

A further step towards the characterisation of Terebellides (Annelida, Trichobranchidae) diversity in the Northeast Atlantic, with the description of a new species

María Barroso¹, Juan Moreira², María Capa³, Arne Nygren^{4,5}, Julio Parapar¹

I Departamento de Bioloxía, Universidade da Coruña, A Coruña, Spain 2 Departamento de Biología (Zoología) & Centro de Investigación en Biodiversidad y Cambio Global (CIBC-UAM), Facultad de Ciencias, Universidad Autónoma de Madrid, Madrid, Spain 3 Departament de Biologia, Universitat de les Illes Balears, Mallorca, Spain 4 Sjöfartmuseet Akvariet, Göteborg, Sweden 5 Institutionen för marina vetenskaper, Göteborgs Universitet, Göteborg, Sweden

Corresponding author: María Barroso (maria.p.barroso@udc.es)

Academic editor: Greg Rouse Received 4 August 2022 Accepted 23 October 2022 Published 28 November 2022
https://zoobank.org/4168C32E-37A7-4912-A909-4912E69030AA

Citation: Barroso M, Moreira J, Capa M, Nygren A, Parapar J (2022) A further step towards the characterisation of *Terebellides* (Annelida, Trichobranchidae) diversity in the Northeast Atlantic, with the description of a new species. ZooKeys 1132: 85–126. https://doi.org/10.3897/zookeys.1132.91244

Abstract

Several new species of genus *Terebellides* Sars, 1835 (Annelida, Trichobranchidae) have been recently described from the Northeast Atlantic Ocean after the detection of a large complex of species based on DNA sequence data from previous research. Some of those species (belonging to the so-called Group A) have already been described elsewhere. In this paper, we revise several *Terebellides* clades belonging to Groups B, C and D resulting in the identification of five nominal species: *Terebellides gracilis* Malm, 1874, *Terebellides atlantis* Williams, 1984, *Terebellides williamsae* Jirkov, 1989, *Terebellides irinae* Gagaev, 2009, and *Terebellides shetlandica* Parapar, Moreira & O'Reilly, 2016, plus one new species described here as *Terebellides lavesquei* **sp. nov.** All these species are characterised by a combination of morphological features complemented with a nucleotide diagnostic approach (specific COI nucleotides in the alignment position). Morphological characters used to discriminate between taxa refer to the branchial shape, presence/absence of ciliated papillae dorsal to thoracic notopodia and the morphology of thoracic and abdominal uncinal teeth. An updated identification key to all described species of this genus in NE Atlantic waters is also included.

Keywords

DNA barcoding, DNA species delineation, identification key, integrative taxonomy, Northeast Atlantic, polychaetes, SEM, systematics

Copyright María Barroso et al. This is an open access article distributed under the terms of the Creative Commons Attribution License (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Introduction

The genus *Terebellides* Sars, 1835 (Annelida) is distinguishable from other members of Trichobranchidae by the unique mid-dorsal stalk bearing the characteristic branchial lobes, provided with packed branchial lamellae. This taxon is morphologically homogenous and boundaries between species are difficult to assess because relevant characters rely on some microscopic details. These include features of branchiae, chaetae and uncini that need examination with Scanning Electron Microscopy (Parapar et al. 2016c, 2020a; Lavesque et al. 2019). In addition, further work is still needed to assess morphological intra- and interspecific variability for some characters.

A recent comprehensive molecular survey and a set of analytical methods (Nygren et al. 2018) revealed that the actual diversity of *Terebellides* is much higher than expected increasing from 5 to 25 the total number of species present in the NE Atlantic. Furthermore, molecular-based phylogenetic analyses by Nygren et al. (2018) facilitated the search of diagnostic characters. Thus, three nominal species have been identified, including the type species, *Terebellides stroemii* Sars, 1835, and six have been described new to science (Parapar et al. 2020a). All these belonged to the so-called Group A (sensu Nygren et al. 2018). In addition, Lavesque et al. (2019) described eight new species from France based in a molecular survey as well. Many of these recently described species had been usually misidentified as *T. stroemii*; this taxon was previously thought as having a cosmopolitan distribution and resulted therefore in many species being overlooked worldwide.

Nygren et al. (2018) stablished four main groups of species: A, B, C, and D, which differ from each other by several morphological characters. Parapar et al. (2020b) studied group A species that are characterised by greater overall body length (10.0–50.0 mm), branchial lobes almost completely fused with ventral lobes that were partially or totally obscured, and the presence of papillae on margins of branchial lamellae in some species.

The aim of the present study is the morphological and molecular characterisation of members of Groups B, C, and D after Nygren et al. (2018). A total of five nominal species was identified and a new species is here described as *Terebellides lavesquei* sp. nov.

Materials and methods

This paper is based on the study of 215 *Terebellides* specimens belonging to Groups B, C, and D as defined in Nygren et al. (2018) (Table 1); five correspond to previously described species and one is new to science. This material is deposited in the Zoological Museum Bergen (**ZMBN**, Bergen, Norway), Goteborg Natural History Museum (**GNM**, Goteborg, Sweden), and the Norwegian University of Science and Technology, Science Museum (**NTNU-VM**, Trondheim, Norway; Bakken et al. 2020).

These specimens are part of a large collection of *Terebellides* specimens (table S1 in Nygren et al. 2018) mostly collected in the Norwegian and Swedish continental shelf

but also from the Irish and Celtic seas, North Sea, Barents Sea, Greenland Sea, South Icelandic coast, and the Arctic Ocean.

Light microscope photographs were done using an Olympus SZX12 stereomicroscope equipped with an Olympus C-5050 digital camera. Line drawings were made with a Wacom CTL-4100K-S pen tablet based on photographs and observations made with an Olympus BX40 stereomicroscope. Specimens for Scanning Electron Microscopy (SEM) were prepared by critical point drying, covered with gold and examined and photographed under a JEOL JSM-6400 electron microscope at the Servizos de Apoio á Investigación (**SAI**, Universidade da Coruña, Spain).

For staining procedures, 10 mg of Methyl Green (MG) colourant were dissolved in 5 ml of 20% ethanol and specimens were held in there for 30 s. MG staining patterns and thoracic uncini morphology were characterised based on the classifications proposed by Schüller and Hutchings (2010, 2013) and Parapar et al. (2020b), respectively; only specimens of similar/comparable size were considered.

For each species, the list of the museum registration numbers and collection details (geographic area, locality, coordinates, depth, collecting date and habitat) is provided in Suppl. material 1. Unless specified, each registration number holds a single specimen; associated GenBank DNA sequence accession numbers are provided in Suppl. material 2.

The correspondence between species numerals (Nygren et al. 2018) and names is as follows: species 1 – *Terebellides shetlandica* Parapar, Moreira & O'Reilly, 2016; species 5 – *Terebellides lavesquei* sp. nov.; species 16 – *Terebellides atlantis* Williams, 1984; species 24 – *Terebellides irinae* Gagaev, 2009; species 2 – *Terebellides williamsae* Jirkov, 1989; species 3 – *Terebellides gracilis* Malm, 1874.

The present study deals with the main Terebellides groups B, C, and D, proposed by Nygren et al. (2018) after phylogenetic analyses of nuclear (28S rDNA and internal transcriber spacer 2, ITS2) and mitochondrial (cytochrome C oxidase I and 16S r DNA) markers from specimens of Northeast Atlantic (NEA) Terebellides, representing a follow-up to Parapar et al. (2020a) who characterised the species within Group A. In this way, additional analyses with only the COI dataset have been performed in order to assess diagnostic nucleotides for each of the species and the genetic distances between them. Phylogenetic analyses of COI Terebellides sequences in GenBank generated by Nygren et al. (2018) and Lavesque et al. (2019) were performed, using Trichobranchus roseus (Malm, 1874), Polycirrus sp., and Pista cf. cristata (Müller, 1776) as outgroups (Nygren et al. 2018). Methodology followed that described by Parapar et al. (2020a) and included alignment of 471 sequences with MAFFT version 7.017 (Katoh et al. 2002), calculation of the best substitution model (TVM+F+I+G4), according to Bayesian information criterion - BIC with IQTREE version 1.6.11 (Nguyen et al. 2015). Maximum likelihood phylogenetic analyses were run in IQTREE version 1.6.11 (Nguyen et al. 2015), with ultrafast bootstrap (Hoang et al. 2018). Unequivocal nucleotide diagnostic characters are provided as the positions in the alignment, shown in Suppl. material 2.

The most distinctive taxonomic morphological characters for Terebellides include morphology of branchiae (sensu Parapar et al. 2016a), type of thoracic uncini (sensu Parapar et al. 2020b) and abdominal uncini (sensu Parapar et al. 2020a); Methyl Green (MG) staining pattern (sensu Schüller and Hutchings 2010, 2013) and geographic and bathymetric distribution data are also useful to discern species. Regarding branchiae, Parapar et al. (2016a) proposed four types: type 1, with large lobes almost completely fused; type 2, with lobes fused ~ 50% of their length; type 3, with lobes only fused at base; and type 4, with small lobes not fused and reduced dorsal lobes. Parapar et al. (2020a, b) also defined four types of thoracic uncini and three types of abdominal uncini based on the rostrum vs. capitium length ratio (RvC), and the relative size of the capitium teeth. In the species studied here, three types of thoracic uncini have been identified: type 1 - RvC = 2(3)/1, capitium with two or three large teeth and subsequent ones much smaller; type 3 - RvC = 1/1, capitium with four or five mid-sized teeth followed by slightly smaller teeth; type 4 - RvC = 1/1, capitium with 5–7 small teeth and remaining ones similar in size at least in two rows. Two types of abdominal uncini were also identified: type 1A - RvC = 1/0.7, capitium with 3-5 large teeth in first row and one or two in a second row; type 2 - RvC = 1/0.9, capitium with four or five teeth and remaining ones smaller.

Schüller and Hutchings (2010, 2013) defined several types of Methyl Green staining patterns. The patterns observed in the species studied here are similar to the following patterns: pattern 1, segments (SG) 1–6 solid and SG 7–14 striped; pattern 2, SG 1–5 solid, SG 6 white and SG 7–14 striped; and pattern 9, SG 1–5 solid, SG 3 with J-shape glandular region, SG 6 dark solid and SG 7–18 striped.

Abbreviations used in text, table, and figures:

abl	anterior branchial lobe (lobe #5);
bdl	branchial dorsal lobes;
bdltp	branchial dorsal lobe terminal papilla;
bf	branchial filament;
bvl	branchial ventral lobes;
cap	capitium;
сор	copepod;
cr	ciliary row;
ct	ciliary tuft;
dpn	dorsal projection of notopodium;
gc	geniculate chaetae;
MG	Methyl Green;
NEA	Northeast Atlantic;
nc	notochaetae;
00C	oocytes;
RvC	rostrum vs. capitium length ratio;
SEM	Scanning Electron Microscope;

SG	segment;
STM	stereomicroscope;
TC	thoracic chaetiger;
tdp	thoracic dorsal papilla;
tll	thoracic lateral lappets;
tm	tentacular membrane;
TU	thoracic unciniger;
wTC	white thoracic chaetiger.
	-

Systematic account

Five lineages of the *Terebellides* Groups B, C, and D (sensu Nygren et al. 2018) were identified as nominal species already reported in the Northeast Atlantic: *Terebellides gracilis* Malm, 1874, *T. atlantis* Williams, 1984, *T. williamsae* Jirkov, 1989, *T. irinae* Gagaev, 2009, and *T. shetlandica* Parapar, Moreira & O'Reilly, 2016. In addition, one of the lineages did not match any of the previously known *Terebellides* species and is herein describes as new: *Terebellides lavesquei* sp. nov. (Fig. 1).

Following Nygren et al. (2018) nomenclature and grouping, species have been here grouped as follows: Group B – species 1, 5, and 16, Group C – species 24, and Group D – species 2 and 3; description of each taxon will follow this order. Species 17, 27 (Group B), and 22 (Group C) are represented by 1–4 specimens each (see appendix S36 in Nygren et al. 2018) and are pending formal description until more material is available. Species 4, 14, and 26 (Group B), 25 (Group C), and 15 (Group D) will be described by D. Gaeva and I. Jirkov (Shirshov Institute of Oceanology, Russia). For correspondence between species names and numerals see below.

Family Trichobranchidae Malmgren, 1866

Genus Terebellides Sars, 1835 emended by Schüller & Hutchings, 2013

Type species. *Terebellides stroemii* Sars, 1835, redescribed by Parapar and Hutchings (2014) and neotype deposited.

Terebellides Group B (sensu Nygren et al. 2018)

Figs 2–12

Description. The morphological features shared by all examined species in Group B in this paper (clades 1, 5, and 16) are itemised below. Some of these are also shared by Groups A, C, and D as defined in Nygren et al. (2018) (see Remarks below). Clades 4, 14, and 26 will be studied elsewhere; formal descriptions of clades 17 and 27 will wait until more material is available.

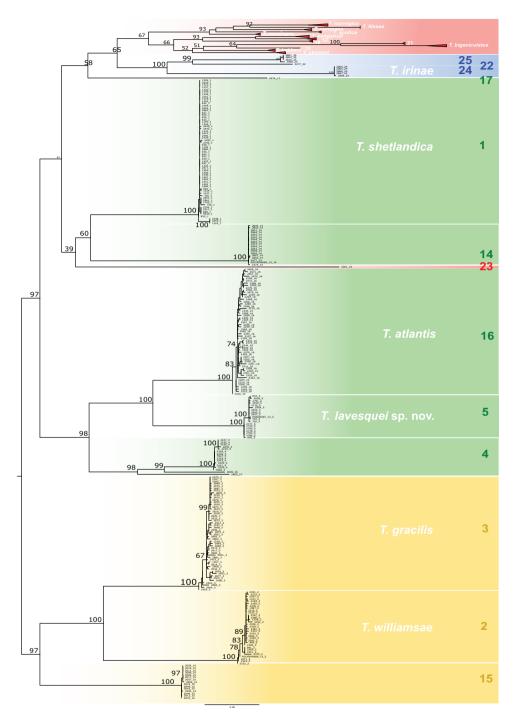


Figure 1. Phylogenetic tree after Maximum Likelihood analyses on a dataset of all COI sequences in Nygren et al. (2018) and Lavesque et al. (2019). Bootstrap support values above nodes. Species other than members of Groups B, C, and D are collapsed.

Body appearance. Complete individuals ranging from 5.0–35.0 mm in length. Body tapering posteriorly with segments increasingly shorter and crowded towards pygidium. Prostomium compact; large tentacular membrane surrounding mouth (Figs 2B, 3B, C), with typical buccal tentacles with expanded tips (Figs 2A, B, 3A, B). SG 1 as an expanded structure below tentacular membrane in a lower lip.

Branchiae. Branchiae arising as single structure from SG 3, with a single stalked mid-dorsal stem (Figs 2A, B, 3B, C, 4B), lobes not fused or partially fused, ventral ones obscured or not by dorsal ones (Figs 2A, B, 3A–C, 4A, B). Dorsal lobes ending posteriorly in short terminal papilla and ventral lobes ending in long filaments (Figs 2B, 3A, B). Anterior projection of dorsal lobes (fifth lobe) normally absent but present only in clade 5 (Fig. 3B). Posterior end of dorsal lobes reaching TC 4 (Figs 2A, B, 3A, B). Branchial lamellae provided with several parallel rows of cilia in inner face (Fig. 6C) and ciliary tufts not observed. Ciliary papillae absent in branchial lamellae margin.

Thorax. Eighteen pairs of notopodia (SG 3–20), those of TC 1 ca. as long as subsequent ones or slightly shorter (Figs 2A, 3A–C). Lateral lappets and dorsal projections of notopodia in anterior thoracic chaetigers with different degree of development depending on size and preservation conditions, but both more conspicuous on TC 1–4 (Fig. 3A–C). All notochaetae as simple capillaries (Fig. 5B). Size of notochaetae of TC 1 similar to subsequent ones. Neuropodia as sessile pinnules from TC 6 to body end, with uncini in single or double rows, from TC 7 throughout. Neuropodia on TC 6, provided with several sharply bent, acute-tipped, geniculate chaetae with minute teeth forming a capitium only visible with SEM (Figs 5C, 7A, 8C). From TC 7, neuropodia with one or several rows of uncini per torus (Figs 5D, 7B, C, 8D), with long shafted denticulate hooks, with large main fang (rostrum) longer than upper crest of teeth (capitium), rostrum/capitium length ratio of ~ 2:1, capitium composed by several teeth above main fang of decreasing length (Figs 5D, 7B, 8D).

Abdomen and pygidium. Approximately half as long as thorax and progressively thinner (Fig. 2A–C). Neuropodia ranging from 18–34 chaetigers and forming erect pinnules (Figs 5E, 8E) with several uncini per torus, number depending on specimen size. Uncini provided with several teeth above rostrum surmounted by a capitium composed of several teeth of decreasing length (Figs 5F, 7D, 8F). Pygidium blunt, as funnel-like depression.

Colour pattern. Colour in preserved specimens whitish or pale brown (Fig. 3A–C). MG staining pattern characterised by 1) compact green colourant in SG 1–6, then turning into striped pattern in SG 7–14 and fading in following segments or 2) by compact green colourant in SG 1–6, J-shaped glandular region in SG 3–5, and striped pattern in SG 7–14 (Fig. 12).

Remarks. Among the aforementioned characters, branchial features might serve to distinguish most of Group B species from those of Groups A, C, and D. Those include size of branchial lobes, lobes not fused, presence of long filaments on ventral ones, and presence of ciliary rows on branchial lamellae. Other taxa described or reported worldwide bear the same type of branchiae (type 3), including *Terebellides ehlersi* McIntosh, 1885, *T. intoshi* Caullery, 1915, *T. lobatus* Hartman & Fauchald, 1971, *T. mundora* Hutchings & Peart, 2000 and *T. sepultura* Garraffoni & Lana, 2003 (Parapar et al. 2016a, b).

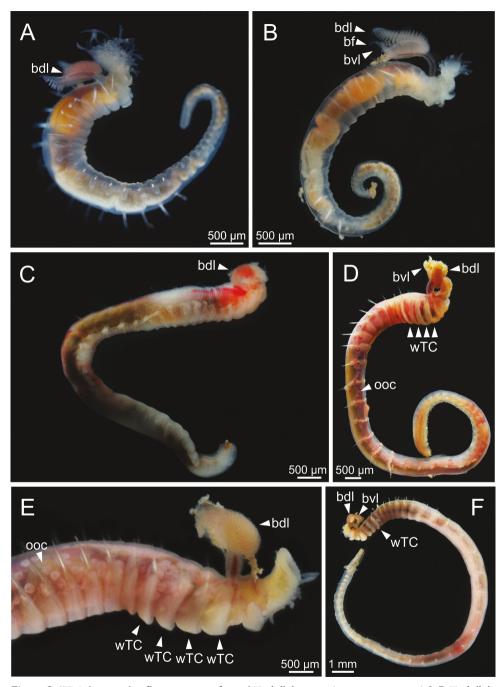


Figure 2. STM photographs of live specimens of several *Terebellides* species (non-type specimens) A, B *Terebellides* shetlandica Parapar, Moreira & O'Reilly, 2016 (species 1; A ZMBN116171 B ZMBN116181) C *Terebellides* lavesquei sp. nov. (species 5; GNM15112) D, E *Terebellides williamsae* Jirkov, 1989 (species 2; D GNM15108
E GNM15109) F *Terebellides gracilis* Malm, 1874 (species 3; GNM15111). Abbreviations: bdl – branchial dorsal lobe; bf – branchial filament; bvl – branchial ventral lobe; ooc – oocytes; wTC – white thoracic chaetiger.

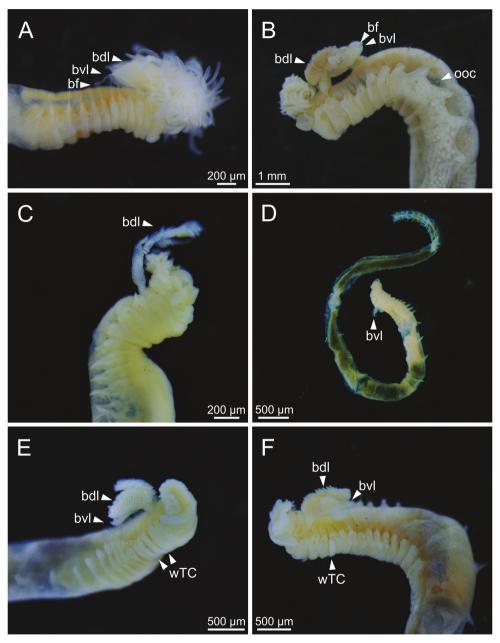


Figure 3. STM photographs of several *Terebellides* species (A, C-F non-type specimens) A *Terebellides shetlandica* Parapar, Moreira & O'Reilly, 2016 (species 1; ZMBN116186) B *Terebellides lavesquei* sp. nov. (species 5; holotype, ZMBN116322) C *Terebellides atlantis* Williams, 1984 (species 16; ZMBN116472) D *Terebellides irinae* Gagaev, 2009 (species 24; ZMBN116498) E *Terebellides williamsae* Jirkov, 1989 (species 2; ZMBN116269) F *Terebellides gracilis* Malm, 1874 (species 3; ZMBN116283). Abbreviations: bdl – branchial dorsal lobe; bf – branchial filament; bvl – branchial ventral lobe; ooc – oocytes; wTC – white thoracic chaetiger.

Terebellides shetlandica Parapar, Moreira & O'Reilly, 2016

Figs 2A, B, 3A, 4A, 5, 9, 10A, 11, 12, Table 1, Suppl. materials 1, 2

Terebellides shetlandica Parapar, Moreira & O'Reilly, 2016a: 211–225, figs 1–9, 11. Species 1 – Nygren et al. 2018: 18–22, figs 6, 10.

Material examined. 30 specimens (Suppl. material 1), Skagerrak (GNM14640); Swedish coast (ZMBN116171, ZMBN116181, ZMBN116185, ZMBN116186, ZMBN116187, ZMBN116188, ZMBN116191, ZMBN116192, ZMBN116193, ZMBN116196, ZMBN116198, ZMBN116200, ZMBN116201, ZMBN116202, ZMBN116203, ZMBN116204, ZMBN116206); Norwegian coast (ZMBN116207, ZMBN116208, ZMBN116214, ZMBN116216, ZMBN116219, ZMBN116220, ZMBN116221, ZMBN116226, ZMBN116227, ZMBN116228, ZMBN116235, ZMBN116242).

GenBank accession numbers of material examined (COI). MG024894, MG024895, MG024896, MG024897, MG024898, MG024899, MG024900, MG024901, MG024902, MG024903, MG024904, MG024905, MG024906, MG024907, MG024908, MG024909, MG024910, MG024911, MG024912, MG024913, MG024914, MG024915, MG024916, MG024917, MG024918, MG024919, MG024920, MG024921, MG024922, MG024923, MG024924, MG024925, MG024926, MG024927, MG024928, MG024929, MG024930, MG024931, MG024932, MG024933, MG024934, MG024935, MG024936, MG024937, MG024938, MG024939, MG024940, MG024941, MG024942, MG024943, MG024944, MG024945, MG024946, MG024947, MG024948, MG024949, MG024950, MG024951, MG024952, MG024953, MG024954, MG024955, MG024956.

Diagnostic features of studied material. Complete individuals ranging from 5.0–16.0 mm in length (Fig. 9). Branchial dorsal lobes lamellae provided with well-developed papillary projections and branchial ventral lobes provided with long filaments, ranging from 175.0–225.0 μ m in length (Figs 2A, B, 4A, 5A, B). Between 22–26 lamellae on dorsal lobes (Fig. 5A, B). Lateral lappets present on TC 1–4; dorsal projections of thoracic notopodia on TC 2 and TC 3 (Fig. 5B). Geniculate chaetae in TC 5, acutely bent, with poorly marked capitium (Fig. 5C). Ciliated papilla dorsal to thoracic notopodia not observed. From TC 7, neuropodia with one row of type 4 thoracic uncini per torus, with rostrum/capitium length ratio of ~ 2:1 and capitium with a first row of small teeth, followed by several smaller teeth (Fig. 5D). Abdomen with 25–34 pairs of neuropodia with type 2 uncini (Fig. 5E, F). Copepods attached to body surface in three specimens (Fig. 5B).

Colour pattern. MG staining pattern characterised by compact green colourant in SG 1–6, then turning into striped pattern in SG 7–14 and fading in following segments (Fig. 12). Similar to pattern 1.

Nucleotide diagnostic features. All sequences of *Terebellides shetlandica* share and are distinguished from other available *Terebellides* sequences in unique combinations

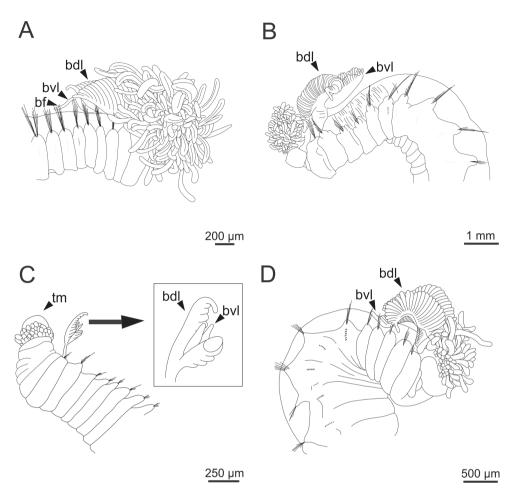


Figure 4. Line drawings of several *Terebellides* species (**A**, **C**, **D** non-type specimens) **A** *Terebellides shetlandica* Parapar, Moreira & O'Reilly, 2016 (species 1; ZMBN116186), anterior end, right lateral view **B** *Terebellides lavesquei* sp. nov. (holotype; ZMBN116322), anterior end, left lateral view **C** *Terebellides irinae* Gagaev, 2009 (species 24; ZMBN116498), anterior end, ventral view **D** *Terebellides gracilis* Malm, 1874 (species 3; ZMBN116283), anterior end, right lateral view. Abbreviations: bdl – branchial dorsal lobe; bf – branchial filament; bvl – branchial ventral lobe; tm – tentacular membrane.

of nucleotides (underlined) at the given position of our alignment: 78–98: CCAAC-CCGGAGCCTATTTAGGT, 186–192: CGGAAAC, 210–219: GCTAGGCGCC, 228–234: GGCATTC, 264–276: TCTCCCGCCTGCC, 288–292: CGTT, 306: C, 333–342: CGTCTACCCT, 351–369: AGACAATATGGCACACGCC, 381–402: AGATCTGGCTATTTCTCCCCTA, 453–459: AGTAATA, 511–522: TCAGC-TATAATC, 535–558: TTACTTCTTCTCTCGCCAGTTCTG.

Type locality. NW Hutton Oilfield, between Shetland Islands and Norway, 61°10'N, 01°12'E (Parapar et al. 2016a).

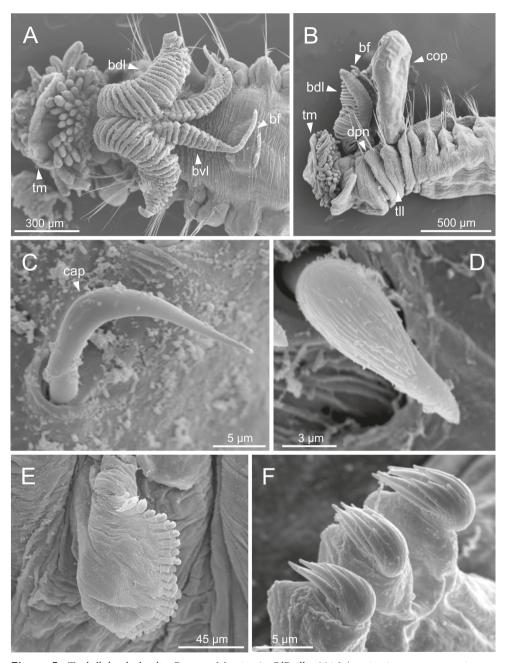


Figure 5. *Terebellides shetlandica* Parapar, Moreira & O'Reilly, 2016 (species 1; non-type specimens, ZMBN116181, ZMBN116204 and ZMBN116219), SEM micrographs **A** anterior end, dorsal view **B** anterior end, left lateral view **C** TC 6 (TU1), geniculate chaeta **D** thoracic uncinus **E** abdominal neuropodium **F** abdominal uncini. Abbreviations: bdl – branchial dorsal lobe; bf – branchial filament; bvl – branchial ventral lobe; cap – capitium; cop – copepod; dpn – dorsal projection of notopodium; TC – thoracic chaetiger; tll – thoracic lateral lobes; tm – tentacular membrane; TU – thoracic unciniger.

Distribution and bathymetry. Norwegian coast and shelf, North Sea, Skagerrak, Kattegat; 25–375 m deep; 92.7% of specimens present at depths below 200 m (Figs 10A, 11, Suppl. material 1).

Remarks. *Terebellides shetlandica* is a small species, reaching up to 16 mm length and is characterised by having branchiae of type 3 and long filaments in ventral branchial lobes, thoracic uncini of type 4, abdominal uncini of type 2 and lacking papillae on margins of branchial lamellae (Table 1). Parapar et al. (2016a) pointed out that *T. atlantis* is the most similar species to *T. shetlandica*; this is confirmed here according to molecular analyses and morphological examination. Both species are small sized (length: *T. shetlandica*, 5–16 mm; *T. atlantis*, 10–16 mm) and have branchiae of type 3, with free branchial lobes. However, the branchiae of *T. shetlandica* have a high number (22–26) of tightly packed branchial lamellae, all lobes are similar in shape and length and ventral ones bear long filaments whereas *T. atlantis* has a fewer number of branchiae (10–11), lamellae are not packed, lobes differ in shape and size and ventral lobes bear shorter filaments. Furthermore, the range of abdominal chaetigers number is higher in *T. shetlandica* than in *T. atlantis* (25–34 vs. 23–28 respectively).

Terebellides lavesquei sp. nov.

https://zoobank.org/2D993190-50A3-42C0-B11E-508EA59276B6 Figs 2C, 3B, 4B, 6, 7, 9, 10B, 11, 12, Table 1, Suppl. materials 1, 2

Species 5 – Nygren et al. 2018: 18–22, figs 6, 10.

Material examined. Type material. *Holotype*: ZMBN116322. *Paratypes* (16 specimens): Skagerrak (GNM15112); Norwegian coast (NTNU–VM61386, NTNU–VM61387, NTNU–VM68252, ZMBN116319, ZMBN116320, ZMBN116321, ZMBN116323, ZMBN116324, ZMBN116325, ZMBN116326, ZMBN116327, ZMBN116328, ZMBN116329, ZMBN116330, ZMBN116331, ZMBN116332).

Holotype. Complete specimen, 34.0 mm long and 2.0 mm wide (Fig. 4B); female with oocytes in body cavity.

GenBank accession numbers of material examined (COI). MG025054, MG025055, MG025056, MG025057, MG025058, MG025059, MG025060, MG025061, MG025062, MG025063, MG025064, MG025065, MG025066, MG025067, MG025068, MG025069, MG025070.

Diagnostic features of type material. Complete individuals ranging from 5.0–35.0 mm in length (Fig. 9). Branchial dorsal lobes lamellae provided with well-developed papillary projections and branchial ventral lobes provided with long filaments, ranging from 125.0–250.0 μ m in length (Fig. 6A). Between 17–42 lamellae on dorsal lobes (Fig. 6A, B). Ciliary rows present on lamellae inner face (Fig. 6B, C). Ventral branchial lobes hidden in between dorsal ones but sometimes discernible below (Fig. 3B). Lateral lappets present on T C1–4; dorsal projection of thoracic notopodia on TC 2–4 (Fig. 3B). Geniculate chaetae in TC 5, acutely bent, with well-defined capitium (Fig. 7A). Ciliated papilla dorsal to

thoracic notopodia not observed. From TC 7, neuropodia with one or two rows of type 3 thoracic uncini per torus, with rostrum/capitium length ratio of ~ 2:1 and capitium with a first row of four or five medium-sized teeth, followed by several smaller teeth (Fig. 7B, C). Abdomen with 30–31 pairs of neuropodia with type 2 uncini (Fig. 7D). Copepods observed attached to body dorsal surface in one specimen (Fig. 6D, E).

Colour pattern. MG staining pattern characterised by compact green colourant in SG 1–6, J-shaped glandular region in SG 3–5 and striped pattern in SG 7–14 (Fig. 12). Similar to pattern 9.

Nucleotide diagnostic features. All sequences of *Terebellides lavesquei* sp. nov. share and are distinguished from other available *Terebellides* sequences in unique combinations of nucleotides (underlined) at the given position of our alignment: 78–99: TCAACCCGGTGCTTACCTCGGT, 156–174: TTTAGTTATGC-CAGTCTTC, 261–264: GTTA, 270–279: TCCAGCACTT, 315–336: AGTTGG-GACCGGTTGAACCGTT, 351–369: AGACAATATAGCTCATGCG, 405–411: CTTGGCT, 426–447: CCTAGGATCAATTAACTTTATC, 459–483: CAACATACGCTGAAAAGGTTTACGA, 510–525: GTCCGCGGGTTATCACA, 534–558: ACTTCTTTATCCCTTCCAGTCTTG, 573–580: CATGCTTC, 606–627: CTTTTTCGACCCAGCTGGTGGG.

Type locality. Hordaland, Lysefjord (Norway), 60°07'N, 05°04'E; 119 m deep.

Distribution and bathymetry. Norwegian coast and shelf, Skagerrak; 115–534 m deep; ~ 50% of specimens collected at depths above 200 m (Figs 10B, 11, Suppl. material 1).

Etymology. This species is dedicated to Nicolas Lavesque, Station Marine d'Arcachon, CNRS (France) for his remarkable recent contributions to the diversity of Terebellidae and Trichobranchidae in Atlantic waters.

Groups Putative species sensu Nygren et al. (2018) Species (as reported/described herein)		В			С	D	
		1	5	16	24	2	3
		T. shetlandica Parapar, Moreira & O'Reilly, 2016	T. lavesquei sp. nov.	<i>T. atlantis</i> Williams, 1984	<i>T. irinae</i> Gagaev, 2009	T. <i>williamsae</i> Jirkov, 1989	<i>T. gracilis</i> Malm, 1874
Branchiae	type (1)	3	2	3	4	2	2
	papillae on lamellae edge	no	no	no	no	no	no
Thorax	ciliated papilla dorsal to notopodium	no (?)	no (?)	no (?)	no (?)	yes	yes
	chaetiger with geniculate chaetae	TC 6	TC 6	TC 6	TC 6	TC 6	TC 6
	uncini type (2)	4	3	3	3	1	1
Abdomen	uncini type (3)	2	2	2	2	1A	1A
Bathymetry – Above (A) / Below (B) 200 m depth (4)		A/B	A/B	В	В	В	В
Distribution – North (N) /South (S) of $60^{\circ}N^{(4)}$		N/S ⁽⁵⁾	N/S	Ν	Ν	N/S ⁽⁵⁾	N/ S ⁽⁵⁾

Table 1. Comparison of discriminating taxonomic characters of the species studied in this work. Cells in italics show discriminatory characters of each subgroup. ⁽¹⁾ sensu Parapar et al. (2016a); ⁽²⁾ sensu Parapar et al. (2020b); ⁽³⁾ sensu Parapar et al. (2020a); ⁽⁴⁾ dominant trend in bold; ⁽⁵⁾ Skagerrak.

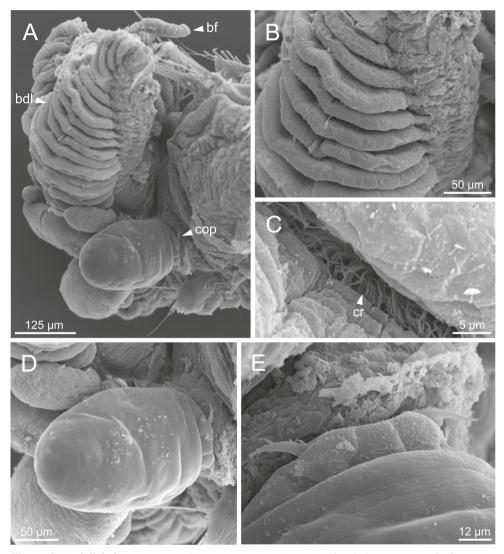


Figure 6. *Terebellides lavesquei* sp. nov. (non-type specimen, ZMBN116332), SEM micrographs **A** anterior end, left lateral view **B** branchial lamellae, detail **C** ciliary row, detail **D** copepod **E** copepod, anterior end. Abbreviations: bdl – branchial dorsal lobe; bf – branchial filament; cop – copepod; cr – ciliary row.

Remarks. *Terebellides lavesquei* sp. nov. is a medium-sized species, reaching up to 35 mm in length. It is characterised by the lack of papillae on margins of branchial lamellae and by having branchiae of type 2, filaments on ventral branchial lobes, thoracic uncini of type 3 and abdominal uncini of type 2 (Table 1). *Terebellides lavesquei* sp. nov. is genetically close to *T. shetlandica* and *T. atlantis* but mostly differs from them regarding branchiae features (Table 1). Lobes are partially fused and have many

tightly packed lamellae (17–42) in comparison with these species. *Terebellides lavesquei* sp. nov. is also similar to *Terebellides parapari* Lavesque, Hutchings, Daffe, Nygren & Londoño-Mesa, 2019 in having filaments in ventral branchial lobes and the presence of glandular regions, but they differ in the branchial morphology, with lobes fused ca. half of their length in *T. lavesquei* sp. nov. and fused only at the base in *T. parapari*. They also differ in TC 1 notochaetae length, being all similar in *T. lavesquei* sp. nov. but longer than those in following chaetigers in *T. parapari*.

Branchial shape of *T. lavesquei* sp. nov. is similar to that of *Terebellides narribri* Hutchings & Peart, 2000, because both lobes are fused to each other for ca. half their length and have a high number of tightly packed lamellae. However, *T. narribri* have thoracic uncini of type 1 whereas *T. lavesquei* sp. nov. have thoracic uncini of type 3. Furthermore, *T. lavesquei* sp. nov. and *T. shetlandica* seem to have a more restricted bathymetric distribution in shallow waters (down to 534 and 375 m, respectively) whereas *T. atlantis* reaches depths of 2750 m (see below).

Terebellides atlantis Williams, 1984

Figs 3C, 8, 9, 10C, 11, 12, Table 1, Suppl. materials 1, 2

Terebellides atlantis Williams, 1984: 121–123, fig. 4, table 1. Species 16 – Nygren et al. 2018: 18–22, figs 6, 10.

Material examined. 15 specimens (Suppl. material 1), Barents Sea (ZMBN116454, ZMBN116455, ZMBN116458, ZMBN116459, ZMBN116460, ZMBN116462, ZMBN116463, ZMBN116465, ZMBN116467, ZMBN116468, ZMBN116470, ZMBN116471, ZMBN116472, ZMBN116474); Norwegian coast (ZMBN116476).

GenBank accession numbers of material examined (COI). MG025258, MG025259, MG025260, MG025261, MG025262, MG025263, MG025264, MG025265, MG025266, MG025267, MG025268, MG025269, MG025270, MG025271, MG025272, MG025273, MG025274, MG025275, MG025276, MG025277, MG025278, MG025279, MG025280, MG025281, MG025282, MG025283, MG025284, MG025285, MG025286, MG025287, MG025288, MG025289, MG025290, MG025291, MG025292, MG025293, MG025294, MG025295, MG025296, MG025297, MG025298, MG025299, MG025300, MG025301, MG025302, MG025303, MG025304, MG025305, MG025306, MG025307, MG025308, MG025309, MG025310, MG025311, MG025312.

Diagnostic features of studied material. Complete individuals ranging from 10.0–16.0 mm in length (Fig. 9). Branchial dorsal lobes lamellae provided with well-developed papillary projections and branchial ventral lobes (Fig. 8A, B) provided with long filaments (sometimes broken), 175.0 µm in length. Between 10–11 lamellae on dorsal lobes. Lateral lappets present on TC 1–4; dorsal projection of thoracic notopodia on TC 2–4 (Fig. 8A). Geniculate chaetae in TC 5, acutely bent, with well-defined capitium (Fig. 8C). Ciliated papilla dorsal to thoracic notopodia not observed. From

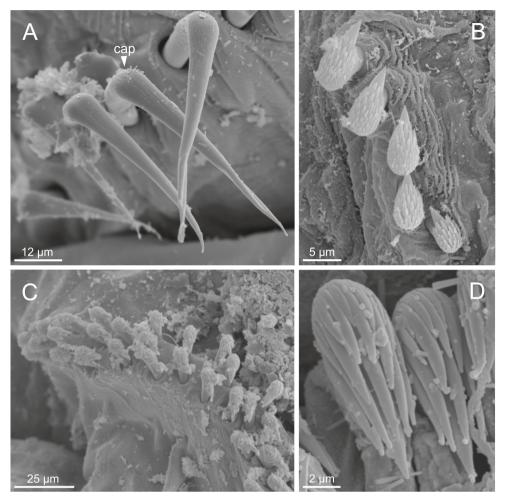


Figure 7. *Terebellides lavesquei* sp. nov. (non-type specimens, NTNU–VM61387 and ZMBN116332), SEM micrographs **A** TC 6 (TU1), geniculate chaetae **B** thoracic uncini **C** double row of thoracic uncini **D** abdominal uncini. Abbreviations: cap – capitium; TC – thoracic chaetiger; TU – thoracic unciniger.

TC 7, neuropodia with one row of type 3 thoracic uncini per torus, with rostrum/ capitium length ratio of ~ 2:1 and capitium with a first row of three or four medium-sized teeth, followed by several smaller teeth (Fig. 8D). Abdomen with 23–28 pairs of neuropodia with type 2 uncini (Fig. 8E, F).

Colour pattern. MG staining pattern characterised by compact green colourant in SG 1–6, J-shaped glandular region in SG 3–5 and striped pattern in SG 7–14 (Fig. 12). Similar to pattern 9.

Nucleotide diagnostic features. All sequences of *Terebellides atlantis* share and are distinguished from other available *Terebellides* sequences in unique combinations of nucleotides (underlined) at the given position of our alignment: 60–84: TATTCGTATTGAGCTAGGGCAACCT, 132–150: ACATGCATTTT-TAATAATC, 171–189: TTTTATTGGTGGATTTGGT, 213–231: GGGAGCTC-CTGATATAGCC, 264–294: ACTACCACCAGCCTTAATCTTATTAGTAAGC, 345–363: ATTATCTGATAATATGGCT, 384–399: CCTTGCTATTTTTTCA, 477–484: GCTACGAC, 549–573: TCCAGTCTTAGCTGGTGCAATCACT, 558– 591: CCGT, 615–630: TCCAGCTGGTGGTGGT.

Type locality. Atlantic Ocean, off New England, 39°56.5'N, 70°39.9'W (Williams 1984).

Distribution and bathymetry. Barents Sea, Greenland Sea, South Iceland, Norwegian coast and shelf; 219–2750 m deep (Figs 10C, 11, Suppl. material 1).

Remarks. *Terebellides atlantis* is a small species, reaching up to 16 mm in length. It is characterised by the lack of papillae on margins of branchial lamellae, and by having branchiae of type 3 and filaments in ventral branchial lobes, thoracic uncini of type 3 and abdominal uncini of type 2 (Table 1). The most similar species to *T. atlantis* are *T. shetlandica* and *T. lavesquei* sp. nov. but *T. atlantis* differs from the latter in the size and type of branchiae (see remarks for *T. lavesquei* sp. nov. above). Branchial lobes are often missing as previously highlighted by Parapar et al. (2011). Finally, *T. atlantis* has the widest geographical distribution and depth range (219 to 2750 m) among Group B species.

Terebellides Group C (sensu Nygren et al. 2018)

Figs 3, 4, 9–13

Description. The morphological features of the examined species in Group C in this paper (clade 24) are itemised below. Some of these are also shared by Groups A, B, and D as defined in Nygren et al. (2018) (see Remarks below). Clade 25 will be studied elsewhere; formal description of clade 22 will wait until more material is available.

Body appearance. Incomplete individuals ranging from 10.0–17.0 mm in length. Body tapering posteriorly with segments increasingly shorter and crowded towards pygidium. Prostomium compact; large tentacular membrane surrounding mouth (Fig. 3D), with typical buccal tentacles with expanded tips (Fig. 3D). SG 1 as an expanded structure below tentacular membrane in a lower lip (Fig. 3D).

Branchiae. Branchiae arising as single structure from SG 3, with a single stalked mid-dorsal stem, lobes not fused (Fig. 4C). Dorsal lobes ending posteriorly in short terminal papilla (Fig. 3D) and ventral lobes ones ending in long filaments. Anterior projection of dorsal lobes (fifth lobe) present. Posterior end of dorsal lobes reaching TC 4. Ciliary rows of cilia and ciliary tufts in inner face of branchial lamellae not observed. Ciliary papillae absent in branchial lamellae margin.

Thorax. Eighteen pairs of notopodia (SG 3–20) (Fig. 3D), those of TC 1 approximately as long as subsequent ones (Fig. 4C). Lateral lappets and dorsal projections of notopodia in anterior thoracic chaetigers with different degree of development depending on size and preservation conditions, but both more conspicuous on

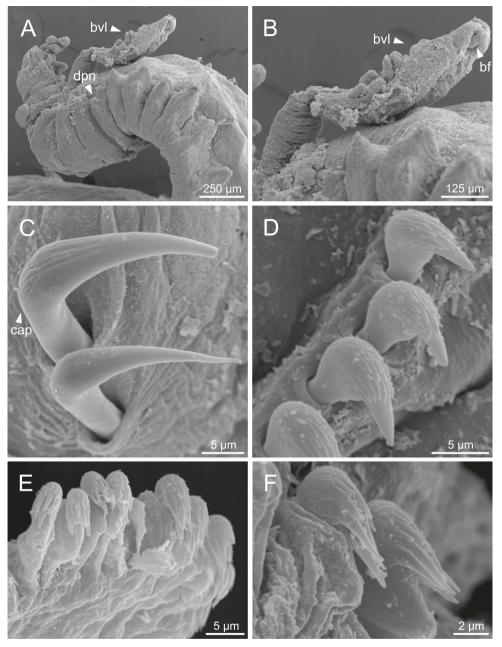


Figure 8. *Terebellides atlantis* Williams, 1984 (species 16; non-type specimens, ZMBN116454 and ZMBN116459), SEM micrographs **A** anterior end, left lateral view **B** branchiae, detail **C** TC 6 (TU1), geniculate chaetae **D** thoracic uncini **E** abdominal neuropodium **F** abdominal uncini. Abbreviations: bf – branchial filament; bvl – branchial ventral lobe; cap – capitium; dpn – dorsal projection of notopodium; TC – thoracic chaetiger; TU – thoracic unciniger.

TC 1–5. All notochaetae as simple capillaries. Size of notochaetae of TC 1 similar to subsequent ones. Neuropodia as sessile pinnules from TC 6 to body end, with uncini in single rows, from TC 7 throughout. Neuropodia on TC 6, provided with several sharply bent, acute-tipped, geniculate chaetae (Fig. 13B) with minute teeth forming a capitium only visible with SEM (Fig. 13B). From TC 7, neuropodia with one row of uncini per torus (Fig. 13C), with long shafted denticulate hooks, with large main fang (rostrum) longer than upper crest of teeth (capitium), rostrum/capitium length ratio of ~ 2:1, capitium composed by several teeth above main fang of decreasing length (Fig. 13D).

Abdomen and pygidium. Approximately half as long as thorax and progressively thinner (Fig. 3D). Neuropodia ranging from 18–20 chaetigers and forming erect pinnules with several uncini per torus, number depending on specimen size. Uncini provided with several teeth above rostrum surmounted by a capitium composed of several teeth of decreasing length (Fig. 13E, F). Pygidium blunt, as funnel-like depression.

Colour pattern. Colour in preserved specimens whitish (Fig. 3D). MG staining pattern characterised by compact green colourant in SG 1–4, then turning into striped pattern in SG 5–14 and fading in following segments (Fig. 12).

Remarks. Among the above-mentioned characters, branchial features might serve to distinguish most of Group C species from those of Groups A, B and D. Those include branchial lobes size, presence of filaments in ventral ones and lobes which are not fused. Other taxa such as *Terebellides mira* Schüller & Hutchings, 2013 and *T. rigel* Schüller & Hutchings, 2013 also bear branchiae with similar shape (Parapar et al. 2016a).

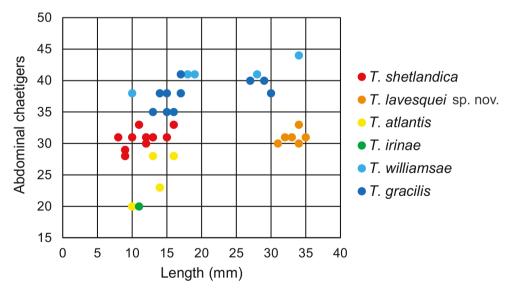


Figure 9. Relationship between number of abdominal chaetigers and body length (complete specimens considered except for *T. irinae*).

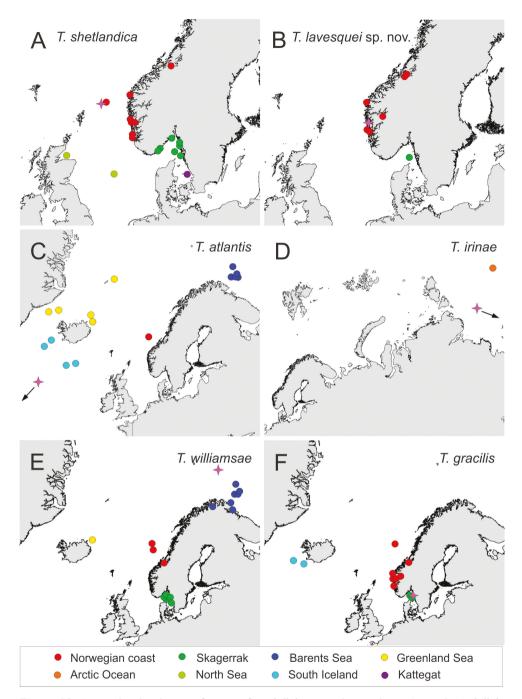


Figure 10. Geographic distribution of species of *Terebellides* in Northeast Atlantic Ocean **A** *Terebellides shetlandica* Parapar, Moreira & O'Reilly, 2016 **B** *Terebellides lavesquei* sp. nov. **C** *Terebellides atlantis* Williams, 1984 **D** *Terebellides irinae* Gagaev, 2009 **E** *Terebellides williamsae* Jirkov, 1989 **F** *Terebellides gracilis* Malm, 1874. Pink star denotes the type locality of each taxon.

Terebellides irinae Gagaev, 2009

Figs 3D, 4C, 9, 10D, 11-13, Table 1, Suppl. materials 1, 2

Terebellides irinae Gagaev, 2009: 474–478. Species 24 – Nygren et al. 2018: 18–22, figs 6, 10.

Material examined. 6 specimens (Suppl. material 1), Arctic Ocean (ZMBN116496, ZMBN116497, ZMBN116498, ZMBN116499, ZMBN116500, ZMBN116501).

GenBank accession numbers of material examined (COI). MG025340, MG025341, MG025342, MG025343, MG025344.

Diagnostic features of studied material. Incomplete individuals ranging from 10.0–17.0 mm in length (Fig. 9). Branchial dorsal lobes provided with filaments, 75.0 μ m in length (Fig. 3D) and branchial ventral lobes reduced, distinctly smaller than dorsal ones (Fig. 4C). Dorsal lobes provided with seven lamellae (Fig. 4C). Lateral lappets present on TC 1–4; dorsal projection of thoracic notopodia on TC 2–5 (Fig. 3D). Geniculate chaetae in TC 5, acutely bent and provided with hardly distinguishable capitium (Fig. 13B). Ciliated papilla dorsal to thoracic notopodia not observed. From TC 7, neuropodia with one row of type 3 thoracic uncini per torus, with rostrum/capitium length ratio of ~ 2:1 and capitium with a first row of four or five medium-sized teeth, followed by several smaller teeth (Fig. 13C, D). Abdomen with at least 20 pairs of neuropodia with type 2 uncini (Fig. 13E, F).

Colour pattern. MG staining pattern characterised by compact green colourant in SG 1–4, then turning into striped pattern in SG 5–14 and fading in following segments (Fig. 12). Similar to pattern 1.

Nucleotide diagnostic features. All sequences of *Terebellides irinae* share and are distinguished from other available *Terebellides* sequences in unique combinations of nucleotides (underlined) at the given position of our alignment: 177–204: CGGGGGGTTTGGAAACTGGTTAATCCCC, 213–225: TGGGGC-CCCAGAC, 249–258: CATAAGGTTC, 273–303: GGCCCTCATCCTAC-TAGTCAGCTCAGCTGCT, 305–321: GGCTGGT, 327–336: ATGAACTGTA, 342–372: ACCACTTTCAGACAACATCGCTCATGCCGGA, 381–399: AG-ATCTAGCAATTTTCTCA, 426: CCTAGGTTCTATTAACTTCATCACAACA-GTC, 483–499: TCTAGAACGAATCCCAC, 535–573: TTATTACTATCACTAC-CAGTGCTAGCCGGAGCTATTACC, 594–612: CATTAACACATCATTCTC, 618–636: AGCCGGTGGTGGTGATCCT.

Type locality. Arctic Ocean, 73°04'N, 157°12'W (Gagaev 2009).

Distribution and bathymetry. Arctic Ocean; 4038–4380 m deep (Figs 10D, 11, Suppl. material 1).

Remarks. *Terebellides irinae* is a small species, reaching up to 17 mm in length and is characterised by the lack of papillae on margins of branchial lamellae, and by having branchiae of type 4, filaments in ventral branchial lobes, thoracic uncini of type 3 and abdominal uncini of type 2 (Table 1). Jirkov and Leontovich (2013) proposed

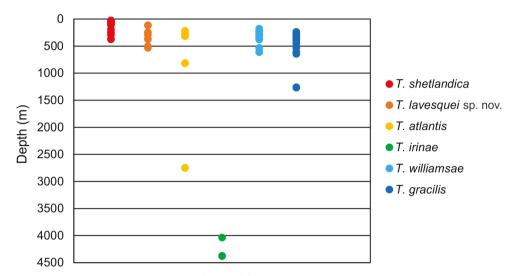


Figure 11. Bathymetric distribution of *Terebellides* species studied in this work.

T. irinae as synonym of *T. stroemii* because it fit within the variability of the latter. However, Parapar and Hutchings (2014) redescribed *T. stroemii* designating a neotype and *T. irinae* not fit in this concept. Later, Nygren et al. (2018) recognised *T. irinae* as different from *T. stroemii* after molecular analyses and pointed out that *T. irinae* is the only species present in the Arctic Ocean at depths below 4000 m (Fig. 11). Furthermore, *T. irinae* is the only species in Northeast Atlantic Ocean bearing branchiae of type 4 and therefore is also considered as a valid species in this work. Other taxa from elsewhere such as *T. mira* and *T. rigel* also bear the same branchial type, these two species have branchial lobes free from each other with few numbers of not packed lamellae and ventral lobes are also distinctly smaller than the dorsal ones.

Terebellides Group D (sensu Nygren et al. 2018)

Figs 2-4, 9-18

Description. The morphological features shared by all examined species in Group D in this paper (clades 2 and 3) are itemized below. Some of these are also shared by Groups A, B, and C as defined in Nygren et al. (2018) (see Remarks below). Clade 15 will be studied elsewhere.

Body appearance. Complete individuals ranging from 5.0–34.0 mm in length. Body tapering posteriorly with segments increasingly shorter and crowded towards pygidium. Prostomium compact; large tentacular membrane surrounding mouth (Figs 2D–F, 3E, F), with typical buccal tentacles with expanded tips (Figs 2E, F, 3E, F). SG 1 as an expanded structure below tentacular membrane in a lower lip (Figs 2D, E, 3E, F).

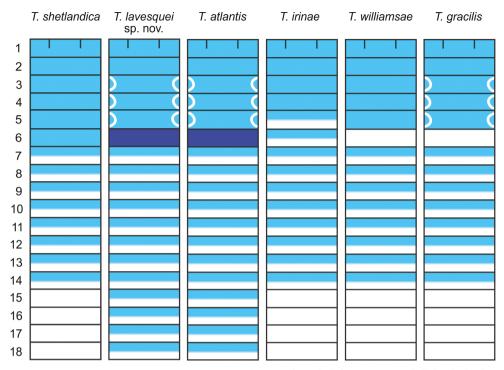


Figure 12. Body MG staining patterns in ventral view of *Terebellides* species. *Terebellides shetlandica* Parapar, Moreira & O'Reilly, 2016, *Terebellides lavesquei* sp. nov., *Terebellides atlantis* Williams, 1984, *Terebellides irinae* Gagaev, 2009, *Terebellides williamsae* Jirkov, 1989 and *Terebellides gracilis* Malm, 1874. Segments indicated in Arabic numbers.

Branchiae. Branchiae arising as single structure from SG 3, with a single stalked mid-dorsal stem (Figs 2D, E, 3E, F), one pair of dorsal (upper) partially fused lobes (Figs 2D, E, 3E, F), and a pair of shorter ventral (lower) lobes (Fig. 3E, F) obscured or not by dorsal ones (Figs 2D–F, 3E, F). Dorsal lobes ending posteriorly in short terminal papilla (Fig. 3E, F) and ventral lobes ending in long filaments. Anterior projection of dorsal lobes (fifth lobe) present (Fig. 2D–F). Posterior end of dorsal lobes reaching TC 4–5 (Figs 2D–F, 3E, F). Branchial lamellae provided with several parallel rows of cilia and ciliary tufts present in inner face (Figs 14B, C, 16B, C, 17B). Ciliary papillae absent on the margin of branchial lamellae.

Thorax. Eighteen pairs of notopodia (SG 3–20) (Fig. 2D, E), those of TC 1 approximately as long as subsequent ones (Fig. 2D, E). Lateral lappets and dorsal projections of notopodia in anterior thoracic chaetigers with different degree of development depending on size and preservation conditions, but both more conspicuous on TC 1–5 (Figs 2D–F, 3E, F). White ventral colouration present on TC 1–4 (Figs 2D, 3E) or only on TC 4 (Figs 2E, F, 3F). All notochaetae as simple capillaries (Fig. 15A). Size of notochaetae of TC 1 similar to subsequent ones. Neuropodia as sessile pinnules from TC 6 to body end, with uncini in single or double rows, from TC 7 throughout. Neuropodia on TC 6, provided with several sharply bent, acute-tipped, geniculate

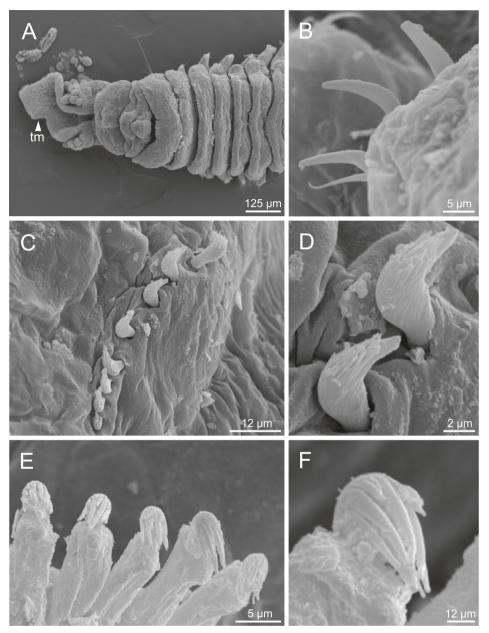


Figure 13. *Terebellides irinae* Gagaev, 2009 (species 24; non-type specimen, ZMBN116501), SEM micrographs. **A** anterior end, ventral view **B** TC 6 (TU1), geniculate chaetae **C** row of thoracic uncini **D** thoracic uncini **E** abdominal uncini **F** abdominal uncinus, detail. Abbreviations: tc – thoracic chaetiger; tm – tentacular membrane; tu – thoracic unciniger.

chaetae (Figs 15B, 18A) with minute teeth forming a capitium only visible with SEM (Fig. 18A, B). From TC 7, neuropodia with one row of uncini per torus (Figs 15C, 18C), with long shafted denticulate hooks, with large main fang (rostrum) longer than

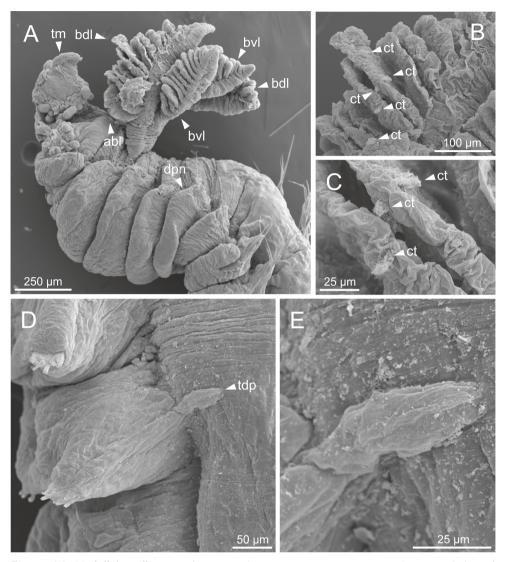


Figure 14. *Terebellides williamsae* Jirkov, 1989 (species 2; non-type specimens, ZMBN116249 and ZMBN116251), SEM micrographs **A** anterior end, left lateral view **B** branchial lamellae, detail **C** ciliary tufts, detail **D** TC and thoracic dorsal papilla **E** thoracic dorsal papilla, detail. Abbreviations: abl – anterior branchial lobe; bdl – branchial dorsal lobe; bvl – branchial ventral lobe; ct – ciliary tuft; dpn – dorsal projection of notopodium; TC – thoracic chaetiger; tdp – thoracic dorsal papilla; tm – tentacular membrane.

upper crest of teeth (capitium), rostrum/capitium length ratio of ~ 2:1, capitium composed by several teeth above main fang of decreasing length (Figs 15D, 18D).

Abdomen and pygidium. Approximately half as long as thorax and progressively thinner (Fig. 2D, E). Neuropodia ranging from 18–44 chaetigers and forming erect pinnules (Figs 15E, 18E) with several uncini per torus, number depending on specimen size. Uncini provided with several teeth above rostrum surmounted by a capitium

composed of several teeth of decreasing length (Figs 15F, 18F). Pygidium blunt, as funnel-like depression.

Colour pattern. Colour in preserved specimens whitish or pale brown (Fig. 3E, F). MG staining pattern characterised by compact green colourant in SG 1–5 and SG 7–13, SG 6 white and SG 14 striped (Fig. 12).

Remarks. Among the aforementioned characters, the white ventral colouration in anterior thoracic chaetigers may be a useful character to distinguish Group D species from those of Groups A–C. Other taxa described or reported worldwide showing this colouration pattern are *Terebellides distincta* Williams, 1984 and *T. ceneresi* Lavesque, Hutchings, Daffe, Nygren & Londoño-Mesa, 2019.

Terebellides williamsae Jirkov, 1989

Figs 2D, 3E, 9, 10E, 11, 12, 14, 15, Table 1, Suppl. materials 1, 2

Terebellides williamsae Jirkov, 1989: 124. Species 2 – Nygren et al. 2018: 18–22, figs 6, 10.

Material examined. 20 specimens (Suppl. material 1), Skagerrak (GNM14639, GNM15107, GNM15108); Barents Sea (ZMBN116246, ZMBN116247, ZMBN116248, ZMBN116249, ZMBN116251, ZMBN116252, ZMBN116253, ZMBN116254, ZMBN116255, ZMBN116257, ZMBN116260, ZMBN116262, ZMBN116263, ZMBN116266, ZMBN116269, ZMBN116270, ZMBN116271).

GenBank accession numbers of material examined (COI). MG024957, MG024958, MG024959, MG024960, MG024961, MG024962, MG024963, MG024964, MG024965, MG024966, MG024967, MG024968, MG024969, MG024970, MG024971, MG024972, MG024973, MG024974, MG024975, MG024976, MG024977, MG024978, MG024979, MG024980, MG024981, MG024982, MG024983, MG024984, MG024985, MG024986, MG024987, MG024988.

Diagnostic features of studied material. Complete individuals ranging from 9.0–34.0 mm in length (Fig. 9). Branchial dorsal lobes lamellae provided with well-developed papillary projections and branchial ventral lobes provided with short posterior filaments, 50.0 µm in length (Figs 3E, 14A). Between 16–18 lamellae on dorsal lobes (Fig. 14A, B). Ciliary tufts present in inner face of lamellae (Fig. 14B, C). Ventral branchial lobes hidden in between dorsal ones but sometimes discernible below (Fig. 14A). Lateral lappets present on TC 1–4; dorsal projection of thoracic notopodia on TC 2–4 (Fig. 14A). White ventral colouration present on TC 1–4 (Figs 2D, 3E). Geniculate chaetae in TC 5, acutely bent, with well-marked capitium (Fig. 15B). Ciliated papilla dorsal to thoracic notopodia observed in TC 7 (Fig. 14D, E). From TC 7, neuropodia with one row of type 1 thoracic uncini per torus, with rostrum/capitium length ratio of ~ 2:1 and capitium with a first row of two or three large teeth, followed by many smaller teeth (Fig. 15C, D). Abdomen with 38–44 pairs of neuropodia with type 1A uncini (Fig. 15E, F).

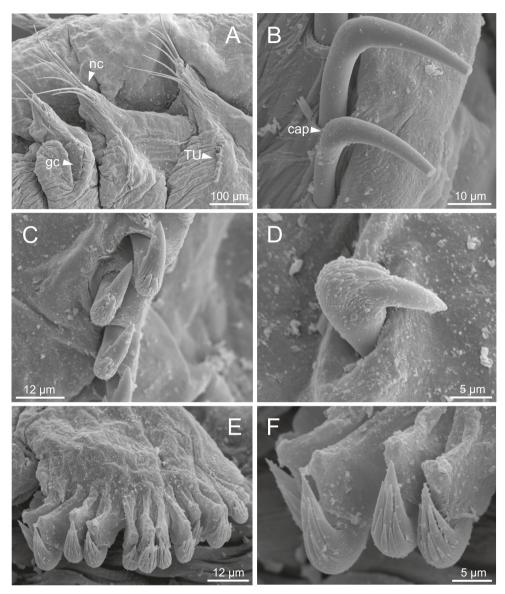


Figure 15. *Terebellides williamsae* Jirkov, 1989 (species 2; non-type specimen, ZMBN116249), SEM micrographs **A** TC 5–7, lateral view **B** TC 6 (TU1), geniculate chaetae **C** row of thoracic uncini **D** thoracic uncinus **E** abdominal neuropodium **F** abdominal uncini. Abbreviations: cap – capitium; gc – geniculate chaetae; nc – notochaetae; TC – thoracic chaetiger; TU – thoracic unciniger.

Colour pattern. MG staining pattern characterised by compact green colourant in SG 1–5 and SG 7–13, SG 6 white and SG 14 striped, J-shaped glandular regions in SG 3–5 (Fig. 12). Similar to pattern 2.

Nucleotide diagnostic features. All sequences of *Terebellides williamsae* share and are distinguished from other available *Terebellides* sequences in unique combinations of nucleotides (underlined) at the given position of our alignment: 59–62: TATC,

75–96: TGGACAACCTGGGGCATTCCTG, 132–144: TCATGCTTTTTA, 153–157: TTTCC, 216–234: TGCTCCTGATATAGCTTTC, 264–277: CCTCC-CTCCAGCTT, 315–318: GGTT, 327–342: CTGAACAGTATACCCC, 381–399: AGATTTGGCTATTTTTTCT, 414–432: TATCTCCTCTATTCTTGGC, 450–454: TACA, 515–529: AAAAATCACTACCA, 543–573: TTCACTTCCTGTATT-AGCAGGAGCTATTACA, 600–609: CACTTCCTTT, 630–640: CGACCCAATTT.

Type locality. Barents Sea, Norway, 74°30'N, 28°00'E (Jirkov 1989).

Distribution and bathymetry. Barents Sea, Greenland Sea, Norwegian coast and shelf, Skagerrak; at depths of 178–612 m but most of the specimens (97%) were collected above 200 m (Figs 10E, 11, Suppl. material 1).

Remarks. *Terebellides williamsae* is a medium-sized species, reaching up to 34 mm in length; it is characterised by the lack of papillae on margins of branchial lamellae and by having branchiae of type 2 and posterior filaments in ventral branchial lobes, thoracic uncini of type 1 and abdominal uncini of type 1A (Table 1). All these features are shared with *T. gracilis*; in fact, Parapar et al. (2011) suggested this species as a synonym to *T. gracilis* after examining specimens from Iceland. Nygren et al. (2018) pointed out that there were no morphological differences between both species, but their molecular analyses indicate that specimens from the Barents Sea ("Species 2") would correspond to *T. williamsae*. Nygren et al. (2018) suggested therefore that *T. williamsae* might be a valid species and different from *T. gracilis* ("Species 3", see below). Here, examination of specimens of *T. williamsae* show that they differ from *T. gracilis* in the number of chaetigers with white ventral colouration, i.e., in *T. williamsae* white colouration is present in TC 1–4 while in *T. gracilis* it is only present on TC 4.

Terebellides gracilis Malm, 1874

Figs 2E, F, 3F, 4D, 9, 10F, 11, 12, 16–18, Table 1, Suppl. materials 1, 2

Terebellides gracilis Malm, 1874: 67–105, p. 100. Species 3 – Nygren et al. 2018: 18–22, figs 6, 10.

Material examined. 20 specimens (Suppl. material 1), Skagerrak (GNM15110, GNM15111); Norwegian coast (ZMBN116276, ZMBN116278, ZMBN116282, ZMBN116283, ZMBN116284, ZMBN116285, ZMBN116287, ZMBN116289, ZMBN116293, ZMBN116295, ZMBN116297, ZMBN116298, ZMBN116301, ZMBN116306, ZMBN116307, ZMBN116309, ZMBN116310, 116313).

GenBank accession numbers of material examined (COI). MG024583, MG024584, MG024585, MG024586, MG024587, MG024588, MG024589, MG024590, MG024591, MG024592, MG024593, MG024594, MG024595, MG024596, MG024597, MG024598, MG024599, MG024600, MG024601, MG024602, MG024603, MG024604, MG024605, MG024606, MG024607, MG024608, MG024609, MG024610, MG024611, MG024612, MG024613, MG024614, MG024615, MG024616, MG024617, MG024618, MG024619, MG024620, MG024621, MG024622, MG024623, MG024624, MG024625,

MG024626, MG024627, MG024628, MG024629, MG024630, MG024631, MG024632, MG024633, MG024634, MG024635, MG024636, MG024637.

Diagnostic features of studied material. Complete individuals ranging from 5.0–29.0 mm in length (Fig. 9). Branchial dorsal lobes lamellae provided with well-developed papillary projections and branchial ventral lobes provided with long posterior filaments, ranging from 125.0–175.0 μ m in length (Fig. 16D, E). Between 23–32 lamellae on dorsal lobes (Figs 4C, 16A, D, E, 17A). Ciliary rows and ciliary tufts on inner branchial lamellae present (Figs 16B, C, 17B). Ventral branchial lobes hidden in between dorsal ones but sometimes discernible below (Figs 16A, D, 17A). Lateral lappets present on TC 1–4; dorsal projection of thoracic notopodia on TC 1–5 (Fig. 16D). White ventral colouration presents only on TC 4 (Figs 2E, F, 3F). Geniculate chaetae present in TC 5,

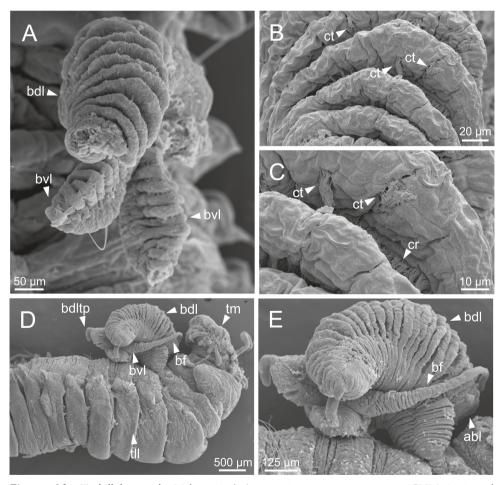


Figure 16. *Terebellides gracilis* Malm, 1874 (species 3; non-type specimens, GNM15110 and ZMBN116313), SEM micrographs **A** branchiae, dorsal view **B** branchial lamellae, detail **C** ciliary tufts, detail **D** anterior end, right lateral view **E** branchiae, lateral view. Abbreviations: abl – anterior branchial lobe; bdl – branchial dorsal lobe; bdlp – branchial dorsal lobe terminal papilla; bf – filament; bvl – branchial ventral lobe; cr – ciliary row; ct – ciliary tuft; tll – thoracic lateral lobes; tm – tentacular membrane.

acutely bent, with marked capitium (Fig. 18A, B). Ciliated papilla dorsal to thoracic notopodia observed in TC 2–4 (Fig. 17A, C, D). From TC 7, neuropodia with one row of type 1 thoracic uncini per torus, with rostrum/capitium length ratio of ~ 2:1 and capitium with a first row of two or three large teeth, followed by many smaller teeth (Fig. 18C, D). Abdomen with 34–41 pairs of neuropodia with type 1A uncini (Fig. 18E, F).

Colour pattern. MG staining characterised by compact green colourant in SG 1–5 and SG 7–13, SG 6 white and SG 14 striped (Fig. 12). Similar to pattern 2.

Nucleotide diagnostic features. All sequences of *Terebellides gracilis* share and are distinguished from other available *Terebellides* sequences in unique combinations of nucleotides (underlined) at the given position of our alignment: 39–63: TG-GTACTTCAATAAGACTTCTTATC, 84–96: TGGGGCATTCCTG, 111–132: TTATAACACAAATTGTTACTGCT, 138–157: TTTTTTAATAATTTTTTTTCC, 216–234: TGCTCCTGATATAGCTTTC, 264–277: CCTCCCTCCAGCTT, 315–327: AGCTGGGACAGGT, 333–351: AGTCTACCCTCCTTTATCT, 381–399: AGATTTGGCTATTTTTTCT, 414–432: TATCTCCTCTATTCTTGGC, 450–545: TACA, 516–529: AAAAATCACTACCA, 543–552: TTCACTTCCT, 600–609: CACTTCCTTT, 630–640: CGACCCAATTT.

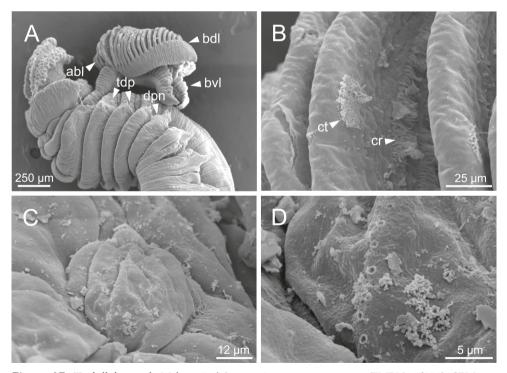


Figure 17. *Terebellides gracilis* Malm, 1874 (species 3; non-type specimen, ZMBN116282), SEM micrographs **A** anterior end, left lateral view **B** ciliary tufts and ciliary row, detail **C** dorsal projection of notopodium **D** pores of dorsal projection of notopodium, detail. Abbreviations: abl – anterior branchial lobe; bdl – branchial dorsal lobe; bvl – branchial ventral lobe; cr – ciliary row; ct – ciliary tuft; dpn – dorsal projection of notopodium; tdp – thoracic dorsal papilla.

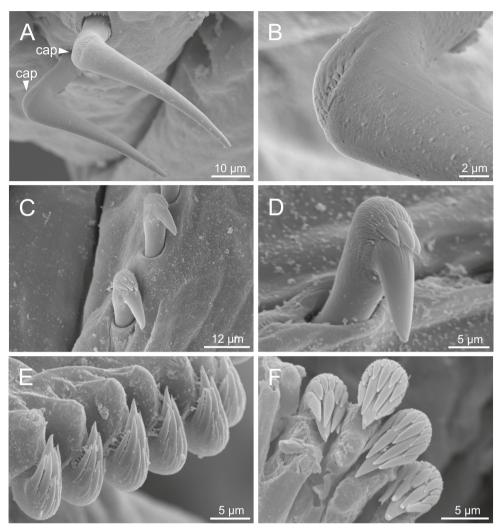


Figure 18. *Terebellides gracilis* Malm, 1874 (species 3; non-type specimens, ZMBN116282 and ZMBN116313), SEM micrographs **A** TC 6 (TU1), geniculate chaetae **B** capitium of geniculate chaeta, detail **C** thoracic uncini **D** thoracic uncinus **E** abdominal neuropodium **F** abdominal uncini. Abbreviations: cap – capitium; TC – thoracic chaetiger; TU – thoracic unciniger.

Type locality. Atlantic Ocean, Norway (Malm 1874).

Distribution and bathymetry. South Iceland, Norwegian coast and shelf, Skagerrak; 237–1268 m deep (Figs 10F, 11, Suppl. material 1).

Remarks. *Terebellides gracilis* is a medium-sized species, reaching up to 29 mm in length and is characterised by the lack of papillae on margins of branchial lamellae, having branchiae of type 2 and filaments in ventral branchial lobes, presence of thoracic uncini of type 1 and abdominal uncini of type 1A (Table 1). As stated above, these features are shared with *T. williamsae* but both species differ in the pattern of white ventral thoracic colouration. Besides, they show a MG pattern close to type 2 but only *T. gracilis* showed J-shaped glandular regions in SG 3–5 as observed in the specimens studied here. *Terebellides gracilis* has apparently a more restricted geographical distribution than *T. williamsae* but reaching deeper depths (down to 1268 m).

Key to Northeast Atlantic Ocean species of Terebellides

The following key of European species of *Terebellides* is based on those by Lavesque et al. (2019) and Parapar et al. (2020a) but has been updated to include the species belonging to Groups B, C and D studied herein. The order of the presentation of the discriminating characters and the taxa has been changed to fit better with the clades recovered in the phylogenetic trees by Nygren et al. (2018) and Lavesque et al. (2019).

The characters considered were the ventral pigmentation of anterior thoracic chaetigers in live and fixed specimens, types of thoracic uncini (sensu Parapar et al. 2020b), morphology of branchiae (sensu Parapar et al. 2016a), morphology of the abdominal uncini (sensu Parapar et al. 2020a), the size of species (small species: < 20 mm in length; medium: 20–40 mm; large: > 40 mm), the presence of geniculate chaetae in TC 5–6 or only in TC 6, the presence or absence of papillae in branchial lamellae margins, the shape of glandular region in TC 3, and the presence or absence of ciliary tufts in branchial lamellae. In those cases where two species are considered as cryptic and only distinguished by molecular characters, geographic and bathymetric distribution has been provided instead.

1	White ventral colouration on anterior thoracic chaetigers
_	No distinct ventral colouration on anterior thoracic chaetigers4
2	Medium/large species (>20 mm in length); 5 th branchial lobe present; no- tochaetae of TC 1 similar to subsequent ones; main fang of thoracic uncini
	straight; thoracic uncini with capitium composed of 2-3 large teeth and sub-
	sequent ones much smaller
-	Small species (< 20 mm in length); 5 th branchial lobe absent; notochaetae of
	TC 1 absent or shorter than subsequent ones; thoracic uncini with capitium
	composed of 4 or 5 mid-sized teeth and following of slightly smaller teeth
	T. ceneresi Lavesque, Hutchings, Daffe, Nygren & Londoño-Mesa, 2019
3	White ventral colouration on TC 1 to TC 4 T. williamsae Jirkov, 1989
_	White ventral colouration only on TC 4 T. gracilis Malm, 1874
4	Branchial lobes all small and not fused; reduced dorsal lobes
	T. irinae Gagaev, 2009
_	Branchiae otherwise
5	Lower branchial lobes with posterior projections as filaments; branchiae with
	lobes fused ~ 50% of their length or with lobes only fused at base; small/
	medium species (<40 mm in length)
_	Lower branchial lobes with posterior projections; branchiae with large lobes
	almost completely fused; large species (> 40 mm in length)9

6	Thoracic uncini with capitium composed of 5–7 small teeth, remaining ones
	similar in size at least in two rows
	<i>T. shetlandica</i> Parapar, Moreira & O'Reilly, 2016
_	Thoracic uncini with capitium composed of 4-5 mid-sized teeth and fol-
	lowed by slightly smaller teeth7
7	Branchiae with lobes fused ~ 50% of their length; medium-sized species
	(> 20 mm in length)
_	Branchiae with lobes only fused at base; small species (< 20 mm in length) 8
8	Glandular region in TC 3 present; notochaetae from TC 1 longer than
	subsequent ones
	T. parapari Lavesque, Hutchings, Daffe, Nygren & Londoño-Mesa, 2019
_	Glandular region in TC 3 not observed; all notochaetae of similar size
	<i>T. atlantis</i> Williams, 1984
9	Geniculate chaetae in TC 5 and TC 6; abdominal uncini with $RvC = 1/0.7$,
	capitium with 4–5 teeth and remaining ones smaller
	<i>T. bigeniculatus</i> Parapar, Moreira & Helgason, 2011
-	Geniculate chaetae in TC 6 only10
10	Branchial lamellae margins lacking papillae11
_	Branchial lamellae margins with papillae13
11	Branchiae with lobes fused ~ 50% of their length
	T. gralli Lavesque, Hutchings, Daffe, Nygren & Londoño-Mesa, 2019
_	Branchiae with large lobes almost completely fused12
12	Abdominal uncini with $RvC = 1/0.7$, capitium with 4–5 teeth and remaining
	ones smaller
_	Abdominal uncini with $RvC = 1/0.9$, capitium composed of 3–5 large teeth in
	first row and 1–2 in a second row <i>T. kongsrudi</i> Parapar, Capa, Nygren &
	Moreira, 2020 and <i>T. bakkeni</i> Parapar, Capa, Nygren & Moreira, 2020
13	Glandular region in TC 3 round or oval
_	Glandular region in TC 3 otherwise
14	Glandular region in TC 3 remained white with MG; branchial lamellae with
	rounded papillae; TC 1–3 without conspicuous dorsal projection
	T. lilasae Lavesque, Hutchings, Daffe, Nygren & Londoño-Mesa, 2019
_	Glandular region in TC 3 stained blue with MG; branchial lamellae with
	conical papillae; TC 1–3 with conspicuous dorsal projection
1.5	T. bonifi Lavesque, Hutchings, Daffe, Nygren & Londoño-Mesa, 2019
15	Branchial ciliary tufts present
	T. gentili Lavesque, Hutchings, Daffe, Nygren & Londoño-Mesa, 2019
-	Branchial ciliary tufts absent
16	Most branchial lamellae with marginal papillae; mouth with upper lip elongated
	<i>T. resomari</i> Lavesque, Hutchings, Daffe, Nygren & Londoño-Mesa, 2019
_	Only anterior branchial lamellae with marginal papillae; upper lip not
	elongated17

17	Thoracic uncini with capitium composed of 2-3 large teeth and subsequent
	ones much smaller T. ronningae Parapar, Capa, Nygren & Moreira, 2020
_	Thoracic uncini with capitium composed of 4 or 5 mid-sized teeth and fol-
	lowing slightly smaller ones
18	Deep-water species; usually at depths below 200 m
_	Shallow-water species; mostly at depths above 100 m
	doño-Mesa, 2019 and T. scotica Parapar, Capa, Nygren & Moreira, 2020

Discussion

Species groups

According to Nygren et al. (2018), *Terebellides* is divided in four main groups of species: A, B, C and D, which differ from each other by several morphological characters. Species of Group A were studied by Parapar et al. (2020a). Species in Group B are characterised by small-medium body length (5.0–35.0 mm), branchiae of type 2 or 3 with incompletely fused or free lobes (only fused at their base), long posterior filaments in ventral lobes, thoracic uncini of type 3 or 4 and abdominal uncini of type 2. *Terebellides lavesquei* sp. nov. belongs to this group and is also characterised by the lack of papillae on margins of branchial lamellae and by having branchiae of type 2 and thoracic uncini of type 3. Group C is defined by having thoracic uncini of type 3 only and abdominal uncini of type 2. Finally, Group D species are of medium length and bear white ventral colouration in anterior chaetigers, branchiae of type 2 with incompletely fused lobes, ventral branchial lobes with posterior filaments, ciliary tufts in the inner face of branchial lamellae, thoracic uncini of type 1 and abdominal uncini of type 1A.

Integrative taxonomy

The apparently morphological homogeneity of members of *Terebellides* has hidden an unexpected species richness in the Northeast Atlantic. Species delimitation analyses of DNA sequence data have allowed to reveal some of the *Terebellides* species that otherwise would have gone unnoticed (Nygren et al. 2018; Lavesque et al. 2019; Parapar et al. 2020a). Moreover, molecular data have provided further evidence of species hypothesis diagnosed solely based on morphological features (e.g., Gagaev 2009; Parapar et al. 2011, 2016c; Parapar and Hutchings 2014).

An integrative approach, that aims at considering different sources of evidence, has become a common and grounded method for general species delineation (Dayrat 2005; Schlick-Steiner et al. 2010) including marine annelids (Capa et al. 2010, 2013; Capa and Murray 2015; Aguado et al. 2019; Kara et al. 2020; Teixeira et al. 2020). In

addition, the integration of a variety of types of data has been used in formal species descriptions, a crucial step that includes providing a new name and facilitates communication about these entities (Goldstein and DeSalle 2011) and their diagnoses, which allows the correct identification for the species (Pante et al. 2015). In fact, the inclusion of DNA sequence information in formal species descriptions or diagnoses has been revealed as a useful practice to identify taxonomic groups (Renner 2016), especially with a high proportion of cryptic and pseudo-cryptic species such as it happens in annelids (Halt et al. 2009; Nygren and Pleijel 2011; Parapar et al. 2020a; Grosse et al. 2021).

In the present study, following the aims and methods of the similar previous study by Parapar et al. (2020a), who dealt with species belonging to Group A, several of the Northeast Atlantic Ocean *Terebellides* molecular lineages recovered within Groups B, C and D and compatible with a hypothetical species rank (after Nygren et al. 2018) are newly described, including morphological and COI sequence traits allowing to distinguish them from other congeners. In addition, a new species, *Terebellides lavesquei* sp. nov. is also described following the same approach. The difference between the present work and the previous (Parapar et al. 2020a) relies on the recognition of the diagnostic COI nucleotides for the species described. Parapar et al. (2020a) provided a list of unequivocal single nucleotides (autapomorphies) in specific positions of the alignment while in the present study a short sequence of nucleotides is provided to ease the identification along the alignment, and these include a unique combination of single nucleotides together with others that do not show variation within the sequences available.

Group B comprises eight species; one of them was identified herein as *T. atlantis*, matching the diagnostic characters and distribution of *T. atlantis*, originally described by Williams (1984) and from specimens collected in deep Icelandic waters by Parapar et al. (2011). A second species was recognised as *T. shetlandica* according to the description by Parapar et al. (2016a). The remaining six species represent undescribed taxa that will be dealt with elsewhere. Group C was composed by three species; one species was identified as *T. irinae* and the other two represent undescribed taxa that will be dealt with elsewhere. Finally, Group D comprises three species: *T. gracilis*, that matches the diagnostic characters and distribution originally described by Malm (1874), *T. williamsae* following the original description by Jirkov (1989), and one undescribed taxon that will be dealt with elsewhere.

Species distributions

The range of distribution of all nominal species identified here is expanded. Nygren et al. (2018) pointed out that species such as *T. shetlandica* and *T. atlantis* have a wide distribution and were more frequent in samples, while *T. lavesquei* sp. nov. seems restricted to the Norwegian and Swedish coast. Some species were found at shallow depths, reaching the continental shelf border (0–200 m) such as *T. shetlandica*, while *T. atlantis* and *T. lavesquei* sp. nov. were found at depths below 200 m. *Terebellides atlantis* showed the wider bathymetric distribution (219–2750 m deep) among the species of this group. *Terebellides irinae* appeared at depths below 4000 m and its distribution seems to be restricted to the Arctic Ocean. Species of Group D, *T. williamsae* and *T. gracilis*, show a wide geographic and bathymetric distribution.

Comparisons with other NEA species of the genus Terebellides

Lavesque et al. (2019) described eight species from the Atlantic and Mediterranean coasts of France (see Key above), six of them belong to Group A sensu Nygren et al. (2018) and two are morphologically similar to those of Groups B and D, namely *T. ceneresi* and *T. parapari*. Lavesque et al. (2019) and Parapar et al. (2020a) considered *T. parapari* as related to Group B and particularly to *T. shetlandica*. *Terebellides parapari* shares with *T. shetlandica* and *T. atlantis* the presence of branchiae of type 3 and branchial lobes that are free from each other; *T. parapari* also shares with *T. shetlandica*, *T. lavesquei* sp. nov. and *T. atlantis* the presence of posterior filaments in lower branchial lobes, thoracic uncini of type 3 and abdominal uncini of type 2. However, branchial filaments in *T. shetlandica*, *T. lavesquei* sp. nov. and *T. atlantis* are longer and all notochaetae are of similar length while in *T. parapari* notochaetae in TC 1 are longer than subsequent ones.

On the other hand, *T. ceneresi* shares many morphological similarities with *T. williamsae* and *T. gracilis* and therefore was related to Group D by Lavesque et al. (2019) and Parapar et al. (2020a). These three species show white ventral colouration in anterior thoracic chaetigers, but in *T. williamsae* it is present in TC 1–4 while in *T. ceneresi* and *T. gracilis* is present only in TC 4. Other shared characters are the presence of ciliary tufts in the inner face of branchial lamellae and abdominal uncini of type 1A. However, *T. ceneresi* lacks the anterior branchial lobe (5th lobe) that is present in *T. williamsae* and *T. gracilis* and *T. gracilis* they are partially fused. Finally, *T. ceneresi* bears thoracic uncini of type 3 whereas *T. williamsae* and *T. gracilis* bear type 1.

Among the remaining clades that will be described elsewhere, clades 4, 14 and 26 do not correspond either to *T. atlantis* or *T. shetlandica* because of differences in the branchiae type (i.e., type 2: incompletely fused lobes) and the absence of posterior filaments in branchial ventral lobes; they also differ in geographic distribution, being the aforementioned clades restricted to some areas in NEA. Likewise, clade 25 does not fit within *T. irinae* due to being medium sized and by having branchiae of type 1 (=large lobes almost totally fused). Finally, clade 15 does not match either to *T. williamsae* or *T. gracilis* because of having a ventral colouration extending across more segments (TC 1–10 vs TC 1–4 or TC 4 in *T. gracilis* and *T. williamsae* respectively).

Characters and identification key

In this work, the following characters have been studied in all specimens: morphology of branchiae (sensu Parapar et al. 2016a), types of thoracic uncini (sensu Parapar et al. 2020b), abdominal uncini (sensu Parapar et al. 2020a), MG staining patterns (sensu Schüller and Hutchings 2010, 2013), and geographic and bathymetric distributions.

Among the species studied here, branchiae of *T. lavesquei* sp. nov., *T. williamsae*, and *T. gracilis* correspond to type 2, *T. shetlandica* and *T. atlantis* to type 3, and *T. irinae* to type 4. Regarding thoracic uncini, *T. williamsae* and *T. gracilis* have type 1, *T. lavesquei* sp. nov., *T. atlantis*, and *T. irinae* have type 3 and *T. shetlandica* bears

type 4. Finally, considering abdominal uncini, *T. williamsae* and *T. gracilis* have type 1A and *T. shetlandica*, *T. lavesquei* sp. nov., *T. atlantis*, and *T. irinae* have type 2.

Schüller and Hutchings (2010, 2013) defined several types of MG staining patterns according to the presence or absence of coloured bands in the segments along the body and their solid/striped appearance. The patterns observed in the species studied here are similar to those patterns described by Schüller and Hutchings (2010, 2013): *T. shetlandica* and *T. irinae* agree to pattern 1, *T. williamsae* and *T. gracilis* to pattern 2, and *T. lavesquei* sp. nov. and *T. atlantis* to pattern 9. However, we found that species with types 2 and 9 bear a J-shaped glandular region that is composed of three segments instead of only one as reported by Schüller and Hutchings (2010, 2013). These variations suggest that new staining patterns with taxonomic relevance might be determined when specimens from elsewhere are studied.

The species key is an update to those by Lavesque et al. (2019) and Parapar et al. (2020b) but still does not allow for morphological discrimination between three species (*T. norvegica*, *T. europaea*, and *T. scotica*). At present, these species can be differentiated only genetically and according to their geographical or bathymetric distributions.

Conclusions

A total of five nominal species has been identified as belonging to *Terebellides* Groups B, C, and D (according to Nygren et al. 2018): *Terebellides gracilis* Malm, 1874, *Terebellides atlantis* Williams, 1984, *Terebellides williamsae* Jirkov, 1989, *Terebellides irinae* Gagaev, 2009 and *Terebellides shetlandica* Parapar, Moreira & O'Reilly, 2016, and a new species is here described as *Terebellides lavesquei* sp. nov. Other species outlined by species delimitation analyses within these Groups will be either described elsewhere or would require additional material to be found.

The five species identified herein have been characterised based on morphological and molecular characters. The most relevant morphological features discriminating between species are branchial shape, ventral pigmentation of anterior thoracic chaetigers in live and fixed specimens, and the morphology of thoracic and abdominal uncini. For the molecular recognition of the species described, short sequences of nucleotides among the COI alignment have been provided as diagnostic to ease the identification.

Acknowledgements

We would like to thank all people involved in the paper by Nygren et al. (2018), for providing the specimens studied herein. Thanks also to Ada Castro and Catalina Sueiro (Servizos de Apoio á Investigación, Universidade da Coruña) for SEM assistance, and to María Candás (Estación de Bioloxía Mariña da Graña-Ferrol, Universidade de Santiago de Compostela, Spain) for assistance with the stereomicroscope photographs. This study was partly supported by the FAUNA IBÉRICA research project Polychaeta VII, Palpata, Canalipalpata II (PGC2018–095851–B–C64) funded by the Agencia Estatal

123

de Investigación, Ministerio de Ciencia e Innovación, and coordinated by JP. Funding was also provided from the Ramón y Cajal program (RYC-2016- 20799) funded by Spanish MINECO, Agencia Estatal de Investigación, Comunidad Autónoma de las Islas Baleares and the European Social Fund to MC. Financial support was also provided by the Norwegian Taxonomy Initiative: Cryptic polychaete species in Norwegian waters, knr 49-13, project no. 70184228 to AN; Polychaetes in the Norwegian Sea, project no. 70184227; Polychaetes in Skagerrak, project no.70184216; and the MAREANO program. We are deeply grateful to the reviewers of the manuscript, Pat Hutchings and Nicolas Lavesque, as well as Greg Rouse, ZooKeys Subject Editor and Nathalie Yonow, ZooKeys Copy Editor, for their constructive suggestions, which greatly improved the quality of the manuscript. We also thank Yordanka Banalieva and Tsvetelina Milenkova for their valuable help in the efficient manuscript editorial and accounting management processes.

References

- Aguado MT, Capa M, Lago-Barcia D, Gil J, Pleijel F, Nygren A (2019) Species delimitation in *Amblyosyllis* (Annelida, Syllidae). PLoS ONE 14(4): e0214211. https://doi.org/10.1371/ journal.pone.0214211
- Bakken T, Hårsaker K, Daverdin M (2020) Marine invertebrate collection NTNU University Museum. Version 1.535. NTNU University Museum. [Occurrence dataset:] https://doi. org/10.15468/ddbs14 [accessed on 26 June 2020]
- Capa M, Murray A (2015) Integrative taxonomy of *Parasabella* and *Sabellomma* (Sabellidae: Annelida) from Australia: description of new species, indication of cryptic diversity, and translocation of some species out of their natural distribution range. Zoological Journal of the Linnean Society 175(4): 764–811. https://doi.org/10.1111/zoj.12308
- Capa M, Bybee DR, Bybee SM (2010) Establishing species and species boundaries in Sabellastarte Krøyer, 1856 (Annelida: Sabellidae): an integrative approach. Organisms, Diversity & Evolution 10(5): 351–371. https://doi.org/10.1007/s13127-010-0033-z
- Capa M, Pons J, Hutchings P (2013) Cryptic diversity, intraspecific phenetic plasticity and recent geographical translocations in *Branchiomma* (Sabellidae, Annelida). Zoologica Scripta 42(6): 637–655. https://doi.org/10.1111/zsc.12028
- Caullery M (1915) Sur les *Terebellides* Malmgren du Siboga et les Terébelliens voisins. Bulletin de la Société Zoologique de France 40: 111–116.
- Dayrat B (2005) Towards integrative taxonomy. Biological Journal of the Linnean Society. Linnean Society of London 85(3): 407–417. https://doi.org/10.1111/j.1095-8312.2005.00503.x
- Gagaev SY (2009) *Terebellides irinae* sp. n., a new species of *Terebellides* (Polychaeta: Terebellidae) from the Arctic basin. Russian Journal of Marine Biology 35(6): 474–478. https://doi.org/10.1134/S1063074009060042
- Garraffoni ARS, Lana PC (2003) Species of *Terebellides* (Polychaeta, Terebellidae, Trichobranchinae) from the Brazilian coast. Iheringia. Série Zoologia 93(4): 355–363. https://doi.org/10.1590/S0073-47212003000400002

- Goldstein PZ, DeSalle R (2011) Integrating DNA barcode data and taxonomic practice: Determination, discovery, and description. BioEssays 33(2): 135–147. https://doi. org/10.1002/bies.201000036
- Grosse M, Capa M, Bakken T (2021) Describing the hidden species diversity of *Chaetozone* (Annelida, Cirratulidae) in the Norwegian Sea using morphological and molecular diagnostics. ZooKeys 1039: 139–176. https://doi.org/10.3897/zookeys.1039.61098
- Halt MN, Kupriyanova EK, Cooper SJ, Rouse GW (2009) Naming species with no morphological indicators: species status of *Galeolaria caespitosa* (Annelida: Serpulidae) inferred from nuclear and mitochondrial gene sequences and morphology. Invertebrate Systematics 23(3): 205–222. https://doi.org/10.1071/IS09003
- Hartman O, Fauchald K (1971) Deep-water benthic polychaetous annelids off New England to Bermuda and other North Atlantic Areas. Part II. Allan Hancock Monographs in Marine Biology 6: 1–327.
- Hoang DT, Chernomor O, Von Haeseler A, Minh BQ, Vinh LS (2018) UFBoot2: Improving the ultrafast bootstrap approximation. Molecular Biology and Evolution 35(2): 518–522. https://doi.org/10.1093/molbev/msx281
- Hutchings P, Peart R (2000) A revision of the Australian Trichobranchidae (Polychaeta). Invertebrate Systematics 14(2): 225–272. https://doi.org/10.1071/IT98005
- Jirkov IA (1989) Bottom fauna of the USSR. Polychaeta. Moscow State University Press, Moscow, 141 pp. [English translation from Russian]
- Jirkov IA, Leontovich MK (2013) Identification keys for Terebellomorpha (Polychaeta) of the eastern Atlantic and the North Polar Basin. Invertebrate Zoology 10(1): 217–243. https:// doi.org/10.15298/invertzool.10.2.02
- Kara J, Molina-Acevedo IC, Zanol J, Simon C, Idris I (2020) Morphological and molecular systematic review of *Marphysa* Quatrefages, 1865 (Annelida: Eunicidae) species from South Africa. PeerJ 8: e10076. https://doi.org/10.7717/peerj.10076
- Katoh K, Misawa K, Kuma KI, Miyata T (2002) MAFFT: A novel method for rapid multiple sequence alignment based on fast Fourier transform. Nucleic Acids Research 30(14): 3059–3066. https://doi.org/10.1093/nar/gkf436
- Lavesque N, Hutchings P, Daffe G, Nygren A, Londoño-Mesa MH (2019) A revision of the French Trichobranchidae (Polychaeta), with descriptions of nine new species. Zootaxa 4664(2): 151–190. https://doi.org/10.11646/zootaxa.4664.2.1
- Malm AW (1874) Annulater i hafvet utmed Sverges vestkust och omkring Göteborg. Kongelige Vetenskaps och Viterrhets Samhällets Göteborgs Handlingar 14: 71–105.
- McIntosh WC (1885) Report on the Annelida Polychaeta collected by HMS Challenger during the years 1873–76. Report of the Scientific Results of the Voyage of HMS Challenger 1873–76(12): 1–554.
- Müller OF (1776) Zoologiae Danicae prodromus, seu, Animalium Daniae et Norvegiae indigenarum characteres, nomina, et synonyma imprimis popularium. Impensis auctoris. https://doi.org/10.5962/bhl.title.63795
- Nguyen LT, Schmidt HA, Von Haeseler A, Minh BQ (2015) IQ-TREE: A fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. Molecular Biology and Evolution 32(1): 268–274. https://doi.org/10.1093/molbev/msu300

- Nygren A, Pleijel F (2011) From one to ten in a single stroke-resolving the European *Eumida sanguinea* (Phyllodocidae, Annelida) species complex. Molecular Phylogenetics and Evolution 58(1): 132–141. https://doi.org/10.1016/j.ympev.2010.10.010
- Nygren A, Parapar J, Pons J, Meißner K, Bakken T, Kongsrud JA, Oug E, Gaeva D, Sikorski A, Johansen RA, Hutchings PA, Lavesque N, Capa M (2018) A mega-cryptic species complex hidden among one of the most common annelids in the North-East Atlantic. PLoS ONE 13(6): e0198356. https://doi.org/10.1371/journal.pone.0198356
- Pante E, Schoelinck C, Puillandre N (2015) From integrative taxonomy to species description: One step beyond. Systematic Biology 64(1): 152–160. https://doi.org/10.1093/sysbio/syu083
- Parapar J, Hutchings P (2014) Redescription of *Terebellides stroemii* (Polychaeta, Trichobranchidae) and designation of a neotype. Journal of the Marine Biological Association of the United Kingdom 95(2): 323–337. https://doi.org/10.1017/S0025315414000903
- Parapar J, Moreira J, Helgason GV (2011) Taxonomy and distribution of *Terebellides* (Polychaeta, Trichobranchidae) in Icelandic waters, with the description of a new species. Zootaxa 2983(1): 1–20. https://doi.org/10.11646/zootaxa.2983.1.1
- Parapar J, Moreira J, O'Reilly M (2016a) A new species of *Terebellides* (Polychaeta: Trichobranchidae) from Scottish waters with an insight into branchial morphology. Marine Biodiversity 46(1): 211–225. https://doi.org/10.1007/s12526-015-0353-5
- Parapar J, Moreira J, Martin D (2016b) On the diversity of the SE Indo-Pacific species of *Terebellides* (Annelida; Trichobranchidae), with the description of a new species. PeerJ 4: e2313. https://doi.org/10.7717/peerj.2313
- Parapar J, Moreira J, Gil J, Martin D (2016c) A new species of the genus *Terebellides* (Polychaeta, Trichobranchidae) from the Iranian coast. Zootaxa 4117(3): 321–340. https://doi. org/10.11646/zootaxa.4117.3.2
- Parapar J, Capa M, Nygren A, Moreira J (2020a) To name but a few: Descriptions of five new species of *Terebellides* (Annelida, Trichobranchidae) from the North-East Atlantic. ZooKeys 992: 1–58. https://doi.org/10.3897/zookeys.992.55977
- Parapar J, Martin D, Moreira J (2020b) On the diversity of *Terebellides* (Annelida, Trichobranchidae) in West Africa, seven new species and the redescription of *T. africana* Augener, 1918 stat. prom. Zootaxa 4771(1): 1–61. https://doi.org/10.11646/zootaxa.4771.1.1
- Renner SS (2016) A return to Linnaeus's focus on diagnosis, not description: The use of DNA characters in the formal naming of species. Systematic Biology 65(6): 1085–1095. https://doi.org/10.1093/sysbio/syw032
- Sars M (1835) Beskrivelser og iagttagelser over nogle mærkelige eller nye i havet ved den bergenske kyst levende dyr af polypernes, acalephernes, radiaternes, annelidernes, og molluskernes classer: med en kort oversigt over de hidtil af forfatteren sammesteds fundne arter og deres forekommen. Dahl. https://doi.org/10.5962/bhl.title.13017
- Schlick-Steiner BC, Steiner FM, Seifert B, Stauffer C, Christian E, Crozier RH (2010) Integrative taxonomy: A multisource approach to exploring biodiversity. Annual Review of Entomology 55(1): 421–438. https://doi.org/10.1146/annurev-ento-112408-085432
- Schüller M, Hutchings PA (2010) New insights in the taxonomy of Trichobranchidae (Polychaeta) with the description of a new *Terebellides* from Australia. Zootaxa 2395(1): 1–16. https://doi.org/10.11646/zootaxa.2395.1.1

- Schüller M, Hutchings PA (2013) New species of *Terebellides* (Polychaeta: Trichobranchidae) from the deep Southern Ocean, with a key to all described species. Zootaxa 3619(1): 1–45. https://doi.org/10.11646/zootaxa.3619.1.1
- Teixeira MA, Vieira PE, Pleijel F, Sampieri BR, Ravara A, Costa FO, Nygren A (2020) Molecular and morphometric analyses identify new lineages within a large *Eumida* (Annelida) species complex. Zoologica Scripta 49(2): 222–235. https://doi.org/10.1111/zsc.12397
- Williams SJ (1984) The status of *Terebellides stroemi* (Polychaeta; Trichobranchidae) as a cosmopolitan species, based on a worldwide morphological survey, including description of new species. In: Hutchings PA (Ed.) Proceedings of the First International Polychaete Conference, Sydney, Australia, 1984. The Linnean Society of New South Wales, 118–142.

Supplementary material I

Abiotic data and more information of the material used

Authors: María Barroso, Juan Moreira, María Capa, Arne Nygren, Julio Parapar Data type: Abiotic data

- Explanation note: The table shows the abiotic data and more information of the material used: Specimen voucher, Site, Geographic area, Locality, Latitude, Longitude, Depth, Collecting date, Habitat, Remarks and figures.
- Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/zookeys.1132.91244.suppl1

Supplementary material 2

List of COI sequences considered in present study

Authors: María Barroso, Juan Moreira, María Capa, Arne Nygren, Julio Parapar Data type: Molecular data

- Explanation note: List of COI sequences considered in present study (Group B, C and D), museum vouchers and GenBank accession numbers.
- Copyright notice: This dataset is made available under the Open Database License (http://opendatacommons.org/licenses/odbl/1.0/). The Open Database License (ODbL) is a license agreement intended to allow users to freely share, modify, and use this Dataset while maintaining this same freedom for others, provided that the original source and author(s) are credited.

Link: https://doi.org/10.3897/zookeys.1132.91244.suppl2