1 Saltbush (*Atriplex* sp.)

Sukirtha Srivarathan^{1,2,3*}, Anh Dao Thi Phan^{1,2}, Olivia Wright^{1,2,4}, Daniel Cozzolino^{1,2}, Yasmina Sultanbawa^{1,2} and Michael E. Netzel^{1,2}

¹ARC Industrial Transformation Training Centre for Uniquely Australian Foods, Queensland Alliance for Agriculture and Food Innovation, The University of Queensland, Queensland, Australia; ²Centre for Nutrition and Food Sciences, Queensland Alliance for Agriculture and Food Innovation, The University of Queensland, Queensland, Australia; ³Department of Biosystems Technology, Faculty of Technology, University of Jaffna, Kilinochchi, Sri Lanka; ⁴School of Human Movement and Nutrition Sciences, The University of Queensland, Queensland, Australia

Abstract

Salinity and shrinkage of water tables are among the major crises in the world which adversely affect food production. It has been reported that about one-fifth of total farming land (45 million ha) is salt-affected worldwide. Currently, immense attention is granted to the domestication of halophytes (salt-tolerant plants) due to their evolved adaptive mechanisms to salinity including the production of secondary compounds. Halophytes are ecologically important plants, of which saltbush (*Atriplex* sp.) has been traditionally used as medicine, food, animal feed and as a 'tool' for rehabilitation of degraded land. This plant species has the potential to be used as a source of functional food/functional food ingredients, but is underutilized due to lack of popularity or limited knowledge. This chapter will give an overview of the traditional use of saltbush, an important edible halophyte, its nutritional composition, bioactive properties and potential food applications.

Introduction

Indigenous plants have gained immense interest as a source of nutrients and bioactive phytochemicals, satisfying both food demand and health needs (Pinela *et al.*, 2017; Sõukand, 2016). One such is saltbush (Fig. 1.1), an indigenous edible halophyte, member of the genus *Atriplex*, of which 400 species are found around the world (Ortíz-Dorda *et al.*, 2005). The highest number of saltbush species was found in South America (70–80 species), while the lowest number was reported in the Mediterranean basin and Europe (40–50 species) (O'Leary and Glenn, 1994; Osmond *et al.*, 1980). *Atriplex* species belong to the subfamily Chenopodioideae of the family Amaranthaceae and are distributed in arid and semi-arid regions with 200 to 400 mm of mean annual rainfall (Ortíz-Dorda *et al.*, 2005). Most species are halophytes and are found in salt marshes, salt deserts, mangroves and sand dunes near coastal areas (Wei *et al.*, 2019). *Atriplex* species are ecologically important plants and have been used for rehabilitation of degraded land (saline/alkaline soils, mine waste, shallow soils and bare saline soil) due to

© CAB International 2022. Handbook of Phytonutrients in Indigenous Fruits and Vegetables (eds D. Sivakumar et al.) DOI: 10.1079/9781789248067.0001

^{*}Corresponding author: s.srivarathan@uq.edu.au



Fig. 1.1. Saltbush (Artiplex sp.).

their ability to tolerate salinity-stressed environments through various biological mechanisms. However, they can also absorb salt from the soil to improve soil structure and increase soil fertility (Le Houérou, 1992; Ortíz-Dorda *et al.*, 2005; Piovan *et al.*, 2014). Though they have been used as human food or animal feed since ancient times, information on their dietary relevance is very limited. Therefore, this chapter reviews the nutritional profiles and dietary relevance of *Atriplex* species and safety issues related to their utilization in the human diet.

Botany

The most common saltbushes are shown in Table 1.1. Atriplex species consist of various sizes (1.0-2.5 m) and morphology (Table 1.1). These saltbush species are generally dioecious or monoecious. However, Atriplex sp. have specialized salt bladders on their leaves for storing surplus NaCl (Glenn et al., 1998). Most of the Atriplex sp. leaves have a green/grey colour (including A. cinera, A. nummularia and A. lentiformis), while leaves of A. hortensis can be green, yellow-green, faintly red, dark red and purple. This makes A. hortensis an especially attractive addition to salads (Carlsson and Hallqvist, 1981). It is interesting to note that the fruits of Atriplex sp. occur in different shapes, from triangular (A. amnicola, A. cinera) to flat and round (A. lentiformis) (Table 1.1).

Traditional Use

Atriplex species have had diverse applications for food and non-food uses from ancient times. For instance, A. cinera was popular as a potherb among the Indigenous people of Australia (Maiden, 1889). Furthermore, Atriplex sp. have been used in traditional cuisine and as bush food, and are still used raw or cooked in modern recipes. For instance, leaves of A. hortensis are used in stews, salads and vegetable spread (Carlsson and Hallqvist, 1981; Bussmann et al., 2017). Some of the Atriplex species were consumed not only by humans but also by livestock (A. nummularia, A. halimus, A. amnicola and A. semibaccata), with two species identified as nitrogen-rich fodder (A. halimus and A. nummularia) (Mehdiyeva et al., 2017). It has also been reported that saltbushes have been widely utilized as fodder reserves, particularly in famine periods or periods of forage shortage (in either drought or cold season) and as a supplementary fodder in arid and semi-arid regions (Ortíz-Dorda et al., 2005; Abbeddou et al., 2011). Furthermore, these shrubs have been used as 'emergency' livestock fodder due to their favourable crude protein content (Khan et al., 2000) in addition to their use as a source of fuel wood (Belkheiri and Mulas, 2013). The most important Atriplex sp. used for the rehabilitation of degraded lands are A. nummularia and A. canescens, respectively. These species are also used in ornamental plantations, particularly in seaside resorts (Le Houérou, 1992).

Scientific name	Common name	Botany	References
Atriplex halimus	Mediterranean saltbush	A perennial native shrub, $1-3 \times 3$ m, monoecious, inflorescences are in dense spikes, male flowers are at the top of the spike and female flowers at the base	(Ortíz-Dorda <i>et al.</i> , 2005; Clauser <i>et al.</i> , 2013)
Atriplex amnicola	River saltbush	A perennial shrub, $1 \times 1-2.5 \times 4$ m, dioecious, elongated and spear- shaped leaves (1–3 cm), fruits may be woody (2–6 mm), triangular or spherical	(Barrett-Lennard, 2003)
Atriplex cinera	Grey saltbush	Sprawling, semi-erect, woody, heavily branched, leafy shrub, 1.5×4 m and dioecious or subdioecious. Branchlets stout, rooting freely on contact with soil; stems at first ridged and angular, soon becoming terete and woody with age, grey- green leaves (2 cm), hard, fruits (2–6 mm), triangular	(De Lange <i>et al</i> ., 1998; Barrett-Lennard, 2003)
Atriplex hortensis	Garden orache or simply orache, red orache, mountain spinach, French spinach or orache	Shrub, 1–2.5 m, large leaves, $(14 \times 10 \text{ cm})$, flowers in leafless spikes, leaves can be green, yellow-green, faintly red, dark red and purple	(Carlsson, 1975; Carlsson and Hallqvist, 1981; Mehdiyeva <i>et al.</i> , 2017)
Atriplex nummularia	Oldman saltbush	Upright shrub, 2×1.5 m, irregularly shaped grey-green leaves (2–4 cm), fruits (6 mm), fan shape	(Barrett-Lennard, 2003)
Atriplex semibaccata	Australian saltbush, berry saltbush or creeping saltbush	Sprawling undershrub, xero-hypo halophyte	(Le Houérou, 1992)
Atriplex cana	-	A perennial shrub, multi-branched, 20–50 cm, stems become more or less woody	(Cai <i>et al.</i> , 2015; Wei <i>et al.</i> , 2019)
Atriplex mollis	-	Shrub, 1 m, pink, erect panicles of bladdery, fruit bracts (1 cm)	(Le Houérou, 1992)
Atriplex undulata	Wavy leaf saltbush	A perennial shrub, 1 × 2 m, wavy or crinkly leaves (0.5–1.5 cm), fruits (1–3 mm), soft and round	(Barrett-Lennard, 2003)
Atriplex lentiformis	Quailbrush	A perennial shrub, 2.5×2.5 m, silvery blue-green leaves (2 cm), fruits (2–5 mm), flat and round	(Barrett-Lennard, 2003)

Table 1.1. Botanical information about selected Atriplex species.

It is interesting to note that *A. hortensis* can be used as part of landscaping gardens and parks since these plants are decorative and beautiful, particularly during the period of fructification (Mehdiyeva *et al.*, 2017). These plants are also considered as annual spinach in North and Middle Asia (Mueller, 1880). According

to Shinn (1899), the Australian saltbushes were mentioned for the first time in the Report of the College of Agriculture in 1882. Seeds of *A. vesicaria* and *A. nummularia* were sent from Australia to California for propagation. According to Mueller (1880), *A. nummularia* was mentioned as 'one of the tallest, more fattening and wholesome of Australian pastoral saltbushes, recommended for artificial rearing' and was found from Queensland through the desert tracts to Victoria and South Australia (Shinn, 1899). In 1894, the nutritional analyses on saltbush were compared with those of other green fodders and the findings suggested that the nutrients were comparable with alfalfa.

Nutritional Composition

A. halimus can be a complement forage with favourable nutritive value during the drought and famine period (Benhammou et al., 2009). Humans have consumed this Atriplex sp. since ancient times, but only a few studies have explored its nutritional composition. The leaves of A. halimus were found to have 25.1 g/100 g (dry weight: DW) carbohydrate and 16.2 g/100 g (DW) protein (Abbeddou et al., 2011). Interestingly, Atriplex species are rich sources of fibre irrespective of their growing locations. The fibre content of A. halimus was reported to range from 25.8 to 36.7 g/100 g DW (neutral detergent fibre) and 16.5 to 23.1 g/100 g DW (acid detergent fibre), which is comparable to olive leaves (Abbeddou et al., 2011).

A. nummularia and A. amnicola are the most important species utilized for grazing and land rehabilitation in Australia (Kumara Mahipala et al., 2009). As for A. halimus, only very little scientific information is available on its nutritional profile and dietary relevance. A recent study with Australian grown Oldman saltbush (A. nummularia) showed that saltbush leaves were rich in protein (20.1 g/100 g DW), fibre (41.5 g/100 g DW) and minerals (particularly Ca (1.4 g/100 g DW), Na (4.1 g/100 g DW), Mg (0.9 g/100 g DW) and Fe (11.7 mg/100 g DW); Srivarathan et al., 2020). The reported protein content in A. nummularia ranged from 13.4 to 25.2 g/100 g DW and was comparable to that of Australian grown Oldman saltbush (Khalil et al., 1986; Yousif et al., 1994; Ben Salem et al., 2010; Revell et al., 2013; Srivarathan et al., 2020). However, A. amnicola and A. cinera had a considerably lower protein content (9.0-9.7 g/100 g)g DW) than A. nummularia, whereas their fibre contents were in the same range as found in other Atriplex sp. (neutral detergent fibre (25.2–48.8 g/100 g DW); acid detergent fibre (16.1–33.5 g/100 g DW); Warren and Casson, 1993; Kumara Mahipala *et al.*, 2009; Revell *et al.*, 2013).

A. hortensis is considered to be one of the ancient cultivated plants, originated in Europe (Stevens, 1994), and has been used as a potherb in conventional medicine. Although it has been used in the food and pharmaceutical industry (Livadariu, 2013; Bussmann et al., 2017), its nutritional composition and bioactive properties are not well reported (Wright et al., 2002). According to Carlsson and Clarke (1983), leaves of A. hortensis are a rich source of protein. The content of crude protein in the leaves was higher than that of spinach (Spinacia oleracea) and A. hortensis has the potential to be used as a substitute for spinach due to its high yield and prolonged harvest season. Palmitic, oleic, linoleic and α -linolenic acids could be identified as the main fatty acids in the leaves of A. hortensis (Carlsson and Clarke, 1983).

The seeds of A. hortensis were found to have a relatively high content of carbohydrate (62.0g/ 100 g DW), followed by protein (28.3 g/100 g DW) and dietary fibre (13.3 g/100 g DW)g DW) (Wright et al., 2002). The fibre and protein content of A. hortensis seeds was higher than that of sweet and bitter quinoa (Wright et al., 2002). The protein content in the seeds was also higher than that found in the main 'food-cereals' including rice (7.6 g/100 g DW), wheat (14.3 g/100 g DW), maize (10.2 g/100 g DW)g DW) and barley (10.8 g/100 g DW) (Duke and Atchley, 1986; Wright et al., 2002). Furthermore, the amino acid profile in both, seeds and leaves, was found to be favourable compared to maize, rice and wheat, having a higher content of lysine, the limiting amino acid in cereals (Wright et al., 2002).

Phytochemicals and Bioactive Properties

The presence of different phytochemicals including saponins, flavonoids (especially gly-cosides), tannins, terpenoids and alkaloids were reported for *Atriplex* sp. (Table 1.2) (Boutaoui *et al.*, 2018). Halophytes can have relatively high levels of phenolic compounds since they develop

Plant species	Bioactive compounds/phytochemicals	Bioactive properties	Ethnopharmacological use/ general applications	References
A. halimus	Saponins, betaines, mineral salts, alkaloids, phytoecdysteroids, flavonoids (isorhamnetin- 3-O-glucopyranoside, rutin-4',7-dimethylether and kaempferol-3,7-dirhaminoside-4'- methyloxide, isoorientin, 4'-methoxy-7- glucoside-5-hydroxyisoflavone, naringenin-4'-O- rhamnopyranoside, hesperidin), phenolic acids (<i>p</i> - hydroxybenzoic acid, chlorogenic acid, cinnamic acid, ferulic acid and salicylic acid)	Antioxidant, anti- acetylcholinesterase, hypoglycaemic	Leaves are used to treat heart disease, diabetes, rheumatism	(Bayoumi and El-Shaer, 1992; Dinan <i>et al.</i> , 1998; Said <i>et al.</i> , 2008; Benhammou <i>et al.</i> , 2009; Emam, 2011; Clauser <i>et al.</i> , 2013)
A. mollis	Rutin, catechin, epicatechin, naringenin, <i>p</i> - coumaric acid, gallic acid, chlorogenic acid, <i>p</i> - hydroxybenzoic acid, sinapinic acid, syringic acid, ferulic acid and vanillic acid	Anti-inflammatory, antioxidant, hepatoprotective	Phytopharmaceutical use	(Boutaoui <i>et al.</i> , 2018)
A. hortensis	Condensed tannins and other phenolic compounds	Antioxidant	Potherb in traditional medicine, used as diuretic and anastasis	(Carlsson and Hallqvist, 1981; Stevens, 1994; Wright <i>et al.</i> , 2002; Mehdiyeva <i>et al.</i> , 2017)
A.nummularia	Vitamin Ε, β-carotene	Antioxidant	ı	(Pearce <i>et al.</i> , 2005; Ben Salem <i>et al.</i> , 2010)
A. lentiformis	Quercetin, kaempferol, quercetin and kaemferol glycosides	Antioxidant		(Awaad <i>et al.</i> , 2012)

Table 1.2. Bioactive compounds/phytochemicals identified in selected Atriplex species.

a biological antioxidant system as a response to abiotic stress (Ramani *et al.*, 2004; Ksouri *et al.*, 2012; Boutaoui *et al.*, 2018).

A. halimus has been used as a medicinal plant in conventional Arabic medicine for glycaemic control in diabetic patients and to treat heart disease (Table 1.2) (Said et al., 2002). It has also been used as a treatment for rheumatism and to combat parasites in veterinary use (Bayoumi and El-Shaer, 1992). Decoction and/ or a remedy bath are the usual administration. A. hortensis is used as a diuretic, anastasis and against haemorrhage (Mehdiyeva et al., 2017). It was also reported that it has higher levels of total phenolics and condensed tannins than S. oleracea (Carlsson and Hallqvist, 1981). Microwave-assisted extraction of A. mollis recovered several phytochemicals including gallic acid, catechin, chlorogenic acid, p-hydroxybenzoic acid, rutin, sinapinic acid, ferulic acid, naringenin and benzoic acid (Boutaoui et al., 2018). No reported bioactivities or therapeutic application of A. mollis could be found in the literature. A. mollis and A. nummularia could be also identified as valuable sources of antioxidants having relatively high levels of vitamin E(139 mg/kg dry matter)(DM)) and beta carotene (34.5 mg/kg DM) (Pearce et al., 2005; Ben Salem et al., 2010). According to White and Rewell (2007), A. nummularia was used in Australia to treat weaner sheep with vitamin E deficiency. A recent study on A. lentiformis showed that the isolated quercetin-6,4'-dimethoxy-3polyphenols, fructo-rhamnoside and quercetin-4'-methoxy-3-fructo-rhamnoside, exhibited a strong antioxidant activity in rats with no side effects on liver and kidney function (Awaad et al., 2012).

Antinutritive Compounds

According to Watson *et al.* (1987), oxalate presents an important anti-nutrient in *Atriplex* leaves. For instance, *A. hortensis* was reported with an oxalate content of up to 8.7% DM (Carlsson and Hallqvist, 1981). However, the toxicity of saltbush is regarded as low since its high salt content can control the intake of oxalate before toxicity is attained (Ben Salem

et al., 2010). As for other plant ingredients, oxalate levels can vary significantly, depending on the maturity, harvest season and specific environmental conditions (Abu-Zanat *et al.*, 2003). It was also reported that oxalate might have lowered the calcium absorption in tilapia fish fed with dehydrated *Atriplex* leaves, resulting in suppressed growth (Yousif *et al.*, 1994). Osmond (1963), Sandberg *et al.* (1967) and Duke and Williams (1977) reported low levels of quaternary alkaloids in *Atriplex* sp. in addition to saponins, tannins and flavonoids. However, it was surprising that no saponins could be found in the seeds of *A. hortensis* (Wright *et al.*, 2002).

Bioaccessibility and Bioavailability

Several *in vitro* studies on the digestibility of halophytes in the context of their 'efficiency' as livestock feed have been conducted (Weston *et al.*, 1970; Abbeddou *et al.*, 2011; Tosto *et al.*, 2015). However, no reports about human studies to assess the actual bioaccessibility and/ or bioavailability of the main nutrients and phytochemicals in halophytes could be found.

Products/Formulations/Applications

Based on their nutritional composition, Atriplex sp. have the potential to be used as functional foods/functional food ingredients. The seeds of A. hortensis could be used to produce saponin-free food products. A. hortensis seeds and leaves, with their high protein content and well-balanced amino acid profiles, may have the potential to be used as a novel/alternative protein source (Carlsson and Clarke, 1983; Wright et al., 2002). Since Atriplex sp. are generally rich in protein and most of them can be consumed raw, the addition to a meal as a complementary vegetable or incorporation into salads are other possible food 'applications'. Furthermore, due to their salty taste and unique aroma, leaves (fresh and dried) of Atriplex sp. can be added to a meal as a potential salt substitute or served together with seafood. Another 'avenue' is the typical functional food and/or phytopharmaceutical applications of Atriplex sp., due to their broad range of bioactive phytochemicals such as chlorophylls, carotenoids, betalains and phenolic compounds (Suhaj, 2006; Ksouri *et al.*, 2012).

Conclusions and Future Trends

This chapter has provided an overview of *Atriplex* sp., from their botany to potential (functional) food applications, to showcase their nutritional and bioactive potential to a broader audience.

However, the showcased potential needs to be investigated further in human (clinical) studies to substantiate the actual bioavailability and health benefits of the main nutrients, bioactive phytochemicals and derived products. Another focus should be led on the development of different *Atriplex* sp.-based products suitable for the mainstream food market (e.g. salt replacement), but also for specific functional food and/or phytopharmaceutical applications (e.g. supporting diabetes management).

Acknowledgements

The authors acknowledge the Traditional Owners of the lands on which the indigenous plant species were harvested and respect the knowledge and experience the Traditional Owners hold regarding the care, harvest and use of these plants. This research is funded by the Australian Research Council (ARC) Industrial Transformation Training Centre (ITTC) for Uniquely Australian Foods (grant number: IC180100045).

References

- Abbeddou, S., Rihawi, S., Hess, H.D., Iñiguez, L., Mayer, A.C. et al. (2011) Nutritional composition of lentil straw, vetch hay, olive leaves, and saltbush leaves and their digestibility as measured in fat-tailed sheep. Small Ruminant Research 96(2–3), 126–135. DOI: 10.1016/j.smallrumres.2010.11.017.
- Abu-Zanat, M.M., Al-Hassanat, F.M., Alawi, M. and Ruyle, G.B. (2003) Oxalate and tannins assessment in *Atriplex halimus* L. and *A. nummularia* L. *Journal of Range Management* 56(4), 370. DOI: 10.2307/4004041.
- Awaad, A.S., Maitland, D.J., Donia, A.E.R.M., Alqasoumi, S.I. and Soliman, G.A. (2012) Novel flavonoids with antioxidant activity from a Chenopodiaceous plant. *Pharmaceutical Biology* 50(1), 99–104. DOI: 10.3109/13880209.2011.591806.
- Barrett-Lennard, E.G. (2003) Pasture plants for saltland. In: *Saltland Pastures in Australia- A Practical Guide*, 2nd edn. CRC for Plant-based Management of Dryland Salinity, Western Australia.
- Bayoumi, M. and El-Shaer, H. (1992) Impact of halophytes on animal health and nutrition. In: Squires, V. and Ayoub, A.T. (eds) Halophytes as a Resource for Livestock and for Rehabilitation of Degraded Lands, 1st edn. Springer, Dordrecht. DOI: 10.1007/978-94-011-0818-8.
- Belkheiri, O. and Mulas, M. (2013) The effects of salt stress on growth, water relations and ion accumulation in two halophyte *Atriplex* species. *Environmental and Experimental Botany* 86, 17–28. DOI: 10.1016/j.envexpbot.2011.07.001.
- Ben Salem, H., Norman, H.C., Nefzaoui, A., Mayberry, D.E., Pearce, K.L. et al. (2010) Potential use of oldman saltbush (*Atriplex nummularia* Lindl.) in sheep and goat feeding. *Small Ruminant Research* 91(1), 13–28. DOI: 10.1016/j.smallrumres.2009.10.017.
- Benhammou, N., Bekkara, F.A. and Kadifkova Panovska, T. (2009) Antioxidant activity of methanolic extracts and some bioactive compounds of *Atriplex halimus*. *Comptes Rendus Chimie* 12(12), 1259–1266. DOI: 10.1016/j.crci.2009.02.004.
- Boutaoui, N., Zaiter, L., Benayache, F., Benayache, S., Cacciagrano, F. et al. (2018) Atriplex mollis Desf. aerial parts: Extraction procedures, secondary metabolites and color analysis. *Molecules* 23(8), E1962. DOI: 10.3390/molecules23081962.

- Bussmann, R.W., Paniagua-Zambrana, N.Y., Sikharulidze, S., Kikvidze, Z., Kikodze, D. et al. (2017) Plant and fungal use in Tusheti, Khevsureti, and Pshavi, Sakartvelo (Republic of Georgia), Caucasus. Acta Societatis Botanicorum Poloniae 86(2), 1. DOI: 10.5586/asbp.3517.
- Cai, D., Yan, C. and Wei, Y. (2015) Seed germination characteristics of the desert subshrub *Atriplex cana* and its ecological significance. *Acta Prataculturae Sinica* 24, 131–138.
- Carlsson, R. (1975) Selection of Centrospermae and Other Species for Production of Leaf Protein Concentrates. Verlag nicht ermittelbar.
- Carlsson, R. and Clarke, E.M.W. (1983) *Atriplex hortensis* L. as a leafy vegetable, and as a leaf protein concentrate plant. *Qualitas Plantarum Plant Foods for Human Nutrition* 33(2–3), 127–133. DOI: 10.1007/BF01091298.
- Carlsson, R. and Hallqvist, C.W. (1981) *Atriplex hortensis* L. revival of a spinach plant. *Acta Agriculturae Scandinavica* 31, 229–234. DOI: 10.1080/00015128109435702.
- Clauser, M., Dall'Acqua, S., Loi, M.C. and Innocenti, G. (2013) Phytochemical investigation on Atriplex halimus L. from Sardinia. Natural Product Research 27(20), 1940–1944. DOI: 10.1080/14786419.2013.793684.
- De Lange, P.J., Murray, B.G. and Gardner, R.O. (1998) *Atriplex cinerea*(Chenopodiaceae) in New Zealand. *New Zealand Journal of Botany* 36, 521–529.
- Dinan, L., Whiting, P. and Scott, A.J. (1998) Taxonomic distribution of phytoecdysteroids in seeds of members of the Chenopodiaceae. *Biochemical Systematics and Ecology* 26(5), 553–576. DOI: 10.1016/S0305-1978(98)00005-2.
- Duke, J.A. and Atchley, A.A. (1986) Handbook of Proximate Analysis Tables of Higher Plants. CRC Press.
- Duke, J.A. and Williams, M.C. (1977) Phytotoxin tables. *CRC Critical Reviews in Toxicology* 5(3), 189–237. DOI: 10.3109/10408447709082600.
- Emam, S.S. (2011) Bioactive constituents of Atriplex halimus plant. Journal of Natural Products 4, 25-41.
- Glenn, E.P., Jed Brown, J. and O'Leary, J.W. (1998) Irrigating crops with seawater. *Scientific American* 279(2), 76–81. DOI: 10.1038/scientificamerican0898-76.
- Khalil, J.K., Sawaya, W.N. and Hyder, S.Z. (1986) Nutrient composition of *Atriplex* leaves grown in Saudi Arabia. *Journal of Range Management* 39(2), 104. DOI: 10.2307/3899277.
- Khan, M., Ungar, I.A. and Showalter, A.M. (2000) Effects of Salinity on Growth, Water Relations and Ion Accumulation of the Subtropical Perennial Halophyte, *Atriplex griffithii* var. stocksii. *Annals of Botany* 85(2), 225–232. DOI: 10.1006/anbo.1999.1022.
- Ksouri, R., Ksouri, W.M., Jallali, I., Debez, A., Magné, C. et al. (2012) Medicinal halophytes: potent source of health promoting biomolecules with medical, nutraceutical and food applications. *Critical Reviews* in Biotechnology 32(4), 289–326. DOI: 10.3109/07388551.2011.630647.
- Kumara Mahipala, M.B.P., Krebs, G.L., McCafferty, P. and Gunaratne, L.H.P. (2009) Chemical composition, biological effects of tannin and in vitro nutritive value of selected browse species grown in the West Australian Mediterranean environment. *Animal Feed Science and Technology* 153(3–4), 203–215. DOI: 10.1016/j.anifeedsci.2009.06.014.
- Le Houérou, H.N. (1992) The role of saltbushes (*Atriplex* spp.) in arid land rehabilitation in the Mediterranean Basin: a review. *Agroforestry Systems* 18(2), 107–148. DOI: 10.1007/BF00115408.
- Livadariu, O. (2013) In vitro experimental researches regarding the treatment with phytoregulators of orach (atriplex hortensis L.). Bulletin of the University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca Animal Science and Biotechnologies 70, 289–295.
- Maiden, J.H. (1889) *The Useful Native Plants of Australia (including Tasmania)*. Turner and Henderson, London. DOI: 10.5962/bhl.title.120959.
- Mehdiyeva, N.P., Fayvush, G., Aleksanyan, A., Alizade, V.M., Batsatsashvili, K. et al. (2017) Atriplex hortensis L. Atriplex tatarica L. Amaranthaceae. In: Bussmann, R.W. (ed.) Ethnobotany of the Caucasus. Springer International Publishing, Cham, Switzerland. DOI: 10.1007/978-3-319-49412-8.
- Mueller, F. von (1880) Select Extra-tropical Plants Readily Eligible for Industrial Culture or Naturalisation, with Indications of Their Native Countries and Some of their Uses. Office of the Superintendent of Government, Calcutta, India. DOI: 10.5962/bhl.title.54192.
- O'Leary, J. and Glenn, E.P. (1994) Global distribution and potential for halophytes. In: *Halophytes as a Resource for Livestock and for Rehabilitation of Degraded Lands*. Springer, Dordrecht, Netherlands. DOI: 10.1007/978-94-011-0818-8.
- Ortíz-Dorda, J., Martínez-Mora, C., Correal, E., Simón, B. and Cenis, J.L. (2005) Genetic structure of *Atriplex halimus* populations in the Mediterranean Basin. *Annals of Botany* 95(5), 827–34. DOI: 10.1093/aob/mci086.

- Osmond, B. (1963) Oxalates and ionic equilibria in Australian saltbushes (*Atriplex*). *Nature* 198(4879), 503–504. DOI: 10.1038/198503a0.
- Osmond, C.B., Björkman, O. and Anderson, D.J. (1980) *Physiological Processes in Plant Ecology: Toward a Synthesis with Atriplex.* Springer Science & Business Media, Berlin, Heidelberg. DOI: 10.1007/978-3-642-67637-6.
- Pearce, K.L., Masters, D.G., Smith, G.M., Jacob, R.H. and Pethick, D.W. (2005) Plasma and tissue α-tocopherol concentrations and meat colour stability in sheep grazing saltbush (*Atriplex* spp.). *Australian Journal of Agricultural Research* 56(7), 663. DOI: 10.1071/AR05001.
- Pinela, J., Carvalho, A.M. and Ferreira, I.C.F.R. (2017) Wild edible plants: Nutritional and toxicological characteristics, retrieval strategies and importance for today's society. *Food and Chemical Toxicology* 110, 165–88. DOI: 10.1016/j.fct.2017.10.020.
- Piovan, M.J., Zapperi, G.M. and Pratolongo, P.D. (2014) Seed germination of *Atriplex undulata* under saline and alkaline conditions. *Seed Science and Technology* 42(2), 286–292. DOI: 10.15258/ sst.2014.42.2.17.
- Ramani, B., Zorn, H. and Papenbrock, J. (2004) Quantification and fatty acid profiles of sulfolipids in two halophytes and a glycophyte grown under different salt concentrations. *Zeitschrift Für Naturforschung* C 59(11–12), 835–42. DOI: 10.1515/znc-2004-11-1212.
- Revell, D.K., Norman, H.C., Vercoe, P.E., Phillips, N., Toovey, A. et al. (2013) Australian perennial shrub species add value to the feed base of grazing livestock in low- to medium-rainfall zones. Animal Production Science 53(11), 1221. DOI: 10.1071/AN13238.
- Said, O., Fulder, S., Khalil, K., Azaizeh, H., Kassis, E. et al. (2008) Maintaining a physiological blood glucose level with "glucolevel", a combination of four anti-diabetes plants used in the traditional Arab herbal medicine. Evidence-Based Complementary and Alternative Medicine 5(4), 421–428. DOI: 10.1093/ecam/nem047.
- Said, O., Khalil, K., Fulder, S. and Azaizeh, H. (2002) Ethnopharmacological survey of medicinal herbs in Israel, the Golan Heights and the West Bank region. *Journal of Ethnopharmacology* 83(3), 251–265. DOI: 10.1016/s0378-8741(02)00253-2.
- Sandberg, F., Michel, K.H., Staf, B. and M, T. (1967) Screening of plants of family chenopodiaceae for alkaloids. *Acta Pharmaceutica Suecica* 4, 51.
- Shinn, C.H. (1899) Australian Salt-bushes: Results of Eighteen Years' Tests, Characteristics, Propagation, and Field Experiments. University Press.
- Sõukand, R. (2016) Perceived reasons for changes in the use of wild food plants in Saaremaa, Estonia. *Appetite* 107, 231–241. DOI: 10.1016/j.appet.2016.08.011.
- Srivarathan, S., Netzel, M., Thi Phan, A.D. and Sultanbawa, Y. (2020) Exploring the Nutritional Profile and Bioactive Potential of Australian Grown Saltbush (*Atriplex* sp.). In: *Proceedings of The third International Tropical Agriculture Conference (TROPAG 2019)*, MDPI, Basel, Switzerland, (pp. 83–84). DOI: 10.3390/proceedings2019036083.
- Stevens, J. (1994) Orach-Atriplex hortensis L. Fact sheet HS-637. Horticultural Sciences Department, Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida, Gainesville, Florida.
- Suhaj, M. (2006) Spice antioxidants isolation and their antiradical activity: a review. *Journal of Food Composition and Analysis* 19(6–7), 531–537. DOI: 10.1016/j.jfca.2004.11.005.
- Tosto, M.S.L., Araújo, G.G.L., Ribeiro, L.G.P., Henriques, L.T., Menezes, D.R. et al. (2015) In vitro rumen fermentation kinetics of diets containing oldman saltbush hay and forage cactus, using a cattle inoculum. Arquivo Brasileiro de Medicina Veterinária e Zootecnia 67(1), 149–158. DOI: 10.1590/1678-6937.
- Warren, B. and Casson, T. (1993) Saltbush quality and sheep performance. ACIAR proceedings, Australian Centre for International Agricultural Research 71.
- Watson, M.C., O'Leary, J.W. and Glenn, E.P. (1987) Evaluation of Atriplex lentiformis (Torr.) S. Wats. and Atriplex nummularia Lindl. as irrigated forage crops. Journal of Arid Environments 13(3), 293–303. DOI: 10.1016/S0140-1963(18)31119-4.
- Wei, C., Zhou, S., Li, W., Jiang, C., Yang, W. et al. (2019) Chemical composition and allelopathic, phytotoxic and pesticidal activities of *Atriplex cana* Ledeb. (Amaranthaceae) essential oil. Chem Biodivers 16(4), e1800595-n. DOI: 10.1002/cbdv.201800595.
- Weston, R., Hogan, J. and Hemsley, J. (1970) Some aspects of the digestion of *Atriplex nummularia* (saltbush) by sheep. In: *Proceedings of the Australian Society of Animal Production*, (pp. 517–521).

- White, C.L. and Rewell, L. (2007) Vitamin E and selenium status of sheep during autumn in Western Australia and its relationship to the incidence of apparent white muscle disease. *Australian Journal of Experimental Agriculture* 47(5), 535. DOI: 10.1071/EA05123.
- Wright, K.H., Pike, O.A., Fairbanks, D.J. and Huber, C.S. (2002) Composition of Atriplex hortensis, Sweet and Bitter Chenopodium quinoa Seeds. *Journal of Food Science* 67(4), 1383–1385. DOI: 10.1111/j.1365-2621.2002.tb10294.x.
- Yousif, O.M., Alhadrami, G.A. and Pessarakli, M. (1994) Evaluation of dehydrated alfalfa and salt bush (*Atriplex*) leaves in diets for tilapia (*Oreochromis aureus* L.). *Aquaculture* 126(3–4), 341–347. DOI: 10.1016/0044-8486(94)90050-7.