Surname				Centre Number		Candidate Number
Other Names						2
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wjec	A400U20-1		517-A400U20-1			duqas
cbac	BIOLOGY – A level component 2 Continuity of Life		Part of	WJEC		
	TUESDAY, 20 JUNE	E 2017 –	MORNIN	G		

2 hours

For Examiner's use only				
Question	Maximum Mark	Mark Awarded		
1.	15			
2.	18			
3.	17			
4.	17			
5.	11			
6.	13			
7.	9			
Total	100			

ADDITIONAL MATERIALS

In addition to this examination paper, you will need a calculator and a ruler.

INSTRUCTIONS TO CANDIDATES

Use black ink or black ball-point pen. Do not use gel pen. Do not use correction fluid.

Write your name, centre number and candidate number in the spaces at the top of this page.

Answer **all** questions.

Write your answers in the spaces provided in this booklet. If you run out of space, use the continuation pages at the back of the booklet, taking care to number the question(s) correctly.

INFORMATION FOR CANDIDATES

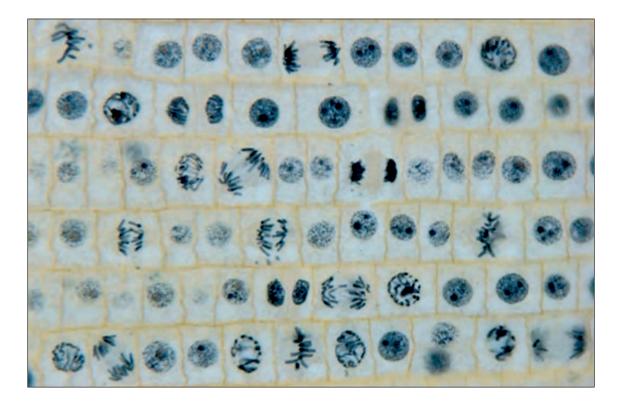
The number of marks is given in brackets at the end of each question or part-question.

The assessment of the quality of extended repsonse (QER) will take place in question **7**. The quality of written communication will affect the awarding of marks.



Answer all questions.

1. The photomicrograph below shows cells from a root tip undergoing cell division.



(a) (i) Name the type of cell division taking place in this tissue.

[1]

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(ii) Count the number of cells in metaphase and anaphase. Enter the values in the table below: [2]

.....

Phase	Number of cells	Time in minutes	% of cell cycle (by time)
Interphase	35	394	55
Prophase	9	101	14
Metaphase			
Anaphase			
Telophase/cytokinesis	10	112	16
Totals	64	720	100



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(iii)	Calculate the time taken for the two phases, using the formula.	Examir only
Time for a p	particular phase = $\frac{\text{number of cells in the phase}}{\text{total number of cells counted}} \times \text{total time taken for cell cycles}$	
	Enter the values in the table opposite.	[2]
(iv)	Calculate the percentage time spent in each phase.	
	Enter the values in the table opposite.	[2]
(b) Des	cribe two ways in which the process of cell division shown differs between plar animal cells.	nt cells [2]



In humans, unrestricted cell division can lead to the development of abnormal growths called What term is applied to the formation of malignant growths by unrestricted cell

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15

[1]

Studies have shown that the plasma membranes of cells in these growths have (ii) some transmembrane proteins with shorter carbohydrate side chains.

In the boxes below, draw diagrams of a short section of the plasma membrane from a normal and an abnormal cell. Include labels for the following structures: phospholipid bilayer, transmembrane protein and carbohydrate side chains. [3]

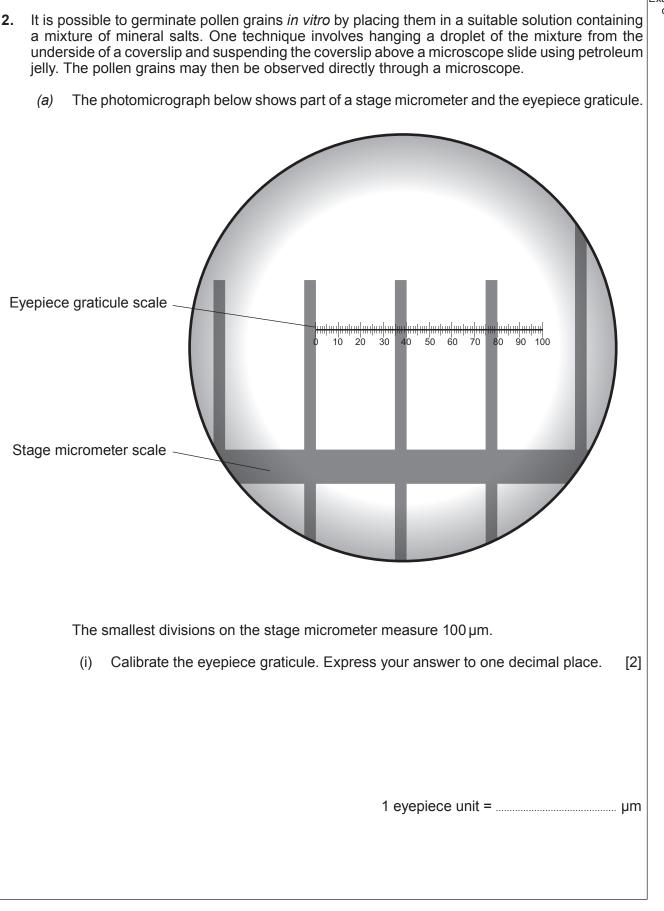
Normal	Abnormal
(iii)	How would the difference described in (ii) affect the properties of the abnormal cells?
······	
.	

tumours.

(i)

division?

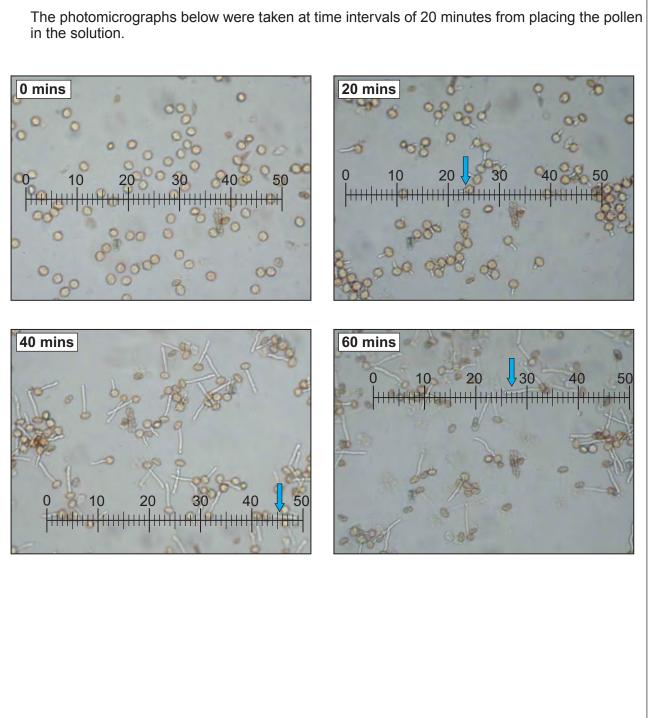
(C)



05

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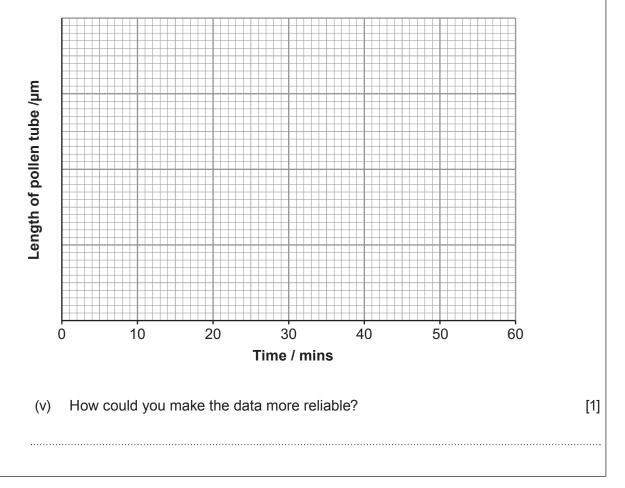


Examiner only (ii) Use the eyepiece graticule to measure the length of the pollen tube indicated by an arrow from each photomicrograph. Measure each pollen tube to the nearest whole number. **Enter the values in the table below.** [2]

Time / mins	Length of pollen tube / eyepiece graticule units	Actual length of pollen tube /μm
0	0	0
20		
40		
60		

(iii) Calculate the actual length of the pollen tubes in µm. Write your answers in the table above. [2]

(iv) Plot a graph of the results using a suitable scale for the y-axis.





[3]

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(b)	Using the technique described on page 5, design an experiment to investigate the effect of pH on the growth of pollen tubes.	Examiner only
	Give a brief description of your method. [4]	

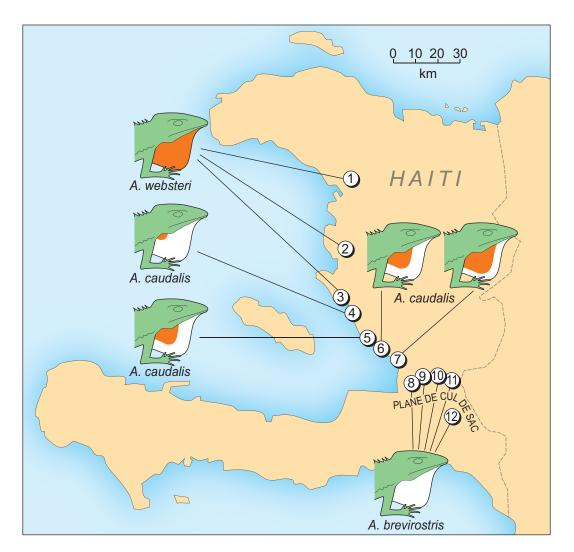
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Examiner only The photomicrographs below show pollen tubes. Photomicrograph \bf{A} shows a clear zone near the tip with an arrow; this is a vesicle. Electron micrograph \bf{B} shows organelles present in a region further back from the tip. Α mitochondrion rough endoplasmic reticulum vesicle Golgi body 0.5 µm 5µm Describe the roles of the vesicle and the labelled organelles in the passage of the pollen tubes through the style of a flower. [4] (C) A400U201 09 18



3. In the Caribbean there is a group of lizards called Anoles. The males possess a retractable flap of skin under the throat, termed a dewlap, which is used in courtship, aggressive interactions and even encounters with predators. The patterns of orange colouration on the dewlaps of three species of Anole lizard and their distribution in Haiti are shown below.



Zone	Sites	Habitat
North	1,2,3 Beach	
Central	4,5,6,7	Beach
South	8,9,10,11,12	Forest



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<i>(a)</i> (i) Describe	<i>(a)</i> (i) Describe the patterns of orange and white dewlap colouration for each species.				
Species	Pattern of orange and white				
Anolis brevirostris					
Anolis websteri					
Anolis caudalis					
(ii) Describe	how the three species are distributed between the three zones. [1]				
(iii) Describe	how the dewlap colouration varies from north to south in the central zone. [1]				



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		∃Examiner			
Breed Menc	ding experiments in the lab have revealed that dewlap colour-pattern is inherited as a lelian trait, in which orange (D) is dominant to white (d).	only			
The g	The gene pool of the <i>A.brevirostris</i> population contains no D alleles.				
	Hardy-Weinberg equations may be used to calculate the frequencies of the alleles D and d gene pool of the population of <i>A.websteri</i>				
p + q	= 1				
p ² + 2	$2pq + q^2 = 1$				
(b)	In the north zone population of 400 lizards, four were found to have white dewlaps.				
	Calculate the frequencies of the two alleles in this population, and calculate how many of the lizards with orange dewlaps were heterozygous. [4]				
••••••					



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(ii)	Explain why after many generations the frequency of allele D is zero in the <i>A.brevirostris</i> population but the frequency of allele d in <i>A.websteri</i> will never the zero.
The <i>l</i> colou	Anole lizards found at sites 3 and 4 are different species. Explain how their dewla uration may have resulted in this speciation.
	The <i>i</i>



4. In mice, coat colour is determined by genes at more than one locus.

Locus I.

This gene determines the colour of pigment. There are two major alleles: **B** coding for black pigment and **b** for brown.

Locus II.

This gene determines the distribution of pigment in the hair. Allele **A** produces a phenotype called agouti. The agouti phenotype is due to yellow bands on the otherwise dark hair shaft. In the non-agouti phenotype (determined by the allele **a**), the yellow bands are absent, so the hair shaft is a solid dark colour.

The table below shows the genotypes and corresponding phenotypes.

Locus I genotype	Locus II genotype	Phenotype (coat colour)
BB or Bb AA or Aa		Normal agouti
BB or Bb	аа	Solid black
bb AA or Aa		Cinnamon (brown with yellow bands)
bb	аа	Solid brown

(a) (i) Complete the genetic diagram below to show the expected outcome of crossing mice heterozygous at both loci. [4]

Parental phenotype	 х	
Parental genotype	 х	
Gametes	 х	



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A series of crosses between mice which were heterozygous at both loci produced 32 offspring. The results are shown in the table below. A null hypothesis was proposed that there was *'no significant difference between the observed and expected numbers of offspring'*.

-			n	
Phenotype	Observed O	Expected <i>E</i>		
Normal Agouti	22			
Solid black	4			
Cinnamon	3			
Solid brown	3			

(b) (i) Using the table provided, calculate χ^2 to test this null hypothesis.

$$\chi^2 = \sum \frac{(O-E)^2}{E}$$

(ii) Choose an appropriate probability level and circle the critical value for χ^2 in the table below: [1]

Degrees					Probability	/			
of freedom	0.9	0.8	0.7	0.5	0.2	0.1	0.05	0.02	0.01
1	0.016	0.064	0.15	0.46	1.64	2.71	3.84	5.41	6.64
2	0.21	0.45	0.71	1.39	3.22	4.60	5.99	7.82	9.21
3	0.58	1.00	1.42	2.37	4.64	6.25	7.82	9.84	11.34
4	1.06	1.65	2.20	3.36	5.99	7.78	9.49	11.67	13.28

(iii) State whether you accept or reject the null hypothesis, and explain why.

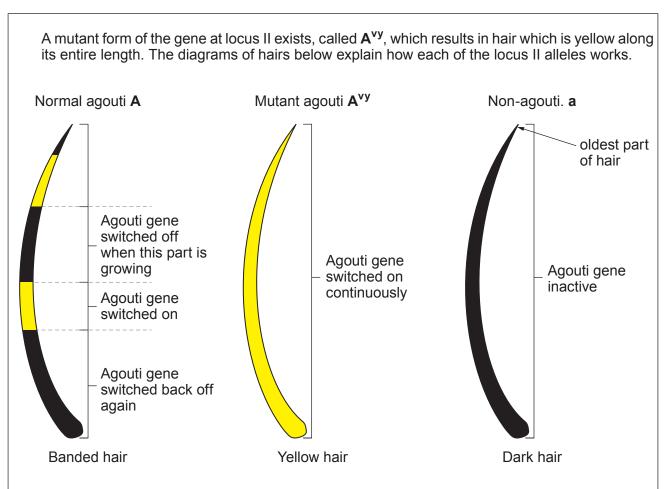


[2]

[3]

χ² =

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Scientists have bred a strain of mice with the genotype $A^{vy}a$.

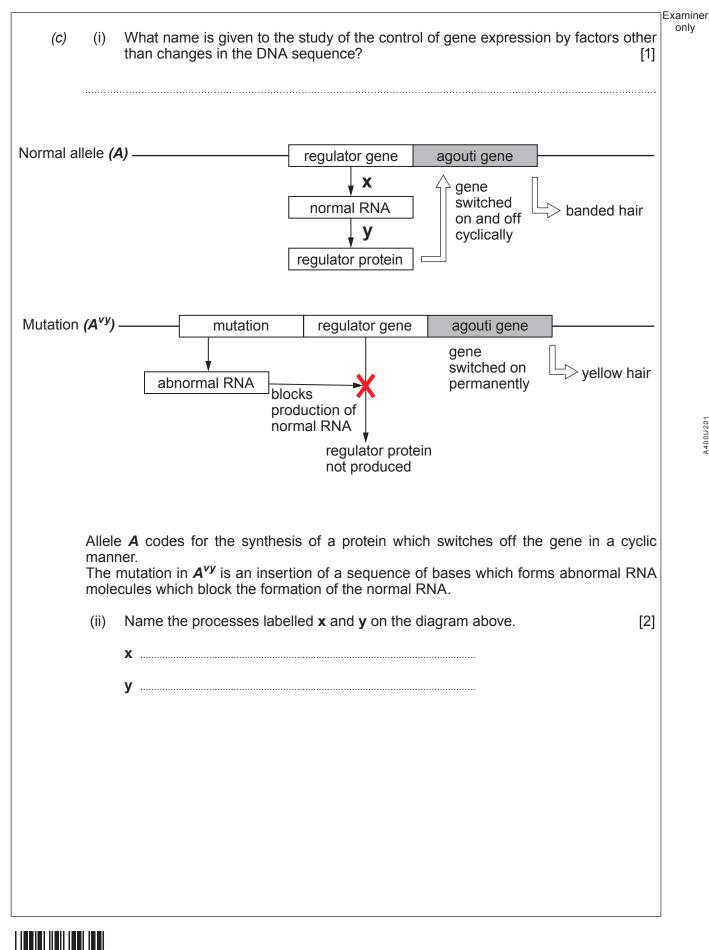
Since A^{vy} is dominant to **a**, all of the mice with genotype $A^{vy}a$ could be expected to be yellow.

In fact, some are yellow, some are normal agouti and some are shades in between.



These mice have identical DNA base sequences in the agouti gene locus but the gene is being expressed differently.

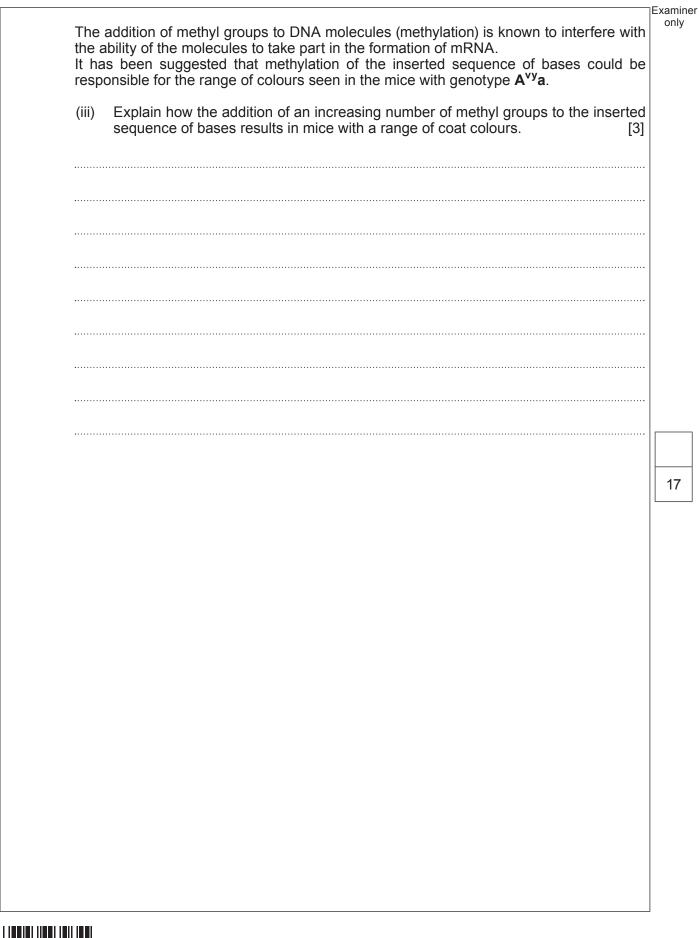




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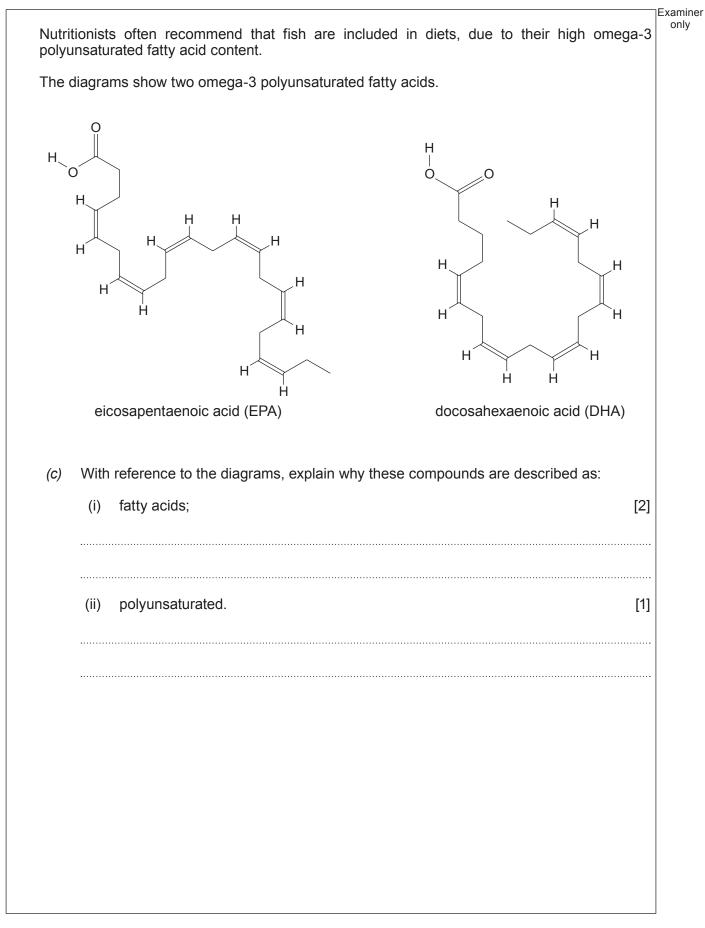
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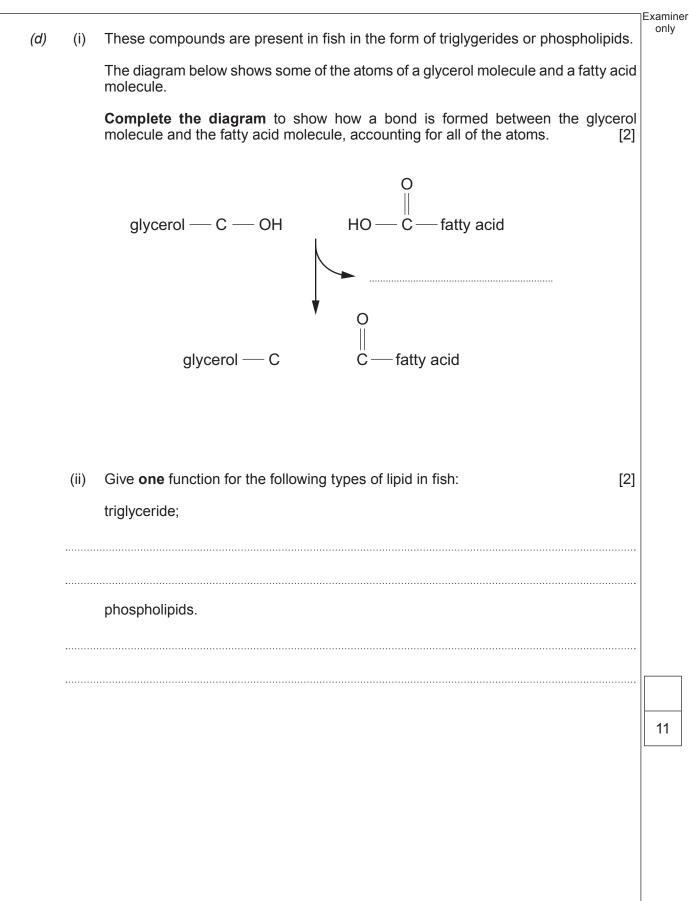


used	ently, a million tons of wild fish are harvested from the oceans annually. Of these, 80% are to feed fish reared in farms.
to be	do not make their own omega-3 fatty acids, they get these by eating algae. For farmed fish as nutritious as wild fish, they need to be fed a diet rich in omega-3 fatty acids. Algae are to culture on a large scale, so wild fish provide a more convenient source.
of on	ntists have identified and spliced genes from algae that code for producing high levels nega-3 fatty acids into <i>Camelina</i> plants. The genetically modified <i>Camelina</i> plants now uce seeds that contain 26% omega-3 fatty acids, so are useful for fish farm feed.
(a)	Explain how named enzymes would be used to cut genes from algae and splice them into <i>Camelina</i> plants. [3]
••••••	
(b)	Explain why feeding genetically modified <i>Camelina</i> seed to farmed fish could benefit biodiversity. [1]

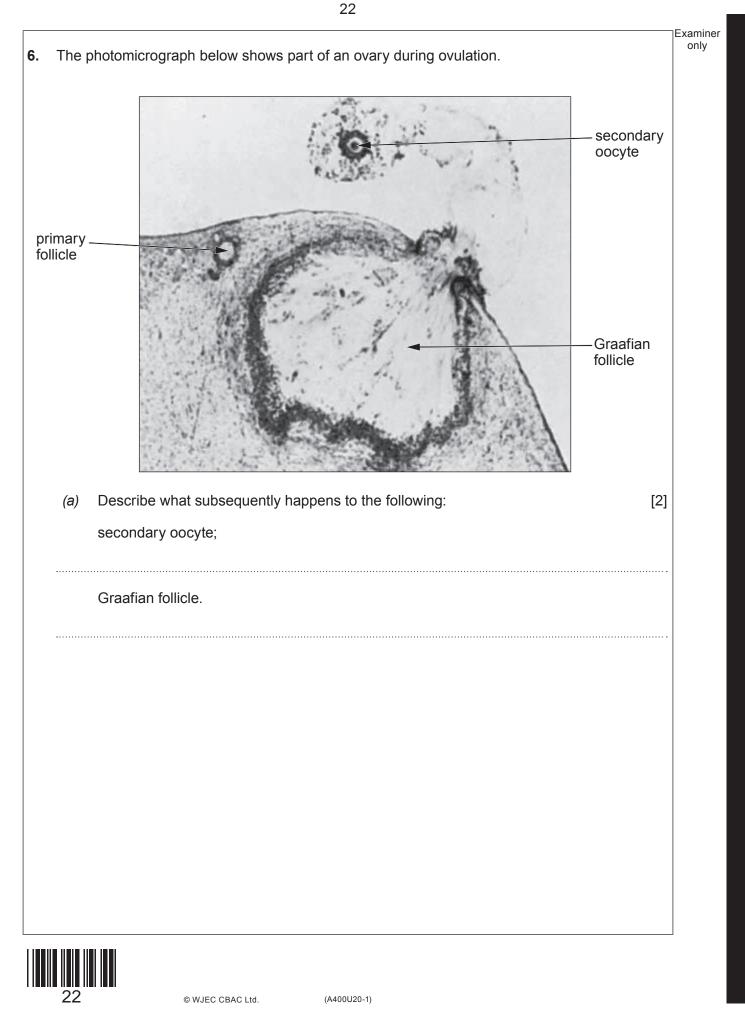










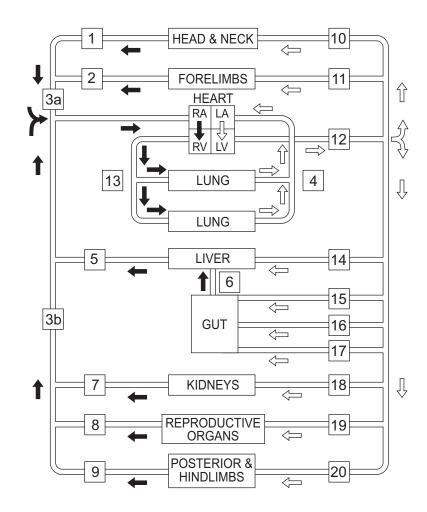


b)	(i)	Describe how the hormones FSH and oestrogen illustrate the principle of negative feedback in controlling ovulation. [4]
	(ii)	Explain why oestrogen may therefore be used as a contraceptive. [3]
	••••••	



(c) For use as a contraceptive, oestrogen can be incorporated into a porous patch which may be applied to the skin on a woman's arm. The diagram below is a schematic map of the human circulatory system.

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oxygenated blood \Rightarrow deoxygenated blood



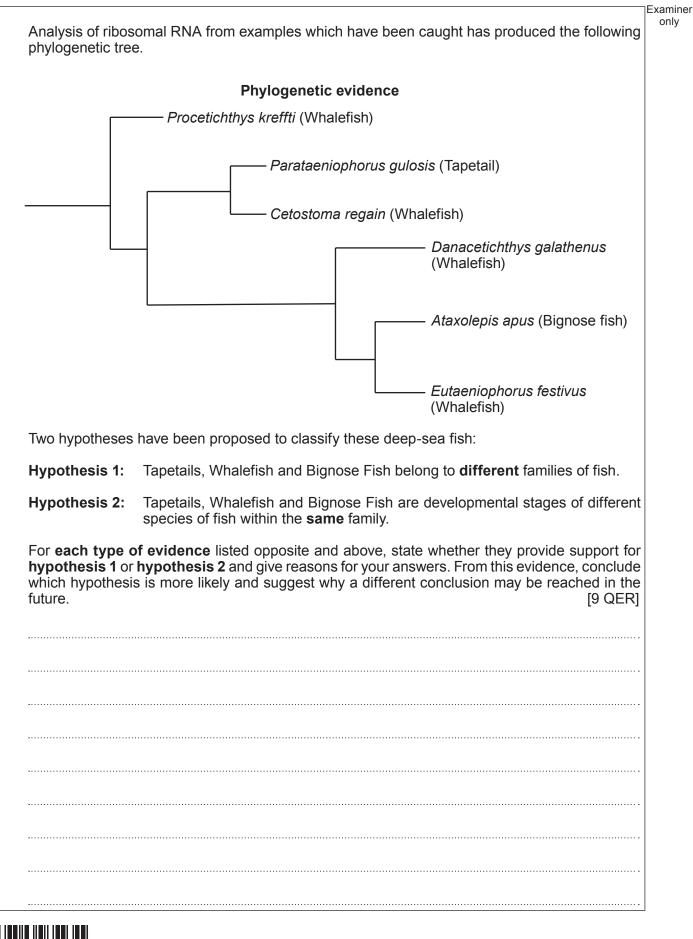
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(i)	Using numbers for blood vessels and letters for the chambers of heart, describe the route that oestrogen would take from the patch to the organ where it would exert its contraceptive effect. [2]	Examiner only
(ii)	Apart from containing more oestrogen, describe one other difference in the blood leaving the patch and arriving at the target organ and state where the change would have taken place. [2]	
		13



The deep regions of the oceans (1000 – 4000 m) are nutrient-poor habitats. In order to survive, 7. the fish that live there have evolved remarkable morphological and life-cycle strategies. The larvae of many deep-sea species develop in the nutrient-rich surface waters (0-200m). Very few specimens of these fish have been caught, which makes them difficult to classify. This illustrates the tentative nature of classification. The table below provides information on specimens of three types of deep-sea fish. Type of **Example specimen** Evidence Fish Depth Caught: 0 – 200 m Number caught: 120 **Size:** 5 – 56 mm Age: juvenile Tapetail Gender: unknown Feeding: large numbers of animal plankton; mouth is upturned, jaws are small Depth Caught: below 1000 m Number caught: 600 **Size:** 26 – 408 mm Age: adult Whalefish Gender: all female Feeding: eat large prey; jaws are long and can open very wide to swallow prey whole Depth Caught: below 1000 m Number caught: 65 **Size:** 34 – 68 mm Bignose Age: adult Fish Gender: all male Feeding: do not feed; have a very large liver which acts as a store of energy and nutrients





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