



Can environmental complexity predict functional trait composition of ground-dwelling ant assemblages? A test across the Amazon Basin

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ABSTRACT

Environmental gradients may influence species distributions by filtering their functional traits, resulting in a correspondence between community functional composition and local environmental conditions. We used a vegetation gradient as an indicator for environmental complexity to test whether it predicted the morphological composition of ground-dwelling ant assemblages across geographic extents. The sampling design covered 126 plots distributed across eight sampling sites along a broad environmental gradient in the Amazon Basin. Plots covered different phytophysiognomies that have a strong relation with forest biomass and, consequently, litter production. We selected six morphological traits related to ant foraging strategies and behavior. Generalized linear mixed models were used to predict how environmental complexity affects trait composition of ground-dwelling ant assemblages. Structurally less complex environments (eg. Amazonian savannah) harboured more species of smaller ants, with relatively smaller mandibles and relatively larger eyes. In more complex environments (eg. dense ombrophylous forest), there were more ant species of larger size, with relatively larger mandibles and relatively smaller eyes. No relationship was detected between relative femur length and the environmental gradient investigated. The functional approach focused on individual traits may illuminate which ant foraging strategies are best adapted to a particular habitat. Our data reveal that the morphological composition of ground-dwelling ant assemblages responds clearly to environmental complexity suggesting that certain ant characteristics offer ecological advantages to particular species in particular habitats.

1. Introduction

One of the main objectives of ecology is to understand the relationship between organisms and the environment (Vellend, 2016). To address this, ecologists try to identify which mechanisms structure assemblages in different habitats and scales (Levin, 1992; McGill, 2010; Sobral and Cianciaruso, 2012). Locally, environmental conditions may play a key role in selecting species exhibiting similar morphological, behavioral or reproductive traits (Keddy, 1992). This suggests that along a gradient with distinct environmental conditions, the functional traits of species should gradually change so that assemblages have different functional compositions on each side of the environmental gradient (Weiher et al., 2011; Sommer et al., 2014; Bishop et al., 2016; Peters et al., 2016). This process forms the basis of functional ecology (McGill et al., 2006).

Several studies have investigated how habitat conditions can play a role in the selection of functional traits in invertebrates and sometimes showing strong associations between morphology, habitat type (Barton et al., 2011; Gibb and Parr, 2013; Yates et al., 2014; Schofield et al., 2016; Peters et al., 2016; Graça et al., 2017). In particular, ants are a useful group for investigating this question given their extensive morphological variation and wide ecological distribution (Silva and Brandão, 2010; Arnan et al., 2017). For example, habitat complexity can often filter ant species based on morphological traits (Gibb and Parr, 2013; Yates et al., 2014; Schofield et al., 2016). Ants with proportionally longer legs may move faster and more efficiently in less complex habitats. Otherwise, proportionally smaller legs allow better access to microhabitats in rugose substrates, such as litter interstices (Kaspari and Weiser, 1999; Farji-Brener et al., 2004; Gibb and Parr, 2013). However, a field test found smaller ant species with

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proportionally smaller legs being more abundant in habitats with lower litter amount (Parr et al., 2003). Another trait often related to environmental complexity is eye size (Schofield et al., 2016). Ant species with proportionally smaller eyes are associated in habitats with low light availability (Weiser and Kaspari, 2006; Schofield et al., 2016). However, in more complex environments such as dense forests, where there is lower light availability at ground layer, larger eyes may indicate a greater demand in the capacity of perception of the environment (Yates et al., 2014). Similarly, the average mandible size of ant assemblages may respond to increased availability of resources. Ants with larger mandible may access prey of different shapes and sizes (Fowler et al., 1991; Schofield et al., 2016). Thus, these studies reveal the link between ant traits and the environment and show the ambiguity in ant responses.

While there are clear differences between the functional composition of ant assemblages inhabiting contrasting biomes (Schofield et al., 2016), and disturbed vs. undisturbed habitats (Yates et al., 2014), biomes are not homogeneous and may also encompass significant variation in ant functional composition. Moreover, ant species density peaks in the tropics (Dunn et al., 2009) but the drivers of ant functional composition in tropical forest such as the Amazon are largely unknown. For instance, the Amazon Basin shows a wide landscape structure variation ranging from plant- and nutrient-rich non-flooded ombrophilous forest (Chauvel et al., 1987; ter Steege et al., 2013) to white-sand forest on nutrient-poor soil which is under the influence of periodic flooding (do Vale et al., 2014). Thus, phytophysiognomy can vary from Amazonian savannah to dense forest resulting in many vegetation classes with different levels of environmental complexity (Instituto Brasileiro de Geografia e Estatística-IBGE, 2012; Emilio et al., 2010). These vegetation classes have a strong relation with forest biomass (Saatchi et al., 2007) which in turn correlates with the production of litter (Aragão et al., 2009), a key factor for several ant functional traits (Kaspari and Weiser, 1999; Weiser and Kaspari, 2006; Schofield et al., 2016). In the present study, we used a large standardized database (Baccaro et al., 2013; Oliveira et al., 2009; Souza et al., 2012, 2016) that covers several phytophysiognomies in the Brazilian Amazonia to ask how the environmental complexity gradient of the Amazon Basin may affect the functional trait composition of ground-dwelling ant assemblages.

We selected six ant morphological traits (Weber's length, head length and width, hind femur length, eye length and mandible length) related to foraging behavior and predatory specialization (Silva and Brandão, 2010; Schofield et al., 2016) to test the following hypotheses:

- (i) less complex habitats favor larger ants species with a longer femur length, while more complex environments favor smaller body size and relative femur length, as predicted by the size-grain hypothesis (Kaspari and Weiser, 1999);
- (ii) the average eye size is relatively smaller in ant assemblages in dense forests than in ant assemblages in more open vegetation, as predicted by the light level-eye size hypothesis (Schofield et al., 2016);
- (iii) more complex environments favor more specialized predator species, represented by ants with relatively large mandibles, thus, dense forests should harbor more ground-dwelling ant species with relatively larger mandibles, as predicted by the foraging specialization hypothesis (Gronenberg et al., 1997; Schofield et al., 2016; Silva and Brandão, 2010).

2. Materials and methods

2.1. Study site

Ground-dwelling ants were sampled at eight sampling sites maintained by the Brazilian Biodiversity Research Program, PPBio (Costa and Magnusson, 2010). Three of these sites (Maracá Ecological Station, Cauamé Campus and Viruá National Park) are located in the Roraima



Fig. 1. Map of the study area. Location of the eight sampling sites (white balloons) along a broad environmental gradient in the Brazilian Amazon.

State (extreme north of Brazil) and the other five (Ducke Reserve, Manaquiri, Orquestra, Capanã and Jari) are located in the Amazonas State (Fig. 1).

The Maracá Station (Maracá) is located on an island on the Uraricoera River which is an ecotone zone between savannas and Amazonian forests (Souza et al., 2012). Cauamé Campus (Cauamé) belongs to the Federal University of Roraima and presents typical savanna vegetation with a markedly dry period (Instituto Brasileiro de Geografia e Estatística-IBGE, 2012). The Viruá Park (Viruá) is located in low plains subject to flood, with some hills with moderate altitudes (Souza et al., 2012).

The Ducke Reserve (Ducke) is covered by undisturbed dense forest on moderately rugged terrain, with small perennial streams in the valleys (Chauvel et al., 1987). The sites located along the BR-319 highway in the interfluvium between the Purus and Madeira rivers harbor open and dense forests mosaics (Magnusson et al., 2013). The Manaquiri site is characterized by seasonal floods in soils with small intermittent streams (Baccaro et al., 2013).

These sites encompass a broad environmental gradient of approximately 1,050 km of extent (between the first and last sampling point, in the North/South direction) (Fig. 1). Sampling sites, geographic coordinates, phytophysiognomies found at each site, number of plots and number of samples per plot is summarized in Table 1.

2.2. Experimental design

The experimental design of the sampling sites followed the RAPELD system (Magnusson et al., 2013). Each grid-shaped system gives access to permanent plots where several organisms and environmental variables can be surveyed (Costa and Magnusson, 2010). In the research sites the grid consists of parallel trails of 5 km located 1 km apart. In each trail, five permanent plots are distributed. Each plot has 250 m of extension per 1 m of width, following the altitudinal contours to minimize within plot topo-edaphic variation. The number plots per sampling site ranged from five to 30 totaling 127 plots. The sampling effort and the spatial distribution of the subsamples were the same among all plots (Table 1).

2.3. Ant sampling

The ants were collected between September 2006 and June 2012 and all the collections were carried out in the respective dry season. The

Table 1

Sampling sites ordered from the north to the south in Brazil, their geographic coordinates, phytophysiognomy, mean annual cumulative rainfall, number of plots and number of samples per plot.

Sites	Geographical coordinates	Phytophysiognomies	Number of plots	Samples per plot
Maracá	03° 23' 45.60" N 61° 28' 24.61" W	Open ombrophylous forest, Semideciduous forest, Deciduous forest, Campinarana forest	30	10
Cauamé	02° 52' 01.20" N 60° 38' 02.40" W	Open savana	11	10
Viruá	01° 27' 01.72" N 61° 01' 28.96" W	Open ombrophylous forest, Campinarana forest Seasonal campinarana, Seasonal shrubby campinarana	30	10
Ducke	02° 58' 17.51" S 59° 57' 28.43" W	Dense ombrophylous forest	30	10
Manaquiri	03° 41' 24.00" S 60° 18' 36.00" W	Open ombrophylous forest	10	10
Orquestra	04° 59' 04.16" S 61° 34' 14.20" W	Dense ombrophylous forest	5	10
Capanã	05° 37' 31.92" S 62° 10' 56.92" W	Dense ombrophylous forest	5	10
Jari	05° 57' 26.94" S 62° 29' 20.51" W	Dense ombrophylous forest	5	10

ants were sampled with pitfall traps, placed at 25 m intervals of each other, totaling 10 samples per plot. The pitfalls (plastic cups 8 cm long by 9.5 cm in diameter) were buried until their edge remained at the same level as the ground and were filled with 1/3 of 70% alcohol and a few drops of detergent. After 48 h the traps were collected, the ants were sorted and placed in 90% alcohol to preserve the material in the laboratory (Souza et al., 2016). Ants were classified up to genus level with the use of taxonomic keys (Baccaro et al., 2015). Later, the ants were morphotyped and, when possible, were identified to the species level using available taxonomic keys or by specialists and by comparison with specimens deposited in zoological collections. The voucher specimens were deposited at the INPA Invertebrate Collection.

2.4. Functional traits

Six morphological traits related to foraging strategies were measured from each ant species/morphospecies. We followed the measurement protocol described in Schofield et al. (2016):

1. Weber's length - it was measured from the anterodorsal margin of the pronotum to the posteroventral margin of the propodeum (Hölldobler and Wilson, 1990). This measure is an indicator of body size, which is a key life history trait (Kaspari, 1996; Kaspari and Weiser, 1999).
2. Hind femur length - straight-line distance from the femur's insertion into the coxae, and its attachment to the tibia (Kaspari and Weiser, 1999). The femur length is related to habitat complexity. Larger legs may allow faster and more efficient locomotion in relatively flat habitats; however, they become a disadvantage by increasing the cross-sectional area of the ant's body and limiting its access to interstitial environments (Hurlbert et al., 2008; Kaspari and Weiser, 1999).
3. Eye length - the measurement was made from the upper to lower border covering the maximum length of the eye. Eye size is an important feature in the search for food resources (Weiser and Kaspari, 2006).
4. Mandible length - in front view, it was measured at the point of the clypeus insertion to the apex of the mandible. Mandible size is related to the diet and predatory specialization of ants (Schofield et al., 2016). Larger mandible allows access to different shapes and sizes of prey (Fowler et al., 1991).
5. Head length - it was measured from the maximum length from the apex of the head to the ventral/anterior-most portion of the clypeus. Head length is a proxy for body mass and may contribute to the cross-sectional area of the ant body (Kaspari and Weiser, 1999).
6. Head width - in frontal view, the measurement was made of the maximum width of the head, without considering the eyes. Head width is related to ants' foraging strategy so that larger heads can support larger mandibles and thereby prey upon larger prey (Fowler et al., 1991).

All morphological traits, except for mandible and Weber's length, were standardized by head length (trait value/head length) to obtain the value of the trait relative to the size of the ant species (Schofield et al., 2016). Head length was used for standardization because it is used to measure the cross-sectional area of the ant's body (how much the leg of the ant can exceed the height of its head), in addition to being highly correlated with Weber's length (Pearson correlation = 0.975). Mandible length was standardized by head width because the muscular tissue of the mandible, which occupies much of the internal volume of the head, divides space with the mandible glands which are located on the lateral margins of the ant's head (Fowler et al., 1991; Bishop et al., 2015). Head length and head width were used only for the standardization of the traits and were not tested against the environmental gradient. The number of individuals measured by species ranged from one to six specimens, depending on the abundance of the species in the samples. For the most abundant species we sampled individuals from different locations and vegetations to encompass the natural variation that may exist in functional traits along the environmental gradient (Appendix Table B.1). Measurements were only taken for workers and, for the polymorphic species, only minor workers were sampled. Measurements were used to obtain trait means for each species. All measurements were made using Leica M 125 stereomicroscope with the aid of Leica Application Suite, Version: 4.8.0.

2.5. Environmental gradient

To evaluate how the environmental gradient may influence the functional composition of ant assemblages, we used the phytophysiognomy (Table 1) as a proxy to the amount and heterogeneity of litter. The phytophysiognomy unit is defined as a vegetation type with its own characteristics of structure and species composition. In the Amazon, the phytophysiognomy has a strong connection with precipitation, tree height and forest biomass (Saatchi et al., 2007; IBGE, 2012). Soil litter accumulation is a factor that can vary greatly within a few meters within the forest, but on average, it changes more between vegetations than within vegetation types (Villela and Proctor, 1999). Thus, given the scale of the work, we used the phytophysiognomy of the area sampled as an indicator of the level of environmental complexity for ground-dwelling ants. We used information on vegetation and climate characteristics, average height of trees and average percentage of vegetation cover of each phytophysiognomies unit to represent this gradient on an ordinal scale (Fig. 2).

Based on information from these descriptors, the phytophysiognomy were ordered according to the level of environmental complexity. Less complex vegetations were assigned smaller values; the more complex the vegetation the higher the value assigned as seen in Appendix Table A. The data on phytophysiognomy for each site were taken from the PPBio site {<https://ppbiodata.inpa.gov.br/metacatui/>} and were based on the Brazilian Vegetation Classification System developed by the Brazilian Institute of Geography and Statistics (in



Fig. 2. Ordination of phytophysiognomies according to environmental complexity. 1. Open savanna, 2. Seasonal shrubby campinarana, 3. Seasonal campinarana, 4. Campinarana forest, 5. Deciduous forest, 6. Semideciduous forest, 7. Open ombrophylous forest and 8. Dense ombrophylous forest. Adapted from Graça et al. (2017).

Table 2

Summary of Generalized Linear Mixed Models (GLMM) results relating community-trait mean (CTM) of ground-dwelling ants to environmental complexity. Except for Weber's length, all traits were standardized relative to head size (see Methods for details). a: model intercept; b: model slope, R^2_m : variance explained by predictor; R^2_c : variance explained by predictor plus random factor; P: test probability.

Functional traits	Models				
	a	b	r^2_m	r^2_c	P
Weber's length	1.531	-0.517	0.22	0.59	< 0.01
Hind femur length	1.224	-0.048	0.02	0.17	0.258
Eye length	0.204	0.051	0.32	0.46	< 0.001
Mandible length	0.658	-0.090	0.44	0.44	< 0.001

Portuguese, Instituto Brasileiro de Geografia e Estatística -IBGE). Data on mean tree height, average vegetation cover and vegetation and climate characteristics of each phytophysiognomy were taken from the Technical Manual of Brazilian Vegetation of Instituto Brasileiro de Geografia e Estatística-IBGE (2012).

2.6. Statistical analysis

All statistical analyzes were performed in the R environment for statistical computing (R Core Team, 2017; version 3.4.3), with support of packages “lmerTest” (Kuznetsova et al., 2017), “MuMIn” (Barton, 2009) and “visreg” (Breheny and Burchett, 2016). First, to understand how the environment can select species based on their traits we calculated the community-trait mean (CTM) in each plot, i. e. the mean trait value of species co-occurring in a given plot:

$$CTM_j = \frac{\sum_i^k x_i}{\sum_i^k s}$$

where x is the trait value of species i , s is number of species collected in the plot and j represents a given trait. We used a linear mixed effects model (GLMM) to evaluate whether variation in CTM across plots could be explained by environmental complexity while controlling for possible effects related to the location of the collection sites. We included sampling site as a random factor in the models to control for the

potential spatial autocorrelation between nearby collection points. Thus, collection site was declared as a random factor and environmental complexity was declared as a fixed factor in the models. One model was created for the CTM of each trait. For each model, the marginal r^2_m (variation explained only by the fixed factor) and conditional r^2_c (variation explained by the fixed and random factors) were calculated (Nakagawa and Schielzeth, 2013). The models were built with the predictor variable being environmental complexity (x) and the response variable the morphological traits (y). To evaluate which models best express the shape of the relationships, linear models were compared with models that assumed a rationalization of the standard linear equation ($Y = a + b \times 1/X$) to account for asymptotic relationships (i.e. functional composition changing along part of the environmental gradient, and then remaining constant). The Akaike's Information Criterion corrected for sample size (AICc) (Burnham and Anderson, 2002) was used to choose the best model and as the asymptotic models best represented all the relationships, only they were included in the main text (Appendix Table C and Figures). The intercept (the value of the response variable when the predictor variable is equals zero) and the slope (how much the response variable changes due to the increase of one unit in the predictor variable) of the models were calculated.

3. Results

A total of 994 ants belonging to 271 species/morphospecies were measured in the 127 plots sampled along the environmental gradient studied (Appendix Table B.2). The number of species varied considerably between the assemblages, from 4 species captured in a plot of Viruá Park to 56 species in a plot in Maracá Station. A total of 5,964 morphological measurements of the ants were made. Weber's length, relative eye length, and relative mandible length were related to the environmental complexity gradient (Table 2). Weber's length and the relative length mandible responded positively to environmental complexity. More complex areas harboured larger ants (Weber's length) and ants with relative longer mandibles (Fig. 3 a, d). The relative length eye responded negatively to the gradient, meaning that the increase in complexity decreased the relative length of the ants' eyes (Fig. 3c). Relative mandible length had the best-fitted model, followed by the relative eye length and Weber's length (Table 2). No significant

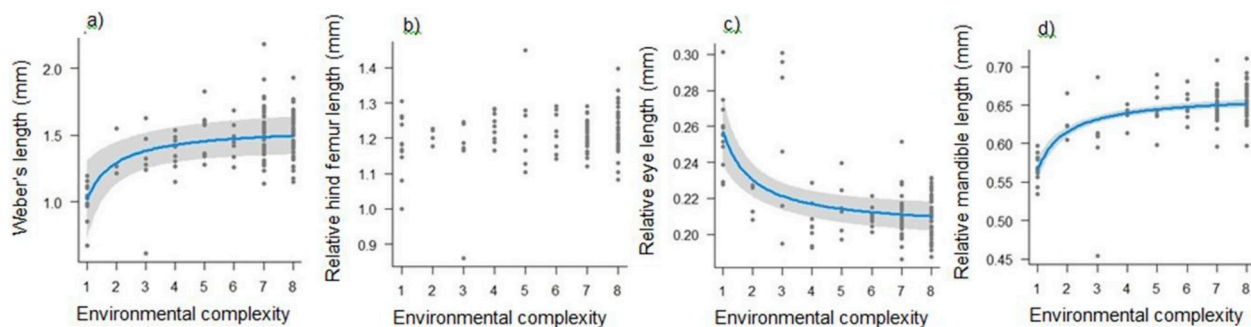


Fig. 3. Relationship between abundance-weighted trait average and environmental complexity across ant assemblages for different traits. Each point represents one ant assemblage sampled at a given plot. The blue line represents model predictions and the surrounding gray area represents the 95% confidence interval of the significant models. The figure of the hind femur length (b) was included for illustration although it has no relationship with the environmental complexity. (For interpretation of the references to color in this figure legend, the reader is referred to the Web version of this article.)

Table 3

The minimum, maximum and mean of the trait values at each level of environmental complexity are given in table.

Environmental complexity	Weber's length			Relative hind femur length			Relative eye length			Relative mandible length		
	Min	Max	Mean	Min	Max	Mean	Min	Max	Mean	Min	Max	Mean
1	0.803	1.322	1.137	1.008	1.311	1.187	0.224	0.289	0.250	0.543	0.597	0.570
2	1.464	1.775	1.563	1.178	1.228	1.208	0.219	0.237	0.229	0.604	0.655	0.622
3	0.903	1.912	1.532	0.862	1.247	1.147	0.206	0.311	0.267	0.453	0.682	0.593
4	1.354	1.746	1.582	1.159	1.284	1.228	0.204	0.238	0.214	0.606	0.651	0.634
5	1.313	1.800	1.534	1.064	1.410	1.189	0.196	0.237	0.215	0.594	0.690	0.643
6	1.318	1.741	1.501	1.104	1.253	1.179	0.199	0.219	0.208	0.620	0.675	0.647
7	1.159	2.281	1.581	1.081	1.291	1.203	0.192	0.259	0.211	0.587	0.703	0.645
8	0.949	1.920	1.292	1.087	1.425	1.238	0.181	0.240	0.207	0.597	0.707	0.649

relationship was detected between the relative hind femur length and the environmental gradient studied (Table 2). The minimum, maximum and average values of the observations at each level of environmental complexity for each trait are given in Table 3.

4. Discussion

Our results show that habitat complexity can promote significant changes in the functional composition of ground-dwelling ant assemblages. The functional approach focused on each morphological trait individually provides information about how ant foraging strategy responds to environmental variation. We found that ant species inhabiting less complex habitats were smaller than ant species inhabiting more complex habitats, and on average, relative hind femur length was invariant along the environmental gradient studied. We also found that with increasing environmental complexity the ants exhibited relatively smaller eyes and relatively larger mandibles.

Taken together, the results of body size and relative hind femur length are partially in line with the size-grain hypothesis. The size-grain hypothesis states that ants in less complex habitats would be larger assuming they have longer legs, allowing for more efficient movement through fissures and obstacles of a relatively “flat” habitat. On the other hand, the greater complexity generated by the debris and leaves stacked on the surface of the habitat would benefit smaller ants with relatively shorter legs because they can exploit resources more efficiently and shelter in these environments (Kaspari and Weiser, 1999; Farji-Brener et al., 2004; Gibb and Parr, 2013; Yates et al., 2014; Schofield et al., 2016). Three possible explanations could account for the observed pattern of body size with the environmental gradient.

The first explanation is related to thermoregulatory capacity of ants. Body size has an important role in heat conservation in both vertebrate (Amado et al., 2018) and invertebrate ectotherms (Pereboom and Biesmeijer, 2003; Chown and Gaston, 2010). Surface-to-volume ratio increases as body size decreases, so that insects with small body size tend to exchange heat more easily (Chapman, 1998). Thus, ground-dwelling ant assemblages in environments more exposed to sunlight, such as open vegetation, may harbour smaller species because they avoid overheating. In addition, the body size of ants is related to cuticle coloration (Bishop et al., 2016), which plays an important thermoregulatory role in insects (Clusella Trullas et al., 2007). Assemblages of ants composed of small-sized species have a lighter coloration (Bishop et al., 2016), which can also prevent overheating through low heat retention (Clusella Trullas et al., 2007). It is possible that thermoregulatory requirements are more important than the size-grain hypothesis explaining the size composition of ground-dwelling ant assemblages in hot environments. Another, alternative explanation would be associated with the availability of resources. The greater supply of food in more complex habitats may favor larger ants because they have larger food requirements, as in butterflies (Barlow, 1994) and other arthropods (Chown and Gaston, 2010). However, studies evaluating the role of body size in ants indicate that larger size may confer protection against desiccation in extremely hot habitats and greater resistance to

starvation in unproductive habitats (Cerdá and Retana, 2000; Kaspari and Vargo, 1995). Therefore, the explanations of thermoregulatory capacity and availability of resources have little support for ant responses to environmental complexity.

The third explanation is related to a possible bias in the measure used to estimate the environmental complexity gradient in our study. For instance, some plots in classified as less complex in our study have the soil covered by grasses (shrubby campinarana and open savanna), which may lead the ants to experience a relatively complex habitat and buffer the thermal variation. This could explain the small size of the ants' body in less complex environments, assuming that they can move better through the interstices of the grasses. However, the lack of relationship between hind femur length and environmental complexity contradicts this “grass hypothesis”. Further studies directly evaluating the effect of the substrate on a scale corresponding to the scale of ant's activity may clarify how these traits (body size and leg length) are related with the environmental gradient.

In line with our second and third predictions, ant species in dense forests had relatively smaller eyes and relatively large mandibles. Two hypotheses can explain these relationships: the light level-eye size hypothesis and the foraging specialization hypothesis (Schofield et al., 2016). In environments with high luminosity, ants can orient themselves and navigate better using vision and, conversely, in places with lower luminosity, the visual orientation becomes less relevant (Schofield et al., 2016). The absence of eyes or reduction in eye size is a common feature in Neotropical ant genera inhabiting the subsoil or at the interface between soil and litter such as *Acropyga*, *Prionopelta*, *Tranopelta*, *Nomamyrmex*, and others. Thus, sites with a large accumulation of litter may favor ant species inhabiting this microhabitat, and therefore, these sites harbor ants with smaller eyes, on average. In addition, many foraging species within the litter have predatory behavior (Weiser and Kaspari, 2006). If low light levels in the litter select ants with smaller eyes, then the predatory ants would have to use other attributes in searching for prey in these environmental conditions. The larger antennal scape found in ants inhabiting more complex environments (Yates et al., 2014) or mechanosensitive bristles related to prey detection (Brown and Wilson, 1959), are examples of how some features may compensate for the decrease in visual perception while foraging for prey in the litter layer.

The positive relationship between relative mandible length and environmental complexity is possibly associated with specialization in species foraging. Ants with longer mandibles can capture prey with variable shapes and sizes (Fowler et al., 1991), and this advantage can be increased in habitats with greater resource availability due to the greater possibility of finding prey of large sizes. Thus, investment in the construction of a large mandible allowing greater amplitude in prey size becomes an important factor for predatory ants in more productive environments (Schofield et al., 2016). In addition, analyzes of guild classification based on ant morphology have shown that generalist species, such as omnivorous or granivorous species, have relatively smaller mandibles (Silva and Brandão, 2010), and this strategy has been related to more simple and open habitats (Bishop et al., 2015;

Schofield et al., 2016). Taken as a whole, this indicates that although many ant species belonging to guilds such as fungus-growers or some invasive species occur in almost all habitats, some foraging behaviors may be associated with specific habitat types. We suggest that generalist species may represent the common strategy to survive in simple environmental conditions and that more complex habitats may support more specialized foraging due to the greater diversity and abundance of prey.

A pattern that was repeated in all traits investigated was that the change in trait composition along the environmental gradient occurred mainly between the less complex vegetation (Amazonian savannah) and other vegetations (white-sand forests and terra-firme forests). On average, assemblages had similar trait compositions between intermediate and more complex vegetation. Possibly the availability of niches created by the litter on the ground of these intermediate to more complex habitats favors species functionally equivalent to those living in the more complex habitats, making the asymptotic model a better descriptor of the observed trait relations.

5. Conclusions

In general, this work shows clear relationships between the functional composition (community-trait mean) of ground-dwelling ant assemblages and environmental complexity. We suggest that at the investigated spatial scale and extent, environmental complexity acts as a “filter” on certain morphological characteristics, so that species having certain morphologies - namely, larger bodies with relatively small eyes and large mandibles - tend to be favoured in complex

Appendix B. Supplementary data

Supplementary data related to this article can be found at <https://doi.org/10.1016/j.actao.2019.05.004>.

Appendix

Table A

Data used as criteria for ordering phytophysiognomy at levels of environmental complexity. Data on the characteristics of the vegetation were taken from the Brazilian Vegetation Classification System of the Brazilian Institute of Geography and Statistics (Instituto Brasileiro de Geografia e Estatística-IBGE, 2012).

Phytophysiognomies ordered at levels of complexity	Number of plots in each level of complexity	Criteria used for the classification of phytophysiognomies		
		Characteristics of vegetation and climate	Height of trees (m)	Percentage of plant cover (%)
1 - Open savana	12	Widely open area with some grass on the ground	0	0
2 - Seasonal shrubby campinarana	4	Purely herbaceous vegetation with waterlogging during the rainy season	~2	<5%
3 - Seasonal campinarana	6	More densely planted or wooded vegetation than Seasonal shrubby	~20	<50
4 - Campinarana forest	9	More densely planted or wooded vegetation than Seasonal campinarana	~20	<50
5 - Deciduous forest	7	Seasonal climate with the deciduity of much of the vegetation cover	between 20 and 40	~50
6 - Semideciduous forest	8	Seasonal climate with semi-deciduous vegetation cover	between 20 and 40	between 60 and 80%
7 - Open ombrophylous forest	36	It exhibits less dense vegetation, with around 60 dry days	~40	<80%
8 - Dense ombrophylous forest	45	Dense vegetation with well-distributed rainfall throughout the year	~40	~80%

Table B1

Sampling site and phytophysiognomy from which the individuals were taken to the measurements of each species.

Taxon	Site	Phytophysiognomy
<i>Acanthognathus ocellatus</i>	Viruá	Seasonal shrubby campinarana
<i>Acanthostichus bentoni</i>	Ducke	Dense ombrophylous forest
<i>Acanthostichus bentoni</i>	Ducke	Dense ombrophylous forest
<i>Acanthostichus bentoni</i>	Ducke	Dense ombrophylous forest

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habitats. Our work corroborates some hypotheses although others need further investigation, bearing in mind that other factors may be acting simultaneously on ant species assemblages.

Author's contribution

J.L.P.S., F.B.B., E.F. and D.R.G. conceived the idea; A.C. and D.R.G. performed the morphological measurements; P.A.C.L.P., F.B.B. and D.R.G. analyzed the data and D.R.G. wrote the article with substantial collaboration from all authors.

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Table B1 (continued)

Taxon	Site	Phytophysiology
<i>Acanthostichus bentoni</i>	Ducke	Dense ombrophylous forest
<i>Acanthostichus bentoni</i>	Ducke	Dense ombrophylous forest
<i>Acanthostichus bentoni</i>	Ducke	Dense ombrophylous forest
<i>Acromyrmex</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Acromyrmex</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Acromyrmex subterraneus</i>	Maracá	Semideciduous forest
<i>Acropyga</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Acropyga</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Acropyga</i> sp. 01	Manaquiri	Open ombrophylous forest
<i>Acropyga</i> sp. 01	Manaquiri	Open ombrophylous forest
<i>Acropyga</i> sp. 01	Maracá	Semideciduous forest
<i>Acropyga</i> sp. 01	Maracá	Semideciduous forest
<i>Acropyga</i> sp. 02	Ducke	Dense ombrophylous forest
<i>Anochetus diegensis</i>	Ducke	Dense ombrophylous forest
<i>Anochetus diegensis</i>	Ducke	Dense ombrophylous forest
<i>Anochetus diegensis</i>	Ducke	Dense ombrophylous forest
<i>Anochetus diegensis</i>	Manaquiri	Open ombrophylous forest
<i>Anochetus diegensis</i>	Manaquiri	Open ombrophylous forest
<i>Anochetus diegensis</i>	Manaquiri	Open ombrophylous forest
<i>Anochetus emarginatus</i>	Ducke	Dense ombrophylous forest
<i>Anochetus horridus</i>	Ducke	Dense ombrophylous forest
<i>Anochetus horridus</i>	Ducke	Dense ombrophylous forest
<i>Anochetus horridus</i>	Ducke	Dense ombrophylous forest
<i>Anochetus horridus</i>	Ducke	Dense ombrophylous forest
<i>Anochetus horridus</i>	Manaquiri	Open ombrophylous forest
<i>Anochetus horridus</i>	Manaquiri	Open ombrophylous forest
<i>Apterostigma pilosum</i>	Ducke	Dense ombrophylous forest
<i>Apterostigma pilosum</i>	Maracá	Open ombrophylous forest
<i>Apterostigma pilosum</i>	Viruá	Campinarana forest
<i>Apterostigma</i> sp. 03	Ducke	Dense ombrophylous forest
<i>Apterostigma</i> sp. 04	Ducke	Dense ombrophylous forest
<i>Apterostigma urichii</i>	Maracá	Semideciduous forest
<i>Apterostigma urichii</i>	Ducke	Dense ombrophylous forest
<i>Apterostigma urichii</i>	Ducke	Dense ombrophylous forest
<i>Apterostigma urichii</i>	Ducke	Dense ombrophylous forest
<i>Atta cephalotes</i>	Maracá	Deciduous forest
<i>Atta cephalotes</i>	Viruá	Seasonal campinarana
<i>Atta cephalotes</i>	Ducke	Dense ombrophylous forest
<i>Atta sexdens</i>	Maracá	Deciduous forest
<i>Atta sexdens</i>	Viruá	Open ombrophylous forest
<i>Atta sexdens</i>	Viruá	Open ombrophylous forest
<i>Atta sexdens</i>	Ducke	Dense ombrophylous forest
<i>Atta sexdens</i>	Ducke	Dense ombrophylous forest
<i>Atta sexdens</i>	Ducke	Dense ombrophylous forest
<i>Azteca</i> sp. 01	Viruá	Campinarana forest
<i>Azteca</i> sp. 01	Viruá	Seasonal campinarana
<i>Azteca</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Azteca</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Blepharidatta brasiliensis</i>	Jari	Dense ombrophylous forest
<i>Blepharidatta brasiliensis</i>	Jari	Dense ombrophylous forest
<i>Blepharidatta brasiliensis</i>	Ducke	Dense ombrophylous forest
<i>Blepharidatta brasiliensis</i>	Ducke	Dense ombrophylous forest
<i>Blepharidatta brasiliensis</i>	Ducke	Dense ombrophylous forest
<i>Blepharidatta brasiliensis</i>	Ducke	Dense ombrophylous forest
<i>Brachymyrmex heeri</i>	Viruá	Seasonal shrubby campinarana
<i>Brachymyrmex heeri</i>	Viruá	Seasonal shrubby campinarana
<i>Brachymyrmex heeri</i>	Viruá	Seasonal shrubby campinarana
<i>Brachymyrmex heeri</i>	Maracá	Semideciduous forest
<i>Brachymyrmex heeri</i>	Manaquiri	Open ombrophylous forest
<i>Brachymyrmex</i> sp. 01	Manaquiri	Open ombrophylous forest
<i>Brachymyrmex</i> sp. 01	Manaquiri	Open ombrophylous forest
<i>Brachymyrmex</i> sp. 01	Cauamé	Open savanna
<i>Brachymyrmex</i> sp. 01	Cauamé	Open savanna
<i>Brachymyrmex</i> sp. 01	Cauamé	Open savanna
<i>Brachymyrmex</i> sp. 01	Cauamé	Open savanna
<i>Brachymyrmex</i> sp. 02	Capana	Dense ombrophylous forest
<i>Brachymyrmex</i> sp. 02	Capana	Dense ombrophylous forest
<i>Brachymyrmex</i> sp. 02	Cauamé	Open savanna
<i>Brachymyrmex</i> sp. 03	Manaquiri	Open ombrophylous forest
<i>Camponotus ager</i>	Cauamé	Open savanna
<i>Camponotus ager</i>	Cauamé	Open savanna
<i>Camponotus ager</i>	Viruá	Seasonal shrubby campinarana
<i>Camponotus ager</i>	Viruá	Open ombrophylous forest
<i>Camponotus ager</i>	Ducke	Dense ombrophylous forest
<i>Camponotus atriceps</i>	Ducke	Dense ombrophylous forest

(continued on next page)

Table B1 (continued)

Taxon	Site	Phytophysognomy
<i>Camponotus atriceps</i>	Ducke	Dense ombrophylous forest
<i>Camponotus atriceps</i>	Capana	Dense ombrophylous forest
<i>Camponotus atriceps</i>	Viruá	Seasonal shrubby campinarana
<i>Camponotus atriceps</i>	Viruá	Seasonal campinarana
<i>Camponotus atriceps</i>	Viruá	Open ombrophylous forest
<i>Camponotus balzani</i>	Orquestra	Dense ombrophylous forest
<i>Camponotus balzani</i>	Orquestra	Dense ombrophylous forest
<i>Camponotus balzani</i>	Orquestra	Dense ombrophylous forest
<i>Camponotus crassus</i>	Cauamé	Open savanna
<i>Camponotus crassus</i>	Cauamé	Open savanna
<i>Camponotus crassus</i>	Ducke	Dense ombrophylous forest
<i>Camponotus crassus</i>	Viruá	Seasonal campinarana
<i>Camponotus crassus</i>	Viruá	Seasonal campinarana
<i>Camponotus fastigatus</i>	Cauamé	Open savanna
<i>Camponotus fastigatus</i>	Ducke	Dense ombrophylous forest
<i>Camponotus fastigatus</i>	Ducke	Dense ombrophylous forest
<i>Camponotus femoratus</i>	Manaquiri	Open ombrophylous forest
<i>Camponotus femoratus</i>	Manaquiri	Open ombrophylous forest
<i>Camponotus femoratus</i>	Viruá	Campinarana forest
<i>Camponotus femoratus</i>	Ducke	Dense ombrophylous forest
<i>Camponotus femoratus</i>	Ducke	Dense ombrophylous forest
<i>Camponotus femoratus</i>	Ducke	Dense ombrophylous forest
<i>Camponotus leydigi</i>	Cauamé	Open savanna
<i>Camponotus leydigi</i>	Cauamé	Open savanna
<i>Camponotus leydigi</i>	Viruá	Seasonal shrubby campinarana
<i>Camponotus novogranadensis</i>	Cauamé	Open savanna
<i>Camponotus novogranadensis</i>	Cauamé	Open savanna
<i>Camponotus novogranadensis</i>	Jari	Dense ombrophylous forest
<i>Camponotus novogranadensis</i>	Ducke	Dense ombrophylous forest
<i>Camponotus novogranadensis</i>	Viruá	Seasonal shrubby campinarana
<i>Camponotus rapax</i>	Ducke	Dense ombrophylous forest
<i>Camponotus rapax</i>	Manaquiri	Open ombrophylous forest
<i>Camponotus rapax</i>	Viruá	Open ombrophylous forest
<i>Camponotus rapax</i>	Viruá	Seasonal shrubby campinarana
<i>Camponotus rapax</i>	Viruá	Seasonal campinarana
<i>Camponotus rapax</i>	Viruá	Seasonal shrubby campinarana
<i>Camponotus sericeiventris</i>	Viruá	Campinarana forest
<i>Camponotus sericeiventris</i>	Viruá	Campinarana forest
<i>Camponotus</i> sp. 14	Maracá	Open ombrophylous forest
<i>Carebara escherichi</i>	Ducke	Dense ombrophylous forest
<i>Carebara escherichi</i>	Ducke	Dense ombrophylous forest
<i>Carebara escherichi</i>	Ducke	Dense ombrophylous forest
<i>Carebara escherichi</i>	Ducke	Dense ombrophylous forest
<i>Carebara lignata</i>	Maracá	Open ombrophylous forest
<i>Carebara urichi</i>	Ducke	Dense ombrophylous forest
<i>Carebara urichi</i>	Ducke	Dense ombrophylous forest
<i>Carebara urichi</i>	Ducke	Dense ombrophylous forest
<i>Carebara urichi</i>	Ducke	Dense ombrophylous forest
<i>Carebara urichi</i>	Manaquiri	Open ombrophylous forest
<i>Centromyrmex gigas</i>	Maracá	Open ombrophylous forest
<i>Centromyrmex</i> sp. 01	Jari	Dense ombrophylous forest
<i>Cephalotes pusillus</i>	Maracá	Semideciduous forest
<i>Cephalotes pusillus</i>	Maracá	Semideciduous forest
<i>Cephalotes pusillus</i>	Viruá	Seasonal campinarana
<i>Cephalotes atratus</i>	Capana	Dense ombrophylous forest
<i>Cephalotes atratus</i>	Capana	Dense ombrophylous forest
<i>Cephalotes maculatus</i>	Maracá	Semideciduous forest
<i>Cephalotes marginatus</i>	Jari	Dense ombrophylous forest
<i>Crematogaster brasiliensis</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster brasiliensis</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster brasiliensis</i>	Maracá	Semideciduous forest
<i>Crematogaster brasiliensis</i>	Viruá	Open ombrophylous forest
<i>Crematogaster brasiliensis</i>	Viruá	Campinarana forest
<i>Crematogaster brasiliensis</i>	Viruá	Seasonal shrubby campinarana
<i>Crematogaster tenuicula</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster tenuicula</i>	Maracá	Open ombrophylous forest
<i>Crematogaster tenuicula</i>	Maracá	Seasonal campinarana
<i>Crematogaster tenuicula</i>	Viruá	Seasonal shrubby campinarana
<i>Crematogaster tenuicula</i>	Viruá	Campinarana forest
<i>Crematogaster tenuicula</i>	Viruá	Campinarana forest
<i>Crematogaster limata</i>	Manaquiri	Open ombrophylous forest
<i>Crematogaster limata</i>	Manaquiri	Open ombrophylous forest
<i>Crematogaster limata</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster limata</i>	Viruá	Campinarana forest

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Table B1 (continued)

Taxon	Site	Phytophysognomy
<i>Crematogaster limata</i>	Viruá	Campinarana forest
<i>Crematogaster sotobosque</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster sotobosque</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster sotobosque</i>	Orquestra	Dense ombrophylous forest
<i>Crematogaster sotobosque</i>	Manaquiri	Open ombrophylous forest
<i>Crematogaster sotobosque</i>	Manaquiri	Open ombrophylous forest
<i>Crematogaster flavosensitiva</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster flavosensitiva</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster flavosensitiva</i>	Manaquiri	Open ombrophylous forest
<i>Crematogaster flavosensitiva</i>	Viruá	Open ombrophylous forest
<i>Crematogaster flavosensitiva</i>	Viruá	Seasonal shrubby campinarana
<i>Crematogaster curvispinosa</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster stollii</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster stollii</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster erecta</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster erecta</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster nigropilosa</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster nigropilosa</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster evallans</i>	Viruá	Seasonal campinarana
<i>Crematogaster evallans</i>	Viruá	Seasonal campinarana
<i>Crematogaster evallans</i>	Viruá	Seasonal campinarana
<i>Crematogaster longispina</i>	Ducke	Dense ombrophylous forest
<i>Crematogaster longispina</i>	Maracá	Deciduous forest
<i>Crematogaster rochai</i>	Maracá	Deciduous forest
<i>Crematogaster jardínero</i>	Cauamé	Open savanna
<i>Crematogaster jardínero</i>	Cauamé	Open savanna
<i>Crematogaster jardínero</i>	Cauamé	Open savanna
<i>Crematogaster jardínero</i>	Cauamé	Open savanna
<i>Cyphomyrmex laevigatus</i>	Ducke	Dense ombrophylous forest
<i>Cyphomyrmex laevigatus</i>	Ducke	Dense ombrophylous forest
<i>Cyphomyrmex laevigatus</i>	Ducke	Dense ombrophylous forest
<i>Cyphomyrmex laevigatus</i>	Ducke	Dense ombrophylous forest
<i>Cyphomyrmex laevigatus</i>	Ducke	Dense ombrophylous forest
<i>Cyphomyrmex laevigatus</i>	Maracá	Open ombrophylous forest
<i>Cyphomyrmex peltatus</i>	Orquestra	Dense ombrophylous forest
<i>Cyphomyrmex peltatus</i>	Orquestra	Dense ombrophylous forest
<i>Cyphomyrmex peltatus</i>	Orquestra	Dense ombrophylous forest
<i>Cyphomyrmex peltatus</i>	Viruá	Campinarana forest
<i>Cyphomyrmex peltatus</i>	Viruá	Campinarana forest
<i>Cyphomyrmex peltatus</i>	Ducke	Dense ombrophylous forest
<i>Cyphomyrmex rimosus</i>	Manaquiri	Open ombrophylous forest
<i>Cyphomyrmex rimosus</i>	Manaquiri	Open ombrophylous forest
<i>Cyphomyrmex rimosus</i>	Viruá	Campinarana forest
<i>Cyphomyrmex rimosus</i>	Viruá	Campinarana forest
<i>Cyphomyrmex rimosus</i>	Viruá	Campinarana forest
<i>Cyphomyrmex rimosus</i>	Viruá	Campinarana forest
<i>Discothyrea denticulata</i>	Maracá	Seasonal shrubby campinarana
<i>Dolichoderus bispinosus</i>	Maracá	Semideciduous forest
<i>Dolichoderus bispinosus</i>	Maracá	Semideciduous forest
<i>Dolichoderus bispinosus</i>	Jari	Dense ombrophylous forest
<i>Dolichoderus bispinosus</i>	Orquestra	Dense ombrophylous forest
<i>Dolichoderus bispinosus</i>	Capana	Dense ombrophylous forest
<i>Dolichoderus bispinosus</i>	Ducke	Dense ombrophylous forest
<i>Dolichoderus imitator</i>	Ducke	Dense ombrophylous forest
<i>Dolichoderus imitator</i>	Orquestra	Dense ombrophylous forest
<i>Dolichoderus imitator</i>	Jari	Dense ombrophylous forest
<i>Dolichoderus imitator</i>	Maracá	Open ombrophylous forest
<i>Dolichoderus decollatus</i>	Maracá	Deciduous forest
<i>Dolichoderus decollatus</i>	Maracá	Open ombrophylous forest
<i>Dorymyrmex bicolor</i>	Cauamé	Open savanna
<i>Dorymyrmex bicolor</i>	Cauamé	Open savanna
<i>Dorymyrmex bicolor</i>	Cauamé	Open savanna
<i>Dorymyrmex goeldii</i>	Cauamé	Open savanna
<i>Dorymyrmex goeldii</i>	Viruá	Open ombrophylous forest
<i>Dorymyrmex goeldii</i>	Viruá	Seasonal shrubby campinarana
<i>Dorymyrmex goeldii</i>	Viruá	Seasonal shrubby campinarana
<i>Dorymyrmex richteri</i>	Cauamé	Open savanna
<i>Dorymyrmex richteri</i>	Cauamé	Open savanna
<i>Dorymyrmex richteri</i>	Cauamé	Open savanna
<i>Dorymyrmex richteri</i>	Cauamé	Open savanna
<i>Eciton burchellii</i>	Maracá	Open ombrophylous forest
<i>Eciton burchellii</i>	Maracá	Open ombrophylous forest
<i>Eciton burchellii</i>	Maracá	Open ombrophylous forest
<i>Eciton burchellii</i>	Maracá	Open ombrophylous forest

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Table B1 (continued)

Taxon	Site	Phytophysognomy
<i>Eciton dulcius</i>	Ducke	Dense ombrophylous forest
<i>Eciton dulcius</i>	Ducke	Dense ombrophylous forest
<i>Eciton rapax</i>	Ducke	Dense ombrophylous forest
<i>Eciton rapax</i>	Ducke	Dense ombrophylous forest
<i>Eciton rapax</i>	Manaquiri	Open ombrophylous forest
<i>Eciton rapax</i>	Manaquiri	Open ombrophylous forest
<i>Ectatomma brunneum</i>	Viruá	Open ombrophylous forest
<i>Ectatomma brunneum</i>	Viruá	Open ombrophylous forest
<i>Ectatomma brunneum</i>	Viruá	Seasonal campinarana
<i>Ectatomma brunneum</i>	Viruá	Campinarana forest
<i>Ectatomma edentatum</i>	Ducke	Dense ombrophylous forest
<i>Ectatomma edentatum</i>	Ducke	Dense ombrophylous forest
<i>Ectatomma edentatum</i>	Ducke	Dense ombrophylous forest
<i>Ectatomma edentatum</i>	Maracá	Open ombrophylous forest
<i>Ectatomma edentatum</i>	Viruá	Campinarana forest
<i>Ectatomma edentatum</i>	Viruá	Campinarana forest
<i>Ectatomma edentatum</i>	Viruá	Open ombrophylous forest
<i>Ectatomma lugens</i>	Ducke	Dense ombrophylous forest
<i>Ectatomma lugens</i>	Ducke	Dense ombrophylous forest
<i>Ectatomma lugens</i>	Viruá	Seasonal campinarana
<i>Ectatomma lugens</i>	Viruá	Seasonal campinarana
<i>Ectatomma lugens</i>	Ducke	Dense ombrophylous forest
<i>Ectatomma lugens</i>	Ducke	Dense ombrophylous forest
<i>Ectatomma ruidum</i>	Cauamé	Open savanna
<i>Ectatomma ruidum</i>	Cauamé	Open savanna
<i>Ectatomma ruidum</i>	Cauamé	Open savanna
<i>Gigantiops destructor</i>	Ducke	Dense ombrophylous forest
<i>Gigantiops destructor</i>	Ducke	Dense ombrophylous forest
<i>Gigantiops destructor</i>	Ducke	Dense ombrophylous forest
<i>Gigantiops destructor</i>	Viruá	Campinarana forest
<i>Gigantiops destructor</i>	Viruá	Open ombrophylous forest
<i>Gnamptogenys acuminata</i>	Manaquiri	Open ombrophylous forest
<i>Gnamptogenys acuminata</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys acuminata</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys acuminata</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys curvoclypeata</i>	Maracá	Seasonal campinarana
<i>Gnamptogenys curvoclypeata</i>	Maracá	Seasonal campinarana
<i>Gnamptogenys curvoclypeata</i>	Maracá	Campinarana forest
<i>Gnamptogenys strigata</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys strigata</i>	Maracá	Open ombrophylous forest
<i>Gnamptogenys haenschi</i>	Manaquiri	Open ombrophylous forest
<i>Gnamptogenys haenschi</i>	Manaquiri	Open ombrophylous forest
<i>Gnamptogenys haenschi</i>	Manaquiri	Open ombrophylous forest
<i>Gnamptogenys horni</i>	Jari	Dense ombrophylous forest
<i>Gnamptogenys horni</i>	Orquestra	Dense ombrophylous forest
<i>Gnamptogenys horni</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys horni</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys horni</i>	Manaquiri	Open ombrophylous forest
<i>Gnamptogenys horni</i>	Manaquiri	Open ombrophylous forest
<i>Gnamptogenys moelleri</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys moelleri</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys moelleri</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys moelleri</i>	Capana	Dense ombrophylous forest
<i>Gnamptogenys regularis</i>	Maracá	Open ombrophylous forest
<i>Gnamptogenys regularis</i>	Maracá	Open ombrophylous forest
<i>Gnamptogenys regularis</i>	Maracá	Open ombrophylous forest
<i>Gnamptogenys regularis</i>	Maracá	Open ombrophylous forest
<i>Gnamptogenys sulcata</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys sulcata</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys sulcata</i>	Manaquiri	Open ombrophylous forest
<i>Gnamptogenys sulcata</i>	Maracá	Open ombrophylous forest
<i>Gnamptogenys tortuolosa</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys tortuolosa</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys tortuolosa</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys tortuolosa</i>	Ducke	Dense ombrophylous forest
<i>Gnamptogenys tortuolosa</i>	Maracá	Semideciduous forest
<i>Gnamptogenys tortuolosa</i>	Maracá	Open ombrophylous forest
<i>Hylomyrma immanis</i>	Capana	Dense ombrophylous forest
<i>Hylomyrma immanis</i>	Capana	Dense ombrophylous forest
<i>Hylomyrma immanis</i>	Manaquiri	Open ombrophylous forest
<i>Hylomyrma immanis</i>	Manaquiri	Open ombrophylous forest
<i>Hypoconera</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 01	Ducke	Dense ombrophylous forest

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Table B1 (continued)

Taxon	Site	Phytophysognomy
<i>Hypoconera</i> sp. 01	Manaquiri	Open ombrophylous forest
<i>Hypoconera</i> sp. 01	Maracá	Deciduous forest
<i>Hypoconera</i> sp. 02	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 02	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 03	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 03	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 03	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 04	Maracá	Open ombrophylous forest
<i>Hypoconera</i> sp. 04	Maracá	Deciduous forest
<i>Hypoconera</i> sp. 04	Maracá	Open ombrophylous forest
<i>Hypoconera</i> sp. 04	Maracá	Deciduous forest
<i>Hypoconera</i> sp. 04	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 04	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 05	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 05	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 05	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 05	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 05	Maracá	Semideciduous forest
<i>Hypoconera</i> sp. 06	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 06	Maracá	Open ombrophylous forest
<i>Hypoconera</i> sp. 06	Maracá	Semideciduous forest
<i>Hypoconera</i> sp. 06	Maracá	Semideciduous forest
<i>Hypoconera</i> sp. 07	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 07	Ducke	Dense ombrophylous forest
<i>Hypoconera</i> sp. 07	Maracá	Open ombrophylous forest
<i>Hypoconera</i> sp. 07	Maracá	Semideciduous forest
<i>Hypoconera</i> sp. 07	Maracá	Semideciduous forest
<i>Hypoconera</i> sp. 08	Capana	Dense ombrophylous forest
<i>Hypoconera</i> sp. 08	Capana	Dense ombrophylous forest
<i>Hypoconera</i> sp. 08	Capana	Dense ombrophylous forest
<i>Hypoconera</i> sp. 08	Capana	Dense ombrophylous forest
<i>Hypoconera</i> sp. 09	Jari	Dense ombrophylous forest
<i>Hypoconera</i> sp. 09	Jari	Dense ombrophylous forest
<i>Hypoconera</i> sp. 09	Orquestra	Dense ombrophylous forest
<i>Hypoconera</i> sp. 09	Maracá	Open ombrophylous forest
<i>Hypoconera</i> sp. 11	Jari	Dense ombrophylous forest
<i>Hypoconera</i> sp. 11	Jari	Dense ombrophylous forest
<i>Hypoconera</i> sp. 11	Jari	Dense ombrophylous forest
<i>Hypoconera</i> sp. 14	Manaquiri	Open ombrophylous forest
<i>Hypoconera</i> sp. 14	Manaquiri	Open ombrophylous forest
<i>Hypoconera</i> sp. 14	Manaquiri	Open ombrophylous forest
<i>Hypoconera</i> sp. 14	Manaquiri	Open ombrophylous forest
<i>Kalathomyrmex emryi</i>	Cauamé	Open savanna
<i>Labidus coecus</i>	Ducke	Dense ombrophylous forest
<i>Labidus coecus</i>	Ducke	Dense ombrophylous forest
<i>Labidus coecus</i>	Viruá	Campinarana forest
<i>Labidus coecus</i>	Viruá	Campinarana forest
<i>Labidus coecus</i>	Maracá	Seasonal campinarana
<i>Labidus coecus</i>	Manaquiri	Open ombrophylous forest
<i>Labidus mars</i>	Ducke	Dense ombrophylous forest
<i>Labidus praedator</i>	Jari	Dense ombrophylous forest
<i>Labidus praedator</i>	Ducke	Dense ombrophylous forest
<i>Labidus praedator</i>	Manaquiri	Open ombrophylous forest
<i>Labidus praedator</i>	Manaquiri	Open ombrophylous forest
<i>Labidus praedator</i>	Ducke	Dense ombrophylous forest
<i>Labidus praedator</i>	Viruá	Campinarana forest
<i>Labidus spininodis</i>	Ducke	Dense ombrophylous forest
<i>Labidus spininodis</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys gaigei</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys gaigei</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys gaigei</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys gaigei</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys gaigei</i>	Viruá	Open ombrophylous forest
<i>Leptogenys gaigei</i>	Viruá	Open ombrophylous forest
<i>Leptogenys pusilla</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys pusilla</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys wheeleri</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys wheeleri</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys wheeleri</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys wheeleri</i>	Ducke	Dense ombrophylous forest
<i>Leptogenys wheeleri</i>	Ducke	Dense ombrophylous forest
<i>Mayaponera constricta</i>	Viruá	Seasonal campinarana
<i>Mayaponera constricta</i>	Viruá	Open ombrophylous forest
<i>Mayaponera constricta</i>	Ducke	Dense ombrophylous forest

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Table B1 (continued)

Taxon	Site	Phytophysiology
<i>Mayaponera constricta</i>	Ducke	Dense ombrophylous forest
<i>Mayaponera constricta</i>	Orquestra	Dense ombrophylous forest
<i>Mayaponera constricta</i>	Orquestra	Dense ombrophylous forest
<i>Megalomyrmex balzani</i>	Ducke	Dense ombrophylous forest
<i>Megalomyrmex balzani</i>	Manaquiri	Open ombrophylous forest
<i>Megalomyrmex balzani</i>	Jari	Dense ombrophylous forest
<i>Megalomyrmex balzani</i>	Jari	Dense ombrophylous forest
<i>Megalomyrmex balzani</i>	Jari	Dense ombrophylous forest
<i>Megalomyrmex driftii</i>	Jari	Dense ombrophylous forest
<i>Megalomyrmex driftii</i>	Jari	Dense ombrophylous forest
<i>Megalomyrmex driftii</i>	Jari	Dense ombrophylous forest
<i>Megalomyrmex driftii</i>	Jari	Dense ombrophylous forest
<i>Megalomyrmex incisus</i>	Maracá	Open ombrophylous forest
<i>Megalomyrmex incisus</i>	Maracá	Open ombrophylous forest
<i>Megalomyrmex leoninus</i>	Maracá	Deciduous forest
<i>Megalomyrmex leoninus</i>	Maracá	Deciduous forest
<i>Megalomyrmex leoninus</i>	Viruá	Semideciduous forest
<i>Megalomyrmex leoninus</i>	Viruá	Open ombrophylous forest
<i>Megalomyrmex silvestrii</i>	Jari	Dense ombrophylous forest
<i>Megalomyrmex silvestrii</i>	Jari	Dense ombrophylous forest
<i>Megalomyrmex silvestrii</i>	Jari	Dense ombrophylous forest
<i>Megalomyrmex</i> sp. 04	Ducke	Dense ombrophylous forest
<i>Monomorium pharaonis</i>	Ducke	Dense ombrophylous forest
<i>Monomorium pharaonis</i>	Ducke	Dense ombrophylous forest
<i>Monomorium pharaonis</i>	Ducke	Dense ombrophylous forest
<i>Monomorium pharaonis</i>	Ducke	Dense ombrophylous forest
<i>Monomorium pharaonis</i>	Ducke	Dense ombrophylous forest
<i>Monomorium pharaonis</i>	Ducke	Dense ombrophylous forest
<i>Mycocepurus smithii</i>	Maracá	Open ombrophylous forest
<i>Mycocepurus smithii</i>	Maracá	Open ombrophylous forest
<i>Mycocepurus smithii</i>	Maracá	Semideciduous forest
<i>Mycocepurus smithii</i>	Maracá	Semideciduous forest
<i>Mycocepurus</i> sp. 01	Maracá	Semideciduous forest
<i>Mycocepurus</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Mycocepurus</i> sp. 01	Maracá	Semideciduous forest
<i>Myrmicocrypta</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Myrmicocrypta</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Myrmicocrypta</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Myrmicocrypta</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Myrmicocrypta</i> sp. 02	Viruá	Open ombrophylous forest
<i>Myrmicocrypta</i> sp. 02	Viruá	Campinarana forest
<i>Myrmicocrypta</i> sp. 02	Maracá	Open ombrophylous forest
<i>Neivamyrmex adnepos</i>	Jari	Dense ombrophylous forest
<i>Neivamyrmex angustinodis</i>	Ducke	Dense ombrophylous forest
<i>Neivamyrmex angustinodis</i>	Ducke	Dense ombrophylous forest
<i>Neivamyrmex angustinodis</i>	Ducke	Dense ombrophylous forest
<i>Neivamyrmex angustinodis</i>	Ducke	Dense ombrophylous forest
<i>Neivamyrmex gibbatus</i>	Ducke	Dense ombrophylous forest
<i>Neivamyrmex gibbatus</i>	Ducke	Dense ombrophylous forest
<i>Neivamyrmex gibbatus</i>	Viruá	Open ombrophylous forest
<i>Neivamyrmex gibbatus</i>	Viruá	Open ombrophylous forest
<i>Neivamyrmex iridescens</i>	Ducke	Dense ombrophylous forest
<i>Neivamyrmex orthonotus</i>	Ducke	Dense ombrophylous forest
<i>Neivamyrmex orthonotus</i>	Ducke	Dense ombrophylous forest
<i>Neivamyrmex pilosus</i>	Maracá	Semideciduous forest
<i>Neivamyrmex pilosus</i>	Maracá	Semideciduous forest
<i>Neivamyrmex swainsonii</i>	Maracá	Deciduous forest
<i>Neivamyrmex swainsonii</i>	Maracá	Deciduous forest
<i>Neivamyrmex swainsonii</i>	Maracá	Deciduous forest
<i>Neoponera apicalis</i>	Ducke	Dense ombrophylous forest
<i>Neoponera apicalis</i>	Ducke	Dense ombrophylous forest
<i>Neoponera apicalis</i>	Ducke	Dense ombrophylous forest
<i>Neoponera apicalis</i>	Viruá	Seasonal campinarana
<i>Neoponera apicalis</i>	Maracá	Semideciduous forest
<i>Neoponera apicalis</i>	Manaquiri	Open ombrophylous forest
<i>Neoponera commutata</i>	Ducke	Dense ombrophylous forest
<i>Neoponera commutata</i>	Ducke	Dense ombrophylous forest
<i>Neoponera commutata</i>	Manaquiri	Open ombrophylous forest
<i>Neoponera commutata</i>	Manaquiri	Open ombrophylous forest
<i>Neoponera commutata</i>	Viruá	Open ombrophylous forest
<i>Neoponera commutata</i>	Viruá	Open ombrophylous forest

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Table B1 (continued)

Taxon	Site	Phytophysognomy
<i>Neoponera laevigata</i>	Virúá	Open ombrophylous forest
<i>Neoponera verenae</i>	Orquestra	Dense ombrophylous forest
<i>Neoponera verenae</i>	Orquestra	Dense ombrophylous forest
<i>Neoponera verenae</i>	Orquestra	Dense ombrophylous forest
<i>Neoponera verenae</i>	Orquestra	Dense ombrophylous forest
<i>Nomamyrmex esenbeckii</i>	Ducke	Dense ombrophylous forest
<i>Nomamyrmex esenbeckii</i>	Ducke	Dense ombrophylous forest
<i>Nomamyrmex esenbeckii</i>	Ducke	Dense ombrophylous forest
<i>Nomamyrmex hartigi</i>	Ducke	Dense ombrophylous forest
<i>Nomamyrmex hartigi</i>	Ducke	Dense ombrophylous forest
<i>Nylanderia caeciliae</i>	Ducke	Dense ombrophylous forest
<i>Nylanderia caeciliae</i>	Ducke	Dense ombrophylous forest
<i>Nylanderia caeciliae</i>	Capana	Dense ombrophylous forest
<i>Nylanderia caeciliae</i>	Jari	Dense ombrophylous forest
<i>Nylanderia caeciliae</i>	Jari	Dense ombrophylous forest
<i>Nylanderia caeciliae</i>	Jari	Dense ombrophylous forest
<i>Nylanderia guatemalensis</i>	Cauamé	Open savanna
<i>Nylanderia guatemalensis</i>	Maracá	Open ombrophylous forest
<i>Nylanderia guatemalensis</i>	Ducke	Dense ombrophylous forest
<i>Nylanderia guatemalensis</i>	Ducke	Dense ombrophylous forest
<i>Nylanderia guatemalensis</i>	Jari	Dense ombrophylous forest
<i>Nylanderia guatemalensis</i>	Jari	Dense ombrophylous forest
<i>Nylanderia sp. 01</i>	Ducke	Dense ombrophylous forest
<i>Nylanderia sp. 01</i>	Ducke	Dense ombrophylous forest
<i>Nylanderia sp. 01</i>	Virúá	Campinarana forest
<i>Nylanderia sp. 01</i>	Virúá	Campinarana forest
<i>Nylanderia sp. 01</i>	Virúá	Seasonal campinarana
<i>Nylanderia sp. 04</i>	Manaquiri	Open ombrophylous forest
<i>Nylanderia sp. 04</i>	Manaquiri	Open ombrophylous forest
<i>Nylanderia sp. 04</i>	Maracá	Campinarana forest
<i>Nylanderia sp. 04</i>	Maracá	Campinarana forest
<i>Ochetomyrmex semipolitus</i>	Orquestra	Dense ombrophylous forest
<i>Ochetomyrmex semipolitus</i>	Orquestra	Dense ombrophylous forest
<i>Ochetomyrmex semipolitus</i>	Manaquiri	Open ombrophylous forest
<i>Ochetomyrmex semipolitus</i>	Manaquiri	Open ombrophylous forest
<i>Ochetomyrmex semipolitus</i>	Virúá	Campinarana forest
<i>Ochetomyrmex semipolitus</i>	Virúá	Open ombrophylous forest
<i>Octostruma balzani</i>	Ducke	Dense ombrophylous forest
<i>Octostruma balzani</i>	Ducke	Dense ombrophylous forest
<i>Octostruma balzani</i>	Capana	Dense ombrophylous forest
<i>Octostruma balzani</i>	Orquestra	Dense ombrophylous forest
<i>Octostruma balzani</i>	Jari	Dense ombrophylous forest
<i>Odontomachus bauri</i>	Capana	Dense ombrophylous forest
<i>Odontomachus bauri</i>	Virúá	Open ombrophylous forest
<i>Odontomachus bauri</i>	Virúá	Campinarana forest
<i>Odontomachus bauri</i>	Virúá	Campinarana forest
<i>Odontomachus brunneus</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus caelatus</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus caelatus</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus caelatus</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus caelatus</i>	Virúá	Open ombrophylous forest
<i>Odontomachus caelatus</i>	Virúá	Open ombrophylous forest
<i>Odontomachus caelatus</i>	Jari	Dense ombrophylous forest
<i>Odontomachus haematodus</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus haematodus</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus haematodus</i>	Virúá	Campinarana forest
<i>Odontomachus haematodus</i>	Virúá	Campinarana forest
<i>Odontomachus haematodus</i>	Maracá	Deciduous forest
<i>Odontomachus haematodus</i>	Manaquiri	Open ombrophylous forest
<i>Odontomachus laticeps</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus laticeps</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus laticeps</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus laticeps</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus meinerti</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus meinerti</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus meinerti</i>	Maracá	Semideciduous forest
<i>Odontomachus meinerti</i>	Maracá	Open ombrophylous forest
<i>Odontomachus meinerti</i>	Maracá	Open ombrophylous forest
<i>Odontomachus meinerti</i>	Maracá	Open ombrophylous forest
<i>Odontomachus opaciventris</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus opaciventris</i>	Ducke	Dense ombrophylous forest

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Table B1 (continued)

Taxon	Site	Phytophysiology
<i>Odontomachus opaciventris</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus opaciventris</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus opaciventris</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus opaciventris</i>	Manaquiri	Open ombrophylous forest
<i>Odontomachus scalptus</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus scalptus</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus scalptus</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus scalptus</i>	Ducke	Dense ombrophylous forest
<i>Odontomachus scalptus</i>	Ducke	Dense ombrophylous forest
<i>Oxyepoecus</i> sp. 01	Maracá	Deciduous forest
<i>Pachycondyla crassinoda</i>	Ducke	Dense ombrophylous forest
<i>Pachycondyla crassinoda</i>	Ducke	Dense ombrophylous forest
<i>Pachycondyla crassinoda</i>	Ducke	Dense ombrophylous forest
<i>Pachycondyla crassinoda</i>	Maracá	Campinarana forest
<i>Pachycondyla crassinoda</i>	Manaquiri	Open ombrophylous forest
<i>Pachycondyla crassinoda</i>	Manaquiri	Open ombrophylous forest
<i>Pachycondyla harpax</i>	Ducke	Dense ombrophylous forest
<i>Pachycondyla harpax</i>	Ducke	Dense ombrophylous forest
<i>Pachycondyla harpax</i>	Ducke	Dense ombrophylous forest
<i>Pachycondyla harpax</i>	Manaquiri	Open ombrophylous forest
<i>Pachycondyla harpax</i>	Viruá	Campinarana forest
<i>Pachycondyla impressa</i>	Capana	Dense ombrophylous forest
<i>Pheidole biconstricta</i>	Ducke	Dense ombrophylous forest
<i>Pheidole biconstricta</i>	Ducke	Dense ombrophylous forest
<i>Pheidole biconstricta</i>	Orquestra	Dense ombrophylous forest
<i>Pheidole biconstricta</i>	Viruá	Campinarana forest
<i>Pheidole biconstricta</i>	Viruá	Campinarana forest
<i>Pheidole biconstricta</i>	Ducke	Dense ombrophylous forest
<i>Pheidole cataractae</i>	Ducke	Dense ombrophylous forest
<i>Pheidole cataractae</i>	Ducke	Dense ombrophylous forest
<i>Pheidole cataractae</i>	Maracá	Open ombrophylous forest
<i>Pheidole cataractae</i>	Maracá	Campinarana forest
<i>Pheidole cataractae</i>	Maracá	Campinarana forest
<i>Pheidole embolopyx</i>	Manaquiri	Open ombrophylous forest
<i>Pheidole embolopyx</i>	Manaquiri	Open ombrophylous forest
<i>Pheidole exigua</i>	Manaquiri	Open ombrophylous forest
<i>Pheidole flavens</i>	Ducke	Dense ombrophylous forest
<i>Pheidole flavens</i>	Ducke	Dense ombrophylous forest
<i>Pheidole flavens</i>	Ducke	Dense ombrophylous forest
<i>Pheidole flavens</i>	Maracá	Seasonal campinarana
<i>Pheidole flavens</i>	Maracá	Seasonal campinarana
<i>Pheidole meinerti</i>	Manaquiri	Open ombrophylous forest
<i>Pheidole nitella</i>	Ducke	Dense ombrophylous forest
<i>Pheidole nitella</i>	Ducke	Dense ombrophylous forest
<i>Pheidole nitella</i>	Ducke	Dense ombrophylous forest
<i>Pheidole radoszkowskii</i>	Orquestra	Dense ombrophylous forest
<i>Pheidole radoszkowskii</i>	Maracá	Semideciduous forest
<i>Pheidole radoszkowskii</i>	Ducke	Dense ombrophylous forest
<i>Pheidole radoszkowskii</i>	Viruá	Seasonal shrubby campinarana
<i>Pheidole radoszkowskii</i>	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 02	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 02	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 02	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 02	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 04	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 04	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 09	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 09	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 09	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 09	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 10	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 10	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 10	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 10	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 11	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 11	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 11	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 11	Maracá	Seasonal campinarana

(continued on next page)

Table B1 (continued)

Taxon	Site	Phytophysiology
<i>Pheidole</i> sp. 11	Maracá	Seasonal campinarana
<i>Pheidole</i> sp. 12	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 12	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 12	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 12	Maracá	Deciduous forest
<i>Pheidole</i> sp. 12	Maracá	Deciduous forest
<i>Pheidole</i> sp. 13	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 13	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 13	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 13	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 13	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 14	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 14	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 14	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 14	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 14	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 14	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 14	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 15	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 15	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 15	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 15	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 15	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 16	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 16	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 16	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 16	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 16	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 17	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 17	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 17	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 19	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 19	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 19	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 19	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 20	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 20	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 20	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 20	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 20	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 23	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 23	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 23	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 23	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 23	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 24	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 24	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 24	Virúá	Seasonal campinarana
<i>Pheidole</i> sp. 24	Virúá	Seasonal campinarana
<i>Pheidole</i> sp. 24	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 25	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 25	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 26	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 26	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 26	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 26	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 26	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 27	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 27	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 27	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 27	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 27	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 27	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 28	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 28	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 28	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 28	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 28	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 28	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 29	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 29	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 29	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 30	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 31	Ducke	Dense ombrophylous forest

(continued on next page)

Table B1 (continued)

Taxon	Site	Phytophysiology
<i>Pheidole</i> sp. 31	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 31	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 31	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 32	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 32	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 32	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 32	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 34	Viruá	Campinarana forest
<i>Pheidole</i> sp. 34	Viruá	Campinarana forest
<i>Pheidole</i> sp. 36	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 36	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 37	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 37	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 37	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 38	Viruá	Open ombrophylous forest
<i>Pheidole</i> sp. 38	Viruá	Open ombrophylous forest
<i>Pheidole</i> sp. 41	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 41	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 43	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 43	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 44	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 44	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 44	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 45	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 45	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 45	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 46	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 46	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 46	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 46	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 47	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 47	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 47	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 48	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 48	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 49	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 49	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 49	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 49	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 49	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 51	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 51	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 51	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 51	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 51	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 52	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 52	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 52	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 52	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 53	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 53	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 53	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 54	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 54	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 54	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 55	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 55	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 55	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 55	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 55	Ducke	Dense ombrophylous forest
<i>Pheidole</i> sp. 57	Viruá	Seasonal shrubby campinarana
<i>Pheidole</i> sp. 57	Viruá	Seasonal shrubby campinarana
<i>Pheidole</i> sp. 57	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 57	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 59	Maracá	Semideciduous forest
<i>Pheidole</i> sp. 60	Maracá	Semideciduous forest
<i>Pheidole</i> sp. 60	Maracá	Semideciduous forest
<i>Pheidole</i> sp. 60	Orquestra	Dense ombrophylous forest
<i>Pheidole</i> sp. 60	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 60	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 61	Maracá	Semideciduous forest

(continued on next page)

Table B1 (continued)

Taxon	Site	Phytophysiology
<i>Pheidole</i> sp. 61	Maracá	Semideciduous forest
<i>Pheidole</i> sp. 61	Viruá	Open ombrophylous forest
<i>Pheidole</i> sp. 62	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 62	Maracá	Deciduous forest
<i>Pheidole</i> sp. 62	Maracá	Deciduous forest
<i>Pheidole</i> sp. 63	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 63	Maracá	Deciduous forest
<i>Pheidole</i> sp. 63	Maracá	Deciduous forest
<i>Pheidole</i> sp. 64	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 64	Maracá	Semideciduous forest
<i>Pheidole</i> sp. 64	Maracá	Semideciduous forest
<i>Pheidole</i> sp. 70	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 70	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 70	Maracá	Deciduous forest
<i>Pheidole</i> sp. 70	Viruá	Campinarana forest
<i>Pheidole</i> sp. 70	Viruá	Campinarana forest
<i>Pheidole</i> sp. 71	Maracá	Semideciduous forest
<i>Pheidole</i> sp. 71	Maracá	Semideciduous forest
<i>Pheidole</i> sp. 71	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 71	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 73	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 75	Maracá	Campinarana forest
<i>Pheidole</i> sp. 75	Maracá	Campinarana forest
<i>Pheidole</i> sp. 75	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 75	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 75	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 76	Viruá	Seasonal campinarana
<i>Pheidole</i> sp. 76	Viruá	Seasonal campinarana
<i>Pheidole</i> sp. 76	Viruá	Open ombrophylous forest
<i>Pheidole</i> sp. 76	Maracá	Deciduous forest
<i>Pheidole</i> sp. 78	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 78	Maracá	Semideciduous forest
<i>Pheidole</i> sp. 78	Viruá	Deciduous forest
<i>Pheidole</i> sp. 78	Viruá	Deciduous forest
<i>Pheidole</i> sp. 79	Maracá	Open ombrophylous forest
<i>Pheidole</i> sp. 87	Orquestra	Open ombrophylous forest
<i>Pheidole</i> sp. 87	Orquestra	Open ombrophylous forest
<i>Pheidole</i> sp. 87	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 87	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 87	Capana	Dense ombrophylous forest
<i>Pheidole</i> sp. 91	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 91	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 91	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 97	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 97	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 105	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 105	Jari	Dense ombrophylous forest
<i>Pheidole</i> sp. 111	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 111	Manaquiri	Open ombrophylous forest
<i>Pheidole</i> sp. 120	Jari	Dense ombrophylous forest
<i>Pheidole vorax</i>	Manaquiri	Open ombrophylous forest
<i>Pheidole vorax</i>	Manaquiri	Open ombrophylous forest
<i>Pheidole vorax</i>	Ducke	Dense ombrophylous forest
<i>Pheidole vorax</i>	Orquestra	Dense ombrophylous forest
<i>Pogonomyrmex naegeli</i>	Cauamé	Open savanna
<i>Pogonomyrmex naegeli</i>	Cauamé	Open savanna
<i>Pogonomyrmex naegeli</i>	Cauamé	Open savanna
<i>Pogonomyrmex naegeli</i>	Cauamé	Open savanna
<i>Pogonomyrmex naegeli</i>	Cauamé	Open savanna
<i>Pogonomyrmex naegeli</i>	Cauamé	Open savanna
<i>Pogonomyrmex naegeli</i>	Cauamé	Open savanna
<i>Prionopelta punctulata</i>	Ducke	Dense ombrophylous forest
<i>Prionopelta punctulata</i>	Ducke	Dense ombrophylous forest
<i>Prionopelta punctulata</i>	Ducke	Dense ombrophylous forest
<i>Prionopelta punctulata</i>	Ducke	Dense ombrophylous forest
<i>Prionopelta punctulata</i>	Ducke	Dense ombrophylous forest
<i>Prionopelta punctulata</i>	Ducke	Dense ombrophylous forest
<i>Pseudomyrmex flavidulus</i>	Cauamé	Open savanna
<i>Pseudomyrmex flavidulus</i>	Cauamé	Open savanna
<i>Pseudomyrmex flavidulus</i>	Maracá	Deciduous forest
<i>Pseudomyrmex</i> sp. 05	Viruá	Seasonal campinarana

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Table B1 (continued)

Taxon	Site	Phytophysiology
<i>Pseudomyrmex</i> sp. 05	Viruí	Seasonal campinarana
<i>Pseudomyrmex</i> sp. 05	Cauamé	Open savanna
<i>Pseudomyrmex</i> sp. 05	Cauamé	Open savanna
<i>Pseudomyrmex tenuis</i>	Orquestra	Dense ombrophylous forest
<i>Pseudomyrmex tenuis</i>	Capana	Dense ombrophylous forest
<i>Pseudomyrmex tenuis</i>	Jari	Dense ombrophylous forest
<i>Rasopone arhuaca</i>	Maracá	Semideciduous forest
<i>Rasopone arhuaca</i>	Maracá	Semideciduous forest
<i>Rasopone arhuaca</i>	Maracá	Open ombrophylous forest
<i>Rasopone arhuaca</i>	Ducke	Dense ombrophylous forest
<i>Rasopone arhuaca</i>	Ducke	Dense ombrophylous forest
<i>Rasopone arhuaca</i>	Ducke	Dense ombrophylous forest
<i>Rogeria alzatei</i>	Ducke	Dense ombrophylous forest
<i>Rogeria alzatei</i>	Ducke	Dense ombrophylous forest
<i>Rogeria alzatei</i>	Ducke	Dense ombrophylous forest
<i>Rogeria alzatei</i>	Ducke	Dense ombrophylous forest
<i>Rogeria alzatei</i>	Ducke	Dense ombrophylous forest
<i>Rogeria cornuta</i>	Capana	Dense ombrophylous forest
<i>Rogeria cornuta</i>	Capana	Dense ombrophylous forest
<i>Rogeria cornuta</i>	Capana	Dense ombrophylous forest
<i>Rogeria cornuta</i>	Capana	Dense ombrophylous forest
<i>Rogeria foreli</i>	Maracá	Deciduous forest
<i>Rogeria leptonana</i>	Maracá	Deciduous forest
<i>Rogeria</i> sp. 01	Capana	Dense ombrophylous forest
<i>Sericomyrmex</i> sp. 01	Manaquiri	Open ombrophylous forest
<i>Sericomyrmex</i> sp. 01	Manaquiri	Open ombrophylous forest
<i>Sericomyrmex</i> sp. 01	Maracá	Semideciduous forest
<i>Sericomyrmex</i> sp. 01	Maracá	Semideciduous forest
<i>Sericomyrmex</i> sp. 01	Maracá	Campinarana forest
<i>Sericomyrmex</i> sp. 01	Maracá	Dense ombrophylous forest
<i>Solenopsis brevicornis</i>	Maracá	Dense ombrophylous forest
<i>Solenopsis brevicornis</i>	Ducke	Dense ombrophylous forest
<i>Solenopsis brevicornis</i>	Ducke	Dense ombrophylous forest
<i>Solenopsis brevicornis</i>	Ducke	Dense ombrophylous forest
<i>Solenopsis castor</i>	Maracá	Deciduous forest
<i>Solenopsis castor</i>	Maracá	Deciduous forest
<i>Solenopsis castor</i>	Jari	Dense ombrophylous forest
<i>Solenopsis castor</i>	Jari	Dense ombrophylous forest
<i>Solenopsis clytemnestra</i>	Cauamé	Open savanna
<i>Solenopsis clytemnestra</i>	Cauamé	Open savanna
<i>Solenopsis clytemnestra</i>	Manaquiri	Open ombrophylous forest
<i>Solenopsis clytemnestra</i>	Ducke	Dense ombrophylous forest
<i>Solenopsis clytemnestra</i>	Ducke	Dense ombrophylous forest
<i>Solenopsis geminata</i>	Ducke	Dense ombrophylous forest
<i>Solenopsis geminata</i>	Ducke	Dense ombrophylous forest
<i>Solenopsis geminata</i>	Maracá	Deciduous forest
<i>Solenopsis geminata</i>	Maracá	Deciduous forest
<i>Solenopsis geminata</i>	Maracá	Deciduous forest
<i>Solenopsis saevissima</i>	Cauamé	Open savanna
<i>Solenopsis saevissima</i>	Cauamé	Open savanna
<i>Solenopsis saevissima</i>	Ducke	Dense ombrophylous forest
<i>Solenopsis saevissima</i>	Ducke	Dense ombrophylous forest
<i>Solenopsis saevissima</i>	Ducke	Dense ombrophylous forest
<i>Solenopsis</i> sp. 06	Cauamé	Open savanna
<i>Solenopsis</i> sp. 06	Manaquiri	Open ombrophylous forest
<i>Solenopsis</i> sp. 06	Ducke	Dense ombrophylous forest
<i>Solenopsis</i> sp. 06	Ducke	Dense ombrophylous forest
<i>Solenopsis</i> sp. 09	Ducke	Dense ombrophylous forest
<i>Solenopsis</i> sp. 09	Ducke	Dense ombrophylous forest
<i>Solenopsis</i> sp. 09	Ducke	Dense ombrophylous forest
<i>Solenopsis</i> sp. 09	Ducke	Dense ombrophylous forest
<i>Solenopsis</i> sp. 11	Maracá	Open ombrophylous forest
<i>Solenopsis</i> sp. 11	Maracá	Deciduous forest
<i>Solenopsis</i> sp. 11	Maracá	Deciduous forest
<i>Strumigenys appretiata</i>	Maracá	Semideciduous forest
<i>Strumigenys appretiata</i>	Maracá	Semideciduous forest
<i>Strumigenys appretiata</i>	Maracá	Semideciduous forest
<i>Strumigenys beebei</i>	Capana	Dense ombrophylous forest
<i>Strumigenys beebei</i>	Capana	Dense ombrophylous forest

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Table B1 (continued)

Taxon	Site	Phytophysognomy
<i>Strumigenys beebei</i>	Capana	Dense ombrophylous forest
<i>Strumigenys beebei</i>	Capana	Dense ombrophylous forest
<i>Strumigenys beebei</i>	Capana	Dense ombrophylous forest
<i>Strumigenys cosmostela</i>	Manaquiri	Open ombrophylous forest
<i>Strumigenys cosmostela</i>	Manaquiri	Open ombrophylous forest
<i>Strumigenys denticulata</i>	Capana	Dense ombrophylous forest
<i>Strumigenys denticulata</i>	Capana	Dense ombrophylous forest
<i>Strumigenys denticulata</i>	Capana	Dense ombrophylous forest
<i>Strumigenys denticulata</i>	Jari	Dense ombrophylous forest
<i>Strumigenys denticulata</i>	Jari	Dense ombrophylous forest
<i>Strumigenys denticulata</i>	Jari	Dense ombrophylous forest
<i>Strumigenys elongata</i>	Maracá	Open ombrophylous forest
<i>Strumigenys elongata</i>	Maracá	Open ombrophylous forest
<i>Strumigenys elongata</i>	Maracá	Open ombrophylous forest
<i>Strumigenys elongata</i>	Maracá	Open ombrophylous forest
<i>Strumigenys infidelis</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys infidelis</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys infidelis</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys infidelis</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys inusitata</i>	Jari	Dense ombrophylous forest
<i>Strumigenys perparva</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys perparva</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys perparva</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys perparva</i>	Manaquiri	Open ombrophylous forest
<i>Strumigenys perparva</i>	Manaquiri	Open ombrophylous forest
<i>Strumigenys</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Strumigenys</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Strumigenys</i> sp. 02	Ducke	Dense ombrophylous forest
<i>Strumigenys</i> sp. 02	Ducke	Dense ombrophylous forest
<i>Strumigenys</i> sp. 02	Ducke	Dense ombrophylous forest
<i>Strumigenys</i> sp. 02	Ducke	Dense ombrophylous forest
<i>Strumigenys</i> sp. 08	Maracá	Open ombrophylous forest
<i>Strumigenys</i> sp. 08	Maracá	Open ombrophylous forest
<i>Strumigenys</i> sp. 08	Maracá	Semideciduous forest
<i>Strumigenys</i> sp. 08	Maracá	Semideciduous forest
<i>Strumigenys stenotes</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys stenotes</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys stenotes</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys trinidadensis</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys trinidadensis</i>	Ducke	Dense ombrophylous forest
<i>Strumigenys trinidadensis</i>	Maracá	Open ombrophylous forest
<i>Strumigenys trinidadensis</i>	Maracá	Open ombrophylous forest
<i>Strumigenys tridifera</i>	Jari	Dense ombrophylous forest
<i>Strumigenys tridifera</i>	Jari	Dense ombrophylous forest
<i>Strumigenys tridifera</i>	Jari	Dense ombrophylous forest
<i>Strumigenys tridifera</i>	Jari	Dense ombrophylous forest
<i>Strumigenys tridifera</i>	Jari	Dense ombrophylous forest
<i>Strumigenys tridifera</i>	Jari	Dense ombrophylous forest
<i>Strumigenys villiersi</i>	Capana	Dense ombrophylous forest
<i>Strumigenys villiersi</i>	Capana	Dense ombrophylous forest
<i>Strumigenys villiersi</i>	Jari	Dense ombrophylous forest
<i>Strumigenys zeteki</i>	Orquestra	Dense ombrophylous forest
<i>Strumigenys zeteki</i>	Orquestra	Dense ombrophylous forest
<i>Strumigenys zeteki</i>	Jari	Dense ombrophylous forest
<i>Strumigenys zeteki</i>	Jari	Dense ombrophylous forest
<i>Tapinoma</i> sp. 01	Cauamé	Open savanna
<i>Tapinoma</i> sp. 01	Cauamé	Open savanna
<i>Tapinoma</i> sp. 01	Cauamé	Open savanna
<i>Tapinoma</i> sp. 01	Cauamé	Open savanna
<i>Tapinoma</i> sp. 01	Cauamé	Open savanna
<i>Trachymyrmex bugnioni</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex bugnioni</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex bugnioni</i>	Virúá	Campinarana forest
<i>Trachymyrmex cornetzi</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex cornetzi</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex cornetzi</i>	Manaquiri	Open ombrophylous forest
<i>Trachymyrmex cornetzi</i>	Manaquiri	Open ombrophylous forest
<i>Trachymyrmex cornetzi</i>	Maracá	Semideciduous forest
<i>Trachymyrmex cornetzi</i>	Maracá	Semideciduous forest
<i>Trachymyrmex diversus</i>	Manaquiri	Open ombrophylous forest

(continued on next page)

Table B1 (continued)

Taxon	Site	Phytophysiology
<i>Trachymyrmex diversus</i>	Manaquiri	Open ombrophylous forest
<i>Trachymyrmex diversus</i>	Manaquiri	Open ombrophylous forest
<i>Trachymyrmex diversus</i>	Manaquiri	Open ombrophylous forest
<i>Trachymyrmex farinosus</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex farinosus</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex farinosus</i>	Maracá	Open ombrophylous forest
<i>Trachymyrmex farinosus</i>	Maracá	Open ombrophylous forest
<i>Trachymyrmex farinosus</i>	Maracá	Open ombrophylous forest
<i>Trachymyrmex isthmicus</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex isthmicus</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex mandibulares</i>	Manaquiri	Open ombrophylous forest
<i>Trachymyrmex opulentus</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex opulentus</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex opulentus</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex opulentus</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex opulentus</i>	Ducke	Dense ombrophylous forest
<i>Trachymyrmex ruthae</i>	Maracá	Semideciduous forest
<i>Trachymyrmex ruthae</i>	Maracá	Semideciduous forest
<i>Trachymyrmex</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 01	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 02	Manaquiri	Open ombrophylous forest
<i>Trachymyrmex</i> sp. 04	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 04	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 04	Manaquiri	Open ombrophylous forest
<i>Trachymyrmex</i> sp. 04	Maracá	Open ombrophylous forest
<i>Trachymyrmex</i> sp. 05	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 05	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 05	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 05	Maracá	Open ombrophylous forest
<i>Trachymyrmex</i> sp. 05	Maracá	Open ombrophylous forest
<i>Trachymyrmex</i> sp. 06	Virúá	Campinarana forest
<i>Trachymyrmex</i> sp. 06	Maracá	Semideciduous forest
<i>Trachymyrmex</i> sp. 07	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 07	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 07	Manaquiri	Open ombrophylous forest
<i>Trachymyrmex</i> sp. 07	Maracá	Open ombrophylous forest
<i>Trachymyrmex</i> sp. 07	Virúá	Campinarana forest
<i>Trachymyrmex</i> sp. 07	Virúá	Campinarana forest
<i>Trachymyrmex</i> sp. 08	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 08	Ducke	Dense ombrophylous forest
<i>Trachymyrmex</i> sp. 08	Maracá	Open ombrophylous forest
<i>Trachymyrmex</i> sp. 08	Maracá	Open ombrophylous forest
<i>Tranopelta gilva</i>	Maracá	Campinarana forest
<i>Wasmannia auropunctata</i>	Manaquiri	Open ombrophylous forest
<i>Wasmannia auropunctata</i>	Manaquiri	Open ombrophylous forest
<i>Wasmannia auropunctata</i>	Ducke	Dense ombrophylous forest
<i>Wasmannia auropunctata</i>	Ducke	Dense ombrophylous forest
<i>Wasmannia auropunctata</i>	Jari	Dense ombrophylous forest
<i>Wasmannia auropunctata</i>	Jari	Dense ombrophylous forest
<i>Wasmannia iheringi</i>	Ducke	Dense ombrophylous forest
<i>Wasmannia rochai</i>	Maracá	Open ombrophylous forest
<i>Wasmannia rochai</i>	Maracá	Open ombrophylous forest
<i>Wasmannia scrobifera</i>	Ducke	Dense ombrophylous forest
<i>Wasmannia scrobifera</i>	Ducke	Dense ombrophylous forest
<i>Wasmannia scrobifera</i>	Ducke	Dense ombrophylous forest
<i>Wasmannia scrobifera</i>	Ducke	Dense ombrophylous forest
<i>Wasmannia scrobifera</i>	Ducke	Dense ombrophylous forest

Table B2

Total number of species/morphospecies collected, taxon and number of specimens measured of each species and total amount of ants measured.

Number of species/morphospecies	Taxon	Number of specimens measured for each species
1	<i>Acanthognathus ocellatus</i>	1
2	<i>Acanthostichus bentoni</i>	6
3	<i>Acromyrmex</i> sp. 01	2
4	<i>Acromyrmex subterraneus</i>	1
5	<i>Acropyga</i> sp. 01	6
6	<i>Acropyga</i> sp. 02	1
7	<i>Anochetus diegensis</i>	6
8	<i>Anochetus emarginatus</i>	1
9	<i>Anochetus horridus</i>	6
10	<i>Apterostigma pilosum</i>	3
11	<i>Apterostigma</i> sp. 03	1
12	<i>Apterostigma</i> sp. 04	1
13	<i>Apterostigma urichii</i>	4
14	<i>Atta cephalotes</i>	3
15	<i>Atta sexdens</i>	6
16	<i>Azteca</i> sp. 01	4
17	<i>Blepharidatta brasiliensis</i>	6
18	<i>Brachymyrmex heeri</i>	5
19	<i>Brachymyrmex</i> sp. 01	5
20	<i>Brachymyrmex</i> sp. 02	3
21	<i>Brachymyrmex</i> sp. 03	1
22	<i>Camponotus ager</i>	5
23	<i>Camponotus atriceps</i>	6
24	<i>Camponotus balzani</i>	3
25	<i>Camponotus crassus</i>	5
26	<i>Camponotus fastigatus</i>	3
27	<i>Camponotus femoratus</i>	6
28	<i>Camponotus leydigi</i>	3
29	<i>Camponotus novogranadensis</i>	5
30	<i>Camponotus rapax</i>	6
31	<i>Camponotus sericeiventris</i>	2
32	<i>Camponotus</i> sp. 14	1
33	<i>Carebara escherichi</i>	4
34	<i>Carebara lignata</i>	1
35	<i>Carebara urichi</i>	5
36	<i>Centromyrmex gigas</i>	1
37	<i>Centromyrmex</i> sp. 01	1
38	<i>Cephalotes atratus</i>	2
39	<i>Cephalotes maculatus</i>	2
40	<i>Cephalotes marginatus</i>	1
41	<i>Cephalotes pusillus</i>	3
42	<i>Crematogaster brasiliensis</i>	6
43	<i>Crematogaster curvispinosa</i>	1
44	<i>Crematogaster erecta</i>	2
45	<i>Crematogaster evallans</i>	3
46	<i>Crematogaster flavosensitiva</i>	5
47	<i>Crematogaster jardineri</i>	4
48	<i>Crematogaster limata</i>	5
49	<i>Crematogaster longispina</i>	2
50	<i>Crematogaster nigropilosa</i>	2
51	<i>Crematogaster rochai</i>	1
52	<i>Crematogaster sotobosque</i>	5
53	<i>Crematogaster stollii</i>	2
54	<i>Crematogaster tenuicula</i>	6
55	<i>Cyphomyrmex laevigatus</i>	6
56	<i>Cyphomyrmex peltatus</i>	6
57	<i>Cyphomyrmex rimosus</i>	6
58	<i>Discothyrea denticulata</i>	1
59	<i>Dolichoderus bispinosus</i>	6
60	<i>Dolichoderus decollatus</i>	2
61	<i>Dolichoderus imitator</i>	4
62	<i>Dorymyrmex bicolor</i>	3
63	<i>Dorymyrmex goeldii</i>	4
64	<i>Dorymyrmex richteri</i>	4
65	<i>Eciton burchellii</i>	4
66	<i>Eciton dulcius</i>	2
67	<i>Eciton rapax</i>	4
68	<i>Ectatomma brunneum</i>	4
69	<i>Ectatomma edentatum</i>	7

(continued on next page)

Table B2 (continued)

Number of species/morphospecies	Taxon	Number of specimens measured for each species
70	<i>Ectatomma lugens</i>	6
71	<i>Ectatomma ruidum</i>	3
72	<i>Gigantiops destructor</i>	5
73	<i>Gnamptogenys acuminata</i>	4
74	<i>Gnamptogenys curvoclypeata</i>	3
75	<i>Gnamptogenys haenschii</i>	3
76	<i>Gnamptogenys horni</i>	6
77	<i>Gnamptogenys moelleri</i>	4
78	<i>Gnamptogenys regularis</i>	4
79	<i>Gnamptogenys strigata</i>	2
80	<i>Gnamptogenys sulcata</i>	4
81	<i>Gnamptogenys tortuolosa</i>	6
82	<i>Hylomyrma inmanis</i>	4
83	<i>Hypoponera</i> sp. 01	5
84	<i>Hypoponera</i> sp. 02	2
85	<i>Hypoponera</i> sp. 03	4
86	<i>Hypoponera</i> sp. 04	5
87	<i>Hypoponera</i> sp. 05	5
88	<i>Hypoponera</i> sp. 06	4
89	<i>Hypoponera</i> sp. 07	5
90	<i>Hypoponera</i> sp. 08	4
91	<i>Hypoponera</i> sp. 09	4
92	<i>Hypoponera</i> sp. 11	3
93	<i>Hypoponera</i> sp. 14	4
94	<i>Kalathomyrmex emryi</i>	1
95	<i>Labidus coecus</i>	6
96	<i>Labidus mars</i>	1
97	<i>Labidus praedator</i>	6
98	<i>Labidus spininodis</i>	2
99	<i>Leptogenys gaigei</i>	6
100	<i>Leptogenys pusilla</i>	2
101	<i>Leptogenys wheeleri</i>	5
102	<i>Mayaponera constricta</i>	6
103	<i>Megalomyrmex balzani</i>	4
104	<i>Megalomyrmex driftii</i>	4
105	<i>Megalomyrmex incisus</i>	2
106	<i>Megalomyrmex leoninus</i>	4
107	<i>Megalomyrmex silvestrii</i>	3
108	<i>Megalomyrmex</i> sp. 04	1
109	<i>Monomorium pharaonis</i>	6
110	<i>Mycocarpus smithii</i>	4
111	<i>Mycocarpus</i> sp. 01	3
112	<i>Myrmicocrypta</i> sp. 01	4
113	<i>Myrmicocrypta</i> sp. 02	3
114	<i>Neivamyrmex adnepos</i>	1
115	<i>Neivamyrmex argustinodis</i>	4
116	<i>Neivamyrmex gibbatus</i>	4
117	<i>Neivamyrmex iridescens</i>	1
118	<i>Neivamyrmex orthonotus</i>	2
119	<i>Neivamyrmex pilosus</i>	2
120	<i>Neivamyrmex swainsonii</i>	3
121	<i>Neoponera apicalis</i>	6
122	<i>Neoponera commutata</i>	6
123	<i>Neoponera laevigata</i>	1
124	<i>Neoponera verena</i>	4
125	<i>Nomamyrmex esenbeckii</i>	3
126	<i>Nomamyrmex hartigi</i>	2
127	<i>Nylanderia caeciliae</i>	6
128	<i>Nylanderia guatemalensis</i>	6
129	<i>Nylanderia</i> sp. 01	5
130	<i>Nylanderia</i> sp. 04	4
131	<i>Ochetomyrmex semipolitus</i>	6
132	<i>Octostruma balzani</i>	5
133	<i>Odontomachus bauri</i>	4
134	<i>Odontomachus brunneus</i>	1
135	<i>Odontomachus caelatus</i>	6
136	<i>Odontomachus haematodus</i>	6
137	<i>Odontomachus laticeps</i>	3
138	<i>Odontomachus meinerti</i>	6
139	<i>Odontomachus opaciventris</i>	6

(continued on next page)

Table B2 (continued)

Number of species/morphospecies	Taxon	Number of specimens measured for each species
140	<i>Odontomachus scalptus</i>	5
141	<i>Oxyepoecus</i> sp. 01	1
142	<i>Pachycondyla crassinoda</i>	6
143	<i>Pachycondyla harpax</i>	5
144	<i>Pachycondyla impressa</i>	1
145	<i>Pheidole biconstricta</i>	6
146	<i>Pheidole cataractae</i>	5
147	<i>Pheidole embolopyx</i>	2
148	<i>Pheidole exigua</i>	1
149	<i>Pheidole flavens</i>	5
150	<i>Pheidole meinerti</i>	1
151	<i>Pheidole nitella</i>	3
152	<i>Pheidole radoszkowskii</i>	5
153	<i>Pheidole</i> sp. 01	2
154	<i>Pheidole</i> sp. 02	4
155	<i>Pheidole</i> sp. 04	2
156	<i>Pheidole</i> sp. 09	4
157	<i>Pheidole</i> sp. 10	4
158	<i>Pheidole</i> sp. 105	2
159	<i>Pheidole</i> sp. 11	5
160	<i>Pheidole</i> sp. 111	2
161	<i>Pheidole</i> sp. 12	5
162	<i>Pheidole</i> sp. 120	1
163	<i>Pheidole</i> sp. 13	5
164	<i>Pheidole</i> sp. 14	6
165	<i>Pheidole</i> sp. 15	5
166	<i>Pheidole</i> sp. 16	4
167	<i>Pheidole</i> sp. 17	3
168	<i>Pheidole</i> sp. 19	4
169	<i>Pheidole</i> sp. 20	5
170	<i>Pheidole</i> sp. 23	5
171	<i>Pheidole</i> sp. 24	5
172	<i>Pheidole</i> sp. 25	2
173	<i>Pheidole</i> sp. 26	4
174	<i>Pheidole</i> sp. 27	6
175	<i>Pheidole</i> sp. 28	5
176	<i>Pheidole</i> sp. 29	3
177	<i>Pheidole</i> sp. 30	1
178	<i>Pheidole</i> sp. 31	4
179	<i>Pheidole</i> sp. 32	4
180	<i>Pheidole</i> sp. 34	2
181	<i>Pheidole</i> sp. 36	2
182	<i>Pheidole</i> sp. 37	3
183	<i>Pheidole</i> sp. 38	2
184	<i>Pheidole</i> sp. 41	2
185	<i>Pheidole</i> sp. 43	2
186	<i>Pheidole</i> sp. 44	3
187	<i>Pheidole</i> sp. 45	3
188	<i>Pheidole</i> sp. 46	3
189	<i>Pheidole</i> sp. 47	3
190	<i>Pheidole</i> sp. 48	2
191	<i>Pheidole</i> sp. 49	5
192	<i>Pheidole</i> sp. 51	5
193	<i>Pheidole</i> sp. 52	4
194	<i>Pheidole</i> sp. 53	3
195	<i>Pheidole</i> sp. 54	3
196	<i>Pheidole</i> sp. 55	5
197	<i>Pheidole</i> sp. 57	4
198	<i>Pheidole</i> sp. 59	1
199	<i>Pheidole</i> sp. 60	5
200	<i>Pheidole</i> sp. 61	3
201	<i>Pheidole</i> sp. 62	3
202	<i>Pheidole</i> sp. 63	3
203	<i>Pheidole</i> sp. 64	3
204	<i>Pheidole</i> sp. 70	5
205	<i>Pheidole</i> sp. 71	4
206	<i>Pheidole</i> sp. 73	1
207	<i>Pheidole</i> sp. 75	5
208	<i>Pheidole</i> sp. 76	4
209	<i>Pheidole</i> sp. 78	4
210	<i>Pheidole</i> sp. 79	1
211	<i>Pheidole</i> sp. 87	5
212	<i>Pheidole</i> sp. 91	3
213	<i>Pheidole</i> sp. 97	2

(continued on next page)

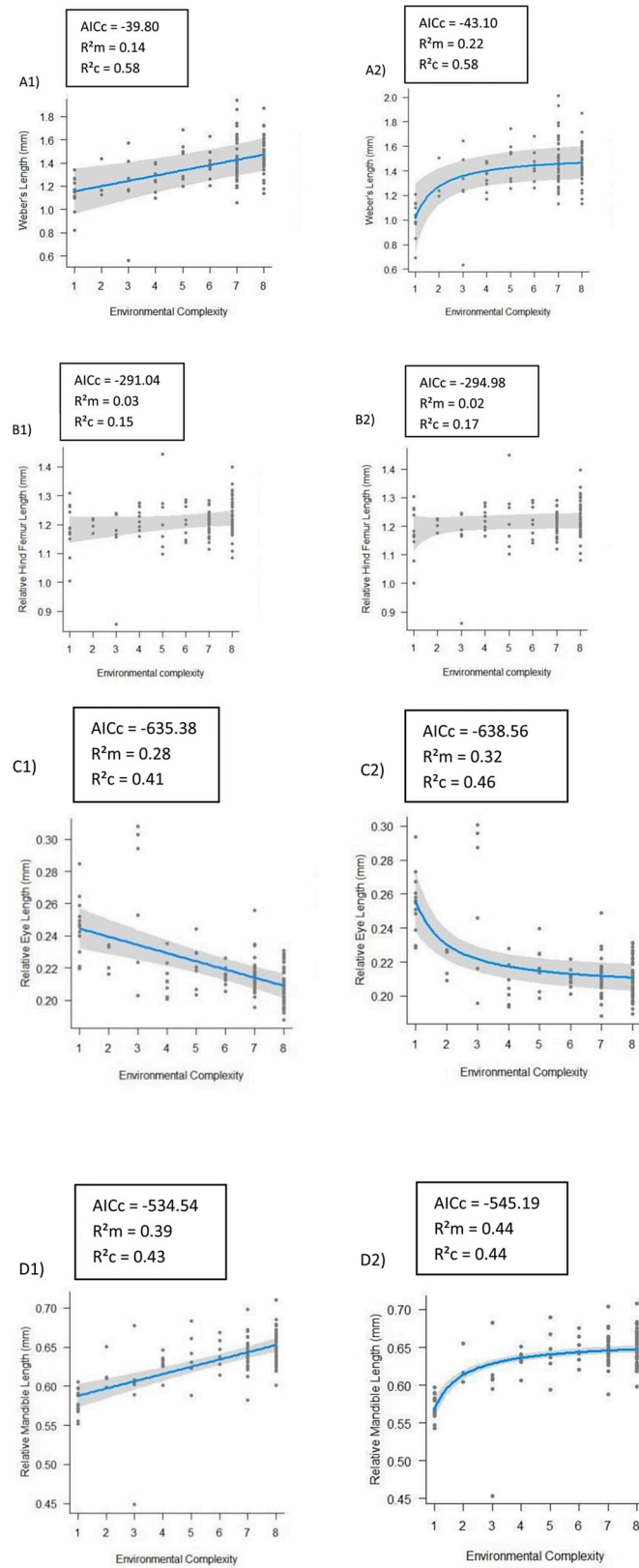
Table B2 (continued)

Number of species/morphospecies	Taxon	Number of specimens measured for each species
214	<i>Pheidole vorax</i>	4
215	<i>Pogonomyrmex naegelii</i>	6
216	<i>Prionopelta punctulata</i>	6
217	<i>Pseudomyrmex flavidulus</i>	3
218	<i>Pseudomyrmex</i> sp. 05	4
219	<i>Pseudomyrmex tenuis</i>	3
220	<i>Rasopone arhuaca</i>	6
221	<i>Rogeria alzatei</i>	5
222	<i>Rogeria cornuta</i>	5
223	<i>Rogeria foreli</i>	1
224	<i>Rogeria leptonana</i>	1
225	<i>Rogeria</i> sp. 01	1
226	<i>Sericomyrmex</i> sp 01	6
227	<i>Solenopsis brevicornis</i>	4
228	<i>Solenopsis castor</i>	4
229	<i>Solenopsis clytemnestra</i>	5
230	<i>Solenopsis geminata</i>	5
231	<i>Solenopsis saevissima</i>	5
232	<i>Solenopsis</i> sp. 06	4
233	<i>Solenopsis</i> sp. 09	4
234	<i>Solenopsis</i> sp. 11	3
235	<i>Strumigenys appretiata</i>	3
236	<i>Strumigenys beebei</i>	5
237	<i>Strumigenys cosmostela</i>	2
238	<i>Strumigenys denticulata</i>	6
239	<i>Strumigenys elongata</i>	4
240	<i>Strumigenys infidelis</i>	4
241	<i>Strumigenys inusitata</i>	1
242	<i>Strumigenys perparva</i>	5
243	<i>Strumigenys</i> sp. 01	2
244	<i>Strumigenys</i> sp. 02	4
245	<i>Strumigenys</i> sp. 08	4
246	<i>Strumigenys stenotes</i>	3
247	<i>Strumigenys trinidadensis</i>	4
248	<i>Strumigenys trudifera</i>	6
249	<i>Strumigenys villiersi</i>	3
250	<i>Strumigenys zeteki</i>	4
251	<i>Tapinoma</i> sp. 01	5
252	<i>Trachymyrmex bugnioni</i>	3
253	<i>Trachymyrmex cornetzi</i>	6
254	<i>Trachymyrmex diversus</i>	4
255	<i>Trachymyrmex farinosus</i>	5
256	<i>Trachymyrmex isthmicus</i>	2
257	<i>Trachymyrmex mandibulares</i>	1
258	<i>Trachymyrmex opulentus</i>	5
259	<i>Trachymyrmex ruthae</i>	2
260	<i>Trachymyrmex</i> sp. 01	5
261	<i>Trachymyrmex</i> sp. 02	1
262	<i>Trachymyrmex</i> sp. 04	4
263	<i>Trachymyrmex</i> sp. 05	5
264	<i>Trachymyrmex</i> sp. 06	2
265	<i>Trachymyrmex</i> sp. 07	6
266	<i>Trachymyrmex</i> sp. 08	4
267	<i>Tranopelta gilva</i>	1
268	<i>Wasmannia auropunctata</i>	6
269	<i>Wasmannia iheringi</i>	1
270	<i>Wasmannia rochai</i>	2
271	<i>Wasmannia scrobifera</i>	5

Table C

Results of Akaike Information Criterion corrected (AICc) for both models (linear and asymptotic models) and *P* value. The lower the value of AICc the more supported is the fit of the model. More negative values indicate lower AICc values.

	Linear models		Asymptotic models	
	AICc	<i>P</i>	AICc	<i>P</i>
Weber length	-39.80	< 0.05	-43.10	< 0.05
Femur length	-291.04	0.146	-294.98	0.258
Eye length	-635.38	< 0.05	-638.56	< 0.05
Mandible length	-534.54	< 0.05	-545.19	< 0.05



Figures. Figures of partial of the linear and asymptotic models with the values of AICc with R²m and R²c value of each model in the upper left corner. Figure A1, B1, C1 and D1 are the results of the linear models of the Weber Length, Relative Hind Femur Length, Relative Eye Length and Relative Mandible Length, respectively. Figures A2, B2, C2 and D2 are the results of the asymptotic models for each trait respectively

References

- Amado, T.F., Bidau, C.J., Olalla-Tárraga, M.Á., 2018. Geographic Variation of Body size in New World Anurans: energy and water in a balance. *Ecography*. <https://doi.org/10.1111/ecog.03889>.
- Aragão, L.E.O.C., Malhi, Y., Metcalfe, D.B., Silva-Espejo, J.E., Jiménez, E., Navarrete, D., Almeida, S., Costa, A.C.L., Salinas, N., Phillips, O.L., Anderson, L.O., Alvarez, E., Baker, T.R., Goncalves, P.H., Huamán-Ovalle, J., Mamani-Solórzano, M., Meir, P., Monteagudo, A., Patiño, S., Peñuela, M.C., Prieto, A., Quesada, C.A., Rozas-Dávila, A., Rudas, A., Silva Jr., J.A., Vásquez, R., 2009. Above- and below-ground net primary productivity across ten Amazonian forests on contrasting soils. *Biogeosciences* 6, 2759–2778. <https://doi.org/10.5194/bg-6-2759>.
- Arnan, X., Cerdá, X., Retana, J., 2017. Relationships among taxonomic, functional, and phylogenetic ant diversity across the biogeographic regions of Europe. *Ecography* 40, 448–457. <https://doi.org/10.1111/ecog.01938>.
- Baccaro, F.B., Feitosa, R.M., Fernandez, F., Fernandes, I.O., Izzo, T.J., Souza, J.L.P., Solar, R., 2015. Guia para os gêneros de formigas do Brasil. Editora INPA, Manaus. <https://doi.org/10.5281/zenodo.32912>.
- Baccaro, F.B., Rocha, I.F., Aguilá, B.E.G., Schietti, J., Emilio, T., Pinto, J.L.P.V., Lima, A.P., Magnusson, W.E., 2013. Changes in ground-dwelling ant functional diversity are correlated with water-table level in an amazonian terra firme forest. *Biotropica* 45, 755–763. <https://doi.org/10.1111/btp.12055>.
- Barlow, N.D., 1994. Size distributions of butterfly species and the effect of latitude on species sizes. *Oikos* 71, 326–332. <https://doi.org/10.2307/3546281>.
- Barton, K., 2009. MuMIn: Multi-Model Inference, R Package Version 0.12.0. <http://r-forge.r-project.org/projects/mumin/>.
- Barton, P.S., Gibb, H., Manning, A.D., Lindenmayer, D.B., Cunningham, S.A., 2011. Morphological traits as predictors of diet and microhabitat use in a diverse beetle assemblage 301–310. *J. Biol. Life Sci.* 102, 301–310. <https://doi.org/10.1111/j.1095-8312.2010.01580.x>.
- Bishop, T.R., Robertson, M.P., Gibb, H., van Rensburg, B.J., Braschler, B., Chown, S.L., Foord, S.H., Munyai, T.C., Okey, I., Tshivhandekano, P.G., Werenkraut, V., Parr, C.L., 2016. Ant assemblages have darker and larger members in cold environments. *Glob. Ecol. Biogeogr.* 25, 1489–1499. <https://doi.org/10.1111/geb.12516>.
- Bishop, T.R., Robertson, M.P., van Rensburg, B.J., Parr, C.L., 2015. Contrasting species and functional beta diversity in montane ant assemblages. *J. Biogeogr.* 42, 1776–1786. <https://doi.org/10.1111/jbi.12537>.
- Breheny, P., Burchett, W., 2016. visreg: visualization of regression models. R package version 2.
- Brown, W.L., Wilson, E.O., 1959. The evolution of the dacetine ants. *Q. Rev. Biol.* 34, 278–294. <https://doi.org/10.1086/516403>.
- Burnham, K.P., Anderson, D.R., 2002. *Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach*. Springer, Berlin.
- Cerdá, X., Retana, J., 2000. Alternative strategies by thermophilic ants to cope with extreme heat: individual versus colony level traits. *Oikos* 89, 155–163. <https://doi.org/10.1034/j.1600-0706.2000.890117.x>.
- Chapman, R.F., 1998. *The Insects: Structure and Function*, fourth ed. Cambridge University Press, New York.
- Chauvel, A., Lucas, Y., Boulet, R., 1987. On the genesis of the soil mantle of the region of Manaus, Central Amazonia, Brazil. *Experientia* 43 234–24. <https://doi.org/10.1007/BF01945546>.
- Chown, S.L., Gaston, K.J., 2010. Body size variation in insects: a macroecological perspective. *Biol. Rev.* 85, 139–169. <https://doi.org/10.1111/j.1469-185X.2009.00097.x>.
- Clusella Trullas, S., Wyk, J.H.V., Spotila, J.R., 2007. Thermal melanism in ectotherms. *J. Therm. Biol.* 32, 235–245. <https://doi.org/10.1016/j.jtherbio.2007.01.013>.
- Costa, F.R.C., Magnusson, W.E., 2010. The need for large-scale, integrated studies of biodiversity – the experience of the Program for biodiversity research in Brazilian Amazonia. *Nat. Conserv.* 8, 3–12. <https://doi.org/10.4322/natcon.00801001>.
- do Vale, J.D.D., Zuanon, J., Magnusson, W.E., 2014. The influence of rain in limnological characteristics of Viruá wetlands, Brazilian Amazon. *Acta Limnol. Bras.* 26, 254–267. <https://doi.org/10.1590/S2179-975X2014000300005>.
- Dunn, R.R., Guenard, B., Weiser, M.D., Sanders, N.J., 2009. *Geographic gradients*. In: Lack, L., Parr, C., Abbott, K.L. (Eds.), *Ant Ecology*. Oxford University Press Inc., New York, pp. 38–58.
- Emilio, T., Nelson, B.W., Schietti, J., Desmoulière, S.J.M., Santo, E., Helder, M.V., Costa, F.R., 2010. Assessing the relationship between forest types and canopy tree beta diversity in Amazonia. *Ecography* 33, 738–747. <https://doi.org/10.1111/j.1600-0587.2009.06139.x>.
- Farji-Brener, A.G., Barrantes, G., Ruggiero, A., 2004. Environmental rugosity, body size and access to food: a test of the size-grain hypothesis in tropical litter ants. *Oikos* 104, 165–171. <https://doi.org/10.1111/j.0030-1299.2004.12740.x>.
- Fowler, H.G., Forti, L.C., Brandão, C.R.F., Delabie, J.H.C., Vasconcelos, H.L., 1991. *Ecologia nutricional de formigas*. In: Panizzi, A.R., Parra, J.R.P. (Eds.), *Ecologia nutricional de insetos*. Editora Manule, Sao Paulo, Brazil, pp. 131–223. <https://doi.org/10.1590/S0102-311X1993000100013>.
- Gibb, H., Parr, C.L., 2013. Does structural complexity determine the morphology of assemblages? An experimental test on three continents. *PLoS One* 8, 1–7. <https://doi.org/10.1371/journal.pone.0064005>.
- Graça, M.B., Pequeno, P.A.C.L., Franklin, E., Souza, J.L.P., Morais, J.W., 2017. Taxonomic, functional, and phylogenetic perspectives on butterfly spatial assembly in northern Amazonia. *Ecol. Entomol.* 42, 816–826. <https://doi.org/10.1111/een.12454>.
- Gronenberg, W., Paul, J., Just, S., Hölldobler, B., 1997. Mandible muscle fibers in ants: fast or powerful? *Cell Tissue Res.* 289, 347–361. <https://doi.org/10.1007/s004410050882>.
- Hölldobler, B., Wilson, E.O., 1990. *The Ants*. Springer, Berlin.
- Hurlbert, A.H., Ford, B., Powell, S., 2008. Shaking a leg and hot to trot: the effects of body size and temperature on running speed in ants. *Ecol. Entomol.* 33, 144–154. <https://doi.org/10.1111/j.1365-2311.2007.00962.x>.
- Instituto Brasileiro de Geografia e Estatística-IBGE, 2012. *Manual Técnico da Vegetação Brasileira/Brazilian Vegetation Classification System 2^o*. Retrieved from. <https://biblioteca.ibge.gov.br/visualizacao/livros/liv63011.pdf>.
- Kaspari, M., 1996. Worker size and seed size selection by harvester ants in a Neotropical forest. *Oecologia* 105, 397–404. <https://doi.org/10.1007/BF00328743>.
- Kaspari, M., Vargo, E.L., 1995. Colony size as a buffer against seasonality: Bergmann's rule in social insects. *Am. Nat.* 145, 610–632. <https://doi.org/10.1086/285758>.
- Kaspari, M., Weiser, M.D., 1999. The size-grain hypothesis and interspecific scaling in ants. *Funct. Ecol.* 13, 530–538. <https://doi.org/10.1046/j.1365-2435.1999.00343.x>.
- Keddy, P.A., 1992. Assembly and response rules: two goals for predictive community ecology. *J. Veg. Sci.* 3, 157–164. <https://doi.org/10.2307/3235676>.
- Kuznetsova, A., Brockhoff, P.B., Christensen, R.H.B., 2017. lmerTest package: tests in linear mixed effects models. *J. Stat. Softw.* 82, 1–26. <http://doi.org/10.18637/jss.v082.i13>.
- Levin, S.A., 1992. The problem of pattern and scale in ecology: the Robert H. MacArthur award lecture. *Ecol.* 73, 1943–1967. <https://doi.org/10.2307/1941447>.
- Magnusson, W.E., Braga-Neto, R., Pezzini, F., Baccaro, F.B., Bergallo, H., Penha, J., Rodrigues, D., Verdade, L.M., Lima, A., Albernaz, A.L., Hero, J.M., Lawson, B., Castilho, C., Drucker, D., Franklin, E., Mendonça, F., Costa, F., Galdino, G., Castley, G., Zuanon, J., do Vale, J., Santos, J.L.C., Luizão, R., Cintra, R., Barbosa, R.L., Lisboa, A., Krolbitz, R.V., Cunha, C.N., Pontes, A.R.M., 2013. Biodiversidade e monitoramento ambiental integrado/Biodiversity and integrated environmental monitoring. Bilingual edition: Portuguese/English. Attema Editorial, Assessoria e Design, Manaus, Brazil. Retrieved from. https://ppbio.inpa.gov.br/sites/default/files/Biodiversidade_e_monitoramento_ambiental_integrado.pdf.
- McGill, B.J., 2010. Matters of scale. *Science* 328, 575–576. <https://doi.org/10.1126/Science.1188528>.
- McGill, B.J., Enquist, B.J., Weiher, E., Westoby, M., 2006. Rebuilding community ecology from functional traits. *Trends Ecol. Evol.* 21, 178–185. <https://doi.org/10.1016/j.tree.2006.02.002>.
- Nakagawa, S., Schielzeth, H., 2013. A general and simple method for obtaining R² from generalized linear mixed-effects models. *Methods Ecol. Evol.* 4, 133–142. <https://doi.org/10.1111/j.2041-210x.2012.00261.x>.
- Oliveira, P.Y., Souza, J.L.P., Baccaro, F.B., 2009. Ant species distribution along a topographic gradient in a "terra-firme" forest reserve in Central Amazonia. *Pesq. agropec. bras.* 44, 852–860. <https://doi.org/10.1590/S0100-204X2009000800008>.
- Parr, Z.J.E., Parr, C.L., Chown, S.L., 2003. The size-grain hypothesis: a phylogenetic and field test. *Ecol. Entomol.* 28, 475–481. <https://doi.org/10.1046/j.1365-2311.2003.00529.x>.
- Pereboom, J.J.M., Biesmeijer, J.C., 2003. Thermal constraints for stingless bee foragers: the importance of body size and coloration. *Oecologia* 137, 42–50. <https://doi.org/10.1007/s00442-003-1324-2>.
- Peters, M.K., Peisker, J., Steffan-Dewenter, I., Hoiss, B., 2016. Morphological traits are linked to the cold performance and distribution of bees along elevational gradients. *J. Biogeogr.* 43, 2040–2049. <https://doi.org/10.1111/jbi.12768>.
- Saatchi, S.S., Houghton, R.A., Alvala, R.C.S., Soares, J.V., Yu, Y., 2007. Distribution of aboveground live biomass in the Amazon basin. *Glob. Chang. Biol.* 13, 816–837. <https://doi.org/10.1111/j.1365-2486.2007.01323.x>.
- Schofield, S.F., Bishop, T.R., Parr, C.L., 2016. Morphological characteristics of ant assemblages (Hymenoptera: formicidae) differ among contrasting biomes. *Myrmecol. News* 23, 129–137. <https://doi.org/10.1111/j.1439-0388.2011.00951.x>.
- Silva, R.R., Brandão, C.R.F., 2010. Morphological patterns and community organization in leaf-litter ant assemblages. *Ecol. Monogr.* 80, 107–124. <https://doi.org/10.1890/08-1298.1>.
- Sobral, F.L., Cianciaruso, M.V., 2012. Phylogenetic and functional assembly structure: (re)assembling the community ecology on different spatial scales. *J. Biosci.* 28, 617–631. <https://doi.org/10.3828/tp.2012.6>.
- Sommer, B., Harrison, P.L., Beger, M., Pandolfi, J.M., 2014. Trait-mediated environmental filtering drives assembly at biogeographic transition zones. *Ecology* 95, 1000–1009. <https://doi.org/10.1890/13-1445.1>.
- Souza, J.L.P., Baccaro, F.B., Landeiro, V.L., Franklin, E., Magnusson, W.E., 2012. Trade-offs between complementarity and redundancy in the use of different sampling techniques for ground-dwelling ant assemblages. *Appl. Soil Ecol.* 56, 63–73. <https://doi.org/10.1016/j.apsoil.2012.01.004>.
- Souza, J.L.P., Baccaro, F.B., Landeiro, V.L., Franklin, E., Magnusson, W.E., Pequeno, P.A.C.L., Fernandes, I.O., 2016. Taxonomic sufficiency and indicator taxa reduce sampling costs and increase monitoring effectiveness for ants. *Divers. Distrib.* 22, 111–122. <https://doi.org/10.1111/ddi.12371>.
- ter Steege, H., Pitman, N.C., Sabatier, D., Baraloto, C., Salomão, R.P., Guevara, J.E., Phillips, O.L., Castilho, C.V., Magnusson, W.E., Molino, J.F., Monteagudo, A., Núñez Vargas, P., Montero, J.C., Feldpausch, T.R., Coronado, E.N., Killeen, T.J., Mostacedo, B., Vasquez, R., Assis, R.L., Terborgh, J., Wittmann, F., Andrade, A., Laurance, W.F., Laurance, S.G., Marimon, B.S., Marimon, B.H., Guimarães Jr., V., Vieira, I.C., Amaral, L.L., Brienen, R., Castellanos, H., Cárdenas López, D., Duivenvoorden, J.F., Mogollón, H.F., Matos, F.D., Dávila, N., García-Villacorta, R., Stevenson Diaz, P.R., Costa, F., Emilio, T., Levis, C., Schietti, J., Souza, P., Alonso, A., Dallmeier, F., Montoya, A.J., Fernandez Piedade, M.T., Araujo-Murakami, A., Arroyo, L., Gribel, R., Fine, P.V., Peres, C.A., Toledo, M., Aymard, C.G.A., Baker, T.R., Cerón, C., Engel, J., Henkel, T.W., Maas, P., Petronelli, P., Stropp, J., Zartman, C.E., Daly, D., Neill, D., Silveira, M., Paredes, M.R., Chave, J., Filho, A.L., Jorgensen, P.M., Fuentes, A., Schongart, J., Cornejo Valverde, F., Di Fiore, A., Jimenez, E.M., Peñuela Mora, M.C., Phillips, J.F.,

- Rivas, G., van Andel, T.R., von Hildebrand, P., Hoffman, B., Zent, E.L., Malhi, Y., Prieto, A., Rudas, A., Ruschell, A.R., Silva, N., Vos, V., Zent, S., Oliveira, A.A., Schutz, A.C., Gonzales, T., Nascimento, M.T., Ramirez-Angulo, H., Sierra, R., Tirado, M., Umaña Medina, M.N., van der Heijden, G., Vela, C.I., Vilanova Torre, E., Vriesendorp, C., Wang, O., Young, K.R., Baidar, C., Balslev, H., Ferreira, C., Mesones, I., Torres-Lezama, A., Urrego Giraldo, L.E., Zagt, R., Alexiades, M.N., Hernandez, L., Huamantupa-Chuquimaco, I., Milliken, W., Palacios Cuenca, W., Pauletto, D., Valderrama Sandoval, E., Valenzuela Gamarra, L., Dexter, K.G., Feeley, K., Lopez-Gonzalez, G., Silman, M.R., 2013. Hyperdominance in the Amazonian tree flora. *Science* 342. <https://doi.org/10.1126/science.1243092>.
- Vellend, M., 2016. *The Theory of Ecological Communities*. Princeton University Press, New Jersey.
- Villela, D., Proctor, J., 1999. Mass litterfall, chemistry, and nutrient retranslocation in a Monodominant Forest on Maraca Island, Roraima, Brazil. *Biotropica* 31, 198–211. Retrieved from: <http://www.jstor.org/stable/2663783>.
- Weiher, E., Freund, D., Bunton, T., Stefanski, A., Lee, T., Bentivenga, S., 2011. Advances, challenges and a developing synthesis of ecological community assembly theory. *Philos. Trans. R. Soc. B Biol. Sci.* 366, 2403–2413. <https://doi.org/10.1098/rstb.2011.0056>.
- Weiser, M.D., Kaspari, M., 2006. Ecological morphospace of new world ants. *Ecol. Entomol.* 31, 131–142. <https://doi.org/10.1111/j.0307-6946.2006.00759.x>.
- Yates, M.L., Andrew, N.R., Binns, M., Gibb, H., 2014. Morphological traits: predictable responses to macrohabitats across a 300 km scale. *Peer J.* 2, e271. <https://doi.org/10.7717/peerj.271>.