



# Phylogeny, diversification patterns and historical biogeography of euglossine orchid bees (Hymenoptera: Apidae)

SANTIAGO R. RAMÍREZ<sup>1\*</sup>, DAVID W. ROUBIK<sup>2</sup>, CHARLOTTE SKOV<sup>3</sup> and NAOMI E. PIERCE<sup>1</sup>

<sup>1</sup>Museum of Comparative Zoology, Harvard University, 26 Oxford St., Cambridge, MA 02138, USA

<sup>2</sup>Smithsonian Tropical Research Institute, Apartado 0843-03092, Balboa, Ancón, Panamá, República de Panamá, Unit 0948, APO AA 34002-0948, USA

<sup>3</sup>Rungsted Gymnasium, Stadionalle 14, 2960 Rungsted Kyst, Denmark

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The orchid bees constitute a clade of prominent insect pollinators distributed throughout the Neotropical region. Males of all species collect fragrances from natural sources, including flowers, decaying vegetation and fungi, and store them in specialized leg pockets to later expose during courtship display. In addition, orchid bees provide pollination services to a diverse array of Neotropical angiosperms when foraging for food and nesting materials. However, despite their ecological importance, little is known about the evolutionary history of orchid bees. Here, we present a comprehensive molecular phylogenetic analysis based on ~4.0 kb of DNA from four loci [cytochrome oxidase (*COI*), elongation factor 1- $\alpha$  (*EF1- $\alpha$* ), arginine kinase (*ArgK*) and RNA polymerase II (*Pol-II*)] across the entire tribe Euglossini, including all five genera, eight subgenera and 126 of the approximately 200 known species. We investigated lineage diversification using fossil-calibrated molecular clocks and the evolution of morphological traits using disparity-through-time plots. In addition, we inferred past biogeographical events by implementing model-based likelihood methods. Our dataset supports a new view on generic relationships and indicates that the cleptoparasitic genus *Exaerete* is sister to the remaining orchid bee genera. Our divergence time estimates indicate that extant orchid bee lineages shared a most recent common ancestor at 27–42 Mya. In addition, our analysis of morphology shows that tongue length and body size experienced rapid disparity bursts that coincide with the origin of diverse genera (*Euglossa* and *Eufriesea*). Finally, our analysis of historical biogeography indicates that early diversification episodes shared a history on both sides of Mesoamerica, where orchid bees dispersed across the Caribbean, and through a Panamanian connection, thus reinforcing the hypothesis that recent geological events (e.g. the formation of the isthmus of Panama) contributed to the diversification of the rich Neotropical biota. © 2010 The Linnean Society of London, *Biological Journal of the Linnean Society*, 2010, 100, 552–572.

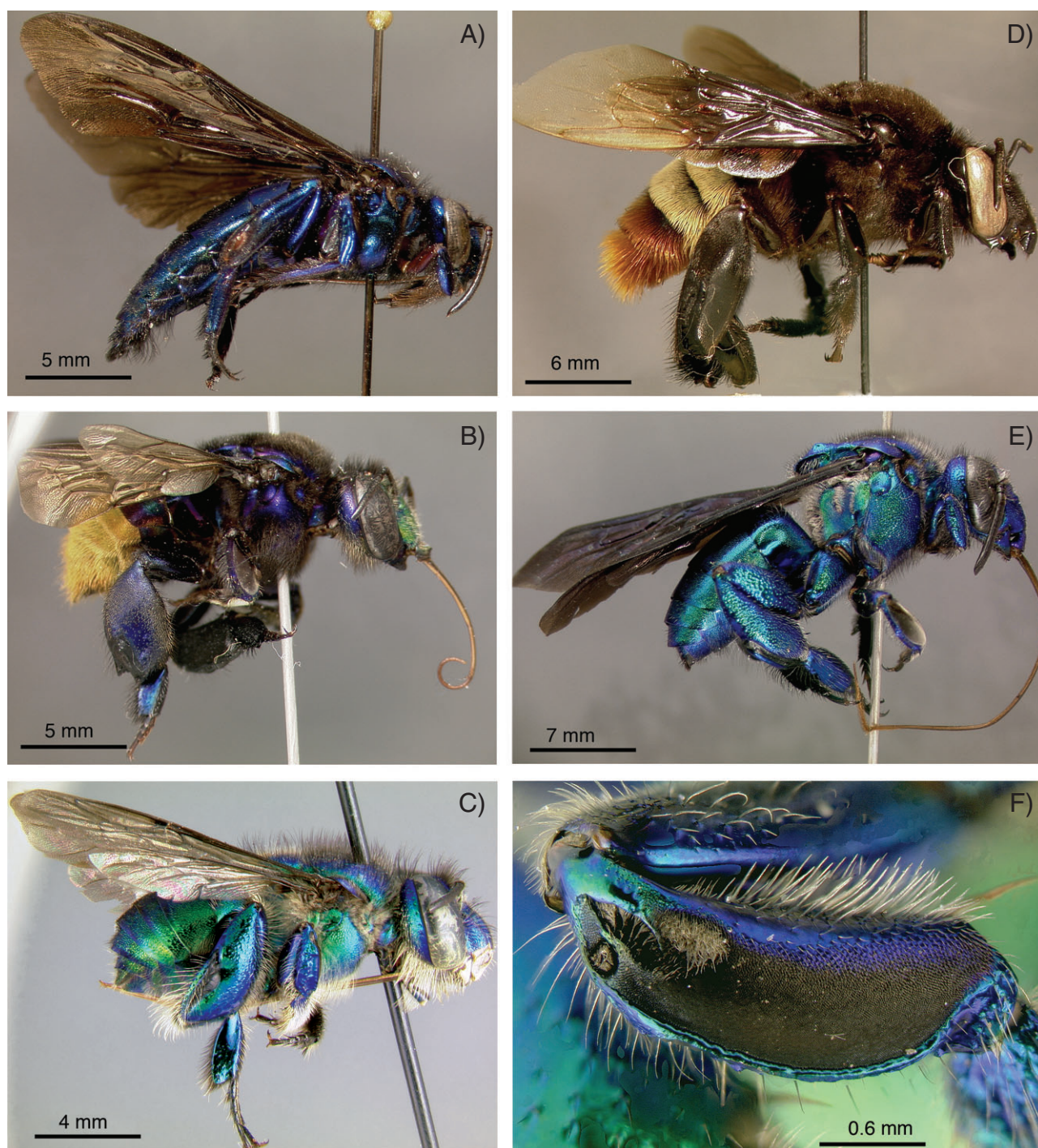
**ADDITIONAL KEYWORDS:** *Aglae* – Amazon – Andes – DTT plots – *Eulaema* – Miocene – Oligocene – penalized likelihood – relaxed molecular clocks.

## INTRODUCTION

For more than 150 years, orchid bees (tribe Euglossini) have caught the attention of scientists and naturalists alike, mainly as a result of the

intricate association of male bees as pollinators of orchid flowers (Dressler, 1967, 1968, 1982a; Roubik & Hanson, 2004). Male orchid bees collect aromatic compounds from orchids (and other nonfloral sources) and store them in specialized pouches located in their hind tibiae (Fig. 1). They later expose such fragrances by presenting small samples of scent to females during courtship (Eltz *et al.*, 1999; Eltz, Roubik & Whitten, 2003; Bembé, 2004; Eltz, Sager & Lunau 2005a, Eltz, Roubik & Lunau, 2005b). Fragrances are

\*Corresponding author. Current address: University of California Berkeley, 137 Mulford Hall #3114, Berkeley, CA 94720, USA.  
E-mail: sramirez@post.harvard.edu



**Figure 1.** Representative species of each of the five orchid bee genera within the tribe Euglossini: A, *Aglae caerulea*; B, *Eufriesea lucifera*; C, *Euglossa paisa*; D, *Eulaema meriana*; E, *Exaerete smaragdina*. F, An example of a midtibia felty patch (of *Euglossa paisa*), showing the characters routinely used for species-level diagnosis based on male bees. Note the hindleg opening slit through which male bees deposit and store chemical fragrances.

thus thought to act as species-specific pheromone analogues that evolved in response to reproductive interference among closely related lineages (Zimmermann, Ramírez & Eltz, 2009). Whilst collecting fra-

grances, male orchid bees visit and pollinate a large diversity of orchids (Williams, 1982; Williams & Whitten, 1983), as well as other plant families (Ramírez, Dressler & Ospina, 2002). Since the 1960s,

when fragrance-gathering behaviour was discovered (Vogel, 1963, 1966; Dodson *et al.*, 1969), synthetic chemical baits that attract male bees have been widely used in ecological surveys across the Neotropics (Roubik & Ackerman, 1987; Roubik & Hanson, 2004). These surveys revealed a great deal about orchid bee ecology, population biology and natural history (Dressler, 1982a; Ramírez *et al.*, 2002; Cameron, 2004; Roubik & Hanson, 2004; Ramírez, 2009).

The tribe Euglossini belongs to the bee family Apidae, and is contained within a clade commonly known as the corbiculate bees (Michener, 2007). Female corbiculate bees have basket-shaped hind tibiae (corbiculae) for transporting pollen and nesting materials to the nest (Roubik, 1989), a trait that has been secondarily lost in nest-parasites (Michener, 2007). In addition to orchid bees, corbiculate bees also include the primitively eusocial bumble bees (tribe Bombini) and the highly eusocial honey bees (tribe Apini) and stingless bees (tribe Meliponini) (Michener, 2007). However, unlike their close relatives, orchid bees do not display eusocial traits, such as worker sterility or cooperative brood care, and instead exhibit solitary, communal or parasocial nesting (Zucchi, Sakagami & Camargo, 1969; Cameron & Ramírez, 2001; Soucy, Giray & Roubik, 2003; Augusto & Garófalo, 2004; Roubik & Hanson, 2004).

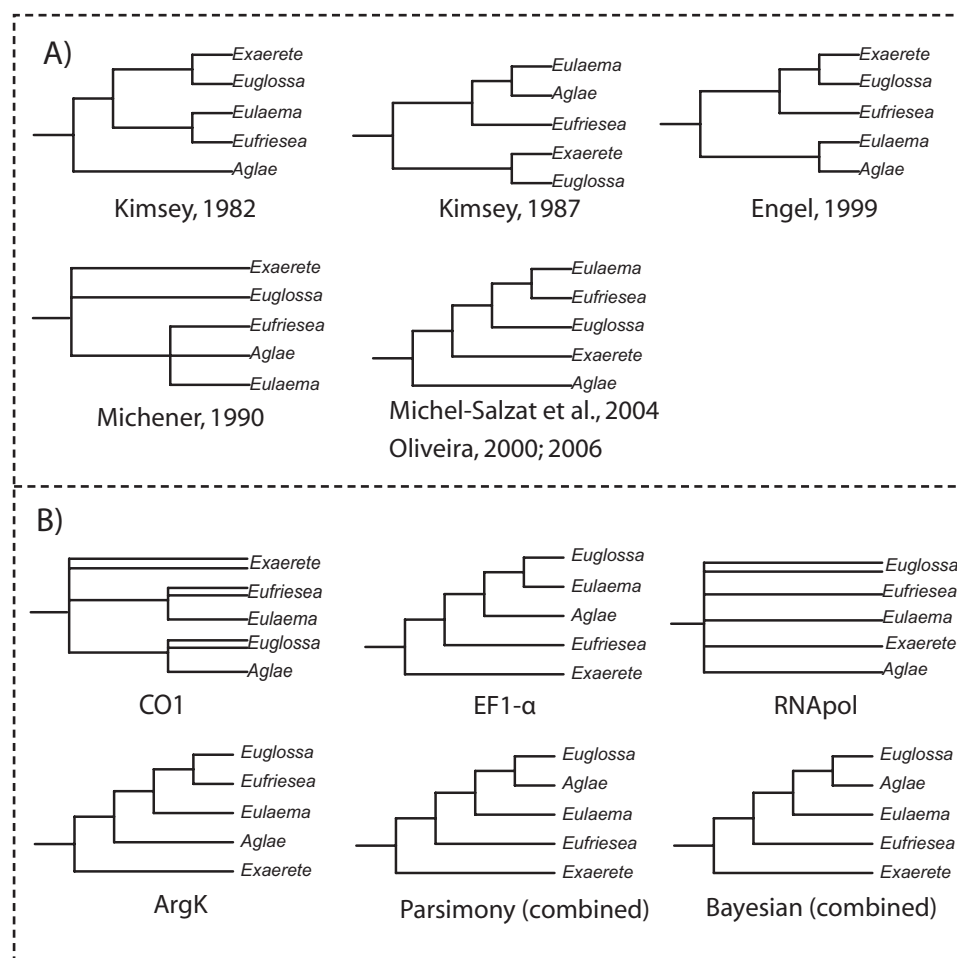
Corbiculate bees are contained within the monophyletic long-tongued bees (Michener, 1990; Danforth *et al.*, 2006), a status supported by both morphological and molecular characters (Michener, 2007; Kawakita *et al.*, 2008), but the phylogenetic position of Euglossini within corbiculate bees has been a subject of considerable debate. Because all tribes of corbiculate bees, except Euglossini, exhibit some degree of advanced eusociality (Michener, 2007), the position of orchid bees with respect to the remaining corbiculate bee lineages can potentially establish the number of origins of eusocial behaviour in this group (for a most up-to-date review, see Kawakita *et al.* 2008). Whereas hypotheses based on morphological characters tend to favour orchid bees as sister to the rest of the corbiculate bees, and thus suggest a single origin of eusociality among corbiculate clades (Michener, 1944; Engel, 2001; but see Winston & Michener, 1977), most studies based on molecular characters favour the hypothesis of orchid bees as sister to honey bees, thus implying a dual origin of highly eusocial behaviour (Cameron & Mardulyn, 2001; Kawakita *et al.*, 2008).

The Euglossini contains five (Fig. 1) well-delineated extant genera (*Aglae*, *Eufriesea*, *Euglossa*, *Exaerete* and *Eulaema*) and over 200 described species (Moure, Melo & Faria, 2007). Several alternative hypotheses regarding the relationships between orchid bee genera have been advanced on the basis of both

molecular and morphological characters (Fig. 2A). Moreover, classification schemes and phylogenetic hypotheses from morphology have been proposed for portions of all genera.

The genus *Aglae* contains a single large species (*A. caerulea*), which is an exclusive cleptoparasite (or 'cuckoo' parasite) of *Eulaema* (Dressler, 1982a). *Aglae caerulea* is restricted to the lowland wet forests of the Chocó region and Amazon basin of South America (Ramírez *et al.*, 2002; Anjos-Silva, Camilo & Garófalo, 2006). The genus *Eufriesea* contains about 60 medium-sized species distributed throughout the Neotropical region. Although no subgeneric classifications exist for this genus, several species' groups were identified by Kimsey (1982) based on subtle morphological characters, including tongue length, integument coloration, genitalic characters and head shape. The genus *Euglossa* is the most diverse within the tribe, containing approximately 110 species of primarily small bees distributed throughout the Neotropics. This genus is currently subdivided into six subgenera based on external morphological characters (Moure, 1969; Dressler, 1978a, b, 1982a, b, c, d), as well as genitalic characters (Ospina-Torres, Parra & Gonzalez, 2006). Bembé (2007) used morphological characters to infer the phylogenetic relationships among species of the *cordata* group in the subgenus *Euglossa s.s.* Darveau *et al.* (2005) used a single mitochondrial marker to infer the relationships among 32 species of orchid bees, including 24 in the genus *Euglossa*. Several new species of *Euglossa* have been described in recent years (Bembé, 2004, 2007; Roubik, 2004; Ramírez, 2005, 2006; Parra, Ospina-Torres & Ramírez, 2006; Rasmussen & Skov, 2006; Hinojosa-Diaz & Engel, 2007; Nemésio, 2007, 2009a, b), and additional taxa probably remain undescribed, especially from poorly sampled areas of South America (e.g. Colombia, Ecuador and Peru). The genus *Eulaema* contains 15–20 species of large bees (Fig. 1) that are also distributed across the Neotropics. *Eulaema* is subdivided into two morphologically distinct subgenera (Moure, 1950, 2000). Oliveira (2006) used morphological characters to infer phylogenetic relationships among species, which supported the monophyletic status of the subgenera mentioned above. Finally, *Exaerete* contains seven cleptoparasitic species that use nests of either *Eulaema* or *Eufriesea* as their hosts. Species in this genus are arranged into two species' groups based on morphological traits (Kimsey, 1979; Anjos-Silva, Engel & Andena, 2007).

In summary, the tribe Euglossini encompasses a diverse, but comparatively well-studied group of bees that inhabits the New World tropics. However, despite the widespread interest and extensive literature concerning natural history, behavioural ecology,



**Figure 2.** Summary of alternative hypotheses of the generic relationships among orchid bees proposed in previous studies (A) and those supported by individual genes in this study (B).

taxonomy, chemical ecology and population biology – as well as the suitability for comparative study – a comprehensive phylogenetic analysis of the entire tribe is still lacking. Here, we investigate the internal phylogenetic relationships of orchid bees, and explore their morphological evolution, time of divergence, lineage diversification and historical biogeography using comparative methods.

## METHODS

### TAXONOMIC SAMPLING

We provide an extensive sampling of the tribe Euglossini, including all five genera, all eight subgenera, 25 of the 26 species' groups and a total of 126 of the approximately 200 species. We sampled nine out-group taxa, including all corbiculate bee tribes, and two noncorbiculate genera in the family Apidae (*Centris* and *Epicharis*). Our DNA matrix contained a total of 146 terminals. Most specimens were collected

between 2002 and 2006 from 12 Neotropical countries (Supporting Information Appendix S1). Tissue samples (thoracic muscle or forelegs) or whole specimens were preserved in 100% ethanol and kept at  $-20^{\circ}\text{C}$  for DNA extraction. For each sample, we sequenced a total of  $\sim 4.0$  kb from fragments of four different loci, including the mitochondrial protein-coding gene cytochrome oxidase (*CO1*, 1.2 kb) and the nuclear protein-coding genes elongation factor 1- $\alpha$  (*EF1- $\alpha$* , F2 copy,  $\sim 1.2$  kb), arginine kinase (*ArgK*,  $\sim 0.7$  kb) and RNA polymerase II (*Pol-II*, 0.8 kb). Voucher specimens for the sampled taxa (listed in Supporting Information Appendix S1) are currently deposited in the Museum of Comparative Zoology at Harvard University.

### MOLECULAR PROCEDURES

We extracted genomic DNA from individual bees using the Qiagen DNA Extraction Kit (Qiagen Inc.,

Valencia, CA, USA), and ran polymerase chain reactions (PCRs) on a Bio-Rad DNA Engine Dyad Peltier thermal cycler (Bio-Rad Laboratories Inc., Hercules, CA, USA). All reactions were prepared in volumes of 25  $\mu\text{L}$  with 2.5  $\text{mmol L}^{-1}$   $\text{MgCl}_2$ , 2.5  $\text{mmol L}^{-1}$  PCR buffer and *Taq* polymerase (Qiagen Inc.) using available primer sets (Danforth *et al.*, 2004a). We purified PCR products by incubating samples at 37 °C for 35 min with the *Escherichia coli* enzyme exonuclease I (New England Biolabs, Hanover, MD, USA), and subsequently raised the temperature to 80 °C for 20 min. We performed cycle sequencing using a BigDye™ Terminator v3.1 Ready Reaction Cycle Sequencing Kit (Applied Biosystems Inc., Foster City, CA, USA). Samples were sequenced on an Applied Biosystems Inc. 3100 Genetic Analyzer using forward and reverse strands. Complementary strands were assembled with the software package Sequencher v4.2 (Gene Codes Corp., Ann Arbor, MI, USA).

#### PHYLOGENETIC METHODS

We assembled a DNA matrix containing all four loci using the software packages MacClade v4.06 and Mesquite v2.01 (Maddison & Maddison, 2003). We implemented parsimony analyses in the software package PAUP\* v4.10b (Swofford, 2003) by assuming unordered transitions and weighting all characters equally. Heuristic tree searches consisted of 100 random addition sequences, using the tree bisection–reconnection (TBR) swapping algorithm; bootstrap support values were estimated via nonparametric bootstrapping in 1000 replicates. We implemented Bayesian phylogenetic analyses in the software package MRBAYES v3.1.1 (Ronquist & Huelsenbeck, 2003). Tree searches were performed assuming both single and multiple models of sequence evolution for each locus, and Markov chain Monte Carlo (MCMC) searches were made for 10 million generations, sampling every 1000 generations, for a total of 10 000 trees. We estimated model parameters during runs, and estimated Bayesian posterior probabilities as the proportion of trees sampled; the trees obtained in the first one million generations were discarded.

#### MOLECULAR CLOCK CALIBRATION

We performed molecular clock analyses via penalized likelihood (PL) using the software package r8s v1.71 (Sanderson, 2002) and via mean path length (MPL) using the software package PATHd8 v1.0 (Britton *et al.*, 2007). Unlike PL, the MPL method does not assume autocorrelation between parent and child branches, and thus assumes heterogeneous substitution rates across lineages. We calibrated a molecular phylogenetic tree obtained via Bayesian analyses

using both fossil and biogeographical data. Two fossil orchid bees are known from Miocene Dominican amber deposits, and several corbiculate bees are known from amber and shale deposits elsewhere. The amber-preserved orchid bee *Euglossa moronei* (Engel, 1999a) (15–20 Myr old) (Iturralde-Vinent & MacPhee, 1996) bears synapomorphic characters that place it unambiguously within extant lineages of *Euglossa*. We used its age as a minimum age constraint for the genus *Euglossa*. The other known fossil orchid bee, *Paleoeglossa mellisiflora* (Poinar, 1998), cannot be assigned unequivocally to an extant orchid bee lineage (D. W. Roubik & S. Ramírez, pers. observ.), and thus we did not include it in this analysis. We calibrated our molecular clock analysis with other corbiculate bee fossils, including the giant honey bee *Apis lithohermaea* from the Pliocene–Pleistocene (14–16 Myr old) olivine basalts of Japan (Engel, 2006), and the stingless bee *Proplebeia dominicana* from Miocene (15–20 Myr old) lignite, sandy clay beds of Dominican amber deposits and Chiapas amber (Wille & Chandler, 1964; Camargo, Grimaldi & Pedro, 2000). In addition, because orchid bees are restricted to the tropical forests in the New World, we used the Gondwana break-up between tropical America and Africa (~70–100 Mya; Smith, Smith & Funnell, 1994; Danforth *et al.*, 2004b) as a maximum age constraint for the node connecting Euglossini and Apini. In fact, the fossil record suggests that orchid bees have been restricted to the New World Tropics since the Eocene (Engel, 1999a, b; 2001). On the other hand, extant members of the genus *Apis* are largely distributed in the Old World, but a recent fossil discovery revealed that honey bees were present in North America (Nevada) during the Miocene (Engel, Hinojosa-Díaz & Rasnitsyn, 2009). We note that molecular clock calibrations based on geographical distributions should be used with caution. The estimated age for corbiculate bees in our analysis is congruent with that suggested by Hines (2008). We estimated branch lengths using the majority rule consensus tree obtained via Bayesian analyses optimized with likelihood using the model of sequence evolution GTR + I +  $\Gamma$ .

#### ANALYSIS OF LINEAGE DIVERSIFICATION

We investigated the timing and patterns of diversification in orchid bees with lineage-through-time (LTT) plots. LTT plots depict the cumulative (log) number of nodes as a function of time. We estimated LTT plots on the basis of fossil-calibrated molecular clock chronograms obtained via both PL and MPL. In addition, we tested whether significant changes occurred in the diversification rates throughout the evolutionary history of orchid bees by implementing various likelihood-based statistical methods in the software

packages APE v2.0 (Paradis, Claude & Strimmer, 2004) and GEIGER v1.2-02 (Harmon *et al.*, 2008).

#### ANALYSIS OF HISTORICAL BIOGEOGRAPHY

Extensive studies of plants and animals have identified recurrent areas of endemism across the Neotropical region. We assigned taxa to biogeographical regions of endemism described by Morrone (2006), and coded the distribution (presence/absence) of orchid bee taxa into discrete geographical regions by surveying both the literature (Kimsey, 1982; Ramírez *et al.*, 2002; Roubik & Hanson, 2004; Nemésio & Silveira, 2007; Nemésio, 2009a, b) and museum collections. For consistency, we used the same nomenclature of biogeographical regions as in Kimsey (1982). In addition, we coded the altitudinal distribution of orchid bee taxa as either predominantly lowland (< 800–900 m), montane (> 900 m) or widespread (Ramírez *et al.*, 2002; Roubik & Hanson, 2004). We analysed the two distributional datasets in a model-based likelihood framework using the software package LAGRANGE v2.0.1 (Ree & Smith, 2008). This flexible platform permits the formulation of dispersal–extinction–cladogenesis (DEC) models by specifying instantaneous transition rates between geographical ranges (or altitudinal zones). Unlike previous methods used in the analysis of historical biogeography (e.g. DIVA), LAGRANGE can incorporate information on past geological events (e.g. land bridge connections, range uplifts) and time (branch lengths), and thus provides a more robust framework for the reconstruction of ancestral areas (Clark *et al.*, 2008).

We built two separate DEC models. In the first model (M0), we allowed all transition probabilities between geographical ranges and altitudinal zones to be equally likely at all times. In the second model (M1), we restricted the geographical ranges and dispersals to reflect geological events that may have affected either vicariance or dispersal events. In M1, lineages were allowed to disperse to and from the Panama endemic zone + Choco region only after the isthmus of Panama was formed gradually as an archipelago at 3.1–3.7 Mya (Coates & Obando, 1996; Coates *et al.*, 2004). Likewise, lineages were allowed to disperse to and from the Andes after the major Andean uplift took place at 2.7 Mya (Gregory-Wodzicki, 2000). The remaining instantaneous transition probabilities were otherwise left unmodified.

#### ANALYSIS OF MORPHOLOGICAL DIVERSIFICATION

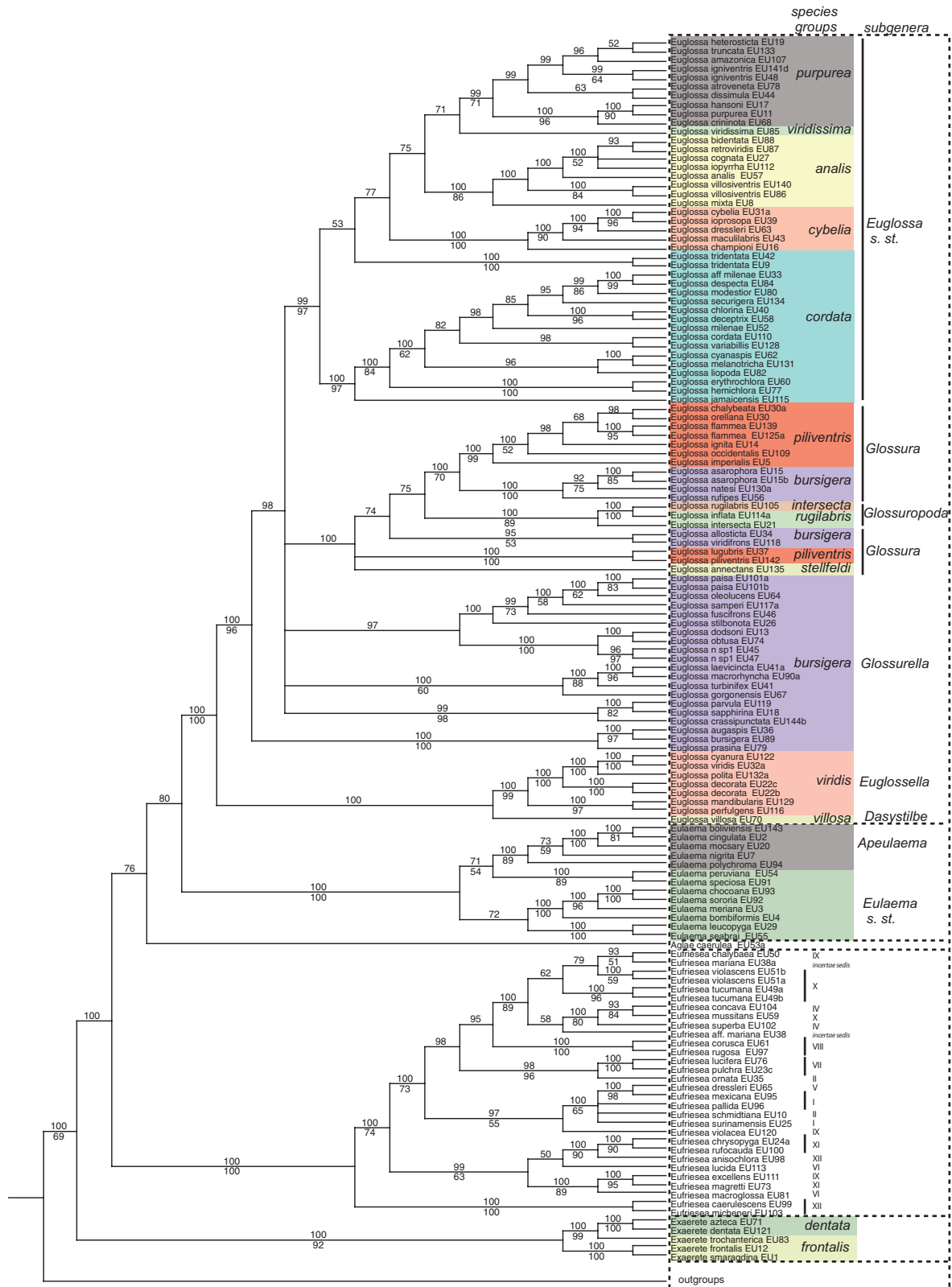
To investigate the morphological diversification of functional traits in orchid bees, we used a subset of the species included in the phylogenetic analysis. At

least three individuals from a total of 75 orchid bee species (Supporting Information Appendix S2) were measured from specimens deposited in the Museum of Comparative Zoology (Harvard University). We measured total body length, body width (intertegular distance), body mass (dry weight) and tongue length (basal to distal prementum length). We investigated morphological disparity by implementing disparity-through-time (DTT) plots (Harmon *et al.*, 2003). This method calculates dissimilarity as the mean square (pair-wise) Euclidian distance among points in morphospace for subclades relative to the variance of the entire clade. Disparity is calculated for individual nodes by moving up the phylogeny from the root of the tree. This methodology was first implemented by palaeontologists to investigate morphological divergence across geological time scales, and has the advantage of being insensitive to sample size (Foote, 1997). To compare the observed empirical morphological disparity of traits (analysed separately and simultaneously) with respect to a null model, we performed 1000 simulations using both Brownian and speciation models of character evolution. Then, we averaged the simulated values for each node, plotted them against the relative lineage divergence times, and calculated the difference between the empirical and simulated curves by estimating the area between the curves. Brownian models of character evolution assume that a continuous character changes along the branches of a tree following a Brownian motion process, whereas speciation models of character evolution assume that changes on continuous characters occur during branching processes only (Pagel, 1999). These analyses were carried out using the package GEIGER v1.2-02 (Harmon *et al.*, 2008).

## RESULTS

### PHYLOGENETIC ANALYSES

Our maximum parsimony and Bayesian inference analyses – based on four loci – resolved most phylogenetic relationships within and between orchid bee lineages. Both methods yielded informative phylogenetic trees (Fig. 3) that were congruent with each other under different optimization schemes and model parameters. Our parsimony analysis yielded 552 shortest trees (TL = 6899), with a strict consensus almost identical to the result of the Bayesian analysis (Fig. 3). The Bayesian phylogenetic analysis also yielded well-supported trees that varied little regardless of whether we used a single (GTR +  $\Gamma$  + I) or partitioned (locus-specific) model of sequence evolution (Supporting Information Table S1). All our combined phylogenetic analyses returned all genera as monophyletic, *Exaerete* as the sister to the remaining



**Figure 3.** Bayesian cladogram of tribe Euglossini with mapped taxonomic schemes (subgenera and species' groups). Support values below the branches were obtained via parsimony bootstrap; values above the branches correspond to Bayesian posterior probabilities.

genera, *Eufriesea* as sister to *Euglossa* + *Eulaema* + *Aglae*, and *Aglae* clustered within Euglossini (never basal). The relative positions of *Eulaema* and *Eufriesea* were inconsistent, depending on the phylogenetic method and model of sequence evolution used.

Our phylogenetic analyses indicated that four of the six subgenera recognized on the basis of morphological characters within *Euglossa* (*Euglossa s.s.*, *Euglossella*, *Dasystilbe* and *Glossuropoda*) are monophyletic, whereas the remaining two (i.e. *Glossura* and *Glossurella sensu* Ramírez *et al.*, 2002) constitute paraphyletic groups (Fig. 3). Recently, Moure *et al.* (2007) and Faria & Melo (2007) proposed the transfer of several species from *Glossurella* to *Glossura*, thus making both subgenera more congruent with our phylogenetic results. However, *Glossura* remains paraphyletic because of the position of *Glossuropoda* (Fig. 3). In addition, the subgenus *Glossurella* was not recovered as monophyletic, but this was the result of unresolved nodes (Fig. 3). The remaining species' groups and subgenera in *Euglossa*, *Eulaema* and *Exaerete* were also recovered as monophyletic, but this was not the case for the genus *Eufriesea*, where most species' groups were recovered as paraphyletic (Figs 3, 4).

#### DIVERGENCE TIMES AND DIVERSIFICATION PATTERNS

The results from our PL and MPL analyses indicated that Euglossini shared a most recent common ancestor during the Miocene–Eocene periods (27–42 Mya), depending on whether we use the oldest or youngest ages of the fossil calibrations (Supporting Information Table S2). Time estimates obtained via PL and MPL were comparable (Fig. 5).

Our analysis of lineage diversification using LTT plots indicated that orchid bee lineages accumulated rapidly and steadily during the Miocene (15–22 Mya) (Figs 5, 6). The shape of the curve describing lineage accumulation did not change significantly under either PL or MPL, but the slope differed depending on which fossil calibrations were used (Fig. 7). We detected various nodes in which diversification rates increased. Most of these nodes were concentrated in *Euglossa*.

The probability of detecting changes in the diversification patterns of orchid bees suggested that a model of constant diversification (model A) had a log-likelihood of  $-366.764$  [Akaike information criterion (AIC) = 735.527;  $\delta = 0.183$ ]. A model in which diversification constantly increased or decreased following the Weibull law (model B) had a log-likelihood of  $-359.083$  (AIC = 722.167,  $\alpha = 0.168$ ,  $\beta = 1.30$ ). A model in which diversification abruptly changed with a breakpoint (model C) at 4.0 Mya had a log-likelihood of  $-390.523$  (AIC = 785.045;  $\delta 1 = 0.157$ ,

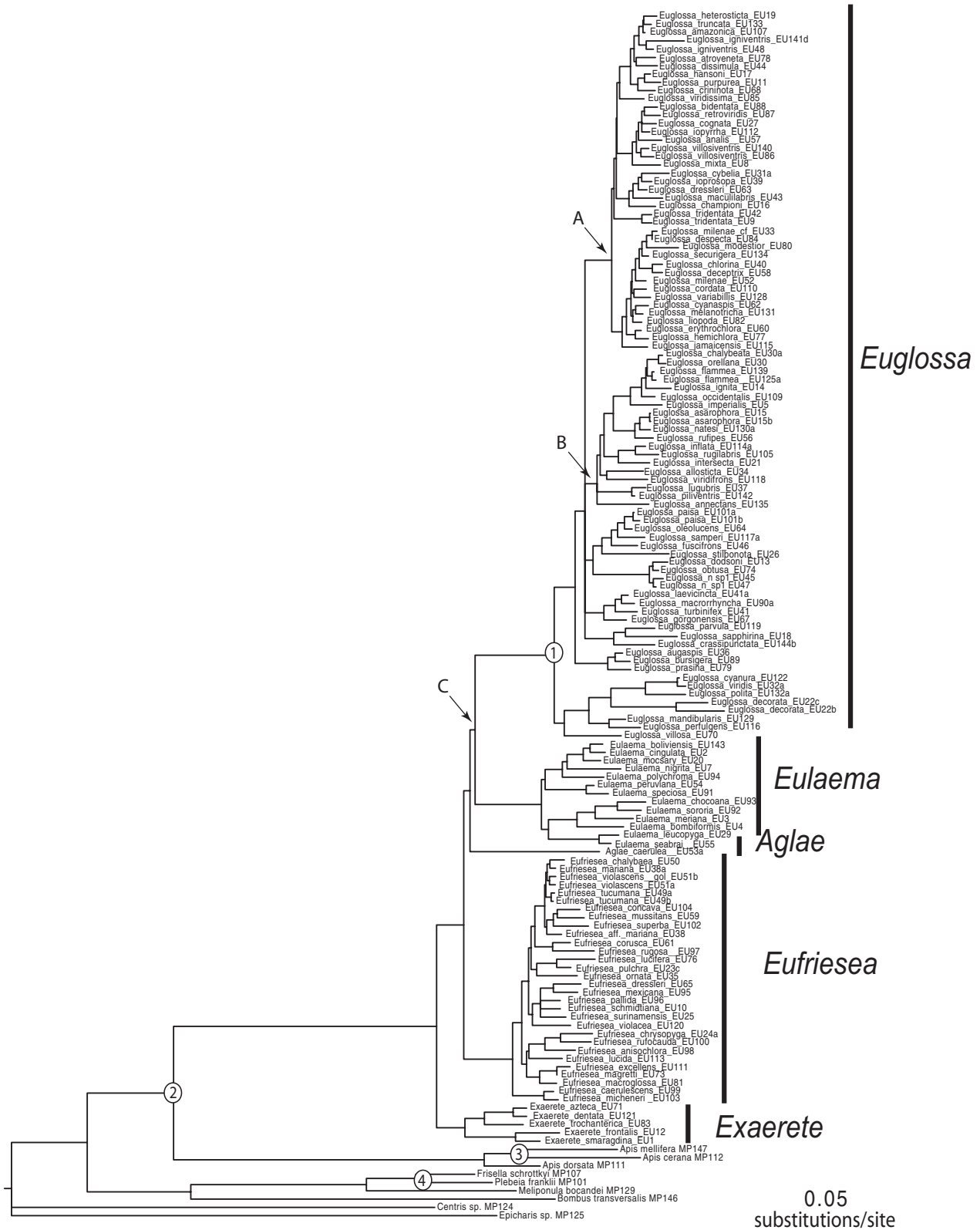
$\delta 2 = 0.079$ ). To discern among these alternative hypotheses, we performed likelihood ratio tests, which suggested that diversification following a Weibull law (model B) gave a significantly better fit to our data than the other models ( $\chi^2 = 15.361$ , d.f. = 1,  $P = 1 \times 10^{-4}$ ). Our parameter estimates for model B suggested that  $\beta > 1$ , and thus the diversification of orchid bees has decreased towards the Recent (Supporting Information Table S3). We also carried out a relative cladogenesis test for all slices through the tree, and found that several nodes in the genus *Euglossa* exhibited an increased diversification rate, even when applying a Bonferroni correction (Fig. 6).

#### HISTORICAL BIOGEOGRAPHY

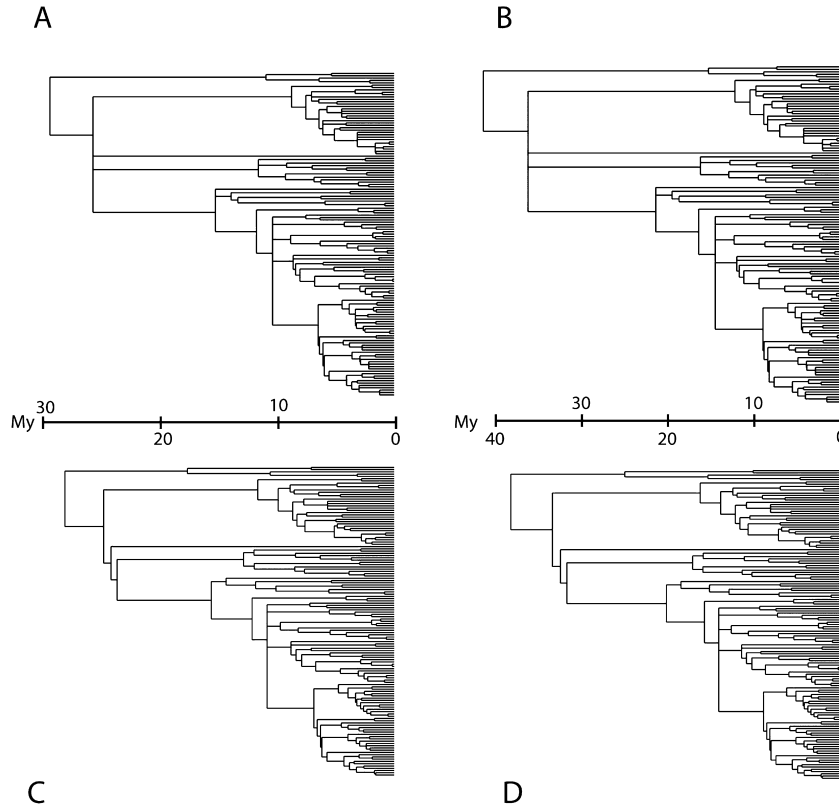
We identified seven areas of endemism for orchid bee taxa across the Neotropical region: (1) Central America; (2) Panama Endemic Zone; (3) Choco; (4) Andes; (5) Amazon; (6) Atlantic Forest; and (7) the Paraguayan Corridor (Fig. 8). The Amazon region exhibited the highest species' diversity (68 species), and the Paraguayan Corridor the lowest (17 species). On average, each species spanned  $2.41 \pm 1.77$  geographical regions. Our dataset contained 56 species endemic to a single region, and eight widespread species whose distribution spanned six or more regions (Fig. 8). Our likelihood analysis of ancestral geographical ranges based on both unrestricted (M0) and restricted (M1) DEC models yielded identical estimates of the global likelihood at the root node ( $\ln L = -438.7$ ). Similarly, dispersal rates ( $d = 0.05635$ ) and extinction rates ( $e = 0.01082$ ) did not vary between DEC models M0 and M1. Recurrent range splits between distinct geographical regions were observed. In five cases, single species restricted to Central America were recovered as sister to larger clades that were either restricted to South America or had widespread distributions (Fig. 8; nodes highlighted with red circles). In addition, we observed recurrent range splits connecting the Amazon and the Paraguayan Corridor, the Amazon and the Atlantic Forest, and the Amazon and the Andes (Fig. 8).

Our altitudinal dataset contained 57 lowland species, 23 montane species and 43 widespread species (Fig. 8). The likelihood range reconstruction of altitudinal ranges using DEC models M0 and M1 yielded identical estimates for the global likelihood at the root node ( $\ln L = -152.6$ ), dispersal rates ( $d = 0.1066$ ) and extinction rates ( $e = 8.136 \times 10^{-4}$ ). Our analysis using the DEC model M1 yielded 14 montane monophyletic lineages (Fig. 8). In all but two cases (*Euglossa ioprosopa* and *E. laevicincta*), montane lineages either gave rise to additional montane lineages, or failed to diversify further. On average, montane clades contained two lineages per clade. Montane





**Figure 4.** Phylogram of consensus tree obtained from a Bayesian tree search using a partitioned optimization model (see text for details). Branch lengths were calculated via maximum likelihood under a GTR + I +  $\Gamma$  model of sequence evolution. Numbers indicate the position of fossil calibrations; letters indicate nodes referred to in Supporting Information Table S2.



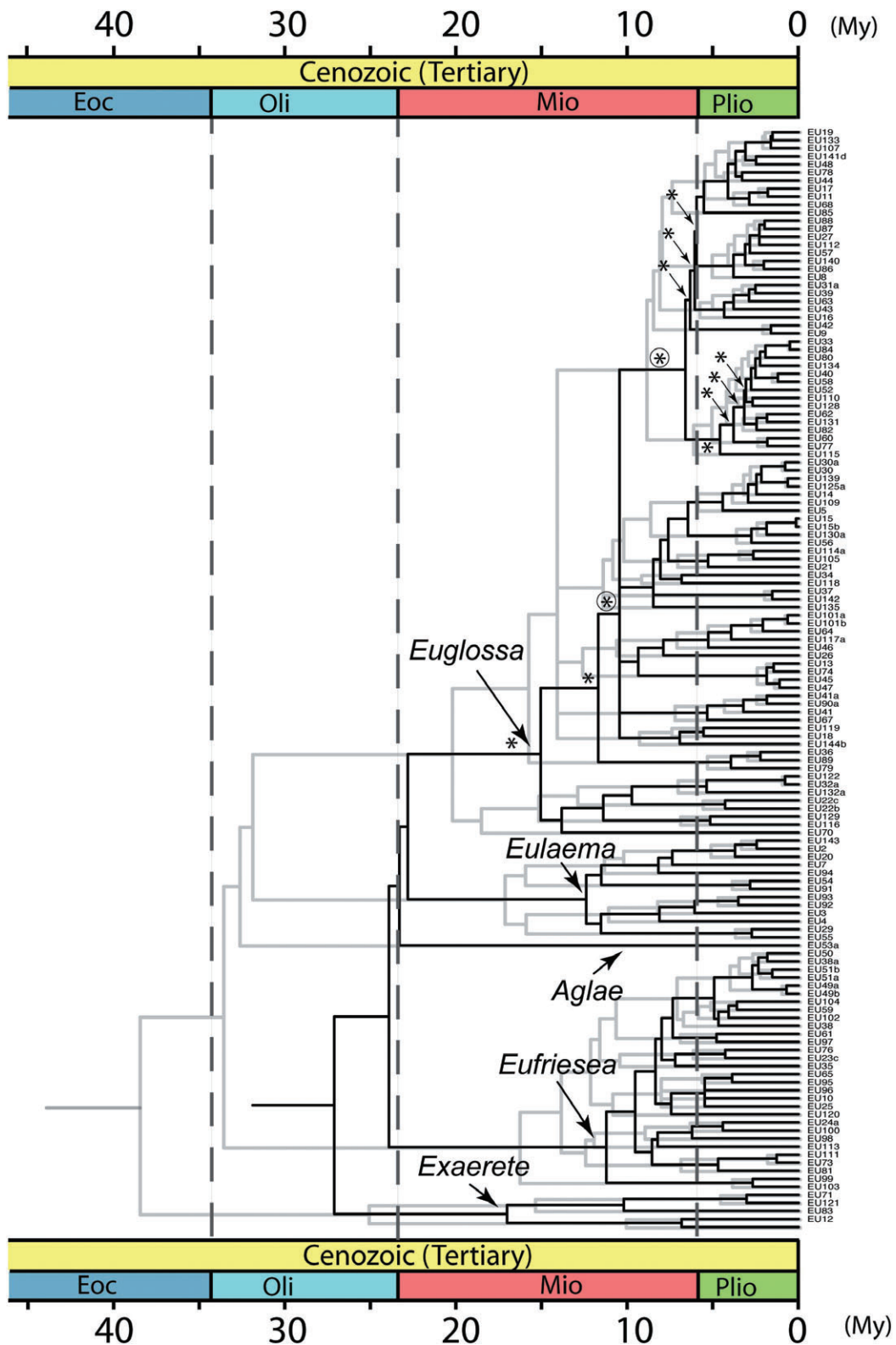
**Figure 5.** Chronograms of orchid bees obtained via minimum path length (A, B) and penalized likelihood (C, D) employing the younger minimum ages (A, C) and older ages (B, D) of fossils used to calibrate molecular clocks.

colonization events coincided with either geographical range shifts from the Amazon to the Andes, or included nonrange shifts within Central America (Fig. 8; grey circles). The species *E. ioprosopa* and *E. laevicincta* were unambiguously reconstructed as secondary lowland colonization events derived from exclusively montane lineages (relative probabilities of 0.98 and 0.99, respectively; Fig. 8). On average, the ages of the most recent common ancestor of exclusive montane lineages were  $3.88 \pm 2.7$  Myr and  $5.9 \pm 4.2$  Myr, depending on whether we used the younger or older set of fossil calibrations. Among the 14 montane lineages recovered in our analyses, three were reconstructed as derived from widespread lineages and 10 from exclusive lowland lineages (Fig. 8).

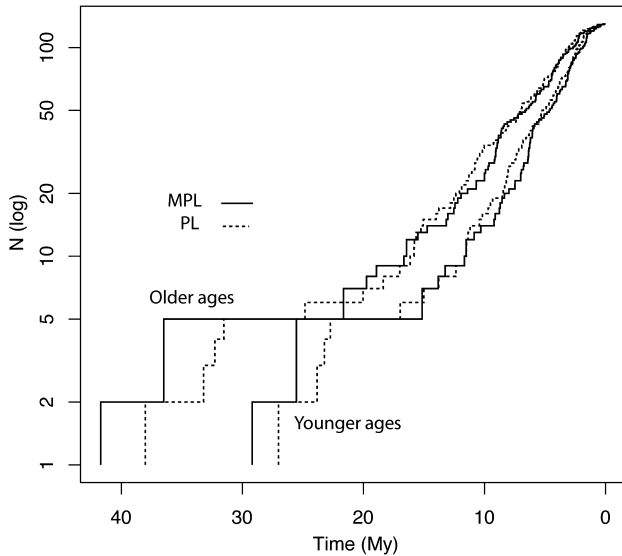
#### MORPHOLOGICAL DISPARITY

We obtained DTT plots for both individual morphological traits and all traits simultaneously. The results showed substantial variation in the average subclade disparity among traits (Fig. 9). Body width (as measured via intertegular distance) had the lowest average trait disparity with respect to simulated null models of character evolution (inferred

using Brownian motion and speciation models of character evolution), as indicated by the lowest values of the area between the observed and null curves (Supporting Information Table S4; Fig. 9). The average empirical disparity calculated for tongue length was lower between subclades (early on during diversification), but showed a marked disparity burst against both Brownian and speciation null curves at the middle of the time scale (Fig. 9). This time period corresponds well with the origin of the species-rich and morphologically diverse genus *Euglossa*. Moreover, the observed disparity in tongue length decreased towards the tip branches, where the average dissimilarity was substantially lower than predicted by both Brownian and speciation null models (Fig. 9). Conversely, body mass had an intermediate average subclade disparity, as indicated by the close proximity of both the empirical and simulated curves (Fig. 9). However, as with the patterns observed for tongue length, there was a spike in body mass disparity around the middle of the relative time scale. Although the observed disparity in body mass decreased below the null closer to the tips, a large disparity burst was recovered (Fig. 9).



**Figure 6.** Fossil-calibrated chronogram of the tribe Euglossini, inferred from a single topology obtained via Bayesian methods (see text), and calibrated via penalized likelihood with two sets of age constraints corresponding to the younger (darker lines) and older (lighter lines) fossil age boundaries. Asterisks subtend the nodes for which a diversification increase was noted using the relative cladogenesis test, even when applying a Bonferroni correction (circled asterisks).



**Figure 7.** Semi-logarithmic lineage-through-time (LTT) plot depicting diversification patterns of orchid bees. LTT plots obtained via penalized likelihood (PL) and minimum path length (MPL) are superimposed for comparison. Two sets of ages were used to calibrate the molecular clock analysis: those corresponding to older and younger age boundaries for each fossil record used in the calibration of the molecular clock.

When we used all the morphological variables simultaneously to calculate DTT, the observed average trait disparity accumulated at a slower rate than expected by the Brownian null model, but faster than suggested by the speciation null curve (Fig. 9). Finally, we observed an inverse relationship between both body mass and body size and the species' diversity within each genus (Fig. 10).

## DISCUSSION

The phylogenetic analyses presented here spanned the entire tribe Euglossini, including all genera, all subgenera, most species' groups and about two-thirds of the known species. Our findings shed new light on the phylogenetic relationships, time of divergence, morphological diversification and historical biogeography in orchid bees. Our analyses illustrate the complex interplay between geological events, lineage diversification and trait evolution in the New World tropics.

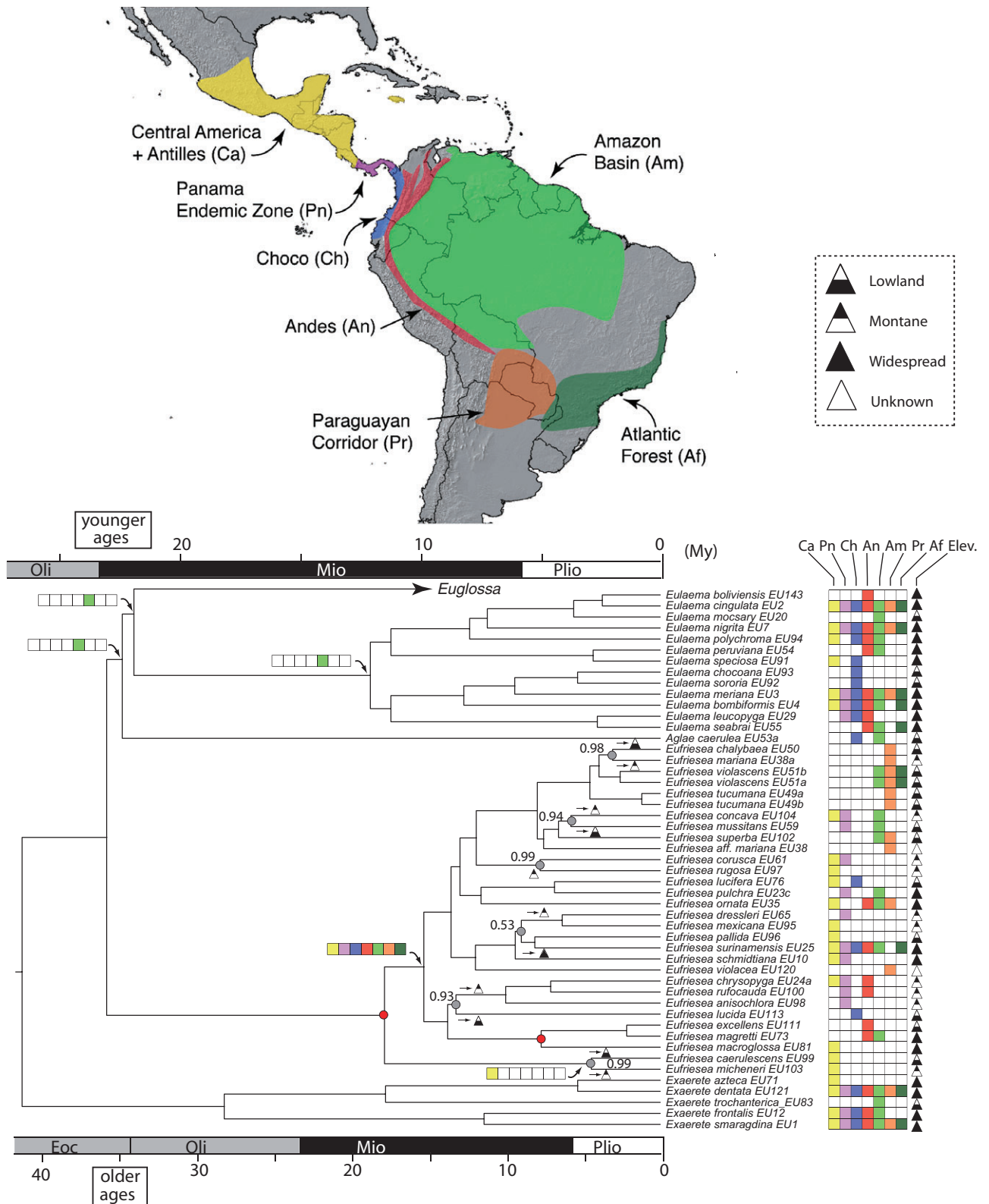
### ORCHID BEE SYSTEMATICS

Our results indicate that the cleptoparasitic genus *Exaerete* is sister to the remaining orchid bee genera. Because a reversal from nest-parasitism to nest-building is unlikely (Danforth, 2002; Michener, 2007)

and cleptoparasites depend on nest-building species to lay their own eggs, it is unlikely that the most recent common ancestor of Euglossini was a nest-parasite. Thus, our results require either the existence of basal nest-building orchid bee lineage(s) that went extinct, or stem lineage(s) leading to extant *Exaerete* that switched from nest-building to cleptoparasitism. We speculate that the former scenario is more likely.

Previous systematic studies have addressed the relationships among orchid bee genera, but none of these have suggested the scenario proposed here (Fig. 2). Our results differ from those using morphological characters (Kimsey, 1987; Engel, 1999a; Oliveira, 2006), particularly in the relative position of *Exaerete* (basal in our study). Such a discrepancy most probably stems from the differences in taxonomic sampling and the number of characters used (< 37). Notably, our results are incongruent with the generic relationships proposed by Michel-Salzat, Cameron & Oliveira (2004). Their study, which was based on both previously available morphological characters and four gene fragments, indicated that *Aglae* is sister to the remaining orchid bee genera. Although our molecular dataset overlapped partially with two of the gene fragments used by them, none of our gene partitions or combined analyses returned such topology. This incongruence may be explained by their limited sample size (18 species, half in the genus *Eulaema*).

Our phylogenetic results support previously proposed hypotheses concerning the internal relationships within orchid bee genera, including those for *Euglossa*, *Eulaema* and *Exaerete*. The internal relationships recovered for *Euglossa* correspond well with several infrageneric groups identified previously on the basis of external morphology (Dressler, 1978a, b). For instance, *Euglossa s.s.* and all of its species' groups (except *E. tridentata*) were recovered as monophyletic. Dressler (1978a) defined *Euglossa s.s.* to include male-specific synapomorphic characters, such as the rhomboid hind tibiae and the comma-shaped anterior tuft of the mid-tibia. Similarly, our results support the monophyletic status of the subgenus *Euglossella* (+ *E. villosa*), which was recovered as sister to the rest of *Euglossa*. The subgenus *Euglossella* contains some of the most atypical species in the genus, including *E. decorata*, *E. perfulgens* and related species, which constitute the only *Euglossa s.l.* with a brown or black integument that resembles the stingless bees of the genus *Melipona* (Hinojosa-Diaz & Engel, 2007). *Euglossa villosa* constitutes the monotypic subgenus *Dasystilbe*, and is restricted to Central America and Mexico. This species was recovered as sister to the subgenus *Euglossella* using a locus-specific model of sequence evolution, but as



**Figure 8.** Dated chronogram of orchid bees (split into two panels) showing extant and selected reconstructed geographical and altitudinal range distributions. Estimated model-based LAGRANGE ancestral splits are shown only for selected nodes, together with their relative probabilities.

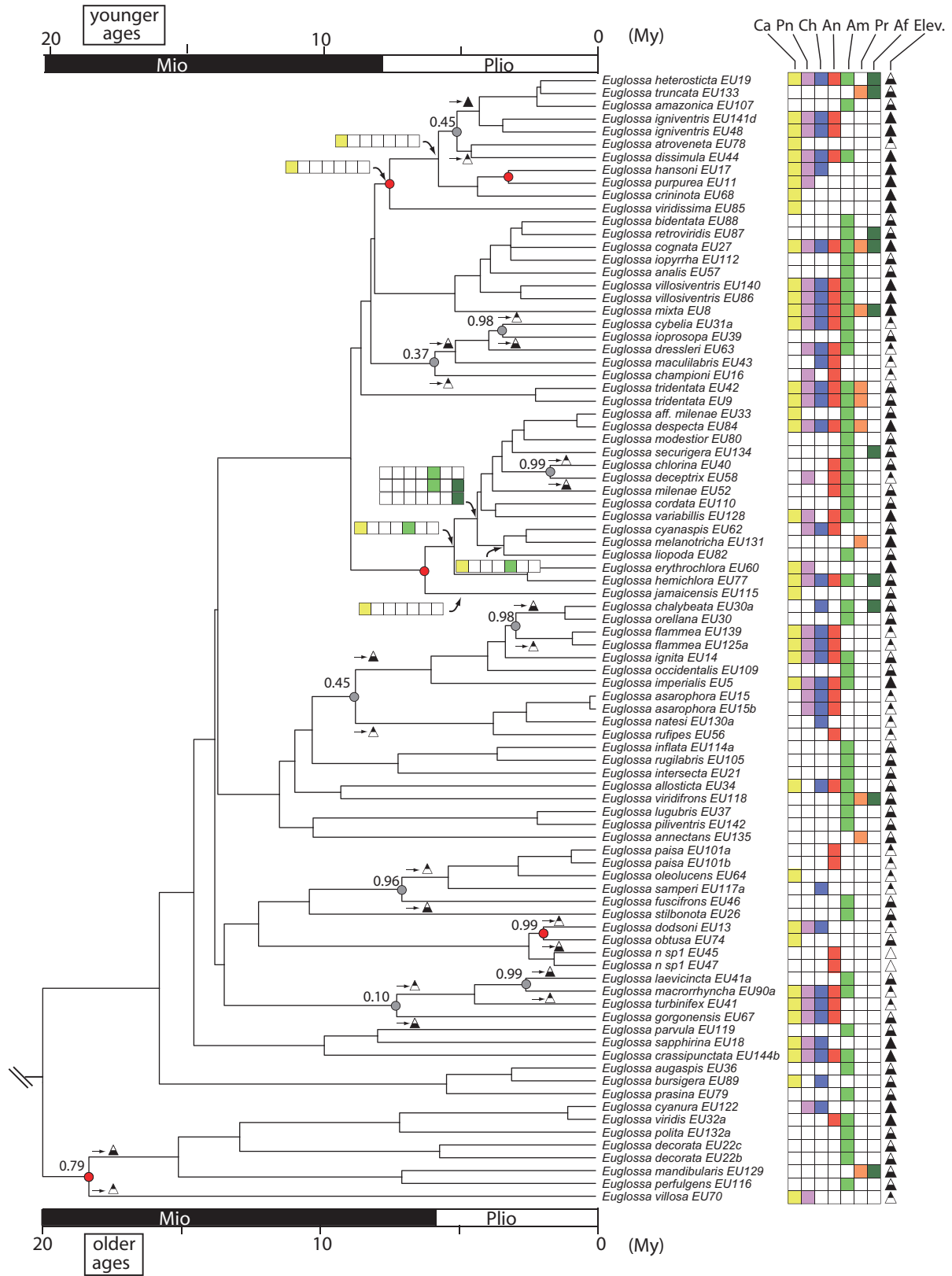
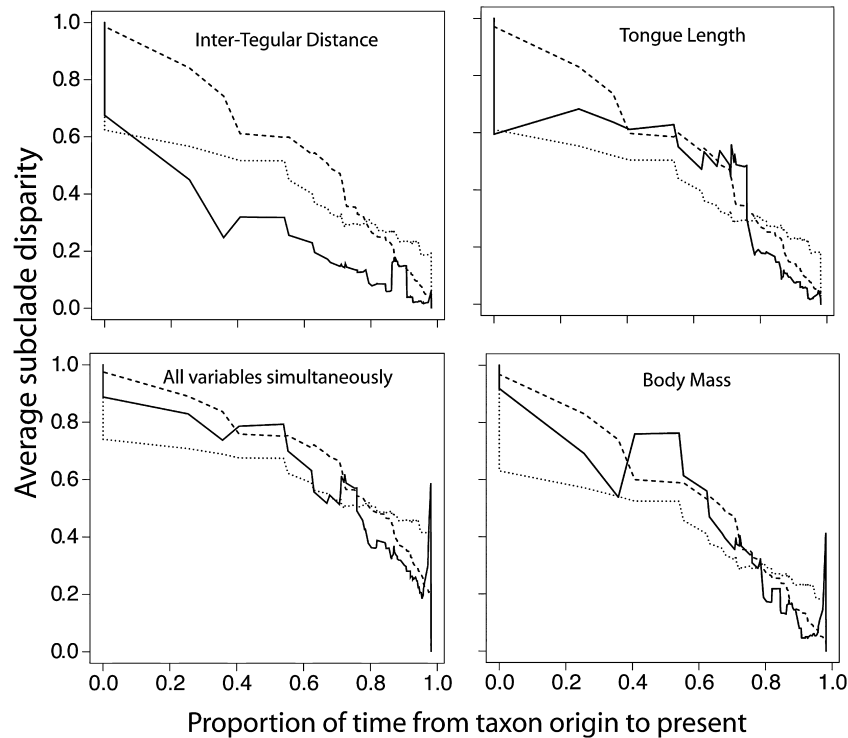


Figure 8. Continued



**Figure 9.** Disparity-through-time plot of morphological traits analysed both separately and simultaneously. Full lines correspond to the empirical values, broken lines to the simulated Brownian motion null model and dotted lines to the simulated speciation null model.

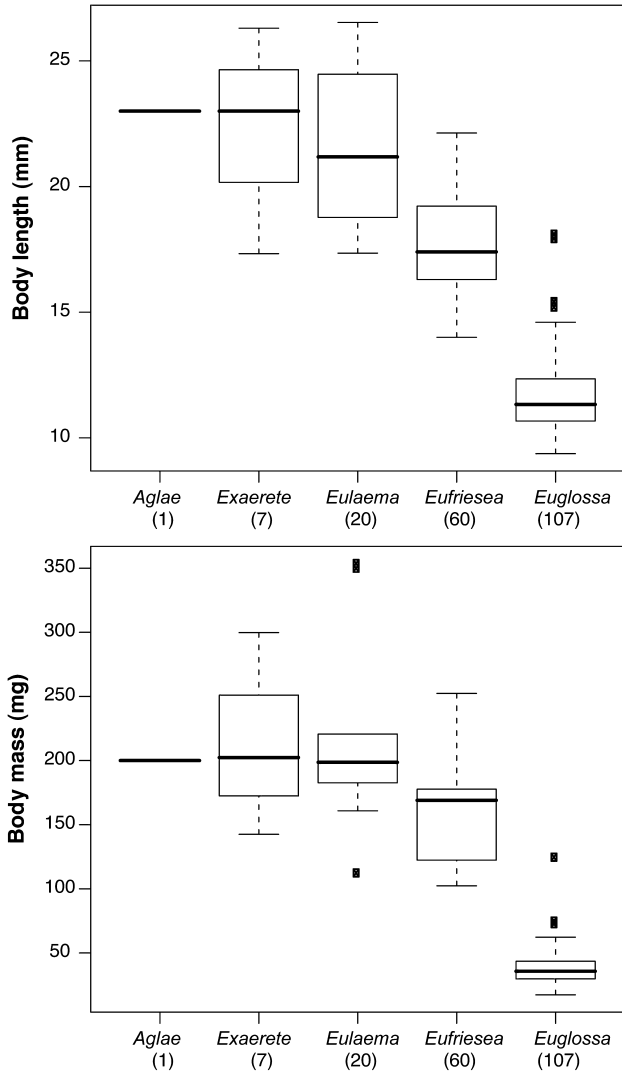
sister to the rest of *Euglossa* using a single (GTR + I +  $\Gamma$ ) model of sequence evolution for all loci. Our phylogenetic analyses also suggest that the subgenera *Glossura* and *Glossurella* in the genus *Euglossa* are paraphyletic. The subgenus *Glossura* is characterized as containing large bees with long mouthparts, labrum longer than wide and biconvex scutellum (Moure, 1967, 1969), and this was later revised to include species with long, slender bodies and elongated mouthparts (Dressler, 1978a). *Glossurella* was erected by Dressler (1982a, b) to contain both small and large bees with relatively long tongues, slender body sizes and semicircular depressions on the second sternite of males. Interestingly, both *Glossura* and *Glossurella* are not reciprocally monophyletic. Instead, our results suggest that all species from each group possessing a relatively large body size and tongue length exceeding the body length fall into a single, highly supported monophyletic clade (Fig. 3). Moreover, this clade is further supported by the bilobed gonostylus of the male genitalia (Ospina-Torres *et al.*, 2006). The remaining species in the subgenus *Glossura*, which constitute the *bursigera* species' group (Dressler, 1982c), were recovered as paraphyletic.

The two subgenera proposed previously for *Eulaema* were recovered as monophyletic, except for

the placement of the species pair composed of *E. peruviana* and *E. speciosa*. These results concord well with previously proposed hypotheses based on morphological characters (Moure, 2000; Oliveira, 2006), and are congruent with the intermediate position of *E. peruviana* between the two subgenera (Oliveira, 2006).

The phylogenetic relationships within the clepto-parasitic *Exaerete* reflect previous hypotheses about the internal relationships of the genus (Engel, 1999a; Anjos-Silva *et al.*, 2007). *Exaerete trochanterica* was formerly included in the *frontalis* species' group (Kimsey, 1979), but our analyses concord with its placement in the *dentata* group (Anjos-Silva *et al.*, 2007).

Most of the species' groups proposed for the genus *Eufriesea* were not recovered as monophyletic, which is not surprising given the taxonomic difficulties in this genus and the intergradation of species' groups. In fact, species' groups were originally conceived to aid in species' identifications and not as potential subgenera (Kimsey, 1982). Only a handful of the species' pairs identified by Kimsey (1982) were recovered as closely related in our phylogenetic analyses. In addition, a similar pattern to that found in *Euglossa* was obtained for *Eufriesea*. The first branching of the genus corresponds to that between the



**Figure 10.** Boxplots of body mass and body size grouped by orchid bee genera. Numbers in parentheses indicate the approximate number of total species within each clade.

species' pair *Eufriesea caerulescens* + *E. micheneri* (both restricted to Central America and Mexico) and the rest of the genus, which is distributed widely across the Neotropical region.

DIVERSIFICATION PATTERNS

The molecular clock analyses presented here suggest that orchid bees are a relatively young group, with all extant lineages sharing a most recent common ancestor during the Miocene–Eocene periods, approximately 27–42 Mya. Our age estimates, combined with the species' diversity for each genus, indicate that orchid bees diversified rapidly, particularly at 15–20 Mya.

Our analysis of lineage diversification also indicates that net diversification rates in Euglossini

declined through time, as implied by our corrected estimates of the gamma statistic (Paradis, 1997). Numerous recent studies have used species-level molecular phylogenies to explore net diversification rates (speciation rate–extinction rate) and, in most cases, a decrease in the net diversification rate has been observed through time. This pervasive pattern has been attributed to density-dependent diversification processes that may result from a decrease in speciation rates (in the absence of extinction rates) rather than increasing extinction rates (Rabosky & Lovette, 2008). Our study adds weight to this emerging paradigm.

All species of *Exaerete* are nest-parasites of species of *Eulaema* or *Eufriesea*. However, our results suggest that extant lineages in this genus shared a common ancestor before the appearance of these current hosts. Not only has nest-parasitism in *Exaerete* potentially been stable for 20–25 Myr, but our data also suggest that ancestral host lineages of parasitic orchid bees may have gone extinct. Both *Aglae* (one species) and *Exaerete* (seven species) have the lowest species' diversity in the tribe, which may imply that cleptoparasitism imposed constraints on net diversification, as it has appeared to do in other taxa (Wiegmann, Mitter & Farrell, 1993; Pierce *et al.*, 2002). This hypothesis deserves further examination with a more robust phylogenetic sampling of groups across all bees.

Our results also suggest that a negative correlation exists between body size and the number of species per lineage independent of age (Fig. 9). For instance, *Aglae* (one species), *Eulaema* (15–20 species) and *Exaerete* (seven species) have the largest body size among orchid bees, whereas *Eufriesea* (60 species) and *Euglossa* (> 100 species) are composed of mostly medium and small bees, respectively. This could be explained by the fact that larger species have greater dispersal capabilities (Janzen, 1971; Dick *et al.*, 2004), and thus could be less prone to geographical isolation, extinction or allopatric speciation. To further explore the role of geography in determining lineage diversification, we mapped the geographical distributions of orchid bees and reconstructed their patterns of historical biogeography using explicit models of dispersal, extinction and cladogenesis.

HISTORICAL BIOGEOGRAPHY AND MORPHOLOGICAL DIVERSIFICATION

Our analysis of historical biogeography provides insights into the impact of geological events on the diversification patterns of orchid bees, and reveals a complex interaction between dispersal, local extinction and vicariance at a continental scale. The implementation of explicit DEC models had negligible effects on our estimates of global dispersal and extinction rates,



which we interpreted as evidence of congruence between our divergence time estimates and the imposed model constraints (Clark *et al.*, 2008). We observed low local extinction rates ( $e = 0.01082$ ) relative to dispersal rates ( $d = 0.05635$ ), which may suggest that extant geographical distributions and diversity patterns in orchid bees were influenced strongly by long-distance dispersal across geographical barriers.

Our analysis of ancestral range reconstruction revealed episodic dispersal events across regions that were disjunct during the Miocene–Pliocene. For example, the most basal node in the genus *Eufriesea* corresponds to a split between Central American and widespread South American lineages. Similarly, we observed several cases in which early divergences in the genus *Euglossa* coincided with splits between Central American lineages (including *Euglossa jamaicensis*; the only described strictly insular species among orchid bees) and South American or widespread lineages (red circles in Fig. 8). Assuming that extant geographical distributions reflect ancestral occurrences, it is tempting to suggest that some early diversification in orchid bees took place after populations moved across the Isthmian archipelago, or even across the Greater and Lesser Antilles, or a proto-Antillean archipelago, with subsequent dispersal and diversification events during the closing of the Panama isthmus at 2.7 Mya (Coates *et al.*, 2004). A similar scenario has been suggested for other groups of animals, including stingless bees (Camargo, Moure & Roubik, 1988). Our likelihood analysis of ancestral range reconstruction also indicates that many lineages of *Euglossa* diversified within lowland Amazonian forests, and then dispersed across the proto-Antillean Archipelago to give rise to taxa endemic to Mexico. Because we coded the presence of taxa in the Amazon region as binary characters, we ignored distinct distributions within this region. Thus, our dataset is not suited to investigate the fine-grained patterns of historical biogeography within Amazonia (e.g. Dick & Heuertz, 2008; Miller *et al.*, 2008; Santos *et al.*, 2009). Future studies should take into account known areas of endemism for Amazonian orchid bees to investigate the potential impact of Miocene marine incursions (Rossetti, Toledo & Goés, 2005) or Pleistocene forest fragmentation (Mayle, 2004) in the diversification of Euglossini.

We coupled our likelihood estimates of altitudinal ancestral distributions with fossil-calibrated divergence times, and found that montane lineages have diverged from lowland sister clades in the past ~4–8 Myr. This time range corresponds closely with the estimated time frame of the northern Andean uplift. In fact, approximately 50% of the current Andean altitude was achieved in the past 10 Myr

(Gregory-Wodzicki, 2000). Likewise, several Mesoamerican mountain ranges were formed during the Miocene, including the Sierra Madre Occidental and the Neovolcanic Axis (Becerra, 2003 and references therein). An exception to this pattern corresponds to the montane Central American and Mexican species *Euglossa villosa*, which, based on our clock estimates, diverged approximately 13–17 Mya from its Amazonian sister clade, the subgenus *Euglossella*. Interestingly, our analyses also indicate that the colonization of montane habitats by orchid bees was relatively rare, and suggest that montane lineages failed to diversify significantly. In contrast with recent studies of vertebrates (Brumfield & Edwards, 2007; Santos *et al.*, 2009), our results indicate that the colonization of the Amazon by Andean montane lineages was exceedingly rare.

Orchid bees exhibit extensive overlap in the functional traits measured across taxa. Our analysis of morphological diversification suggests that changes in body size and tongue length were small between closely related species, and thus were recovered as phylogenetically conserved. Moreover, our analysis suggests that most of the change in body size and tongue length occurred early in the diversification of orchid bees, synchronously with the origin of diverse genera. Although tongue length (Borrell, 2005) and body size (Roubik, 1989, 1992) have been shown to play a role in resource utilization in bees, our analyses suggest that such traits were not involved in the speciation of orchid bees, at least not in the recent past. However, future studies using more fine-grained analyses should consider morphological differentiation in the context of geographical distributions and community assembly.

Euglossine bees account for an important fraction of the pollination services in the Neotropical region (Janzen, 1971; Dressler, 1982a; Williams, 1982; Ramírez, 2009). For instance, several angiosperm plant families, including Costaceae, Euphorbiaceae, Orchidaceae and Zingiberaceae, are known to contain lineages that either rely heavily or depend exclusively on male and female euglossine bees for pollination (Dressler, 1982a). However, the potential impact of such plant–pollinator mutualisms on the evolutionary history of diverse angiosperm groups remains largely unexplored. Orchid bees pose an ideal case to investigate fundamental questions about the role of mutualistic interactions in the origin, evolution and diversification of diverse tropical lineages.

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## REFERENCES

- Anjos-Silva EJ, Camilo E, Garófalo CA. 2006.** Occurrence of *Aglae caerulea* Lepeletier & Serville (Hymenoptera: Apidae: Euglossini) in the Parque Nacional da Chapada dos Guimarães, Mato Grosso State, Brazil. *Neotropical Entomology* **35**: 868–870.
- Anjos-Silva EJD, Engel MS, Andena SR. 2007.** Phylogeny of the cleptoparasitic bee genus *Exaerete* (Hymenoptera: Apidae). *Apidologie* **38**: 1–7.
- Augusto SC, Garófalo CA. 2004.** Nesting biology and social structure of *Euglossa (Euglossa) townsendi* Cockerell (Hymenoptera, Apidae, Euglossini). *Insectes Sociaux* **51**: 400–409.
- Becerra JX. 2003.** Synchronous coadaptation in an ancient case of herbivory. *Proceedings of the National Academy of Sciences of the United States of America* **100**: 12 804–12 807.
- Bembé B. 2004.** Functional morphology in male euglossine bees and their ability to spray fragrances (Hymenoptera, Apidae, Euglossini). *Apidologie* **35**: 283–291.
- Bembé B. 2007.** Revision der *Euglossa cordata*-gruppe und untersuchungen zur funktionsmorphologie und faunistik der Euglossini (Hymenoptera, Apidae). *Entomofauna* **14** (suppl): 1–146.
- Borrell BJ. 2005.** Long tongues and loose niches: evolution of euglossine bees and their nectar flowers. *Biotropica* **37**: 664–669.
- Britton T, Anderson CL, Jacquet D, Lindqvist S, Bremer K. 2007.** Estimating divergence times in large phylogenetic trees. *Systematic Biology* **56**: 741–752.
- Brumfield RT, Edwards SV. 2007.** Evolution into and out of the Andes: a Bayesian analysis of historical diversification in *Thamnophilus* antshrikes. *Evolution* **61**: 346–367.
- Camargo JMF, Grimaldi D, Pedro SRM. 2000.** The extinct fauna of stingless bees (Hymenoptera: Apidae: Meliponini) in Dominican amber: two new species and redescription of the male of *Proplebeia dominicana* (Wille and Chandler). *American Museum Novitates* **3293**: 1–24.
- Camargo JMF, Moure JS, Roubik DW. 1988.** *Melipona yucatanica* new species (Hymenoptera: Apidae: Meliponinae); stingless bee dispersal across the Caribbean Arc and post-Eocene vicariance. *The Pan-Pacific Entomologist* **64**: 147–157.
- Cameron SA. 2004.** Phylogeny and biology of neotropical orchid bees (Euglossini). *Annual Review of Entomology* **49**: 377–404.
- Cameron SA, Mardulyn P. 2001.** Multiple molecular data sets suggest independent origins of eusocial behavior in bees (Hymenoptera: Apinae). *Systematic Biology* **50**: 194–214.
- Cameron SA, Ramírez S. 2001.** Nest architecture and nesting ecology of the orchid bee *Eulaema meriana* (Hymenoptera: Apinae: Euglossini). *Journal of the Kansas Entomological Society* **74**: 142–165.
- Clark JR, Ree RH, Alfaro ME, King MG, Wagner WL, Roalson EH. 2008.** A comparative study in ancestral range reconstruction methods: retracing the uncertain histories of insular lineages. *Systematic Biology* **57**: 693–707.
- Coates AG, Collins LS, Aubry MP, Berggren WA. 2004.** The geology of the Darien, Panama, and the late Miocene–Pliocene collision of the Panama arc with northwestern South America. *Geological Society of America Bulletin* **116**: 1327–1344.
- Coates AG, Obando JA. 1996.** The geologic evolution of the Central American isthmus. In: Jackson JBC, Budd AF, Coates AG, eds. *Evolution and environments in tropical America*. Chicago, IL: Chicago University Press, 21–56.
- Danforth BN. 2002.** Evolution of sociality in a primitively eusocial lineage of bees. *Proceedings of the National Academy of Sciences of the United States of America* **99**: 286–290.
- Danforth BN, Brady SG, Sipes SD, Pearson A. 2004b.** Single-copy nuclear genes recover Cretaceous-age divergences in bees. *Systematic Biology* **53**: 309–326.
- Danforth BN, Fang J, Sipes S, Brady SG, Almeida E. 2004a.** Phylogeny and molecular systematics of bees (Hymenoptera: Apoidea). Cornell University, Ithaca, NY. Available at <http://www.entomology.cornell.edu/BeePhylogeny/>.
- Danforth BN, Sipes S, Fang J, Brady SG. 2006.** The history of early bee diversification based on five genes plus morphology. *Proceedings of the National Academy of Sciences of the United States of America* **103**: 15 118–15 123.
- Darveau CA, Hochachka PW, Kenneth J, Welch C, Roubik DW, Suarez RK. 2005.** Allometric scaling of flight

- energetics in Panamanian orchid bees: a comparative phylogenetic approach. *Journal of Experimental Biology* **208**: 3581–3591.
- Dick CW, Heuertz M. 2008.** The complex biogeographic history of a widespread tropical tree species. *Evolution* **62**: 2760–2774.
- Dick CW, Roubik DW, Gruber KF, Bermingham E. 2004.** Long-distance gene flow and cross-Andean dispersal of lowland rainforest bees (Apidae: Euglossini) revealed by comparative mitochondrial DNA phylogeography. *Molecular Ecology* **13**: 3775–3785.
- Dodson CH, Dressler RL, Hills HG, Adams RM, Williams NH. 1969.** Biologically active compounds in orchid fragrances. *Science* **164**: 1243–1249.
- Dressler RL. 1967.** Why do euglossine bees visit orchid flowers? *Atas do Simpósio sobre a Biota Amazonica* **5**: 171–180.
- Dressler RL. 1968.** Pollination by euglossine bees. *Evolution* **22**: 202–210.
- Dressler RL. 1978a.** An infrageneric classification of *Euglossa*, with notes on some features of special taxonomic importance (Hymenoptera; Apidae). *Revista de Biología Tropical* **26**: 187–198.
- Dressler RL. 1978b.** New species of *Euglossa* from Mexico and Central America. *Revista de Biología Tropical* **26**: 167–185.
- Dressler RL. 1982a.** Biology of the orchid bees (Euglossini). *Annual Review of Ecology and Systematics* **13**: 373–394.
- Dressler RL. 1982b.** New species of *Euglossa*. II. (Hymenoptera: Apidae). *Revista de Biología Tropical* **30**: 121–129.
- Dressler RL. 1982c.** New species of *Euglossa*. III. The *burgigera* species group (Hymenoptera: Apidae). *Revista de Biología Tropical* **30**: 131–140.
- Dressler RL. 1982d.** New species of *Euglossa*. IV. The *cordata* and *purpurea* species groups (Hymenoptera: Apidae). *Revista de Biología Tropical* **30**: 141–150.
- Eltz T, Sager A, Lunau K. 2005a.** Juggling with volatiles: exposure of perfumes by displaying male orchid bees. *Journal of Comparative Physiology A: Neuroethology, Sensory, Neural, and Behavioral Physiology* **191**: 575–581.
- Eltz T, Roubik DW, Lunau K. 2005b.** Experience-dependent choices ensure species-specific fragrance accumulation in male orchid bees. *Behavioural Ecology and Sociobiology* **59**: 149–156.
- Eltz T, Roubik DW, Whitten MW. 2003.** Fragrances, male display and mating behaviour of *Euglossa hemichlora*: a flight cage experiment. *Physiological Entomology* **28**: 251–260.
- Eltz T, Whitten WM, Roubik DW, Linsenmair KE. 1999.** Fragrance collection, storage, and accumulation by individual male orchid bees. *Journal of Chemical Ecology* **25**: 157–176.
- Engel MS. 1999a.** The first fossil *Euglossa* and phylogeny of the orchid bees (Hymenoptera: Apidae, Euglossini). *American Museum Novitates* **3272**: 1–14.
- Engel MS. 1999b.** The taxonomy of recent and fossil honey bees (Hymenoptera: Apidae; Apis). *Journal of Hymenoptera Research* **8**: 165–196.
- Engel MS. 2001.** A monograph of the Baltic amber bees and evolution of the Apoidea (Hymenoptera). *Bulletin of the American Museum of Natural History* **259**: 1–192.
- Engel MS. 2006.** A giant honey bee from the Middle Miocene of Japan (Hymenoptera: Apidae). *American Museum Novitates* **3504**: 1–12.
- Engel MS, Hinojosa-Díaz IA, Rasnitsyn AP. 2009.** A honey bee from the Miocene of Nevada and the biogeography of *Apis* (Hymenoptera: Apidae: Apini). *Proceedings of the California Academy of Sciences* **60**: 23–38.
- Faria LRR Jr, Melo GAR. 2007.** Species of *Euglossa* (*Glossura*) in the Brazilian Atlantic forest, with taxonomic notes on *Euglossa stellfeldi* Moure (Hymenoptera, Apidae, Euglossina). *Revista Brasileira de Entomologia* **51**: 275–284.
- Footo M. 1997.** The evolution of morphological diversity. *Annual Review of Ecology and Systematics* **28**: 129–152.
- Gregory-Wodzicki KM. 2000.** Uplift history of the Central and Northern Andes: a review. *Geological Society of America Bulletin* **112**: 1091–1105.
- Harmon LJ, Schulte JA, Larson A, Losos JB. 2003.** Tempo and mode of evolutionary radiation in iguanian lizards. *Science* **301**: 961–964.
- Harmon LJ, Weir JT, Brock CD, Glor RE, Challenger W. 2008.** GEIGER: investigating evolutionary radiations. *Bioinformatics* **24**: 129–131.
- Hines HM. 2008.** Historical biogeography, divergence times, and diversification patterns of bumble bees (Hymenoptera: Apidae: *Bombus*). *Systematic Biology* **57**: 58–75.
- Hinojosa-Díaz IA, Engel MS. 2007.** Two new orchid bees of the subgenus *Euglossella* from Peru (Hymenoptera: Apidae). *Beiträge zur Entomologie* **57**: 93–104.
- Iturralde-Vinent MA, MacPhee RDE. 1996.** Age and paleogeographical origin of Dominican amber. *Science* **273**: 1850–1852.
- Janzen DH. 1971.** Euglossine bees as long-distance pollinators of tropical plants. *Science* **171**: 203–205.
- Kawakita A, Ascher JS, Sota T, Kato M, Roubik DW. 2008.** Phylogenetic analysis of the corbiculate bee tribes based on 12 nuclear protein-coding genes (Hymenoptera: Apoidea: Apidae). *Apidologie* **39**: 163–175.
- Kimsey LS. 1979.** An illustrated key to the genus *Exaerete* with descriptions of male genitalia and biology (Hymenoptera: Euglossini, Apidae). *Journal of the Kansas Entomological Society* **52**: 735–746.
- Kimsey LS. 1982.** Systematics of bees of the genus *Eufriesea* (Hymenoptera, Apidae). *University of California Publications, Entomology* **95**: 1–125.
- Kimsey LS. 1987.** Generic relationships within the Euglossini. *Systematic Entomology* **12**: 63–72.
- Maddison DR, Maddison WP. 2003.** *MacClade v.4.06* [computer program]. Sunderland, MA: Sinauer.
- Mayle FE. 2004.** Assessment of the Neotropical dry forest refugia hypothesis in the light of palaeoecological data and vegetation model simulations. *Journal of Quaternary Science* **19**: 713–720.
- Michel-Salzat A, Cameron SA, Oliveira ML. 2004.** Phylogeny of the orchid bees (Hymenoptera: Apinae:

- Euglossini): DNA and morphology yield equivalent patterns. *Molecular Phylogenetics and Evolution* **32**: 309–323.
- Michener CD. 1944.** Comparative external morphology, phylogeny, and a classification of the bees. *Bulletin of the American Museum of Natural History* **82**: 151–326.
- Michener CD. 1990.** Classification of the Apidae. *University of Kansas Science Bulletin* **54**: 75–119.
- Michener CD. 2007.** *The bees of the World*. Baltimore, MD: Johns Hopkins University Press.
- Miller MJ, Bermingham E, Klicka J, Escalante P, Do Amaral FSR, Weir JT, Winker K. 2008.** Out of Amazonia again and again: episodic crossing of the Andes promotes diversification in a lowland forest flycatcher. *Proceedings of the Royal Society London B* **275**: 1133–1142.
- Morrone JJ. 2006.** Biogeographic areas and transition zones of Latin America and the Caribbean islands based on pan-biogeographic and cladistic analyses of the entomofauna. *Annual Review of Entomology* **51**: 467–494.
- Moure JS. 1950.** Contribuição para o conhecimento do gênero *Eulaema* Lepeletier. *Dusenía* **1**: 191–200.
- Moure JS. 1967.** A check-list of the known euglossine bees (Hymenoptera, Apidae). *Atas do Simpósio sobre a Biota Amazonica* **5**: 395–415.
- Moure JS. 1969.** The Central American species of *Euglossa* subgenus *Glossura* Cockerell, 1917 (Hymenoptera, Apidae). *Revista de Biología Tropical* **15**: 227–247.
- Moure JS. 2000.** As espécies do gênero *Eulaema* Lepeletier, 1841 (Hymenoptera, Apidae, Euglossinae). *Acta Biologica Paranaense, Curitiba* **29**: 1–70.
- Moure JS, Melo GAR, Faria LRR Jr. 2007.** Euglossini Latreille, 1802. In: Moure JS, Urban D, Melo GAR, eds. *Catalogue of Bees (Hymenoptera, Apoidea) in the Neotropical Region*. Curitiba, Brazil: Sociedade Brasileira de Entomologia, 214–255.
- Nemésio A. 2007.** Three new species of *Euglossa* Latreille (Hymenoptera: Apidae) from Brazil. *Zootaxa* **1547**: 21–31.
- Nemésio A. 2009a.** Orchid bees (Hymenoptera: Apidae) of the Brazilian Atlantic Forest. *Zootaxa* **2041**: 1–242.
- Nemésio A. 2009b.** Taxonomic notes on *Euglossa* (*Glossurotopoda*) with a key to the known species (Hymenoptera: Apidae: Euglossini). *Zootaxa* **2142**: 45–56.
- Nemésio A, Silveira FA. 2007.** Diversity and distribution of orchid bees (Hymenoptera: Apidae) with a revised checklist of species. *Neotropical Entomology* **36**: 874–888.
- Oliveira ML. 2006.** Nova hipótese de relacionamento filogenético entre os gêneros de Euglossini e entre as espécies de *Eulaema* Lepeletier, 1841 (Hymenoptera: Apidae: Euglossini). *Acta Amazonica* **36**: 273–286.
- Ospina-Torres R, Parra HA, Gonzalez VH. 2006.** The male gonostylus of the orchid bee genus *Euglossa* (Apidae: Euglossini). *Zootaxa* **1320**: 49–55.
- Page M. 1999.** Inferring the historical patterns of biological evolution. *Nature* **401**: 877–884.
- Paradis E. 1997.** Assessing temporal variations in diversification rates from phylogenies: estimation and hypothesis testing. *Proceedings of the Royal Society of London B* **264**: 1141–1147.
- Paradis E, Claude J, Strimmer K. 2004.** APE: Analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**: 289–290.
- Parra HA, Ospina-Torres R, Ramírez S. 2006.** *Euglossa natesi* n. sp., a new species of orchid bee from the Chocó region of Colombia and Ecuador (Hymenoptera: Apidae). *Zootaxa* **1298**: 29–36.
- Pierce NE, Braby MF, Heath A, Lohman DJ, Mathew J, Rand DB, Travassos MA. 2002.** The ecology and evolution of ant association in the Lycaenidae (Lepidoptera). *Annual Review of Entomology* **47**: 733–771.
- Poinar G. 1998.** *Paleoeuglossa melissiflora* gen. n., sp. n. (Euglossinae: Apidae), fossil orchid bees in Dominican amber. *Journal of the Kansas Entomological Society* **71**: 29–34.
- Rabosky DL, Lovette IJ. 2008.** Density-dependent diversification in North American wood warblers. *Proceedings of the Royal Society of London B* **275**: 2363–2371.
- Ramírez S. 2005.** *Euglossa paisa*, a new species of orchid bee from the Colombian Andes (Hymenoptera: Apidae). *Zootaxa* **1065**: 51–60.
- Ramírez S. 2006.** *Euglossa samperi* n. sp., a new species of orchid bee from the Ecuadorian Andes (Hymenoptera: Apidae). *Zootaxa* **1272**: 61–68.
- Ramírez S. 2009.** Quick guide: orchid bees. *Current Biology* **19**: 1061–1063.
- Ramírez S, Dressler RL, Ospina M. 2002.** Orchid bees (Hymenoptera: Apidae: Euglossini) from the Neotropical Region: a species checklist with notes on their biology. *Biota Colombiana* **3**: 7–118.
- Rasmussen C, Skov C. 2006.** Description of a new species of *Euglossa* (Hymenoptera: Apidae: Euglossini) with notes on comparative biology. *Zootaxa* **1210**: 53–67.
- Ree RH, Smith SA. 2008.** Maximum likelihood inference of geographic range evolution by dispersal, local extinction, and cladogenesis. *Systematic Biology* **57**: 4–14.
- Ronquist F, Huelsenbeck JP. 2003.** MRBAYES 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* **19**: 1572–1574.
- Rossetti DF, Toledo PM, Goés AM. 2005.** New geological framework for western Amazonia (Brazil) and implications for biogeography and evolution. *Quaternary Research* **63**: 78–89.
- Roubik DW. 1989.** *Ecology and natural history of tropical bees*. Cambridge: Cambridge University Press.
- Roubik DW. 1992.** Loose niches in tropical communities: why are there so few bees and so many trees? In: Hunter MD, Ohgushi T, Price PW, eds. *Effects of resource distribution on animal-plant interactions*. San Diego, CA: Academic Press, 327–354.
- Roubik DW. 2004.** Sibling species of *Glossura* and *Glossurotopoda* in the Amazon Region (Hymenoptera: Apidae: Euglossini). *Journal of the Kansas Entomological Society* **77**: 235–253.
- Roubik DW, Ackerman JD. 1987.** Long-term ecology of euglossine orchid-bees (Apidae: Euglossini) in Panama. *Oecologia* **73**: 321–333.
- Roubik DW, Hanson PE. 2004.** *Orchid bees of tropical*

- America: biology and field guide*. Heredia, Costa Rica: INBIO.
- Sanderson MJ. 2002.** Estimating absolute rates of molecular evolution and divergence times: a penalized likelihood approach. *Molecular Biology and Evolution* **19**: 101–109.
- Santos JC, Coloma LA, Summers K, Caldwell JP, Ree R, Cannatella DC. 2009.** Amazonian amphibian diversity is primarily derived from late Miocene Andean lineages. *PLoS Biology* **7**: 448–461.
- Smith AG, Smith DG, Funnell BM. 1994.** *Atlas of Mesozoic and Cenozoic coastlines*. Cambridge: Cambridge University Press.
- Soucy SL, Giray T, Roubik DW. 2003.** Solitary and group nesting in the orchid bee *Euglossa hyacinthina* (Hymenoptera, Apidae). *Insectes Sociaux* **50**: 248–255.
- Swofford DL. 2003.** *PAUP\* v4b* [computer program]. Sunderland, MA: Sinauer Associates.
- Vogel S. 1963.** Das sexuelle Anlockungsprinzip der Catasetinen- und Stanhopeen-Blüten und die wahre Funktion ihres sogenannten Futtergewebes. *Plant Systematics and Evolution* **110**: 308–337.
- Vogel S. 1966.** Pollination neotropischer Orchideen durch Duftstoff-höselnde Prachtbienen-Männchen. *Naturwissenschaften* **7**: 1–2.
- Wiegmann BM, Mitter C, Farrell B. 1993.** Diversification of parasitic insects: extraordinary radiation or specialized dead end? *The American Naturalist* **142**: 737–754.
- Wille A, Chandler LC. 1964.** A new stingless bee from the Tertiary of the Dominican Republic (Hymenoptera; Meliponini). *Revista de Biología Tropical* **12**: 187–195.
- Williams NH. 1982.** The biology of orchids and euglossine bees. In: Arditti J, ed. *Orchid biology: reviews and perspectives, II*. Ithaca, NY: Cornell University Press, 119–171.
- Williams NH, Whitten WM. 1983.** Orchid floral fragrances and male euglossine bees: methods and advances in the last sesquidecade. *Biological Bulletin* **164**: 355–395.
- Winston ML, Michener CD. 1977.** Dual origin of highly social behavior among bees. *Proceedings of the National Academy of Sciences of the United States of America* **74**: 1135–1137.
- Zimmermann Y, Ramírez SR, Eltz T. 2009.** Chemical niche differentiation among sympatric species of orchid bees. *Ecology* **90**: 2994–3008.
- Zucchi RS, Sakagami F, Camargo JMFd. 1969.** Biological observations on a neotropical parasocial bee, *Eulaema nigrita*, with a review on the biology of Euglossinae (Hymenoptera, Apidae). A comparative study. *Journal of the Faculty of Science, Hokkaido University Series VI, Zoology* **17**: 271–380.

## SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article:

**Table S1.** Models of sequence evolution used for individual loci (determined via likelihood ratio test) employed in the locus-specific partitions for Bayesian phylogenetic analyses.

**Table S2.** Mean and standard deviation divergence times (in Myr) calculated for individual nodes of Euglossini via penalized likelihood (PL) and mean path length (MPL). Two sets of age constraints corresponding to the younger and older fossil age boundaries were implemented (see text for details). Numbered nodes indicate constrained nodes, and letters indicate selected nodes for which summary statistics are provided.

**Table S3.** Estimated parameters and hypothesis testing using diversification survival models of orchid bee lineages. An asterisk indicates the best-fit model for each subclade. Parameter values in parentheses correspond to the estimated errors, unless otherwise indicated.

**Table S4.** The area between the curves corresponds to the difference between the observed and the null disparity-through-time plots from Fig. 9. The average null disparity curves in each case were based on simulated data using two models of character evolution (Brownian and speciation). Positive areas indicate a higher than expected overall morphological disparity, whereas negative areas indicate that the observed disparity was lower than expected via simulations.

**Appendix S1.** Collection data, taxonomic information and National Center for Biotechnology Information (NCBI) GenBank accession numbers of samples used in the present study. Species with incomplete sequences from more than a single specimen were merged, and are indicated with an asterisk (\*).

**Appendix S2.** Mean values  $\pm$  SD of morphological measurements for a subset of orchid bee species.

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