

Fraction of Inspired Oxygen With Low-Flow Versus High-Flow Devices: A Simulation Study

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Abstract

Purpose: The fraction of inspired oxygen while administering oxygen to patients must be measured as it represents the alveolar oxygen concentration, which is important from a respiratory physiology viewpoint. Therefore, the purpose of this study was to compare the fractions of inspired oxygen obtained through different oxygen delivery devices.

Methods: A simulation model of spontaneous respiration was used. The fractions of inspired oxygen obtained through low- and high-flow nasal cannulas and a simple oxygen mask were measured. The fraction of inspired air was measured every second for 30 s after 120 s of oxygen administration. This was measured three times under each condition.

Results: With a low-flow nasal cannula, airflow reduced both the intratracheal fraction of inspired oxygen and extraoral oxygen concentration, indicating that exhalatory respiration occurred during rebreathing and may be involved in increasing the intratracheal fraction of inspired oxygen.

Conclusion: Oxygen administration during expiratory flow may lead to an increased oxygen concentration in the anatomical dead space, which may be involved in the increase in the fraction of inspired oxygen. With a high-flow nasal cannula, a high fraction of inspired oxygen can be achieved even at a flow rate of 10 L/min. When determining the optimum amount of oxygen, it is necessary to set an appropriate flow rate for patients and specific conditions without being bound by the fraction of inspired oxygen values alone. It might be difficult to estimate the fraction of inspired oxygen while using a low-flow nasal cannula and simple oxygen mask in clinical situations.

Categories: Anesthesiology, Emergency Medicine, Public Health

Keywords: fraction of inspired oxygen, respiratory system, respiration, oxygen, cannula

Introduction

Oxygen administration, performed in both acute and chronic phases of respiratory failure, is a common procedure in clinical medicine. The various modes of oxygen administration include an intubation tube, nasal cannula, oxygen mask, reservoir face mask, venturi mask, and high-flow nasal cannula (HFNC) [1-5]. The fraction of inspired oxygen (FiO₂) represents the percentage of oxygen in the inspired air that is involved in alveolar gaseous exchange. The degree of oxygenation (P/F ratio) is the ratio of partial pressure of oxygen in arterial blood (PaO₂) to FiO₂. Although the diagnostic utility of the P/F ratio remains controversial, it is a widely used clinical indicator of oxygenation [6-8]. Therefore, it is clinically important to know the FiO₂ value when administering oxygen to patients.

During intubation, FiO₂ can be accurately measured using an oxygen monitor that includes a ventilator circuit, whereas when employing nasal cannulas and oxygen masks for oxygen delivery, only an “estimated value” of the FiO₂ can be measured based on the inspiratory time. This “estimated value” is the ratio of the amount of oxygen supplied to the tidal volume. However, it does not consider some factors from the viewpoint of respiratory physiology. Studies have shown that multiple factors affect FiO₂ measurements [2,3]. Although oxygen administration during expiratory flow may lead to an increased oxygen concentration in the anatomical dead space, such as the oral cavity, pharynx, and trachea, there are no reports on this topic in the current literature. Nevertheless, some clinicians argue that these factors are less important in practice, and the “estimated value” is sufficient to overcome clinical challenges.

In recent years, HFNC has attracted particular attention in emergency medicine and intensive medical care [9]. HFNC achieves a high FiO₂ and oxygen flow, and has two major advantages - pharyngeal dead space washout and reduction in nasopharyngeal resistance which should not be ignored when considering oxygen administration [10,11]. Additionally, it may be necessary to assume that the measured FiO₂ is the oxygen concentration within the respiratory tract or alveoli because the alveolar oxygen concentration during

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inspiration is important from the viewpoint of the P/F ratio.

Oxygen delivery methods other than intubation are commonly employed in routine clinical practice. Therefore, it is important to gather more evidence on FiO_2 , as measured using these oxygen delivery devices, to prevent unnecessary over-administration of oxygen and to understand the respiratory status during oxygen administration in terms of safety. However, measuring FiO_2 in the trachea of the human body is difficult. Some researchers have attempted to simulate FiO_2 with a spontaneous respiration model [4,12,13]. Thus, in this study, we aimed to measure FiO_2 using a simulation model of spontaneous respiration.

Materials And Methods

Spontaneous breathing simulation model

This is a simulation study and ethical approval was not required as this study did not involve human subjects. To simulate spontaneous breathing, we prepared a model of spontaneous breathing with reference to the model created by Hsu et al. (Figure 1) [12]. A ventilator from an anesthesia apparatus (Fabius Plus; Lübeck, Germany: Draeger, Inc.) and test lungs (Dual adult TTL; Grand Rapids, MI: Michigan Instruments, Inc.) were prepared to reproduce spontaneous respiration. The two devices were manually linked with a rigid metal strap. One bellow (the driving side) of the test lung was connected to a ventilator. The other bellow (the passive side) of the test lung was connected to the “oxygen administration model.” Once the ventilator delivered fresh gas to the lung (the driving side), the bellow expanded, forcibly pulling the other bellow (passive side). This movement drew in gas through the trachea of the manikin, thus simulating spontaneous breathing.

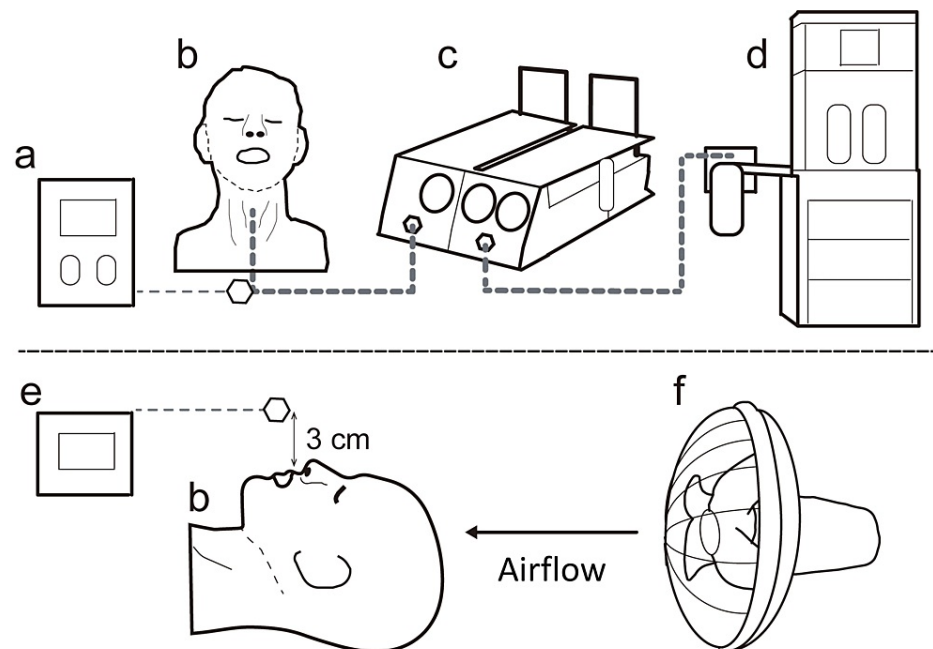


FIGURE 1: Experimental schema and actual experiment.

(a) Oxygen monitor, (b) human manikin, (c) test lungs, (d) anesthesia apparatus, (e) oxygen monitor, and (f) electric fan.

The ventilator settings were as follows: tidal volume, 500 mL; respiration rate, 10 breaths/min; and inspiratory-to-expiratory ratio (I:E ratio), 1:2 (respiration time = 1 s). The compliance setting of the test lungs was set to 0.5 for the experiment.

Oxygen administration model

An oxygen monitor (MiniOx 3000; Pittsburgh, PA: Medical Services of America, Inc.) and human manikin (MW13; Kyoto, Japan: Kyoto Kagaku Co., Ltd.) were used for the oxygen administration model. Pure oxygen was administered at 1, 2, 3, 4, and 5 L/min, and each FiO_2 was measured. For HFNC (MaxVenturi; Coleraine, Northern Ireland: Armstrong Medical), oxygen/air mixed gas was administered at 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, and 60 L, and FiO_2 was evaluated under each condition. For HFNC, experiments were conducted at oxygen concentrations of 45%, 60%, and 90%.

Evaluation of the Effect of Exhaled Respiration During Nasal Oxygen Administration

The extraoral oxygen concentration was measured (BSM-6301; Tokyo, Japan: Nihon Kohden Corp.) 3 cm above the maxillary incisors when oxygen was administered via a nasal cannula (Finefit; Osaka, Japan: Japan Medicalnext Co.) (Figure 1). Through the nasal cannula, air was blown from the head side of the manikin using an electric fan (HEF-33YR; Tokyo, Japan: Hitachi) to eliminate expiratory rebreathing, and FiO_2 was measured after 2 min.

FiO_2 Measurement

FiO_2 was measured every second for 30 s, after 120 s of oxygen administration. After each measurement, the manikin and laboratory were ventilated. FiO_2 was measured three times under each condition. The experiment began after calibrating each measuring instrument.

Estimated FiO_2 Value (Nasal Cannula)

Traditionally, nasal cannula oxygen inhalation is assessed to allow for the measurement of FiO_2 . The calculation method used in this experiment changed depending on the content of spontaneous breathing (Table 1). The estimated value was calculated based on the respiratory conditions (tidal volume: 500 mL; respiration rate: 10 breaths/min; inspiratory-to-expiratory ratio {I:E ratio} = 1:2) set in the anesthesia device.

□	Intratracheal FiO_2				Estimated value	Extraoral FiO_2			
	Without airflow		During airflow			Without airflow		During airflow	
	Average	SD	Average	SD		Average	SD	Average	SD
1 L	48.3	± 1.5	25.3	± 1.7	26.3	22.5	± 0.4	20.3	± 0.4
2 L	61.6	± 0.5	40.1	± 2.0	31.5	23.7	± 1.4	20.6	± 0.3
3 L	69.4	± 0.5	52.7	± 1.3	36.8	25.1	± 1.1	20.7	± 0.4
4 L	71.6	± 0.4	58.7	± 0.6	42.1	25.0	± 1.5	20.6	± 0.7
5 L	71.4	± 0.3	62.4	± 1.2	47.3	24.2	± 2.0	20.8	± 0.3

TABLE 1: Results of low-flow nasal cannula (LFNC).

The "estimated value" is calculated at each oxygen flow rate. For LFNC oxygen administration, a nasal cannula was used.

SD: standard deviation; FiO_2 : fraction of inspired oxygen.

Statistical analysis

All analyses were performed using Origin software (Northampton, MA: OriginLab Corporation). The results are expressed as mean ± standard deviation (SD) for the number of tests (N) [12]. We have rounded up at the second decimal point in all results.

Results

FiO_2 measurement for low-flow nasal cannula

To calculate the "estimated value," the oxygen taken into the lungs in a single inspiration was the amount of oxygen within the nasal cannula and the rest was outside air. Therefore, when the respiration time was 2 s, the oxygen administered via the nasal cannula in 2 s was 1000/30 mL. The oxygen dose taken from the outside air was 21% of the tidal volume (1000/30 mL). The final FiO_2 was the amount of oxygen supplied to the tidal volume. Therefore, the "estimated value" of FiO_2 could be calculated by dividing the total amount of oxygen consumed by the tidal volume.

Before every measurement, the calibration of the intratracheal oxygen monitor was 20.8% and that of the extraoral oxygen monitor was 21%. Table 1 shows the mean FiO_2 values of the LFNC at each flow rate. These values were 1.5-1.9 times higher than the "estimated value" (Table 1). The extraoral oxygen concentration

was higher than that of room air (21%). The mean values decreased before the introduction of airflow from an electric fan. These values were similar to the “estimated value.” During airflow while the extraoral oxygen concentration was approximated to that of the room air, the intratracheal FiO_2 values were higher than the “estimated value” at more than 2 L/min. Regardless of the presence or absence of airflow, as the flow rate increased, the FiO_2 difference decreased (Figure 2).

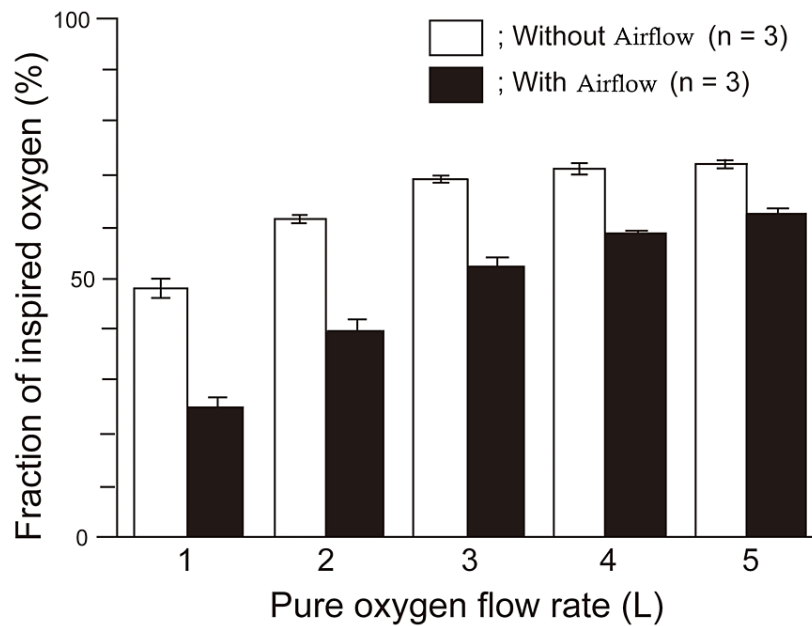


FIGURE 2: FiO_2 with/without airflow in LFNC, the FiO_2 differed with/without airflow.

LFNC: low-flow nasal cannula; FiO_2 : fraction of inspired oxygen

FiO_2 measurement for a simple oxygen mask

Table 2 shows the mean FiO_2 values of the simple oxygen mask (Ecolite oxygen mask; Osaka, Japan: Japan Medicalnext Co., Ltd.) at each oxygen concentration. These values increased as the oxygen concentration increased (Table 2). LFNC had a higher FiO_2 than did the simple oxygen mask at the same oxygen flow rate. The difference of FiO_2 was approximately 11-24% at 1-5 L/min.

□	Intratracheal FiO ₂	
	Average	SD
1 L	30.9	± 0.9
2 L	37.7	± 0.7
3 L	47.0	± 1.9
4 L	54.9	± 1.2
5 L	60.6	± 1.2
6 L	68.6	± 0.5
7 L	71.6	± 0.6
8 L	74.7	± 0.9
9 L	76.8	± 0.6
10 L	79.6	± 0.6

TABLE 2: FiO₂ measurement of the simple oxygen mask.

For oxygen administration, a simple oxygen mask was used.

SD: standard deviation; FiO₂: fraction of inspired oxygen

FiO₂ measurement for high-flow nasal cannula

Table 3 shows the mean value of FiO₂ for HFNC at each flow rate and oxygen concentration. These values were close to the set oxygen concentrations, regardless of whether the flow rate was low or high (Table 3).

□	45%		60%		90%	
	Average	SD	Average	SD	Average	SD
10 L	41.1	± 0.8	54.1	± 1.5	88.6	± 1.4
15 L	42.6	± 0.4	56.7	± 0.4	89.5	± 0.6
20 L	42.5	± 0.2	56.9	± 1.2	89.6	± 1.0
25 L	42.3	± 0.2	57.1	± 1.8	89.6	± 0.7
30 L	42.1	± 0.1	57.2	± 2.1	89.3	± 1.0
35 L	42.1	± 0.1	57.2	± 1.8	89.3	± 1.1
40 L	41.8	± 0.3	57.3	± 1.3	89.2	± 0.6
45 L	41.8	± 0.3	57.1	± 1.4	88.6	± 0.4
50 L	41.7	± 0.2	57.1	± 1.3	88.2	± 0.2
55 L	41.5	± 0.2	57.2	± 1.3	88.0	± 0.5
60 L	41.4	± 0.3	57.2	± 1.4	87.7	± 0.3

TABLE 3: FiO₂ measurement with HFNC at each oxygen concentration.

For HFNC, a high-flow system and flow driver were prepared.

HFNC: high-flow nasal cannula; SD: standard deviation; FiO₂: fraction of inspired oxygen.

Discussion

While using the LFNC, the intratracheal FiO_2 values were higher than the “estimated values” and extraoral FiO_2 values were higher than those at room air. Flowing air was found to reduce both intratracheal FiO_2 and extraoral FiO_2 . These results indicate that exhalatory respiration occurred during rebreathing with LFNC. Regardless of the presence or absence of airflow, as the flow rate increased, the FiO_2 difference decreased. This result showed that another factor might be related to the increased intratracheal FiO_2 . In addition, they also indicated that oxygen administration increased the oxygen concentration in the anatomical dead space, which may be involved in the increase in FiO_2 [2]. Conventional wisdom purports that LFNC does not cause expiratory rebreathing. It is speculated that this may have a significant effect on the difference between the measured value from the nasal cannula and the “estimated value.”

FiO_2 was lower for a simple mask than for a nasal cannula at low flow rates of 1-5 L/min, possibly because the oxygen concentration does not increase easily as the mask part becomes an anatomical dead space. In more than 5 L/min condition, flow oxygen minimized room air dilution, making FiO_2 steady [12]. At less than 5 L/min, a low point was noted in the FiO_2 value because of room air dilution and dead space rebreathing [12]. Indeed, the accuracy of the oxygen flowmeter can be highly variable. A MiniOx 3000 was used to monitor oxygen concentration; however, this device does not possess an adequate temporal resolution to measure variation in oxygen concentration over the breath (manufacturer quotes 20 s for a 90% response). This requires an oxygen monitor with much faster time response.

In actual clinical practice, the morphology of the nasal cavity, oral cavity, and pharynx differs among individuals, and the FiO_2 value may differ from the results obtained in this study. In addition, respiratory conditions vary among patients, and higher oxygen consumption will result in lower oxygen levels of exhalatory respiration. These conditions may result in a decrease in FiO_2 values. Therefore, it is difficult to estimate a reliable FiO_2 while using an LFNC and simple oxygen mask in actual clinical situations. However, this experiment suggests that the concept of anatomical dead space and rebreathing exhalatory respiration probably influences FiO_2 . Given this finding, it is possible that FiO_2 may be sufficiently increased, even at low flow rates, depending on conditions rather than the “estimated values.”

According to the British Thoracic Society guidelines, clinicians should prescribe oxygen according to a target saturation range and monitor whether the patient’s condition is maintained within the target saturation range [14]. Although the “estimated values” of FiO_2 were very low in this study, it is possible to achieve an actual FiO_2 that is higher than the “estimated value” depending on the patient’s condition.

While using HFNC, the FiO_2 values were close to the set oxygen concentrations, regardless of whether the flow rate was low or high. The results of this study indicate that high FiO_2 may have been achieved even at a flow rate of 10 L/min. Similar studies have suggested that FiO_2 did not change between 10 and 30 L [12,15]. Reportedly, the high flow rate with HFNC eliminates the need to consider anatomical dead space [2,16]. It is possible that the anatomical dead space was washed out with an oxygen flow rate of more than 10 L/min. Dysart et al. posited that the principal mechanism of action of HFT may be flushing of the dead space of the nasopharyngeal cavity, thereby reducing overall dead space and increasing the fraction of minute ventilation that is alveolar ventilation [17].

A previous study of HFNC measured FiO_2 of the nasopharynx with a catheter, but the FiO_2 was lower than that in this experiment [15,18-20]. Ritchie et al. reported that the calculated FiO_2 approached 0.60 as gas flow rates increased above 30 L/min during nose breathing [15]. In practice, a flow rate of 10-30 L/min or higher is required while using HFNC. Because of the characteristics of HFNC, the conditions in the nasal cavity have a considerable effect, and HFNC is often started at high flow rates. If respiratory conditions improve, it may also be necessary to reduce flow rates, as FiO_2 may be sufficient.

These results are based on simulations and do not imply that the findings of FiO_2 can be applied directly to actual patients. However, based on these results, in case of devices other than intubation or HFNC, it can be expected that the FiO_2 value will change significantly depending on the conditions. When oxygen is administered using an LFNC or simple oxygen mask in clinical situations, treatment is often evaluated by only the value of “oxygen saturation of the peripheral artery” (SpO_2) using a pulse oximeter. In case the patient is anemic, it is suggested that the patient cannot be strictly managed without considering the values of SpO_2 , PaO_2 , and arterial oxygen content. In addition, Downs et al. and Beasley et al. have argued that unstable patients may actually be placed at risk with the precautionary use of high-concentration oxygen therapy [21-24]. During physiological deterioration, a patient who is administered high-concentration oxygen therapy would have a high pulse oximeter reading, which would mask the progressive decline in the P/F ratio, and therefore, the staff may not be alerted at the right time, leading to impending deterioration requiring mechanical support. It was previously believed that a high FiO_2 is protective and provides patients

a margin of safety; however, the theory is not applicable in a clinical situation [14].

Therefore, care should be taken even when oxygen is administered perioperatively or in the early stages of respiratory failure. The findings of the study suggest that an accurate measurement of FiO_2 cannot be obtained, except with intubation or HFNC. When using LFNC or a simple oxygen mask, it is necessary to ensure that oxygen is administered prophylactically to prevent mild respiratory failure. These devices may not be suitable when the respiratory status needs to be rigorously evaluated, especially in cases where FiO_2 results are crucial. Even at low flow rates, FiO_2 increases with an increase in oxygen flow and may mask respiratory failure. Moreover, even when performing postoperative management with SpO_2 , it is desirable to have as low a flow rate as possible. This is necessary for the early detection of respiratory failure. A high oxygen flow increases the risk of early detection failure. It is desirable to determine the oxygen dose after confirming what vitals will be improved by administering oxygen. Based on the results of this study alone, it is not advisable to change the concept of oxygen administration. However, we believe that the novel ideas presented in this study should be considered in terms of the approach taken in clinical practice. In addition, when determining the amount of oxygen proposed by the guidelines, it is necessary to set an appropriate flow rate for patients without being bound by the FiO_2 value measured using the conventional inspiratory flow rate.

We suggest that the concept of FiO_2 should be reconsidered, considering the range of applications of oxygen therapy and the clinical setting, as FiO_2 is an indispensable parameter for guiding oxygen administration. However, there were few limitations to this study. If FiO_2 in the human trachea can be measured, more accurate values can be obtained. However, it is currently difficult to make such measurements without being invasive. Further research should be carried out with non-invasive measuring devices developed in the future.

Conclusions

In this study, we measured intratracheal FiO_2 with a simulation model of spontaneous respiration using an LFNC, a simple oxygen mask, and an HFNC. Oxygen administration during expiratory flow may lead to an increased oxygen concentration in the anatomical dead space, which may be involved in an increase in the fraction of inspired oxygen. With an HFNC, a high fraction of inspired oxygen can be achieved, even at a flow rate of 10 L/min. When determining the optimum amount of oxygen, it is necessary to set an appropriate flow rate for patients and specific conditions, without being bound by the fraction of inspired oxygen values alone. It might be challenging to estimate the fraction of inspired oxygen while using an LFNC and simple oxygen mask in clinical situations.

The study results suggest that exhalatory respiration is involved in the increase in intratracheal FiO_2 with LFNC. When determining the amount of oxygen proposed by the guidelines, it is necessary to set an appropriate flow rate for patients without being bound by the FiO_2 values, which were measured using the conventional inspiratory flow rate.

Additional Information

Disclosures

Human subjects: All authors have confirmed that this study did not involve human participants or tissue.

Animal subjects: All authors have confirmed that this study did not involve animal subjects or tissue.

Conflicts of interest: In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

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References

1. Boumphrey SM, Morris EA, Kinsella SM: 100% inspired oxygen from a Hudson mask—a realistic goal? . *Resuscitation*. 2003, 57:69-72. [10.1016/s0300-9572\(02\)00436-7](https://doi.org/10.1016/s0300-9572(02)00436-7)
2. Leigh JM: Variation in performance of oxygen therapy devices . *Anaesthesia*. 1970, 25:210-22. [10.1111/j.1365-2044.1970.tb00192.x](https://doi.org/10.1111/j.1365-2044.1970.tb00192.x)
3. Waldau T, Larsen VH, Bonde J: Evaluation of five oxygen delivery devices in spontaneously breathing subjects by oxygraphy. *Anaesthesia*. 1998, 53:256-63. [10.1046/j.1365-2044.1998.00318.x](https://doi.org/10.1046/j.1365-2044.1998.00318.x)
4. Wagstaff TA, Soni N: Performance of six types of oxygen delivery devices at varying respiratory rates .

- Anaesthesia. 2007, 62:492-503. [10.1111/j.1365-2044.2007.05026.x](https://doi.org/10.1111/j.1365-2044.2007.05026.x)
5. Goudra BG, Chandramouli M, Singh PM, Sandur V: Goudra ventilating bite block to reduce hypoxemia during endoscopic retrograde cholangiopancreatography. *Saudi J Anaesth*. 2014, 8:299-301. [10.4103/1658-354X.130756](https://doi.org/10.4103/1658-354X.130756)
 6. Maiolo G, Collino F, Vasques F, et al.: Reclassifying acute respiratory distress syndrome. *Am J Respir Crit Care Med*. 2018, 197:1586-95. [10.1164/rccm.201709-1804OC](https://doi.org/10.1164/rccm.201709-1804OC)
 7. Ranieri VM, Rubenfeld GD, Thompson BT, et al.: Acute respiratory distress syndrome: the Berlin Definition. *JAMA*. 2012, 307:2526-33. [10.1001/jama.2012.5669](https://doi.org/10.1001/jama.2012.5669)
 8. Ray S, Rogers L, Pagel C, Raman S, Peters MJ, Ramnarayan P: PaO₂/FiO₂ ratio derived from the SpO₂/FiO₂ ratio to improve mortality prediction using the pediatric index of mortality-3 score in transported intensive care admissions. *Pediatr Crit Care Med*. 2017, 18:151-6. [10.1097/PCC.0000000000001075](https://doi.org/10.1097/PCC.0000000000001075)
 9. Lee JH, Rehder KJ, Williford L, Cheifetz IM, Turner DA: Use of high flow nasal cannula in critically ill infants, children, and adults: a critical review of the literature. *Intensive Care Med*. 2013, 39:247-57. [10.1007/s00134-012-2743-5](https://doi.org/10.1007/s00134-012-2743-5)
 10. Gotera C, Díaz Lobato S, Pinto T, Winck JC: Clinical evidence on high flow oxygen therapy and active humidification in adults. *Rev Port Pneumol*. 2013, 19:217-27. [10.1016/j.rppneu.2013.05.005](https://doi.org/10.1016/j.rppneu.2013.05.005)
 11. Kernick J, Magarey J: What is the evidence for the use of high flow nasal cannula oxygen in adult patients admitted to critical care units? A systematic review. *Aust Crit Care*. 2010, 23:53-70. [10.1016/j.aucc.2010.01.001](https://doi.org/10.1016/j.aucc.2010.01.001)
 12. Hsu WC, Orr J, Lin SP, Yu L, Tsou MY, Westenskow DR, Ting CK: Efficiency of oxygen delivery through different oxygen entrainment devices during sedation under low oxygen flow rate: a bench study. *J Clin Monit Comput*. 2018, 32:519-25. [10.1007/s10877-017-0023-1](https://doi.org/10.1007/s10877-017-0023-1)
 13. Lamb K, Piper D: Southmedic OxyMask™ compared with the Hudson RCI® non-rebreather mask™: safety and performance comparison. *Can J Respir Ther*. 2016, 52:13-5.
 14. O'Driscoll BR, Howard LS, Earis J, Mak V: BTS guideline for oxygen use in adults in healthcare and emergency settings. *Thorax*. 2017, 72:ii1-ii90. [10.1136/thoraxjnl-2016-209729](https://doi.org/10.1136/thoraxjnl-2016-209729)
 15. Ritchie JE, Williams AB, Gerard C, Hockey H: Evaluation of a humidified nasal high-flow oxygen system, using oxygraphy, capnography and measurement of upper airway pressures. *Anaesth Intensive Care*. 2011, 39:1103-10. [10.1177/0310057X1103900620](https://doi.org/10.1177/0310057X1103900620)
 16. Frizzola M, Miller TL, Rodriguez ME, et al.: High-flow nasal cannula: impact on oxygenation and ventilation in an acute lung injury model. *Pediatr Pulmonol*. 2011, 46:67-74. [10.1002/ppul.21326](https://doi.org/10.1002/ppul.21326)
 17. Dysart K, Miller TL, Wolfson MR, Shaffer TH: Research in high flow therapy: mechanisms of action. *Respir Med*. 2009, 103:1400-5. [10.1016/j.rmed.2009.04.007](https://doi.org/10.1016/j.rmed.2009.04.007)
 18. Oberg B, Waldau T, Larsen VH: The effect of nasal oxygen flow and catheter position on the accuracy of end-tidal carbon dioxide measurements by a pharyngeal catheter in unintubated, spontaneously breathing subjects. *Anaesthesia*. 1995, 50:695-8. [10.1111/j.1365-2044.1995.tb06096.x](https://doi.org/10.1111/j.1365-2044.1995.tb06096.x)
 19. Lodeserto FJ, Lettich TM, Rezaie SR: High-flow nasal cannula: mechanisms of action and adult and pediatric indications. *Cureus*. 2018, 10:[10.7759/cureus.3639](https://doi.org/10.7759/cureus.3639)
 20. Şancı E, Coşkun FE, Bayram B: Impact of high-flow nasal cannula on arterial blood gas parameters in the emergency department. *Cureus*. 2020, 12:[10.7759/cureus.10516](https://doi.org/10.7759/cureus.10516)
 21. Downs JB: Has oxygen administration delayed appropriate respiratory care? Fallacies regarding oxygen therapy. *Respir Care*. 2003, 48:611-20.
 22. Beasley R, Aldington S, Robinson G: Is it time to change the approach to oxygen therapy in the breathless patient?. *Thorax*. 2007, 62:840-1. [10.1136/thx.2006.068866](https://doi.org/10.1136/thx.2006.068866)
 23. Niesters M, Mahajan RP, Aarts L, Dahan A: High-inspired oxygen concentration further impairs opioid-induced respiratory depression. *Br J Anaesth*. 2013, 110:837-41. [10.1093/bja/aes494](https://doi.org/10.1093/bja/aes494)
 24. Fu ES, Downs JB, Schweiger JW, Miguel RV, Smith RA: Supplemental oxygen impairs detection of hypoventilation by pulse oximetry. *Chest*. 2004, 126:1552-8. [10.1378/chest.126.5.1552](https://doi.org/10.1378/chest.126.5.1552)