

Universidade Federal de Minas Gerais
Instituto de Ciências Biológicas
Departamento de Morfologia

Tese de Doutorado

**EFEITOS DA CONTAMINAÇÃO POR METAIS PESADOS E
BIOMARCADORES DE IMPACTO AMBIENTAL EM PEIXES DA
BACIA DO RIO SÃO FRANCISCO, MG**

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Instituto de Ciências Biológicas
Universidade Federal de Minas Gerais
Setembro/2019

LOURENÇO ALMEIDA SAVASSI

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BIOMARCADORES DE IMPACTO AMBIENTAL EM PEIXES DA
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Tese apresentada ao Programa de Pós-Graduação em Biologia Celular do Departamento de Morfologia, do Instituto de Ciências Biológicas, da Universidade Federal de Minas Gerais, como requisito parcial para obtenção do título de Doutor em Ciências.

Área de concentração: Biologia Celular

Orientador: Dr. Nilo Bazzoli

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Às **oito horas** do dia **30 de setembro de 2019**, reuniu-se, no Instituto de Ciências Biológicas da UFMG, a Comissão Examinadora da Tese, indicada pelo Colegiado do Programa, para julgar, em exame final, o trabalho final intitulado: "**EFEITOS DA CONTAMINAÇÃO POR METAIS PESADOS E BIOMARCADORES DE IMPACTO AMBIENTAL EM PEIXES DA BACIA DO RIO SÃO FRANCISCO, MG**", requisito final para obtenção do grau de Doutor em Biologia Celular. Abrindo a sessão, o Presidente da Comissão, **Dr. Nilo Bazzoli**, após dar a conhecer aos presentes o teor das Normas Regulamentares do Trabalho Final, passou a palavra ao candidato, para apresentação de seu trabalho. Seguiu-se a arguição pelos examinadores, com a respectiva defesa do candidato. Logo após, a Comissão se reuniu, sem a presença do candidato e do público, para julgamento e expedição de resultado final. Foram atribuídas as seguintes indicações:

Prof./Pesq.	Instituição	Indicação
Dr. Nilo Bazzoli	UFMG	APROVADO
Dr. José Enemir dos Santos	PUC MINAS	APROVADO
Dr. Lucas Marcon	UEMG	APROVADO
Dr. Guilherme Mattos Jardim Costa	UFMG	APROVADO
Dra. Gleide Fernandes de Avelar	UFMG	APROVADO

Pelas indicações, o candidato foi considerado: _____

O resultado final foi comunicado publicamente ao candidato pelo Presidente da Comissão. Nada mais havendo a tratar, o Presidente encerrou a reunião e lavrou a presente ATA, que será assinada por todos os membros participantes da Comissão Examinadora. **Belo Horizonte, 30 de setembro de 2019.**

Dr. Nilo Bazzoli (Orientador) _____

Dr. José Enemir dos Santos _____

Dr. Lucas Marcon _____

Dr. Guilherme Mattos Jardim Costa _____

Drª. Gleide Fernandes de Avelar _____

Prof. Vanessa Pinho da Silva
Sub-Coordenadora do Programa de
Pós Graduação em Biologia Celular ICB/UFMG

Obs: Este documento não terá validade sem a assinatura e carimbo do Coordenador

Vanessa Pinho da Silva

O presente trabalho foi desenvolvido no laboratório de Ictiohistologia, do Departamento de Morfologia, do Programa de Pós-Graduação em Biologia Celular, Instituto de Ciências Biológicas, UFMG, coordenado pela Prof^a. Dr^a. Elizete Rizzo, para desenvolvimento geral do projeto, adicionalmente incluindo todo o apoio técnico disponibilizado pelo laboratório de Ictiologia, do Programa de Pós-Graduação em Biologia de Vertebrados, PUC Minas, coordenado pelo Prof. Dr. Nilo Bazzoli.

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RESUMO

Metais pesados são elementos inorgânicos que podem provocar sérios danos à saúde dos organismos vivos em geral. Através de resíduos industriais, rurais e domésticos, metais como Cádmio (Cd), Chumbo (Pb), Cobre (Cu), Cromo (Cr), Ferro (Fe) e Zinco (Zn), atingem constantemente os ambientes aquáticos. Esses poluentes geralmente não são degradados naturalmente e possuem alta capacidade de bioacumulação em tecidos e órgãos gerando graves patologias. Diante desse problema ambiental o objetivo do presente estudo foi avaliar, através de análises histológicas e imunohistoquímicas, o estado de contaminação do dourado, *Salminus franciscanus*, espécie de peixe de grande porte, endêmico da bacia do rio São Francisco, MG, que representa alto valor comercial e é amplamente consumido na alimentação da população. Foram avaliados os níveis de Cd, Cr, Cu, Fe, Pb e Zn, no fígado e no tecido muscular, de 68 exemplares de *S. franciscanus* coletados em dois rios da bacia do rio São Francisco, MG, Brasil: Abaeté (pouca atividade antrópica) e Paraopeba, trechos A e B (alta atividade antrópica). Em conjunto foram avaliadas alterações histopatológicas no fígado e baço (órgãos de intenso metabolismo), e expressão de três proteínas consideradas atualmente biomarcadoras de impacto ambiental: Metalotioneína (MT), Proteína de Choque Térmico-70 (HSP70) e Citocromo P450-1A (CYP1A). Os resultados mostraram que os peixes do rio Paraopeba estão impróprios ao consumo humano, ultrapassando os limites seguros ao consumo no filé (Cd, Cr, Cu e Pb acima do limite permitido) estabelecidos pela Organização Mundial de Saúde (OMS) e pela Agência Nacional de Vigilância Sanitária (ANVISA). Além dos elevados níveis de metais pesados na musculatura dos peixes, no fígado foram registrados valores acima do limite legal para os metais Cd, Cr, Cu, Fe, Pb e Zn (todos os metais analisados) sendo registradas e quantificadas alterações histopatológicas no fígado e baço dos peixes, indicando diferenças significativas à pior qualidade ambiental e ao cenário de maior contaminação por metais pesados, encontrado no rio Paraopeba. Também foram registradas diferenças significativas nos níveis de expressão das proteínas MT, HSP70 e CYP1A, entre os rios Abaeté e Paraopeba, evidenciando a alta contaminação em ambos os trechos de amostragem no rio Paraopeba em relação ao rio Abaeté.

Palavras-chave: Bioindicadores; Poluição aquática; Toxicologia; Teleósteos.

ABSTRACT

Heavy metals are inorganic elements that can cause serious health problems in living organisms in general. Through industrial, rural and domestic waste, metals such as Cadmium (Cd), Lead (Pb), Copper (Cu), Chromium (Cr), Iron (Fe) and Zinc (Zn) constantly reach aquatic environments. These pollutants are usually not naturally degraded and have a high capacity of bioaccumulation in tissues and organs inducing pathologies. Due this environmental problem, the objective of the present study was to evaluate, through histological and immunohistochemical analyzes, the condition of contamination of dourado, *Salminus franciscanus*, a large fish species, endemic to the São Francisco River Basin, MG, which represents a high commercial value and is widely consumed in population feeding. Levels of Cd, Cr, Cu, Fe, Pb and Zn was quantified in the liver and muscle tissue, of 68 specimens of *S. franciscanus* collected from two rivers of the São Francisco River Basin, MG, Brazil: Abaeté (low anthropic activity) and Paraopeba, sites A and B (high anthropic activity). Histopathological alterations in the liver and spleen (organs of intense metabolism) and expression of three proteins currently considered biomarkers of environmental impact, were evaluated: Metallothionein (MT), Heat Shock Protein-70 (HSP70) and Cytochrome P450-1A (CYP1A). The results evidenced that fish from the Paraopeba River are inappropriate for human consumption, exceeding the safe limits for muscle consumption (Cd, Cr, Cu and Pb above the permitted limit) established by the World Health Organization (WHO) and the Agência Nacional de Vigilância Sanitária (ANVISA). In addition to the high levels of heavy metals registered in fish muscles, in liver the values were recorded above the legal limit for Cd, Cr, Cu, Fe, Pb and Zn metals (all metals analyzed) and histopathological alterations in liver and spleen were recorded and quantified, indicating significant differences in the poor environmental quality and the scenario of high contamination by heavy metals found in Paraopeba River. Significant differences in expression levels of MT, HSP70 and CYP1A proteins, were also recorded between the Abaeté and Paraopeba Rivers, evidencing the high contamination in both sampling sites of Paraopeba River in relation to Abaeté River.

Keywords: Bioindicators; Teleosts; Toxicology; Water pollution.

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1. INTRODUÇÃO GERAL

O crescimento mundial da população provocou a expansão de atividades urbanas e rurais, desencadeando um aumento nas atividades industriais e na produção de resíduos químicos, que atualmente são produzidos em larga escala, constituindo fontes poluidoras dos recursos hídricos, fauna e flora (Cao et al., 2017; Kwok et al., 2014; Jiang et al., 2018). A poluição dos corpos d'água implica em substâncias tóxicas que afetam a saúde dos peixes e conseqüentemente da população que utiliza essa carne na alimentação (Barone et al., 2013; Yi et al., 2017; Zhong et al., 2018).

O aumento da contaminação aquática por metais pesados, provenientes de atividades antrópicas como minerações, agropecuária e indústrias, atualmente vem se tornando um problema ambiental cada vez mais reportado em estudos toxicológicos (Ribeiro et al., 2012, Squadrone et al., 2013). Metais pesados são persistentes e não se degradam, acumulando-se nos tecidos de peixes e outros organismos aquáticos (Marques et al., 2009; D'Costa et al., 2017; Lunardelli et al., 2018). Neste sentido, a capacidade dos peixes em assimilar compostos inorgânicos no organismo, os tornam importantes bioindicadores ambientais (Gomes & Sato, 2011; Tabinda et al., 2013; Luczynska et al., 2018). A capacidade bioacumulativa dos peixes os tornam interessantes modelos de estudo para eventos da biologia celular, tais como a expressão de proteínas bioindicadoras de impacto ambiental, assim como ferramentas ecológicas para uma avaliação da qualidade do meio ambiente refletida no organismo vivo (De la Torre et al., 2007; Thomé et al., 2009, Monferran et al., 2016).

A carne dos peixes para a alimentação humana é reconhecida como excelente fonte de vitaminas, proteínas e minerais, porém se torna necessária uma eficaz avaliação das concentrações de metais pesados e outros contaminantes na carne, para um consumo seguro da população (Souza, 2003; Reis et al., 2009; Taweel et al., 2013). Nos peixes, os metais pesados podem causar distúrbios no crescimento e na reprodução, além de alterações histopatológicas na pele, brânquias, fígado, baço e rins (Vitek et al., 2007). O fígado e o baço participam do metabolismo e detoxificação de substâncias no organismo, e o tecido muscular, amplamente utilizado na alimentação da população, sendo assim, ambos devem ser analisados em relação ao nível de contaminação antes do consumo humano (Mendil et al., 2005; Castro-González & Méndez-Armenta, 2008; Subotić et al., 2013).

Para documentar e quantificar os efeitos de poluentes nos ambientes aquáticos, alguns biomarcadores fisiológicos e histopatológicos de peixes vêm sendo utilizados, tais

como: apoptose, proteínas do choque térmico, metalotioneínas, alterações hormonais e reprodutivas, além de alterações histopatológicas em órgãos alvo, tais como: baço, brânquias, fígado e gônadas (Santos et al., 2005; Fishelson, 2006; Thomé et al., 2009; Prado et al., 2011; Arantes et al., 2016; Savassi et al., 2016; Paschoalini et al., 2019). A utilização desses biomarcadores de impacto ambiental em conjunto às análises de metais pesados, fornecem informações cruciais para o entendimento da influência de metais pesados sobre o organismo, assim como ajudar agências governamentais a criar atualizados limites de contaminantes, na carne dos peixes.

1.1 Metais pesados

Metais pesados são definidos como elementos metálicos que apresentam elevada massa e número atômico, alta densidade, capacidade de formar sulfetos e hidróxidos insolúveis em água, serem de fácil absorção por um organismo vivo, além de alta capacidade tóxica em baixas concentrações (Fergusson, 1990; Tchounwou et al., 2012). Nos últimos anos, tem surgido uma crescente preocupação ecológica, e de saúde pública global, associada à contaminação ambiental por metais pesados. Devido ao aumento exponencial do uso desses metais em várias aplicações industriais, agrícolas, domésticas e tecnológicas, tem sido reportado em estudos científicos um aumento da exposição humana a esses elementos (Jia et al., 2018; Hossain et al., 2018; Luczyska et al., 2018).

Estudos recentes apontam metais pesados amplamente distribuídos nos ambientes aquáticos (Figura 1), dentre eles Arsênio (As), Cádmio (Cd), Chumbo (Pb), Cobre (Cu), Mercúrio (Hg) e Zinco (Zn) que são altamente bioacumulativos e tendem a aumentar a concentração no organismo dos animais com o passar dos anos, pois não conseguem ser eliminados fisiologicamente (Squadrone et al., 2013; Protano et al., 2014). O Cromo (Cr) apesar de não indicar ser bioacumulativo, tem efeitos tóxicos sobre o organismo, assim como os outros elementos (Arantes et al., 2016; Masindi & Muedi, 2018). Esse acúmulo de compostos tóxicos no organismo provoca a formação de patologias em órgãos alvos, como a esteatose no fígado, fibrose no baço, lesões na porção cranial dos rins, necroses e câncer em diversos órgãos (Vieira et al., 2011; Thévenod & Lee, 2015; Paschoalini et al., 2019). Em concentrações elevadas, estes metais também podem interromper a homeostase de íons, induzir a danos no DNA e distúrbios em vários órgãos (Vieira et al., 2011; Nai et al., 2015; Li et al., 2015), afetando organelas celulares e componentes como a membrana celular, mitocôndria, lisossomo, retículo endoplasmático, núcleo e enzimas

envolvidas no metabolismo, desintoxicação e reparo de danos no organismo (Wang & Shi, 2001).

Intensificando a contaminação por metais pesados através da bioacumulação, quando um animal de cadeia alimentar superior se alimenta de um animal posicionado em posição inferior contaminado, ocorre a propagação da contaminação através da cadeia alimentar, fenômeno denominado de Biomagnificação (Suedel et al., 1994). Esse processo intensifica a contaminação pois os metais pesados têm alto poder bioacumulativo, aumentando sua concentração no organismo com o passar dos anos e conseqüentemente elevando seu potencial tóxico (Ali & Khan, 2018; Liu et al., 2019).

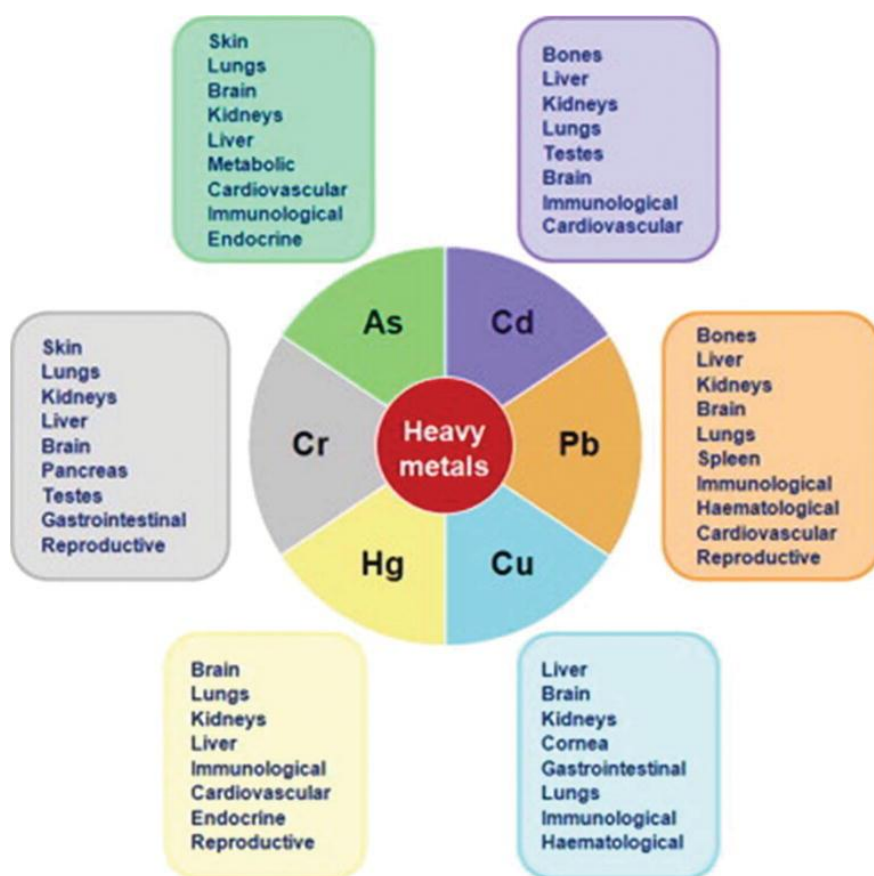


Figura 1. Sistemas e órgãos afetados pelos diferentes tipos de metais pesados, no organismo. Reproduzido de Masindi & Muedi, 2018.

1.2 Metalotioneínas

Metalotioneínas (MT) são uma classe de proteínas citosólicas, ricas em cisteínas, caracterizadas pelo baixo peso molecular e afinidade por cátions divalentes. As funções biológicas das MT envolvem desintoxicação e homeostase de íons metálicos, sendo importantes no metabolismo intracelular de cobre (Cu), zinco (Zn) e cádmio (Cd),

atuando na proteção contra o dano oxidativo e toxicidade resultante da exposição excessiva a metais-traço (Stillman, 1995; Sevcikova et al., 2013). Ainda se torna necessário elucidar o tipo de resposta de cada metal sobre os níveis de MT no organismo. A exposição crônica a metais pesados aumenta a produção de espécies reativas de oxigênio (ROS), como o peróxido de hidrogênio (H_2O_2), radical superóxido (O_2^-) e radical hidroxila (OH), podendo levar ao estresse oxidativo e causar peroxidação lipídica, danos no DNA e oxidar moléculas em membranas biológicas e tecidos, assim como demonstrado na Figura 2 (Atli & Canli, 2010; Mittler, 2002; Ruttkay-Nedecky et al., 2013). Órgãos relacionados ao intenso metabolismo e desintoxicação, como fígado e baço, são conhecidos locais de acúmulo de metais pesados e por isso devem ser direcionados a estudos que analisam a expressão de MT nesses órgãos de peixes teleósteos (Carvalho et al., 2004; Marijic & Raspor, 2006).

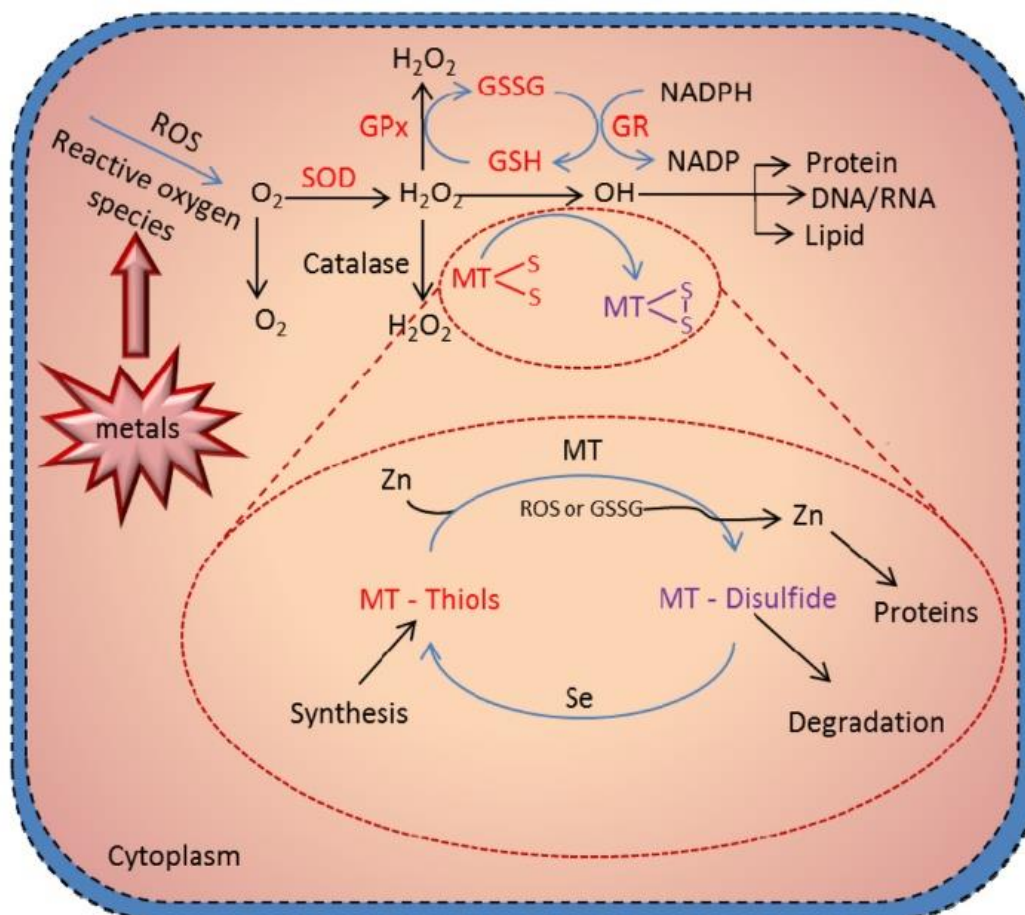


Figura 2. Desenho esquemático sobre a capacidade da MT como eliminadora de radicais livres e à indução a espécies reativas de oxigênio (ROS) nas células, causadas pelos metais pesados (Ruttkay-Nedecky et al., 2013).

Nos organismos aquáticos as concentrações de MT nos tecidos aumentam rapidamente, quando induzidos por exposição a metais-traço, como Cd, Cu, Zn e As, tanto em experimentos de laboratório quanto na natureza (Viarengo et al., 1999). Devido a sua especificidade e capacidade de se ligar a íons metálicos, a MT torna-se um excelente biomarcador, utilizado no monitoramento de ambientes aquáticos contaminados por metais-traço (Chan, 1995; El-Shehawi et al., 2007; Knapen et al., 2007; Rajeshkumar et al., 2013). Apesar de ser muito recomendada em estudos de contaminação por metais pesados, por confirmar a contaminação por metais pesados no organismo, a utilização de metalotioneínas em estudos toxicológicos, principalmente brasileiros, ainda é pouco estabelecida e necessita ser melhor investigada na ictiofauna.

1.3 Proteínas do choque térmico

Proteínas do choque térmico (Heat Shock Proteins) são altamente conservadas durante a evolução e funcionam como chaperonas moleculares que interferem na síntese proteica, promovendo a correta organização molecular de proteínas que se formaram de maneira incorreta (Feder & Hofmann, 1999; Parcellier et al., 2003). As HSP podem pertencer a diversas famílias e dentre elas a HSP70 é conhecida por interagir diretamente com elementos da via apoptótica, seja ela intrínseca ou extrínseca, inibindo a cascata de eventos que culminam com a morte celular (Mosser et al., 1997, Parcellier et al., 2003). Dentre os genes da família de resposta ao choque térmico, a HSP70 é um dos genes altamente conservados e o primeiro a ser induzido em resposta a diversos fatores estressantes gerados pelo ambiente sobre o organismo (Mukhopadhyay et al., 2003).

Estudos realizados em peixes teleósteos têm mostrado um aumento da expressão de HSP70, em locais onde a qualidade ambiental não é favorável à saúde dos peixes, devido a presença de poluentes ambientais, como metais pesados, sugerindo que o aumento destas proteínas está associado com a proteção celular, incluindo o aumento da apoptose em peixes submetidos experimentalmente a agentes tóxicos (Janz et al., 2001; Weber & Janz, 2001; Rajeshkumar et al., 2013). Sugere-se também, que metais pesados se acumulam nas células vivas e podem interagir com o grupo tiol das proteínas, influenciando na formação de proteínas anormais, as quais sinalizam para a indução de genes HSP70 (Figura 3) (Somasundaram et al., 2018). Estudos recentes apresentam resultados em que pode ocorrer um aumento na expressão da proteína HSP70 em casos de contaminação por metais pesados, porém essa relação ainda não é bem elucidada (Rajeshkumar et al., 2013; 2017).

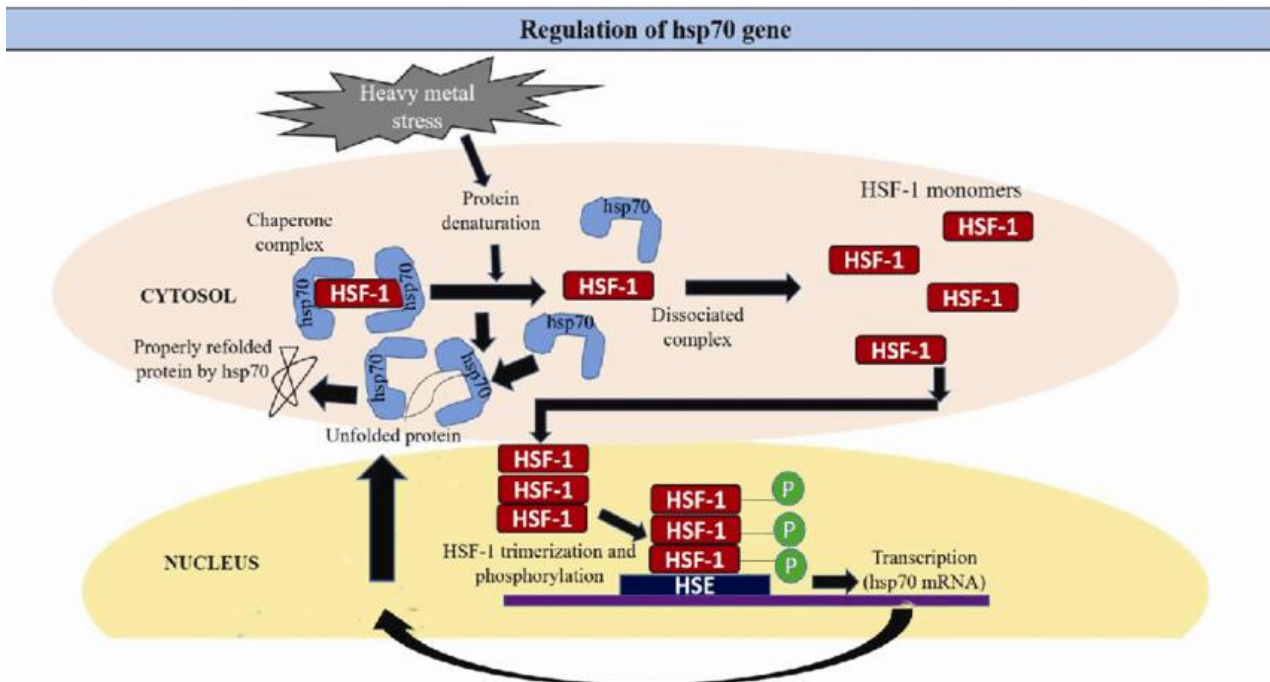


Figura 3. Desenho esquemático da indução de genes HSP70 sob contaminação por metais pesados (Somasundaram et al., 2018).

1.4 Citocromo P450

O citocromo P450 (CYP) representa uma família de enzimas responsável pela biotransformação de compostos orgânicos. A sub-família CYP1A tem alta afinidade por hidrocarbonetos policíclicos aromáticos que dependem basicamente do receptor AHR (Aryl Hydrocarbon Receptor). A indução inicia-se pela ligação do xenobiótico específico ao AHR, que atrai uma proteína de choque térmico 90 (HSP90), e se movimenta do citoplasma ao núcleo da célula (Kawajiri & Fujii-Kuriyama, 2007). No núcleo, a HSP90 se dissocia do complexo receptor-indutor. Após a liberação da HSP90, o complexo receptor-indutor liga-se a outra proteína, o translocador de receptor de hidrocarbonetos aromáticos (ARNT), que também se localiza no núcleo da célula. No núcleo, o complexo ARNT-AHR liga-se a uma região específica do DNA, o elemento de resposta à xenobiótico (XRE). Fatores transcricionais são utilizados para acessar a região promotora do gene CYP1A, e o RNA mensageiro é então sintetizado. Essa ativação gênica provoca um aumento na expressão e na atividade da enzima CYP1A, levando então a biotransformação, que resulta na detoxificação/bioativação dos contaminantes orgânicos (Figura 4) (Safe & Krishnan, 1995; Kawajiri & Fujii-Kuriyama, 2007).

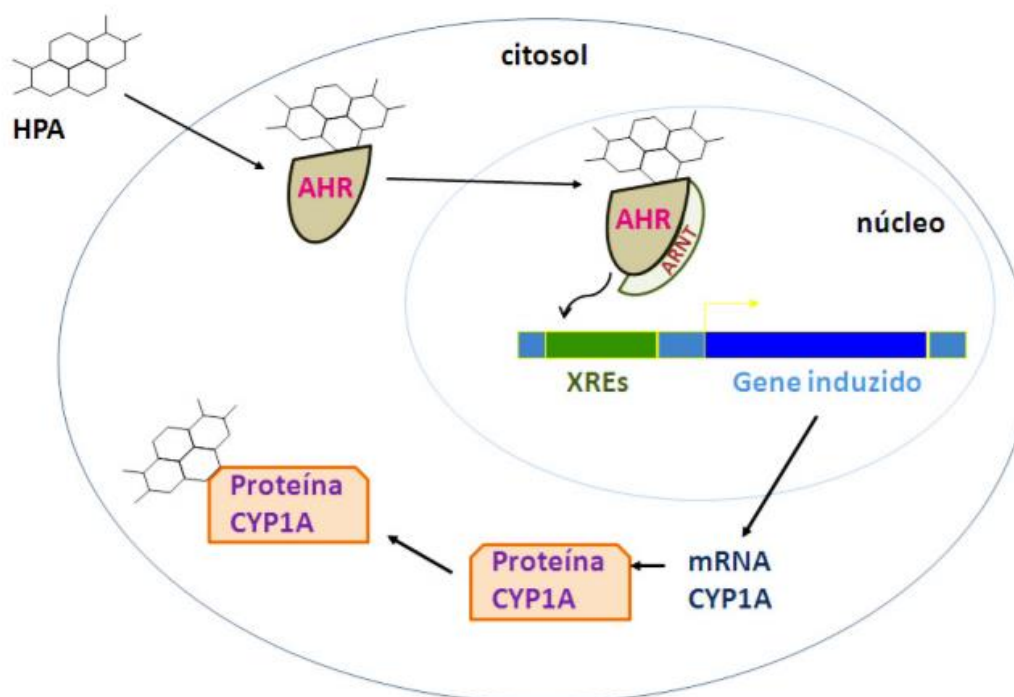


Figura 4. Desenho esquemático da indução do gene CYP1A na presença de um hidrocarboneto policíclico aromático (HPA) (Kawajiri & Fujii-Kuriyama, 2007).

1.5 Espécie em estudo

O *Salminus franciscanus* (Lima & Britski, 2007), anteriormente classificado como *Salminus brasiliensis* (Cuvier, 1816), é conhecido popularmente como dourado (Figura 5). É um peixe da família Bryconidae, sub-família Salmininae, endêmico da bacia do rio São Francisco, Minas Gerais. O dourado tem grande importância comercial e ecológica, podendo atingir mais de 1,0 metro de comprimento total, ultrapassando 25kg de peso corporal, com hábito alimentar essencialmente piscívoro, ocupando o topo da cadeia alimentar nos rios, o que favorece a bioacumulação de metais pesados. Em relação à biologia reprodutiva do dourado Freitas et al. (2013) observou que o peixe reproduz no alto e médio dos rios, no período de outubro a fevereiro, onde o fotoperíodo é maior, a água está mais quente e com maior taxa de oxigênio dissolvido, favorecendo a desova do tipo total, com ovos não-adesivos e alta fecundidade. Atualmente constitui um dos principais alvos da pesca comercial e esportiva (Petrere 1989; Sato & Godinho, 2003) e devido à crescente degradação ambiental e barramentos nos rios da bacia do rio São Francisco por usinas hidrelétricas e termelétricas, suas populações têm sido ameaçadas (Sato & Sampaio, 2005, Freitas et al., 2013).



Figura 5 – Exemplar de dourado, *Salminus franciscanus* (comprimento total - 55,0cm).

1.6 Área de estudo

O rio Paraopeba é considerado um dos principais afluentes do rio São Francisco, MG, sua nascente está localizada no município de Cristiano Otoni, MG, e sua foz na represa de Três Marias, município de Felixlândia, MG. Possui uma área de 13.643 km² e extensão aproximada de 510 km (IGAM, 2012^a). É classificado como uma área prioritária para a conservação, devido à alta diversidade de espécies endêmicas de peixes, reprodução de espécies de piracema e/ou por ser ambiente único no Estado (Costa et al., 1998), porém atualmente vêm sendo ameaçado pela contaminação por resíduos de galvanoplastia das indústrias de aço, resíduos da agricultura, curtumes, esgoto (doméstico e industrial) e rejeitos minerários, incluindo contaminantes como metais pesados, que excedem os níveis de segurança para a saúde humana, além da presença em seu curso, de uma usina termoeletrica e uma hidrelétrica.

Entretanto o rio Abaeté, com sua nascente localizada na Serra da Canastra, município de São Gotardo, MG e sua foz no rio São Francisco, MG, em um local denominado como Pontal do Abaeté, apresenta melhor qualidade ambiental, sendo um ambiente mais preservado de atividades antrópicas, apresentando pouca contaminação ambiental (IGAM, 2012^b). Também apresenta parâmetros físico-químicos adequados para o sucesso reprodutivo dos peixes, criando um ambiente propício e único para a manutenção da ictiofauna na bacia do rio São Francisco, MG (Arantes et al., 2010; Weber, et al., 2013; Nunes et al., 2015). Neste contexto, fica evidente a necessidade de estudos voltados ao diagnóstico da atual situação da ictiofauna na bacia do rio São Francisco, MG, e de sua sanidade. Devendo estes estudos, fornecer informações sobre como os

contaminantes estão presentes, e suas consequências, para com o ambiente natural, possibilitando constituir ações prioritárias de instituições de pesquisa e fiscalização ambiental.

2. JUSTIFICATIVA

O ambiente aquático está cada vez mais contaminado por resíduos industriais e domésticos, incluindo metais pesados, que posteriormente tendem a ser bioacumulados nos peixes e em toda a fauna aquática. Essa bioacumulação promove diversas alterações biológicas negativas no organismo vivo e os efeitos, ainda pouco descritos na literatura, precisam ser melhor investigados. A carência de estudos dessa natureza no Brasil, demonstra a necessidade de um maior número de pesquisas científicas nessa área de toxicologia ambiental e seus efeitos.

Na bacia do rio São Francisco, MG, estudos recentes vêm sendo realizados registrando altas concentrações de metais pesados bioacumulativos em peixes de interesse comercial, tornando os peixes impróprios para consumo humano, além do desenvolvimento de diversas alterações histopatológicas em órgãos alvo, e alterações reprodutivas. Estudos toxicológicos em ambientes contaminados, como aqui realizado, nos proporcionam utilizar o peixe como modelo experimental para se avaliar efeitos dos poluentes aquáticos sobre o organismo dos peixes, avaliando em conjunto a qualidade ambiental e fatores que afetam a saúde humana, como o consumo da carne contaminada por metais pesados.

3. OBJETIVOS

3.1 Objetivo geral

Avaliar a contaminação por metais pesados sobre alterações histopatológicas e expressão de proteínas biomarcadoras de impacto ambiental, em órgãos-alvo como fígado, baço e músculo, do dourado, *Salminus franciscanus*, coletados em dois rios da bacia do rio São Francisco, MG: Paraopeba e Abaeté.

3.2 Objetivos específicos

- Calcular os Índices: Gonodassomático (IGS), Hepatossomático (IHS) e Fator de Condição de Fulton (K), em machos e fêmeas;
- Comparar os valores de IGS, IHS e K entre os trechos de amostragem A e B do rio Paraopeba, com o rio Abaeté;
- Dosar as concentrações de metais pesados (Chumbo, Cobre, Cromo, Cádmio, Ferro e Zinco) em fragmentos de fígado e músculo dos peixes coletados nos diferentes trechos de amostragem;
- Correlacionar as concentrações de metais pesados obtidas com os valores seguros ao consumo humano (OMS/ANVISA);
- Detectar e quantificar as principais alterações histopatológicas no fígado e baço dos peixes;
- Avaliar as alterações na expressão de Metalotioneína (MT), Proteínas de Choque Térmico-70 (HSP70) e Citocromo P450-1A (CYP1A) no fígado dos peixes, via Imunohistoquímica Peroxidase;
- Imunolocalizar MT, HSP70 e CYP1A no fígado, através de técnicas Imunohistoquímicas Peroxidase;
- Comparar alterações histopatológicas e expressão de biomarcadores de qualidade ambiental entre os trechos de amostragem do presente estudo.

4. ARTIGO SUBMETIDO

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Title

Assessment of environmental heavy metal pollution and potential human health risk in a highly consumed fish, *Salminus franciscanus*, using an immunohistochemical and histopathological approach

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04-Sep-2019

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1 **ASSESSMENT OF ENVIRONMENTAL HEAVY METAL POLLUTION AND**
2 **POTENTIAL HUMAN HEALTH RISK IN A HIGHLY CONSUMED FISH,**
3 ***SALMINUS FRANCISCANUS*, USING AN IMMUNOHISTOCHEMICAL AND**
4 **HISTOPATHOLOGICAL APPROACH**

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12 **ABSTRACT**

13 Due to production of industrial, rural and domestic waste, heavy metals such as cadmium
14 (Cd), chrome (Cr), lead (Pb), zinc (Zn), copper (Cu) and iron (Fe) constantly reach aquatic
15 environments. The present study uses histological and immunohistochemical analyses to
16 evaluate the contamination status of *Salminus franciscanus*, a large fish of great economic
17 importance. Levels of Cd, Cr, Pb, Zn, Cu and Fe were evaluated for liver and muscle
18 tissue of 68 fish, from two tributaries of the upper São Francisco River Basin, Brazil:
19 Abaeté and Paraopeba. In addition, histopathological alterations and expression of three
20 biomarkers — metallothionein (MT), heat shock protein-70 (HSP70) and cytochrome
21 P450-1A (CYP1A) — were assessed. Fish from the Paraopeba River were found to be
22 inappropriate for human consumption because they exceed safe limits established by the
23 World Health Organization (WHO). Histopathological alterations in the liver and spleen
24 were significantly more frequently in fish from the Paraopeba River ($P < 0.05$). Significant
25 differences were also observed in the levels of expression of MT, HSP70 and CYP1A
26 proteins, regarding the heavy metal contamination levels. Fish from Abaeté River, on the
27 other hand, had higher values for IGS and low levels of metal contamination in liver and
28 muscle, and thus can be considered healthy for consumption and population
29 sustainability. We emphasize the socio-environmental importance of *S. franciscanus* as
30 an excellent environmental bioindicator.

31 **Keywords:** Bioaccumulation; Histopathology; Metallic ions; Teleosts; Trace elements;
32 Water contamination.

33 INTRODUCTION

34 Toxicological studies have recently reported increases in the contamination of
35 aquatic environments by heavy metals, mainly from activities such as mining, agriculture
36 and industry (Squadrone et al. 2013; Santolin et al. 2015; Zhong et al. 2018; Jiang et al.,
37 2019). Heavy metals do not degrade and thus tend to accumulate in organs and tissues of
38 fish and other aquatic organisms (D'Costa et al. 2017; Lunardelli et al. 2018). At high
39 concentrations these metals can also disrupt ion homeostasis, induce DNA damage, and
40 cause histopathological alterations and disorders in different organs (Vieira et al. 2011;
41 Nai et al. 2015). This accumulation of toxic compounds in organisms causes pathologies
42 in target organs, such as steatosis in the liver, fibrosis in the spleen, lesions in the cranial
43 portion of the kidneys and necrosis and cancer in several organs (Vieira et al. 2011;
44 Thévenod and Lee, 2015). Chronic exposure to heavy metals can also increase levels of
45 reactive oxygen species (ROS), such as hydrogen peroxide (H₂O₂), superoxide radical
46 (O₂⁻) and hydroxyl radical (OH⁻), thus triggering oxidative stress and cell death (Ercal et
47 al. 2001; Ratn et al. 2018; Ibor et al. 2019).

48 Liver and spleen are organs involved in intense metabolism and detoxification. As
49 such, they can accumulate certain pollutants, especially heavy metals, and thus are
50 suitable targets for toxicological studies (Alm-Eldeen et al. 2018). The expression of
51 certain liver proteins, especially metallothioneins (MTs) can be altered by exposure to
52 metals. These proteins are characterized by having low molecular weight and being
53 cysteine-rich. Moreover, they are important for homeostasis of essential metals and
54 detoxification of toxic metals by protecting against oxidative damage and toxicity
55 resulting from excessive exposure to heavy metals (Chan, 1995; Ruttkay-Nedecky et al.
56 2013, Sevcikova et al. 2013; Le et al. 2016). Due to their specificity and ability to bind to
57 metal ions, MTs are considered suitable biomarkers for monitoring aquatic environments
58 contaminated by heavy metals (Rajeshkumar et al. 2013).

59 Several biomarkers are widely used in toxicological and biomonitoring studies to
60 assess environmental stress in aquatic organisms, including heat shock proteins (HSPs)
61 and some cytochrome P450 family proteins, such as CYP1A. Heat shock proteins are
62 molecular chaperones that act in the formation and restructuring of proteins in the body
63 (Feder and Hofmann, 1999; Parcellier et al. 2003). Among the proteins of this family,
64 HSP70 is a highly conserved within vertebrates and the first to be induced in response to
65 several stressors (Mukhopadhyay et al. 2003). Heavy metals can accumulate in cells and

66 interact with the thiol group of proteins, generating an abnormal or denatured protein,
67 consequently up-regulating the expression of HSP70 (Somasundaram et al. 2018).
68 CYP1A is a member of the xenobiotic family of metabolic enzymes, acts on the
69 detoxification of contaminants in organs of intense metabolism (i.e., liver), and has high
70 affinity for polycyclic aromatic hydrocarbons that basically depend on the aryl
71 hydrocarbon receptor-AHR (Safe and Krishnan 1995; Kawajiri and Fujii-Kuriyama,
72 2007).

73 The Abaeté and Paraopeba Rivers are important tributaries for the maintenance of
74 native fish stocks in the São Francisco River, an important river basin in South-America
75 (Domingos et al. 2012). The Abaeté River has low levels of environmental contamination
76 and adequate water quality parameters for the successful reproduction of migratory fish,
77 and thus contributes to maintaining ichthyofauna (Arantes et al. 2010; Weber et al. 2013;
78 Nunes et al. 2015; Paschoalini et al. 2019). However, high levels of domestic and
79 agricultural sewage, and effluents from mining operations and tanneries have been
80 reported for the Paraopeba River, which negatively affect fish species of high commercial
81 value (IGAM 2012A; Arantes et al. 2016; Savassi et al. 2016; Paschoalini et al. 2019).

82 *Salminus franciscanus* (Lima and Britski, 2007), popularly known as dourado, is a
83 large piscivorous and highly consumed fish. The species breeds in the rainy season, from
84 October to February, exhibiting total spawning, non-adhesive eggs and high fecundity
85 (Freitas et al. 2013). The goal of the present study was to use histopathological analyses
86 and protein expression of biomarkers of environmental impact to evaluate heavy metal
87 contamination of *S. franciscanus* from two tributaries of the São Francisco River Basin
88 in southeastern Brazil.

89 MATERIALS AND METHODS

90 Sampling

91 Specimens of *S. franciscanus* were sampled from the Abaeté and Paraopeba
92 Rivers during the breeding seasons (October to February) of 2010, 2011 and 2012.
93 Samples from the Paraopeba River were taken at two distinct sites: Site A - downstream
94 of the Igarapé thermoelectric plant (19°57'50.24"S, 44°16'52.42"W); and Site B -
95 downstream of the Retiro Baixo power plant (18°52'28.64"S, 44°46'50.27"W). Samples
96 from the Abaeté River were taken from a single site, considered reference site in present
97 study (18°05'07.8"S, 45°17'00.1"W). All procedures performed during fish collection

98 followed the ethical principles established by the Conselho Nacional de Controle de
99 Experimentação Animal (CONCEA). The study was developed with approval by the
100 Animal Use Ethics Committee of the Pontifical Catholic University of Minas Gerais
101 (CEUA-PUC Minas-protocol 021/2015).

102 A total of 68 adult specimens were used in this study: 36 fish from the Paraopeba
103 River (18 from Site A and 18 from Site B), and 32 from the Abaeté River. All fish were
104 euthanized with a lethal dose of Eugenol (200mg /l). Total length (TL), body weight
105 (BW), weight of gonads (GW) and liver weight (LW) were measured for each specimen.

106 To standardize the analyses, and due to the sexual dimorphism exhibited by this
107 species, only females with total length (TL) between 60 and 87 cm and body weight (BW)
108 between 2250 and 8400g, and males with TL between 58 and 84 cm and BW between
109 2050 and 7980g, were used.

110 **Water quality**

111 Water quality data for the sampled sites were obtained from quarterly reports of
112 Instituto Mineiro de Gestão das Águas (IGAM), which has monitoring points coinciding
113 with the sites sampled in the present study (Tables 1 and 2). The raw data obtained from
114 IGAM are available for public consultation and were used in the present study as water
115 quality parameters for the two rivers (IGAM, 2012B). Legislation established by the
116 Canadian Environment Council (CCME, 2011) was adopted as a reference for
117 permissible levels of heavy metals in water because it is among the most rigorous and
118 current of such legislations.

119 **Biological indices**

120 Gonadosomatic ($GSI = GW / BW \times 100$) and hepatosomatic ($HSI = LW / BW \times$
121 100) indices were calculated to analyze changes in the volume of gonads and liver related
122 to body weight. Only males and females in the advanced maturation stage were used in
123 this study. The Fulton condition factor ($K = BW / TL^3 \times 100$) was also calculated to
124 compare the health status of males and females between the rivers.

125 **Histology and immunohistochemistry**

126 Liver and spleen samples from 21 specimens, seven per sampled river Site, of *S.*
127 *franciscanus* were used for immunohistochemical reactions. The samples were fixed in

128 Bouin solution for 24 h at room temperature, embedded in paraffin, submitted microtomy
129 at 5- μ m thickness, and stained with hematoxylin-eosin and Gomori trichrome.

130 The following primary antibodies were used: anti-MT (Abcam 1:50); anti-HSP70
131 (Sigma 1: 100); and anti-CYP1A (Biosense 1:50). The sections were hydrated and
132 washed for 60 min under running water. Blocking of endogenous peroxidase was
133 performed using 3% hydrogen peroxide (H₂O₂) diluted in TBS.

134 Antigenic recovery occurred in sodium citrate buffer (pH 6.0) for 30 min in a
135 water bath at 60 ° C. Blocking buffer (2% BSA + 0.05% Triton X-100 + 0.01% Tween
136 20) was used for permeabilization and blocking of non-specific reactions. Primary
137 antibodies were applied using their respective dilutions and remained in the humid
138 chamber for 15 h (overnight) at 4 ° C. After washing with TBS, Dako EnVision + Dual
139 Link System HRP, (100 μ l per slide) was applied. The reaction was revealed using 3'-
140 diaminobenzidine (DAB) and the sections were counterstained with hematoxylin for 30
141 s. To ensure success in the procedure, the primary antibodies were not used in the negative
142 controls.

143 **Histopathological assessment**

144 For quantification of histopathological alterations in liver, 10 fish per river site
145 were chosen at random. Six random images of each specimen were analyzed at 200X
146 magnification, for a total of 60 images per river site. Using the grid function with 494
147 points of intersection of Image J software, the following histopathological changes were
148 counted: sinusoidal congestion, sinusoidal dilatation, pigmented macrophages and
149 vacuolization of hepatocytes. In addition, blood vessels and parenchyma were also
150 quantified.

151 For quantification of fibrosis in spleen, 10 fish per river site were chosen at
152 random. Six random images of each specimen were analyzed at 100X magnification, for
153 a total of 60 images per river site. The images were analyzed using the IHC Toolbox
154 function of Image J software. The Image J software dropper tool was used to remove the
155 background and isolate the region to be quantified (connective tissue stained by light
156 green). The image was then transformed into a binary image, and the program read the
157 specific wavelength for the chosen pigment. The results of this analysis were expressed
158 as percentage of marked area.

159

160 **Immunohistochemical assessment**

161 Immunoperoxidase reactions for MT, HSP70 and CYP1A were analyzed using
162 the IHC Toolbox, function of Image J software (Gupta et al. 2007). The software provides
163 a dropper tool function that permits manual identification of the pigment to be analyzed
164 (regions stained by dark brown from positive reactions to DAB). The software function
165 was used to remove the background and isolate the region of interest. Images were then
166 transformed into binary images for quantification. The results are expressed as percentage
167 of marked area out of the total area of the image.

168 **Heavy metal analysis**

169 Concentrations of copper (Cu), lead (Pb) cadmium (Cd), zinc (Zn), chromium (Cr)
170 and iron (Fe) were determined by submitting samples of liver and muscle to acid
171 decomposition by microwave (US-EPA 351-A) StartD (Milestone) with controlled
172 heating. Samples of approximately 1.5 g, in triplicate, were weighed and placed in PTFE-
173 TFM pumps with 20 mL of nitric acid (65% HNO₃) and digested for 25 min at 220°C.
174 After acid digestion, the samples and blank solutions were analyzed in an atomic
175 absorption spectrophotometer with flame and graphite furnace (model iCE 3500 - Thermo
176 Fisher Scientific). To ensure analytical quality of the results and validation of the method,
177 standard fish protein DORM 2 (fish protein with certified reference material for trace
178 metals) was analyzed together with the study material.

179 **Statistical analyses**

180 Statistical analyses were performed using GraphPad Prism software version 5.0
181 for Windows (GraphPad Software, San Diego, California, USA, www.graphpad.com).
182 The results were expressed as mean \pm standard error (S.E.M.) and considered significant
183 in the 95% confidence interval ($p < 0.05$). Since both biological data and metal
184 concentrations were not normally distributed, non-parametric Kruskal-Wallis tests were
185 used followed by Dunn and Mann-Whitney post-tests (Wilcoxon test).

186 **RESULTS**

187 The physico-chemical water quality parameters of the Paraopeba River were
188 similar to those of the Abaeté River with the exception of conductivity, which was
189 significantly higher in the Paraopeba River (Table 1). In addition, metals, including as
190 Pb, Zn, Cr and Fe, were at higher concentrations than the limits allowed by the Canadian

191 Environmental Council (CCME 2011). In contrast, none of the analyzed heavy metals
192 exceeded the established safe limits in the Abaeté River, (Table 2).

193 Regarding biological indices, mean GSI values were higher for males and females
194 of the Abaeté River than for fish from the Paraopeba River ($p < 0.05$). Likewise, HSI was
195 significantly higher for fish of the Paraopeba River than for fish of the Abaeté River (p
196 < 0.05). The Fulton K condition factor was closer to 1.00 for fish of the Abaeté River,
197 differing significantly from the values for Paraopeba fish ($p < 0.05$). No significant
198 differences were observed for HIS and K between sites A and B of the Paraopeba River
199 (Table 3).

200 Histopathological analyses revealed morphological alterations in hepatic and
201 splenic tissues, with higher incidences for fish caught in the two sites of the Paraopeba
202 River. Alterations to the liver included areas of congestion and dilation of sinusoidal
203 capillaries, large numbers of pigmented macrophages associated with blood vessels,
204 infiltration of inflammatory cells in vascularized regions, and hepatocytes with
205 cytoplasmic vacuolization, displacement and nuclear flattening (Fig. 1).

206 Quantification of histopathologies revealed that congestion of the sinusoids varied
207 statistically among the three sampled river sites, with there being a greater amount for
208 fish from site A of the Paraopeba River, while the highest percentage for dilation of
209 sinusoids was observed in fish from PBR B. Pigmented macrophages were not detected
210 in samples from the Abaeté River, but a significant percentage of these cells was observed
211 in the two sites of the Paraopeba River. Cytoplasmic vacuolization in hepatocytes also
212 differed significantly between the Abaeté and Paraopeba Rivers, higher in Paraopeba
213 sites. The occurrence of blood vessels and liver parenchyma were also quantified and had
214 higher percentage in the Abaeté River (Fig. 2).

215 Histological analysis of spleen revealed extensive areas of fibrosis in the
216 trabeculae of the organ, indicating an increase in interstitial stroma and a decrease in the
217 parenchyma in fish from the two sites of the Paraopeba River. In addition, the presence
218 of pigmented macrophages associated with blood vessels and increased connective tissue
219 was observed in fish from Paraopeba sites (Fig. 3). There were more areas of fibrosis in
220 the spleen of fish from the Paraopeba River (Fig. 4), and more in PBR A than PBR B.

221 Molecular analysis of biomarkers also revealed significant differences between
222 the sampled sites of the present study. The expression of MT occurred in granules
223 dispersed in the cytoplasm of hepatocytes and in some cases in close association with
224 blood vessels and near areas of vacuolation in hepatocytes (Fig. 5). The MT levels were

225 higher in site A than in site B of the Paraopeba River, with significant differences in
226 relation to those of Abaeté River (Fig. 6). The markers HSP70 and CYP1A presented
227 different patterns of expression than MT, with more scattered markers in the cytoplasm
228 (Fig. 5). Quantification of HSP70 revealed no significant differences between sites of the
229 Paraopeba River, but significantly higher levels in the Abaeté River ($p < 0.05$) (Fig. 6).
230 As with MT and HSP70, CYP1A had higher levels in the Paraopeba River, with the levels
231 being higher in PBR A presented higher values than PBR B ($P < 0.05$) (Fig. 6). Negative
232 immunoperoxidase controls did not show any protein expression, ensuring the reliability
233 of the technique (Fig. 5).

234 The analysis of heavy metals in fragments of liver and muscle of *S. franciscanus*
235 revealed values that exceeded the safe limits for human consumption established by the
236 World Health Organization (WHO 2002) and the National Health Surveillance Agency
237 (ANVISA 2013) (Table 4). For the Abaeté River, only Cr, in both liver and muscle, was
238 detected at levels above the legal limit. In the Paraopeba River, on the other hand, heavy
239 metals including Cu, Pb, Cd, Cr and Fe had values that exceeded the safe limits allowed
240 by the two agencies. The concentrations of all metals in the liver of fish from the
241 Paraopeba River were at levels above safe limits, and were higher than the levels in
242 muscular tissue, which were still, with the exception of zinc, above the safe limits (Table
243 4).

244 **DISCUSSION**

245 A major contemporary concern worldwide is the correct disposal of pollutants
246 produced on a large scale (Jaishankar et al. 2014; Li and Xie 2016). Untreated effluents
247 released directly into water bodies cause damage to several organisms, especially in fish
248 (Wilhelm et al. 2017). This scenario was apparent in the Paraopeba River, where the
249 results of the present study corroborated other studies conducted in the same River, such
250 as Arantes et al. (2016), Savassi et al. (2016) and Paschoalini et al. (2019), who
251 demonstrated morphophysiological alterations in different species of fish due to heavy
252 metal contamination. However, the Abaeté River, also analyzed in the present work,
253 presents better environmental conditions, with low levels of pollutants (IGAM 2012A;
254 Procópio et al. 2014; Paschoalini et al. 2019). We emphasizing that in the present study
255 innovations on the expression of biomarkers of environmental pollution and other aspects
256 related to histopathology were obtained.

257 Physico-chemical parameters and biometric data did not differ significantly
258 among the river sites sampled in the present study. Standardization of these parameters
259 in comparative toxicological analyses results in more reliable data, since the metabolic
260 capacity and the appearance of histopathologies in fish may be related to water
261 temperature, pH, dissolved oxygen, size and body weight (Canli and Atli 2003; Polat et
262 al. 2015; Sassi et al. 2010). Among the biological indices evaluated, GSI had higher
263 values in the Abaeté River, which is in agreement with recently published studies that
264 showed this river to possess favorable conditions for successful reproductive of different
265 species of teleosts (Arantes et al. 2010; Weber et al. 2013; Nunes et al. 2015; Paschoalini
266 et al. 2019). On the other hand, HSI, unlike GSI, was found to have higher values in
267 environments contaminated by metals, namely sites A and B of the Paraopeba River. This
268 increased HSI may be related to increased organ detoxification activity (Pyle et al. 2005;
269 Querol et al. 2002). The Fulton condition factor tends to be lower in higher quality
270 environments (Fang et al. 2009; Weber et al. 2017), as observed in the present study.

271 Histopathological analyses are widely used to detect physiological changes and to
272 diagnose the health of fish exposed to pollutants in a chronic or acute manner (Camargo
273 and Martinez 2007; Costa et al. 2011; Kaur et al. 2018). Histopathological findings such
274 as those for the liver of fish from site A and B of the Paraopeba River have also been
275 reported for other freshwater species from other environments impacted by heavy metals
276 (Jaishankar et al. 2014; Savassi et al. 2016; Ratn et al. 2018). Sinusoid congestion,
277 dilatation, and infiltration of inflammatory cells in the liver of fish contaminated by heavy
278 metals occurred at significantly higher levels, indicating the negative influence of these
279 elements on the organism as showed by Poleksic et al. (2010). Likewise, high levels of
280 fibrosis like those recorded for the spleens of fish from Paraopeba River have been
281 reported in other environments, where fish are exposed to heavy metals in recent studies,
282 significantly associating heavy metal contamination on the incidence of fibrosis in fish
283 spleen (Handy et al. 2002; Salim et al. 2015).

284 The results of the histopathological analyses and the concentrations of heavy
285 metals in liver and muscle coincide with the results obtained by the immunohistochemical
286 tests, with MT, HSP70 and CYP1A levels being higher in fish from the Paraopeba River.
287 These results demonstrate that histopathological analyses associated with molecular
288 evaluation methods are necessary for accurately diagnosing the class of pollutants acting
289 on organisms, as reported by Rajeshkumar et al. (2017). Studies of the effects of heavy
290 metal contamination on histopathological alterations and protein expression in organism,

291 as performed here, reflect the actual condition of the environmental contamination status
292 and can be applied worldwide, contributing to a better understanding of this
293 environmental problem.

294 Hepatic expression of MT coincided with the levels of heavy metals detected in
295 the fish collected in the two studied rivers. Heavy metal contamination in Paraopeba River
296 was higher in site A than site B, corroborating with MT expression, which was higher in
297 site A, followed by site B and Abaeté River, respectively. The significantly higher levels
298 of MT observed in sites A and B of the Paraopeba River indicate greater exposure of *S.*
299 *franciscanus* to metallic ions and confirm the effectiveness of this biomarker in the
300 detection of metallic pollutants in aquatic environments (Le et al. 2016; Mani et al. 2014).

301 The results for the environmental stress biomarkers reinforce the idea that heavy
302 metals, detected at high concentrations in Paraopeba River, may promote the formation
303 of abnormal proteins, since significant increases in the expression of HSP70 in PBR were
304 detected. Similar results were observed by Rajeshkumar et al. (2013) and Somasundaram
305 et al. (2018), who found heavy metals to be responsible for triggering the activation
306 cascade of this protein. In contrast to HSP70, CYP1A was expressed in a manner similar
307 to MT, with significant differences between the sites of the Paraopeba River. Despite the
308 lack of studies correlating heavy metals and CYP1A in freshwater fish, our results
309 indicate an increase in the expression of this biomarker in environments contaminated by
310 metals. These results corroborate with Anwar-Mohamed et al. (2009), who showed that
311 heavy metals can regulate the expression of CYP1A in different aryl-hydrocarbon
312 receptor (AHR) signaling pathways in a metal-dependent manner.

313 The analysis of heavy metals in hepatic and muscular tissues revealed higher
314 levels of contamination of the liver than of the muscle, which is expected since organs of
315 intense metabolism tend to exhibit higher levels of contamination (Eneji et al. 2011;
316 Taweel et al. 2013). In addition, levels of heavy metals that exceed the limits allowed for
317 food by the World Health Organization (WHO 2002), have been previously recorded in
318 the muscle tissue of fish from the Paraopeba River (Arantes et al. 2016; Paschoalini et al.
319 2019), which represents a potential danger to the health of any population that consumes
320 *S. franciscanus* extensively.

321 The Paraopeba River was classified as a priority conservation area due to its high
322 diversity of native fish species (Drummond et al. 2005). Nonetheless, the data of the
323 present study, along with those of other studies, reveal that the toxic condition of this
324 river negatively influences the biology of different species of fish (Arantes et al. 2016;

325 Savassi et al. 2016; Paschoalini et al. 2019). Additionally, on January 24, of 2019, a
326 mining tailings dam on the Paraopeba River broke, releasing a massive load of heavy
327 metals into the river, which will certainly contribute to increasing the toxic condition of
328 the river with negative impacts for the conservation and maintenance of its ichthyofauna.
329 In summary, the results of the present study are essential for the continuity of research in
330 aquatic ecosystems contaminated by heavy metals, highlighting the importance of studies
331 about the use of piscivorous fish as an experimental model to evaluate the levels of heavy
332 metal contamination and its effects on the organism, since the natural environment is
333 increasingly contaminated by these inorganic pollutants.

334 **CONCLUSIONS**

335 The present study indicates that the bioaccumulation of heavy metals may be
336 related to the higher occurrence of histopathological alterations in the liver and spleen of
337 fish. These changes, associated with significant increase expression of MT, HSP70 and
338 CYP1A in fish liver, suggest that these biomarkers can be important in the assessment of
339 environmental quality in ecosystems contaminated by heavy metals. The study provides
340 important and current information of the action of heavy metals on the organism,
341 especially in the use of environmental impact biomarkers associated with histological
342 tools. In summary, the data obtained in the present study show that heavy metals may
343 negatively influence the biology of *Salminus franciscanus* and human health through the
344 consumption of contaminated fish.

345 **ACKNOWLEDGMENTS**

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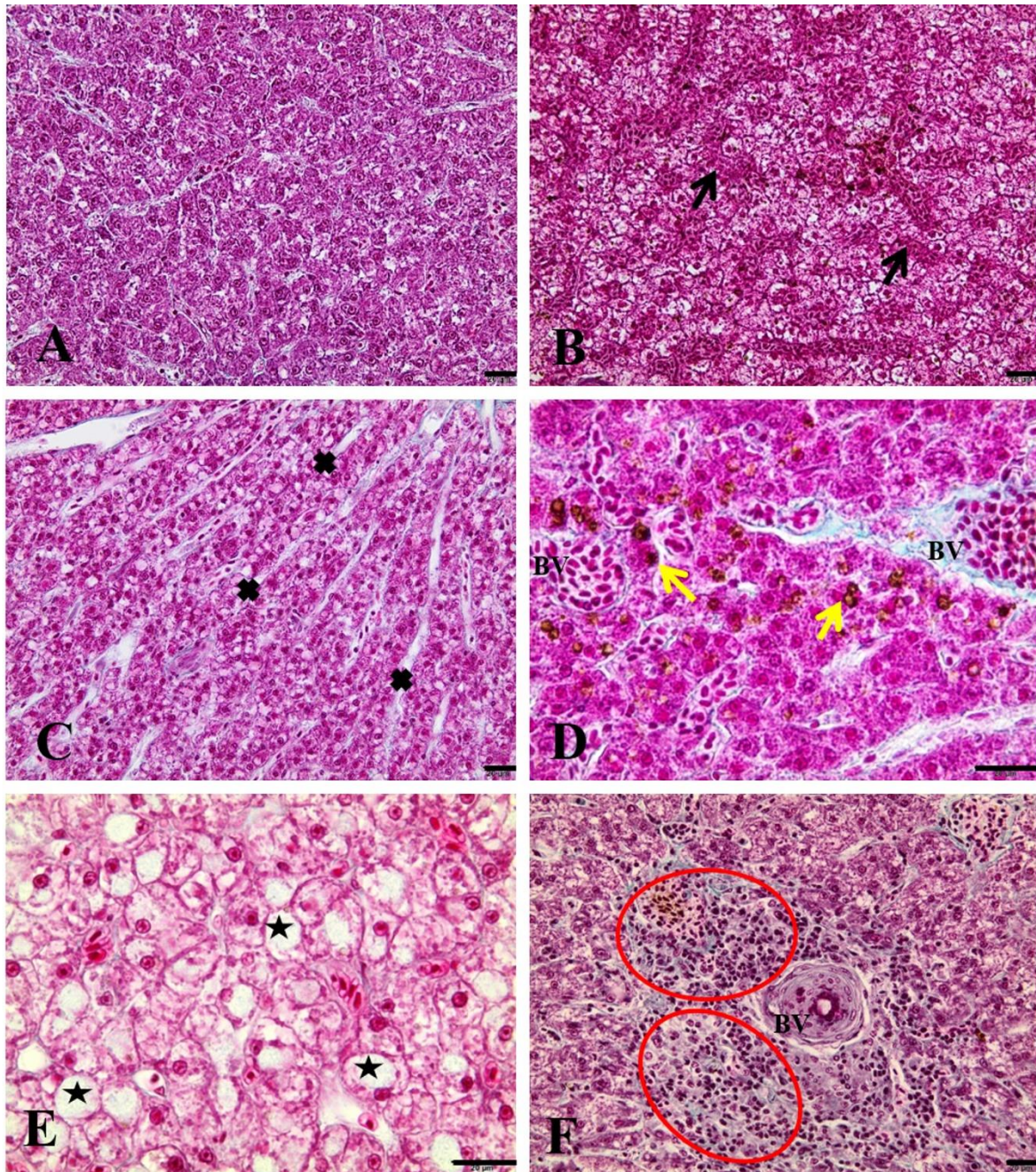
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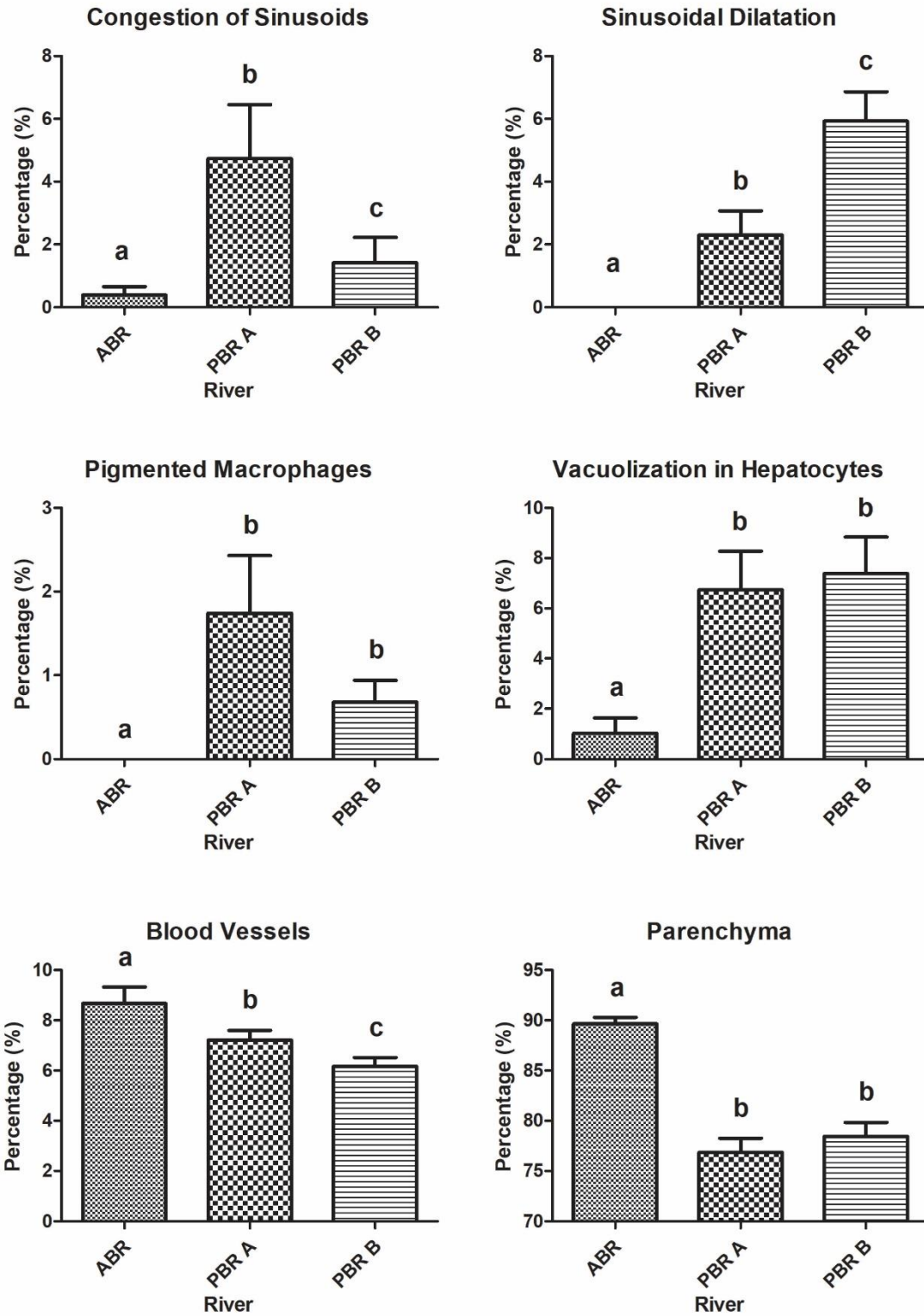
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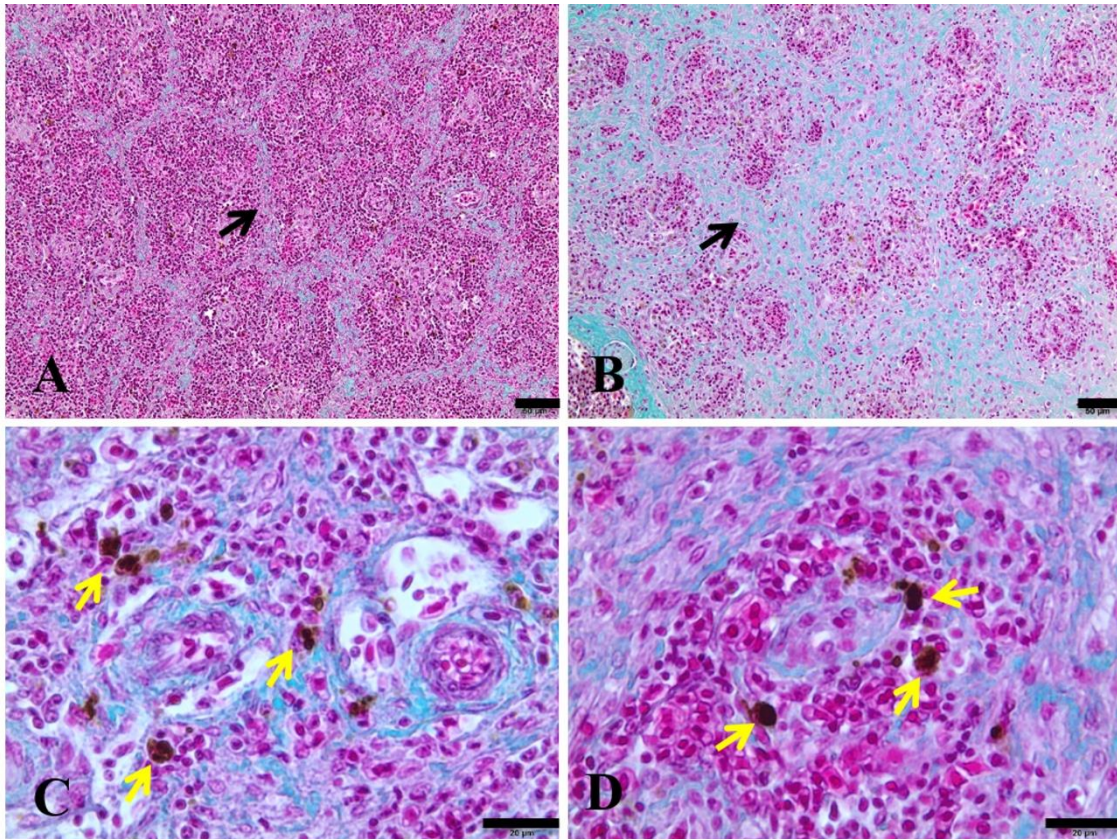
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595 **Figure 1.** Histopathological assessment of the liver of *S. franciscanus* from Abaeté and
 596 Paraopeba rivers, São Francisco River Basin, Brazil. (A) Liver tissue showing normal
 597 morphology of a fish from the Abaeté River (200x). B, C, D, E and F represent
 598 histopathological alterations in fish from the Paraopeba River: (B) congestion of the sinusoid
 599 capillaries (black arrow) (200x); (C) dilatation of the sinusoid capillaries (black cross)
 600 (D) accumulation of pigmented macrophages (yellow arrow), aggregated among blood vessels
 601 (bv) (400x); (E) cytoplasmic vacuolization with nuclear displacement (black star) (400x); (F)
 602 infiltration of immune cells (red circle) next to blood vessels (bv) (100x). Scale bar = 20µm.



603

604 **Figure 2.** Quantification of histopathological alterations detected in liver of *S.*
 605 *francicanus* captured in the Abaeté (ABR) and Paraopeba (PBR A and PBR B) rivers,
 606 São Francisco River basin, Brazil. Different letters indicate statistical differences between
 607 the sampled sites ($p < 0.05$; Kruskal-Wallis test).



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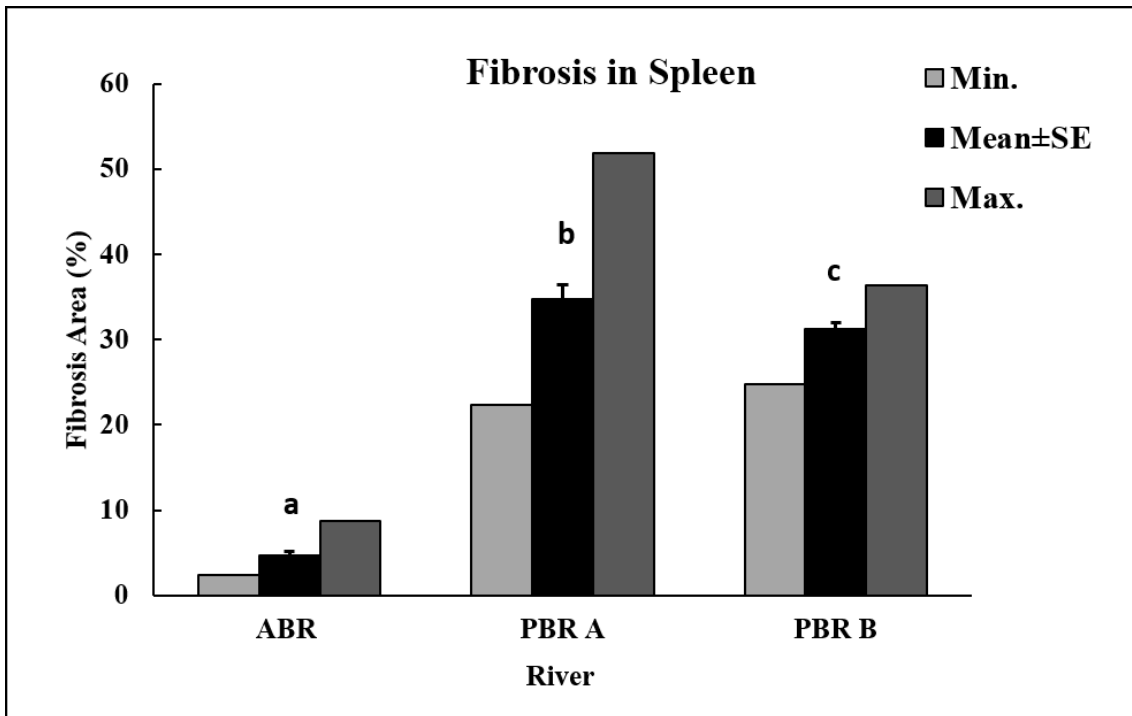
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Figure 3. Histopathological assessment of the spleen of *S. franciscanus* from the Abaete and Paraopeba rivers, São Francisco River basin, Brazil. (A) Normal aspect of splenic tissue. Normal distribution between the stromal area (stained in green, identified by the black arrow) and the parenchymal area (stained in pink / purple) for fish from the Abaeté River (100x), scale bar = 50µm. B, C, and D represent alterations in the spleen of fish from Paraopeba River: (B) excessive connective tissue production, denominated fibrosis (100x), scale bar = 50µm; (C and D) accumulation of pigmented macrophages, distributed in parenchymal area (yellow arrow), scale bar = 20µm (400x).



629

630 **Figure 4.** Quantification of fibrosis area in the spleen of *S. franciscanus* collected in the
 631 Abaeté (ABR) and Paraopeba (sites PBR A and PBR B) rivers. Different letters indicate
 632 statistical differences between sampled sites ($p < 0.05$; Kruskal-Wallis test).

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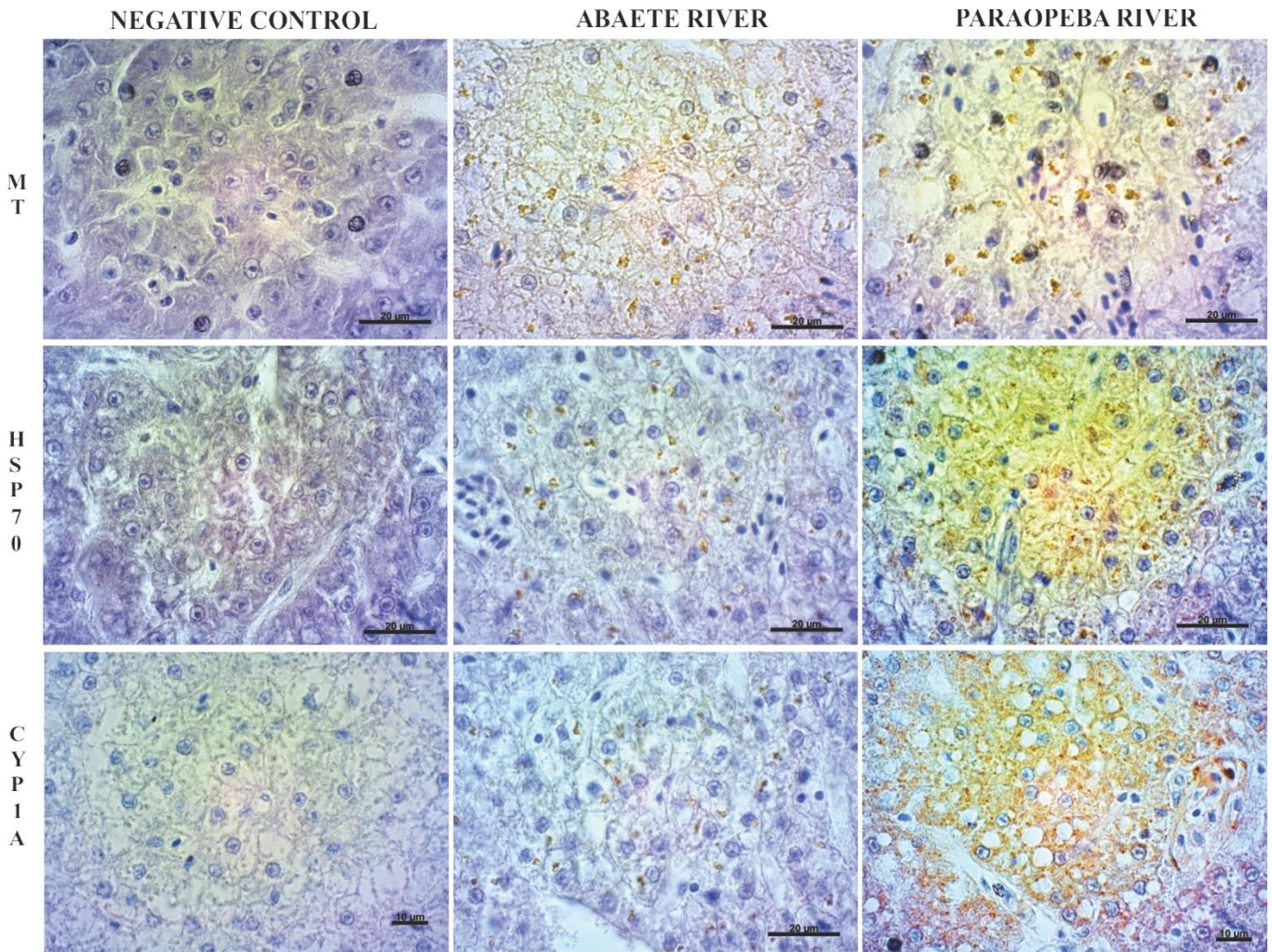
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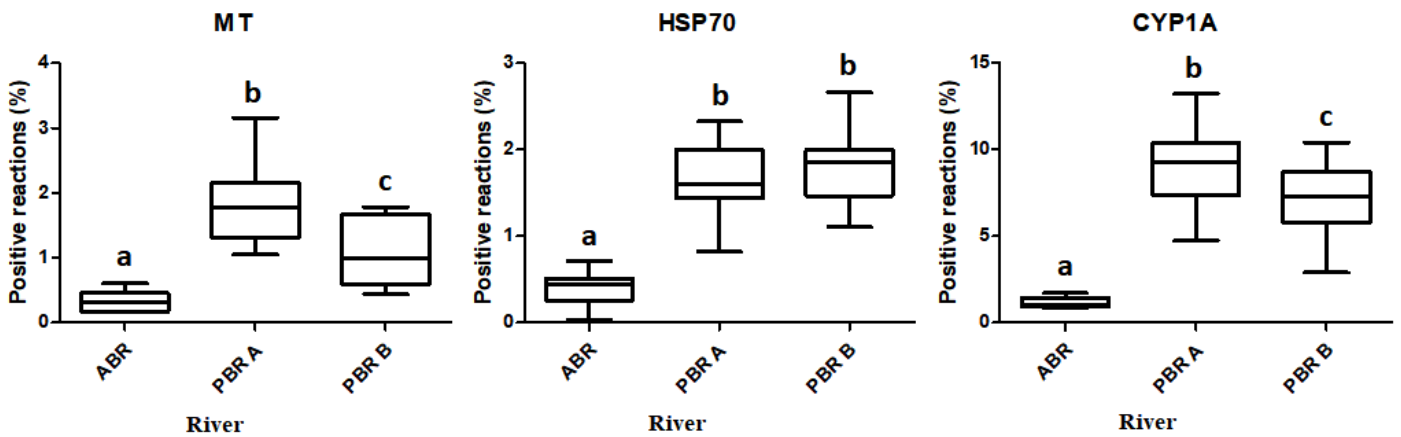
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654 **Figure 5.** Immunohistochemical reactions of biomarkers in liver of *S. franciscanus*, from
 655 Abaeté and Paraopeba rivers: metallothionein (MT), heat shock protein 70 (HSP70) and
 656 cytochrome P4501A (CYP1A).

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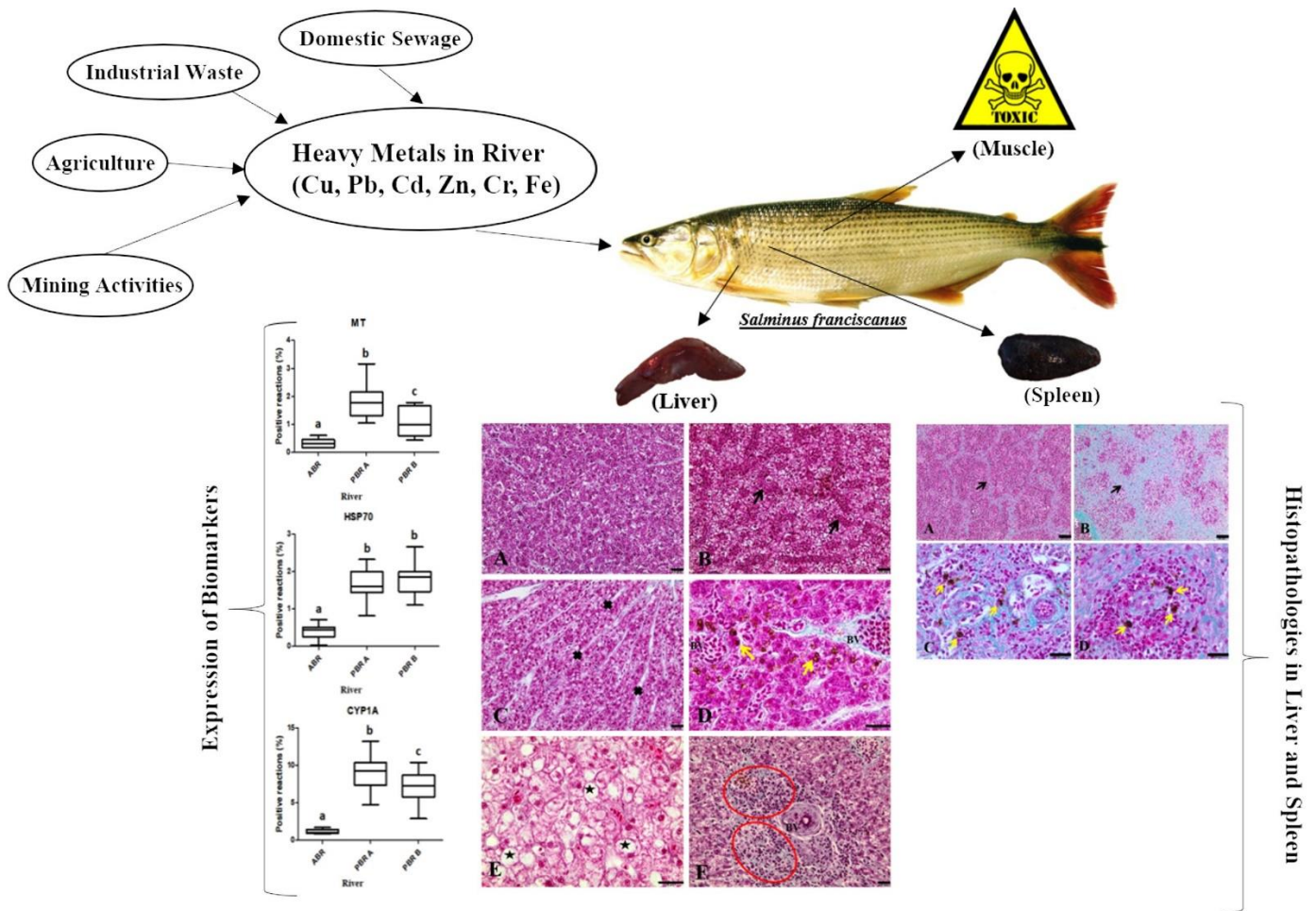


673 **Figure 6.** Quantification of positive reactions to immunohistochemistry peroxidase for
 674 metallothionein (MT), heat shock protein 70 (HSP70) and cytochrome P4501A (CYP1A)
 675 in liver of *S. franciscanus* from the Abaeté (ABR) and Paraopeba (sites PBR A and PBR
 676 B) rivers. Different letters indicate statistical differences between sampled sites ($p < 0.05$;
 677 Kruskal-Wallis test).

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682 **Graphical Abstract:** Heavy metals contamination, from diverse polluting sources, are
 683 bioaccumulated by fish and cause severe histopathologies, in target organs, including
 684 differences on expression of immunohistochemical environmental biomarkers, which
 685 will protect the organism against injury resulting from this environmental chemical
 686 pollution.

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698 **Tables**

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700 **Table 1** - Physico-chemical parameters of the water of Abaeté and Paraopeba Rivers
 701 (Sites ABR, PBR A and PBR B), São Francisco River Basin, Brazil, for the years 2010,
 702 2011 and 2012.

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Sites	Oxygen	Temperature	pH	Conductivity
ABR	7.07 ± 1.07 ^A	26.34 ± 0.47 ^A	7.02 ± 0.14 ^A	64.67 ± 2.54 ^A
PBR A	7.25 ± 0.3 ^A	24.64 ± 1.03 ^A	6.77 ± 0.14 ^A	84.12 ± 6.07 ^B
PBR B	7.13 ± 0.25 ^A	25.09 ± 0.78 ^A	6.89 ± 0.16 ^A	77.67 ± 4.86 ^B

Data from Minas Gerais Institute of Water Management (IGAM, 2012^B). Different letters indicate statistical differences between the sampling sections (p<0.05).

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708 **Table 2** – Heavy metal contamination of the water of Abaeté and Paraopeba Rivers (Sites
 709 ABR, PBR A and PBR B), São Francisco River basin, Brazil, for the years 2010, 2011
 710 and 2012.

Sites	Heavy Metals (mg/ml)					
	Cu	Pb	Cd	Zn	Cr	Fe
ABR	nd	nd	nd	0.005 ± 0.001 ^a	nd	0.1 ± 0.05 ^a
PBR A	nd	0.002 ± 0.001 ^a	nd	0.07 ± 0.01 ^b	0.04 ± 0.01 ^a	1.32 ± 0.05 ^b
PBR B	nd	0.002 ± 0.001 ^a	0.0005	0.02 ± 0.01 ^b	0.04 ± 0.01 ^a	0.8 ± 0.05 ^c
Water limits						
CCME (2011)	0.002	0.001	0.00009	0.03	0.001	0.3

Data obtained by IGAM in October to February 2010, 2011 and 2012. Values are expressed as mean±S.E Different letters indicate statistical differences between the sampling sections (p < 0.05). nd = not detected.

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720 **Table 3** - Biometric data and biological indices for male and female fish of *Salminus*
 721 *franciscanus* from Abaeté and Paraopeba Rivers (Sites ABR, PBR A and PBR B), São
 722 Francisco River Basin, Brazil.

Sex	Sites	TL(cm)	BW(g)	GSI(%)	HSI(%)	K
Males	ABR	61.5 ± 13.9 ^a	2822.4 ± 1550.7 ^a	1.27 ± 0.76 ^a	0.64 ± 0.77 ^a	1.06 ± 0.07 ^a
	PBR A	65.0 ± 6.6 ^a	3316.6 ± 1224.2 ^a	1.01 ± 0.58 ^a	0.77 ± 0.15 ^b	1.18 ± 0.10 ^b
	PBR B	63.0 ± 8.3 ^a	2975.1 ± 1203.4 ^a	0.85 ± 0.15 ^b	0.70 ± 0.07 ^b	1.27 ± 0.11 ^b
Females	ABR	77.5 ± 5.7 ^a	5744.9 ± 1270.0 ^a	5.07 ± 1.85 ^a	0.77 ± 0.15 ^a	1.24 ± 0.11 ^a
	PBR A	79.0 ± 13.0 ^a	7353.3 ± 1603.0 ^a	3.03 ± 0.93 ^b	0.90 ± 0.18 ^b	1.47 ± 0.17 ^b
	PBR B	73.06 ± 3.7 ^a	5253.8 ± 848.9 ^a	2.52 ± 0.11 ^b	0.84 ± 0.12 ^b	1.41 ± 0.07 ^b

723 Different letters indicate statistical differences between the sampling sections (p<0.05).

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728 **Table 4** - Concentrations of heavy metals in liver and muscle of *Salminus franciscanus*
 729 from Abaeté and Paraopeba Rivers (Sites ABR, PBR A and PBR B), São Francisco River
 730 Basin, Brazil.

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	Heavy metals (mg/kg)					
	Cu	Pb	Cd	Zn	Cr	Fe
Abaeté River						
Liver	19.12 ± 4.52 ^a	0.18 ± 0.3 ^a	0.07 ± 0.06 ^a	19.54 ± 4.31 ^a	0.65 ± 0.15 ^a	90.92 ± 14.53 ^a
Muscle	5.45 ± 0.41 ^a	0.12 ± 0.08 ^a	0.02 ± 0.01 ^a	8.88 ± 1.55 ^a	0.56 ± 0.12 ^a	7.13 ± 1.47 ^a
Paraopeba River - site A						
Liver	108.58 ± 22.42 ^b	2.28 ± 0.21 ^b	0.43 ± 0.05 ^b	173.87 ± 35.54 ^b	1.77 ± 0.08 ^b	1639.24 ± 502.09 ^b
Muscle	32.20 ± 0.21 ^b	2.24 ± 0.24 ^b	0.37 ± 0.02 ^b	13.40 ± 2.39 ^b	1.73 ± 0.10 ^b	210.15 ± 41.27 ^b
Paraopeba River - site B						
Liver	71.49 ± 15.70 ^c	1.42 ± 0.11 ^b	0.33 ± 0.22 ^b	160.48 ± 30.07 ^b	0.48 ± 0.23 ^a	1028.23 ± 388.59 ^c
Muscle	18.40 ± 5.66 ^c	0.73 ± 0.07 ^c	0.25 ± 0.03 ^b	27.84 ± 9.54 ^c	0.24 ± 0.04 ^a	186.68 ± 32.68 ^b
Tissue limits (mg/kg)						
OMS (2002)	30.0	0.2	0.05	nf	nf	109.0
ANVISA (2013)	30.0	0.3	0.05	50.0	0.1	nf

733 Data are expressed as mean ± SEM. Different letters indicate statistical differences between the sampling sections

734 (p<0.05) / nf = not found. Copper (Cu), Lead (Pb), Cadmium (Cd), Zinc (Zn), Chromium (Cr) and Iron (Fe).

5. CONSIDERAÇÕES FINAIS

O presente estudo fornece informações suficientes para atestar que metais pesados foram detectados em concentrações elevadas na musculatura dos peixes do rio Paraopeba (Cu, Pb, Cd, Cr), excedendo os limites seguros de contaminantes em alimentos, estabelecido pela Agência Nacional de Vigilância Sanitária (ANVISA, 2013). Em órgãos de intenso metabolismo, tal como fígado, essa concentração de metais pesados (Cu, Pb, Cd, Zn, Cr e Fe, todos os metais analisados) foram registradas em valores muito acima dos permitidos pela ANVISA também, indicando o papel do fígado em bioacumular e metabolizar esses elementos, assim justificando a importância de estudos toxicológicos e seus efeitos sobre o organismo. A bioacumulação de metais pesados no *Salminus franciscanus* está relacionada à maior ocorrência de alterações histopatológicas no fígado e baço dos peixes, expostos a esta classe de contaminantes. A expressão de proteínas biomarcadoras de impacto ambiental tais como Metalotioneínas (MT), Proteínas do Choque Térmico (HSP70) e Citocromo P450 (CYP1A) se mostraram eficazes na avaliação de ambientes contaminados por metais pesados, e quando utilizadas em conjunto à outras ferramentas de avaliação ambiental apresentam uma maior confiabilidade e eficiência na avaliação dos impactos ambientais. As alterações histopatológicas, associadas às elevações significativas na expressão de MT, HSP70 e CYP1A no fígado dos peixes, sugerem o importante papel da utilização destes biomarcadores na avaliação da qualidade ambiental em ecossistemas contaminados por metais pesados, embora os estudos sobre o tema ainda sejam escassos. Em suma, os dados obtidos no presente estudo, evidenciam que metais pesados podem influenciar negativamente a biologia de *Salminus franciscanus* e à saúde humana, através do consumo de peixes contaminados pela população.

6. ANEXO I

- Participação na coautoria do seguinte trabalho recentemente publicado (Paschoalini et al., 2019):

Ecotoxicology and Environmental Safety 169 (2019) 539–550



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Heavy metals accumulation and endocrine disruption in *Prochilodus argenteus* from a polluted neotropical river



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ABSTRACT

Heavy metals are considered major pollutants of aquatic environments due to the difficulty of metabolization and the bioaccumulative potential in tissues of aquatic organisms, especially fish muscle that is often used as food worldwide. In addition to causing cell damage, some metals such as aluminium (Al), cadmium (Cd), copper (Cu), and lead (Pb) can act as endocrine disrupting chemicals in fish. The Paraopeba and Abaete Rivers are important tributaries of the upper São Francisco River basin, but the Paraopeba River receives, along its course, the discharge of many types of effluents that affect fish species, including widely consumed species such as *Prochilodus argenteus*. This study evaluated histological and molecular changes caused by chronic exposure to heavy metals in *P. argenteus* from the Paraopeba River and compared this to fish from the non-impacted Abaete River. Sampled fish from both rivers were used in histological analyses and immunohistochemical assays. The results showed increased incidence of histopathologies and changes in number and morphology of germline cells in both sexes. In addition, up-regulated expression of oestrogens-induced proteins in the liver of males were detected in polluted environment. All the alterations were related to the concentration of metals in water and fish. The high concentration of various metals observed in water and fish from Paraopeba River serves as an alert to the environmental and public health regulatory authorities.

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