



**First phylogenetic analysis of the family Neriidae (Diptera),
with a study on the issue of scaling continuous characters**

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3 **First phylogenetic analysis of the family Neriidae (Diptera), with a study**
4 **on the issue of scaling continuous characters**
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23 Running title: Neriidae phylogeny and the scaling of continuous characters
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Abstract

Neriidae are a small family of acalyptratae flies, mostly distributed along the tropics. Very little is known about their biology, and the evolutionary relationships among species have never been evaluated. We perform the first comprehensive phylogenetic analysis of the family, including 48 species from all biogeographic regions inhabited, as well as five species of Micropezidae and one Cypselosomatidae as outgroups. We build a morphological data matrix of 194 characters, including 72 continuous characters. We first explore ways to deal with the issue of scaling continuous characters, including rescaling ranges to unity and using implied weighting. We find that both strategies result in very different phylogenetic hypotheses, and that implied weighting reduces only partially the issue of scaling. Furthermore, using implied weighting after rescaling characters improves the congruence between partitions and results in higher values of group support. With respect to the Neriidae, we confirm the monophyly of the family and of most its genera, although we do not obtain any of the currently accepted supra-generic groups. We propose to restrict the *Eoneria* and *Nerius* groups exclusively to the Neotropical fauna, and synonymyze *Glyphidops* subgenus *Oncopsia* Enderlein with *Glyphidops* subgenus *Glyphidops* Enderlein, eliminating the subgenera of the latter. This revised phylogeny presents a striking biogeographic consistency, and shows that previous main divisions of the family were based on events of convergence.

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Introduction

Neriidae are a small family of medium to large size acalyptratae flies, with a general morphology characterized by relatively elongated bodies and long and slender legs, generally ornamented with two rows of ventral spines (Aczél, 1951; Carvalho-Filho and Esposito, 2008; Sepúlveda *et al.*, 2013a). Neriid flies can be readily distinguished from all other Acalyptratae by the elongated porrect antennae with an apical to subapical arista, and a pedicel divided into a body and a finger-like projection that connects with the median region of the first flagellomere (Aczél, 1961; Steyskal, 1968; Buck, 2010). Despite being present in all biogeographic regions, most of the family's diversity is concentrated along the tropics (Steyskal, 1968; 1987). So far, 110 species have been described and placed in 19 genera (Sepúlveda *et al.*, 2013a), with almost two-thirds of this diversity occurring in the New World (Eberhard, 1998). Among the American species, three occur in the desertic regions of southwestern United States (Mangan and Baldwin, 1986; Steyskal, 1987), whereas the vast majority dwells throughout Central and South America (Aczél, 1961; Steyskal, 1968). The rest of the species are mainly distributed along the Australian-Oriental (Aczél, 1954a; Steyskal, 1977; Pitkin, 1989) and Afrotropical ecozones (Aczél, 1954b; Steyskal, 1980; Barraclough, 1993a).

Neriidae were historically considered by some as an independent family closely related to the Micropezidae (Hendel, 1922; Cresson, 1930), while others regarded them as a subfamily included within the Micropezidae (Enderlein, 1922; Hennig, 1934; 1936; 1937). Consensus on the placement of this clade at the family level was only reached after the thorough taxonomic work done by Martín L. Aczél, including several monographs on the diversity of neriid flies from all inhabited continents (Aczél, 1951; 1954a; 1954b; 1954c; 1955a; 1955b; 1955c; 1959; 1961). Neriidae are at present recognized as one of the families included within the superfamily Nerioidea, the monophyly of which has been retained in several morphological and molecular higher-level phylogenetic analyses (J. F. McAlpine, 1989; Yeates and Wiegmann, 2005; Yeates *et al.*, 2007; Wiegmann *et al.*, 2011). Several synapomorphies are taken to support the monophyly of the Nerioidea, including the peculiar morphology of the elongated male and female genitalia and the desclerotized lower region of the face (Aczél, 1951; J. F. McAlpine, 1989; D. K. McAlpine, 1996; Yeates *et al.*, 2007; Buck and McAlpine, 2010). However, the internal taxonomic and systematic organization of the clade is still highly controversial. The most conservative view recognizes only three families within the Nerioidea (J. F. McAlpine, 1989): the extremely diverse Micropezidae (often subdivided into several subfamilies, see D. K. McAlpine, 1974; Marshall, 2010), and the less diverse sister taxa Cypselosomatidae and Neriidae. The monophyly of the Cypselosomatidae + Neriidae clade was originally proposed by J. F. McAlpine (1989), and has found subsequent confirmation in a recent molecular phylogeny (Wiegmann *et al.*, 2011). Other authors have proposed to separate Pseudopomyzidae from Cypselosomatidae and to establish this group as a fourth family (D. K. McAlpine, 1966;

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3 Shatalkin, 1994), although many have favored a subfamilial rank for the two (Griffiths,
4 1972; Prado, 1984; J. F. McAlpine, 1987; 1989). D. K. McAlpine (1996) rejected this last
5 stance arguing that the similarities between both clades is due to the retention of
6 symplesiomorphies, yet consensus on their position is still lacking. D. K. McAlpine (1996)
7 has also transferred the Megamerinidae from the Diopsoidea into the Neriioidea, but others
8 have not followed this proposal (Buck, 2010).
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12 The modern systematic structure of the Neriidae was mainly erected by Aczél
13 (1954a; 1961), although it was Enderlein (1922) who first defined some of the suprageneric
14 clades that remain currently valid. As a result, the family is divided in two subfamilies: the
15 Neriinae, characterized by the presence of antennal sockets (also called antennal bases),
16 formed by the protruding and more or less inflated frontal region of the upper face (also
17 referred to as mesofacial plate), into which the antennae are inserted; and the basal
18 Telostylinae, which lack these antennal bases. However, much confusion has subsequently
19 arisen from this division, and many authors doubt that they constitute monophyletic groups
20 (Pitkin, 1989; Barraclough, 1993a; Buck, 2010). The entirety of the American fauna
21 belongs to the subfamily Neriinae, which Aczél (1961) further subdivided into two groups:
22 the *Nerius*-group, among which the dorsal region of the antennal bases is polished and
23 shiny; and the *Eoneria*-group, in which the antennal bases are dull, and have at most a faint
24 greasy luster. Furthermore, some authors (Aczél, 1961; Buck and Marshall, 2004; Buck,
25 2010; Sepúlveda *et al.*, 2013a) have partially discussed morphological similarities between
26 the Neotropical genera *Longina* Wiedemann, *Cerantichir* Enderlein and *Odontoloxozus*
27 Enderlein, although no author has gone as far as to propose these genera as constituting a
28 monophyletic group, and no other phylogenetic hypothesis has been put forward. A
29 summary of all these proposed relationships can be found in Fig. 1.
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38 Ever since the mid 20th century, few taxonomic works have been published on the
39 family. A handful of papers have dealt with the description of some new species (Mangan
40 and Baldwin, 1986; Buck and Marshall, 2004; Sepúlveda *et al.*, 2013b), but the systematic
41 relationships within the family have never been reevaluated nor tested using matrix-based
42 phylogenetic methods. The family does present some peculiarities that have been
43 interpreted by some as impediments towards a morphological evaluation of the
44 relationships among genera. For example, the male genitalia is extremely conserved, and
45 basically useless to determine evolutionary relationships (Buck, 2010). On the other hand,
46 the distribution of setae (chaetotaxy), which has been routinely used with phylogenetic
47 purposes among Diptera (McAlpine, 1987; Simpson *et al.*, 1999; Lambkin *et al.*, 2013) is
48 outstandingly variable within specific limits among neriids (Aczél, 1951; Barraclough,
49 1993a; Buck, 2010), a feature that has even been experimentally addressed (Bonduriansky,
50 2009). The confusion that has arisen from such intra-specific variation is most surely
51 epitomized by Steyskal's (1965) synonymization of the genera *Antillonarius* Hennig and
52 *Imrenerius* Aczél due to the presence in the same specimen of "(...) a well-developed
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3 anterior *ntpl* [notopleural] on one side and a barely distinguishable one on the other", a
4 character that not only represented the main difference distinguishing both genera, but that
5 was also considered by Aczél (1961) as having a high taxonomic importance within the
6 family. These and other issues have led some authors to even doubt the value of cladistic
7 enquiry altogether (see D. K. McAlpine, 1996 for such an argumentation concerning the
8 phylogenetic relationships among the Neriioidea).
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12 Nonetheless, the impressive taxonomic legacy of M. L. Aczél (summarized by
13 Mello, 2010), including the description or redescription of almost half of the known species
14 of neriids, may prove to be a treasure vault for phylogenetic analysis. Taxonomic
15 descriptions generally harbor an outstanding amount of morphological data. This
16 information has a quite direct correlation with the one included in morphological matrices
17 used in cladistic analyses (Winston, 1999), making it plausible that a translation of these
18 descriptions into the codified nature of data matrices will result in valid phylogenetic
19 analyses. In particular, Aczél's taxonomic descriptions are extremely rich in anatomical
20 details, including the registry of pigmentation patterns, chaetotaxy, general morphology and
21 more than 40 body measurements (generally expressed employing ranges of variation) for
22 both male and female specimens (see Aczél, 1959; 1961 for a few examples). The key to a
23 new and revised phylogeny of the neriids may be hidden among this precise record.
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30 One type of character that Aczél regularly insisted in using while delimiting groups
31 within the Neriidae are body measurements and proportions. In his revision of American
32 Neriidae (Aczél, 1961) he classified species according to body size, shape of the eyes,
33 length of postcranium, proportions of the legs, and length of the male and female
34 postabdomen. Such continuous characters have been historically neglected from cladistic
35 analysis both due to theoretical and practical issues concerning their implementation.
36 Objections against their use have included concerns regarding the existence of homologies
37 in quantitative characters (Pimentel and Riggins, 1987), their nature as phenetic data
38 (Cranston and Humphries, 1988) and the unavoidable arbitrariness involved in the methods
39 used to discretize them (Archie, 1985; Crisp and Weston, 1987; Felsenstein, 1988; Farris,
40 1990). Much of the theoretical objections have since been dealt with (Chappill, 1989; Rae,
41 1998), and many authors have argued that the distinction between discrete and continuous
42 characters is actually just a matter of degree (Stevens, 1991; Gift and Stevens, 1997; Wiens,
43 2001; MacLeod, 2002). A new reappraisal of quantitative data has followed after Goloboff
44 *et al.* (2006) implemented the treatment of continuous characters as such in the software
45 TNT (Goloboff *et al.*, 2008b). Since then, factual evidence that continuous characters carry
46 useful phylogenetic information has accumulated (Goloboff *et al.*, 2006; Hornung-Leoni
47 and Sosa, 2008; Pereyra and Mound, 2009; de Bivort *et al.*, 2010; Escapa and Catalano,
48 2013).
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56 Despite proving useful in many cases, some methodological caveats concerning the
57 implementation of continuous characters in parsimony analysis are still poorly explored.
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3 One of such issues is scaling, which has been referred to as “one of the most pervasive
4 problems in the analysis of continuous characters” (Goloboff *et al.*, 2006). As the same
5 authors argued, although it is clear that within a single continuous character a change in
6 state between two species should be proportional to the magnitude of the difference they
7 exhibit, the problem arises when considering the cost of transformations among different
8 characters that potentially vary in widely different magnitudes. When this occurs,
9 characters expressed in higher orders of magnitude will typically dominate the analysis
10 (Thiele and Ladiges, 1988; Wiens, 2001; Goloboff *et al.*, 2006; Baur and Leunberger,
11 2011), resulting in an unbalanced character influence towards determining the optimal
12 topology. Two methodological approaches have been therefore proposed to reduce this
13 phenomenon. The first one is the practice of rescaling continuous characters, standardizing
14 their ranges of variation to a common magnitude, usually unity. This method has been
15 extensively used, both to standardize discretized (Colles, 1980; Cranston and Humphries,
16 1988; Thiele and Ladiges, 1988; Vargas *et al.*, 2010) and non-discretized continuous data
17 (Abdala and Juárez Heredia, 2013; Escapa and Catalano, 2013), although some have
18 objected to this practice on different grounds (Mikevich and Farris, 1981; Farris, 1990;
19 Goloboff *et al.*, 2006). The second approach was proposed by Goloboff *et al.* (2006) who
20 argued that the use of implied weighting against homoplasy (Goloboff, 1993) may be a way
21 to reduce the issue of scaling. Since characters measured on larger scales will most likely
22 have a higher amount of homoplasy than characters measured in smaller scales, the former
23 would receive lower implied weights and vice versa, possibly balancing the overall
24 influence of the different characters. This may reduce the effect of the magnitude of
25 continuous characters on the phylogenetic hypothesis while at the same time elegantly
26 eluding the problem of determining an appropriate scaling factor (Goloboff *et al.*, 2006),
27 and has been recently adopted by some (Mannion *et al.*, 2013). Although the argument is
28 logical, no empirical evidence has been presented to support such claim, and it is not clear
29 whether implied weighting successfully deals with the issue of scaling, therefore
30 eliminating the dominance of large characters. Furthermore, the effect that these two
31 strategies have on the resulting phylogenetic hypothesis has never been tested.

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44 In the present work, we pursue a double objective. First, we analyze different
45 strategies for the use of continuous characters under parsimony, including an analysis of the
46 effects of rescaling and implied weighting. Second, we apply such insights to develop the
47 first phylogenetic study of the Neriidae. For this, we built a 194 character matrix, which
48 includes 72 continuous characters. We include in the analysis 48 species of neriids, with
49 representatives from 14 out of 19 valid genera and from all biogeographic regions
50 inhabited. Also included are 5 species of Micropezidae and one species of
51 Cypselosomatidae, used as outgroups. Our results show that rescaling and applying implied
52 weight to continuous characters result in completely different phylogenetic hypothesis, and
53 that implied weights reduces, yet does not eliminate, the issues of scaling. Regarding the
54 phylogenetic relationships among the Neriidae, we do not obtain many of the classical
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3 subdivisions of the family as monophyletic groups, and therefore discuss the evolutionary
4 and biogeographic history of the family in light of a revised phylogeny.
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8 9 **Materials and methods**

10 11 12 **Taxon sampling**

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15 Fifty-four species are included in the present analysis. The ingroup consists of 48
16 species of Neriidae, representing 44% of the described species. This sampling includes
17 representatives of 14 out of 19 currently valid genera (the remaining five genera are all
18 monotypic), and contains representatives of all major biogeographic regions inhabited.
19 With respect to the outgroup, five species of Micropezidae and one species of
20 Cypselosomatidae were included, therefore testing the monophyly of the Neriidae with
21 respect to the other families conforming the superfamily Neriioidea *sensu* J. F. McAlpine
22 (1989). The five micropezids included were chosen in order to incorporate a wide
23 taxonomic and morphological diversity, and include two representatives of the
24 Taenipterinae, two Micropezinae and one Eurybatinae. All species incorporated in the
25 analysis can be found in Table 1.
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33 34 **Character sampling**

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36 Specimens from 26 of the 54 species included in the study were revised at the
37 collection of the Instituto Superior de Entomología, Fundación Miguel Lillo, Tucumán,
38 Argentina (see Appendix S2 for a list of revised specimens). These include many of the
39 original material deposited by Aczél. For the rest of the species, character states were
40 recorded from taxonomic descriptions. Our sampling resulted in 122 discrete characters that
41 describe the morphology of adult organisms, since immature stages have been described
42 only for a couple of species (Steyskal, 1968). A description of these can be found in
43 Appendix S1. Of these, 77 were coded as binary and 45 as multistate, of which 35 were
44 taken as ordered. Furthermore, a total of 42 measurements were taken for all revised
45 specimens, either by directly measuring them with an Olympus SZ4045 binocular
46 microscope with an ocular micrometer, or from photographs of the specimens taken with an
47 Olympus U-CMAD3 (Infinity 1) digital camera attached to an Olympus SZX7 binocular
48 microscope, and afterwards digitally measuring body features with the software tpsDig2
49 (Rohlf 2010). For species included in the analysis that were not revised, the same
50 measurements were annotated from available taxonomic descriptions. These were later
51 combined into 72 continuous characters, all of which (except for character 1 and 2: male
52 and female body length, respectively) take the shape of simple ratios. This practice,
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3 although subject to some debate (Atchley *et al.*, 1976; Corrucini, 1977; Albrecht, 1978;
4 Atchley, 1978; Hills, 1978) is extremely common both in morphometric taxonomy and
5 phylogenetic analyses (Baur and Leuenberger, 2011; see de Bivort *et al.*, 2010; Lopardo *et*
6 *al.*, 2011; Mannion *et al.*, 2013 for recent phylogenetic analysis that incorporate characters
7 expressed as ratios). The objective behind this practice was either to reduce the effect of
8 body size in morphological measurements (e.g.: chars. 3, 35, etc.), or to numerically
9 represent shapes, proportions or relative positions (e.g.: chars. 5, 22, 49, respectively).
10 From the 72 continuous characters, 5 described the male and female genitalia (chars. 68 and
11 henceforth), and 3 were coded using male and female data indistinctively, due to having
12 access to fragmentary data relating to sexual differences (chars. marked with an asterisk in
13 Appendix S1). The remaining ratios were initially built using male and female
14 measurements separately, resulting in 64 (32 pairs) of sex-specific characters.
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23 **Analysis of continuous characters**

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25 The objective of this section was to study the behavior of quantitative (continuous)
26 characters under different strategies of analysis. Since no phylogenetic conclusions are to
27 be taken from the results, the duplication of phylogenetic information through the use of
28 covarying characters was not considered an issue, and is dealt with latter (see Identification
29 of sexually dimorphic characters). Therefore, the entire set of 72 continuous characters was
30 retained and used without testing their independence. Four strategies for the treatment of
31 continuous characters were explored, involving: the use of continuous data without
32 treatment (strategy A), applying implied weights (strategy B), rescaling (strategy C), and
33 rescaling + applying implied weights (strategy D). When using implied weights, a
34 concavity constant (k) of 6 was always used. When rescaling, the range of all characters
35 was standardized to unity. For this and all subsequent analysis the Willi Hennig Society
36 version of the program TNT (Goloboff *et al.*, 2008b) and the software Statistica (Statsoft
37 2001) were used for phylogenetic and statistical enquiry, respectively.
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44 On the first place, we aimed at proving Goloboff *et al.*'s (2006) assertion that the fit
45 assigned to a given continuous character when using implied weighting negatively
46 correlates with the scale of such character. Therefore, we searched for the most
47 parsimonious tree (henceforth MPT) supported by the continuous data partition under
48 implied weights, with (strategy D) and without (strategy B) rescaling the data. We used a
49 driven search (Goloboff, 2002; Giribet, 2007), starting with five random addition sequences
50 with SPR and TBR branch swapping followed by ratcheting (Nixon, 1999), sectorial
51 searches, tree drifting and fusing (Goloboff, 1999), until minimum length was found 10
52 times. Unless otherwise stated, this strategy was maintained in all subsequent tree searches.
53 A single MPT was found for both strategies, and the fit of all characters in both trees was
54 recorded as the complement of the value obtained with the function `fit` (actually the
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3 “distortion” of a character). These values were used in a regression analysis using the range
4 of variation of each character (maximum – minimum, a proxy for a character’s magnitude)
5 as the independent variable.
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8 Afterwards, the MPTs for the four strategies were topologically compared in a pair-
9 wise manner, in order to study the impact that different strategies had on the resulting
10 phylogenetic hypothesis. For the comparisons we used the number of taxa in the agreement
11 subtree (Gordon, 1980; Eulenstein *et al.*, 2004), the number of internal nodes in the strict
12 consensus (Mickevich, 1978; Swofford, 1991), and the weighted and unweighted SPR
13 distances (Goloboff, 2007) as measures of topological similarity. The weighted SPR
14 distance has been considered a superior measure of topological similarity (Goloboff, 2007),
15 since it measures not only the number of SPR moves needed to transform one tree into the
16 other, but it also weights each move by its distance (*i.e.*, the number of nodes (n) separating
17 the subtree’s location before and after the movement). The weight assigned to each move is
18 then $n / (n + j)$, and a value of 3 was always used for j . For ease of comparison, all
19 measures except for the weighted SPR distance were standardized so that the maximum
20 possible tree similarity obtained using each of them was equal to 1 (since a higher value of
21 SPR distance implies a higher topological difference, rather than a higher resemblance as in
22 the other employed measures, the SPR derived similarity was used instead).
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29 The compatibility between the discrete and continuous character partitions was then
30 explored under the four strategies (when using implied weights, both data partitions were
31 weighted). An incongruence length difference test (ILD, Farris *et al.*, 1995) was used to
32 study the incongruence between partitions, and weighted SPR distances were used to
33 compare the MP trees supported by both partitions. Furthermore, absolute group
34 frequencies under jackknifing (Farris *et al.*, 1996) were estimated for the MPTs of the
35 entire dataset under all strategies, using 500 pseudoreplicate datasets by eliminating
36 characters under $p = 36$ and analyzing each one with 10 runs of TBR + ratcheting with 10
37 iterations each. Higher mean node support values were taken to represent that a particular
38 strategy of analysis resulted in a higher congruence between characters (as discussed by
39 Goloboff, 1997; Ramírez, 2003; Goloboff *et al.*, 2008a).
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45 We also designed an experiment to test the validity of Goloboff *et al.*’s (2006)
46 second assertion relating to the issue of scaling, *i.e.* that implied weighting balances the
47 overall influence of continuous characters due to the negative correlation between a
48 characters magnitude and its fit. We ordered continuous characters with respect to their
49 magnitude (once again, using their range as an estimate of such variable), and built two
50 subsets of characters by selecting the 18 smallest and 18 largest of them (this number was
51 chosen since it represents the top and bottom quartile). Defined in such a way, the smallest
52 character within the “large” subset had a range 3.2 times larger than that of the largest one
53 in the “small” subset. The MPTs for these two subsets of characters were searched and
54 compared using the weighted SPR distance to the MPT supported by the complete
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3 continuous dataset. A strategy that guarantees a balanced influence of all characters should
4 result in a more or less similar distance from the trees supported by both subsets to the tree
5 of the entire continuous partition. On the contrary, under a strategy where the magnitude of
6 a character determines its influence over the optimal topology, the tree supported by the
7 largest characters should be much more similar to the one supported by the entire partition
8 than the one supported by the smallest characters, resulting in asymmetrical SPR distances.
9 This procedure was repeated for all four strategies. To further explore the implications of
10 the values obtained, we built null distributions of SPR distances by generating 1000
11 random subsets of 18 continuous character for each strategy, searching for the MPTs for
12 every subset (using 20 RAS + TBR and holding up to 10 trees of the minimum length), and
13 calculating the weighted SPR distance to the MPT of the entire continuous partition (the
14 script used can be found in Appendix S3). Since many subsets supported more than one
15 optimal tree, we calculated the SPR distance using all of them and retained, for each subset,
16 the smallest distance. The resulting value can be interpreted as the “importance” of these
17 subsets in determining the optimal tree of the entire continuous partition, in a similar way
18 as discussed by DeGusta (2004) for individual characters. Therefore, the shortest the SPR
19 distance, the higher the influence on the topology supported by the entire continuous
20 dataset.
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29 Since the calculation of SPR distances is an NP-complete problem (Bordewich and
30 Semple, 2005), the algorithm implemented in TNT is heuristic and depends on two
31 parameters: the number of replications and the number of stratifications (Goloboff, 2007).
32 Given that SPR distances were intensely used, we first performed a tuning analysis to
33 define appropriate values for those two parameters (the script used can be found in
34 Appendix S3). One hundred Wagner trees were created and coupled into 50 pairs, and the
35 SPR distance between them was explored using different combinations of parameters. We
36 first defined a broad and less intense search, calculating the distance between trees for all
37 combinations of number of replications between 1,000 and 20,000 (with steps every 1,000)
38 and number of stratifications between 0 and 75 (with steps every 5). No major change in
39 the resulting distances was found using stratification values above 30. On the other hand,
40 since computational time increments with an increase in the number of replicates, we found
41 that a number of 13,000 replicates guaranteed finding the shortest distance for more than
42 95% of tree pairs, being a good compromise between computational effort and accuracy. A
43 second, more intense yet restricted search (number of replicates: 1,000 to 13,000, steps of
44 1,000; number of stratifications: 0 to 30, steps of 1) showed that the shortest path between
45 pairs of trees was, on average, attained using 13,000 replicates and 18 stratifications (Fig.
46 2). These parameters were therefore used for all SPR distance calculations
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56 **Outgroup definition**

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3 Historically, the Neriidae were considered to be closely related with the
4 Micropezidae (Aczél, 1951), even being classified as a subfamily included within the later
5 (Hennig, 1936; 1937; Griffith, 1972). After their status as an independent family was
6 recognized, some authors pointed out the existence of morphological similarities between
7 the Neriidae and the Cypselosomatidae (D. K. McAlpine, 1966; 1974), with latter analyses
8 confirming such clade to be monophyletic (J. F. McAlpine, 1989; Wiegmann *et al.*, 2011).
9 Therefore, we rooted our trees with *Taeniptera annulata*, a micropezid of the subfamily
10 Taenipterinae, which have been considered to be morphologically very dissimilar with
11 respect to Neriidae by Hennig (1937), Aczél (1951) and D. K. McAlpine (1974).
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19 **Identification of sexually dimorphic continuous characters**

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21 So far, all continuous characters were *a priori* taken to be independent, including
22 the 32 pairs of male-female characters. The codification of male and female data in separate
23 characters may be useful in many occasions, since by reducing the ranges of variation, this
24 practice results in less overlapping and greater informativeness of each of the resulting
25 characters. Furthermore, it allows a more detailed study of the evolution of sexual
26 dimorphism (Hormiga *et al.*, 2000). However, these advantages are only true if male and
27 female characters are effectively independent, otherwise their codification as separate
28 characters only results in the inclusion of redundant phylogenetic information in the
29 analysis. Therefore, we tested whether our 32 pairs of male-female characters presented
30 differences in their phylogenetic information. This was done following a strategy similar to
31 that of de Bivort *et al.* (2010). Regression analyses were performed using male and female
32 characters corresponding to the same morphological structure. Since some characters were
33 expressed using ranges of variation and others only as single data points, in the first case
34 the mean between the minimum and maximum values was used. In case a species presented
35 missing data for at least one of both characters it was consequently excluded from that
36 particular regression. Two characters were then considered to be dependent when they
37 showed a significant linear regression with a complete absence of outliers. Since the
38 objective of the regression analysis is to study whether a sex-specific character confidently
39 predicts the value of that same character for the opposite sex (therefore showing a lack of
40 independence among all taxa under study), the presence of a single outlier is evidence that,
41 at least for one species, the same attribute is providing different phylogenetic information
42 for males and females (de Bivort *et al.*, 2010). Therefore, for each regression, the residue of
43 each data point was annotated and divided by the corresponding expected value. This
44 resulted in a number that expressed the magnitude of the difference between observed and
45 expected values for each species as a fraction of the expected value. Three nested,
46 progressively stricter confidence intervals were considered, according to which characters
47 were retained as dimorphic only if there was at least a single species for which the observed
48 value differed from the expected one by a 20, 30 or 50% of the latter (Fig. 3). Otherwise, in
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3 the absence of outliers, male and female characters were collapsed into a single character,
4 whose range was defined by using the smallest value of both as a lower limit and the largest
5 one as the upper limit (the script used can be found in Appendix S3). This resulted in the
6 retention as separate characters of approximately 77, 40 and 27% of the 32 original pairs as
7 the criterion became stricter (see Fig. 4c). This analysis included only male-female pairs of
8 identical characters. Different morphological measures were considered independent
9 without further *a priori* confirmation (in a fashion similar as for example Fink and Zelditch,
10 1995; Strait & Grine, 2004).
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17 **Cladistic analysis**

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20 Phylogenetic analysis was undertaken under strategy D, that is, rescaling continuous
21 data to unit range and using implied weighting (see below). Trees were searched using
22 constant of concavity k ranging from 1 to 10. This resulted in 10 ways to weight characters,
23 which coupled with the 3 male-female collapse criteria, determined 30 different forms to
24 analyze the data. All of these resulted in a single MPT. A majority rule consensus (cut-off
25 50%) was employed to perform a sensitivity analysis and identify those groups that were
26 recovered under most conditions of analysis. The phylogenetic hypothesis proposed was
27 chosen by calculating the majority consensus frequency for all groups present in each one
28 of the 30 trees, averaging the values, and looking for the tree with higher mean group
29 frequencies. This hypothesis is therefore the one that combines the groups most frequently
30 present in the entire 30 tree set. As measures of support we used absolute frequencies of
31 jackknife (parameters as defined above, but 1000 pseudoreplicates) and Bremer support
32 (Bremer, 1988; 1994). For the last one, a heuristic calculation was done by searching trees
33 that were increasingly suboptimal by 0.05 units of fit (under $k = 6$, the same value
34 employed for the analysis of continuous characters). Tree search was done by TBR
35 swapping, and up to 1000 trees for each round were retained. Search continued until
36 reaching trees suboptimal by 1.35 units of fit, the point at which all nodes had been
37 contradicted. Bremer supports were then calculated from the 27000 existing suboptimal
38 trees, and plotted as units of fit x 100. TBR swapping of trees suboptimal by 0.05 units of
39 fit produced only 933 trees, not overflowing tree space; hence values below 5 are probably
40 exact.
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51 **Results**

52 **Analysis of continuous characters**

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3 We found a significant and negative correlation ($p < 0.0001$) between the fit
4 assigned by implied weights and a character's range (Fig. 4b). The regression analysis
5 determined that in fact, implied weights is almost exclusively weighting against a
6 character's magnitude, with a value of R^2 of 0.76. The strength of this correlation was not
7 dependent on the few extremely large characters, being equally strong when considering
8 only characters with ranges < 1 (Fig. 4a, see regression values at the epigraph). When
9 applying implied weights after rescaling, the correlation was lost ($p = 0.78$), and characters
10 with similar original scales were now assigned widely different fit values (Fig. 4).
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15 Therefore, implied weighting is in fact dealing with the magnitude of continuous
16 characters, as originally stated by Goloboff *et al.* (2006). To prove whether the resulting
17 effect was similar to rescaling the data, we performed several tests of topological
18 differences between the MPTs obtained by each of the four strategies under analysis. We
19 found that treating the data (strategies B, C and D) resulted in phylogenetic hypothesis that
20 were all profoundly different from the one obtained by using the continuous dataset without
21 treatment (strategy A), as shown by all measures of topological similarity employed (Fig.
22 5). When topologically comparing the MPTs of treated vs. untreated data, both the number
23 of nodes in the strict consensus and the number of taxa in the agreement subtree were
24 always between 0.2 and 0.3 of the value corresponding to an absolute congruence, while
25 tree similarity derived from unweighted SPR distances ranged from 0.49 to 0.58. This sharp
26 topological difference could be pointing out that both rescaling and implied weighting are
27 eliminating, or at least strongly reducing, the dominance of large characters on the optimal
28 topology, a phenomenon that otherwise characterizes the use of continuous characters
29 without treatment. In fact, the mean consistency index (Kluge and Farris, 1969) for the two
30 largest characters (male and female body size, chars. 1 and 2) under strategy A was 0.54,
31 while it dropped to values between 0.27 and 0.32 for the remaining strategies. Similarly, the
32 mean retention index (Farris, 1989) for those two characters showed a decrease in value
33 from 0.88 to 0.61 - 0.68 after the data was treated. Fig. 6 shows the MPTs found using
34 exclusively the continuous partition under strategies A (a), B (b), and C (c), with the
35 optimization of the largest character, male body length, superimposed. The topology of
36 strategy A shows a high degree of dependence on this character (as evident also from the
37 values of CI and RI), which decreases both after rescaling and using implied weighting.
38 Evidence supporting a similar effect of rescaling and implied weighting was also found
39 using the ILD test, which revealed that continuous and discrete partitions were significantly
40 incongruent when compared under strategy A ($p = 0.026$), while the three ways to treat data
41 resulted in congruence between partitions (all $p > 0.315$).
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53 However, it can also be seen that the resulting topologies (Fig. 6b and c) are also
54 quite different from each another. In fact, the topological difference of the MPTs resulting
55 from rescaling and using implied weighting on continuous data (comparison B - C) present
56 somewhat intermediate values of SPR distances (Fig. 5) between the highly divergent
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3 values obtained from comparing treated vs. untreated data, and the more congruent values
4 resulting from comparing both trees obtained from data that was rescaled (comparison B -
5 D) or weighted (comparison C - D). The other measures of topological similarity also show
6 this intermediate placement, with the number of shared internal nodes attaining low values
7 and the number of taxa in the agreement subtree higher ones (Fig. 5a). It is therefore
8 evident that, although implied weighting is dealing with the magnitude of continuous
9 characters, applying lower weights to large characters (Fig. 4) and reducing their influence
10 on the final topology (Fig. 6), it is doing so in a different way than the rescaling of data, and
11 the two strategies result in quite different supported phylogenies (Fig. 5).
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16 Finally, a test was designed to study whether both alternatives were equally
17 allowing a balanced influence of all characters on the final topology (see Materials and
18 methods: Analysis of continuous characters), as originally proposed by Goloboff *et al.*
19 (2006). The resulting null distributions can be seen in Fig. 7. Distributions obtained under
20 strategies B, C and D showed a significant fit to a normal distribution (Chi-Square tests <
21 10.89, $p > 0.2$). Distribution obtained under strategy A showed a clear bimodal pattern, and
22 did not fit a normal distribution (Chi-Square test = 135.11, $p < 0.00001$). The variances of
23 the distributions were all significantly different from each other, with the smallest values
24 corresponding to strategies C and D (3.68 and 3.91 respectively), while the distribution of
25 strategy B showed a variance of 4.37 and that of strategy A of 9.87. Smaller variances can
26 be attributed to a contraction of the distribution towards lower values, with a displacement
27 of the right tail while the location of the left one remains relatively more constant (for
28 example, the position of the value leaving 5% of data to the right varies between
29 distributions 2.5 times as much as the position of the value that leaves 5% of the data to the
30 left). Under all four strategies, the distance obtained using the tree supported by the subset
31 including the 18 smallest characters was always near the median value of the distribution
32 (with 39.4 to 50.8% of obtained distance values being smaller). On the contrary, the
33 position of the distance obtained using the 18 largest characters differed widely among
34 strategies. For strategy A, this subset of characters had an enormous influence in the final
35 topology, showing an SPR distance 1.48 units shorter than the shortest one found by the
36 random subset generation (for the values of SPR distances obtained under each strategy see
37 Fig. 7). For strategy B, the position of the large subset left only 15.2% of values to the left
38 of the distribution, and was therefore considered to be still strongly influencing the overall
39 topology. On the contrary, among the two distributions obtained after rescaling (strategies
40 C and D), the large character subset was placed almost at the mean point of the distribution,
41 leaving 48.9 and 55% of distances to the left. In fact, for these two distributions, the
42 distances separating the position of the small and large subsets not only decreased greatly
43 (Fig. 7) with respect to the ones obtained for strategies A and B, their positions were even
44 inverted, with the large character subset having slightly larger SPR distances than the small
45 character one.
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3 It was therefore concluded that implied weighting was in fact reducing the
4 dominance of large characters on the final topology, although it was doing so in a partial
5 way and was comparatively inefficient to eliminate such influence altogether, as was in fact
6 happening when continuous characters were rescaled (Fig. 7). However, strategies C and D
7 behaved very similarly in all tests developed, and the decision on whether to weight
8 continuous characters against their homoplasy after these were rescaled proved to be the
9 least decisive of all that were explored (SPR distance between nodes C and D in Fig. 5b is
10 the shortest one). However, the MPT for the combined dataset under strategy C proved to
11 be very different than the ones obtained under strategy D for all k values tested.
12 Furthermore, strategy D resulted in a higher topological congruence between the trees
13 obtained for the discrete and continuous partitions when analyzed separately (weighted
14 SPR distances: 11.86 vs. 13.16), and in a higher mean group support for the MPT of the
15 combined dataset (average jackknife resampling frequency: 52.51 vs. 44.47; number of
16 nodes above 50%: 26 vs. 19) than strategy C. As a consequence, only strategy D was
17 employed for the phylogenetic analysis, given that it showed evidence of being able to
18 guarantee a balanced character influence and to increase the congruence between different
19 sources of characters.
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30 Cladistic analysis

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32 As already discussed, the phylogenetic analysis was performed under strategy D,
33 exploring 10 concavity functions of implied weighting ($k = 1$ to 10) and 3 progressively
34 stricter criteria for the collapsing of sexually dimorphic continuous characters. This
35 summed up to 30 forms of analysis, and a search using new technologies driven to find the
36 optimum 10 times always resulted in a single most parsimonious tree for each of them (this
37 phenomenon is common when using continuous characters and should not be taken as
38 evidence of a strong phylogenetic signal, see Bardin *et al.*, 2013). All of the obtained trees
39 were very similar, evidencing low levels of sensitivity to the parameters tested. Most major
40 (generic or supra-generic) groups were very stable and almost uncontradicted in all of the
41 obtained topologies, with 42 out of 48 resolved relationships present in the majority-rule
42 consensus showing frequencies higher than 80% (see Fig. 8 for values of frequency). The
43 few unresolved nodes mostly reflected conflict in the exact placement of single species
44 within those major clades.
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50 For each of these 30 trees, we calculated the frequency of occurrence of each of its
51 groups in the complete set of trees. We then retained the phylogenetic hypothesis that had
52 the highest mean group frequency, since it was the tree that showed the most commonly
53 found, and less sensitive clades. This tree, shown in Fig. 8, was found for k values from 6 to
54 10, in congruence with previous studies that found that mild concavity values resulted in
55 higher topological congruence between different morphological and molecular datasets
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(Ramírez, 2003; Lopardo, 2005; Goloboff, 2008b); and a criterion to retain sexually dimorphic characters without collapsing only if there was a 30% difference between observed and predicted values in the male-female regression analysis. It is worth noting that this criterion retained without collapsing all characters that have been traditionally reported in taxonomic revisions and experimental works as sexually dimorphic: body size and leg length (Bonduriansky, 2006; 2007), length of the scape (Aczél, 1951; 1961), length of the postcranium (Sepúlveda *et al.*, 2013a); as well as some other features of the antennae and the wings (see Appendix S1 for a list of characters retained as dimorphic).

The following supra-generic groups were obtained with relatively strong support and were insensitive to changes in the parameters of the analysis (jackknifing absolute frequencies/Bremer support/majority rule frequencies): Neriidae, clade A (99/130.75/100); American neriids, clade F (56/34.79/100); Neotropical species of the *Eoneria*-group, clade G (70/16.84/100); and the *Longina* - *Cerantichir* - *Odontoloxozus* group, clade J (79/66.37/100). Likewise, the following genera are confirmed to be monophyletic based on the same evidence: *Chaetonerius* Hendel (60/26.57/100), *Glyphidops* Enderlein (82/15.66/100), *Indonesicesa* Koçak and Kemal (99/51.30/100), *Longina* (94/82.45/100) and *Nerius* Fabricius (91/20.39/80). The synapomorphies supporting these and other groups in Fig. 8 are shown in Appendix 1.

As can be seen, most genera are obtained as monophyletic, with only a few clades contradicting the current taxonomy, although some of these are also shown to be largely insensitive to the parameters explored (Fig. 8). Specifically, the monophyly of the two subgenera contained within *Glyphidops sensu* Aczél (1961) is never recovered, although the genera itself is found to be monophyletic. Furthermore, *Eoloxozus sabroskyi* is placed in all trees as the sister group of *Eoneria maldonadoi* and nested within *Eoneria* Aczél. Likewise, the genus *Cerantichir* is found to be paraphyletic in all of the obtained trees. The chosen phylogenetic hypothesis (Fig. 8) shows *C. peruana* and *Odontoloxozus longicornis* conforming a monophyletic group, in congruence with the original description of the first species as *Od. peruanus* (Hennig, 1937) and contradicting Buck's (2010) reassignment. Finally, the genus *Telostylinus* Enderlein is not retained as a natural group, but subdivided into several different (although closely related) clades. Other than these four examples, the resulting phylogeny is highly congruent with the taxonomy of the family.

Discussion

Continuous characters and the issue of scaling

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3 The general apathy towards the use of continuous characters in phylogenetic
4 analyses has long worn off. Few if any object nowadays to their inclusion in cladistic
5 studies. Both from theoretical and practical points of view, quantitative characters have
6 proven to be both valid and useful data for systematists (Rae, 1998; Wiens, 2001; 2004;
7 Goloboff *et al.*, 2006; de Bivort *et al.*, 2010). Several breakthroughs have contributed to
8 this change in general perception. First, if continuous and discrete characters are in fact not
9 that different from each other, then there is no reason for their *a priori* exclusion from a
10 phylogenetic analysis. Many, if not most, of the phenotypic variability found in nature is
11 quantitative (Wiens, 2001; see also Baum, 1988; Chappill, 1989; Stevens, 1991; Thiele,
12 1993; Rae, 1998), independent on the decision to discretize them or not. If this is so, both
13 types of characters are actually representing the same attributes of living organisms, with
14 discrete characters being “informally discretized continuous characters” (de Bivort *et al.*,
15 2010: p. 302; similar arguments in Gift and Stevens, 1997; MacLeod, 2002; Haas, 2003),
16 *i.e.*: continuous variation that is more intuitively assigned to different, non-overlapping
17 categories. It is in fact the overlapping property of continuous data which initially posed a
18 problem for character coding, with much of the subsequent debate stemming from an
19 indiscriminate equation between ‘continuous’ and ‘overlapping’ (Thiele, 1993; Rae, 1998).
20 Nonetheless, if in fact “many so-called qualitative characters are based on a quantitative
21 phenomenological base filtered through the reified semantic discontinues of (...) terminology”
22 (Stevens, 1991: p. 553), many of the original theoretical objections to the use
23 of continuous characters (in the sense of Crisp and Weston, 1987; Pimentel and Riggins,
24 1987; Cranston and Humphries, 1988; Mickevich and Weller, 1990) are unjustified, and
25 discrete characters may even suffer from similar arbitrariness in the circumscription of
26 states.
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29 Another factor leading to the change in perception on the use of quantitative data
30 stems from Goloboff *et al.*'s (2006) incorporation into the software TNT of a set of
31 algorithms to use continuous characters as such. This allowed to entirely eluding all
32 discretization methods, which were considered inappropriate (Reid and Sidwell, 2002) or
33 unavoidably arbitrary (Archie, 1985; Crisp and Weston, 1987; Felsenstein, 1988; Farris,
34 1990; Gift and Stevens, 1997). Subsequent implementations demonstrated that continuous
35 characters carry useful phylogenetic information and formulate hypothesis congruent with
36 other sources of characters (Goloboff *et al.*, 2006; Hornung-Leoni and Sosa, 2008; Pereyra
37 and Mound, 2009; de Bivort *et al.*, 2010; Escapa and Catalano, 2013). Their dismissal from
38 cladistic analyses is therefore unwarranted.
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51 Despite all this progress, certain profound issues concerning the implementation of
52 continuous characters in phylogenetic analysis have received little attention. One of such
53 issues is that of scaling, that is, the differential influence of characters depending on the
54 scale in which they are measured. Although many have acknowledged the importance of
55 this problem, few have discussed alternatives to deal with it (Thiele, 1993; Wiens, 2001;
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3 Goloboff *et al.*, 2006). So far, two different strategies, rescaling and using implied weights,
4 have been proposed. The efficiency of both towards reducing the dominance of large
5 characters, as well as the consequences of each on the phylogenetic reconstruction, had
6 never been discussed. Our aim in the present study was to address these questions.
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10 If implied weighting was in fact dealing with the issue of scaling, as originally
11 proposed by Goloboff *et al.* (2006), it was necessary to demonstrate that characters were
12 receiving weights according to the range they presented, such that larger character were
13 assigned lower weights and small characters larger ones. To such end, we proved that the
14 fit assigned by implied weights to continuous characters presented a significant and
15 negative linear correlation with the range of a character (Fig. 4). This correlation was
16 strong, explaining 76% of differences in fit, and was equally strong after excluding the
17 largest characters in the matrix. Furthermore, the pattern was lost after characters were
18 rescaled to unit range (Fig. 4), demonstrating that the covariation was in fact a consequence
19 of the character's scale and not any other attribute relating to their phylogenetic signal. It
20 appeared therefore plausible that implied weighting was actually dealing with the issue of
21 scaling. As a matter of fact, different evidences showed that both implied weighting and
22 rescaling reduced the dominance of large characters on the optimal topology: both methods
23 eliminated the otherwise significant incongruence between continuous and discrete datasets
24 when partitions were compared untreated, and resulted in trees that were very different than
25 the one supported by the untreated continuous dataset (Fig. 5) and less dependent on the
26 larger characters (Fig. 6). Nonetheless, the topologies supported after rescaling and implied
27 weighting continuous characters were also quite different from each other, as shown by the
28 measures of topological similarity employed (Fig. 5). It was therefore evident that, although
29 both strategies were dealing with the issue of scaling, resulting in a decrease in the
30 dominance of large characters, they were doing so in different ways. Since the choice of
31 strategy had an important impact on the resulting phylogenetic hypothesis, both of them
32 could not be used interchangeably. Furthermore, it was not clear whether implied weighting
33 was as efficient in dealing with the issue of scaling as was standardizing the ranges of
34 continuous characters.
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44 To further evaluate this, we designed an experiment that aimed at detecting the
45 topological dependence of the optimal phylogenetic hypothesis with respect to the scale of
46 groups of characters, and see whether both methods were efficient in balancing the
47 influence of different characters. This was done by comparing, through weighted SPR
48 distances, the topological difference of trees supported by subsets of characters to the most
49 parsimonious tree of the entire continuous dataset. Characters with a strong influence in
50 determining the optimal tree of the entire partition will, when isolated, support a similar
51 tree. On the contrary, characters with a weaker influence will build hypothesis more
52 dissimilar to the one supported by the entire partition once all other characters are
53 inactivated. The resulting histograms are shown in Fig. 7. When continuous data is used
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3 without treatment (strategy A, Fig. 7a), the variance of the SPR distribution is larger, more
4 than twice that of all other histograms. Since differences in variance were mostly due to
5 changes in the position of the right tail of the distribution, it is evident that higher values
6 represent a decrease in the overall balance of influence of the characters, with certain
7 subsets contributing nothing (or very little) to the resulting phylogeny. In fact, only under
8 strategy A did the distribution's right tail advance to values obtained when comparing
9 random generated trees (weighted SPR distances > 24 units, mean = 28.62). The variance
10 of the obtained distributions was significantly reduced by all other strategies of analysis,
11 reaching the lowest values when characters were rescaled (strategies C and D).
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16 An interesting pattern arose after the SPR distances for the trees supported
17 exclusively by the 18 largest and 18 smallest characters were plotted on top of the
18 distributions. The "small" character subset resulted, for all strategies, in values of SPR
19 distances close to the median of the distribution, showing that their influence was relatively
20 insensitive to the chosen strategy. On the contrary, values for the subset of the largest
21 characters varied widely depending on the way continuous characters were analyzed. For
22 untreated data, these characters resulted in a tree that was by far the most similar to the tree
23 of the entire partition (Fig. 7a), showing that their influence in determining such topology
24 was massive. When characters were analyzed under implied weighting (Fig. 7b), such
25 influence became certainly reduced, with the difference in SPR distances between the
26 "large" and "small" subsets diminishing from 10.85 to 1.52. Despite such reduction in the
27 asymmetry of influences, large characters still retained a strong influence, with 84.8% of
28 randomly generated subsamples having larger SPR distances. The asymmetry in influence
29 is only completely eliminated after characters are rescaled. After this, the SPR distance
30 obtained for "large" and "small" subsets differed only by 0.22 and 0.03 units from one
31 another, depending on whether implied weighting was applied or not, respectively (Fig. 7c-
32 d). Furthermore, both subsets were placed very near the mean value for each distribution,
33 showing that rescaling had eliminated all information on the magnitude of the characters
34 and these had become, despite their original differences in scale, two average subsets of
35 characters with respect to their influence in the supported phylogeny.
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45 All evidence points to the same conclusion: implied weighting reduces the issue of
46 scaling, yet it does so only partially. Large characters still show a more than average
47 influence on the resulting phylogeny, and this is shown to be exclusively due to their larger
48 scales. As Farris (1990, p. 91) stated "(...) domination cannot logically be objectionable in
49 itself, without some independent grounds for objecting to the dominant factor." The scale in
50 which a character is measured is a factor that results in asymmetrical influences among
51 different continuous characters, and yet it is completely arbitrarily determined during
52 character coding and has no relationship whatsoever to phylogenetic signal. Objecting to
53 such factor of dominance is therefore logical (hence the so called "issue of scaling"), and
54 measures should be taken to eliminate it from phylogenetic inference. We found that
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standardizing continuous characters to a common range is capable of doing so, while the use of implied weighting with such purposes is only partially successful. Furthermore, the strong correlation between the fit of a character and its scale when the rescaling step is avoided (Fig. 4) interfered with the use of implied weighting for its true purpose, to weight characters according to their homoplasy (Goloboff, 1993). This degree of homoplasy should not, once again, be influenced by a character's scale, since such factor has nothing to do with the degree of discordance between the character and a tree, nor with the lack of hierarchical structure in the character's state transformations. Therefore, characters are only weighted according to their homoplasy after they have been rescaled. When this is done, fit and scale no longer correlate (Fig. 4), and characters with similar original scales receive widely different weights (differences in fit after rescaling were up to 50 times larger than before doing so when comparing pairs of characters with similar scales). Furthermore, we found that such procedure resulted in a higher topological congruence between the discrete and continuous datasets, as well as formulating phylogenetic hypothesis with larger values of group support. Based on all the aforementioned evidence, we advocate that continuous characters should be rescaled before their use in cladistic analysis. We found that this procedure leads to the formulation of phylogenetic hypothesis that are unaffected by the scale in which characters were coded, a factor that is both arbitrary and irrelevant with respect to both the weight a character should receive and its influence on the resulting phylogeny. Rescaling continuous characters also allows for a subsequent use of implied weighting in which weights are assigned exclusively depending on the amount of homoplasy a character presents, a practice that has been shown to improve phylogenetic analyses of morphological datasets and increase the congruence of characters (Goloboff, 1997; Ramírez, 2003; Goloboff *et al.*, 2008a; as well as the present study).

Phylogeny of the Neriidae

After defining the strategy under which characters were used, we performed a sensitivity analysis (Wheeler, 1995; Giribet, 2003) to see which groups were present under different values of k and different criteria for the recognition of sexually dimorphic characters. None of these two parameters are likely to have a "correct" value, so instead of looking for one we decided to determine which groups are less dependent on such decision (as defended by Goloboff *et al.*, 2008a). The 30 different combinations of parameters explored resulted in highly similar trees, with only a few groups being resolved differently among cladograms. Moreover, these groups were mostly the result of differences in placement of single species, with higher-level clades remaining stable (see NT values in Fig. 8).

The monophyly of the family Neriidae was always recovered, and proved to be strongly supported by the data. Many of the synapomorphies retained reflect changes in the

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3 morphology of the head and antennae (Appendix 1), among which are an elongated head
4 with a larger postcranial region and a prolonged superior region of the mesofacial plate,
5 porrect and elongated antennae with a longer scape and pedicel, the presence of an inner
6 process of the pedicel, an apical positioning of the arista, which presents long pubescence
7 and thickened and differentially colored basal flagellomeres, convergent postvertical
8 bristles, a reduction/absence of anterior notopleural bristles and yellow colored procoxae.
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12 The first species to branch off are always *Telostylus binotatus* and *Gymnonerius*
13 *fuscus*. Both genera inhabit exclusively the Australian-Oriental regions, with their ranges of
14 distribution including the Malay Peninsula, Indochina and most of the islands of the Malay
15 Archipelago. The genus *Telostylus* Bigot is composed of 11 species (of which *Tl. binotatus*
16 is the type species) and was already considered by Aczél (1954a; 1955a) to include some of
17 the most basal neriids. On the other hand, the genus *Gymnonerius* Hendel includes only a
18 single species, *Gy. fuscus*. This species shows extensive variability in both coloration and
19 body proportions, which led Hennig (1937) to create numerous subspecies (Steyskal,
20 1977). Aczél (1955a) did not follow this opinion, arguing that a reexamination of a
21 significant amount of specimens was needed before modifying the taxonomy of the genus.
22 He also considered *Gymnonerius* to be morphologically very different from all other neriids
23 of the Australian and Oriental regions, proposing this was due to its derived nature which
24 brought it closer to the Neotropical neriids. Although in most trees these two genera branch
25 off successively, in two of them they form a monophyletic group, supported by an
26 elongated postcranium, reduced third costal section, shorter preabdomen, a reduction in the
27 number of fronto-orbital bristles, a white antennal arista and long non-apical scutellar
28 bristles. We did not choose this hypothesis given its low frequency in the sensitivity
29 analysis, although the possibility for such basal monophyletic clade remains open for
30 subsequent studies.
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39 The remaining of the Australian-Oriental fauna, represented in the analysis by the
40 genera *Telostylinus*, *Indonesicesa*, and *Paranerius* Bigot, was obtained as a complex group
41 of early and poorly-resolved lineages. Despite the lack of strong support for most of these
42 clades, and the change in position of several species among the obtained trees, some
43 conclusions can be advanced. First of all, evidence is strong with respect to the monophyly
44 of the genus *Indonesicesa*, a clade that was obtained among all trees with high levels of
45 support (for synapomorphies see Appendix 1). All trees also suggested a close relationship
46 between this genus and *Paranerius*, the two of them forming a monophyletic group in most
47 of them. Both genera inhabit the western regions of the island of New Guinea (Aczél,
48 1954a; Pitkin, 1989), and share several synapomorphies of the female sex (larger size,
49 shorter postcrania, smaller fourth costal section, wider ovipositor, 2-3 dorsal bristles in the
50 procoxae), as well as a brown frontal vitta and short and spiniform bristles. On the other
51 hand, the genus *Telostylinus* is never obtained as a monophyletic group, consisting instead
52 of several clades whose exact number and composition differs among the different trees.
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3 The pattern nonetheless shows a striking biogeographic consistency, with the early
4 branches (*T. papuanus* and *T. spinicoxa*, as well as the two species that group with
5 *Indonesicesa* and *Paraneri*, *T. zonalis* and *T. longipennis*) restricted to the island of New
6 Guinea, while the more derived species form a monophyletic group (clade D of Figure 8,
7 which includes the type species of the genus, *T. lineolatus*) whose distribution is almost
8 entirely Micronesian (except for *T. lineolatus* which is widespread through the entire
9 Southeast Asia and Oceania; Aczél, 1954a; 1955a; Pitkin, 1989). Further study is needed to
10 precisely determine natural groups among this morphologically and geographically
11 heterogeneous group.
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16 The relationships among the remaining groups of neriids were more resolved and
17 many of the groups obtained showed higher levels of stability and support (Fig. 8). We
18 recovered a single origin for all species inhabiting outside the Australian-Oriental regions.
19 This (almost entirely) Afrotropical + Neotropical clade (clade E) was present in 100% of
20 trees, yet it is not taxonomically defined here due to poor values of jackknifing frequencies.
21 Synapomorphies of this group include an enlargement of the frontal and genal regions of
22 the head, longer and taller thorax, several changes in wing venation, a reduction in femoral
23 length, thinner epandria, triangular and thin inner processes of the pedicel, the presence of
24 anterior notopleural bristles and strong rows of spines on the male forecoxae. This group is
25 further subdivided into the genus *Chaetonerius*, including all of the African fauna as well
26 as some species from the Oriental region, of which only *Ch. inermis* is included in the
27 analysis (Steyskal, 1977; Steyskal, 1980; Pitkin, 1989; Barraclough, 1993a); and the entire
28 American fauna (clade F), including all species from both Neotropical and Nearctic regions
29 (Steyskal, 1968; 1987; Buck, 2010). The two clades are strongly supported, and defining
30 synapomorphies are listed in Appendix 1. Aczél's insistence on the condition of
31 *Chaetonerius* as a "phylogenetically homogenous group" (Aczél, 1955b: 9) is therefore
32 confirmed, as well as his subdivision of the genus into a *Ch. apicalis* – *Ch. collarti* – *Ch.*
33 *ghesquierei* group and one including *Ch. brachialis* and *Ch. latifemur*.
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42 Among neriids from the Americas, species are divided into two main groups, both
43 of which include all American genera contained in the *Eoneria* and *Nerius*-groups defined
44 by Aczél. We therefore propose to restrict these names to the Neotropical-Nearctic neriids.
45 Defined in such a way, the *Eoneria*-group (clade G), including the genera *Antillonerius*,
46 *Eoloxozus* and *Eoneria* is supported by an elongated head, a shorter postpedicel, an
47 increase in the number of fronto-orbital bristles, the presence of katepisternal bristles and
48 the continuation of the mesonotal pruinosity into the scutellum. Within this group, the basal
49 position of *Antillonerius* is strongly inferred, while the relationships between *Eoneria*
50 Aczél and *Eoloxozus* Aczél would imply the need to synonymize both genera. However, a
51 conservative approach is adopted, since the clade conformed by *Eoneria maldonadoi* and
52 *Eoloxozus sabroskyi* shows very low values of Bremer support. On the other hand, the
53 *Nerius*-group (clade H), including the genera *Nerius*, *Longina*, *Odontoloxozus*, *Cerantichir*
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3 and *Glyphidops*, is supported by a strong increase in female size, an enlargement of the
4 region of the thorax before the transverse suture, several changes in wing venation, a
5 polished and shiny dorsal surface of the antennal bases, a reduction in the length of bristles
6 (which become spiniform), the lack of occipital bristles, and a reduction in the number of
7 bristles of both basicosta and male procoxae. The clade is further subdivided into the genus
8 *Glyphidops* on one side, the most diverse of all Neotropical genera (Steyskal, 1968), and
9 the remaining genera on the other (clade I). Aczél (1961) first proposed the close
10 relationship between the genera *Glyphidops* and *Oncopsia* Enderlein (as defined by
11 Enderlein, 1922), including both as subgenera of the former. Although the decision to
12 include both in a single genus is validated by our results, we do not obtain the subgenera as
13 monophyletic clades. In fact, the dense and whitish antennal pubescence that defined the
14 subgenus *Glyphidops* (Aczél, 1961) is shown to be the result of at least two events of
15 convergence. We consequently eliminate the subgeneric divisions within *Glyphidops*, with
16 the subgenus *Oncopsia* becoming a junior synonym of the subgenus *Glyphidops*.

23 On the other hand, the relationships between the genera *Odontoloxozus*, *Longina*
24 and *Cerantichir* are the only ones that have received recent attention. Buck and Marshall
25 (2004) proposed the existence of a monophyletic clade including *Longina* and *Cerantichir*,
26 citing the bare regions at the base of wings and the strong suprahumeral (first dorsocentral)
27 bristles as possible synapomorphies. Buck (2010) afterwards transferred *Od. peruanus* to
28 the genus *Cerantichir*, given the common apomorphic condition of an elongated
29 antepronotal ridge and scutum which end beyond the level of humeral carina. Nonetheless,
30 as discussed by Sepúlveda *et al.* (2013a), the genus *Longina* also shares this peculiar
31 configuration of the anterior region of the thorax, and the other characteristics used to
32 describe the genus *Cerantichir* by Buck (2010) are either also present in *Longina*, or absent
33 from *C. peruana*. Aczél (1961) on the other hand, favored a close relationship between
34 *Longina* and *Odontoloxozus*. Our results strongly confirm the monophyly of a *Cerantichir*
35 + *Odontoloxozus* + *Longina* group (clade J), as well as the validity and derived status
36 within the group of the genus *Longina* (Fig. 8). On the other hand, the monophyly of the
37 genus *Cerantichir* as currently defined was never obtained. From all the combination of
38 parameters analyzed, 80% of conditions resulted in the groupings shown in Fig. 8, with *C.*
39 *enderleini* as the first species to branch off followed by a clade that divides into the
40 *Longina* and a *C. peruana* + *Od. longicornis* group. This topology conflicts with Buck's
41 (2010) reassignment. However, 20% of trees supported the following different
42 configuration: (*Od. longicornis*, (*C. peruana*, (*C. enderleini*, (*Longina*))). In such case,
43 placing *C. peruana* in either genus would result in non-natural groups. Once again, a
44 conservative approach is favored, without modifying the taxonomy of the involved genera
45 until their relationships are completely clarified.

55 The phylogeny resulting from our analysis has little in common with the taxonomic
56 higher-level divisions of the family as proposed by Aczél (1961). The basal split between
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3 the subfamilies Neriinae and Telostylinae is conclusively rejected. In fact, the lack of
4 antennal bases, a character supposedly uniting the Telostylinae (Enderlein, 1922; Aczél,
5 1961) is determined to be a convergence between the genus *Telostylus*, arising as the most
6 basal split within the family, before the evolutionary origin of the structure, and the genus
7 *Chaetonerius* which have secondarily lost them. Furthermore, the polished and shiny dorsal
8 region of these antennal bases, the character uniting the *Nerius*-group *sensu* Aczél (in
9 contraposition to the dull antennal bases of the *Eoneria*-group) is found to have originated
10 three times independently, in the lineages leading to *Gymnonerius*, *Paranerius* and finally
11 to all Neotropical species presenting such character. Our phylogenetic hypothesis is also
12 more congruent with the biogeography of the neriids. The scheme of relationships derived
13 from Aczél's taxonomy (Fig. 1) proposed a much more complex biogeographic history,
14 uniting in the same groups genera from many different continents. Aczél only referred
15 twice to the question of the geographic origin of the family: first, stating that "We still have
16 no idea where on Earth's surface branched off this family from a common root" (Aczél,
17 1954a: p. 507), and once again a few months later, when apparently he became convinced
18 that the presence of the entire subfamily Telostylinae in the Oriental region was evidence
19 supporting that region as the family's "primary center of distribution" (Aczél, 1954b: p. 2).
20 Our results confirm this, since all species obtained as early branchings are restricted to
21 Southeast Asia. From that ancestral region, several species of *Telostylinus* colonized the
22 Oceanic islands, while their sister group divided into the mainly African *Chaetonerius* and
23 the American fauna.
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33 The lack of information regarding the phylogenetic relationships within Neriidae
34 contrasts dramatically with the otherwise thorough knowledge of both higher and lower
35 level phylogeny of many groups of Diptera (see Lambkin *et al.*, 2013 for a summary of
36 phylogenetic studies of dipterans), a pattern that may well be derived from Hennig himself
37 being a dipterologist (Richter and Meier, 1994; Meier, 2005). We provide here the first
38 phylogenetic reconstruction of this family, one that is mainly the legacy of Martín L. Aczél.
39 His thorough morphological descriptions and taxonomic revisions allowed for the
40 construction of a detailed morphological matrix with which to elucidate the evolutionary
41 history of neriid flies. Furthermore, his insistence in the use of continuous characters, as
42 well as his systematic registry of intraspecific variation allowed us to overpass commonly
43 cited obstacles towards the elucidation of a neriid phylogeny (Buck, 2010), resulting in the
44 retention of a large amount of synapomorphies derived from body proportions, chaetotaxy
45 and even male genitalia (Appendix 1). It is fair to say that Aczél has once again proven "the
46 fundamental importance of morphology for entomology" (Aczél, 1951: p. 483).
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19 References

20
21 Abdala, C. S., Juárez Heredia, V. I. 2013. Taxonomía y filogenia de un grupo de lagartos
22 amenazados: el grupo de *Liolaemus anomalus* (Iguania: Liolaemidae). Cuad. Herpetol. 27,
23 109–153.
24

25
26 Aczél, M. L. 1951. Morfología externa y división sistemática de las «Tanypezidiformes»
27 con sinopsis de las especies argentinas de «Tylidae» («Micropezidae») y «Neriidae»
28 (Dipt.). Acta Zool. Lilloana 11, 483–589.
29

30
31 Aczél, M. L. 1954a. Results of the Archbold Expedition: Neriidae von Neuguinea
32 (Diptera). Treubia 22, 505–531.
33

34
35 Aczél, M. L. 1954b. Neriidae of the Belgian Congo (Diptera, Acalyptratae). Bull. Inst. R.
36 Sc. N. B. 30, 1–23.
37

38
39 Aczél, M. L. 1954c. Neriidae in the collection of the Musée royal du Congo belge. Rev.
40 Zool. Bot. Afr. 49, 161–166.
41

42
43 Aczél, M. L. 1955a. Neriidae von Indonesien (Dipt. Acalyptratae). Treubia 23, 19–40.
44

45
46 Aczél, M. L. 1955b. Neriidae (Diptera, Acalyptrata). Exploration du Parc National de l'
47 Upemba. I. Mission G. F. de Witte 38, 85–92.
48

49
50 Aczél, M. L. 1955c. Neriidae in the collections of the Musée Royal du Congo Belge,
51 Tervuren (Supplement). Rev. Zool. Bot. Afr. 51, 1–2.
52

53
54 Aczél, M. L. 1959. Diptera: Neriidae and Micropezidae (Tylidae). Insects of Micronesia 14,
55 47–90.
56

57
58 Aczél, M. L. 1961. A revision of American Neriidae (Diptera, Acalyptratae). Studia Ent. 4,
59 257–346.
60

- 1
2
3 Albrecht, G. H. 1978. Some comments on the use of ratios. *Syst. Zool.* 27, 67–71.
4
5
6 Archie, J. 1985. Methods for coding variable morphological features for numerical
7 taxonomic analysis. *Syst. Zool.* 34, 326–345.
8
9 Atchley, W. R. 1978. Ratios, regression intercepts, and the scaling of data. *Syst. Zool.* 27,
10 78–83.
11
12 Atchley, W. R., Gaskins, C. T., Anderson, D. 1976. Statistical properties of ratios. I.
13 Empirical results. *Syst. Zool.* 25, 137–148.
14
15
16 Bardin, J., Rouget, I., Yacobucci, M., Cecca, F. 2013. Increasing the number of discrete
17 character states for continuous characters generates well-resolved trees that do not reflect
18 phylogeny. *Integr. Zool.* <http://dx.doi.org/10.1111/1749-4877.12076>.
19
20
21 Barraclough, D. A. 1993a. The southern African species of Neriidae (Diptera). *Ann. Natal*
22 *Mus.* 34, 1–17.
23
24 Barraclough, D. A. 1993b. Review of the type material of African *Chaetonerius* species
25 (Diptera: Neriidae), with lectotype designations and new synonymy. *J Afr. Zool.* 107, 269–
26 278.
27
28
29 Baum, B. 1988. A simple procedure for establishing discrete characters from measurement
30 data, applicable to cladistics. *Taxon* 37, 63–70.
31
32
33 Baur, H., Leuenberger, C. 2011. Analysis of ratios in multivariate morphometry. *Syst. Biol.*
34 60, 813–825.
35
36
37 Bonduriansky, R. 2006. Convergent evolution of sexual shape dimorphism in Diptera. *J.*
38 *Morphol.* 267, 602–611.
39
40 Bonduriansky, R. 2007. The evolution of condition-dependent sexual dimorphism. *Am.*
41 *Nat.* 169, 9–19.
42
43
44 Bonduriansky, R. 2009. Condition dependence of developmental stability in the sexually
45 dimorphic fly *Telostylinus angusticollis* (Diptera: Neriidae). *J. Evol. Biol.* 22, 861–872.
46
47
48 Bordewich, M., Semple, C. 2005. On the computational complexity of the rooted subtree
49 prune and regraft distance. *Ann. Combinatorics* 8, 409–423.
50
51
52 Bremet, K. 1988. The limits of amino acid sequence data in angiosperm phylogenetic
53 reconstruction. *Evolution* 42, 795–803.
54
55
56 Bremer, K. 1994. Branch support and tree stability. *Cladistics* 10, 295–304.
57
58
59
60

1
2
3 Buck, M. 2010. Neriidae. In: Brown, B.V., Borkent, A., Cumming, J.M., Wood, D.M.,
4 Woodley, N.E., Zumbado, M.A. (Eds.), Manual of Central American Diptera, Volume II.
5 NRC Research Press, Ottawa, 56, pp. 815–819.
6
7

8 Buck, M, Marshall, S. A. 2004. A review of the genus *Longina* Wiedemann, with
9 descriptions of two new species (Diptera, Neriidae). Stud. Dipterol. 11, 23–32.
10

11 Buck, M., McAlpine, D. K. 2010. Pseudopomyzidae. In: Brown, B.V., Borkent, A.,
12 Cumming, J.M., Wood, D.M., Woodley, N.E., Zumbado, M.A. (Eds.), Manual of Central
13 American Diptera, Volume II. NRC Research Press, Ottawa, 57, pp. 821–825.
14
15

16 Carvalho-Filho, F. S., Esposito, M. C. 2008. Neriidae (Diptera: Schizophora) of the
17 Brazilian Amazon: New records of genera and species, and key to species. Neotrop.
18 Entomol. 37, 58–62.
19
20

21 Chappill, J. 1989. Quantitative characters in phylogenetic analysis. Cladistics 5, 217–234.
22

23 Colless, D. 1980. Congruence between morphometric and allozyme data for *Menidia*
24 species: A reappraisal. Syst. Zool. 29, 288–299.
25
26

27 Corruccini, R. S. 1977. Correlation properties of morphometric ratios. Syst. Zool. 26, 211–
28 214.
29
30

31 Cox, C. B. 2001. The biogeographic regions reconsidered. J. Biogeogr. 28, 511–523.
32

33 Cranston, P., Humphries, C. 1988. Cladistics and computers: a chironomid conundrum?
34 Cladistics 4, 72–92.
35
36

37 Cresson, E. T. 1930. Notes and descriptions of some neotropical Neriidae and
38 Mycropezidae. T. Am. Entomol. Soc. 56, 307–362.
39
40

41 Crisp, M., Weston, P. 1987. Cladistics and legume systematics, with an analysis of the
42 Bossiaceae, Brongniartieae and Mirbelieae. In: Stirton, C. (Ed.), Advances in Legume
43 Systematics, Part 3. Royal Botanical Gardens, Kew, pp. 65–130.
44

45 de Bivort, B., Clouse, R. M., Giribet, G. 2010. A morphometrics-based phylogeny of the
46 temperate Gondwanan mite harvestmen (Opiliones, Cyphophthalmi, Pettalidae). J. Zool.
47 Syst. Evol. Res. 48, 294–309.
48
49

50 DeGusta, D. 2004. A method for estimating the relative importance of characters in
51 cladistic analyses. Syst. Biol. 53, 529–532.
52
53

54 Eberhard, W. G. 1998. Reproductive behavior of *Glyphidops flavifrons* and *Nerius*
55 *plurivittatus* (Diptera, Neriidae). J. Kansas Entomol. Soc. 71, 89–107.
56
57

58 Enderlein, G. von. 1922. Klassifikation der Mikropeziden. Arch. Naturgesch. 88, 140–229.
59
60

- 1
2
3 Escapa, I. H., Catalano, S. A. 2013. Phylogenetic analysis of Araucariaceae: Integrating
4 molecules, morphology, and fossils. *Int. J. Plant Sci.* 174, 1153–1170.
5
6
7 Eulenstein, O., Chen, D., Burleigh, J. G., Fernández-Baca, D., Sanderson, M. J. 2004.
8 Performance of flip-supertree construction with a heuristic algorithm. *Syst. Biol.* 53, 1–10.
9
10 Farris, J. S. 1989. The retention index and the rescaled consistency index. *Cladistics* 5,
11 417–419.
12
13 Farris, J. S. 1990. Phenetics in camouflage. *Cladistics* 6, 91–100.
14
15 Farris, J. S., Albert, V. A., Källersjö, M., Lipscomb, D., Kluge, A. G. 1996. Parsimony
16 jackknifing outperforms neighbor-joining. *Cladistics* 12, 99–124.
17
18 Farris, J. S., Källersjö, M., Kluge, A. G., Bult, C. 1995. Testing significance of
19 incongruence. *Cladistics* 10, 315–319.
20
21 Felsenstein, J. 1988. Phylogenies and quantitative characters. *Ann. Rev. Ecol. Syst.* 19,
22 445–471.
23
24 Fink, W. L., Zelditch, M. L. 1995. Phylogenetic analysis of ontogenetic shape
25 transformations: a reassessment of the piranha genus *Pygocentrus* (Teleostei). *Syst. Biol.*
26 44, 343–360.
27
28 Gift, N., Stevens, P. F. 1997. Vagaries in the delimitation of character states in quantitative
29 variation - an experimental study. *Syst. Biol.* 46, 112–125.
30
31 Giribet, G. 2003. Stability in phylogenetic formulations and its relationship to nodal
32 support. *Syst. Biol.* 52, 554–564.
33
34 Giribet, G. 2007. Efficient tree searches with available algorithms. *Evol. Bioinform.* 3, 1–
35 16.
36
37 Goloboff, P. A. 1993. Estimating character weights during tree search. *Cladistics* 9, 83–91.
38
39 Goloboff, P. A. 1997. Self-weighted optimization: tree searches and character state
40 reconstructions under implied transformation costs. *Cladistics* 13, 225–245.
41
42 Goloboff, P. A. 1999. Analyzing large data sets in reasonable times: solutions for
43 composite optima. *Cladistics* 15, 415–428.
44
45 Goloboff, P. A. 2002. Techniques for analyzing large data sets. In: DeSalle, R., Giribet, G.,
46 Wheeler, W.C. (Eds.), *Techniques in Molecular Systematics and Evolution*. Birkhäuser,
47 Basel, pp. 70–79.
48
49 Goloboff, P. A. 2007. Calculating SPR distances between trees. *Cladistics* 23, 1–7.
50
51
52
53
54
55
56
57
58
59
60

1
2
3 Goloboff, P. A., Carpenter, J. M., Salvador Arias, J., Miranda Esquivel, D. R. 2008a.
4 Weighting against homoplasy improves phylogenetic analysis of morphological data sets.
5 Cladistics 24, 758–773.
6

7
8 Goloboff, P. A., Farris, J. S., Nixon, K. C. 2008b. TNT, a free program for phylogenetic
9 analysis. Cladistics 24, 774–786.
10

11 Goloboff, P. A., Mattoni, C. I., Quinteros, A. S. 2006. Continuous characters analyzed as
12 such. Cladistics 22, 589–601.
13

14
15 Gordon, A. 1980. On the assessment and comparison of classifications. In: Tommassone,
16 R. (Ed.), Analyse de Données et Informatique. INRIA, LeChesnay, pp. 149–160.
17

18
19 Griffiths, G. C. D. 1972. The phylogenetic classification of Diptera Cyclorrhapha, with
20 special reference to the structure of the male postabdomen. Series Entom. 8, III.
21

22
23 Haas, A. 2003. Phylogeny of frogs as inferred from primarily larval characters (Amphibia:
24 Anura). Cladistics 19, 23–89.
25

26
27 Hendel, F. 1922. Die paläarktischen Muscidae acalyptratae Girsch. = Haplostomata Frey
28 nach ihren Familien und Gattungen. I. Die Familien (Anm.: 1. Teil). Konowia 1, 145–160.
29

30
31 Hennig, W. 1934. Zur Kenntnis d. Kopulationsorgane de Tyliden. Zool. Anz. 107, 67–76.

32
33 Hennig, W. 1936. Beziehungen zwischen geographischer Verbreitung und Systematischer
34 Gliederung bei einigen Dipterenfamilien. Ein Beitrag zum Problem der Gliederung system.
35 Kategorien höherer Ordnung. Zool. Anz. 116, 161–175.
36

37
38 Hennig, W. 1937. Übersichtüber die Arten der Neriiden und über die Zoogeographie
39 dieser Acalyptraten-Gruppe. Stettin. Ent. Ztg. 98, 240–280.

40
41 Hills, M. 1978. On ratios - a response to Atchley, Gaskins, and Anderson. Syst. Zool. 27,
42 61–62.
43

44
45 Hormiga, G., Scharff, N., Coddington, J. A. 2000. The phylogenetic basis of sexual size
46 dimorphism in orb-weaving spiders (Araneae, Orbiculariae). Syst. Biol. 49, 435–462.
47

48
49 Hornung-Leoni, C. T., Sosa, V. 2008. Morphological phylogenetics of *Puya* subgenus *Puya*
(Bromeliaceae). Bot. J. Linn. Soc. 156, 93–110.
50

51
52 Kluge, A. G., Farris, J. S. 1969. Quantitative phyletics and the evolution of anurans. Syst.
53 Zool. 18, 1–32.
54

55
56 Lambkin, C. L., Sinclair, B. J., Pape, T., Courtney, G. W., Skevington, J. H., Meier, R.,
57 Yeates, D. K., Blagoderov, V., Wiegmann, B. M. 2013. The phylogenetic relationships
58
59
60

1
2
3 among infraorders and superfamilies of Diptera based on morphological evidence. *Syst.*
4 *Entomol.* 38, 164–179.

5
6
7 Lopardo, L. 2005. Phylogenetic revision of the genus *Negayan* (Araneae, Anyphaenidae,
8 Amaurobioidinae). *Zool. Scr.* 34, 245–277.

9
10 Lopardo, L., Giribet, G., Hormiga, G. 2011. Morphology to the rescue: molecular data and
11 the signal of morphological characters in combined phylogenetic analyses – a case study
12 from mysmenid spiders (Araneae, Mysmenidae), with comments on the evolution of web
13 architecture. *Cladistics* 27, 278–330.

14
15
16 MacLeod, N. 2002. Phylogenetic signals in morphometric data. In: Macleod, N., Forey,
17 P.L. (Eds.), *Morphology, Shape and Phylogeny*. Taylor and Francis, London, pp. 100–138.

18
19
20 Mangan, R. L., Baldwin, D. 1986. A new cryptic species of *Odontoloxozus* (Neriidae:
21 Diptera) from the Cape Region of Baja California Sur (Mexico). *Proc. Ent. Soc. Wash.* 88,
22 110–121.

23
24
25 Mannion, P. D., Upchurch, P., Barnes, N., Mateus, O. 2013. Osteology of the Late Jurassic
26 Portuguese sauropod dinosaur *Lusotitan atalaiensis* (Macronaria) and the evolutionary
27 history of basal titanosauriforms. *Zool. J. Linn. Soc.* 168, 98–206.

28
29
30 Marshall, S. A. 2010. Micropezidae. In: Brown, B.V., Borkent, A., Cumming, J.M., Wood,
31 D.M., Woodley, N.E., Zumbado, M.A. (Eds.), *Manual of Central American Diptera*,
32 Volume II. NRC Research Press, Ottawa, 55, pp. 805–813.

33
34
35 McAlpine, D. K. 1966. Description and biology of an Australian species of
36 Cypselosomatidae (Diptera), with a discussion of family relationships. *Aust. J. Zool.* 14,
37 673–685.

38
39
40 McAlpine, D. K. 1974. The subfamily classification of the Mycropezidae and the genera of
41 Eurybatinae (Diptera: Schizophora). *J. Entomol. Ser. B* 43, 231–245.

42
43
44 McAlpine, D. K. 1996. Relationships and classification of the Pseudopomyzidae (Diptera:
45 Nerioidea). *Proc. Linn. Soc. N. S. W.* 116, 223–232.

46
47
48 McAlpine, J. F. 1987. Cypselosomatidae. In: McAlpine, J.F., Peterson, B.V., Shewell,
49 G.E., Teskey, H.J., Vockroth, J.R., Wood, D.M. (Eds.), *Manual of Nearctic Diptera*,
50 Volume II. Research Branch, Agriculture Canada, Ottawa, Ontario, 55, pp. 757–760.

51
52
53 McAlpine, J. F. 1989. Phylogeny and classification of the Muscomorpha. In: McAlpine,
54 J.F., Wood, D.M. (Eds.), *Manual of Nearctic Diptera*, Volume III. Research Branch,
55 Agriculture Canada, Ottawa, Ontario, pp. 1397–1518.

- 1
2
3 Meier, R. 2005. Role of Dipterology in phylogenetic systematics: the insight of Willi
4 Hennig. In: Yeates, D.K., Wiegmann, B.M. (Eds.), *The Evolutionary Biology of Flies*.
5 Columbia University Press, New York, pp. 45–62.
6
7
8 Mello, R. L. 2010. The Diptera described by Martín L. Aczél. *Stud. Dipterol.* 17, 223–236.
9
10
11 Mickevich, M. F. 1978. Taxonomic congruence. *Syst. Zool.* 27, 143–158.
12
13 Mickevich, M. F., Farris, J. S. 1981. The implications of congruence in *Menidia*. *Syst.*
14 *Zool.* 30, 351–370.
15
16 Mickevich, M. F., Weller, S. J. 1990. Evolutionary character analysis: tracing character
17 change on a cladogram. *Cladistics* 6, 137–170.
18
19
20 Nixon, K. C. 1999. The parsimony ratchet, a new method for rapid parsimony analysis.
21 *Cladistics* 15, 407–414.
22
23
24 Pereyra, V., Mound, L. A. 2009. Phylogenetic relationships within the genus *Cranothrips*
25 (Thysanoptera, Melanthripidae) with consideration of host associations and disjunct
26 distributions within the family. *Syst. Entomol.* 34, 151–161.
27
28
29 Pimentel, R., Riggins, R. 1987. The nature of cladistic data. *Cladistics* 3, 201–209.
30
31 Pitkin, B. R. 1989. Family Neriidae. In: Evenhuis, N.L. (Ed.), *Catalog of the Diptera of the*
32 *Australasian and Oceanian Regions*. Bishop Museum & E. J. Brill, Honolulu, pp. 468–469.
33
34 Prado, A. P. 1984. Family Cypselosomatidae. In: Papavero, N. (Ed.), *A catalogue of the*
35 *Diptera of the Americas South of the United States*. Departamento de Zoologia, Secretaria
36 de Agricultura, São Paulo, pp. 1–2.
37
38
39 Rae, T. 1998. The logical basis for the use of continuous characters in phylogenetic
40 systematics. *Cladistics* 14, 221–228.
41
42
43 Ramírez, M. J. 2003. The spider subfamily Amaurobioidinae (Araneae, Anyphaenidae): a
44 phylogenetic revision at the generic level. *Bull. Am. Mus. Nat. Hist.* 277, 1–262.
45
46 Reid, G., Sidwell, K. 2002. Overlapping variables in botanical systematics. In: Macleod,
47 N., Forey, P.L. (Eds.), *Morphology, Shape and Phylogeny*. Taylor and Francis, London, pp.
48 53–66.
49
50
51 Richter, S., Meier, R. 1994. The development of phylogenetic concepts in Hennig's early
52 theoretical publications (1947–1966). *Syst. Biol.* 43, 212–221.
53
54
55 Rohlf, F. J. 2010. tpsDig version 2.16. <http://life.bio.sunysb.edu/morph/index.html>.
56
57
58
59
60

1
2
3 Sepúlveda, T. A., Pereira-Colavite, A., de Carvalho, C. J. B. 2013a. Revision of the
4 Neotropical genus *Cerantichir* (Diptera: Neriidae) with new records and a key to species.
5 Rev. Colomb. Entomol. 39, 125–131.
6

7
8 Sepúlveda, T. A., Wolf, M. I., de Carvalho, C. J. B. 2013b. Revision of the Neotropical
9 genus *Eoneria* (Diptera: Neriidae) with description of a new species from Colombia.
10 Zootaxa 3636, 245–256.
11

12
13 Shatalkin, A. 1994. Palearctic species of Pseudopomyzidae (Diptera). Russ. Entomol. J. 3,
14 129–145.
15

16
17 Simpson, P., Woehl, R., Usui, K. 1999. The development and evolution of bristle patterns
18 in Diptera. Development 126, 1349–1364.
19

20
21 StatSoft, Inc. 2001. STATISTICA (data analysis software system), version 6.
22 www.statsoft.com
23

24
25 Stevens, P. F. 1991. Character states, morphological variation, and phylogenetic analysis: A
26 review. Syst. Bot. 16, 553–583.
27

28
29 Steyskal, G. C. 1965. Synonymy of the genera *Antillonarius* and *Imrenerius* (Diptera:
30 Neriidae). Proc. Ent. Soc. Wash. 67, 60.
31

32
33 Steyskal, G. C. 1968. Family Neriidae. In: Papavero, N. (Ed.), A Catalogue of Diptera of
34 the Americas South of the United States. Departamento de Zoologia, Secretaria da
35 Agricultura, São Paulo, pp. 1–7.
36

37
38 Steyskal, G. C. 1977. Family Neriidae. In: Delfinado, M.D., Hardy, D.E. (Eds.), A
39 Catalogue of the Diptera of the Oriental Region, Volume III. Suborder Cyclorhapha
40 (excluding Division Aschiza). University Press of Hawaii, Honolulu, pp. 8–11.
41

42
43 Steyskal, G. C. 1980. Family Neriidae. In: Crosskey, R.W. (Ed.), Catalogue of the Diptera
44 of the Afrotropical Region. British Museum (Natural History), London, p. 578.
45

46
47 Steyskal, G. C. 1987a. Neriidae. In: McAlpine, J.F., Peterson, B.V., Shewell, G.E., Teskey,
48 H.J., Vockroth, J.R., Wood, D.M. (Eds.), Manual of Nearctic Diptera, Volume II. Research
49 Branch, Agriculture Canada, Ottawa, Ontario, 57, pp. 769–771.
50

51
52 Steyskal, G. C. 1987b. Mycropezidae. In: McAlpine, J.F., Peterson, B.V., Shewell, G.E.,
53 Teskey, H.J., Vockroth, J.R., Wood, D.M. (Eds.), Manual of Nearctic Diptera, Volume II.
54 Research Branch, Agriculture Canada, Ottawa, Ontario, 56, pp. 761–768.
55

56
57 Strait, D. S., Grine, F. E. 2004. Inferring hominoid and early hominid phylogeny using
58 craniodental characters: the role of fossil taxa. J. Hum. Evol. 47, 399–452.
59
60

1
2
3 Swofford, D. L. 1991. When are phylogeny estimates of molecular and morphological data
4 incongruent? In: Miyamoto, M.M., Cracraft, K. (Eds.), *Phylogenetic Analysis of DNA*
5 *Sequences*. Oxford Univ. Press, New York, pp. 295–333.
6
7

8 Thiele, K. 1993. The Holy Grail of the perfect character: The cladistic treatment of
9 morphometric data. *Cladistics* 9, 275–304.
10

11 Thiele, K., Ladiges, P. Y. 1988. A cladistic analysis of *Angophora* Cav. (Myrtaceae).
12 *Cladistics* 4, 23–42.
13
14

15 Vargas, S., Breedy, O., Guzman, H. M. 2010. The phylogeny of *Pacifigorgia* (Coelenterata,
16 Octocorallia, Gorgoniidae): a case study of the use of continuous characters in the
17 systematics of the Octocorallia. *Zoosystema* 32, 5–18.
18
19

20 Wheeler, D. C. 1995. Sequence alignment, parameter sensitivity, and the phylogenetic
21 analysis of molecular data. *Syst. Biol.* 44, 321–331.
22
23

24 Wiegmann, B. M., Trautwein, M., Winkler, I., Barr, N., Kim, J.-W., *et al.* 2011. Episodic
25 radiations in the fly tree of life. *Proc. Natl. Acad. Sci. USA* 108, 5690–5695.
26
27

28 Wiens, J. J. 2001. Character analysis in morphological phylogenetics: problems and
29 solutions. *Syst. Biol.* 50, 689–699.
30

31 Wiens, J. J. 2004. The role of morphological data in phylogeny reconstruction. *Syst. Biol.*
32 53, 653–661.
33
34

35 Winston, J. E. 1999. *Describing species: Practical taxonomic procedure for biologists*.
36 Columbia University Press, New York.
37

38 Yeates, D. K., Wiegmann, B. M. 2005. Phylogeny and evolution of Diptera: recent insights
39 and new perspectives. In: Yeates, D.K., Wiegmann, B.M. (Eds.), *The Evolutionary Biology*
40 *of Flies*. Columbia University Press, New York, pp. 14–44.
41
42

43 Yeates, D. K., Wiegmann, B. M., Courtney, G. W., Meier, R., Lambkin, C. & Pape, T.
44 2007. Phylogeny and systematics of Diptera: two decades of progress and prospects.
45 *Zootaxa* 1668, 565–590.
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Appendix 1: Synapomorphies

The following table (Table 2) shows the synapomorphic features of selected genera and supra-generic groups retained in the phylogenetic hypothesis shown in Fig. 8. Character number is stated between parentheses. For synapomorphies derived from uncollapsed (that is, dimorphic) continuous characters, the intervening sex is made explicit. Otherwise, no particular sex is mentioned, and both character numbers are separated by a slash. Continuous characters were mapped with their original scales. See Appendix S1 for character description and abbreviations.

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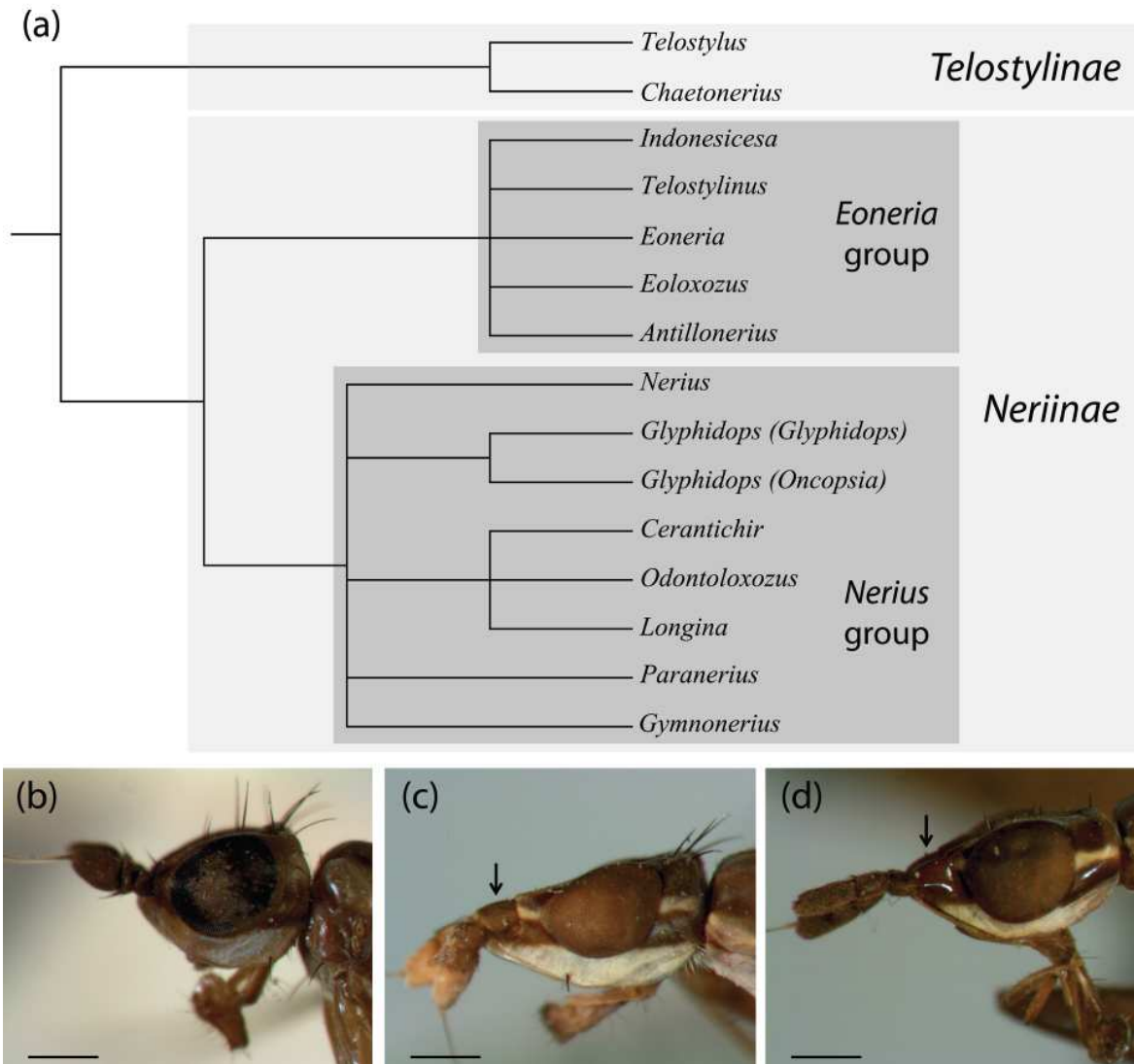


Fig. 1: (a) Current phylogenetic hypothesis within the family Neriidae (only genera included in the present study are shown). (b) *Chaetonerius apicalis* (female), a member of the Telostylinae which lack antennal bases. (c) *Eoneria maldonadio* (female), a member of the Eoneria-group that present antennal bases with a dull dorsal region. (d) *Nerius pilifer* (male), a member of the Nerius-group that present antennal bases with a shiny dorsal region. Black arrows show the position of the antennal bases. Scale bars = 0.5 mm.

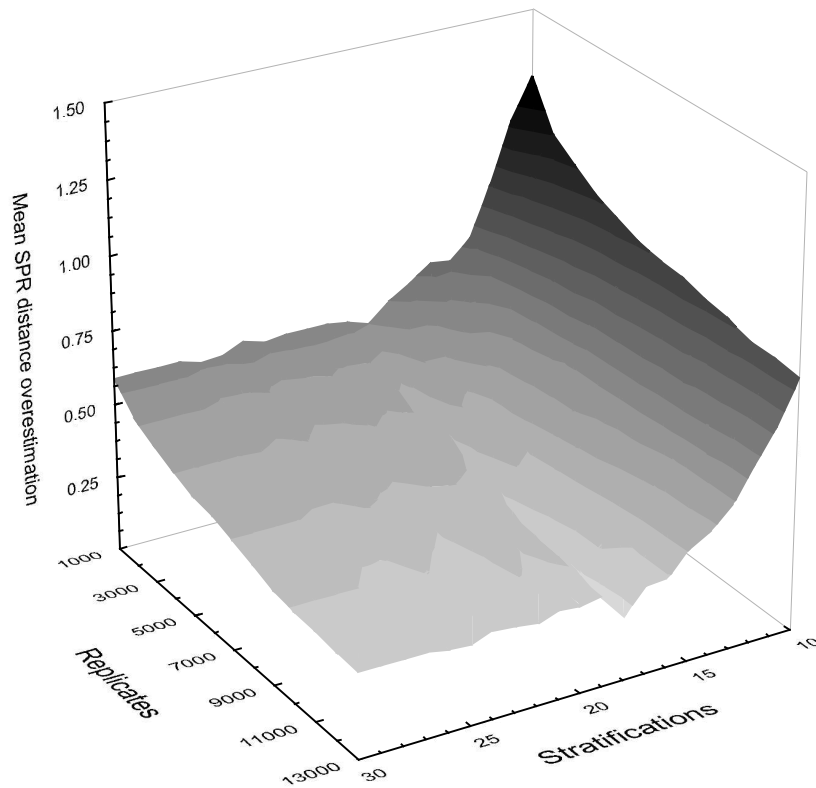


Fig. 2: Efficiency of the weighted SPR distance algorithm ($j = 3$) in finding the minimum distance for several combinations of parameters. The plotted surface shows the mean overestimation of 50 different SPR distances. This was calculated by finding the shortest distance for each one of the 50 pairs of trees, and subtracting that amount to all values obtained for that particular pair (the shortest path for each pair therefore becomes 0, and all other distances are expressed as the positive amount by which they overestimated that same distance). Minimum bias was obtained using 18 stratifications and 13,000 replicates.

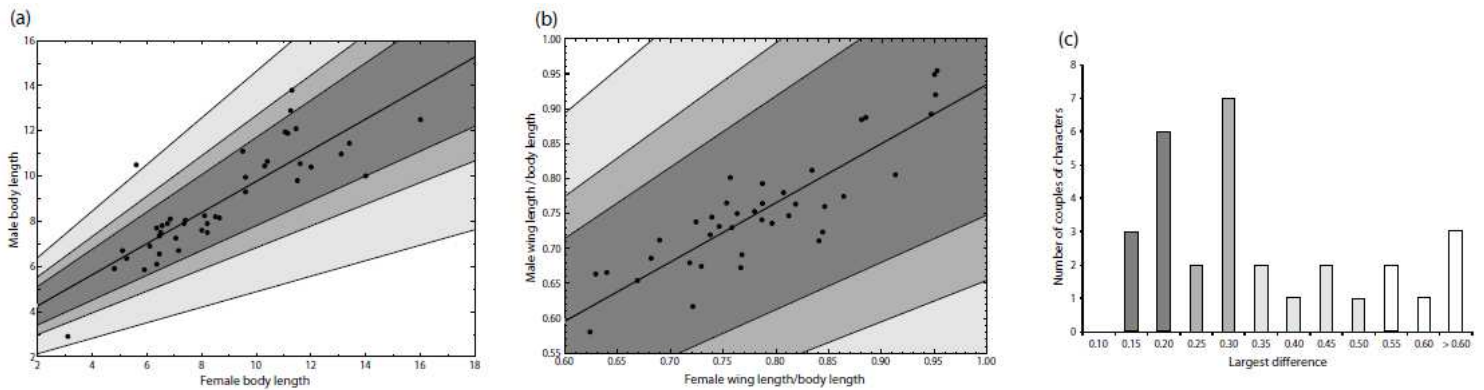


Fig. 3: Independence analysis for male-female couples of characters. **(a, b)** Two examples of the regression analysis performed for all couples of male-female characters. The three stricter criteria for the recognition of sexual dimorphism are shown as more inclusive gray regions. Characters shown in **(a)** (char. numbers 1, 2) are retained as sexually dimorphic for all criteria, while those shown on **(b)** (char. numbers 35, 36) are always collapsed into a single character. **(c)** Histogram grouping all male-female couples of characters into categories (range 0.05) according to the largest difference between observed and expected values, as fraction of the latter, found in the regression analyses. Colors are as in figures **(a)** and **(b)**, and show the number of characters collapsing for each criterion.

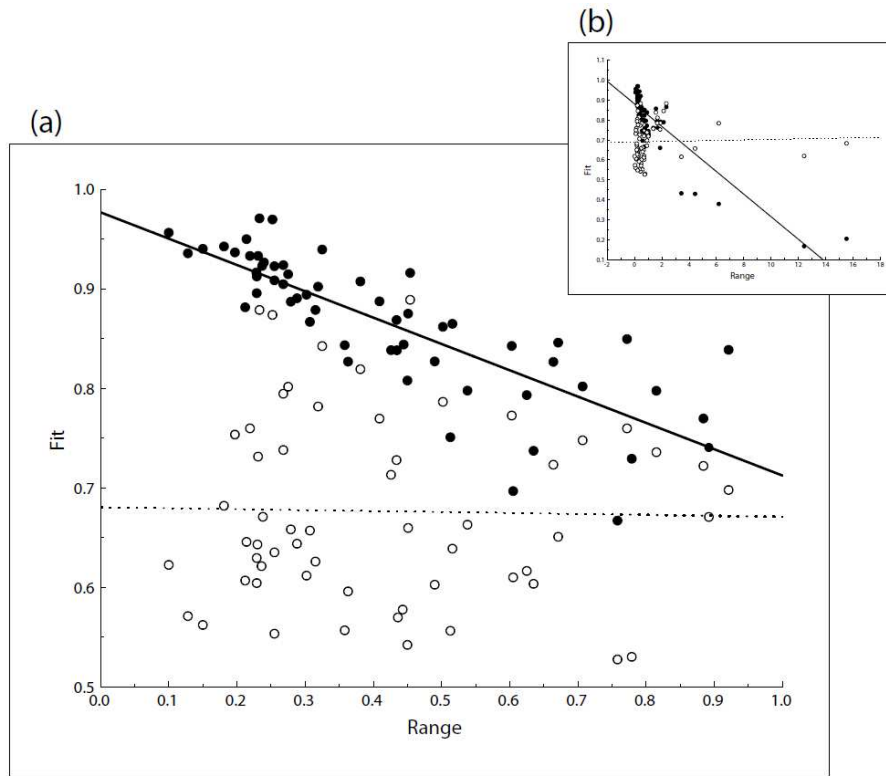


Fig. 4: Regression of the fit assigned by implied weights on the character's range, corresponding to the MPT obtained with (empty circles, dotted line) and without (black circles, solid line) rescaling continuous data. Results are the same whether using the entire continuous dataset (figure **(b)**, statistical values in text), or only characters with ranges < 1 (80% of total characters, figure **(a)**). The significant and strong correlation between both variables ($p < 0.0001$, $R^2 = 0.66$) is lost if implied weighting is applied after rescaling characters ($p = 0.87$, $R^2 < 0.001$).

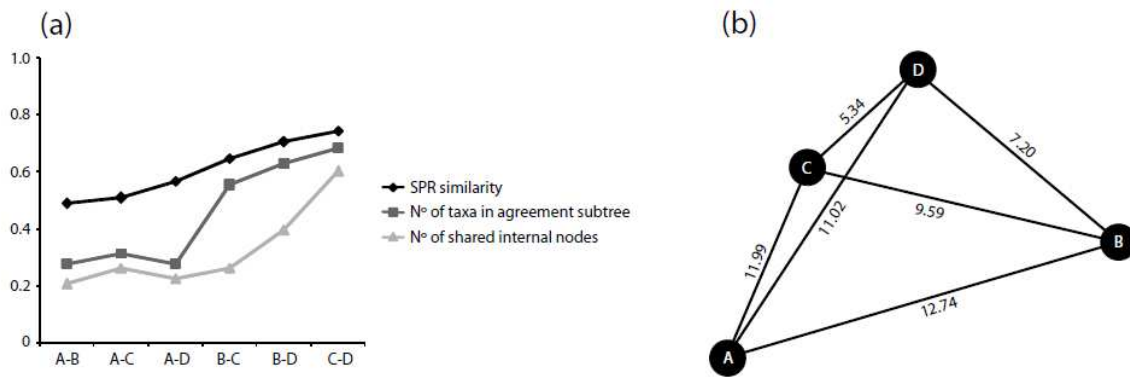


Fig. 5: Topological effect of different strategies of analysis, resulting from the comparison of the resulting MPTs. **(a)** All pair-wise comparisons of MPTs using unweighted SPR derived similarity (black), number of taxa in agreement subtree (dark gray) and number of shared internal nodes (light gray). **(b)** In-scale tetrahedron, with each side representing the weighted SPR distances ($j = 3$) between MPTs (vertex labels according to the strategy employed).

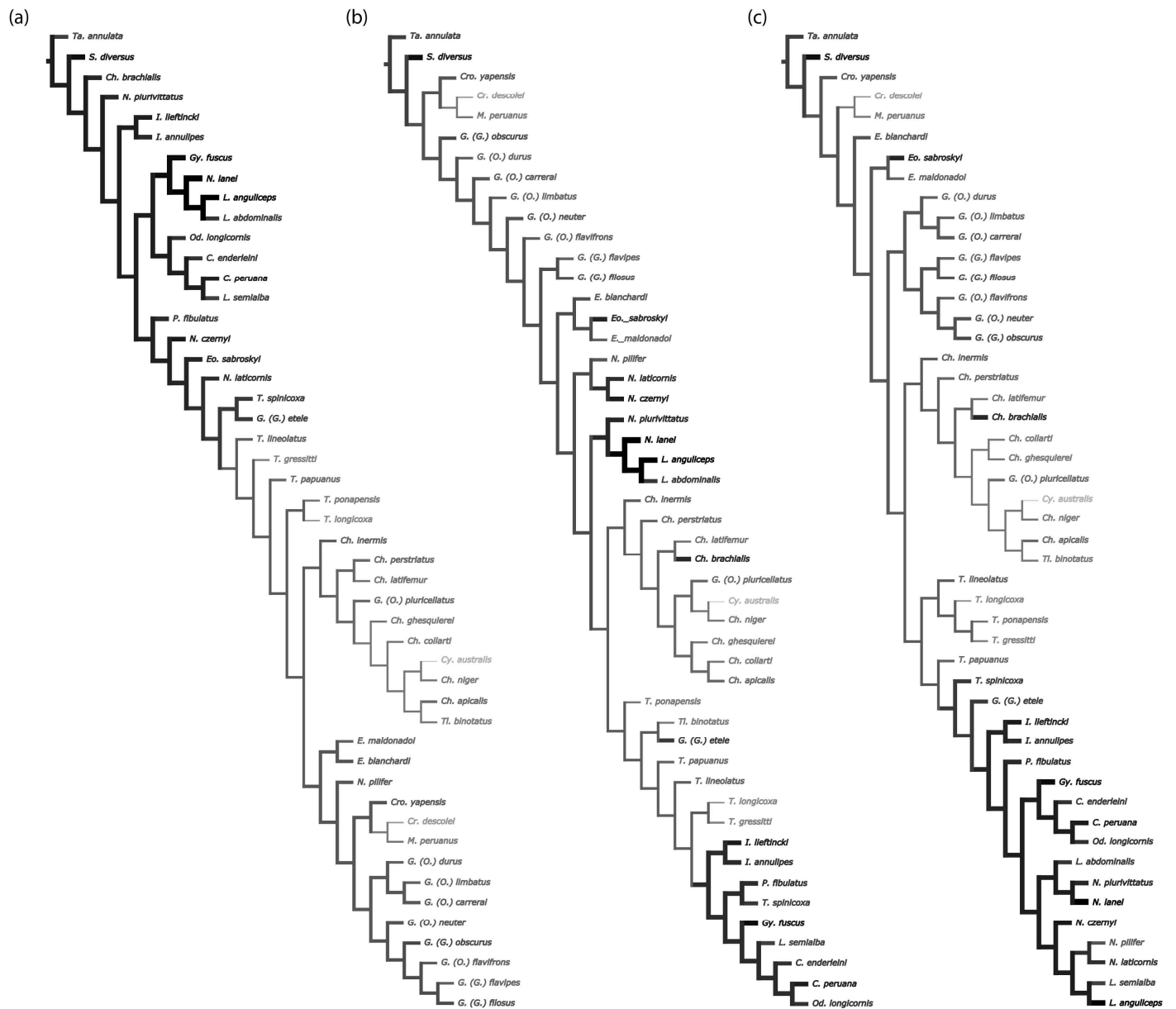


Fig. 6: Single MPTs obtained using the continuous character partition under strategies A (a), B (b) and C (c). The largest character (male body length) has been optimized on each topology, with broader and darker branches representing higher character values. All three topologies differ considerably among each other. Both implied weighting and rescaling reduce the MPT's dependence on the extremely large characters, as seen by the higher homoplasy present in trees (b) and (c).

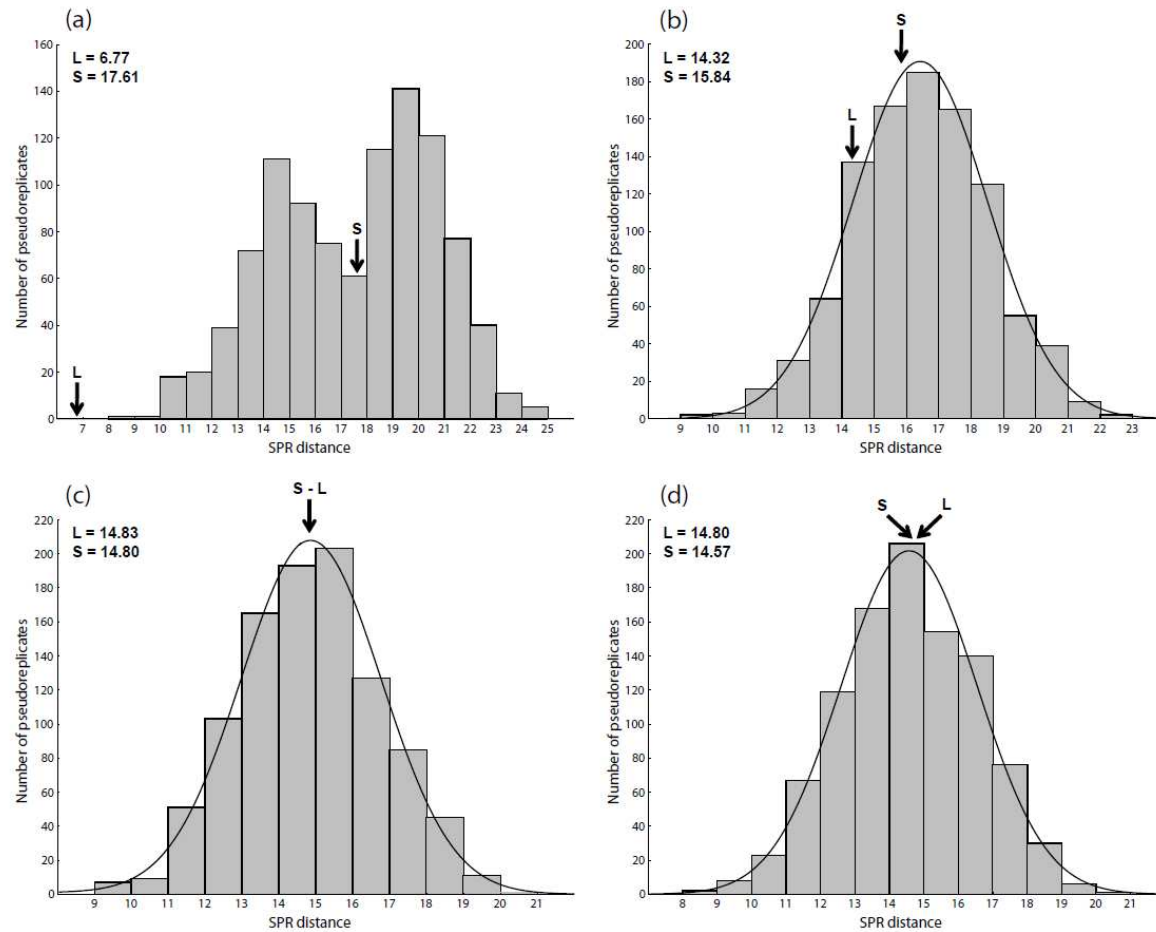


Fig. 7: Distribution of weighted SPR distances comparing 1000 trees obtained from randomly generated subsets of 18 continuous characters, with the MPT from the entire continuous dataset. The letter naming each distribution coincides with the strategy used to obtain it. Distance values are grouped into discrete categories of unit range. A line delineating the inferred normal distribution is shown for those distributions that showed significant fitting. The SPR distances for the large (L) and small (S) character subsets can be seen at the top left corner of each graph, and black arrows show their location on the histogram.

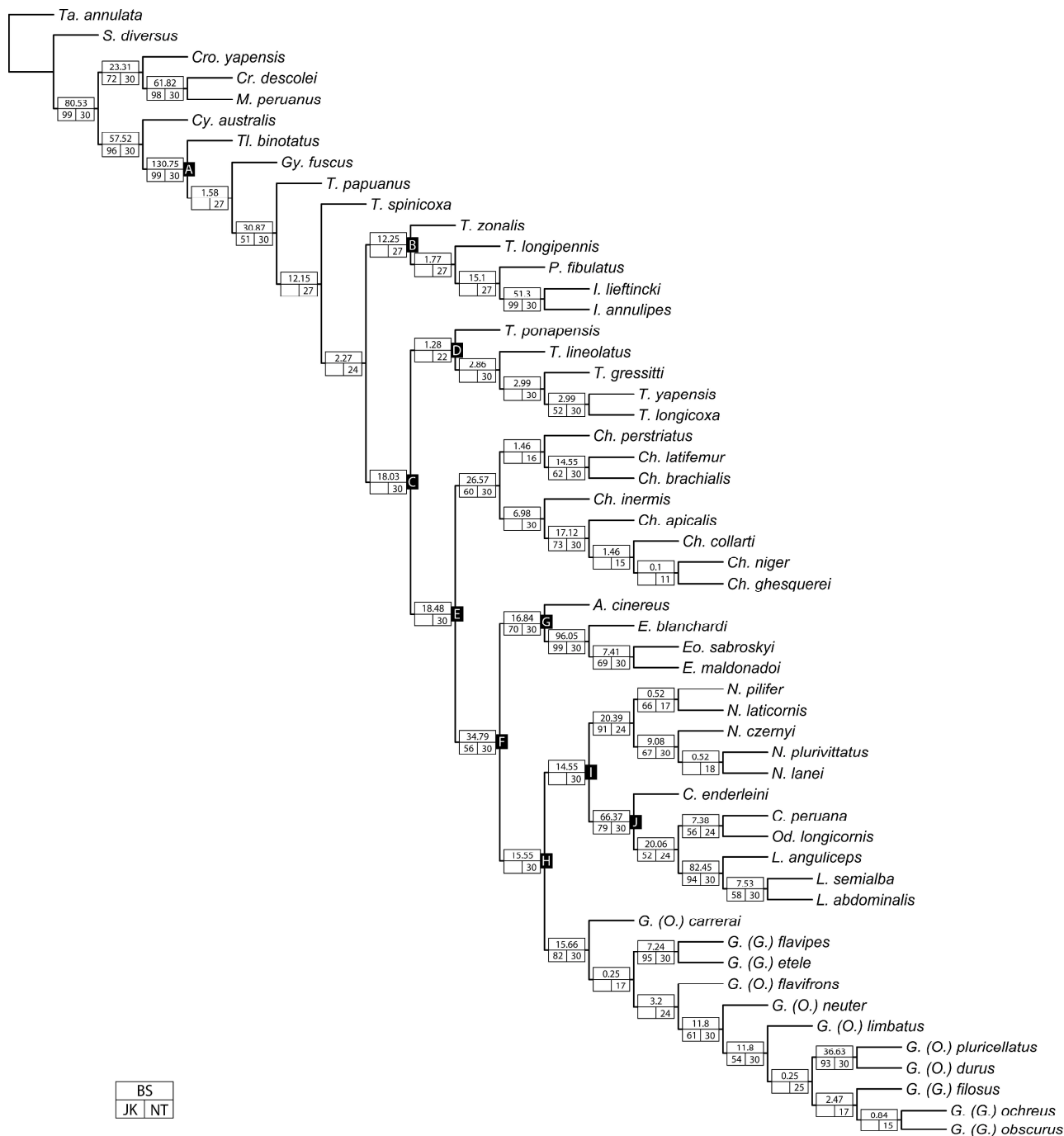


Fig. 8: Preferred topology. The tree was found after rescaling continuous characters, collapsing male-female pairs in the absence of residues > 30% of the expected value, and using implied weights with k from 6 to 10. Fit = 60.72176 (for $k = 6$), length = 847.175, CI = 0.283, RI = 0.623. Numbers above branches represent Bremer support (BS) in units of fit $\times 100$, and numbers below branches are, to the left, jackknifing absolute frequencies (JK, only shown are values > 50), and, to the right, number of trees containing the group (NT, maximum value = 30). Capital letters denote supra-generic clades that are discussed in the text (see Appendix 1 for a list of synapomorphies).

Table 1. List of species included in the analysis, with their taxonomic position and authority (according to Steyskal, 1968; 1980; 1987; Pitkin, 1989; Barraclough, 1993b; Buck, 2010), and geographical distributions (limits following Cox, 2001).

Family	Subfamily	Genus	Species	Author	Distribution		
Micropezidae	Taeniapterinae	<i>Taeniaptera</i>	<i>Ta. annulata</i>	Fabricius, 1787	Neotropical		
		<i>Scipopus</i>	<i>S. diversus</i>	Hendel, 1936	Neotropical		
	Micropezinae	<i>Micropeza</i>	<i>M. (Micropeza) peruanus</i>	Hennig, 1936	Neotropical		
		<i>Cryogonus</i>	<i>Cr. (Cressonius) descolei</i>	Aczél, 1949	Neotropical		
Cypselosomatidae	Eurybatinae	<i>Crosa</i>	<i>Cro. yapensis</i>	Steyskal, 1952	Australian		
		<i>Cypselosoma</i>	<i>Cy. australis</i>	McAlpine, 1966	Australian		
Neriidae	Telostyliinae	<i>Telostylus</i>	<i>Tl. binotatus</i>	Bigot, 1859	Australian/Oriental		
		<i>Chaetonerius</i>	<i>Ch. apicalis</i>	Walker, 1849	Afrotropical		
			<i>Ch. brachialis</i>	Enderlein, 1922	Afrotropical		
			<i>Ch. collarti</i>	Aczél, 1954	Afrotropical		
			<i>Ch. ghesquièrei</i>	Aczél, 1954	Afrotropical		
			<i>Ch. inermis</i>	Schiner, 1868	Oriental		
			<i>Ch. latifemur</i>	Enderlein, 1922	Afrotropical		
			<i>Ch. niger</i>	Czerny, 1932	Afrotropical		
			<i>Ch. perstriatus</i>	Speiser, 1910	Afrotropical		
			Neriinae	<i>Nerius</i>	<i>N. czernyi</i>	Aczél, 1961	Neotropical
					<i>N. lanei</i>	Aczél, 1961	Neotropical
					<i>N. laticornis</i>	Hennig, 1937	Neotropical
					<i>N. plurivittatus</i>	Bigot, 1886	Neotropical
		<i>N. pilifer</i>			Fabricius, 1805	Neotropical	
		<i>Glyphidops</i>			<i>G. (Glyphidops) etele</i>	Aczél, 1961	Neotropical
					<i>G. (G.) filus</i>	Fabricius, 1805	Neotropical
					<i>G. (G.) flavipes</i>	Wiedemann, 1830	Neotropical
					<i>G. (G.) obscurus</i>	Hennig, 1937	Neotropical
					<i>G. (G.) ochreus</i>	Hennig, 1937	Neotropical
			<i>G. (Oncopsia) carrerai</i>	Aczél, 1961	Neotropical		
			<i>G. (O.) durus</i>	Cresson, 1926	Neotropical		
			<i>G. (O.) flavifrons</i>	Bigot, 1886	Neotropical/Nearctical		
		<i>G. (O.)</i>	<i>G. (O.) limbatus</i>	Enderlein, 1922	Neotropical		
			<i>G. (O.) neuter</i>	Hennig, 1937	Neotropical		
			<i>G. (O.) pluricellatus</i>	Schiner, 1868	Neotropical		
			<i>Longina</i>	<i>L. abdominalis</i>	Wiedemann, 1830	Neotropical	
				<i>L. anguliceps</i>	Buck & Marshall, 2004	Neotropical	
				<i>L. semialba</i>	Buck & Marshall, 2004	Neotropical	
	<i>Odontoloxozus</i>			<i>Od. longicornis</i>	Coquillett, 1904	Neotropical/Nearctical	
	<i>Cerantichir</i>		<i>Ce. enderleini</i>	Hennig, 1937	Neotropical		
			<i>Ce. peruana</i>	Hennig, 1937	Neotropical		
	<i>Gymnonerius</i>		<i>Gy. fuscus</i>	Wiedemann, 1984	Oriental		
<i>Paranerius</i>	<i>P. fibulatus</i>		Enderlein, 1922	Australian			
<i>Eoneria</i>	<i>E. blanchardi</i>		Aczél, 1951	Neotropical			
	<i>E. maldonadoi</i>	Aczél, 1961	Neotropical				
<i>Eoloxozus</i>	<i>Eo. sabroskyi</i>	Aczél, 1961	Neotropical				
<i>Antillonerius</i>	<i>A. cinereus</i>	Röder, 1885	Neotropical				
<i>Indonesicesa</i>	<i>I. annulipes</i>	Doleschall, 1858	Australian				
	<i>I. lieftincki</i>	Aczél, 1954	Australian				
	<i>Telostylinus</i>	<i>T. gressitti</i>	Aczél, 1959	Australian			
<i>Telostylinus</i>	<i>T. lineolatus</i>	Wiedemann, 1930	Australian/Oriental				
	<i>T. longicoxa</i>	Thomson, 1869	Australian				
	<i>T. longipennis</i>	Aczél, 1954	Australian				
	<i>T. papuanus</i>	Meijere, 1915	Australian				
	<i>T. ponapensis</i>	Aczél, 1959	Australian				
	<i>T. spinicoxa</i>	Aczél, 1954	Australian				
	<i>T. yapensis</i>	Aczél, 1959	Australian				
	<i>T. zonalis</i>	Aczél, 1954	Australian				

Table 2: List of sinapomorphies.

Group	Synapomorphies
Neriidae (clade A)	Head height/length (7/8): 0.830-0.952 → 0.667
	Male eye length/height (12): 0.930-0.936 → 0.971
	Male postcranium length/head length (16): 0.163-0.173 → 0.202-0.225
	Male length of scape/head length (18): 0.036-0.060 → 0.092
	Length of pedicel/head length (20/21): 0.050-0.051 → 0.267
	Male third costal section/wing length (43): 0.161 → 0.098-0.113
	Ultimate section of M_{1+2} /third section of costal vein (47/48): 2.928 → 3.897-3.917
	Superior region of mesofacial plate (84): short → prolonged
	Postvertical bristles (93): divergent → convergent
	Antenna (97): porrect and short → porrect and elongated
	Shape of scape (98): linear → subglobose to obconical
	Inner process of pedicel (99): absent → present
	Position of antennal arista (104): dorsal → apical
Basal flagellomeres of arista (105): slender → thickened	
Pubescence of antennal arista (106): absent → long	
Differentially pigmented region of antennal arista (109): absent → encompassing basally enlarged region	
Anterior pair of notopleural bristle (121): equal/subequal to posterior pair → absent, or hair-like	
Color of procoxa (171): brown → yellow	
Clade B	Female body length (2): 6.800-8.200 → 8.500-8.700
	Anterior/posterior region of frons (11): 0.974-1.000 → 0.706-0.833
	Wing length/body length (35/35): 0.755-0.772 → 0.796-0.897
	Female second costal section/wing length (42): 0.570-0.579 → 0.605
	Female third costal section/wing length (44): 0.132-0.136 → 0.108-0.118
	Upper margin of dark lateral vitta of occiput (78): running straight → moving downwards
	Occipital bristles (96): 2-3 → absent
Proepisternal bristles (120): one strong and spine-like pair → one short and inconspicuous pair	
Preabdominal marginal longitudinal vittae (189): absent → present	
Clade C	Female body length (2): 6.800-8.200 → 6.400
	Genae height/head height (14/15): 0.108-0.115 → 0.132-0.147
	Female length of pedicel/head length (21): 0.277-0.286 → 0.246-0.247
	Length of postpedicel/head length (24/25): 0.312-0.316 → 0.300
	Male width/length of postpedicel (26): 0.522-0.544 → 0.571-0.581
	Wing width/length (37/38): 0.263-0.264 → 0.277-0.282
Male first costal section/wing length (39): 0.027-0.028 → 0.030-0.035	
Male third costal section/wing length (43): 0.111-0.113 → 0.117-0.123	

	Male fourth costal section/wing width (45): 0.069-0.072 → 0.104-0.114
	Ultimate section of M_{1+2} /third section of costal vein (47/48): 3.897-3.917 → 3.647-3.750
	Female mid femur length/thorax length (55): 1.633 → 1.624-1.629
	Hind femur length/thorax length (56/57): 1.833-1.871 → 1.760-1.786
	Length of epandrium/body length (69): 0.092-0.096 → 0.107-0.111
	Clear vitta in occiput (76): absent → complete
Clade D	Male body length (1): 7.500-7.900 → 7.200-7.400
	Female frons width/head width (10): 0.404-0.412 → 0.350-0.364
	Male length of scape/head length (18): 0.115-0.116 → 0.135-0.147
	Hind tibia length/hind femur length (62/63): 0.829-0.831 → 0.880-0.882
	Color of meso and metacoxae (172): brown → yellow
Clade E	Male frons width/head width (9): 0.387-0.458 → 0.473-0.475
	Genae height/head height (14): 0.132-0.147 → 0.161-0.163
	Thorax length/body length (28/29): 0.311-0.315 → 0.322-0.325
	Thorax height/length (33/34): 0.838-0.839 → 0.841-0.852
	Female first costal section/wing length (40): 0.030-0.031 → 0.035-0.045
	Female second costal section/wing length (42): 0.570-0.576 → 0.547-0.554
	Male third costal section/wing length (43): 0.117-0.123 → 0.135
	Ultimate section of M_{1+2} /third section of costal vein (47/48): 3.647-3.750 → 3.110-3.300
	Length of A_1+CuA_2 /length of CuA_2 (51): 3.455 → 3.369-3.439
	Fore femur length/thorax length (52/53): 1.491-1.544 → 1.226-1.239
	Female mid femur length/thorax length (55): 1.624-1.629 → 1.451-1.550
	Female preabdomen width/length (67): 0.450-0.464 → 0.477-0.481
	Width/length of epandrium (70): 0.320-0.325 → 0.263-0.271
	Shape of inner process of pedicel (100): finger-like → triangular and thin
	Anterior pair of notopleural bristles (121): absent → hair-like, or equal/subequal to posterior pair
	Antero and posteroventral row of spines on fore femur (162): absent, or reduced and hair-like → present
	Apical dark stripe in hind femur (184): absent → present
Clade F	Male postcranium length/head length (16): 0.167-0.169 → 0.176-0.177
	Thorax length before suture/behind suture (30): 0.568-0.571 → 0.670
	Thorax width/length (31/32): 0.583-0.593 → 0.559-0.560
	Female first costal section/wing length (40): 0.035-0.045 → 0.048-0.049
	Female second costal section/wing length (42): 0.547-0.554 → 0.516-0.517
	Length of A_1+CuA_2 /length of CuA_2 (51): 3.369-3.439 → 3.186-3.355
	Fore femur length/thorax length (52/53): 1.226-1.239 → 0.882-1.079
	Male mid femur length/thorax length (54): 1.651-1.786 → 1.159-1.636
	Female mid femur length/thorax length (55): 1.451-1.550 → 1.029-1.446
	Hind femur length/thorax length (56/57): 1.600-1.786 → 1.233-1.365
	Female preabdomen width/length (67): 0.477-0.481 → 0.485-0.500

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3		Length of syntergite 7+8/body length (68): 0.080-0.081 → 0.061-0.063
4		Color of frontal vitta (74): apically yellow → completely yellow
5		Position of ocellar plate (80): before posterior eye margin → on posterior eye margin
6		Shape of posterior margin of head (83): curved → straight
7		Relative lengths of <i>vti</i> and <i>vte</i> bristles (92): <i>vti</i> shorter → equal
8		Position of antennal arista (104): apical → subapical
9		Pubescence of antennal arista (106): long → microscopically short
10		Proepisternal bristles (120): one strong and spine-like pair → one short and inconspicuous pair
11		Relative lengths of <i>sa</i> and <i>pa</i> bristles (131): approximately equal → <i>pa</i> larger
12		Color of pruinosity of mesonotum (137): yellow → gray
13		Longitudinal dustless medial line in mesonotum (138): absent → present
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20		Head length/body length (3/4): 0.208-0.215 → 0.222-0.252
21		Female eye length/height (13): 1.212-1.220 → 1.225-1.229
22		Length of postpedicel/head length (24/25): 0.292-0.300 → 0.266-0.279
23	Clade G	Number of <i>orsa</i> bristles (88): 2 → 3
24		Katepisternal bristle (128): absent → present
25		Lateral pruinosity in scutellum (144): absent → present
26		
27		Female body length (2): 6.400 → 8.000-8.200
28		Thorax length before suture/behind suture (30): 0.670 → 0.914-0.950
29		Male second costal section/wing length (41): 0.554-0.563 → 0.485-0.489
30		Length of $A_1 + CuA_2$ /length of CuA_2 (51): 3.186-3.355 → 2.262-2.315
31		Fore tibia length/fore femur length (58/59): 0.985-1.016 → 0.983
32	Clade H	Texture of dorsal surface of antennal bases (86): not polished → polished and shiny
33		Type of bristles (87): long and bristle-like → short and spiniform
34		Occipital bristles (96): 1, or 2-3 → absent
35		Number of bristles in basicosta (158): 2 → 1
36		Number of dorsal bristles in male procoxae (166): 3 → 2
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41		Male body length (1): 7.900 → 9.800-10.000
42		Female body length (2): 8.000-8.200 → 8.500-9.500
43		Head width/length (5/6): 0.697-0.771 → 0.667-0.691
44		Head height/length (7/8): 0.591-0.634 → 0.556-0.569
45		Male postcranium length/head length (16): 0.176-0.203 → 0.241-0.254
46		Female postcranium length/head length (17): 0.170-0.175 → 0.229
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49	Clade I	Female length of scape/head length (19): 0.119-0.130 → 0.133
50		Male length of process of pedicel/length of pedicel (22): 0.400-0.434 → 0.250-0.263
51		Female length of process of pedicel/length of pedicel (23): 0.417-0.469 → 0.286-0.321
52		Thorax length/body length (28/29): 0.322-0.325 → 0.331-0.337
53		Thorax width/length (31/32): 0.559-0.560 → 0.533-0.556
54		Male third costal section/wing length (43): 0.135 → 0.113-0.127
55		Female third costal section/wing length (44): 0.135-0.136 → 0.133
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3	Prebasal section of M_{1+2} /median section of M_{1+2} (48/49): 0.921-0.950 → 0.879
4	Fore tibia length/fore femur length (58/59): 0.983 → 0.911-0.952
5	Ovipositor width/length (72): 0.399-0.508 → 0.363-0.390
6	Shape of inner process of pedicel (100): triangular and thin → triangular and broad
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9	Female body length (2): 8.500-9.500 → 11.500
10	Head height/length (7/8): 0.556-0.569 → 0.533-0.546
11	Female frons width/head width (10): 0.410-0.412 → 0.459
12	Genae height/head height (14/15): 0.161-0.176 → 0.177-0.198
13	Male postcranium length/head length (16): 0.241-0.254 → 0.266-0.288
14	Female postcranium length/head length (17): 0.229 → 0.235-0.240
15	Male length of pedicel/head length (20): 0.250-0.269 → 0.271-0.389
16	Female length of process of pedicel/length of pedicel (23): 0.286-0.321 → 0.274
17	Male width/length of postpedicel (26): 0.571-0.581 → 0.508
18	Thorax width/length (31/32): 0.533-0.556 → 0.481-0.488
19	Thorax height/length (33/34): 0.841-0.852 → 0.727-0.738
20	Preabdomen length/body length (64/65): 0.375-0.383 → 0.357-0.371
21	Female preabdomen width/length (67): 0.485-0.500 → 0.365
22	Clade J Width/length of epandrium (70): 0.263-0.271 → 0.277-0.283
23	Clear vitta in occiput (71): complete → incomplete
24	Position of clear vitta (77): medial → inferior
25	Shape of posterior margin of head (83): straight → curved
26	<i>vt</i> bristle (91): present → absent
27	Shape of first flagellomere (101): ovate → subrectangular
28	Position of antennal arista (104): subapical → dorsoapical
29	General pattern of coloration of antennal arista (108): brown → white
30	Configuration of anterior region of thorax (116): scutum and anteprenotal ridge ending at level of postpronotal carina → ending beyond level of postpronotal carina
31	Shape of katepisternum (117): higher than wide → as wide as high
32	Postpronotal bristles (119): absent → hair-like
33	Relative lengths of <i>sa</i> and <i>pa</i> bristles (131): <i>pa</i> larger → approximately equal
34	Shape of dorsal face of mid femur (164): straight/slightly convex → concave
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36	Head width/length (5/6): 0.762-0.771 → 0.874-1.000
37	Head height/length (7/8): 0.632-0.676 → 0.693-0.751
38	Anterior/posterior region of frons (11): 0.974-1.000 → 0.933
39	Female postcranium length/head length (16): 0.166-0.170 → 0.134-0.144
40	Male length of process of pedicel/length of pedicel (22): 0.400-0.434 → 0.500-0.622
41	Female length of process of pedicel/length of pedicel (23): 0.439-0.500 → 0.588
42	Thorax length before suture/behind suture (30): 0.568-0.571 → 0.511-0.534
43	Wing length/body length (35/36): 0.755-0.772 → 0.834-0.862
44	Male third costal section/wing length (43): 0.135 → 0.144
45	Preabdomen length/body length (64/65): 0.383 --> 0.400-0.409
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3	Antennal bases (85): present → absent
4	Number of extra dorsocentral bristles posterior to transverse suture (126): 0 → 1
5	Non-apical scutellar bristles (132): absent/vestigial → long
6	Number of lateral bristles on metacoxae (169): 1 → 2
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9	Head length/body length (3/4): 0.208-0.215 → 0.195-0.199
10	Head width/length (5/6): 0.697-0.771 → 0.827-0.843
11	Male frons width/head width (9): 0.473-0.475 → 0.433-0.438
12	Male length of scape/head length (18): 0.115-0.116 → 0.112
13	Male length of process of pedicel/length of pedicel (22): 0.400-0.434 → 0.459
14	Female width/length of postpedicel (27): 0.594-0.605 → 0.561
15	Thorax length/body length (28/29): 0.322-0.325 → 0.316-0.321
16	Female second costal section/wing length (42): 0.516-0.517 → 0.467-0.485
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19	<i>Glyphidops</i> Male third costal section/wing length (43): 0.135 → 0.142-0.149
20	Prebasal section of M_{1+2} /median section of M_{1+2} (49/50): 0.921-0.950 → 1.014-1.064
21	Preabdomen length/body length (64/65): 0.383 → 0.387-0.400
22	Ovipositor length/body length (71): 0.217-0.218 → 0.196
23	Ovipositor width/length (72): 0.399-0.508 → 0.548
24	Upper margin of dark lateral vitta of occiput (78): running straight → fused with ocellar plate
25	Proepisternal bristles (120): one short and inconspicuous pair → absent
26	Number of bristles in basicosta (158): 1 → 0
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31	Female body length (2): 10.300 → 10.600-10.700
32	Head width/length (5/6): 0.762-0.771 → 0.857-0.907
33	Head height/length (7/8): 0.591-0.640 → 0.683-0.716
34	Anterior/posterior region of frons (11): 0.706-0.833 → 0.678
35	Male eye length/height (12): 1.186-1.188 → 1.119
36	Female eye length/height (13): 1.165 → 1.138-1.155
37	Genae height/head height (14/15): 0.074-0.115 → 0.138-0.158
38	Male postcranium length/head length (16): 0.167-0.169 → 0.157-0.166
39	Female postcranium length/head length (17): 0.164 → 0.142-0.148
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42	<i>Indonesicesa</i> Thorax length before suture/behind suture (30): 0.568-0.571 → 0.673
43	Female second costal section/wing length (42): 0.605 → 0.547-0.564
44	Male fourth costal section/wing width (45): 0.061-0.072 → 0.057-0.058
45	Width/length of epandrium (70): 0.320-0.328 → 0.337-0.347
46	Shape of posterior margin of head (83): curved → straight
47	Relative lengths of <i>v_{ti}</i> and <i>v_{te}</i> bristles (92): <i>v_{ti}</i> shorter → equal
48	Occipital bristles (96): absent → 2-3
49	Apex of first flagellomere (103): rounded → pointed
50	Spinules on fore tibia (163): absent → present
51	Club-shaped thickening in apex of male fore tibia (165): absent → present
52	Color of meso and metacoxae (172): brown → yellow
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3	Apical dark stripe in tibiae (185): absent → present
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5	Female body length (2): 11.600-13.100 → 13.400
6	Male length of scape/head length (18): 0.116-0.144 → 0.272
7	Female length of scape/head length (19): 0.133 → 0.236-0.284
8	Color of frontal vitta (74): apically yellow, or completely yellow → brown
9	Clear vitta in occiput (76): incomplete → absent
10	Vibrissae (94): absent → present
11	
12	Shape of scape (98): subglobose to obconical → semicylindrical
13	<i>Longina</i>
14	Subcostal break (150): present → absent
15	Bare areas of wing (157): absent, or small → large
16	Ciliae on upper squamae (159): long → reduced
17	Antero and posteroventral row of spines on fore femur (162): present → reduced and
18	hair-like
19	Number of dorsal bristles in female procoxae (167): 3, or 4 → 5
20	Apical dark stripe in tibiae (185): present → absent
21	
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23	Genae height/head height (14/15): 0.161-0.176 → 0.160
24	Male width/length of postpedicel (26): 0.571-0.581 → 0.588-0.667
25	Wing width/wing length (37/38): 0.274-0.288 → 0.259-0.265
26	Female fourth costal section/wing width (46): 0.094-0.100 → 0.086-0.092
27	Ultimate section of M_{1+2} /third section of costal vein (47/48): 2.878-3.300 → 3.449-3.465
28	Length of syntergite 7+8/body length (68): 0.061-0.063 → 0.070-0.074
29	Length of epandrium/body length (69): 0.115 → 0.148-0.153
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32	<i>Nerius</i>
33	Width/length of epandrium (70): 0.263-0.271 → 0.179-0.194
34	Ovipositor length/body length (71): 0.217-0.224 → 0.243-0.256
35	Position of ocellar plate (80): on posterior eye margin → behind posterior eye margin
36	Katepisternal bristle (128): absent → present
37	Shape of crossvein <i>dm-cu</i> (153): straight to slightly convex → strongly convex
38	Number of central rings in anterior femur (175): 1 → 0
39	Number of central rings in mid femur (179): 1 → 0
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41	Apical dark stripe in tibiae (186): present → absent
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Appendix S1: Character description

Continuous characters

The subsequent list shows all characters initially registered and included in the analysis of continuous characters. Characters marked with an asterisk are those computed using male and female data indistinctively. Contiguous male-female couples of characters that are shaded gray were collapsed into a single character given that they presented a lack of outliers for the criteria that led to the preferred phylogenetic hypothesis, *i.e.* a difference between at least one observed and expected value > 30% of the latter in the regression analysis. Non-shaded characters were retained as such, resulting in a quantitative dataset of 54 continuous characters that was used for the cladistic analysis of Neriidae. Some comments on the way in which certain measurements were taken can be found between brackets next to the respective characters.

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| 1. <i>Male body length.</i> | (Excluding antennae and |
| 2. <i>Female body length.</i> | postabdomen) |
| 3. <i>Male head length/body length.</i> | |
| 4. <i>Female head length/body length.</i> | |
| 5. <i>Male head width/head length.</i> | |
| 6. <i>Female head width/head length.</i> | |
| 7. <i>Male head height/head length.</i> | |
| 8. <i>Female head height/head length.</i> | |
| 9. <i>Male frons width/head width.</i> | (Measured at vertex) |
| 10. <i>Female frons width/head width.</i> | |
| 11. <i>Anterior/posterior region of frons. *</i> | |
| 12. <i>Male eye length/eye height.</i> | |
| 13. <i>Female eye length/eye height.</i> | |
| 14. <i>Male genae height/head height.</i> | (The height of the genae was |
| 15. <i>Female genae height/head height.</i> | measured below the eyes) |
| 16. <i>Male postcranium length/head length.</i> | |
| 17. <i>Female postcranium length/head length.</i> | |
| 18. <i>Male length of scape/head length.</i> | |
| 19. <i>Female length of scape/head length.</i> | |
| 20. <i>Male length of pedicel/head length.</i> | |
| 21. <i>Female length of pedicel/head length.</i> | |
| 22. <i>Male length of process of pedicel/length of pedicel.</i> | |
| 23. <i>Female length of process of pedicel/length of pedicel.</i> | |

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- 3 24. Male length of first flagellomere/head length.
- 4 25. Female length of first flagellomere/head length.
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- 6 26. Male width of first flagellomere/length of first flagellomere.
- 7 27. Female width of first flagellomere/length of first flagellomere.
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- 9 28. Male thorax length/body length. (Including scutellum)
- 10 29. Female thorax length/body length.
- 11 30. Thorax length before transverse suture/thorax length behind suture. *
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- 13 31. Male thorax width/thorax length.
- 14 32. Female thorax width/thorax length.
- 15 33. Male thorax height/thorax length.
- 16 34. Female thorax height/thorax length.
- 17 35. Male wing length/body length.
- 18 36. Female wing length/body length.
- 19 37. Male wing width/wing length.
- 20 38. Female wing width/wing length.
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- 22 39. Male first costal section/wing length.
- 23 40. Female first costal section/wing length.
- 24 41. Male second costal section/wing length.
- 25 42. Female second costal section/wing length.
- 26 43. Male third costal section/wing length.
- 27 44. Female third costal section/wing length.
- 28 45. Male fourth costal section/wing width.
- 29 46. Female fourth costal section/wing width.
- 30 47. Male ultimate section of M_{1+2} /third section of costal vein.
- 31 48. Female ultimate section of M_{1+2} /third section of costal vein.
- 32 49. Male prebasal section of M_{1+2} /median section of M_{1+2} .
- 33 50. Female prebasal section of M_{1+2} /median section of M_{1+2} .
- 34 51. Length of A_1+CuA_2 /length of CuA_2 . *
- 35 52. Male fore femur length/thorax length.
- 36 53. Female fore femur length/thorax length.
- 37 54. Male mid femur length/thorax length.
- 38 55. Female mid femur length/thorax length.
- 39 56. Male hind femur length/thorax length.
- 40 57. Female hind femur length/thorax length.
- 41 58. Male fore tibia length/fore femur length.
- 42 59. Female fore tibia length/fore femur length.
- 43 60. Male mid tibia length/mid femur length.
- 44 61. Female mid tibia length/mid femur length.
- 45 62. Male hind tibia length/hind femur length.
- 46 63. Female hind tibia length/hind femur length.
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64. Male preabdomen length/body length.	
65. Female preabdomen length/body length.	
66. Male preabdomen width/preabdomen length.	(The width of the preabdomen was measured at the posterior margin of the second tergite)
67. Female preabdomen width/preabdomen length.	
68. Length of syntergite 7+8/body length.	
69. Length of epandrium/body length.	
70. Width of epandrium/length of epandrium.	(The width of the epandrium was measured at the apex)
71. Ovipositor length/body length.	
72. Ovipositor width/ovipositor length.	(The width of the ovipositor was taken at the base of the structure)

Discrete characters

All multistate characters were treated as unordered unless otherwise stated. Autapomorphic characters were excluded.

73. *Shape of frons between the eyes: convex (0), concave (1)*. The frons is more or less deeply impressed in all Neriidae and some Micropezinae (sometimes adopting a V-shape), while it is convex and outwardly rounded in other micropezids, as in the Taenipterinae and most Eurybatinae (Aczél, 1951).

74. *Colour of frontal vitta: brown (0), apically yellow (1), completely yellow (2)*. The coloration markings on the frons vary considerably within and among genera. These markings sometimes include totally or partially the fronto-orbital plates, but this distinction was not taken into account. Some species present a characteristic yellowish wedge-shaped spot restricted to the anterior region of the frons (state 1), typical of the *Chaetonerius* but also present in some species of *Telostylinus* (Aczél 1954a, 1954b, 1955b); others, show a yellow marking that extends back at least until the ocellar tubercle (state 2). The character was treated as ordered, since the presence of a completely yellow frontal vitta includes having an apical spot.

75. *Postcranium pigmentation pattern: homogenous (0), dark occiput and clear postgenae (1)*. All neriids present a head laterally divided into a darker upper third, encompassing the occiput behind the eyes, and a whitish postgenal area. Micropezids included in the analysis have laterally homogenous postcrania, while *Cypselosoma australis* has a clearer genal region.

76. *Clear vitta in occiput: absent (0), incomplete (1), complete (2)*. Within the dark region of the occiput (char. 75), a clear yellowish vitta is generally present, more intensely marked

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3 in Neotropical species. This vitta is nonetheless absent (state 0) in some species (eg.: some
4 *Chaetonerius* and *Telostylinus*) most of them from the Australian-Oriental region, while it
5 is incomplete and faint (state 1) in the genera *Odontoloxozus* and *Cerantichir*, not attaining
6 the posterior margin of the head. The character was treated as ordered.
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10 77. *Position of clear vitta in occiput: medial (0), submedial (1), inferior (2)*. Most species
11 with a clear vitta within the dark region of the occiput present it in a medial position (state
12 0). However, within some species of the genus *Nerius*, this vitta is placed submedially
13 (state 1, Aczél, 1961), and in the genera *Cerantichir* and *Odontoloxozus* it is “(...) placed
14 near lower margin of this blackish brown area” (Aczél, 1961: 298). The character was
15 treated as ordered.
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19 78. *Upper margin of dark lateral vitta of occiput: moving downwards (0), running straight*
20 *(1) fused with ocellar plate (2)*. Most species present a yellowish frontal vitta (char. 74),
21 and even among those that do not, a clear stripe develops in the fronto-orbital plates. In
22 both cases, the dark region of the occiput presents a well defined upper margin. This
23 margin can move downwards as it progresses towards the back of the head (state 0), as in
24 the genus *Rhoptrum* (Aczél, 1954a), run straight (state 1), or advance towards the dorsal
25 region of the head, fusing with the ocellar plate, as in some *Glyphidops* (Aczél, 1961). In
26 case this upper margin is absent, the state was codified as inapplicable.
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31 79. *Auxiliary ocellar lamina: absent (0), present (1)*. Among the Micropezinae and
32 Eurybatinae, the ocellar lamina continues backwards up until the vertex, forming an
33 elevation that is similarly strongly quitinized and pigmented as the ocellar tubercle. This
34 structure is absent from the rest of the Neriioidea.
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38 80. *Position of ocellar plate: close to anterior eye margin (0), before posterior eye margin*
39 *(1), on posterior eye margin (2), behind posterior eye margin (3)*. The placement of the
40 ocellar lamina on the frons is clearly anterior among Taenipterinae, while it adopts a more
41 posterior position in Micropezinae, Eurybatinae and Neriidae (Aczél, 1951). However, even
42 among these, the exact position with respect to the posterior eye margin can vary
43 considerably depending on the elongation of the head and the postcranium. This character
44 was treated as ordered.
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48 81. *Ocellar furrow: absent (0), present (1)*. Exclusively in the genus *Longina* (*L.*
49 *abdominalis* and *L. semialba*), the ocellar tubercle is surrounded both laterally and
50 posteriorly by a deep furrow (Aczél, 1961; Buck and Marshall, 2004).
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53 82. *Shape of posterior region of occiput: rounded (0), sharply truncated (1)*. Aczél (1951:
54 490) discussed the shape of the head among this group of flies, stating that only in the
55 Neriidae was the posterior region of the occiput always sharply truncated, given that it does
56 not participate in the elongation of the head. However, some genera (*Gymnonerius*,
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3 *Teloneria*, *Paranerius* and *Cerantichir*) have species with an elongated occiput forming a
4 rounded vertex.
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7 83. *Shape of posterior margin of head: curved (0), straight (1)*. The posterior margin of the
8 head is straight in most Neotropical neriids, continuing the sharp angle of the occiput (char.
9 82). On the other hand, this margin is curved in most Australian-Oriental neriids and the
10 *Cerantichir-Odontoloxozus-Longina* clade, with the maximum length of the head occurring
11 at its center.
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14 84. *Superior region of mesofacial plate: short (0), prolonged (1)*. A character shared by all
15 Neriidae and some Micropezinae is the prolonged mesofacial plate that, although not
16 forming antennal bases in the latter nor in the Telostylinae (char. 85), gives nonetheless the
17 head of both groups a conical and longitudinally elongated morphology (Aczél, 1951).
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21 85. *Antennal bases: absent (0), present (1)*. The anterior region of the mesofacial plate
22 (upper face, or lunule) has become inflated, protruding and medially divided in a group of
23 Neriidae, adopting the appearance of an extra segment of the antenna. This peculiar
24 configuration of the head, usually referred to as antennal bases (Aczél 1951, 1961), divides
25 the family into its two constituting subfamilies (Enderlein, 1922; Aczél, 1954a).
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29 86. *Texture of dorsal surface of antennal bases: not polished (0), polished and shiny (1)*.
30 This character was proposed by Hennig (1937) to constitute the tribal limits of the family,
31 but in Aczél's systematic revision of the family (1954) was downgraded, and subsequently
32 used as the base for dividing the Neriinae into the *Eoneria* and the *Nerius* groups (1961).
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36 87. *Type of bristles: long and bristle-like (0), short and spiniform (1)*. Aczél (1951)
37 considered the presence of short and strong bristles to be characteristic of the Neriidae, with
38 only a few exceptions. However, after analyzing the African and Oceanic faunas, the
39 presence of "long, bristle-like bristles" in many genera of the Old World was interpreted as
40 plesiomorphic, and partly contributed to the conception of the Telostylinae and the
41 *Eoneria*-group as basal clades (Aczél, 1954a; 1961).
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45 88. *Number of superior anterior orbital bristles: range 0 - 4*. The fronto-orbital bristles in
46 all Neriioidea are located in a simple reclinate series (D. K. McAlpine, 2000). The anterior
47 pairs (if present) always stand on a barely quitinized region of the fronto-orbital plate
48 (Aczél, 1951), while a single pair is generally present on the posterior, strongly quitinized
49 region. Although many authors no longer discriminate between the anterior pairs (superior
50 anterior orbital bristles or *orsa* according to Hennig, 1937), and the posterior one (superior
51 superior orbital bristle or *orss*), this distinction was maintained here since it allowed to
52 separate the fronto-orbital bristles into positionally homologous elements. The character
53 was treated as ordered.
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3 89. *Superior superior orbital bristles: absent (0), present (1)*. See char. 88. This pair of
4 bristles is only absent in the Micropezinae.
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7 90. *Vertical interior bristles (vti): absent (0), present (1)*.
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9 91. *Vertical external bristle (vte): absent (0), present (1)*.
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11 92. *Relative length of vti and vte bristles: vti larger (0), equal (1), vti shorter (2)*. The
12 relative lengths of both cephalic and thoracic bristles were often stated by Aczél in his
13 taxonomic descriptions. In particular, *vti* bristles are more commonly elongated (state 0) in
14 micropezids, while among Neriidae they are generally equal (state 1) to *vte* in the
15 Neotropical genera and considerably reduced (and therefore shorter than *vte*, state 2) in
16 most Old World genera. This character was treated as continuous.
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20 93. *Postocellar bristles: divergent (0), convergent (1)*. Postocellar bristles (referred to as
21 postvertical bristles or *pvt* by Aczél) are invariantly present in all included taxa.
22 Nonetheless, in Neriidae these are generally strongly convergent (state 1) and even crossed
23 if long enough, considered by many as an apomorphic state (Aczél, 1951; Hennig, 1958,
24 McAlpine, 1974). Among Micropezidae and Cypselosomatidae they are always divergent
25 (state 0).
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29 94. *Vibrissae: absent (0), present (1)*.
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31 95. *Genal bristles: absent (0), present (1)*.
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34 96. *Occipital bristles: absent (0), 1 (1), 2-3 (2), many (3)*. This character was divided into
35 four states, representing the absence of occipital bristles (state 0), the presence of a single
36 bristle, of only a few or of many (states 1, 2 and 3 respectively). This grouping corresponds
37 to the variability observed in revised specimens, with many of them having 2 occipital
38 bristles on one side of the head and three on the other, whereas the distinction with respect
39 to the presence of many occipital bristles is very clear. This character was treated as
40 ordered.
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44 97. *Antennae: pending and relatively short (0), porrect and short (1), porrect and*
45 *elongated (2)*. The elongated and porrect antenna (state 2) of the Neriidae is one of the most
46 well established synapomorphies of the family (Cresson, 1938; Aczél, 1951, 1961;
47 Steyskal, 1968; Barraclough, 1993a; Buck, 2010). In contraposition, the antennae of the
48 other families within Neriioidea is relatively short (Aczél, 1951), with pending pedicel and
49 first flagellomere (state 0), except in the Cypselosomatidae (J. F. McAlpine, 1981b), which
50 have porrect antennae yet retaining its short length (state 1). The character was treated as
51 ordered.
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55 98. *Shape of scape: linear (0), subglobose to obconical (1), semicylindrical (2)*. This
56 character reflects the relationship between the length and the height of the scape. A given
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scape was taken to be linear (state 0) if it was significantly higher than long (length = 1/3 – 2/3 of height), subglobose to obconical (state 1) if both dimensions were approximately equivalent (length = 0.8 - 1.5 the height), and semicylindrical (state 2) when the length was at least twice the height (proposed as a synapomorphy of the *Longina* by Buck and Marshall, 2004).

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99. *Inner process of pedicel: absent (0), present (1)*. The inner face of the pedicel of the neriid antennae presents a projection that attaches to the median region of the first flagellomere (Steyskal, 1987a). This effectively separates the pedicel into a membered shaped at the base (referred to as body) and an apical prolongation (referred to as process; see chars. 22, 23) (Aczél, 1951), a synapomorphic antennal configuration that is not present in any other dipteran family (Aczél, 1954a).

100. *Shape of inner process of pedicel: triangular and broad (0), triangular and thin (1), finger-like (2)*. A lot has been written on the shape, length, width and apex of the inner process of the pedicel of the different genera of neriid flies. However, many authors have used the same terminology to describe slightly different morphologies, as is the case of the term “finger-like” in the work of Enderlein (1922), Aczél (1951, 1954a) and Sepúlveda *et al.* (2013a). Some authors have even changed the way in which they described this structure throughout their work, leading to inconsistent depictions; e.g.: the use of “finger-like” between Aczél 1951 and 1954, the criteria relating to the broadness of the process between Aczél 1954 and 1961, among others. Here, processes were divided into two different general shapes: a triangular morphology, with both sides more or less straightly tapering into an apex, and a finger-like morphology, in which both sides run parallel through some portion of the structure. By this definition, all Neotropical Neriidae present triangular processes, in contrast with what Sepúlveda *et al.* (2013a) described. On the other side, this definition of “finger-like” is more congruent with what Aczél (1954a, 1955) described as processes developing a “longitudinal keel”. Triangular processes were further subdivided into those which are broad (state 0) and those which are thin (state 1), considering broad all processes whose base occupy the entire inner side of the pedicel. Since all finger-like processes are thin, the character was treated as ordered, being therefore equivalent to a couple of characters, one relating to its shape and the other to its broadness.

101. *Shape of first flagellomere: ovate (0), obovate (1), subrectangular (2)*. First flagellomeres can be broadly divided into those presenting an ovoid shape, and those whose dorsal and ventral margins run parallel (Sepúlveda *et al.*, 2013a), creating a subrectangular shape. Ovoid postpedicels were further classified by Aczél (1961) as ovate or obovate depending on whether the maximum width is attained near the base or the apex of the segment, respectively, and we maintained this classification.

102. *Base of first flagellomere: rounded (0), truncate (1)*. The base of the first flagellomere is in most species rounded, given that the apex of the pedicel is clearly concave when seen

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3 in a lateral view (state 0). However, in a few species of Neotropical neriids, the apex of the
4 pedicel ends in a sharp and transverse apex (Aczél, 1961), therefore the first flagellomere
5 emerges from a truncate base (state 1).
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10 *103. Apex of first flagellomere: pointed (0), rounded (1), truncate (2).* This important
11 feature has been widely used in taxonomic descriptions and genera definitions (Aczél,
12 1961). Although related to both position of antennal arista (char. 104) and shape of the first
13 flagellomere (char. 101), the shape of the apex of the postpedicel is not unequivocally
14 determined by these other attributes. The apex of this segment was considered pointed
15 when it presents a width equivalent to the arista, and truncate when it ends in a surface
16 more or less perpendicular to the longitudinal axis. On the other hand, the majority of first
17 flagellomeres were considered to present a rounded apex, without discriminating those that
18 were described as “widely rounded” (Aczél, 1961; Sepúlveda *et al.*, 2013a), given that this
19 attribute depends only on the width of the postpedicel near the apex (a variable already
20 included in chars. 26 and 27), and does not represent a difference in morphology.
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25 *104. Position of antennal arista: apical (0), subapical (1), dorsoapical (2), dorsal (3).* One
26 of the most distinguishing (and profoundly plesiomorphic) feature of the Neriidae is the
27 apical/subapical placement of the antennal arista (Aczél, 1951; Steyskal, 1968; McAlpine,
28 1981). In his monography on American Neriidae, Aczél (1961) further subdivided this trait,
29 discriminating those species for which the antenna is placed at the dorsoapical angle of the
30 first flagellomere (state 2), and others have followed this distinction (Buck and Marshall,
31 2004; Sepúlveda *et al.*, 2013a; 2013b). This character was treated as ordered, given that the
32 states represent discrete points of a positional cline.
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37 *105. Basal flagellomeres of arista: slender (0), thickened (1).* Another of the
38 synapomorphic characters of the neriid antenna is the enlarged configuration of the three
39 (sometimes only two) most basal flagellomeres of the arista (Aczél, 1961).
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43 *106. Pubescence of antennal arista: absent (0), microscopically short (1), long (2).* The
44 degree of development of the pubescence of the antennal arista has been proposed as a key
45 feature in the delineation of groups within the family (Aczél, 1961). Three major types of
46 pilosity occur within the neriids: a completely naked arista, typical of the genera *Nerius* and
47 *Glyphidops* (with some exceptions); an arista covered by microscopically short yet
48 conspicuous pubescence, only visible at 30-40x magnification (e.g.: American branch of
49 the *Eoneria*-group); or covered with long and fine hairs, that although varying in length, are
50 always visible at naked eye or using lower magnifications. Among the analyzed
51 Micropezidae the arista is pilose exclusively in the Eurybatinae (Aczél, 1959), while the
52 extremely short and sparse pubescence of the included Cypselosomatidae (only visible at
53 200x magnification; McAlpine, 1966) was also codified as absent. This character was
54 treated as ordered.
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107. *Pattern of pubescence of antennal arista: basally dense and apically sparse/naked (0), homogenously dense (1).* Among species with a hairy arista, excluding the basal enlarged aristomeres which are always naked, the pubescence can either homogenously cover the structure, or vary in density along it. In this last case, the density of hairs is invariably reduced towards the apex of the arista.

108. *General pattern of coloration of antennal arista: brown (0), white (1), basally white and apically brown (2).* Aczél (1961) originally described two general patterns of coloration of the antennal arista among the Neotropical Neriidae. According to him, a group of genera present a dark brown arista, bare or covered with brown pubescence, while the other present a white arista, always covered in hair. However, he noted that in case the arista gets sparcely covered towards the apex (char. 107), this region can develop a brown coloration. Due to this last pattern being also common among Old World neriids (Aczél, 1954a; 1955a), it was taken as a third state. Although a few taxonomic descriptions acknowledge different colors for the arista and for the pubescence it carries, the modularity of these two characters is doubtful, and in most species both change in a congruent fashion. This character therefore simply describes the general coloration of the antennal arista as a whole.

109. *Differentially pigmented region of antennal arista: absent (0), encompassing basally enlarged region (1), distad to enlarged region (2).* Apart from the general coloration described previously (char. 108), most species of Neriidae present a small region of differentially pigmented region at the base of the antennal arista. This region attains a color that varies between testaceous yellow and dark brown, and stands out as a darker region in whitish aristae or as a bright region in brown ones. In most cases, the differentially pigmented zone is found at the base of the arista, where the enlarged aristomeres are (state 1), although in some members of the genus *Telostylinus* it is placed right after this enlarged region (state 2).

110. *Size of buccal cavity: small (0), large (1).* All groups included in the analysis have oval-shaped buccal cavities, of which that of Neriidae and Taenipterinae is much bigger, with a total surface at least 3 times (and up to 10 times) larger than that of Micropezinae and Eurybatinae (Aczél, 1951).

111. *Shape of prementum: as long as wide (0), slightly longer (1), considerably longer (2).* The prementum of Micropezinae is approximately as long as wide, 1.25 – 1.5 times longer than wide in the Taenipterinae and Eurybatinae, and becoming noticeably thin and elongated in the Neriidae, 2 – 3 longer than wide according to Aczél (1951: 495), although it can even be 4 times longer than wide in some *Chaetonerius* and *Telostylinus*. This character was treated as ordered.

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112. *Shape of clypeus: U-shaped (0), shield-shaped (1)*. Only in the Micropezinae one finds the small and U-shaped clypeus that is typical of the Muscomorpha (McAlpine, 1981), whereas in Neriidae, Taeniapterinae and Eurybatinae the clypeus is relatively large and shield-shaped (Aczél, 1951).

113. *Lateral borders of the clypeus: thickening backwards (0), uniform (1), thickening forwards (2)*. Despite both Neriidae and Taeniapterinae having similarly shaped clypeus (char. 101), they differ in the structure of the lateral borders of the sclerite, which become strongly thicker posteriorly in Neriidae and anteriorly in Taeniapterinae (Aczél, 1951). The Micropezinae and Eurybatinae present an intermediate state, with the lateral borders of the clypeus being uniform. The character was treated as ordered.

114. *Length of maxillary palpi: reaching clypeus' anterior margin (0), not reaching (1)*. Another feature shared between Neriidae and Taeniapterinae is the presence of elongated maxillary palpi (state 0), which almost invariantly reach the anterior margin of the clypeus. Micropezinae and Eurybatinae on the other hand, always present short maxillary palpi (state 0).

115. *Shape of maxillary palpi: compressed (0), subcylindrical (1)*. Contrary to what happens with the length of the maxillary palpi (char. 103), the shape of this structure allies the Neriidae and Micropezinae, both of which have subcylindrical maxillary palpi, while it is always compressed in the Taeniapterinae (Aczél, 1951).

116. *Configuration of anterior region of thorax: scutum and anteprenotal ridge ending at level of postpronotal carina (0), scutum and anteprenotal ridge ending beyond level of postpronotal carina (1)*. Among some species with an elongated thorax, the presutural scutellum and the anteprenotal ridge have an anterior ending beyond the level of the postpronotal (humeral) carina. This was taken to be an apomorphic configuration of the thorax by Buck (2010), and the basis for his transfer of *Odontoloxozus peruanus* to the genus *Cerantichir*. However, as Sepúlveda *et al.* (2013a) noticed, this character is also shared by the genus *Longina*. Among Old World neriids it is also present in the species *Gymnonerius fuscus* (Aczél, 1955a: 35, Fig. 4).

117. *Shape of katepisternum: as wide as high (0), higher than wide (1)*. The katepisternum of all Neotropical taxa with elongated thorax (except the genus *Nerius*) is about as wide as it is high, being higher than wide in the rest (Aczél, 1961), as well as in the entire Old World fauna.

118. *Copulatory processes constituted by fifth sternite: absent (0), present (1)*. In general, most male micropezids present forceps-like processes of variable sizes and shapes, which arise from the sternite 5 (Steyskal, 1987b). These processes, which are involved in the copula, are absent from Neriidae and Cypselosomatidae, and from some genera of

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3 Micropezinae and Taeniapterinae (Aczél, 1951), although present in those included in the
4 analysis.
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7 *119. Postpronotal bristles: absent (0), hair-like (1), strong (2).* This pair of bristles, also
8 referred to as humeral bristles, were cited as absent from the Neriidae and the
9 Mycropezidae by Aczél (1951: 486). However, he afterwards described the presence of this
10 bristles as a synapomorphic condition of the genus *Odontoloxozus* (Aczél, 1961). Since
11 then, other authors (Buck and Marshall, 2004; Sepúlveda *et al.*, 2013a) described the
12 presence of small bristles on the postpronotal lobe for species of the genera *Cerantichir* and
13 *Longina*. These were codified as reduced (state 1), with only *Od. longicornis* and *C.*
14 *peruana*, former *Od. peruanus*, having strong (state 2) postpronotal bristles. The character
15 was treated as ordered.
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20 *120. Proepisternal bristles: absent (0), one short and inconspicuous pair (1), one strong*
21 *and spine-like pair (2), many (3).* The presence of proepisternal bristles (referred to as
22 propleural by Aczél) is an important feature in the taxonomy of the higher Diptera
23 (McAlpine, 1981). Most species of the Australian-Oriental and Afrotropic regions present
24 strong proepisternal bristles, contrasting with the inconspicuous pair present in Neotropical
25 Neriidae, which is furthermore absent in many species. A patch of bristles in this region is
26 common among the Taeniapterinae.
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30 *121. Anterior pair of notopleural bristles: absent (0), hair-like (1), equal/subequal to*
31 *posterior pair (2).* The posterior pair of notopleural bristles is invariantly present and strong
32 in all species included in the analysis (Aczél, 1951). However, the anterior pair has a
33 tendency towards reduction (Aczél, 1954a, 1961), being totally absent (state 0) in a few
34 genera of the Australian-Oriental region (e.g.: *Telostylinus*, *Rhoptrum*, etc.); hair-like (state
35 1) in many genera of the Neotropics (e.g.: *Glyphidops*, *Nerius*, etc.); and as strong as the
36 posterior pair, although generally shorter in length, in genera from both the Neotropical and
37 Afrotropical regions (e.g.: *Longina*, *Eoneria*, *Chaetonerius*, etc.). The character was treated
38 as ordered.
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43 *122. Suprahumeral protuberances: absent (0), present (1).* Aczél (1961) acknowledged a
44 close relationship between *Cerantichir peruana* and *Longina abdominalis* given the shared
45 presence of a suprahumeral protuberance on which a strong spine sits. Later, Buck and
46 Marshall (2004) described the presence of this structure in *L. anguliceps* as well. They
47 furthermore discussed the presence of an enlarged first dorsocentral (lacking a
48 protuberance) as a possible synapomorphy of a *Longina* - *Cerantichir* clade. However,
49 suprahumeral bristles are common in many genera, and their strength is very polymorphic
50 within species, so only the presence of the protuberance was codified.
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55 *123. Presutural dorsocentral bristle: absent (0), present (1).* The Neriidae present a wide
56 variety of dorsocentral bristle patterns, ranging from only one to as much as nine (six if one
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3 leaves out the small bristles before the presutural pair in *Eoneria* and *Eoloxozus*, char. 127).
4 The presence of such “complete series of dorsocentrals” (D. K. McAlpine, 1974: 232)
5 common to some Neriidae, Cypselosomatidae and Pseudopomyzidae, have been regarded
6 as plesiomorphic with respect to the reduced pattern present in most micropezids.
7 Dorsocentral bristles were codified separately based on their position (chars. 123 to 127),
8 following the same principle adopted for the fronto-orbital bristles (chars. 88, 89). This
9 character refers to the bristle present before the transverse suture.
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14 *124. Postsutural dorsocentral bristle: absent (0), present (1).* See char. 117.

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16 *125. Prescutellar dorsocentral bristle: absent (0), present (1).* This bristle pair, situated just
17 before the scutellum, is only absent in the genera *Micropeza* (Aczél, 1951).
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20 *126. Number of extra dorsocentral bristles posterior to transverse suture: range 0 - 2.*
21 Between the postsutural and prescutellar bristle pairs, one or two more pairs of bristles may
22 be present, as large as the former. In case one bristle pair is present, this one is situated in
23 the middle of the postsutural area of the scutellum (characteristic of *Chaetonerius*); if two
24 are present, these divide this area into three more or less equally spaced regions
25 (characteristic of *Eoneria* and *Eoloxozus*). The character was treated as ordered.
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29 *127. Number of extra dorsocentral bristles anterior to transverse suture: range 0 - 3.* A
30 patch of small dorsocentral bristles are present in the genera *Eoneria* and *Eoloxozus* in front
31 of the prescutellar pair. This bristles are short yet strong and bristle-like, unlike the
32 inconspicuous hairs that are present in many other species of neriids, as for example
33 *Longina anguliceps* (Buck and Marshall, 2004) and some *Telostylinus* species (Aczél,
34 1959). The character was treated as ordered.
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38 *128. Katepisternal bristle: absent (0), present (1).*

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40 *129. Horizontal line of bristles in posterior margin of katepisternum: absent (0), present*
41 *(1).* The presence of this line of small bristles (referred to as hypopleural by Hennig, 1934;
42 and sternopleural by Aczél, 1951) is restricted to the Taeniopterinae and was taken to be a
43 symplesiomorphic character by Aczél (1951), who did not considered them to be
44 homologous to the single katepisternal bristle (char. 128) of the other Neriioidea, given that
45 they stand at different positions. In the Eurybatinae *Crosa yapensis*, he found what he
46 considered to be an intermediate form, having well differentiated katepisternal bristles
47 (char. 128) while retaining a reduced version of the taeniopterine “fan of erect bristle-like
48 hairs” (Aczél, 1959: 89; D. K. McAlpine, 1974).
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53 *130. Supra-alar bristle: absent (0), present (1).* Unlike the post-alar (*pa*) bristle, which is
54 always present, the supra-alar (*sa*) bristle is absent in *Glyphidops filosus* and *G. ochreus*
55 (Aczél, 1961).
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131. *Relative length of sa and pa bristles: approximately equal (0), pa larger (1).* Although the *pa* bristles are generally a little bit longer than the *sa* bristles, only in certain genera of Neotropical Neriidae do the *pa* bristles attain lengths considerably longer (30 - 50%) than that of the *sa* bristles.

132. *Non-apical scutellar bristles: absent/vestigial (0), long (1).* In the scutellum there is always an apical bristle pair, which is also generally the longest bristle pair on the thorax. However, in many species an extra bristle may be present. This can adopt the shape of a vestigial bristle, very small and hair-like (whose presence is generally polymorphic within species, and was therefore not codified as a separate state), or be strong and well-developed, although always shorter than the apical pair. This long non-apical scutellar pair is exclusively found in the Telostylinae and in *Gymnonerius fuscus*, all of which are described as having 2 pairs of scutellar bristles (Aczél, 1954a; 1955a), as well as in some Cypselosomatidae (D. K. McAlpine, 1966).

133. *Protuberances on apical scutellar bristles: absent (0), present (1).* These protuberances represent projections of the surface of the scutellum, in the shape of a cone or semisphere, in which the apical bristles stand, sometimes even longer than the bristle itself (Aczél, 1961).

134. *Protuberances on hind notopleural bristles: absent (0), present (1).*

135. *Coloration of katatergite: brown (0), yellow (1).* Aczél (1961) discussed the presence of an intensely yellowish coloration on the katatergite (or inferior pleurotergite) of some species of the genus *Glyphidops*, proposing that this character may be of taxonomic importance (Aczél, 1961). Among Old World neriids, the horizontal yellow vitta of the thorax is broader than in Neotropical species, sometimes including the katatergite. However, among many species in which this horizontal vitta encompasses the entire postnotum, the katatergite is nonetheless dark brown in colour, showing that the coloration of this region of the thorax is independent of the rest. Consequently, the presence of a yellow katatergite was also acknowledged for some species of the genus *Telostylus* and *Chaetonerius* (Aczél 1954a, 1959).

136. *Central stripe of pruinosity on mesonotum: absent (0), present (1).* A very complex and important feature of the neriid thorax is the pattern of pigmentation on the mesonotum (Aczél, 1961). The vast majority of species (except the genera *Gymnonerius*, *Teloneria* and *Telostylus*) have a central stripe covered by intense clear dusting, covering approximately the central third of the mesonotum.

137. *Color of pruinosity of mesonotum: yellow (0), gray (1).* Most Neotropical neriids have the central region of the mesonotum (char. 136) covered with whitish to grayish dusting, except in the genera *Cerantichir* and in *Glyphidops limbata*. In these and all species of the Old World, the pruinosity of the mesonotum is distinctly yellow.

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138. *Longitudinal dustless medial line in mesonotum: absent (0), present (1).* The central stripe of pruinosity of the mesonotum (char. 136) may be interrupted by a medial line which lacks the clear dusting and therefore presents the same coloration as the lateral regions of the mesonotum.

139. *Position of dustless medial line: anterior to transverse suture (0), posterior to transverse suture (1), complete (2).*

140. *Margins of dustless medial line: converging backwards (0), straight (1), diverging backwards (2).* Among some species of the genus *Glyphidops*, the dustless medial line of the mesonotum varies in shape along the longitudinal axis, sometimes becoming narrower posteriorly (state 0), sometimes becoming wider (state 2). Among all other species that possess this medial line, the borders of the line are straight, therefore not changing shape along the thorax.

141. *Longitudinal dustless lateral lines in mesonotum: absent (0), present (1).* A different pattern of pigmentation in the mesonotum results from the presence of two dustless lateral lines in the pruinose central stripe. This leads to a three-stripe pattern if the medial line is absent (char. 138), as in the *Telostylinus* or some *Nerius*, or a four-stripe pattern if the medial line is present, as in *Antillonerius*.

142. *Position of dustless lateral lines: anterior to transverse suture (0), posterior to transverse suture (1), complete (2).*

143. *Central yellowish vitta in scutellum: absent (0), present (1).* Among most species, the central region of the scutellum has a yellowish vitta, which presents diverse degrees of wideness. Among species with yellowish mesonotal pruinosity (char. 137) this vitta develops as a prolongation of the posterior region of the central stripe, but it is also present in many species with grayish pruinosity.

144. *Lateral pruinosity in scutellum: absent (0), present (1).* As has been described before, some species with grayish mesonotal pruinosity nonetheless develop a yellow vitta in the center of the scutellum. However, the two lateral stripes of grayish pruinosity among these species also continue into the lateral sides of the scutellum (state 1), which are therefore very different in color from the lateral dark brown regions of the mesonotum.

145. *Dustless regions at the base of bristles and hairs: absent (0), present (1).* Aczél (1961) drew attention to the similarities between *Eoloxozus sabroskyi* and *Odontoloxozus longicornis*, a resemblance based on many distinguishing characters, one of which is the insertion of the bristles and hairs of the pleurae, mesonotum and preabdominal tergites on dustless dots. This character is not present in any other species.

146. *Anal lobe and alula: reduced (0), developed (1).* The proximal region of the wing, including the anal lobe and the alula, is considerably reduced in the species of

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3 Micropezinae, resulting in a quite different wing morphology that significantly narrows
4 towards the wing stalk (see Aczél, 1951: lamina III). The wings of Cypselosomatidae and
5 Neriidae retain the plesiomorphic well-developed alula and anal lobe, while the same wing
6 configuration is interpreted as a consequence of a secondarily broadened wing-base in some
7 Taenipterinae and Eurybatinae (D. K. McAlpine, 1974).
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11 *147. Crossvein bm-cu: absent (0), vestigial/weak (1), conspicuous (2).* The crossvein bm-cu
12 is according to Aczél (1951) the only wing vein whose presence is variable within the
13 Neriioidea. Among the groups included in the analysis, this vein is completely absent in the
14 Micropezinae and in *Cypselosoma astralis*, reduced or vestigial in many Taenipterinae and
15 all Neriidae, and conspicuous only in some genera of Taenipterinae (e.g.: *Taeniptera*)
16 and Eurybatinae (Aczél, 1959). The character was treated as ordered.
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20 *148. First costal section: absent (0), present (1).* The apical parts of the Sc and R₁ veins are
21 divergent in the Taenipterinae and in all Neriidae, determining the first costal section
22 (char. 39, 40) or pterostigma according to Aczél (1951, 1954b). On the other hand, the
23 apical parts of these veins run parallel, never diverging, in the Micropezinae, the eurybatine
24 fly *Crosa yapensis* and the Cypselosomatidae (state 0).
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28 *149. Longitudinal veins R₄₊₅ and M₁₊₂: fusing at or near wing apex (0), not fusing (1).*
29 Among all families of Neriioidea included in the analysis, the veins R₄₊₅ and M₁₊₂ are
30 convergent, resulting in an r₄₊₅ cell that narrows towards wing apex. In some genera of
31 Micropezidae (e.g.: *Taeniptera*, *Micropeza*) these veins fuse with each other, sometimes at
32 the level of costal vein, sometimes before it, resulting in an apical crossvein (Aczél, 1951).
33 These species therefore lack a fourth costal section (chars. 45 and 46).
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37 *150. Subcostal break: absent (0), present (1).* The costal vein is more or less strongly
38 interrupted just proximal to where the subcostal vein joins it. This character was first
39 reported by Aczél (1951) for all revised species of Neriidae except for the genus *Longina*.
40 The character was further used as evidence to elevate Neriidae to the level of family
41 (McAlpine, 1974), and latter considered as a groundplan plesiomorphy of the Neriioidea,
42 given its shared presence in Pseudopomyzidae and Cypselosomatidae (McAlpine, 2000).
43 Aczél (1961: 270) stated that the subcostal break was present in most genera of neriids, and
44 some trace of it could be found in all studied specimens except for the genera *Longina*, for
45 which it was taken to be absent.
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50 *151. Direction of vein CuA₂: distal (0), proximal (1).* Among the Taenipterinae and
51 Eurybatinae, the vein CuA₂ meets the A₁ vein at a position that is distal with respect to the
52 point where it originates from CuA₁, therefore having a distal direction of development
53 (state 0). This direction is inverted in the rest of the included taxa (state 1), with the CuA₂
54 vein developing towards the wing base (Aczél, 1951; D. K. McAlpine, 1974).
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3 152. Length of A_1+CuA_2 vein: reaching posterior wing margin (0), short and not reaching
4 posterior margin (1), vestigial (2). The vein A_1+CuA_2 (pedicel of the anal cell in Aczél,
5 1951) is long among the Taeniopterinae and the Micropezinae, attaining the posterior
6 margin of the wing (state 0). On the other hand, among the Neriidae, Eurybatinae and
7 Cypselosomatidae it is always short (Aczél, 1951; D. K. McAlpine, 1966), generally having
8 about half the length it would take to attain hind wing margin (state 1), except in the genera
9 *Teloneria*, in which it is “vanishingly small” (Aczél, 1955a: 32 and plate 2), almost absent
10 (state 2). The character was treated as ordered.

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15 153. Shape of crossvein dm-cu: straight to slightly convex (0), strongly convex (1),
16 undulated (2). The shape of the crossvein dm-cu is typically straight (state 0), sometimes a
17 little bit outwardly convex, in most species studied. Only does this vein attain a relatively
18 high degree of curvature in the genus *Nerius* (Aczél, 1961) and in *Gymnonerius fuscus*
19 (Aczél, 1955a). Both in *Eoneria* and in *Eoloxozus*, and especially in the later, the vein is
20 undulated (state 2), adopting an S-shape (Aczél, 1961; Sepúlveda *et al.*, 2013b). This
21 feature, although never specified as such, is also shared by *Odontoloxozus longicornis* (see
22 Buck, 2010).

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27 154. Angle of crossvein dm-cu: transverse (0), slightly oblique (1), strongly oblique (2).
28 The Neotropical genera *Loxozus*, *Odontoloxozus* and *Eoloxozus* have a dm-cu crossvein
29 that is placed strongly oblique (state 2) with respect to the longitudinal veins, almost
30 moving parallel to posterior wing margin (Aczél, 1961). The rest of the species, have a vein
31 that is either normally placed (state 0), or only slightly oblique (state 1). This character was
32 treated as ordered.

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36 155. Infuscation of crossvein dm-cu: absent (0), present (1). Only one included species,
37 *Telostylinus zonalis*, presents an isolated brownish infuscation in the dm-cu cross vein
38 (Aczél, 1954a). However, both species that present supernumerary crossveins (char. 156)
39 also present a brown lined dm-cu vein, as part of a broader pattern of infuscation (Aczél,
40 1961; Sepúlveda *et al.*, 2013b).

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44 156. Supernumerary crossveins: absent (0), present (1). Only three species within the
45 Neriioidea have been described with supernumerary crossveins arising from the R_{2+3} and
46 M_{1+2} veins, all of which are neriids (Aczél, 1961). Two of these species, *Eoneria*
47 *maldonadoi* and *Glyphidops pluricellata*, have been included in the analysis.

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50 157. Bare areas of wing: absent (0), small (1), large (2). Buck and Marshall (2004)
51 described the absence of microtrichia in most of the basal region of the wings of the genus
52 *Longina* (state 2), extending beyond level of r-m cross vein in the r_1 , r_{2+3} and r_{4+5} cells, and
53 proposed it as a putative synapomorphy of the genus. However, they also discussed the
54 presence of less pronounced bare areas in *Cerantichir enderleini* (state 1). The rest of
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3 Neotropical Neriidae, as well as all Old World species revised, have completely
4 microtrichose wings (state 0). The character was treated as ordered.
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7 *158. Number of bristles in basicosta: range 0 - 3.* The number of bristles in the wide
8 basicosta of the Neriioidea varies considerably. It is always 3 in the Taenipterinae, 0 in the
9 Micropezinae and Eurybatinae, and a number between 0 and 2 in the Neriidae. Among the
10 last, the tendency towards reduction is stronger among Neotropical neriids, and generally
11 circumscribed to the *Nerius*-group, while the *Eoneria*-group and the Telostylinae retain the
12 presence of 2 bristles. The character was treated as ordered.
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16 *159. Ciliae on upper calypter: absent (0), short (1), long (2).* The general shape of the
17 calypteres does not vary within the Neriidae, being the upper calypter rounded or oval and
18 the inferior one reduced to a linear strip (or frenulum squamulare). However, some degree
19 of variation in the length of the marginal hairs present in the calypteres can be found. In
20 general, the upper calypter has long hairs (state 2), while the lower one has short ones (state
21 1), or they are completely absent (state 0). However, in the species *Longina anguliceps* this
22 tendency is reversed (Buck and Marshall, 2004), while in *Glyphidops ochreus* both
23 calypteres have equally long ciliae (Aczél, 1961). This character was treated as ordered.
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28 *160. Ciliae on lower calypter: absent (0), short (1), long (2).* See char. 159. This character
29 was treated as ordered.
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32 *161. Shape of halteres: lobulose capitulum and short pedicel (0), compact capitulum and*
33 *long pedicel (1).* The morphology of the halteres is modified only in the Taenipterinae
34 (state 1), which posses a small and compact capitulum (or knob) that is supported by a
35 relatively longer pedicel (or stem).
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38 *162. Antero and posteroventral row of spines on fore femur: absent (0), reduced and hair-*
39 *like (1), present (2).* Another of the distinguishing features of the Neriidae is the presence
40 of two rows of conspicuous spines on the ventral region of the femora (Aczél, 1951;
41 McAlpine, 1981; Buck, 2010). These bristles are nonetheless not present in all species,
42 having been lost (state 0) in some species of *Telostylinus*, *Teloneria* and *Antillonarius*. In
43 some other species, these rows only bear weak and hair-like bristles (state 1, present also in
44 the Cypselosomatidae and Eurybatinae), a pattern very common for Old World species, in
45 contrast with the stout spines of many Neotropical genera, which often sit on cylindrico-
46 conical protuberances. This character is profoundly sexually dimorphic, being involved in
47 the male-male aggressive displays (Eberhard, 1998), and was therefore only codified for
48 male specimens of each species. The presence of spines on mid and hind femora covaries
49 with the strength of their development on the fore femur, and where therefore also excluded
50 from the data. The character was treated as continuous.
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3 163. *Spinules on fore tibia: absent (0), present (1)*. Among some few species of neriids the
4 antero and posteroventral rows of spines of the fore femur (char. 162) continue into the
5 tibia, in the shape of smaller spinules.
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8 164. *Shape of dorsal face of mid femur: concave (0), straight to slightly convex (1)*. The
9 dorsal face of the mid femur is only concave in the *Logina - Odontoloxozus - Cerantichir*
10 clade, giving this segment an arched shape (Aczél, 1961). In the rest of the included
11 species, the mid femur presents a straight to convex shape, being more strongly curved on
12 species with short femora. This character also determines the place at which the femur
13 attains its largest width, a character often described by Aczél (1955a, 1961), being always
14 apical in species with concave mid femur and medial to distomedial in the rest.
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18 165. *Club-shaped thickening in apex of male fore tibia: absent (0), present (1)*. The
19 presence of a thickened apex of the fore tibiae was first described by Hennig (1937) as a
20 characteristic of the genus *Rhoptrum*. Aczél (1954a) later concluded that this character is
21 sexually dimorphic, being only present in the male sex.
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25 166. *Number of dorsal bristles in male procoxae: range 0 - 8*. All neriids have 1-2 apical
26 bristles on the procoxae. However, the number of dorsal bristles present is extremely
27 variable, and according to Aczél (1961) may be of taxonomic value. This character presents
28 an enormous degree of intraspecific variation, with larger specimens always having more
29 dorsal procoxal bristles than smaller ones. However, if this variability is included in the
30 analysis, phylogenetic signal may be retrieved. The character also presents a high degree of
31 sexual dimorphism, and was therefore codified separately for males and females (char.
32 167). Characters 166 and 167 were treated as ordered.
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37 167. *Number of dorsal bristles in female procoxae: range 0 - 6*. See char. 166.
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39 168. *Number of lateral bristles in mesocoxae: range 0 - 3*. The number of lateral bristles
40 (that is, excluding the apical ones) on the meso and metacoxae is very constant within
41 species and has a very interesting pattern of variation among taxa (Aczél, 1961). Characters
42 167 and 168 were treated as ordered.
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45 169. *Number of lateral bristles in metacoxae: range 1 - 3*. See char. 168.
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47 170. *Small dorsal bristles in mid and hind tibiae: absent (0), present (1)*. All along the
48 length of the mid and hind tibiae of the studied Micropezidae and Taenipaterinae there is a
49 row of tiny black bristles, that is always absent in Neriidae (Aczél, 1951).
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52 171. *Color of procoxa: yellow (0), brown (1)*. Among all species studied, the coxae can
53 present the same light to dark brown coloration of the pleurae (state 0), or they can be
54 yellowish, clearly different from the surrounding coloration (state 1). However, the color of
55 the meso and metacoxae is always the same (char. 172), whereas the color of the procoxa
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3 varies independently. Therefore, the color of the procoxae was divided into two separate
4 characters.
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7 *172. Color of meso and metacoxae: yellow (0), brown (1).* See char. 171.
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9 *173. Color of fore femur: yellow (0), brown (1).* The coloration of the legs is an important
10 taxonomic character, being commonly used in several generic keys (Aczél, 1954b, 1961).
11 Two attributes of the femora coloration were used, the base color and the presence of bands
12 or rings of differential coloration. Since the positional homology of these rings was often
13 difficult to establish, they were divided into three categories: basal (char. 174) and apical
14 (char. 176) stripes, usually broad and always attaining the base or apex of the
15 corresponding femur; and the number of central rings (char. 175), thinner, usually lacking a
16 conspicuous margin, and restricted to the central region of the femur. Furthermore, since
17 differences in patterning of the three femora are not uncommon, these four character were
18 codified separately for each femur (chars. 173-184).
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23 *174. Basal stripe in fore femur: absent (0), present (1).* See char. 173.
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25 *175. Number of central rings in fore femur: range 0 - 3.* See char. 173. The character was
26 treated as ordered.
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28 *176. Apical stripe in fore femur: absent (0), present (1).* See char. 173.
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30 *177. Color of mid femur: yellow (0), brown (1).* See char. 173.
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32 *178. Basal stripe in mid femur: absent (0), present (1).* See char. 173.
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34 *179. Number of central rings in mid femur: range 0 - 3.* See char. 173. The character was
35 treated as ordered.
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38 *180. Apical stripe in mid femur: absent (0), present (1).* See char. 173.
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40 *181. Color of hind femur: yellow (0), brown (1).* See char. 173.
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42 *182. Basal stripe in hind femur: absent (0), present (1).* See char. 173.
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44 *183. Number of central rings in hind femur: range 0 - 3.* See char. 173. The character was
45 treated as ordered.
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48 *184. Apical stripe in hind femur: absent (0), present (1).* See char. 173.
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50 *185. Color of tibiae: yellow (0), brown (1).* Unlike what happens with the femora, the color
51 of the tibiae is very homogenous, and was therefore merged into a single character. This
52 character nonetheless has shown to vary independently with respect to the coloration of the
53 femora.
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3 186. *Apical dark stripe in tibiae: absent (0), present (1)*. Most species have a pattern of
4 tibial coloration in which it becomes darker towards the tip (state 1). This happens both on
5 yellowish or brown tibiae (char. 185). Among other species, the tibiae are homogenously
6 dark brown (state 0).
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10 187. *Preabdominal median longitudinal vitta: absence (0), presence (1)*. Some species of
11 the genera *Telostylinus* and *Chaetonerius* have been inconsistently described by Aczél as
12 having brown or yellow background coloration in the preabdominal tergites (see for
13 example Aczél, 1954b and 1954c). Here, we adopt the hypothesis that the background
14 coloration of all neriid species (as well as the included Micropezidae and
15 Cypselosomatidae) is brown, since even among genera described as having yellow
16 background coloration there are species with entirely brown preabdominal tergites (e.g.:
17 *Ch. niger* and *T. papuanus*). Therefore, the pigmentation pattern of all neriid species can be
18 described by the presence/absence of 3 types of longitudinal vittae: a median thin vitta, two
19 wide lateral vittae (char. 188) and two marginal vittae, which are very wide and usually
20 divide the preabdomen into thirds (char. 189); all of these markings are whitish to
21 yellowish in color. When defined in this way, the presence of the median longitudinal vitta
22 is exclusive of the *Eoneria* – *Eoloxozus* clade (although it is also present as polymorphism
23 in some *Glyphidops*), while the lateral vittae are only found among those species of
24 *Chaetonerius* and *Telostylinus* that were described as having yellow tergites.
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31 188. *Preabdominal lateral longitudinal vittae: absent (0), present (1)*. See char. 187.
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33 189. *Preabdominal marginal longitudinal vittae: absent (0), present (1)*. See char. 187.
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35 190. *Shape of epandrium: flattened to hemispheric (0), semicylindrical (1)*. The epandrium
36 of the Micropezidae is generally shorter than that of the Neriidae (char. 69), yet its shape is
37 also quite different, with the epandrium of the Micropezidae lacking the clearly
38 semicylindrical shape common to all Neriidae (state 1). The shape of the epandrium of
39 Cypselosomatidae is much closer to that of the Neriidae than to the rest of included taxa
40 (McAlpine, 1974), and was therefore also codified as semicylindrical.
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44 191. *Lateral border of semicylindrical epandria: narrowing at the center (0), nearly*
45 *straight (1)*. The only significant difference in structure in the genitalia of all known neriid
46 species is present in the epandrium of three *Chaetonerius* species: *Ch. apicalis*, *Ch. collarti*
47 and *Ch. ghesquièrei* (known as the *apicalis*-group, Aczél, 1954b). These have a modified,
48 bulky epandrium (state 1), whose lateral margins run more or less straightly from base to
49 apex, and with enlarged and folded lateroapical regions. The rest of the family has a more
50 slender epandrium, clearly narrowing at the center, with a distinctive hour-glass shape.
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54 192. *Width of ventral furrow of epandrium: thin (0), intermediate (1), broad (2)*. The entire
55 ventral surface of the epandrium presents a longitudinal cleft in which the folded aedeagus
56 reposes. This “genital pouch” (Aczél, 1961: 272) is thin and linear in the Taeniapterinae,
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3 intermediately wide in Tylinae and Eurybatinae, and attaining its maximum broadness
4 among Neriidae (Aczél, 1951). The character was treated as ordered.
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7 *193. Surstyli: absent (0), vestigial/reduced (1), big (2).* The presence and development of
8 the surstyli are features that show significant differences among groups of Neriodea
9 (Aczél, 1951). This lobes are completely absent in the Taenipaterinae, while present in
10 other groups included in the analysis. However, the surstyli of Neriidae are much reduced,
11 almost vestigial, while this structure is well-developed in the Tylinae, Eurybatinae and
12 Cypselosomatidae. The character was treated as ordered.
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16 *194. Shape of ventral face of ovipositor: concave (0), convex (1).* The ventral surface of the
17 ovipositor is concave and sunken in most species studied. However, Aczél (1951) noted
18 that the ovipositor of the genera *Eoneria* and *Longina* was convex. Although this character
19 was not mentioned in subsequent descriptions, the ventral surface of the ovipositor proved
20 to be concave in all other revised species. The state of *L. anguliceps* and *L. semialba*, not
21 known by Aczél, were codified as missing data, since the shape of the female terminalia
22 was not specified in their taxonomic descriptions.
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Appendix S2: Material examined

All revised material is deposited in the Colección Entomológica, Instituto y Fundación Miguel Lillo, Tucumán, Argentina (curator Emilia Pérez). Acronyms are: BPBM, Bernice P. Bishop Museum, Honolulu; IRSN, Institut Royal des Sciences Naturelles de Belgique, Brussels; KUEC, Kyushu University, Fukuoka; MRAC, Musée royal de l'Afrique Centrale, Tervuren.

Cerantichir peruana – **Peru**: Valle Chanchamayo, 25.II.1929, L. Weyrauch, 1 male.

Chaetonerius apicalis – **Democratic Republic of the Congo**: Eala, I.1935 (ex IRSN, I. G. 10.482), J. Ghesquière, 1 male. Eala, VII.1935 (ex IRSN, I. G. 10.482), J. Ghesquière, 2 male and 1 female.

Chaetonerius brachialis – **Democratic Republic of the Congo**: Bambesa, 10.XII.1938 (ex IRSN, I. G. 12.234), J. Vrydagh, 1 male.

Chaetonerius collarti – **Democratic Republic of the Congo**: Eala, III.1936 (ex IRSN, I. G. 10.482), J. Ghesquière, 2 males (paratypes).

Chaetonerius inermis – **Indonesia**: West Java, Idjen plateau, Blawan, 950 m, VI.1924, K. W. Dammerman, 1 male and 1 female.

Chaetonerius latifemur – **Democratic Republic of the Congo**: Eala, 12.II.1935 (Ex IRSN, I.G. 10.482), J. Ghesquière, 1 female. Eala, III.1935 (ex IRSN, I .G. 10.482), J. Ghesquière, 1 male and 1 female. Eala, 7.IV.1935 (ex IRSN, I .G. 10.482), J. Ghesquière, 2 females. Eala, VII.1935 (ex IRSN, I .G. 10.482), J. Ghesquière, 1 male. Eala, 6.V.1936 (ex IRSN, I .G. 10.482), J. Ghesquière, 1 male. Rutshuru, XI.1937 (ex IRSN, I .G. 10.482), J. Ghesquière, 2 males. Rutshuru, 4.XII.1937 (ex IRSN, I .G. 10.482), J. Ghesquière, 1 female.

Cheatonerius niger – **Democratic Republic of the Congo**: Rutshuru, Kilinga, VI.1936 (ex MRAC), L. Lippens, 1 female. Kivu, Nzombe Amont, 200 m near Mwana, 1952 (ex MRAC), A. Froidebise, 1 male.

Chaetonerius perstriatus – **Democratic Republic of the Congo**: Upemba National Park, Kalule-Nord, near Klamalwa, 3-4.III.1949 (Mis. G. F. de Witte 2401a), R. Bowa, 1 male and 1 sex unknown (postabdomen missing; paratypes, labeled *Chaetonerius wittei*).

Eoloxozus sabroskyi – **Peru**: Quebrada Verde, Lurín, 10.X.1950, L. Weyrauch, 1 female.

1
2
3 *Eoneria blanchardi* – **Argentina**: Chacho, II.1974, 1 male. Corrientes, I.1950, D'Angelo, 1
4 female (paratype).
5

6
7 *Eoneria maldonadoi* – **Argentina**: La Rioja, XI.1952, 1 female (holotype). Catamarca,
8 Andaluallas, 2000 m, 19.I.1968, R. Golbach, A. L. Terán and H. Willink, 1 female.
9 “Vinagre D, La S., 9.IX.1965” (?), 1 female.
10

11 *Glyphidops neuter* – **Argentina**: Dpto. San Pedro, Carumbé, 28.I-10.III.1963, R. Golbach,
12 1 female (labeled *Oncopsia neutra*).
13

14
15 *Gymnonerius fuscus* – **Indonesia**: Mentawai Is., Sipora, 14.X.1924, H. M. Karny, 1 female.
16 North Sumatra, Medan, Saengei Krio, IV.1928, J.C. v.d. Meer Mohr, 1 female. Western
17 Java, Djampang-Tengah, Goenoeng Tjisoeroe, 600-800 m, III.1933, M. E. Walsh, 2 males
18 and 1 female. Western Java, Djampang-Tengah, Goenoeng Tjisoeroe, 600-800 m, IX.1933,
19 M. E. Walsh, 1 male.
20
21

22
23 *Longina abdominalis* – **Argentina**: Misiones, Puerto Bemberg, 12-29.I.1945, H. Willink
24 and R. Golbach, 1 male (labeled *Longina peletieri*). **Paraguay**: Caaguazú, Paso Yobai, 280
25 m, VI.1951, J. Foerster, 3 males (2 labeled *Longina vittata*, 1 labeled *Longina peletieri*).
26 Dpto. San Pedro, Carumbé, 28.I-10.III.1963, R. Golbach, 8 males and 3 females.
27
28

29 *Nerius czernyi* – **Argentina**: Misiones, Puerto Aguirre, 19.I.1931, K. Hayward, 1 female
30 (holotype).
31

32
33 *Nerius pilifer* – **Argentina**: Misiones, Puerto Bemberg, 12-29.I.1945, K. Hayward, H.
34 Willink and R. Golbach, 3 males and 3 female. Tucumán, Burruyacú, Villa Padre Monti,
35 17.I-7.II.1948, R. Golbach, 1 female. Jujuy, Caimancito, 26.V.1949, N. Kusnecow. Jujuy,
36 “Monrés, La-Minb” (?), 11.II.1951, 3 males. Salta, Orán, Abra Grande, 10.I-28.II-1967, R.
37 Golbach, 1 female. **Bolivia**: Santa Cruz, El Cidral, 1-28.I.1962, R. Golbach, 1 female.
38 **Brasil**: São Paulo, Paraná River, Porto Cabral, 1-25.IV.1944, M. Carrera and E. Dente, 2
39 males. São Paulo, Paraná River, Porto Cabral, 1-25.IV.1944, M. Carrera, 1 male and 3
40 females. São Paulo, Paraná River, Porto Cabral, 20-31.III.1944, Trav. Fo., M. Carrera and
41 E. Dente, 1 male and 4 females. **Paraguay**: Dpto. San Pedro, Carumbé, 28.I-10.III.1963, R.
42 Golbach, 1 male and 1 female.
43
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47 *Paranerius fibulatus* – **Indonesia**: New Guinea, Papua, Araucaria Camp, 800 m,
48 25.III.1939 (Archbold's Netherland Indian – American New Guinea Expedition), L. J.
49 Toxopeus, 1 male.
50
51

52 *Rhoptrum annulipes* – **Indonesia**: New Guinea, Papua, Bernhard Camp, 50 m, 12.IX.1938
53 (Archbold's Netherland Indian – American New Guinea Expedition), J. Olthof, 3 males and
54 2 females. New Guinea, Papua, Araucaria Camp, 800 m, 7.III.1939 (Archbold's Netherland
55 Indian – American New Guinea Expedition), L. J. Toxopeus, 1 female.
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3 *Rhoptrum lieftincki* – **Indonesia**: New Guinea, Papua, Rattan Camp, 1500 m, 12.III.1939
4 (Archbold's Netherland Indian – American New Guinea Expedition), L. J. Toxopeus, 1
5 male (paratype). New Guinea, Papua, Araucaria Camp, 800 m, 20.II.1939 (Archbold's
6 Netherland Indian – American New Guinea Expedition), L. J. Toxopeus, 1 female
7 (paratype).
8
9

10
11 *Telostylinus gressitti* – **Federal States of Micronesia**: Faraulep atoll, Faraulep Is., 21-
12 .1952, N. Krauss, 1 male (paratype). Ifaluk atoll, Ifaluk Is., 4.XI.1953, M. Bates, 1 male
13 (paratype). **Palau**: Babelthuap Is., Ulimang, 10.XII.1947, H. S. Dybas, 1 female (paratype).
14 Peleliu Is., Mt. Amiangel, 22.XII.1952, J. L. Gressitt, 1 male (paratype). Koror, 18-
15 20.IV.1955, J. W. Beardeley, 1 female (paratype).
16
17

18
19 *Telostylinus lineolatus* – **Federal States of Micronesia**: Kusaie, Mutunlik, 22 m,
20 15.II.1953, J. F. G. Clarke, 1 female. **Indonesia**: Western Java, Bandoeng, 700 m,
21 17.III.1940, J. Olthof, 2 males.
22

23
24 *Telostylinus longicoxa* – **Federal States of Micronesia**: Kusaie, Lelu Is., Mt. Fenkol,
25 30.I.1936, Z. Ono, 1 male (ex BPBM). Truk (Chuuk) State, Weno Is., S. Valley, Mt.
26 Tonaachau, 4.IV.1949, R. W. L. Potts, 1 male. Pohnpei (Ponape) Is., Kolonia, VI-IX.1950,
27 P. A. Adams, 1 male. Kusaie, Mutunlik, 22 m, 15.II.1953, J. F. G. Clarke, 1 female. Truk
28 (Chuuk) State, Weno Is, Mt. Teroken, Nantaruil, J. L. Gressitt, 1 female. **Marshall**
29 **Islands**: Arno Atoll, Ine Is., 28.VII.1950, I. La Rivers, 1 male. Namu atoll, Namu Is.,
30 24.X.1953, J. W. Beardley, 1 female. Lae atoll, Lae Is., 24.X.1953, J. W. Beardley, 1
31 female. **Northern Mariana Islands**: Saipan Is., Matansha-Calabera, 3.V.1940, Yasumatsu
32 and Yoshimura, 1 male (ex KUEC).
33
34
35

36
37 *Telostylinus papuanus* – **Indonesia**: New Guinea, Papua, Bernhard Camp, 50 m, 7.XI.1938
38 (Archbold's Netherland Indian – American New Guinea Expedition), J. Olthof , 1 female.
39 New Guinea, Papua, Sigi Camp, 1500 m, 2.II.1939 (Archbold's Netherland Indian –
40 American New Guinea Expedition), L. J. Toxopeus, 1 female. New Guinea, Papua, Rattan
41 Camp, 1200 m, 4.II.1939 (Archbold's Netherland Indian – American New Guinea
42 Expedition), L. J. Toxopeus, 2 males.
43
44

45
46 *Telostylinus ponapensis* – **Federal States of Micronesia**: Pohnpei (Ponape) Is., southeast
47 of Nanpohnmal, 70 m, 11.I.1953, 1 female (paratype). Pohnpei (Ponape) Is., Mt.
48 Tamatamansakir, 180 m, 17.I.1953, J. L. Gressitt, 1 male (paratype).
49

50
51 *Telostylinus spinicoxa* – **Indonesia**: New Guinea, Papua, Jayapura (ex Hollandia),
52 VII.1938 (Archbold's Netherland Indian – American New Guinea Expedition), L. J.
53 Toxopeus, 1 male (paratype).
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Telostylinus yapensis – **Federal States of Micronesia**: Yap, Yap Is., Mt. Matade, 95 m, 1.XII.1952, J. L. Gressitt, 1 female (paratype). Yap, Yap. Is., 1952, N. L. H. Krauss, 1 female.

For Peer Review

Appendix S3: Scripts

SPR distance tuning (SPRsensitivity.run)

The following script generates 100 Wagner trees, groups them in pairs, and calculates the same 50 weighted SPR distances under a variety of combinations of the two parameters that determine its efficiency in finding the shortest distance, *i.e.* number of replicates and levels of stratification. Results are saved to a comma-delimited CSV file which can be opened directly with MS-Excel. The weight assigned to moves can be easily modified (or eliminated for unweighted SPR distances), as well as the values of the parameters explored.

```

macro = ;

mult 100 = wagner keepall ;

var =
    + numrepl
    + numstrat
    + sprmoves
    + tree1
    + tree2 ;

sprdiff [3 ;

report - ;
silent = file ;
log SPRdistances.csv ;
silent - file ; quote Tree1, Tree2, Stratifications, Replicates,
Movements ; silent = file ;

loop 0 30
    set numstrat #1 ;

    loop 1 20
        set numrepl #2 * 1000 ;

        loop 0 ntrees
            if ( #3 < 50 )
                set tree1 #3 * 2 ;
                set tree2 #3 * 2 + 1 ;
                if ( #1 == 0 )
                    set sprmoves sprdiff [ 'tree1' 'tree2'
'numrepl' ] ;
            else

```

```

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```

```

                                set sprmoves sprdiff [ 'tree1' 'tree2'
'numrepl'x'numstrat' ] ;
                                end
                                silent - file ; quote 'tree1', 'tree2',
'numstrat', 'numrepl', 'sprmoves' ; silent = file ;
                                else
                                continue ;
                                end
                                stop
                                stop
                                stop
proc/;

```

Collapsing of continuous character (collapsing.run)

This script allows collapsing couples of continuous characters into a single one without modifying the original matrix. To do so, the smallest and largest values of both are stored as a range on one of the characters, with the other becoming inactivated. In its present form, the script does so only for contiguous couples of characters, but can be modified for different matrix configurations. The number of characters being collapsed can be modified easily by stating a list of exceptions (collexceptions.txt) that is read by the script, and the number of exceptions has to be stated as argument.

```

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```

```

macro = ;

var =
    + exceptions [%1]
    + numExceptions
    + numcont
    + min
    + max
    + exception
    + continueLoop
    + nextLoop
    + secondchar ;

proc collexceptions2.txt ;

set numExceptions %1 ;
set numExceptions -- ;

loop 0 nchar
if ( !iscont[#1] ) set numcont #1 - 1 ;
endloop end
stop

```



```

1
2
3
4 loop 0 ('numcont' - 1)
5
6   quote #1;
7
8   set continueLoop 0 ;
9
10  loop 0 'numExceptions'
11    set exception 'exceptions[#2]' ;
12    if(#1 == 'exception')
13      set continueLoop 1 ;
14      continue;
15    end
16  stop
17
18  if ('continueLoop' == 1) continue; end
19
20  loop 0 ntax
21
22    if ( contmaxs[#1 #2] == 65.535 )
23      if ( contmaxs[(#1+1) #2] == 65.535 )
24        continue ;
25      else
26        set min contmins[(#1+1) #2] ;
27        set max contmaxs[(#1+1) #2] ;
28      end
29    else
30      set min contmins[#1 #2] ;
31      set max contmaxs[#1 #2] ;
32      if ( !(contmaxs[(#1+1) #2] == 65.535) )
33        if(contmins[(#1+1) #2] < 'min') set min
34        contmins[(#1+1) #2] ; end
35        if(contmaxs[(#1+1) #2] > 'max') set max
36        contmaxs[(#1+1) #2] ; end
37      end
38    end
39
40    quote MIN-MAX ;
41    quote 'min'-'max' ;
42
43    xread = #1 #2 'min'-'max' ;
44
45  stop
46
47  set secondchar #1 + 1 ;
48
49  ccode ] 'secondchar' ;
50
51  if (#1 < ('numcont' - 1))
52    set nextLoop #1 + 2 ;
53    setloop 'nextLoop' ;
54  end
55
56
57
58
59
60

```

```

1
2
3
4     stop
5
6     loop 0 'numExceptions'
7         set exception 'exceptions[#1]' ;
8         ccode [ 'exception' ;
9     stop
10
11
12 proc/;
13
14
15

```

collexceptions.run:

```

16
17
18 macro = ;
19
20
21 set exceptions[0] 10;
22 set exceptions[1] 29;
23 set exceptions[2] 50;
24 set exceptions[3] 67;
25
26 ...
27 proc/;
28
29

```

Balance of characters (subsampleSPR.run)

This script searches for the optimal tree of the complete continuous partition and saves it to a TRE file. It then generates n pseudoreplicates of m continuous characters by activating them randomly (n and m have to be stated as arguments), searches for up to 10 optimal trees supported by each pseudoreplicate, calculates the weighted SPR distance of all of the trees found to the saved one, and retains the shortest value. This is done in order to avoid the use of consenses, which may result in the subestimation of SPR distances. Values are subsequently saved to a comma-delimited CSV file which can be opened directly with MS-Excel. The script can be easily modified to be used for entire matrices or different kinds of partitions. Tree search and SPR distance parameters can be also changed with ease.

```

46
47
48
49 macro = ;
50
51
52 var =
53     + numrepl
54     + numcharac
55     + randomCharac
56     + sprmove
57     + sprparcial
58
59
60

```

```

1
2
3         + sprsimilarity ;
4
5     set numrepl %1 ;
6     set numcharac %2 ;
7
8     macfloat 8 ;
9     taxname - ;
10    col 0 ;
11    rseed 0 ;
12
13
14    sprdiff [3 ;
15
16    loop 0 nchar
17        if ( !iscont [#1] )
18            ccode ] #1 ;
19        else
20            continue ;
21        end
22    stop
23
24    mult = tbr ratchet replic 10 hold 10 ;
25    tsave *trees.tre ;
26    save ;
27    tsave/ ;
28
29
30    ccode ] . ;
31
32    report - ; silent = file ; log SPRresults.csv ; silent - file ;
33    quote Moves ; silent = file ;
34
35    loop 1 'numrepl'
36
37        keep 0 ;
38
39        loop 1 'numcharac'
40            set randomCharac getrandom [ 0 nchar ] ;
41            if ( (iscont['randomCharac']) &&
42                (!isact['randomCharac']) )
43                ccode [ 'randomCharac' ;
44            else
45                setloop #2 - 1 ;
46            end
47        stop
48
49        mult = tbr noratchet replic 20 hold 10 ;
50        reroot 0 ;
51
52
53        proc trees.tre ;
54        set sprmove 10000 ;
55
56        loop 0 (ntrees - 1)
57            set sprparcial sprdiff [ #2 ntrees 13000x18 ] ;
58
59
60

```

```
1
2
3         if ( 'sprparcial' < 'sprmove' )
4             set sprmove 'sprparcial' ;
5         end
6     stop
7
8     silent - file ; quote 'sprmove' ; silent = file ;
9
10    ccode ] . ;
11
12 stop
13
14 proc/;
```

For Peer Review