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Bipolarity in sea slugs: a new species of *Doridunculus* (Mollusca: Nudibranchia: Onchidoridoidea) from Antarctica

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Abstract Bipolar distributions of benthic taxa have intrigued many biologists since the first Antarctic expeditions. Records of taxa, either at species or higher taxonomic levels, encompassing this peculiar distribution have been regularly reported since then. Moreover, the study of heterobranch mollusks from remote areas, such as Antarctica, often provides key taxa so far unknown, which might be essential for systematics. Here, we describe a new species of nudibranch from the eastern Weddell Sea using micro-computed tomography (micro-CT), namely, Doridunculus punkus n. sp. (Akiodorididae). Hitherto, the genus Doridunculus was only reported from the Arctic region. We provide an extensive description of morphological and anatomical characters of D. punkus n. sp., thereby offering new insights into the anatomy of the understudied Akiodorididae. A comparison of all Akiodorididae species shows that D. punkus n. sp. exhibits intermediate characters between some Onchidoridoidea families (i.e., hook-shaped innermost lateral teeth). Furthermore, the detailed study of its reproductive system suggests a close relationship of both Akiodorididae and Goniodorididae. We

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suggest an Antarctic origin of Akiodorididae followed by dispersion and a subsequent speciation in the Arctic.

Keywords Akiodorididae · Phanerobranchia · Heterobranchia · Weddell Sea · Disjunct distribution · Micro-CT

Introduction

A disjunct distribution of sister taxa covering the northern and southern hemispheres is a phenomenon known as bipolarity (Stepanjants et al. 2006). Bipolar distributions can occur either at the species, genus, or higher taxonomic levels (Allcock and Griffiths 2015). In mollusks, approximately 30% of living Antarctic bivalve and gastropod families are characterized by a bipolar distribution, including heterobranch genera such as Philine Ascanius, 1772 and Toledonia Dall, 1902 (Rudman 1972; Warén 1989; Dell 1990; Crame 1993). The wide fossil record of mollusks suggests at least three paleontological periods in which bipolar and/or amphitropical (i.e., on both sides of the tropics) events occurred, Late Jurassic (~150 Mya), Paleogene-Neogene (~23 Mya), and Neogene-Pleistocene (~2.6 Mya; Crame 1993). Current disjunct distributions might be the result of transequatorial dispersal during glacial maximum cooling or, alternatively, a prior cosmopolitan species isolated vicariantly in high latitudes during interglacial periods (Allcock and Griffiths 2015). Vicariant cases imply that species once placed in the tropics might have sheltered in deep waters during interglacial periods, a phenomenon called equatorial submergence (Stepaniants et al. 2006). This is applicable for Philine for example, which is distributed in deep waters of all oceans (OBIS 2016). However, knowledge of deep equatorial species is often lacking, probably due to the scarce sampling done so far in deep tropical waters (Allcock and Griffiths 2015).





Antarctic glacial and interglacial periods during the early Cenozoic (~65 Mya) triggered species migration toward warmer temperatures in lower latitudes or in deep-sea shelters (Thatje et al. 2005). The Antarctic final breakup during the late Cenozoic allowed the formation of the Antarctic Circumpolar Current (~25 Mya). This thermal and hydrographic barrier hampered marine organisms' dispersion from north to south and above 1000 m depth at the Southern Ocean (SO; Barker and Thomas 2004). However, deep-sea organisms from the Weddell Sea may have dispersed through the Antarctic Bottom Water, which flows as part of the global thermohaline circulation system (Stepanjants et al. 2006; Pawlowski et al. 2007). Thereby, organisms with high dispersal capabilities may have colonized distant regions, either by planktonic larvae or by attaching to floating debris (Raguá-Gil et al. 2004), even contemporarily (Stepanjants et al. 2006; Pawlowski et al. 2007; Jun et al. 2012).

Wägele et al. (2008) suggested that basal members of some major Nudibranchia lineages may have an Antarctic origin. Several families and genera are only found in the Southern Ocean, and these might be crucial for understanding the evolution of heterobranch lineages. Nudibranchia, with approximately 35 described species from Antarctica, is the most speciose heterobranch lineage (De Broyer et al. 2016). Among them, the Onchidoridoidea Gray, 1827 of the SO are understudied phanerobranch dorids, characterized for being unable to fully retract their gills. This taxon comprises five families, namely, Akiodorididae Millen & Martynov, 2005; Calycidorididae Roginskaya, 1972; Corambidae Bergh, 1871; Goniodorididae Adams & Adams, 1854; and Onchidorididae Gray, 1827 (WoRMS Editorial Board 2015). Onchidoridoidea were formerly assigned to Suctoria due to the possession of a buccal pump (Bergh 1892). Akiodorididae is considered to be a basal family within Onchidoridoidea, presently related to Goniodorididae based on the reproductive system (Hallas and Gosliner 2015). In fact, Akiodorididae was recently established and includes genera previously placed in Goniodorididae and Onchidorididae (Millen and Martynov 2005). The family Akiodorididae currently contains five genera, all of them restricted to polar waters. Akiodoris Bergh, 1892 confined to the North Pacific, Armodoris Minichev, 1972 from the SO, and Doridunculus Sars, 1878 from the North Pacific and North Atlantic, each having two described species. Echinocorambe Valdés & Bouchet, 1998 in the Norwegian Sea and Prodoridunculus Thiele, 1912 from the Davies Sea are both monotypic. Prodoridunculus was described based on juvenile specimens, and it was suggested to be the senior synonym of Armodoris antarctica Minichev, 1972 (Millen and Martynov 2005). The synapomorphies of Akiodorididae are a smooth, thin, lip disk; two or more inner lateral teeth; and rectangular, reduced outer lateral teeth (Millen and Martynov 2005). When present, the rachidian tooth ranges from a small plate to a wide archshaped structure, sometimes with a central cusp (Millen and Martynov 2005). The inner lateral teeth have a strong cusp directed downward, with several denticles along the margins. The marginal teeth are rectangular and progressively decrease in size toward the edge of the radula (Millen and Martynov 2005). Externally, these species have a spiculated notum covered by tubercles. The gills are dorsal and arranged in a semicircle, with the anus lying posteriorly; gills are not found in a pocket as in Onchidorididae (Hallas and Gosliner 2015). However, in *Echinocorambe*, gills are reduced to one simple leaf and the anus is ventral (Valdés and Bouchet 1998).

The genus *Doridunculus* was first described by Sars (1878) from Risvær (Norway) at 100-m depth based on a single specimen, named D. echinulatus Sars, 1878. The species presents two dorsal keels lying side by side, and its dorsum is covered by conical spiculated tubercles. Later, Odhner (1907) described D. pentabranchus Odhner, 1907, which was found from muddy bottoms of Skagerrak, Norway, at 335-m depth. This species differs from the type species only by having five gills instead of three. Therefore, Odhner (1922) himself further considered D. pentabranchus as a probable junior synonym of D. echinulatus. Decades later, a new species, named D. unicus Martynov & Roginskaya, 2005, was described from abyssal waters (3000–3620 m) of the Sea of Japan (Martynov and Roginskaya 2005). Contrary to D. echinulatus, D. unicus lacks the dorsal keels and presents more gills, larger oral tentacles, and a rachidian tooth. Presently, these are the only described species of *Doridunculus*, both inhabiting exclusively the northern hemisphere (Martynov and Roginskaya 2005).

Here, we describe Doridunculus punkus n. sp., collected in the Antarctic eastern Weddell Sea, based on a single specimen collected in Austasen at 228-m depth in 2003. Since the analysis of a potentially new nudibranch species and its description require dissection, radula preparation, and examination of internal anatomy, usually, at least one specimen is destroyed in doing so. In order to save the holotype, we performed X-ray micro-computed tomography scans followed by 3D reconstruction to describe the new Doridunculus species. Micro-CT analysis allows non-destructive investigation of type material from regions that are difficult to survey, as is the case here (see Moles et al. 2016a). Moreover, we provide a comparative anatomical description of *Doridunculus* and the other Akiodorididae genera. A discussion on the bipolar distribution of this family is also included and extended to other examples found in the literature.

Material and methods

Sample collection

One specimen of *D. punkus* n. sp. was collected in the eastern Weddell Sea (Antarctica) during the Antarctic cruise ANT





XXI/2 (November 2003–January 2004) of RV Polarstern (Alfred Wegener Institute, Bremerhaven, Germany). The specimen was collected with Agassiz trawl in Austasen at 228-m depth (PS65/280–1), later photographed alive, and transferred to 70% ethanol for morphological analysis.

Morphological analysis

For micro-CT analysis, D. punkus n. sp. was contrasted with 1% iodine metal (I₂) dissolved in 100% ethanol (I₂E) for 24 h. Subsequently, the specimen was transferred to a pipette tip containing 100% ethanol, in which the specimen was arrested. The pipette tip was then mounted on a pin with superglue. Three X-ray tomography scans were performed with an XRadia Micro XCT-200 (Carl Zeiss X-ray Microscopy Inc.). For the macro-scan, the 0.4× object lens unit was used, at 40 kV and 200 μA, with a pixel size of 15.35 μm. For scanning the anterior body region, the 4× object lens unit, at 90 kV and 88 µA, was used, with a pixel size of 3.70 µm. For the micro-scan of the radula and spicules, the 10× object lens unit, at 40 kV and 200 µA, was used with a pixel size of 2.02 µm. All scans were performed by using Binning 2 (summarizing 4 pixels) and subsequently reconstructed by using Binning 1 (full resolution) as given in Sombke et al. (2015). Tomography projections were then reconstructed using the reconstruction software XMReconstructor software (Carl Zeiss Microscopy GmbH), resulting in image stacks (Tiff format). We used the software platform Amira® 5.4. (FEI, Visualization Science Group) for image segmentation. Two graphical 3D PDF reconstructions of D. punkus n. sp. were performed using Deep Exploration. 3D PDFs can be opened in Adobe Acrobat Reader and activated by clicking on it (see Supporting Information).

Results

Systematics

Euctenidiacea Tardy, 1970.

Doridacea Thiele, 1912.

Onchidoridoidea Gray, 1827.

Akiodorididae Millen & Martynov, 2005.

Doridunculus Sars, 1878.

Type species. Doridunculus echinulatus Sars, 1878.

Diagnosis of the genus Color translucent white. Body elongated, notum rounded anteriorly, posteriorly tapered, slightly shorter than tail. Velum not clearly separated from ventral notum side. Oral tentacles small, conical, or folded. Notal longitudinal keel(s) present or absent. Notum covered by numerous conical tubercles. Tail ridged. Gills dorsal, minute, 3 to 10, disposed in semicircle. Network of elongated spicules

visible through epidermis. Radular formula 1–10.1.(0–1).1.10–1 (with or without rachidian); inner lateral hookshaped or with pronounced, big cusp pointing downward; rectangular marginal teeth forming interlocking pointed plates, decreasing in size toward edge.

The genus contains only the following three species (Martynov and Roginskaya 2005): *D. echinulatus*, *D. unicus*, and *D. punkus* n. sp. described herein.

D. punkus n. sp. (Figs. 1, 2, 3, 4, and 5) (See 3D PDF of the reconstructed anatomy of the whole specimen and a more detailed anterior part in Supplementary Material 1 and 2, respectively) http://zoobank.org/NomenclaturalActs/69A41 F49-FAEC-4A6F-ADD6-0657084A5745.

Holotype Adult specimen, 16 mm. Austasen, eastern Weddell Sea (71° 7.15′ S, 11° 26.23′ W) collected by Agassiz trawl on 29 December 2003, at 228-m depth (PS65/280–1). Deposited in SNSB Zoologische Staatssammlung München (catalog number ZSM Moll 2016113).

Type locality Austasen, eastern Weddell Sea (Antarctica).

External morphology (Figs. 1 and 2a, b) Live animal measured 16 mm (Fig. 1a), when preserved $15 \times 6 \times 6$ mm (length:width:height). Color translucent white, brownish digestive gland seen postero-laterally by transparency. Body high, elongated, pentagonal in transverse section (Fig. 1b); mantle rim protruding; oral veil rounded, slightly folded laterally; oral tentacles grooved; notum rounded posteriorly (Fig. 1c); foot narrow, notched anteriorly, tapering posteriorly, mid-dorsal ridge present in tail (Fig. 1d). Posterior part not covered by notum. Rhinophores bearing seven diagonal

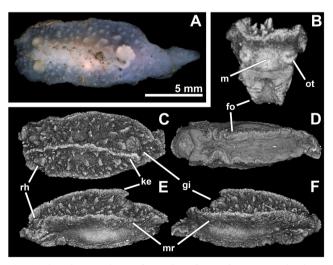


Fig. 1 Live photograph (a) of *Doridunculus punkus* n. sp. and volume rendering (b-f) of the preserved animal. a Dorsal view. b Frontal view. c Dorsal view d Ventral view. e Left-lateral view. f Right-lateral view. fo foot, gi gills, ke keel, m mouth, mr mantle rim, ot oral tentacles, rh rhinophores



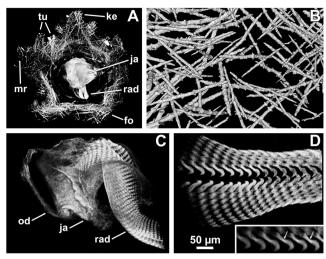


Fig. 2 Micro-CT photographs of sclerotized structures of *Doridunculus punkus* n. sp. **a** Transverse section of the cephalic region showing spicules' arrangement in the body wall. **b** Detail of the notal spicules. **c** Lateral view of the buccal mass. **d** Distal part of the radula (approximate scale); close-up showing inner lateral teeth, note longitudinal denticles (*white arrows*). *fo* foot, *ja* jaw, *ke* keel, *mr* mantle rim, *od* oral disk, *rad* radula, *tu* tubercles

lamellae, retractile within smooth cavities. Notum dorsally keeled, starting right in front of rhinophores, extending posteriorly to gills, forming a protuberance tilted toward left side of gills (Fig. 1e); average height of keel about 2 mm. Dorsal

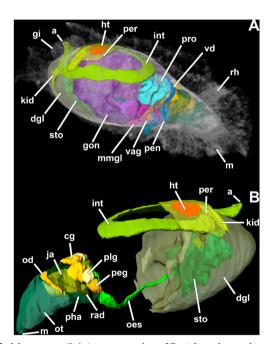


Fig. 3 Macro-scan $(0.4\times)$ reconstruction of *Doridunculus punkus* n. sp. a Right-lateral, general view of all reconstructed organs. b Left-lateral view of the digestive system. a anus, cg cerebral ganglia, dgl digestive gland, gi gills, ht heart, int intestine, ja jaws, gon gonad, kid kidney, m mouth, mmgl membrane + mucus glands, od oral disk, oes oesophagus, ot oral tube, pen penis, peg pedal ganglia, per pericardium, pha pharynx, plg pleural ganglia, pro prostate, rad radula, rh rhinophore, sto stomach, vag vagina, vd vas deferens

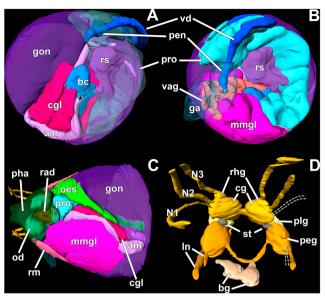


Fig. 4 Micro-CT reconstruction of the anterior body region of *Doridunculus punkus* n. sp. (4×). a Antero-lateral view of the reproductive system; membrane + mucus glands and vagina not depicted; *asterisk* showing connection between distal ampulla and prostate. b Frontal view of the reproductive system. c Ventral view of digestive, nervous, and reproductive systems. d Frontal view of the nervous system; missing nerves are depicted with *white-dashed lines.* am ampulla, bc bursa copulatrix, bg buccal ganglia, cg cerebral ganglia, cgl capsule gland, ga genital atrium, gon gonad, ln lateral nerves, mmgl membrane + mucus glands, N1 oral nerve, N2 labial nerve, N3 rhinophoral nerve, od odontophore, oes oesophagus, pen penis, peg pedal ganglia, pha pharynx, plg pleural ganglia, pro prostate, rad radula, rhg rhinophoral ganglia, rm retractor muscles, rs receptaculum seminis, st statocyst, vag vagina, vd vas deferens

notum heterogeneously covered by pointed, conical, spiculated papillae; smaller in notal periphery and at sides of dorsal keel (Fig. 1c). Gills five, shortly pinnate, arranged in semicircle (Fig. 1a, c, f); anal papilla small, placed just behind gills. Genital pore small, situated antero-laterally beneath

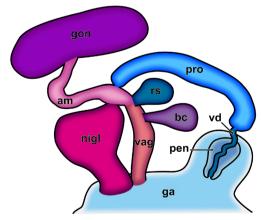


Fig. 5 Schematic outline of the reproductive system of *Doridunculus punkus* n. sp. *am* ampulla, *bc* bursa copulatrix, *ga* genital atrium, *gon* gonad, *nigl* nidamental glands (capsule, membrane, and mucus glands), *pen* penis, *pro* prostate, *rs* receptaculum seminis, *vag* vagina, *vd* vas deferens





notum rim. Spicules conspicuous, fusiform, irregularly multiknobbed, sometimes with median crown of knobs (Fig. 2a); densely lying within notum, imparting rough texture, forming dense network toward epidermis (Fig. 2b).

Digestive system (Figs. 2c, d, 3, and 4c) Mouth lying ventrally, opening at base of oral veil, right in front of foot edge, in vertical furrow. Oral tube pyriform, surrounded by follicular oral glands. Oral disk thin, smooth, cuticular; demarcating transition into pharynx. Pharynx bulbous ventrally; two thick longitudinal muscles arise from postero-lateral part reaching gonad laterally (Fig. 4c). Jaws ear-shaped, without distinct ornamentation. Odontophore placed at rear part of pharynx. Radular formula 41 × 10.1.0.1.10 (Fig. 2c, d). Innermost lateral hook-shaped, with one longitudinal denticle at middle part (see white arrows in Fig. 2d), flatened at base. Marginal teeth romboid in shape, presenting outer pointed cusp; decreasing in size toward lateral rim. Rachidian not observed. Salivary glands paired, saccular, small; lying behind pharynx, connected to it through small, thin, tubular ducts. Oesophagus thin, originating from posterior part of pharynx; passing through nerve ring, widening toward ventral side, reaching stomach far posteriorly; partly covered by gonad. Stomach completely surrounded by holohepatic digestive gland, situated in posterior third of viscera (Fig. 3a, b). Digestive gland presenting reticulated pattern, more pronounced toward periphery. Intestine thin, originating mid-dorsally from stomach, widening, becoming compressed, forming pronounced loop to right side, returning toward left side, ending in anal papilla.

Reproductive system (Figs. 4a–c and 5) Triaulic. Gonad placed in middle region of body in front of digestive gland; globular in appearance, presenting up to 60 follicles. Gonoducts large connecting in front of gonad, converging into thin, tubular ampulla. Ampulla lying on gonad, extending ventrally (Fig. 4a). Vas deferens originating from distal gonoduct close to ampulla, leading into big, folded prostate; prostate making counterclockwise loop alongside body wall, connecting dorsally to thick distal vas deferens, proximally making short loop (Fig. 4b). Penis small, short, apparently unarmed; placed inside large, globe-shaped, muscular, penial sheath, lying close to genital atrium in right antero-lateral side of body (Fig. 4a, b).

Vaginal duct originating from proximal oviduct, close to ampulla forming a very short uterine duct, entering saccular receptaculum seminis. Bursa copulatrix saccular, roundish, originating from thin duct distally from vagina, placed in central transverse position. Vagina relatively short, opening into genital atrium. Oviduct short, placed at distal gonoduct leading to capsule gland. Capsule gland extending far posteriorly into gonad. Membrane and mucus glands not clearly distinct from each other, embracing capsule gland, extending latero-

ventrally close to body wall (Fig. 4c). See overall schematic outline of reproductive system in Fig. 5.

Nervous system (Fig. 4d) Cerebral ganglia large, interconnected by short commissure; laterally oral nerve (N1) reaching mouth; labial nerve (N2) reaching anterior part of oral tube; rhinophoral nerve (N3) connected through small rhinophoral ganglia to cerebral ganglia, innervating rhinophores. Neither optical nerve, optic ganglion, nor eyes detected. Pleural ganglia attached to cerebral ganglia, without visible connectives. Statocysts present on ventral side of pleural ganglia, close to pharynx. Pedal ganglia large, interconnected by long commissure; with two lateral nerves running to ventral side. Two buccal ganglia, small, neighboring, lying at base of salivary glands, partly surrounded by them.

Circulatory and excretory systems (Fig. 3) Pericardium flattened, placed at posterior part of body, above kidney, intestine, and digestive gland. Heart placed in longitudinal axis, no obvious distinction between auricle and ventricle. Aorta and blood gland not observed. Kidney lying below pericardium, extending back behind digestive gland; no nephroduct observed.

Ecology The single specimen of *D. punkus* n. sp. was found in a detritus-rich benthic bottom at 228-m depth. The community was dominated by sessile phyla, such as sponges (*Cinachyra*, *Clathria*, *Isodyctia*, *Iophon*, *Tedania*), bryozoans (*Alcyonidium*, *Carbasea*, *Isoschizoporella*, *Notoplites*, *Reteporella*), gorgonians (*Thouarella*, *Primnoisis*), ascidians (*Aplidium*, *Cnemidocarpa*), and pterobranchs (*Cephalodiscus*). Vagile fauna such as nudibranchs (*Doris*, *Doto*, *Tritonia*, *Tritoniella*), sea cucumbers, and polychaetes were also collected at the same station.

Etymology *D. punkus* n. sp. is named after the Mohican hair-style of the punks, referring to the presence of one keel in the dorsum.

Remarks

D. punkus n. sp. differs from its congeners by the small folded oral tentacles, rounded (instead of bilobed) posterior notum, and by having a notal rim not covering the foot. The new species presents a single dorsal keel, while D. echinulatus presents two keels, more or less separated (Sars 1878; Odhner 1907), and D. unicus lacks them (Martynov and Roginskaya 2005). Both D. punkus n. sp. and D. echinulatus lack a rachidian tooth. In D. unicus, the rachidian is present and it is similar to that of Akiodoris and Armodoris (Millen and Martynov 2005). A hook-shaped inner lateral tooth is an autapomorphy of the herein described species, not found in





any of the described species of Akiodorididae. The stomach of *D. punkus* n. sp. is completely enveloped by the digestive gland, while it is fully free in *D. unicus* (Martynov and Roginskaya 2005). Cerebral and pleural ganglia are separated in *D. punkus* n. sp., similarly to *Armodoris* (Millen and Martynov 2005). In *D. punkus* n. sp., eyes were not observed; this character was not specified in *D. echinulatus* (Sars 1878; Odhner 1907), while *D. unicus* possesses eyes (Millen and Martynov 2005).

Discussion

D. punkus n. sp. is the first record of the genus in the southern hemisphere, since the two other species were only recorded previously in the North Atlantic and the Sea of Japan (Sars 1878; Martynov and Roginskaya 2005). It seems plausible that the whole family is restricted to either polar and/or deep waters. We were able to reconstruct both hard and soft tissues of the single specimen collected using micro-CT techniques. The new species is placed within the genus *Doridunculus* since it conforms with most of the characters of the hitherto described species (see Table 1). For instance, D. punkus n. sp. and its congeners exclusively possess a dorsal ridge on the tail. Their notum does not cover the foot posteriorly, as in Armodoris and Prodoridunculus (Minichev 1972; Valdés and Bouchet 1998). However, D. punkus n. sp. differs from congeners in having a rounded notum posteriorly, like in *Prodoridunculus*, and not bilobed as in D. echinulatus, D. unicus, and Echinocorambe. Moreover, D. punkus n. sp. presents a notal rim extending laterally forming flaps, as in all other Akiodorididae. A single dorsal keel similar in position and height to the two present in D. echinulatus is also a diagnostic character of the new species (named after it). The gills are arranged in a semicircle in all akiodoridids (Millen and Martynov 2005). The lack of a branchial pocket to withdraw the gills, a feature also typical for the new species, was considered a synapomorphy of Akiodorididae, separating this group from other Onchidoridoidea (Millen and Martynov 2005).

In the buccal apparatus, a thin and smooth lip disk is shared among akiodoridids. The radular structure is quite different in *D. punkus* n. sp.; it lacks a rachidian tooth, although a rudimentary thin plate could have possibly been unnoticed in the micro-CT scan. In fact, the other species lacking the rachidian teeth were not analyzed using scanning electron microscopy (SEM), i.e., *D. echinulatus* and *P. gaussianus* (Sars 1878; Thiele 1912); thus, this absence is not well documented (Martynov and Roginskaya 2005). Remarkably, *D. punkus* n. sp. is the first member of the Akiodorididae that possess a large hook-shaped inner lateral tooth, suggesting that multiple inner lateral teeth evolved once within Onchidoridoidea (Hallas and

Gosliner 2015). Indeed, the inner lateral tooth of most akiodoridids still presents a strong curved cusp, thus resembling a rudimentary hook. Likewise, the whole family seems to have square marginal teeth decreasing in size toward the outer edge. The radula of *D. punkus* n. sp., as well as many Goniodorididae taxa, is similar to that of the Polyceridae, although the latter are lacking the innermost reduced lateral tooth (e.g., Vallès et al. 2000). Therefore, Wägele and Willan (2000) postulated that the innermost lateral tooth of the Goniodorididae actually could represent the second lateral of the Polyceridae. This might also be the case in *D. punkus* n. sp.

Our tomographic analyses reveal that the pharynx of D. punkus n. sp. is bulged and forms a buccal pump, as for all Akiodorididae, although the presence of two wide longitudinal retractor muscles at each side of the pharynx was never reported in this family before. The stomach is completely enclosed by the digestive gland in D. punkus n. sp., contrary to Akiodoris and Armodoris (Millen and Martynov 2005) and probably D. unicus as depicted by Martynov and Roginskaya (2005), albeit not reported therein. The intestine is dorsal and forms a pronounced loop forward, as in all studied species of Akiodorididae. The anus is dorsal in all species, except for the aberrant Echinocorambe brattegardi (Valdés and Bouchet 1998), where it is ventral, and thus, strongly resembling the species of the genus Corambe (Martynov and Schrödl 2011). The digestive gland is overlapping the gonad in *Doridunculus*, Akiodoris, and Armodoris (Martynov and Roginskaya 2005; Millen and Martynov 2005; Valdés et al. 2011). However, only in D. punkus n. sp., the gonad is restricted to the midlongitudinal section of the animal, which could be attributed to the ontogenetic stage. In the new species, the ampulla is thin and tubular and connects with a receptaculum seminis by a short uterine duct, similar to Akiodoris (Millen 1985). The ampulla leads to a vaginal duct which has a saccular bursa copulatrix placed distally. The prostate can be tubular and club-shaped, and it is always wide and voluminous in Akiodorididae. The penis is normally unarmed, except in Akiodoris and Armodoris, where it is densely covered with spines (Millen and Martynov 2005).

In conclusion, *D. punkus* n. sp. shares most of the characters with Akiodorididae. However, it also looks externally similar to *Aegires albus* Thiele, 1912 found in the same waters (Wägele 1987a). Both species are of similar color, size, and shape, presenting dorsal, irregularly scattered tubercles and spicules (Wägele 1987b). Nevertheless, *D. punkus* n. sp. presents a rather squared anterior part, the mantle rim protruding laterally, and a dorsal keel. On the other hand, the external appearance of the new species (i.e., elongated notum, trailing ridged foot) strongly resembles that of the genus *Diaphorodoris* Iredale & O'Donoghue, 1923 (Millen 1985). *Diaphorodoris* is presently assigned to the family Calycidorididae together with the Arctic monotypic genus *Calycidoris* Abraham, 1876 (Hallas and Gosliner 2015).





 Table 1
 Comparative table of diagnostic characters of the Akiodorididae genera, including all the species of Doridunculus

	Doridunculus punkus n. sp.	Doridunculus echinulatus G.O. Sars, 1878	Doridunculus unicus Martynov & Roginskaya, 2005	Akiodoris Bergh, 1892	Armodoris Minichev, 1972	Echinocorambe Valdés & Bouchet, 1998	Prodoridunculus Thiele, 1912
Notum Rim Tail Posteriorly Dorsal tubercles	Elevated Expanded laterally Not covered Rounded Conical	Elevated Expanded ventrally Not covered Bilobed Elongated, conical	Elevated Expanded ventrally Not covered Bilobed Elongated, cylindro-conical	Elevated Expanded Not covered Rounded Elongated or rounded	Elevated Expanded ventrally Covered Rounded Rounded	Flattened Expanded ventrally Not covered Bilobed Elongated	Flattened Expanded ventrally Covered Rounded Small and big, conical in four longitudinal
Ridge Spicules Rhinophores Sheath Gills Position Shape	One, mid-dorsal Rod-like, tuberculate Conical Smooth 5, semicircle Dorsal Pinnate	Two, mid-dorsal Present Large Smooth 3–5, semicircle Dorsal Pinnate	Absent Rod-like, normally hollow Large Few tubercles 6–10, semicircle Dorsal Unipinnate and bipinnate	Absent Rod-like or quadrate Conical Tuberculated 4–17, semicircle Dorsal Bipinnate, tripinnate, or	Absent Rod-like, straight Short, wide Tuberculated 5, semicircle Dorsal Unipinnate	Absent Absent Conical Smooth 1 Ventral Smooth	rows Absent Present n.s. n.s. n.s.
Tentacles Radular formula Rachidian Inner lateral(s)	Small, folded 10.1.0.1.10 Absent Hook-shaped	Club-shaped 1–6.0.6–1 Absent Inner cusp, 4–5	Large, flattened 4-9.1.1.1.1.9-4 Trapezoidal, long cusp Inner cusp, 2-6 denticles	quadripinnate Rounded or flap-like 3–13.2.1.2.13–3 Wide, arch-shaped, or very small Inner cusp, 2–3 denticles	Rounded 4-8.4-6.14-6.8-4 Plate-like, with or without central cusp Inner rounded cusp,	Large 4.3.1.3.4 Cuspless plate Inner cusp, 3–4 denticles	n.s. 2.2.0.2.2 Absent Inner cusp, 3-4 denticles
Marginal Stomach Reproductive	Squared, one large outer cusp Fully free from digestive gland Triaulic	Squared, one large outer cusp n.s.	Squared, one large outer cusp Partly covered by digestive gland? Triaulic	Squared Fully free from digestive gland Triaulic	Inner cusp, 6–0 denticles Partly covered by digestive gland Triaulic	Squared, one large outer cusp n.s.	Squared, one large outer cusp n.s.
System Penis Ampulla Gonad Bursa copulatrix Receptaculum	Unarmed Tubular Free, not covered by digestive gland Saccular Saccular	n.s. n.s. n.s. n.s.	Unarmed Voluminous, club-shaped Partly covered by digestive gland? Flattened, saccular Long and narrow	Armed Tubular Partly covered by stomach and digestive gland Wide, saccular	Armed Tubular or voluminous bean-shaped Partly covered by stomach and digestive gland Oval, large Small or long	Unarmed n.s. n.s. n.s.	n.s. n.s. n.s. n.s.
Cereborpleural ganglia Eyes Distribution Depth range (m)	Separated Not observed Eastern Weddell Sea (Antarctica) 228 Present study	n.s. n.s. Norwegian Sea 80–100 Sars 1878	Fused Present Sea of Japan 3000–3620 Martynov and Roginskaya 2005	Fused Present Sea of Okhotsk and British Columbia (North Pacific) 10–780 Millen and Martynov 2005	Separated Present Davies Sea, Ross Sea, and South Shetland Islands (Antarctica) 25–40 Millen and Martynov 2005, Valdés et al. 2011	Absent Norwegian Sea 2538–3016 Valdés and Bouchet 1908	n.s. Davies Sea (Antarctica) n.s. Thiele 1912

n.s. not specified



But, *D. punkus* n. sp., as well as all akiodoridids, lacks a semicontractible branchial pocket into which gills can be retracted, a feature typical for Calycidorididae and some Onchidorididae (Fahey and Valdés 2005; Martynov et al. 2009). Morphological and phylogenetic studies placed Akiodorididae as sister group of Goniodorididae, sharing a receptaculum seminis that is connected to the uterine duct (Hallas and Gosliner 2015). Instead, in Calycidorididae, the uterine duct is situated independently on the vagina, and in Corambidae and Onchidorididae, the receptaculum is twice connected to the vagina and the uterine duct (Millen 1985; Fahey and Valdés 2005; Millen and Martynov 2005; Martynov and Schrödl 2011). Overall, all these facts support the placement of the new species within Akiodorididae and altogether within Onchidoridoidea.

Our finding of a representative of *Doridunculus* in the SO strongly suggests a bipolar distribution of the genus. Current bipolar disjunct distributions may have been the result of periods of dispersal and/or vicariant isolation, which have occurred several times in Earth's history (Crame 1993). Meridional deep flows were stronger in periods of climate cooling and may have formed dispersion bridges of animals from temperate and cold zones of one hemisphere to another (Vinogradova 1997). Alternatively, a prior cosmopolitan distribution in cooler times may have caused vicariant isolation during interglacial periods (Crame 1993; Allcock and Griffiths 2015). Nudibranchs, as well as other heterobranch taxa, may have originated in Antarctica (Wägele et al. 2008; Göbbeler and Klussmann-Kolb 2010; Moles et al. 2016b). This assumption is based on the presence of basal members of several heterobranch lineages in these waters. Therefore, it is plausible to assume an Antarctic origin of Akiodorididae, and the later dispersion out of Antarctica in glacial periods, a common pattern observed for other different taxa, such as cnidarians, priapulids, polychaetes, amphipods, copepods, isopods, tanaidaceans, holothuroids, and ophiuroids (Vinogradova 1997; Stepanjants et al. 2006; Brandt et al. 2007; Clarke 2008; Allcock and Griffiths 2015). Glacial periods implied periods of environmental stress for Antarctic fauna, resulting in a dramatic decrease in diversity (Zinsmeister 1982). Shelf fauna was completely impoverished by grounded ice masses during glacial maxima, inducing the sheltering migration into marine oases (polynyas) and deepsea waters (Thatje et al. 2005, 2008). Consequently, species could have migrated using deep-water gateways, such as the Antarctic Bottom Water, as a part of the global thermohaline circulation system (Stepanjants et al. 2006; Pawlowski et al. 2007). The lack of molecular data of Akiodorididae precludes answering the mechanisms of such distribution. Therefore, a thorough taxon sampling in remote areas and deep-sea waters as well as molecular clock analyses are essential for revealing the phylogeographic history, including origin, dispersion, and speciation of Akiodorididae.

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References

- Allcock, A.L., & Griffiths, H.J. (2015). Bipolarity. In *Biogeographic Atlas of the Southern Ocean* (pp. 431–436).
- Barker, P. F., & Thomas, E. (2004). Origin, signature and palaeoclimatic influence of the Antarctic circumpolar current. *Earth-Science Reviews*, 66, 143–162.
- Bergh, R. (1892). Malacologische Untersuchungen, 3. System der Nudibranchiaten Gasteropoden. In Carl Semper, Reisen im Archipel der Philippinen, Zweiter Theil. Wissenschaftliche Resultate Band 2, Theil 3, Heft 18 (pp. 995–1165).
- Brandt, A., Gooday, A. J., Brandão, S. N., Brix, S., Brökeland, W., Cedhagen, T., Choudhury, M., Cornelius, N., Danis, B., De Mesel, I., Diaz, R. J., Gillan, D. C., Ebbe, B., Howe, J. A., Janussen, D., Kaiser, S., Linse, K., Malyutina, M., Pawlowski, J., Raupach, M., & Vanreusel, A. (2007). First insights into the biodiversity and biogeography of the Southern Ocean deep sea. *Nature*, 447, 307–311. doi:10.1038/nature05827.
- Clarke, A. (2008). Antarctic marine benthic diversity: patterns and processes. *Journal of Experimental Marine Biology and Ecology*, 366, 48–55.
- Crame, J. A. (1993). Bipolar molluscs and their evolutionary implications. *Journal of Biogeography*, 20, 145–161.
- De Broyer, C., Clarke, A., Koubbi, P., Pakhomov, E., Scott, F., Vanden Berghe, W & Danis, B., (eds). (2016). The SCAR-MarBIN Register of Antarctic Marine Species (RAMS), [06/04/2016]. World Wide Web electronic publication. Available online at http://www.scarmarbin.be/scarramsabout.php.
- Dell, R.K. (1990). Antarctic Mollusca, with special reference to the fauna of the Ross Sea. *Bulletin of the Royal Society of New Zealand*, 27.
- Fahey, S. J., & Valdés, A. (2005). Review of Acanthodoris gray, 1850 with a phylogenetic analysis of Onchidorididae Alder and Hancock, 1845 (Mollusca, Nudibranchia). Proceeding of the California Academy of Sciencies, 56, 213–273.
- Göbbeler, K., & Klussmann-Kolb, A. (2010). Out of Antarctica?—new insights into the phylogeny and biogeography of the Pleurobranchomorpha (Mollusca, Gastropoda). *Molecular Phylogenetics and Evolution*, 55, 996–1007. doi:10.1016/j. ympev.2009.11.027.
- Hallas, J. M., & Gosliner, T. M. (2015). Family matters: the first molecular phylogeny of the Onchidorididae gray, 1827 (Mollusca, Gastropoda, Nudibranchia). *Molecular Phylogenetics and Evolution*, 1827, 16–27. doi:10.1016/j.ympev.2015.03.015.
- Jun, W., Oliver, T. A., Ducklow, H. W., Amaral-Zettler, L. A., & Sogin, M. L. (2012). Marine bacteria exhibit a bipolar distribution. doi:10.1073/pnas.1212424110.
- Martynov, A. V., & Roginskaya, I. S. (2005). A new species of the genus *Doridunculus* G.O. Sars, 1878 (Mollusca, Nudibranchia): a hydroid-feeding dorid from the abyssal depths of the sea of Japan. *Ruthenica*, *14*, 135–145.





- Martynov, A. V., & Schrödl, M. (2011). Phylogeny and evolution of corambid nudibranchs (Mollusca: Gastropoda). Zoological Journal of the Linnean Society, 163, 585–604. doi:10.1111/j.1096-3642.2011.00720.x.
- Martynov, A. V., Korshunova, T., Sanamyan, N., & Sanamyan, K. (2009). Description of the first cryptobranch onchidoridid. *Zootaxa*, 43, 1–43.
- Millen, S. V. (1985). The nudibranch genera Onchidoris and Diaphorodoris (Mollusca, Opisthobranchia) in the northeastern Pacific. The Veliger, 28, 80–93.
- Millen, S. V., & Martynov, A. (2005). Redescriptions of the nudibranch genera Akiodoris Bergh, 1879 and Armodoris Minichev, 1972 (suborder Doridacea), with a new species of Akiodoris and a new family Akiodorididae. Proceeding of the California Academy of Sciencies, 56, 1–22.
- Minichev, Y. (1972). Opisthobranchiate molluscs of the Davis Sea. Issledovaniya Fauny Morey, 19, 358–382.
- Moles, J., Wägele, H., Ballesteros, M., Pujals, Á., Uhl, G., & Avila, C. (2016a). The end of the cold loneliness: 3D reconstruction of *Doto antarctica* (Heterobranchia: Nudibranchia) and description of the sympatric *D. carinova* n. Sp. *PloS One, 11*, e0157941.
- Moles, J., Wägele, H., Schrödl, M., & Avila, C. (2016b). An Antarctic opisthobranch clade is sister to all other Cephalaspidea (Gastropoda: Heterobranchia). *Zoologica Scripta*. doi:10.1111/zsc.12199.
- OBIS. (2016). Global biodiversity indices from the Ocean Biogeographic Information System. Intergovernmental Oceanographic Commission of UNESCO. http://www.iobis.org.
- Odhner, N. H. J. (1907). Northern and Arctic invertebrates in the collection of the Swedish State Museum (Rijksmuseum). III. Opisthobranchia and Pteropoda. *Kungliga Svenska Vetenskaps-Akademiens Handlingar*, 41, 1–118.
- Odhner, N. H. J. (1922). Norwegian opisthobranchiate Mollusca in the collections of the zoological museum of Kristiania. Nyt Magazin for Naturvidenskaberne, 60, 1–47.
- Pawlowski, J., Fahrni, J., Lecroq, B., Longet, D., Cornelius, N., Excoffier, L., Cedhagen, T., & Gooday, A. J. (2007). Bipolar gene flow in deep-sea benthic foraminifera. *Molecular Ecology*, 16, 4089–4096.
- Raguá-Gil, J. M., Gutt, J., Clarke, A., & Arntz, W. E. (2004). Antarctic shallow-water mega-epibenthos: shaped by circumpolar dispersion or local conditions. *Marine Biology*, 144, 829–839.
- Rudman, W. B. (1972). The genus *Philine* (Opisthobranchia, Gastropoda). *Proceedings of the Malacological Society of London*, 40, 171–187.
- Sars, G. O. (1878). Bidrag til kundskaben om Norges Arktiske Fauna. I. Mollusca regions arcticae Norvegiae. Christiania, 1–426.
- Sombke, A., Lipke, E., Michalik, P., Uhl, G., & Harzsch, S. (2015). Potential and limitations of X-ray micro-computed tomography in arthropod

- neuroanatomy: a methodological and comparative survey. *Journal of Comparative Neurology*, 523, 1281–1295. doi:10.1002/cne.23741.
- Stepanjants, S. D., Cortese, G., Kruglikova, S. B., & Bjørklund, K. R. (2006). A review of bipolarity concepts: history and examples from Radiolaria and Medusozoa (Cnidaria). *Marine Biology Research*, 2, 200–241. doi:10.1080/17451000600781767.
- Thatje, S., Hillenbrand, C.-D., & Larter, R. (2005). On the origin of Antarctic marine benthic community structure. TRENDS in Ecology and Evolution, 20, 534–540.
- Thatje, S., Hillenbrand, C.-D., Mackensen, A., & Larter, R. (2008). Life hung by a thread: endurance of Antarctic fauna in glacial periods. *Ecology*, 89, 682–692.
- Thiele, J. (1912). Die Antarktischen Schnecken und Muscheln. Deutsche Südpolar-Expedition (1901-1903) Im auftrage des Reichsamtes des Innern Herausgegeben von Erich von Drygalski Leiter der Expedition. Zoologi, 13, 183–286.
- Valdés, Á., & Bouchet, P. (1998). A blind abyssal Corambidae (Mollusca, Nudibranchia) from the Norwegian Sea, with a reevaluation of the systematics of the family. *Sarsia*, 83, 15–20. doi:10.1080/00364827.1998.10413665.
- Valdés, Á., Moran, A. L., & Woods, H. A. (2011). A new species of Armodoris (Mollusca, Gastropoda, Nudibranchia, Akiodorididae) from McMurdo sound, Antarctica. Polar Biology, 34, 459–463. doi:10.1007/s00300-010-0889-6.
- Vallès, Y., Valdés, Á., & Ortea, J. (2000). On the phanerobranch dorids of Angola (Mollusca, Nudibranchia): a crossroads of temperate and tropical species. *Zoosystema*, 22, 15–30.
- Vinogradova, N. G. (1997). Zoogeography of the abyssal and hadal zones. *Advances in Marine Biology*, 32, 325–387.
- Wägele, H. (1987a). The distribution of some Antarctic nudibranchs (Opisthobranchia). *Journal of Molluscan Studies*, 53, 179–188.
- Wägele, H. (1987b). Redescription and anatomy of Aegires (Anaegires) albus Thiele, 1912 (Opisthobranchia, Doridacea) and synonymy with A. protectus Odhner, 1934. Polar Biology, 7, 267–272.
- Wägele, H., & Willan, R. C. (2000). Phylogeny of the Nudibranchia. Zoological Journal of the Linnean Society, 130, 83–181. doi:10.1006/zjls.
- Wägele, H., Klussmann-Kolb, A., Vonnemann, V., & Medina, M. (2008).
 Heterobranchia I. The Opisthobranchi. In W. F. Ponder & D. R.
 Lindberg (Eds.), *Phylogeny and evolution of the Mollusca* (pp. 385–408). Berkeley: University of California Press.
- Warén, A. (1989). New and little known mollusca from Iceland. *Sarsia*, 74, 1–28.
- WoRMS Editorial Board. (2015). World Register of Marine Species. http://www.marinespecies.org at VLIZ. Accessed 05 Nov 2015.
- Zinsmeister, W. J. (1982). Late Cretaceous-early Tertiary molluscan biogeography of the southern Circum-Pacific. *Journal of Paleontology*, 56, 84–102.

