
Chapter 1. Introduction

1.1. Background

Agriculture has had an enormous impact on the world's ecosystems. Natural habitats on arable land have been reduced by more than 50% by clearing for cropping or permanent pasture (Green et al. 2005), and much of the remaining natural habitats have been altered by livestock grazing (Groombridge & Jenkins 2002). Habitat destruction and modification for agriculture is one of the greatest global threats to biodiversity (Foley et al. 2005), yet global food demand is projected to double by 2050 (Tilman et al. 2011). Rapid rates of clearing to establish new industrial agricultural practices are mostly confined to developing countries in tropical and subtropical regions (Vandermeer & Perfecto 2007), though in areas where the extent of agricultural expansion is relatively complete, biodiversity loss continues through agricultural intensification (Donald 2004) and the continuing impacts of habitat fragmentation (Fischer & Lindenmayer 2007; Krauss et al. 2010). Nonetheless, agricultural landscapes can also be crucial repositories of biodiversity (Perfecto et al. 2009). A challenge facing humanity is to understand how biodiversity is changed by agricultural practices (Vandermeer 2011), and how biodiversity conservation can be integrated into farming practices and agricultural landscapes (Harvey et al. 2008; Henle et al. 2008; Rands et al. 2010; Phalan et al. 2011).

This thesis presents research into patterns of plant diversity in one agricultural region in eastern Australia—the North-West Slopes of New South Wales (NSW). This chapter introduces the concepts of biodiversity and its conservation, reviews current ecological knowledge of biodiversity patterns in agricultural landscapes—including the wheat–sheep zone of eastern Australia—and identifies knowledge gaps to be addressed by this

research. The chapter concludes with a statement of thesis objectives and an outline of thesis structure.

1.2. Biodiversity

1.2.1. Definition

The term ‘biodiversity’ has been used to describe many aspects of the living world (Mace 2005), and can mean many things to different people. The term is a contraction of ‘biological diversity’, and first came into use around 25 years ago (Wilson & Peter 1988). Biodiversity has been defined many times, and has been regarded variously as a concept, a measurable entity, and as a social or political construct (Gatson 1996). It is necessary to be specific about the definition when using the term in a scientific context (Lovejoy 1997). The most prevalent use of the term defines it as the variety of life in all its forms and at all levels of organisation (Hunter & Gibbs 2007). Biodiversity is often considered at three levels: genetic diversity (the genetic information of all species), species diversity (the variety of species on Earth) and ecosystem diversity (the variety of habitats, biotic communities and ecological processes). Biodiversity as a field of study, however, can represent an area of interest—the ‘scientific study of the patterns in, and the determinants and consequences of, the variety of life’ (Gatson 1998).

1.2.2. Why conserve biodiversity?

The importance of biodiversity to people varies greatly. The case for conserving it ranges from ethical and philosophical viewpoints to strict utilitarianism (New 2006). There are widespread views that humans have a moral and ethical responsibility of stewardship towards the variety of life, to opposing the extinction of species, and to supporting conservation efforts (Callicot 1986), though these arguments cannot be defended on scientific grounds. Other popular arguments for biodiversity conservation

include aesthetic values, or the usefulness of biodiversity for food, medicines and industrial products, including uses not yet discovered (Kunin & Lawton 1996; Díaz et al. 2006).

An additional argument for biodiversity conservation relates to the ecological roles that species play in the world. Humanity relies on other organisms to photosynthesise, form soils, pump water, fix nutrients, transform gases and regulate climate (Ehrlich & Ehrlich 1992). Species also impact on each other through competition, mutualisms and trophic interactions. There is increasing evidence that changes to biodiversity alter ecosystem processes and resilience, and have profound consequences for the services derived from ecosystems (Tilman 1999; Naeem 2002). Nevertheless, our knowledge of the impact of biodiversity loss is incomplete, and conserving biodiversity preserves options for solutions to future environmental problems (Chapin et al. 2000). The urgency of the need to conserve biodiversity is heightened by the concept of extinction debts—a condition in which populations persist in habitats following environmental change, sometimes for an extended period, prior to their extinction (Kuussaari et al. 2009; Krauss et al. 2010). In such cases, current occupancy of habitats overestimates carrying capacity and underestimates species loss due to past environmental change (Hanski & Ovaskainen 2002).

Despite the variety of justifications for conserving biodiversity (or because of it), 150 countries signed the Convention on Biological Diversity (UNCBD 1992), a treaty that pledges to conserve biodiversity, to use it sustainably, and to promote fair and equitable sharing of its benefits. Similarly, *Australia's biodiversity conservation strategy 2010–2030* (Commonwealth of Australia 2010a) details the commitment of the federal and all state and territory governments of Australia to the conservation of biological diversity.

1.2.3. *Plant diversity and conservation*

At the heart of many biodiversity conservation issues—and the focus of this thesis—is plant diversity and conservation. Plants are fundamental components of many of the Earth’s ecosystems, providing physical structures and resources for many other organisms. At any location, plants and vegetation influence local microclimate, hydrology, soil erosion and the types of animals that exist (Hamilton & Hamilton 2006). The loss of plant species inevitably results in the loss of other species. Many of the world’s charismatic animals are endangered by habitat loss and degradation, and efforts to save them are exercises in plant conservation (Dietz et al. 1994).

Research has most commonly focussed on plant *species* diversity, though there is increasing evidence of the importance of genetic diversity within plant populations (Wimp et al. 2004). This thesis considers plant species diversity predominantly from a plant conservation perspective. Such a perspective recognises the cultural, aesthetic and intrinsic values of Australia’s flora, the potential role of plant diversity in ecosystem functioning and other ecological interactions, as well as the policy of all Australian governments to conserve biodiversity (Commonwealth of Australia 2010).

1.2.4. *Australia’s biological diversity*

The flora of Australia is diverse and unique. Australia is one of 12 megadiverse countries that together harbour up to 70% of the world’s species (McNeely et al. 1990). Australia has a particularly high level of endemic species—including around 85% of its approximately 20 000 vascular plant species (Hnatiuk 1990)—which is related to the continent’s long evolutionary history of isolation subsequent to its separation from Antarctica beginning in the late Cretaceous (Frakes 1999; Murthy et al. 2011). Increasing aridity since the mid Tertiary resulted in the development of many

sclerophyllous (hard-leaved) species (Attiwill & Wilson 2003) and the evolution of diverse taxa such as the eucalypts (~800 species, Wilson 2003) and acacias (~970 species). The uniqueness of Australia's biodiversity is of global interest and significance.

1.3. Biodiversity patterns in agricultural landscapes

Pre-existing biodiversity is threatened in agricultural landscapes when natural ecosystems are subject to destruction, modification and fragmentation (Foley et al. 2005). The study of biodiversity in agricultural landscapes has presented new challenges for ecologists. A common ecological approach to studying agricultural landscapes has been to adopt landscape models that are conceptual summaries of how species are distributed through space. Most landscape models are based on ecological processes that influence species distribution patterns (Fischer 2004). This section summarises the landscape models commonly used by ecologists to conceptualise and study biodiversity patterns in agricultural landscapes.

1.3.1. Fragmentation models

The modification of landscapes for agriculture results in habitat loss (the reduction of the proportion of a landscape that is composed of habitat, Ewers & Didham 2006) and habitat fragmentation (the division of habitat into smaller, more isolated patches, Lindenmayer & Fischer 2006). The two processes are related, though their distinction is important—their effect on populations can vary in magnitude (Smith et al. 2009), and fragmentation changes the properties of the remaining habitat (Fahrig 2003). Habitat is a species-specific concept. What constitutes habitat loss for one species may not be for another. In this sense, equating habitat loss to the loss of native vegetation can be misleading (Fischer & Lindenmayer 2007). Nevertheless, many species in agricultural

landscapes are restricted to remnant native vegetation patches, and fragmentation has frequently been modelled in terms of remnant vegetation patches (Debinski & Holt 2000). The effects of habitat loss and fragmentation on pre-existing populations and ecosystems have become a prominent research focus in ecology.

The analogy of remnant vegetation in agricultural landscapes to islands in the ocean led to the adoption of Island Biogeography Theory (IBT, MacArthur & Wilson 1967) as the first theoretical framework to explain biodiversity patterns in fragmented landscapes. When applied to fragmented landscapes, IBT proposes that species richness in patches tends towards an equilibrium representing the rate of colonisation of new species from other patches, and the rate of local extinction within the patch (Figure 1.1). Extinction rate is determined largely by patch size, and colonisation rate is determined largely by the proximity of the patch to other patches. Hence, species richness increases with island size and decreases with degree of isolation. IBT is the most widely used model for conceptualising fragmented landscapes (Haila 2002), and is commonly advocated for the design of reserve networks (Diamond 1975; Cabeza & van Teeffelen 2001; Sax & Gaines 2011).

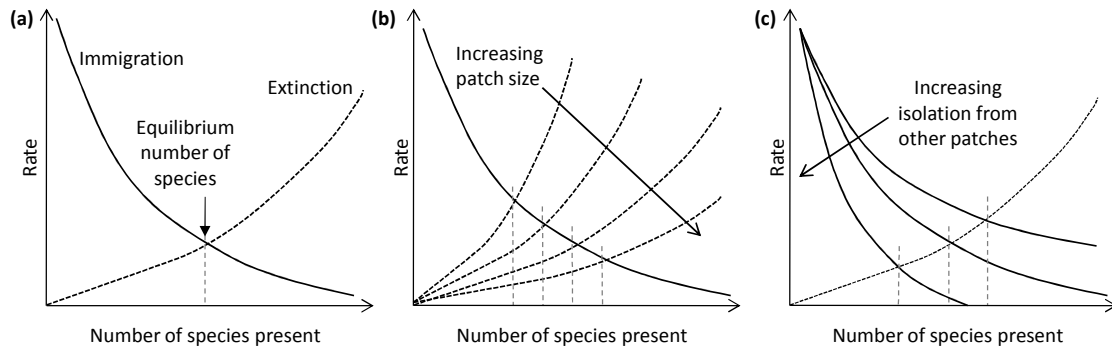


Figure 1.1 The equilibrium theory of island biogeography: (a) the number of species present on an island represents equilibrium between the rate of immigration of new species on to the island and the rate of extinction of previously established species. The equilibrium number of species is predicted to (b) increase with increasing island size, as the rate of extinction decreases, and (c) decrease with increasing isolation from other islands, as the rate of immigration decreases (modified from MacArthur & Wilson 1967; Vandermeer 2011).

The second classical paradigm used to study fragmented landscapes is Metapopulation Theory (MT), conceived by Levins (1969) and elaborated by others (Hanski & Gilpin 1991; McCullough 1996; Hanski & Simberloff 1997). MT is closely related to IBT, as it envisages the same processes of colonisation and extinction at work (Hanski 1991). MT conceptualises the dispersal and recolonisation interactions among subpopulations, and the persistence of the aggregate of subpopulations (i.e. the metapopulation). MT applies only to those species that demonstrate (1) subdivision into many local populations, (2) limited and random dispersal, and (3) relatively frequent local extinction. As this applies to many species whose ranges have been fragmented by human alteration, MT has been applied to many conservation problems in fragmented landscapes (McCullough 1996).

Forman's (1995) Patch–Matrix–Corridor Model was a modification of IBT for terrestrial environments that included corridors, in addition to remnant vegetation patches and the agricultural matrix, in its classification of landscape elements. This

model recognised that landscapes that retain corridors of habitat connecting remnant vegetation patches are more likely to retain species due to greater connectivity and recolonisation between patches.

1.3.2. Incorporating the matrix into agricultural landscape models

In reality, the agricultural matrix—the land surrounding isolated remnants of native vegetation—is not always analogous to an ocean, as assumed in IBT and MT (Haila 2002). The agricultural matrix is rarely hostile to all species. The matrix in many agricultural landscapes can be rich in biodiversity and can influence diversity patterns and the persistence of populations (Perfecto & Vandermeer 2002; Ries et al. 2004; Hendrickx et al. 2007; Franklin & Lindenmayer 2009).

To overcome the limitations of the binary classification of habitat and non-habitat of IBT and MT, McIntyre & Hobbs (1999) proposed an alternative model for conceptualising human-modified landscapes. Their Variegation Model recognises that landscapes can vary in their extent of modification, and defines a continuum of modification states that includes variegated landscapes as an intermediate state between intact and fragmented landscapes. In variegated landscapes, the original habitat is variously modified rather than largely destroyed. They highlighted many regions that fit this description. This approach considers the degree of habitat modification of different parts of the landscape and the extent to which different species are precluded from variously modified habitat states. The variegation model has proved useful for classifying and describing agricultural landscapes, and for overcoming the pitfalls of applying other fragmentation models to these landscapes.

Fischer et al. (2004) conceptualised landscapes as overlaid species-specific habitat contour maps. Their model acknowledges that different species respond differently to

the same environmental variables, and hence habitat suitability is modelled in relation to environmental gradients rather than human-defined patches and patch boundaries. While this model is more complex and unlikely to be directly applied to landscapes for large numbers of species, it is useful for conceptualising the complexity of habitat suitability, particularly in variegated landscapes, where variously modified habitat may extend the effective size of habitat for some species.

The Mosaic Concept, proposed by Duelli (1997), is a landscape model designed specifically for studying biodiversity in agricultural landscapes. The model again uses human-defined habitat types, but views agricultural landscapes as a mosaic of discrete land use types that result from human modification of natural habitat. This model assumes that patches of the same land use type generally have relatively homogeneous living conditions for plant species resulting from previous human land use decisions and interaction of these with the physical environment (Lunt & Spooner 2005; Simmering et al. 2006). According to Duelli (1997), biodiversity in an agricultural landscape is most determined by habitat variability (the number of land use types per unit area), habitat heterogeneity (structural and compositional variability within habitats) and the proportions of each land use type in the landscape. This model is useful for determining how landscape diversity is generated by the various land use types in agricultural landscapes.

While studies based on the above models have often superseded studies based on fragmentation models, the relevance and importance of fragmentation models for particular groups of species are still largely unknown. To evaluate the relevance of fragmentation models we need to know, firstly, how fragmented species are in the

landscape, and secondly, do populations in fragments act as metapopulations via colonisations and extinctions?

1.4. Effect of agricultural intensity on biodiversity

One of the key questions concerning the role of the matrix in conserving landscape biodiversity is: what effect does agricultural intensity have on biodiversity (Vandermeer 2011)? Two hypotheses describe the extremes of possible relationships between biodiversity and agricultural intensity (Figure 1.2). The first assumes that biodiversity follows a negative exponential trend with intensification, so that even agriculture at low intensity results in dramatic decreases in biodiversity. The alternative hypothesis states that biodiversity is relatively tolerant to low or moderate agricultural intensities, and only declines markedly under high agricultural intensity. The nature of this relationship has not been determined for most agricultural landscapes (Vandermeer 2011), or for most taxa, though greater understanding could improve our ability to manage landscapes for conservation.

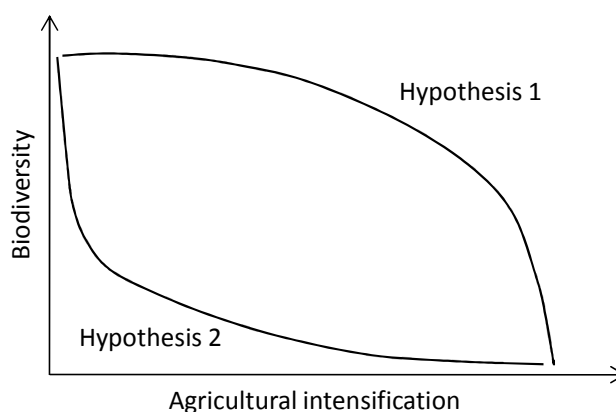


Figure 1.2 Two hypotheses about the relationship between agricultural management intensity and biodiversity (adapted from Green et al. 2005).

The biodiversity–intensity relationship assumes that agricultural yield is directly related to intensity. This is not always the case, and the x -axis of the biodiversity–intensity

relationship could also be conceptualised as yield, farm profit, or any value that is in conflict with biodiversity. Similarly, the measure of biodiversity used on the y-axis requires careful consideration. As many potential farming systems could apply in a given region, each with complex relationships to biodiversity, the reality of the biodiversity–intensity relationship is inevitably more complex than represented by Figure 1.2. This relationship has value in that it prompts us to consider the sensitivity of different species in the landscape to agriculture. However, the relationship could also form the basis of empirical evaluations, in which the biodiversity measures being investigated are clearly defined, and observed over a particular gradient of agricultural intensities, as has been done by Flohre et al. (2011).

The nature of the biodiversity–intensification relationship in different agricultural regions has often been used to provide support for one of two proposed conservation strategies for human-dominated landscapes (Balmford et al. 2003; Green et al. 2005; Fischer et al. 2008): (1) land sparing, in which increased intensification and yields on parts of the farm can free up land for conservation management on other parts of the farm, and (2) wildlife-friendly farming, which involves extensive, but lower intensity production that incorporates biodiversity conservation within the farming system. Generally, wildlife-friendly farming is thought to be the more appropriate biodiversity conservation strategy where Hypothesis 1 (Figure 1.2.) is true, while land sparing is more appropriate where Hypothesis 2 is true. However, many authors have argued that while land sparing may be a potential means of conservation, it is unlikely in reality. Matson & Vitousek (2006) suggested that intensification begets extensive land use, and successful intensive land use provides the economic incentive for more intensive farming, rather than maintaining total landscape yields and sparing land for

conservation. They also argue that the many effects of intensive farming on vegetation and wildlife beyond farming areas are not taken into account in most evaluations.

Perfecto & Vandermeer (2010) have argued that the land sparing vs. wildlife-friendly farming debate is neither useful for optimising production outcomes or successfully conserving biodiversity. They argue that current conventional agriculture involves much overconsumption and waste. On a caloric, per capita basis, global food demands require less intensification than many estimates suggest, and addressing socioeconomic inequities will improve options for wildlife-friendly farming and on-farm biodiversity conservation (Vandermeer & Perfecto 2007).

1.5. The wheat–sheep zone of eastern Australia

The study region for this research into plant diversity in agricultural landscapes—the North-West Slopes of NSW—forms part of the wheat–sheep zone of eastern Australia (Figure 1.3a). The extent of this zone coincides with Australia’s woodlands and open woodlands (Figure 1.3b), which are characterised by extensive tree clearing and landscape modification, livestock grazing and fertilisation. Research on the effects of these processes is substantial (McIntyre & Lavorel 1994a; Prober & Thiele 1995; Yates & Hobbs 1997a; Kirkpatrick 1999; McIntyre et al. 2002; Landsberg et al. 2003; Dorrough & Ash 2004; Lunt & Spooner 2005; Dorrough et al. 2007), though due to their complexity, many of the effects of land use are still poorly understood. One impediment to the ecological understanding of these systems is that the findings in one region of the wheat–sheep zone do not always translate to other regions due to the complex interactions between climate, productivity, the degree of landscape modification, and other agricultural management practices, both past and present (McIntyre & Martin 2001; McIntyre et al. 2003; Lunt & Spooner 2005; Dorrough et al.

2006). Consensus on the influences of agricultural management practices will require recognition and understanding of these differences. Despite the differences between agricultural regions, our understanding of ecosystem dynamics in any particular region of the wheat–sheep zone is largely based on research conducted in other parts of the zone. With this in mind, this section briefly reviews the impacts of agriculture on grassy ecosystems in the wheat–sheep zone, with particular reference to the effects on plant diversity.

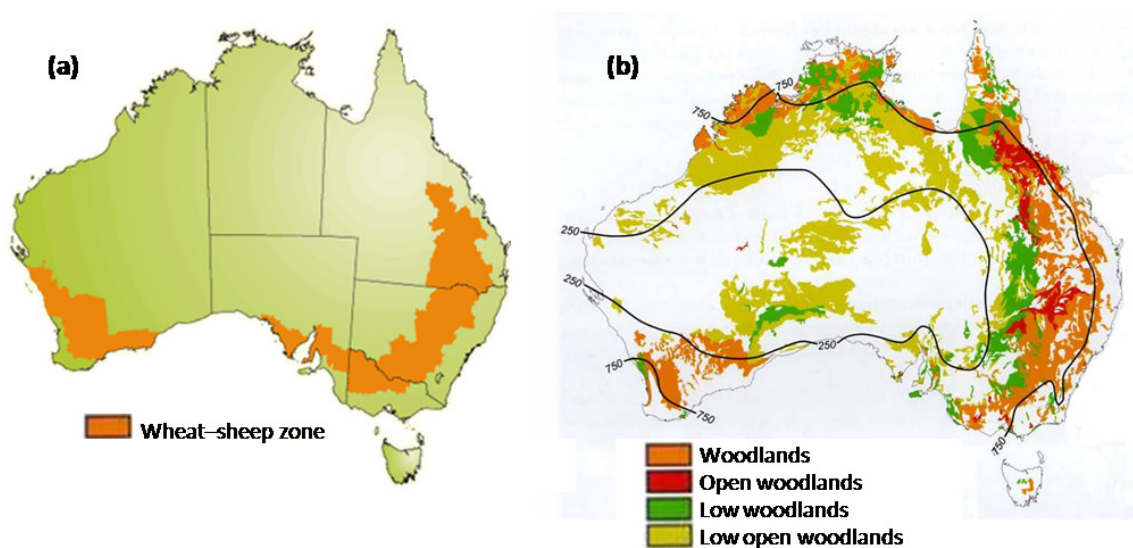


Figure 1.3 (a) Wheat–sheep zone of eastern Australia and south-west Western Australia (ARNA 2007); (b) woodlands of Australia, showing the 250 mm and 750 mm annual rainfall isohyets. Source: Lindenmayer et al. (2005), redrawn from Johnson (2003).

1.5.1. Management history

The Australian continent has a long history of management by indigenous Australians. The time of arrival of the first humans on the continent was around 50 000 years ago (Summerhayes et al. 2010; Rasmussen et al. 2011). There is evidence that many Australian landscapes are the ecological outcomes of management activities of the Aboriginal people (Kohen 1995). The history and role of fire in Australia long pre-dates the arrival of Aboriginal people, but Aboriginal burning regimes have been argued to

have strongly influenced – and possibly determined – the composition of vegetation in eastern Australia (Bowman 1998; Gammage 2011a, b).

Despite this long history of management, it was the introduction of agricultural practices by Europeans from 1788 that brought about extensive and rapid ecological changes in south-eastern Australia. The dispossession of Aboriginal people from their lands by Europeans, and the associated suppression of Aboriginal burning regimes, had a large influence on the distribution and structure of some vegetation types, and possibly the intensity of fires (Bowman 1998; Bowman & Davies 2003).

Following European occupation, grassy woodlands and grasslands quickly became an important pastoral resource (Yates & Hobbs 1997b). The introduction of livestock had an enormous impact on the groundstorey vegetation of the wheat–sheep belt (Yates & Hobbs 1997a). Australian ecosystems have not evolved with hard-hooved ungulate herbivores (Mack & Thompson 1982), and the introduction of cattle and sheep had a major impact on soils, resulting in loss of biological soil crusts, reduced soil water infiltration rates and increased soil surface erosion, resulting in a loss of ecosystem function (Noble & Tongway 1983; Wilson 1990; Yates & Hobbs 1997a; Kirkpatrick 1999; Prober et al. 2002; Gale & Haworth 2005). In addition to the influence of ungulate hooves, changes in total grazing pressure, herbivore densities and the differing foraging behaviour of sheep and cattle compared to native herbivores have influenced understorey composition. In the wheat–sheep zone, livestock grazing is believed to have caused the decline or local extinction of many grazing-sensitive native plants (Lunt et al. 2007a). Other changes associated with livestock grazing in these systems include a decline in the diversity of native forb species (Trémont & McIntyre 1994), a shift toward cool-season growth (Moore 1970), and an increasing dominance of annual

plants (Pettit et al. 1995; Prober & Thiele 1995). The absence of soil seed banks of native forbs makes many of these changes irreversible in the absence of human-mediated restoration measures (Lunt 1997; Morgan 1998a).

In addition to the effects of changed grazing regimes, clearing and fragmentation have also had a large influence on native vegetation. In what is now the wheat–sheep zone, tree thinning by felling and ringbarking to increase productivity in the grassy layer was extensive (McDougall & Kirkpatrick 1994). Denser eucalypt woodlands with shrubby understoreys were less productive, and hence were less extensively cleared (Johnson 2003). Clearing was primarily focussed on the woodlands dominated by box eucalypts, which previously covered over 10 million ha across south-eastern Australia (Beadle 1981). Over 90% of these woodlands have been cleared, and are now largely restricted to remnants of varying size, quality and isolation (McIntyre 1994; McIntyre & Lavorel 1994b; Prober & Thiele 1995).

The extent of landscape modification varies across the wheat–sheep zone, from variegated landscapes to highly fragmented landscapes, depending on the predominant agricultural land uses. Fragmentation of these communities has led to reduced species diversity through insularisation and altered ecosystem processes (see Yates & Hobbs 1997a for a review of the past and continuing impacts of fragmentation on eucalypt woodlands). The vegetation of the cleared areas varies from completely transformed crops to derived native grasslands and savannas dominated by native grasses and in which many native plant species survive. In some regions, commercially grazed pastures contain a high richness of native species, including rare species (Gilfedder & Kirkpatrick 1994; McIntyre & Lavorel 1994b; Chilcott et al. 1997).

The use of fertiliser in native vegetation is also associated with the transformation of the wheat–sheep zone (Dorrough et al. 2006). Many native species have coevolved with nutrient-poor soils (Beadle 1954). Losses of endangered plants and declines in species richness have been linked to increases in phosphorus (P) and nitrogen (N). In some areas, fertiliser application rates have increased over time in response to agricultural intensification and declining productivity of soils (Passioura 2002). These nutrients can accumulate in native vegetation and be transported across the landscape by livestock (Taylor & Hedges 1984; Taylor et al. 1987; Duncan et al. 2008) and by wind and erosion (Leys & McTainsh 1994; Morgan 1998b). In many cases, the relative influence of the confounded impacts of clearing, grazing and fertiliser on native vegetation have not been disentangled (but see Dorrough et al. 2006).

Another major continuing threat to plant diversity in the wheat–sheep zone is invasions of exotic species, in particular exotic grasses and herbs (McIntyre & Lavorel 1994b). Small vegetation remnants in agricultural landscapes are particularly susceptible to invasions due to high exposure to invasive propagules (Adair 1995) and a high edge-to-area ratio (Panetta & Hopkins 1991). Exotic species abundance has been shown to increase with grazing intensity (Pettit et al. 1995; Dorrough et al. 2007), soil disturbance (McIntyre & Lavorel 1994b) and nutrient enrichment (Morgan 1998b; Allcock 2002), and invasions are generally associated with declines in native species richness (Yates & Hobbs 1997a; Clarke 2003).

1.5.2. North-West Slopes of NSW

The vegetation of the North-West Slopes of NSW (the study region) is reviewed in detail in Chapter 2, including a comparison of several features such as the climate and extent of landscape modification with other regions in the wheat–sheep zone.

1.6. Thesis objectives and aims

The main objective of this research was to examine patterns of species distribution across farms grazing sheep and cattle in the study region, and to determine the value of various land uses in agricultural landscapes to plant diversity conservation. Traditionally, ecologists and conservation biologists have focussed on the distribution and abundance of different taxa in remnants of intact or semi-natural native vegetation in agricultural landscapes, with fewer studies addressing the role of anthropogenic land uses, such as pastures, in contributing to landscape diversity (Bennett et al. 2006). This research addresses this knowledge gap, with a particular focus on the plant diversity of derived native pastures grazed for livestock production. In doing so, I test the nature of the biodiversity – agricultural intensification relationship—that is, what is the relationship of plant diversity to agricultural intensity in the region? Specific aims were to: (1) document the diversity, phenology and ecology of native perennial pastures, (2) investigate the biophysical and management determinants of vegetation composition in the study region, (3) provide plant conservation recommendations for the North-West Slopes of NSW, and (4) evaluate the likely economic impact on farmers of implementing different plant conservation strategies. Such research is essential if effective conservation measures and policies are to be developed and implemented in agricultural landscapes.

1.7. Thesis structure

This thesis consists of five research papers (Chapters 2–6) addressing plant diversity in the study region, plus a synthesis chapter (Chapter 7) summarising the findings and recommendations. The following paragraphs introduce each of the chapters, and explain how the sequence of chapters, and the research questions they address, are designed to

build on the knowledge of plant diversity and its conservation in agricultural landscapes. Each research paper will be submitted for publication in the scientific literature, and hence has been written as a stand-alone contribution. However, some introductory material common to two or more papers has been removed to avoid excessive repetition. References to other chapters in the thesis are cited by chapter number.

Chapter 2 describes the North-West Slopes of NSW, and analyses a database of existing vegetation survey data to characterise the native vegetation of the region. This chapter includes an analysis of the occurrence, frequency and conservation status of understorey plant species in the region, and the patterns in native vegetation composition that are relevant to farmland plant diversity. It highlights some important conservation challenges, and sets the scene for the subsequent research.

Chapter 3 addresses botanical and ecological knowledge gaps regarding plant diversity in native pastures. The study documents the seasonal variation in species composition of native pastures in the region, as well as the short-term (2.5-year) response of native pasture to the exclusion of grazing by livestock. The results of this study are pivotal to the design, data collection and interpretation of the subsequent studies in the project, and also provide useful reference material for the design and interpretation of future research into plant diversity in both native pastures and agricultural landscapes.

Chapter 4 is a case study of a single farm and some of the land immediately adjacent to it in the surrounding agricultural landscape. It reports the vegetation composition of the different land uses. Two landscape ecological models are applied to evaluate how landscape diversity is generated across the various land uses, at different spatial scales.

This chapter demonstrates the importance of spatial scale to the evaluation of conservation value of native pastures at the farm scale.

In Chapter 5, the scale of the analysis broadens from the farm to the region. Vegetation was surveyed and biophysical and management data were compiled to determine the key determinants of vegetation composition for on-farm vegetation. Sample-based rarefaction curves and habitat specificity indices were used to determine the landscape and regional-scale effects of agricultural land uses on landscape plant diversity.

Chapter 6 uses a scenario and model farm approach to determine the likely opportunity costs of implementing plant conservation management on farms grazed by sheep in the region. The scenarios are based on landscape ecology and woodland restoration principles, and the conservation management recommendations from Chapter 5.

Chapter 7 synthesises the findings of Chapters 2–6, details conclusions regarding plant diversity patterns in the study region, and summarises the management recommendations arising from these studies.

STATEMENT OF AUTHORS' CONTRIBUTION

We, the PhD candidate and the candidate's Principal Supervisor, certify that all co-authors have consented to their work being included in the thesis and they have accepted the candidate's contribution as indicated in the *Statement of Originality*.

	Author's Name (please print clearly)	% of contribution
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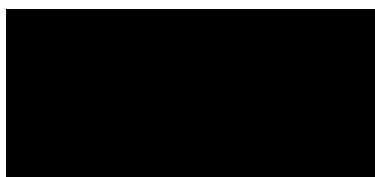
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Type of work	Page number/s
Text	19–44
Tables and figures	22–40

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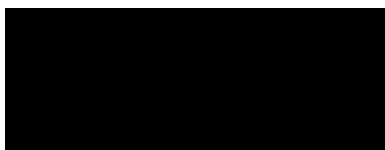
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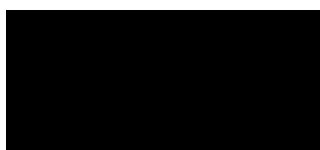
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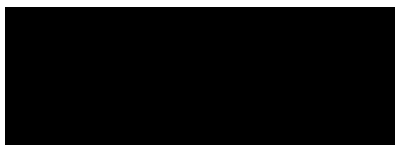
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Type of work	Page number/s
Text	47–70
Tables and figures	52–64

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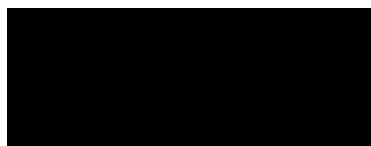
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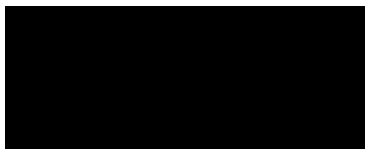
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Text	73–96
Tables and figures	77–90

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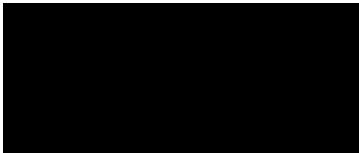
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However, many native grass and herb species are restricted to the now limited extent of woodlands not grazed by livestock. These habitats make a high contribution to regional plant diversity.

STATEMENT OF AUTHORS' CONTRIBUTION

We, the PhD candidate and the candidate's Principal Supervisor, certify that all co-authors have consented to their work being included in the thesis and they have accepted the candidate's contribution as indicated in the *Statement of Originality*.

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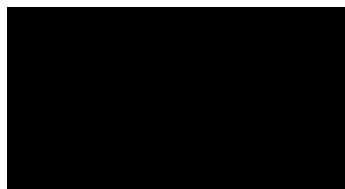
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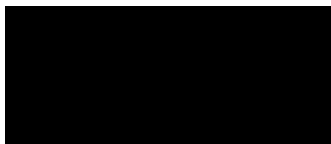
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Type of work	Page number/s
Text	99–133
Tables and figures	103–123

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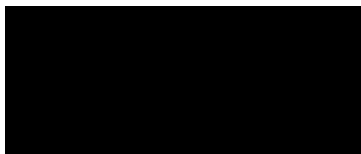
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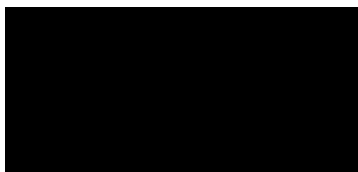
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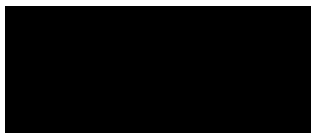
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Type of work	Page number/s
Text	136–157
Tables and figures	142–150

Name of Candidate: Nicholas Schultz

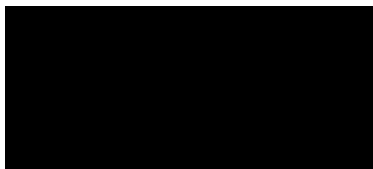
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Chapter 7. Synthesis and conclusions

7.1. Introduction

Biodiversity is under threat in the world's agricultural landscapes due to the modification and destruction of natural habitat, and the continuing impacts of habitat fragmentation (Tilman et al. 2001; Foley et al. 2005). Globally, there are increasing calls by conservationists to integrate biodiversity conservation and production systems (Harvey et al. 2008; Perfecto & Vandermeer 2008). However, the gap in understanding of ecological interactions and diversity patterns in agricultural landscapes must first be addressed (Vandermeer 2011). This study addressed this knowledge gap for the North-West Slopes of NSW by determining the plant conservation value of vegetation on sheep farms grazed by both sheep and cattle. In particular, the study aimed to: (1) provide new information on the diversity, phenology and ecology of native perennial pastures; (2) investigate the biophysical and management determinants of grassy vegetation composition in the region; (3) provide region-specific plant conservation recommendations, and (4) evaluate the likely economic impact of implementing different plant conservation strategies.

This chapter summarises and explores the implications of the research findings presented in this thesis. The objectives are to: (1) synthesise the main findings of the research; (2) highlight the major theoretical advances of the work; (3) summarise the main management recommendations, and (4) highlight and discuss future research implications.

7.2. Summary of main findings

In Chapter 2, a large dataset of existing vegetation surveys was analysed. This highlighted some key conservation issues in the study region. Canonical Correspondence Analysis (CCA) revealed that native vegetation composition in the study region was controlled by two major gradients: a climate gradient and a slope – soil depth gradient. Furthermore, the study demonstrated that the location of the region’s conservation reserves was predominantly in hilly terrain at higher altitude, with few reserves in gently sloping areas at lower elevation. Hence, as farmland occupies the majority of low sloping, fertile land, plant conservation in the region will require either widespread uptake of on-farm conservation initiatives, or for a large number of farms to be bought and added to the reserve system. To allow for the conservation of plant diversity at the regional scale, agricultural lands must successfully integrate biodiversity conservation, or a suite of reserves will be needed in areas of low relief. These findings provided context and direction for studies of groundstorey plant conservation reported in subsequent chapters.

In Chapter 3, vegetation composition in paired grazed and ungrazed plots was monitored in native pastures over 2.5 years, in which time composition varied greatly with season and interannual rainfall. Spring surveys in non-drought years recorded the highest number of species at each site, but even the periods of maximum species density (SD) revealed only 60–72% of the species recorded at each site over the 2.5 years. Nevertheless, compositional differences were much greater among sites than seasons— that is, seasonal and interannual variability was not as influential in determining composition as the factors that determine spatial differences (such as biophysical variability and management). Exclusion of livestock grazing did not influence

composition over 2.5 years, but within-site species turnover should be considered when using one-off vegetation survey data to address research questions in plant ecology and conservation.

In Chapter 4, the mosaic concept (Duelli 1997) was applied to a single case study farm to observe how different land uses contributed to landscape plant diversity. Additive partitioning was used to observe how landscape richness was partitioned across four spatial scales (quadrat, patch, land use and landscape) for five land uses in the landscape (grazed and ungrazed woodlands, native pastures, roadsides and sown crops). Grazed and ungrazed woodlands had similar species richness to native pastures at the quadrat and patch scale, but both had greater dissimilarity between patches than pastures. Hence, woodlands were more species rich at the landscape scale. A habitat specificity measure (gamma diversity contribution, GDC) was employed to identify patches contributing high numbers of infrequent species to landscape diversity. GDC was significantly greater for ungrazed woodland patches than all other land uses, showing that these sites contributed a higher effective number of species to landscape diversity. Native pastures, sown forage crops and grazed woodlands made low contributions to landscape diversity, suggesting that synergies were unlikely to exist between pastoral practices and plant diversity conservation in this region. The study showed that those parts of the farm that were not grazed by livestock were important for understorey plant diversity at the landscape scale.

In Chapter 5, 127 sites were surveyed across four land use categories—previously cultivated native pastures, never-cultivated native pastures, grazed woodlands and ungrazed woodlands—to test the influence of land management and biophysical variables on vegetation. Livestock grazing, fertiliser application and tree cover were not

well correlated with total species densities (i.e. total species richness per unit area), but were correlated with native and exotic species densities. CCA showed that vegetation composition was strongly correlated with agricultural intensity (covarying gradients of decreasing tree cover and time-since-cultivation, and increasing amount and duration of fertiliser use and grazing intensity). Climate was secondary to agricultural intensity in explaining farmland plant composition. This contrasted with the native vegetation surveys analysed in Chapter 2, where climate best explained compositional patterns.

The rates of species accumulation across woodlands and pastures observed at the landscape scale in Chapter 4 were accentuated at the regional scale in Chapter 5. Regional species rarefaction curves showed that ungrazed woodlands accumulated the most species across the region, and native pastures accumulated the least, reflecting a high degree of compositional homogeneity in native pastures. The contribution of sites to regional diversity was negatively correlated with land use intensity: mean GDC was significantly higher in ungrazed woodlands than in other land uses. This suggested that both grazing and clearing may be influential in the homogenisation of pasture composition. Overall, this study showed that pastoral management had a consistent influence on vegetation composition that was not conducive to the conservation of a large proportion of native vascular plant species in the region. Native pastures with no cultivation history were comprised of a larger proportion of the regional native species than pastures that had been previously cultivated, and unfertilised pastures showed greater floristic affinity with the woodland understorey vegetation. For the suite of species that persist in them, native pastures provide extensive habitat that ensures large, viable populations that are well connected in the landscape. Nevertheless, woodland

vegetation that was not grazed by livestock made the largest contribution to regional plant diversity, despite its limited extent in the landscape.

In Chapter 6, a modelling approach was used to determine the opportunity costs of setting aside woodland on farms to be managed for conservation without livestock grazing. Although management options such as increasing fertiliser use on other native pastures or sowing an additional area of forage oats could partially offset the opportunity costs of set-aside land, these options were not a convincing means of on-farm conservation for two reasons. First, a high proportion of the opportunity costs were associated with the capital required to establish and maintain set-asides (initial fencing and on-going weed control), so that offsets through intensification represented a small proportion of the total cost compared with the option of simply reducing flock size to account for lost grazing area. Second, detrimental environmental and biodiversity outcomes associated with intensification options were not factored into the analysis. The potential for these disadvantages to be large was incongruent with conservation management objectives, and did not warrant their selection over management options with no adverse environmental impacts. The cost of on-farm conservation management was substantial (opportunity costs of \$173 ha⁻¹ yr⁻¹ for reducing flock size and managing a 25-ha set-aside woodland). Incentives and stewardship payments of \$100–200 ha⁻¹ yr⁻¹ will be required if conservation management through set-aside land is to be widely adopted, unless cost-sharing arrangements for capital expenditure can be made, or suitable existing fences are in place.

7.3. Main contributions of this study

7.3.1. Contribution to scientific theory and practice

This study has contributed to scientific theory and practice by: (1) providing evidence of the importance of farm vegetation to plant conservation; (2) increasing the knowledge of seasonality and interannual variability in native pasture composition; (3) providing new information about the partitioning of landscape diversity across different land uses in an agricultural landscape; (4) identifying the biophysical and management determinants of grassy vegetation composition in the study region; (5) demonstrating the regional homogeneity of grassy vegetation in grazed and treeless native pastures; (6) identifying areas that contribute high numbers of unique plant species to the regional species pool, and (7) providing a basis for the investigation of the ecological processes that create landscape diversity patterns.

The analyses of existing vegetation survey data (Chapter 2) contributed to the knowledge of conservation issues in the study region. Other authors (Hosking & James 1998) have highlighted that existing reserves in the region do not adequately represent the range of regional vegetation communities, a finding commonly reported in other regions (Pressey et al. 1993; Pressey 1994; Fischer et al. 2010). The current study supported the findings of these authors by: (1) providing evidence for the importance of strategic and representative reserve creation, and (2) highlighting the importance of farmland in biodiversity conservation, due to its occurrence on land of high conservation priority—gently sloping, fertile areas.

Chapter 3 added substantially to the knowledge of seasonal and interannual variability in the species composition of native pastures in the region. Prior to this study, knowledge of these patterns was based on: (1) seasonal vegetation monitoring from a

study by Burrows (2004) from a different vegetation type and different climate; (2) a study by Williams (1979) of the region's native pastures that included winter and spring repeat surveys only, and (3) observations of seasonal ground cover and herbage mass of the dominant grasses in native pastures in several studies (Lodge 1981; Lodge & Whalley 1981; Lodge et al. 2003). The current study was the first to provide detailed data on seasonal variation of total plant species composition of native pastures, across years of varying rainfall. Most vegetation research is based on a single survey conducted at each survey site, without knowledge or consideration of the proportion of site species diversity not captured by a single survey. These data improve our ability to estimate the proportion of site species richness not recorded by single vegetation surveys, and to compare surveys conducted in different seasons or years. The study challenged the assumption of many plant demographic studies that almost all species present in a vegetation community are detected in a single survey.

The extensive survey of the case study farm based on the mosaic concept (Duelli 1997) contributed new insights into how plant diversity is partitioned across different land uses and spatial scales in an agricultural landscape. In Europe, this approach has allowed a better understanding of diversity patterns in agricultural landscapes (Wagner et al. 2000; Duelli & Obrist 2003; Simmering et al. 2006), but it has not been applied in Australia. The approach recognises that one land use type might be more species rich than another at the patch scale, but less species rich at the landscape scale. This study contributed knowledge of landscape patterns that have major conservation significance but have received little attention in Australian plant diversity studies in agricultural landscapes. The study considered only a single landscape, but revealed the landscape-

scale diversity of several land uses, highlighting the potential for such analyses to further understanding of diversity patterns in farmland.

Chapter 5 identified the determinants of species density of total, native and exotic species in the region's grassy vegetation. The determinants of site-scale density across environment and management gradients in grassy vegetation have been the subject of much research in other parts of the southern and eastern Australian wheat–sheep zone, including the Northern Tablelands of NSW (McIntyre & Lavorel 1994b; Clarke 2003), the Monaro Tablelands of NSW (Dorrrough et al. 2004a), the Tasmanian midlands (Kirkpatrick et al. 2005) and subtropical south-east Queensland (McIntyre et al. 2003). However, this was the first such study on the North-West Slopes of NSW. Strong differences were evident between regions in the effects of grazing, clearing and fertiliser application on groundstorey species densities (McIntyre et al. 2002; Dorrrough et al. 2004a). Hence, this study was important for determining how the relationships on the North-West Slopes of NSW differ from those in other regions.

Chapter 5 also demonstrated the regional homogeneity of grassy vegetation in grazed and treeless native pastures. These findings have major implications for how grazing is viewed as a process acting at landscape and regional scales. While several other studies have observed the effects of grazing at broad landscape scales (Landsberg et al. 2002; McIntyre et al. 2002; Dorrrough et al. 2007), this study revealed a homogenising effect of grazing and clearing on grassy vegetation more clearly than other Australian studies. This may be partly because grazing does not have the same influence in other regions (e.g. McIntyre et al. 2002). Nonetheless, the broad-scale effects of grazing remain untested in most agricultural landscapes in south-eastern Australia, and this study highlighted their importance for plant conservation.

The analyses of GDC contributed information about landscape diversity patterns that differed from the diversity patterns observed by additive partitioning and species accumulation curves. Habitat specificity indices such as GDC consider which species a site holds, rather than simply how many species, and hence provided information about the uniqueness of the flora of particular sites, which is not evident from species numbers and abundances alone. No other studies of plant diversity in Australia have used specificity indices in an agricultural context. The information provided by GDC can: (1) identify diversity patterns of conservation significance; (2) help identify ecological processes that create these patterns, and (3) prioritise vegetation patches in the landscape for reservation or conservation management.

7.4. Management recommendations

Conservation efforts at the ecological community level need to be focussed on those communities that are underrepresented in the reserve system due to their occurrence on low sloping, fertile land—in particular, Box Gum Grassy Woodlands (BGGW), which are listed as critically endangered under the Commonwealth EPBC Act 1999, and have been cleared from an estimated 94% of their pre-1750 distribution on the North-West Slopes of NSW (NPWS 2000). Specifically:

- landholders should consider any BGGW remnant for conservation management, and
- this vegetation should be prioritised for additional stewardship payments by the Namoi and Border Rivers – Gwydir Catchment Management Authorities and for land acquisitions by the federal government and non-government organisations. The federal government, through their BGGW Environmental Stewardship Project (BGGW ESP) and Multiple Ecological Communities project, should

continue to prioritise conservation in these communities, and provide further rounds of funding for additional stewardship contracts.

Furthermore, BGGW vegetation on fertile land of low slope (i.e. land of high agricultural capability) is of particularly high conservation value and should be prioritised for conservation management. This will require the:

- conservation and protection of existing high quality remnant vegetation, particularly the region's Travelling Stock Routes on low sloping ground, which provide some of the highest quality remnants of BGGW. The future management of these TSRs is currently being debated;
- restoration of remnant vegetation that has been degraded by livestock grazing, and
- regeneration of woodland in areas of native pasture that have remained free of fertilisation and cultivation.

These recommendations can be seen as an overarching goal for all stakeholders (landholders, policy makers, catchment management authorities, etc.). However, socioeconomic and political factors related to the production nature of the landscape for which this recommendation is intended are likely to constrain its realisation, unless viable farm practices involving reduced stocking rate are adopted.

These management recommendations differ from those stemming from other research in several important ways. For instance, Fischer et al. (2009) suggested that the next generation of conservation initiatives should consider a suite of measures in addition to the traditional land-sparing conservation management. They surmised that there was 'overwhelming evidence that high grazing pressure is substantially more damaging to plants than low grazing pressure', and as such proposed that lowering stocking rates and

implementing cell grazing may provide broad-scale biodiversity benefits. While I agree that these initiatives warrant research, of which there is currently little (Dorrough et al. 2004b), the results of this thesis suggest that both high and low grazing pressures may fail to provide high conservation value habitat for plant conservation, and so support for alternative grazing regimes as a conservation strategy may be premature, unless the grazing regime includes not grazing some of the farm.

These management recommendations also differ from a global trend in ecological theory and biodiversity conservation in agricultural landscapes. Authors such as Perfecto et al. (2009) and Vandermeer & Perfecto (2007) argue that the quality of the agricultural matrix is far more important to biodiversity conservation than the extent and quality of conservation reserves. They proposed a return to traditional agro-ecological landscapes, or the implementation of alternative agro-ecological techniques, to create high quality agricultural matrices. I agree that systems in which species are not confined to remnants and not subject to fragmentation and isolation are much preferable to 'island-sea' landscapes. However, these ideas were developed mostly from research in Mesoamerica and South America—areas with a longer history of agro-ecological management practices, and an evolutionary history of grazing by ungulates. In south-eastern Australia, the equivalent agro-ecological alternatives are less well defined. although grazing systems such as cell grazing (Earl & Jones 1996; Richards & Lawrence 2009) aim to improve the quality of the agricultural matrix, but have yet to demonstrate positive plant conservation outcomes empirically. These practices are still based on livestock grazing, and results from this thesis suggest that they may not be as effective at conserving the regional plant species pool as increasing the network of woodland habitats not grazed by livestock.

The studies in this thesis provided empirical evidence (which has previously been lacking: Dorrough 2010) that the approach of the BGGW ESP (a government scheme that provides stewardship payments to landholders for managing set-aside land for conservation outcomes) may be a suitable option for conserving the regional plant species pool. As there is no generally agreed grazing regime to incorporate biodiversity conservation with production in temperate Australia (and very little empirical evidence to recommend any regime), the BGGW ESP is successful in the current socioeconomic and political climate and can potentially deliver positive conservation outcomes. However, there may be management alternatives that increase farm profit at lower stocking rates or flock sizes, due to lower variable costs of production and higher quality product. Such alternatives could have greater synergies with farm conservation and could allow more cost-effective conservation gains.

The management recommendation of stock exclusion from woodland patches for groundstorey plant conservation has synergies with other taxonomic groups. For example, declining woodland bird species are more likely to inhabit large patches of trees (Seddon et al. 2003) and often respond negatively to grazing (Martin & McIntyre 2007). The exclusion of domestic grazing can also increase the abundance of small native mammals (Read & Cunningham 2010). Also, tree regeneration can improve substantially when stock are excluded (Spooner & Briggs 2008).

Increasing the extent of BGGW is the primary management recommendation. However, for the suite of species that persist in them, native pastures provide extensive habitat that ensures large, viable populations that are well connected in the landscape. This is a preferable agricultural matrix to agricultural regions in which the matrix is either crops or pastures dominated by annual cool-season, exotic grasses. Since native pastures that

have not been previously cultivated support a greater number of the region's native plant species, these pastures should not be considered for intensification of pastoral activities, such as cultivation for forage crops.

7.5. Further research

7.5.1. Plant diversity in the wheat–sheep zone

The scope of this research was to identify landscape diversity patterns and evaluate the role of native pastures in the conservation of groundstorey plant diversity on the North-West Slopes of NSW. A major conclusion was that many species are restricted to ungrazed woodland patches that are infrequent in the landscape, indicating that these are spatially arranged in patches in a matrix of unsuitable habitat akin to the island–sea model of island biogeography theory (IBT, MacArthur & Wilson 1967). These species may also form metapopulations and behave as predicted by metapopulation theory (MT, Levins 1969; Hanski & Gilpin 1991). As such, further research questions remain: are the predictions of IBT and MT realised for the suite of species that are restricted to woodland habitats? Is the configuration of these ungrazed woodland patches important for species persistence? Ecological knowledge of the suite of woodland-restricted, grazing-sensitive species that are most likely to represent the extinction debt in this regional ecosystem is also required. Patches that are reproductively connected for some species can be isolated for others. Seed rain studies may reveal the dispersal ability of many woodland-restricted herbaceous species and demonstrate the likely connectivity of patches. Patches of remnant vegetation that have previously been comprehensively surveyed for species incidence may also provide an opportunity for revisitation studies (e.g. Sutton & Morgan 2009) that could reveal colonisation rates of species restricted to such habitats, to test for functional connectivity between fragments.

Research questions regarding the potential to restore native pastures and heavily grazed grassy woodland to high quality grassy woodland habitat are vital to the development of conservation strategies in the wheat–sheep zone. While the protection of high quality patches is a high priority for plant conservation, improving the connectivity of these patches in the landscape may also be vital in ensuring the viability of many species. This will require successful restoration and regeneration of high conservation value habitat, though many questions remain regarding how to achieve this: when grazing is removed, under what circumstances, and over what temporal scales will degraded woodlands provide suitable habitat for grazing-sensitive species? What degree of active management, such as weed control, is needed to ensure positive conservation outcomes? Monitoring the outcomes of existing conservation efforts and research on alternative conservation management techniques is needed, and research in south-eastern Australia has begun to answer some of these questions (Smallbone et al. 2007; Spooner & Briggs 2008; Lindenmayer et al. 2010; McIntyre et al. 2010).

The ecological processes that determine landscape diversity patterns require further research. The landscape methods used in this thesis, such as additive partitioning, species accumulation curves and habitat specificity measures, helped to identify the landscape-scale patterns evident in the study region. The knowledge of such patterns may enable ecologists to more readily identify the processes that are responsible for them (Gering et al. 2003). The present study highlighted landscape-scale patterns and hypothesised which ecological processes caused them. However, these hypotheses require experimental verification. For example, a replicated grassland experiment with a variety of disturbance treatments in large experimental plots (≥ 1 ha) could help to reveal the mechanisms by which grazing increases the homogeneity of grassy vegetation. Such

research could also improve our ability to develop management strategies that successfully maintain heterogeneity, and extend the applicability and theoretical advances made from the observation of broad-scale diversity patterns.

In agreement with Whalley (2005), further research is needed to provide objective evidence of the biodiversity outcomes of alternate grassland management systems, such as cell grazing implemented by some livestock producers (e.g. Wright & Wright 2000). This research must incorporate the issues of spatial scale highlighted in this thesis—local-scale comparisons of species richness between management techniques will be insufficient to substantiate claims of improved conservation value. In particular, the effects of grazing systems such as cell grazing on broad-scale plant diversity patterns requires further research: does providing extra watering points and creating smaller paddock sizes to increase the degree of utilisation of pastures by livestock increase or decrease the homogeneity of native pastures? My results suggest that such management may create low heterogeneity grassland environments with low accumulation of species across broad spatial scales.

A review of grazing impacts (Lunt et al. 2007a) provided a decision tree to predict the effects of livestock grazing on the conservation values of native vegetation. Though broad-scale ecological processes were factored into the decision tree, consideration of the impact of grazing on plant diversity was limited to species richness effects at small spatial scales. Future research should aim to develop a similar tool for predicting the broad-scale impacts of grazing and agricultural intensification on conservation values, including the interactions among grazing management, climate, soil type and fertility. It is likely that many opportunities exist to re-analyse large survey datasets that have previously been used to identify site-scale diversity patterns, to test hypotheses

regarding landscape scale diversity patterns across different land uses. Analytical methods such as habitat specificity indices (including GDC), additive partitioning and sample-based rarefaction (such as Mao Tao and Coleman) can potentially be applied to existing survey datasets that have been well-stratified across environmental gradients of interest to reveal important information for biodiversity conservation that has not yet been revealed by site-scale analyses.

7.6. Conclusions

Biodiversity conservation in agricultural landscapes may face many political and socioeconomic challenges; however, robust ecological research is a prerequisite for designing sustainable agricultural landscapes, influencing land use policy and management decisions, and ultimately avoiding species losses. This research has provided new insight into the conservation of plant diversity in farmland on the North-West Slopes of NSW, in particular regarding the homogeneous composition of native pastures across broad spatial scales, and the importance of observing management and land use influences at multiple spatial scales. The study challenges ecologists and conservation biologists to strive to understand the ecological mechanisms and interactions underlying the diversity patterns of different groups of taxa in agricultural landscapes around the world.

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APPENDIX 1

Species list for all taxa represented in metadataset

Origin (O): N = native; E = exotic. **Life form (LF):** C = climber or creeper; F = forb; G = grass; Gl = grass-like; LS = low shrub; P = parasite, Pt = pteridophyte; TS = tall shrub; T = tree. **Life cycle (LC):** AN = annual; PE = perennial; PA = perennial bulb with annual shoots; SP = short-lived perennial.

Species	O	LF	LC	Species	O	LF	LC
PTERIDOPHYTES				CYCAD			
ADIANTACEAE				ZAMIACEAE			
<i>Adiantum atroviride</i>	N	Pt	PE	<i>Macrozamia concinna</i>	N	TS	PE
<i>Adiantum formosum</i>	N	Pt	PE	<i>Macrozamia glaucophylla</i>	N	TS	PE
<i>Adiantum hispidulum</i>	N	Pt	PE	<i>Macrozamia heteromera</i>	N	TS	PE
<i>Cheilanthes austrotenuifolia</i>	N	Pt	PE	<i>Macrozamia stenomera</i>	N	LS	PE
<i>Cheilanthes distans</i>	N	Pt	PE	CONIFER			
<i>Cheilanthes sieberi</i>	N	Pt	PE	CUPRESSACEAE			
<i>Pellaea calidirupium</i>	N	Pt	PE	<i>Callitris columellaris</i>	N	T	PE
<i>Pellaea falcata</i>	N	Pt	PE	<i>Callitris endlicheri</i>	N	T	PE
<i>Pellaea nana</i>	N	Pt	PE	MONOCOTYLEDON			
<i>Pellaea paradoxa</i>	N	Pt	PE	ALLIACEAE			
ASPLENIACEAE				<i>Nothoscordum borbonicum</i>	E	GL	PA
<i>Asplenium flabellifolium</i>	N	Pt	PE	AMARYLLIDACEAE			
<i>Asplenium trichomanes</i>	N	Pt	PE	<i>Crinum flaccidum</i>	N	GL	PA
<i>Pleurosorus subglandulosus</i>	N	Pt	PE	ANTHERICACEAE			
AZOLLACEAE				<i>Arthropodium fimbriatum</i>	N	Gl	PA
<i>Azolla filiculoides</i>	N	Pt	PE	<i>Arthropodium milleflorum</i>	N	Gl	PA
BLECHNACEAE				<i>Arthropodium sp. A</i>	N	Gl	PA
<i>Blechnum cartilagineum</i>	N	Pt	PE	<i>Arthropodium sp. B</i>	N	Gl	PA
<i>Blechnum minus</i>	N	Pt	PE	<i>Arthropodium strictum</i>	N	Gl	PA
<i>Blechnum nudum</i>	N	Pt	PE	<i>Caesia parviflora</i>	N	Gl	PA
<i>Doodia aspera</i>	N	Pt	PE	<i>Laxmannia compacta</i>	N	Gl	PA
<i>Doodia australis</i>	N	Pt	PE	<i>Laxmannia gracilis</i>	N	Gl	PA
<i>Doodia caudata</i>	N	Pt	PE	<i>Thysanotus tuberosus</i>	N	GL	PA
CYATHEACEAE				<i>Tricoryne elatior</i>	N	GL	PA
<i>Cyathea australis</i>	N	Pt	PE	ASPARAGACEAE			
DENNSTAEDTIACEAE				<i>Asparagus aethiopicus</i>	E	C	PA
<i>Histiopteris incisa</i>	N	Pt	PE	<i>Asparagus asparagoides</i>	E	C	PA
<i>Hypolepis glandulifera</i>	N	Pt	PE	<i>Asparagus officinalis</i>	E	F	PA
<i>Pteridium esculentum</i>	N	Pt	PE	ASPHODELACEAE			
DICKSONIACEAE				<i>Asphodelus fistulosus</i>	E	Gl	SP
<i>Calochlaena dubia</i>	N	Pt	PE	<i>Bulbine bulbosa</i>	N	Gl	PE
<i>Dicksonia antarctica</i>	N	Pt	PE	<i>Bulbine semibarbata</i>	N	GL	AN
DRYOPTERIDACEAE				COLCHICACEAE			
<i>Lastreopsis decomposita</i>	N	Pt	PE	<i>Wurmbea biglandulosa</i>	N	GL	PA
<i>Lastreopsis microsora subsp. microsora</i>	N	Pt	PE	<i>Wurmbea dioica subsp. dioica</i>	N	GL	PA
<i>Polystichum fallax</i>	N	Pt	PE	COMMELINACEAE			
MARSILEACEAE				<i>Commelina cyanea</i>	N	F	PE
<i>Marsilea drummondii</i>	N	Pt	AN	<i>Murdannia graminea</i>	N	GL	PA
<i>Marsilea hirsuta</i>	N	Pt	AN	<i>Tradescantia fluminensis</i>	E	F	PE
OPHIOGLOSSACEAE				CRASSULACEAE			
<i>Ophioglossum lusitanicum</i>	N	Pt	PE	<i>Kalanchoe sexangularis</i>	E	LS	PE
POLYPODIACEAE				CYPERACEAE			
<i>Pyrrosia confluens var. confluens</i>	N	Pt	PE	<i>Bulbostylis densa</i>	N	GL	AN
<i>Pyrrosia rupestris</i>	N	Pt	PE	<i>Carex appressa</i>	N	GL	PE
PTERIDACEAE				<i>Carex breviculmis</i>	N	GL	PE
<i>Pteris tremula</i>	N	Pt	PE	<i>Carex gaudichaudiana</i>	N	GL	PE
THELYPTERIDACEAE				<i>Carex inomitata</i>	N	GL	PE
<i>Christella dentata</i>	N	Pt	PE	<i>Carex inversa</i>	N	GL	PE
				<i>Carex longibrachiata</i>	N	GL	PE
				<i>Caustis flexuosa</i>	N	GL	PE

Species	O	LF	LC	Species	O	LF	LC
<i>Cyperus bifax</i>	N	GL	PE	ORCHIDACEAE			
<i>Cyperus brevifolius</i>	E	GL	PE	<i>Caladenia capillata</i>	N	GL	PA
<i>Cyperus difformis</i>	N	GL	AN	<i>Caladenia carnea</i>	N	GL	PA
<i>Cyperus eragrostis</i>	E	GL	PE	<i>Caladenia fuscata</i>	N	GL	PA
<i>Cyperus exaltatus</i>	N	GL	PE	<i>Caladenia gracilis</i>	N	GL	PA
<i>Cyperus fulvus</i>	N	GL	PE	<i>Calochilus campestris</i>	N	GL	PA
<i>Cyperus gracilis</i>	N	GL	PE	<i>Calochilus robertsonii</i>	N	GL	PA
<i>Cyperus gunnii</i> subsp. <i>gunnii</i>	N	GL	PE	<i>Cryptostylis leptochila</i>	N	GL	PA
<i>Cyperus lucidus</i>	N	GL	PE	<i>Cyanicula caerulea</i>	N	GL	PA
<i>Cyperus papyrus</i>	E	GL	PE	<i>Cymbidium canaliculatum</i>	N	GL	PA
<i>Cyperus sanguinolentus</i>	N	GL	SP	<i>Cyrtostylis reniformis</i>	N	GL	PA
<i>Cyperus sphaeroides</i>	N	GL	PE	<i>Dipodium punctatum</i>	N	GL	PA
<i>Cyperus subulatus</i>	N	GL	PE	<i>Dipodium roseum</i>	N	GL	PA
<i>Cyperus vaginatus</i>	N	GL	PE	<i>Dipodium variegatum</i>	N	GL	PA
<i>Eleocharis acuta</i>	N	GL	PE	<i>Eriochilus cucullatus</i>	N	GL	PA
<i>Eleocharis cylindrostachys</i>	N	GL	PE	<i>Microtis parviflora</i>	N	GL	PA
<i>Eleocharis pallens</i>	N	GL	PE	<i>Microtis unifolia</i>	N	GL	PA
<i>Eleocharis plana</i>	N	GL	PE	<i>Orthoceras strictum</i>	N	GL	PA
<i>Eleocharis pusilla</i>	N	GL	PE	<i>Pterostylis arenicola</i>	N	GL	PA
<i>Fimbristylis dichotoma</i>	N	GL	PE	<i>Pterostylis coccinea</i>	N	GL	PA
<i>Gahnia aspera</i>	N	GL	PE	<i>Pterostylis curta</i>	N	GL	PA
<i>Isolepis habra</i>	N	GL	PE	<i>Pterostylis longicurva</i>	N	GL	PA
<i>Isolepis inundata</i>	N	GL	PE	<i>Pterostylis mutica</i>	N	GL	PA
<i>Isolepis subtilissima</i>	N	GL	PE	<i>Pterostylis parviflora</i>	N	GL	PA
<i>Lepidosperma laterale</i>	N	GL	PE	<i>Pterostylis praetermissa</i>	N	GL	PA
<i>Schoenoplectus mucronatus</i>	N	GL	PE	<i>Pterostylis setifera</i>	N	GL	PA
<i>Schoenoplectus validus</i>	N	GL	PE	<i>Pterostylis species B</i>	N	GL	PA
<i>Schoenus apogon</i>	N	GL	PE	PHORMIACEAE			
<i>Scirpus polystachyus</i>	N	GL	PE	<i>Dianella 'admixta'</i>	N	GL	PE
<i>Scleria mackaviensis</i>	N	GL	PE	<i>Dianella caerulea</i> var. <i>caerulea</i>	N	GL	PE
HYDROCHARITACEAE				<i>Dianella caerulea</i> var. <i>cinerascens</i>	N	GL	PE
<i>Vallisneria nana</i>	N	GI	AN	<i>Dianella longifolia</i> var. <i>longifolia</i>	N	GL	PE
HYPOXIDACEAE				<i>Dianella longifolia</i> var. <i>stenophylla</i>	N	GL	PE
<i>Hypoxis exilis</i>	N	F	PE	<i>Dianella revoluta</i> var. <i>revoluta</i>	N	GL	PE
<i>Hypoxis hygrometrica</i> var. <i>villosisepala</i>	N	GL	PA	<i>Dianella revoluta</i> var. <i>vinosa</i>	N	GL	PE
IRIDACEAE				<i>Stypandra glauca</i>	N	GL	PE
<i>Libertia paniculata</i>	N	GL	PE	POACEAE			
<i>Patersonia sericea</i>	N	GL	PE	<i>Aira cupaniana</i>	E	G	AN
JUNCACEAE				<i>Amphibromus nervosus</i>	N	G	PE
<i>Juncus articulatus</i>	E	GL	PE	<i>Ancistrachne uncinulata</i>	N	G	PE
<i>Juncus australis</i>	N	GL	PE	<i>Anthoxanthum odoratum</i>	E	G	PE
<i>Juncus bufonius</i>	E	GL	AN	<i>Aristida acuta</i>	N	G	PE
<i>Juncus continuus</i>	N	GL	PE	<i>Aristida benthamii</i>	N	G	PE
<i>Juncus filicaulis</i>	N	GL	PE	<i>Aristida calycina</i> var. <i>calycina</i>	N	G	PE
<i>Juncus firmus</i>	N	GL	PE	<i>Aristida calycina</i> var. <i>praealta</i>	N	G	PE
<i>Juncus fockei</i>	N	GL	PE	<i>Aristida caput-medusae</i>	N	G	PE
<i>Juncus homalocaulis</i>	N	GL	PE	<i>Aristida echinata</i>	N	G	PE
<i>Juncus ochrocoleus</i>	N	GL	PE	<i>Aristida gracilipes</i>	N	G	PE
<i>Juncus pauciflorus</i>	N	GL	PE	<i>Aristida jerichoensis</i> var. <i>jerichoensis</i>	N	G	PE
<i>Juncus planifolius</i>	N	GL	AN	<i>Aristida jerichoensis</i> var. <i>subspinulifera</i>	N	G	PE
<i>Juncus prismatocarpus</i>	N	GL	PE	<i>Aristida latifolia</i>	N	G	PE
<i>Juncus radula</i>	N	GL	PE	<i>Aristida leichhardtiana</i>	N	G	PE
<i>Juncus remotiflorus</i>	N	GL	PE	<i>Aristida leptopoda</i>	N	G	PE
<i>Juncus sarophorus</i>	N	GL	PE	<i>Aristida personata</i>	N	G	PE
<i>Juncus subsecundus</i>	N	GL	PE	<i>Aristida psammophila</i>	N	G	PE
<i>Juncus usitatus</i>	N	GL	PE	<i>Aristida queenslandica</i> var. <i>queenslandica</i>	N	G	PE
<i>Juncus vaginatus</i>	N	GL	PE	<i>Aristida ramosa</i>	N	G	PE
<i>Luzula densiflora</i>	N	GL	PE	<i>Aristida vagans</i>	N	G	PE
<i>Luzula flaccida</i>	N	GL	PE	<i>Arundinella nepalensis</i>	N	G	PE
<i>Luzula meridionalis</i>	N	GL	PE	<i>Austroanthonia bipartita</i>	N	G	PE
LOMANDRACEAE				<i>Austroanthonia caespitosa</i>	N	G	PE
<i>Lomandra confertifolia</i>	N	GL	PE	<i>Austroanthonia eriantha</i>	N	G	PE
LUZURIAGACEAE				<i>Austroanthonia fulva</i>	N	G	PE
<i>Eustrephus latifolius</i>	N	C	PE	<i>Austroanthonia laevis</i>	N	G	PE
<i>Geitonoplesium cymosum</i>	N	C	PE	<i>Austroanthonia monticola</i>	N	G	PE
ORCHIDACEAE				<i>Austroanthonia penicillata</i>	N	G	PE
<i>Acianthus collinus</i>	N	GL	PA				

Species	O	LF	LC	Species	O	LF	LC
POACEAE				POACEAE			
<i>Austrodanthonia pilosa</i>	N	G	PE	<i>Digitaria diffusa</i>	N	G	PE
<i>Austrodanthonia racemosa</i> var. <i>obtusata</i>	N	G	PE	<i>Digitaria divaricatissima</i>	N	G	PE
<i>Austrodanthonia racemosa</i> var. <i>racemosa</i>	N	G	PE	<i>Digitaria hubbardii</i>	N	G	PE
<i>Austrodanthonia richardsonii</i>	N	G	PE	<i>Digitaria hystrichoides</i>	N	G	PE
<i>Austrodanthonia setacea</i>	N	G	PE	<i>Digitaria parviflora</i>	N	G	PE
<i>Austrodanthonia tenuior</i>	N	G	PE	<i>Digitaria ramularis</i>	N	G	PE
<i>Austrostipa aristiglumis</i>	N	G	PE	<i>Echinochloa colona</i>	N	G	AN
<i>Austrostipa densiflora</i>	N	G	PE	<i>Echinochloa crusgalli</i>	E	G	AN
<i>Austrostipa nitida</i>	N	G	PE	<i>Echinopogon caespitosus</i>	N	G	PE
<i>Austrostipa nodosa</i>	N	G	PE	<i>Echinopogon cheelii</i>	N	G	PE
<i>Austrostipa ramosissima</i>	N	G	PE	<i>Echinopogon intermedius</i>	N	G	PE
<i>Austrostipa rudis</i> subsp. <i>nervosa</i>	N	G	PE	<i>Echinopogon mckiei</i>	N	G	PE
<i>Austrostipa rudis</i> subsp. <i>rudis</i>	N	G	PE	<i>Echinopogon nutans</i>	N	G	PE
<i>Austrostipa scabra</i> subsp. <i>falcata</i>	N	G	PE	<i>Echinopogon ovatus</i>	N	G	PE
<i>Austrostipa scabra</i> subsp. <i>scabra</i>	N	G	PE	<i>Eleusine tristachya</i>	E	G	SP
<i>Austrostipa setacea</i>	N	G	PE	<i>Elymus scaber</i> var. <i>plurinervis</i>	N	G	PE
<i>Austrostipa tuckeri</i>	N	G	PE	<i>Elymus scaber</i> var. <i>scaber</i>	N	G	PE
<i>Austrostipa verticillata</i>	N	G	PE	<i>Enneapogon gracilis</i>	N	G	PE
<i>Avena barbata</i>	E	G	AN	<i>Enneapogon lindleyanus</i>	N	G	PE
<i>Avena fatua</i>	E	G	AN	<i>Enneapogon nigricans</i>	N	G	PE
<i>Avena sterilis</i> subsp. <i>ludoviciana</i>	E	G	AN	<i>Enteropogon acicularis</i>	N	G	PE
<i>Axonopus fissifolius</i>	E	G	PE	<i>Enteropogon ramosus</i>	N	G	PE
<i>Bothriochloa biloba</i>	N	G	PE	<i>Entolasia stricta</i>	N	G	PE
<i>Bothriochloa bladhii</i> subsp. <i>bladhii</i>	N	G	PE	<i>Eragrostis benthamii</i>	N	G	PE
<i>Bothriochloa decipiens</i>	N	G	PE	<i>Eragrostis brownii</i>	N	G	PE
<i>Bothriochloa ewartiana</i>	N	G	PE	<i>Eragrostis cilianensis</i>	E	G	AN
<i>Bothriochloa macra</i>	N	G	PE	<i>Eragrostis curvula</i>	E	G	PE
<i>Briza maxima</i>	E	G	AN	<i>Eragrostis elongata</i>	N	G	SP
<i>Briza minor</i>	E	G	AN	<i>Eragrostis lacunaria</i>	N	G	PE
<i>Bromus alopecuroides</i>	E	G	AN	<i>Eragrostis leptocarpa</i>	N	G	PE
<i>Bromus brevis</i>	E	G	AN	<i>Eragrostis leptostachya</i>	N	G	PE
<i>Bromus catharticus</i>	E	G	SP	<i>Eragrostis megalosperma</i>	N	G	PE
<i>Bromus diandrus</i>	E	G	AN	<i>Eragrostis molybdea</i>	N	G	PE
<i>Bromus hordeaceus</i>	E	G	AN	<i>Eragrostis parviflora</i>	N	G	AN
<i>Bromus secalinus</i>	E	G	AN	<i>Eragrostis trachycarpa</i>	N	G	PE
<i>Bromus tectorum</i>	E	G	AN	<i>Eriochloa australiensis</i>	N	G	AN
<i>Capillipedium parviflorum</i>	N	G	PE	<i>Eriochloa crebra</i>	N	G	PE
<i>Capillipedium spicigerum</i>	N	G	PE	<i>Eriochloa procera</i>	N	G	SP
<i>Catapodium rigidum</i>	E	G	AN	<i>Eriochloa pseudoacrotricha</i>	N	G	SP
<i>Cenchrus caliculatus</i>	N	G	PE	<i>Eulalia aurea</i>	N	G	PE
<i>Chloris gayana</i>	E	G	PE	<i>Festuca asperula</i>	N	G	PE
<i>Chloris truncata</i>	N	G	PE	<i>Festuca elatior</i>	E	G	PE
<i>Chloris ventricosa</i>	N	G	PE	<i>Heteropogon contortus</i>	N	G	PE
<i>Chloris virgata</i>	E	G	SP	<i>Holcus lanatus</i>	E	G	PE
<i>Cleistochloa rigida</i>	N	G	PE	<i>Homopholis belsonii</i>	N	G	PE
<i>Cymbopogon obtectus</i>	N	G	PE	<i>Homopholis proluta</i>	N	G	PE
<i>Cymbopogon refractus</i>	N	G	PE	<i>Hordeum geniculatum</i>	E	G	AN
<i>Cynodon dactylon</i>	N	G	PE	<i>Hordeum glaucum</i>	E	G	AN
<i>Cynodon incompletus</i>	N	G	PE	<i>Hordeum leporinum</i>	E	G	AN
<i>Dactylis glomerata</i>	E	G	PE	<i>Hyparrhenia hirta</i>	E	G	PE
<i>Dactyloctenium aegyptium</i>	N	G	PE	<i>Imperata cylindrica</i> var. <i>major</i>	N	G	PE
<i>Dactyloctenium radulans</i>	N	G	PE	<i>Joycea pallida</i>	N	G	PE
<i>Desmazeria rigida</i>	N	G	PE	<i>Lachnagrostis filiformis</i>	N	G	AN
<i>Deyeuxia gunniana</i>	N	G	PE	<i>Lamarckia aurea</i>	E	G	AN
<i>Dichanthium sericeum</i>	N	G	PE	<i>Leptochloa decipens</i> subsp. <i>decipens</i>	N	G	PE
<i>Dichanthium setosum</i>	N	G	PE	<i>Leptochloa divaricatissima</i>	N	G	PE
<i>Dichelachne crinita</i>	N	G	PE	<i>Lolium perenne</i>	E	G	SP
<i>Dichelachne inaequiglumis</i>	N	G	PE	<i>Lolium rigidum</i>	E	G	AN
<i>Dichelachne micrantha</i>	N	G	PE	<i>Microlaena stipoides</i>	N	G	PE
<i>Dichelachne rara</i>	N	G	PE	<i>Notodanthonia longifolia</i>	N	G	PE
<i>Dichelachne sieberiana</i>	N	G	PE	<i>Oplismenus aemulus</i>	N	G	PE
<i>Digitaria ammophila</i>	N	G	PE	<i>Oplismenus imbecillis</i>	N	G	PE
<i>Digitaria breviglumis</i>	N	G	PE	<i>Panicum capillare</i> var. <i>capillare</i>	E	G	AN
<i>Digitaria brownii</i>	N	G	PE	<i>Panicum decompositum</i>	N	G	PE
<i>Digitaria ciliaris</i>	E	G	AN	<i>Panicum effusum</i>	N	G	PE
				<i>Panicum gilvum</i>	E	G	AN
				<i>Panicum laevinode</i>	N	G	PE

Species	O	LF	LC	Species	O	LF	LC
POACEAE				XANTHORRHOEACEAE			
<i>Panicum miliaceum</i>	E	G	AN	<i>Lomandra longifolia</i>	N	GL	PE
<i>Panicum paludosum</i>	N	G	PE	<i>Lomandra multiflora</i> subsp. <i>multiflora</i>	N	GL	PE
<i>Panicum queenslandicum</i> var. <i>acuminatum</i>	N	G	PE	<i>Lomandra patens</i>	N	GL	PE
<i>Panicum queenslandicum</i> var. <i>queenslandicum</i>	N	G	PE	<i>Xanthorrhoea acaulis</i>	N	TS	PE
<i>Panicum simile</i>	N	G	PE	<i>Xanthorrhoea glauca</i> subsp. <i>angustifolia</i>	N	TS	PE
<i>Paspalidium aversum</i>	N	G	PE	<i>Xanthorrhoea glauca</i> subsp. <i>glauca</i>	N	TS	PE
<i>Paspalidium gausum</i>	N	G	PE	<i>Xanthorrhoea johnsonii</i>	N	TS	PE
<i>Paspalidium gracile</i>	N	G	PE	DICOTYLEDON			
<i>Paspalum dilatatum</i>	E	G	PE	ACANTHACEAE			
<i>Paspalum distichum</i>	N	G	PE	<i>Brunoniella australis</i>	N	F	PE
<i>Pennisetum alopecuroides</i>	N	G	PE	<i>Brunoniella pumilio</i>	N	F	PE
<i>Pennisetum clandestinum</i>	E	G	PE	<i>Pseuderanthemum variabile</i>	N	F	PE
<i>Phalaris aquatica</i>	E	G	PE	<i>Rostellularia adscendens</i> subsp. <i>adscendens</i>	N	F	PE
<i>Phalaris minor</i>	E	G	AN	AIZOACEAE			
<i>Phalaris paradoxa</i>	E	G	AN	<i>Glinus lotoides</i>	N	F	AN
<i>Phragmites australis</i>	N	G	PE	<i>Tetragonia tetragonioides</i>	N	F	AN
<i>Poa annua</i>	E	G	AN	<i>Zaleya galericulata</i>	N	F	SP
<i>Poa fax</i>	N	G	AN	ALISMATACEAE			
<i>Poa labillardierei</i> var. <i>labillardierei</i>	N	G	PE	<i>Damasonium minus</i>	N	F	SP
<i>Poa queenslandica</i>	N	G	PE	AMARANTHACEAE			
<i>Poa sieberiana</i> var. <i>hirtella</i>	N	G	PE	<i>Alternanthera denticulata</i>	N	F	AN
<i>Poa sieberiana</i> var. <i>sieberiana</i>	N	G	PE	<i>Alternanthera pungens</i>	E	F	SP
<i>Polypogon monspeliensis</i>	E	G	AN	<i>Alternanthera species A</i>	N	F	PE
<i>Rostraria cristata</i>	E	G	AN	<i>Amaranthus hybridus</i>	E	F	AN
<i>Rytidosperma vickeryae</i>	N	G	PE	<i>Amaranthus macrocarpus</i> var. <i>macrocarpus</i>	N	F	AN
<i>Sarga leiocladum</i>	E	G	PE	<i>Amaranthus mitchellii</i>	N	F	AN
<i>Secale cereale</i>	E	G	AN	<i>Amaranthus powellii</i>	E	F	AN
<i>Setaria constricta</i>	E	G	PE	<i>Amaranthus viridis</i>	E	F	AN
<i>Setaria criniforme</i>	E	G	PE	<i>Gomphrena celosioides</i>	N	F	AN
<i>Setaria distans</i>	E	G	PE	<i>Nyssanthes diffusa</i>	N	F	SP
<i>Setaria gracilis</i>	E	G	PE	<i>Nyssanthes erecta</i>	N	F	AN
<i>Setaria jubiflorum</i>	E	G	PE	AMYGDALACEAE			
<i>Setaria paspalidioides</i>	E	G	PE	<i>Prunus cerasus</i>	E	T	PE
<i>Setaria pumila</i>	E	G	AN	ANACARDIACEAE			
<i>Setaria verticillata</i>	E	G	AN	<i>Schinus molle</i> var. <i>areira</i>	E	T	PE
<i>Sorghum bicolor</i>	E	G	SP	APIACEAE			
<i>Sorghum halepense</i>	E	G	PE	<i>Actinotus helianthi</i>	N	F	AN
<i>Sporobolus caroli</i>	N	G	SP	<i>Ammi majus</i>	E	F	SP
<i>Sporobolus creber</i>	N	G	PE	<i>Centella asiatica</i>	N	F	PE
<i>Sporobolus elongatus</i>	N	G	PE	<i>Conium maculatum</i>	E	F	SP
<i>Themeda avenacea</i>	N	G	PE	<i>Cyclospermum leptophyllum</i>	E	F	AN
<i>Themeda triandra</i>	N	G	PE	<i>Daucus carota</i>	E	F	AN
<i>Tragopogon porrifolius</i>	E	G	SP	<i>Daucus glochidiatus</i>	N	F	AN
<i>Tragus australianus</i>	N	G	AN	<i>Eryngium ovinum</i>	N	F	SP
<i>Triodia mitchellii</i>	N	G	PE	<i>Foeniculum vulgare</i>	E	F	SP
<i>Triodia scariosa</i>	N	G	PE	<i>Hydrocotyle acutiloba</i>	E	F	PE
<i>Tripogon loliiformis</i>	N	G	SP	<i>Hydrocotyle laxiflora</i>	N	F	PE
<i>Urochloa advena</i>	E	G	PE	<i>Hydrocotyle peduncularis</i>	N	F	PE
<i>Urochloa panicoides</i>	E	G	AN	<i>Hydrocotyle tripartita</i>	N	F	PE
<i>Vulpia bromoides</i>	E	G	AN	<i>Oreomyrrhis ciliata</i>	N	F	PE
<i>Vulpia muralis</i>	E	G	AN	<i>Oreomyrrhis eriopoda</i>	N	F	PE
<i>Vulpia myuros</i> forma <i>megalura</i>	E	G	AN	<i>Platysace lanceolata</i>	N	LS	PE
<i>Walwhalleya subxerophila</i>	N	G	PE	<i>Trachymene incisa</i>	N	F	PE
RESTIONACEAE				<i>Xanthosia spp.</i>	N	TS	PE
<i>Baloskion stenocoleum</i>	N	GL	PE	APOCYNACEAE			
STYLIDIACEAE				<i>Alstonia constricta</i>	N	TS	PE
<i>Stylidium graminifolium</i>	N	GL	PE	<i>Carissa ovata</i>	N	LS	PE
TYPHACEAE				<i>Parsonsia eucalyptophylla</i>	N	C	PE
<i>Typha domingensis</i>	N	GL	PE	<i>Parsonsia lanceolata</i>	N	C	PE
<i>Typha orientalis</i>	N	GL	PE	ARALIACEAE			
XANTHORRHOEACEAE				<i>Astrotricha longifolia</i>	N	TS	PE
<i>Lomandra filiformis</i> subsp. <i>coriacea</i>	N	GL	PE	<i>Hedera helix</i>	E	C	PE
<i>Lomandra filiformis</i> subsp. <i>filiformis</i>	N	GL	PE	<i>Polyscias sambucifolia</i>	N	TS	PE
<i>Lomandra filiformis</i> subsp. <i>flavior</i>	N	GL	PE				
<i>Lomandra leucocephala</i>	N	GL	PE				

Species	O	LF	LC	Species	O	LF	LC
ASCLEPIADACEAE				ASTERACEAE			
<i>Gomphocarpus fruticosus</i>	E	F	PE	<i>Eclipta platyglossa</i>	N	F	AN
<i>Marsdenia viridiflora</i> subsp. <i>viridiflora</i>	N	C	PE	<i>Euchiton gymnocephalus</i>	N	F	PE
<i>Oxyptalum coeruleum</i>	E	LS	PE	<i>Euchiton involucratus</i>	N	F	PE
<i>Rhyncharrhena linearis</i>	N	C	PE	<i>Euchiton sphaericus</i>	N	F	SP
<i>Tylophora paniculata</i>	N	C	PE	<i>Gamochoaeta americana</i>	E	F	AN
ASTERACEAE				<i>Gamochoaeta calviceps</i>	E	F	AN
<i>Actinobole uliginosum</i>	N	F	AN	<i>Gamochoaeta spicata</i>	E	F	PE
<i>Ammobium alatum</i>	N	F	PE	<i>Glossogyne tannensis</i>	N	F	PE
<i>Arctotheca calendula</i>	E	F	AN	<i>Hedypnois rhagadioloides</i> subsp. <i>cretica</i>	E	F	AN
<i>Aster subulatus</i>	E	F	SP	<i>Helichrysum scorpioides</i>	N	F	PE
<i>Bidens pilosa</i>	E	F	AN	<i>Hypochaeris glabra</i>	E	F	AN
<i>Bidens subalternans</i>	E	F	AN	<i>Hypochaeris microcephala</i> var. <i>albiflora</i>	E	F	PE
<i>Brachyscome aculeata</i>	N	F	PE	<i>Hypochaeris radicata</i>	E	F	PE
<i>Brachyscome angustifolia</i> var. <i>angustifolia</i>	N	F	PE	<i>Ixiolaena leptolepis</i>	N	F	PE
<i>Brachyscome angustifolia</i> var. <i>heterophylla</i>	N	F	PE	<i>Ixiolaena tomentosa</i>	N	F	PE
<i>Brachyscome ciliaris</i>	N	F	PE	<i>Lactuca saligna</i>	E	F	SP
<i>Brachyscome curvicaarpa</i>	N	F	AN	<i>Lactuca serriola</i>	E	F	SP
<i>Brachyscome dentata</i>	N	F	PE	<i>Lagenophora gracilis</i>	N	F	PE
<i>Brachyscome dissectifolia</i>	N	F	PE	<i>Lagenophora stipitata</i>	N	F	PE
<i>Brachyscome diversifolia</i> var. <i>dissecta</i>	N	F	PE	<i>Leptorhynchus panaetioides</i>	N	F	PE
<i>Brachyscome gracilis</i>	N	F	AN	<i>Leptorhynchus squamatus</i>	N	F	PE
<i>Brachyscome melanocarpa</i>	N	F	PE	<i>Leucochrysum albicans</i>	N	F	PE
<i>Brachyscome microcarpa</i>	N	F	PE	<i>Microseris lanceolata</i>	N	F	PA
<i>Brachyscome multifida</i>	N	F	AN	<i>Minuria leptophylla</i>	N	F	PE
<i>Brachyscome nova-anglica</i>	N	F	AN	<i>Minuria scoparia</i>	N	F	PE
<i>Brachyscome procumbens</i>	N	F	PE	<i>Olearia canescens</i>	N	LS	PE
<i>Brachyscome smithwhitei</i>	N	F	PE	<i>Olearia elliptica</i>	N	LS	PE
<i>Brachyscome spathulata</i>	N	F	PE	<i>Olearia microphylla</i>	N	LS	PE
<i>Brachyscome stuartii</i>	N	F	PE	<i>Olearia myrsinoides</i>	N	LS	PE
<i>Calocephalus citreus</i>	N	F	PE	<i>Olearia ramosissima</i>	N	LS	PE
<i>Calotis anthemoides</i>	N	F	PE	<i>Olearia ramulosa</i>	N	LS	PE
<i>Calotis cuneifolia</i>	N	F	PE	<i>Olearia rosmarinifolia</i>	N	LS	PE
<i>Calotis dentex</i>	N	F	PE	<i>Olearia viscidula</i>	N	LS	PE
<i>Calotis hispidula</i>	N	F	AN	<i>Ozothamnus adnatus</i>	N	LS	PE
<i>Calotis lappulacea</i>	N	F	PE	<i>Ozothamnus bidwillii</i>	N	LS	PE
<i>Carduus tenuiflorus</i>	E	F	AN	<i>Ozothamnus diosmifolius</i>	N	LS	PE
<i>Carthamus lanatus</i>	E	F	AN	<i>Ozothamnus obcordatus</i>	N	LS	PE
<i>Cassinia arcuata</i>	N	LS	PE	<i>Picris angustifolia</i> subsp. <i>carolorum-</i> <i>henricorum</i>	N	F	AN
<i>Cassinia compacta</i>	N	LS	PE	<i>Picris hieracioides</i>	N	F	AN
<i>Cassinia laevis</i>	N	LS	PE	<i>Podolepis jaceoides</i>	N	F	PE
<i>Cassinia leptocephala</i>	N	LS	PE	<i>Podolepis neglecta</i>	N	F	PE
<i>Cassinia quinquefaria</i>	N	LS	PE	<i>Pseudognaphalium luteo-album</i>	N	F	AN
<i>Cassinia species B</i>	N	LS	PE	<i>Pycnosorus globosus</i>	N	F	PE
<i>Cassinia species D</i>	N	LS	PE	<i>Rhodanthe anthemoides</i>	N	F	PE
<i>Cassinia tegulata</i>	N	LS	PE	<i>Schkuhria pinnata</i> var. <i>abrotanoides</i>	E	F	AN
<i>Cassinia theodori</i>	N	LS	PE	<i>Senecio australis</i>	N	F	PE
<i>Centaurea calcitrapa</i>	E	F	SP	<i>Senecio bathurstianus</i>	N	F	PE
<i>Centaurea melitensis</i>	E	F	AN	<i>Senecio daltonii</i>	N	F	PE
<i>Centaurea solstitialis</i>	E	F	SP	<i>Senecio diaschides</i>	N	F	PE
<i>Centipeda minima</i> var. <i>minima</i>	N	F	AN	<i>Senecio gunnii</i>	N	F	PE
<i>Chondrilla juncea</i>	E	F	PE	<i>Senecio hispidulus</i>	N	F	PE
<i>Chrysocephalum apiculatum</i>	N	F	PE	<i>Senecio macranthus</i>	N	F	PE
<i>Chrysocephalum semipapposum</i>	N	F	PE	<i>Senecio madagascariensis</i>	E	F	SP
<i>Cichorium intybus</i>	E	F	PE	<i>Senecio minimus</i>	N	F	PE
<i>Cirsium vulgare</i>	E	F	SP	<i>Senecio prenanthoides</i>	N	F	PE
<i>Conyza bonariensis</i>	E	F	AN	<i>Senecio quadridentatus</i>	N	F	PE
<i>Conyza parva</i>	E	F	AN	<i>Senecio spanomerus</i>	N	F	PE
<i>Conyza sumatrensis</i>	E	F	AN	<i>Senecio tenuiflorus</i>	N	F	PE
<i>Cosmos bipinnatus</i>	E	F	AN	<i>Sigesbeckia australiensis</i>	N	F	AN
<i>Craspedia canens</i>	N	F	SP	<i>Sigesbeckia orientalis</i> subsp. <i>orientalis</i>	N	F	AN
<i>Craspedia variabilis</i>	N	F	SP	<i>Silybum marianum</i>	E	F	SP
<i>Crepis capillaris</i>	E	F	SP	<i>Solenogyne bellioides</i>	N	F	PE
<i>Cymbonotus lawsonianus</i>	N	F	PE	<i>Solenogyne dominii</i>	N	F	PE

Species	O	LF	LC	Species	O	LF	LC
ASTERACEAE				CAMPANULACEAE			
<i>Solenogyne gunnii</i>	N	F	PE	<i>Wahlenbergia gracilis</i>	N	F	PE
<i>Soliva anthemifolia</i>	E	F	AN	<i>Wahlenbergia graniticola</i>	N	F	PE
<i>Sonchus asper</i> subsp. <i>glaucescens</i>	E	F	AN	<i>Wahlenbergia luteola</i>	N	F	PE
<i>Sonchus oleraceus</i>	E	F	AN	<i>Wahlenbergia planiflora</i> subsp. <i>longipila</i>	N	F	PE
<i>Stuartina hamata</i>	N	F	AN	<i>Wahlenbergia planiflora</i> subsp. <i>planiflora</i>	N	F	PE
<i>Tagetes minuta</i>	E	F	AN	<i>Wahlenbergia stricta</i> subsp. <i>alterna</i>	N	F	PE
<i>Taraxacum officinale</i>	E	F	PE	<i>Wahlenbergia stricta</i> subsp. <i>stricta</i>	N	F	PE
<i>Triptilodiscus pygmaeus</i>	N	F	AN	CANNABACEAE			
<i>Verbesina encelioides</i> subsp. <i>encelioides</i>	E	F	AN	<i>Cannabis sativa</i>	N	F	PE
<i>Vernonia cinerea</i>	N	F	AN	CAPPARACEAE			
<i>Vittadinia cervicularis</i> var. <i>cervicularis</i>	N	F	PE	<i>Capparis michellii</i>	N	T	PE
<i>Vittadinia cervicularis</i> var. <i>subcervicularis</i>	N	F	PE	CAPRIFOLIACEAE			
<i>Vittadinia condyloides</i>	N	F	PE	<i>Lonicera japonica</i>	E	C	PE
<i>Vittadinia cuneata</i> var. <i>cuneata</i>	N	F	PE	CARYOPHYLLACEAE			
<i>Vittadinia cuneata</i> var. <i>hirsuta</i>	N	F	PE	<i>Arenaria leptoclados</i>	E	F	AN
<i>Vittadinia dissecta</i>	N	F	PE	<i>Arenaria serpyllifolia</i>	E	F	AN
<i>Vittadinia hispidula</i> var. <i>hispidula</i>	N	F	PE	<i>Cerastium glomeratum</i>	E	F	AN
<i>Vittadinia muelleri</i>	N	F	PE	<i>Gypsophila tubulosa</i>	E	F	AN
<i>Vittadinia pterochaeta</i>	N	F	PE	<i>Moenchia erecta</i>	E	F	AN
<i>Vittadinia pustulata</i>	N	F	PE	<i>Paronychia brasiliensis</i>	E	F	PE
<i>Vittadinia sulcata</i>	N	F	AN	<i>Petrorhagia dubia</i>	E	F	AN
<i>Xanthium spinosum</i>	E	F	AN	<i>Petrorhagia nanteuilii</i>	E	F	AN
<i>Xanthium strumarium</i>	E	F	AN	<i>Polycarpaea corymbosa</i> var. <i>minor</i>	N	F	AN
<i>Xerochrysum bracteatum</i>	N	F	PE	<i>Polycarpon tetraphyllum</i>	E	F	AN
<i>Xerochrysum viscosum</i>	N	F	PE	<i>Sagina apetala</i>	E	F	AN
<i>Zinnia peruviana</i>	E	F	AN	<i>Scleranthus biflorus</i>	N	F	PE
BIGNONIACEAE				<i>Scleranthus diander</i>	N	F	PE
<i>Pandorea pandorana</i> subsp. <i>austracaledonica</i>	N	C	PE	<i>Scleranthus pungens</i>	N	F	PE
<i>Pandorea pandorana</i> subsp. <i>pandorana</i>	N	C	PE	<i>Silene dioica</i>	E	F	PE
BORAGINACEAE				<i>Silene gallica</i>	E	F	AN
<i>Austrocynoglossum latifolium</i>	N	F	PE	<i>Spergularia rubra</i>	E	F	SP
<i>Buglossoides arvensis</i>	E	F	AN	<i>Stellaria angustifolia</i>	N	F	PE
<i>Cynoglossum australe</i>	N	F	PE	<i>Stellaria flaccida</i>	N	F	PE
<i>Cynoglossum suaveolens</i>	N	F	PE	<i>Stellaria media</i>	E	F	SP
<i>Echium plantagineum</i>	E	F	AN	<i>Stellaria multiflora</i>	N	F	AN
<i>Echium vulgare</i>	E	F	AN	<i>Stellaria pallida</i>	E	F	AN
<i>Ehretia membranifolia</i>	N	TS	PE	<i>Stellaria pungens</i>	N	F	PE
<i>Halgania brachyrhyncha</i>	N	LS	PE	CASUARINACEAE			
<i>Myosotis laxa</i> subsp. <i>caespitosa</i>	E	F	AN	<i>Allocasuarina littoralis</i>	N	T	PE
BRASSICACEAE				<i>Allocasuarina luehmannii</i>	N	T	PE
<i>Brassica nigra</i>	E	F	AN	<i>Allocasuarina torulosa</i>	N	T	PE
<i>Brassica oleracea</i>	E	F	PE	<i>Casuarina cunninghamiana</i> subsp. <i>cunninghamiana</i>	N	T	PE
<i>Brassica rapa</i> subsp. <i>campestris</i>	E	F	AN	CELASTRACEAE			
<i>Capsella bursa-pastoris</i>	E	F	AN	<i>Celastrus australis</i>	N	C	PE
<i>Cardamine hirsuta</i>	E	F	AN	<i>Maytenus bilocularis</i>	N	T	PE
<i>Cardamine paucijuga</i>	N	F	AN	<i>Maytenus cunninghamii</i>	N	TS	PE
<i>Coronopus didymus</i>	E	F	SP	<i>Maytenus silvestris</i>	N	TS	PE
<i>Hirschfeldia incana</i>	E	F	SP	CHENOPODIACEAE			
<i>Lepidium africanum</i>	E	F	SP	<i>Atriplex semibaccata</i>	N	F	PE
<i>Lepidium pseudohyssopifolium</i>	N	F	SP	<i>Atriplex spinibractea</i>	N	F	PE
<i>Rapistrum rugosum</i>	E	F	SP	<i>Atriplex suberecta</i>	N	F	PE
<i>Rorippa laciniata</i>	N	F	PE	<i>Chenopodium album</i>	E	F	AN
<i>Sisymbrium erysimoides</i>	E	F	SP	<i>Chenopodium ambrosioides</i>	E	LS	PE
<i>Sisymbrium irio</i>	E	F	SP	<i>Chenopodium auricomum</i>	N	F	PE
<i>Sisymbrium officinale</i>	E	F	SP	<i>Chenopodium carinatum</i>	N	F	AN
CACTACEAE				<i>Chenopodium desertorum</i> subsp. <i>desertorum</i>	N	F	PE
<i>Opuntia aurantiaca</i>	E	F	PE	<i>Chenopodium melanocarpum</i>	N	F	AN
<i>Opuntia stricta</i>	E	F	PE	<i>Chenopodium murale</i>	E	F	AN
CAMPANULACEAE				<i>Chenopodium pumilio</i>	N	F	AN
<i>Wahlenbergia communis</i>	N	F	PE	<i>Einadia hastata</i>	N	F	PE
<i>Wahlenbergia fluminalis</i>	N	F	PE	<i>Einadia nutans</i> subsp. <i>linifolia</i>	N	F	PE
<i>Wahlenbergia gracilentia</i>	N	F	PE	<i>Einadia nutans</i> subsp. <i>nutans</i>	N	F	PE
				<i>Einadia polygonoides</i>	N	F	PE

Species	O	LF	LC	Species	O	LF	LC
CHENOPODIACEAE				EPACRIDACEAE			
<i>Einadia trigonos subsp. leiocarpa</i>	N	F	PE	<i>Acrothamnus hookeri</i>	N	LS	PE
<i>Einadia trigonos subsp. stellulata</i>	N	F	PE	EUPHORBIACEAE			
<i>Enchylaena tomentosa</i>	N	LS	PE	<i>Adriana urticoides</i>	N	TS	PE
<i>Maireana aphylla</i>	N	LS	PE	<i>Bertya mollissima</i>	N	LS	PE
<i>Maireana brevifolia</i>	N	LS	PE	<i>Bertya oleifolia</i>	N	LS	PE
<i>Maireana enchylaenoides</i>	N	F	PE	<i>Beyeria viscosa</i>	N	LS	PE
<i>Maireana microcarpa</i>	N	LS	PE	<i>Breynia cernua</i>	N	LS	PE
<i>Maireana microphylla</i>	N	LS	PE	<i>Breynia oblongifolia</i>	N	LS	PE
<i>Rhagodia parabolica</i>	N	LS	PE	<i>Chamaesyce australis</i>	N	F	PE
<i>Salsola tragus</i>	N	F	SP	<i>Chamaesyce dallachyana</i>	N	F	PE
<i>Sclerolaena birchii</i>	N	F	PE	<i>Chamaesyce drummondii</i>	N	F	PE
<i>Sclerolaena divaricata</i>	N	LS	PE	<i>Chamaesyce species A</i>	N	F	PE
<i>Sclerolaena muricata var. muricata</i>	N	F	PE	<i>Claoxylon australe</i>	N	TS	PE
<i>Sclerolaena muricata var. semiglabra</i>	N	F	PE	<i>Croton phebaloides</i>	N	LS	PE
<i>Sclerolaena muricata var. villosa</i>	N	F	PE	<i>Euphorbia eremophila</i>	N	F	SP
CHLOANTHACEAE				<i>Euphorbia lathyris</i>	E	F	SP
<i>Spartothammella juncea</i>	N	LS	PE	<i>Euphorbia peplus</i>	E	F	AN
CLUSIACEAE				<i>Homalanthus populifolius</i>	N	TS	PE
<i>Hypericum gramineum</i>	N	F	PE	<i>Phyllanthus carpentariae</i>	N	LS	PE
<i>Hypericum japonicum</i>	N	F	PE	<i>Phyllanthus gunnii</i>	N	LS	PE
<i>Hypericum perforatum</i>	E	F	PE	<i>Phyllanthus subcrenulatus</i>	N	LS	PE
CONVOLVULACEAE				<i>Phyllanthus virgatus</i>	N	F	SP
<i>Calystegia marginata</i>	N	C	PE	<i>Poranthera microphylla</i>	N	F	AN
<i>Convolvulus arvensis</i>	E	C	PE	FABACEAE			
<i>Convolvulus erubescens</i>	N	F	PE	<i>Acacia blakei subsp. diphylla</i>	N	T	PE
<i>Convolvulus remotus</i>	N	F	PE	<i>Acacia brownii</i>	N	LS	PE
<i>Cuscuta campestris</i>	E	P	PE	<i>Acacia buxifolia</i>	N	TS	PE
<i>Dichondra repens</i>	N	F	PE	<i>Acacia caesiella</i>	N	TS	PE
<i>Dichondra species A</i>	N	GI	PA	<i>Acacia cheelii</i>	N	TS	PE
<i>Evolvulus alsinoides var. decumbens</i>	N	F	PE	<i>Acacia dealbata</i>	N	T	PE
<i>Evolvulus alsinoides var. villosicalyx</i>	N	F	PE	<i>Acacia deanei</i>	N	T	PE
CRASSULACEAE				<i>Acacia decora</i>	N	LS	PE
<i>Crassula sieberiana</i>	N	F	AN	<i>Acacia doratoxylon</i>	N	T	PE
CUCURBITACEAE				<i>Acacia falcata</i>	N	T	PE
<i>Citrullus lanatus</i>	E	F	AN	<i>Acacia falciformis</i>	N	TS	PE
<i>Cucumis myriocarpus subsp. leptodermis</i>	E	F	AN	<i>Acacia gladiiformis</i>	N	TS	PE
DILLENIACEAE				<i>Acacia harpophylla</i>	N	T	PE
<i>Hibbertia acicularis</i>	N	LS	PE	<i>Acacia homalophylla</i>	N	TS	PE
<i>Hibbertia dentata</i>	N	C	PE	<i>Acacia implexa</i>	N	T	PE
<i>Hibbertia hermanniifolia</i>	N	LS	PE	<i>Acacia leiocalyx subsp. leiocalyx</i>	N	T	PE
<i>Hibbertia linearis</i>	N	LS	PE	<i>Acacia leptoclada</i>	N	TS	PE
<i>Hibbertia obtusifolia</i>	N	LS	PE	<i>Acacia maidenii</i>	N	T	PE
<i>Hibbertia riparia</i>	N	LS	PE	<i>Acacia melanoxylon</i>	N	T	PE
<i>Hibbertia rufa</i>	N	LS	PE	<i>Acacia myrtifolia</i>	N	TS	PE
<i>Hibbertia serpyllifolia</i>	N	LS	PE	<i>Acacia neriiifolia</i>	N	TS	PE
DROSERACEAE				<i>Acacia paradoxa</i>	N	TS	PE
<i>Drosera auriculata</i>	N	F	AN	<i>Acacia pendula</i>	N	T	PE
<i>Drosera peltata</i>	N	F	PE	<i>Acacia penninervis</i>	N	T	PE
ELATINACEAE				<i>Acacia pruinosa</i>	N	TS	PE
<i>Elatine gratioloides</i>	N	F	AN	<i>Acacia rubida</i>	N	TS	PE
EPACRIDACEAE				<i>Acacia salicina</i>	N	TS	PE
<i>Acrotriche serrulata</i>	N	LS	PE	<i>Acacia triptera</i>	N	T	PE
<i>Brachyloma daphnoides</i>	N	LS	PE	<i>Acacia ulicifolia</i>	N	TS	PE
<i>Epacris pulchella</i>	N	LS	PE	<i>Acacia venulosa</i>	N	TS	PE
<i>Leucopogon attenuatus</i>	N	LS	PE	<i>Acacia viscidula</i>	N	LS	PE
<i>Leucopogon lanceolatus</i>	N	LS	PE	<i>Aotus subglaucula</i>	N	LS	PE
<i>Leucopogon microphyllus</i>	N	LS	PE	<i>Bossiaea buxifolia</i>	N	LS	PE
<i>Leucopogon muticus</i>	N	LS	PE	<i>Bossiaea neo-anglica</i>	N	LS	PE
<i>Leucopogon neo-anglicus</i>	N	LS	PE	<i>Bossiaea obcordata</i>	N	LS	PE
<i>Leucopogon virgatus</i>	N	LS	PE	<i>Bossiaea prostrata</i>	N	LS	PE
<i>Lissanthe strigosa subsp. strigosa</i>	N	LS	PE	<i>Chamaecytisus palmensis</i>	E	T	PE
<i>Lissanthe strigosa subsp. subulata</i>	N	LS	PE	<i>Crotalaria agatiflora subsp. agatiflora</i>	E	LS	PE
<i>Melichrus erubescens</i>	N	LS	PE	<i>Crotalaria dissitiflora subsp. dissitiflora</i>	N	F	PE
<i>Melichrus urceolatus</i>	N	LS	PE	<i>Crotalaria mitchellii subsp. laevis</i>	N	F	PE
<i>Monotoca scoparia</i>	N	LS	PE	<i>Cullen tenax</i>	N	F	PE
Ericaceae							

Species	O	LF	LC	Species	O	LF	LC
FABACEAE				FABACEAE			
<i>Daviesia acicularis</i>	N	LS	PE	<i>Swainsona behriana</i>	N	F	PE
<i>Daviesia genistifolia</i>	N	LS	PE	<i>Swainsona bracteata</i>	N	F	PE
<i>Daviesia latifolia</i>	N	LS	PE	<i>Swainsona galegifolia</i>	N	F	PE
<i>Daviesia pubigera</i>	N	LS	PE	<i>Swainsona luteola</i>	N	F	PE
<i>Daviesia ulicifolia</i>	N	LS	PE	<i>Swainsona microphylla</i>	N	F	PE
<i>Desmodium brachypodum</i>	N	F	PE	<i>Swainsona monticola</i>	N	F	PE
<i>Desmodium gunnii</i>	N	F	PE	<i>Swainsona murrayana</i>	N	F	PE
<i>Desmodium varians</i>	N	F	PE	<i>Swainsona parviflora</i>	N	F	PE
<i>Dillwynia phyllicoides</i>	N	LS	PE	<i>Swainsona queenslandica</i>	N	F	PE
<i>Dillwynia retorta species complex</i>	N	LS	PE	<i>Swainsona reticulata</i>	N	F	PE
<i>Dillwynia sericea</i>	N	LS	PE	<i>Templetonia stenophylla</i>	N	LS	PE
<i>Gleditsia triacanthos</i>	E	T	PE	<i>Tephrosia brachyodon</i>	N	F	PE
<i>Glycine canescens</i>	N	C	PE	<i>Trifolium angustifolium</i>	E	F	AN
<i>Glycine clandestina</i>	N	C	PE	<i>Trifolium arvense</i>	E	F	AN
<i>Glycine microphylla</i>	N	C	PE	<i>Trifolium campestre</i>	E	F	AN
<i>Glycine stenophylla</i>	N	C	PE	<i>Trifolium cernuum</i>	E	F	AN
<i>Glycine tabacina</i>	N	C	PE	<i>Trifolium dubium</i>	E	F	AN
<i>Glycine tomentella</i>	N	C	PE	<i>Trifolium fragiferum</i>	E	F	AN
<i>Gompholobium huegelii</i>	N	LS	PE	<i>Trifolium glomeratum</i>	E	F	AN
<i>Hardenbergia violacea</i>	N	C	PE	<i>Trifolium pratense</i>	E	F	PE
<i>Hovea heterophylla</i>	N	LS	PE	<i>Trifolium repens</i>	E	F	PE
<i>Hovea lanceolata</i>	N	LS	PE	<i>Trifolium subterraneum</i>	E	F	AN
<i>Hovea linearis</i>	N	LS	PE	<i>Trifolium tomentosum</i>	E	F	AN
<i>Hovea purpurea</i>	N	LS	PE	<i>Vicia hirsuta</i>	E	C	AN
<i>Indigofera adesmiifolia</i>	N	LS	PE	<i>Vicia monantha</i>	E	C	AN
<i>Indigofera australis</i>	N	LS	PE	<i>Vicia sativa</i>	E	C	AN
<i>Isotropis foliosa</i>	N	LS	PE	<i>Vigna lanceolata var. lanceolata</i>	N	C	PE
<i>Jacksonia scoparia</i>	N	TS	PE	<i>Vigna vexillata var. angustifolia</i>	N	C	PE
<i>Kennedia rubicunda</i>	N	C	PE	<i>Zornia dyctiocarpa var. dyctiocarpa</i>	N	F	PE
<i>Lespedeza juncea subsp. sericea</i>	N	LS	PE	<i>Zornia floribunda</i>	N	F	PE
<i>Lotus australis</i>	N	F	PE	FUMARIACEAE			
<i>Lotus cruentus</i>	N	F	PE	<i>Fumaria capreolata subsp. capreolata</i>	E	F	AN
<i>Medicago arabica</i>	E	F	AN	GENTIANACEAE			
<i>Medicago laciniata</i>	E	F	AN	<i>Centaurium erythraea</i>	E	F	AN
<i>Medicago lupulina</i>	E	F	SP	<i>Centaurium tenuiflorum</i>	E	F	AN
<i>Medicago minima</i>	E	F	AN	<i>Schenkia australis</i>	N	F	AN
<i>Medicago orbicularis</i>	E	F	AN	GERANIACEAE			
<i>Medicago polymorpha</i>	E	F	AN	<i>Geranium molle subsp. molle</i>	E	F	SP
<i>Medicago sativa</i>	E	F	PE	<i>Geranium neglectum</i>	N	F	PE
<i>Medicago truncatula</i>	E	F	AN	<i>Geranium potentilloides var. potentilloides</i>	N	F	PE
<i>Melilotus albus</i>	E	F	SP	<i>Geranium retrorsum</i>	E	F	PE
<i>Melilotus indicus</i>	E	F	AN	<i>Geranium solanderi var. grande</i>	N	F	PE
<i>Mirbelia oxylobioides</i>	N	LS	PE	<i>Geranium solanderi var. solanderi</i>	N	F	PE
<i>Mirbelia pungens</i>	N	LS	PE	<i>Pelargonium inodorum</i>	N	F	AN
<i>Neptunia gracilis forma gracilis</i>	N	F	PE	GOODENIACEAE			
<i>Pultenaea campbellii</i>	N	LS	PE	<i>Brunonia australis</i>	N	F	PE
<i>Pultenaea cuneata</i>	N	LS	PE	<i>Dampiera stricta</i>	N	LS	PE
<i>Pultenaea foliolosa</i>	N	LS	PE	<i>Goodenia bellidifolia subsp. argentea</i>	N	F	PE
<i>Pultenaea linophylla</i>	N	LS	PE	<i>Goodenia bellidifolia subsp. bellidifolia</i>	N	F	PE
<i>Pultenaea microphylla</i>	N	LS	PE	<i>Goodenia cycloptera</i>	N	F	AN
<i>Pultenaea myrtooides</i>	N	LS	PE	<i>Goodenia fascicularis</i>	N	F	PE
<i>Pultenaea scabra</i>	N	LS	PE	<i>Goodenia glabra</i>	N	F	PE
<i>Pultenaea setulosa</i>	N	LS	PE	<i>Goodenia hederacea</i>	N	F	PE
<i>Pultenaea spinosa</i>	N	LS	PE	<i>Goodenia ovata</i>	N	F	PE
<i>Rhynchosia minima</i>	N	C	PE	<i>Goodenia pinnatifida</i>	N	F	AN
<i>Robinia pseudoacacia</i>	E	T	PE	<i>Goodenia rotundifolia</i>	N	F	PE
<i>Senna aciphylla</i>	N	LS	PE	<i>Scaevola albida</i>	N	LS	PE
<i>Senna artemisioides nothosubsp. coriacea</i>	N	LS	PE	<i>Scaevola hookeri</i>	N	F	PE
<i>Senna artemisioides subsp. zygophylla</i>	N	LS	PE	<i>Velleia paradoxa</i>	N	F	PE
<i>Senna barclayana</i>	N	F	PE	HAEMODORACEAE			
<i>Senna baronfieldii</i>	N	LS	PE	<i>Haemodorum planifolium</i>	N	F	PE
<i>Senna clavigera</i>	N	LS	PE	HALORAGACEAE			
<i>Senna coronilloides</i>	N	LS	PE	<i>Gonocarpus elatus</i>	N	F	PE
<i>Senna occidentalis</i>	E	LS	PE	<i>Gonocarpus humilis</i>	N	F	PE
<i>Sesbania cannabina var. cannabina</i>	N	LS	PE				

Species	O	LF	LC	Species	O	LF	LC
HALORAGACEAE				MALVACEAE			
<i>Gonocarpus micranthus</i>	N	F	PE	<i>Abutilon malvifolium</i>	N	LS	PE
<i>Gonocarpus tetragynus</i>	N	F	PE	<i>Abutilon oxycarpum</i> var. <i>oxycarpum</i>	N	LS	PE
<i>Gonocarpus teucroides</i>	N	F	PE	<i>Abutilon oxycarpum</i> var. <i>subsagittatum</i>	N	LS	PE
<i>Haloragis aspera</i>	N	F	PE	<i>Abutilon theophrasti</i>	E	F	AN
<i>Haloragis glauca</i> forma <i>glauca</i>	N	F	PE	<i>Abutilon tubulosum</i>	N	LS	PE
<i>Haloragis heterophylla</i>	N	F	PE	<i>Hibiscus sturtii</i>	N	LS	PE
<i>Haloragis serra</i>	N	F	PE	<i>Hibiscus trionum</i>	E	F	SP
<i>Myriophyllum verrucosum</i>	N	F	PE	<i>Malus pumila</i>	E	T	PE
IRIDACEAE				<i>Malva neglecta</i>	E	F	AN
<i>Sisyrinchium exile</i>	E	F	AN	<i>Malva parviflora</i>	E	F	AN
LAMIACEAE				<i>Malvastrum americanum</i>	E	F	SP
<i>Ajuga australis</i>	N	F	PE	<i>Malvastrum coromandelianum</i>	N	F	AN
<i>Lamium amplexicaule</i>	E	F	AN	<i>Modiola caroliniana</i>	E	F	SP
<i>Marrubium vulgare</i>	E	F	PE	<i>Pavonia hastata</i>	E	LS	PE
<i>Mentha australis</i>	N	F	PE	<i>Sida corrugata</i>	N	F	PE
<i>Mentha diemenica</i>	N	F	PE	<i>Sida cunninghamii</i>	N	F	PE
<i>Mentha satureioides</i>	N	F	PE	<i>Sida filiformis</i>	N	F	PE
<i>Plectranthus graveolens</i>	N	F	PE	<i>Sida rhombifolia</i>	N	F	PE
<i>Plectranthus parviflorus</i>	N	F	PE	<i>Sida spinosa</i>	E	LS	PE
<i>Plectranthus suaveolens</i>	N	F	PE	<i>Sida subspicata</i>	N	LS	PE
<i>Prostanthera cruciflora</i>	N	LS	PE	<i>Sida trichopoda</i>	N	F	PE
<i>Prostanthera lasianthos</i>	N	LS	PE	MELIACEAE			
<i>Prostanthera nivea</i>	N	LS	PE	<i>Melia azedarach</i>	N	T	PE
<i>Prostanthera ovalifolia</i>	N	LS	PE	MENISPERMACEAE			
<i>Prostanthera scutellarioides</i>	N	LS	PE	<i>Stephania japonica</i>	N	C	PE
<i>Prunella vulgaris</i>	E	F	PE	MORACEAE			
<i>Salvia reflexa</i>	E	F	AN	<i>Ficus coronata</i>	N	T	PE
<i>Salvia verbenaca</i>	E	F	PE	<i>Ficus rubiginosa</i>	N	T	PE
<i>Scutellaria humilis</i>	N	F	PE	<i>Maclura pomifera</i>	E	T	PE
<i>Teucrium argutum</i>	N	F	PE	<i>Morus alba</i>	E	T	PE
<i>Teucrium species D</i>	N	F	PE	MYOPORACEAE			
LAURACEAE				<i>Eremophila debilis</i>	N	LS	PE
<i>Cassytha pubescens</i>	N	C	PE	<i>Eremophila mitchellii</i>	N	TS	PE
LENTIBULARIACEAE				<i>Eremophila polyclada</i>	N	LS	PE
<i>Utricularia dichotoma</i>	N	F	PE	<i>Myoporum montanum</i>	N	LS	PE
LINACEAE				MYRTACEAE			
<i>Linum marginale</i>	N	F	PE	<i>Acmena smithii</i>	N	T	PE
LOBELIACEAE				<i>Angophora floribunda</i>	N	T	PE
<i>Isotoma axillaris</i>	N	F	PE	<i>Callistemon pityoides</i>	N	TS	PE
<i>Isotoma fluviatilis</i>	N	F	PE	<i>Callistemon pungens</i>	N	TS	PE
<i>Lobelia gibbosa</i>	N	F	AN	<i>Callistemon viminalis</i>	N	T	PE
<i>Lobelia gracilis</i>	N	F	PE	<i>Calytrix tetragona</i>	N	LS	PE
<i>Pratia concolor</i>	N	F	PE	<i>Eucalyptus acaciiformis</i>	N	T	PE
<i>Pratia pedunculata</i>	N	F	PE	<i>Eucalyptus albens</i>	N	T	PE
<i>Pratia purpurascens</i>	N	F	PE	<i>Eucalyptus andrewsii</i>	N	T	PE
LOGANIACEAE				<i>Eucalyptus banksii</i>	N	T	PE
<i>Logania albiflora</i>	N	LS	PE	<i>Eucalyptus blakelyi</i>	N	T	PE
LORANTHACEAE				<i>Eucalyptus bridgesiana</i>	N	T	PE
<i>Amyema bifurcatum</i>	N	P	PE	<i>Eucalyptus caleyi</i> subsp. <i>caleyi</i>	N	T	PE
<i>Amyema cambagei</i>	N	P	PE	<i>Eucalyptus caleyi</i> subsp. <i>ovendenii</i>	N	T	PE
<i>Amyema linophylla</i> subsp. <i>orientalis</i>	N	P	PE	<i>Eucalyptus caliginosa</i>	N	T	PE
<i>Amyema longifolia</i>	N	P	PE	<i>Eucalyptus camaldulensis</i>	N	T	PE
<i>Amyema miquelii</i>	N	P	PE	<i>Eucalyptus chloroclada</i>	N	T	PE
<i>Amyema pendula</i>	N	P	PE	<i>Eucalyptus conica</i>	N	T	PE
<i>Amyema quandang</i> var. <i>bancroftii</i>	N	P	PE	<i>Eucalyptus crebra</i>	N	T	PE
<i>Amyema quandang</i> var. <i>quandang</i>	N	P	PE	<i>Eucalyptus dalrympleana</i>	N	T	PE
<i>Dendrophthoe glabrescens</i>	N	P	PE	<i>Eucalyptus dealbata</i>	N	T	PE
<i>Dendrophthoe vitellina</i>	N	P	PE	<i>Eucalyptus dwyeri</i>	N	T	PE
<i>Lysiana subfalcata</i>	N	P	PE	<i>Eucalyptus elliptica</i>	N	T	PE
<i>Muellerina bidwillii</i>	N	P	PE	<i>Eucalyptus laevopinea</i>	N	T	PE
<i>Muellerina eucalyptoides</i>	N	P	PE	<i>Eucalyptus macrorhyncha</i>	N	T	PE
LYTHRACEAE				<i>Eucalyptus malacoxyton</i>	N	T	PE
<i>Lythrum hyssopifolia</i>	N	F	AN	<i>Eucalyptus mckieana</i>	N	T	PE
<i>Lythrum salicaria</i>	N	F	PE	<i>Eucalyptus melanophloia</i>	N	T	PE
MALACEAE				<i>Eucalyptus melliodora</i>	N	T	PE
<i>Cotoneaster glaucophyllus</i>	E	TS	PE	<i>Eucalyptus microcarpa</i>	N	T	PE

Species	O	LF	LC	Species	O	LF	LC
MYRTACEAE				PLANTAGINACEAE			
<i>Eucalyptus moluccana</i>	N	T	PE	<i>Plantago hispida</i>	N	F	PE
<i>Eucalyptus nobilis</i>	N	T	PE	<i>Plantago lanceolata</i>	E	F	SP
<i>Eucalyptus nortonii</i>	N	T	PE	<i>Plantago turrifera</i>	N	F	AN
<i>Eucalyptus obliqua</i>	N	T	PE	<i>Plantago varia</i>	N	F	PE
<i>Eucalyptus pauciflora</i>	N	T	PE	POLYGALACEAE			
<i>Eucalyptus pilligaensis</i>	N	T	PE	<i>Comesperma sylvestre</i>	N	LS	PE
<i>Eucalyptus populnea</i> subsp. <i>bimbil</i>	N	T	PE	<i>Polygala japonica</i>	N	F	PE
<i>Eucalyptus prava</i>	N	T	PE	POLYGANACEAE			
<i>Eucalyptus quinniorum</i>	N	T	PE	<i>Fallopia convolvulus</i>	E	F	AN
<i>Eucalyptus sideroxylon</i>	N	T	PE	<i>Rumex brownii</i>	N	F	PE
<i>Eucalyptus stannicola</i>	N	T	PE	<i>Rumex crispus</i>	E	F	PE
<i>Eucalyptus stellulata</i>	N	T	PE	POLYGONACEAE			
<i>Eucalyptus subtilior</i>	N	T	PE	<i>Muehlenbeckia florulenta</i>	N	LS	PE
<i>Eucalyptus viminalis</i>	N	T	PE	<i>Muehlenbeckia gracillima</i>	N	C	PE
<i>Eucalyptus volcanica</i>	N	T	PE	<i>Muehlenbeckia rhyticarya</i>	N	LS	PE
<i>Eucalyptus youmanii</i>	N	T	PE	<i>Persicaria attenuata</i>	N	F	AN
<i>Kunzea opposita</i>	N	LS	PE	<i>Persicaria decipiens</i>	N	F	AN
<i>Kunzea</i> sp. ' <i>Mt Kaputar</i> '	N	LS	PE	<i>Persicaria hydropiper</i>	N	F	AN
<i>Leptospermum brevipes</i>	N	TS	PE	<i>Persicaria lapathifolia</i>	N	F	AN
<i>Leptospermum microcarpum</i>	N	T	PE	<i>Persicaria prostrata</i>	N	F	PE
<i>Leptospermum novae-angliae</i>	N	T	PE	<i>Polygonum arenastrum</i>	E	F	SP
<i>Leptospermum polygalifolium</i>	N	TS	PE	<i>Polygonum aviculare</i>	E	F	SP
<i>Melaleuca bracteata</i>	N	TS	PE	<i>Polygonum patulum</i>	N	F	AN
<i>Micromyrtus sessilis</i>	N	LS	PE	<i>Rumex acetosella</i>	E	F	PE
<i>Micromyrtus striata</i>	N	LS	PE	<i>Rumex crystallinus</i>	N	F	AN
NYCTAGINACEAE				<i>Rumex dumosus</i>	N	F	PE
<i>Boerhavia dominii</i>	N	F	PE	PORTULACACEAE			
<i>Boerhavia reptata</i>	N	F	PE	<i>Calandrinia eremaea</i>	N	F	AN
OLEACEAE				<i>Portulaca oleracea</i>	N	F	AN
<i>Jasminum lineare</i>	N	LS	PE	PRIMULACEAE			
<i>Jasminum suavisissimum</i>	N	LS	PE	<i>Anagallis arvensis</i>	E	F	SP
<i>Ligustrum lucidum</i>	E	TS	PE	PROTEACEAE			
<i>Notelaea microcarpa</i> var. <i>microcarpa</i>	N	T	PE	<i>Grevillea floribunda</i> subsp. <i>floribunda</i>	N	LS	PE
<i>Notelaea microcarpa</i> var. <i>velutina</i>	N	T	PE	<i>Grevillea ramosissima</i> subsp. <i>ramosissima</i>	N	LS	PE
<i>Olea europaea</i>	E	T	PE	<i>Grevillea triternata</i>	N	LS	PE
ONAGRACEAE				<i>Hakea eriantha</i>	N	TS	PE
<i>Epilobium billardioreanum</i> subsp. <i>cinereum</i>	N	F	PE	<i>Hakea laevipes</i> subsp. <i>graniticola</i>	N	TS	PE
<i>Epilobium billardioreanum</i> subsp. <i>hydrophilum</i>	N	F	PE	<i>Hakea microcarpa</i>	N	TS	PE
<i>Epilobium hirtigerum</i>	N	F	PE	<i>Lomatia arborescens</i>	N	T	PE
<i>Ludwigia peploides</i> subsp. <i>montevidensis</i>	N	F	PE	<i>Persoonia chamaepeuce</i>	N	TS	PE
<i>Oenothera rosea</i>	E	LS	PE	<i>Persoonia cornifolia</i>	N	TS	PE
OXALIDACEAE				<i>Persoonia fastigiata</i>	N	LS	PE
<i>Oxalis chnoodes</i>	N	F	PE	<i>Persoonia sericea</i>	N	LS	PE
<i>Oxalis corniculata</i>	N	F	PE	<i>Petrophile canescens</i>	N	LS	PE
<i>Oxalis thompsoniae</i>	N	F	PE	RANUNCULACEAE			
PAPAVERACEAE				<i>Clematis aristata</i>	N	C	PE
<i>Argemone ochroleuca</i> subsp. <i>ochroleuca</i>	N	F	AN	<i>Clematis glycinoides</i> var. <i>glycinoides</i>	N	C	PE
PHYTOLACCACEAE				<i>Clematis glycinoides</i> var. <i>submutica</i>	N	C	PE
<i>Phytolacca octandra</i>	E	F	SP	<i>Clematis microphylla</i> var. <i>leptophylla</i>	N	C	PE
PITTOSPORACEAE				<i>Clematis microphylla</i> var. <i>microphylla</i>	N	C	PE
<i>Billardiera scandens</i>	N	LS	PE	<i>Ranunculus lappaceus</i>	N	F	PE
<i>Bursaria longisepala</i>	N	TS	PE	<i>Ranunculus meristus</i>	N	F	PE
<i>Bursaria spinosa</i>	N	TS	PE	<i>Ranunculus pumilio</i>	N	F	AN
<i>Cheiranthra cyanea</i> var. <i>cyanea</i>	N	LS	PE	<i>Ranunculus sessiliflorus</i> var. <i>pilulifer</i>	N	F	AN
<i>Pittosporum angustifolium</i>	N	T	PE	<i>Ranunculus sessiliflorus</i> var. <i>sessiliflorus</i>	N	F	AN
<i>Pittosporum undulatum</i>	N	T	PE	<i>Ranunculus undosus</i>	N	F	PE
<i>Rhytidosporum diosmoides</i>	N	LS	PE	RHAMNACEAE			
<i>Rhytidosporum procumbens</i>	N	LS	PE	<i>Alphitonia excelsa</i>	N	T	PE
PLANTAGINACEAE				<i>Cryptandra amara</i> var. <i>amara</i>	N	LS	PE
<i>Plantago cunninghamii</i>	N	F	PE	<i>Cryptandra amara</i> var. <i>floribunda</i>	N	LS	PE
<i>Plantago debilis</i>	N	F	SP	<i>Cryptandra amara</i> var. <i>longiflora</i>	N	LS	PE
<i>Plantago gaudichaudii</i>	N	F	PE	<i>Cryptandra propinqua</i>	N	LS	PE
				<i>Discaria pubescens</i>	N	LS	PE

Species	O	LF	LC	Species	O	LF	LC
RHAMNACEAE				SAPINDACEAE			
<i>Pomaderris angustifolia</i>	N	TS	PE	<i>Cupaniopsis baileyana</i>	N	T	PE
<i>Pomaderris betulina</i>	N	TS	PE	<i>Dodonaea boroniifolia</i>	N	LS	PE
ROSACEAE				<i>Dodonaea heteromorpha</i>	N	TS	PE
<i>Acaena agnipila</i>	N	F	PE	<i>Dodonaea rhombifolia</i>	N	TS	PE
<i>Acaena echinata</i>	N	F	PE	<i>Dodonaea sinuolata</i>	N	LS	PE
<i>Acaena novae-zelandiae</i>	N	F	PE	<i>Dodonaea triquetra</i>	N	TS	PE
<i>Acaena ovina</i>	N	F	PE	<i>Dodonaea viscosa</i> subsp. <i>angustifolia</i>	N	TS	PE
<i>Rosa rubiginosa</i>	E	LS	PE	<i>Dodonaea viscosa</i> subsp. <i>angustissima</i>	N	TS	PE
<i>Rubus discolor</i>	E	LS	PE	<i>Dodonaea viscosa</i> subsp. <i>cuneata</i>	N	TS	PE
<i>Rubus fruticosus</i> sp. agg.	E	LS	PE	<i>Dodonaea viscosa</i> subsp. <i>mucronata</i>	N	TS	PE
<i>Rubus parvifolius</i>	N	LS	PE	<i>Dodonaea viscosa</i> subsp. <i>spatulata</i>	N	TS	PE
<i>Rubus pyramidalis</i>	E	LS	PE	SCROPHULARIACEAE			
<i>Rubus rosifolius</i>	N	LS	PE	<i>Derwentia arcuata</i>	N	F	PE
<i>Rubus ulmifolius</i>	E	LS	PE	<i>Derwentia arenaria</i>	N	F	PE
<i>Rubus vestitus</i>	E	LS	PE	<i>Derwentia derwentiana</i>	N	F	PE
RUBIACEAE				<i>Derwentia velutina</i>	N	F	PE
<i>Asperula conferta</i>	N	F	PE	<i>Euphrasia collina</i>	N	F	PE
<i>Asperula cunninghamii</i>	N	F	PE	<i>Gratiola pedunculata</i>	N	F	PE
<i>Asperula scoparia</i>	N	F	PE	<i>Gratiola peruviana</i>	N	F	PE
<i>Asperula subulifolia</i>	N	F	PE	<i>Linaria arvensis</i>	E	F	AN
<i>Coprosma hirtella</i>	N	LS	PE	<i>Linaria pelisseriana</i>	E	F	AN
<i>Coprosma quadrifida</i>	N	LS	PE	<i>Mimulus gracilis</i>	N	F	PE
<i>Galium aparine</i>	E	F	AN	<i>Mimulus prostratus</i>	N	F	PE
<i>Galium binifolium</i>	N	F	AN	<i>Mimulus repens</i>	N	F	PE
<i>Galium ciliare</i>	N	F	PE	<i>Misopates orontium</i>	E	F	AN
<i>Galium gaudichaudii</i>	N	F	PE	<i>Verbascum thapsus</i> subsp. <i>thapsus</i>	E	F	SP
<i>Galium migrans</i>	N	F	PE	<i>Verbascum virgatum</i>	E	F	SP
<i>Galium murale</i>	E	F	AN	<i>Veronica anagallis-aquatica</i>	E	F	PE
<i>Galium propinquum</i>	N	F	AN	<i>Veronica arvensis</i>	E	F	AN
<i>Opercularia aspera</i>	N	LS	PE	<i>Veronica calycina</i>	E	F	AN
<i>Opercularia diphylla</i>	N	F	PE	<i>Veronica plebeia</i>	N	F	PE
<i>Opercularia hispida</i>	N	F	PE	SIMAROUBACEAE			
<i>Pomax umbellata</i>	N	LS	PE	<i>Ailanthus altissima</i>	E	T	PE
<i>Psydrax odorata</i>	N	T	PE	SMILACACEAE			
<i>Psydrax oleifolia</i>	N	T	PE	<i>Smilax australis</i>	N	C	PE
<i>Sherardia arvensis</i>	E	F	AN	SOLANACEAE			
RUTACEAE				<i>Capsicum annuum</i> var. <i>glabriusculum</i>	E	LS	PE
<i>Boronia microphylla</i>	N	LS	PE	<i>Cestrum parqui</i>	E	LS	PE
<i>Boronia polygalifolia</i>	N	LS	PE	<i>Datura ferox</i>	E	F	AN
<i>Boronia rubiginosa</i>	N	LS	PE	<i>Datura stramonium</i>	E	F	AN
<i>Correa alba</i> var. <i>alba</i>	N	LS	PE	<i>Lycium ferocissimum</i>	E	TS	PE
<i>Correa glabra</i>	N	LS	PE	<i>Nicotiana suaveolens</i>	N	F	PE
<i>Correa reflexa</i>	N	LS	PE	<i>Physalis ixocarpa</i>	E	F	AN
<i>Geijera parviflora</i>	N	TS	PE	<i>Solanum americanum</i>	N	LS	PE
<i>Melicope micrococca</i>	N	T	PE	<i>Solanum aviculare</i>	N	TS	PE
<i>Phebalium squamulosum</i> subsp. <i>squamulosum</i>	N	LS	PE	<i>Solanum brownii</i>	N	LS	PE
<i>Zieria aspalathoides</i>	N	LS	PE	<i>Solanum chenopodioides</i>	E	F	AN
<i>Zieria cytisoides</i>	N	TS	PE	<i>Solanum cinereum</i>	N	F	PE
SALICACEAE				<i>Solanum ellipticum</i>	N	F	PE
<i>Populus nigra</i>	E	T	PE	<i>Solanum esuriale</i>	N	F	PE
<i>Salix babylonica</i>	E	T	PE	<i>Solanum ferocissimum</i>	N	LS	PE
SAMBUCACEAE				<i>Solanum opacum</i>	N	F	AN
<i>Sambucus australasica</i>	N	LS	PE	<i>Solanum papaverifolium</i>	N	F	PE
<i>Sambucus gaudichaudiana</i>	N	LS	PE	<i>Solanum parvifolium</i>	N	LS	PE
SANTALACEAE				<i>Solanum prinophyllum</i>	N	F	SP
<i>Exocarpos cupressiformis</i>	N	HP	PE	<i>Solanum pseudocapsicum</i>	E	LS	PE
<i>Exocarpos strictus</i>	N	TS	PE	<i>Solanum semiarmatum</i>	N	LS	PE
<i>Leptomeria drupacea</i>	N	TS	PE	STACKHOUSIACEAE			
<i>Santalum lanceolatum</i>	N	HP	PE	<i>Stackhousia monogyna</i>	N	F	PE
<i>Thesium australe</i>	N	F	PE	<i>Stackhousia muricata</i>	N	F	PE
SAPINDACEAE				<i>Stackhousia viminea</i>	N	F	PE
<i>Alectryon forsythii</i>	N	TS	PE	STERCULIACEAE			
<i>Alectryon oleifolius</i>	N	T	PE	<i>Brachychiton populneus</i> subsp. <i>populneus</i>	N	T	PE
<i>Alectryon subdentatus</i> forma <i>subdentatus</i>	N	T	PE	<i>Rulingia dasyphylla</i>	N	TS	PE
<i>Atalaya hemiglauca</i>	N	T	PE				

Species	O	LF	LC	Species	O	LF	LC
THYMELAEACEAE				VERBENACEAE			
<i>Pimelea curviflora</i>	N	LS	PE	<i>Verbena bonariensis</i>	E	F	PE
<i>Pimelea curviflora</i> var. <i>divergens</i>	N	LS	PE	<i>Verbena gaudichaudii</i>	E	F	PE
<i>Pimelea curviflora</i> var. <i>gracilis</i>	N	LS	PE	<i>Verbena hispida</i>	E	F	PE
<i>Pimelea curviflora</i> var. <i>sericea</i>	N	LS	PE	<i>Verbena incompta</i>	E	F	PE
<i>Pimelea glauca</i>	N	LS	PE	<i>Verbena litoralis</i>	E	F	PE
<i>Pimelea latifolia</i>	N	LS	PE	<i>Verbena officinalis</i>	E	F	PE
<i>Pimelea linifolia</i> subsp. <i>collina</i>	N	LS	PE	<i>Verbena quadrangularis</i>	E	F	PE
<i>Pimelea linifolia</i> subsp. <i>linifolia</i>	N	LS	PE	<i>Verbena rigida</i>	E	F	PE
<i>Pimelea micrantha</i>	N	LS	PE	VIOLACEAE			
<i>Pimelea neo-anglica</i>	N	LS	PE	<i>Hybanthus monopetalus</i>	N	F	PE
<i>Pimelea simplex</i>	N	LS	PE	<i>Melicytus dentatus</i>	N	TS	PE
<i>Pimelea stricta</i>	N	LS	PE	<i>Viola banksii</i>	N	F	PE
<i>Pimelea strigosa</i>	N	LS	PE	<i>Viola betonicifolia</i>	N	F	PE
ULMACEAE				<i>Viola caleyana</i>	N	F	PE
<i>Trema tomentosa</i> var. <i>viridis</i>	N	T	PE	VISCACEAE			
URTICACEAE				<i>Korthalsella rubra</i> subsp. <i>geijericola</i>	N	P	PE
<i>Parietaria debilis</i>	N	F	AN	<i>Notothixos cornifolius</i>	N	P	PE
<i>Urtica incisa</i>	N	F	PE	VITACEAE			
<i>Urtica urens</i>	E	F	AN	<i>Cayratia clematidea</i>	N	C	PE
VERBENACEAE				<i>Clematicissus opaca</i>	N	C	PE
<i>Glandularia aristigera</i>	E	F	PE	ZYGOPHYLLACEAE			
<i>Oncinocalyx betchei</i>	N	F	PE	<i>Tribulus micrococcus</i>	N	F	AN
<i>Phyla canescens</i>	E	F	PE	<i>Tribulus minutus</i>	N	F	AN
<i>Phyla nodiflora</i> var. <i>nodiflora</i>	E	F	PE	<i>Tribulus terrestris</i>	E	F	AN

APPENDIX 2

Species list for taxa represented in seasonal monitoring dataset (Chapter 3)

Plant species incidence data for six grazed pasture sites for 10 monitoring periods over two years from October 2008 to October 2010, showing origin (O, N = native, E = exotic) and Raunkier's life-form classification (R, H = hemicryptophyte, G = geophyte, T = therophyte (annual), Ph = Phanerophyte, C = Chamaephyte). The presence (+) or absence (-) of each species is indicated at each site for each monitoring period, in chronological order.

	O	R	Arlemont	Bareela	Gemma	Marathon	Werona	Wooli
Monocotyledons								
ANTHERICACEAE								
<i>Arthropodium minus</i>	N	H	-----	-----	-----	-----	-----	++-----+
<i>Dichopogon fimbriatus</i>	N	H	-----	-----	-----	-----	-----	++-+---+
ASPHODELACEAE								
<i>Bulbine bulbosa</i>	N	G	-----	-----	+-----	-----	-----	+---+---+
CYPERACEAE								
<i>Carex inversa</i>	N	H	++++-++++	+++++-----	+++++-----	+++++-----	-----	+++++-----
<i>Cyperus gracilis</i>	N	H	+-----	-----	-----	-----	+-----	-----
JUNCACEAE								
<i>Juncus subsecundus</i>	N	H	-----	+++++-----	-----	-----	-----	-----
<i>Juncus usitatus</i>	N	H	+++++-----	-----	-----	-----	-----	-----
POACEAE								
<i>Aristida leptopoda</i>	N	H	-----	-----	+---+---+	-----	-----	-----
<i>Aristida ramosa</i>	N	H	+++++-----	+++++-----	+++++-----	+++++-----	+++++-----	+++++-----
<i>Austrodanthonia bipartita</i>	N	H	-----	+++++-----	++-+---+	+++++-----	+++++-----	+++++-----
<i>Austrodanthonia caespitosa</i>	N	H	-----	++-+---+	-----	-----	+++++-----	-----
<i>Austrodanthonia racemosa</i>	N	H	+++---+---	-----	-----	-----	-----	-----
<i>Austrostipa aristiglumis</i>	N	H	-----	-----	+++++-----	-----	-----	-----
<i>Austrostipa scabra</i>	N	H	-----	++-----	-----	+++++-----	+++++-----	+++++-----
<i>Austrostipa setacea</i>	N	H	-----	-----	-----	-----	-----	+++-----
<i>Austrostipa verticillata</i>	N	H	-----	-----	-----	+++++-----	-----	+++++-----
<i>Avena barbata</i>	E	T	-----	-----	+-----	-----	-----	-----
<i>Bothriochloa macra</i>	N	H	+++++-----	+++++-----	++-+---+	+++++-----	+++++-----	+++++-----
<i>Bromus hordeaceus</i>	E	T	++-----	++-----	+++---+---	-----	+++---+---	---+-----
<i>Chloris truncata</i>	N	H	---+---+---	+++++-----	+++++-----	-----	+++++-----	+++++-----
<i>Chloris ventricosa</i>	N	H	-----	-----	-----	+++---+---	-----	-----
<i>Cynodon dactylon</i>	N	H	++-----	-----	++-----	++-----	-----	++-----
<i>Dichanthium sericeum</i>	N	H	-----	+++++-----	+++++-----	-----	+++++-----	-----
<i>Dichelachne micrantha</i>	N	H	-----	+++-----	-----	-----	-----	-----
<i>Digitaria brownii</i>	N	H	-----	-----	-----	-----	-----	++-----
<i>Digitaria divaricatissima</i>	N	H	-----	-----	-----	-----	-----	-----
<i>Eleusine tristachya</i>	E	T	+++-----	+++++-----	-----	-----	-----	-----
<i>Elymus scaber</i>	N	H	+++-----	---+-----	-----	-----	-----	---+-----
<i>Enneapogon gracilis</i>	N	H	-----	-----	-----	+++++-----	-----	-----
<i>Enteropogon acicularis</i>	N	H	-----	-----	-----	+++++-----	-----	+++++-----
<i>Eragrostis cilianensis</i>	E	T	-----	-----	-----	+++-----	-----	---+-----
<i>Eragrostis leptostachya</i>	N	H	+++++-----	+++++-----	-----	+++++-----	+++++-----	+++++-----
<i>Hordeum leporinum</i>	E	T	+-----	-----	++-----	+++---+---	-----	+++---+---
<i>Lolium perenne</i>	E	H	---+---+---	---+---+---	+++---+---	+++---+---	+++++-----	---+---+---
<i>Microlaena stipoides</i>	N	H	---+---+---	-----	-----	-----	-----	-----
<i>Panicum effusum</i>	N	H	-----	---+---+---	---+---+---	---+---+---	+++++-----	---+---+---
<i>Paspalum dilatatum</i>	E	H	-----	---+---+---	-----	-----	-----	-----

<i>Rostraria cristata</i>	E	T	-----	-----	-----	-----	+++--+--+	+--+--++
<i>Sporobolus creber</i>	N	H	+++++++	+++++++	-----	++++--+	+++++++	+--+--+
<i>Tragus australianus</i>	N	T	-----	-----	-----	-----	-----	-----
<i>Urochloa panicoides</i>	E	T	++-----	-----	-----	++--++++	++-----	-----
<i>Vulpia myuros</i>	E	T	+++--+---	++-----	+-----	-----	+++--+--+	++-----+
Dicotyledons								
AMARANTHACEAE								
<i>Alternanthera nana</i>	N	T	-----	--+--+--	-----	-----	-----	-----
<i>Alternanthera pungens</i>	E	H	-----	-----	-----	+-----	-----	-----
<i>Amaranthus macrocarpus</i>	E	T	-----	-----	--+-----	-----	-----	-----
<i>Amaranthus retroflexus</i>	E	T	-----	-----	-----	++-----	-----	--+-----
APIACEAE								
<i>Cyclospermum leptophyllum</i>	E	T	--+-----+	++-----	--+--+--+	-----	--+--+---	-----
<i>Daucus glochidiatus</i>	N	T	+-----	+++++--+	++--++++	++--++-	+++++++	+-----++
<i>Hydrocotyle laxiflora</i>	N	H	-----	++-----+	-----	-----	-----	-----
ASTERACEAE								
<i>Bidens subalternans</i>	E	T	-----	-----	-----	+++-----	-----	-----
<i>Calotis lappulacea</i>	N	H	-----	-----	-----	+++++++	+++--+---	+++++++
<i>Carthamus lanatus</i>	E	T	+++++++	+++-----	+++++++	-----+	+++++++	+++++++
<i>Chrysocephalum apiculatum</i>	N	H	-----	+++++++	-----	-----	-----	-----
<i>Cirsium vulgare</i>	E	T	+++++--+	+++++++	+++++---	--+---	++-----	-----
<i>Conyza bonariensis</i>	E	H	-----	+++++--+	--+-----	+++++--+	--+-----	--+-----
<i>Conyza sumatrensis</i>	E	H	--+++--+	-----	-----	-----	-----	-----
<i>Cotula australis</i>	N	T	-----+	++--+--+	-----	---+--+	+-----+	---+--+
<i>Cymbonotus lawsonianus</i>	N	H	-----	+++++++	+--+-----	-----	+++++++	+++--+---
<i>Euchiton involucratus</i>	N	H	-----	+++++++	++--+--+	+++++--+	+++++--+	+-----
<i>Glossocardia bidens</i>	N	H	-----	--+-----	-----	+-----	-----	-----
<i>Hedynois rhagadioloides</i> <i>subsp. cretica</i>	E	T	-----	-----+	++--++++	-----	++--++++	-----+
<i>Hypochaeris microcephala</i>	E	H	-----	---+--+	-----	-----	-----	-----
<i>Hypochaeris radicata</i>	E	H	+++--+--+	++--+--+	++--+--+	---+--+	++--+--+	++--+--+
<i>Lactuca serriola</i>	E	T	-----	++--+--+	--+-----	--+-----	-----	-----
<i>Onopordum acanthium subsp.</i> <i>acanthium</i>	E	T	+-----	-----	-----	-----	-----	-----
<i>Schkuhria pinnata var.</i> <i>abrotanoides</i>	E	T	-----	-----	-----	+++++---	-----	-----
<i>Silybum marianum</i>	E	T	-----	-----	++--++++	---+--+	-----	++-----+
<i>Solenogyne bellioides</i>	N	H	-----	-----+	-----	-----	-----	---+---
<i>Sonchus oleraceus</i>	E	T	-----	--+-----+	++--+--+	+++++--+	-----	-----++
<i>Stuartina hamata</i>	N	T	-----	-----	-----	-----	-----	+-----+
<i>Taraxacum officinale</i>	E	H	-----	++--+---	-----	---+---	+-----	++-----
<i>Triptilodiscus pygmaeus</i>	N	T	-----	-----	-----	-----	-----	+-----+
<i>Vittadinia muelleri</i>	N	H	-----	-----	---+---	-----	+++++++	+++++++
<i>Vittadinia sulcata</i>	N	T	-----	-----	-----	+++++++	++--++++	-----
<i>Xanthium spinosum</i>	E	T	+-----	+++++---	+++++++	-----	-----	---+---
BRASSICACEAE								
<i>Capsella bursa-pastoris</i>	E	H	-----	++--+--+	---+--+	---+--+	-----	---+--+
<i>Lepidium africanum</i>	E	H	-----	+++++++	-----	+++++--+	--+-----	+++++---
<i>Sisymbrium irio</i>	E	T	-----	-----	-----+	+-----+	-----	-----
<i>Sisymbrium officinale</i>	E	T	-----	-----	-----	+-----	-----	-----
CACTACEAE								
<i>Opuntia stricta</i>	E	Ph	-----	-----	-----	-----	-----	-----+++++
CAMPANULACEAE								
<i>Wahlenbergia communis</i>	N	H	-----	+++++---	-----	-----+	---+--+	---+--+
CARYOPHYLLACEAE								
<i>Arenaria leptoclados</i>	E	T	-----+	-----	-----+	-----	-----	-----+
<i>Cerastium glomeratum</i>	E	T	-----	-----+	-----	-----	-----	-----
<i>Paronychia brasiliana</i>	E	H	--+++---	+++++---	-----	-----	-----	-----
<i>Petrorhagia nanteuilii</i>	E	T	+++++---	+++++---	--+--+---	-----	----+---	-----
<i>Polycarpon tetraphyllum</i>	E	T	-----	--+-----	-----	-----	--+-----	--+--+
<i>Silene gallica</i>	E	T	--+-----	-----	-----	--+-----	-----	-----
CHENOPODIACEAE								
<i>Atriplex spinibractea</i>	N	Ph	-----	-----	-----	---+---	-----	-----+---
<i>Chenopodium melanocarpum</i>	N	T	-----	-----	---+---	-----	-----	-----

<i>Chenopodium pumilio</i>	N	T	-----+++	+-----	-----	-----	-----	--+----+
<i>Einadia nutans</i>	N	C	-----	+++++++	-----	+++++++	+++++++	+++++++
<i>Maireana microphylla</i>	N	Ph	-----	-----	-----	+++++++	-----	-----
<i>Salsola kali</i>	N	T	-----	-----	-----	+++++	-----	-----
CONVOLVULACEAE								
<i>Convolvulus erubescens</i>	N	H	-----	-++-+-	-++-+-	-----	-++-+-	-----
<i>Dichondra repens</i>	N	H	+++++--++	+++++++	+++++++	+++++++	+++++	+++++
CRASSULACEAE								
<i>Crassula sieberiana</i>	N	T	-----	++-----	-+-----	+-----	+-----	+-----
CUCURBITACEAE								
<i>Cucumis myriocarpus</i>	E	T	-----	-----	-----	-----	+-----	-----
EUPHORBIACEAE								
<i>Chamaesyce drummondii</i>	N	H	-----	++++-++-	+++++++	-----	+++-----	-----
FABACEAE								
<i>Glycine tabacina</i>	N	H	-++-+-	++++-++-	-----	-----	-----	-++-+-
<i>Medicago laciniata</i>	E	T	-----	-----	-----	-----	-----	-----
<i>Medicago lupulina</i>	E	H	-----	-----	-----	-----	-++-+-	-----
<i>Medicago minima</i>	E	T	-----	-----	++-++-+	-----	-----	-----
<i>Medicago polymorpha</i>	E	T	---++-+	++++-++-	++-++-+	+++++++	++-++-+	++-++-+
<i>Medicago truncatula</i>	E	T	-----	-----	-----	-++-+-	-----	-----
<i>Trifolium angustifolium</i>	E	T	-----	-----	-----	-----	-----	-----
<i>Trifolium arvense</i>	E	T	-----	-----	-+-----	-+-----	-++-+-	-+-----
<i>Trifolium campestre</i>	E	T	-+-----	-+-----	-+-----	-----	-----	-----
<i>Trifolium glomeratum</i>	E	T	-+-----	-+-----	-----	-++-+-	++++-++-	-----
<i>Trifolium repens</i>	E	H	-----	-----	-----	-----	-----	-----
<i>Trifolium subterraneum</i>	E	T	++++-++-	-+-----	-+-----	-----	-----	++++-++-
<i>Trifolium tomentosum</i>	E	T	-----	-----	-+-----	-----	-----	-+-----
<i>Trifolium vesiculosum</i>	E	T	-----	-----	-+-----	-----	-----	-----
GENTIANACEAE								
<i>Centaurium erythraea</i>	E	T	-+-----	-+-----	-+-----	-+-----	-+-----	-----
<i>Erodium cicutarium</i>	E	H	-----	+-----	-----	-----	-----	-----
<i>Erodium crinitum</i>	E	H	-----	-----	-----	-----	-----	++++-++-
<i>Geranium solanderi</i>	N	H	++++-++-	++++-++-	-+-----	-----	-----	+-----
GOODENIACEAE								
<i>Goodenia pinnatifida</i>	N	H	-----	-++-+-	-----	-----	-----	-----
<i>Scaevola humilis</i>	N	H	-----	-+-----	-----	-----	-----	-----
LAMIACEAE								
<i>Lamium amplexicaule</i>	E	T	-----	-----	++-++-+	++++-++-	-----	-----
<i>Marrubium vulgare</i>	E	Ph	-----	-----	-+-----	-+-----	-----	-----
<i>Mentha saturoioides</i>	N	H	-----	+++--+-	++++-++-	-----	+++--+-	-----
<i>Salvia reflexa</i>	E	T	-----	-----	-----	-+-----	-----	-----
<i>Salvia verbenaca</i>	E	H	-----	---++-+	-----	-----	-----	-----
MALVACEAE								
<i>Malva parviflora</i>	E	T	-----	-----	-+-----	-+-----	-----	++++-++-
<i>Modiola caroliniana</i>	E	C	++++-++-	-----	-----	-----	-----	-----
<i>Sida corrugata</i>	N	H	-----	-----	-----	-----	-----	---++-+
<i>Sida spinosa</i>	E	T	-----	-----	-----	-----	-----	+++--+-
MYOPORACEAE								
<i>Eremophila debilis</i>	N	Ph	-----	---+-----	-----	-----	-----	+-----
NYCTAGINACEAE								
<i>Boerhavia dominii</i>	N	H	-----	-----	-++-+-	-++-+-	-----	-++-+-
OXALIDACEAE								
<i>Oxalis corniculata</i>	N	C	+++--+-	++++-++-	++++-++-	+++--+-	+++--+-	+++--+-
PLANTAGINACEAE								
<i>Plantago debilis</i>	N	H	-----	++++-++-	++++-++-	-+-----	-+-----	++++-++-
<i>Plantago lanceolata</i>	E	H	-----	-----	-+-----	-----	-----	-----
POLYGANACEAE								
<i>Rumex brownii</i>	N	H	++++-++-	++++-++-	---+-----	-----	+-----	++++-++-
<i>Polygonum aviculare</i>	E	c	++++-++-	+++--+-	-+-----	-+-----	-----	-----
PORTULACACEAE								
<i>Portulaca oleracea</i>	N	T	-----	-----	-----	-----	-----	-+-----
PRIMULACEAE								
<i>Anagallis arvensis</i>	E	T	-+-----	+-----	++-++-+	-----	-----	+-----
RANUNCULACEAE								

<i>Ranunculus sessiliflorus</i>	N	H	-----	-----	-----	-----+	-----	-----
ROSACEAE								
<i>Acaena novae-zelandiae</i>	N	C	-----	-----	-----	-----	-----	+---+---
RUBIACEAE								
<i>Asperula conferta</i>	N	C	-----	++++++++	++++++++	-----	-----	-----
SCROPHULARIACEAE								
<i>Kickxia spuria subsp. integrifolia</i>	E	H	-----	-----	--+-----	-----	-----	-----
<i>Linaria arvensis</i>	E	T	-----	-+-----+	-----	-----	-----	-----
<i>Verbascum thapsus</i>	E	T	+++-----	-----	-----	-----	-----	-----
<i>Veronica arvensis</i>	E	T	-----	-----	-----	-----+	-----	-----
<i>Veronica peregrina</i>	E	T	-----	-+-----	-----	-----	-----	-----
URTICACEAE								
<i>Urtica incisa</i>	N	H	-----	-----	-----	-+-----	-----	-----
ZYGOPHYLLACEAE								
<i>Tribulus terrestris</i>	E	T	-----	-----	-----+--	--++-+--	-----	--++-+--

APPENDIX 3

Species list for taxa represented in the farm case study (Chapter 4), Showing origin (O, N = native, E = exotic) and species occurrence frequency in each of six land uses (U = ungrazed woodland, G = grazed woodland, P = native pasture, R = roadside, C = crop, D = dry creek bed).

Species	O	U	G	P	R	C	D
Pteridophytes							
ADIANTACEAE							
<i>Cheilanthes</i>	N	1	1	1	-	-	-
<i>Cheilanthes sieberi</i>	N	1	4	1	1	-	1
Cycads							
ZAMIACEAE							
<i>Macrozamia</i>	N	-	1	-	-	-	-
Monocotyledons							
ANTHERICACEAE							
<i>Arthropodium minus</i>	N	2	1	-	-	-	-
<i>Dichopogon</i>	N	1	3	1	1	-	1
CYPERACEAE							
<i>Carex breviculmis</i>	N	1	2	-	-	-	-
<i>Carex inversa</i>	N	2	3	5	4	-	2
<i>Cyperus gracilis</i>	N	2	5	1	-	-	2
<i>Scleria mackaviensis</i>	N	-	1	-	-	-	-
IRIDACEAE							
<i>Sisyrinchium sp. A</i>	N	-	2	-	-	-	-
JUNCACEAE							
<i>Juncus usitatus</i>	N	-	-	2	-	1	-
ORCHIDACEAE							
<i>Pterostylis sp.</i>	N	1	-	-	-	-	-
PHORMIACEAE							
<i>Dianella longifolia</i>	N	3	-	-	1	-	-
POACEAE							
<i>Aristida leptopoda</i>	N	1	-	-	-	-	-
<i>Aristida ramosa</i>	N	3	5	5	5	3	2
<i>Austroanthonia</i>	N	-	1	4	4	3	-
<i>Austroanthonia</i>	N	3	5	2	2	-	2
<i>Austroanthonia</i>	N	-	2	1	-	-	-
<i>Austrostipa</i>	N	-	-	-	3	-	-
<i>Austrostipa scabra</i>	N	3	5	5	4	1	2
<i>Austrostipa setacea</i>	N	-	-	-	2	-	-
<i>Austrostipa</i>	N	1	5	3	-	2	2
<i>Avena barbata</i>	E	-	-	1	5	3	-
<i>Bothriochloa macra</i>	N	3	5	5	4	1	2
<i>Briza minor</i>	E	-	-	1	-	-	-
<i>Bromus catharticus</i>	E	-	-	1	3	-	1
<i>Bromus molliformis</i>	E	-	1	4	5	3	1
<i>Chloris truncata</i>	N	-	-	4	2	3	1
<i>Chloris ventricosa</i>	N	3	2	2	-	1	2
<i>Cymbopogon</i>	N	3	2	-	-	-	-
<i>Cynodon dactylon</i>	N	-	1	2	2	2	-
<i>Dichanthium</i>	N	2	2	5	5	2	1
<i>Dichelachne</i>	N	-	2	1	1	-	1
<i>Elymus scaber</i>	N	1	2	3	4	-	2
<i>Enneapogon</i>	N	1	1	-	-	-	-
<i>Enteropogon</i>	N	1	1	-	-	-	-
<i>Eragrostis</i>	N	-	-	2	-	-	-
<i>Eriochloa</i>	N	-	-	-	1	-	-
<i>Hordeum leporinum</i>	E	-	-	3	2	7	-
<i>Hyparrhenia hirta</i>	E	-	-	-	3	-	-
<i>Lolium perenne</i>	E	1	2	5	5	7	-
<i>Panicum effusum</i>	N	1	1	-	1	-	2
<i>Paspalidium gracile</i>	N	3	4	-	-	-	1
<i>Paspalum dilatatum</i>	E	-	-	-	5	-	-
<i>Poa sieberiana var. sieberiana</i>	N	3	2	-	-	-	-
<i>Rostraria cristata</i>	E	-	-	4	2	3	-
<i>Sarga leiocladum</i>	E	2	-	-	1	-	-
<i>Sorghum bicolor</i>	E	1	-	-	-	1	-
<i>Sporobolus creber</i>	N	2	1	5	3	-	1
<i>Themeda triandra</i>	N	1	-	-	-	-	-
<i>Triticum aestivum</i>	E	-	-	-	-	2	-
<i>Vulpia myuros</i>	E	-	-	4	3	-	-
XANTHORRHOEACEAE							
<i>Lomandra filiformis</i>	N	3	2	-	-	-	1
<i>Lomandra longifolia</i>	N	2	3	-	2	-	-
<i>Lomandra multiflora</i>	N	2	4	-	1	-	1
Dicotyledons							
ACANTHACEAE							
<i>Brunoniella australis</i>	N	3	3	-	-	-	2
<i>Rostellularia</i>	N	2	-	-	-	-	1
AMARANTHACEAE							
<i>Alternanthera</i>	E	-	-	1	-	-	-
APIACEAE							
<i>Cyclospermum</i>	E	-	-	4	3	5	-
<i>Daucus glochidiatus</i>	N	1	1	5	4	-	1
<i>Foeniculum vulgare</i>	E	-	-	-	1	-	-
<i>Hydrocotyle laxiflora</i>	N	1	3	1	-	-	1
ASCLEPIADACEAE							
<i>Marsdenia viridiflora</i>	N	-	-	-	-	-	1
ASTERACEAE							
<i>Brachyscome</i>	N	-	2	-	-	-	-
<i>Calotis hispidula</i>	N	-	-	3	-	-	-
<i>Calotis lappulacea</i>	N	2	2	4	1	3	-
<i>Carthamus lanatus</i>	E	-	-	5	2	2	1
<i>Centaurea solstitialis</i>	E	1	-	3	4	3	1
<i>Chrysocephalum</i>	N	2	-	-	3	-	1
<i>Cirsium vulgare</i>	E	1	1	-	-	-	-
<i>Conyza bonariensis</i>	E	-	-	2	-	1	-
<i>Cotula australis</i>	N	-	-	2	-	2	-
<i>Cymbonotus</i>	N	1	1	3	1	-	2
<i>Euchiton</i>	N	2	-	4	2	1	2
<i>Hedypnois</i>	E	-	-	3	4	2	1
<i>subsp. cretica</i>							
<i>Hypochaeris</i>	E	-	-	1	-	-	-
<i>Hypochaeris radicata</i>	E	-	-	5	1	-	-
<i>Lactuca saligna</i>	E	-	-	-	1	-	1
<i>Microseris</i>	N	1	-	-	-	-	-
<i>Senecio</i>	N	-	-	-	2	-	1
<i>Silybum marianum</i>	E	-	-	-	-	2	-
<i>Solenogyne</i>	N	1	2	-	-	-	2
<i>Soliva sessilis</i>	E	-	-	1	-	-	-
<i>Sonchus oleraceus</i>	E	-	-	2	2	3	-
<i>Vittadinia muelleri</i>	N	3	4	3	3	3	2
<i>Xanthium spinosum</i>	E	-	1	-	-	2	-
<i>Cassinia laevis</i>	N	1	1	-	-	-	-

ASTERACEAE							
<i>Vittadinia sulcata</i>	N	-	1	4	3	2	2
BIGNONIACEAE							
<i>Pandorea pandorana</i>	N	1	-	-	-	-	-
<i>Pandorea pandorana pandorana</i>	N	-	2	-	-	-	-
BORAGINACEAE							
<i>Buglossoides</i>	E	-	-	-	1	1	-
<i>Echium</i>	E	-	-	-	4	-	-
BRASSICACEAE							
<i>Capsella bursa-</i>	E	-	-	1	1	4	-
<i>Lepidium africanum</i>	E	1	1	2	-	4	-
<i>Rapistrum rugosum</i>	E	-	-	-	1	-	-
<i>Sisymbrium</i>	E	-	-	-	2	3	-
CACTACEAE							
<i>Opuntia aurantiaca</i>	E	-	1	-	-	-	1
<i>Opuntia stricta</i>	E	-	-	1	1	1	1
CAMPANULACEAE							
<i>Wahlenbergia</i>	N	2	3	5	2	-	2
<i>Wahlenbergia</i>	N	-	-	-	-	-	1
CARYOPHYLLACEAE							
<i>Arenaria leptoclados</i>	E	-	1	2	3	-	-
<i>Paronychia</i>	E	-	-	1	-	1	-
<i>Petrorhagia</i>	E	1	2	4	3	-	1
<i>Polycarpon</i>	E	-	-	2	-	3	-
CHENOPODIACEAE							
<i>Atriplex suberecta</i>	N	-	-	1	1	2	-
<i>Einadia hastata</i>	N	-	2	-	-	1	-
<i>Einadia nutans</i>	N	1	4	5	2	5	1
<i>Maireana</i>	N	1	-	-	1	-	-
<i>Salsola kali</i>	N	-	-	2	1	4	-
<i>Maireana</i>	N	-	2	3	3	4	-
CLUSIACEAE							
<i>Hypericum</i>	N	-	1	-	-	-	-
CONVOLVULACEAE							
<i>Convolvulus</i>	N	2	1	2	2	1	2
<i>Dichondra repens</i>	N	3	5	5	2	1	2
CRASSULACEAE							
<i>Crassula sieberiana</i>	N	-	1	5	-	5	1
EUPHORBIACEAE							
<i>Chamaesyce</i>	N	-	1	5	-	2	-
<i>Phyllanthus virgatus</i>	N	-	-	1	-	-	-
FABACEAE							
<i>Glycine clandestina</i>	N	1	3	-	-	-	-
<i>Glycine tabacina</i>	N	3	5	3	5	-	2
<i>Hardenbergia</i>	N	1	-	-	-	-	-
<i>Cullen tenax</i>	N	-	-	-	3	-	-
<i>Desmodium</i>	N	3	4	-	-	-	2
<i>Desmodium varians</i>	N	2	-	-	-	-	-
<i>Lotus australis</i>	N	-	-	-	1	-	-
<i>Medicago lupulina</i>	E	-	-	3	1	-	-
<i>Medicago</i>	E	3	3	5	5	3	2
<i>Medicago sativa</i>	E	-	-	-	1	5	-
<i>Swainsona</i>	N	-	1	1	1	-	-
<i>Swainsona reticulata</i>	N	-	-	-	1	-	-
<i>Trifolium</i>	E	-	-	-	3	-	-
<i>Trifolium arvense</i>	E	-	1	5	5	1	1
<i>Trifolium</i>	E	-	-	5	1	1	-
<i>Senna occidentalis</i>	E	-	-	-	2	-	1
GENTIANACEAE							
<i>Geranium solanderi</i>	N	-	1	3	5	-	-
GOODENIACEAE							
<i>Goodenia pinnatifida</i>	N	-	-	1	1	-	-
LAMIACEAE							
<i>Ajuga australis</i>	N	3	1	-	-	-	-
<i>Marrubium vulgare</i>	E	-	-	1	2	3	-
<i>Mentha satureioides</i>	N	1	-	1	3	-	1
LAMIACEAE							
<i>Salvia reflexa</i>	E	-	-	1	3	-	-
<i>Teucrium sp. D</i>	N	1	2	-	-	-	-
MALVACEAE							
<i>Malva parviflora</i>	E	-	-	1	-	5	-
<i>Madiola caroliniana</i>	E	-	-	1	1	-	-
<i>Sida corrugata</i>	N	1	4	4	1	-	1
<i>Sida cunninghamii</i>	N	1	-	-	-	-	-
MALVACEAE							
<i>Sida trichopoda</i>	N	1	-	-	-	-	1
<i>Sida spinosa</i>	E	2	1	2	1	1	-
MYOPORACEAE							
<i>Eremophila debilis</i>	N	1	-	-	-	-	-
NYCTAGINACEAE							
<i>Boerhavia dominii</i>	N	1	2	5	-	3	-
OLEACEAE							
<i>Jasminum</i>	N	1	1	-	1	-	-
<i>Notelaea microcarpa microcarpa</i>	N	2	2	-	-	-	-
OXALIDACEAE							
<i>Oxalis corniculata</i>	N	2	5	5	3	3	2
PITOSPORACEAE							
<i>Bursaria spinosa</i>	N	-	3	-	1	-	2
PLANTAGINACEAE							
<i>Plantago debilis</i>	N	1	2	5	-	1	2
<i>Plantago lanceolata</i>	E	-	1	-	2	-	-
POLYGANACEAE							
<i>Rumex brownii</i>	N	-	2	5	3	-	2
<i>Polygonum aviculare</i>	E	-	-	-	-	3	-
PRIMULACEAE							
<i>Anagallis arvensis</i>	E	-	2	1	3	-	2
RUBIACEAE							
<i>Asperula conferta</i>	N	3	1	1	3	-	-
<i>Galium aparine</i>	E	1	1	-	2	1	-
<i>Galium migrans</i>	N	-	-	-	-	-	1
SAPINDACEAE							
<i>Dodonaea sinuolata sinuolata</i>	N	-	-	-	-	-	1
<i>Dodonaea viscosa angustissima</i>	N	1	3	-	-	-	1
SCROPHULARIACEAE							
<i>Veronica arvensis</i>	E	-	-	4	1	-	-
SOLANACEAE							
<i>Solanum nigrum</i>	E	-	-	1	-	-	-
THYMELAEACEAE							
<i>Pimelea curviflora</i>	N	-	-	-	-	-	1
<i>Pimelea neo-anglica</i>	N	2	-	-	-	-	-
URTICACEAE							
<i>Urtica incisa</i>	N	-	-	2	-	-	-
VERBENACEAE							
<i>Verbena bonariensis</i>	E	-	1	1	3	-	-
ZYGOPHYLLACEAE							
<i>Tribulus terrestris</i>	E	-	-	-	-	2	-

APPENDIX 4

Species list for taxa represented in regional survey dataset (Chapter 5), showing origin (O, N = native, E = exotic), life-form (LF, Pt = pteridophyte, Gl = grass like, G = Grass, F = forb, C = climber, LS = low shrub, G = geophytes), life cycle (LC, PE = perennial, PA = perennial with annual shoots, SP = short-lived perennial, A = annual), and land use (PC = pasture, previously cultivated, PU = pasture, never cultivated, G = woodland grazed by livestock, U = woodland not grazed by livestock)

Species	O	LF	LC	PC	PU	G	U											
PTERIDOPHYTES																		
ADIANTACEAE																		
<i>Cheilanthes distans</i>	N	Pt	PE	0	0	0	5											
<i>Cheilanthes lasiophylla</i>	N	Pt	PE	0	0	1	1											
<i>Cheilanthes sieberi</i>	N	Pt	PE	6	13	14	23											
MONOCOTYLEDON																		
ANTHERICACEAE																		
<i>Arthropodium fimbriatum</i>	N	Gl	PA	6	11	15	9											
<i>Arthropodium minus</i>	N	Gl	PA	2	3	8	4											
<i>Arthropodium sp. B</i>	N	Gl	PA	0	0	1	0											
<i>Dichopogon fimbriatus</i>	N	Gl	PA	0	1	1	5											
<i>Tricoryne elatior</i>	N	Gl	PA	0	1	3	0											
ASPHODELACEAE																		
<i>Asphodelus fistulosus</i>	E	Gl	AN	0	0	0	1											
<i>Bulbine bulbosa</i>	N	Gl	PE	0	2	3	5											
COLCHICACEAE																		
<i>Wurmbea biglandulosa</i>	N	Gl	PA	1	3	5	2											
COMMELINACEAE																		
<i>Commelina cyanea</i>	N	Gl	PE	0	0	2	1											
CYPERACEAE																		
<i>Carex breviculmis</i>	N	Gl	PE	0	0	0	1											
<i>Carex incomitata</i>	N	Gl	PE	0	0	0	1											
<i>Carex inversa</i>	N	Gl	PE	26	23	26	15											
<i>Cyperus fulvus</i>	N	Gl	PE	0	0	1	1											
<i>Cyperus gracilis</i>	N	Gl	PE	2	4	18	16											
<i>Fimbristylis dichotoma</i>	N	Gl	PE	0	0	0	1											
<i>Lepidosperma laterale</i>	N	Gl	PE	0	0	0	5											
<i>Schoenus apogon</i>	N	Gl	PE	0	0	0	1											
<i>Schoenus subaphyllus</i>	N	Gl	PE	0	1	0	0											
<i>Scleria mackaviensis</i>	N	Gl	PE	0	1	2	6											
HYPOXIDACEAE																		
<i>Hypoxis hygrometrica</i>	N	F	PE	0	1	1	0											
IRIDACEAE																		
<i>Romulea minutiflora</i>	E	F	PA	0	0	0	0											
<i>Romulea rosea</i>	E	F	PA	1	0	0	0											
<i>Sisyrinchium sp. A</i>	E	Gl	AN	0	0	1	0											
JUNCACEAE																		
<i>Juncus bufonius</i>	E	Gl	AN	1	0	1	0											
<i>Juncus capitatus</i>	E	Gl	AN	0	1	0	0											
<i>Juncus continuus</i>	N	Gl	PE	1	0	0	0											
<i>Juncus filicaulis</i>	N	Gl	PE	0	0	1	0											
<i>Juncus firmus</i>	N	Gl	PE	0	0	1	0											
<i>Juncus homalocalis</i>	N	Gl	PE	0	0	2	0											
<i>Juncus pauciflorus</i>	N	Gl	PE	1	0	1	0											
<i>Juncus subglauca</i>	N	Gl	PE	1	0	1	0											
<i>Juncus subsecundus</i>	N	Gl	PE	1	0	0	0											
<i>Juncus usitatus</i>	N	Gl	PE	7	6	6	1											
<i>Luzula densiflora</i>	N	Gl	PE	0	0	1	1											
<i>Luzula flaccida</i>	N	Gl	PE	0	1	0	1											
LUZURIAGACEAE																		
<i>Eustrephus latifolius</i>	N	C	PE	0	0	0	3											
ORCHIDACEAE																		
<i>Microtis unifolia</i>	N	Gl	PA	1	2	2	0											
<i>Pterostylis curta</i>	N	Gl	PA	0	0	0	1											
PHORMIACEAE																		
<i>Dianella caerulea var. cinerascens</i>	N	Gl	PE	0	0	0	3											
<i>Dianella longifolia var. longifolia</i>	N	Gl	PE	0	0	1	8											
<i>Dianella revoluta var. revoluta</i>	N	Gl	PE	1	1	1	8											
<i>Dianella revoluta var. vinosa</i>	N	Gl	PE	0	0	0	1											
POACEAE																		
<i>Aira cupaniana</i>	E	G	AN	0	3	4	3											
<i>Anthoxanthum odoratum</i>	E	G	PE	1	0	0	0											
<i>Aristida acuta</i>	N	G	PE	0	0	0	1											
<i>Aristida caput-medusa</i>	N	G	PE	0	0	1	0											
<i>Aristida jerichoensis</i>	N	G	PE	0	0	1	0											
<i>Aristida leptopoda</i>	N	G	PE	0	0	1	0											
<i>Aristida ramosa</i>	N	G	PE	30	23	18	26											
<i>Aristida vagans</i>	N	G	PE	0	0	0	1											
<i>Austrodanthonia bipartita</i>	N	G	PE	23	10	8	9											
<i>Austrodanthonia caespitosa</i>	N	G	PE	9	4	7	1											
<i>Austrodanthonia racemosa var. obtusata</i>	N	G	PE	1	1	0	3											
<i>Austrodanthonia racemosa var. racemosa</i>	N	G	PE	4	5	8	15											
<i>Austrostipa scabra</i>	N	G	PE	24	17	22	21											
<i>Austrostipa setacea</i>	N	G	PE	1	1	1	1											
<i>Austrostipa verticillata</i>	N	G	PE	9	8	17	8											
<i>Avena barbata</i>	E	G	AN	10	2	1	1											
<i>Avena sativa</i>	E	G	AN	1	0	0	0											
<i>Bothriochloa macra</i>	N	G	PE	35	20	17	17											
<i>Briza minor</i>	E	G	AN	7	6	4	0											

<i>Isoetopsis graminifolia</i>	N	F	AN	1	0	0	1	<i>Cerastium glomeratum</i>	E	F	AN	11	6	6	0
<i>Lactuca saligna</i>	E	F	SP	15	9	6	2	<i>Gypsophila tubulosa</i>	E	F	AN	1	0	0	0
<i>Lactuca serriola</i>	E	F	SP	11	6	14	3	<i>Paronychia brasiliana</i>	E	F	PE	14	3	8	0
<i>Leiocarpa panaetioides</i>	N	F	PE	1	0	0	1	<i>Petrorhagia nanteuillii</i>	E	F	AN	22	24	20	12
<i>Leptorhynchus squamatus</i>	N	F	PE	0	0	1	1	<i>Polycarpon tetraphyllum</i>	E	F	AN	13	10	7	0
<i>Microseris lanceolata</i>	N	F	PA	1	0	0	2	<i>Sagina apetala</i>	E	F	AN	0	6	2	2
<i>Olearia elliptica subsp. elliptica</i>	N	LS	PE	0	2	2	12	<i>Silene gallica</i>	E	F	AN	1	1	1	0
<i>Olearia viscidula</i>	N	LS	PE	1	1	3	3	<i>Spergularia rubra</i>	E	F	SP	2	0	1	0
<i>Onopordum acanthium</i>	E	F	AN	5	1	2	0	<i>Stellaria media</i>	E	F	SP	1	0	3	1
<i>Pseudognaphalium luteoalbum</i>	N	F	AN	2	1	0	1	CHENOPODIACEAE							
<i>Rhodanthe anthemoides</i>	N	F	PE	0	0	1	0	<i>Atriplex spinibractea</i>	N	F	PE	1	2	1	0
<i>Schkuhria pinnata var. abrotanoides</i>	E	F	AN	2	1	1	0	<i>Chenopodium album</i>	E	F	AN	0	1	0	0
<i>Senecio diaschides</i>	N	F	PE	1	0	0	0	<i>Chenopodium pumilio</i>	N	F	AN	2	3	5	3
<i>Senecio madagascariensis</i>	E	F	SP	1	1	1	3	<i>Einadia hastata</i>	N	F	PE	4	1	9	4
<i>Senecio quadridentatus</i>	N	F	PE	5	4	7	3	<i>Einadia nutans</i>	N	F	PE	20	11	12	11
<i>Sigesbeckia australiensis</i>	N	F	AN	1	1	8	8	<i>Einadia trigonos</i>	N	F	PE	1	0	3	1
<i>Silybum marianum</i>	E	F	SP	12	6	5	2	<i>Maireana enchylaenoides</i>	N	F	PE	0	0	0	1
<i>Solenogyne bellioides</i>	N	F	PE	0	4	12	6	<i>Maireana microphylla</i>	N	LS	PE	7	3	5	4
<i>Soliva sessilis</i>	E	F	AN	6	6	5	1	<i>Salsola kali</i>	N	F	AN	3	3	0	1
<i>Sonchus asper</i>	E	F	AN	0	1	1	0	<i>Sclerolaena birchii</i>	N	F	PE	3	0	0	0
<i>Sonchus oleraceus</i>	E	F	AN	22	19	22	9	CLUSIACEAE							
<i>Stuartina hamata</i>	N	F	AN	1	5	2	3	<i>Hypericum gramineum</i>	N	F	PE	3	4	12	10
<i>Taraxacum officinale</i>	E	F	PE	6	1	3	1	<i>Hypericum perforatum</i>	E	F	PE	0	1	0	0
<i>Triptilodiscus pygmaeus</i>	N	F	AN	5	6	1	1	CONVOLVULACEAE							
<i>Vittadinia cervicularis</i>	N	F	PE	0	0	0	3	<i>Convolvulus erubescens</i>	N	F	PE	8	8	1	2
<i>Vittadinia cuneata var. hirsuta</i>	N	F	PE	0	0	2	4	<i>Dichondra repens</i>	N	F	PE	19	21	26	20
<i>Vittadinia muelleri</i>	N	F	PE	19	12	10	13	<i>Dichondra sp. A</i>	N	F	PE	0	0	0	9
<i>Vittadinia sulcata</i>	N	F	AN	5	5	5	7	<i>Evolvulus alsinoides var. decumbens</i>	N	F	PE	0	0	0	3
<i>Xanthium spinosum</i>	E	F	AN	2	0	0	0	CRASSULACEAE							
<i>Xerochrysum bracteatum</i>	N	F	PE	0	2	2	0	<i>Crassula sieberiana</i>	N	F	AN	21	13	13	8
<i>Zinnia peruviana</i>	E	F	AN	1	0	0	0	CUCURBITACEAE							
BIGNONIACEAE								<i>Cucumis myriocarpus</i>	E	F	AN	0	0	1	0
<i>Pandorea pandorana</i>	N	C	PE	0	0	1	3	DILLENIACEAE							
<i>Amsinckia intermedia</i>	E	F	AN	0	1	0	0	<i>Hibbertia obtusifolia</i>	N	LS	PE	0	3	3	4
<i>Anchusa arvensis</i>	E	F	AN	0	1	0	0	<i>Hibbertia riparia</i>	N	LS	PE	0	0	0	1
<i>Buglossoides arvensis</i>	E	F	AN	5	2	6	3	EPACRIDACEAE							
<i>Cynoglossum australe</i>	N	F	PE	0	0	0	1	<i>Lissanthe strigosa</i>	N	LS	PE	0	1	0	2
<i>Cynoglossum suaveolens</i>	N	F	PE	0	1	0	2	<i>Melichrus urceolatus</i>	N	LS	PE	0	1	1	8
<i>Echium plantagineum</i>	E	F	AN	1	1	2	1	EUPHORBIACEAE							
<i>Echium vulgare</i>	E	F	AN	1	0	0	0	<i>Chamaesyce drummondii</i>	N	F	PE	6	5	10	6
<i>Heliotropium amplexicaule</i>	E	F	PE	1	0	0	0	<i>Phyllanthus gunnii</i>	N	LS	PE	1	4	3	0
BRASSICACEAE								<i>Phyllanthus virgatus</i>	N	F	SP	0	1	0	3
<i>Capsella bursa-pastoris</i>	E	F	AN	24	11	9	1	<i>Poranthera microphylla</i>	N	F	AN	0	1	0	3
<i>Coronopus didymus</i>	E	F	SP	1	0	1	0	FABACEAE							
<i>Lepidium africanum</i>	E	F	AN	21	9	9	3	<i>Acacia decora</i>	N	LS	PE	0	0	0	2
<i>Lepidium bonariense</i>	E	F	SP	4	1	4	1	<i>Acacia juvenile</i>	N	LS	PE	0	0	0	1
<i>Rapistrum rugosum</i>	E	F	SP	1	0	0	0	<i>Cullen tenax</i>	N	F	PE	0	1	0	3
<i>Sisymbrium erysimoides</i>	E	F	SP	2	0	0	0	<i>Desmodium brachypodum</i>	N	F	PE	0	2	7	24
<i>Sisymbrium irio</i>	E	F	SP	2	6	3	0	<i>Desmodium gunnii</i>	N	F	PE	0	0	0	4
<i>Sisymbrium officinale</i>	E	F	SP	6	1	12	0	<i>Desmodium varians</i>	N	F	PE	5	10	13	17
<i>Sisymbrium orientale</i>	E	F	SP	0	0	2	0	<i>Glycine clandestina</i>	N	C	PE	0	2	6	13
CACTACEAE								<i>Glycine tabacina</i>	N	C	PE	13	18	24	28
<i>Opuntia aurantiaca</i>	E	F	PE	0	1	0	10	<i>Glycine tomentella</i>	N	C	PE	0	0	0	1
<i>Opuntia stricta</i>	E	F	PE	4	4	3	8	<i>Hardenbergia violacea</i>	N	C	PE	0	0	0	1
CAMPANULACEAE								<i>Lespedeza juncea subsp. sericea</i>	N	LS	PE	0	1	0	0
<i>Wahlenbergia communis</i>	N	F	PE	26	24	26	24	<i>Lotus australis</i>	N	F	PE	0	0	1	0
<i>Wahlenbergia gracilentia</i>	N	F	PE	1	1	0	3	<i>Lupinus angustifolius</i>	E	F	AN	0	0	0	1
CARYOPHYLLACEAE								<i>Medicago arabica</i>	E	F	AN	0	0	0	0
<i>Arenaria leptoclados</i>	E	F	AN	19	14	11	6								

<i>Medicago laciniata</i>	E	F	AN	1	0	0	0
<i>Medicago lupulina</i>	E	F	SP	17	7	2	1
<i>Medicago minima</i>	E	F	AN	0	0	2	1
<i>Medicago orbicularis</i>	E	F	AN	0	1	2	0
<i>Medicago polymorpha</i>	E	F	AN	27	16	19	6
<i>Medicago sativa</i>	E	F	PE	3	0	1	0
<i>Medicago trunculata</i>	E	F	AN	0	0	1	0
<i>Pultenaea foliolosa</i>	N	LS	PE	0	0	0	1
<i>Pultenaea sp. G</i>	N	LS	PE	0	1	0	3
<i>Rhynchosia minima</i>	N	C	PE	0	0	0	2
<i>Swainsona galegifolia</i>	N	F	PE	4	3	18	14
<i>Swainsona monticola</i>	N	F	PE	0	2	1	0
<i>Swainsona reticulata</i>	N	F	PE	2	0	2	1
<i>Templetonia stenophylla</i>	N	LS	PE	0	0	0	1
<i>Trifolium angustifolium</i>	E	F	AN	10	7	8	2
<i>Trifolium arvense</i>	E	F	AN	26	20	20	8
<i>Trifolium campestre</i>	E	F	AN	15	18	20	6
<i>Trifolium cernuum</i>	E	F	AN	9	3	0	0
<i>Trifolium glomeratum</i>	E	F	AN	32	16	24	5
<i>Trifolium hirtum</i>	E	F	AN	0	3	1	0
<i>Trifolium repens</i>	E	F	PE	17	10	6	1
<i>Trifolium striatum</i>	E	F	AN	5	2	3	0
<i>Trifolium subterraneum</i>	E	F	AN	3	7	0	0
<i>Trifolium tomentosum</i>	E	F	AN	5	5	4	1
<i>Trifolium vesiculosum</i>	E	F	AN	4	4	5	0
<i>Vicia sativa</i>	E	C	AN	1	1	1	0
<i>Zornia dyctiocarpa</i> var. <i>dyctiocarpa</i>	N	F	PE	0	0	5	0
GENTIANACEAE							
<i>Centaurium erythraea</i>	E	F	AN	6	5	4	1
<i>Sebaea ovata</i>	N	F	AN	0	0	1	0
GERANIACEAE							
<i>Erodium cicutarium</i>	E	F	AN	3	3	0	1
<i>Erodium crinitum</i>	E	F	AN	2	6	3	1
<i>Geranium retrosum</i>	N	F	PE	2	1	0	1
<i>Geranium solanderi</i>	N	F	PE	23	16	24	10
GOODENIACEAE							
<i>Goodenia bellidifolia</i> subsp. <i>argentea</i>	N	F	PE	0	0	0	0
<i>Goodenia glabra</i>	N	F	PE	0	1	1	0
<i>Goodenia hederacea</i> var. <i>hederacea</i>	N	F	PE	1	0	2	3
<i>Goodenia pinnatifida</i>	N	F	AN	3	2	9	7
<i>Scaevola humilis</i>	N	F	PE	0	1	1	0
LAMIACEAE							
<i>Ajuga australis</i>	N	F	PE	0	2	6	15
<i>Lamium amplexicaule</i>	E	F	AN	9	7	5	1
<i>Marrubium vulgare</i>	E	F	PE	3	2	9	3
<i>Mentha diemenica</i>	N	F	PE	0	0	0	2
<i>Mentha satuireioides</i>	N	F	PE	5	7	9	2
<i>Salvia reflexa</i>	E	F	AN	5	1	4	0
<i>Salvia verbenaca</i>	E	F	PE	1	0	0	0
<i>Teucrium sp. D</i>	N	F	PE	1	1	2	3
MALVACEAE							
<i>Abutilon tubulosum</i>	N	LS	PE	5	0	0	1
<i>Malva parviflora</i>	E	F	AN	10	5	6	0
<i>Malvastrum americanum</i>	E	F	SP	0	0	0	5
<i>Modiola caroliniana</i>	E	F	SP	11	3	5	0
<i>Sida corrugata</i>	N	F	PE	11	10	10	10
<i>Sida cunninghamii</i>	N	F	PE	0	3	1	6
<i>Sida spinosa</i>	E	LS	PE	3	4	3	2
<i>Sida trichopoda</i>	N	F	PE	0	0	2	0
MYOPORACEAE							
<i>Eremophila debilis</i>	N	LS	PE	1	0	1	13
NYCTAGINACEAE							
<i>Boerhavia dominii</i>	N	F	PE	11	9	10	8
OLEACEAE							
<i>Jasminum lineare</i>	N	LS	PE	0	0	0	4
<i>Jasminum suavisissimum</i>	N	LS	PE	0	0	5	6
OXALIDACEAE							
<i>Oxalis corniculata</i>	N	F	PE	15	14	22	19
<i>Oxalis perennans</i>	N	F	PE	21	8	2	2
PAPAVERACEAE							
<i>Agremone ochroleuca</i>	N	F	AN	0	3	0	0
<i>Papaver hybridum</i>	E	F	AN	3	1	1	0
<i>Papaver somniferum</i>	E	F	AN	6	1	1	1
PLANTAGINACEAE							
<i>Plantago cunninghamii</i>	N	F	PE	0	0	0	1
<i>Plantago debilis</i>	N	F	SP	17	13	21	11
<i>Plantago lanceolata</i>	E	F	SP	4	0	1	2
POLYGANACEAE							
<i>Acetosella vulgaris</i>	E	F	PE	1	1	1	0
<i>Fallopia convolvulus</i>	E	F	AN	0	1	0	0
<i>Rumex brownii</i>	N	F	PE	35	27	26	10
<i>Rumex crispus</i>	E	F	PE	1	0	0	0
<i>Polygonum aviculare</i>	E	F	SP	15	2	4	0
<i>Portulaca oleracea</i>	N	F	AN	2	0	2	2
PRIMULACEAE							
<i>Anagallis arvensis</i>	E	F	SP	18	15	17	9
RANUNCULACEAE							
<i>Clematis glycinoides</i> var. <i>glycinoides</i>	N	C	PE	0	0	0	6
<i>Clematis microphylla</i>	N	C	PE	0	0	0	3
<i>Ranunculus lappaceus</i>	N	F	PE	0	1	1	1
<i>Ranunculus sessiliflorus</i> var. <i>sessiliflorus</i>	N	F	AN	5	7	16	6
ROSACEAE							
<i>Acaena agnipila</i>	N	F	PE	0	0	1	0
<i>Acaena novae-zelandiae</i>	N	F	PE	0	0	1	2
<i>Acaena ovina</i>	N	F	PE	5	4	6	3
<i>Rosa rubiginosa</i>	E	LS	PE	0	1	1	2
<i>Rubus anglocandicans</i>	E	LS	PE	0	0	2	0
<i>Rubus parvifolius</i>	N	LS	PE	0	1	0	1
RUBIACEAE							
<i>Asperula conferta</i>	N	F	PE	14	6	13	11
<i>Galium aparine</i>	E	F	AN	0	3	5	8
<i>Galium gaudichaudii</i>	N	F	PE	0	0	0	2
<i>Galium migrans</i>	N	F	PE	0	1	0	1
<i>Opercularia hispida</i>	N	F	PE	0	0	0	1
<i>Sherardia arvensis</i>	E	F	AN	0	0	1	0
RUTACEAE							
<i>Correa reflexa</i>	N	LS	PE	0	0	0	1
SCROPHULARIACEAE							
<i>Kickxia spuria</i> subsp. <i>Integrifolia</i>	E	F	PE	0	0	0	0
<i>Linaria arvensis</i>	E	F	AN	9	11	7	3
<i>Linaria pelisseriana</i>	E	F	AN	3	1	0	0
<i>Misopates orontium</i>	E	F	AN	0	0	1	0
<i>Verbascum thapsus</i>	E	F	SP	5	5	0	1
<i>Verbascum virgatum</i>	E	F	SP	1	0	0	1
<i>Veronica arvensis</i>	E	F	AN	19	10	3	1

<i>Veronica calycina</i>	E	F	AN	0	0	1	0												
<i>Veronica plebeia</i>	N	F	PE	0	1	0	3												
SOLANACEAE																			
<i>Solanum esuriale</i>	N	F	PE	0	0	1	1												
<i>Solanum nigrum</i>	E	LS	SP	0	1	4	0												
<i>Solanum parviflorum</i>	N	LS	PE	0	0	0	1												
<i>Solanum prinophyllum</i>	N	F	SP	0	1	0	1												
STACKHOUSIACEAE																			
<i>Stackhousia monogyna</i>	N	F	PE	0	0	0	8												
<i>Stackhousia viminea</i>	N	F	PE	1	0	3	3												
THYMELAEACEAE																			
<i>Pimelea curviflora</i>	N	LS	PE	5	5	13	3												
<i>Pimelea linifolia</i> subsp. <i>linifolia</i>	N	LS	PE	0	0	0	1												
<i>Pimelea micrantha</i>	N	LS	PE	1	2	0	0												
<i>Pimelea neo-anglica</i>	N	LS	PE	3	3	13	11												
URTICACEAE																			
<i>Parietaria australis</i>	N	F	AN	0	0	0	1												
<i>Urtica incisa</i>	N	F	PE	10	6	17	1												
VERBENACEAE																			
<i>Glandularia aristigera</i>	E	F	PE	0	0	2	0												
<i>Oncinocalyx betchei</i>	N	F	PE	0	0	1	4												
<i>Verbena bonariensis</i>	E	F	PE	2	1	1	1												
<i>Verbena brasiliensis</i>	E	F	PE	0	0	0	1												
<i>Verbena gaudichaudii</i>	E	F	PE	0	1	1	2												
<i>Verbena incompta</i>	E	F	PE	2	1	2	2												
<i>Verbena officinalis</i>	E	F	PE	2	0	2	0												
<i>Verbena quadrangularis</i>	E	F	PE	4	0	0	0												
VIOLACEAE																			
<i>Viola betonicifolia</i>	N	F	PE	1	1	2	2												
ZYGOPHYLLACEAE																			
<i>Tribulus terrestris</i>	E	F	AN	1	2	2	2												

APPENDIX 5

Thirty five climatic variables derived from ANUCLIM (Hutchinson et al. 2000)

1	Annual mean temperature
2	Mean diurnal range
3	Isothermality
4	Temperature seasonality
5	Max temperature of warmest period
6	Min temperature of coldest period
7	Temperature annual range
8	Mean temperature of wettest quarter
9	Mean temperature of driest quarter
10	Mean temperature of warmest quarter
11	Mean temperature of coldest quarter
12	Annual precipitation
13	Precipitation of wettest period
14	Precipitation of driest period
15	Precipitation seasonality
16	Precipitation of wettest quarter
17	Precipitation of driest quarter
18	Precipitation of warmest quarter
19	Precipitation of coldest quarter
20	Annual mean radiation
21	Highest period radiation
22	Lowest period radiation
23	Radiation seasonality
24	Radiation of wettest quarter
25	Radiation of driest quarter
26	Radiation of warmest quarter
27	Radiation of coldest quarter
28	Annual mean moisture index
29	Highest period moisture index
30	Lowest period moisture index
31	Moisture index seasonality
32	Mean moisture index of highest quarter
33	Mean moisture index of lowest quarter
34	Mean moisture index of warmest quarter
35	Mean moisture index of coldest quarter

APPENDIX 6

Gross margin – Merino ewes (19.5 micron) x terminal rams

Flock size	1,300		
Flock mortality	2%	Ram	2%
Productive life	5 years	Marking	90%
Ewe body weight	50 kg	Weaning	85%
DSE rating /ewe	2.1	Weaning	3 months

INCOME					
Wool	number	class	kg hd⁻¹	\$ kg⁻¹	Budget
Shear	1,274	ewes	4.8	6.50	39,759.68
	26	rams	3.5	6.50	591.66
	1,105	lambs	1.5	5.18	8,585.85
Crutch	2,407	mixed ages	0.4	3.45	3,318.01
Sheep Sales	number	class	\$ hd⁻¹	cwt	
	245	CFA ewes	60.00	22.60	14,700.00
	5	CFA rams	61.00		305.00
	1,094	mixed sex store	101.20	20.24	110,712.80
A. Total Income:					177,973.01

VARIABLE COSTS					
Replacements	number	class	cost (\$)	reps	
	5	rams	800.00		4,000.00
	271	ewes	90.00		24,390.00
Cartage	271	ewes	2.00		542.00
Wool Harvesting & Selling Costs					
Shearing	1,274	ewes	5.98	1	7,618.52
	26	rams	8.43	1	219.18
Crutching	1,287	ewes	0.85	1	1,098.20
	26	rams	1.71	1	44.37
	1,094	weaners	0.85	1	933.51
Wool tax			0.02		1,045.10
Commission, warehouse, testing charges	49	bales	42.99		2,106.51
Wool cartage and packs	49	bales	38.19		1,362.39
Sheep Health					
<i>Drenching following DrenchPlan</i>	number	class			
Broadspectrum	1,300	adults	0.42	2	1,092.00
	1,170	lambs	0.21	3	737.10
Dipping	1,300	adults	1.07	1	1,391.00
Jetting	1,300	adults	0.58	1	754.00
Vaccination- 6 in 1	1,300	adults	0.24	1	312.00
	1,170	lambs	0.24	1	280.80
Mark	1,170	lambs	1.35	1	1,579.50
Scanning	1,274	ewes	0.90	1	1,146.60
Livestock Selling Costs					
Livestock cartage	1,344	sale sheep	2.00		2,688.00
Commission on sheep sales			0.05		6,285.89
Yard dues	245	CFA ewes	0.50	per	122.50
	5	CFA rams	0.50	per	2.50
	547	mixed 20kg	0.50	per	273.50
	547	mixed 22kg	0.50	per	273.50
Transaction levy	245	CFA ewes	0.20	per	49.00
	5	CFA rams	0.20	per	1.00
	547	mixed 20kg	0.20	per	109.40
	547	mixed 22kg	0.20	per	109.40
Rural Lands Protection Board rates			0.24	per	666.94
Grain/Fodder/Forage Crop/Pasture inputs					
Grain	1,274	ewes	0.63	8	6,420.96
Grazing oats	70	@	210		14,700.00
Pasture maintenance	110	@	76.75		8,442.50
Note: Pasture cost calculated at 125kg of single super applied every 4th year - \$550/ton and \$8/ha application cost.					
B. Total Variable Costs:					90,798.17
GROSS MARGIN (A-B)					87,174.84
GROSS MARGIN /EWE					67.06
GROSS MARGIN /DSE					31.93
GROSS MARGIN /HA					127.73

APPENDIX 7

Gross margin – trade cattle

Herd size	100	trade cattle
Stocking rate	4.4	DSE ha ⁻¹
Mortality	2	%
Target body weight:	500	kg
DSE rating	11.3	DSE hd ⁻¹

INCOME						
Stock sales - number	class	kg hd⁻¹	c kg⁻¹ LWT	\$ hd⁻¹	Budget (A\$)	
98	steers	500	1.95	975	95,550.00	
A. Total Income:					95,550.00	
VARIABLE COSTS						
Replacements	class	kg hd⁻¹	c kg⁻¹ LWT	\$ hd⁻¹		
100	steers	400	1.75	700	70,000.00	
Purchasing Commission	100	head	5		500.00	
Health						
Broadspectrum	100	head	1.5	1	150.00	
Vaccination- 5 in 1	100	head	0.25	1	25.00	
Bloat capsules	100	head	18.25	1	1,825.00	
Livestock Selling Costs						
Livestock cartage	98	head	10		980.00	
Levies	98	head	5		490.00	
Yard dues	98	head	4			
Commission on sales	0.05	on	1	of sales	4,777.50	
Fodder						
Supplementary fodder	0	@	190		0.00	
Lucerne	71	@	145		10,295.00	
Improved pasture	34		76.75		2,609.50	
Native pasture	150				0.00	
B. Total Variable Costs:					91,652.00	
					GROSS MARGIN (A-B)	
					3,898.00	
					GROSS MARGIN /HEAD	
					38.98	
					GROSS MARGIN /DSE	
					3.46	
					GROSS MARGIN /HA	
					15.25	