

Ultrastructure of feathered triancre in the Thaumastodermatidae and the description of a new species of *Tetranchyroderma* (Gastrotricha: Macrodasyida) from Australia

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Tetranchyroderma adeleae sp. nov. is described from North Stradbroke Island, eastern Australia, where it is present in low abundance in the clean, coarse sediments of Cylinder beach. The species is characterized by the presence of epidermal glands on the oral hood and three pairs of dorsolateral cirrata. This is also the first described species of *Tetranchyroderma* with mixed ancre types that consist of both ordinary triancre and feathered triancre. Ordinary triancre are present on the ventrolateral margins of the body while feathered triancre adorn the dorsal side. Details of the ultrastructure of the cuticle in *T. adeleae* sp. nov. and a species of *Pseudostomella* are compared to each other and the cuticle of additional species of Thaumastodermatidae. At the ultrastructural level, feathered triancre are composed of a thickened endocuticular base with three tines (shafts) that give rise to pointed, feather-shaped scales. The structure of the base, tines and scales reveals a finely granular construction without noticeable substructure. The ultrastructure of feathered triancre is similar to that of ordinary triancre and broadly similar to feathered triancre in other species. However, differences are noted in the contour of the feathered scales that are not detected with light microscopy. Future studies of the thaumastodermatid cuticle using scanning electron microscopy are warranted to provide enhanced three-dimensional resolution and insight into their evolution.

Keywords: ultrastructure, feathered triancre, Thaumastodermatidae, new species, *Tetranchyroderma*, Australia

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INTRODUCTION

Marine gastrotrichs are microscopic, worm-like invertebrates that inhabit the interstices of littoral and subtidal sediments in all oceans and seas. While they are rarely the most abundant form of meiofauna (but see Coull, 1985; Hochberg, 1999), their morphological diversity can be overwhelming as a result of the various and often exquisite elaborations of their cuticle. The Thaumastodermatidae is the most speciose family of macrodasyidan gastrotrichs and one that shows the largest diversity in cuticular morphology and ultrastructure (see Ruppert, 1988). The construction of the cuticle is highly elaborate, resulting from modifications of both endo- and exocuticle, and forming structures such as flat and bowl-shaped scales, spines, and multi-pronged hooks called ancre (Rieger & Rieger, 1977). The shape of the cuticular structures may have strong phylogenetic signal, and some taxa within the Thaumastodermatidae can be grouped based on cuticular sculpture (Rieger & Rieger, 1977; Hochberg & Litvaitis, 2001). The diversity of cuticular ornamentation (spines, scales, triancre, tetrancres and pentancre) is well-characterized from over 100 species distributed among five genera. Ruppert (1970) added a unique form of sculpture to

our knowledge of cuticular diversity when he described *Pseudostomella plumosa* Ruppert, 1970 from North Carolina, USA. The dorsal surface of *P. plumosa* is composed of highly modified triancre that form feather-like scales (feathered triancre). Since then, three species of *Pseudostomella* have been described with a similar cuticular covering; *P. klauserae* and *P. megapalpator* from Australia (Hochberg, 2002b) and *P. faroensis* from the Faroe Bank (Clausen, 2004). No comparable cuticular structures have been described from other species in the family, and to date, nothing is known about their ultrastructure.

Research on marine interstitial gastrotrichs of Australia has yielded several new species in the past few years (Hochberg, 2002b, c, 2003; Nicholas & Todaro, 2005, 2006). Among them is a species of *Tetranchyroderma* with a cuticular sculpture similar to the feathered triancre of species of *Pseudostomella*. Here, I describe this new species of *Tetranchyroderma* and provide an ultrastructural account of the cuticle. These observations are compared to the ultrastructure of feathered triancre in *P. megapalpator* to arrive at a better understanding of the fine structural differences in their construction.

MATERIALS AND METHODS

Sediment samples from the littoral zone of Cylinder beach, North Stradbroke Island (27° 26'S 153° 32'E) were collected

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in July and August, 2001. Four sediment samples (100 cm³ each) were taken from the upper 10 cm of sand in the low and middle tide region. Fauna was extracted using the anaesthetization/decantation technique with 7.5% MgCl₂ (Pfannkuche & Thiel, 1988). Gastrotrichs were analysed with DIC/Nomarski optics on an Olympus BH2 microscope equipped with ocular micrometer and a Sanyo HiRes Color CCD camera. Approximately 12 specimens of a new species of *Tetranchyroderma* were measured and photographed. The positions of organs are expressed in reference to percentage body units (total body length = 100 units (U)). Two specimens were fixed in 5% formalin, mounted whole in de Faures fluid, and deposited as types in the Queensland Museum.

Specimens of *Tetranchyroderma* and additional gastrotrichs were prepared for electron microscopy. Specimens of *Pseudostomella megapalpatator* Hochberg, 2002 were collected from Frenchman's beach, Australia (27° 01'S 153° 35'E) as previously described (Hochberg, 2002b). *Tetranchyroderma* cf. *polypodium* Luporini, Magagnini & Tongiorgi, 1971 was collected from Green Turtle beach, South Hutchinson Island, Fort Pierce, FL, USA (27° 25.177'N; 80° 16.368'W) in September 2004. All gastrotrichs were prepared for transmission electron microscopy (TEM) after relaxation in 7.5% MgCl₂ for 20 minutes. Specimens were fixed in 3% glutaraldehyde in 0.1 M cacodylate buffer (pH 7.2) for 24 hours, followed by three buffer rinses (15 minutes each) and post fixation in 1.5% OsO₄ in 0.1 M cacodylate buffer for 1 hour. After three more buffer rinses (15 minutes each), gastrotrichs were dehydrated in a graded ethanol series, transferred to propylene oxide and embedded in an Araldite/Epon mixture. Resin blocks for TEM were sectioned at 60–70 nm with a Diatome diamond knife on a Reichert or Sorvall ultramicrotome, collected on coated grids, and stained with uranyl acetate and lead citrate. Grids were examined with a JEOL 1010 TEM at 60 kV at the University of Queensland's Centre for Microscopy and Microanalysis and with a 100 CX II TEM at 60 kV at the Smithsonian Marine Station in Fort Pierce, Florida. One specimen of *T. cf. polypodium* was examined with scanning electron microscopy (SEM). After dehydration as in the above protocol, the specimen was dried in a critical point dryer, sputter coated with gold, and examined on a JEOL 6400 SEM at 10 kV at the Smithsonian Marine Station.

The descriptive terminology of Todaro (2002) is adopted. Abbreviations used in the text are the following: ci, cilium; co, caudal organ; CrD, dorsal cirrata; en, endocuticle; eg, epidermal gland; ep, epidermis; ex, exocuticle; fo, frontal organ; mr, midrib of scale; mi, mitochondrion; mu, muscle; mv, microvilli; ph, pharynx; sb, scale base; sc, feathered scale; TbA, anterior adhesive tubes; TbD, dorsal adhesive tubes; TbL, lateral adhesive tubes; TbP, posterior adhesive tubes; tc, tactile cilia; te, testis; ti, tine.

RESULTS

SYSTEMATICS

Phylum GASTROTRICHA

Order MACRODASYIDA Remane, 1925 [Rao & Clausen, 1970]

Family THAUMASTODERMATIDAE Remane, 1929

Genus *Tetranchyroderma* Remane, 1926

Tetranchyroderma adeleae new species
(Figures 1–3)

TYPE MATERIAL

Holotype: an adult specimen 448 μm long, formalin-de Faure's wholemount, collected from the low tide region of Cylinder beach, North Stradbroke Island, Queensland, Australia (27° 01'S 148° 35'E) on July 10, 2001. Repository: Queensland Museum (G218766).

Paratype: an adult 333 μm long, wholemount, same data as holotype. Repository: Queensland Museum (G218772).

Other material: twelve specimens studied alive and measured. One slightly damaged specimen prepared for TEM.

ETYMOLOGY

The species is dedicated to my wife.

ECOLOGY AND DISTRIBUTION

Tetranchyroderma adeleae sp. nov. has only been recorded from the type locality. Cylinder beach is located on the north-east side of North Stradbroke Island, Australia. It is a long stretch of sand beach open to high-energy swells from the north-east. The beach consists of well-oxygenated and clean, medium-grained sand and containing some shell fragments. Sympatric interstitial fauna included: Amphipoda and Ostracoda (Crustacea), *Diurodrilus* sp. and *Protodrillus* sp. (Annelida), Nematoda, *Neodasys* sp., *Paraturbanella* sp., *Tetranchyroderma* sp., *Turbanella* sp., and *Xenotrichula* sp. (Gastrotricha), and several species of Kalyptorhynchia and Proseriata (Platyhelminthes) and Acoela.

DIAGNOSIS

Tetranchyroderma with body up to 448 μm long and 76 μm wide. Head with fringed oral hood bearing four epidermal glands. Mouth to 35 μm wide, its margin surrounded by small papillae. Pharynx to 104 μm long and 38 μm wide. Four pairs of epidermal glands from mid-pharynx to caudal end along lateral margin of body. Four TbA per side, 9–10 μm long; one pair of tubes at midline. Fourteen to sixteen TbL per side, 6–9 μm long. Three pairs of CrD (dorsal cirrata) to 32 μm long. Three TbP per side, 5–6 μm long. Cuticular sculpture consists of ordinary triancre and feathered triancre, extending from head to caudal end. Tactile cilia present sparsely at the mouth margin and along lateral margin of body. Transverse rows of locomotory cilia cover most of the ventral surface. Single testis on right side with vas deferens connecting to caudal organ. Frontal organ present.

DESCRIPTION

General appearance: the description is based on an adult specimen with a body length of 448 μm (Figures 1–3). The body is highly opaque and somewhat reflective under transmitted light due to the cuticle. The oral hood is up to 48 μm long and bears small papillae with cilia on its anterior margin (Figure 1A). Four elongate epidermal glands, 6–7 μm long, are present at the anterior portion of the hood (Figure 1A). Ventrally, the mouth margin bears few small papillae and is lined with short tactile cilia (2–3 μm; Figure 1B). In the trunk region, four pairs of epidermal glands are present dorsolaterally beginning at ~U24 and extending to U85. All glands are slightly elongate, asymmetrical in shape, and up to 11 μm long. Tactile cilia are present on the lateral margin of the body beginning at the base of the oral hood.

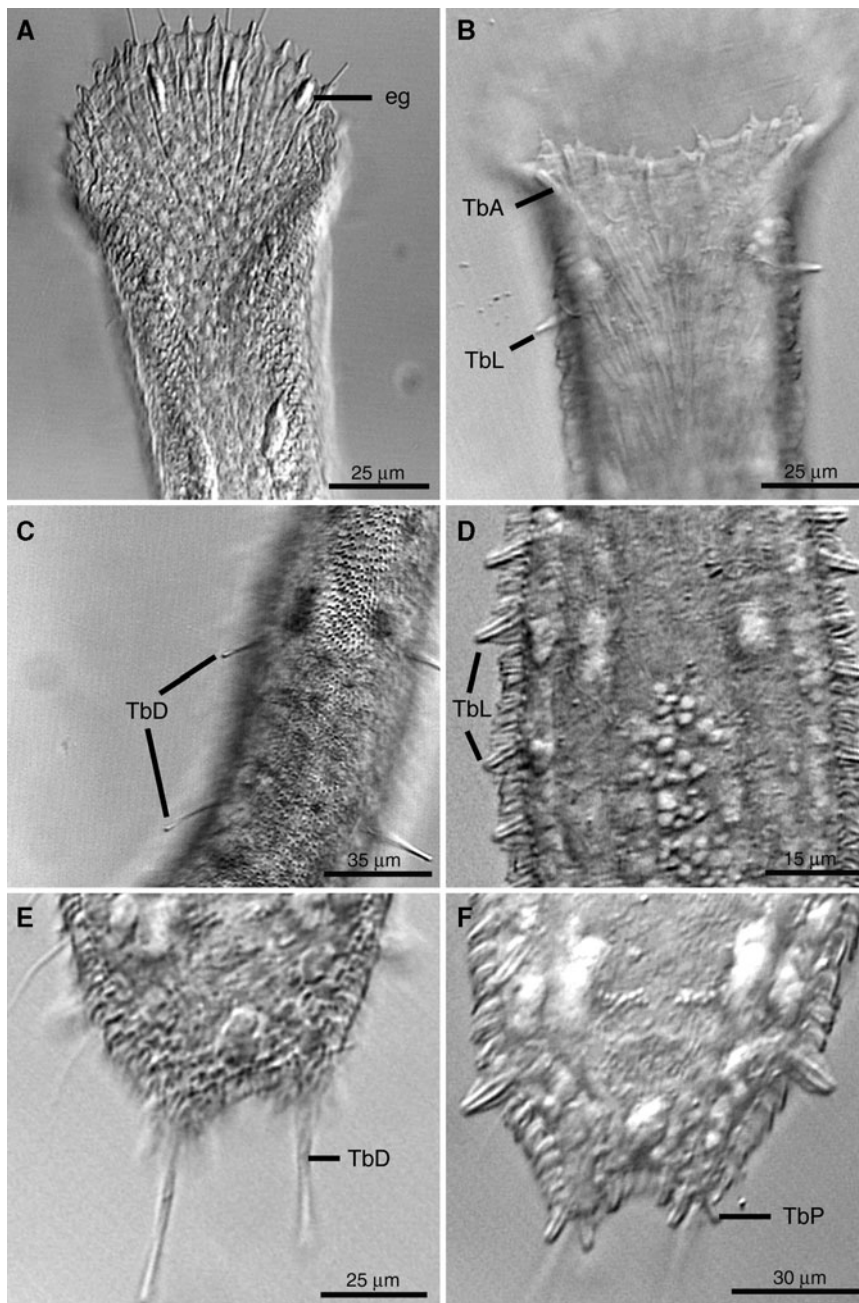


Fig. 1. Light micrographs of *Tetranchyroderma adeleae* sp. nov. Holotype: (A) dorsal view of anterior end; (B) ventral view of anterior end; (C) dorsal view of trunk; (D) ventral view of trunk; (E) dorsal view of caudal end; (F) ventral view of caudal end TbD mislabelled (=CrD).

Digestive tract: the digestive tract begins with a large terminal mouth to 35 μm wide. The pharynx, from the ventral mouth margin to the pharyngointestinal junction, is 104 μm long and 18 μm wide. Pharyngeal pores are present at U₃₂ and the pharyngointestinal junction is present at U₃₄. The intestine is up to 24 μm wide and glandular in appearance. A ventral anus is present at U₈₈.

Cuticle: two types of cuticular structures are present (Figures 2 & 3). Ordinary triancres with a slight posterior curvature are present on the ventrolateral margins of the body (Figures 2C, D & 3E). Ordinary triancres consist of three tines that arise from a common base. Each tine is of equal length, with shorter tines $\sim 3 \mu\text{m}$ long close to the lateral margins of the oral hood, and longer tines $\sim 5 \mu\text{m}$ long on

the lateral margins of the main trunk. All tines display a caudal bend but do not broaden into feather-shaped scales (Figures 2C & 3E). Ordinary triancres close to the caudal lobes have a more pronounced caudal bend than those at the anterior end of the body (Figure 2D). Feathered triancres are present on the dorsal surface from the middle of the oral hood to the caudal end (Figure 1A, B). Feathered triancres consist of three caudally-directed tines that arise from a common base (Figures 1A, B & 3C, D). On the trunk, each tine is made of a central shaft that rises vertically for $\sim 3 \mu\text{m}$ and immediately broadens as it bends caudally to form a feathered scale $\sim 5 \mu\text{m}$ long (Figure 3D). A midrib is apparent down the centre of each feathered scale (Figure 3D). The length of feathered scales is noticeably shorter at the anterior

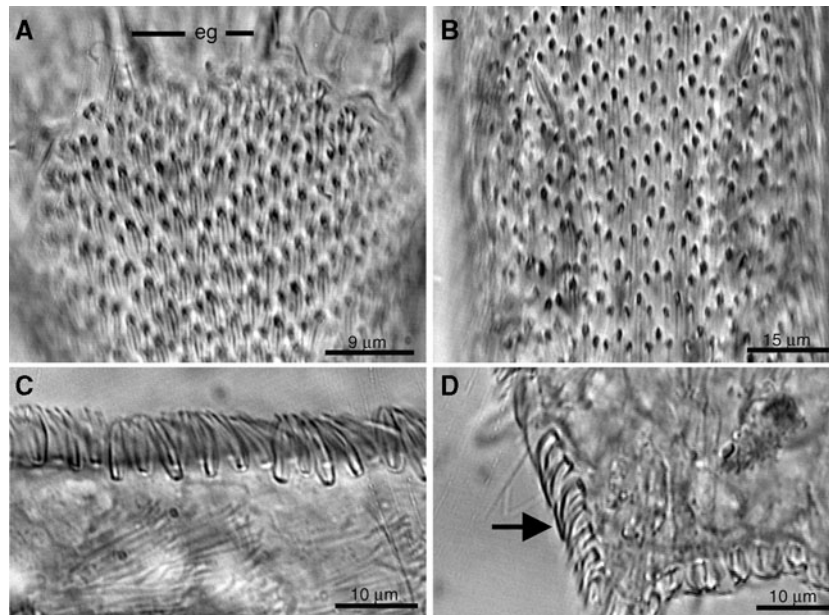


Fig. 2. DIC micrograph of the cuticle of the paratype of *Tetranchyroderma adeleae* sp. nov. (A) Dorsal portion of oral hood; (B) dorsal view of trunk; (C) view of the ventrolateral body wall; (D) view of the ventrolateral body wall at the posterior end of the trunk. Arrow points to ordinary triancre with significant caudal bend.

end of the body (2–3 μm). There are approximately sixteen longitudinal columns of feathered triancre on the dorsal trunk.

Adhesive organs: four anterior adhesive tubes (TbA) per side are present ventrally at the lateral mouth margin (Figures 1B & 3A). All tubes are duogland in appearance and of equal length, $\sim 8 \mu\text{m}$. Two tube-like structures to 7 μm long are present at the midline of the mouth margin, the others being placed laterally. Sixteen lateral adhesive tubes (TbL) are slightly sublateral in position. The first

pair of tubes, at U22, does not appear to be duogland (Figure 1B). All other lateral adhesive tubes are duogland and begin at U33 (Figures 1D & 3A). The last pair of adhesive tubes is highly cuticular and broader than the rest (Figure 1F). Three pairs of dorsal cirrata (CrD), $\sim 32 \mu\text{m}$ long, occur along the trunk and contain small refractive secretions (Figures 1C, E & 3B). The first pair of tubes is present at U35, second pair at U46, and last pair inserts between and dorsal to the paired posterior adhesive tubes (TbP) on the caudal lobes. The caudal lobes are short and bear two adhesive tubes (TbP)

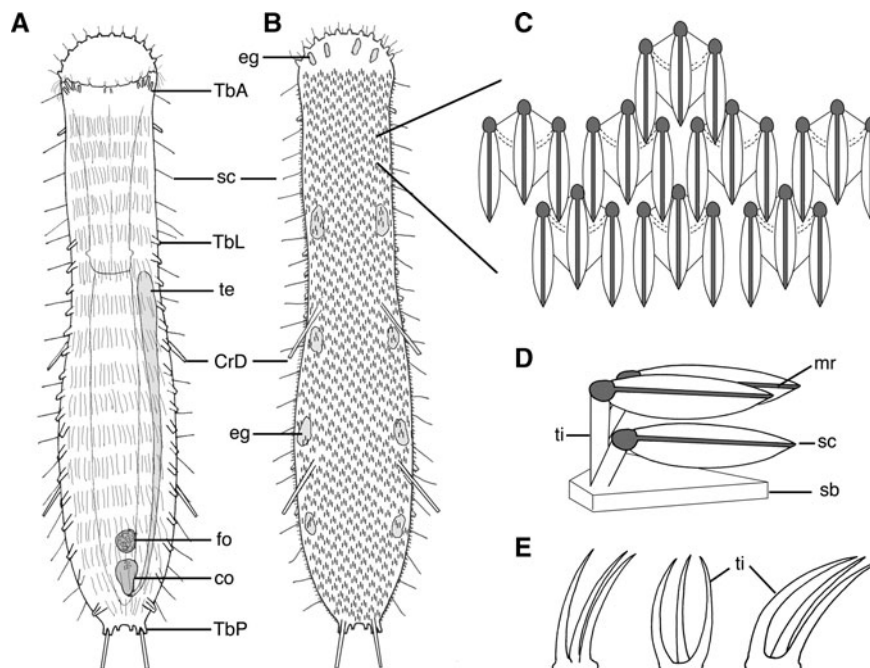


Fig. 3. Schematic illustration of *Tetranchyroderma adeleae* sp. nov. (A) Ventral view; (B) dorsal view; (C) dorsal view showing the organization of the feathered triancre; (D) hypothetical lateral view of feathered triancre; (E) lateral views of the ordinary triancre present on the ventrolateral body wall.

~4 μm long. A third pair of adhesive tubes, 4 μm long, is positioned between the caudal lobes (Figure 1F).

Ciliature: ventral locomotory cilia begin below the mouth margin and form a continuous field of transverse rows down to the base of the trunk (Figures 1B & 3A). Tactile cilia are present on the oral hood and trunk. Up to 25 pairs of lateral tactile cilia, to 20 μm in length, protrude from the ventrolateral body margin (Figure 3B).

Reproductive system: all specimens are simultaneous hermaphrodites (Figure 3A). A single testis is present on right side beginning at the pharyngo-intestinal junction and leading caudally into an elongate vas deferens. The vas deferens curves medially to connect to the base of the male caudal organ. The male caudal organ (co, Figure 3A), at ~U84, is an inverted pear-shape containing refractive spheres and wrapped in circular muscles. Anterior to the caudal organ at U73 is the frontal organ (fo, Figure 3A), 15 μm in diameter, and containing sperm. Paired ovaries are present at U88.

CUTICLE ULTRASTRUCTURE OF *T. ADELEAE* SP. NOV.

The following descriptions of the cuticle are based on a single, slightly damaged specimen. Sections through individual triancres are at an oblique angle relative to the longitudinal axis of the triancres (Figure 4). Most TEM sections were taken through the middle of the trunk region. The cuticle is divided into an outer lamellar layer, or exocuticle, and a basal layer, or endocuticle (Figure 5; see terminology by Rieger & Rieger, 1977; Ruppert, 1991). The thickness of the entire cuticle excluding the ancre is 165–226 nm depending on the body region (min: 165 nm in the ventral trunk; max: 226 nm in the dorsal trunk). The maximum thickness of the exocuticle in the dorsal and dorsolateral trunk is ~42 nm, with thinner regions ~32 nm around sensory and locomotory cilia on the ventral and lateral body walls. Individual locomotory cilia are covered with a single layer of exocuticle ~9 nm thick. The organization of the exocuticular layers into bilayers

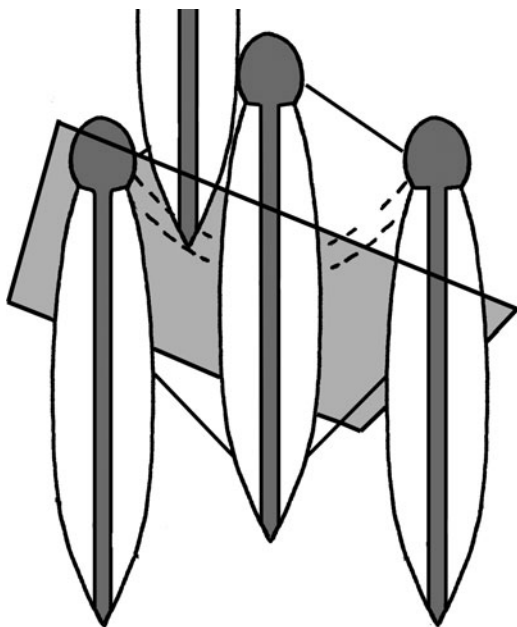


Fig. 4. Schematic illustration of the approximate TEM plane of section (shaded rectangle) through the feathered triancres.

(*sensu* Rieger & Rieger, 1977) could not be determined, nor could the number of bilayers on the trunk.

The endocuticle has a slightly granular and homogeneous appearance; it is ~108–169 nm thick in the ventral midbody region and ~121–182 nm thick in the dorsal midbody region. All ancre (ordinary triancres and feathered triancres) are specializations of the endocuticle and appear as slightly lighter regions embedded within it (Figure 5A, B). The entire base of an ordinary triancre was not captured in section. Individual triancre tines have a maximum width of 384 nm, which consists mostly of endocuticle; only a single exocuticular layer is observed around individual tines (Figure 5D). Feathered triancres consist of three individual tines (shafts) with a distinct base that is embedded within the endocuticle (Figure 5A). The ancre base is up to 3.95 μm wide in cross-section and has a maximum height of 398 nm (max exocuticle of 37 nm; max endocuticle of 361 nm). All ancre bases narrow at their posterior apex to ~180–205 nm high. The tines that arise from the base and support the feathered scales are ~550–562 nm wide. The connection between the tine and an individual feathered scale was not captured in section. Feathered scales are similar in appearance to the tine and base, with a distinctly homogeneous appearance and composed of both endocuticle and exocuticle (Figure 5A, C). There are at least two exocuticular sheets around each feathered scale. The feathered scales have a slight concavity on their outer surface (Figure 5A, C) and a maximum width (edge to edge) of 2.4 μm , though this might be an overestimation due to the oblique plane of section. Each feathered scale has a distinct midrib on its ventral surface (Figure 5C). The maximum height of a feathered scale (total cuticle) from the ventral base of the midrib to the dorsal surface of the feathered scale is ~562 nm. The outer edges of the feathered scales are ~200–226 nm high.

Sections of several sensory cilia that project from the lateral body wall revealed a robust construction (Figure 5E). These structures are composed from inside to outside of the following: a central cilium with a $9 \times 2 + 2$ axoneme; a space with membrane-like structures; a homogeneous endocuticle up to 450 nm thick; and a single layer of exocuticle. Embedded within the endocuticle are ten symmetrically disposed microvilli.

CUTICLE ULTRASTRUCTURE OF *PSEUDOSTOMELLA MEGAPALPATOR*

The following description is based on a single specimen sectioned transversely. The sculptured cuticle consists of both exocuticle and endocuticle. Individual exocuticle bilayers are ~7–9 nm thick. The exocuticle on the dorsal and ventral sides of the body has a thickness of 30–45 nm. Up to six exocuticular bilayers surround each locomotory cilium, with a maximum thickness to 40 nm. On the dorsal surface of the trunk are feathered triancres similar in appearance to those of *T. adeleae* sp. nov. The feathered triancres are constructed of individual tines that are supported by an ancre base embedded in endocuticle (Figure 6B, C & D). The base has a similar granular and homogeneous appearance to the endocuticle of the trunk. Sections through the base reveal its maximum height to be 320 nm. Individual tines arise from the base and are ~160–180 nm wide and 1.6–1.9 μm high. The tines are similar in appearance to the base. Feathered scales arise from individual tines (Figure 6C). All scales are

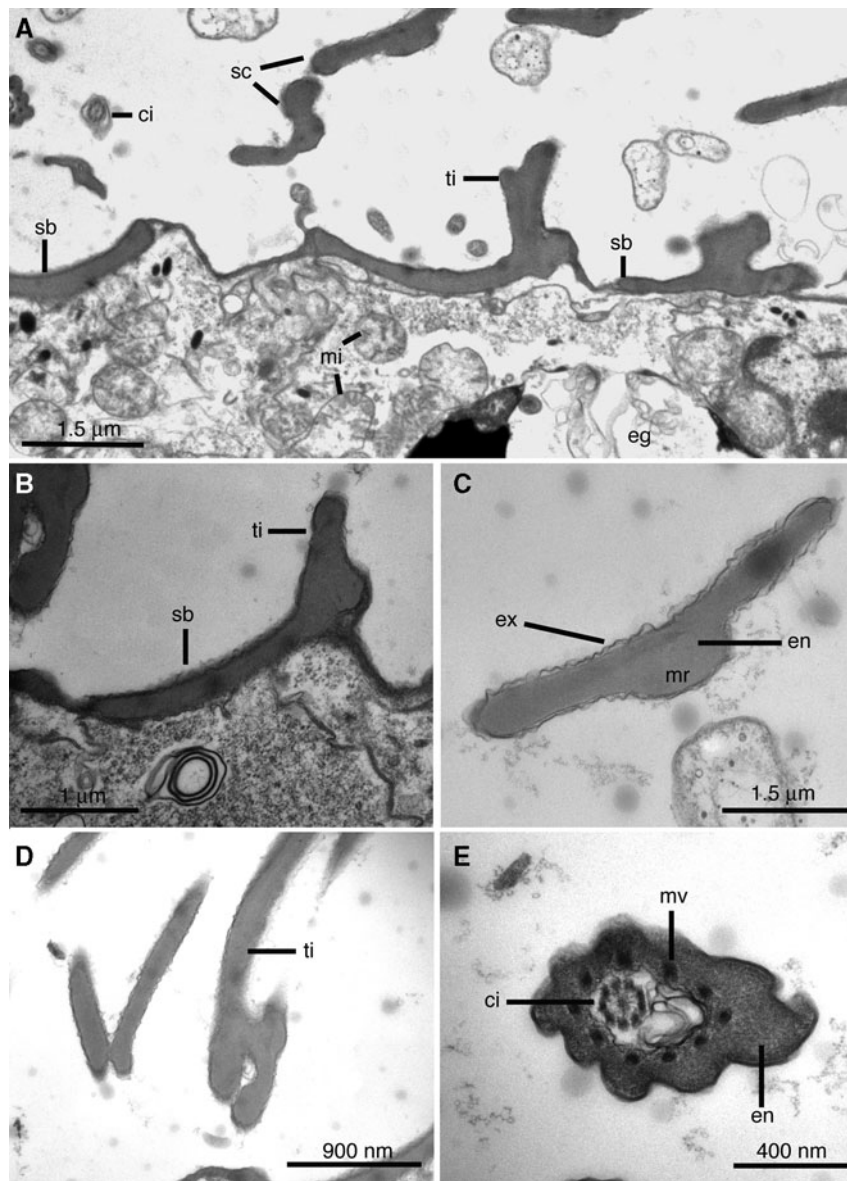


Fig. 5. TEM photomicrographs of the cuticle of *Tetranchyroderma adeleae* sp. nov. (A) Section revealing multiple feathered scales and their bases; (B) single scale base with a single tine in longitudinal section; (C) cross-section through a feathered scale; (D) longitudinal sections through ordinary triancre; (E) cross-section of a sensory cilium from the ventrolateral body wall.

concave on their outer surface; this concavity gives each scale a distinct u-shaped appearance (Figure 6C, E). A single exocuticular sheet surrounds each feathered scale. At the centre of each scale is a small depressed region. This depression is observed in all sections of scales where the scale is connected to its respective tine (Figure 6C); however, sections that show free scales may or may not have this depression evident (Figure 6E). Individual scale widths (edge to edge) and scale contours are highly variable due to the plane of section and their orientation. Some scales are extremely concave on their outer surface while others appear relatively flat.

TAXONOMIC REMARKS

Tetranchyroderma adeleae sp. nov. is the third species of *Tetranchyroderma* and the tenth species of marine Gastrotricha described from Australia (Hochberg, 2002b, c, 2003; Nicholas & Todaro, 2005, 2006). To date, there is a total of 62 described species of *Tetranchyroderma* reported

from around the globe including all continents except Antarctica; 29 species possess five-pronged spines (pentancre), 29 species possess four-pronged spines (tetrancre), one species possesses three-pronged spines (triancre), and three species possess a mixture of ancre types (Nicholas & Todaro, 2006). Within Australia, only two species of *Tetranchyroderma* have been described previously: *T. australiense* Nicholas & Todaro, 2006 and *T. pentaspersus* Nicholas & Todaro, 2007. *Tetranchyroderma adeleae* sp. nov. is the first described species in the genus to possess feathered triancre. Ruppert (1970) reports that feathered triancre are known from other species in the genus, but no formal descriptions are published. The presence of both ordinary and feathered triancre is a consistent feature of this new species.

Thus far, the only other species of *Tetranchyroderma* with a triancre covering is *T. tribulosum* Clausen, 1965 described from Norway. Apart from the difference in cuticular morphology, *T. adeleae* sp. nov. differs from *T. tribulosum* in

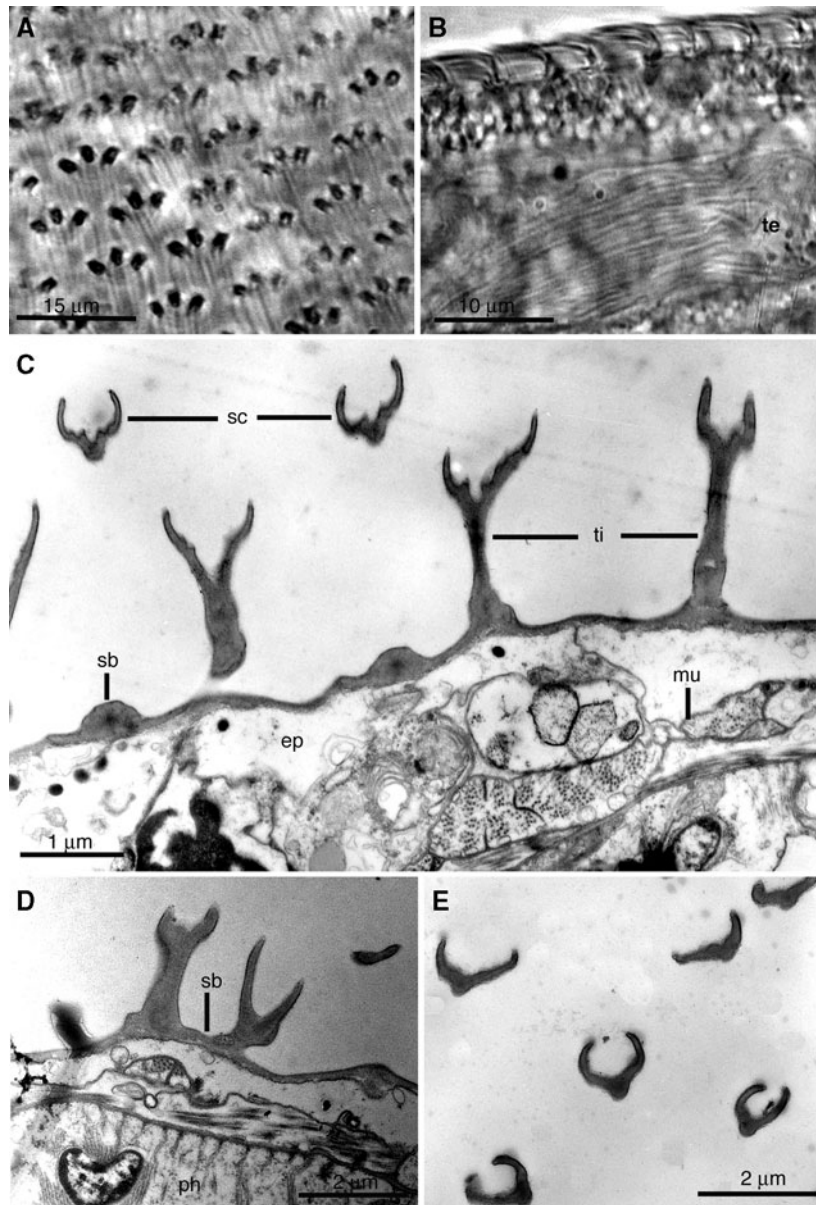


Fig. 6. Cuticle of *Pseudostomella megapalpatator*. (A) Light micrograph of dorsal cuticle; (B) light micrograph of lateral body wall; (C) TEM cross-section through the dorsal body wall; (D) TEM cross-section through the lateral body wall; (E) cross-section through individual feathered scales.

possessing a larger body size, different distribution of adhesive tubes and epidermal glands, absence of piston pits, and presence of papillae and epidermal glands on the oral hood. In general appearance, *T. adeleae* sp. nov. most closely approaches the pentancrous *T. polyprobolostomum* Hummon, Todaro, Balsamo & Tongiorgi, 1996. Both species possess elongate dorsolateral cirrata (CrD) and an oral hood with papillae along the free margin. However, *T. adeleae* sp. nov. lacks the TbD that are prevalent along the midline of the body in *T. polyprobolostomum*.

The most distinctive feature of *T. adeleae* sp. nov. is the dorsal cuticle, which under transmitted light gives a sheen-like appearance due to the imbricated feather-like scales. The gross morphology of the feathered triancres bears strong resemblance to similar ancrets described for *Pseudostomella plumosa* Ruppert, 1970, *P. klauserae* Hochberg, 2002, *P. megapalpatator* Hochberg, 2002 (Figure 6), and *P. faroensis* Clausen, 2004. However, differences are present in the relative shape of the feathered scales and

the organization of their tines. For example, the scales of both *P. plumosa* and *P. faroensis* are slightly rounded at their apex while those of *P. klauserae*, *P. megapalpatator* (Figure 6A), and *T. adeleae* sp. nov. have a more pointed appearance. Also, the latter four species appear to have feathered scales that arise independently from the base (on individual tines) as opposed to the condition in *P. plumosa* where all three feathered scales arise from a common tine.

DISCUSSION

The morphology of the cuticle in the Thaumastodermatidae is perhaps the most structurally complex within the Gastrotricha, only rivalled by members of the Chaetonotidae (order Chaetonotida). Rieger & Rieger (1977) examined the ultra-structure of the cuticle in thaumastodermatids and hypothesized that the current diversity in forms (scales, spines,

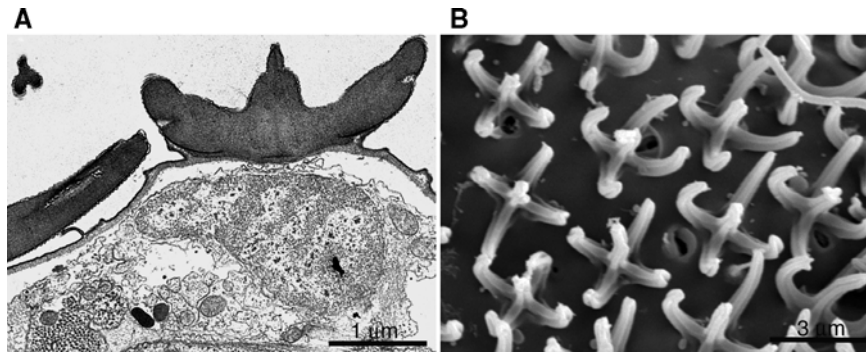


Fig. 7. Pentancre of *Tetranchyroderma* cf. *polypodium*. (A) TEM micrograph of a pentancre. Note the bowl-shaped appearance and central spine; (B) SEM micrograph revealing multiple pentancre.

triancres, tetrancres and pentancre) was derived from an ancestral bowl-shaped scale with a central spine (Figure 7A). Evolutionary modification of the bowl into a circler of spines around a central spine (pentancre, Figure 7B) is hypothesized to be the plesiomorphic condition for the subfamily Thaumastodermatinae Ruppert, 1978. Within the subfamily is a monophyletic clade containing *Pseudostomella*, *Tetranchyroderma* and *Thaumastoderma* (Hochberg & Litvaitis, 2001). Species of *Thaumastoderma* only possess tetrancres, while species of *Pseudostomella* and *Tetranchyroderma* possess pentancre, tetrancres, ordinary triancres or feathered triancres. Few species show mixed ancre types, though some show variation in the amount of ancre coverage (e.g. *Tetranchyroderma hypopsylancrum* Hummon, Tongiorgi & Todaro, 1993) and tine length (e.g. *Tetranchyroderma tanymesatherum* Hummon, Todaro, Balsamo & Tongiorgi, 1996).

The presence of more than one ancre type in species of *Tetranchyroderma* is exceptional, with only three other species known to possess mixed ancre: *T. paradoxum* Thane-Fenchel, 1970 possesses pentancre, tetrancres and ordinary triancres, and *T. paralittoralis* Rao, 1991 and *T. tentaculatum* Rao, 1993 possess pentancre and tetrancres. *Tetranchyroderma adeleae* sp. nov. is now the fourth known species to possess at least two different ancre types. The significance of mixed ancre types in a single species, or different ancre types within the Thaumastodermatinae, is difficult to ascertain without a better knowledge of species-level relationships within the subfamily. Important questions regarding the origins of the different ancre types include: (1) what is the ancestral ancre type of the subfamily?; (2) what are the selective pressures that favour different ancre types?; and (3) is the presence of mixed ancre in a single species a more primitive or derived trait relative to a single ancre type? Interestingly, no other species with feathered triancres also possess ordinary triancres; these include *Pseudostomella plumosa* (Ruppert, 1970) from the USA (Ruppert, 1970), *P. klauserae* and *P. megapalpator* from Australia (Hochberg, 2002b), and *P. faroensis* from the Faroe Bank (Clausen, 2004).

To gain insight into the differences between ordinary and feathered triancres, I examined the fine structure of both types in *T. adeleae* sp. nov. and compared them to the feathered triancres in *P. megapalpator*. Results indicate that all triancres are broadly similar in fine structure to the variety of cuticular formations described for other species of Thaumastodermatidae (Rieger & Rieger, 1977); all are solid modifications of the endocuticle with a finely granular appearance. Moreover, there is no distinctive substructure (e.g.

striations and honeycombs) in the scales as present in species of *Lepidodasys*, which are hypothesized to be closely related to Thaumastodermatidae based on morphological grounds (Ruppert, 1991; Hochberg & Litvaitis, 2001). Instead, scale appearance is comparable to the tine (shaft) and base, though there are slight differences in staining quality between the feathered triancres and the rest of the endocuticle, i.e. the ancre are slightly more electron lucent than the general body endocuticle. In terms of cuticle thickness, both the exocuticle and endocuticle of *T. adeleae* sp. nov. fall into the range known for other species of *Tetranchyroderma* (Rieger & Rieger, 1977). Moreover, the differences between species examined here with feathered triancres, *T. adeleae* sp. nov. and *P. megapalpator*, are relatively minimal, which is not unexpected considering the homogeneity of the sculptured cuticle within the family (see Rieger & Rieger, 1977; Ruppert, 1991). However, the current investigation did reveal two differences worth noting: (1) the thickness of the tines between ordinary and feathered triancres; and (2) the contour of the feathered scales among species. First, ordinary tines appear much thinner and less robust than the tines that support feathered triancres, suggesting that the evolution of feathered scales involved more than just a caudal bend and flattening of ordinary tines. Second, the cross-sectional shape of the feathered scales examined in *T. adeleae* sp. nov. are relatively flat (Figure 5), while the feathered scales of *P. megapalpator* have a heavily convex surface (Figure 6). Significantly, these differences are not apparent at the light microscopic level (compare Figure 2A, B with Figure 6A), revealing that there is likely to be more variation in cuticular sculpture of these (and other) species than is currently reported. In addition, the structure and organization of the scale bases and tines are likely to be more variable than current assessments would indicate, warranting the future use of scanning electron microscopy to provide increased three-dimensional resolution.

In addition, transmission electron microscopy indicates that the stiff sensory cilia on the trunk of *T. adeleae* sp. nov. conform to the structure of sensory cilia described in other gastrotrichs by Rieger & Rieger (1977) and Hochberg (2002a). Individual sensory cilia are enwrapped in a sleeve of endocuticle at their base (and for several microns beyond the body wall), and within the endocuticle is a ten-part symmetrical arrangement of microvilli (Figure 5E). A similar condition is also noted for *Lepidodermella squamata* (Chaetonotida), which prompted Hochberg (2002a) to hypothesize that the structure of these cilia is an autapomorphy of Gastrotricha. Species of other

taxa such as Loricifera (Kristensen, 1991), Kinorhyncha (Kristensen & Higgins, 1991) and Platyleminthes (Rohde & Watson, 1995) have sensory cilia with an eight-part arrangement of microvilli.

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