# Species of the Parasitic Isopod Genera 

 Ceratothoa and Glossobius (Crustacea: Cymothoidae) from the Mouths of Flying Fishes and Halfbeaks (Beloniformes)NIEL L. BRUCE and

THOMAS E. BOWMAN

## SERIES PUBLICATIONS OF THE SMITHSONIAN INSTITUTION

Emphasis upon publication as a means of "diffusing knowledge" was expressed by the first Secretary of the Smithsonian. In his formal plan for the Institution, Joseph Henry outlined a program that included the following statement: "It is proposed to publish a series of reports, giving an account of the new discoveries in science, and of the changes made from year to year in all branches of knowledge." This theme of basic research has been adhered to through the years by thousands of titles issued in series publications under the Smithsonian imprint, commencing with Smithsonian Contributions to Knowledge in 1848 and continuing with the following active series:

Smithsonian Contributions to Anthropology<br>Smithsonian Contributions to Astrophysics<br>Smithsonian Contributions to Botany<br>Smithsonian Contributions to the Earth Sciences<br>Smithsonian Contributions to the Marine Sciences<br>Smithsonian Contributions to Paleobiology<br>Smithsonian Contributions to Zoology<br>Smithsonian Folklife Studies<br>Smithsonian Studies in Air and Space<br>Smithsonian Studies in History and Technology

In these series, the Institution publishes small papers and full-scale monographs that report the research and collections of its various museums and bureaux or of professional colleagues in the world of science and scholarship. The publications are distributed by mailing lists to libraries, universities, and similar institutions throughout the world.

Papers or monographs submitted for series publication are received by the Smithsonian Institution Press, subject to its own review for format and style, only through departments of the various Smithsonian museums or bureaux, where the manuscripts are given substantive review. Press requirements for manuscript and art preparation are outlined on the inside back cover.

Robert McC. Adams Secretary
Smithsonian Institution

# Species of the Parasitic Isopod Genera Ceratothoa and Glossobius (Crustacea: Cymothoidae) from the Mouths of Flying Fishes and Halfbeaks (Beloniformes) 

Niel L. Bruce and Thomas E. Bowman

SMITHSONIAN INSTITUTION PRESS
Washington, D.C.
1989


#### Abstract

Bruce, Niel L., and Thomas E. Bowman. Species of the Parasitic Isopod Genera Ceratothoa and Glossobius (Crustacea: Cymothoidae) from the Mouths of Flying Fishes and Halfbeaks (Beloniformes). Smithsonian Contributions to Zoology, number 489, 28 pages, 17 figures, 1989.-New diagnoses are given for Ceratothoa and Glossobius. Three species of Ceratothoa and four species of Glossobius are reported from the mouths of four species of flying fishes and six species of halfbeaks, as follows: Ceratothoa angulata from the Philippines (no host) and Borneo (on Hyporhamphus dussumieri (Valenciennes,1846)); C. guttata from the Philippines (on "flying fish"), the Gulf of Carpentaria, Australia, Taiwan, and Madagascar (on Parexocoetus brachypterus (Richardson, 1846)); C. retusa from Mozambique (no host), Gulf of Carpentaria, Australia (on Hemiramphus robustus Günther, 1866), Cobourg Peninsula, Northern Territory, Australia (on Hemiramphus far (Forssk̊l, 1775)), West Irian (on H. far), and Durban, South Africa (on H. far); Glossobius anctus, new species, from Western Australia, New South Wales, Hawaii, and Japan (on Euleptorhamphus viridis (van Hasselt, 1823)); G. auritus from the Bahamas and the Caribbean (on Cypselurus comatus (Mitchill, 1815)), tropical eastern Pacific (no host), Japan (on Cypselurus agoo (Temminck and Schlegel, 1854)), "Ost Indien" (no host), and Thailand (on Cypselurus sp.); G. hemiramphi from Georgia, Florida, the Bahamas, the Caribbean, the west coast of Africa from Dakar, Senegal, to Luanda, Angola (all on Hemiramphus brasiliensis (Linnaeus, 1758)), and Bermuda (on Hemiramphus bermudensis Collette, 1962); G. impressa, western Atlantic from New Jersey to Rio de Janeiro, Brazil, but not in the Caribbean, and eastern Atlantic from Dakar, Senegal, to off Angola at $10^{\circ} 48^{\prime} \mathrm{S}$ (host not identified except 1 record from Hirundichthyes speculiger (Valenciennes, 1846)). All 7 isopod species are described and illustrated in detail, and a key is given to the species of Glossobius. Cteatessa and Rhexanella are reduced to synonyms of Ceratothoa. Ceratothoa hemiramphi and C. venusta are placed in synonymy with C. retusa and C. guttata, respectively. Glossobius albinae is considered a junior synonym of $G$. auritus.

Glossobius hemiramphi has two disjunct populations, a western Atlantic population ranging from Georgia to the Yucatan Peninsula and in the Caribbean; a west African coastal population ranging from Dakar, Senegal, to Luanda, Angola. Western Atlantic specimens are distinctly longer than those from West Africa.


Official publication date is handstamped in a limited number of initial copies and is recorded in the Institution's annual report, Smithsonian Year. SERIES COVER DESIGN: The coral Montastrea cavernosa (Linnaeus).

[^0]
## Contents

Page
Introduction ..... 1
Abbreviations ..... 1
Acknowledgments ..... 1
Ceratothoa Dana, 1852 ..... 1
Ceratothoa angulata (Richardson, 1910), new combination ..... 2
Ceratothoa guttata (Richardson, 1910), new combination ..... 4
Ceratothoa retusa (Schioedte and Meinert, 1883), new combination ..... 8
Glossobius Schioedte and Meinert, 1883 ..... 12
Key to the Species of Glossobius ..... 13
Glossobius anctus, new species ..... 13
Glossobius auritus Bovallius, 1885 ..... 16
Glossobius hemiramphi Williams and Williams, 1985 ..... 19
Glossobius impressus (Say, 1818) ..... 22
Literature Cited ..... 27

# Species of the Parasitic Isopod Genera Ceratothoa and Glossobius (Crustacea: Cymothoidae) from the Mouths of Flying Fishes and Halfbeaks (Beloniformes) 

Niel L. Bruce and Thomas E. Bowman

## Introduction

Beloniform fishes are infested by cymothoid isopods on the body surface, in the gill chambers, and in the mouth. Only flying fishes have these isopods on the body surface: Nerocila exocoeti Pillai, 1954, on Exocoetus brachypterus, and Nerocila trichiura (Miers, 1877) on Exocoetus evolans, E. volitans, and Cypselurus nigricans (see Trilles, 1975; Kurochkin, 1980; Bruce and Harrison-Nelson, 1988). The gill-infesting isopods, all members of the genus Mothocya, have been treated in detail by Bruce (1986) in a revision of Mothocya that included material from non-Beloniform host families. The mouthinfesting isopods, except for a few specimens of the taxonomically difficult genus Cymothoa not dealt with here, are species of Ceratothoa and Glossobius. Glossobius is fully revised here, and appears to be restricted to hosts in the families Exocoetidae and Hemiramphidae. Members of the genus Ceratothoa infest a much wider range of fish species, and only three of the more than 40 nominal species are recorded herein from hosts in the Exocoetidae and Hemiramphidae.

AbBREVIATIONS.-Abbreviations for institutions listed in the "Material" sections are given in the "Acknowledgments" section below, except for the following: AM, Australian Museum, Sydney; ANSP, Academy of Natural Sciences of Philadelphia; CSIRO, Commonwealth Scientific and Industrial Research Organization; FMNH, Field Museum of Natural History, Chicago; MCZ, Museum of Comparative Zoology, Harvard University, Cambridge; USNM, Collections of the former United States National Museum, now in the National

[^1]Museum of Natural History, Smithsonian Institution, Washington, D.C.; UZMC, Universitets Zoologiske Museum, Copenhagen.

In the "Material" sections, isopod specimens separated by commas were collected from the same fish specimen; isopod specimens separated by semicolons were collected from different fish specimens.

ACKNOWLEDGMENTS.-This work was supported by a Smithsonian Postdoctoral Fellowship to N.L. Bruce. For the loan of material used in this study we thank Åke Anderson, Naturhistoriska Riksmuseet, Stockholm (RMS), A.J. Bruce, Northern Territory Museum, Darwin (NTM); P.J.F. Davie, Queensland Museum, Brisbane (QM); H.-E. Gruner, Museum für Naturkunde der Humboldt-Universităt zu Berlin (MHUB); L.B. Holthuis, Rijksmuseum van Natuurlijke Historie, Leiden (RNHL); R.J. Lincoln and J. Ellis, British Museum (Natural History), London (BM(NH)); and M.G. van der Merwe, South African Museum, Cape Town (SAM). Our special thanks go to B.B. Collette, Bureau of Commercial Fisheries Systematics Laboratory, Washington, D.C., who collected and donated the bulk of the material that made this study possible. This manuscript benefitted from careful reviews by R.C. Brusca and E.H. Williams.

## Genus Ceratothoa Dana, 1852

Ceratothoa Dana, 1852:303; 1853:747-752.-Schioedte and Meinert, 1883:322-323.-Bowman, 1978:217.-Brusca, 1981:177.
Codonophilus Haswell, 1881:471; 1882:283.-Hale, 1926:223.
Rhexana Schioedte and Meinert, 1883:289.
Cteatessa Schioedte and Meinert, 1883:296.
Meinertia Stebbing, 1893:354; 1900:642; 1910a:103.-Richardson, 1905:236.-Menzies, 1962:116.—Schultz, 1969:156.
Rhexanella Stebbing, 1911:179.

Not Ceratothoa.-Richardson, 1905:236.—Schultz, 1969:155.—Kussakin, 1979:287. [= Glossobius Schioedte and Meinert, 1883.]
Diagnosis of Female (provisional).-Body narrow, about 2.5 to 3.5 times longer than wide, frontal margin of cephalon extended ventrally as short, blunt, triangular rostrum, not separating antennules. Pereonite 1 with anterolateral margins encompassing cephalon. Pleonite 1 much narrower than pleonites 2-5. Antennules with basal articles in contact, proximal articles broad and flattened. Pereopod 1 short, pereopods 2 and 3 longer and more slender than 1; pereopods 5-7 basis with prominent posterior expansion. Pleopods decreasing in size posteriorly, rami with pockets or depressions separated by ridge; pleopod 1 not operculate.

Male.-The same as Glossobius male.
Type Species.-As Bowman (1978) indicated, the type species, not designated by Dana (1852), must be either Cymothoa gaudichaudii Milne-Edwards, 1840 or Cymothoa parallela Otto, 1828, the two species originally assigned to Ceratothoa by Dana. The presumed holotype of C. gaudichaudii is damaged, unrecognizable, and held at the Muséum National d'Histoire Naturelle, Paris (Trilles, 1973b). We do not know where the type material of $C$. parallela is held, or indeed if it is extant. Consequently we do not designate a type species for Ceratothoa; this action is best deferred until a generic revision is undertaken.

REMARKS.-Since the majority of species of Ceratothoa are still poorly known, and since we have not studied a designated type species for the genus, the diagnosis given here is provisional.

Ceratothoa and Glossobius are very similar. However, the characters that separate adult females of the two genera are unambiguous, and it seems prudent to maintain their integrity.

Examination of type material of Cteatessa retusa, type species of the genus Cteatessa, reveals no characters by which Cteatessa can be distinguished from Ceratothoa. Schioedte and Meinert (1883) proposed a monotypic new genus for $C$. retusa. A comparison of their diagnoses of Cteatessa and Ceratothoa shows that they distinguished these two genera by the head size (large to moderate in Ceratothoa, small in Cteatessa), the eyes (distinctly developed in Ceratothoa, obscure in Cteatessa), antenna 1 (dilated, rarely compressed in Ceratothoa, compressed in Cteatessa), and the dactyls of the pereopods (usually short in Ceratothoa, very long in Cteatessa). These are mainly quantitative rather than qualitative differences, and we do not believe them to be significant at the generic level. Hence, we consider Cteatessa to be a junior synonym of Ceratothoa. Although we have not examined type material of Rhexana verrucosa Schioedte and Meinert (placed in Rhexanella by Stebbing, 1911), Schioedte and Meinert's (1883) diagnosis fails to distinguish their genus from Ceratothoa. We also place Rhexanella in synonymy with Ceratothoa.

Within the genus Ceratothoa species can be grouped according to their pereopod morphology. A feature of the
genus, generally considered typical, is the prominent expansion of the basis of the posterior pereopods. Most Ceratothoa species, including the two initially placed in the genus by Dana (1852), have this character. But Ceratothoa gilberti (Richardson, 1904) (see Brusca, 1981) has no trace of this basal expansion, and it is only weakly developed in Ceratothoa steindachneri Koelbel, 1879 (see Trilles, 1973a). Another pereopodal character that is unequally distributed within the genus is the expansion of the ischium of the posterior pereopods. This occurs in Ceratothoa guttata (Richardson, 1910) and C. carinata (Bianconi, 1870) (see Trilles, 1972).

## Ceratothoa angulata (Richardson, 1910), new combination

## Figures 1, 2

Meinertia angulata Richardson, 1910:22, fig. 21.
Codonophilus angulatus (Richardson).-Nierstrasz, 1931:132.
Material.-Pacific ocean: Philippines: Luzon, Port San Pio (= Port San Pio Quinto, $18^{\circ} 54^{\prime} \mathrm{N}, 121^{\circ} 51^{\prime} \mathrm{E}$ ), near mouth of a small stream, 20 ft . seine, ovigerous $\varphi, 21.5 \mathrm{~mm}$ (holotype, USNM 41008). Indonesia: Borneo, ex Hyporhamphus dussumieri (Valenciennes) (Division of Fishes, USNM 218692), non-ovigerous $\varphi, 17.5 \mathrm{~mm}$ and $\sigma^{7}, 7.0 \mathrm{~mm}$ (USNM 240023).

TYPE.-Holotype, USNM 41008.
Type Locality.-See "Material."
DESCRIPTION of FEmale.-Length/width: holotype 2.5, Borneo 3.2. Width, including coxae, greatest at pereonite 5. Cephalon triangular, rounded anteriorly, lateral margins anterior to eyes slightly concave in holotype, nearly straight in Borneo $\$$. Eyes moderate-size, outlines fairly distinct. Pereonite 1 shoulders reaching level of anterior margins of eyes, anterior margins round-truncate, lateral margins shallowly sinuate, anterior one-third to one-half raised into ridge; medial to each ridge is a subcircular depression. Pereonites 2-5 subequal, about two-thirds length of pereonite 2, pereonites 6 and 7 progressively shorter; coxae rather narrow, shorter than pereonites, margins not elevated. Length of pleon as percent of total length: holotype 15.0, Borneo 13.4. All pleonites visible and subequal in length; pleonite 5 equal in width to pleotelson anterior margin. Pleotelson length/width: holotype 0.53, Borneo 0.55 , widest at base; lateral margins evenly rounded, posterior margin shallowly emarginate.

Antennule composed of 7 dorsoventrally flattened articles. Antenna about as long as antennule, more slender, composed of 8 articles. Antennules and antennae extending to middle of eye, posterior to anterior margin of pereonite 1 shoulders.

Labrum crescent-shape. Mandible palp articles all distinct. Maxillule with 2 longer and 2 shorter apical spines. Maxilla with partly fused medial and lateral lobes each armed with 6 recurved spines. Maxilliped palp article 3 with 2 recurved apical spines and 2 more slender straight spines on medial margin.


Figure 1.-Ceratothoa angulata (non-ovigerous $\$, 17.5 \mathrm{~mm}$, Borneo): $a$, dorsal view; $b$, pereon, lateral view; $c$, buccal area; $d$, maxillule apex; $e$, maxilla; $f$, maxilliped; $g-m$, pereopods $1-7 ; n-q$, pleopods $1-4 ; r$, pleopod 5 , anterior view; $s$, pleopod 5 , posterior view.


FIGURE 2.-Ceratothoa angulata ( $a, \sigma^{7}, 7.0 \mathrm{~mm}$, Borneo; $b-c$, $\%$, holotype, 21.5 mm , Philippines): $a$, dorsal view; $b$, cephalon and pereonite 1 , dorsal view; $c$, right uropod, ventral view.

Pereopod 1 robust, basis rectangular; merus about one-third length of ischium and one-third wider than carpus; carpus short, triangular; propodus half length of ischium, palm straight. Pereopod 2 similar to pereopod 1, but merus less produced. Pereopods 5-7 with basis progressively more expanded posteriorly.

Pleopods with depression on anterior surface and corresponding elevation on posterior surface of exopod; elevation fits into depression on anterior surface of endopod; posterior surface of endopod with corresponding elevation.

Uropods extending to or slightly beyond posterior margin of pleotelson; rami subequal in length, both with straight medial margin and convex lateral margin; exopod spatulate, endopod narrowing distally.

DESCRIPTION OF MALE.-Length/width: 3.0. Cephalon relatively shorter than in $\%$. Eyes more densely pigmented. Pereonite 1 shoulders obtuse anteriorly. Pereonite 1 lateral margins not elevated. Pleon length $15.3 \%$ of total length. Pleotelson length/width: 0.56 .

COLOR.-Cephalon, pereonites 1-3, and anterior half of pereonite 4 colored with black chromatophores. Posterior to this, body is unpigmented in holotype; Borneo $q$ is pigmented on posterior pereonites, pleon, and anterior part of pleotelson with more widely scattered chromatophores; rest of pleotelson is unpigmented. $\sigma^{7}$ is nearly unpigmented, but has black chromatophores on antennules and very widely scattered on body.

Size.-See "Material."
REMARKS.-The distinguishing features of $C$. angulata are the truncate shoulders of pereonite 1 , the slightly emarginate pleotelson, and the spatulate exopod of the uropod.

Host.-Unknown for the holotype. The Borneo specimens were taken from Hyporhamphus dussumieri (Valenciennes).

DISTRIBUTION.-Known from the Philippines and Borneo. The host, Hyporhamphus dussumieri, is a "wide-spread tropical species found from Madagascar through the East Indies to the Philippines, New Guinea, Australia and Oceania" (Collette, 1974:89). It is possible that Ceratothoa angulata has a similar widespread distribution.

## Ceratothoa guttata (Richardson, 1910), new combination

## Figures 3, 4

Meinertia guttata Richardson, 1910:20-21,fig. 19.
Codonophilus guttatus.-Nierstrasz, 1931:132.
Meinertia venusta Avdeev, 1978:30-32, fig. 1.
Ceratothoa venusta.-Avdeev, 1981:1160, 1164, fig. 3 [part].
Material.-Indian ocean: Madagascar: Ex Parexocoetus brachypterus (Richardson), ovigerous $9,20.0$ mm (USNM 216589). PACIFIC OCEAN: Taiwan: Off Keelung, ex Parexocoetus brachypterus, coll. H. Teng, 6 Jun 1957, ovigerous $\%, 23.0 \mathrm{~mm}$ and $\sigma^{\circ}, 5.8 \mathrm{~mm}$ (USNM 216587). Philippines: Jolo Island, from mouths of flying fish, coll. U.S. Bureau of Fisheries Albatross Philippine Expedition


FIGURE 3.-Ceratothoa guttata (a-e, $s$, syntypes; $f-r$, ovigerous $q$, topotype, 17.5 mm ): $a, q, 18.6 \mathrm{~mm}$, with mancas; $b$, non-ovigerous $\$, 15.5 \mathrm{~mm}$; $c$, buccal area, non-ovigerous $\$, 15.5 \mathrm{~mm} ; d$, ventral view of pleopods, non-ovigerous $q, 15.5 \mathrm{~mm} ; e$, pereopod 1 , non-ovigerous $\phi, 15.5 \mathrm{~mm} ; f$, antennule; $g$, antenna; $h$, lateral view; $i$, pereopod 1 ; $j$, pereopod 3; $k$, pereopod 7; $l$, uropod (broken), non-ovigerous 9 ; m, maxilliped; $n$, maxilla; $o$, maxillule apex; $p$, mandible; $q$, maxilla apex; $r$, maxilliped article 3 ; $s$, maxilliped (broken), non-ovigerous $q, 15.5$ mm . (Scale line $=4.0 \mathrm{~mm}$.)


FIgURE 4.-Ceratothoa guttata (all topotypic; a-e, ovigerous $9,17.5 \mathrm{~mm} ; f-v, \sigma^{7}, 6.5 \mathrm{~mm}$ ): a, pleopod 1 , ventral view; $b$, pleopod 1 , dorsal view; $c-e$, pleopods $2,3,5$, respectively; $f$, dorsal view; $g$, buccal area; $h$, pereon and pleon, lateral view; $i$, antennule; $j$, antenna; $k$, maxilla apex; $l$, maxilliped; $m$, pereopod $1 ; n$, pereopod 7; 0 , maxilliped article 3; $p$, penes; $q$, mandible palp; $r$ - $u$, pleopods $1-3,5$, respectively; $v$, uropod. (Scale line $=2.0 \mathrm{~mm}$.)

1907-1909, 7 Feb 1908, 4 ovigerous ㅇ, 14.5, 14.8, 16.4, 18.6 $\mathrm{mm} ; 2$ non-ovigerous $\uparrow, 15.5$ and $\sim 16.5 \mathrm{~mm} ; 3 \sigma^{\top}, 5.5,6.0,7.4$ $\mathrm{mm} ; 100+$ mancas (all syntypes, USNM 40914). Jolo Island, from mouths of Parexocoetus brachypterus, coll. U.S. Bureau of Fisheries Albatross Philippine Expedition 1907-1909, 7 Feb 1908, 4 ovigerous $\uparrow, 15.0,17.5,18.8,19.2 \mathrm{~mm} ; 1$ broken
 mm (USNM 216588). Australia: Queensland, Gulf of Carpentaria, off Mornington Island, from mouth of Parexocoetus brachypterus, coll. A. Harris, 16 Mar 1983, ovigerous $q$, 14.5 mm , CSIRO (QM W11752).

Types.-Syntype series, USNM 40914.
TyPE LOCALITY.-Jolo Island, Philippines, $\sim 5^{\circ} 58^{\prime} \mathrm{N}$, $121^{\circ} 06^{\prime} \mathrm{E}$.

Description of Female.-Body about 2.7 times longer than wide, widest at pereonite 5; pereonite 1 longest, pereonites 2-6 about equal in length, pereonite 7 less than half (0.38) length of pereonite 6 ; median broad longitudinal ridge runs from pereonite $2-5$ or 6 . Cephalon with posterolateral margins weakly overlapped by anterolateral margins of pereonite 1 ; anterior margin produced to form narrowly rounded rostrum; eyes small. Pereonite 1 narrower than pereonite 2, anterolateral margins weakly produced anteriorly, weak dorsal depression at angle formed by indentation of anterior margin. Pereonites 2 and 3 with dorsolateral bosses; coxae large, all shorter than segments; coxa on one side of pereonite 7 much smaller than opposite coxa. Pleon short, about $12 \%$ of total body length; pleonite 1 narrow; all pleonites visible, subequal in length; pleonite 5 wider than pleotelson anterior margin, posterior margin with 2 ill-defined submedian lobes. Pleotelson 0.72 as long as wide, subrectangular, posterior margin with median emargination.

Antennule robust, composed of 7 articles, first 3 of which are long, expanded, and flattened. Antenna less robust, composed of 8 articles, first 4 of which are long.

Labrum small, set posterior to antennal bases; mandible palps lie anterior to labrum. Mandible incisor robust, palp prominent, greatly extending beyond incisor, articles 1 and 2 fused. Maxillule with 1 broad-based and 3 slender spines. Maxilla endopod and lateral lobe each with 4 recurved spines. Maxilliped composed of 3 articles and obscurely segmented basal article; article 1 with laminar oostegital lobe, second oostegital lobe arises from basal article; palp article 3 oval, with 2 spines.

Pereopod 1 robust, basis rectangular; merus less than half length of ischium, posterior margin dilated; carpus short, triangular; propodus about equal to combined lengths of merus and carpus, palm straight. Pereopods 2 and 3 longer than pereopod 1 , all articles proportionally more slender, merus not expanded. Pereopods 4-7 basis with posterior expansion, becoming progressively larger toward posterior. Pereopods 5-7 ischium anterior margin dilated, merus set into lateral side of ischium; carpus short, triangular. Brood pouch made up of 5 pairs of alternately overlapping oostegites; those arising from sternite 1 not forming part of egg chamber but lie flat over
mouthparts; remaining oostegites arise from sternites $2,3,4$, and 6.
Pleopods all lamellar, all rami except pleopod 1 endopod with 2 marked depressions; all peduncles without developed lobes; pleopods 3-5 endopods with weakly developed proximomedial lobe. Pleopod 1 largest, curves laterally and dorsally to partly cover lateral margins of pleon, covers other pleopods; endopod much smaller than exopod; pleopods 2-5 becoming much smaller toward posterior. Uropod short, held under posterior margin of pleotelson, rami curve medially, subequal in length, apices narrowly rounded to acute.

DESCRIPTION OF MALE.-Much smaller than female, body straight with smoothly arched dorsum. Coxae all as long as segment. Pleon very narrow, pleonite 1 distinctly narrower than others. Pereopod 1 merus not dilated, carpus proportionally longer than in female; pereopods 1-3 similar. Pereopods 5-7 similar, basis with anterior expansion, ischium not dilated. Mandible palp with 3 unfused articles, article 3 with setae. Maxilla endopod and lateral lobe each with 2 recurved spines. Maxilliped without basal article, article 3 narrow with 2 terminal stout recurved spines. Pleopods simple, without folds or depressions; peduncles longer than in female. Pleopod 2 without appendix masculina. Pleopods 1-5 progressively decreasing in size.

COLOR.-The syntypes have faded and lack color; the topotypic series and the more recent material from Madagascar and Taiwan show a dense covering of chromatophores over the anterior segments and their appendages which lessens toward the posterior. Pereonite 7, pleon, and pleotelson are virtually without chromatophores.
Size.-Ovigerous females between 14.5 and 23.0 mm , non-ovigerous females between 15.5 and $\sim 16.5 \mathrm{~mm}$, males between 5.4 and 7.4 mm .

VARIATION.-Non-ovigerous females lack the oostegital lobe of the maxilliped, but the maxilliped structure is the same. Not all females have the coxae of pereonites 5-7 laterally and dorsally displaced, this being restricted to females with mancas near release.

Remarks.-This species has not been recorded since it was first described, and is here placed in the genus Ceratothoa. Meinertia venusta from the mouth of Parexocoetus brachypterus from the Red Sea (Avdeev, 1978) appears, from Avdeev's drawings of the unique specimen (an 11 mm female), not to differ from the present material and is here placed in synonymy with C. guttata. Our request for the loan of Avdeev's holotype was not answered.
This species belongs to the group of Ceratothoa species in which pereopods 5-7 have strongly developed expansions on the basis. Within that group two species have the ischium expanded posteriorly, Ceratothoa guttata and C. carinata (see Trilles, 1973b).

Host.-Recorded only from Parexocoetus brachypterus (Richardson).
Distribution.-Red Sea; Madagascar; Taiwan; Philippines; Gulf of Carpentaria, Australia.

Ceratothoa retusa (Schioedte and Meinert, 1883), new combination

## Figures 5-8

Cymothoa (Ceratothoa) retusa Schioedte and Meinert in Hilgendorf, 1879:847 [nomen nudum].
Cteatessa retusa Schicedte and Meinert, 1883:297-299, pl. 11, figs.11-13.Stebbing, 1910b:424.-Barnard, 1925:393.-Nierstrasz,1931:131.Kensley, 1978:80, fig. 32G, H.-Trilles, 1986:625, fig. 2a [from Schioedte and Meinert].
Codonophilus hemiramphi Pillai, 1954:14-15.
Material.-Indian ocean: Mozambique: Host unknown (MHUB 1 708), leg. W. Peters, ovigerous $\%, 33.8 \mathrm{~mm}$ (syntype). South Africa: Durban Bay, ex Hemirhamphus far (Forsskål) (SAM-A6063), coll. H.W. Bell-Marley, 1919, ovigerous $\%$ in 2 pieces, estimated length 31.5 mm ( 33 mm , Barnard, 1925) and $\sigma^{\prime}, 10.7 \mathrm{~mm}(12 \mathrm{~mm}$, Barnard, 1925). pacific ocean: West Irian (formerly Dutch New Guinea): Ex Hemiramphus far (AM I-1339), of with empty marsupium, 27.5 mm and $\sigma^{\circ}, 9.1 \mathrm{~mm}$ (USNM 235308). Australia: Northern Territory, Coburg Peninsula, Point Danger, ex tongue Hemiramphus far (NTM Cr. 002318), coll. B.C. Russell and H. Larson, 30 Apr 1982, non-ovigerous 9 , 22.8 mm . Northern Territory, Gulf of Carpentaria, Groote Eylandt area, ex Hemiramphus robustus Günther (NTM Cr. 002319), coll. Northern Territory Fisheries, 4 Mar 1983, non-ovigerous $\rho, 22.3 \mathrm{~mm}$.

Types.-Syntypes, Stockholm Museum, 2 \& from Durban ("Port Natal"); Berlin Museum, 1 \& from Mozambique (figured by Schioedte and Meinert, 1883, pl. 11, figs. 11-13).

DESCRIPTION OF FEMALE.-Length/width: Mozambique 3.2, West Irian 2.1, Point Danger 2.3, Durban 2.9, Groote Eylandt 3.2. Width, including coxae, greatest at pereonite 6. Cephalon triangular, narrowly rounded or (in Mozambique and Durban specimens) pointed anteriorly. Eyes small, outlines vague, poorly pigmented, sometimes covered by shoulders of pereonite 1. Pereonite 1 dorsum concave on either side of cephalon, lateral margin elevated into ridge recurving medially. Pereonite 1 longest; pereonites $2-4$ subequal, about half length of pereonite 1; pereonites 5-7 progressively shorter; coxae with dorsal process curved medially, better developed on posterior pereonites. Length of pleon as percent of total length: Mozambique 12.4, West Irian 15.6, Point Danger 20.2, Durban 14.3, Groote Eylandt 15.4. All pleonites visible; pleonites 1-4 subequal in length; pleonite 5 distinctly longer, wider than pleotelson anterior margin. Pleotelson length/width: Mozambique 0.54, West Irian 0.29, Point Danger 0.39, Durban 0.41, Groote Eylandt 0.47; lateral margins evenly rounded, not bulging; posterior margin shallowly emarginate.

Antennule composed of 7 articles; first 3 articles expanded, flattened; last 3 articles may be fused. Antennule of Mozambique syntype composed of 4 articles, distal articles partly fused. Antenna about as long as antennule, more slender, composed of 6-9 articles depending on extent of fusion of distal articles.

Labrum forming $U$-shaped arch around mouth. Articles of mandibular palp all distinct; 3rd article with 2 blunt apical spines. Maxillule with 2 longer and 2 shorter curved apical spines. Maxilla with partly fused medial and lateral lobes armed with 6 and 10 or more recurved spines respectively. Maxilliped article 3 with 2 curved apical spines and 2 small straight spines on medial margin.

Pereopod 1 robust, basis rectangular; merus about half length of ischium, somewhat expanded posteriorly; carpus short, subtriangular; propodus slightly shorter than combined lengths of merus and carpus, palm straight. Pereopods 2-4 much longer than pereopod 1; all articles proportionally more slender, merus not expanded, dactyls very long. Pereopods 5-7 with basis expanded posteriorly, expansion progressively larger posteriorly.

Pleopods decreasing in size posteriorly; rami with 2 depressions separated by oblique ridge. Uropod rami longer than peduncle, oval, narrowing slightly distally; curving slightly medially; rami subequal in length, or 1 ramus (outer or inner) slightly longer than other.

DESCRIPTION OF MALE.-Length/width: West Irian 2.5, Durban 3.0. Cephalon more rounded than in $q$. Eyes well developed, fully pigmented. Pereonite 1 shoulders narrower and shorter than in 9 . Pereonites 2-6 subequal in width, pereonite 7 slightly narrower. Length of pleon as percent total length: West Irian 17.2, Durban 15.8. Pleotelson relatively longer; length/width: West Irian 0.76, Durban 0.79.

COLOR.-Anterior part varying from tan to nearly black in alcohol. Posterior part much paler, with little or no pigment. Boundary between dark and pale part varies from posterior margin of pereonite 3 to pereonite 5 .

SIzE.-Ovigerous females between 27.5 and 33.8 mm , non-ovigerous females between 22.3 and 31 mm , males between 9.1 and 10.7 mm .

Variation.-The cephalon is most pointed in the South African female, least in the Point Danger female. The shoulders of pereonite 1 are broadest and most elevated laterally in the West Irian female, less so in the Point Danger female, and least in the South African and Groote Eylandt females. Recurving of the coxae is most strongly developed in the West Irian female. The merus of pereopod 1 and 2 is most inflated in the South African female.

REMARKS.-Pillai's (1954) preliminary diagnosis of Codonophilus hemiramphi was brief, and no illustrations accompanied it. A later report (Pillai, 1964) gave full descriptions and illustrations of three of the isopods diagnosed in 1954, but did not include C. hemiramphi. His diagnosis (Pillai, 1954:15) of C. hemiramphi was based largely on pereonite 1: "The anterior half of the lateral border is very prominently ridge like and a little internal to this is another equally prominent dorsal ridge. The space in between these two ridges is flat or even slightly concave giving it the appearance of an independent facet." If his dorsal ridge is the continuation of the lateral ridge medially onto the dorsal surface of pereonite


Figure 5.-Ceratothoa retusa ( $a-j, \$, 27.5 \mathrm{~mm}, l-n, \sigma^{*}, 9.1 \mathrm{~mm}$, West Irian): $a$, dorsal view; $b$, lateral view; $c$, antennule; $d-j$, pereopods 1-7; $k$, uropod; $l$, dorsal view; $m$, cephalon, dorsal view; $n$, penes.


Figure 6.-Ceratothoa retusa ( $\$, 22.8 \mathrm{~mm}$, Point Danger, Northern Territories, Australia, NTM Cr. 002318): $a$, dorsal view; $b$, buccal area; $c$, pereonite 1 , lateral view; $d$, antennule; $e$, antenna; $f$, left mandible; $g$, maxillule apex; $h$, maxilla; $i$, maxilliped; $j$ - $p$, pereopods $1-7 ; q$, pleopod 1 , posterior view; $r$, pleopod 2 endopod, posterior view; $s$, pleopod 2 exopod, posterior view; $t-v$, pleopods 3-5, posterior view; $w$, left uropod, ventral view.


FIgURE 7.-Ceratothoa retusa ( $a-i, q, \sim 31.5 \mathrm{~mm}, j-n, \sigma^{7}, 10.7 \mathrm{~mm}$, from Durban, South Africa; $0,9,22.3 \mathrm{~mm}$ from Groote Eylandt area, Gulf of Carpentaria, Australia): $a$, anterior body, dorsal view; $b$, posterior body, dorsal view; c-e, pereopods $1-3 ; f$, pereopod 5; $g-i$, coxa and basis of left pereopods 5-7; j, dorsal view; $k-m$, pereopods $1-3 ; n$, pereopod $6 ; o$, dorsal view.


Figure 8.-Ceratothoa retusa (syntype of Cteatessa retussa Schioedte and Meinert from Berlin Museum): $a$, head and pereonite 1, dorsal view; $b$, pleon and pleotelson, dorsal view; $c-8$, pereonites 1-5; $h$, pereonite 7.

1 , this description fits our specimens.
That the host of C. hemiramphi is Hemiramphus far strengthens the case for considering this isopod a synonym of Ceratothoa retusa.

Hosts.-Not known for Schioedte and Meinert's (1883) specimens. Pillai's specimen(s) (1954) and all our specimens were taken from Hemiramphus far except the Groote Eylandt specimen, which came from Hemiramphus robustus.

Distribution.-Northern Australia; West Irian; coast of Kerala state, southwestern India; Durban, South Africa; Mozambique.

## Genus Glossobius Schioedte and Meinert, 1883

## Glossobius Schioedte and Meinert, 1883:299.

Ceratothoa.-Richardson, 1905:283.-Schultz, 1969:155.-Kussakin, 1979:287. [Not Ceratothoa Dana, 1852, sensu Brusca, 1981.]
DIAGNOSIS of FEMALE.-Body narrow, about 2.5 to 3.5 times longer than wide; cephalon rostrum as in Ceratothoa. Pereonite 1 lateral margins lobed, anterior margin not recessed or only weakly recessed to accommodate cephalon. Pereonites 6 and 7 shortest, 7 markedly so; pleonite 1 distinctly narrower than pleonites 2-5. Antennule basal articles in contact, proximal articles broad and flattened. Pereopod 1 short, pereopods 2 and 3 longer and more slender than 1; pereopods 5-7 basis with prominent anterior expansion. Pleopods all lamellar, decreasing in size toward posterior; pleopod 1 indurate, exopod forming operculum.

DESCRIPTION OF FEMALE.-Body straight, usually between 2.5 and 3.5 times longer than wide. Cephalon with prominent rostrum; eyes small, subtriangular, facets indistinct. Pereonite 1 lateral margins weakly or prominently lobed; anterior margin not recessed or only weakly recessed to accommodate cephalon. Pereonite 1 longest, pereonites 6 and 7 short. All coxae shorter than their segments. Pleonites subequal in length; pleonite 1 distinctly narrower than pleonites $2-5$ which are subequal in width.

Basal articles of antennules in contact, antennule proximal articles broad and flattened. Mandible palp large, folding across anterior to prominent labrum. Maxillule styliform, with 3 or 4 terminal spines. Maxilla broad with abundant spines on lateral lobe ( 12 to 21 ) and endopod ( 9 to 11). Maxilliped set distinctly posterior to other mouthparts, with 2 oostegital lobes. Pereopod 1 short, pereopods 2 and 3 longer and more slender than pereopod 1, pereopods 5-7 basis with flattened posterior expansion. Brood pouch with 5 pairs of oostegites arising from sternites 1-4 and 6, anterior pair overlying maxillipeds. Pleopods all lamellar, decreasing in size toward posterior; pleopod 1 indurate, covering all other pleopods; pleopod rami with depressions which may form pockets or folds.

DESCRIPTION OF MALE.-Body approximately rectangular in shape; pleonite 1 much narrower than pleonites 2-5. Appendages similar to female but antennule and antenna less robust, maxilla with fewer spines, pereopods 1-3 similar to each other, and pereopods 5-7 expansions of basis less developed. Penes prominent, apices lie between pleopod
peduncles. Pleopod rami all lamellar; pleopod 2 without appendix masculina.

TYpe Species.-Ceratothoa linearis Dana, 1853. Schioedte and Meinert (1883) included two of Dana's (1853) species, Ceratothoa crassa and C. linearis, when they established Glossobius. To our knowledge, a type species for the genus has not been designated previously, and we select Glossobius linearis as type species. Glossobius lineatus is generally considered to be a junior synonym of $G$. impressus.

REMARKS.-The major problem in diagnosing and discriminating Glossobius, Ceratothoa, and related genera is the lack of modern descriptions for species or diagnoses to genera. The only recent generic treatment is that of Brusca (1981) diagnosing Ceratothoa and Cymothoa. Bowman (1978) re-
solved the nomenclatural problems but was not concerned with how to distinguish the genera. At present, differences between Glossobius and Ceratothoa seem only to be that Ceratothoa has the anterolateral margins of pereonite 1 projecting forward, the anterior margin of pereonite 1 recessed, and pereonite 6 longer than in Glossobius.

Another similar genus is Lobothorax Bleeker, 1857. Lobothorax is said to differ from Glossobius in having large eyes, a broad flat rostrum, pereonite 1 produced into spoonlike shoulders, a median dorsal carina in some species, pereonites 5-7 very short, and antennule bases that are set apart.

Species currently included in Glossobius are G. impressus, G. auritus, G. hemiramphi, and Glossobius anctus, new species.

## Key to the Species of Glossobius

| 1. Pereonite 1 with large bulbous lateral lobe . . . . . . . . . . . . . . . G. impressus |  |
| :--- | :--- |
|  | Pereonite 1 without bulbous lobe . . . . . . . . . . . . . . . . . . . . . . . . . . 2 |

## Glossobius anctus, new species

## Figures 9, 10

MATERIAL.-All from Euleptorhamphus viridis (van Hasselt). PACIFIC OCEAN: Australia: Western Australia, North West Shelf, $18^{\circ} 25^{\prime} \mathrm{S}, 118^{\circ} 52^{\prime} \mathrm{E}, 150 \mathrm{~m}$ depth, coll. F.R.V. Soela, 17 May 1979, ovigerous $\uparrow, 28.0 \mathrm{~mm}$ (holotype, AM P35743) and $\sigma^{\top}, 9.5 \mathrm{~mm}$ (paratype, AM P35744). New South Wales, Bryon Bay, $28^{\circ} 38^{\prime}$ S, $153^{\circ} 37^{\prime}$ E, 26 Jun 1910, $\sigma^{\top}, 8.0$ mm (AM P35745). Japan: Iozu Sea, non-ovigerous $\uparrow, 26.0$ mm (paratype, USNM 227113). Wahasa Bay, Yoroo, 1929, ovigerous $8,30.0 \mathrm{~mm}$ and immature, 4.0 mm (paratypes, USNM 227110). Hawaii: Honolulu, ovigerous $\uparrow, 25.5 \mathrm{~mm}$ and $\sigma^{\prime}, 8.2 \mathrm{~mm}$ (paratypes, USNM 227112). Honolulu, coll. Albatross, ovigerous $\rho$, approximately 34 mm , head missing, (USNM 227111).

TYPES.-Holotype, AM P35743; paratypes, AM P35744, USNM 227110, $227112,227113$.

Type Locality.-North West Shelf of Western Australia, $18^{\circ} 25^{\prime} \mathrm{S}, 118^{\circ} 52^{\prime} \mathrm{E}$.

ETYMOLOGY.-From the Latin anctus, meaning choke, alluding to the buccal-filling size of this species.

DESCRIPTION OF FEMALE.-Body straight, about 4 times as long as wide, sides subparallel; dorsum strongly vaulted. Rostrum anterior margin bluntly rounded; eyes very small, triangular, facets indistinct. Pereonite 1 anterolateral angles scarcely produced. Pereonites 1-3 subequal in length; pereonites 4-6 progressively decreasing in length, pereonite 7
distinctly shorter than pereonite 6 , about $25 \%$ length of pereonite 1. Coxae all shorter than pereonites; widest anteriorly, between half and two-thirds as wide as long. Pleonites 1 and 5 subequal in length, longer than pleonites 2-4; posterior margin of pleonite 5 not lobed. Pleotelson lateral margins converging slightly; posterior margin truncate, with medial emargination.

Antennule extending to posterior of cephalon, composed of 7 articles. Antenna extending to posterior of cephalon, composed of 9 articles.

Labrum prominent, fleshy, anterior margin rounded, posterolateral margins produced. Mandible palp article 3 with 6 terminal setae, medial margins with microtrichs. Maxillule with 4 terminal spines. Maxilla with 14 spines on lateral lobe, 10 on endopod. Maxilliped article 3 with 2 terminal spines.

Pereopods 1 and 2 of similar proportions, but pereopods 2 and 3 longer than pereopod 1 and pereopod 3 less robust than 1 and 2.

Pleopods same as $G$. impressus. Uropod rami both bluntly rounded, extending just beyond posterior of pleotelson.

DESCRIPTION OF MALE.-About one-third length of female. Antennule with 6 or 7 articles, antenna with 7 or 8 . Maxilla lateral lobe with 4 spines, endopod with 3. Mandible palp article 2 with about 6 stout setae at mediodistal angle; article 3 medially constricted, with 8 stiff setae on mediodistal margin. Maxillule with 4 terminal spines. Maxilliped article 3 with 4 terminal hooked spines; medial margins with fine serrate scales. Pereopods less robust than in female; pereopods 5-7


Figure 9.-Glossobius anctus (a-c, e, ovigerous $q, 28.0 \mathrm{~mm}$, Western Australia, holotype, AM P35743; d, ovigerous $q, 25.0 \mathrm{~mm}$, Hawaii, ANSP 91844; $f-o$, ovigerous $q, 30.0 \mathrm{~mm}$, Japan, paratype, USNM 227110; $p$, non-ovigerous $\rho, 26 \mathrm{~mm}$, Japan, paratype, USNM 227113): $a$, dorsal view; $b$, lateral view; $c$, cephalon and pereonite 1, dorsal view; $d$, lateral view; $e$, cephalon, ventral view, $f$, pereopod $1 ; g$, pereopod $2 ; h$, pereopod 7; $i$, maxillule apex; $j$, maxilliped article 3 ; $k$, maxilla apex; $l$, mandible; $m$, antennule; $n$, antenna; $o$, antenna, terminal articles; $p$, maxilliped. (Scale line $=5.0 \mathrm{~mm}$.)


FIgure 10.-Glossobius anctus ( $a-g, q, 30.0 \mathrm{~mm}$, Japan, paratype, USNM 227110; $h-q, \sigma^{7}, 9.5 \mathrm{~mm}$, Western Australia, paratype, AM P35744): $a$, pleopod 1, exopod; b, pleopod 1, posterior view; $c$, pleopod 2; $d$, pleopod 3; e, pleopod 5, anterior view; $f$, pleopod 5, posterior view; $g$, uropod, in situ; $h$, habitus, dorsal view; $i$, pereon and pleon, lateral view; $j$, cephalon, ventral view; $k$, pereopod $1 ; l$, pereopod $7 ; m$, maxilliped; $n$, maxilla; $o$, mandible palp; $p$, maxillule apex; $q$, penes; $r$, uropod. (Scale line $=2.0 \mathrm{~mm}$.)
basis with moderately developed expansions. Penes elongate.
Color.-Pale to tan in alcohol.
SIZE.-Ovigerous females between 25.5 and 34.0 mm , one non-ovigerous female was 26.0 mm , males between 8.0 and 9.5 mm .
REMARKS.-This species is readily identified by the large coxal plates, large labrum, and uropods with both rami bluntly rounded and of approximately equal length.
HosT.-Recorded only from Euleptorhamphus viridis.
DISTRIBUTION.-Tropical and subtropical Pacific and Indian Ocean, with records from Hawaii, Japan, and eastern and western Australia.

## Glossobius auritus Bovallius, 1885

## Figures 11, 12

Glossobius auritus Bovallius, 1885:12-17, pl. 3: figs. 24-33.-Stebbing, 1893:354, pl. 15.
Ceratothoa laticauda.-Gerstaecker, 1882:258.-Richardson, 1904:23.Trilles, 1973b:1252, pl. 2: figs. 12, 13.-Kurochkin, 1980:289.
Glossobius laticauda.-Stebbing, 1893:354.-Van Name, 1936:490.-Brian and Dartevelle, 1949:181.-Trilles, 1979:258.
Codonophilus laticauda.-Nierstrasz, 1931:131.
Codonophilus auritus.-Nierstrasz, 1931:132.
Glossobius laticaudus.-Avdeev, 1982b:66.
Glossobius albinae Kononenko, 1986:331.
Not Cymothoa laticauda Milne-Edwards, 1840:274 [nomen dubium].
Not Ceratothoa crassa Dana, 1853:753-754, pl. 50: figs. 2a,b,b',c.
MAterial-Pacific ocean: Thailand: Gulf of Thailand, off Paknam, mouth of flying fish Cypselurus sp., coll. H.M. Smith, 6 Aug 1953, ovigerous $q, 21.9 \mathrm{~mm}$ and $\sigma^{7}, 8.4$ mm (USNM 216605). Japan: Off Honshu Island, ex Cypselurus agoo (Temminck and Schlegel), coll. Jordan and Snyder, Stanford University Expedition, ca. 1900, ovigerous $q, 26.0$ mm (AM P35746). Mexico: Off Mexico, $15^{\circ} 36^{\prime} \mathrm{N}, 98^{\circ} 33^{\prime} \mathrm{W}$, from stomach of Coryphaena hippurus Linnaeus, coll. W.L. Klaw, 31 Mar 1957, ovigerous $\rho, 26.0 \mathrm{~mm}$ (USNM 104866). atlantic ocean: Bahamas: 3 miles west of Bimini, ex Cypselurus comatus (Mitchill), 23-24 Jul 1963, ovigerous $\rho$, 29.5 mm and $\sigma^{\prime \prime}, 12.2 \mathrm{~mm}$ (USNM 216576). Atlantic, $23^{\circ} 35^{\prime} \mathrm{N}, 68^{\circ} 18^{\prime} \mathrm{W}$, Shell Canada Tanker Pinnacles, coll. G.F. Burgess, 20 May 1955, non-ovigerous $\%, 24.5 \mathrm{~mm}$ (USNM 98987). Lesser Antilles: Saba Bank, $1 / 2$ mile north of $17^{\circ} 27^{\prime} \mathrm{N}, 63^{\circ} 13^{\prime} \mathrm{W}$, Smithsonian Bredin sta 108-56, ex Cypselurus comatus, coll. D.V. Nicholson, 1956, ovigerous $\rho$, 26.5 mm and $\sigma^{\prime \prime}, 10.5 \mathrm{~mm}$ (USNM 216580). $15^{\circ} 27^{\prime} \mathrm{N}$, $45^{\circ} 00^{\prime} \mathrm{W}$, mouth of flying fish on board Swedish vessel Monarch, coll. Capt. George von Scheile, ovigerous $\uparrow, 32.8$ mm and ơ, 10.5 mm (syntypes, RMS Isopod 4971). "Ost Indien," bought from Carl Wessel at Hamburg, ovigerous 9 , 28.5 mm (syntype, RMS Isopod 3475); several mancas (syntypes, RMS Isopod 3476). Southwest of Barbados, $12^{\circ} 35^{\prime} \mathrm{N}, 59^{\circ} 54^{\prime} \mathrm{W}$, Geronimo sta $7-23$, ex Cypselurus comatus, 6 Feb 1966, ovigerous $\rho, 26.0 \mathrm{~mm}$ and $\sigma^{\circ}, 10.2 \mathrm{~mm}$ (USNM 216579). Honduras: Off Honduras, $17^{\circ} 35$ 'N,
$82^{\circ}{ }^{\prime} 2^{\prime}$ W, Geronimo sta 6-218, ex Cypselurus comatus, 23 Oct 1965, ovigerous $q, 27.5 \mathrm{~mm}$ and $\sigma^{7}, 9.4 \mathrm{~mm}$ (USNM 216578). North of Cabo Falso, ex flying fish at 100 fathom line, 15 Jun 1972, ovigerous $\uparrow, 26.8 \mathrm{~mm}$ (USNM 216577).

TYPES.-Syntypes, Naturhistoriska Riksmuseet, Stockholm, Isopod nos. 4971, 3475, and 3476.
TyPE Locality.-North Atlantic Ocean at $15^{\circ} 26^{\prime} \mathrm{N}$, $45^{\circ} 00^{\prime} \mathrm{W}$.

Description of Female.-Body straight, between 2.06 and 2.41 times longer than maximum width, widest at pereonite 5 ; pereon with ill-defined longitudinal mediodorsal ridge. Cephalon lateral margins converge abruptly in front of eyes to form rostrum which appears truncate in dorsal view; eyes small, subtriangular, facets indistinct. Pereonite 1 longest, anterior margin convex, not recessed to accommodate cephalon; anterolateral margins form into 2 bosses which do not project anteriorly. Pereonites 2-4 subequal in length, pereonite 5 slightly shorter than 4 ; pereonites 6 and 7 markedly shorter than other pereonites, pereonite 7 about $15-18 \%$ length of pereonite 1. Coxae all shorter than pereonites; coxae of pereonites 6 and 7 smaller on one side than other, each with distinct acute dorsal ridge. Pleonites $1-4$ subequal in length; pleonite 5 slightly longer, posterior margin with 2 low submedian lobes. Pleotelson nearly as long as wide, lateral margins converging slightly; posterior margin truncate, with median emargination.

Antennule extending to posterior of cephalon, composed of 8 articles, first 3 of which are broad and flattened. Antenna extends to pereonite 1 , composed of 9 articles, first 4 of which are broad.

Mandible palp large, folding across anterior to labrum. Maxillule styliform, with 4 terminal spines. Maxilla lateral lobe with about 17 spines, endopod with 8 . Maxilliped as in $G$. impressus, but article 3 with 2-4 spines.

Pereopod 1 short, merus posterior margin not dilated, dactylus extending to middle of carpus; pereopods 2 and 3 longer and less robust than pereopod 1. Pereopods 5-7 basis with posterior expansion well developed, anterior expansion weakly developed; merus with weak anterior expansion.

Pleopods as in G. impressus. Uropod curving medially, endopod slightly longer than exopod, both rami with apices narrowly rounded or acute.

DESCRIPTION OF MALE.-Much shorter than female ( $38 \%$ on average), body approximately rectangular in shape. Antennule with 6 or 7 articles, antenna with 8 or 9 . Maxilla lateral lobe with 9 spines, endopod with 6 . Maxilliped with 3 stout recurved terminal spines on article 3. Pereopods less robust than in female, pereopods 5-7 basis with weakly developed expansions. Penes elongate, apices lie between pleopod peduncles. Pleopods all simple; appendix masculina absent. Uropods similar to female, exopod slightly longer than endopod.

COLOR.-Anterior of animal and anterior appendages dark brown to nearly black, becoming lighter toward posterior; pleon, pleotelson, and pereopods 6 and 7 virtually without chromatophores.


FIgURE 12.-Glossobius auritus (a-f, ovigerous $9,26.5 \mathrm{~mm}$, Saba Bank, Lesser Antilles, USNM 216580; g-t, $\sigma^{\prime \prime}, 10.5 \mathrm{~mm}$, Saba Bank, Lesser Antilles, USNM 216580): $a$, pleopod 1, ventral view; $b$, pleopod 1, dorsal view; $c-e$, pleopods $2,4,5$, respectively; $f$, pleopod 5 , dorsal view; $g$, antennule; $h$, antenna; $i$, dorsal view; $j$, pereon and pleon, lateral view; $k$, maxilliped; $l$, maxilliped article 3; $m$, maxilla apex; $n$, penes; $o$, pereopod $1 ; p$, pereopod 7; $q-s$, pleopods $1,2,5$, respectively; $t$, uropod. (Scale line $=3.0 \mathrm{~mm}$.)

SIZE.-Ovigerous females between 21.9 and 32.8 mm , one non-ovigerous female was 24.5 mm , males between 8.4 and 12.2 mm .

REMARKS.-Identification of Glossobius auritus is not problematic. Glossobius impressus is far more elongate and has large bulbous lateral lobes on pereonites 1 and 2. Glossobius hemiramphi, so far found only on hemiramphid fishes, is also more elongate and has an acute rostrum, while G. anctus has exceedingly large coxae and a pleotelson nearly as long as wide.

There is a problem in deciding which name to use for this species. In the past this species has been referred to (in various genera) as Glossobius laticauda (Milne-Edwards, 1840). Glossobius crassa (Dana, 1853) and Glossobius auritus were placed in synonymy with C. laticauda. Milne-Edwards'(1840) description lacked figures, he gave no host, no locality, and there are apparently no types. Trilles (1973b) did not list any type material in the Paris Museum. Furthermore, the description of G. laticauda specifically disagrees with the material subsequently figured under that name and with the present material. Milne-Edwards (1840:274) stated, and we paraphrase, "First segment ... with large lamellar elongations which advance on either side of the head." This agrees more with certain species of Ceratothoa or Lobothorax than with Glossobius. As there is little chance of the identity of $G$. laticauda being resolved we here regard it as a nomen dubium.

Glossobius crassa was described by Dana (1853) from a single dry specimen about 28 mm in length, from the "Southwestern Pacific." Dana illustrated the body in dorsal view, pereopod 6, and the uropod. The very short uropodal rami and the proportions of pereonite 1 and the pleotelson clearly distinguish it from G. auritus.

We have examined the type material of Glossobius auritus and can positively identify the present material as belonging to that species.

We have not seen the recently described Glossobius albinae Kononenko, 1986, but Kononenko's figure agrees completely with the material examined here, and we have no hesitation in placing his species in synonymy with $G$. auritus.

HosTs.-In most records the host was not identified; in our material, where the host has been identified, it is Cypselurus comatus in the Atlantic and Cypselurus agoo in Japan. Kononenko (1986) recorded the host as Cheilopogon heterurus (=Cypselurus heterurus (Rafinesque)).

DISTRIBUTION.-Present material is from the Caribbean, tropical East Pacific, Japan, and Thailand. Trilles (1973b) summarizes previous records from the Pacific, Indian, and Atlantic Oceans. Kononenko's specimen was collected in the central South Atlantic ( $4^{\circ} 35^{\prime} \mathrm{S}, 22^{\circ} 30^{\circ} \mathrm{W}$ ).

## Glossobius hemiramphi Williams and Williams, 1985

Figures 13, 14
Glossobius hemiramphi Williams and Williams, 1985:147-152, figs. 1-26.

Ceratothoa impressa (Say).-Berkeley and Houde, 1978:636 [misidentification].

Material.-With one exception, all from Hemiramphus brasiliensis (Linnaeus).
western atlantic: Georgia: Off Georgia, M/V Theodore $N$. Gill cruise 3 , sta $36,31^{\circ} 40^{\prime} \mathrm{N}, 80^{\circ} 37^{\prime} \mathrm{W}$, non-ovigerous ¢, 11.8 mm and postmanca, 3.7 mm ; non-ovigerous $\rho, 9.7 \mathrm{~mm}$; juvenile, 4.9 mm ; non-ovigerous $9,10.4 \mathrm{~mm}$ and postmanca, 3.2 mm . Off Georgia, Gill cruise 7, sta $34,31^{\circ} 00^{\prime} \mathrm{N}, 80^{\circ} 58^{\prime} \mathrm{W}$, non-ovigerous $¢, 17.4 \mathrm{~mm}$. Florida: Off Florida, Gill cruise 7, sta $17,29^{\circ} 38^{\prime} \mathrm{N}, 79^{\circ} 36^{\prime} \mathrm{W}$, ovigerous $\rho, 15.5 \mathrm{~mm}$ and $\sigma^{\circ}, 5.4$ mm. New Smyrna Beach, ex USNM 57124, $\sigma^{7}, 7.3 \mathrm{~mm}$. Dade Co., coll. Steve Berkeley, non-ovigerous $\rho, 29.6 \mathrm{~mm}$ and $\sigma^{\circ}$, 9.7 mm ; $\%$ with mancas, 36.9 mm ; $\%$ with mancas, 25.5 mm , ovigerous $\varnothing, 27.8 \mathrm{~mm}, \sigma^{7}, 9.4 \mathrm{~mm}, \sigma^{7}, 9.6 \mathrm{~mm}$ ( 4 specimens in 1 vial); non-ovigerous $9,26.8 \mathrm{~mm}$ and $\sigma^{7}, 8.4 \mathrm{~mm}$. Key West, ex USNM 30948, $\varnothing$ with mancas, 28.2 mm and $\sigma^{\circ}, 7.9$ mm ; ex USNM 35044, non-ovigerous $q, 24.9 \mathrm{~mm}$ and $\sigma^{\circ}, 9.1$ mm; ex USNM 120519 , ovigerous $q, 27.4 \mathrm{~mm}$ and $\sigma^{7}, 8.4 \mathrm{~mm}$. Dry Tortugas, ex MCZ 5203, ovigerous $\uparrow, 27.2 \mathrm{~mm}$ and $\sigma^{\prime \prime}, 6.9$ mm. Bahamas: Grand Bahama Island, West End, ex Univ. Miami Marine Lab no. 11667, manca, 3.3 mm . Green Cay, non-ovigerous $\$, 5.2 \mathrm{~mm}$ and $\sigma^{\prime}, 3.7 \mathrm{~mm}$. Mexico: Quintana Roo, Puerto Morelos, ex USNM 50476, ovigerous $\boldsymbol{q}, 32.2 \mathrm{~mm}$. West Indies: Haiti, ex USNM 164798, ovigerous $q, 23.4 \mathrm{~mm}$ and $\sigma^{7}, 9.4 \mathrm{~mm}$. Puerto Rico, off Guanica Bay, $\%$ (holotype, USNM 212532), $\sigma^{7}$ (allotype, USNM 213533), $3 \phi$ and $5 \sigma^{7}$ (paratypes, USNM 213534-213541). St. Thomas, ex MCZ 5203 no .8 , ovigerous $\%, 29.2 \mathrm{~mm}$ and $\sigma^{\circ}, 8.9 \mathrm{~mm}$; ex MCZ 5203 no. 11, ovigerous $\rho, 28.6 \mathrm{~mm}$ and $\sigma^{\prime}, 9.9 \mathrm{~mm}$. St. Croix, ex BM(NH) 1863.8.7.20, ovigerous $9,30.2 \mathrm{~mm}$. Bermuda: Ex Hemiramphus bermudensis Collette, 1962, nonovigerous $\uparrow, 22.1 \mathrm{~mm}$ and $\sigma^{\prime \prime}, 6.8 \mathrm{~mm}$.

EASTERN ATLANTIC: Senegal: Dakar, Galathea sta 10, ex UZMC P-34190, ovigerous $\rho, 21.0 \mathrm{~mm}$ and $\sigma^{\circ}, 3.6 \mathrm{~mm}$; non-ovigerous $\varphi, 20.3 \mathrm{~mm}$ and $\sigma^{7}, 5.3 \mathrm{~mm}$; ovigerous $\uparrow, 17.4$ mm and $\sigma^{\circ}, 5.1 \mathrm{~mm}$; non-ovigerous $\phi, 19.0 \mathrm{~mm}$ and $\sigma^{\circ}, 5.2$ mm . Guinea: Off Guinea, $10^{\circ} 44^{\prime}-11^{\circ} 46$ N, $17^{\circ} 06^{\prime}-$ $16^{\circ} 58^{\prime} \mathrm{W}$, non-ovigerous $\varnothing, 10.3 \mathrm{~mm}$ and manca 2.8 mm . Sierra Leone: Freetown, ovigerous $\phi, 23.7 \mathrm{~mm}$ and $\sigma^{7}, 6.1$ mm ; non-ovigerous $q, 18.7 \mathrm{~mm}$ and manca, 2.9 mm ; $q$ with mancas, 19.7 mm and $\sigma^{\top}, 5.5 \mathrm{~mm}$; non-ovigerous $\uparrow, 16.4 \mathrm{~mm}$ and $\sigma^{\prime \prime}, 6.2 \mathrm{~mm}$; ovigerous $q, 17.3 \mathrm{~mm}$ and $\sigma^{\prime}, 5.4 \mathrm{~mm} ; ~ \%$ with empty marsupium, $15.1 \mathrm{~mm}, \sigma^{7}, 5.6 \mathrm{~mm}$, and manca, 2.9 mm ; non-ovigerous $\$, 17.7 \mathrm{~mm}$ and $\sigma^{\circ}, 4.7 \mathrm{~mm} ; \%$ with mancas, 16.2 mm and $\sigma^{7}, 6.0 \mathrm{~mm}$; non-ovigerous $\varphi, 18.6 \mathrm{~mm}$ and postmanca, 2.8 mm ; ovigerous $\rho, 16.3 \mathrm{~mm}$ and $\sigma^{\prime}, 5.3 \mathrm{~mm}$; $\rho$ with mancas, 16.3 mm and $\sigma^{\circ}, 5.1 \mathrm{~mm}$; non-ovigerous $q, 13.3$ mm and $\sigma^{\prime \prime}, 3.8 \mathrm{~mm}$. Liberia: Off Monrovia, non-ovigerous $q, 13.3 \mathrm{~mm}$ and $\sigma^{7}, 4.6 \mathrm{~mm}$; Atlantide sta 52 , immature $q, 7.8$ mm ; immature $\uparrow, 7.3 \mathrm{~mm}$; postmanca, 4.3 mm ; postmanca, 3.0 mm . Off Port Marshall, Atlantide sta 53; non-ovigerous $\rho, 13.0$ $\mathrm{mm} ; \sigma^{7}, 7.4 \mathrm{~mm}$; non-ovigerous $\%, 8.2 \mathrm{~mm}$. Ghana:


Figure 13.-Glossobius hemiramphi (q, a-c, Haiti; $d-h$, St. Thomas; $i-j$, Liberia; $k-p$, Accra): a, head and peronite 1, dorsal view; $b$, pereonite 1, lateral view; $c$, pleon and pleotelson, dorsal view; $d-j$, pereopods 1-7; $k-0$, pleopods 1-5; p, uropod.


Sekondi-Takoradi, ex UZMC P-341915, $\sigma^{7}, 4.8 \mathrm{~mm}$. Accra, Atlantide sta 77, ex UZMC P-341934, non-ovigerous $9,15.3$ mm and $\sigma^{7}, 5.1 \mathrm{~mm}$; ex UZMC P-341933, non-ovigerous $q$, 13.1 mm and $\sigma^{7}, 4.8 \mathrm{~mm}$; ex UZMC P-341936, $\odot$ with mancas, 25.0 mm . Angola: Luanda, Atlantide sta 137, non-ovigerous ¢, 11.8 mm ; non-ovigerous $\$, 9.6 \mathrm{~mm}$.

TYPES.-Holotype, USNM 213532; allotype, USNM 213533; 8 paratypes ( $3 \circ, 5 \sigma^{*}$ ), USNM 213534-213541, 23 paratypes in Williams and Williams collection. For measurements of type material see Williams and Williams (1985).

Type Locality.-One mile offshore of Guanica Bay, La Maruca Reef, Puerto Rico, $17^{\circ} 56.5^{\prime} \mathrm{N}, 66^{\circ} 54.5^{\prime} \mathrm{W}$.

DESCRIPTION OF FEMALE.-Length/width of $\rho$ with oostegites 2.8-3.6, of $q$ without oostegites 3.0-3.8. Width greatest at pereonite 6. Cephalon abruptly narrowed anterior to eyes into narrowly rounded triangular rostrum. Eyes small, oval. Pereonite 1 longest, anterior one-third to one-half of lateral margin elevated dorsally into ridge. Lengths of pereonites 2-6 gradually decreasing; pereonite 7 abruptly shorter (relative lengths in percent of pereonites $1-7$ of 27.8 mm 9 from Dade Country, Florida: $25.5,16.0,15.2,14.8,13.2,10.3,5.0)$. Coxae shorter than their pereonites; coxa 7 may have concave dorsal surface and lateral margin elevated into ridge. Pereonite 7 not overlapping any pleonites. Pleonites $1-4$ subequal in length; pleonite 5 about twice as long, wider than pleotelson. Pleotelson length/width $0.56-0.71$, narrowing slightly posteriorly, lateral margins slightly convex, posterior margin slightly concave.

Antennule composed of 7 articles, article 3 longest. Antenna composed of 7-8 articles, slightly longer than antennule, article 4 longest. Mouthparts as described by Williams and Williams (1985).

Pereopods similar to those of Glossobius impressus; pereopod 3 with longest dactyl.

Pleopods 2-5 with depressions separated by ridges (shown only for endopods in Figure 13l-o). Uropods reaching slightly beyond posterior margin of pleotelson; rami subequal in length
or exopod slightly longer.
Description of Male.-As described by Williams and Williams (1985).

COLOR.-Head and pereonites 1-4 covered with black chromatophores, sparser on pereonite 4. Pereonites 5-7, pleon, and pleotelson unpigmented.

SLZE.-Females with oostegites ranged in length from 15.1 to 36.9 mm (Figure 14). Western Atlantic specimens are distinctly longer than the Eastern Atlantic specimens. There is a slight overlap between 23 and 25.5 mm , and the second smallest specimen is a 15.5 mm female from Florida.

REMARKS.-The distinguishing features of $G$. hemiramphi are the obtusely pointed rostrum, pereonite 7 not overlapping any pleonites, pleon wider than pleotelson, and uropods extending beyond posterior margin of pleotelson.

Hemiramphus brasiliensis is limited to coastal waters, but has pelagic larvae that could be carried between the two populations by ocean currents. However, Collette (1965) found differences between the populations in numbers of gill rakers on the first and second arches and in dorsal and anal fin ray counts suggesting some restriction of gene flow. Glossobius hemiramphi is not known to infest larval halfbeaks, and the free-swimming juvenile stage of the isopod is probably too short-lived and vulnerable to predators to allow transport by ocean currents between the two host populations. It seems probable that the two populations are effectively isolated from each other, with little possibility of gene exchange between them.

Hosts.-Hemiramphus brasiliensis (Linnaeus) and H. bermudensis Collette.

Distribution.-Western Atlantic: Georgia, Florida, Bermuda, Bahamas, Haiti, Puerto Rico, Virgin Islands, Yucatan Peninsula, Mexico. Eastern Atlantic: From Dakar, Senegal, south to Luanda, Angola. The host, H. brasiliensis, has a wider known distribution, extending north to the Cape Verde Islands in the Eastern Atlantic, and from Woods Hole, Massachusetts, to Rio de Janeiro in the Western Atlantic (Collette, 1965).

## Glossobius impressus (Say, 1818)

## Figures 15-17

Cymothoa impressa Say, 1818:397.-De Kay, 1844:48.
Ceratothoa linearis Dana, 1853:752, pl. 50: figs 1a-1d.-Gerstaecker, 1882:260.—Stebbing, 1893:354.-Richardson, 1900:221; 1901:529.
Ceratothoa exocoeti Cunningham, 1871:499, pl. 59: fig. 5.-Gerstaecker, 1882:260.
Glossobius linearis.-Schioedte and Meinert, 1883:301, pl. 12: figs. 1-9.Hansen, 1895:18, pl 2: figs. 2-2d.-Brian and Dartevelle, 1949:176.
Ceratothoa impressa.-Richardson, 1905:234-236, figs. 236-240; 1913:2, 6.-Fowler, 1912:292-293, pl. 83.-Tattersall, 1921:214.-Stephensen, 1948:43-44, fig. 9.-Schultz, 1969:155-156, fig. 233.-Anonymous, 1971:16-17.-Lincoln, 1971a:103-104, photograph facing p. 90; 1971b:184.-Trilles, 1972:7-9, figs. 3-24, photographs 5, 6; 1973b:12531255, pl. 2: figs. 14-16.-Bowman, 1978:217.—Kussakin, 1979:287, fig. 153.-Kurochkin, 1980:289.

Meinertia impressa.-Nierstrasz, 1915:89-90; 1918:119.
Codonophilus impressus.-Nierstrasz, 1931:131.
Glossobius impressa.-Avdeev, 1981:1160, 1164; 1982a:70.
Glossobius impressus.-Avdeev, 1982b:65-67, fig. 2, no. 4; 1985:89.
Not Ceratothoa impressa.-Berkeley and Houde, 1978:636 [= Glossobius hemiramphi Williams and Williams, 1985].

MATERIAL_-ATLANTIC OCEAN: $60^{\circ} 42^{\prime} \mathrm{N}, 70^{\circ} 00^{\prime} \mathrm{W}$, from (stomach ?) Thunnus albacares (Bonnaterre), R/V Delaware, 3 Oct 1957, non-ovigerous $\circ+29.9 \mathrm{~mm}$ (USNM 216583). New Jersey: Off Cape May, ovigerous $\uparrow, 24.5 \mathrm{~mm}$ (holotype, ANSP 1608). $33^{\circ} 20^{\prime} \mathrm{N}, 59^{\circ} 50^{\prime} \mathrm{W}$, from mouth of flying fish, coll. Dr. R.P. Campbell, 1954, ovigerous $q$, approximately 28 mm , cephalon missing, and $\sigma^{0}, 11.0 \mathrm{~mm}$. West Indies: Anguilla, Bat Caves, electric light off Crocus Bay, sta 58-56, from T. albacares, 13 Apr 1958, immature $\circ, 19.8 \mathrm{~mm}$ (USNM 216582). North Atlantic: No other data, fish catalog no. 7272 and 6296, ovigerous $9,34.5 \mathrm{~mm}$ (USNM 216584). $6^{\circ} 1^{\prime} \mathrm{N}, 41^{\circ} 31^{\prime} \mathrm{W}, 77 \mathrm{~m}$ depth, 2020-2355 hrs, Isaacs-Kidd midwater trawl, field no. RHB 961, Chain Cruise No. 35, ex Hirundichthys speculiger (Valenciennes) (MCZ 250414), coll. Dr. G.W. Mead, 12 Feb 1963, non-ovigerous $\circ, 36.0 \mathrm{~mm}$ (USNM 216581). Brazil: North of Sâo Luis (= Maranhâo), $2^{\circ} 23^{\prime} \mathrm{N}, 39^{\circ} 38^{\prime} \mathrm{W}$, ex flying fish on board S.S. Vasari, coll. S.G. Davis, no date, non-ovigerous $q, 38.5 \mathrm{~mm}$ (USNM 46107). Off Rio de Janeiro, from mouth of flying fish Exocoetus sp., fish catalog no. 49090 , coll. J.B. Hatcher, 1897, ovigerous $q, 33.5 \mathrm{~mm}$ and $\sigma^{\prime}, 8.3 \mathrm{~mm}$ (USNM 22797). West Africa: Senegal, Dakar Harbor, no host, Geronimo sta 4-123, 5 Sep 1964, non-ovigerous $\circ, 27.9 \mathrm{~mm}$ (USNM 119491). $7^{\circ} 20^{\prime} \mathrm{N}, 25^{\circ} 20^{\prime} \mathrm{W}$, catalogued 1884 , don. W. Hower, ovigerous $\uparrow, 40.0 \mathrm{~mm}(\mathrm{BM}(\mathrm{NH})) .5^{\circ} 0^{\circ} \mathrm{S}, 27^{\circ} 15^{\prime} \mathrm{W}$, Terra Nova Expedition, ovigerous $\uparrow$, approximately 40.0 mm , pleotelson detached (BM(NH) 1921.11.29.150). $10^{\circ} 48^{\prime} \mathrm{S}, 2^{\circ} 07^{\prime} \mathrm{W}$, during voyage of Northumberland from Durban to Tenerife, ovigerous $q, 35.0 \mathrm{~mm}$ and $\sigma^{2}, 8.5 \mathrm{~mm}$ (BM(NH)). No data except "from flying fish," don. A.M. Norman, non-ovigerous $\uparrow, 34.0$ mm (BM(NH)).

Types.- Holotype, Academy of Natural Sciences, Philadelphia, Pennsylvania, USA, ANSP 1608.

Type Locality.-Cape May, New Jersey.

DESCRIPTION OF FEMALE.-Body from 3.0 to 3.5 times longer than wide, sides subparallel. Cephalon lateral margin converging slightly toward anterior, abruptly narrowed in front of eyes to form broad rostrum, appearing rounded or truncate in dorsal view, more acute in anterior perspective; eyes lateral, small, narrow, subtriangular, facets indistinct. Pereonite 1 longest, pereonites 2-4 subequal in length, slightly shorter than 1 , pereonite 5 shorter than 4 ; pereonites 6 and 7 markedly shorter than other pereonites; pereonite 7 about $20 \%$ length of pereonite 1. Pereonite 1 not encompassing cephalon, anterior margin straight, not sinuate or lobed; lateral margins produced laterally to form bulbous lobe with lateral laminar flange; pereonite 2 with similar flanged lobe formed from coxae. Coxae of pereonites $2-5$ far shorter than segment, coxae of pereonites 6 and 7 nearly as long as segment, those of pereonite 7 shorter on one side than on other. Pleonites of about equal length, pleonite 1 very narrow, pleonites $2-5$ subequal in width. Pleotelson approximately rectangular, lateral margins nearly straight; posterior margin with median emargination, dorsal surface with median longitudinal depression.
Antennule composed of 7 articles, 1-3 broad, flattened, tending to fuse on dorsal surfaces. Antenna with 8 articles, 1-5 broad and flattened. Mandible palp large, 3 -articled, folding across anterior to labrum. Maxillule slender, styliform, with 3-4 terminal spines. Maxilla broad, folded (Figure 16d) or simple (Figure 16 m ); lateral lobe with 12 (Figure $16 d$ ) or 20 (Figure 16 m ) spines, endopod with $9-11$ spines. Maxilliped set distinctly posterior to other buccal appendages, with 2 large laminar oostegital lobes, proximal part of basal article fleshy, swollen; article 3 rounded, with 2 small spines.
Pereopod 1 short, merus weakly expanded posteriorly, dactylus extending to middle of carpus, pereopods 2 and 3 longer than 1 , less robust, articles proportionally longer, dactylus much longer than in pereopod 1 , extending to posterior of merus, pereopods 5-7 with merus anterior margin slightly dilated.
Pleopods all laminar, surfaces formed into fine nodular ridges except pleopod 1 endopod anterior surface which is smooth; peduncles narrow, without prominent lobes. Pleopod 1 largest, covering all other pleopods; pleopods becoming progressively smaller toward posterior; all rami (except pleopod 1 endopod) with depressions, in some cases forming folds or pockets. Pleopods 3-5 endopods with weakly developed proximomedial lobe. Uropods short, usually held under pleotelson posterior margin; exopod distinctly shorter than endopod.

Description of Male.-Very much smaller than female, body rectangular in shape, lacking distinctive pereonal morphology of female. Antennule with 7 articles, antenna with 9, neither with flattened articles. Mandible palp with setae; maxilla lateral lobe with 4 spines, endopod with 2 . Maxilliped article 3 narrow, with 2 recurved terminal spines.

Pereopods similar to female but pereopod 1 not as robust, posterior pereopods with expansions of basis less developed.


FIGURE 15.-Glossobius impressus ( $a, g, h$, ovigerous $\%, 24.5 \mathrm{~mm}$, Cape May, New Jersey, holotype, ANSP 1608; b, $c, f-n$, non-ovigerous $\boldsymbol{q}, 36.0 \mathrm{~mm}$, Chain sta. 35; $d$, ovigerous $\$, 40.0 \mathrm{~mm}$, Terra Nova Expedition; $e$, non-ovigerous $¢, 38.5 \mathrm{~mm}$, Maranhâo, Brazil): $a$, dorsal view; $b$, dorsal view; $c$, cephalon, ventral view; $d$, cephalon and pereonite 1 , dorsal view; $e$, cephalon and pereonite 1 , dorsal view; $f$, pleon, ventral view of right side; $g$, cephalon, dorsal view; $h$, rostrum, dorsal view; $i$, pereopod $1 ; j$, pereopod $3 ; k$, lateral view; $l$, pereopod $6 ; m$, pereopod 7, lateral view; $n$, pereopod 7, medial view. (Scale line $=8.0 \mathrm{~mm}$.)


Figure 16.-Glossobius impressus ( $a-g, i-l, n-t$, non-ovigerous $q, 36.0 \mathrm{~mm}$, Chain sta $35 ; h, m$, ovigerous $q$, 34.5 mm , USNM 216584): $a$, maxilliped; $b$, maxilliped article 3; $c$, maxillule; $d$, maxilla apex; $e$, maxilla; $f$, mandible; $g$, uropod; $h$, maxilliped; $i$, antennule, dorsal view; $j$, antenna, dorsal view; $k$, antennule, dorsal view; $l$, antenna, dorsal view; $m$, maxilla apex; $n$, pleopod $1 ; o$, pleopod 1 , dorsal view; $p$, pleopod $2 ; q$, pleopod 3 ; $r$, pleopod 4; $s$, pleopod 5; $t$, pleopod 5 endopod, posterior view.


FIGURE 17.-Glossobius impressus ( $\sigma^{7}, 8.3 \mathrm{~mm}$, Rio de Janeiro, USNM 22797): $a$, dorsal view; $b$, pereon and pleon, lateral view; $c$, cephalon, dorsal view; $d$, buccal region; $e$, antennule; $f$, antenna; $g$, uropod; $h$, mandible; $i$, maxilliped; $j$, maxilla apex; $k$, maxilliped article $3, l$, penes; $m$, pereopod $1 ; n$, pereopod $7 ; o-r$, pleopods $1-3,5$, respectively. (Scale line $=\mathbf{3 . 0} \mathrm{mm}$.)

Penes elongate, apices lie between pleopod peduncles. Pleopods all simple, appendix masculina absent. Uropod exopod slightly longer than endopod.

COLOR.-Specimens all with cephalon, pereonites 1-3 and appendages appearing dark brown to nearly black, sometimes gradually lightening toward posterior; pleon, pleotelson, and pereopods 6 and 7 virtually without chromatophores.

Size.-Ovigerous females range from $24.5-40.0 \mathrm{~mm}$, non-ovigerous females from 27.9-38.5 mm, males between 8.3 and 11.0 mm .

VARIATION.-There is considerable variation present in the detailed shape of the lateral extensions of pereonites 1 and 2 , the blade like flanges varying from a large scoop to a weakly developed ridge. There is a substantial change in the appearance of the maxilliped between ovigerous and nonovigerous females, that of the non-ovigerous female being far more heavily covered with scales and spinules and having the 2 spines on article 3 more prominent. The maxilla of some non-ovigerous females was folded and lobed (Figure 16d) and had fewer spines on the lateral lobe (12) than did ovigerous females (20).

REMARKS.-No figure has ever been published of the holotype, a dried specimen; a dorsal view of it is given here (Figure $15 a$ ). This species can be easily identified by the prominent lateral lobes on pereonites 1 and 2.

Hosts.- Of the material examined only one is from an identified host, Hirundichthys speculiger. Other records from "Espadon" (swordfish, Xiphius gladius Linnaeus) (Trilles, 1972) or Thunnus albacares (present material) are almost surely from exocoetid fishes eaten by those fishes. One immature pair of specimens (USNM 216585) that we could not identify was taken from a specimen of Hirundichthys affinis (Günther) collected in the Philippines.

DISTRIBUTION.-Tropical to subtropical Atlantic, but not yet recorded from the Caribbean. Trilles (1972) recorded this species from New Caledonia, and Nierstrasz (1915) reported it from Makassar Strait. Two records from higher latitudes need to be confirmed by more collections before they can be accepted: Køge Bugt, just south of Copenhagen (Stephensen, 1948); south of the Cape of Good Hope, $52^{\circ} 58^{\prime} \mathrm{S}, 22^{\circ} 58^{\prime} \mathrm{E}$ (Nierstrasz, 1918). Cymothoids probably do not occur in such cold waters.

## Literature Cited

Anonymous
1971. Fish Parasite, South Atlantic Ocean. Marine Observer (London), 41:16-17.
Avdeev, V.V.
1978. [Parasitic Isopods of the Family Cymothoidae (Crustacea, Flabellifera) from the Red Sea.] Biologiya Morya (Vladivostok), 1978(4):30--35. [In Russian.]
1981. [Crustaceans of the Family Cymothoidae (Isopoda): Mesoparasites of Fishes.] Zoologicheskii Zhurnal, 60(8):1160-1167. [In Russian.]
1982a. [Peculiarities of the Geographic Distribution and the History of Marine Isopod Fauna Formation (the Family Cymothoidae s. str.).] Parazitologiya, 16(1):69-77. [In Russian.]
1982b. [Some Ecological and Geographical Pecularities of Isopods of the Genus Glossobius, Parasites of Fishes of the World Ocean's Epipelagic Zone.] Biologiya Morya (Vladivostok), 1982(3):65-67. [In Russian.]
1985. Specific Features of the Distribution of Marine Parasitic Isopod Crustaceans of the Family Cymothoidae (Isopoda, Flabellifera). In W.J. Hargis, Jr., editor, Parasitology and Pathology of Marine Organisms of the World Ocean. National Oceanic and Atmospheric Administration Technical Report, National Marine Fisheries Service, 25:89-92.
Barnard, K.H.
1925. Contributions to the Crustacean Fauna of South Africa, No. 9: Further Additions to the List of Isopoda. Annals of the South African Museum, 20(5):381-412.
Berkeley, S.A., and E.D. Houde
1978. The Biology of Two Exploited Species of Halfbeaks, Hemirhamphus brasiliensis and H. balao from Southeast Florida. Bulletin of Marine Science, 28(4):624-644.
Bianconi, J.J.
1870. Specimina zoologica Mosambicana, Fasc. XVII. Memorie dell'Accademia delle Scienze dell'Istituto di Bologna, series 2, 9(2):199-222, plates 1-4.
Bleeker, P.
1857. Recherches sur les Crustacés de l'Inde Archipélagique, II: Sur les Isopodes Cymothoadiens de l'Archipel Indien. Natuurkundige Vereeniging in Nederlandsche-Indie, Batavia, 2:20-40, plates 1, 2.
Bovallius, C.
1885. New or Imperfectly Known Isopoda, I: Bihang till Kongliga Svenska Vetenskapsakademiens Handlingar, 10(11):1-32, plates 1-5.
Bowman, T.E.
1978. Nomenclatural Problems in the Cymothoid Isopod Genera Ceratothoa, Codonophilus, Glossobius and Meinertia: Their Solution by Applying the Law of Priority. Crustaceana, 34:217-219.
Brian, A., and E. Dartevelle
1949. Contribution à l'étude des Isopodes marins et fluviatiles du Congo. Annales du Musée du Congo Belge, C (Zoologie), series 3(3), volume 1, fascicle 2:77-208.
Bruce, N.L.
1986. Revision of the Isopod Crustacean Genus Mothocya Costa, in Hope, 1851 (Cymothoidae: Flabellifera), Parasitic on Marine Fishes. Journal of Natural History, 20(5):1089-1192.
Bruce, N.L., and E.B. Harrison-Nelson
1988. New Records of Fish Parasitic Marine Isopod Crustaceans (Cymothoidae, Anilocrinae) from the Indo-West Pacific. Proceedings of the Biological Society of Washington, 101(3):585-602.

Brusca, R.C.
1981. A Monograph on the Isopoda Cymothoidae (Crustacea) of the Eastern Pacific. Zoological Journal of the Linnean Society, 73(2):117-199.
Collette, B.B.
1965. Hemiramphidae (Pisces, Synentognathi) from Tropical West Africa. Atlantide Report, 8:217-235.
1974. The Garfishes (Hemiramphidae) of Australia and New Zealand. Records of the Australian Museum, 29(2):11-105.
Cunningham, R.O.
1871. Notes on the Reptiles, Amphibia, Fishes, Mollusca, and Crustacea Obtained during the Voyage of H.M.S. Nassau in the Years 1866-1869. Transactions of the Linnean Society of London, 27(4):465-502, plates 58-59.
Dana, J.D.
1852. On the Classification of the Crustacea Choristopoda or Tetradecapoda. American Journal of Science and Arts, series 2, 14(41):297316.
1853. Crustacea, Part II. In United States Exploring Expedition, 14:6891618, atlas, plates 1-99 [published in 1855].
De Kay, J.E.
1844. Zoology of New York or the New York Fauna, Part 6: Crustacea. In Natural History of New York, 1: 70 pages, 13 plates. Albany: Carroll and Cook.
Fowler, H.B.
1912. The Crustacea of New Jersey. Annual Report of the New Jersey State Museum for 1911, 2:29-650, plates 1-150.
Gerstaecker, A.
1882. Sechste Ordnung: Isopoda-Asseln. In H.G. Bronn, Klassen und Ordnungen des Thier-Reichs, 5(2):8-278.
Hale, H.M.
1926. Review of Australian Isopods of the Cymothoid Group, Part II. Transactions of the Royal Society of South Australia, 50:201-234.
Hansen, H.J.
1895. Isopoden, Cumaceen und Stomatopoden der Plankton-Expedition. In Ergebnisse der Plankton-Expedition der Humboldt-Stiftung, 2.G.c.:1-105, plates 1-8.

Haswell, W.A.
1881. On Some New Australian Marine Isopoda, Part 1. Proceedings of the Linnean Society of New South Wales, 5:470-481.
1882. Catalogue of the Australian Stalk- and Sessile-Eyed Crustacea. xxiv +324 pages. Sydney: The Australian Museum.
Hilgendorf, F .
1879. Die von Hm. W. Peters in Moçambique gesammelten Crustaceen. Monatsberichte der Königlich Akademie der Wissenschaften zu Berlin, Physikalisch-mathematischen Klasse, 1878 (25 No-vember):782-851, plates 1-4.
Kensley, B.
1978. Guide to the Marine Isopods of Southern Africa. 173 pages. Cape Town: South African Museum.
Koelbel, C.
1879. Uber einige neue Cymothoiden. Sitzungsberichte der MathematischNaturwissenschafllichen Klasse der Kaiserichen Akademie der Wissenschaften, Wien, 78(2):401-416, plates 1, 2.
Kononenko, A.F.
1986. [A New Parasitic Isopod of Flying Fishes, Glossobius albinae sp. n. (Isopoda, Cymothoidae), from the Atlantic.] Parazitologiya,

20(4):329-332. [In Russian.]
Kurochkin, Y.V.
1980. [On the Parasitic Fauna of Flying Fishes (Family Exocoetidae) of the World Ocean.] Trudy Instituta Okeanologii, 97:278-295. [In Russian.]
Kussakin, O.G.
1979. [Marine and Brackish Water Isopods (Isopoda) of Cold and Temperate Waters of the Northern Hemisphere, I: Suborder Flabellifera.] Opredeliteli po Faune SSSR, 122:1-472. [In Russian.]
Lincoln, R.J.
1971a. Postscript. Marine Observer, 41(233):103-104.
1971b. Isopod Fish Parasites. Marine Observer, 41(234):184-186, figures 1-5.
Menzies, R.J.
1962. The Lund University Chile Expedition, 1948-1949, No. 42: The Zoogeography, Ecology and Systematics of the Chilean Marine Isopods. Lunds Universitets Arskriffer, n.f., avd 2, bd 57, nr 11, pages 1-162.
Miers, E.J.
1877. On a Collection of Crustacea, Decapoda, and Isopoda, Chiefly from South America, with Descriptions of New Genera and Species. Proceedings of the Zoological Society of London, 1877, 43:652679, plates 66-69.
Milne-Edwards, H .
1840. Histoire naturelle des Crustacés comprenent l'anatomie, la physiologie et la classification de ces animavx. Volume 3, 638 pages, 42 plates. Paris.
Nierstrasz, H.F.
1915. Die Isopoden-sammlung in Naturhistorischen Reichsmuseum zu Leiden, 1: Cymothoidae. Zoologische Mededeelingen, 1:71-108, plates 3,4 .
1918. Alte und neue Isopoden. Zoologische Mededeelingen, 4:103-142, plates 9,10 .
1931. Die Isopoden der Siboga Expedition, 3: Isopoda Genuina, 2: Flabellifera. Siboga-Expeditie, 32c:123-233, plates 10, 11.
Oto, A.W.
1828. Beschreibung einiger neven in den Jahren 1818 und 1819 im Mittelländischen Meere gefundener Crustaceen. Nova Acta Academiae Caesareae Leopoldino Carolinae, 14:329-354, plates 20-22.
Pillai, N.K.
1954. A Preliminary Note on the Tanaidacea and Isopoda of Travancore. Bulletin of the Central Research Institute, University of Travancore, Trivandrum, 3C:1-21.
1964. Parasitic Isopods of the Family Cymothoidae from South Indian Fishes. Parasitology, 54:211-223.
Richardson, H.
1900. Synopses of North American Invertebrates, VIII: The Isopoda, Part I: Chelifera, Flabellifera, Valvifera. American Naturalist, 34(399):207-230.
1901. Keys to the Isopods of the Atlantic Coast of North America, with Descriptions of New and Litule-Known Species. Proceedings of the United States National Museum, 23:493-579.
1904. Contribution to the Natural History of the Isopoda. Proceedings of the United States National Museum, 27:1-89.
1905. A Monograph on the Isopods of North America. Bulletin of the United States National Museum, 54:1-727.
1910. Marine Isopods Collected in the Philippines by the U.S. Fisheries Steamer "Albatross" in 1907-1908. U.S. Bureau of Fisheries Document, 736:1-44.
1913. Crustacés Isopodes. In Deuxième Expédition Antarctique Française (1908-1910) commandé par le Dr. Jean Charcot, pages 1-24. Paris: Masson et Cie.
Say, T
1818. An Account of the Crustacea of the United States. Journal of the Academy of Natural Sciences of Philadelphia, volume 1, part 2, numbers 5, 6:374-401.

Schioedte, J.C., and Fr. Meinert
1883. Symbolae ad monographium cymothoarum crustaceorum isopodum familiae, III: Saophridae, IV: Ceratothoinae. Naturhistorisk Tidsskrift, series 3, 13:281-378, plates 11-16.
Schultz, G.A.
1969. How to Know the Marine Isopod Crustaceans. vii +354 pages. Dubuque, Iowa: Wm. C. Brown.
Stebbing, TR.R.
1893. A History of Crustacea: Recent Malacostraca. xvii +466 pages. London: Kegan, Paul and Trench.
1900. On Crustacea Brought by Dr. Willey from the South Seas. In A. Willey, editor, Zoological Results Based on Material from New Britain, New Guinea, Loyalty Islands, and Elsewhere, Collected during the Years of 1895, 1896 and 1897, 5(33):605-690. Cambridge: University Press
1910a. No. VI: Isopoda from the Indian Ocean and British East Africa. In The Percy Sladen Trust Expedition in the Indian Ocean in 1905, under the Leadership of Mr. J. Stanley Gardiner, Vol. 1. Transactions of the Linnean Society of London (Zoology), 14:83-123, plates 5-10.
1910b. General Catalogue of South African Crustacea (Part V of South African Crustacea, for the Marine Investigations in South Africa). Annals of the South African Museum, 6(4):281-593, plates 15-22. [Plates 41-48 of Crustacea.]
1911. Indian Isopods. Records of the Indian Museum, 6(4):179-191,plates 10-12.
Stephensen, K.
1948. Storkrebs, IV: Ringkrebs, 3: Tanglus (Marine Isopoder) og Tanaider. In Danmarks Fauna, 53:1-187.
Tatersall, W.M.
1921. Tanaidacea and Isopoda. In British Antarctic "Terra-Nova" Expedition 1910, Zoology, 3(8):191-258.
Trilles, J.-P.
1972. Sur quatre isopodes cymothoides du Pacifique (Nouvelle Caledonie). Cahiers de l'Office de la Recherche Scientifique et Technique Outre-Mer (Océanographie), 10:3-17.
1973a. Les Cymothoidae (Isopoda, Flabellifera) des côtes françaises (Systematique, faunistique écologie et répartition g6ographique), I: Les Ceratothoinae Schioedte and Meinert, 1883. Bulletin du Muséum National d'Histoire Naturelle, Paris, series 3, 91 (Zoologie 70):1191-1230.

1973b. Les Cymothoidae (Isopoda, Flabellifera) du Museum national d'Histoire naturelle de Paris. Étude critique accompagné de précisions en particulieur sur la répartition géographique et l'Ecologie des différentes espèces représentées, I: Les Ceratothoinae Schioedte et Meinert, 1883. Bulletin du Muséum National d'Histoire Naturelle, Paris, series 3, 91 (Zoologie 70):1231-1267.
1975. Les Cymothoidae (Isopoda, Flabellifera) des côtes françaises, II: Les Anilocridae Schioedte et Meinert, 1881, Genres Anilocra Leach, 1818, et Nerocila Leach, 1818. Bulletin du Muséwm National d'Histoire Naturelle, Paris, series 3, 290 (Zoologie 200):347-378.
1979. Les Cymothoidae (Isopoda, Flabellifera; parasites de poissons) du Rijksmuseum van Natuurlijke Historie de Leiden II: Afrique, Amérique et régions Indo-Ouest-Pacifiques. Zoologische Mededelingen, 54(17):245-275.
1986. Les Cymothoidae (Crustacea, Isopoda, Flabellifera) d'Afrique. Bulletin du Muséum National d'Histoire Naturelle, Paris, series 4, 8(3):617-636.
Van Name, W.G.
1936. The American Land and Freshwater Crustacea. Bulletin of the American Musewm of Natural History, 21:1-535.
Williams, E.H., and L.B. Williams
1985. A New Cymothoid Isopod, Glassobius hemiramphi, from the Mouth of the Ballyhoo, Hemiramphus brasiliensis (Linnaeus) (Exocoetidae), in the Caribbean Sea. Crustaceana, 48(2):147-152.

## REQUIREMENTS FOR SMITHSONIAN SERIES PUBLICATION

Manuscripts intended for series publication receive substantive review (conducted by their originating Smithsonian museums or offices) and are submitted to the Smithsonian Institution Press with Form SI-36, which must show the approval of the appropriate authority designated by the sponsoring organizational unit. Requests for special treatment-use of color, foldouts, case-bound covers, etc.-require, on the same form, the added approval of the sponsoring authority.

Review of manuscripts and art by the Press for requirements of series format and style, completeness and clarity of copy, and arrangement of all material, as outlined below, will govern, within the judgment of the Press, acceptance or rejection of manuscripts and art.

Copy must be prepared on typewriter or word processor, double-spaced, on one side of standard white bond paper (not erasable), with $11 / 4^{\prime \prime}$ margins, submitted as ribbon copy (not carbon or xerox), in loose sheets (not stapled or bound), and accompanied by original art. Minimum acceptable length is 30 pages.

Front matter (preceding the text) should include: title page with only title and author and no other information, abstract page with author, title, series, etc., following the established format; table of contents with indents reflecting the hierarchy of heads in the paper; also, foreword and/or preface, if appropriate.

First page of text should carry the title and author at the top of the page; second page should have only the author's name and professional mailing address, to be used as an unnumbered footnote on the first page of printed text.

Center heads of whatever level should be typed with initial caps of major words, with extra space above and below the head, but no other preparation (such as all caps or underline, except for the underline necessary for generic and specific epithets). Run-in paragraph heads should use period/dashes or colons as necessary.

Tabulations within text (lists of data, often in parallel columns) can be typed on the text page where they occur, but they should not contain rules or numbered table captions.

Formal tables (numbered, with captions, boxheads, stubs, rules) should be submitted as carefully typed, double-spaced copy separate from the text; they will be typeset unless otherwise requested. If camera-copy use is anticipated, do not draw rules on manuscript copy.

Taxonomic keys in natural history papers should use the aligned-couplet form for zoology and may use the multi-level indent form for botany. If cross referencing is required between key and text, do not include page references within the key, but number the keyed-out taxa, using the same numbers with their corresponding heads in the text.

Synonymy in zoology must use the short form (taxon, author, year:page), with full reference at the end of the paper under "Literature Cited." For botany, the long form (taxon, author, abbreviated journal or book title, volume, page, year, with no reference in "Literature Cited") is optional.

Text-reference system (author, year:page used within the text, with full citation in "Literature Cited" at the end of the text) must be used in place of bibliographic footnotes in all Contributions Series and is strongly recommended in the Studies Series: "(Jones. 1910:122)" or ". . Jones (1910:122)." If bibliographic
footnotes are required, use the short form (author, brief title, page) with the full citation in the bibliography.

Footnotes, when few in number, whether annotative or bibliographic, should be typed on separate sheets and inserted immediately after the text pages on which the references occur. Extensive notes must be gathered together and placed at the end of the text in a notes section.
Bibliography, depending upon use, is termed "Literature Cited," "References," or "Bibliography." Spell out titles of books, articles, journals, and monographic series. For book and article titles use sentence-style capitalization according to the rules of the language employed (exception: capitalize all major words in English). For journal and series titles, capitalize the initial word and all subsequent words except articles, conjunctions, and prepositions. Transliterate languages that use a non-Roman alphabet according to the Library of Congress system. Underline (for italics) titles of journals and series and titles of books that are not part of a series. Use the parentheses/colon system for volume (number): pagination: "10(2):5-9." For alignment and arrangement of elements, follow the format of recent publications in the series for which the manuscript is intended. Guidelines for preparing bibliography may be secured from Series Section. SI Press.

Legends for illustrations must be submitted at the end of the manuscript, with as many legends typed, double-spaced, to a page as convenient.
Illustrations must be submitted as original art (not copies) accompanying, but separate from, the manuscript. Guidelines for preparing art may be secured from Series Section, SI Press. All types of illustrations (photographs, line drawings, maps, etc.) may be intermixed throughout the printed text. They should be termed Figures and should be numbered consecutively as they will appear in the monograph. If several illustrations are treated as components of a single composite figure, they should be designated by lowercase italic letters on the illustration; also, in the legend and in text references the italic letters (underlined in copy) should be used: "Figure $9 \mathbf{b}$." Illustrations that are intended to follow the printed text may be termed Plates, and any components should be similarly lettered and referenced: "Plate 9b." Keys to any symbols within an illustration should appear on the art rather than in the legend.
Some points of style: Do not use periods after such abbreviations as "mm, ft, USNM, NNE." Spell out numbers "one" through "nine" in expository text, but use digits in all other cases if possible. Use of the metric system of measurement is preferable; where use of the English system is unavoidable, supply metric equivalents in parentheses. Use the decimal system for precise measurements and relationships, common fractions for approximations. Use day/month/year sequence for dates: "9 April 1976." For months in tabular listings or data sections, use three-letter abbreviations with no periods: "Jan, Mar, Jun," etc. Omit space between initials of a personal name: "J.B. Jones."
Arrange and paginate sequentially every sheet of manuscript in the following order: (1) title page. (2) abstract, (3) contents, (4) foreword and/or preface, (5) text, (6) appendixes, (7) notes section, (8) glossary. (9) bibliography. (10) legends, (11) tables. Index copy may be submitted at page proof stage, but plans for an index should be indicated when manuscript is submitted.



[^0]:    Library of Congress Cataloging in Publication Data
    Bruce, Niel L.
    Species of the parasitic Isopod genera Ceratothoa and Glossobius (Crustacea: Cymothoidac) from the mouths of flying fishes and halfbeaks (Beloniformes) / Niel L. Bruce and Thomas E. Bowman.
    p. cm.-(Smithsonian contributions to zoology ; no. 489)

    Includes bibliographical references.
    Supt. of Docs. no.: SI 1.27:489

    1. Ceratothoa-Classification. 2. Glossobius-Classification. 3. Fishes-Parasites. I. Bowman, Thomas E. II. Title. III. Series.
    QL1.S54 no. 489
    [QL444.M34]
    591 s-dc20
    [595.3 ${ }^{\circ} 72$ ]
[^1]:    Niel L. Bruce, CSIRO, P.O. Box 89, East Melbourne, Victoria, 3002, Australia. Thomas E. Bowman, Department of Invertebrate Zoology, National Museum of Natural History, Smithsonian Institution, Washington, D.C. 20560.

