



Vaccinium vitis-idaea L.: Chemical Contents, Pharmacological Activities

Arnold Alexeevich Shamilov^{1*}, Valentina Nikolaevna Bubenchikova², Maxim Valentinovich Chernikov³, Dmitry Igorevich Pozdnyakov⁴, Ekaterina Robertovna Garsiya¹

¹Department of Pharmacognosy, Botany and Technology of Phytopreparations, Pyatigorsk Medical-Pharmaceutical Institute (PMPI), Branch of Volgograd State Medical University, Ministry of Health of Russia, 11, Kalinina Prospect, Pyatigorsk, Russian Federation, 357532.

²Department of Pharmacognosy and Botany, Kursk State Medical University (KSMU), Ministry of Health of Russia, 3, K. Marx Street, Kursk, Russian Federation, 305041.

³Department of Biology and Physiology, Pyatigorsk Medical-Pharmaceutical Institute (PMPI), Branch of Volgograd State Medical University, Ministry of Health of Russia, 11, Kalinina prospect, Pyatigorsk, Russian Federation, 357532.

⁴Department of Pharmacology with course of clinical Pharmacology, Pyatigorsk Medical-Pharmaceutical Institute (PMPI), Branch of Volgograd State Medical University, Ministry of Health of Russia, 11, Kalinina prospect, Pyatigorsk, Russian Federation, 357532.

Article Info

Article History:

Received: 15 February 2020

Accepted: 9 July 2020

ePublished: 25 December 2020

Keywords:

-Phytochemistry

-Phytotherapy

-Vaccinium

-Vaccinium vitis-idaea L

Abstract

One of the most known species of the genus *Vaccinium* (Ericaceae) is *Vaccinium vitis-idaea* L. or lingonberry. Leaves are included in the State Pharmacopoeia of the Russian Federation (XIV-th edition) and the State Pharmacopoeia of the Republic of Belarus (II-nd edition). The aim of this review is an analysis of data about a chemical content and types of pharmacological activities of *Vaccinium vitis-idaea* L. to discuss the tendency of future investigations on this plant. The main parts of works describe researches of chemical contents of fruits as medicinal and edible plant material. The majority of researches describe results of *in vitro* experiments. A significant interest is the study of the neuroprotective activity of the *Vaccinium vitis-idaea* extracts as well as their anti-cytokine and antiapoptotic properties and metabolic effects. The main biologically active compounds are phenolglycosides, tannins, proanthocyanes, saponins etc. These results will be of great significance for the development of new drugs from this plant and use along with the fruits of other parts of the plant.

Introduction

Some well-known medicinal plants can be interesting for a detailed investigation. These plants are included in the State Pharmacopoeias of different countries and have a broad medical application, but a study of qualitative and quantitative chemical compositions of the groups of substances other than those responsible for the main pharmacological effect, as well as the study of new and not well-known therapeutic effects, is promising and relevant. Many species of genus *Vaccinium* L. are known as medicinal and edible plants.

The genus *Vaccinium* comprises more than 100 species, 11 of them grow among the flora of Russia.¹ In a lot of countries, a pharmacopoeia species is *Vaccinium vitis-idaea* L. According to the data of The Plant List (2019), the synonyms of *Vaccinium vitis-idaea* L. are: *Myrtilus exigua* Bubani., *Rhodococcum vitis-idaea* Avrorin., *Vaccinium jesoense* Miq., *Vitis-idaea punctata* Moench., *Vitis-idaea punctifolia* Gray.² In different countries, this plant is also named as lingonberry (English), preiselbeere (German), airelles (French).

According to the taxonomy and phylogeny of flowering

plants, worked out by A. L. Takhtajan, the genus *Vaccinium* has the following systematic positions:³ phylum - Magnoliophyta, class - Magnoliopsida, subclass - Dilleniidae, superorder - Dilleniales, order - Ericales; family - Ericaceae, subfamily - Vaccinioideae (tribe: Oxydendreae, Vaccinieae, Gaultherieae), tribe Vaccinieae [genus: *Gaylussacia*, *Vaccinium* (including *Oxycoccus*), *Symphysia*, *Thibaudia*, *Satyria*, *Agapetes* (including *Pentapterigium*), *Didonica*, *Lateropora*, *Notopora*, *Utleya*, *Gonocalyx*, *Dimorphanthera*, *Paphia*, *Cavendishia*, *Orthaea*, *Macleania*, *Psammisia*, *Mycerinus*, *Polyclita*, *Anthopteropsis*, *Ceratostema*, *Semiramisia*, *Oreanthes*, *Siphonandra*, *Pellegrinia*, *Disterigma*, *Sphyrospermum*, *Diogenesia*, *Rusbya*, *Themistoclesia*, *Plutarchia*, *Demostenesia*, *Anthopterus*, *Costera*].³

The latest version of the Angiosperm Phylogeny Group (APG) is APG IV (2016). APG uses the clades which include all taxa in the hierarchic sequence and they have only one match. The family Ericaceae matches the clade Asterids, the order Ericales.⁴ *Vaccinium vitis-idaea* L. matches the clade *Vaccinium*.⁵

*Corresponding Author: Arnold Alexeevich Shamilov, E-mail: shamilovxii@yandex.ru

©2020 The Author(s). This is an open access article and applies the Creative Commons Attribution License (<http://creativecommons.org/licenses/by-nc/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, as long as the original authors and source are cited.

According to S. K.Cherepanov's data^{1,11} species occur in the Russian flora, among them 4 species grow in the Caucasus, 4 – in the European part of Russia, 4 – in West Siberia, 10 – in the Russian Far East.

V. vitis-idaea L. is an evergreen subshrub with a height of 2.5-25 cm. It has round whitish hairy stems and coriaceous, obovate to elliptic leaves. Their tops are flat or daedalous, finely toothed or smooth-margin. The leaves are induplicate, 5-27 mm long and 3-12 mm wide, on a short pubescent petiole 0.5-3 mm long. The leaf upper surface is dark green; the lower surface is pale green, waxy with deep-brown scattered glandular dots. Flowers have short reddish pubescent peduncles; at the ends of last-year's branches there are inflorescences in racemes, in short, but thick drooping clusters of 2-8 with a faint but pleasant smell. Calyxes are quadridentate with four short rounded reddish notches, 0.75-1.25 mm long and 0.75-1 mm wide. Corolla is bell-shaped, pale pink, 4-6.5 mm long. It has four petals. There are 8 stamens with pilose anther filaments. Fruits are globose berries carmine in color when ripe. Fruits are edible. *V. vitis-idaea* L.'s main bloom time is early summer and late spring (June-July). Its ripening time is September.⁶

Plants with larger leaves (10-30 mm long) occur in Europe and Asia. This variety is named *V. vitis-idaea* var. *vitis-idaea* L., *V. vitis-idaea* subsp. *vitis-idaea* or cowberry. In the North America, lingonberry or *V. vitis-idaea* var. *minus* Lodd. (*V. vitis-idaea* subsp. *minus* (Lodd.) Hultén), has leaves 5–18 mm long.⁷

Use in Traditional Medicine

Fruits of lingonberry are used as medicinal and edible plant materials. Fruits are used in jams, juices, vines, pies.⁸ Other plant parts are also used in medicine.

Vaccinium vitis-idaea L. was included in the first Russian Pharmacopoeia.⁹ Leaves are included in the Russian State Pharmacopoeia (XIVth edition). At the pharmaceutical marker there is "Brunsiver", a polyherbal drug. It includes *Vaccinium vitis-idaea* L. leaves (50%), herbs of *Hypericum perforatum* L. and *Bidens tripartita* L. (20% and 10%, respectively), fruits of *Rosa* spp. L. (10%). The data on clinical trials of this complex drug are insufficient. Usually, it is used as infusion for treatment of diseases of the genitourinary system, cystitis, genitourinary infection, urethritis.¹⁰

In the Encyclopedia of Traditional Chinese Medicine, *V. vitis-idaea* is represented as a source of 6-O-acetyl arbutin, arbutin (its content in leaves is about 4%), avicularin, 2-O-caffeoyl arbutin, cyanidin-3-O-β-D-galactoside, (-)-epicatechin, hydroquinone, 4-hydroxyphenyl-β-gentiobioside, proanthocyanidin A2, procyanidin B1, B3, B7, rhodioloside. To reduce strangury and urinary disorders, leaves of the plant are used as an antitoxic agent.¹¹ Leaves have astringent, antiseptic, diuretic, anti-fever, anti-inflammatory effects; fruits and leaves can show a metabolite activity, have anti-fever, reparative, analgesic, sedative, detoxifying effects. Shoots are known as a

homeopathic remedy.¹² Fruits are used to treat diabetes, gastric and intestinal ulcers, liver diseases, periodontitis, hypertension, eczema, dermatophytosis, diathesis, respiratory infections, gastric and skin cancers, eye diseases, rheumatism, neurasthenia.^{12,13}

Chemical Composition

In different aerial parts of *Vaccinium vitis-idaea* L., phenologlycosides, phenylpropanoids, anthocyanes, saponins have been found out by researches from different countries; and in fruits, the content of anthocyanes has been established.

Phenols, phenologlycosides, coumarins

Munich scientists Friedrich and Schoenert¹⁴ collected *Vaccinium vitis-idaea* L. fruits and leaves from April to August. Aqueous and methanol extracts were prepared from fruits and leaves and then analyzed by thin-layer chromatography (TLC) and column chromatography (CC). It turned out impossible to isolate hydroquinone-β-D-glucopyranoside (arbutin) and hydroquinone using either polyamide or silica gel. The presence of these compounds was established by TLC after post processing of the chromatogram with diazotized sulfanilic acid and saturated sodium carbonate solution. The investigation resulted in the conclusion that *Vaccinium vitis-idaea* L. leaves contain arbutin and hydroquinone. The optimal period of harvesting is August. The authors also claim that *Vaccinium vitis-idaea* L. can hybridize with other species of the *Vaccinium* genus and form intermediate hybrid forms. In 1979 Swiss scientists Sticher *et al.*¹⁵ developed methods of phenolic analysis using high-performance liquid chromatography (HPLC). In *Vaccinium vitis-idaea* L. leaves they detected 3-5% of arbutin and 0.14-0.20% of hydroquinone. It was established that if the amount of hydroquinone increased, the amount of arbutin decreased. Neither methylarbutin nor hydroquinone monomethyl were detected in the leaves.

In 2005-2006 Pyka *et al.*¹⁶ collected *Vaccinium vitis-idaea* L. leaves which are a pharmacopoeia plant material in Poland. The leaves were air-dried. A spectrodensitometric method for the determination of arbutin was worked out by the scientists. The aqueous-methanol extract (1:1, v/v) of leaves was applied onto the chromatographic plate and detected at $\lambda_{\max} = 285$ nm. The amount of arbutin in 1 g of the raw material (air-dried leaves) was 35 mg/g (2005) and 47 mg/g (2006). This method was proposed as a pharmacopoeia method (the Polish Pharmacopoeia, IV-th edition).

Wang *et al.*¹⁷, scientists of *Heilongjiang University* (2005), obtained an aqueous-alcohol extract from *V. vitis-idaea* leaves. Its analysis resulted in the isolation of arbutin and coumarin fraxetin-8-O-glucoside (fraxin).

Liu *et al.*¹⁸ (2014) collected *V. vitis-idaea* leaves in the flora areas of Turkey and Finland from the end of June to the middle of September. The leaves were collected every 10-th day and subjected to freezing with liquid

nitrogen. The acetone, chloroform, aqueous-alcohol extracts from the leaves were subjected to the analytic investigation. In the obtained fractions, the entire phenolic complex, including phenolic glycosides, was analyzed. The use of HPLC resulted in the detection of arbutin and 2-O-caffeoylarbutin. The lowest amount of arbutin was detected on 30 June, the highest – on 18 July; the lowest amount of 2-O-caffeoylarbutin was registered on 29 August (9 mg/g of dry mass), the highest – on 18 July (14 mg/g of dry mass).

Raudone *et al.*¹⁹ (2019) published the results of the investigated phenolic profile in the several cultivars of *V. vitis-idaea* L. and the antioxidant activity of cultivars leaves. In their opinion, in the accumulation of phenolic compounds, an important role is played by such external factors as light, sun rays, temperature, type of soil, methods of cultivation, insects; and the internal factor - the genetic background. These factors and, to a greater extent, the genetic diversity, cause variability even within the same species. That was why in this work 10 different cultivars were analyzed. They were: «Sussi», «Erntekrone», «Masovia», «Koralle», «Kostromskaja rozovaja», «Sanna», «Rubin», «Kostromicka», «Erntesegen», «Erntedank»; 1 subspecies (*V. vitis-idaea* subsp. *minus*) and 1 *Vaccinium vitis-idaea* L. variety (*V. vitis-idaea* var. *leucocarpum*). Qualitative and quantitative analyses of phenolic compounds in *Vaccinium vitis-idaea* L. leaves, were carried out by HPLC Photo-Diode Array (HPLC-PDA). The analyses have resulted in the conclusion that in *Vaccinium vitis-idaea* L. leaves the main compounds are simple phenols, flavonoids, proanthocyanidines and hydroxycinnamic acids. The highest amount of arbutin (mg/g dry weight or mg/g DW) was established in the following cultivars: «Kostromskaja rozovaja» (56.968±2325.73 mg/g DW),

«Rubin» (36.059±1472.12 mg/g DW) and *V. vitis-idaea* subsp. *minus* (28.708±1172.03 mg/g DW). Arbutin was the main compound in all *Vaccinium vitis-idaea* cultivars and taxa. Its quantity was about 41-78% of the total amount of phenolic compounds (Figure 1).

Flavonoids

Several flavonoids: quercetin, quercitrin, isoquercitrin and quercetin-3-arabinoside, were isolated from *Vaccinium vitis-idaea* L. leaves by Friedrich and Schönerert.¹⁴

Ek *et al.*²⁰ published data about the chemical composition of *Vaccinium vitis-idaea* L. fruits, leaves and shoots collected in the west of Finland in summer and autumn of 2004. Ethanol, acetonitrile and methanol extracts were obtained by the authors. The methanol extract contained the highest amount of phenolic compounds from *Vaccinium vitis-idaea* L. fruits, leaves and shoots. The ultrasound homogenisator was used for the best yield of phenolic compounds. After that the methanol extract was subjected to centrifugation. Before the analysis by HPLC, the obtained extracts were diluted in water in the ratio of 1:2.

According to the mass-spectrometry (MS) data and the subsequent proton nuclear magnetic resonance (proton NMR) detection of the substances, 28 compounds of the phenolic nature were detected. Ten of them referred to flavonoids: [hyperoside (quercetin-3-O-β-galactoside), isoquercitrin (quercetin-3-O-β-glucoside), quercetin-O-(hexose-deoxyhexoside), reynoutrin (quercetin-3-O-β-xyloside), quercetin-3-O-α-arabinopyranoside, avicularin (quercetin-3-O-α-arabinofuranoside), quercitrin (quercetin-3-O-α-rhamnoside), quercetin-3-O-(4''-3-hydroxy-3-methylglutaryl)-α-rhamnoside, kaempferol-pentoside, kaempferol-deoxyhexoside, kaempferol-(3-hydroxy-3-methylglutaryl)]. The above-listed compounds

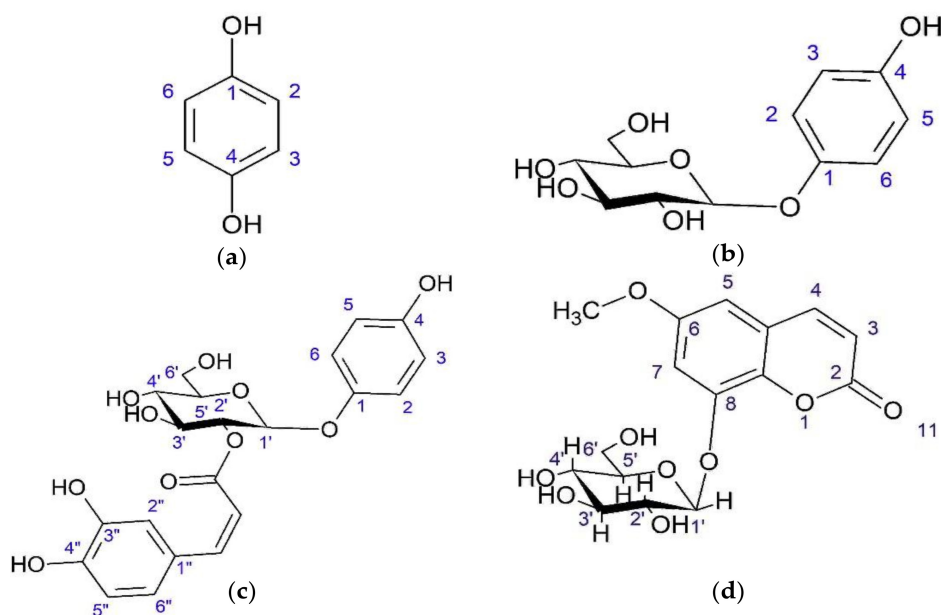


Figure 1. Phenols, phenologlycosides, coumarins from *Vaccinium vitis-idaea* L.: (a) hydroquinone; (b) arbutin; (c) 2-o-caffeoylarbutin; (d) fraxin.

were also detected in the fruits and leaves with stems.

Liu *et al.*¹⁸ identified 12 compounds referring to flavonoids, by methods of HPLC-diode array (HPLC-DAD), HPLC-MS, gas chromatography-flame ionization detector (GC-FID), proton NMR. Ten of them were previously discovered and three were detected in this plant material for the first time: rutin (quercetin-3-O-rutinoside), quercetin-3-O-glucuronide, kaempferol-3-O-glucuronide. The lowest total amount of flavonoids was detected on 29 August, the highest – on 12 September. In content, the main compounds in *Vaccinium vitis-idaea* L. leaves were the following: hyperoside, quercetin-3-O-arabinoside, quercetin-3-O-rhamnoside. The total amount of flavonoids was 15-20 mg/g.

Lätti *et al.*²¹ identified flavonoids in *Vaccinium vitis-idaea* L. leaves. They detected the above listed compounds and also discovered 3,5,7-trihydroxy-2-(4-hydroxy-3,5-methoxyphenol)-4H-chromen-4-one (syringetin hexoside). The authors claim that when grown together, *Vaccinium myrtillus* and *Vaccinium vitis-idaea* can form a natural species hybrid *Vaccinium x intermedium* Ruthe.

Riihinen *et al.*²² extracted juice from *Vaccinium vitis-idaea* L. fruits collected in Finland. After the isolation of flavonoids by CC with the help of Sephadex LH-20 gel, methods of HPLC-PDA and NMR were used. Seven flavonoids such as quercetin-3-O-[4''-(3-hydroxy-3-methylglutaroil)]- α -rhamnoside, quercetin-3-O- α -rhamnoside, quercetin-3-O- β -glucoside, quercetin-3-O- β -galactoside, quercetin-3-O- α -arabinofuranoside, quercetin-3-O- α -xylopyranoside, quercetin-3-O- α -arabinopyranoside, were identified in *Vaccinium vitis-idaea* L. juice. Later the authors identified quercetin-3-O- β -galactopyranoside.²³

Drózdź *et al.*²⁴ extracted two substances from *V. vitis-idaea* fruits using ethanol (deionization water and ethanol 60:40, v/v). In the aqueous extract, the total amount of flavonoids was higher (2.55-3.53 μ mole catechin equivalent to (CE)/g fresh weight of fruit) than anthocyanes (0.34-0.41 mg of cyanidin-3-glucoside equivalent to (C3G)/g fresh weight of fruit). The yield of flavonoids was twice higher when the aqueous-ethanol solvent was used.

Later the authors obtained ethanol (ethanol-water 60:40 v/v) and ethyl acetate extracts from *V. vitis-idaea* fruits cultivated in Poland.²⁵ The total amount of flavonoids was 53.2-67.8 μ mole CE/100 g from the fruits using the ethanol-water mixture and 40.7-62.9 μ mole CE/100 g using the ethyl acetate solvent.

In *V. vitis-idaea* L. fruits,²⁶ the portion of flavonoids is 7% of all phenols.

Raudone *et al.*¹⁹ established that in the above listed *Vaccinium vitis-idaea* L. cultivars and taxa, the principle components of the flavonoids are hyperoside, avicularin, quercetin and, in smaller amounts, astragalín. The highest amount of hyperoside (mg/g dry weight or mg/g DW) was registered in *V. vitis-idaea* subsp. *minus* (5771.34 \pm 235.61 mg/g of DW) and *V. vitis-idaea* var. *leucocarpum* (5503.44 \pm 224.6 mg/g DW); avicularin in *V. vitis-idaea* var. *leucocarpum* (5503.44 \pm 224.68 mg/g DW; quercetin

in "Koralle" cultivar (7826.68 \pm 319.52 mg/g DW); astragalín in "Rubin" cultivar (120.60 \pm 4.92 mg/g DW) and "Kostromskaja rozovaja" (114.67 \pm 4.68 mg/g DW).

The data about flavonoids from *V. vitis-idaea* are presented in Table 1.

Proanthocyanes

Ho *et al.*²⁷ isolated such tannins as procyanidin B1, procyanidin B3, procyanidin A1, cinnamtannin B1, epicatechin-(4 β \rightarrow 8)-epicatechin-(4 β \rightarrow 8, 2 β \rightarrow O \rightarrow 7)-catechin, epicatechin-(4 β \rightarrow 6)-epicatechin-(4 β \rightarrow 8, 2 β \rightarrow O \rightarrow 7)-catechin from *Vaccinium vitis-idaea* L. leaves and fruits by Morimoto's method (1988). In *Vaccinium vitis-idaea* L. fruits, the amount of proanthocyanines was 71% of the total amount of phenols; the major compound was trimer A-type.²² Later Riihinen *et al.*^{28,29} identified (+)-catechin and (-)-epicatechin in the *Vaccinium vitis-idaea* L. fruit juice (Figure 2).

Raudone *et al.*¹⁹ detected qualitative and quantitative contents of (+)-catechin, (-)-epicatechin, procyanidin C1, procyanidin A2 in *Vaccinium vitis-idaea* L. cultivars and taxa listed above. In "Masovia" cultivar, the amount of (+)-catechin was 2954.64 \pm 120.62 mg/g (DW) and in "Kostromskaja rozovaja" it was 2273.92 \pm 92.83 mg/g (DW); (-)-epicatechin was detected in "Kostromskaja rozovaja" in the amount of 743.67 \pm 30.36 mg/g (DW) and in "Rubin" - 669.11 \pm 27.32 mg/g (DW); in *V. vitis-idaea* var. *leucocarpum* procyanidin C1 was in the amount of 4645.97 \pm 189.67 mg/g; in *V. vitis-idaea* subsp. *minus* procyanidin A2 was in the amount of 7074.60 \pm 288.82 mg/g; in "Erntekrone" - 3609.57 \pm 147.36 mg/g (DW). and in "Masovia" - 3554.68 \pm 145.12 mg/g (DW).

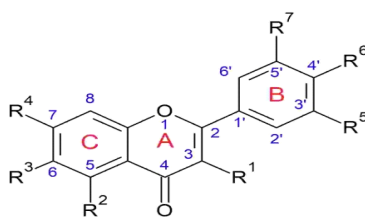
Anthocyanes

Viljanen *et al.*³⁰ detected such anthocyanes as cyaniding-3-glucoside, cyaniding-3-galactoside, cyaniding-3-arabinoside in *Vaccinium vitis-idaea* L. fruits by HPLC. Ek *et al.*²⁰ detected proanthocyanidines and cyanidines in *Vaccinium vitis-idaea* L. fruits and leaves. Anthocyanes were not detected in the leaves with shoots.

Later Dudonne *et al.*³¹ confirmed these data and also added new information on the quantitatively content of each anthocyanes in their total amount. Cyanidin-3-galactoside was represented by 58%, cyaniding-3-arabinoside – by 35% and cyaniding-3-glucoside – by 7% from the total amount of anthocyanes. According to the data in one of the works,²⁶ paeonidin-3-glucoside was also detected in the amount 4.2%. Anthocyanes are presented by 15% from the total amount of phenols.²⁶

The United States Department of Agriculture (USDA), Agricultural Research Service (ARS), Horticultural Crops Research Unit (HCRU) recommended to standardize *Vaccinium vitis-idaea* L. fruits by the amounts of the three anthocyanes described above because there are already several cases of adulteration of these valuable plant materials.³²

Brown *et al.*³³ used 1% formic acid in methanol to extract

Table 1. Flavonoids from *Vaccinium vitis-idaea* L.

No	Name	Type of flavonoids	R ¹	R ²	R ³	R ⁴	R ⁵	R ⁶	R ⁷	Glycoside
1	Quercetin	Flavonol quercetin	OH	OH	OH	OH	OH	OH		
2	Quercitrin		3-O- α -L-rhamnoside	OH	OH	OH	OH	OH		O-glycoside
3	Isoquercitrin		3-O- β -D-glucopyranoside	OH	OH	OH	OH	OH		O-glycoside
4	Guaijaverin, quercetin-3-O- α -L-arabinoside		3-O- α -L-arabinopyranoside	OH	OH	OH	OH	OH		O-glycoside
5	Hyperoside (quercetin-3-O-galactoside)		3-O- β -D-galactoside	OH	OH	OH	OH	OH		O-glycoside
6	Reynoutrin (quercetin-3-O- β -D-xyloside)		3-O- β -D-xyloside	OH	OH	OH	OH	OH		O-glycoside
7	Avicularin (quercetin-3-O- α -L-arabinofuranoside)		3-O- α -L-arabinofuranoside	OH	OH	OH	OH	OH		O-glycoside
8	Quercetin-3-O-[4''-(3-hydroxy-3-methylglutaroyl)]-R-rhamnose		3-O-[4''-(3-hydroxy-3-methylglutaroyl)]-R-rhamnose	OH	OH	OH	OH	OH		O-glycoside
9	Rutin (quercetin-3-rutinoside)		3-O- β -D-rutinoside	OH	OH	OH	OH	OH		O-glycoside
10	Quercituron (quercetin 3-O- β -D-glucuronopyranoside)		3-O- β -D-glucuronopyranoside	OH	OH	OH	OH	OH		O-glycoside
11	Isorhamnetin-3-O-glucoside	Flavonol 3-methylquercetin	3-O- β -D-glucopyranoside	OH	OH	OCH ₃	OH	OH		O-glycoside
12	Kaempferol-pentoside	Flavonol kaempferol	3-O-pentoside	OH	OH	OH	OH	OH		O-glycoside
13	Kaempferol-deoxyhexoside		3-O-hexoside	OH	OH	OH	OH	OH		O-glycoside
14	Kaempferol-(3-hydroxy-3-methylglutaroyl) rhamnoside		(3-hydroxy-3-methylglutaroyl)rhamnoside	OH	OH	OH	OH	OH		O-glycoside
15	Nicotiflorin (kaempferol-3-O- β -rutinoside)		3-O- β -rutinoside	OH	OH	OH	OH	OH		O-glycoside
16	Astragaln (kaempferol-3-O- β -D-glucopyranoside)		3-O- β -D-glucopyranoside	OH	OH	OH	OH	OH		O-glycoside
17	Kaempferol-3-O-glucuronide		3-O- β -D-glucuronopyranoside	OH	OH	OH	OH	OH		O-glucoside
18	Syringetin hexoside (3',5'-O-dimethylmyricetin-hexoside)		Flavonol myricetin	Hexoside	OH	OH	OCH ₃	OH	OCH ₃	

anthocyanes from *Vaccinium vitis-idaea* L. fruits, then the extract was centrifuged. The residue was separated, subjected to the process of extraction again by the same solvent, and then centrifuged. All the supernatants were united and evaporated to dryness under vacuum. Then the

residue was analyzed by HPLC-MS/MS. The anthocyanes were detected at the wavelength of 520 nm.

The study resulted in the identification of cyaniding-3-O-galactoside, cyaniding-3-O-glucoside, cyaniding-3-O-arabinoside. Other anthocyanes were found out

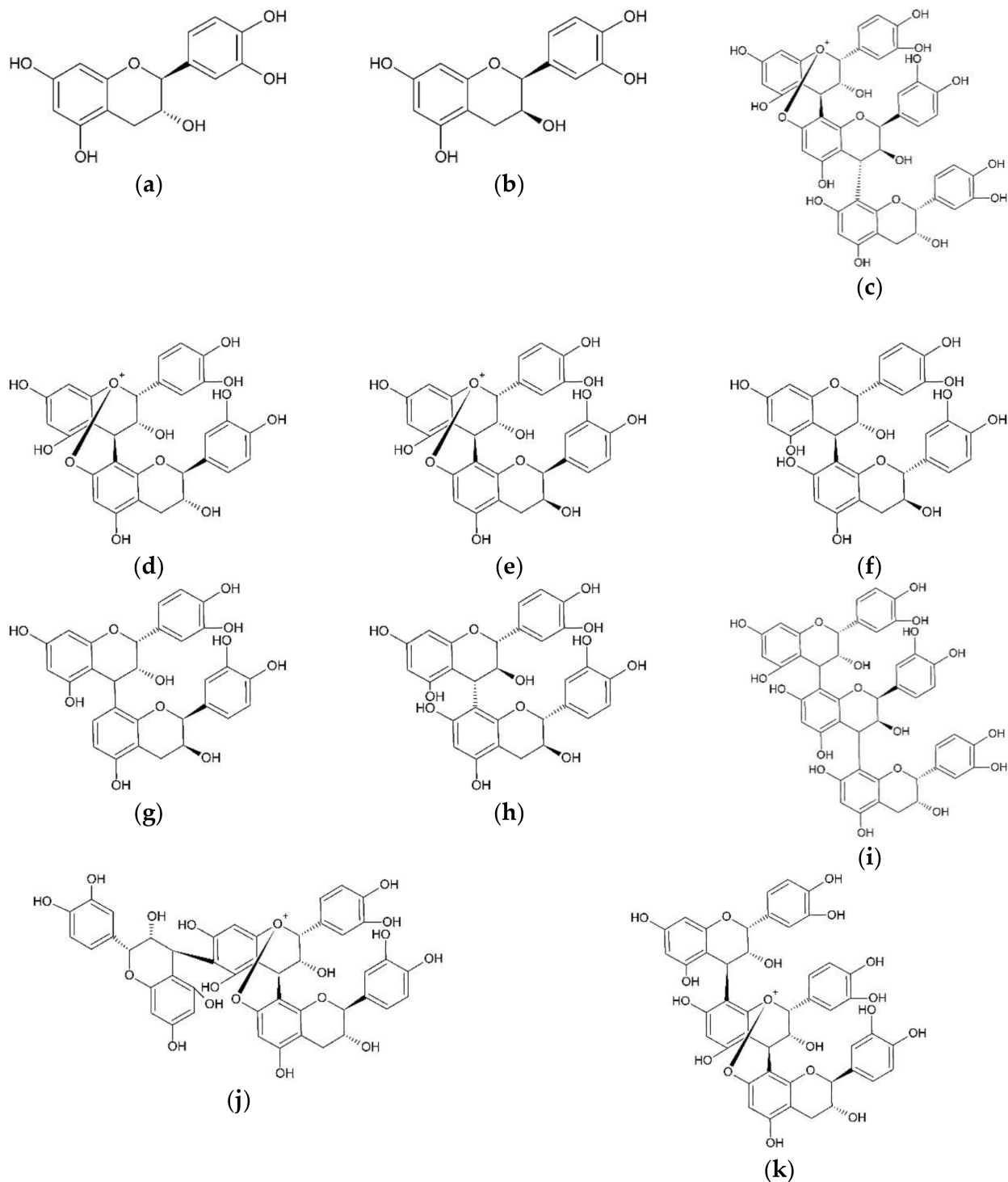


Figure 2. Proanthocyanines from *Vaccinium vitis-idaea* L.: (a) (+)-catechin; (b) (-)-epicatechin; (c) cinnamtannin B1; (d) proanthocyanidin A1; (e) proanthocyanidin A2; (f) proanthocyanidin B1; (g) proanthocyanidin B2; (h) proanthocyanidin B3; (i) proanthocyanidin C1; (j) epicatechin-(4 β →6)-epicatechin-(4 β →8, 2 β →O→7)-catechin; (k) epicatechin-(4 β →8)-epicatechin-(4 β →8, 2 β →O→7)-catechin.

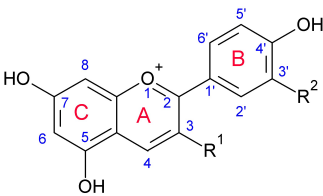
in low concentrations: cyaniding-3-O-sambubioside, pelargonidin-3-O-glucoside, paeonidin-3-O-galactoside, paeonidin-3-O-glucoside, cyanidin-3-O-xyloside, cyanidin-3-O-(6''-O-acetyl)glucoside (Table 2).

Hydroxycinnamic acids

Ek *et al.*²⁰ isolated 4 hydroxycinnamic acids (ferulic acid

[3-methoxy-4-hydroxycinnamic acid], caffeoyl-hexose-hydroxyphenol, coumaroyl-hexose-hydroxyphenol, 2''-caffeoylarbutin] from *V. vitis-idaea* fruits, leaves and shoots. The last compound was found only in the fruits.

Hokkanen *et al.*³⁴ investigated chemical contents of *V. vitis-idaea* and the intermediate hybrid form *Vaccinium x intermedium* Ruthe L. (hybrid bilberry). The leaves with

Table 2. Anthocyanes from *Vaccinium vitis-idaea* L.


No	Name	R ¹	R ²
	Cyanidin-3-O-β-D-glucoside	3-O-β-D-glucopyranoside	OH
	Cyanidin-3-O-β-D-galactoside	3-O-β-D-galactoside	OH
	Cyanidin-3-O-β-D-arabinoside	3-O-β-D-arabinoside	OH
	Peonidin-3-O-β-D-glucoside	3-O-β-D-glucopyranoside	OCH ₃
	Cyanidin-3-O-sambubioside (Cyanidin 3-xyloglucoside)	3-O-β-D-xylopyranosyl-(1-2)-β-D-glucopyranoside	OH
	Pelargonidin 3-glucoside (callistephin)	3-O-β-D-glucopyranoside	H
	Peonidin 3-O-β-D-galactoside	3-O-β-D-galactoside	OCH ₃
	Cyanidin 3-O-xyloside	3-O-β-xylofuranoside	OH
	Cyanidin-3-O-(6''-acetyl)glucoside)	3-O-(6-O-acetyl-β-D-glucopyranoside)	OH

stems were collected in summer and autumn of 2005-2006 in the flora area of southern Finland. The plant materials were extracted by methanol. The analysis was carried out by ultra-performance liquid chromatography coupled with time-of-flight mass spectrometry (UPLC-TOF-MS).

In total, 51 phenolic compounds were identified. For the first time ever, caffeoyl-shikimic acid and caffeic acid, trans-chlorogenic acid, cis-chlorogenic acid, p-cumaric acid and iridoid-coumaroyl iridoid (vaccinoside) were detected. In *V. vitis-idaea* L. leaves with shoots, the contents of hydroxycinnamic acids was 24.4% from the total amount of phenolic compounds; the total amount of iridoids was 7.5%.

Ieri *et al.*³⁵ collected a few species of the genus *Vaccinium* (Ericaceae) including *V. vitis-idaea* in the period from 2009 to 2011. The plants were collected in the flora area of Italy, the plant materials were young and mature leaves of *V. vitis-idaea* L. They were used for the preparation of aqueous-glycerol-alcohol extracts which were analyzed then by HPLC-DAD. In these extracts the following phenolic compounds were identified: flavonoids, phenylpropanoids, phenologlycosides, coumarines. Several new flavonoids were also identified.

In the leaves and shoots, there were the following compounds: trans-5-O-caffeoyl-quinic acid (trans-chlorogenic acid), caffeoyl-hexose-hydroxyphenol, p-coumaroyl arbutin, p-coumaroyl acetyl arbutin, 2-O-caffeoyl arbutin, 4-O-caffeoyl-quinic acid (cryptochlorogenic acid), caffeoyl iridoid, coumaroyl iridoid. Coumarin-fraxetin-8-O-glucoside which had been identified before, was also found out in the leaves and shoots.

As for the content of biologically active substances in *Vaccinium vitis-idaea* L. leaves, the main group of them was represented by phenologlycosides. The main compound was arbutin, its highest amount was detected in the young leaves. The second group was represented by hydroxycinnamic acids, the third one – by flavonoids. Anthocyanes were not identified in the leaves. Previously,

Ehala *et al.*³⁶ found out cinnamic acid (41.2 ± 2.36 µg/g of frozen berry extract) in *Vaccinium vitis-idaea* L. fruits by methods of capillary electrophoresis.

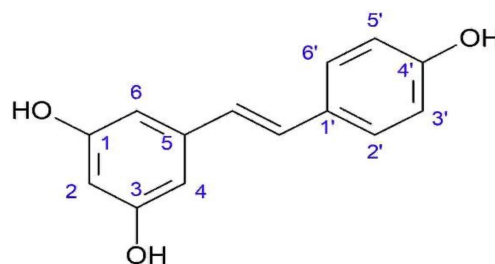
According to the data in the work by Zheng W,³⁷ in *V. vitis-idaea* L. berries caffeic acid was also present (6.2%). Hydroxycinnamic acids are represented by 7% from the total amount of phenolic compounds in *Vaccinium vitis-idaea* L. fruits.

Raudone *et al.*¹⁹ established the contents of chlorogenic acid and cryptochlorogenic acid in *V. vitis-idaea* L. cultivars and taxa listed above. The highest amount of chlorogenic and cryptochlorogenic acids was in *V. vitis-idaea* var. *leucocarpum* leaves (954.03 ± 38.95 mg/g DW and 1025.34 ± 41.86 mg/g DW, respectively).

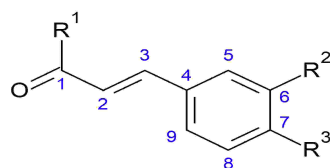
The data about hydroxycinnamic acids from *V. vitis-idaea* are given in Table 3.

Stilbenes

Rimando *et al.*³⁸ published experimental data on the content of stilbenes in *V. vitis-idaea* fruits (Figure 3).

**Figure 3.** Stilbenes from *Vaccinium vitis-idaea* L.: Resveratrol.

Lyophilisate from fresh fruits was extracted by the mixture of methanol-acetone-acetic acid (40:40:20:0.1, v/v/v/v), then the extract was condensed under vacuum. The obtained aqueous extract was then repeatedly subjected to extraction by ethyl acetate. The ethyl acetate extracts were mixed up and evaporated to dryness under vacuum.

Table 3. Hydroxycinnamic acids from *Vaccinium vitis-idaea* L.

No	Name	R ¹	R ²	R ³
1	Ferulic acid (4-hydroxy-3-methoxycinnamic acid)	OH	OCH ₃	OH
2	Cinnamic acid	OH	H	H
3	Caffeic acid	OH	OH	OH
4	Caffeoyl-hexose-hydroxyphenol (6-O-Caffeoylarbutin)	6-O-arbutin	OH	OH
5	Coumaroyl-hexose-hydroxyphenol (4-O-E-coumaroylarbutin)	4-O-arbutin	H	OH
6	2-O-Caffeoylarbutin	2-O-arbutin	OH	OH
7	Caffeoylshikimic acid (dactylifric acid)	3-O-4,5-dihydroxycyclohex-1-ene-1-carboxylic acid	OH	OH
8	Trans-chlorogenic acid (trans-5-caffeoyl-quinic acid)	5-O-1,3,5-trihydroxycyclohex-1-carboxylic acid	OH	OH
9	Cis-chlorogenic acid (cis-5-caffeoyl-quinic acid)	5-O-cis-1,3,4-trihydroxycyclohex-1-carboxylic acid	OH	OH
10	Chlorogenic acid (3-O-caffeoylquinic acid)	trans-3-O-quinic acid	OH	OH
11	P-coumaric acid (4-hydroxycinnamic acid)	OH	H	OH
12	Coumaroyl iridoid (Vaccinoside, 10-p-trans-coumaroyl-1S-monotropein)	1S-monotropein	H	OH
13	Cryptochlorogenic acid (4-O-trans-caffeoylquinic acid)	4-O-1,3,5-trihydroxycyclohex-1-carboxylic acid	OH	OH

The evaporated residue was diluted in water, and in the form of suspension it was placed into column C18 (Waters Scientific, Mississauga, ON). The process of elution was carried out, first, with water used as eluent, then with a mixture of methanol-acetone-water-formic acid (40:40:20:0.1, v/v/v/v). The fractions were analyzed by gas-liquid chromatography-mass spectrometry (GLC-MS). The amount of resveratrol was 5884 ng/g dry sample.

Ehala *et al.*³⁶ collected *Vaccinium vitis-idaea* L. fruits in the vicinities of Tallinn (Estonia) in 2005. The fruits were subjected to extraction by the mixtures of methanol-water (50:50 and 70:30, v/v) and ethanol-water (70:30, v/v). The obtained extracts from the fresh fruits were analyzed by capillary electrophoresis, using sodium tetraborate (pH 9.3) with 5% methanol as a buffer solution, 20 kV voltage, the temperature of 25°C and the detection at 210 nm. The analysis time was 5 minutes. The amount of trans-resveratrol in *Vaccinium vitis-idaea* L. frozen fruits was 30.00±2,8 µg/g.

Saponins

Szakiel and Mroczek³⁹ collected *Vaccinium vitis-idaea* L. rhizomes, stems, fruits and leaves in Poland (2004). The plant materials were shade dried at 40°C, ground and then placed into the Soxhlet apparatus. All the parts of the plant were subjected to the sequential extraction separately using diethyl ether and methanol. Then the methanol extract was condensed, diluted in water and extracted by n-butanol. The compounds were separated by CC using silica gel. The compounds were eluted and then hydrolyzed by Kiliani mixture (hydrochloric acid/acetic acid/water in the ratio of 10: 35: 55, v/v/v) at 100°C for 2 hours. Then the compounds

were identified by MS and GC. At all parts of *Vaccinium vitis-idaea* L. there was 3β-hydroxy-olea-12-en-28-oic acid (oleanolic acid) and its isomer 3β-hydroxy-urs-12-en-28-oic acid (ursolic acid).

In *Vaccinium vitis-idaea* L., the content of free triterpene acids, i.e. oleanolic acid, was the following: in fruits - 1.005%, in leaves - 0.61%, in shoots - 0.32%, in rhizomes - 0.35%. The content of ursolic acid at the lingonberry was 0.71% (fruits), 0.41% (leaves), 0.24% (shoots), 0.29% (rhizome).³⁹

Szakiel *et al.*⁴⁰ identified triterpene acids in the young and mature leaves collected in the flora area of Finland and Poland in the August (fruits and leaves) and December (only leaves) in 2010. The plant materials were extracted with diethyl ether. The extract was then hydrolyzed with 10% NaOH in 80% MeOH at 80 °C for 3 hours. The residue was diluted with water (1:5, v/v), then the solution was neutralized with 5% CH₃COOH. After that the solution was extracted with diethyl ether. The isolated triterpene acids and sterines were analyzed by TLC and identified by GC-MS. Lupeol and α-amyrin were identified by HPLC. Thus, the following compounds were identified in the young and mature leaves from Finland (1) and Poland (2) (Figures 4 and 5):

1. Neutral triterpenoids (α-amyrin, β-amyrin, betulin, cycloartanol, erythrodiol, fer-7-en-3β-ol, friedelin, lupeol, swert-9(11)-en-3β-ol, taraxasterol, urs-12-en-28-al, uvaol): their total amount was (1) 800.0 and (2) 1282.5 µg/g dry weight;
2. Triterpenic acids (oleanolic acid, ursolic acid); their total amount was (1) 8911.6 and (2) 8132.8 µg/g dry weight;
3. Steroids (campesterol, sitosterol, stigmasta-3,5-dien-7-

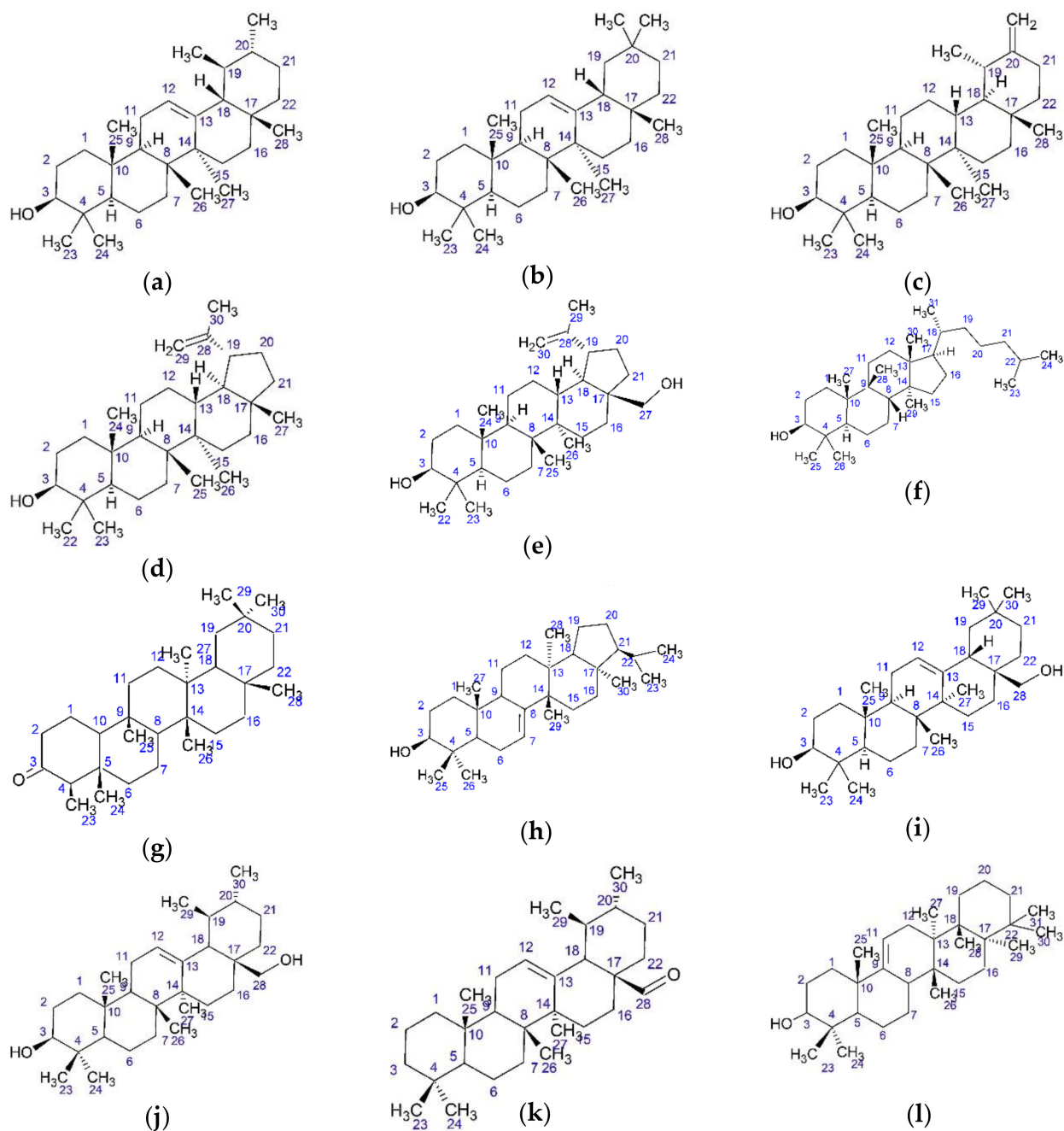


Figure 4. Triterpenes from *Vaccinium vitis-idaea* L.: (a) α -amyrin; (b) β -amyrin; (c) taraxasterol; (d) lupeol; (e) betulin; (f) cycloartanol; (g) friedelin; (h) fern-7-en- 3β -ol; (i) erythrodiol; (j) uvaol; (k) urs-12-en-28-al; (l) swert-9(11)-en- 3β -ol.

one, stigmasterol); their total amount was (1) 1591.0 and (2) 1813.6 $\mu\text{g/g}$ dry weight;

4. Ethers (α -amyrin, β -amyrin, fern-7-en- 3β -ol, taraxasterol, uvaol, oleanolic acid, ursolic acid, campesterol, sitosterol, stigmasterol); the total amount of the ethers was (1) 948.8 and (2) 431.9 $\mu\text{g/g}$ dry weight.

The total amount of all saponins was the following: in the young leaves (1) 8446.9 $\mu\text{g/g}$ dry weight and in the mature leaves 13693.9 $\mu\text{g/g}$ dry weight; in the young leaves (2) 5732.6 and in the mature leaves 13537.09 $\mu\text{g/g}$ dry weight. The plant materials were collected in August 2010. The

total amount of saponins in the young leaves was (1) 840.4 $\mu\text{g/g}$ dry weight and in the mature leaves - 14998.5 $\mu\text{g/g}$ dry weight; (2) the young leaves contained 10514.6 $\mu\text{g/g}$ and the mature leaves - 14075.5 $\mu\text{g/g}$ dry weight.

Qualitative and quantitative contents of saponins depended on both - the area of collection and the time of collection. Taraxasterol was not detected in the young leaves collected in December (Finland), while it was detected in all the materials collected in the flora area of Poland. Uvaol accumulated in the young leaves collected in Finland.⁴⁰

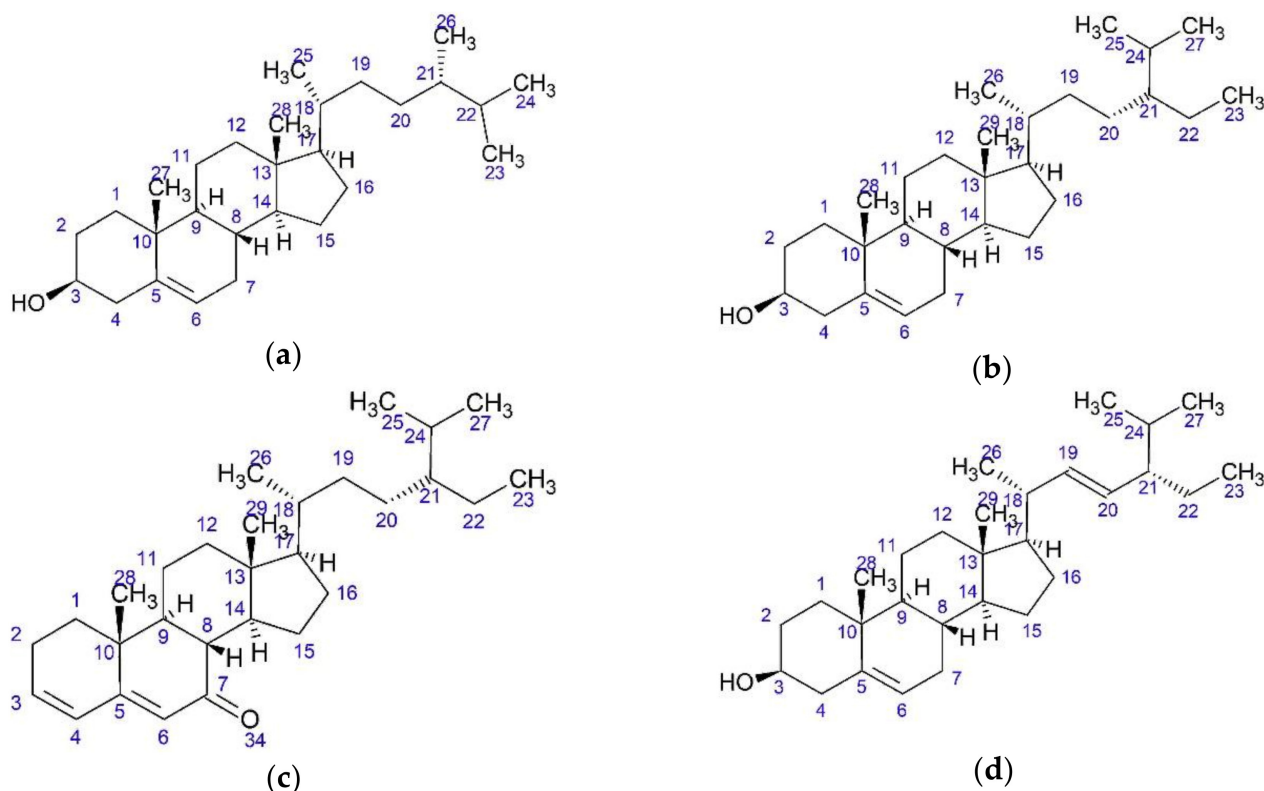


Figure 5. Steroids from *Vaccinium vitis-idaea* L.: (a) campesterol; (b) sitosterol; (c) stigmasta-3,5-dien-7-one; (d) stigmasterol.

Essential oil

Radulović *et al.*⁴¹ investigated the composition analysis of *Vaccinium vitis-idaea* L. essential oil. In its leaves 338 compounds were identified (90.4 from the total peaks area - by GLC). The main compounds were: α -terpineol (17.0%), pentacosane (6.4%), (E,E)- α -farnesene (4.9%), linalool (4.7%) and 3-hexen-1-ol (4.4%), hexadecanoic acid (4.5%), (E)-geranylacetone (4.1%), p-menthane (19.8%), farnesane (5.7%), caryophyllane (3.7%), eudesmane (3.0%), 10-epi- γ -eudesmol (3.0%) β -caryophyllen (2.9%), heptacosane (2.9%), nonacosane (2.0%) and geraniol (1.5%) (Figure 6). Monoterpenes account for 27.4 %, sesquiterpenes – for 18.7% of the total.

Organic acids and saccharides

Mikulic-Petkovsek *et al.*⁴² collected 18 species including *Vaccinium vitis-idaea* L. in the central part of Slovenia . The fruits were subjected to extraction with bidistillation water at the room temperature. Then the homogenate was centrifuged. The supernatant was filtrated and analyzed by HPLC-UV. In *Vaccinium vitis-idaea* L. fruits, glucose (37.9 g/kg), fructose (29.2 g/kg) and saccharose (0.45 g/kg) were identified. As for the organic acids, in *Vaccinium vitis-idaea* L. fruits they were represented by citric acid (20.0 g/kg), mallic acid (2.7 g/kg), tartaric acid (3.379 g/kg), fumaric acid (35.6 g/kg), shikimic acid (40.9 g/kg).⁴²

Pharmacological Properties

It has been established that the extracts obtained from

Vaccinium vitis-idaea, have a wide range of pharmacological activities, including antioxidant,²⁷ anti-inflammatory,⁴³ anti-neoplastic,⁴⁴ lipid corrective,⁴⁵ neuroprotective,⁴⁶ hypoglycemic,⁴⁷ antiapoptotic,⁴⁸ antiviral⁴⁹ and antibacterial⁵⁰ types of effects.

Antioxidant activity

A significant number of experimental researches have been devoted to the investigation of antioxidant properties of *Vaccinium vitis-idaea* extracts. So, according to Ho *et al.*²⁷ (dedicated to the evaluation of antioxidant activity, preclinical study), the tannins obtained from *Vaccinium vitis-idaea*, have the ability to inhibit the processes of lipid peroxidation, which was confirmed by a decrease in the formation of thiobarbituric acid-active products (thiobarbituric acid reactive substances or TBARS). In this study, the inhibitory properties of *Vaccinium vitis-idaea* tannins against superoxide radical (xanthine oxidase inhibition test) have also been established.

In another study conducted by Määttä-Riihinen *et al.*²⁹ (dedicated to the evaluation of antioxidant activity and hypolipidemic properties, preclinical *in vitro* study), it was established that *Vaccinium vitis-idaea* ethylacetate extract contains phenolic acids, catechins, procyanidins, and phenologlycosides. These substances *in vitro* suppressed the oxidation of low-density lipoproteins, which can indirectly indicate the presence of anti-atherosclerotic and angioprotective properties of the extract.

In addition, antiradical and chelating activity is specific for

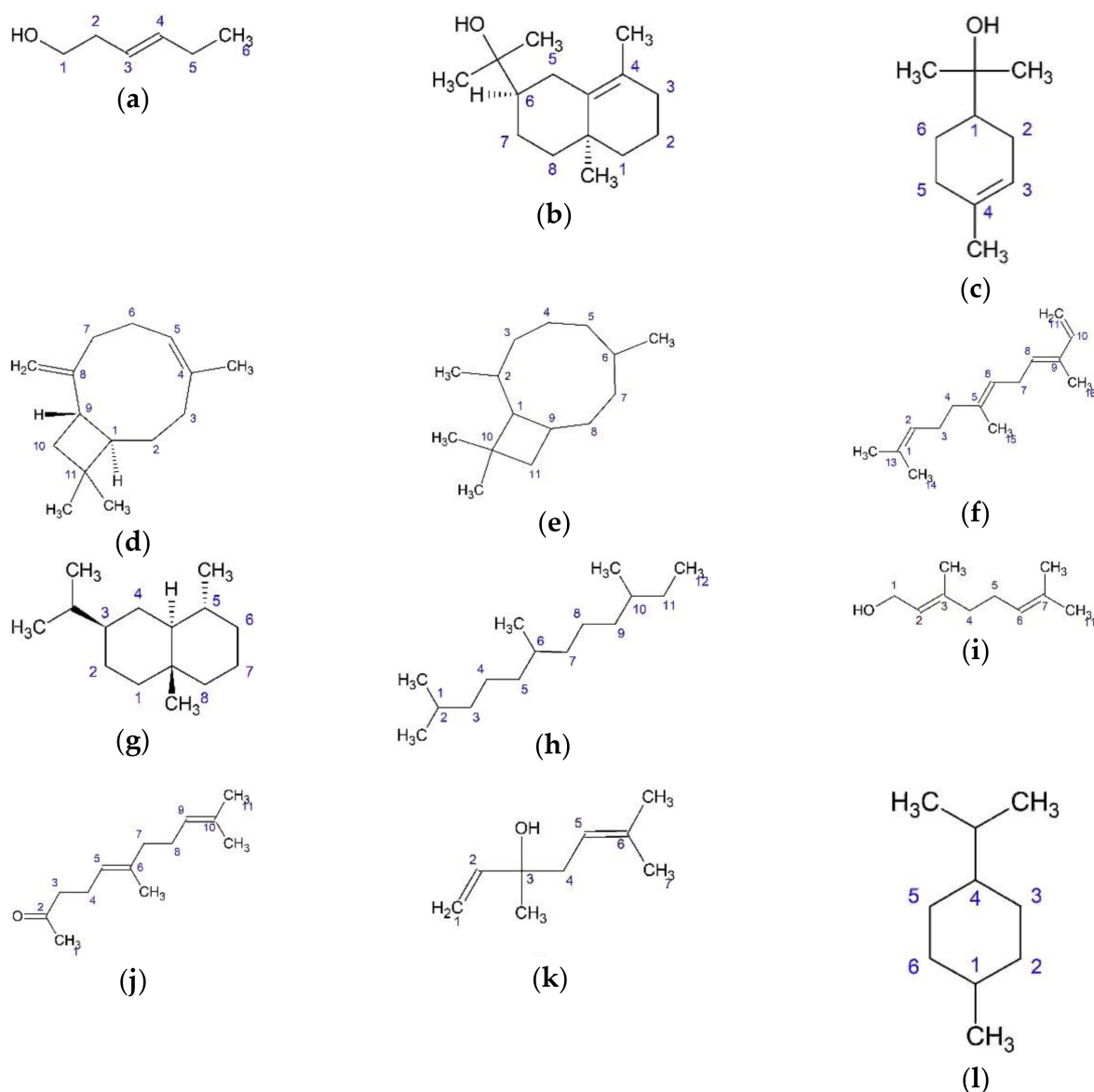


Figure 6. The main compounds of essential oil from *Vaccinium vitis-idaea* L.: (a) 3-hexen-1-ol; (b) 10-epi- γ -eudesmol; (c) α -terpineol; (d) β -caryophyllen; (e) caryophyllane; (f) E,E- α -farnesene; (g) eudesmane; (h) farnesane; (i) geraniol; (j) geranylacetone; (k) linalool; (l) p-menthane.

various extracts from *Vaccinium vitis-idaea*. Meanwhile, it was established that the ethanol extract from *Vaccinium vitis-idaea* L. leaves has scavenger properties relative to hydroxyl and ABTS-radicals (2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid), this extract was also characterized by the presence of metal chelating activity.¹⁹ The study of Wang *et al.*⁵¹ (dedicated to the evaluation of antioxidant activity, preclinical *in vitro* study) proved *in vitro* antiradical properties of *Vaccinium vitis-idaea* L. extract against DPPH (2,2-diphenyl-1-picrylhydrazyl), peroxy (ROO), hydroxyl (OH) and superoxide - anion radical. In this study, it was also shown that *Vaccinium vitis-idaea* extracts inhibit the activity of nuclear factor- κ B (NF- κ B), mitogen-activated protein kinases (MAPK) and activator protein-1 (AP-1) in mice's epidermal cell culture.

In addition, an experimental study conducted by Viljanen *et al.*⁵² (dedicated to the evaluation of antioxidant activity, preclinical *in vitro* study) showed that *Vaccinium vitis-idaea* L. fruit extract inhibits the oxidation of lipoprotein liposomes induced by lactalbumin.

The presence of direct antioxidant activity in the extracts obtained from *Vaccinium vitis-idaea* L. is quite important, as it can provide the basis for the anti-apoptotic action. It was proven in the study by Isaak *et al.*⁴⁸ (dedicated to the evaluation of antioxidant activity, preclinical *in vitro* study). In this study, the standardized *Vaccinium vitis-idaea* L. extract in the *in vitro* test, dose-dependently suppressed a cascade of reactive oxygen species (ROS)-induced apoptosis reactions in human cardiomyocyte cell cultures (H_2O_2 was used as an apoptosis-induce substance),

while this action was associated by the authors with a high content of anthocyanins in the analyzed extract.

Antibacterial activity

In addition to antioxidant properties - a "classic" type of pharmacological activity for natural origin objects - *Vaccinium vitis-idaea* L. extracts are characterized by the presence of antibacterial properties. Ho *et al.*⁵³ provides evidence that tannins isolated from *Vaccinium vitis-idaea* L. (epicatechin-(4 β →8)-epicatechin-(4 β →8, 2 β →O→7)-catechin) showed antimicrobial activity against *Porphyromonas gingivalis* and *Prevotella intermedia* - pathogens of periodontal infections. At the same time, the test objects were only slightly inferior in terms of the therapeutic effect of the tetracycline, the antibiotic that is most widely used for the treatment of infectious periodontitis.

Besides, the work carried out by Toivanen *et al.*,⁵⁴ (dedicated to the evaluation of antibacterial activity, preclinical study) demonstrated a high level of antimicrobial activity (comparable to the referent - streptavidin) of *Vaccinium vitis-idaea* L. extract in relation to *Neisseria meningitidis* Pili. This study also provides data on the potential mechanism of action of the studied extract. So, apparently, the antibacterial activity of *Vaccinium vitis-idaea* L. berries extract is associated with the inhibition of adhesion of *Neisseria meningitidis* Pili to human epithelial cells.

In addition to *Vaccinium vitis-idaea* L. berries extract, the extract obtained from *Vaccinium vitis-idaea* leaves, has also antibacterial properties. That was proved in the study by Wojnicz *et al.*⁵⁰ (devoted to the evaluation of antibacterial activity, preclinical study) where the aqueous extract from *Vaccinium vitis-idaea* L. leaves, had a bactericidal effect on *E. coli* strain isolated from urine of the patients suffering from bacterial pyelonephritis.

Vaccinium vitis-idaea L. extracts have also antitussive and mucolytic effects, which make these objects effective means of adjuvant therapy of upper respiratory tract infections.¹⁷ In this case, the antimicrobial effect of *Vaccinium vitis-idaea* L. extracts can be associated with suppression of bacterial hemagglutinin expression.⁵⁵ It is important that the endophytes of *Vaccinium vitis-idaea* have an antimicrobial activity (strains of such bacteria as *Mycobacterium aurum*, *M. tuberculosis* H37Rv), which opens up certain prospects for the use of herbal medicines based on *Vaccinium vitis-idaea* L., in the treatment of one of the main infectious causes of mortality in the world - tuberculosis.⁵⁶

Antifungal and antiviral activities

In addition to antibacterial activity, the extracts obtained from *Vaccinium vitis-idaea* L., are characterized by the presence of antifungal and antiviral properties. Cioch *et al.*⁵⁷ show the fungicidal activity of *Vaccinium vitis-idaea* L. fruit extract against pathogenic fungi of the genera *Aspergillus*, *Penicillium* and *Saccharomyces*. (antifungal and antiviral activity, preclinical study) The study by

Ermis *et al.*⁵⁸ (antifungal properties, preclinical study) demonstrated the fungicidal activity of *Vaccinium vitis-idaea* L. fruits, which, when powdered, acted as a dietary supplement to commercial juices with reduced sugar content, against pathogenic fungi *Absidia glauca*, *Penicillium brevicompactum*, *Saccharomyces cerevisiae* and *Zygosaccharomyces bailii*.

A study conducted by Nikolaeva-Glomb *et al.*⁴⁹ (antiviral properties, preclinical study) revealed a high level of antiviral activity of *Vaccinium vitis-idaea* L. fruit extract. In the course of this study, it was established that *in vitro* methanolic *Vaccinium vitis-idaea* L. fruit extract inhibited the replication of highly common and virulent pathogens: poliovirus type 1 (PV-1) and Coxsackie B1 virus (CV-B1), human respiratory syncytial virus A2 (HRSV-A2) and influenza A / H3N2.

Neuroprotective activity

The neuroprotective properties of some plant extracts are widely known, for example, *Ginkgo biloba* extract (Egb 761) and drugs based on it, which opens up the prospect of finding effective neuroprotectors among products of natural origin.⁵⁹ In this regard, a number of experimental studies on the evaluation of the neuroprotective activity of extracts from *Vaccinium vitis-idaea* were conducted.

Kelly *et al.*⁴⁶ (devoted to the evaluation of neuroprotective activity, preclinical study) report that *Vaccinium vitis-idaea* L. fruit extract has a pronounced neuroprotective effect under conditions of experimental glutamate-mediated excitotoxicity and β -amyloid-related neurotoxicity, which in turn opens up certain prospects for the application of these extracts in the treatment of neurodegenerative brain diseases such as Alzheimer's and Parkinson's diseases. At the same time, for *Vaccinium vitis-idaea* L. extracts, a complex mechanism of action is implied, which includes regulation of calcium concentration and NF- κ B signaling pathway, restoration of the mitochondrial membrane potential, reduction of cytokine activity (tumor necrosis factor alpha, TNF- α). In addition, in the culture of cortical cells, *Vaccinium vitis-idaea* L. extract reduced glutamate-mediated excitotoxicity.⁶⁰

Anti-inflammatory and immunotropic activity

Due to the presence of proven anti-cytokine properties in *Vaccinium vitis-idaea* L. extracts, the evaluation of the anti-inflammatory and immunotropic potential of these objects is promising. That was studied by Shaikh *et al.*⁴³ (preclinical study, *in vitro*) The experiments performed by Shaikh *et al.* (2015) made it possible to establish cyclooxygenase (COX)-inhibiting properties of *Vaccinium vitis-idaea* L. extract and, in addition, the anticytokine properties of this object were confirmed. This assumes a high therapeutic efficacy of the studied extract in the treatment of both acute and sluggish current chronic inflammatory processes.

In the work carried out by Vogl *et al.*⁶¹ (devoted to the evaluation of anti-inflammatory action mechanism, preclinical study) it was shown that *Vaccinium vitis-*

idaea L fruits extract *in vitro* suppresses the formation of interleukin-8, E-selectin and tumor necrosis factor – α , as well as promotes the activation of the peroxisome proliferator-activated receptors (PPAR α and PPAR γ), which may indicate the anti-inflammatory properties of *Vaccinium vitis-idaea* extracts, the presence of which was the basis for the use of these extracts in the treatment of inflammatory diseases of the gastrointestinal and urogenital tracts. Besides, the anti-inflammatory action of *Vaccinium vitis-idaea* L extracts, in particular a fruit extract, may be based on a decrease in vascular permeability, suppression of exudation and inflammatory edema, realized by inhibiting the activity of enzymes of the metalloproteinase family, which, in turn, has been proven in the work by Kondo *et al.*⁶² (devoted to the evaluation of anti-inflammatory action mechanism, preclinical study).

Cytotoxic activity

Among the drugs based on plant extracts, there are not only drugs for the treatment of various types of brain diseases - neuroprotectors - but also antineoplastic drugs intended for the treatment of cancer.

In the study by Du *et al.*⁶³ (devoted to the evaluation of anti-proliferative properties preclinical study), an anti-cancer activity of 4-[(Tetrahydro-2H-pyran-2-yl) oxy] phenol was established. This is an analogue of hydroquinone, identified as one of the active components in the extracts obtained from *Vaccinium vitis-idaea*,

In the C26 cell culture of colon cancer 4-[(Tetrahydro-2H-pyran-2-yl) oxy], phenol activated apoptosis by suppressing the activity of antiapoptotic B-cell lymphoma 2 (Bcl-2) proteins and activating proapoptotic Bax proteins and, accordingly, the caspase-dependent pathway of apoptosis.

Antineoplastic activity

The work carried out by Misikangas *et al.*⁶⁴ (anti-cancer action, preclinical *in vivo* study), showed that adding food enriched with the powder of *Vaccinium vitis-idaea* L berries to the diet of animals, reduces the growth of malignant colon carcinomas in mice by more than 60% ($p < 0.05$) relative to the control group. At the same time, researchers associate the anticancer activity of this nutrient with a decrease in the activity of oncogenes, such as: β -catenin, cyclin D1, adenosine deaminase, ecto-5-nucleotidase and prostaglandin E2 (in the case of affinity for EP4 receptors). In the study by Fan *et al.*⁶⁵ (radioprotective properties, preclinical *in vivo* study) the radioprotective effect of anthocyanins extracted from *Vaccinium vitis-idaea* L fruits. It is expressed in the restoration of all hematopoiesis sprouts, normalization of phagocytic activity in animals, in conditions of acute radiation exposure, which, in turn, can also be a reference to antineoplastic properties of the extracts obtained from *Vaccinium vitis-idaea* L.

Hypoglycemic activity

In addition to the above listed types of pharmacological activity, *Vaccinium vitis-idaea* L. extracts are characterized

by the presence of hypoglycemic properties.

In the current conditions of extreme prevalence of insulin-resistant diabetes mellitus and the practical absence of safe and cost-effective methods for pharmacocorrection of this type of diabetes, the use of *Vaccinium vitis-idaea* L. extracts can be an effective alternative method for treating hyperglycemic conditions. The work carried out by Eid *et al.*⁶⁶ (hypoglycemic action, preclinical *in vivo* study), demonstrated a significant potential for glycemic control of *Vaccinium vitis-idaea* L. in C57BL/6 mice under conditions of insulin resistant diabetes. At the same time, the potential mechanisms of the hypoglycemic action of the studied object, including an increase in the activity of glucose-transporting metabolic systems (GLUT-4), 5' adenosine monophosphate-activated protein kinase (AMPK) and serine/threonine-specific protein kinase (Akt) in skeletal muscles, were established. In addition, AMPK-activating properties of extracts from *Vaccinium vitis-idaea* were proven in the study by Eid *et al.*⁶⁷ (hypoglycemic action mechanisms evaluation, preclinical *in vivo* study), hereby it was found out that activation of AMPA in striated muscle caused by the use of *Vaccinium vitis-idaea* L. berries, was not accompanied by the dissociation of electron transfer reactions in the mitochondrial respiratory chain, therefore the risk of hyperlactataemia when using products based on *Vaccinium vitis-idaea*, is minimal, which distinguishes this object from those traditionally used in insulin-resistant diabetes activators of AMPK, for example, metformin.

It is quite important that, in addition to the directly hypoglycemic effect, *Vaccinium vitis-idaea* L. extracts exhibit a pleiotropic pharmacological action in diabetes mellitus. So, in the work by Reichert *et al.*⁶⁸ (devoted to the evaluation of pleiotropic hypoglycemic properties preclinical study) it was shown that the use of *Vaccinium vitis-idaea* L. fruits in experimental diabetes mellitus, contributed to the reduction of *central* nervous system complications in diabetes mellitus. The study by Zhu *et al.*⁴⁵ (devoted to the evaluation of hypoglycemic properties preclinical study) demonstrated that while using *Vaccinium vitis-idaea* L., there is a decrease in the concentration of low density lipoprotein (LDL) and an increase in the high density lipoprotein (HDL) fraction in the *blood serum*. It should be notified that the hypoglycemic properties of *Vaccinium vitis-idaea* L. extracts were confirmed by clinical data.⁶⁹ The *in vitro* studies also confirm the presence of hypoglycemic activity in *Vaccinium vitis-idaea* L. Thus, the work by Quek and Henry⁷⁰ (hypoglycemic properties preclinical *in vivo* study), shows that crushed *Vaccinium vitis-idaea* L. fruits improve glycemic control by reducing the breakdown of complex carbohydrates to monosaccharides. In addition, a high hypoglycemic activity of *Vaccinium vitis-idaea* L., has been confirmed in a number of clinical studies.

According to Rokka *et al.*⁷¹ (hypoglycemic properties clinical study), the use of powdered *Vaccinium vitis-idaea* L. berries in 9 healthy volunteers at the median age of 35 years (20-54 years), contributed to a significant decrease

in basal blood glucose levels. Hypoglycemic effect is associated with the modulation of the PPAR γ function and, as a result, the elimination of insulin resistance.

Meanwhile, two studies conducted by Törrönen *et al.*⁷² and Törrönen *et al.*⁷³ (hypoglycemic properties clinical study), reported a decrease in postprandial glucose levels and insulin secretion in healthy female volunteers. However, it should be notified that in the first case, the patients received

150 g of *Vaccinium vitis-idaea* L. in the form of berry puree, and in the second case, the therapy was carried out with a multi-component polyphenolic composition, which included equal amounts of strawberries, blueberries, cranberries and black currants.

The summarized data characterizing the spectrum of the pharmacological activity of *Vaccinium vitis-idaea* L., are presented in Table 4.

Table 4. Types of pharmacological activity of *Vaccinium vitis-idaea* L.

Pharmacological activity	Extract	Type of study	The dose/concentration IC ₅₀ range for <i>in vitro</i> tests	Methods	Ref.
Antioxidant, chellating	Tanins	Preclinical	IC ₅₀ – 7.76-630.0	Thiobarbituric acid test, oxidase inhibition test, cytochrome C test	27
	Ethyl acetate extract	Preclinical	IC ₅₀ – 13.0-86.0	DPPH-test, oxidation of methyl linoleate emulsion test, LDL oxidation test	28
	Ethanol extract	Preclinical	Inhibition percent – 12.08-88.5%	ABTS Radical inhibition test, Ferric Reducing Antioxidant Power test, Ferrous Ions Chelating Activity test	19
Antiapoptotic	Standardized extract	Preclinical	20 nmol/L	Hydrogen peroxyde induced apoptosis of H9c2 cells	48
Anti-inflammatory	Aqueous extract of fruits	Preclinical	1 mg/ml	Carrageenan and Phorbol Myristate Acetate induced edema	43
	Berries	Preclinical	Not indicated	DU145 prostate tumor cells.	62
Antineoplastic	(4-[(Tetrahydro-2H-pyran-2-yl) oxy] phenol)	Preclinical	2.5, 5, 10, 20 or 40 µg/ml	Inhibition of C26 murine colon carcinoma cell proliferation	63
	Berries Powder	Preclinical	10%	Multiple intestinal neoplasia/+ mice biological model	64
Radioprotective	Aqueous extract of fruits	Preclinical	50, 100, 200 mg/kg	Irradiation γ -radiation (6 Gy), at a dose rate of 1.01 Gy/min	65
Hypolipidemic	Lingonberry juice	Clinical	2.1 mg/day	Hypolipidemic activity investigated on patients with diabetes mellitus	45
	Lingonberry juice cocktail		69.5 mg/day		
	Lingonberry drink		375 mg/day		
	Lingonberry juice		24.8 mg/day		
Neuroprotective	Aqueous extract of fruits	Preclinical	25 mg/kg; 50 mg/kg	Transgenic model of Alzheimer disease	46
	Berries Powder	Preclinical	125, 250 and 500 mg/kg	Streptozotocin induced diabetes mellitus	68
Hypoglycemic	Berries Powder	Preclinical	125, 250 and 500 mg/kg	Dietary mouse model / diet-induced obesity	66
	Berries Powder	Clinical	Not indicated	Activity investigated on 9 healthy volunteers, glucose control	71
	Berries puree	Clinical	150 g	Activity investigated on healthy femalevolunteers, postprandial glucose control	72
	Berries (in combination)	Clinical	Not indicated	Activity investigated on healthy female volunteers, postprandial glucose and insulin control	73
Antiviral	Methanol extract of fruits	Preclinical	Not indicated	<i>In vitro</i> antiviral tests	49
Antibacterial	Tanins	Preclinical	0.5%	Agar dilution method	53
	Aqueous extract of fruits		0.5–8 µg/ml	Dot Binding and Binding Inhibition Assay	54
	Aqueous extract of leaves		0.125-20 mg/ml	Agar dilution test	50
Antifungal	Methanol, aqueous and ethanol extracts	Preclinical	Not indicated	Diffusion paper-disc assay	57

Discussion

The analysis of the literature data made it possible to arrive at the conclusion that *Vaccinium vitis-idaea* L. is a polymorphic species growing all over the area of Russia and it is a species typical of European countries. It also occurs in the USA. When grown together, *Vaccinium myrtillus* and *Vaccinium vitis-idaea* can form a natural infraspecies hybrid *Vaccinium x intermedium* Ruthe. *Vaccinium vitis-idaea* L. leaves are a pharmacopoeia medicinal plant material in Russia and in the Republic of Belarus.

The main biologically active compounds of lingonberry are phenylpropanoids, simple phenols such as arbutin and hydroquinone as the main components, flavonoids, proanthocyanidines, anthocyanidines, catechins, saponins, etc. The majority of works describe chemical contents of fruits because berries are used as a medicinal and edible plant material. Several authors claim that the chemical contents depend on cultivars of *Vaccinium vitis-idaea* L., the time of harvesting and the flora area. The quality control of *V. vitis-idaea* raw materials is carried out only by the total amount of phenolglycosides. Metabolomics research and developing the standardized finger prints plants from different habitats and different growing season are the actual future aim.

The analysis of the types of the pharmacological activity of *Vaccinium vitis-idaea* extracts, made it possible to establish that the majority of studies in which the biological activity of this plant is studied, are limited to the works performed *in vitro*. Thus, despite a sufficient number of researches devoted to the evaluation of the antioxidant activity of *Vaccinium vitis-idaea* extracts, all of them are usually focused on the evaluation of antiradical and chelating properties of these objects.

Meanwhile, the undoubted scientific and practical interest can be presented by the study of the effect of *Vaccinium vitis-idaea* L. extracts on the processes of peroxidation *in vivo*, with the assessment of changes in the activity of endogenous antioxidant enzymes such as superoxide dismutase, catalase, enzymes of glutathione series (glutathione peroxidase, glutathione-S-transferase), and also with the determination of the concentration of lipid peroxidation products (TBARS). Taking into account the fact that an oxidative stress is a "universal" pathophysiological process and underlies pathogenesis of a number of diseases, *in vivo* evaluation of the antioxidant activity of *Vaccinium vitis-idaea* L. extracts, will significantly expand the therapeutic potential of drugs obtained from *Vaccinium vitis-idaea* L.. The study of the neuroprotective activity of *Vaccinium vitis-idaea* extracts is also of significant interest. Thus, the established anti-cytokine and antiapoptotic properties in combination with the ability of these extracts to reduce the cytotoxicity of β -amyloid may be a prerequisite for the use of *Vaccinium vitis-idaea* extracts in the treatment of a number of progressive neurodegenerative diseases. That requires, however, a more detailed study. Hereby, this type of pharmacological activity (AMPK activation), opens up certain perspectives of medicines based on the application

of *Vaccinium vitis-idaea* L.: correction of muscle fatigue and ischemic/hypoxic state. It can also be important to study various aspects of the possible mechanisms of action of these extracts. It may be appropriate to assess the effect of *Vaccinium vitis-idaea* L. extracts on the activity of metabolic transport systems (various subclasses of GLUT), PPAR, caspases and other effectors.

Conclusion

The carried out analysis of the literature data on the chemical contents and pharmacological studies of *Vaccinium vitis-idaea* L. revealed unresolved issues. There are other groups of natural compounds, such as polysaccharides, phenylpropanoids, tannins, the quantitative contents of which is has not been mentioned in the literature. The main part of the work contains the results of *Vaccinium vitis-idaea* L. fruits research. Such parts of the plant as leaves, are not fully investigated, though they have been used for a long time in traditional medicine. In addition, according to the above presented data, the flora area affects both the qualitative and quantitative composition of biologically active compounds.

There are no reliable data on the regularity of accumulation of biologically active substances in various habitats. The neuroprotective effect is a potentially new type of activity. In most studies, the activity of *Vitis-idaea* extracts is tested *in vitro*. In the absence of *in vivo* test results, it cannot be considered sufficient. In addition, it seems scientifically justified to study the mechanisms of action, extracts from lingonberries. Thus, it is possible to study the mechanisms of implementation of cerebrotropic, diuretic and antioxidant types of activity, with the establishment of specific pharmacotherapeutic targets at the molecular level. It is also advisable to further evaluate the hypoglycemic and hypolipidemic properties of extracts, while their PPAR – agonistic activity is seen as interesting, which can serve as a prerequisite for the creation of drugs with polyvalent pharmacological activity.

All these aspects establish the high interest for future investigations *Vaccinium vitis-idaea* L. as a source of natural drugs with wide range of activities.

Acknowledgments

The authors acknowledge Pyatigorsk Medical-Pharmaceutical Institute (PMPI), Branch of Volgograd State Medical University, Ministry of Health of Russia, Pyatigorsk, the Russian Federation, Kursk State Medical University (KSMU), Ministry of Health of Russia, Kursk.

Conflict of Interest

The authors declare they have no conflict of interest.

References

1. Czerepanov SK. Vascular plants of Russia and neighboring countries (within the former USSR). Russian Edition. Saint-Petersburg: Word and Family-95; 1995.

2. The Plant List. The Royal Botanic Gardens, Kew and Missouri Botanical Garden. Available at: <http://www.theplantlist.org> (Accessed 14 February 2020)
3. Takhtadjan AL. The system of magnoliophytes. Leningrad: Nauka;1987.
4. The Angiosperm Phylogeny Group. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. Bot J Linn Soc. 2016;181(1): 1–20. doi:10.1111/boj.12385
5. Kron KA, Powell EA, Luteyn JL. Phylogenetic relationships within the blueberry tribe (Vaccinieae, Ericaceae) based on sequence data from MATK and nuclear ribosomal ITS regions, with comments on the placement of Satyria. Am J Bot. 2002;89(2):327-36. doi:10.3732/ajb.89.2.327
6. Komarov VL, Shishkin BK. Flora of the USSR. In: Komarov VL, Shishkin BK, Bobrov EG, editors. Volume 18. Leningrad: Publishing of Academy of Sciences of USSR; 1952.
7. Tutin TG, Heywood VH, Burges NA, Moore DM, Valentine DH, Walters SM, et al. Flora Europaea: Diapensiaceae to Myoporaceae. Vol. 3. London: Cambridge University Press; 1972.
8. Kole K. Wild Crop Relatives: Genomic and Breeding Resources Temperate Fruits. Berlin, Heidelberg: Springer-Verlag;2011.p.197-221. doi:10.1007/978-3-642-16057-8
9. Pharmacopoea Rossica. Petropoli. 1782. Available at: https://books.google.ru/s?id=uL2rvC5GRvMC&dq=Pharmacopoea+Rossica&hl=ru&source=gbs_navlinks_s. (Accessed 14 February 2020).
10. Shikov AN, Pozharitskaya ON, Makarov VG, Wagner H, Verpoorte R, Heinrich M. Medicinal plants of the Russian Pharmacopoeia; their history and applications. J Ethnopharmacol. 2014;154(3):481-536. doi:10.1016/j.jep.2014.04.007
11. Zhou J, Xie G, Yan X. Encyclopedia of Traditional Chinese Medicines — Molecular Structures, Pharmacological Activities, Natural Sources and Applications. Volume 5. Isolated Compounds T-Z. Berlin, Heidelberg: Springer-Verlag; 2011. doi:10.1007/978-3-642-16741-6
12. Abisheva LN, Belenovskaya LM, Bobileva NS, Bikova OP, Kondratenkova TD, Kudrova EL, et al. Useful wild plants of Russia. Budancev AL, Lesiovskaya EE, editors. St. Petersburg: SPCPA: 2001.
13. Budancev AL. Plant resources of Russia. Flowering wild plants, their phytochemical constituents and biological activities. Family Actinidiaceae – Malvaceae, Euphorbiaceae – Haloragaceae. Volume 2. Saint Peterburg: KMK PRESS; 2009. p. 54-6.
14. Friedrich H, Schönert J. Untersuchungen über einige Inhaltsstoffe der Blätter und Früchte von *Vaccinium myrtillus* [Phytochemical investigation of leaves and fruits of *Vaccinium myrtillus* (author's transl)]. Planta Med. 1973;24(1):90-100. German. doi:10.1055/s-0028-1099474
15. Sticher O, Soldati F, Lehmann D. High-performance liquid chromatographic separation and quantitative determination of arbutin, methylarbutin, hydroquinone and hydroquinone-monomethylether in *Arctostaphylos*, *Bergenia*, *Calluna* and *Vaccinium* species (author's transl). Planta Med. 1979;35(3):253-61. German. doi:10.1055/s-0028-1097213
16. Pyka A, Bober K, Stolarczyk A. Densitometric determination of arbutin in cowberry leaves (*Vaccinium vitis idaeae*). Acta Pol Pharm. 2007;64(5):395-400
17. Wang X, Sun H, Fan Y, Li L, Makino T, Kano Y. Analysis and bioactive evaluation of the compounds absorbed into blood after oral administration of the extracts of *Vaccinium vitis-idaea* in rat. Biol Pharm Bull. 2005;28(6):1106-8. doi:10.1248/bpb.28.1106
18. Liu P, Lindstedt A, Markkinen N, Sinkkonen J, Suomela JP, Yang B. Characterization of metabolite profiles of leaves of bilberry (*Vaccinium myrtillus* L.) and lingonberry (*Vaccinium vitis-idaea* L.). J Agric Food Chem. 2014;62(49):12015-26. doi:10.1021/jf503521m
19. Raudone L, Vilkičkyte G, Pitkauskaitė L, Raudonis R, Vainoriene R, Motiekaitė V. Antioxidant activities of *Vaccinium vitis-idaea* L. leaves within cultivars and their phenolic compounds. Molecules. 2019;24(5):844. doi:10.3390/molecules24050844
20. Ek S, Kartimo H, Mattila S, Tolonen A. Characterization of phenolic compounds from lingonberry (*Vaccinium vitis-idaea*). J Agric Food Chem. 2006;54(26):9834-42. doi:10.1021/jf0623687
21. Lätti AK, Riihinen KR, Jaakola L. Phenolic compounds in berries and flowers of a natural hybrid between bilberry and lingonberry (*Vaccinium* × *intermedium* Ruthe). Phytochemistry 2011;72(8):810–5. doi:10.1016/j.phytochem.2011.02.015
22. Riihinen KR, Gödecke T, Pauli GF. Purification of berry flavonol glycosides by long-bed gel permeation chromatography. J Chromatogr A. 2012;1244:20-7. doi:10.1016/j.chroma.2012.04.060
23. Riihinen KR, Mihaleva VV, Gödecke T, Soinenen P, Laatikainen R, Vervoort JM, et al. ¹H-NMR fingerprinting of *Vaccinium vitis-idaea* flavonol glycosides. Phytochem Anal. 2013;24(5):476-83. doi:10.1002/pca.2444
24. Drózd P, Šežienė V, Pyrzyńska K. Phytochemical properties and antioxidant activities of extracts from wild blueberries and lingonberries. Plant Foods Hum Nutr. 2017;72(4):360-4. doi:10.1007/s11130-017-0640-3
25. Drózd P, Šežienė V, Wójcik J, Pyrzyńska K. Evaluation of bioactive compounds, minerals and antioxidant activity of lingonberry (*Vaccinium vitis-idaea* L.) Fruits. Molecules. 2017;23(1):53. doi:10.3390/molecules23010053
26. Jurikova T, Skrovankova S, Mlcek J, Balla S, Snopek L. Bioactive compounds, antioxidant activity, and

- biological effects of European cranberry (*Vaccinium oxycoccos*). *Molecules*. 2018;24(1):24. doi:10.3390/molecules24010024
27. Ho KY, Huang JS, Tsai CC, Lin TC, Hsu YF, Lin CC. Antioxidant activity of tannin components from *Vaccinium vitis-idaea* L. *J Pharm Pharmacol*. 1999;51(9):1075-8. doi:10.1211/0022357991773410
28. Määttä-Riihinen KR, Kähkönen MP, Törrönen AR, Heinonen IM. Catechins and procyanidins in berries of vaccinium species and their antioxidant activity. *J Agric Food Chem*. 2005;53(22):8485-91. doi:10.1021/jf050408l
29. Riihinen KR, Ou ZM, Gödecke T, Lankin DC, Pauli GF, Wu CD. The antibiofilm activity of lingonberry flavonoids against oral pathogens is a case connected to residual complexity. *Fitoterapia*. 2014;97:78-86. doi:10.1016/j.fitote.2014.05.012
30. Viljanen K, Kylli P, Hubbermann EM, Schwarz K, Heinonen M. Anthocyanin antioxidant activity and partition behavior in whey protein emulsion. *J Agric Food Chem*. 2005;53(6):2022-7. doi:10.1021/jf047975d
31. Dudonné S, Vitrac X, Coutière P, Woillez M, Mérillon JM. Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. *J Agric Food Chem*. 2009;57(5):1768-74. doi:10.1021/jf803011r
32. Lee J. Anthocyanin analyses of *Vaccinium* fruit dietary supplements. *Food Sci Nutr*. 2016;4(5):742-52. doi:10.1002/fsn3.339
33. Brown EM, Nitecki S, Pereira-Caro G, McDougall GJ, Stewart D, Rowland I, et al. Comparison of in vivo and in vitro digestion on polyphenol composition in lingonberries: potential impact on colonic health. *Biofactors*. 2014;40(6):611-23. doi:10.1002/biof.1173
34. Hokkanen J, Mattila S, Jaakola L, Pirttilä AM, Tolonen A. Identification of phenolic compounds from lingonberry (*Vaccinium vitis-idaea* L.), bilberry (*Vaccinium myrtillus* L.) and hybrid bilberry (*Vaccinium x intermedium* Ruthe L.) leaves. *J Agric Food Chem*. 2009;57(20):9437-47. doi:10.1021/jf902254z
35. Ieri F, Martini S, Innocenti M, Mulinacci N. Phenolic distribution in liquid preparations of *Vaccinium myrtillus* L. and *Vaccinium vitis-idaea* L. *Phytochem Anal*. 2013;24(5):467-75. doi:10.1002/pca.2462
36. Ehala S, Vaher M, Kaljurand M. Characterization of phenolic profiles of northern European berries by capillary electrophoresis and determination of their antioxidant activity. *J Agric Food Chem*. 2005;53(16):6484-90. doi:10.1021/jf050397w
37. Zheng W, Wang SY. Oxygen radical absorbing capacity of phenolics in blueberries, cranberries, chokeberries, and lingonberries. *J Agric Food Chem*. 2003;51(2):502-09. doi:10.1021/jf020728u
38. Rimando AM, Kalt W, Magee JB, Dewey J, Ballington JR. Resveratrol, pterostilbene, and piceatannol in vaccinium berries. *J Agric Food Chem*. 2004;52(15):4713-9. doi:10.1021/jf040095e
39. Szakiel A, Mroczek A. Distribution of triterpene acids and their derivatives in organs of cowberry (*Vaccinium vitis-idaea* L.) plant. *Acta Biochim Pol*. 2007;54(4):733-40.
40. Szakiel A, Pączkowski C, Koivuniemi H, Huttunen S. Comparison of the triterpenoid content of berries and leaves of lingonberry *Vaccinium vitis-idaea* from Finland and Poland. *J Agric Food Chem*. 2012;60(19):4994-5002. doi:10.1021/jf300375b
41. Radulović N, Blagojević P, Palić R. Comparative study of the leaf volatiles of *Arctostaphylos uva-ursi* (L.) Spreng. and *Vaccinium vitis-idaea* L. (Ericaceae). *Molecules*. 2010;15(9):6168-85. doi:10.3390/molecules15096168
42. Mikulic-Petkovsek M, Schmitzer V, Slatnar A, Stampar F, Veberic R. Composition of sugars, organic acids, and total phenolics in 25 wild or cultivated berry species. *J Food Sci*. 2012;77(10):C1064-70. doi:10.1111/j.1750-3841.2012.02896.x
43. Shaikh RU, Pund MM, Gacche RN. Evaluation of anti-inflammatory activity of selected medicinal plants used in Indian traditional medication system in vitro as well as in vivo. *J Tradit Complement Med*. 2016;6(4):355-61. doi:10.1016/j.jtcme.2015.07.001
44. McDougall GJ, Ross HA, Ikeji M, Stewart D. Berry extracts exert different antiproliferative effects against cervical and colon cancer cells grown in vitro. *J Agric Food Chem*. 2008;56(9):3016-23. doi:10.1021/jf073469n
45. Zhu Y, Miao Y, Meng Z, Zhong Y. Effects of *Vaccinium* berries on serum lipids: A meta-analysis of randomized controlled trials. *Evid Based Complement Alternat Med*. 2015;2015:790329. doi:10.1155/2015/790329
46. Kelly E, Vyas P, Weber JT. Biochemical properties and neuroprotective effects of compounds in various species of berries. *Molecules*. 2017;23(1):26. doi:10.3390/molecules23010026
47. Harbilas D, Martineau LC, Harris CS, Adeyiwola-Spoor DC, Saleem A, Lambert J, et al. Evaluation of the antidiabetic potential of selected medicinal plant extracts from the Canadian boreal forest used to treat symptoms of diabetes: part II. *Can J Physiol Pharmacol*. 2009;87(6):479-92. doi:10.1139/y09-029
48. Isaak CK, Petkau JC, Blewett H, O K, Siow YL. Lingonberry anthocyanins protect cardiac cells from oxidative-stress-induced apoptosis. *Can J Physiol Pharmacol*. 2017;95(8):904-10. doi:10.1139/cjpp-2016-0667
49. Nikolaeva-Glomb L, Mukova L, Nikolova N, Badjakov I, Dincheva I, Kondakova V, et al. In vitro antiviral activity of a series of wild berry fruit extracts against representatives of Picorna-, Orthomyxo- and Paramyxoviridae. *Nat Prod Commun*. 2014;9(1):51-4.
50. Wojnicz D, Kucharska AZ, Sokół-Łętowska A, Kicia M, Tichaczek-Goska D. Medicinal plants extracts affect virulence factors expression and biofilm formation by the uropathogenic *Escherichia coli*. *Urol Res*. 2012

- Dec;40(6):683-97. doi:10.1007/s00240-012-0499-6
51. Wang SY, Feng R, Bowman L, Penhallegon R, Ding M, Lu Y. Antioxidant activity in lingonberries (*Vaccinium vitis-idaea* L.) and its inhibitory effect on activator protein-1, nuclear factor-kappaB, and mitogen-activated protein kinases activation. *J Agric Food Chem.* 2005;53(8):3156-66. doi:10.1021/jf048379m
52. Viljanen K, Kylli P, Kivikari R, Heinonen M. Inhibition of protein and lipid oxidation in liposomes by berry phenolics. *J Agric Food Chem.* 2004;52(24):7419-24. doi:10.1021/jf049198n
53. Ho KY, Tsai CC, Huang JS, Chen CP, Lin TC, Lin CC. Antimicrobial activity of tannin components from *Vaccinium vitis-idaea* L. *J Pharm Pharmacol.* 2001;53(2):187-91. doi:10.1211/0022357011775389
54. Toivanen M, Rynnänen A, Huttunen S, Duricová J, Riihinen K, Törrönen R, et al. Binding of *Neisseria meningitidis* Pili to berry polyphenolic fractions. *J Agric Food Chem.* 2009; 57(8): 3120-7. doi:10.1021/jf803488s
55. Kylli P, Nohynek L, Puupponen-Pimiä R, Westerlund-Wikström B, Leppänen T, Welling J, et al. Lingonberry (*Vaccinium vitis-idaea*) and European cranberry (*Vaccinium microcarpon*) proanthocyanidins: isolation, identification, and bioactivities. *J Agric Food Chem.* 2011;59(7):3373-84. doi:10.1021/jf104621e
56. Gordien AY, Gray AI, Ingleby K, Franzblau SG, Seidel V. Activity of Scottish plant, lichen and fungal endophyte extracts against *Mycobacterium aurum* and *Mycobacterium tuberculosis*. *Phytother Res.* 2010;24(5):692-8. doi:10.1002/ptr.2988
57. Cioch M, Satora P, Skotniczny M, Semik-Szczurak D, Tarko T. Characterisation of antimicrobial properties of extracts of selected medicinal plants. *Pol J Microbiol.* 2017;66(4):463-72. doi:10.5604/01.3001.0010.7002
58. Ermis E, Hertel C, Schneider C, Carle R, Stintzing F, Schmidt H. Characterization of in vitro antifungal activities of small and American cranberry (*Vaccinium oxycoccos* L. and *V. macrocarpon* Aiton) and lingonberry (*Vaccinium vitis-idaea* L.) concentrates in sugar reduced fruit spreads. *Int J Food Microbiol.* 2015;204:111-7. doi:10.1016/j.ijfoodmicro.2015.03.017
59. Wang N, Chen X, Geng D, Huang H, Zhou H. Ginkgo biloba leaf extract improves the cognitive abilities of rats with D-galactose induced dementia. *J Biomed Res.* 2013;27(1):29-36. doi:10.7555/JBR.27.20120047
60. Vyas P, Kalidindi S, Chibrikova L, Igamberdiev AU, Weber JT. Chemical analysis and effect of blueberry and lingonberry fruits and leaves against glutamate-mediated excitotoxicity. *J Agric Food Chem.* 2013;61(32):7769-76. doi:10.1021/jf401158a
61. Vogl S, Picker P, Mihaly-Bison J, Fakhruddin N, Atanasov AG, Heiss EH, et al. Ethnopharmacological in vitro studies on Austria's folk medicine--an unexplored lore in vitro anti-inflammatory activities of 71 Austrian traditional herbal drugs. *J Ethnopharmacol.* 2013;149(3):750-71. doi:10.1016/j.jep.2013.06.007
62. Kondo M, MacKinnon SL, Craft CC, Matchett MD, Hurta RA, Neto CC. Ursolic acid and its esters: occurrence in cranberries and other *Vaccinium* fruit and effects on matrix metalloproteinase activity in DU145 prostate tumor cells. *J Sci Food Agric.* 2011;91(5):789-96. doi:10.1002/jsfa.4330
63. Du Q, Xin G, Niu H, Huang W. Hydroquinone analog 4-[(Tetrahydro-2H-pyran-2-yl) oxy] phenol induces C26 colon cancer cell apoptosis and inhibits tumor growth in vivo. *Mol Med Rep.* 2015;11(6):4671-7. doi:10.3892/mmr.2015.3300
64. Misikangas M, Pajari AM, Päivärinta E, Oikarinen SI, Rajakangas J, Marttinen M, et al. Three Nordic berries inhibit intestinal tumorigenesis in multiple intestinal neoplasia/+ mice by modulating beta-catenin signaling in the tumor and transcription in the mucosa. *J Nutr.* 2007;137(10):2285-90. doi:10.1093/jn/137.10.2285
65. Fan ZL, Wang ZY, Zuo LL, Tian SQ. Protective effect of anthocyanins from lingonberry on radiation-induced damages. *Int J Environ Res Public Health.* 2012;9(12):4732-43. doi:10.3390/ijerph9124732
66. Eid HM, Ouchfoun M, Brault A, Vallerand D, Musallam L, Arnason JT, Haddad PS. Lingonberry (*Vaccinium vitis-idaea* L.) Exhibits Antidiabetic Activities in a Mouse Model of Diet-Induced Obesity. *Evid Based Complement Alternat Med.* 2014;2014:645812. doi:10.1155/2014/645812
67. Eid HM, Martineau LC, Saleem A, Muhammad A, Vallerand D, Benhaddou-Andaloussi A, et al. Stimulation of AMP-activated protein kinase and enhancement of basal glucose uptake in muscle cells by quercetin and quercetin glycosides, active principles of the antidiabetic medicinal plant *Vaccinium vitis-idaea*. *Mol Nutr Food Res.* 2010;54(7):991-1003. doi:10.1002/mnfr.200900218
68. Reichert KP, Schetinger MRC, Gutierrez JM, Pelinson LP, Stefanello N, Dalenogare DP, et al. Lingonberry extract provides neuroprotection by regulating the purinergic system and reducing oxidative stress in diabetic rats. *Mol Nutr Food Res.* 2018;62(16):e1800050. doi:10.1002/mnfr.201800050
69. Linderborg KM, Järvinen R, Lehtonen HM, Viitanen M, Kallio HP. The fiber and/or polyphenols present in lingonberries null the glycemic effect of the sugars present in the berries when consumed together with added glucose in healthy human volunteers. *Nutr Res.* 2012;32(7):471-8. doi:10.1016/j.nutres.2012.06.004
70. Quek R, Henry CJ. Influence of polyphenols from lingonberry, cranberry, and red grape on in vitro digestibility of rice. *Int J Food Sci Nutr.* 2015;66(4):378-82. doi:10.3109/09637486.2015.1042849
71. Rokka S, Ketoja E, Järvenpää E, Tahvonen R. The glycaemic and C-peptide responses of foods rich in dietary fibre from oat, buckwheat and lingonberry. *Int J Food Sci Nutr.* 2013;64(5):528-34. doi:10.3109/09637486.2013.763914
72. Törrönen R, Kolehmainen M, Sarkkinen E, Mykkänen

H, Niskanen L. Postprandial glucose, insulin, and free fatty acid responses to sucrose consumed with blackcurrants and lingonberries in healthy women. *Am J Clin Nutr.* 2012;96(3):527-33. doi:[10.3945/ajcn.112.042184](https://doi.org/10.3945/ajcn.112.042184)

73. Törrönen R, Kolehmainen M, Sarkkinen E, Poutanen K, Mykkänen H, Niskanen L. Berries reduce postprandial insulin responses to wheat and rye breads in healthy women. *J Nutr.* 2013;143(4):430-6. doi:[10.3945/jn.112.169771](https://doi.org/10.3945/jn.112.169771)