

## An overview of North America's diminutive freshwater fish fauna

Micah G. Bennett\* and Kevin W. Conway\*

Based on a review of the literature, we compile a table of North American freshwater fishes with maximum standard length (SL) less than or equal to 50 mm and information on size at sexual maturity. Using previously published criteria for miniature fishes (<26 mm maximum SL, or those that mature at SL <20 mm), we found evidence for seven miniature fishes on the North American continent: *Elassoma alabamae*, *E. zonatum*, *Leptolucania ommata*, *Heterandria formosa*, *Megupsilon aporus*, *Cyprinodon macularius* and *Notropis saladonis*, the smallest being *E. alabamae* (Spring Pygmy Sunfish, maximum size 25.5 mm SL). We compare the number of miniature species in North America to those from Africa, South Asia and South America and discuss patterns of distribution, habitat preference and imperilment amongst North America's smallest freshwater fishes.

### Introduction

The North American continent contains one of the most diverse temperate freshwater fish faunas in the World (Abell et al., 2000; Lundberg et al., 2000), with over 1050 described and numerous undescribed species (Page & Burr, 1991; Lundberg et al., 2000). At the turn of the 20<sup>th</sup> century the North American ichthyofauna was credited with the distinction of having some of the "smallest known fishes and at the same time the smallest known vertebrates" (Smith, 1902). In more recent years the North American continent has lost its "smallest vertebrate" status, which has passed back and forth between Indo-Pacific, Australasian and South East Asian countries (Herre, 1929; Winterbottom & Emery, 1981; Watson & Walker, 2004). Among these is the recent description of the cypriniform genus *Paedocypris* (Kottelat et al., 2006), which contains the smallest living verte-

brate species (*P. progenetica*), maturing at sizes under 8 mm in standard length (SL).

Though small-bodied fishes have always been of interest, ichthyologists have only fairly recently began to undertake continent-wide assessments of small-bodied species. Weitzman & Vari (1988) undertook the first such survey documenting the numerous miniature freshwater fish species of South America. They defined miniature fishes as those no greater than 26 mm SL or maturing at sizes smaller than 20 mm SL and identified 85 such species. Costa & Le Bail (1999) added 26 more to the count for South America. Using Weitzman & Vari's criterion, Kottelat & Vidthayanon (1993) provided a list of miniature fishes for south and southeast Asia and, more recently, Conway & Moritz (2006) compiled a similar list for Africa. Besides providing a more thorough knowledge of continental biodiversity, identifying and tabulating the number of small-

\* Department of Biology, Saint Louis University, 3507 Laclede Avenue, St. Louis, MO 63103, USA.  
 E-mail: micahgbennett@yahoo.com, conwaykw@gmail.com

bodied fishes is important to largescale ecological and evolutionary investigations such as those focused on geographic gradients in body size (e.g., Belk & Houston, 2002; Knouft, 2004), phylogenetic trends in body size (e.g., Knouft & Page, 2003; Ruber et al., 2007), life history evolution (e.g., Weitzman & Vari, 1988), and issues of conservation concern, such as size-biased extinction risks (e.g., Olden et al., 2007).

In this paper we provide a list of all North American freshwater fishes with a maximum SL less than or equal to 50 mm in order to assess whether the North American ichthyofauna contains any miniature species of fish (sensu Weitzman & Vari, 1988) and to determine the smallest North American fish species.

### Methods

We conducted surveys of peer-reviewed literature and books on fishes and fish biology using four standard publication databases (Web of Science, Biological Abstracts, Google Scholar, JSTOR) and compiled a table of all freshwater fishes with a maximum standard length (SL) of less than or equal to 50 mm that inhabit North America (including Mexico). This was necessarily an arbitrary cut-off point, but it was chosen in order to include the lower tail of the size distribution of North American freshwater fishes (sensu Knouft & Page, 2003) and to include any miniature species (< 26 mm maximum SL, sensu Weitzman & Vari, 1988). We used the University of Michigan Museum of Zoology online catalogue of the Division of Fishes (UMMZ, 2009) which includes SL information to determine if specimens larger than those documented in the literature survey had been collected for species for which maximum standard lengths had been published. That catalogue also served as a source of information for species for which no literature could be found. Literature information was also used to compile data on the minimum SL at maturity of all species with standard lengths of less than 50 mm in order to assess whether any species included in our survey matured at sizes below 20 mm SL. Lastly, we utilized the most recent status rankings of the American Fisheries Society (Jelks et al., 2008) to examine patterns of imperilment among diminutive fishes.

### Results

Our survey documented 146 species of North American freshwater fishes with a maximum SL less than, or equal to, 50 mm, 6 of which mature at sizes of less than 20 mm SL (Table 1). The pool of diminutive fishes (equal to or below 50 mm SL) covers a broad range of North American fish families but is made up largely of Poeciliidae (32.0 %), Cyprinidae (18.4 %), Cyprinodontidae (17.0 %), and Percidae (14.3 %). Three of these, the Cyprinidae, Percidae and Poeciliidae are the most diverse families of fishes in North America, making up about 26 %, 16 % and 8 %, respectively, of that fauna (Page & Burr, 1991; Lundberg et al., 2000; Jelks et al., 2008), whereas the Cyprinodontidae accounts for a much smaller portion of that fauna (less than 4 %; Jelks et al., 2008). Thus the composition of Table 1 is not merely reflective of the overall pattern of North American freshwater fish diversity. Using the definition of Weitzman & Vari (1988) seven of these 146 species are considered miniature: *Elassoma alabamiae* (Fig. 1), *Leptolucania ommata* (Fig. 2), *Heterandria formosa*, *Megupsilon aporus* (Fig. 3), *Cyprinodon macularius*, *Notropis saladonis* and *Elassoma zontatum*. *Elassoma alabamiae* is the only species considered a miniature based on maximum known size (25.5 mm SL). The remaining miniatures are defined as such based on minimum size at maturity as all achieve maximum standard lengths greater than 26 mm.

### Discussion

In comparison to other continents for which lists of miniature freshwater fishes have been compiled (South America – Weitzman & Vari, 1988; Southeast Asia – Kottelat & Vidthayanon, 1993; Africa – Conway & Moritz, 2006) North America has few truly miniature freshwater fish species. Africa contains 24 miniature freshwater fish species (Conway & Moritz, 2006), South America harbors well over 100 such species (Weitzman & Vari, 1988; Costa & Le Bail, 1999), and Southeast Asia contains over 50 species (Kottelat & Vidthayanon, 1993; Kottelat et al., 2006), compared to only seven species in North America (Table 1). There is currently no published list of European miniature species but based on Kottelat & Freyhof (2007) there appears to be just two: *Economidichthys trichonis* (maximum size 25 mm SL) and

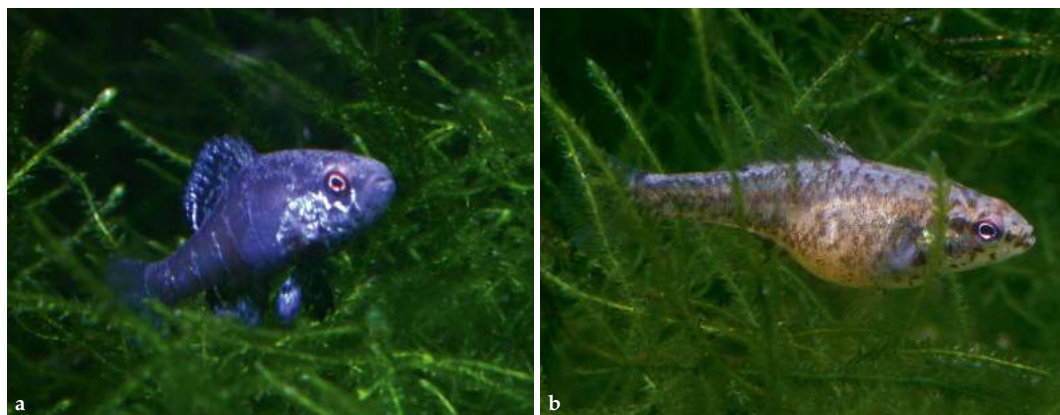


Fig. 1. *Elassoma alabamae*; U.S.A.: Alabama: Limestone County: Beaverdam Creek. a, male; b, female. Photographs courtesy of P. Rakes and J. R. Shute.



Fig. 2. *Leptolucania ommata*; locality unknown. a, male. b, female. Photographs courtesy of P. Loiselle.

**Table 1.** North American freshwater fishes with a maximum known standard length (SL) less than or equal to 50.0 mm. All measurements are in SL unless otherwise stated. Taxa highlighted in bold are miniature species (sensu Weitzman and Vari 1988). Conservation status abbreviations (following Jelks et al. 2008): E, endangered; T, threatened; V, vulnerable; X, extinct; Xn, extirpated in nature; Xp, possibly extinct; –, not assessed.

order – family/species	max SL (mm)	min SL at maturity (mm)	reference (maximum known SL; minimum known SL at maturity)	Conservation Status
<b>Cypriniformes – Cyprinidae</b>				
<i>Cyprinella alvarezdelvillari</i>	43.8	25.0 (♂)	Miller et al., 2005: 110	E
<i>Dionda diaboli</i>	48	?	Miller et al., 2005: 116	E
<i>Dionda erimyzonops</i>	39	21.0 (♂)	Miller et al., 2005: 117; Hubbs & Miller, 1974	–
<i>Evarra tlahuacensis</i>	48	?	Miller et al., 2005: 120	X
<i>Extrarius</i> n. sp. cf. <i>aestivalis</i>	50	?	Gilbert, 1992	rare?
<i>Hybopsis amecae</i>	41	?	Miller et al., 2005:127	–
<i>Hybopsis aulidion</i>	39	?	Miller et al., 2005: 130	–
<i>Iotichthys phlegethontis</i>	41	?	Miller & Behnke, 1985	E
<i>Lythrurus alegnotus</i>	50	?	Boschung & Mayden, 2004	–
<i>Lythrurus snelsoni</i>	43.2	?	Robison, 1985	V
<i>Notropis alborus</i>	50	?	Hubbs & Raney, 1947: 11	–
<i>Notropis ammophilus</i>	'ca.' 48.0	29.0 (♂); 31.0 (♀)	Boschung & Mayden, 2004	–
<i>Notropis anogenus</i>	48	?	Bailey, 1959	T
<i>Notropis buchanani</i>	41	?	Miller et al., 2005: 132	–
<i>Notropis calbazas</i>	26.8	?	Lyons & Mercado-Silva, 2004	E
<i>Notropis chalybaeus</i>	50	?	Boschung & Mayden, 2004	V
<i>Notropis harperi</i>	49	25.0 (♀)	Boschung & Mayden, 2004	–
<i>Notropis melanostomus</i>	38	21	Boschung & Mayden, 2005; Suttkus & Bailey, 1990	T
<i>Notropis ortenburgeri</i>	50	?	UMMZ, 2009	V
<i>Notropis rafinesquei</i>	'ca.' 43.0	22.5	Haag et al., 2007	–
<i>Notropis sabinae</i>	49	38.0 (♀; TL)	Heins, 1981; Williams & Bonner, 2006	–
<b><i>Notropis salaudonis</i></b>	<b>38</b>	<b>19.0 (♂); 21.0 (♀)</b>	Miller et al., 2005: 135; Hubbs & Hubbs, 1958	Xp
<i>Notropis tropicus</i>	41	27.0 (♂)	Miller et al., 2005:137	–
<i>Oregonichthys kalawatseti</i>	49.2	?	Markle et al., 1991: 280	V
<i>Stypodon signifer</i>	40	?	Miller et al., 2005: 140	X
<i>Lythrurus roseipinnis</i>	'ca.' 50.0	28	Boschung & Mayden, 2004	–
<b>Characiformes – Characidae</b>				
<i>Hyphessobrycon compresus</i>	46	?	Miller et al., 2005: 158	–
<b>Siluriformes – Ictaluridae</b>				
<i>Noturus stanauli</i>	36.2	?	Etnier & Jenkins, 1980	E
<b>Percopsiformes – Amblyopsidae</b>				
<i>Amblyopsis rosae</i>	43	?	UMMZ, 2009	T
<i>Chologaster cornuta</i>	48	20.0 (♀)	Ross & Rhode, 2003: 110	–
<b>Atheriniformes – Atherinopsidae</b>				
<i>Atherinella lisa</i>	45	?	Miller et al., 2005: 189	E
<b>Cyprinodontiformes – Cyprinodontidae</b>				
<i>Cualac tessellatus</i>	45.5	?	Miller et al., 1956: 9	E
<i>Cyprinodon arcuatus</i>	42	?	Minckley et al., 2003	Xp
<i>Cyprinodon atrorus</i>	35	?	Miller et al., 2005: 298	E
<i>Cyprinodon beltrani</i>	41	?	Miller et al., 2005: 299	V
<i>Cyprinodon diabolis</i>	28	?	UMMZ, 2009	E
<i>Cyprinodon elegans</i>	49	?	Suttkus & Williams, 2008	E
<i>Cyprinodon eremus</i>	41.5	?	Minckley et al., 2005	E
<i>Cyprinodon eximius</i>	46	?	Miller et al., 2005: 303	T
<i>Cyprinodon fontinalis</i>	50	?	Miller et al., 2005: 303	E
<i>Cyprinodon labiosus</i>	40	?	Miller et al., 2005: 304	E

Table 1. (continued).

<i>Cyprinodon latifasciatus</i>	43	?	Miller et al., 2005: 304	X
<i>Cyprinodon macrolepis</i>	44	?	Miller et al., 2005: 305	E
<b><i>Cyprinodon macularius</i></b>	<b>37.8</b>	<b>15</b>	Minckley et al., 2004; Schoenherr, 1988	E
<i>Cyprinodon meeki</i>	49	?	Miller et al., 2005: 307	E
<i>Cyprinodon nazas</i>	46	?	Miller et al., 2005: 308	T
<i>Cyprinodon pecosensis</i>	'ca.' 50.0	?	Garrett, 1982	E
<i>Cyprinodon pisteri</i>	49.3	23.0-25.0 (♂)	Minckley et al., 2002: 688, 689	E
<i>Cyprinodon rubrofluviatilis</i>	48	?	UMMZ, 2009	–
<i>Cyprinodon simus</i>	33	?	Miller et al., 2005: 309	E
<i>Cyprinodon suaviium</i>	45.5	?	Strecker, 2003	–
<i>Cyprinodon variegatus hubbsi</i>	35	?	Gilbert, 1992	V
<i>Cyprinodon verecundus</i>	35	?	Humphries, 1984: 63	E
<i>Garmanella pulchra</i>	38	?	Miller et al., 2005: 315	–
<i>Jordanella floridae</i>	39	?	McEachran & Fechhelm, 1998:899	–
<b><i>Megupsilon aporus</i></b>	<b>36</b>	<b>15.0 (♂); 18.0 (♀)</b>	Miller et al., 2005: 317	Xn
<b>Cyprinodontiformes – Fundulidae</b>				
<i>Adenia xenica</i>	42	?	Boschung & Mayden, 2004	–
<b><i>Leptolucania ommata</i></b>	<b>27</b>	<b>12.0 (♂); 15.0 (♀)</b>	Boschung & Mayden, 2004	–
<i>Lucania goodei</i>	42	?	Boschung & Mayden, 2004	–
<i>Lucania interioris</i>	~35.0	?	Miller et al., 2005: 316	E
<b>Cyprinodontiformes – Goodeidae</b>				
<i>Allodontichthys polylepis</i>	50	?	Miller et al., 2005: 269	E
<i>Allotoca maculata</i>	48	?	Miller et al., 2005: 274	E
<i>Allotoca meeki</i>	50	?	Miller et al., 2005: 274	E
<i>Allotoca zacapuensis</i>	50	?	Miller et al., 2005: 275	E
<i>Characodon garmani</i>	27	?	Miller et al., 2005: 279	X
<i>Crenichthys nevadae</i>	44	?	Hubbs, 1932	T
<i>Cyprinodon pachycephalus</i>	50	?	Miller et al., 2005: 308	E
<i>Skiffia bilineata</i>	42	?	Miller et al., 2005: 284	E
<i>Skiffia francesae</i>	43	?	Miller et al., 2005: 284	Xn
<b>Cyprinodontiformes – Poeciliidae</b>				
<i>Brachyrhaphis hartwegi</i>	43	?	Miller et al., 2005: 219	–
<i>Gambusia alvarezii</i>	38	?	Miller et al., 2005: 221	E
<i>Gambusia atrora</i>	40	?	Miller et al., 2005: 221	–
<i>Gambusia aurata</i>	34	?	Miller et al., 2005: 222	–
<i>Gambusia clarkhubbsi</i>	43	?	Garrett & Edwards, 2003	E
<i>Gambusia eurystoma</i>	36	?	Miller et al., 2005: 223	V
<i>Gambusia hurtadoi</i>	36	?	Miller et al., 2005: 223	E
<i>Gambusia krumholzi</i>	38	?	Miller et al., 2005: 223	V
<i>Gambusia longispinis</i>	29	?	Miller et al., 2005: 224	E
<i>Gambusia regani</i>	42	32.0 (♀)	Miller et al., 2005: 225	–
<i>Gambusia senilis</i>	46	?	Miller et al., 2005: 226	T
<i>Gambusia sexradiata</i>	43	?	Miller et al., 2005: 227	–
<i>Gambusia speciosa</i>	48	?	Miller et al., 2005: 227	T
<i>Gambusia vittata</i>	37	25.0 (♀)	Miller et al., 2005: 228	–
<b><i>Heterandria formosa</i></b>	<b>30</b>	<b>15.0 (♀)</b>	Boschung & Mayden, 2004	–
<i>Heterophallus echeagarayii</i>	35	?	Miller et al., 2005: 230	–
<i>Heterophallus milleri</i>	31	?	Miller et al., 2005: 231	–
<i>Heterophallus rachovii</i>	32	?	Miller et al., 2005: 231	–
<i>Phallichthys fairweatheri</i>	46	?	Miller et al., 2005: 232	–
<i>Poecilia chica</i>	46	?	Miller et al., 2005: 234	V
<i>Poecilia sulphuraria</i>	50	?	Miller et al., 2005: 239	T
<i>Poeciliopsis baenschi</i>	44	?	Miller et al., 2005: 241	–
<i>Poeciliopsis balsas</i>	43	?	Miller et al., 2005: 242	–
<i>Poeciliopsis fasciata</i>	45	?	Miller et al., 2005: 242	–
<i>Poeciliopsis latidens</i>	48	?	Miller et al., 2005: 245	T

Table 1. (continued).

<i>Poeciliopsis lucida</i>	~42.0	?	Miller et al., 2005: 246	–
<i>Poeciliopsis monacha</i>	44	?	Miller et al., 2005: 246	–
<i>Poeciliopsis occidentalis</i>	48	?	Miller et al., 2005: 246	E
<i>Poeciliopsis presidionis</i>	~45.0	?	Miller et al., 2005: 248	–
<i>Poeciliopsis prolifica</i>	42	?	Miller et al., 2005: 248	–
<i>Poeciliopsis viriosa</i>	46	?	Miller et al., 2005: 251	–
<i>Priapella bonita</i>	45	?	Miller et al., 2005: 251	X
<i>Priapella intermedia</i>	45	?	Miller et al., 2005: 253	–
<i>Xiphophorus andersi</i>	40	?	Miller et al., 2005: 255	–
<i>Xiphophorus clemenciae</i>	46	?	Miller et al., 2005: 255	T
<i>Xiphophorus continens</i>	~30.0	?	Miller et al., 2005: 256	–
<i>Xiphophorus cortezi</i>	50	?	Miller et al., 2005: 257	–
<i>Xiphophorus couchianus</i>	37	?	Miller et al., 2005: 257	E
<i>Xiphophorus evelynae</i>	48	?	Miller et al., 2005: 257	–
<i>Xiphophorus gordonii</i>	30	?	Miller et al., 2005: 258	E
<i>Xiphophorus maculatus</i>	45	?	Miller et al., 2005: 259	–
<i>Xiphophorus meyeri</i>	46.2	?	Miller et al., 2005: 260	E
<i>Xiphophorus milleri</i>	35	?	Miller et al., 2005: 261	E
<i>Xiphophorus multilineatus</i>	~45.0	?	Miller et al., 2005: 262	–
<i>Xiphophorus nigrensis</i>	42	?	Miller et al., 2005: 263	–
<i>Xiphophorus pygmaeus</i>	37.2	?	Morris & Ryan, 1995	–
<i>Xiphophorus xiphidium</i>	49	?	Miller et al., 2005: 264	–
<b>Cyprinodontiformes – Rivulidae</b>				
<i>Millerichthys robustus</i>	33	?	Miller et al., 2005: 207	E
<i>Rivulus tenuis</i>	40	?	Miller et al., 2005: 208	–
<b>Perciformes – Elasmomatidae</b>				
<i>Elassoma alabamae</i>	25.5	?	Mayden, 1993	E
<i>Elassoma boehlkei</i>	27	?	Rohde & Arndt, 1987	T
<i>Elassoma evergladei</i>	28	?	Boschung & Mayden, 2004	–
<i>Elassoma okatie</i>	28.7	?	Rohde & Arndt, 1987	V
<i>Elassoma okefenokee</i>	28	?	Boschung & Mayden, 2004	–
<i>Elassoma zonatum</i>	38.5	19.2(♀)	Walsh & Burr, 1984: 36, 41	–
<b>Perciformes – Percidae</b>				
<i>Etheostoma australe</i>	50	?	Miller et al., 2005: 335	E
<i>Etheostoma chlorosoma</i>	50	?	Page, 1983	–
<i>Etheostoma denoncourti</i>	28.6	?	Stauffer & Van Sink, 1997	V
<i>Etheostoma ditrema</i>	45	?	Page, 1983	T
<i>Etheostoma edwini</i>	49	27.0(♀)	Boschung & Mayden, 2004	–
<i>Etheostoma fonticola</i>	35.5	24.4(♀ TL)	Burr, 1978; Schenk & Whiteside, 1977	E
<i>Etheostoma fusiforme</i>	46	32.0(TL)	Boschung & Mayden, 2004	–
<i>Etheostoma gracile</i>	49	35.0(TL)	Boschung & Mayden, 2004	–
<i>Etheostoma grahamsi</i>	44	?	Miller et al., 2005: 336	T
<i>Etheostoma lugoi</i>	37.6	?	Norris & Minckley, 1997: 162	E
<i>Etheostoma microperca</i>	37	~28.0	Etnier & Starnes, 1993	–
<i>Etheostoma nuchale</i>	45	?	Boschung & Mayden, 2004	E
<i>Etheostoma okaloosae</i>	48	?	Burkhead et al., 1992	E
<i>Etheostoma pottsi</i>	50	?	Miller et al., 2005: 337	T
<i>Etheostoma proeliare</i>	40	29.2(♂); 30.7(♀)	Boschung & Mayden, 2004	–
<i>Etheostoma rubrum</i>	46	27.0(♀)	Raney & Suttkus, 1966; Knight & Ross, 1992:102	E
<i>Etheostoma segrex</i>	43.8	?	Norris & Minckley, 1997: 171	E
<i>Etheostoma stigmaeum</i>	48.9	35.6(♂); 35.7(♀)	Boschung & Mayden, 2004	–
<i>Etheostoma tippecanoe</i>	36	22.5(♂); 21.2(♀)	Stauffer & Van Sink, 1997; Warren et al., 1986	V
<i>Etheostoma trisella</i>	49	37	Boschung & Mayden, 2004; Ryon, 1986: 81	E
<i>Etheostoma zonifer</i>	38	?	Boschung & Mayden, 2004	–
<b>Scorpaeniformes – Cottidae</b>				
<i>Cottus paulus</i>	38	25	Williams, 2002; Boschung & Mayden, 2004	E

*Knipowitschia goerneri* (maximum size 22 mm SL).

Any explanation for this disparity is at this time highly speculative; however, Africa, Asia and South America differ greatly from North America in several key aspects that may promote miniaturization and diversification of diminutive fishes. The equatorial position of much of Africa, South and Southeast Asia and South America is likely a major factor in diversification on those continents in general (Lundberg et al., 2000). The large number of species in the tropics has also been suggested to make miniature species simply more likely to evolve (Weitzman & Vari, 1988). Collette (1962) suggested a link between the acidic environments inhabited by *Etheostoma fusiforme* (Swamp Darter) and its reduced lateral line system and overall small body size. Similarly Weitzman & Vari (1988) noted that many South American miniatures occur in this environment, which is common across many portions of the continent. They also recognized that most of the miniature freshwater fishes included in their table, if not found explicitly in acidic waters, inhabited still or slow-flowing habitats or microhabitats. Kottelat & Vidthayanon (1993) and Kottelat et al. (2006) also commented that most miniature Southeast Asian freshwater fishes occupy lentic or slow-flowing habitats. Our review further supports the notion that miniature fishes are more likely to occur in habitat with little flow. *Cyprinodon macularius*, *Elassoma alabamae* and *Megupsilon aporus* occur in springs or spring-fed pools with little or no flow (Hendrickson & Romero, 1989; Mayden, 1993; Miller & Walters, 1972). *Leptolucania ommata* and *Elassoma zonatum* are found in vegetated swamps and backwaters or streams with little flow (Boschung & Mayden, 2004). *Heterandria formosa* prefers pools of sluggish, heavily-vegetated streams (Boschung & Mayden, 2004) and *Notropis saladonis* inhabits slow-flowing, silty streams (Hubbs & Hubbs, 1958). In addition, many of the smallest species in Table 1, while not miniature by the definition of Weitzman & Vari (1993), are found mostly in swamp and spring habitats (e.g. *Elassoma*, *Xiphophorus*). These habitats, as well as smaller streams (even swift-flowing), likely limit dispersal, and thus gene flow, among populations of small-bodied fishes, promoting diversification (Knouff & Page, 2003).

While this general model encompasses most of the diminutive species we examined (and indeed those studied on other continents), there is



Fig. 3. *Megupsilon aporus*, Mexico: Matinal de El Potosi Nuevo Leon. Male. Photograph courtesy of P. Loisel.

also a small component of diminutive North American freshwater fish diversity that inhabits fast flowing, large and medium-sized upland rivers and streams with gravel, cobble, boulder and bedrock as predominant sediments (e.g. *Noturus stanauli*, *Etheostoma tippecanoe*, *E. denoncourti*). Similar habitat preferences have been reported for a small number of miniature taxa from other continents, including the catfishes *Hoplosternon papillatus* (Stewart, 1985), *Horiomyzon retropinnatus* (Stewart, 1986), and the cichlid *Nanochromis minor* (Roberts & Stewart, 1976). The microhabitats associated with the cryptobenthic behavior of these latter species may be characterized by reduced flow and analogous to the slow velocities of oxbows, springs and swamp.

Imperilment was high among diminutive fishes, with 58 % of those listed in Table 1 labeled as vulnerable (9 % of total), threatened (10.2 %), endangered (32.0 %) or extinct (6.8 %) according to Jelks et al. (2008). Only barely over 42 % of these fishes did not warrant listing by Jelks et al. (2008) (i.e. not threatened). *Elassoma alabamae* is particularly threatened and in need of further protection (Conway & Mayden, 2006). Our review also highlights the lack of critical life history information (minimum size at maturity) for many of the fish species we examined. We anticipate that with more study of reproductive biology, additional species on our list will be identified as miniature based on size at maturity (less than 20 mm SL) in the future.

Small bodied freshwater fishes are particularly faced with a great threat of extinction, but may get less attention than their larger commercially-harvested relatives (Olden et al., 2007). Without a better knowledge of the processes and conditions that gave rise to and help maintain the

diversity of small bodied freshwater fishes, and the habitats that they rely on, there will be little hope for preserving these interesting and unique components of global biodiversity.

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