species (Stål 1874). After that, Mackerras (1967b) did not agree with Melanozosteria as a synonym of Platyzosteria owing to the misidentification of one species of Platyzosteria by Shelford (1909), and placed Melanozosteria and Leptozosteria in Platyzosteria as subgenera. Roth (2003) re-established the taxonomic status of Melanozosteria as a genus. Currently 44 species are known of Melanozosteria, which are mainly distributed in Australia (Beccaloni 2014). Two Melanozosteria species are currently recorded in China (Melanozosteria nitida Brunner von Wattenwyl, 1865 and Melanozosteria soror Brunner von Wattenwyl, 1865). Melanozosteria nitida from Taiwan was originally determined as Periplaneta polita Walker, 1868. Then Shelford (1909) proposed that *Periplaneta polita* is a synonym of *Cutilia nitida* Brunner von Wattenwyl, 1865. Until now, they have both been considered synonyms of Melanozosteria nitida. In the catalogue, Princis (1966) recorded Melanozosteria nitida from Taiwan, China, but he guestioned its distribution on Mainland China. The other species, Melanozosteria soror, is mainly distributed in Australia and the Pacific Islands. Walker (1868) firstly recorded this species from Taiwan, China (it was originally described as Periplaneta philpotti, but later synonymized under Melanozosteria soror in Princis (1957)). Then Shiraki (1931) recorded this species from Hainan, but no further information was provided.

DNA barcodes have been proven to be a helpful method to identify species and to successfully match male and female. Barcoding has been applied to resolve the problems of sexual dimorphism and even to identify nymphs in cockroaches (Evangelista et al. 2013; Qiu et al. 2017; Che et al. 2017; Wang et al. 2018). To date, members of the Polyzosteriinae have been identified primarily on the basis of morphological characters (Mackerras 1965a, 1965b, 1965c, 1966a, 1966b, 1967a, 1967b, 1968a, 1968b; Rentz 2014) and DNA Barcoding has not been employed to investigate the diversity of Polyzosteriinae. In this paper, *Laevifacies quadrialata* gen. et sp. nov. is described from China and the sexual dimorphism is revealed via DNA barcoding. We also record a specimen from Guangxi, thus proving that *Melanozosteria nitida* is also distributed in Mainland China. A key to the known Polyzosteriinae species from China is provided.

Materials and methods

Morphological study

Morphological terminology used in this paper mainly follows McKittrick (1964), Mackerras (1965a) and Roth (2003). Measurements are based on specimens examined. Genital segments of the examined specimens were macerated in 10% NaOH for 20 minutes and rinsed with distilled water, observed in glycerin jelly using a MOTIC K400 stereomicroscope. Photographs of the specimens were taken using a Canon[®] 50D plus a Canon[®] EF 100mm f/2.8L IS USM Macro lens combined with Helicon Focus[®] software. Photos of other characters were taken using a Leica[®] M205A stereomicroscope. All photographs mentioned above were modified in Adobe Photoshop[®] CS6. The type materials are deposited in the Institute of Entomology, College of Plant Protection, Southwest University, Chongqing, China.

DNA extraction, PCR, and sequencing

We used two cockroach specimens for COI sequencing in this study in order to resolve the sexual dimorphism. Both sequences are deposited in GenBank with the accession numbers: MK798103, MK798104 (Table 1). The extraction procedure was according to the Hipure Tissue DNA Mini Kit (Magen Biotech, Guangzhou). Fragments of COI were amplified using PCR. Primers for the amplifications are LCO1490 (5'-GGTCAACAAATCATAAGATATTGG-3') and HCO2198 (5'-TAAACTTCAGGGTGACCAAAAAAATCA-3') (Folmer 1994). The amplification conditions were: initial denaturation at 94 °C for 3 min, followed by 35 cycles for 30 s at 94 °C, 30 s at 49 °C, and 1 min at 72 °C, with a final extension of 10 min at 72 °C.

Sequence processing and phylogenetic analyses

A total of ten COI sequences were analyzed (two sequences of *Laevifacies* species from our study, six sequences of Blattidae, and two sequences of a mantid outgroup downloaded from GenBank) (Table 1). All COI sequences were aligned using MEGA 7.0 and adjusted visually after translation into amino acid sequences. Finally, for the phylogenetic analysis we acquired COI sequences whose lengths were 658 bp, except for *Angustonicus lifou* whose sequence was only 650 bp. The genetic divergence value was quantified based on the Kimura 2-parameter (K2P) distance model (Kimura 1980), using MEGA 7 (Kumar et al. 2016) with 1000 bootstrap replicates. Maximum Likelihood (ML) analysis was implemented in RAxML 7.3.0 (Stamatakis et al. 2008) using GTRGAMMA model with 1000 bootstrap replicates.

Results

Phylogenetic analysis based on COI

In this study, we acquired two COI sequences, whose length, excluding primers, was 658 bp each. The genetic divergence value between male and female of *Laevifacies quadrialata* sp. nov. is 0.9%; however, the interspecific K2P genetic divergence among *Laevifacies quadrialata* sp. nov. and other species ranged from 10.4 to 13.1%.

The ML phylogenetic tree (Figure 1) revealed that male and female of *Laevifacies quadrialata* sp. nov. grouped together with a high support value (MLB = 100).

Taxonomy

Subfamily Polyzosteriinae Tepper, 1893

Key to Species of Polyzosteriinae in China

1	Sexual dimorphism present. Body small; tegmina and hind wings vestigial in
	male; tegmina vestigial and hind wings absent in female
_	Sexes similar. Body large; tegmina vestigial and hind wings absent2
2	Terga and abdomen uniformly dark reddish brown to black
	Melanozosteria nitida
_	Margin of terga with continuous and broad yellow stripes, the middle black;
	sometimes abdomen with continuous or discontinuous yellow stripes
	Melanozosteria soror

Laevifacies gen. nov. http://zoobank.org/EC93B8A9-1413-4EB1-B139-63AF641FD6E3

Type species. *Laevifacies quadrialata* sp. nov. here designated.

Generic diagnosis. Body small to medium, thinner in male, thorax slightly broader than abdomen. Surface smooth and shining. Pronotum slightly semicircular, vertex barely exposed. Male with vestigial tegmina and hind wings on mesonotum and metanotum respectively, both nearly triangular; female only with vestigial tegmina, its shape similar to that of male, without hind wings. Legs strong but short, coxae with

Polyzosteriinae Tepper, 1893: 32; Princis 1950: 170; Princis 1960: 447; Princis 1966: 561; McKittrick 1964: 66; Mackerras 1965a: 841; Rentz 2014: 121.

Tal	ble	1. St	pecies	used	in	this	stud	y

Family		Species	Accession number	Reference
Outgroups	Mantidae	Mantis religiosa	KM529415	Hebert et al. 2015 (Unpublished)
		Mantis religiosa	KR148854	Hebert et al. 2016
Ingroups	Blattidae	Laevifacies quadrialata sp. nov.	MK798103	
		Laevifacies quadrialata sp. nov.	MK798104	
		Periplaneta australiasiae	KX640825	Ma et al. 2017
		Shelfordella lateralis	KU684413	Cheng et al. 2016
		Neostylopyga rhombifolia	KP986425	Legendre et al. 2015
		Hebardina concinna	KF640073	Yue et al. 2014
		Methana parva	KP986422	Legendre et al. 2015
		Angustonicus lifou	KP986393	Legendre et al. 2015



Figure 1. Maximum likelihood (ML) tree derived from COI gene analysis with 1000 bootstrap replicates. Number above branch indicates MLB.

punctation, front femora Type A_2 . Mid and hind metatarsus with strong spines, claws symmetrical. Cerci strong, short and symmetrical. Styli long and symmetrical. Supraanal plate in male short, triangular; subgenital plate broad and short, slightly quadrilateral and symmetrical. L1 divided into two parts, L3 bifurcated, one branch short, the other one long, R1 nearly claw-like and R2 large, hooked.

Etymology. The name *Laevifacies* is derived from two Latin words *laevis* and *facies*, referring to the smooth and shining surface of terga. The gender of *Laevifacies* is feminine.

Remarks. Based on former studies (Gutiérrez 2013, 2014; Mackerras 1965a, 1965b, 1965c, 1966a, 1966b, 1967a, 1967b, 1968a, 1968b; Rehn and Hebard 1927), the Polyzosteriinae is characterized as follows: species having semicircular pronotum, lobiform vestigial tegmina, angles of T2-T7 produced, tarsi usually short, bare or with hind and sometimes mid metatarsi spiny (Laevifacies with mid and hind metatarsi spiny, while in Mackerras (1968b), Australian species of Blattinae and Polyzosteriinae from other Blattidae with all metatarsi spiny), large pulvilli and arolia, cerci strong, short and symmetrical, L1 with hollow finger-like projection and sclerotized projection and R1 claw-like and margin with projection; thus, *Laevifacies* is placed in the subfamily Polyzosteriinae. Laevifacies has common features with Melanozosteria, Eurycotis, Leptozosteria, and Platyzosteria, such as body small to large, and shining, usually with vestigial tegmina, angles of T5-T6 acute, T6-T7 with punctation and hind metatarsus usually spiny (Gutiérrez 2013, 2014; Mackerras 1965c, 1968b). Laevifacies is similar to the Melanozosteria and Eurycotis in general appearance, but it can be distinguished from *Melanozosteria* by the following characters: 1) body thin and small in male (Figure 2A, B), while in Melanozosteria, it is broad and large (Figure 4A, B, F, G); 2) the surface of terga smooth (Figure 2A), vs. surface with punctation in *Melanozosteria* (Figure 4A, F); 3) male with vestigial tegmina and hind wings (Figure 2A), but in Melanozosteria only with vestigial tegmina or apterous (Figure 4A, F); 4) the margin of L2d smooth and posterior of L2d finger-like with more small spines (Figure 3A), while in Melanozosteria the margin strongly denticulate and posterior of L2d with acute angle (Figure 4J, M); 5) L3 bifurcated, one short and the other long (Figure 3A), however, L3 unbifurcated or bifurcated with branches of equal length in Melanozosteria (Figure 4J, M); 6) R1 fist-shaped (Figure 3C), while in *Melanozosteria* foot-shaped or finger-shaped (Figure 4K, M); and 7) R2 only with one large and long uniform structure (Figure 3C), while in Melanozosteria, two unequal forked structures present (Figure 4K, M); and it can be distinguished from *Eurycotis* by the following characters: 1) tibiae not specialized, while in *Eury*cotis, one group of which species have smooth surface, uniform black body and lateral tegmina, with highly specialized caudal tibiae; 2) R2 is hook-like, while in Eurycotis R2 is pincer-like. In addition, Eurycotis is restricted to South and North America and Cuba, while Laevifacies gen. nov. is found in East Asia. Laevifacies is similar to the Methana in the following genitalia characteristics, the margin of L2d smooth, R1 as a strongly claw-like sclerotized process, both of L1 have two structures, L1 of Methana has strong finger-like sclerotization and a membranous lobe, while *Laevifacies* has a finger-like membrane and a strongly sclerotized lobe (Figure 3A, B).

Geographical distribution. China (Hainan).



Figure 2. A–K *Laevifacies quadrialata* sp. nov. **A–F**, **I–K** male holotype **A** in dorsal view **B** in ventral view **C** pronotum, in dorsal view **D** head, in ventral view **E** femur, in ventral view **F** tibia, in ventral view **I** fore tarsus, in ventral view **J** mid tarsus, in ventral view **K** hind tarsus, in ventral view. **G–H** female paratype **G** in dorsal view **H** in ventral view. Scale bars: 5 mm (**A–B, G–H**); 1 mm (**C–F, I–K**).

Laevifacies quadrialata sp. nov. http://zoobank.org/CB699FB7-9F08-4830-A85D-948A8A48E629

Diagnosis. Sexual dimorphism. Body small and black. Surface smooth and shining except last two terga with punctation. Tegmina and hind wings vestigial in male, tegmina vestigial and hind wings absent in female. Angles of T2–T7 sharp and protruded. Legs strong. Supra-anal plate short and triangular. Styli long and symmetrical.

Description. Measurements. Male, pronotum: length × width $5.5-6.2 \times 7.9-8.0$ mm, overall length: 15.6-17.7 mm. Female, pronotum: length × width $7.0-7.1 \times 10.5-10.7$ mm, overall length: 17.0-21.0 mm.

Body black, smooth, shining. Vertex and frons black. Clypeus to part of labrum brown to dark brown, maxillary palpi and labial palpi dark brown to black. Eyes black when the specimens are fresh, fading after a long time (Figure 2D). Antennae dark brown with near middle segments and tip segments milky white (Figure 2A, B). Pronotum black, surface smooth and shining (Figure 2C). Tegmina and hind wings black, terga smooth except last two terga with punctation (Figure 2A, G). Sterna and legs dark brown to black. Cerci dark brown to black, apex yellowish brown (Figure 2A, B, G, H).

Size small to medium, female larger than male. Body oval, vertex nearly unexposed (Figure 2A, B, G, H). Ocelli present, small and round (Figure 2D). Pronotum nearly semicircular, anterior margin arc-shaped, posterior margin nearly straight, posterior



Figure 3. A–G male genitalia features from holotype **A** left phallomere, in dorsal view **B** L1 of left phallomere, in dorsal view **C** right phallomere, in dorsal view **D** subgenital plate, in ventral view **E** supraanal plate, in dorsal view **F–G** female genitalia features from paratype **F** subgenital plate, in ventral view **G** supra-anal plate and genitalia, in dorsal view. Abbreviations: **a.a.**, anterior arch; **acc.pr.**, accessory process; **bsv.**, basivalvula; **c.a.**, central apodeme; **i.p.p.**, inner posterior process; **lat.st.IX–X**, laterosternal of the ninth-tenth segment; **L.ph.**, left phallomere; **L1–L3**, parts of left phallomere; **o.p.p.**, outer posterior process; **R.ph.**, right phallomere; **R1–R3**, parts of right phallomere; **v.I–III**, first-third valve; **v.ph.**, ventral phallomere. Scale bars: 1 mm (**A, C–G**); 0.5 mm (**B**).

angles blunt (Figure 2C). Small, vestigial tegmina and hind wings present in male, both extending to notal hind margin, only vestigial tegmina in female (Figure 2A, G); angles of T2–T7 sharp and protruded, sterna smooth and shining (Figure 2A, B, G, H). Legs strong, fore coxae with punctation; front femora Type A₂ (Figure 2E); mid and hind metatarsus with a row of spines; hind metatarsus fairly long with pulvillus which occupies nearly one-quarter of its length, remainder of surface with hair, claws moderately symmetrical and unspecialized (Figure 2E, F, I, K). Male: supra-anal plate short, triangular, divided into two round lobes (Figure 3E); subgenital plate broad and

short, posterior margin round; styli long and symmetrical (Figure 3D). Cerci symmetrical and strong, with indistinct segmentation, ends sharp (Figure 3E). Female: supra-anal plate with higher sclerotization (Figure 3G).

Male genitalia. Left phallomere consisting of three parts: L1, L2, and L3. L1 with two parts L1a and L1b, L1a with membranous finger-like projection; L1b with sclerotized projection. L2 consisting of L2d and L2v, L2d strongly sclerotized in anterior part, the posterior part with finger-like and with more small spines; L2va simple and broad, L2vb sclerotized and the posterior with a spinous projection. L3 with a simple hook, elongate to the right and bifurcated (Figure 3A, B). Right phallomere consisting of R1, R2, and R3. R1 large, claw-like, right margin with a prominent spine; R2 large, curved hook-like, the base strong and gradually becoming thinner, bent to the right; R3 large and cucullate, highly sclerotized (Figure 3C).

Female genitalia. The first valve (v.I) long, slightly broad and crescent-shaped, terminal membranous; the second valve (v.II) small, flaky and obscured by the v.I; the third valve (v.III) broader than v.I, terminal membranous; paraprocts (pp.) symmetrical and the middle concave; the middle of anterior arch (a.a.) concave; basivalvula (bsv.) trapezoidal (Figure 3G); inner posterior process of the laterosternal shelf (i.p.p.) divided in two parts, which are connected by hairy membrane; outer posterior process of the laterosternal shelf (o.p.p.) symmetrical, terminal with hairs (Figure 3F).

Material examined. HOLOTYPE: male, CHINA, Hainan Prov., Baisha, Yinggeling Nature Reserve, 20-VIII-2010, Guo Zheng leg. PARATYPES: 1 male, same data as holotype; 1 male, Hainan Prov., Mt. Wuzhishan, 18-21-V-2014, Shunhua Gui, Xinran Li & Jianyue Qiu leg.; 1 male, Hainan Prov., Diaoluoshan, 18-IV-2015, Lu Qiu & Qikun Bai leg. (GenBank accession number: MK798103); 2 females, Hainan Prov., Lingshui, Mt. Diaoluoshan, 22-V-2014, Jianyue Qiu, Xinran Li & Shunhua Gui leg. (GenBank accession number: MK798104).

Etymology. The species epithet comes from the Latin word *quadrialata* in reference to the male having four triangular vestigial wings.

Remarks. In our study the interspecific K2P genetic divergence among *L. quadrialata* sp. nov. and other cockroach species ranged from 10.4 to 13.1%. But the genetic divergence value between male and female of *L. quadrialata* sp. nov. is only 0.9%, so we pair them based on their similar morphology combined with this COI data. Sexual dimorphism occurs in *L. quadrialata* sp. nov.: 1) females without hind wings, but males with vestigial hind wings (Figure 2A, 2G); 2) male with narrower body, while female with broader body (Figure 2A, B, G, H).

Geographical distribution. China (Hainan)

Melanozosteria Stål, 1874

Melanozosteria Stål, 1874: 13; Kirby 1904: 129; Shelford 1909: 265; Shelford 1910: 5 (as synonym of *Polyzosteria*); Princis 1966: 569; Mackerras 1968a: 237 (as subgenus); Roth 2003: 167; Rentz 2014: 151.

- *Cutilia* Stål, 1877: 36; Kirby 1904: 134; Shelford 1909: 289; Shelford 1910: 7; Hanitsch 1915: 99; Princis 1949: 10.
- Symtomaptera Tepper, 1893: 106 (as a subgenus of Periplaneta); Kirby 1904: 129; Shelford 1909: 265 (as a synonym of Polyzosteria); Princis 1949: 10 (as a synonym of Melanozosteria).

Melanozosteria nitida Brunner von Wattenwyl, 1865

Diagnosis. Body broad oval and reddish brown to black. Pronotum slightly arched, surface with punctation. Vestigial tegmina sectorial with punctation, separated from mesonotum for nearly whole length, hind wings absent. Surface with punctation. Angles of T2–T7 protruded and sharp. The medial aspects to the styli with stubby and sharp spines.

Redescription. Measurements. Male, pronotum: length \times width 7.4 \times 12.5 mm, overall length: 26.1 mm.

Body uniformly deep reddish brown to black (Figure 4A, B, F, G). Eyes and ocelli yellowish white. Margin of clypeus and labrum dark brown. Vertex and frons black. Antennae brown or black, middle joints creamy-white (Figure 4D). Pronotum, tegmina, abdomen, legs and cerci all uniformly deep reddish brown to black (Figure 4C).

Body large, broad oval and convex, surface shining. Pronotum slightly arched, surface with punctation. Anterior margin of pronotum roundly protruded, and posterior margin straight (Figure 4C). Tegmina vestigial, sectorial, and separated from mesonotum, surface with punctation. Angles of metanotum protruded. Hind wings absent. Surface of all terga shining and with punctation; angles of T2–T7 protruded and sharp, T9 not protruded (Figure 4A, F). Legs short and thick. Fore coxae with slightly punctation; front femora Type A_2 (anterior with two long spines, posterior with many small and slightly equal spines). Tibiae hair-brushes; hind tibiae with a row of spines, hind metatarsus with pulvillus occupying one-quarter to one-third of its length, remainder of ventral surface with spines (Figure 4E). All pulvilli large, claws symmetrical (Figure 4E). Supra-anal plate long, symmetrical and quadrilateral, side edge at gradient, angles of posterior round, the middle of posterior margin concave and with hair. Cerci thick, with blurry segmentation and the terminal segment spinous distally (Figure 4F, G, L). Subgenital plate nearly quadrilateral, short. The medial aspects to the styli with stubby and sharp spines (Figure 4I).

Male genitalia. Left phallomere includes L1, L2, and L3. L1 with three parts (a, b, c). L1a slightly sclerotized, posterior not sclerotized, membranous and blunt. L1b more sclerotized and posterior sharp. L1c anterior slightly sclerotized and posterior blunt membrane. L2 includes L2d and L2v. L2d with a well-sclerotized, strongly denticulate in anterior margin, while the posterior of the sclerite becomes more delicate and ends in a sharp point; L2v usually single, L3 is a simple hook, but the posterior divides into two small forks which resemble an elephant's nose (Figure 4J, 4M). Right phallomere includes R1, R2, and R3. R1 large, elongate, foot-like with broad down-



Figure 4. A–E, I–L *Melanozosteria nitida* from Guangxi, male A in dorsal view B in ventral view C pronotum, in dorsal view D head, in ventral view E tarsus, in ventral view F–H Lectotype of *Melanozosteria nitida*, male F in dorsal view G in ventral view H labels I subgenital plate, in ventral view J left phallomere, in dorsal view K right phallomere, in dorsal view L supra-anal plate, in dorsal view M genitalia of *Melanozosteria nitida* in Mackerras (1968a) F–H provided by H. Bruckner, Natural History Museum Vienna, NOaS Image Collection. Scale bars: 10 mm (A–B, F–G); 1 mm (C–E, I–M).

turned "thumb" and 5–6 strongly denticulate on medial edge, R2a long, fairly broad, tapering slightly towards medial corner; R2b shorter, more strongly sclerotized and tapering to long narrow elongation. R3 with structure of folded sclerite (Figure 4K, 4M).

Materials examined. 1 male, CHINA, Guangxi Prov., Shangsi, Nadang, 15-XI-1958, Dexiang Gu & Jinting Liang leg.

Type specimen examined. Lectotype of *Polyzosteria nitida*, male, Ternate (Natural History Museum Vienna), "Ternate Jeynalle CoII. Br. V. W.", "LECTOTYPE", "LECTOTYPE of *Polyzosteria nitida* Brunn. Selected by KHL Key, 1963."; holotype of *Periplaneta polita*, male, Taiwan (Natural History Museum), "Holotype", "*Periplaneta polita* Walker", "BMNH (E) #878036", presented by Beccaloni (2014).

Remarks. We compared the lectotype of *M. nitida* (from Ternate, Indonesia) with the specimen from Guangxi and found there are minor differences between them: the styli are straight in the Guangxi individual (Figure 4I), but in the lectotype of *M. nitida*, slightly bent (Figure 4F, G). We also compared the genitalia between the Guangxi individual and the illustration in Mackerras (1968a); they share the typical characters of L1b spinous projection and serration along the margin of L2d, but they are also different in the following characteristics: 1) the terminal of L3 divided into two small forks, which resemble an elephant's nose in the Guangxi individual (Figure 4]), while in the Mackerras (1968a) individual, L3 has one blunt hook (Figure 4M); 2) L2v broad and sclerotized, and posterior of L3 membranous in the Guangxi individual (Figure 4]), while L2v thin, long and with sharp sclerotized terminus in the Mackerras (1968a) individual (Figure 4M). And the variation of supra-anal plate between samples from Queensland and New Guinea were treated as intraspecific differences in different locations (Mackerras 1968a). Considering Mackerras (1968a) also recorded that the *M. nitida* is a widely distributed tropical species from Taiwan, Malaya, Moluccas, and Philippines, and due to our specimens being inadequate, the minor difference in the Guangxi individual and the lectotype of *M. nitida* are temporarily considered as the intraspecific differences of different populations.

Geographical distribution. Australia, Philippines, Malaysia, New Guinea, New Caledonia, New Zealand, China, Thailand.

Discussion

Almost all members in the Polyzosteriinae are brachypterous or apterous (excepting the tribe Methanini), and display high developmental stochasticity (Rentz 2014). The Australian Polyzosteriinae exhibit the best examples of aposematic coloration. They are often being metallically colored, or spotted and barred with bright orange, red, or yellow markings (Rentz 1996; Roach and Rentz 1998). When disturbed, they may first display a warning signal before resorting to defensive measures (Bell et al. 2007). However, *Laevifacies quadrialata* sp. nov. did not attract our attention due to their bland appearance and life in a hidden habitat (usually hidden in bushes, Lu Qiu, pers. obs.), even with sexual dimorphism. Sexual dimorphism is very common in cockroaches, some of which beinng

so extreme that it is a challenge for taxonomists to match the two sexes (Roth 1992). In this study, sexual dimorphism is revealed for the first time in Polyzosteriinae on the basis of COI data, and exhibits mainly in the body size and the vestigial hind wings.

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