



In Cooperation with the University of Arizona, School of Natural Resources

Vascular Plant and Vertebrate Inventory of **Fort Bowie National Historic Site**



Southwest Biological Science Center Open-File Report 2005-1167 February 2007

U.S. Department of the Interior U.S. Geological Survey National Park Service







In cooperation with the University of Arizona, School of Natural Resources

Vascular Plant and Vertebrate Inventory of Fort Bowie National Historic Site

By Brian F. Powell, Cecilia A. Schmidt, and William L. Halvorson

Open-File Report 2005-1167

December 2006



USGS Southwest Biological Science Center Sonoran Desert Research Station University of Arizona School of Natural Resources 125 Biological Sciences East Tucson, Arizona 85721

U.S. Department of the Interior U.S. Geological Survey National Park Service

U.S. Department of the Interior DIRK KEMPTHORNE, Secretary

U.S. Geological Survey

Mark Myers, Director

U.S. Geological Survey, Reston, Virginia: 2006

For product and ordering information:

World Wide Web: http://www.usgs.gov/pubprod

Telephone: 1-888-ASK-USGS

For more information on the USGS-the Federal source for science about the Earth, its natural and living resources, natural hazards, and the environment:

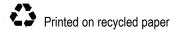
World Wide Web:http://www.usgs.gov Telephone: 1-888-ASK-USGS

Suggested Citation

Powell, B. F, C. A. Schmidt, and W. L. Halvorson. 2006. Vascular Plant and Vertebrate Inventory of Fort Bowie National Historic Site. USGS Open-File Report 2005-1167. U.S. Geological Survey, Southwest Biological Science Center, Sonoran Desert Research Station, University of Arizona, Tucson, AZ.

Cover photo: Ruins of the original military fort, Fort Bowie National Historic Site, Arizona. Photograph by Brian Powell.

Any use of trade, product, or firm names is for descriptive purposes only and does not imply endorsement by the U.S. Government.



Authors

Brian F. Powell & Cecilia Schmidt School of Natural Resources 125 Biological Sciences East, Building 43 The University of Arizona Tucson, AZ 85721

William L. Halvorson USGS SBSC Sonoran Desert Research Station 125 Biological Sciences East, Building 43 The University of Arizona Tucson, AZ 85721

U.S. Geological Survey SBSC Sonoran Desert Research Station Personnel

Charles van Riper III, Station Leader William L. Halvorson, Research Ecologist Cecil R. Schwalbe, Ecologist Michael R. Kunzmann, Ecologist (Emeritus) Kathryn Thomas, Ecologist Pamela Nagler, Physical Scientist

Phil Rosen, Ecologist

Charles van Riper III

Program and Expertise Areas of USGS and UA Personnel

Administration & Outreach
Cori Dolan
Dennis Suhre
Jennifer Meador
Wendy Parrish
Emily Sherbrooke
Fire Management
Dennis Suhre
Cori Dolan
James Feldmann
Bill Halvorson

Invasive Species Research
Avian Ecology
Patricia Guertin
Claire Crow
Jim Malusa
Glenn Johnson
Phil Rosen
Chris O'Brien
Cecil Schwalbe
Brian Powell
Brent Sigafus
Charles van Riper III
Dennis Suhre

Data Management

Brent Sigafus Inventory & Monitoring
Patricia Guertin

Ecology of Amphibians & Reptiles

Kevin Baker

Cristina Jones

Dave Prival

Phil Rosen

Pattleta Sterim

Bill Halvorson

Pamela Nagler

Brian Powell

Cecilia Schmidt

Cecil Schwalbe Vegetation Mapping & Ecology

Brent Sigafus Patricia Guertin
Bill Halvorson
Jim Malusa

Kathryn Thomas

USGS Southwest Biological Science Center http://sbsc.wr.usgs.gov USGS Southwest Biological Science Center, Sonoran Desert Research Station http://sbsc.wr.usgs.gov/sdrs

Kathryn Thomas

Table of Contents

Dedication	viii
Acknowledgements	ix
Executive Summary	xi
Chapter 1: Introduction to the Biological Inventories	1
Project Overview	1
Report Format and Data Organization	2
Verification and Assessment of Results	3
Technical Concepts	4
Chapter 2: Park Overview	5
Park Area and History	5
Natural Resources Overview	5
Natural Resource Management Issues	8
Chapter 3: Plant Inventory	11
Previous Research.	11
Methods	11
Results and Discussion	
Inventory Completeness	12
Chapter 4: Bird Inventory	13
Previous Bird Research	13
Methods	13
Results	19
Inventory Completeness	21
Discussion	
Chapter 5: Mammal Inventory	27
Previous Research	
Methods	
Results and Discussion	
Chapter 6: Management Implications	31
Chapter 7: Additional Inventories	
•	
Chantay 9. Litayatuwa Citad	25

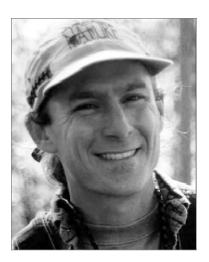
List of Tables

Table 1.	Summary results of plant and vertebrate inventories at Fort Bowle NHS, 2002 to 2004	X1
Table 1.1.	Museums that were queried in 1998 for vertebrate voucher specimens with "Arizona" and "Fort Bowie National Historic Site" in the collection location.	3
Table 2.1.	Average monthly climate data for Bowie, Arizona, 1899–2004.	5
Table 4.1.	Mean density (ha) of the most common tree species at each station along the two VCP transects, Fort Bowie NHS, 2004	15
Table 4.2.	Summary of bird survey effort, Fort Bowie NHS, 2002–2004.	15
Table 4.3.	Total number of observations (Total obs.) and relative abundance (mean \pm SE) of birds during VCP surveys, Siphon Canyon transect, Fort Bowie NHS, 2003 and 2004	20
Table 4.4.	Total number of observations (Total obs.) and relative abundance (mean \pm SE) of birds during VCP surveys, Butterfield transect, Fort Bowie NHS, 2004	22
Table 4.5.	Relative abundance of birds observed during line-transect surveys, Fort Bowie NHS, 2002 and 2003	23
Table 4.6.	Number of observations by breeding behavior for birds, Fort Bowie NHS, 2003 and 2004	24
Table 5.1.	Summary of Trailmaster camera effort, Fort Bowie NHS, 2002 and 2003	29
Table 5.2.	Number of photographs of animals, by Trailmaster camera site, Fort Bowie NHS, 2002 and 2003	29
List of Fig	jures	
Figure 2.1.	Location of Fort Bowie NHS in southern Arizona.	6
Figure 2.2.	Aerial photograph of Fort Bowie NHS showing some of the major features	7
Figure 2.3.	Comparison of monthly weather data during the time of the inventory (2002–2004) compared to the long-term mean (1899–2004), Bowie, Arizona.	8
Figure 4.1.	Photographs taken from bird survey stations, Fort Bowie NHS.	14
Figure 4.2.	Locations of bird surveys, Fort Bowie NHS, 2002–2004.	16
Figure 4.3.	Species accumulation curves, by survey type, for the UA inventory effort, Fort Bowie NHS, 2002–2004	25
Figure 5.1.	Location of Trailmaster camera sites, Fort Bowie NHS, 2002–2003.	27
Figure 5.2.	Diagram of infrared-triggered camera (Trailmaster) set-up.	28

List of Appendices

Appendix A.	Plant species that were observed or collected at Fort Bowie NHS by the University of Arizona inventory personnel (UA; 2002 and 2003) and other studies:	41
Appendix B.	Amphibians and reptiles observed or documented at Fort Bowie NHS by University of Arizona (UA) personnel, Swann et al. (2001; Swann), and Lowe and Johnson (1976; L&J) and from voucher specimens reported in Appendix E (AE).	61
Appendix C.	Number of observations of bird species, by detection type, at Fort Bowie NHS by University of Arizona (UA) inventory personnel, 2002–2004.	63
Appendix D.	Mammals recorded at Fort Bowie NHS by University of Arizona inventory personnel (UA) and/or documented/observed by other researchers or located in the UA mammal collection.	70
Appendix E.	List of voucher specimens collected from Fort Bowie NHS	72
Appendix F.	Summary of vegetation characteristics measured at bird survey stations, Fort Bowie NHS, 2004.	74
Appendix G.	Most abundant bird species at each transect and season based on data published in Russell and Johnson (1976) and the UA inventory.	78
Appendix H.	Number of Trailmaster photos and total number of individuals of each species	80
Appendix I.	Photographic vouchers taken by University of Arizona inventory personnel, Fort Bowie NHS 2002–2004.	80

Dedication



Eric Wells Albrecht 1970-2004

This report, as others in this series, is dedicated to Eric's life and work; he was an extraordinary ecologist, community member, father, and partner. Eric was co-coordinator of the University of Arizona (UA) biological inventory and monitoring program from 2002 until his sudden and unexpected death on September 20, 2004. Eric was near completion of his MS degree in Wildlife Conservation from the UA, which was awarded posthumously in November 2004. In his last year, Eric spearheaded projects to investigate the efficiency of current monitoring programs; he was passionate about using the best available information to guide vertebrate monitoring efforts in the region. He is survived by his partner, Kathy Moore, and their two young children, Elizabeth and Zachary. We hope that the lives of his children will be enriched by Eric's hard work on behalf of the national parks in the Sonoran Desert Network.

Acknowledgements

Thanks to Fort Bowie NHS staff members Larry Ludwig and Barbara Herman-Reese for their on-site support and Superintendent Alan Whalon, Chief of Resources Carrie Dennett, Biological Technician Ruth Olsen, and all the staff at Chiricahua National Monument for their administrative support of our program. This project resulted from the collaboration of many people at the U.S. Geological Survey (USGS), University of Arizona (UA) and the National Park Service (NPS), and was facilitated by the Desert Southwest and Colorado Plateau Cooperative Ecosystem Studies Units (CESUs). The project was funded by the National Park Service Natural Resource Challenge Program. The Southern Arizona Office of NPS facilitated development of the original study plan that led directly to initiation of this project. Andy Hubbard, Network Coordinator of the Sonoran Desert Network (SDN) Inventory and Monitoring (I&M) program, has been a strong and convincing advocate for continuing the role of the USGS/UA Inventory program in the I&M program. Kathy Davis at Tuzigoot and Montezuma Castle national monuments played an instrumental role in this project by providing important early initiative. Larry Norris at the Desert Southwest CESU has provided strong support for our program and spent considerable time and effort providing clear and timely administrative assistance. Matt Goode, Don Swann, and Dale Turner provided much of the early planning for this project; we are indebted to their vision. Special thanks to Lisa Carder and Kathleen Docherty for their years of hard work on all aspects of the project. Eric Albrecht, to whom this report is dedicated, was an outstanding spokesperson and leader of the program; he was an invaluable member of the team and his contributions are sorely missed.

We thank a core group of dedicated field biologists who collected a wealth of data at Fort Bowie NHS: James MacAdam, and Meg Quinn (plants); Eric Albrecht, Gabe Martinez, and Janine McCabe (birds); and Patina Thompson (mammals). Barbara Herman-Reese provided Trailmaster data. We are appreciative of the following people, whose work in the office made the field effort more successful: Debbie Angell, Jennifer Brodsky, Brian Cornelius, Taylor Edwards, Carianne Funicelli, Marina Hernandez, Colleen McClain, Heather McClaren, Lindsay Norpel, Ryan Reese, Jill Rubio, Brent Sigafus, Taffy Sterpka, Jenny Treiber, and Alesha Williams.

Additional administrative support was provided by Valery Catt at the USGS Sonoran Desert Research Station and Terri Rice, Andy Honaman, Jenny Ferry, and especially Cecily Westphal of the School of Natural Resources at the UA. Technical support was graciously given by the following experts: Dan Austin, Michael Chamberland, Phil Jenkins, and Charlotte and John Reeder at the UA Herbarium; Tom Huels of the UA ornithology collection. Thanks to Sharon Megdal and Peter Wierenga, the current and former directors, respectively, of the UA Water Resources Research Center, and all their staff.

We received helpful reviews of an earlier version of this report by: Carrie Dennett, Dennis Fenn, Michele Girard, Andy Hubbard, Larry Laing, Theresa Mau-Crimmons, Larry Norris, Ruth Olsen, and Alan Whalon. All mistakes or omissions are the responsibility of the authors.



Executive Summary

This report summarizes results of an inventory of plants, birds, and mammals of Fort Bowie National Historic Site (NHS) in southeastern Arizona. Surveys at the park were part of a larger effort to inventory vascular plants and vertebrates in eight National Park Service (NPS) units in Arizona and New Mexico. Our inventory efforts build on past research for the park; included in this report is the most comprehensive synthesis of species lists from past studies of plants and vertebrates. Though we did not survey specifically for them, we also include a species lists of amphibians and reptiles that have been observed or documented at the park.

For a park of its size (405 ha), Fort Bowie NHS has extraordinary species richness, especially for plants (638 species) and mammals (57 species). This diversity results from its geographic location at the junction of three biogeographical provinces, its diverse geology, and the presence of year-round water from an active spring.

We found 45 new plant and vertebrate species for the park (Table 1) including:

- Plants: two species representing new families and nine species representing new genera, including one new genus (Euphorbia) represented by three species;
- Birds: zone-tailed hawk and common ground dove;
- Mammals: domestic dog and hooded skunk.

Based on a review of past studies and our own work, we consider the inventories of plants and vertebrates to be 90% complete. With baseline inventories at Fort Bowie NHS among the most complete of the 11 parks in the Sonoran Desert Network, park staff are now in an excellent position to monitor changes in these resources.

Table 1. Summary results of plant and vertebrate inventories at Fort Bowie NHS, 2002 to 2004.

		UA Effort		
Taxonomic group	Number of species recorded	Number of non-native species	Number of new species added to park lista	Total number of species on park listb
Plants	193	13	33	638
Amphibians and Reptiles	8	0	0	40
Birds	109	0	6	189
Mammals	14	1	2	59
Totals	324	14	41	926

^aSpecies that had not been observed or documented by previous studies.

^bFrom all sources (see Appendices A-D for complete lists).



Chapter 1: Introduction to the Biological Inventories

Project Overview

Inventory: A point-in-time effort to document the resources present in an area.

In the early 1990s, responding to criticism that it lacked basic knowledge of natural resources within parks, the National Park Service (NPS) initiated the Inventory and Monitoring Program (NPS 1992). The purpose of the program is to increase scientific research in NPS units and to detect long-term changes in biological resources (NPS 1992). At the time of the program's inception, basic biological information, including lists of plants and animals, were absent or incomplete for many parks (Stohlgren et al. 1995).

Species inventories have both direct and indirect value for management of the park. Species lists facilitate resource interpretation and visitor appreciation of natural resources. Knowledge of which species are present, particularly sensitive species, and where they occur is critical for making management decisions (e.g., locating new facilities). Inventories are also a cornerstone of long-term monitoring. Thorough biological inventories provide a basis for choosing parameters to monitor and can provide initial data (i.e., a baseline) for monitoring ecological populations and communities. Inventories can also test sampling strategies, field methods, data collection protocols, and provide estimates of variation that are essential in prospective power analysis.

Goals

The purpose of this study was to complete basic inventories for vascular plants, birds, and mammals at Fort Bowie National Historic Site (NHS). This effort was part of a larger biological inventory of eight NPS units in southern Arizona and southwestern New Mexico (Davis and Halvorson 2000, Powell et al. 2003, 2004, 2005a). The results presented in this report supersede those reported by Powell et al. (2003, 2004, and 2005b).

The goals of our biological inventory of Fort Bowie NHS were to:

- 1. Conduct field surveys to bring the current species lists for vascular plants, birds, and mammals to at least 90% of the species expected to occur at the park.
- 2. Use repeatable sampling designs and survey methods (when appropriate) that allow estimation of parameters of interest with associated estimates of precision.
- 3. Compile historic occurrence data for plants and vertebrates (including amphibians and reptiles) from three sources: museum records (specimen vouchers), previous studies, and park records.
- 4. Create resources useful to park managers, including detailed species lists, maps of study sites, and high-quality digital images for use in resource interpretation and education.

The bulk of our effort addressed goals number 1 and 2. To maximize efficiency (i.e., the number of species recorded by effort) we used field techniques designed to detect multiple species. We did not undertake single-species surveys for threatened or endangered species.

Administrative History

The original study plan for this project was developed, and an inventory of one Sonoran Desert Network (SDN) park (Tumacácori National Historical Park) was completed, through a cooperative agreement between NPS, UA, and the USGS. This project comprises biological inventories for seven additional parks and was funded through Task Agreements UAZ-03, -05, -06, and -07 (under Colorado Plateau CESU cooperative agreement number 1200-99-009). The National Park Service thereafter obligated additional funds for administration, management, and technical oversight of the biological inventories through the Colorado Plateau CESU (UAZ-07) and the Desert Southwest CESU (cooperative agreement number CA 1248-00-002, reference UAZ39, -77, -87, -97, and 128).

Report Format and Data Organization

This report includes summaries and analyses of data related to vascular plants, birds, and mammals collected from 2002 to 2004 at Fort Bowie NHS. This report is intended to be useful in internal planning processes and outreach and education, and as such we strive to make it relevant, easy to read, and well organized. We report only common names (listed in phylogenetic sequence) unless the species is not listed later in an appendix; in this case we present both common and scientific names. For each taxonomic group we include an appendix of all species that we recorded at the park (Appendices A, C, and D). Although we did not survey specifically for amphibians and reptiles, we made a few incidental observations and we include a list of species observed or documented by others (Appendix B). Species lists are in phylogenetic sequence and include taxonomic order, family, genus, species, subspecies or varieties (if applicable) and common name. Scientific and common names used throughout this document are current according to accepted authorities for each group: Integrated Taxonomic Information System (ITIS 2004) and the PLANTS database (USDA 2004; including designation of plants as "non-native") for plants; Stebbins (2003) for amphibians and reptiles; American Ornithologist Union (AOU 1998, 2003) for birds; and Baker et al. (2003) for mammals. To maintain consistency throughout the document, we do not capitalize the first letter of common names unless they are proper names. In this document we use the International System of Units for measurements.

Previous Amphibian and Reptile Inventories

We did not survey for amphibians and reptiles because there have been two thorough inventories for them: the first by Lowe and Johnson (1976) and more recently by Swann et al. (2001). In addition to re-surveying Lowe and Johnson's line-transects, Swann et al. set up long-term monitoring plots, surveyed road transects, and produced an annotated species list. In total, these studies found one non-native species (American bullfrog), and 31 reptile species (16 lizards, one turtle, and 14 snakes) including Texas horned lizard, a federally listed species of concern. The most notable species is now-extirpated Chiricahua

leopard frog (a federally threatened species; HDMS 2004). In this report, we list the species found by these studies and species that our crews found while conducting surveys for other taxa (Appendix B). We also summarize specimens located in the University of Arizona herpetology collection (Appendix E), and a few photographic vouchers that we took during our course of other field research (Appendix I).

Spatial Data

Most spatial data are geographically referenced to facilitate mapping of study plots and locations of plants or animals. Coordinate storage is the Universal Transverse Mercator (UTM) projection, using North American datum 1983 (NAD 83), Zone 12. We recorded UTM coordinates using hand-held Garmin eMap® Global Positioning System (GPS) units (Garmin International Incorporated, Olathe, KS; horizontal accuracy is about 10–30 m). For each taxon-specific chapter of this document we mapped the location of all plots or stations overlaid on Digital Orthophoto Quarter Quads (DOQQ; produced by the U.S. Geological Survey). All study-site coordinates are stored at the same locations as for data archiving (below).

Species Conservation Designations

We indicate species conservation designations by the following agencies: U.S. Fish and Wildlife Service (responsible for administering the Endangered Species Act), Bureau of Land Management, U.S.D.A. Forest Service, Arizona Game and Fish Department, and Partners in Flight (a partnership of federal, state and local governments, non-governmental organizations, and private industry).

Databases and Data Archiving

We entered field data into taxon-specific databases (Microsoft Access version 97) and checked all data for transcription errors. From these databases we reproduced copies of the original field datasheets using the "Report" function in Access. The output looks similar to the original datasheets but data are easier to read. The databases, printouts, and other data such as digital photographs and GIS layers will be distributed to the park and to the University of Arizona, Special Collections (Main Library, Tucson). Original copies of all datasheets will be given to the NPS SDN I&M program office in

Tucson and may be archived at another location (most likely Western Archaeological Conservation Center, Tucson; Andy Hubbard, *pers. comm.*). This redundancy in data archiving is to ensure that these valuable data are never lost. Along with the archived data we will include copies of the original datasheets and a guide to filling them out. This information, in conjunction with the text of this report, should enable future researchers to repeat our work.

Verification and Assessment of Results

Photographic Vouchers

Whenever possible we documented vertebrate species with analog color photographs. Many of these photos show detail on coloration or other characteristics of visual appearance, and they may serve as educational tools for the park staff and visitors. Photographs will be archived with other data as described above.

Specimen Vouchers

With proper documentation, specimen vouchers are the most indisputable form of evidence of species occurrence. For plants, we searched the University of Arizona Herbarium for existing specimens from Fort Bowie NHS (see Appendix A for results), but we collected herbarium

specimens whenever flowers or fruit were present on plants in the field (Appendix A). All specimens that we collected were accessioned into the University of Arizona Herbarium. Although we did not collect specimen vouchers for vertebrates, we searched for existing vouchers from Fort Bowie NHS in records from 23 natural history museums (Table 1.1 see Appendix E for results).

Assessing Inventory Completeness

We evaluated inventory completeness by (1) examining the rate at which new species were recorded in successive surveys (i.e., species accumulation curves; Hayek and Buzas 1997) and (2) by comparing the list of species we recorded with a list of species likely to be present based on previous research and/or expert opinion. For all species accumulation curves, we randomized the order of the sampling periods to break up clusters of new detections that resulted from temporal conditions (e.g., monsoon initiation) independent of cumulative effort. We used the computer program Species Richness and Diversity III (Pisces Conservation Ltd., IRC House, Pennington, Lymington, UK) to calculate species accumulation curves where the order of samples was shuffled >10 times and the average is plotted, thereby smoothing the curve.

Table 1.1. Museums that were queried in 1998 for vertebrate specimen vouchers with "Arizona" and "Fort Bowie National Historic Site" in the collection location. Collections in bold-faced type had specimens from Fort Bowie NHS. See Appendix E for results.

Collection	Collection cont.
Chicago Academy of Sciences	Peabody Museum, Yale University
Cincinnati Museum of Natural History & Science	Saguaro National Park
Cornell Vertebrate Collections, Cornell University	Strecker Museum, Baylor University, Waco
George Mason University (Fairfax, VA)	Texas Cooperative Wildlife Collection
Marjorie Barrick Museum, University of Nevada-Las Vegas	University of Arizona
Michigan State University Museum (East Lansing)	University of Texas, Arlington
Milwaukee Public Museum	University of Illinois, Champaign-Urbana
Museum of Texas Tech University	University of Colorado Museum
Museum of Vertebrate Zoology, University of California, Berkeley	Walnut Canyon National Monument, Arizona
Museum of Life Sciences, Louisiana State University, Shreveport	Western Archaeological and Conservation Center, Tucson
North Carolina State Museum of Natural Sciences	Wupatki National Monument, Arizona
Oklahoma Museum of Natural History, Norman	

Technical Concepts

Sampling Design

Sampling design is the process of selecting sample units from a population or area of interest (for a review, see Thompson [1992]). Random samples allow inference to the larger population from which those samples were drawn, and estimate the true value of a parameter. Nonrandom samples are less likely to be representative of the entire population, because the sample may (intentionally or not) be biased toward a particular characteristic, perhaps of interest or convenience.

We briefly address sampling design in each chapter. Our surveys were not randomly located because we were more interested in detecting the maximum number of species than in providing inference to a larger area. Thus, abundance estimates (relative abundance, useful as an index to true abundance) detailed in this report may be biased because we surveyed in areas likely to have high species richness; however, the nature or extent of that bias is difficult to characterize or quantify. If population estimates were a higher priority, avoiding this potential bias would have greater importance.

Estimates of Abundance

Estimating population size is a common goal of biologists, generally motivated by the desire to reduce (e.g., pest species), increase (e.g., endangered species), maintain (e.g., game species) or monitor (e.g., indicator species) population size. Our surveys at Fort Bowie NHS were generally focused on detecting species rather than estimating population size. In many cases, however, we present estimates of "relative"

abundance" by species, which is an index to population size; we calculate it as the number of individuals of a species recorded, scaled by survey effort. Some researchers (particularly plant, marine, and invertebrate ecologists) prefer to scale such frequency counts by the number of observations of other species, which provides a measure of community dominance; abundance relative to other species present. If we completed multiple surveys in comparable areas (e.g., anywhere within Fort Bowie NHS), we included a measure of precision (usually standard error) with the mean of those survey results.

Indices of abundance are presumed to correlate with true population size but do not typically attempt to account for variation in detectability among different species or groups of species under different conditions. Metrics (rather than indices) of abundance do consider variation in detection probability, and these include density (number of individuals per unit area; e.g., two black-throated sparrow per hectare of semi-desert grassland), and absolute abundance (population size; e.g., 28 black-throated sparrow at Fort Bowie NHS). These latter techniques are beyond the scope of our research. While it is true that indices to abundance have often been criticized (and with good reason. c.f. Anderson 2001), the abundance information that we present in this report is used to characterize the commonness of different species rather than to quantify changes in abundance through space (e.g., habitat-use studies) or time (e.g., monitoring). As such, relative abundance estimates are more useful than (1) detectability-adjusted estimates of density for only a few species or (2) raw count data for all species without scaling counts by search effort. For a review of methods used to estimate abundance. see Lancia et al. (1996).

Chapter 2: Park Overview

Park Area and History

Fort Bowie National Historic Site (NHS) is located in southeastern Arizona approximately 22 km south of the town of Bowie, Arizona (Fig. 2.1). The park lies in Apache Pass between the Chiricahua Mountains to the south and the Dos Cabezas Mountains to the north.

Fort Bowie NHS was established in 1964 to preserve the historic dwellings of Fort Bowie, a 19th century U.S. military outpost, prehistoric structures associated with Apache Spring, and a portion of the Butterfield Overland Trail and Station (NPS 2001). The park also commemorates the fort's soldiers, the Chiricahua Apaches, and the settlement of the west (NPS 2001). The park, which encompasses 405 ha, is administered by Chiricahua National Monument. Recent annual visitation is approximately 9,500 (NPS 2005).

Natural Resources Overview

Physiography, Geology and Soils

Fort Bowie NHS is located in Apache Pass, which separates the San Simon and the Sulphur Springs Valleys. Elevation at the park ranges from 1,400 m to 1,600 m. The park contains three riparian areas: Siphon Canyon, Cutoff Canyon, and Willow Gulch. The geology of the park is characterized by the Apache Pass Fault made of Pennsylvanian and Cretaceous limestone on top of Precambrian granite (Denney and Peacock 2000). In some areas small fan terraces have formed from the granitic alluvium deposited by drainageways. For a complete soil survey, see Denny and Peacock (2000).

Hydrology

There are no perennial-flowing streams, but two springs, Apache and Mine Tunnel, are found within the park. Three other springs are outside, but near to the boundary of the park: Siphon, Bear, and Goodwin springs. Currently, half of the water flow of Apache Springs can be diverted to adjacent lands for use in cattle tanks, though this is rarely realized (Alan Whalon, *pers. comm.*). Siphon Canyon and Willow Gulch have ephemeral flow during summer (monsoonal) and winter rains (NPS 2001).

Climate

Fort Bowie NHS experiences an annual bimodal pattern of precipitation that is characterized by heavy summer (monsoon) storms from the Gulf of Mexico, and less intense frontal systems from the Pacific Ocean in the winter. On average, approximately one-half of the annual precipitation falls from July through September (Table 2.1; WRCC 2005). The area's hot season occurs from April through October; maximum temperatures in July can exceed 40°C. Winter temperatures dip below freezing and snow is occasional. Based on data from Bowie, Arizona (the closest climate station), average annual precipitation totals during the course of our study were significantly lower than the long-term mean (27.3 cm) in 2002 (21.5 cm) and 2003 (19.3 cm), but similar for 2004 (29.3 cm through October 2004) (Fig 2.3; WRCC 2005). Average annual temperatures during all years of our survey were above the long-term mean of 17.7°C (17.8°C in 2002, 18.3°C in 2003, and 18.9°C through October 2004).

Table 2.1. Average monthly climate data for Bowie, Arizona, 1899–2004. Data from WRCC (2005).

		Month											
Characteristic	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Annual
Maximum temperature (°C)	16.2	18.7	22.2	26.8	31.7	36.8	36.8	35.3	33.2	27.8	20.7	15.7	26.8
Minimum temperature (°C)	-0.7	1.2	3.7	7.0	11.5	16.6	19.7	18.7	15.2	9.0	2.6	-0.5	8.7
Precipitation (cm)	2.1	2.0	1.5	0.7	0.6	0.9	5.2	5.3	2.6	2.2	1.5	2.4	2.3

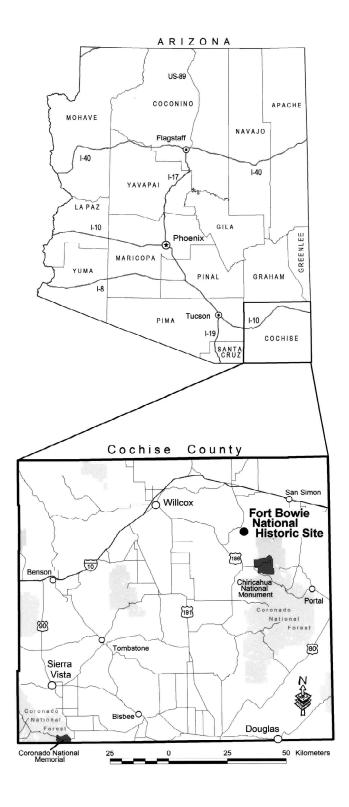


Figure 2.1. Location of Fort Bowie NHS in southern Arizona.

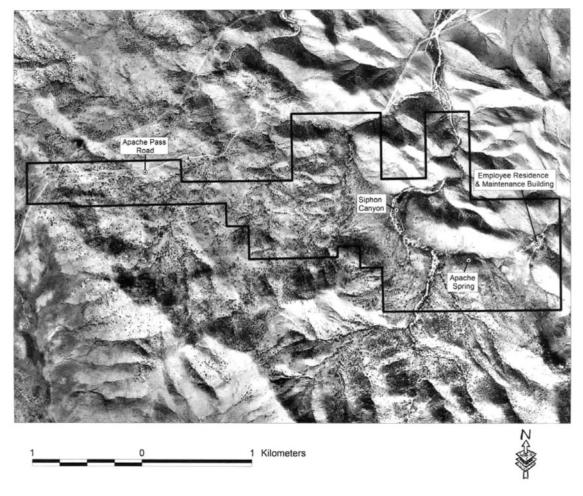


Figure 2.2. Aerial photograph of Fort Bowie NHS showing some of the major features.

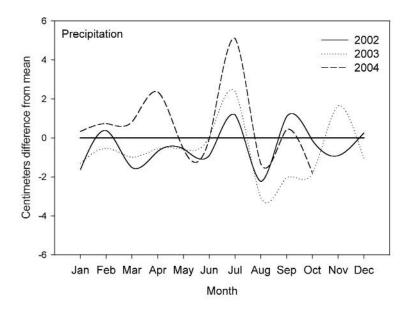
Vegetation

Fort Bowie NHS contains four major vegetation communities (based on Warren et al. [1992]):

- Madrean Evergreen Forest and Woodland containing Emory Oak–Pointleaf Manzanita— Beargrass Association, Emory Oak–Turpentine bush–Grama Grass Association, Scrub Oak–Bear Grass– Oneseed Juniper Association and Desert Deerbrush–Alder-leaf Mountainmohogany–Desert Sumac Association;
- Semi-desert Grassland containing Velvet
 Mesquite—Turpentine bush—Burroweed
 Association, Turpentine bush—Fairy
 Duster—Ocotillo Association,
 Ocotillo—Mariola—Grama Grass
 Association, Velvet Mesquite—Desert
 Sumac—Snakeweed Associations and
 Russian Thistle—Snakeweed—Mixed Grass
 Association;

- Chihuahuan Desertscrub containing Creosote-bush–Velvet Mesquite–Mariola Association; and
- Interior Southwestern Riparian
 Deciduous Forest and Woodland
 containing Arizona Walnut—Netleaf
 Hackberry—Gum Bumelia Association.

Historic photographs from the era of settlement show that juniper and oak trees were not abundant in the area near the fort, presumably because the trees had been cut for firewood (Warren et al. 1992). More recently, mesquite trees have invaded the semi-desert grassland areas of the park, most likely as a result of fire suppression and cattle grazing (NPS 2000b). Park personnel have removed many of the large mesquite trees that encroached into the area near the fort ruins. (NPS 2001).



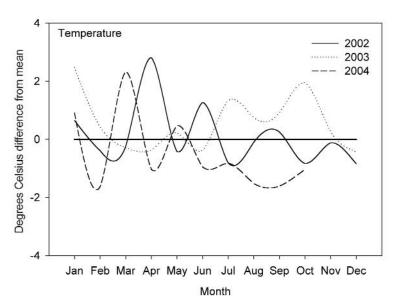


Figure 2.3. Comparison of monthly weather data during the time of the inventory (2002–2004) compared to the long-term mean (1899–2004; thick solid line in both figures), Bowie, Arizona. Data from WRCC (2005).

Natural Resource Management Issues

Cattle Grazing

Two cattle grazing allotments at the park were recently retired, though trespass of cattle onto the park is common. Livestock grazing has degraded an estimated 80% of streams and riparian ecosystems in the western United States through loss of vegetation, stream-bank erosion, soil compaction, flooding, and water pollution (Bahre

1991, BLM 1994, Fleischner 1994). Livestock grazing can also increase the number and extent of non-native plants (Belsky et al. 1999) and can negatively affect wildlife through habitat modification and competition for resources (Saab et al. 1995). At Fort Bowie, cattle have decreased grass cover, damaged historic artifacts, and helped to spread woody plants, such as mesquite, into what was once predominantly semi-desert grassland (NPS 2001).

Non-Native Species

Lehmann lovegrass and other non-native grasses are the most important non-native species issue at the park. The change in species composition in the semi-desert grassland to a community dominated by Lehmann lovegrass, in particular, can alter the fire regime of the area by supporting higher fire frequencies, thereby leading to other changes in vegetation composition and structure including a loss of species richness (Anable et al. 1992).

The presence of American bullfrogs is another important non-native species management issue. Bullfrogs are native to eastern North America but have been introduced throughout the western U.S. for food production and sport (Stebbins 2003). American bullfrog adults and tadpoles are voracious predators (Kiesecker and Blaustein 1997) and are thought to be partially responsible for the decline of many native reptiles (Schwalbe and Rosen 1988) and amphibians (particularly other Ranid frogs; Hayes and Jennings 1986, Lawler et al. 1999) in the southwest.

Undocumented Immigrants

Approximately 20 undocumented immigrants pass through the park each week (NPS 2003). In addition to compromising visitor safety, border crossers also adversely impact the natural resources; they have created trails (including into the fort ruins), damaged vegetation, and have left trash behind. These impacts affect water quality and wildlife movement patterns, though the extent of these impacts has not been established.

Adjacent Land Use

Due to its small size, the park is easily affected by land management practices (e.g., development, mining, grazing, and hunting) outside its boundaries. Currently there is a Buddhist retreat center being built to the north of the park, but because the remainder of the land surrounding the park is managed by the BLM, ex-urban housing development is unlikely to significantly affect the park's resources.

Chapter 3: Plant Inventory

Previous Research

Warren et al. (1992) completed the most thorough inventory of plants at Fort Bowie NHS (Appendix A). Most of the specimen vouchers reported by Warren et al. were collected by Marina Hoy from 1972 to 1977. The collection is comprised of 471 species and subspecies. Warren et al. reported that most of the specimens were located at the park with "duplicates of selected species at the University of Arizona". However, in a recent search of the UA collections, Halvorson (2003) did not find any specimens from this study. Bennett et al. (1996) compiled a species list for the Chiricahua Mountains and vicinity (including Fort Bowie NHS) from a variety of sources including: complete examination of herbaria at Chiricahua National Monument, Fort Bowie NHS, the Southwestern Research Station, and minimal examination of herbaria at the University of Arizona, Arizona State University, and New Mexico State University. In their annotated species list, Bennett et al. indicated if the plant was found at Fort Bowie NHS. Most of the species attributed to Fort Bowie NHS were from the collections made by Marina Hoy, therefore there is considerable overlap between the Bennett et al. list and that by Warren et al. (1992; Appendix A). Ruyle (2001) assessed range conditions at eight permanently marked transects: six in the park and two outside (but near) the park. He found ten species of plants new to the park though he did not, to our knowledge, collect specimens. His work remains the only plot-based plant research at the park. Halvorson and Guertin (2003) mapped the distribution of 22 non-native plant species in the park from the fall of 1999 to the spring of 2001. There are two other poorly documented plant species lists for the park. NPS (2000a) produced an annotated species list for Fort Bowie NHS. This list appears to be compiled from Warren et al. (1992) and Bennett et al. (1996) but these sources do not entirely make up the list (Appendix A). Finally, Hartman and Rottman (1998) compiled a checklist for the park but again, no documentation exists from their work.

Methods

To complete the species list for the park, we used "general botanizing" surveys, during which observers walked throughout the park (particularly around the parade grounds and along trails) and opportunistically collected and recorded plants. In addition to our own results, we present here the first synthesis of findings from past studies and collections.

Spatial Sampling Design

Our survey crews walked throughout the park on each visit. They did not record their search paths, but indicated the location of each collection.

General Botanizing

Field Methods

Whenever possible we collected at least one representative specimen (with reproductive structures) for each plant species that were thought to represent a new species for the park. When we collected a specimen we recorded flower color, associated dominant vegetation, date, collector name(s), and UTM coordinates. We pressed the specimens immediately upon collection. Specimens remained pressed for 2–3 weeks and were frozen for 48 hours to prevent infestation by insects and pathogens. We then mounted the specimens and accessioned them into the University of Arizona Herbarium.

Effort

We made nine day-long visits, typically with two observers, on 28 September and 5, 6, 9, 23, 29, and 31 October 2002 and on 6 and 9 May 2003.

Results and Discussion

We found 193 species and subspecies, including 33 new species for the park (Appendix A). Of these, we found two species representing new families for the park and nine species representing new genera for the park. Of particular note were three species of *Euphorbia*, a new genus for the park. Of the 33 species new for the park, only one was non-native.

Considering all available sources, there have been 638 species and subspecies recorded at the park, of which 38 (6%) are non-native (Appendix A). The percentage of non-native plants in the park's flora is low compared to other sites in southeastern Arizona (e.g., Burgess et al. 1991, Powell et al. 2005a). The number and extent of non-native plants may be buffered by the lack of roads through the park. Roads act as dispersal corridors for non-native plant species, which often thrive in the adjacent disturbed soils. Although the park has a low percentage of non-native species. the percent of area covered by them is greater. Halvorson and Guertin (2003) mapped the distribution of 22 species of non-native plants at the park. Lehmann lovegrass was the most widespread of the non-native species and other non-native species were especially prevalent around the visitor center. Ruyle (2001) established plots and his work remains the only study that is able to quantify the dominance and relative abundance of plants. Yet because of the diversity of vegetation communities and conditions in the park, Ruyle did not survey enough plots to address the dominance of non-native species; he found no nonnative species during his surveys, but clearly did not have plots in the semi-desert grasslands west of Siphon Canyon (near the cemetery) where Lehmann lovegrass dominates. In Chapter 6 we make recommendations for more plot-based survey work.

The number of species documented for the park far outnumbers the expected species richness (330), based on size and topographic relief of the park (Warren et al. 1992). The extraordinary species richness that has been found at the park is due to three main factors: geographic location of park, geologic faults separating different rock substrates, and the presence of permanent water.

Fort Bowie NHS lies at the juncture to four major biogeographical provinces: Madrean, Rocky Mountain, and Sonoran and Chihuahuan deserts. The Madrean biogeographic region in particular is well represented in the flora of the park, and this region has the highest plant species richness in

Arizona (Bowers and McLaughlin 1982). Other floristic influences are from the Great Plains and Chihuahuan regions (Warren et al. 1992).

The high species richness at the park is also the result of local influences from two faults that pass through the park: Apache Pass and Fort Bowie faults (Denny and Peacock 2000). These faults separate major geologic substrates including limestone, shale, sandstone, and granitic and metamorphic rock, all of which give rise to conditions that favor certain plant species. For example, Warren et al. (1992) noted that a number of species with primarily Chihuahuan desert distributions are found on limestone outcrops at Fort Bowie NHS.

Finally, the perennial Apache Spring gives rise to the third determinant of high species richness and vegetation communities at the park. The presence of water is responsible for the vegetation structure in the riparian area, which is in stark contrast to upland areas, and also is an important determinant of vertebrate species richness and abundance (see Chapters 4 and 5).

Inventory Completeness

Fort Bowie NHS has one of the most complete inventories for vascular plants in all of the Sonoran Desert Network parks. Based on our work and that by others, we believe that the inventory is likely 90% complete. We found 33 new species, a 5% increase in the number of plants for the park. Additional surveys, particularly during the late spring, will add more species to the park list, but considering our survey effort, we believe the percentage of new species found will not be significant (see Chapter 7 for additional information on more studies). However, of particular concern in all natural areas is the increase in the abundance and distribution of nonnative species. This will likely happen if Cochise County paves Apache Pass Road, thereby increasing the number of vehicles in the area (see Chapter 6).

Chapter 4: Bird Inventory

Previous Bird Research

The first comprehensive inventory of birds at the park was in 1975 and 1976 by Russell and Johnson (1976) who surveyed five transect routes multiple times in all seasons. They reported abundance of species based on the number of observations per hour of surveys. Although no original data exist from that study (Terry Johnson, pers. comm.), the transect routes were similar to those used by our survey crews and we therefore make gross comparisons between our two studies. Many of the observations of rare birds noted in that report are from Marina Hoy. Fischer (2002) compiled a list of bird species at the park based on: (1) field observations made in the early 1990s (Dan Fischer, pers. comm.) and (2) review of bird lists and specimens from the region. Typically we do not consider species lists credible forms of evidence of species occurrence in an area; often these lists are not well documented. However, the list by Fischer (2002) is well documented and is mostly based on his field experience. We therefore use it to create the species list for the park (Appendix C). In 2002 and 2003 there was a Monitoring Avian Productivity and Survivorship (MAPS; DeSante and O'Grady 2000) banding station at the park, which was operated by staff from the Southeastern Arizona Bird Observatory. Finally, we found specimen vouchers, representing 20 species, which were collected in 1893 and 1894 (Appendix E).

Methods

We surveyed for birds at Fort Bowie NHS in 2003 and 2004. We used four field methods: variable circular-plot (VCP) counts for diurnal birds during the breeding season, nocturnal surveys for owls and nightjars, line transects for birds in the non-breeding season, and incidental observations for all birds in all seasons. Although winter bird surveys were not included in the original study proposal (Davis and Halvorson 2000), we felt they were important in our effort to inventory birds at the park because many species that use the area during the fall and winter may not be present during spring and summer (breeding season) surveys. We concentrated most of our survey effort during the

breeding season because bird distribution is relatively uniform at this time (due to territoriality among most landbird species) (Bibby et al. 2000). Therefore, surveying during the breeding season increased our precision in estimating relative abundance and also enabled us to document breeding activity. Our survey period included peak spring migration times for most species, adding many migratory species to our list. We also sampled vegetation in the vicinity of VCP stations. Vegetation structure and plant species composition are important predictors of bird species richness or the presence of particular species (MacArthur and MacArthur 1961, Rice et al. 1984, Strong and Bock 1990, Powell and Steidl 2000).

In most cases we do not report observations that failed to determine species (e.g., "unknown woodpeckers"). Ravens are an exception. Both Chihuahuan and common ravens occur at the park and both species are difficult to differentiate unless they are viewed at a short range under certain conditions or if they are seen flying together (Bednarz and Raitt 2002). We were not able to positively determine the species for any raven sighting and therefore report all observations as "unknown raven."

Spatial Sampling Design and General Vegetation Characteristics

We subjectively placed the two VCP transects in areas that we believed would have the highest species richness (Siphon Canyon) and would be representative of the dominant vegetation at the park (Butterfield). Some sections of our survey locations correspond to those of Russell and Johnson (1976). The Siphon Canyon transect ran from Apache Spring to where the canyon exits the park (Fig. 4.1). The Butterfield transect began near the junction of the namesake trail and Siphon Canyon and ended near the western boundary of the park (Fig. 4.1; see also Fig. 4.2).

The Butterfield transect follows its namesake trail and it is more xeric than the Siphon Canyon transect. It is dominated by Emory oak and oneseed juniper (Table 4.1) with a wide variety of shrub such as turpentine bush, sotol, manzanita, agave, and yucca (Fig. 4.1). The Siphon Canyon



Figure 4.1. Photographs taken from bird survey stations, Fort Bowie NHS. Photographs A and B are looking east from Butterfield transect station numbers 8 and 6, respectively. Photographs C and D are looking north from Siphon Canyon station numbers 6 and 5, respectively. See Fig. 4.2 for location of stations.

transect has some areas of very dense vegetation, including netleaf hackberry, Arizona oak, and Utah juniper near Apache Spring (Table 4.1). Other riparian vegetation, including desert willow and Arizona walnut, is in the bottom of Siphon Canyon (Fig. 4.1). The upslope areas are similar to the Butterfield transect, but with few Emory oak (Table 4.1).

VCP Surveys Field Methods

We used the variable circular-plot method to survey for diurnally active birds during the breeding season (Reynolds et al. 1980, Buckland et al. 2001). Conceptually, these surveys are similar to traditional "point counts" (Ralph et. al 1995), during which an observer spends a standardized length of time at one location (i.e., station) and records all birds seen or heard and the distance to each bird or group of birds.

We surveyed the Siphon Canyon transect in 2003 and 2004 and the Butterfield transect in

2004 (Table 4.2). Each transect consisted of eight stations, each located a minimum of 250 m apart to maintain independence among observations at each station. We surveyed each year from mid April through early July, the period of peak breeding activity for most species in the area.

On each visit to a transect we alternated the order in which we surveyed stations to minimize bias by time of day and direction of travel. We did not survey when wind speed exceeded 15 km/h or when precipitation exceeded an intermittent drizzle. We began bird surveys approximately 30 minutes before sunrise and concluded no later than three hours after sunrise.

We recorded a number of environmental variables at the beginning of each transect: wind speed (Beaufort scale), presence and severity of rain (qualitative assessment), air temperature (°F), relative humidity (%), and cloud cover (%). After arriving at a station, we waited one minute before beginning the count to allow birds to resume their normal activities. We identified to species all birds

seen or heard during an eight-minute "active" period. For each detection we recorded distance in meters from the observer (measured with laser range finder when possible), time of detection (measured in one-minute intervals beginning at the start of the active period), and the sex and/or age class (adult or juvenile), if known. We did not measure distances to birds that were flying overhead, nor did we use techniques to attract birds (e.g., "pishing"). We made an effort to avoid double-counting individuals that had been recorded at previous stations. During the "passive" count

period (between the eight-minute counts), if we observed a species that had not been recorded previously at a station on that visit, we recorded its distance to the nearest station.

Effort

We surveyed the eight stations of the Siphon Canyon transect five times in 2003 and the Siphon Canyon and Butterfield transects (also eight stations) six times each in 2004 (Table 4.2). We visited each station for eight minutes on each visit.

Table 4.1. Mean density (ha) of the most common tree species at each station along the two VCP transects, Fort Bowie NHS, 2004. Data summarized from Appendix F. Numbers represent the number of individuals observed in the "tree" and "potential cavity-nesting" categories from point-quarter sampling. Species present on >4 stations are included in this summary. See Appendix A for scientific names.

		Species											
Transect	Station	netleaf hackberry	desert willow	Arizona walnut	oneseed juniper	Utah juniper	velvet mesquite	Arizona oak	Emory oak	desert sumac	gum belly		
Butterfield	1	0.7		2.4	0.7	0.7	8.7			2.7			
	2	6.9				6.9	20.8		14.4				
	3					1.9	1.8	1.9	25.9				
	4				4.1		10.1	1.7	23.1				
	5				2.8			3.4	7.4				
	6				11.3		3.8		14.0				
	7				3.5		2.6	1.8	11.5				
	8				6.0		1.5	1.6	3.7				
Siphon Can	yon 1	5.1		2.2	5.1	6.2	8.1	6.3			12.2		
	2	8.6		0.8	6.5		15.5	2.3		4.2	21.0		
	3	7.0	11.0	4.4			11.4		1.3	16.7			
	4	46.8	11.2	5.4			19.3			20.9	15.8		
	5	6.9	3.1	6.2	1.5		6.9				9.2		
	6	8.2	17.0	3.0	2.5		14.0			9.3	0.3		
	7	14.4		3.0	3.1	3.4	3.1			15.7	3.1		
	8	4.2		1.8			6.3	5.8			15.8		

Table 4.2. Summary of bird survey effort, Fort Bowie NHS, 2002–2004. Sample size (*n*; number of visits multiplied by number of stations) was used to calculate relative abundance for each transect and year.

Survey type	Transect name	Year(s)	Visits	Stations/ sections	n
VCP	Butterfield	2004	6	8	48
	Siphon Canyon	2003	5	8	40
		2004	6	8	48
Line transect	Coach	2002-2003	4	6	24
	Siphon Canyon	2002-2003	4	5-6	23
Nocturnal survey	Owl	2003	3	5	15
		2004	3	5	15

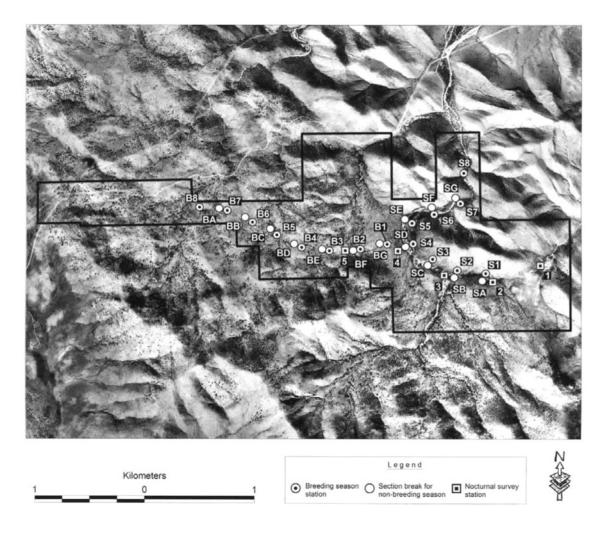


Figure 4.2. Location of bird surveys, Fort Bowie NHS, 2002-2004.

Analyses

We calculated relative abundance of each species along each transect as the number of detections at all stations and visits (including zero values) divided by effort (sample size: total number of visits multiplied by total number of stations). We reduced our full collection of observations for each VCP station (n = 1,986: 1,378 and 608 for Siphon Canyon and Butterfield transects, respectively) to a subset of data (n = 1,093; 793 and 244 for Siphon Canyon and Butterfield transects, respectively) that was more appropriate for estimating relative abundance. We used only those detections that occurred ≤ 75 m from count stations (thereby excluding 344 and 273 observations, respectively) because detectability is influenced by

conspicuousness of birds (i.e., loud, large, or colorful species are more detectable than others) and environmental conditions (dense vegetation can reduce likelihood of some detections).

Truncating detections may reduce the influence of these factors (Verner and Ritter 1983; for a review of factors influencing detectability see Anderson 2001, Farnsworth et al. 2002). We also excluded observations of birds that were flying over the station (145 and 71 observations, respectively), birds observed outside of the eight-minute count period (128 and 45 observations, respectively), and unknown species (38 and 7 observations, respectively). Some observations met more than one of these criteria for exclusion from analysis.

Line-transect Surveys

Field Methods

From 15 October 2002 to 9 January 2003 we surveyed for birds using the line-transect method (Bibby et al. 2002). Line transects differ from VCP transects, in that an observer records birds seen or heard while the observer walks a line, rather than remaining stationary. The transect method is more effective during the non-breeding season because bird vocalizations are less conspicuous and frequent, making birds more difficult to detect (Bibby et al. 2000). This method was once the preferred survey technique for surveys in all seasons and was used by Russell and Johnson (1976) in their surveys of the park.

We established two transects at the park (Fig. 4.2). The transects were broken into sections, with the start and finish locations corresponding to the breeding-season stations. Each section was approximately 250 m in length. As with other survey methods, we alternated direction of travel along transects to reduce biases and did not survey during periods of excessive rain or wind (see breeding-season survey methods for details). We began surveys at sunrise and continued until we completed both transects. As with breeding-season surveys, we recorded weather conditions at the beginning and end of each survey. Prior to beginning a section, we recorded the section name (e.g., "A–B") and the start time.

We timed our travel so that we traversed each section in ten minutes, during which time we assigned all birds seen and/or heard into one of the following distance categories: ≤ 100 m, > 100 m, or "flyover." When possible, we noted the sex and age class of birds. We recorded birds observed before or after surveys as "incidentals" (see section below), and we did not use techniques to attract birds (e.g., "pishing").

Effort

We surveyed all six sections of the Siphon Canyon and Butterfield transects four times each in the fall and winter of 2002 and 2003 (Table 4.2).

<u>Analysis</u>

We used all observations (N = 483), except unknown species, to estimate relative abundance (see Methods section of VCP surveys for more details).

Nocturnal Surveys

Field Methods

To survey for owls we broadcast commercially available vocalizations using a compact disc player and broadcaster (Colver et al. 1999, Bibby et al. 2002) and recorded other nocturnal species (nighthawks and poorwills) when detected. We established one transect from approximately the visitor center to 600 m west of Siphon Canyon, along the Butterfield Trail (Fig. 4.2). The transect had five stations that were a minimum of 300 m apart. As with other survey methods, we varied direction of travel along transects and did not survey during periods of excessive rain or wind. We began surveys approximately 45 minutes after sunset.

We began surveys at each station with a three-minute "passive" listening period during which time we broadcast no calls. We then broadcast vocalizations for a series of two-minute "active" periods. We used vocalizations of species that we suspected, based on habitat and range, might be present: elf, western screech, whiskeredscreech, barn, and (on one occasion) northern pygmy owl. We excluded great horned owl from the broadcast sequence because of its aggressive behavior toward other owls. We broadcast recordings of owls in sequence from smallest to largest size species so that smaller species would not be inhibited by the "presence" of larger predators or competitors (Fuller and Mosher 1987). During active periods, we broadcast owl vocalizations for 30 seconds followed by a 30second listening period. This pattern was repeated two times for each species. During the count period we used a flashlight to scan nearby vegetation and structures for visual detections. If we observed a bird during the three-minute passive period, we recorded the minute of the passive period in which the bird was first observed, the type of detection (aural, visual or both), and the distance to the bird. If a bird was observed during any of the twominute active periods, we recorded in which interval(s) it was detected and the type of detection (aural, visual, or both). As with other survey types, we attempted to avoid double-counting individuals recorded at previous stations. We also used multiple observers, alternated direction of travel along transects, and did not survey during inclement weather.

Effort

We surveyed all five stations on each of the three visits during the breeding season in both 2003 and 2004 (Table 4.2).

Analysis

Because of the low number of detections, we report only the number of detections and do not calculate relative abundance as for the other survey methods.

Incidental and Breeding Observations

Field Methods

When we were not conducting formal surveys and encountered a rare species, a species in an unusual location, or an individual engaged in breeding behavior, we recorded UTM coordinates, time of detection, and (if known) the sex and age class of the bird. We recorded all breeding behavior observations using the standardized classification system (developed by the North American Ornithological Atlas Committee; NAOAC 1990). This system classifies breeding behavior into one of nine categories: adult carrying nesting material, nest building, adult performing distraction display, used nest, fledged young, occupied nest, adult carrying food, adult feeding young, or adult carrying a fecal sac. We made breeding observations during both standardized surveys and incidental observations.

Analysis

We report frequency counts of incidental and breeding observations; we cannot calculate relative abundance because we did not standardize survey effort.

Vegetation Sampling at VCP Stations

In 2004 we sampled vegetation associated with each of the breeding-season stations along the Siphon Canyon and Butterfield transects. We sampled vegetation at five subplots located at a modified random direction and distance from each station. Each plot was located within a 72° range of the compass from the station (e.g., Plot 3 was located between 145° and 216°) to reduce clustering of plots. We randomly placed plots within 75 m of the stations to correspond with truncation of data used in estimating relative abundance.

At each plot we used the point-quarter method (Krebs 1998) to sample vegetation by

dividing the plot into four quadrants along cardinal directions. We applied this method to plants in three height categories: sub-shrubs (0.5–1.0 m), shrubs (> 1.0–2.0 m), trees (> 2.0 m), and one size category: potential cavity-bearing vegetation (> 20 cm diameter at breast height). If there was no vegetation in a given category within 25 m of the plot center, we indicated this in the species column. For each individual plant, we recorded its distance from the plot center, species, height, and maximum canopy diameter (including errant branches). Association of a plant to a quadrant was determined by the location of its trunk, regardless of which quadrant the majority of the plant was in; no plant was recorded in more than one quadrant. Standing dead vegetation was only recorded in the "potential cavity-bearing tree" category. On rare occasions when plots overlapped we repeated the selection process for the second plot.

Within a 5-m radius around the center of each plot, we visually estimated (1) percent ground cover by type (bare ground, litter, or rock); and (2) percent aerial cover of vegetation in each quadrant using three height categories: 0-0.5 m, > 0.5-2.0 m, and > 2.0 m. For both estimates we used one of six categories for percent cover: 0 (0%), 10 (1-20%), 30 (21-40%), 50 (41-60%), 70 (61-80%), and 90 (81-100%).

Analysis

Using point-quarter data, we calculated mean density (number of stems/ha) for all species in each of the four height/size categories using the computer program "Krebs" (Krebs 1998). We collected these data to quantify vegetation characteristics around survey stations (Appendix F).

Comparisons to Surveys by Russell and Johnson (1976)

We summarized data from Russell and Johnson (1976) and made limited comparisons between their results and ours. To facilitate these comparisons, we used only those data from our surveys that corresponded to the study sites used by Russell and Johnson: their "wash-riparian" corresponds to our VCP stations numbers 1–4 and line transects sections A-B to D-E of our Siphon Canyon transect; their "mesquite-grassland" corresponds to our VCP stations numbers 1–4 and line-transect sections A-B to D-E of our Butterfield

transect; and their "oak-juniper woodland" corresponds to our VCP stations numbers 5–8 and line-transect sections E-F to I-J of our Butterfield transect. We combined relative abundance scores from Russell and Johnson for the spring and summer surveys and compared these data to our VCP surveys. We excluded fall surveys from comparison (they visited the park only twice in the fall of 1975) and made direct comparisons between their "winter" surveys and our line-transect surveys. Because of the different field methods (and therefore inappropriateness of comparing relative abundance estimates), we compared ranks of only the most abundant species.

Results

We found 109 species during surveys from October 2002 to July 2004 (Appendix C). We found 77 species during VCP surveys, 51 species during line transect surveys, four species during nocturnal surveys, and 72 species by incidental observation during all seasons. We found no non-native species. We found a number of species of high conservation concern including peregrine falcon, loggerhead shrike, and Baird's sparrow. Based on a review of other studies at the park, there have been 189 species observed at the park (Appendix C).

VCP Survevs

We found three species on the Butterfield transect that we did not find on the Siphon Canyon transect, and 36 species on the Siphon Canyon transect that we did not find on the Butterfield transect (Tables 4.3, 4.4). Among the 36 species found only on the Siphon Canyon transect, some were among the most abundant on the transect: the Cassin's and western kingbirds, Bell's vireo, summer tanager, Bullock's and hooded orioles, and northern cardinal (Table 4.3). Most of these species are riparian obligate species. Among species that we found on both transects, and for which we were able to calculate relative abundance, there were some notable differences among transects: 10 species had higher mean relative abundance on the Butterfield transect, while 12 species had higher mean relative abundance on the Siphon Canyon transect (using the mean relative abundance scores from 2003 and 2004). There were no surprises in

these numbers. Abundant species on the Butterfield transect are known to be associated with drier conditions: the black-throated sparrow, rufous-crowned sparrow, Bewick's wren, Scott's oriole, and canyon towhee. Similarly, the Siphon Canyon transect had species that reach their highest abundance in areas of dense vegetation: the white-winged dove, Gambel's quail, verdin, and Lucy's warbler. In the Siphon Canyon transect in both 2003 and 2004 the northern cardinal, Gambel's quail, and white-winged dove were the most abundant (Table 4.3). On the Butterfield transect the mourning dove, black-throated sparrow, and Bewick's wren were the most abundant (Table 4.4).

We observed 69 species along the Siphon Canyon transect in 2003 and 2004, of which we were able to calculate relative abundance for 47 species (Table 4.3). We found 12 species in each year that we did not find the other year. With the exception of the crissal thrasher, the species not found in the other year were not among the most abundant (Table 4.3). Almost all relative abundance estimates were higher in 2004 (mean \pm SE = 0.24 \pm 0.040) than in 2003 (0.18 \pm 0.030; t = 6.3, P < .0001 on log transformed data). Among the most common species, only ash-throated flycatcher and verdin had much greater relative abundance estimates in 2003 than in 2004. Conversely, many abundant species such as mourning dove, Cassin's kingbird, Bewick's wren, and northern mockingbird had much higher relative abundance estimates in 2004 than in 2003.

Line-transect Surveys

We observed 50 species during line-transect surveys in 2002 and 2003, 35 species on the Butterfield transect and 41 species on the Siphon Canyon transect (Table 4.5). Nine species were unique to the Butterfield transect, while 16 species were unique to the Siphon Canyon transect. Excluding sandhill crane (observed only on one occasion flying over the park), Gambel's quail, chipping sparrow, and white-crowned sparrow were most abundant on the Butterfield transect (Table 4.5). On the Siphon Canyon transect, white-crowned sparrow, Gambel's quail, American robin, and spotted towhee were the most abundant.

Table 4.3. Total number of observations (Total obs.) and relative abundance (mean ± SE) of birds observed during VCP surveys, Siphon Canyon transect, Fort Bowie NHS, 2003 and 2004. Total number of observations includes all birds observed during surveys whereas relative abundance estimates exclude birds observed > 75 m from stations, flyovers, and observations made outside of the eight-minute count period. Sum is the number of observations used in calculating relative abundance estimates. See Methods section for additional details on estimation of relative abundance and effort used in those calculations.

		Relative abundance by year								
Species		2003 (n = 40)			2004 (n = 48)					
	Total obs.	Sum	Mean	SE	Sum	Mean	SE			
Gambel's quail	101	27	0.68	0.169	30	0.63	0.128			
urkey vulture	21				2	0.04	0.029			
Cooper's hawk	8	5	0.13	0.082	2	0.04	0.029			
one-tailed hawk	1									
ed-tailed hawk	4									
golden eagle	1									
vhite-winged dove	122	22	0.55	0.101	34	0.71	0.126			
nourning dove	84	15	0.38	0.117	43	0.90	0.158			
vhite-throated swift	7									
black-chinned hummingbird	8	3	0.08	0.042						
proad-tailed hummingbird	14	1	0.03	0.025	4	0.08	0.040			
adder-backed woodpecker	23	5	0.13	0.053	8	0.17	0.062			
reater pewee	1									
vestern wood-pewee	6	2	0.05	0.035						
gray flycatcher	2				1	0.02	0.021			
lusky-capped flycatcher	1									
sh-throated flycatcher	54	25	0.63	0.142	14	0.29	0.079			
prown-crested flycatcher	3	1	0.03	0.025		0.20	0.0.0			
Cassin's kingbird	61	8	0.20	0.082	33	0.69	0.130			
vestern kingbird	13	3	0.08	0.055	5	0.10	0.054			
Bell's vireo	14	5	0.13	0.082	6	0.13	0.048			
olumbeous vireo	2	<u>U</u>	0.10	0.002	2	0.04	0.029			
varbling vireo	3					0.04	0.023			
curve-billed thrasher	8	1	0.03	0.025	5	0.10	0.054			
crissal thrasher	18	I	0.00	0.023	11	0.10	0.068			
vestern scrub-jay	22	8	0.20	0.082	12	0.25	0.082			
Mexican jay	1	0	0.20	0.002	12	0.23	0.002			
inknown raven	17									
riolet-green swallow	5	1 <i>F</i>	0.00	0.070		0.40	0.057			
verdin	32	15	0.38	0.078	9	0.19	0.057			
oushtit	5	2	0.05	0.050	11	0.23	0.124			
cactus wren	71	19	0.48	0.095	33	0.69	0.130			
ock wren	2				11	0.02	0.021			
canyon wren	3	40	0.05	0.000	1	0.02	0.021			
Bewick's wren	57	10	0.25	0.069	28	0.58	0.111			
nouse wren	2	1	0.03	0.025						
uby-crowned kinglet	13	6	0.15	0.057	6	0.13	0.057			
lue-gray gnatcatcher	4	1	0.03	0.025	2	0.04	0.029			
merican robin	11	1	0.03	0.025						
orthern mockingbird	72	14	0.35	0.084	35	0.73	0.139			
hainopepla	10				6	0.13	0.048			
ucy's warbler	14	1	0.03	0.025	13	0.27	0.071			
rellow-rumped warbler	4	1	0.03	0.025	1	0.02	0.021			
black-throated gray warbler	4				3	0.06	0.046			
Townsend's warbler	1				1	0.02	0.021			
Nilson's warbler	8	2	0.05	0.035	2	0.04	0.029			

		Relative abundance by year								
			2003 (n = 40	0)		2004 (n = 48)				
Species	Total obs.	Sum	Mean	SE	Sum	Mean	SE			
summer tanager	31	4	0.10	0.048	13	0.27	0.088			
western tanager	7	1	0.03	0.025						
green-tailed towhee	8	6	0.15	0.057	1	0.02	0.021			
spotted towhee	7	3	0.08	0.042	3	0.06	0.046			
canyon towhee	40	20	0.50	0.139	26	0.54	0.111			
Botteri's sparrow	1	1	0.03	0.025						
rufous-crowned sparrow	14	2	0.05	0.035	6	0.13	0.064			
chipping sparrow	2	1	0.03	0.025						
Brewer's sparrow	2	4	0.10	0.078						
black-throated sparrow	40	7	0.18	0.071	22	0.46	0.123			
Oregon junco	1	1	0.03	0.025						
northern cardinal	112	38	0.95	0.129	56	1.17	0.113			
pyrrhuloxia	1	1	0.03	0.025						
black-headed grosbeak	7				1	0.02	0.021			
blue grosbeak	2	2	0.05	0.050	1	0.02	0.021			
lazuli bunting	1				1	0.02	0.021			
bronzed cowbird	1	2	0.05	0.050						
brown-headed cowbird	33	5	0.13	0.053	13	0.27	0.077			
hooded oriole	36	15	0.38	0.111	13	0.27	0.083			
Bullock's oriole	25	6	0.15	0.067	11	0.23	0.074			
Scott's oriole	6	2	0.05	0.035	3	0.06	0.035			
house finch	40	8	0.20	0.073	14	0.29	0.079			
lesser goldfinch	3				2	0.04	0.029			

Nocturnal Surveys

We recorded four species of nocturnal birds: one observation each of the western screech owl and great-horned owl, three observations of the elf owl, and 12 observations of the common poorwill.

Incidental and Breeding Observations

We made incidental observations of 72 species, including 12 species that were not detected during any other survey type (Appendix C). These species included: long-eared owl, common ground dove, greater roadrunner, Bendire's thrasher, Baird's sparrow, and grasshopper sparrow. We made 84 observations of breeding behavior, representing 32 species (Table 4.6). We made the most breeding observations of mourning dove (14 observations). We confirmed breeding for Bell's vireo and summer tanager, two riparian-obligate species.

Inventory Completeness

Based on the results from our surveys and the list by Fischer (2002), the inventory of birds that

regularly use the park is probably close to completion. A look at the species accumulation curve for our work indicates that our effort alone was not sufficient to document all of the species that occur in the park; the cumulative number of new species for this study was not approaching an asymptote (Fig. 4.3). We found 6 species that were new to the park list: Baird's sparrow, Hutton's vireo, common ground-dove, dusky-capped flycatcher, greater pewee, and zone-tailed hawk (Appendix C). Of these species, only Hutton's vireo may nest at the park. Because of the location of the park near to the species-rich Chiricahua Mountains (one of the most popular destinations in the United States for bird watchers), we expect that new species of birds will be added to the list for years to come, but that these species will likely be uncommon or rare at the park. Only through major modification of the vegetation community (either by the use of fire or removal of mesquite) will additional species, such as grassland-associated sparrows (e.g., grasshopper, Cassin's, and Botteri's), nest in the park.

Table 4.4. Total number of observations (Total obs.) and relative abundance (mean ± SE) of birds during VCP surveys, Butterfield transect, Fort Bowie NHS, 2004. Total number of observations includes all birds observed during surveys whereas relative abundance estimates exclude birds observed > 75 m from stations, flyovers, and observations made outside of the eightminute count period. Sum is the number of observations used in calculating relative abundance estimates. See Methods section for additional details on estimation of relative abundance and Table 4.2 for sample size.

	Relative abundance (n = 48)						
Species	Total obs.	Sum	Mean	SE			
Gambel's quail	34	7	0.15	0.089			
turkey vulture	17	1	0.02	0.021			
red-tailed hawk	3						
white-winged dove	38	2	0.04	0.029			
mourning dove	71	15	0.31	0.095			
black-chinned hummingbird	3						
ladder-backed woodpecker	20	8	0.17	0.069			
western wood-pewee	1						
Hammond's flycatcher	1	1	0.02	0.021			
ash-throated flycatcher	42	19	0.40	0.093			
Cassin's kingbird	6						
western kingbird	1						
crissal thrasher	12	8	0.17	0.062			
western scrub-jay	17	5	0.10	0.045			
unknown raven	5						
bridled titmouse	1	1	0.02	0.021			
juniper titmouse	3	4	0.08	0.065			
verdin	2	2	0.04	0.029			
bushtit	4	11	0.23	0.124			
cactus wren	39	23	0.48	0.094			
rock wren	7	4	0.08	0.050			
Bewick's wren	50	32	0.67	0.113			
ruby-crowned kinglet	2	2	0.04	0.029			
northern mockingbird	28	4	0.08	0.050			
phainopepla	4	3	0.06	0.046			
Lucy's warbler	3	1	0.02	0.021			
green-tailed towhee	1	1	0.02	0.021			
canyon towhee	36	31	0.65	0.117			
rufous-crowned sparrow	29	17	0.35	0.070			
chipping sparrow	1	1	0.02	0.021			
black-throated sparrow	67	47	0.98	0.141			
northern cardinal	2						
black-headed grosbeak	5						
blue grosbeak	3	3	0.06	0.046			
brown-headed cowbird	18	11	0.23	0.068			
Scott's oriole	18	10	0.21	0.073			
house finch	10	2	0.04	0.029			

Table 4.5. Relative abundance of birds observed during line-transect surveys, Fort Bowie NHS, 2002 and 2003.

		D. 44 - 46 - 1-1 / 0.4	Transe		Cinhan C /	. 00)
Species		Butterfield (n = 24)	SE		Siphon Canyon (n =	: <u>23)</u> SE
Species	Sum	Mean		Sum	Mean	
Gambel's quail	80	3.33	2.383	40	1.74	0.796
Montezuma quail	1	0.04	0.042		0.04	0.040
Cooper's hawk		0.04	0.040	1	0.04	0.043
ed-tailed hawk	11	0.04	0.042			
sandhill crane ^a	52	2.17	2.167			
ed-naped sapsucker				1	0.04	0.043
adder-backed woodpecker	4	0.17	0.078	7	0.30	0.132
northern flicker	8	0.33	0.098	12	0.52	0.187
oggerhead shrike	1	0.04	0.042			
solitary vireo type ^b				1	0.04	0.043
vestern scrub-jay	12	0.50	0.170	21	0.91	0.259
ınknown raven				2	0.09	0.087
Steller's jay	2	0.08	0.083			
curve-billed thrasher				2	0.09	0.060
rissal thrasher	6	0.25	0.090	10	0.43	0.138
oridled titmouse	10	0.42	0.232	1	0.04	0.043
uniper titmouse	2	0.08	0.083			
erdin	6	0.25	0.090	4	0.17	0.102
pushtit	28	1.17	0.809			
prown creeper				1	0.04	0.043
actus wren	6	0.25	0.109	12	0.52	0.165
ock wren		0.20	0.100	3	0.13	0.072
Bewick's wren	3	0.13	0.092	8	0.35	0.072
uby-crowned kinglet	11	0.46	0.159	21	0.91	0.113
vestern bluebird	18	0.75	0.391	9	0.39	0.100
Vesterri bidebird Fownsend's solitaire	4	0.75	0.078	y	0.39	0.212
	4	0.17	0.070	4	0.04	0.040
nermit thrush	0	0.00	0.050	1	0.04	0.043
American robin	2	0.08	0.058	37	1.61	0.838
northern mockingbird	3	0.13	0.069	7	0.30	0.117
edar waxwing				12	0.52	0.522
phainopepla	7	0.29	0.127	9	0.39	0.137
ellow-rumped warbler	2	0.08	0.083	11	0.04	0.043
plack-throated gray warbler				1	0.04	0.043
green-tailed towhee	6	0.25	0.090	28	1.22	0.259
potted towhee	15	0.63	0.189	37	1.61	0.249
canyon towhee	13	0.54	0.170	27	1.17	0.249
ufous-crowned sparrow				1	0.04	0.043
chipping sparrow	54	2.25	0.738	29	1.26	0.389
Brewer's sparrow	21	0.88	0.641	30	1.30	0.531
incoln's sparrow				6	0.26	0.113
plack-chinned sparrow	1	0.04	0.042	4	0.17	0.136
esper sparrow	4	0.17	0.130	29	1.26	1.171
lack-throated sparrow	9	0.38	0.145	3	0.13	0.072
white-crowned sparrow	78	3.25	1.277	56	2.43	0.612
lark-eyed junco	6	0.25	0.090	2	0.09	0.060
orthern cardinal				13	0.57	0.197
pyrrhuloxia				1	0.04	0.043
nouse finch				4	0.17	0.043
pine siskin	1	0.04	0.042	т	0.17	0.001
esser goldfinch	1	0.04	0.042	6	0.26	0.180

 $^{^{\}rm a}\text{All}$ observed flying over the park on one occasion. $^{\rm b}\text{Either}$ solitary or cordilleran.

Table 4.6. Number of observations by breeding behavior for birds, Fort Bowie NHS, 2003 and 2004. Breeding behaviors follow standards set by NAOAC (1990).

			Nest		Adults	carrying		Other		
								Feeding		
								recently	Recently	
Common name	Building	With	With	Occupied	Food	Nesting material	Distraction displays	fledged	fledged	Totals
	Dulluling	eggs	young	Occupied	roou	materiai	uispiays	young	young	
Gambel's quail								1		1
Cooper's hawk				2						2
white-winged dove		1		4						5
mourning dove				14						14
black-chinned hummingbird				1						1
broad-tailed hummingbird	1			3			2			6
ladder-backed woodpecker				1						1
Say's phoebe								2	1	3
brown-crested flycatcher				1						1
Cassin's kingbird	1			2					1	4
western kingbird								1		1
Bell's vireo				1				1		2
western scrub-jay									1	1
barn swallow	1									1
verdin	1									1
cactus wren				1	1			2		4
rock wren					1					1
canyon wren					1			1		2
Bewick's wren					1				1	2
black-tailed gnatcatcher								1		1
northern mockingbird					1			2		3
curve-billed thrasher									1	1
crissal thrasher									1	1
phainopepla				1						1
summer tanager								2		2
canyon towhee				1	2	2		1	1	7
black-throated sparrow			1		1			1		4
northern cardinal				1	1				1	3
brown-headed cowbird									1	1
hooded oriole	1			1		1				3
Bullock's oriole			1	1				1		3
house finch				1						1
Totals	5	1	2	35	10	3	2	16	9	84

Discussion

Based on our research and that by others, Fort Bowie NHS has a diverse bird community (n = 189 species) for a small area (405 ha) with little topographic relief. Although the pattern of extraordinary species richness that we observed in plants (Chapter 3) was not mirrored in the bird community, the diversity of vegetation communities at the sites clearly plays a role in

determining the bird community. For example, there were notable differences in the bird communities along the two repeat-visit VCP transects where the Siphon Canyon transect had almost twice as many species as the Butterfield transect (Tables 4.3, 4.4).

Many of the species that we found in the Siphon Canyon transect are known to occur primarily in riparian areas or areas of dense vegetation: Bell's vireo, summer tanager, hooded

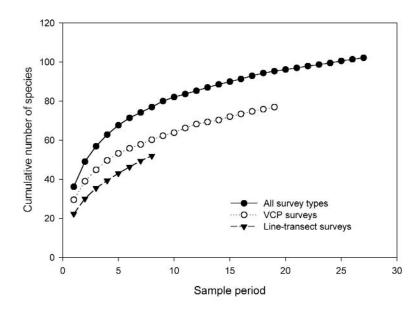


Figure 4.3. Species accumulation curves, by survey type, for the UA bird inventory effort, Fort Bowie NHS, 2002–2004. Each sample period for all survey types represents a randomized ordering of 101 observations (*N* = 2,821; a completely randomized combination of the four survey types). Each sample period for VCP and line-transect surveys represents one survey day.

oriole, and northern cardinal (Powell and Steidl 2000). Although not as species rich as in Siphon Canyon, the bird community along the Butterfield transect had species that are associated with the oak savanna and chaparral including juniper titmouse, Crissal thrasher, and rufous-crowned sparrow. Each bird species is closely tied to gross vegetation characteristics such as (1) vertical structure (MacArthur and MacArthur 1961, Cody 1981), (2) horizontal patchiness (Roth 1976, Kotliar and Weins 1990), and (3) floristics (Rice et al. 1984, Strong and Bock 1990, Powell and Steidl 2002). Given the differences in vegetation structure and composition between the two transects, the differences that we noted in the bird communities were not surprising.

The dense riparian vegetation near Apache Spring plays an important role in providing nesting habitat for Cooper's hawk and summer tanager; they were found nesting in that area. Research in the southwestern U.S. has consistently shown that areas with riparian trees have bird communities that are more diverse than adjacent sites (Carothers et al. 1974, Szaro and Jakle 1985, Strong and Bock 1990), which is due, in part, to the variety of microhabitats that riparian vegetation provides for

nesting (Powell and Steidl 2002), cover, and foraging.

Park managers are interested in the impact that the invasion of the native velvet mesquite is having on the visitors experience in the park (NPS) 2000b) because mesquite was not identified in historic photographs of the area. Velvet mesquite has increased in density and distribution in the region since the late 1800s, primarily due to disruption of historical fire regimes and the introduction of cattle grazing (Humphrey 1974, Brown 1994, Van Auken 2000). The conversion of semi-desert grasslands to mesquite woodlands has important implications for the bird community by favoring shrub-associated species such as northern cardinal, verdin, and black-throated sparrow (Lloyd et al. 1998) while not favoring many species of management concern such as Botteri's, Cassin's, and grasshopper sparrows. The loss of native grassland has been identified as a primary factor in population declines of grassland birds as a group (Herkert 1994, Knopf 1994, Peterjohn and Sauer 1999).

Comparisons to Russell and Johnson (1976)

The research by Russell and Johnson (1976) was the first comprehensive survey of birds at the park. Unfortunately we can only make very gross comparisons between our two studies because we did not use the same survey methods and exactly the same area. Yet a number of species and communities were quite different between the two studies. We found 14 species that Russell and Johnson did not find and they found 62 species that we did not find (Appendix C). We found blackthroated sparrow to be among the most common species in the oak-juniper woodland, but Russell and Johnson did not find them to be very abundant (Appendix G), Conversely, in Siphon Canvon, they found black-chinned hummingbird to be the most common species, whereas it was among the least common species in 2003 and was not found at all in 2004 (Table 4.3). There is little indication of a regional population decline in this species (Sauer et al. 2004), though banding data from Hummingbird

Monitoring Network sites is showing some decline for 2005 (Larry Norris, *pers. comm.*). Nevertheless, the decline that we saw may have been an artifact of different field methods; species such as hummingbirds may be more conspicuous while walking a transect line.

During the non-breeding season the mesquite-grassland vegetation community had the most bird species that did not rank as abundant for the other study (Appendix G). This may have been an artifact of small sample size from both studies. It could also reflect the variability of non-breeding season birds. For example, species such as white-crowned, black-throated, Brewer's, and chipping sparrows can form large, sometimes mixed-species flocks which, if encountered, can radically affect the relative abundance estimates for a transect.

Chapter 5: Mammal Inventory

Previous Research

The documented species list of mammals from the park is largely complete. The first inventory of mammals at Fort Bowie was completed by Roth and Cockrum (1976). More recently Petryszyn (1999) and Hermann-Reese (unpublished data) completed surveys for rodents, and Krebbs (2005) surveyed for bats from 2001 to 2004. Hermann-Reese also surveyed for medium and large mammals using infrared-triggered cameras and in this report we summarize that work (Appendix H). Swann et al. (2001) noted mammals seen incidentally to their surveys of reptiles and amphibians. In all, there have been 57 species documented (including specimens at the University of Arizona mammal collection) for the park: 12

bats, 30 small mammals (Orders Insectivora, Rodentia, and Lagomorpha), and 15 medium to large mammals (Orders Carnivora and Artiodactyla; Appendix D).

Methods

Because mammals have been surveyed extensively, we surveyed only for medium and large mammals using infrared-triggered cameras at three sites (Fig. 5.1). For this report our purpose was to (1) augment the infrared-triggered camera effort of Herman-Reese and (2) synthesize species lists from the previously mentioned survey efforts. We refer the reader to the other inventory efforts for more detailed species accounts.

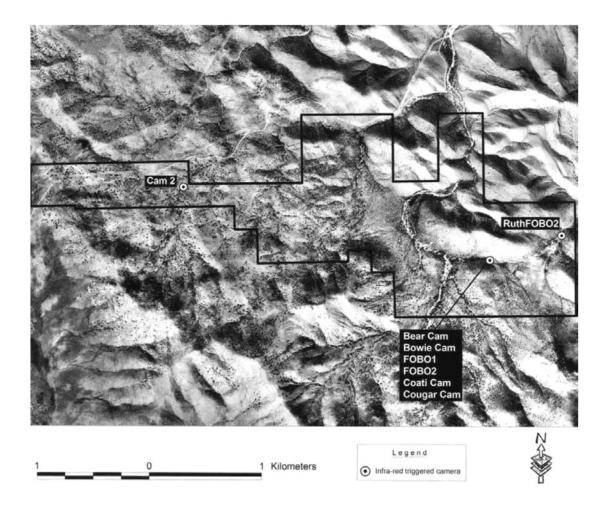


Figure 5.1. Location of Trailmaster camera sites, Fort Bowie NHS, 2002–2003.

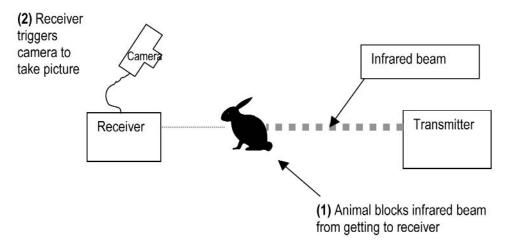


Figure 5.2. Diagram of infrared-triggered camera (Trailmaster) set-up. Image based on Swann et al. (2004).

Trailmaster Cameras

We used infrared-triggered cameras (herein referred to as "Trailmaster"; Trailmaster®; model 1500, Goodman and Associates, Inc, Lenexa, KS; Kucera and Barrett 1993) to record the presence of medium and large mammals. Trailmasters have three components: receiver, transmitter, and camera (Fig. 5.2). The transmitter sends an infrared beam to the receiver at a specified rate (5 times per second for this study). The receiver then sends a signal (via cable) to a camera mounted on a tripod 6–8 m away. When an animal blocks the infrared beam the camera takes a picture.

We set the receiver and transmitter approximately 8 m apart and 20 cm above the ground so that medium and large mammals were captured on film but smaller animals such as rodents and birds were not. We set cameras to take no more than one photograph every five minutes to reduce the chances of recording the same individual more than once (on the same occasion). We placed cameras in three areas of the park (Fig. 6.1; UTM coordinates for Willow Gulch = 644390 N, 3558289 E; Apache Spring = 647101 N, 3557644 E; Visitor Center Road = 647729 N, 3557866 E) that we thought would record the highest number of species; typically these were in areas of dense vegetation. We baited camera sites with a commercial scent lure (ingredients included synthetic catnip oil, bobcat musk, beaver castorium, and propylene glycol as a preservative) or canned cat food. We checked cameras approximately every two weeks to change film and batteries and to ensure their proper function. We

photographed a placard documenting the date and camera location on the first exposure of every new roll of film.

Spatial Sampling Design

We selectively placed cameras in areas that we felt would have the most success, primarily near Apache Spring.

Effort

We operated Trailmaster cameras at three locations from May 2002 to May 2003 for a total of 278 days of camera operation (Table 5.1). Of the three locations, Apache Spring had the most effort (47%), followed by visitor center road (38%) and Willow Gulch (15%).

Analysis

Trailmaster cameras are the most cost-effective method for recording the presence of medium and large mammal species (Kucera and Barrett 1993, Cutler and Swann 1999). However, one drawback to this method is an inability to differentiate individuals, which precludes any estimates of abundance (i.e., one must be able to determine whether one animal has been photographed repeatedly or whether more than one individual has been photographed). In some cases, size or physical abnormality may differentiate individuals of any species, but this was not evident in our photographs. Also, each species is more or less likely to be attracted to the camera area. Therefore, we report the number of times a species was photographed to indicate species that may be common, based on the number of photographs.

Table 5.1. Summary of Trailmaster camera effort, Fort Bowie NHS, 2002 and 2003.

General location	Camera name	Year	Start date	End date	Number of days open
Willow Gulch	Cam 2	2002	6 Oct	19 Oct	13
		2002	19 Oct	17 Nov	29
Apache Spring	Bowie Cam	2002	12 May	25 May	13
	FOBO2	2002	2 Jun	16 Jun	14 ^a
	Coati Cam	2002	30 Jun	7 Jul	7 ^a
	Cougar Cam	2002	14 Jul	21 Jul	7 ^a
		2002	28 Jul	31 Jul	3 ^a
		2002	22 Sep	4 Oct	12
		2002	19 Oct	13 Nov	25
		2002	17 Nov	28 Nov	11
	Bear Cam	2002	25 Aug	21 Sep	27
	FOBO1	2002	6 Oct	18 Oct	12
Visitor Center Road	RuthFOBO2	2002-2003	19 Dec	3 May	105

^aDue to improper documentation, this number is an estimate of the number of days the camera was available to take photographs.

Incidental Observations and Signs

As with other taxa, we recorded UTM coordinates of mammal sightings made outside of formal surveys. Observers from all field crews (e.g., bird crew) recorded mammal sightings.

Results and Discussion

We documented two birds (common black hawk and American robin) and 11 species of mammals using Trailmaster cameras at Fort Bowie NHS in 2002 and 2003 (Table 5.2). We documented two new species of mammals for the park: hooded

skunk and feral dog (Appendix D). The photo of the dog is the first documentation of a non-native mammal in the park. The most photographs were of the white-nosed coati (Table 5.2). Collared peccary and striped skunk were also in many of the photographs.

As we expected, most of the photos were from Apache Spring (Table 5.2). In general, this was an extremely dry period (Fig. 2.3) and the spring was an important resource, drawing in many animals including an American black bear, which was likely dispersing from the Pinaleño or Chiricahua Mountains. We did not document any animals at the Willow Gulch site.

Table 5.2. Number of photographs of animals, by Trailmaster camera site, Fort Bowie NHS, 2002 and 2003. Number in parentheses is the total number of individuals if >1 individual was seen in a picture.

			Site	
		A	pache Spring	Visitor Center Road
Group	Species	Number of photographs	Number of individuals in photographs	Number of photographs and individuals
Bird	common black-hawk	1	1	
	American robin			4
Mammal	American black bear	4	4	
	white-nosed coati	29	55	
	striped skunk	18	19	2
	hooded skunk	2	2	
	white-backed hog-nosed skunk			1
	feral dog	1	1	
	common gray fox	6	6	
	mountain lion	11	12	2
	bobcat	6	6	
	collared peccary	15	20	6
	mule deer	9	12	

Chapter 6: Management Implications

Based on the data from this study and our knowledge of the natural resource issues at the park, we highlight issues that affect the park's natural resources. Coordination with other agencies, non-governmental organizations, and/or adjacent landowners may prove the best route to resolving some of these challenges.

Development Adjacent to the Park

One of the most serious threats to the biological diversity of Fort Bowie NHS may be residential development to the north of the park. The development may lead to an increase in the number and extent of non-native plants (Seabloom et al. 2003) and may disrupt animal movement patterns and result in the loss and/or fragmentation of habitat (Mills et al. 1989, Theobald et al. 1997, Riley et al. 2003), particularly for larger mammals. Also, free-roaming pets, normally associated with development, can negatively impact native vertebrates through harassment and mortality (Coleman and Temple 1993). This may not be a problem because the Buddhist retreat, on which most of the adjacent development is taking place, does not allow pets.

Cattle Grazing

The impact of cattle grazing on the park's natural resources has never been documented. As mentioned in the Park Introduction chapter, cattle grazing can have harmful impacts on the native biota by causing changes in the distribution, abundance, and composition of plant populations as well as soil erosion and compaction. Ruyle (2001) assessed the range condition on eight sites; his results indicate that conditions ranged from fair

to good, but those conditions were only assessed using vegetation measures. We suggest that if grazing were to continue at the park, managers establish more long-term monitoring protocols and sites to determine the impact of grazing on the natural resources of the park, particularly soils, vegetation, and vertebrates.

Perhaps the most damaging practice relating to grazing is the use of water from Apache Spring to supply water to adjacent cattle allotments. Although the allotment of one-half of the water from the spring is rarely realized (Larry Ludwig, *pers. comm.* to Michele Gerard), there is a potential for that amount of water to be diverted. The water from the spring is a vital resource for the park and is responsible for the riparian area that is so valuable for plants and wildlife in the area. A hydrologist could help park managers determine the impact of water diversion on the riparian area.

Visitor Impacts

If the road over Apache Pass is paved, there will likely be increased mortality of reptiles, particularly snakes, which seek out the pavement on hot summer nights to bask (Rosen and Lowe 1994). Increased vehicular volume and speed will also likely increase the modification of animal behavior (Trombulak and Frissell 2000). Modification of behavior probably already takes place at Apache Spring, which is the only source of perennial water for animals, but is also a popular resting spot for hikers. The effect of this conflict has never been quantified, but given the high abundance and species richness of birds and large mammals in that area (see Tables 4.3, 5.2), conflicts are inevitable.

Chapter 7: Additional Inventories

No inventory is ever truly complete; species distributions expand and contract across boundaries, particularly at smaller parks such as Fort Bowie NHS. In general, we feel that the inventories for vascular plants and vertebrates are nearly complete and that the park is in a good position to monitor changes in species composition. Through the continuous collection of data, such as specimens from road kill, photographs, and through research studies at the park, managers can continue to monitor changes. An additional step would be to coordinate additional inventory-like field efforts with adjacent landowners to increase the spatial scope of the inventory effort. Below we suggest ways to complete the species inventories and/or implement monitoring on the park.

Plants

A number of woody invasive species such as velvet mesquite and burroweed are of concern to park managers (NPS 2000b) and there is currently a program to remove mesquite. We suggest that a more rigorous, plot-based vegetation monitoring program be established at the park (e.g., Powell et al. 2005a), including a number of plots in the area of mesquite removal. This monitoring would inform the park managers of the effectiveness of the program as well as document changes in the plant community concurrent to this removal (of particular concern is the spread of non-native grasses such as Lehmann lovegrass). Repeat inventories for plants, particularly the early detection of non-native plants, should be carried out at least every five years.

Specimen vouchers from the park may be residing in the herbarium collections at Arizona State University and Northern Arizona University. In early 2005 it became possible to access information about these collections. We suggest that these databases be searched for specimens from the park. However, care should be taken in accepting the list of species without confirming the proper identification of species or updating taxonomy (Halvorson 2003).

Many of the specimens cited in the report by Warren et al. (1992) are currently housed at the park, but because the park does not have a natural resource staff member who is responsible for the proper curation of that collection, it is unlikely that the plant specimens are receiving proper care. Therefore, we recommend removing them to a collection that has proper archival conditions, such as the University of Arizona.

Reptiles and Amphibians

Swann et al. (2001) surveyed for amphibians and reptiles in 1997 and 1998 and their study remains one of the best examples of a biological inventory in the region. Because they designed their study to form the basis for long-term monitoring, we suggest repeating their effort (or some portion) at least every ten years. It will especially important to survey for amphibians and aquatic reptiles because these groups are experiencing sharp declines in distribution and abundance (Wake 1990).

Birds

Additional surveys during the winter season and during the spring and fall migrations will pick up species missed by efforts at other times. It is important to note, however, that bird lists are difficult to complete because birds are highly mobile. Only sites that are visited regularly by avid bird watchers (e.g., Madera Canyon, Ramsey Canyon, and Patagonia-Sonoita Creek Preserve in southern Arizona) have bird lists that can be considered to be "complete."

Mammals

Herman-Reese did not write up the results of either the small mammal trapping or Trailmaster studies that she undertook. Although we published an appendix of her Trailmaster camera results (Appendix H), this effort is insufficient to properly document and archive data from that project. We encourage park staff to work with her to complete that effort before too much time passes; her field effort was considerable and it would be unfortunate not to have a report to document her effort.

The one group of mammals that may require additional inventory work is bats. Krebbs (2005) netted at the park from 2001-2004 for a total of five nights of netting. It did not appear to

be sufficient for documenting all of the species of insectivorous bats at the park; new species continued to be found. We therefore recommend additional netting at Apache Spring. The use of ultrasonic detectors to identify bat species is increasing, and many researchers are refining the field techniques and improving the technology (e.g., Johnson et al. 2002, Gannon et al. 2003). These technologies may become more useful in the coming years with these refinements.

Chapter 8: Literature Cited

- American Ornithologists' Union (AOU). 1998. Checklist of North American birds, seventh edition. American Ornithologists' Union and Allen Press Inc., Lawrence, KS.
- American Ornithologists' Union (AOU). 2003. Forty-second supplement to the American Ornithologists' Union checklist of North American birds. Auk 117:847–858.
- Anable, M. E., M. P. McClaren, and G. B. Ruyle. 1992. Spread of introduced Lehmann's lovegrass Eragrostis lehmanniana Nees. in southern Arizona, USA. Biological Conservation 61:181–188.
- Anderson, D. R. 2001. The need to get the basics right in wildlife field studies. Wildlife Society Bulletin 29:1294–1297.
- Bahre, C. J. 1991. A legacy of change: historic human impact on vegetation of the Arizona borderlands. University of Arizona Press, Tucson, AZ.
- Baker, R. J., L. C. Bradley, R. D. Bradley, J. W.
 Dragoo, M. D. Engstrom, R. S. Hoffmann, C.
 A. Jones, F. Reid, D. W. Rice, and C. Jones.
 2003. Revised checklist of North American mammals north of Mexico, 2003. Occasional Papers of the Museum of Texas Tech
 University 229:1–23.
- Bednarz, J. C., and R. J. Raitt. 2002. Chihuahuan raven (Corvus cryptoleucus). In The birds of North America, No. 606. A. Poole and F. Gill, editors. The Birds of North America, Inc., Philadelphia, PA.
- Belsky, A. J., A. Matzke, and S. Uselman. 1999. Survey of livestock influences on stream and riparian ecosystems in the western United States. Journal of Soil and Water Conservation 54:419–431.
- Bennett, P. S., R. R. Johnson, and M. R. Kunzman. 1996. An annotated list of vascular plants of the Chiricahua Mountains, Including Pedregosa Mountains, Swisshelm Mountains, Chiricahua National Monument, and Fort Bowie National Historic Site. USGS

- Cooperative Park Studies Unit. Special Report No. 12. School of Natural Resources, University of Arizona, Tucson, AZ
- Bibby, C. J., N. D. Burgess, and D. A. Hill. 2000. Bird census techniques. Academic Press, London, England.
- Bowers, J. E., and S. P. McLaughlin. 1982. Plant species diversity in Arizona. Madroño 29: 227–233.
- Brown, D. E. 1994. Biotic communities: southwestern United States and northwestern Mexico. University of Utah Press, Salt Lake City, UT.
- Buckland, S. T., D. R. Anderson, K. P. Burnham, J. L. Laake, D. L. Borchers, and L. Thomas. 2001. Introduction to Distance Sampling: estimating abundance of biological populations. Oxford University Press, London, England.
- Bureau of Land Management (BLM). 1994. Rangeland reform 1994. Draft environmental impact statement. U. S. Department of the Interior, Washington, D.C.
- Burgess, T. L., J. E. Bowers, and R. M. Turner. 1991. Exotic plants at the Desert Laboratory, Tucson, Arizona. Madroño 38:96–114.
- Carothers, S.W., R. R. Johnson, and S.W. Atchinson. 1974. Population structure and social organization of southwestern riparian birds. American Zoologist 14:97–108.
- Cody, M. L. 1981. Habitat selection in birds: the roles of vegetation structure, competitors, and productivity. Bioscience 31:107–113.
- Coleman, J. S., and S. A. Temple. 1993. Rural residents' free-ranging domestic cats: a survey. Wildlife Society Bulletin 21:381–390.
- Colver, K. J., D. Stokes, and L. Stokes. 1999. Stokes field guide to bird songs. Time Warner Trade, New York, NY.
- Cutler, T. L., and D. E. Swann. 1999. Using remote photography in wildlife ecology: a review. Wildlife Society Bulletin 27:571–581.

- Davis, K., and W. L. Halvorson. 2000. A study plan to inventory vascular plants and vertebrates: Sonoran Desert Network. Southern Arizona Office, National Park Service, Phoenix AZ.
- Denny, D. W., and C. R. Peacock. 2000. Soil Survey of Fort Bowie National Historic Site, Arizona. United States Geological Survey Technical Report No. 64. University of Arizona, Tucson, Arizona.
- DeSante, D. F., and D. R. O'Grady. 2000. The monitoring avian productivity and survivorship (MAPS) program: 1997 and 1998 report. Bird Populations 5:49–101.
- Farnsworth, G. L., K. H. Pollack, J. D. Nichols, T.
 R. Simons, J. E. Hines, and J. R. Sauer. 2002.
 A removal model for estimating detection probabilities from point-count surveys. The Auk 119:414–425.
- Fischer, D. 2002. A checklist of birds of Chiricahua National Monument & Fort Bowie National Historic Site. Southwest Parks and Monuments Association, Tucson, AZ.
- Fleischner, T. L. 1994. Ecological costs of livestock grazing in western North America. Conservation Biology 8:629–644.
- Fuller, M. R., and J. A. Mosher. 1987. Raptor survey techniques. Pp. 37–66. In B. A.
 Geron-Pendleton, B. A. Millsap, K. W. Cline, and D. M. Bird, editors. Raptor management techniques manual. National Wildlife Federation, Washington, D.C.
- Gannon, W. L., R. E. Sherwin, and S. Haymond. 2003. On the importance of articulating assumptions when conducting acoustic studies of habitat use by bats. Wildlife Society Bulletin 31:45–61.
- Halvorson, W. L. 2003. Verification of plant specimens from Sonoran desert parks. Report to the Desert Southwest Cooperative Ecosystems Studies Unit, University of Arizona, Tucson, AZ.
- Halvorson, W. L., and P. Guertin. 2003. USGSWeeds in the west project: status of introduced plants in southern Arizona parks. U.S.Geological Survey, Southwest Biological

- Science Center, Sonoran Desert Research Station, University of Arizona, Tucson, AZ.
- Hartman, E. L., and M. L. Rottman. 1998. A checklist of the vascular plants of Fort Bowie National Historic Site. Unpublished report.
- Hayek, L. C., and M. A. Buzas. 1997. Surveying natural populations. Columbia University Press, New York, NY.
- Hayes, M. P., and M. R. Jennings. 1986. Decline of Ranid frog species in western North America: are bullfrogs (Rana catesbeiana) responsible? Journal of Herpetology 20:490–509.
- Heritage Data Management System (HDMS). 2004. Arizona Game and Fish Department, Phoenix, AZ. Accessed 5 March from: http://www.gf.state.az.us/w_c/edits/hdms_spec ies_lists.html.
- Hermann-Reese, B. No Date. Nocturnal rodents survey count, Fort Bowie National Historic Site (1997–2002). Unpublished data to Fort Bowie National Historic Site.
- Herkert, J. R. 1994. Breeding bird communities of midwestern prairie fragments the effects of prescribed burning and habitat-area. Natural Areas Journal 14:128-135.
- Humphrey, R. R. 1974. Fire in the deserts and desert grasslands of North America. Pages 365-400 In T. T. Kozlowski, and C. E. Algren, editors. Fire and ecosystems. Academic Press, London, England.
- Integrated Taxonomic Information System (ITIS). 2004. Accessed on 20 March 2004 from: http://www.itis.usda.gov/index.html.
- Johnson, J. B., M. A. Menzel, J. W. Edwards, and W. M. Ford. 2002. A comparison of 2 acoustical bat survey techniques. Wildlife Society Bulletin 30:931–936.
- Kiesecker, J. M., and A. R. Blaustein. 1997. Population differences in responses of redlegged frogs (Rana aurora) to introduced bullfrogs. Ecology 78:1752–1760.
- Knopf, F. L. 1994. Avian assemblages on altered grasslands. Studies in avian biology 15:247–257.

- Kotliar, N.B., and J. A. Weins. 1990. Multiple scales of patchiness and patch structure: a hierarchical framework for the study of heterogeneity. Oikos 59:253–260.
- Krebbs, K. 2005. Bat species richness and abundance at the Chiricahua National Monument and Fort Bowie Historic Site.
 Unpublished report to Western National Parks Association and the National Park Service, Tucson, AZ.
- Krebs, C. J. 1998. Ecological Methodology. Second Edition. Addison-Welsey Educational, Menlo Park, CA.
- Kucera, T. E., and R. H. Barrett. 1993. The Trailmaster camera system for detecting wildlife. Wildlife Society Bulletin 21:505–508.
- Lancia, R. A., J. D. Nichols, and K. H. Pollock. 1996. Estimating the number of animals in wildlife populations. Pp. 215–253. In T. A. Bookhout, editor. Research and management techniques for wildlife and habitats. Fifth edition. The Wildlife Society, Bethesda, MD.
- Latta, M. J., C. J. Beardmore, and T. E. Corman. 1999. Arizona Partners in Flight conservation plan. Technical Report 142. Nongame and Endangered Wildlife Program, Arizona Game and Fish Department, Phoenix, AZ.
- Lawler, S. P., D. Dritz., T. Strange, and M. Holyoak. 1999. Effects of introduced mosquitofish and bullfrogs on the threatened California red-legged frog. Conservation Biology 13:613–622.
- Lloyd, J., R. W. Mannan, S. DeStefano, and C. Kirkpatrick. 1998. The effects of mesquite invasion on a southeastern Arizona grassland bird community. Wilson Bulletin 110:403–408.
- Lowe, C. H., and T. B. Johnson. 1976. A survey of the reptiles and amphibians of the Fort Bowie National Historic Site. In Survey of vertebrate fauna of Fort Bowie Historic Site, Arizona. Cooperative National Park Resources Studies Unit. Technical Report No. 2. University of Arizona, Tucson, AZ.

- MacArthur, R. H., and J. W. MacArthur. 1961. On bird species diversity. Ecology 42:594–598.
- Mills, G. S., J. B. Dunning Jr., and J. M. Bates. 1989. Effects of urbanization of breeding bird community structure in southwestern desert habitats. Condor 91:416–428.
- National Park Service (NPS). 1992. NPS-75: Natural resources inventory and monitoring guidelines. U.S. Department of the Interior, Washington, D.C.
- National Park Service (NPS). 2000a. Annotated checklist of vascular plants for Fort Bowie National Historic Site. Unpublished report.
- National Park Service (NPS). 2000b. Natural and cultural resources management plan. Fort Bowie National Historic Site, AZ.
- National Park Service (NPS). 2001. Final environmental impact statement general management plan. Fort Bowie National Historic Site, AZ.
- National Park Service (NPS). 2003. Resource issues in southern U.S. border parks from drug trafficking and undocumented alien activity. Southern Arizona Office, Phoenix, AZ.
- National Park Service (NPS). 2005. NPS Visitation Database Reports. Accessed January 24, 2005 from: http://www2.nature. nps.gov/NPstats/npstats.cfm.
- North American Ornithological Atlas Committee (NAOAC). 1990. Handbook for atlasing North American breeding birds. C. Smith, editor. Accessed 13 July 2001 from: http://americanbirding.org/norac/atlascont.htm.
- Peterjohn, B. G., and J. R. Sauer. 1999.

 Population status of North American grassland birds from the North American Breeding Bird Survey, 1966-1996. Studies in Avian Biology 19:27-44.
- Petryszyn, Y. 1999. Rodent monitoring at Fort Bowie National Historic Site. Unpublished report to Fort Bowie National Historic Site.
- Powell, B. F., and R. J. Steidl. 2000. Nesting habitat and reproductive success of southwestern riparian birds. Condor 102:823–831.

- Powell, B. F., and R. J. Steidl. 2002. Habitat selection by riparian songbirds breeding in southern Arizona. Journal of Wildlife Management 66:1096–1103.
- Powell, B. F., E. W. Albrecht, W. L. Halvorson, C.
 A. Schmidt, P. Anning, and K. Docherty.
 2005a. Vascular plant and vertebrate
 inventory of Tumacácori National Historical
 Park. USGS Open-File report 2005-1142.
 U.S. Geological Survey, Southwest Biological
 Science Center, Sonoran Desert Research
 Station, University of Arizona, Tucson, AZ.
- Powell, B. F., C. A. Schmidt, and W. L. Halvorson. 2005b. Vascular plant and vertebrate inventory of Fort Bowie National Historic Park. Final report to the National Park Service, Sonoran Desert Inventory and Monitoring Program, Tucson, AZ.
- Powell, B. F., E. W. Albrecht, and W. L. Halvorson. 2004. Biological inventory report for the Sonoran Desert Network: 2003. Annual Report No. 3. Sonoran Desert Network Inventory Program. U.S. Geological Survey, Sonoran Desert Research Station and School of Natural Resources, University of Arizona, Tucson, AZ.
- Powell, B. F., E. W. Albrecht, W. L. Halvorson, and K. Docherty. 2003. Biological inventory report for the Sonoran Desert Network: 2002. Annual Report No. 2. Sonoran Desert Network Inventory Program. U.S. Geological Survey, Sonoran Desert Field Station and School of Natural Resources, University of Arizona, Tucson, AZ.
- Ralph, C. J., J. R. Sauer, S. Droege, technical editors. 1995. Monitoring bird populations by point counts. Gen. Tech. Rep. PSW-GTR-149. Pacific Southwest Research Station, Forest Service, U.S. Department of Agriculture, Albany, CA.
- Rappole, J. H. 1995. The ecology of migrant birds: a neotropical perspective. Smithsonian Institution Press, Washington D.C.
- Reynolds, R. T., J. M. Scott, and R. A. Nussbaum. 1980. A variable circular-plot method for estimating bird numbers. Condor 82:309–313.

- Rice, J., B. W. Anderson, and R. D. Ohmart. 1984. Comparison of the importance of different habitat attributes to avian community organization. Journal of Wildlife Management 48:895–911.
- Riley, S. P. D., R. M. Sauvajot, T. K. Fuller, E. C. York, D. A. Kamradt, C. Bromley, and R. K. Wayne. 2003. Effects of urbanization and habitat fragmentation on bobcats and coyotes in southern California. Conservation Biology 17:566–576.
- Rosen, P. C., and C. H. Lowe. 1994. Highway mortality of snakes in the Sonoran desert of southern Arizona. Biological Conservation 68:143–148.
- Roth, R. L. 1976. Spatial heterogeneity and bird species diversity. Ecology 57:773–782.
- Roth, E. L., and E. L. Cockrum. 1976. A survey of the mammals of the Fort Bowie National Historic Site. In Survey of vertebrate fauna of Fort Bowie Historic Site, Arizona. Technical Report No. 2. Cooperative National Park Resources Studies Unit. University of Arizona, Tucson, AZ.
- Russell, S. M. and T. B. Johnson. 1976. A survey of the birds of the Fort Bowie National Historic Site. In Survey of vertebrate fauna of Fort Bowie Historic Site, Arizona. Technical Report No. 2. Cooperative National Park Resources Studies Unit. University of Arizona, Tucson, AZ.
- Ruyle, G. B. 2001. Range resources inventory and vegetation monitoring for Fort Bowie National Historic Site. Unpublished report from School of Renewable Natural Resources, University of Arizona, Tucson, AZ.
- Saab, V. A., C. E. Bock, T. D. Rich, and D. S.
 Dobkin. 1995. Livestock grazing effects in western North America. Pp. 311–353. In T.
 E. Martin and D. M. Finch, editors. Ecology and management of neotropical migratory birds: a synthesis and review of critical issues. Oxford University Press, London, England.
- Sauer, J. R., J. E. Hines, and J. Fallon. 2004. The North American Breeding Bird Survey, results and analysis 1966–2003. Version 2004.1.

- Patuxtant Wildlife Research Center, Laurel MD. http://www.mbr-pwrc.usgs.gov/bbs/bbs.html.
- Schwalbe, C. R., and P. C. Rosen. 1988.
 Preliminary report on effects of bullfrogs on wetland herpetofauna in southeastern Arizona.
 Pp. 166–173. In R. C. Szaro, K. E. Severson, and D. R. Patton, editors. Management of amphibians, reptiles, and small mammals in North America. Gen. Tech. Rep. RM-166, U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station, Fort Collins, CO.
- Seabloom, E. W., E. T. Borer, V. L. Boucher, R. S. Burton, K. L. Cottingham, L. Goldwasser, W. K. Gram, B. E. Kendall, and F. Micheli.
 2003. Competition, seed limitation, disturbance, and reestablishment of California native annual forbs. Ecological Applications 13:575–592.
- Stebbins, R. C. 2003. A field guide to western reptiles and amphibians. Third edition. Houghton Mifflin, New York, NY.
- Stohlgren, T. J., J. F. Quinn, M. Ruggiero, and G.S. Waggoner. 1995. Status of biotic inventories in U.S. national parks. Biological Conservation 71:97–106.
- Strong, T. R., and C. E. Bock. 1990. Bird species distribution patterns in riparian habitats in southeastern Arizona. Condor 92:511–519.
- Swann, D. E., M. J. Goode, and C. R. Schwalbe. 2001. Inventory and recommendations for long-term monitoring of reptiles and amphibians at Fort Bowie National Historic Site, Arizona. Unpublished report to Southern Arizona Office, National Park Service, Phoenix, Arizona.
- Szaro, R. C., and M. D. Jakle. 1985. Avian use of a desert riparian island and its adjacent scrub habitat. Condor 87:511–519.
- Theobald, D. M., J. R. Miller, and N. T. Hobbs. 1997. Estimating the cumulative effects of development on wildlife habitat. Landscape and Urban Planning 39:25–36.
- Thompson, S. K. 1992. Sampling. John Wiley and Sons, New York, NY.

- Trombulak, S. C., and C. A. Frissell. 2000. Review of ecological effects of roads on terrestrial and aquatic communities. Conservation Biology 14:18–30.
- United States Department of Agriculture (USDA). 2004. The PLANTS Database, Version 3.5 (http://plants.usda.gov). National Plant Data Center, Natural Resources Conservation Service, Baton Rouge, LA.
- Van Auken, O. W. 2000. Shrub invasions of North American semiarid grasslands. Annual Review of Ecology and Systematics 31:197–215.
- Verner, J., and L. V. Ritter. 1983. A comparison of transects and point counts in oak-pine woodlands of California. Condor 87:47–68.
- Wake, D. B. 1990. Declining amphibian populations. Science 40:418.
- Warren, P. L., M. S. Hoy, and W. E. Hoy. 1992.
 Vegetation and flora of Fort Bowie National
 Historic Site, Arizona. National Park Service
 Technical Report NPS/WRUA/NRTR-92/43.
- Western Regional Climate Center (WRCC). 2005. Arizona climate sumaries from Bowie, Arizona: http://www.wrcc.dri.edu/summary/climsmaz.html.

Appendix A. Plant species that were observed or collected at Fort Bowie NHS by the University of Arizona inventory personnel (UA; 2002 and 2003) and other studies: Warren et al. (1992; WEA), Bennett et al. (1996; BEA), Ruyle (1996; RUL), Hartman and Rottman (1998; H&R), NPS (2000; NPS), and Halvorson and Guertin (2003; H&G).^a Species in bold-faced type are non-native.

·	adda type are non nauve.			UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Acanthaceae	Anisacanthus thurberi (Torr.) Gray	Thurber's desert honeysuckle	Χ		Χ	Χ	Χ	Χ	Χ	
	Carlowrightia arizonica Gray	Arizona wrightwort			Χ	Χ		Χ	Χ	
Agavaceae	Agave palmeri Engelm.	Palmer's century plant			Χ	Χ		Χ	Χ	
	Agave parryi Engelm.	Parry's agave	X		Χ	Χ		Χ	Χ	
	Yucca baccata Torr.	banana yucca	Χ		Χ			Χ	Χ	
	Yucca baccata var. brevifolia (Schott ex Torr.) L. Benson & Darrow	Spanish dagger				Χ				
	Yucca elata (Engelm.) Engelm.	soaptree yucca			Χ	Χ		Χ	Χ	
Aizoaceae	Trianthema portulacastrum L.	desert horsepurslane	Χ		Χ	Χ		Χ	Χ	
Amaranthaceae	Amaranthus fimbriatus (Torr.) Benth. ex S. Wats.	fringed amaranth				Χ			Χ	
	Amaranthus palmeri S. Wats.	carelessweed			Χ	Χ		Χ	Χ	
	Froelichia arizonica Thornb. ex Standl.	Arizona snakecotton	Χ		Χ	Χ		Χ	Χ	
	Gomphrena caespitosa Torr.	tufted globe amaranth			Χ	Χ		Χ	Χ	
	Guilleminea densa (Humb. & Bonpl. ex Willd.) Moq.	small matweed	Χ	Χ	Χ				Χ	
	Guilleminea densa var. densa (Humb. & Bonpl. ex Willd.) Moq.	small matweed				Χ		Χ		
Anacardiaceae	Rhus aromatica Ait.	fragrant sumac	Χ							
	Rhus microphylla Engelm. ex Gray	littleleaf sumac	Χ	Χ	Χ	Χ		Χ	Χ	
	Rhus trilobata Nutt.	skunkbush sumac			Χ			Χ	Χ	
	Rhus trilobata var. pilosissima Engelm.	pubescent squawbush				Χ		Χ	Χ	
	Rhus virens var. choriophylla (Woot. & Standl.) L. Benson	evergreen sumac			Χ	Χ		Χ	Χ	
	Toxicodendron radicans (L.) Kuntze	eastern poison ivy			Χ				Χ	
	Toxicodendron radicans ssp. divaricatum (Greene) Gillis	eastern poison ivy				Χ				
	Toxicodendron radicans ssp. radicans (L.) Kuntze	eastern poison ivy	Χ					Χ		
	Toxicodendron rydbergii (Small ex Rydb.) Greene	western poison ivy				Χ				
Apiaceae	Cymopterus multinervatus (Coult. & Rose) Tidestrom	purplenerve springparsley			Χ	Χ		Χ	Χ	
	Daucus pusillus Michx.	American wild carrot			Χ	Χ		Χ	Χ	
	Lomatium nevadense (S. Wats.) Coult. & Rose	Nevada biscuitroot			Χ			Χ	Χ	
	Lomatium nevadense var. parishii (Coult. & Rose) Jepson	Parish's biscuitroot				Χ				
	Pseudocymopterus montanus (Gray) Coult. & Rose	alpine false springparsley			Χ	Χ		Χ	Χ	
	Spermolepis echinata (Nutt. ex DC.) Heller	bristly scaleseed			Χ	Χ		Χ	Χ	
Apocynaceae	Macrosiphonia brachysiphon (Torr.) Gray	Huachuca Mountain rocktrumpet			Χ	Χ		Χ	Χ	
Aristolochiaceae	Aristolochia watsonii Woot. & Standl.	Watson's dutchman's pipe			Χ	Χ		Χ	Χ	
Asclepiadaceae	Asclepias asperula (Dcne.) Woods.	spider milkweed	Χ		Χ			Χ	Χ	
1	Asclepias asperula (Dcne.) Woods. ssp. asperula	spider milkweed				Χ				

California goldfields

hoary tansyaster

Common name

New Mexico thistle

UA Herb-UA arium^b WEA BEA

H&R NPS H&G

Χ

RUL

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Family

Asteraceae

Scientific name

Cirsium neomexicanum Gray

Lasthenia californica DC. ex Lindl.

Machaeranthera canescens var. incana (Lindl.) Gray

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Asteraceae	Machaeranthera gracilis (Nutt.) Shinners	slender goldenweed				Χ				
	Machaeranthera pinnatifida (Hook.) Shinners	lacy tansyaster			Χ				Χ	
	Machaeranthera pinnatifida ssp. gooddingii (A. Nels.) B.L.	Goodding's tansyaster				Χ				
	Machaeranthera pinnatifida var. pinnatifida (Hook.) Shinners	lacy tansyaster				Χ		Χ		
	Machaeranthera tagetina Greene	mesa tansyaster			Χ	Χ		Χ	Χ	
	Malacothrix fendleri Gray	Fendler's desertdandelion			Χ	Χ		Χ	Χ	
	Packera neomexicana var. neomexicana (Gray) W.A. Weber & A. Löve	New Mexico groundsel			Χ	Χ		Χ	Χ	
	Packera quercetorum (Greene) C. Jeffrey	Oak Creek ragwort			Χ	Χ		Χ	Χ	
	Parthenium incanum Kunth	mariola	Χ		Χ	Χ	Χ	Χ	Χ	
	Pectis filipes Harvey & Gray	fivebract cinchweed			Χ			Χ	Χ	
	Pectis filipes var. subnuda Fern.	fivebract cinchweed				Χ				
	Pectis longipes Gray	longstalk cinchweed			Χ	Χ		Χ	Χ	
	Pectis prostrata Cav.	spreading cinchweed			Χ	Χ		Χ	Χ	
	Pseudognaphalium canescens ssp. canescens (DC.) W.A. Weber	Wright's cudweed	Χ		Χ	Χ		Χ	Χ	
	Psilostrophe cooperi (Gray) Greene	whitestem paperflower				Χ	Χ		Χ	
	Psilostrophe sparsiflora (Gray) A. Nels.	greenstem paperflower			Χ	Χ		Χ	Χ	
	Psilostrophe tagetina (Nutt.) Greene	woolly paperflower	Χ		Χ			Χ	Χ	
	Psilostrophe tagetina (Nutt.) Greene var. tagetina	woolly paperflower				Χ				
	Rafinesquia neomexicana Gray	New Mexico plumseed			Χ	Χ			Χ	
	Ratibida columnifera (Nutt.) Woot. & Standl.	upright prairie coneflower			Χ	Χ		Χ	Χ	
	Sanvitalia abertii Gray	Albert's creeping zinnia	Χ		Χ	Χ		Χ	Χ	
	Schkuhria wislizenii Gray		Χ							
	Senecio flaccidus var. douglasii (DC.) B.L. Turner & T.M. Barkl.	Douglas' ragwort	Χ			Χ				
	Senecio flaccidus var. flaccidus Less.	threadleaf ragwort			Χ	Χ		Χ	Χ	
	Senecio flaccidus var. monoensis (Greene) B.L. Turner & T.M. Barkl.	Mono ragwort			Χ			Χ	Χ	
	Senecio spartioides var. multicapitatus (Greenm. ex Rydb.) Welsh	broomlike ragwort	Χ		Χ	Χ		Χ	Χ	
	Solidago velutina DC.	threenerve goldenrod			Χ	Χ		Χ	Χ	
	Sonchus oleraceus L.	common sowthistle			Χ	Χ			Χ	
	Stephanomeria pauciflora (Torr.) A. Nels.	brownplume wirelettuce	Χ		Χ	Χ		Χ	Χ	
	Tagetes micrantha Cav.	licorice marigold	Χ							
	Thelesperma longipes Gray	longstalk greenthread			Χ	Χ		Χ	Χ	
	Thelesperma megapotamicum (Spreng.) Kuntze	Hopi tea greenthread	Χ		Χ	X		Χ	Χ	
	Thymophylla acerosa (DC.) Strother	pricklyleaf dogweed			Χ	Χ		Χ	Χ	
	Thymophylla pentachaeta var. pentachaeta (DC.) Small	fiveneedle pricklyleaf			X	X		Χ	X	
	Trixis californica Kellogg	American threefold			X	X		Χ	X	
	Uropappus lindleyi (DC.) Nutt.	Lindley's silverpuffs			X	X		Χ	X	

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Asteraceae	Verbesina encelioides (Cav.) Benth. & Hook. f. ex Gray	golden crownbeard			Χ	Χ		Χ	Χ	
	Verbesina rothrockii Robins. & Greenm.	Rothrock's crownbeard	Χ	Χ	Χ	Χ		Χ	Χ	
	Viguiera deltoidea Gray	Parish's goldeneye			Χ				Χ	
	Viguiera dentata (Cav.) Spreng.	toothleaf goldeneye			Χ	Χ		Χ	Χ	
	Viguiera parishii Greene	Parish's goldeneye				Χ		Χ		
	Zinnia acerosa (DC.) Gray	desert zinnia	Χ		Χ	Χ		Χ	Χ	
	Zinnia grandiflora Nutt.	Rocky Mountain zinnia			Χ	Χ		Χ	Χ	
Bignoniaceae	Chilopsis linearis (Cav.) Sweet	desert willow	Χ		Χ		X	Χ	Χ	
	Chilopsis linearis (Cav.) Sweet ssp. linearis	desert willow				Χ				
Boraginaceae	Amsinckia menziesii var. intermedia (Fisch & C.A. Mey.) Ganders	common fiddleneck			Χ	Χ		Χ	Χ	
	Cryptantha crassisepala (Torr. & Gray) Greene	thicksepal cryptantha			Χ	Χ		Χ	Χ	
	Cryptantha micrantha (Torr.) I.M. Johnston	redroot cryptantha			Χ	Χ		Χ	Χ	
	Cryptantha pterocarya (Torr.) Greene	wingnut cryptantha	Χ							
	Lappula occidentalis (S. Wats.) Greene	flatspine stickseed	Χ							
	Lappula occidentalis var. cupulata (Gray) Higgins	flatspine stickseed			Χ			Χ	Χ	
	Lappula occidentalis var. occidentalis (S. Wats.) Greene	flatspine stickseed			Χ	Χ		Χ	Χ	
	Lithospermum cobrense Greene	smooththroat stoneseed			Χ	Χ		Χ	Χ	
	Lithospermum incisum Lehm.	narrowleaf stoneseed				Χ				
	Pectocarya platycarpa (Munz & Johnston) Munz & Johnston	broadfruit combseed				Χ				
	Pectocarya recurvata I.M. Johnston	curvenut combseed			Χ	Χ		Χ	Χ	
	Plagiobothrys arizonicus (Gray) Greene ex Gray	Arizona popcornflower			Χ	Χ		Χ	Χ	
	Tiquilia canescens (DC.) A. Richards.	woody crinklemat			Χ			Χ	Χ	
	Tiquilia canescens (DC.) A. Richards. var. canescens	woody crinklemat				Χ				
Brassicaceae	Arabis perennans S. Wats.	perennial rockcress	Χ		Χ	Χ		Χ	Χ	
	Descurainia pinnata (Walt.) Britt.	western tansymustard	Χ		Χ			Χ	Χ	
	Descurainia pinnata ssp. glabra (Woot. & Standl.) Detling	western tansymustard				Χ				
	Descurainia sophia (L.) Webb ex Prantl	herb sophia			Χ	Χ		χ	Χ	Х
	Draba cuneifolia Nutt. ex Torr. & Gray	wedgeleaf draba			Χ			Χ	Χ	
	Draba cuneifolia Nutt. ex Torr. & Gray var. cuneifolia	wedgeleaf draba				Χ				
	Draba standleyi J.F. Macbr. & Payson	Standley's draba				Χ				
•••••	Lepidium lasiocarpum Nutt.	shaggyfruit pepperweed			Χ			Χ	Χ	
	Lepidium lasiocarpum Nutt. var. lasiocarpum	shaggyfruit pepperweed				Χ				
	Lepidium thurberi Woot.	Thurber's pepperweed			Χ	Χ		Χ	Χ	
	Lepidium virginicum var. medium (Greene) C.L. Hitchc.	medium pepperweed			Χ	X		X	Χ	
	Lesquerella fendleri (Gray) S. Wats.	Fendler's bladderpod			Χ	X		X	X	
	Lesquerella gordonii (Gray) S. Wats.	Gordon's bladderpod			Χ	X		X	X	

				UA Herb-						
Family	Scientific name		JA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Brassicaceae	Lesquerella tenella A. Nels.	Moapa bladderpod X	(
	Pennellia longifolia (Benth.) Rollins	longleaf mock thelypody			Χ			Χ	Χ	
	Schoenocrambe linearifolia (Gray) Rollins	slimleaf plainsmustard			Χ	Χ		Χ	Χ	
	Schoenocrambe linifolia (Nutt.) Greene	flaxleaf plainsmustard X	(
	Sisymbrium irio L.	London rocket X	(Χ	Χ		Χ	Χ	Χ
	Streptanthella longirostris (S. Wats.) Rydb.	longbeak streptanthella			Χ	Χ		Χ	Χ	
	Streptanthus carinatus ssp. arizonicus (S. Wats.) Kruckeberg, Rodman & Worthington	lyreleaf jewelflower			Χ	Χ		Χ	Χ	
	Thelypodium wrightii Gray	Wright's thelypody			Χ			Χ	Χ	
	Thelypodium wrightii Gray ssp. wrightii	Wright's thelypody				Χ				
Cactaceae	Echinocereus coccineus Engelm. var. coccineus	scarlet hedgehog cactus							Χ	
	Echinocereus fendleri (Engelm.) F. Seitz	pinkflower hedgehog cactus X	(Χ	Χ	
	Echinocereus fendleri var. fasciculatus (Engelm. ex B.D. Jackson) N.P. Taylor	pinkflower hedgehog cactus							Χ	
	Echinocereus fendleri var. ledingii (Peebles) N.P. Taylor	Leding's hedgehog cactus			Χ	Χ			Χ	
	Echinocereus fendleri var. rectispinus (Peebles) L. Benson	pinkflower hedgehog cactus			Χ	Χ		Χ	Χ	
	Echinocereus pectinatus (Scheidw.) Engelm.	rainbow cactus				Χ				
	Echinocereus rigidissimus (Engelm.) Haage f.	rainbow hedgehog cactus			Χ			Χ	Χ	
	Echinocereus triglochidiatus Engelm.	kingcup cactus							Χ	
	Escobaria vivipara var. bisbeeana (Orcutt) D.R. Hunt	Bisbee spinystar			Χ	Χ			Χ	
	Escobaria vivipara var. vivipara (Nutt.) Buxbaum	spinystar X	(Χ		
	Ferocactus wislizeni (Engelm.) Britt. & Rose	candy barrelcactus X	(Χ	Χ	Χ	Χ	Χ	
	Mammillaria grahamii var. grahamii Engelm.	Graham's nipple cactus			Χ	Χ		Χ	Χ	
	Mammillaria grahamii var. oliviae (Orcutt) L. Benson	Graham's nipple cactus				Χ			Χ	
	Mammillaria heyderi var. macdougalii (Rose) L. Benson	Macdougal's nipple cactus			Χ	Χ		Χ	Χ	
	Opuntia chlorotica Engelm. & Bigelow	dollarjoint pricklypear			Χ	Χ		Χ	Χ	
	Opuntia engelmannii Salm-Dyck	cactus apple X	(Χ	
	Opuntia engelmannii Salm-Dyck var. engelmannii	cactus apple			Χ	Χ		Χ	Χ	
	Opuntia kleiniae DC.	candle cholla			Χ				Χ	
	Opuntia leptocaulis DC.	Christmas cactus			Χ	Χ		Χ	Χ	
	Opuntia macrocentra Engelm.	purple pricklypear							Χ	
	Opuntia macrocentra var. macrocentra Engelm.	purple pricklypear X	(Χ	Χ		Χ	Χ	
	Opuntia martiniana (L. Benson) Parfitt	seashore cactus							Χ	
	Opuntia phaeacantha var. major Engelm.	Mojave pricklypear X	(Χ	Χ		Χ	Χ	
	Opuntia santa-rita (Griffiths & Hare) Rose	Santa Rita pricklypear				Χ				
	Opuntia spinosior (Engelm.) Toumey	walkingstick cactus X	(Χ	Χ		Χ	Χ	
	Opuntia ?tetracantha Toumey (pro sp.)	[acanthocarpa ? leptocaulis]				Χ		Χ		
	Peniocereus greggii var. greggii (Engelm.) Britt. & Rose	nightblooming cereus			Χ	Χ		Χ	Χ	

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Cactaceae	Peniocereus greggii var. greggii (Engelm.) Britt. & Rose	nightblooming cereus			Χ	Χ		Χ	Χ	
Campanulaceae	Nemacladus glanduliferus Jepson	glandular threadplant	Χ							
	Triodanis perfoliata (L.) Nieuwl.	clasping Venus' looking-glass			Χ			Χ	Χ	
	Triodanis perfoliata var. perfoliata (L.) Nieuwl.	clasping Venus' looking-glass				Χ				
Capparaceae	Polanisia dodecandra (L.) DC.	redwhisker clammyweed			Χ				Χ	
	Polanisia dodecandra ssp. trachysperma (Torr. & Gray) Iltis	sandyseed clammyweed				Χ		Χ		
Caryophyllaceae	Silene antirrhina L.	sleepy silene	Χ							
Chenopodiaceae	Atriplex canescens (Pursh) Nutt.	fourwing saltbush			Χ	Χ		Χ	Χ	
	Atriplex elegans (Moq.) D. Dietr.	wheelscale saltbush			Χ			Χ	Χ	
	Atriplex elegans (Moq.) D. Dietr. var. elegans	wheelscale saltbush				Χ				
	Chenopodium desiccatum A. Nels.	aridland goosefoot			Χ				Χ	
	Chenopodium fremontii S. Wats.	Fremont's goosefoot			Χ			Χ	Χ	
	Chenopodium fremontii S. Wats. var. fremontii	Fremont's goosefoot				Χ				
	Chenopodium leptophyllum (Moq.) Nutt. ex S. Wats.	narrowleaf goosefoot				Χ				
	Chenopodium murale L.	nettleleaf goosefoot				Χ			Χ	
	Chenopodium pratericola Rydb.	desert goosefoot	Χ					Χ		
	Chenopodium watsonii A. Nels.	Watson's goosefoot			Χ	Χ		Χ	Χ	
	Krascheninnikovia lanata (Pursh) A.D.J. Meeuse & Smit	winterfat	Χ		Χ	Χ	Χ	Χ	Χ	
	Salsola kali L.	Russian thistle	Χ			Χ				
	Salsola tragus L.	prickly Russian thistle			Χ			Χ	Χ	
Commelinaceae	Commelina dianthifolia Delile	birdbill dayflower			Χ	Χ		Χ	Χ	
Convolvulaceae	Convolvulus arvensis L.	field bindweed	Х		Χ	Χ			Χ	
	Convolvulus equitans Benth.	Texas bindweed						Χ	Χ	
	Evolvulus nuttallianus J.A. Schultes	shaggy dwarf morning-glory	Χ							
	Evolvulus sericeus Sw.	silver dwarf morning-glory			Χ			Χ	Χ	
	Evolvulus sericeus var. sericeus Sw.	silver dwarf morning-glory				Χ				
	Ipomoea barbatisepala Gray	canyon morning-glory			Χ	Χ		Χ	Χ	
	Ipomoea coccinea L.	redstar			Χ	••••••		χ	Χ	
	Ipomoea hederacea Jacq.	ivyleaf morning-glory						Χ		
	Ipomoea hederifolia L.	scarletcreeper				Χ		Χ		
	Ipomoea purpurea (L.) Roth	tall morning-glory			χ	X		, ,	Х	Χ
Crassulaceae	Sedum cockerellii Britt.	Cockerell's stonecrop			Χ	X		Χ	X	^
Cucurbitaceae	Apodanthera undulata Gray	melon loco			X	X		X	X	
Guduibilaceae	Cucurbita digitata Gray	fingerleaf gourd			X	X		X	Χ	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,
	Outurbita Ulyitata Oray	iiigeneai godiu			Λ	^		^	χ	

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Cucurbitaceae	Marah gilensis Greene	Gila manroot			Χ	X		Χ	Χ	
Cupressaceae	Juniperus coahuilensis (Martinez) Gaussen ex R.P. Adams	redberry juniper			Χ	Χ			Χ	
	Juniperus deppeana Steud.	alligator juniper			Χ	Χ		Χ	Χ	
	Juniperus monosperma (Engelm.) Sarg.	oneseed juniper	Χ				Χ	Χ		
Cyperaceae	Carex sp. L.	sedge					Χ			
	Cyperus esculentus L.	chufa flatsedge			Χ	Χ		Χ	X	Χ
	Cyperus sphaerolepis Boeckl.	Rusby's flatsedge			Χ	Χ		Χ	Χ	
	Cyperus squarrosus L.	bearded flatsedge			Χ	Χ		Χ	Χ	
Dryopteridaceae	Woodsia mexicana Fée	phanerophlebia			Χ			Χ	Χ	
Ephedraceae	Ephedra trifurca Torr. ex S. Wats.	longleaf jointfir			Χ	Χ	Χ	Χ	Χ	
Ericaceae	Arctostaphylos pringlei Parry	Pringle manzanita				Χ			Χ	
	Arctostaphylos pungens Kunth	pointleaf manzanita	Χ		Χ	Χ		Χ	Χ	
Euphorbiaceae	Acalypha neomexicana MuellArg.	New Mexico copperleaf	Χ		Χ	Χ		Χ	Χ	
	Chamaesyce albomarginata (Torr. & Gray) Small	whitemargin sandmat	Χ		Χ	Χ		Χ	Χ	
	Chamaesyce hyssopifolia (L.) Small	hyssopleaf sandmat			Χ	Χ		Χ	Χ	
	Chamaesyce revoluta (Engelm.) Small	threadstem sandmat	Χ		Χ	Χ		Χ	Χ	
	Chamaesyce serpyllifolia ssp. serpyllifolia (Pers.) Small	thymeleaf sandmat	Χ							
	Chamaesyce serrula (Engelm.) Woot. & Standl.	sawtooth sandmat	Χ							
	Chamaesyce stictospora (Engelm.) Small	slimseed sandmat			Χ	Χ		Χ	Χ	
	Croton pottsii var. pottsii (Klotzsch) MuellArg.	leatherweed	Χ		Χ	Χ		Χ	Χ	
	Euphorbia bilobata Engelm.	blackseed spurge	Χ							
	Euphorbia exstipulata Engelm.	squareseed spurge	Χ							
	Euphorbia heterophylla L.	Mexican fireplant	Χ							
	Tragia nepetifolia Cav.	catnip noseburn			Χ				Χ	
	Tragia ramosa Torr.	branched noseburn	Χ			Χ		Χ	Χ	
Fabaceae	Acacia angustissima (P. Mill.) Kuntze	prairie acacia			Χ			Χ	Χ	
	Acacia angustissima var. suffrutescens (Rose) Isely	prairie acacia				Χ				
	Acacia constricta Benth.	whitethorn acacia			Χ	Χ	Χ	Χ	Χ	
	Acacia greggii Gray	catclaw acacia			Χ	Χ	Χ	Χ	Χ	
	Amorpha fruticosa L.	desert false indigo			Χ	Χ		Χ	Χ	
	Astragalus allochrous Gray	halfmoon milkvetch			Χ	Χ		Χ	Χ	
•••••	Astragalus allochrous var. playanus Isely	halfmoon milkvetch			Χ	Χ		Χ	Χ	
	Astragalus arizonicus Gray	Arizona milkvetch			Χ	Χ		Χ	Χ	
	Astragalus calycosus Torr. ex S. Wats.	Torrey's milkvetch			Χ			Χ	Χ	
	Astragalus calycosus Torr. ex S. Wats. var. calycosus	Torrey's milkvetch			••••••	Χ				
	Astragalus nothoxys Gray	sheep milkvetch			Χ	Χ		Χ	Χ	

				UA Herb-						
Family	Scientific name	Common name	UA	arium ^b	WEA	BEA	RUL	H&R	NPS	H&G
Fabaceae	Astragalus nuttallianus DC.	smallflowered milkvetch			X			X	X	
	Astragalus nuttallianus var. austrinus (Small) Barneby	smallflowered milkvetch				Χ				
	Astragalus tephrodes Gray	ashen milkvetch			Χ	Χ		Χ	Χ	
	Astragalus thurberi Gray	Thurber's milkvetch			Χ	Χ		Χ	Χ	
	Caesalpinia drepanocarpa (Gray) Fisher	sicklepod holdback			Χ	Χ		Χ	Χ	
	Calliandra eriophylla Benth.	fairyduster	Χ		Χ	Χ	Χ	Χ	Χ	
	Calliandra humilis Benth.	dwarf stickpea						Χ		
	Calliandra humilis Benth. var. humilis	dwarf stickpea			Χ	Χ			Χ	
	Calliandra humilis var. reticulata (Gray) L. Benson	dwarf stickpea			Χ	Χ		Χ	Χ	
	Chamaecrista nictitans (L.) Moench	partridge pea	Χ							
	Crotalaria pumila Ortega	low rattlebox	Χ							
	Dalea albiflora Gray	whiteflower prairie clover			Χ	Χ		Χ	Χ	
	Dalea candida var. oligophylla (Torr.) Shinners	white prairie clover	Χ							
	Dalea formosa Torr.	featherplume	Χ		Χ	Χ	Χ	Χ	Χ	
	Dalea nana Torr. ex Gray	dwarf prairie clover			Χ			Χ	Χ	
	Dalea nana var. carnescens Kearney & Peebles	dwarf prairie clover				Χ				
	Dalea nana Torr. ex Gray var. nana	dwarf prairie clover	Χ							
	Dalea pogonathera Gray	bearded prairie clover			Χ	Χ		Χ	Χ	
	Dalea pringlei Gray	Pringle's prairie clover	Χ							
	Dalea versicolor Zucc.	oakwoods prairie clover			Χ				Χ	
	Dalea versicolor var. sessilis (Gray) Barneby	oakwoods prairie clover	Χ			Χ		Χ		
	Dalea wrightii Gray	Wright's prairie clover	Χ		Χ	Χ		Χ	Χ	
	Desmanthus cooleyi (Eat.) Trel.	Cooley's bundleflower	Χ		Χ	Χ		Χ	Χ	
	Desmanthus virgatus (L.) Willd.	wild tantan							Χ	
	Desmanthus virgatus var. depressus (Humb. & Bonpl. ex Willd.)	B.L. Turner wild tantan				Χ				
	Desmodium neomexicanum Gray	New Mexico ticktrefoil	Χ							
	Desmodium procumbens (P. Mill.) A.S. Hitchc.	western trailing ticktrefoil			Χ			Χ	Χ	
	Desmodium procumbens var. exiguum (Gray) Schub.	western trailing ticktrefoil				Χ				
	Erythrina flabelliformis Kearney	coralbean					Χ			
	Galactia wrightii Gray	Wright's milkpea		Χ	Χ			Χ	Χ	
	Galactia wrightii var. mollissima Kearney & Peebles					Χ				
	Hoffmannseggia glauca (Ortega) Eifert	Indian rushpea			Χ	Χ		Χ	Χ	
	Lotus greenei Ottley ex Kearney & Peebles	Greene's bird's-foot trefoil			Χ	Χ		Χ	Χ	
	Lotus humistratus Greene	foothill deervetch			Χ	Χ			Χ	
	Lotus plebeius (Brand) Barneby	New Mexico bird's-foot trefoil			Χ	Χ		Χ	Χ	
	Lotus rigidus (Benth.) Greene	shrubby deervetch			Χ	Χ		Χ	Χ	

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Fabaceae	Lotus wrightii (Gray) Greene	Wright's deervetch			X	Χ		Χ	Χ	
	Lupinus brevicaulis S. Wats.	shortstem lupine			Χ	Χ		Χ	Χ	
	Lupinus concinnus J.G. Agardh	scarlet lupine			Χ	Χ		Χ	Χ	
	Lupinus sparsiflorus Benth.	Mojave Iupine			Χ	Χ		Χ	Χ	
	Macroptilium gibbosifolium (Ortega) A. Delgado	variableleaf bushbean	Х		Χ	Χ		Χ	Χ	
	Marina calycosa (Gray) Barneby	San Pedro false prairie-clover			Χ	Χ		Χ	Χ	
	Mimosa aculeaticarpa Ortega	catclaw mimosa	Χ							
	Mimosa aculeaticarpa var. biuncifera (Benth.) Barneby	catclaw mimosa			Χ	Χ	Χ	Χ	Χ	
	Phaseolus acutifolius Gray	tepary bean	Χ		Χ			Χ	Χ	
	Phaseolus acutifolius var. tenuifolius Gray	tepary bean				Χ				
	Prosopis glandulosa Torr.	honey mesquite	Χ					Χ		
	Prosopis glandulosa var. torreyana (L. Benson) M.C. Johnston	western honey mesquite			Χ	Χ			Χ	
	Prosopis velutina Woot.	velvet mesquite			Χ	Χ	Χ	Χ	Χ	
	Rhynchosia senna var. texana (Torr. & Gray) M.C. Johnston	Texas snoutbean			Χ	Χ		Χ	Χ	
	Robinia neomexicana Gray	New Mexico locust			Χ			Χ	Χ	
	Robinia neomexicana Gray var. neomexicana	New Mexico locust				Χ				
	Senna bauhinioides (Gray) Irwin & Barneby	twinleaf senna			Χ	Χ		Χ	Χ	
	Senna covesii (Gray) Irwin & Barneby	Coves' cassia				Χ			Χ	
	Vicia Iudoviciana ssp. Iudoviciana Nutt.	Louisiana vetch				Χ		Χ	Χ	
	Vicia pulchella Kunth	sweetclover vetch			Χ	Χ		Χ	Χ	
Fagaceae	Quercus arizonica Sarg.	Arizona white oak	Χ		Χ	Χ		Χ	Χ	
	Quercus dunnii Kellogg	Palmer oak			Χ	Χ		Χ	Χ	
	Quercus emoryi Torr.	Emory oak	Χ		Χ	Χ	Χ	Χ	Χ	
	Quercus grisea Liebm.	gray oak	Χ		Χ	Χ		Χ	Χ	
	Quercus hypoleucoides A. Camus	silverleaf oak			Χ	Χ		Χ	Χ	
	Quercus pungens Liebm.	pungent oak	Χ		Χ	Χ		Χ	Χ	
	Quercus rugosa Née	netleaf oak			Χ	Χ		Χ	Χ	
	Quercus toumeyi Sarg.	Toumey oak			Χ	Χ		Χ	Χ	
	Quercus turbinella Greene	Sonoran scrub oak			Χ	Χ		Χ	Χ	
Fouquieriaceae	e Fouquieria splendens Engelm.	ocotillo	Χ		Χ	Χ	Χ	Χ	Χ	
Fumariaceae	Corydalis aurea Willd.	scrambled eggs	Χ		Χ			Χ	Χ	
	Corydalis curvisiliqua ssp. occidentalis (Engelm. ex Gray) W.A. Weber	curvepod fumewort				Χ				
Garryaceae	Garrya flavescens S. Wats.	ashy silktassel				Χ			Χ	
	Garrya wrightii Torr.	Wright's silktassel	Χ		Χ	Χ		Χ	Χ	
Geraniaceae	Erodium cicutarium (L.) L'Hér. ex Ait.	redstem stork's bill	Χ		Χ			Χ	Χ	Χ
	Erodium cicutarium ssp. jacquinianum (Fisch., C.A. Mey. & Avé-Lall.) Br	ig. redstem stork's bill				Χ				

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Geraniaceae	Erodium texanum Gray	Texas stork's bill			Χ	Χ		Χ	Χ	
Hydrangeaceae	Fendlera rupicola Gray	cliff fendlerbush			Χ	Χ		Χ	Χ	
Hydrophyllaceae	Nama hispidum Gray	bristly nama			Χ	Χ		Χ	Χ	
	Phacelia arizonica Gray	Arizona phacelia			Χ	Χ		Χ	Χ	
	Phacelia congesta Hook.	caterpillars			Χ	Χ			Χ	
	Phacelia crenulata Torr. ex S. Wats.	cleftleaf wildheliotrope	Χ		Χ	Χ		Χ	Χ	
,	Phacelia rupestris Greene	rock phacelia						Χ	Χ	
Juglandaceae	Juglans major (Torr.) Heller	Arizona walnut	Χ		Χ	Χ	Χ	Χ	Χ	
Juncaceae	Juncus bufonius L.	toad rush						Χ	Χ	
	Juncus drummondii E. Mey.	Drummond's rush			Χ	Χ			Χ	
	Juncus saximontanus A. Nels.	Rocky Mountain rush			Χ	Χ		Χ	Χ	
	Juncus tenuis Willd.	poverty rush			Χ			Χ	Χ	
Krameriaceae	Krameria erecta Willd. ex J.A. Schultes	littleleaf ratany					Χ			
	Krameria lanceolata Torr.	trailing krameria			Χ			Χ	Χ	
Lamiaceae	Hedeoma drummondii Benth.	Drummond's false pennyroyal			Χ	Χ		Χ	Χ	
	Hedeoma hyssopifolia Gray	aromatic false pennyroyal				Χ		Χ	Χ	
	Hedeoma nana (Torr.) Briq. ssp. nana	dwarf false pennyroyal				Χ				
	Hedeoma nanum (Torrey) Briq.				Χ			Χ	Χ	
	Hedeoma oblongifolia (Gray) Heller	oblongleaf false pennyroyal	Χ							
Lamiaceae	Lamium amplexicaule L.	henbit deadnettle			Χ	Χ		Χ	Χ	
	Marrubium vulgare L.	horehound	Χ		Χ	Χ		Χ	X	Χ
	Salvia columbariae Benth.	chia	Χ		Χ	Χ		Χ	Χ	
	Salvia henryi Gray	crimson sage			Χ	Χ		Χ	Χ	
	Salvia lemmonii Gray	Lemmon's sage			Χ	Χ		Χ	Χ	
	Salvia subincisa Benth.	sawtooth sage	Χ		Χ	Χ		Χ	Χ	
	Stachys coccinea Ortega	scarlet hedgenettle			Χ	Χ		Χ	Χ	
	Trichostema arizonicum Gray	Arizona bluecurls	Χ		Χ	Χ		Χ	Χ	
Liliaceae	Allium acuminatum Hook.	tapertip onion				Χ			Χ	
	Allium macropetalum Rydb.	largeflower onion			Χ	Χ		Χ	Χ	
	Calochortus ambiguus (M.E. Jones) Ownbey	doubting mariposa lily			Χ	Χ		Χ	Χ	
	Dasylirion wheeleri S. Wats.	common sotol	Χ		Χ	Χ	Х	Χ	Χ	
	Dichelostemma capitatum (Benth.) Wood ssp. capitatum	wild hyacinth			Χ	Χ		Χ	Χ	
	Echeandia flavescens (J.A. & J.H. Schultes) Cruden	Torrey's craglily			Χ	Χ		Χ	Χ	
	Nolina microcarpa S. Wats.	sacahuista	Χ		Χ	Χ	Χ	Χ	Χ	
	Zephyranthes longifolia Hemsl.	copper zephyrlily			Χ	Χ		Χ	Χ	

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Linaceae	Linum lewisii Pursh	prairie flax			Χ			Χ	Χ	
	Linum lewisii Pursh var. lewisii	prairie flax				Χ				
	Linum puberulum (Engelm.) Heller	plains flax			Χ	Χ		Χ	Χ	
	Linum usitatissimum L.	common flax			X	Χ		X	Χ	
Loasaceae	Cevallia sinuata Lag.	stinging serpent	Χ		Χ	Χ		Χ	Χ	
	Mentzelia albicaulis (Dougl. ex Hook.) Dougl. ex Torr. & Gray	whitestem blazingstar	Χ		Χ	Χ		Χ	Χ	
	Mentzelia multiflora (Nutt.) Gray	Adonis blazingstar	Χ		Χ				Χ	
	Mentzelia multiflora var. integra M.E. Jones	Adonis blazingstar				Χ				
	Mentzelia pumila Nutt. ex Torr. & Gray	dwarf mentzelia						Χ		
Malpighiaceae	Janusia gracilis Gray	slender janusia			Χ	Χ	Χ	Χ	Χ	
Malvaceae	Abutilon parvulum Gray	dwarf Indian mallow			Χ	Χ		Χ	Χ	
	Gossypium thurberi Todaro	Thurber's cotton					Χ			
	Malvella lepidota (Gray) Fryxell	scurfymallow			Χ	Χ		Χ	Χ	
	Rhynchosida physocalyx (Gray) Fryxell	buffpetal			Χ	Χ		Χ	Χ	
	Sida abutifolia P. Mill.	spreading fanpetals	Х		Χ	Χ		Χ	Χ	
	Sphaeralcea hastulata Gray	spear globemallow			Χ	Χ		Χ	Χ	
	Sphaeralcea laxa Woot. & Standl.	caliche globemallow	Χ		Χ	Χ		Χ	Χ	
	Sphaeralcea wrightii Gray	Wright's globemallow				Χ			Χ	
Moraceae	Morus microphylla Buckl.	Texas mulberry	Χ	Χ	Χ	Χ		Χ	Χ	
Nyctaginaceae	Allionia incarnata L.	trailing windmills	Χ		Χ	Χ		Χ	Χ	
	Boerhavia coccinea P. Mill.	scarlet spiderling						Χ		Χ
	Boerhavia coulteri (Hook. f.) S. Wats.	Coulter's spiderling				Χ				
	Boerhavia diffusa L.	red spiderling			Χ	Χ			Χ	
	Boerhavia erecta L.	erect spiderling						Χ	Χ	
	Boerhavia intermedia M.E. Jones	fivewing spiderling			Χ	Χ			Χ	
	Boerhavia purpurascens Gray	purple spiderling	Χ		Χ	Χ		Χ	Χ	
	Boerhavia spicata Choisy	creeping spiderling	Χ		Χ	Χ		Χ	Χ	
	Mirabilis albida (Walt.) Heimerl	white four o'clock	Χ		Χ	Χ			Χ	
	Mirabilis bigelovii Gray	wishbone-bush			Χ			Χ	Χ	
	Mirabilis bigelovii Gray var. bigelovii	wishbone-bush				Χ				
	Mirabilis coccinea (Torr.) Benth. & Hook. f.	scarlet four o'clock			Χ	Χ		Χ	Χ	
	Mirabilis comata (Small) Standl.	hairy-tuft four o'clock						Χ		
	Mirabilis linearis (Pursh) Heimerl	narrowleaf four o'clock	Χ		Χ	Χ		Χ	Χ	
	Mirabilis longiflora L.	sweet four o'clock			Χ	Χ		Χ	Χ	
	Mirabilis multiflora (Torr.) Gray	Colorado four o'clock			Χ			Χ	Χ	
	Mirabilis multiflora (Torr.) Gray var. multiflora	Colorado four o'clock				Χ				

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Nyctaginaceae	Mirabilis pumila (Standl.) Standl.	dwarf four o'clock			Χ	Χ		Χ	Χ	
Oleaceae	Fraxinus velutina Torr.	velvet ash	Χ		Χ	Χ	Χ	Χ	Χ	
	Menodora scabra Gray	rough menodora			Χ	Χ		Χ	Χ	
Onagraceae	Calylophus hartwegii (Benth.) Raven	Hartweg's sundrops			Χ			Χ	Χ	
	Calylophus hartwegii ssp. pubescens (Gray) Towner & Raven	Hartweg's sundrops				Χ				
	Camissonia californica (Nutt. ex Torr. & Gray) Raven	California suncup			Χ	Χ		Χ	Χ	
	Camissonia contorta (Dougl. ex Lehm.) Kearney	plains evening-primrose			Χ			Χ	Χ	
	Epilobium canum ssp. latifolium (Hook.) Raven	hummingbird trumpet			Χ	Χ		Χ	Χ	
	Epilobium ciliatum ssp. ciliatum Raf.	fringed willowherb	Χ							
	Gaura hexandra ssp. gracilis (Woot. & Standl.) Raven & Gregory	harlequinbush			Χ	Χ		Χ	Χ	
	Oenothera brachycarpa Gray	shortfruit evening-primrose	Χ		Χ	Χ		Χ	Χ	
	Oenothera caespitosa Nutt.	tufted evening-primrose			Χ			Χ	Χ	
	Oenothera caespitosa ssp. caespitosa Nutt.	tufted evening-primrose				Χ				
	Oenothera elata ssp. hirsutissima (Gray ex S. Wats.) W. Dietr.	Hooker's evening-primrose				Χ				
	Oenothera elata ssp. hookeri (Torr. & Gray) W. Dietr. & W.L. Wagner	Hooker's evening-primrose							Χ	
	Oenothera pallida ssp. runcinata (Engelm.) Munz & W. Klein	pale evening-primrose			Χ	Χ		Χ	Χ	
	Oenothera primiveris Gray	desert evening-primrose			Χ	Χ		Χ	Χ	
Orobanchaceae	Orobanche cooperi (Gray) Heller	desert broomrape			Χ	Χ			Χ	
	Orobanche ludoviciana Nutt.	Louisiana broomrape						Χ		
Papaveraceae	Argemone pleiacantha Greene	southwestern pricklypoppy						Χ		
	Argemone pleiacantha ssp. pinnatisecta G.B. Ownbey	southwestern pricklypoppy			Χ				Χ	
	Argemone pleiacantha Greene ssp. pleiacantha	southwestern pricklypoppy				Χ				
	Eschscholzia californica ssp. mexicana (Greene) C. Clark	California poppy	Χ		Χ	Χ		Χ	Χ	
Pedaliaceae	Proboscidea parviflora (Woot.) Woot. & Standl.	doubleclaw			Χ	Χ		Χ	Χ	
Pinaceae	Pinus cembroides Zucc.	Mexican pinyon						Χ		••••••••••••
	Pinus discolor D.K. Bailey & Hawksworth	border pinyon			Χ	Χ			Χ	
	Pinus edulis Engelm.	twoneedle pinyon	Χ	Χ	Χ	Χ		Χ	Χ	
	Pinus monophylla Torr. & Frém.	singleleaf pinyon			Χ				Χ	
	Pinus monophylla var. fallax (Little) Silba	singleleaf pinyon				Χ				
Plantaginaceae	Plantago patagonica Jacq.	woolly plantain	Χ		Χ	Χ		Χ	Χ	
Platanaceae	Platanus wrightii S. Wats.	Arizona sycamore					Χ			
Poaceae	Aristida adscensionis L.	sixweeks threeawn	Χ		Χ	Χ		Χ	Χ	
	Aristida pansa Woot. & Standl.	Wooton's threeawn		Χ						
	Aristida purpurea Nutt.	purple threeawn			Χ		Χ	Χ	Χ	
	Aristida purpurea var. longiseta (Steud.) Vasey	Fendler threeawn			Χ	Χ		Χ	Χ	
	Aristida purpurea var. nealleyi (Vasey) Allred	blue threeawn		Χ	Χ	Χ		Χ	Χ	

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Poaceae	Aristida purpurea var. purpurea Nutt.	purple threeawn			V	X		V	V	
	Aristida temipes var. gentilis (Henr.) Allred	spidergrass			X	Χ		X	X	
	Aristida temipes Cav.	spidergrass			Χ			Χ	X	
	Aristida ternipes Cav. var. ternipes	spidergrass	X							
	Avena fatua L.	oat								Χ
	Bothriochloa barbinodis (Lag.) Herter	cane bluestem	Х		Χ	Χ	Χ	X	X	
	Bothriochloa saccharoides (Sw.) Rydb.	silver bluestem			Χ			Χ	Χ	
	Bouteloua aristidoides (Kunth) Griseb.	needle grama			Χ	Χ		Χ	Χ	
	Bouteloua barbata Lag.	sixweeks grama			Χ	Χ	Χ	Χ	Χ	
	Bouteloua chondrosioides (Kunth) Benth. ex S. Wats.	sprucetop grama		Χ	Χ	Χ	Χ	Χ	Χ	
	Bouteloua curtipendula (Michx.) Torr.	sideoats grama	X		Χ	Χ	Χ	Χ	Χ	
	Bouteloua eriopoda (Torr.) Torr.	black grama	Х		Χ	Χ	Χ	Χ	Χ	
	Bouteloua gracilis (Willd. ex Kunth) Lag. ex Griffiths	blue grama	Х		Χ	Χ	Χ	Χ	Χ	
	Bouteloua hirsuta Lag.	hairy grama	Х		Χ		Χ	Χ	Χ	
	Bouteloua hirsuta Lag. var. hirsuta	hairy grama				Χ				
	Bouteloua repens (Kunth) Scribn. & Merr.	slender grama	Χ	Χ	Χ	Χ	Χ	Χ	Χ	
	Bromus carinatus Hook. & Arn.	California brome								Χ
	Bromus rubens L.	red brome	X							Χ
	Bromus tectorum L.	cheatgrass			χ	Χ		Χ	Χ	
	Cenchrus spinifex Cav.	coastal sandbur			Χ	Χ		Χ	Χ	
	Chloris virgata Sw.	feather fingergrass	Χ		Χ	Χ		Χ	Χ	Χ
	Cynodon dactylon (L.) Pers.	Bermudagrass			Χ	Χ		Χ	Χ	Χ
	Dasyochloa pulchella (Kunth) Willd. ex Rydb.	low woollygrass	Х		Χ	Χ		Χ	Χ	
	Digitaria californica (Benth.) Henr.	Arizona cottontop	Χ		Χ	Χ	Χ	Χ	Χ	
	Digitaria cognata (J.A. Schultes) Pilger	Carolina crabgrass			Χ	Χ			Χ	
	Digitaria cognata (J.A. Schultes) Pilger var. cognata	Carolina crabgrass					Χ	Χ	Χ	
	Digitaria sanguinalis (L.) Scop.	hairy crabgrass			Χ	Χ		Χ	Χ	
	Echinochloa colona (L.) Link	jungle rice	Χ		χ	Χ		Χ	Χ	Χ
	Echinochloa crus-galli (L.) Beauv.	barnyardgrass			Χ	Χ		Χ	Χ	
	Elymus elymoides (Raf.) Swezey	squirreltail	Χ			Χ				
•	Elymus elymoides ssp. elymoides (Raf.) Swezey	squirreltail			Χ			Χ	Χ	
	Enneapogon desvauxii Desv. ex Beauv.	nineawn pappusgrass	Χ		Χ	Χ		Χ	Χ	
	Eragrostis cilianensis (All.) Vign. ex Janchen	stinkgrass	Χ		Χ	Χ		Χ	Χ	Χ
	Eragrostis curvula (Schrad.) Nees	weeping lovegrass	Χ	Х						Χ
	Eragrostis intermedia A.S. Hitchc.	plains lovegrass	Χ		Χ	Χ	Χ	Χ	Χ	

Texas bluestem

Grisebach's bristlegrass

streambed bristlegrass

burrograss

Common name

tufted lovegrass

desert lovegrass

Lehmann lovegrass

UA Herb-

arium⁵ WEA BEA

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

RUL

H&R

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

NPS

Χ

Χ

Χ

H&G

Χ

UA

Χ

Family

Poaceae

Scientific name

Eragrostis lehmanniana Nees

Eragrostis pectinacea (Michx.) Nees ex Steud.

Schizachyrium cirratum (Hack.) Woot. & Standl.

Setaria leucopila (Scribn. & Merr.) K. Schum.

Scleropogon brevifolius Phil.

Setaria grisebachii Fourn.

Eragrostis pectinacea var. miserrima (Fourn.) J. Reeder

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Poaceae	Setaria vulpiseta (Lam.) Roemer & J.A. Schultes	plains bristlegrass			Χ	Χ	Χ	Χ	Χ	
	Sorghum halepense (L.) Pers.	Johnsongrass								Χ
	Sporobolus airoides (Torr.) Torr.	alkali sacaton	Χ			Χ				
	Sporobolus contractus A.S. Hitchc.	spike dropseed	Χ		Χ	Χ	Χ	Χ	Χ	
	Sporobolus cryptandrus (Torr.) Gray	sand dropseed	Χ		Χ	Χ	Χ	Χ	Χ	
	Sporobolus wrightii Munro ex Scribn.	big sacaton			Χ			Χ	Χ	
	Stipa L.	needlegrass					Χ			
	Trachypogon spicatus (L.) Kuntze	spiked crinkleawn					Χ			
	Tragus berteronianus J.A. Schultes	spiked burr grass			Χ	Χ		Χ	Χ	
	Tridens muticus (Torr.) Nash	slim tridens	Χ	Χ	Χ		Χ	Χ	Χ	
	Tridens muticus var. muticus (Torr.) Nash	slim tridens				Χ				
	Urochloa arizonica (Scribn. & Merr.) O. Morrone & F. Zuloaga	Arizona signalgrass	Χ	Χ						
	Vulpia octoflora (Walt.) Rydb.	sixweeks fescue	Χ		Χ			Χ	Χ	
	Vulpia octoflora var. octoflora (Walt.) Rydb.	sixweeks fescue				Χ			Χ	
Polemoniaceae	Allophyllum gilioides (Benth.) A.& V. Grant	dense false gilyflower			Χ				Χ	
	Allophyllum gilioides (Benth.) A.& V. Grant ssp. gilioides	dense false gilyflower				Χ		Χ		
	Eriastrum diffusum (Gray) Mason	miniature woollystar	Χ		Χ	Χ		Χ	Χ	
	Gilia mexicana A.& V. Grant	El Paso gilia	Χ							
	Gilia ophthalmoides Brand	eyed gilia			Χ	Χ			Χ	
	Gilia sinuata Dougl. ex Benth.	rosy gilia			Χ	Χ		Χ	Χ	
	Ipomopsis longiflora (Torr.) V. Grant	flaxflowered ipomopsis			Χ			Χ	Χ	
	Ipomopsis longiflora (Torr.) V. Grant ssp. longiflora	flaxflowered ipomopsis				Χ				
	Ipomopsis multiflora (Nutt.) V. Grant	manyflowered ipomopsis			Χ	Χ		Χ	Χ	
	Linanthus aureus (Nutt.) Greene	golden linanthus	Χ		Χ	Χ		Χ	Χ	
	Phlox austromontana Coville	mountain phlox			Χ			Χ	Χ	
	Phlox austromontana Coville ssp. austromontana	mountain phlox				Χ				
	Phlox gracilis ssp. gracilis (Hook.) Greene	slender phlox			Χ	Χ		Χ	Χ	
	Phlox nana Nutt.	Santa Fe phlox			Χ				Χ	
	Phlox triovulata ex Torr.	threeseed phlox				Χ		Χ		
Polygalaceae	Polygala barbeyana Chod.	blue milkwort			Χ	Χ		Χ	Χ	
	Polygala barbeyana Chod. ssp. barbeyana		Χ							
	Polygala macradenia Gray	glandleaf milkwort			Χ	Χ		Χ	Χ	
	Eriogonum abertianum Torr.	Abert's buckwheat			Χ			Χ	Χ	
	Eriogonum abertianum Torr. var. abertianum	Abert's buckwheat				Χ				
	Eriogonum deflexum Torr.	flatcrown buckwheat			Χ			Χ	Χ	
	Eriogonum deflexum Torr. var. deflexum	flatcrown buckwheat				Χ				

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Polygonaceae	Eriogonum jamesii Benth.	James' buckwheat						Χ	Χ	
	Eriogonum jamesii var. undulatum (Benth.) S. Stokes ex M.E. Jones	James' buckwheat				Χ				
	Eriogonum polycladon Benth.	sorrel buckwheat	X	X	Χ	Χ		Χ	Χ	
	Eriogonum wrightii Torr. ex Benth.	bastardsage	X	Χ	Χ		Χ	Χ	Χ	
	Eriogonum wrightii var. wrightii Torr. ex Benth.	bastardsage				Χ				
	Polygonum convolvulus L.	black bindweed			Χ			X	Χ	
	Rumex crispus L.	curly dock			Χ	Χ		Χ	X	
	Rumex hymenosepalus Torr.	canaigre dock			Χ	Χ		Χ	Χ	
Portulacaceae	Portulaca halimoides L.	silkcotton purslane			Χ	Χ		Χ	Χ	
	Portulaca oleracea L.	little hogweed			Χ	Χ		Χ	Χ	
	Portulaca suffrutescens Engelm.	shrubby purslane			Χ	Χ		Χ	Χ	
	Portulaca umbraticola Kunth	wingpod purslane	Χ							
	Talinum aurantiacum Engelm.	orange fameflower	Χ		Χ	Χ		Χ	Χ	
	Talinum parviflorum Nutt.	sunbright			Χ	Χ		Χ	Χ	
Primulaceae	Androsace occidentalis Pursh	western rockjasmine				Χ				
Pteridaceae	Argyrochosma limitanea ssp. limitanea (Maxon) Windham	southwestern false cloakfern			Χ	Χ		Χ	Χ	
	Astrolepis sinuata (Lag. ex Sw.) Benham & Windham ssp. sinuata	wavy scaly cloakfern			Χ	Χ		Χ	Χ	
	Bommeria hispida (Mett. ex Kuhn) Underwood	copper fern			Χ	Χ		Χ	Χ	
	Cheilanthes eatonii Baker	Eaton's lipfern		Χ	Χ	Χ		Χ	Χ	
	Cheilanthes lindheimeri Hook.	fairyswords			Χ	Χ		Χ	Χ	
	Cheilanthes wootonii Maxon	beaded lipfern			Χ	Χ		Χ	Χ	
	Cheilanthes wrightii Hook.	Wright's lipfern			Χ	Χ		Χ	Χ	
	Notholaena standleyi Maxon	star cloak fern			Χ	Χ		Χ	Χ	
	Pellaea truncata Goodding	spiny cliffbrake			Χ	Χ		Χ	Χ	
Ranunculaceae	Anemone tuberosa Rydb.	desert anemone			Χ	Χ		Χ	Χ	
	Clematis drummondii Torr. & Gray	Drummond's clematis			Χ	Χ		Χ	Χ	
	Delphinium carolinianum ssp. virescens (Nutt.) Brooks	Carolina larkspur			Χ			Χ	Χ	
	Delphinium wootonii Rydb.	Organ Mountain larkspur				Χ				
Rhamnaceae	Ceanothus fendleri Gray	Fendler's ceanothus					Χ			
	Ceanothus greggii Gray	desert ceanothus			Χ			Χ	Χ	
	Ceanothus greggii var. vestitus (Greene) McMinn	Mojave ceanothus				Χ				
	Condalia spathulata Gray	squawbush				Χ		Χ		
	Condalia warnockii M.C. Johnston	Warnock's snakewood	Χ							
	Condalia warnockii var. kearneyana M.C. Johnston	Kearney's snakewood			Χ				Χ	
	Frangula californica ssp. californica (Eschsch.) Gray	California buckthorn			Χ			Χ	Χ	
	Frangula californica ssp. ursina (Greene) Kartesz & Gandhi	California buckthorn				Χ				

c	3	1
C	X	•

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Rhamnaceae	Ziziphus obtusifolia (Hook. ex Torr. & Gray) Gray	lotebush	Χ		Χ		Χ		Χ	
	Ziziphus obtusifolia var. canescens (Gray) M.C. Johnston	lotebush				Χ				
	Ziziphus obtusifolia var. obtusifolia (Hook. ex Torr. & Gray) Gray	lotebush						Χ		
Rosaceae	Cercocarpus montanus Raf.	alderleaf mountain mahogany	Χ							
	Cercocarpus montanus var. argenteus (Rydb.) F.L. Martin	silver mountain mahogany				Χ				
	Cercocarpus montanus var. glaber (S. Wats.) F.L. Martin	birchleaf mountain mahogany				Χ			Χ	
	Cercocarpus montanus var. paucidentatus (S. Wats.) F.L. Martin	hairy mountain mahogany			Χ			Χ	Χ	
	Fallugia paradoxa (D. Don) Endl. ex Torr.	Apache plume			Χ	Χ	Χ	Χ	Χ	
	Purshia mexicana (D. Don) Henrickson	Mexican cliffrose					Χ			
Rubiaceae	Bouvardia ternifolia (Cav.) Schlecht.	firecrackerbush			Χ	Χ		Χ	Χ	
	Diodia teres Walt.	poorjoe	Χ		Χ			Χ	Χ	
	Diodia teres var. angustata Gray	poorjoe				Χ				
	Galium proliferum Gray	limestone bedstraw	Χ		Χ	Χ		Χ	Χ	
	Galium stellatum Kellogg	starry bedstraw	Χ							
	Galium wrightii Gray	Wright's bedstraw	Χ		Χ	Χ		Χ	Χ	
	Houstonia rubra Cav.	red bluet				Χ		Χ		
Rutaceae	Ptelea trifoliata L.	common hoptree	Χ		Χ				Χ	
	Ptelea trifoliata ssp. angustifolia (Benth.) V. Bailey	common hoptree				Χ		Χ		
	Thamnosma texana (Gray) Torr.	Texas rue	Χ		Χ	Χ		Χ	Χ	
Salicaceae	Populus fremontii S. Wats.	Fremont cottonwood			Χ		Χ	Χ	Χ	
	Populus fremontii S. Wats. ssp. fremontii	Fremont cottonwood				Χ				
	Salix bonplandiana Kunth	Bonpland willow							Χ	
	Salix exigua Nutt.	narrowleaf willow			Χ	Χ	Χ	Χ	Χ	
	Salix gooddingii Ball	Goodding's willow			Χ	Χ	Χ	Χ	Χ	
Santalaceae	Comandra umbellata ssp. pallida (A. DC.) Piehl	pale bastard toadflax			Χ			Χ	Χ	
Sapindaceae	Sapindus saponaria L.	wingleaf soapberry	Χ					Χ		
	Sapindus saponaria var. drummondii (Hook. & Arn.) L. Benson	western soapberry			Χ	Χ	Χ		Χ	
Sapotaceae	Sideroxylon lanuginosum Michx.	gum bully	Χ	Χ						
	Sideroxylon lanuginosum ssp. rigidum (Gray) T.D. Pennington	gum bully			Χ	Χ		Χ	Χ	
Saxifragaceae	Heuchera sanguinea Engelm.	coralbells			Χ	Χ		Χ	Χ	
Scrophulariaceae	Castilleja austromontana Standl. & Blumer	Rincon Mountain Indian paintbrush			Χ	Χ		Χ	Χ	••••••
	Castilleja integra Gray	wholeleaf Indian paintbrush			Χ			Χ	Χ	
	Castilleja integra var. gloriosa (Britt.) Cockerell	wholeleaf Indian paintbrush				Χ				
	Castilleja lanata Gray	Sierra woolly Indian paintbrush	Χ		Χ	Χ		Χ	Χ	
	Castilleja sessiliflora Pursh	downy paintedcup			Χ	Χ		Χ	Χ	
•	Castilleja tenuiflora Benth.	Santa Catalina Indian paintbrush				Χ				

		9							
	Penstemon barbatus (Cav.) Roth	beardlip penstemon		X			Χ	Χ	
	Penstemon barbatus ssp. torreyi (Benth.) Keck	Torrey's penstemon			Χ				
	Penstemon linarioides Gray	toadflax penstemon	X	X			Χ	Χ	
	Penstemon linarioides Gray ssp. linarioides	toadflax beardtongue			Χ				
	Penstemon ramosus Crosswhite	lanceleaf beardtongue		Χ	Χ		Χ	Χ	
	Verbascum virgatum Stokes	wand mullein	Χ						
	Veronica peregrina L.	neckweed		Χ			Χ	Χ	
	Veronica peregrina ssp. xalapensis (Kunth) Pennell	hairy purslane speedwell			Χ				
Solanaceae	Chamaesaracha coronopus (Dunal) Gray	greenleaf five eyes		X	Χ		Χ	Χ	
	Chamaesaracha sordida (Dunal) Gray	hairy five eyes	X	X	Χ		Χ	Χ	
	Datura inoxia P. Mill.	pricklyburr		X				X	
	Datura wrightii Regel	sacred thorn-apple	X		Χ		Χ		
	Lycium fremontii Gray	Fremont's desert-thorn		X	Χ		Χ	Χ	
	Lycium pallidum Miers	pale desert-thorn		Χ	Χ	Χ	Χ	Χ	
	Margaranthus solanaceus Schlecht.	netted globecherry	X						
	Nicotiana obtusifolia Mertens & Galeotti	desert tobacco	X						
	Nicotiana obtusifolia var. obtusifolia Mertens & Galeotti	desert tobacco		X	Χ		Χ	Χ	
	Physalis acutifolia (Miers) Sandw.	sharpleaf groundcherry		X	Χ		Χ	Χ	
	Physalis hederifolia var. fendleri (Gray) Cronq.	Fendler's groundcherry		Χ	Χ		Χ	Χ	
	Physalis longifolia Nutt. var. longifolia	longleaf groundcherry		Χ	Χ			Χ	
	Solanum americanum P. Mill.	American black nightshade		X	Χ		Χ	Χ	
	Solanum elaeagnifolium Cav.	silverleaf nightshade		X	Χ		Χ	Χ	
Sterculiaceae	Ayenia filiformis S. Wats.	TransPecos ayenia	X						
Tamaricaceae	Tamarix sp. L.	tamarisk							Χ

netleaf hackberry

spiny hackberry

Wright's beebrush

Dakota mock vervain

Dakota mock vervain

Davis Mountain mock vervain

Common name

roving sailor

Dalmatian toadflax

broomleaf toadflax

seep monkeyflower

giant lousewort

little redstem monkeyflower

UA Herb-

Χ

Χ

Χ

Χ

Χ

UA arium^b WEA BEA RUL

X

Χ

Χ

Χ

Χ

X

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

H&R NPS

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

Χ

H&G

Ulmaceae

Verbenaceae

Family

Scientific name

Mimulus guttatus DC.

Mimulus rubellus Gray

Pedicularis procera Gray

Linaria genistifolia (L.) P. Mill.

Maurandella antirrhiniflora (Humb. & Bonpl. ex Willd.) Rothm.

Celtis laevigata var. reticulata (Torr.) L. Benson

Glandularia bipinnatifida var. bipinnatifida (Nutt.) Nutt.

Celtis pallida Torr.

Aloysia wrightii Heller ex Abrams

Glandularia wrightii (Gray) Umber

Glandularia bipinnatifida (Nutt.) Nutt.

Scrophulariaceae Linaria dalmatica (L.) P. Mill.

				UA Herb-						
Family	Scientific name	Common name	UA	arium⁵	WEA	BEA	RUL	H&R	NPS	H&G
Verbenaceae	Tetraclea coulteri Gray	Coulter's wrinklefruit			Χ	Χ		Χ	Χ	
Violaceae	Verbena gracilis Desf.	Fort Huachuca vervain			Χ	Χ		Χ	Χ	
	Hybanthus verticillatus (Ortega) Baill.	babyslippers			Χ			Χ	Χ	
	Hybanthus verticillatus (Ortega) Baill. var. verticillatus	babyslippers				Χ				
Viscaceae	Phoradendron bolleanum (Seem.) Eichl.	Bollean mistletoe						Χ	Χ	
	Phoradendron californicum Nutt.	mesquite mistletoe			Χ	Χ		Χ	Χ	
	Phoradendron capitellatum Torr. ex Trel.	downy mistletoe			Χ	Χ			Χ	
	Phoradendron coryae Trel.	Cory's mistletoe			Χ	Χ		Χ	Χ	
	Phoradendron pauciflorum Torr.	fir mistletoe				Χ				
Vitaceae	Vitis arizonica Engelm.	canyon grape	Χ		Χ	Χ	Χ	Χ	Χ	
Zygophyllaceae	Kallstroemia grandiflora Torr. ex Gray	Arizona poppy			Χ	Χ		Χ	Χ	
•••••	Kallstroemia parviflora J.B.S. Norton	warty caltrop			Χ	Χ		Χ	Χ	
•••••	Larrea tridentata (Sessé & Moc. ex DC.) Coville	creosote bush			Χ			Χ	Χ	
	Larrea tridentata var. tridentata (Sessé & Moc. ex DC.) Coville	creosote bush	Χ			Χ				
	Tribulus terrestris L.	puncturevine			Χ	Χ		Χ	Χ	Χ

^a This list includes many species (*n* = 54) for which we included separate lines for specimens that were identified to species only and specimens that were identified to its nominate species (e.g., *Guilleminea densa* and *Guilleminea densa* var. *densa*). In these cases we do not determine these to be separate species in the tally of the number of species for the park or number of new species.

^b Specimens at the University of Arizona Herbarium. Specimens date from 1902–1994.

Appendix B. Amphibians and reptiles observed or documented at Fort Bowie NHS by University of Arizona (UA) personnel, Swann et al. (2001; Swann), Lowe and Johnson (1976; L&J), and from voucher specimens reported in Appendix E (AE). "Possible" species have not been observed or documented in the park, but may occur based on habitat and range (reported in Swann et al. [2001]).

	- "	0 : 45			served or			5 "1
Order Caudata	Family Ambystomatidae	Scientific name Ambystoma tigrinum mavortium	Common name barred tiger salamander	UA	Swann	L&J	AE	Possible X
_	Pelobatidae		Couch's spadefoot		X	Χ		^
Anura	Pelobalidae	Scaphiopus couchii			^	Λ Χ		Χ
		Spea bombifrons	plains spadefoot					^
	D. f id	Spea multiplicata	Mexican spadefoot		X	Χ		
	Bufonidae	Bufo punctatus	red-spotted toad		X	X		
		Bufo cognatus	Great Plains toad		Χ	Χ		
		Bufo alvarius	Sonoran desert toad					X
		Bufo woodhousii	Woodhouse's toad					X
		Bufo debilis	Green toad			X		
	Hylidae	Hyla arenicolor	canyon treefrog		Χ	Χ		
	Ranidae	Rana catesbeiana	American bullfrog					X
		Rana chiricahuensis ^a	Chiricahua leopard frog		Χ	Χ		
		Rana blairi	plains leopard frog					X
Testudines	Kinosternidae	Kinosternon flavescens	yellow mud turtle					X
		Kinosternon sonoriense	Sonoran mud turtle					X
	Emydidae	Terrapene ornata	western box turtle			Χ		Χ
	Testudinidae	Gopherus agassizii sonoran⁵	Sonoran desert tortoise					Χ
Squamata	Eublepharidae	Coleonyx variegatus	western banded gecko		Χ	Χ		
	Crotaphytidae	Crotaphytus collaris	eastern collared lizard		Χ	Χ	Χ	
		Gambelia wislizenii	long-nosed leopard lizard					Χ
	Phrynosomatidae	Holbrookia maculata	lesser earless lizard	Χ				
		Cophosaurus texanus	greater earless lizard	Χ	Χ	Χ		
		Callisaurus draconoides	zebra-tailed lizard					Χ
		Sceloporus clarkii	Clark's spiny lizard		Χ		Χ	
		Sceloporus virgatus	striped plateau lizard	Χ				
		Sceloporus slevini	Slevin's bunchgrass lizard					Χ
		Sceloporus jarrovii	mountain spiny lizard					Χ
		Sceloporus magister	desert spiny lizard					Χ
		Sceloporus undulatus	eastern fence lizard					Χ
		Urosaurus ornatus	ornate tree lizard	Χ	Χ	Χ	Χ	
		Uta stansburiana	common side-blotched lizard					Χ
		Phrynosoma cornutum	Texas horned lizard		Χ	Χ		
		Phrynosoma douglasii	pygmy short-horned lizard		Χ	Χ		
		Phrynosoma modestum	round-tailed horned lizard		Χ	Χ		
		Phrynosoma solare	regal horned lizard					Χ
	Scincidae	Eumeces obsoletus	Great Plains skink		Χ	Χ		
	Teiidae	Cnemidophorus uniparens	desert grassland whiptail		Χ	Χ		
		Cnemidophorus exsanguis	Chihuahuan spotted whiptail		Χ		Χ	
		Cnemidophorus sonorae	Sonoran spotted whiptail		Χ	Χ	···········	
		Cnemidophorus tigris	western whiptail (tiger whiptail)		Χ	Χ		
		Cnemidophorus burti	canyon spotted whiptail					Χ

				Obse	erved or d	ocum	ented	
Order	Family	Scientific name	Common name	UA	Swann	L&J	AE	Possible
Squamata	Teiidae	Cnemidophorus inornatus	little striped whiptail					X
	Anguidae	Elgaria kingii	Madrean alligator lizard		Χ	Χ		
	Helodermatidae	Heloderma suspectum	Gila monster	Χ	Χ	Χ		
		Leptotyphlops humilis	western blind snake					Χ
		Leptotyphlops dulcis	Texas blind snake					X
	Colubridae	Diadophis punctatus	ring-necked snake			Χ		
		Heterodon nasicus	western hog-nosed snake					X
	Colubridae	Masticophis flagellum	coachwhip		Χ	Χ		
		Masticophis bilineatus	Sonoran whipsnake	Χ	Χ	Χ		
		Salvadora hexalepis deserticola	Big Bend patch-nosed snake		Χ	Χ		
		Salvadora grahamiae	mountain patch-nosed snake		Χ	Χ		
		Senticolis triaspis	green rat snake	Χ	Χ	Χ		
		Pituophis catenifer	gopher snake	Χ	Χ	Χ		
		Arizona elegans	western glossy snake					Χ
		Lampropeltis getula	common kingsnake					X
		Lampropeltis pyromelana	Sonoran mountain kingsnake					Χ
		Lampropeltis triangulum	milk snake					Χ
		Rhinocheilus lecontei	long-nosed snake					Χ
		Thamnophis cyrtopsis	black-necked garter snake		Χ	Χ		
		Thamnophis marcianus	checkered garter snake					Χ
		Sonora semiannulata	western ground snake					Χ
		Gyalopion canum	Chihuahuan hook-nosed snake					Χ
		Tantilla yaquia	Yaqui black-headed snake		Χ	Χ		
		Tantilla hobartsmithi	southwestern black-headed snake					Χ
		Tantilla nigriceps	plains black-headed snake					Χ
		Trimorphodon biscutatus	western lyre snake			Χ		
		Hypsiglena torquata	night snake		Χ			
	Elapidae	Micruroides euryxanthus	Sonoran coral snake		Χ			
	Viperidae	Sistrurus catenatus	massasauga					Χ
		Crotalus lepidus	rock rattlesnake					X
		Crotalus viridis cerberus	Arizona black rattlesnake					X
		Crotalus scutulatus	Mojave rattlesnake					X
		Crotalus pricei	twin-spotted rattlesnake					Χ
		Crotalus atrox	western diamond-backed rattlesnake		Χ	Χ	Χ	
		Crotalus molossus	black-tailed rattlesnake		Χ	Χ		

^a Now extirpated from the park. Listed as "Threatened" under the Endangered Species Act, "Sensitive" by the U.S.D.A. Forest Service, and "Wildlife of Special Concern" by the Arizona Game and Fish Department (HDMS 2004).

^b "Species of Concern" by U.S. Fish and Wildlife Service, and "Wildlife of Special Concern" by the Arizona Game and Fish Department (HDMS 2004).

						a			bserved			0	e.		··
					UA surve		d Incid-	d	ocumente	ea		Conse	rvation c		
Order	Family	Scientific name	Common name	VCP	Line transect	Noc- turnal	ental	R&J	FI	AE	ESAª	USFS⁵	ΑZ°	AZ PIF⁴	U.S. FWS ^e
Anseriformes	Anatidae	Branta canadensis	Canada goose					Χ							
Galliformes	Odontophoridae	Callipepla squamata	scaled quail					Χ	Χ						
		Callipepla gambelii	Gambel's quail	146	120		1	Χ	Χ						
		Cyrtonyx montezumae	Montezuma quail		1			Χ	Χ						
Ciconiiformes	Ardeidae	<u>Ardea herodias</u>	great blue heron						Χ						
		Butorides virescens	green heron					Χ							
	Cathartidae	Coragyps atratus	black vulture					Χ							
		Cathartes aura	turkey vulture	38			1	Χ	Χ						
Falconiformes	Accipitridae	<u>Pandion haliaetus</u>	osprey						Χ						
		Haliaeetus leucocephalus	bald eagle						Χ						
		<u>Circus cyaneus</u>	northern harrier				1	Χ	Χ						
		Accipiter striatus	sharp-shinned hawk					Χ	Χ			Χ			
		Accipiter cooperii	Cooper's hawk	9	1		5	Χ	Χ						
		Buteogallus anthracinus	common black-hawk				1	Χ	Χ			Χ	Χ	Χ	
		<u>Buteo swainsoni</u>	Swainson's hawk					Χ	Χ	Χ					
		Buteo albonotatus	zone-tailed hawk	1			1								
		<u>Buteo jamaicensis</u>	red-tailed hawk	8	1		2	Χ	Χ	Χ					
		Buteo regalis	ferruginous hawk						Χ						
		Buteo lagopus	rough-legged hawk						Χ						
		Aquila chrysaetos	golden eagle	1				Χ	Χ						
	Falconidae	<u>Falco sparverius</u>	American kestrel					Χ	Χ						
		Falco columbarius	merlin					Χ	Χ						
		<u>Falco peregrinus</u>	peregrine falcon	1					Χ		SC		Χ		Χ
		Falco mexicanus	prairie falcon					Χ	Χ						
Gruiformes	Gruidae	<u>Grus canadensis</u>	sandhill crane		52				Χ						
Charadriiformes	Charadriidae	Charadrius vociferus	killdeer					Χ	Χ						
	Scolopacidae	Tringa flavipes	lesser yellowlegs					Χ							

					UA surve	/ metho	d		served o			Conse	rvation o	designa	tion
Order	Family	Scientific name	Common name	VCP	Line transect	Noc- turnal	Incid- ental	R&J	FI	AE	ESAª	USFS ^b	ΑZ°	AZ PIF	U.S. FWS ^e
Charadriiformes	Scolopacidae	Actitis macularia	spotted sandpiper					Χ							
Columbiformes	Columbidae	Patagioenas fasciata	band-tailed pigeon					Χ							
		Zenaida asiatica	white-winged dove	180			8	Χ	Χ						
		Zenaida macroura	mourning dove	168			16	Χ	Χ						
		Columbina passerina	common ground-dove				2								
Cuculiformes	Cuculidae	Geococcyx californianus	greater roadrunner				3	Χ	Χ	Χ					
Strigiformes	Tytonidae	Tyto alba	barn owl						Χ						
Strigiformes	Strigidae	Megascops kennicottii	western screech-owl			1		Χ	Χ						
		Bubo virginianus	great horned owl			1		Χ	Χ						
		Micrathene whitneyi	elf owl			3		Χ	Χ						Х
		Strix occidentalis lucida	Mexican spotted owl					Χ			LT	Χ	Χ		
		Asio otus	long-eared owl				1	Χ	Χ						
Caprimulgiforme	s Caprimulgidae	Chordeiles acutipennis	lesser nighthawk				2	Χ	Χ						
		Chordeiles minor	common nighthawk					Χ							
		Phalaenoptilus nuttallii	common poorwill			12	2	Χ	Χ						
		Caprimulgus vociferus	whip-poor-will					Χ							
Apodiformes	Apodidae	Chaetura vauxi	Vaux's swift					Χ	Χ						
		Aeronautes saxatalis	white-throated swift	8				Χ							
	Trochilidae	Cynanthus latirostris	broad-billed hummingbird						Χ						
		Eugenes fulgens	magnificent hummingbird					Χ							
		Calothorax lucifer	Lucifer hummingbird					Χ	Χ						
		Archilochus alexandri	black-chinned hummingbird	14			2	Χ	Χ						
		Calypte anna	Anna's hummingbird					Χ	Χ						
		Calypte costae	Costa's hummingbird					Χ	Χ						X
		Stellula calliope	calliope hummingbird						Χ						
		Selasphorus platycercus	broad-tailed hummingbird	14			4	Χ	Χ						
		<u>Selasphorus rufus</u>	rufous hummingbird					Χ	Χ						
Coraciiformes	Alcedinidae	Ceryle alcyon	belted kingfisher					Χ	Χ					Χ	
Piciformes	Picidae	Melanerpes lewis	Lewis's woodpecker					Χ							
		Melanerpes formicivorus	acorn woodpecker					Χ	Χ						

					UA surve	y metho	d		oserved o			Conse	rvation o	designa	tion
Order	Family	Scientific name	Common name	VCP	Line transect	Noc- turnal	Incid- ental	R&J	FI	AE	ESAª	USFS⁵	ΑZ°	AZ PIF	U.S. FWS [®]
Piciformes	Picidae	Melanerpes uropygialis	Gila woodpecker					Χ	Χ						Χ
		Sphyrapicus thyroideus	Williamson's sapsucker					Χ	Χ						
		Sphyrapicus nuchalis	red-naped sapsucker		1				Χ						
		Sphyrpicus varius	yellow-bellied sapsucker					Χ							
		Picoides scalaris	ladder-backed woodpecker	48	11		1	Χ	Χ						
		Colaptes auratus	northern flicker		20			Χ	Χ						
Passeriformes	Vireonidae	<u>Vireo bellii</u>	Bell's vireo	14			5	Χ	Χ			Χ			Χ
	Tyrannidae	Contopus cooperi	olive-sided flycatcher					Χ			SC				
		Contopus pertinax	greater pewee	1											
		Contopus sordidulus	western wood-pewee	7				Χ	Χ						
		Empidonax traillii	willow flycatcher						Χ						
Passeriformes	Tyrannidae	Empidonax hammondii	Hammond's flycatcher	1			3		Χ						
		Empidonax wrightii	gray flycatcher	2			2	Χ	Χ						
		Empidonax oberholseri	dusky flycatcher						Χ						
		Empidonax occidentalis or difficili	s_western flycatcher					Χ							
		Sayornis nigricans	black phoebe					Χ	Χ						
		Sayornis saya	Say's phoebe	1			4	Χ	Χ	Χ					
		Myiarchus tuberculifer	dusky-capped flycatcher	1											
		Myiarchus cinerascens	ash-throated flycatcher	101				Χ	Χ	Χ					
		Myiarchus tyrannulus	brown-crested flycatcher	3			5	Χ	Χ						
		Tyrannus vociferans	Cassin's kingbird	75			5	Χ	Χ						
		Tyrannus verticalis	western kingbird	14			1	Χ	Χ						
	Laniidae	<u>Lanius Iudovicianus</u>	loggerhead shrike		1			Χ	Χ	Χ	SC	Χ			
	Vireonidae	<u>Vireo vicinior</u>	gray vireo					Χ	Χ						
		Vireo plumbeus	plumbeous vireo	2			2	Χ							
		Vireo huttoni	Hutton's vireo				1								
		<u>Vireo gilvus</u>	warbling vireo	3			4	Χ	Χ						
	Corvidae	Cyanocitta stelleri	Steller's jay		2			Χ							
		Aphelocoma californica	western scrub-jay	48	33		1	Χ	Χ	Χ					
		Aphelocoma ultramarina	Mexican jay	1				Χ	Χ						
		Gymnorhinus cyanocephalus	pinyon jay					Χ	Χ						

					UA surve	/ metho	d		oserved o			Conse	vation	designa	tion
Order	Family	Scientific name	Common name	VCP	Line transect	Noc- turnal	Incid- ental	R&J	FI	AE	ESAª	USFS ^b	ΑZ°	AZ PIF	U.S. FWS°
Passeriformes	Corvidae	Nucifraga columbiana	Clark's nutcracker					Χ							
		Corvus cryptoleucus	Chihuahuan raven					Χ	Χ	Χ					
		Corvus corax	common raven					Χ	Χ						
		Corvus sp.	unknown raven	11	2		1								
	Alaudidae	Eremophila alpestris	horned lark						Χ	Χ					
	Hirundinidae	Tachycineta thalassina	violet-green swallow	5				Χ							
		Stelgidopteryx serripennis	northern rough-winged swallow					Χ	Χ						
		Petrochelidon pyrrhonota	cliff swallow					Χ	Χ						
		Hirundo rustica	barn swallow				2	Χ							
	Paridae	Baeolophus wollweberi	bridled titmouse	1	11			Χ	Χ						
		Baeolphus ridgwayi	juniper titmouse	3	2		1	Χ	Χ						
	Remizidae	Auriparus flaviceps	verdin	40	10		1	Χ	Χ						
	Aegithalidae	Psaltriparus minimus	bushtit	11	28			Χ	Χ	Χ					
	Sittidae	Sitta carolinensis	white-breasted nuthatch					Χ	Χ						
	Certhiidae	Certhia americana	brown creeper		1			Χ							
	Troglodytidae	e Campylorhynchus brunneicapillus	cactus wren	120	18		4	Χ	Χ	Χ					
		Salpinctes obsoletus	rock wren	9	3		1	Χ	Χ						
		Catherpes mexicanus	canyon wren	7			2	Χ	Χ						
		Thryomanes bewickii	Bewick's wren	118	11		2	Χ	Χ						
		Troglodytes aedon	house wren	2				Χ							
	Regulidae	Regulus calendula	ruby-crowned kinglet	15	32		1	Χ	Χ						
	Sylviidae	Polioptila caerulea	blue-gray gnatcatcher	4			1	Χ	Χ						
		Polioptila melanura	black-tailed gnatcatcher	1			1		Χ	Χ					
	Turdidae	<u>Sialia mexicana</u>	western bluebird		27		1	Χ	Χ						
		Sialia currucoides	mountain bluebird					Χ	Χ						
		Myadestes townsendi	Townsend's solitaire		4		1	Χ	Χ						
		Catharus ustulatus	Swainson's thrush					Χ							
		Catharus guttatus	hermit thrush		1		4	Χ	Χ						
		Turdus migratorius	American robin	1	39		1	Χ							
	Mimidae	Mimus polyglottos	northern mockingbird	116	10		2	Χ	Χ						
		Oreoscoptes montanus	sage thrasher					Χ	Χ						

					UA surve	y metho	d		served o			Conse	rvation	designa	tion
Order	Family	Scientific name	Common name	VCP	Line transect	Noc- turnal	Incid- ental	R&J	FI	AE	ESAª	USFS ^b	ΑZ°	AZ PIF	U.S. FWS ^e
Passeriformes	Mimidae	Toxostoma bendirei	Bendire's thrasher				1	Χ	Χ	Χ					
		Toxostoma curvirostre	curve-billed thrasher	9	2		1	Χ	Χ	Χ					
		Toxostoma crissale	crissal thrasher	33	16	1	4	Χ	Χ	Χ					Χ
	Sturnidae	Sturnus vulgaris	European starling					Χ							
	Motacillidae	Anthus rubescens	American pipit						Χ						
	Bombycillidae	Bombycilla cedrorum	cedar waxwing		12			Χ	Χ						
	Ptilogonatidae	Phainopepla nitens	phainopepla	17	16		2	Χ	Χ						
	Parulidae	Vermivora celata	orange-crowned warbler				3	Χ	Χ						
		Vermivora ruficapilla	Nashville warbler					Χ	Χ						
		<u>Vermivora virginiae</u>	Virginia's warbler					Χ							
		<u>Vermivora luciae</u>	Lucy's warbler	17			1	Χ	Χ					Χ	
		<u>Dendroica petechia</u>	yellow warbler						Χ						
		<u>Dendroica coronata</u>	yellow-rumped warbler	4	3			Χ	Χ						
		<u>Dendroica nigrescens</u>	black-throated gray warbler	4	1			Χ	Χ						
		<u>Dendroica townsendi</u>	Townsend's warbler	1				Χ	Χ						
		<u>Dendroica occidentalis</u>	hermit warbler						Χ						
		Seiurus noveboracensis	northern waterthrush					Χ							
		Oporornis tolmiei	MacGillivray's warbler					Χ	Χ						
		Geothlypis trichas	common yellowthroat					Χ							
		<u>Wilsonia pusilla</u>	Wilson's warbler	8			5	Χ	Χ						***************************************
	Parulidae	<u>Icteria virens</u>	yellow-breasted chat					Χ							***************************************
	Thraupidae	<u>Piranga rubra</u>	summer tanager	34			2	Χ	Χ						
		Piranga ludoviciana	western tanager	7			3	Χ	Χ						
	Emberizidae	<u>Pipilo chlorurus</u>	green-tailed towhee	9	34		1	Χ	Χ						
		<u>Pipilo maculatus</u>	spotted towhee	7	52			Χ							
		Pipilo fuscus	canyon towhee	86	40		7	Χ	Χ	Χ					
		Aimophila cassinii	Cassin's sparrow						Χ						
		Aimophila texana or botterrii	Botteri's sparrow	1				Χ							
		Aimophila ruficeps	rufous-crowned sparrow	49	1			Χ	Χ						
		Spizella passerina	chipping sparrow	3	83			Χ	Χ						
		Spizella breweri	Brewer's sparrow	2	51		3	Χ	Χ						

					UA surve	y metho	d		oserved o			Conse	rvation o	designa	tion
Order	Family	Scientific name	Common name	VCP	Line transect	Noc- turnal	Incid- ental	R&J	FI	AE	ESAª	USFS⁵	ΑZ°	AZ PIF	U.S. FWS°
Passeriformes	Emberizidae	Spizella atrogularis	black-chinned sparrow	2	5		1	Χ	Χ						
		Passerculus sandwichensis	savannah sparrow					Χ	Χ						
		Pooecetes gramineus	vesper sparrow		33		2	Χ	Χ						
		<u>Melospiza lincolnii</u>	Lincoln's sparrow		6		1	Χ	Χ						
		Chondestes grammacus	lark sparrow					Χ	Χ						
		Amphispiza bilineata	black-throated sparrow	115	12	1	6	Χ	Χ	Χ					
		Calamospiza melanocorys	lark bunting					Χ	Χ						
		Ammodramus savannarum	grasshopper sparrow				3		Χ						
		Ammodramus bairdii	Baird's sparrow				1				Х		Χ		
		Passerella iliaca	fox sparrow					Χ	Χ						
		Melospiza melodia	song sparrow					Χ	Χ						
		Zonotrichia albicollis	white-throated sparrow					Χ							
		Zonotrichia leucophrys	white-crowned sparrow		134		2	Χ	Χ	Χ					
		Junco hyemalis	dark-eyed junco	Χ	Χ			Χ	Χ						
		Junco hyemalis mearnsi	pink-sided junko ^f	1	2										
		Junco hyemalis dorsalis	gray-headed junco ^f		2			Χ							
		Junco hyemalis oreganus	Oregon junco ^f		4										
		Junco phaeonotus	yellow-eyed junco					Χ							
	Cardinalidae	Cardinalis cardinalis	northern cardinal	124	13	1	4	Χ	Χ						
		Cardinalis sinuatus	pyrrhuloxia	1	1			Χ	Χ						
		Pheucticus Iudovicianus	rose-breasted grosbeak					Χ	Χ						
		Pheucticus melanocephalus	black-headed grosbeak	12			1	Χ		Χ					
		<u>Passerina caerulea</u>	blue grosbeak	6			1	Χ	Χ						
		<u>Passerina amoena</u>	lazuli bunting	1				Χ	Χ						
		Passerina ciris	painted bunting					Χ	Χ						
	Icteridae	Sturnella magna lilianae	eastern meadowlark				1	Χ	Χ						
		Sturnella neglecta	western meadowlark					Χ	Χ						
		Euphagus cyanocephalus	Brewer's blackbird						Χ						
		Quiscalus mexicanus	great-tailed grackle						Χ						
		Molothrus aeneus	bronzed cowbird	1			1	Χ	Χ						
		Molothrus ater	brown-headed cowbird	56			2	Χ	Χ						

					UA surve	y metho	od		oserved o ocumente			Conse	rvation o	designa	tion
Order	Family	Scientific name	Common name	VCP	Line transect	Noc- turnal	Incid- ental	R&J	FI	AE	ESAª	USFS ^b	ΑZ°	AZ PIF⁴	U.S. FWS°
Passeriformes	Icteridae	<u>Icterus cucullatus</u>	hooded oriole	44			5	Χ	Χ	Χ					
		<u>Icterus bullockii</u>	Bullock's oriole	26			2	Χ							
		<u>Icterus parisorum</u>	Scott's oriole	26				Χ	Χ						
	Fringillidae	Carpodacus mexicanus	house finch	54	4		2	Χ	Χ						
		Carduelis pinus	pine siskin		1			Χ	Χ						
		<u>Carduelis psaltria</u>	lesser goldfinch	3	6		1	Χ	Χ						
		Coccothraustes vespertinus	evening grosbeak					Χ							
	Passeridae	Passer domesticus	house sparrow					Χ							

a "SC" = "Species of Concern"; "C" = Candidate for listing under the Endangered Species Act, U.S. Fish and Wildlife Service. (in HDMS 2004)

b "Sensitive Species"; U.S.D.A. Forest Service (HDMS 2004).

c "Wildlife of Special Concern"; Arizona Game and Fish Department (HDMS 2004).

d "Priority species"; Arizona Partners in Flight (Latta et al. 1999).

e "Species of Conservation Concern"; U.S. Fish and Wildlife Service (HDMS 2004).

f We include observations of these subspecies in the appendix because field crew members occasionally made the distinction.

Appendix D. Mammals recorded at Fort Bowie NHS by University of Arizona inventory personnel (UA) and/or documented/observed by other researchers: Roth (1976; ROH), Petryszyn (1999; PZN), Herman-Reese (unpublished data; HR), Swann et al. (2001; SEA), Krebbs (2005; KRB), or specimens located in the UA mammal collection (UAMC). Species in hold-faced type is non-native

Order Family	Scientific name	Common name	UA	UAMC	ROH	PZN	HR	SEA	KRB
Insectivora									
Soricidae	Notiosorex crawfordi	Crawford's desert shrew			Χ				
Chiroptera									
Phyllostomidae	Choeronycteris mexicana	Mexican long-tongued bat		Χ	Χ			••••••	***************************************
	Leptonycteris curasoae yerbabuenae			Χ	Χ				
Vespertilionidae	Myotis occultus	Arizona myotis						••••••	Χ
	Myotis auriculus	southwestern myotis							Χ
	Myotis velifer	cave myotis		Χ	Χ				Χ
	Myotis thysanodes	fringed myotis		Χ	Χ				Χ
	Myotis californicus	California myotis							Χ
	Myotis volans	long-legged myotis							Χ
	Myotis ciliolabrum	western small-footed myotis							Χ
	Pipistrellus hesperus	western pipistrelle		Χ	Χ				
	Eptesicus fuscus	big brown bat		Χ					
	Antrozous pallidus	pallid bat		Χ	Χ				
Carnivora									
Ursidae	Ursus americanus	American black bear	Χ				Χ		
Procyonidae	Procyon lotor	northern raccoon			Χ				
	Nasua narica	white-nosed coati	Χ		Χ		Χ		
	Bassariscus astutus	ringtail			Χ		Χ		
Mustelidae	Taxidea taxus	American badger			Χ			Χ	
	Mephitis macroura	hooded skunk	Χ						
	Spilogale gracilis	western spotted skunk			Χ				
	Mephitis mephitis	striped skunk	Χ		Χ		Χ		
	Conepatus mesoleucus	white-backed hog-nosed skunk	Χ		Χ				
Canidae	Canis familiaris	feral dog	Χ						
	Canis latrans	coyote	Χ		Χ		Χ	Χ	
	Urocyon cinereoargenteus	common gray fox	Χ		Χ		Χ	Χ	
Felidae	Lynx rufus	bobcat	Χ		Χ		Χ	Χ	
	Puma concolor	mountain lion	Χ				Χ	Χ	
Rodentia								••••••	
Sciuridae	Spermophilus variegatus	rock squirrel	Χ		Χ		Χ	Χ	
	Spermophilus tereticaudus	round-tailed ground squirrel		Χ					
	Ammospermophilus harrisii	Harris' antelope squirrel		Χ	Χ			Χ	
	Neotamias dorsalis	cliff chipmunk		Χ	Χ			Χ	
Geomyidae	Thomomys bottae	Botta's pocket gopher		Χ	Χ			.	
Heteromyidae	Perognathus flavus flavus	silky pocket mouse		Χ	Χ		Χ		
	Perognathus amplus	Arizona pocket mouse					Χ		
	Chaetodipus penicillatus	Sonoran Desert pocket mouse		Χ	Χ	Χ	Χ		
	Chaetodipus intermedius	rock pocket mouse		Χ	Χ		X		
	Chaetodipus baileyi	Bailey's pocket mouse		Χ	Χ		X		
	Chaetodipus hispidus	hispid pocket mouse		Χ	Χ		X		***************************************
	Dipodomys spectabilis	banner-tailed kangaroo rat		Χ	Χ				
	Dipodomys ordii	Ord's kangaroo rat		Χ	Χ		Χ		
	Dipodomys merriami	Merriam's kangaroo rat		-	Χ	Χ	X		

Order									
Family	Scientific name	Common name	UA	UAMC	ROH	PZN	HR	SEA	KRB
Rodentia									
Heteromyidae	Reithrodontomys megalotis megalotis	western harvest mouse					Χ		
	Reithrodontomys fulvescens	fulvous harvest mouse		Χ	Χ				
	Peromyscus eremicus	cactus mouse		Χ	Χ		Χ		
Muridae	Peromyscus maniculatus	deer mouse		Χ	Χ		Χ		
	Peromyscus leucopus	white-footed mouse		Χ					
	Peromyscus boylii	brush mouse		Χ	Χ		Χ		
	Baiomys taylori	northern pygmy mouse					Χ		
	Onychomys leucogaster	northern grasshopper mouse		Χ	Χ		Χ		
	Onychomys torridus	southern grasshopper mouse		Χ	Χ	Χ	Χ		
	Neotoma albigula	western white-throated woodrat		Χ	Χ	Χ	Χ	Χ	
	Sigmodon hispidus	hispid cotton rat		Χ	Χ		Χ		
Erethizontidae	Erethizon dorsatum	North American porcupine						Χ	
Lagomorpha									
Leporidae	Lepus alleni	antelope jackrabbit		Χ	Χ				
	Lepus californicus	black-tailed jackrabbit	Χ	Χ	Χ			Χ	
	Sylvilagus audubonii	desert cottontail			Χ			Χ	
Artiodactyla									
Tayassuidae	Pecari tajacu	collared peccary	Χ		Χ		Χ	Χ	
Cervidae	Odocoileus hemionus	mule deer	Χ		Χ		Χ	Χ	
	Odocoileus virginianus	white-tailed deer			Χ			Χ	

Appendix E. List of voucher specimens collected from Fort Bowie NHS. See Table 1.1 for list of collections that were queried for specimens.

Group	Common name	Field collection numbers	Collection ^a	Date	Primary Collector
Reptile	eastern collared lizard	12578, 12579	CAS	08/02/1950	H. K. Gloyd
		022207	USNM	05/22/1894	A. K. Fisher
		12578-16024, 12579-16025	CAS	08/02/1950	H. K. Gloyd
	Clark's spiny lizard	12586, 12587, 125867-160323	CAS	08/02/1950	H. K. Gloyd
	ornate tree lizard	12636-16082	CAS	08/02/1950	H. K. Gloyd
	Chihuahuan spotted whiptail	40890	UCO	07/22/1970	R. L. Holland
	western diamond-backed rattlesnake	022194	USNM	UNKN	A. K. Fisher
Bird	Swainson's hawk	3366, 3367, 3368	UCB	05/25/1894	F. H. Fowler
		68829	UCB	05/18/1936	A. H. Miller
	red-tailed hawk	3359, 3360, 3361, 3362	UCB	04/17/1893	F. H. Fowler
	greater roadrunner	3388, 3389, 3390, 3391	UCB	04/10/1893	F. H. Fowler
	Say's phoebe	3425, 3426	UCB	04/11/1893	F. H. Fowler
	ash-throated flycatcher	78819	UCB	05/22/1894	F. H. Fowler
	loggerhead shrike	3500, 3501, 3502, 3503, 3504, 3505, 3506	UCB	04/01/1893	F. H. Fowler
	western scrub-jay	3441, 3442, 3443, 3444	UCB	04/05/1893	F. H. Fowler
	Chihuahuan raven	3457, 3458, 3459	UCB	05/11/1893	F. H. Fowler
	horned lark	3435, 3436, 3437, 3438, 3439, 3440	UCB	04/22/1893	F. H. Fowler
	bushtit	3461	UCB	05/03/1894	F. H. Fowler
	cactus wren	3463, 3464, 3465, 3466, 3467, 3468, 3469, 3471, 3472, 3473, 3474, 3475	UCB	04/01/1893	F. H. Fowler
,	black-tailed gnatcatcher	78882	UCB	04/13/1894	F. H. Fowler
	Bendire's thrasher	3489	UCB	04/15/1894	F. H. Fowler
	curve-billed thrasher	12573, 12573	UA	UNKN	UNKN
	crissal thrasher	3483, 3484, 3486	UCB	04/05/1893	F. H. Fowler
	canyon towhee	3534, 3535, 3536	UCB	04/13/1893	F. H. Fowler
	black-throated sparrow	3544, 3545	UCB	04/21/1893	F. H. Fowler
	white-crowned sparrow	78975	UCB	05/15/1894	F. H. Fowler
	black-headed grosbeak	78931	UCB	05/15/1894	F. H. Fowler
	hooded oriole	3519	UCB	05/28/1894	F. H. Fowler
/lammal	Sandborn's long-nosed bat	23523	UA	09/22/1976	E. L. Roth
namma	Mexican long-tongued bat	23506	UA	09/22/1976	E. L. Roth
	cave myotis	23503, 23504	UA	08/08/1976	E. L. Roth
	cave myous	23621, 23622, 23623	UA	08/08/1976	T. & P. Vaughan
	fringed myotis	23522	UA	09/22/1976	E. L. Roth
	western pipistrelle	23502	UA	08/08/1976	E. L. Roth
	big brown bat	23505	UA	09/22/1976	E. L. Roth
	pallid bat	23501	UA	08/08/1976	E. L. Roth
	round-tailed ground squirrel	23461, 23462	UA	09/23/1976	E. L. Roth
	Harris' antelope squirrel	23241, 23493, 23498	UA	10/18/1975	E. L. Roth
	cliff chipmunk	23475, 23485, 23520, 23643	UA	10/18/1975	E. L. Roth
	Botta's pocket gopher	23473, 23478, 23473	UA	09/24/1976	E. L. Roth
	Sonoran Desert pocket mouse	23242, 23243, 23244, 23245, 23246, 23247, 23248, 23481, 23482, 23484, 23490, 23508, 23514, 23518, 23524,	UA .	03/24/1310	L. L. NOUI
		23525	UA	10/18/1975	E. L. Roth
	rock pocket mouse	23249, 23480, 23495, 23497, 24065	UA	10/18/1975	E. L. Roth
		23641	UA	11/01/1975	P. L. Dods
		23732	UA	08/29/1976	G. S. Mills
		24041	UA	08/07/1976	T. P. Vaughan
	Bailey's pocket mouse	23463, 23499, 23513, 23730	UA	09/23/1976	E. L. Roth
	hispid pocket mouse	23466, 23469, 23470	UA	09/24/1976	E. L. Roth

Group	Common name	Field collection numbers	Collection ^a	Date	Primary Collector
Mammal	banner-tailed kangaroo rat	23456, 23457	UA	09/23/1976	E. L. Roth
	Ord's kangaroo rat	23458, 23459	UA	09/23/1976	E. L. Roth
	Merriam's kangaroo rat	23228—23240, 23460, 23474, 23483, 23489, 23507, 23512, 23731	UA	10/18/1975	E. L. Roth
	fulvous harvest mouse	23467	UA	09/24/1976	E. L. Roth
	cactus mouse	23258—23266, 23476, 23477, 23491, 23496, 23500, 23509, 23515, 24064	UA	10/18/1975	E. L. Roth
	deer mouse	23521, 23527	UA	10/31/1975	E. L. Roth
	white-footed mouse	23486, 23526	UA	11/02/1975	E. L. Roth
	brush mouse	23267, 23479, 23511, 23517, 23642, 23743, 23744	UA	10/18/1975	E. L. Roth
	northern grasshopper mouse	23464	UA	09/23/1976	E. L. Roth
	southern grasshopper mouse	23250—23257, 23468, 23488, 23510	UA	10/31/1975	E. L. Roth
	western white-throated woodrat	23487, 23492, 23494, 23516, 23519, 23639, 23640, 23644, 24042, 24043,			
		24043	UA	11/02/1975	E. L. Roth
	hispid cotton rat	23471	UA	09/24/1976	E. L. Roth
	antelope jackrabbit	23465	UA	09/23/1976	E. L. Roth
	black-tailed jackrabbit	23439	UA	10/19/1975	E. L. Roth

^a CAS = Chicago Academy of Sciences; USNM = U.S. National Museum; UCO = University of Colorado; UCB = University of California, Berkeley; UA = University of Arizona

Appendix F. Summary of vegetation characteristics measured at bird survey stations, Fort Bowie NHS, 2004. See Appendix A for list of common names of plants.

Transect Station	Category	Species	Mean density	Transect Station	Category	Species	Mean density
Butter-				3	Subshrub	Opuntia spinosior	19.83
field 1	Subshrub	Gutierrezia sarothrae	33.69			Arctostaphylos pungens	118.98
		Haplopappus laricifolius	67.39			Dasylirion wheeleri	19.83
		Isocoma tenuisecta	89.85			Nolina microcarpa	39.66
		Prosopis velutina	11.23		Shrub	Arctostaphylos pungens	153.35
		Sideroxylon lanuginosum	22.46			Mimosa biuncifera	19.17
	Shrub	Rhus microphylla	5.19			Prosopis velutina	57.51
		Haplopappus laricifolius	5.19			Quercus arizonica	19.17
		Opuntia spinosior	5.19			Quercus emoryi	57.51
		Prosopis velutina	23.36			Quercus sp.	19.17
		Dasylirion wheeleri	2.60			Nolina microcarpa	57.51
		Nolina microcarpa	2.60		Tree	Juniperus osteosperma	1.85
		Sideroxylon lanuginosum	7.79	••••••		Arctostaphylos pungens	1.85
	Tree	Rhus microphylla	2.69			Prosopis velutina	1.85
		Juniperus monosperma	0.67			Quercus arizonica	1.85
		Juniperus osteosperma	0.67			Quercus emoryi	27.67
		Prosopis velutina	8.75			Pinus edulis	1.85
		Celtis reticulata	0.67		Cavity	Juniperus osteosperma	2.01
	Cavity	Juglans major	2.38			Quercus arizonica	2.01
		Celtis reticulata	0.79			Quercus emoryi	24.13
	Subshrub	Yucca baccata	121.36			Quercus sp.	2.01
	Cuboinub	Gutierrezia sarothrae	40.45	4	Subshrub	Agave palmeri	31.66
		Haplopappus laricifolius	121.36		Caboniab	Yucca baccata	63.33
		Isocoma tenuisecta	161.82			Haplopappus laricifolius	348.31
		Opuntia spinosior	80.91			Arctostaphylos pungens	63.33
		Arctostaphylos pungens	121.36			Prosopis velutina	94.99
		Mimosa biuncifera	80.91			Dasylirion wheeleri	31.66
		Nolina microcarpa	40.45		Shrub	Yucca baccata	9.91
	Shrub	Yucca baccata	22.40		Official	Arctostaphylos pungens	39.63
	Official	Haplopappus laricifolius	22.40			Prosopis velutina	59.44
		Juniperus monosperma	22.40			Quercus emoryi	19.81
		Arctostaphylos pungens	89.60			Dasylirion wheeleri	9.91
		Mimosa biuncifera	22.40			Nolina microcarpa	59.44
		Prosopis velutina	201.60		Tree	Juniperus monosperma	4.05
			44.80		Hee		10.13
		Quercus emoryi	22.40			Prosopis velutina	
	T	Dasylirion wheeleri			O-1.11.	Quercus emoryi	26.34
	Tree	Juniperus deppeana	3.46		Cavity	Quercus arizonica	1.66
		Juniperus osteosperma	6.93		0	Quercus emoryi	19.90
		Prosopis velutina	20.78	5	Subshrub	Gutierrezia sarothrae	88.61
		Quercus emoryi	24.24			Haplopappus laricifolius	443.07
		Pinus edulis	6.93			Arctostaphylos pungens	88.61
		Celtis reticulata	6.93			Dasylirion wheeleri	132.92
	Cavity	Juniperus deppeana	1.96			Nolina microcarpa	132.92
		Quercus emoryi	4.58		Shrub	Yucca baccata	16.80
		Pinus edulis	3.27			Juniperus deppeana	16.80
	Subshrub	Agave palmeri	39.66			Juniperus monosperma	16.80
		Yucca baccata	39.66			Arctostaphylos pungens	100.80
		Haplopappus laricifolius	79.32			Prosopis velutina	16.80
		Isocoma tenuisecta	19.83			Quercus arizonica	16.80
		Opuntia engelmannii	19.83			Quercus emoryi	16.80

Transect Station	Category	Species	Mean density
5	Shrub	Dasylirion wheeleri	117.60
		Nolina microcarpa	16.80
	Tree	Juniperus monosperma	4.92
		Quercus arizonica	4.92
		Quercus emoryi	7.87
		Garrya flavescens	1.97
	Cavity	Juniperus monosperma	0.63
		Quercus arizonica	1.90
		Quercus emoryi	6.96
		Quercus sp.	1.90
3	Subshrub	Yucca baccata	2.85
<u></u>		Haplopappus laricifolius	14.23
		Opuntia spinosior	1.42
		Arctostaphylos pungens	1.42
		Mimosa biuncifera	1.42
		Quercus emoryi	4.27
		Nolina microcarpa	2.85
	Chruh		
	Shrub	Agave palmeri	14.56
		Arctostaphylos pungens	58.22
		Mimosa biuncifera	29.11
		Prosopis velutina	72.78
		Quercus emoryi	14.56
		Nolina microcarpa	101.89
	Tree	Juniperus monosperma	11.25
		Prosopis velutina	3.75
		Quercus emoryi	22.50
	Cavity	Quercus emoryi	5.54
		Quercus sp.	0.40
7	Subshrub	Yucca baccata	139.84
		Baccharis sarothroides	69.92
		Haplopappus laricifolius	559.34
		Opuntia spinosior	139.84
		Arctostaphylos pungens	209.75
		Mimosa biuncifera	69.92
		Nolina microcarpa	209.75
	Shrub	Yucca baccata	14.99
	240	Juniperus monosperma	14.99
		Arctostaphylos pungens	44.97
		Mimosa biuncifera	14.99
		Prosopis velutina	29.98
			179.86
	Troc	Nolina microcarpa	
	Tree	Juniperus monosperma	6.49
		Prosopis velutina	2.60
	O-1-1-	Quercus emoryi	16.87
	Cavity	Juniperus monosperma	0.61
		Quercus arizonica	1.82
		Quercus emoryi	6.06
		Quercus sp.	0.61
3	Subshrub	Agave palmeri	123.43
		Haplopappus laricifolius	617.13
		Mimosa biuncifera	431.99
		Dasylirion wheeleri	61.71
	Shrub	Agave palmeri	1.46
		Juniperus monosperma	2.91

Transect Station	t Category	Species	Mean density
8	Shrub	Mimosa biuncifera	10.19
<u>.</u>		Prosopis velutina	4.37
		Dasylirion wheeleri	2.91
		Nolina microcarpa	7.28
	Tree	Juniperus monosperma	6.00
	1166		1.50
		Prosopis velutina	
		Quercus arizonica	2.25
		Quercus emoryi	4.50
		Garrya flavescens	0.75
	Cavity	Juniperus deppeana	0.24
		Quercus arizonica	0.94
		Quercus emoryi	2.83
	Canyon		470.00
	Subshrub	Agave palmeri	172.80
		Yucca baccata	691.19
		Chrysothamnus nauseosus	172.80
		Gutierrezia sarothrae	345.59
		Haplopappus laricifolius	518.39
		Parthenium incanum	518.39
		Juniperus osteosperma	172.80
		Mimosa biuncifera	172.80
		Garrya flavescens	172.80
		Nolina microcarpa	172.80
		Sideroxylon lanuginosum	345.59
	Shrub	Rhus microphylla	170.36
	Olliub	Haplopappus laricifolius	56.79
		Juniperus osteosperma	56.79
		Mimosa biuncifera	113.58
			56.79
		Prosopis velutina	
		Quercus arizonica	113.58
		Quercus turbinella	170.36
		Garrya flavescens	113.58
		Sapindus saponaria	56.79
		Sideroxylon lanuginosum	170.36
		Celtis reticulata	56.79
	Tree	Juniperus monosperma	8.13
		Juniperus osteosperma	8.13
		Prosopis velutina	8.13
		Quercus arizonica	4.07
		Quercus turbinella	8.13
•••••		Fouquieria splendens	8.13
		Garrya flavescens	12.20
		Sapindus saponaria	4.07
		Sideroxylon lanuginosum	12.20
		Celtis reticulata	
	Covity		8.13
	Cavity	Juniperus monosperma	2.15
		Juniperus osteosperma	4.31
		Quercus arizonica	8.62
		Quercus sp.	2.15
		Juglans major	2.15
		Fraxinus velutina	10.77
		Celtis reticulata	2.15
<u>)</u>	Subshrub	Unknown species	81.73
		Gutierrezia sarothrae	81.73

Transect Station	Category	Species	Mean density
2	Subshrub	Haplopappus laricifolius	163.46
		Isocoma tenuisecta	245.19
		Opuntia engelmannii	81.73
		Opuntia spinosior	163.46
		Prosopis velutina	81.73
		Sideroxylon lanuginosum	572.10
		Celtis reticulata	163.46
	Shrub	Rhus microphylla	237.17
		Baccharis sarothroides	79.06
		Opuntia engelmannii	158.12
		Prosopis velutina	474.34
		Sideroxylon lanuginosum	474.34
		Celtis reticulata	158.12
	Tree	Rhus microphylla	4.19
	1166	Juniperus monosperma	8.39
		Prosopis velutina	29.36
		•••••••••••••••••••••••••••••••••••••••	8.39
		Fouquieria splendens	20.97
		Sideroxylon lanuginosum	
	O 3-	Celtis reticulata	12.58
	Cavity	Juniperus monosperma	4.66
		Prosopis velutina	1.55
		Quercus arizonica	2.33
		Juglans major	0.78
		Celtis reticulata	4.66
	Subshrub	Rhus microphylla	29.36
		Brickellia sp.	88.08
		Gutierrezia sarothrae	117.44
		Haplopappus laricifolius	29.36
		Isocoma tenuisecta	117.44
		Opuntia engelmannii	58.72
		Opuntia spinosior	58.72
		Datura meteloides	58.72
		Celtis reticulata	29.36
	Shrub	Unknown species	6.01
	Omab	Rhus microphylla	6.01
		Haplopappus laricifolius	18.02
		Chilopsis linearis	24.02
		Opuntia spinosior	12.01
		Prosopis velutina	18.02
		Sideroxylon lanuginosum	6.01
		Lycium pallidum	6.01
	Т	Celtis reticulata	24.02
	Tree	Rhus microphylla	16.68
		Chilopsis linearis	16.68
		Prosopis velutina	20.02
		Juglans major	3.34
		Celtis reticulata	10.01
	Cavity	Chilopsis linearis	5.38
		Prosopis velutina	2.69
		Quercus emoryi	1.35
		Juglans major	5.38
	Cavity	Celtis reticulata	4.04
	Subshrub	Brickellia sp.	41.67

Station	Category	Species	Mean density
4	Subshrub	Haplopappus laricifolius	31.26
		Opuntia spinosior	20.84
		Ephedra sp.	10.42
		Sideroxylon lanuginosum	83.35
	Shrub	Rhus microphylla	105.49
		Brickellia sp.	70.32
		Prosopis velutina	35.16
		Sideroxylon lanuginosum	140.65
	Tree	Rhus microphylla	20.93
		Prosopis velutina	34.88
		Juglans major	6.98
		Sideroxylon lanuginosum	27.90
		Celtis reticulata	48.83
	Cavity	Chilopsis linearis	11.21
		Prosopis velutina	3.74
		Juglans major	3.74
		Sideroxylon lanuginosum	3.74
		Celtis reticulata	44.86
5	Subshrub	Anisacanthus thurberi	15.71
		Rhus microphylla	31.42
		Brickellia sp.	47.13
		Chrysothamnus nauseosus	15.71
		Gutierrezia sarothrae	15.71
		Isocoma tenuisecta	47.13
		Parthenium incanum	31.42
		Prosopis velutina	47.13
		Garrya flavescens	15.71
		Sideroxylon lanuginosum	31.42
		Lycium pallidum	15.71
	Shrub	Rhus microphylla	4.43
		Brickellia sp.	2.22
		Chilopsis linearis	2.22
		Prosopis velutina	13.30
		Sideroxylon lanuginosum	15.52
		Celtis reticulata	4.43
		Larrea tridentata	2.22
	Tree	Chilopsis linearis	3.07
		Juniperus monosperma	1.54
		Prosopis velutina	10.75
		Fouquieria splendens	3.07
		Sideroxylon lanuginosum	9.22
		Celtis reticulata	3.07
	Cavity	Chilopsis linearis	3.08
		Prosopis velutina	3.08
		Juglans major	6.16
		Celtis reticulata	10.78
3	Subshrub	Brickellia sp.	34.33
		Chrysothamnus nauseosus	45.77
		Parthenium incanum	22.89
		Opuntia engelmannii	34.33
		Opuntia spinosior	22.89
		Juglans major	11.44
		Sideroxylon lanuginosum	45.77
		Lycium pallidum	11.44

	Mean density
а	15.12
nauseos	us 90.72
3	30.24
ns	45.36
3	15.12
а	15.12
ginosum	75.60
-	15.12
a	9.33
3	32.65
sperma	4.67
3	13.99
dens	9.33
40110	4.67
kii	4.67
	13.99
3	1.39
sperma	0.35
ρυπια	1.39
	0.35
ginosum	
giriosuri	2.44
9 :	41.00
roides	20.50
: - : C - 1!	20.50
icifolius 	20.50
annii	20.50
sperma	20.50
	61.50
la	41.00
olia	20.50
	20.50
ginosum	41.00
	20.50
	61.50
э	88.51
roides	59.00
	118.01
olia	88.51
ginosum	118.01
	118.01
9	15.73
sperma	3.15
dens	3.15
la	3.15
ginosum	
	25.17
sperma	0.60
	2.97
 3	1.19
	1.19
	3.57
ii	

Transect Station	Category	Species	Mean density
}	Subshrub	Brickellia sp.	20.16
		Gutierrezia sarothrae	20.16
		Haplopappus laricifolius	120.96
		Opuntia engelmannii	120.96
		Acacia greggii	20.16
		Mimosa biuncifera	20.16
		Prosopis velutina	20.16
		Fouquieria splendens	20.16
		Sideroxylon lanuginosum	40.32
	Shrub	Unknown species	12.77
		Agave palmeri	12.77
		Brickellia sp.	12.77
		Opuntia engelmannii	25.55
		Acacia greggii	51.09
		Mimosa biuncifera	25.55
		Prosopis velutina	12.77
		Quercus arizonica	12.77
		Fouquieria splendens	12.77
		Nolina microcarpa	12.77
		Sideroxylon lanuginosum	63.86
	Tree	Acacia greggii	3.17
		Prosopis velutina	6.34
		Quercus arizonica	6.34
		Quercus turbinella	3.17
		Fouquieria splendens	25.34
		Sideroxylon lanuginosum	15.84
		Celtis reticulata	3.17
	Cavity	Quercus arizonica	5.27
		Juglans major	1.76
		Fraxinus velutina	8.79
		Celtis reticulata	5.27

Appendix F, part 2

	Li	tter	Bare Ground		Ro	ck
Transect	Mean	SD	Mean	SD	Mean	SD
Butterfield	30	18.4	68	18.2	1	3.1
	50	29.0	51	30.1	4	5.0
	36	23.5	55	22.4	11	15.0
	35	15.7	61	16.5	7	9.2
	29	22.0	43	18.7	27	14.9
	29	12.1	64	18.5	6	9.4
	52	23.3	47	22.7	3	4.7
	22	16.4	76	16.0	6	7.6
Siphon Canyon	55	24.2	33	19.2	9	8.8
	52	27.5	41	24.6	7	12.3
	59	27.4	38	20.9	4	7.5
	40	32.6	60	32.1	1	3.1
	60	34.3	37	31.2	5	11.0
	28	22.7	62	24.6	8	10.7
	58	32.1	25	24.8	16	25.0
	29	26.2	49	33.4	22	18.2

Appendix G. Most abundant bird species at each transect and season based on data published in Russell and Johnson (1976) and the UA inventory. Relative abundance (RA) estimates from Russell and Johnson are number of individuals per transect kilometer whereas RA estimates for the UA effort are the mean number of individuals per transect station or section, including flyovers and birds seen > 75 m or > 100 m from stations or sections, respectively. See methods section for community descriptions. Species in bold-faced type are species that are not found on the corresponding list of the most common species by season. Relative abundance estimates are for showing abundance ranks and cannot be compared between studies because of different methods of data collection.

			St	tudy	
		Russell and Johnson	UA		
Community	Season	Species	RA	Species	RA
Mesquite-grasslands	Breeding	Gambel's quail	4.3	mourning dove	1.9
		black-throated sparrow	2.7	black-throated sparrow	1.8
		canyon towhee	1.7	turkey vulture	1.7
		cactus wren	1.3	canyon towhee	1.3
		mourning dove	0.8	Gambel's quail	1.1
		Mexican jay	0.6	white-winged dove	1.1
		violet-green swallow	0.6	ash-throated flycatcher	1.1
		verdin	0.5	cactus wren	1.1
		Wilson's warbler	0.5	Bewick's wren	1.0
		western tanager	0.4	northern mockingbird	0.6
		ash-throated flycatcher.	0.4	rufous-crowned sparrow	0.6
	Non-breeding	chipping sparrow	11.0	bushtit	1.2
		Brewer's sparrow	9.9	chipping sparrow	1.0
		black-throated sparrow	6.0	western bluebird	0.8
		white-crowned sparrow	3.7	white-crowned sparrow	0.7
		Gambel's quail	2.6	bridled titmouse	0.4
		canyon towhee	2.4	canyon towhee	0.4
		dark-eyed junco	2.4	phainopepla	0.3
		cactus wren	0.7	western scrub-jay	0.3
		ladder-backed woodpecker	0.7	ruby-crowned kinglet	0.3
Oak-juniper woodland	Breeding	Gambel's quail	5.1	mourning dove	2.4
		Mexican jay	1.7	black-throated sparrow	1.5
		western scrub-jay	1.5	Bewick's wren	1.2
		mourning dove	1.5	ash-throated flycatcher	0.8
		cactus wren	0.7	canyon towhee	0.7
		rufous-crowned sparrow.	0.7	Gambel's quail	0.7
		Bewick's wren	0.6	brown-headed cowbird	0.6
		violet-green swallow	0.6	northern mockingbird	0.6
		northern cardinal	0.5	rufous-crowned sparrow	0.6
		black-chinned hummingbird	0.5	cactus wren	0.5
		ash-throated flycatcher	0.4	white-winged dove	0.5
	Non-breeding	Gambel's quail	5.4	Gambel's quail	3.3
		chipping sparrow	4.2	white-crowned sparrow	2.6
		white-crowned sparrow	4.2	chipping sparrow	1.2
		dark-eyed junco	4.2	Brewer's sparrow	0.7
		Brewer's sparrow	1.5	spotted towhee	0.4
		western scrub-jay	1.5	black-throated sparrow	0.2
		mourning dove	1.0	ruby-crowned kinglet	0.2
		canyon towhee	0.8	western scrub-jay	0.2
		cactus wren	0.8	cactus wren	0.2
		ruby-crowned kinglet	0.8	canyon towhee	0.2
		spotted towhee	0.6	crissal thrasher	0.2
		Bewick's wren	0.6	northern flicker	0.2
Vash-riparian	Breeding	black-chinned hummingbird	5.8	Gambel's quail	1.9
	-	northern cardinal	4.6	white-winged dove	1.4

	Season	Study			
		Russell and Johnson		UA	
Community		Species	RA	Species	RA
Wash-riparian	Breeding	mourning dove	3.6	northern cardinal	1.2
		white-winged dove	2.8	mourning dove	1.1
		cactus wren	2.7	Cassin's kingbird	0.9
		Gambel's quail	2.1	cactus wren	0.9
		canyon towhee	2.0	Bewick's wren	8.0
		verdin	1.8	northern mockingbird	0.8
		hooded oriole	1.8	canyon towhee	0.8
Non-breeding		northern cardinal	6.6	white-crowned sparrow	3.0
		chipping sparrow	6.0	American robin	1.8
		white-crowned sparrow	5.8	vesper sparrow	1.8
		dark-eyed junco	5.3	spotted towhee	1.8
		canyon towhee	3.9	Brewer's sparrow	1.7
		spotted towhee	3.7	chipping sparrow	1.3
		Brewer's sparrow	3.5	canyon towhee	1.1
		Gambel's quail	3.1	Gambel's quail	1.1
		cactus wren	2.9	green-tailed towhee	1.1
		curved-bill thrasher	2.0	western scrub-jay	1.1

Appendix H. Number of Trailmaster photos and total number of individuals of each species. Data from Herman-Reese (unpublished data), Fort Bowie NHS, 2000–2001.

Group	Common name	Number of photographs	Total number of individuals photog	graphed
Reptile	western diamond-backed rattlesnake	1	2	
Bird	white-winged dove	1	2	
	mourning dove	3	3	
	greater roadrunner	1	3	
	Mexican jay	1	1	
	northern mockingbird	1	1	
	lark sparrow	28	>55	
	northern cardinal	1	1	
	pyrrhuloxia	3	3	
	Bullock's oriole	2	4	
Mammal	American black bear	36	37	
	white-nosed coati	8	9	
	ringtail		3	3
	striped skunk	2	2	
	coyote		3	3
	common gray fox	41	43	
	mountain lion	1	1	
	bobcat		2	2
	rock squirrel	1	1	
	mule deer		17	40

Appendix I. Photographic vouchers taken by University of Arizona inventory personnel, Fort Bowie NHS, 2002–2004.

Group	Common name	
Reptile	greater earless lizard	
	Gila monster	
	green rat snake	
Bird	common black-hawk	
	mourning dove	
	Bell's vireo	
	canyon towhee	
Mammal	American black bear	
	white-nosed coati	
	striped skunk	
	hooded skunk	
	feral dog	
	common gray fox	
	mountain lion	
	bobcat	
	collared peccary	
	mule deer	