# 6

# Polychaeta

## Lana Crumrine

Well over 200 species of the class Polychaeta are found in waters off the shores of the Pacific Northwest. Larval descriptions are not available for the majority of these species, though descriptions are available of the larvae for at least some species from most families. This chapter provides a dichotomous key to the polychaete larvae to the family level for those families with known or suspected pelagic larva. Descriptions have been gleaned from the literature from sites worldwide, and the keys are based on the assumption that developmental patterns are similar in different geographical locations. This is a large assumption; there are cases in which development varies with geography (e.g., Levin, 1984).

Identifying polychaetes at the trochophore stage can be difficult, and culturing larvae to advanced stages is advised by several experts in the field (Bhaud and Cazaux, 1987; Plate and Husemann, 1994).

Reproduction, Development, and Morphology Within the polychaetes, the patterns of reproduction and larval development are quite variable. Sexes are separate in most species, though hermaphroditism is not uncommon. Some groups undergo a process called epitoky at sexual maturation; benthic adults develop swimming structures, internal organs degenerate, and mating occurs between adults swimming in the water column. Descriptions of reproductive pattern, gamete formation, and spawning can be found in Strathmann (1987). Larval polychaetes generally develop through three stages: the trochophore, metatrochophore, and nectochaete stages. Trochophores are ciliated larvae (see Fig. 1). A band of cilia, the prototroch, is used for locomotion and sometimes feeding. Trochophore larvae are generally broad anteriorly and taper posteriorly. The anterior and posterior sections of the larva are called the episphere and hyposphere, respectively. They are usually pelagic.

By the metatrochophore stage, two to three segments have usually formed. Parapodia (with or without setae) may by this developmental stage become apparent. Doral and ventral podia are know as neuropodia and notopodia, respectively. The anterior-most segment is called the prostomium. Just posterior

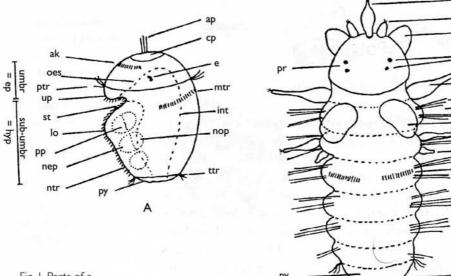


Fig. 1. Parts of a generalized polychaete trochophore (A) and nectochaete (B). (From Korn, 1960) ac, anal cirrus; ak, acrotroch; ap, apical tuft; br, chaetae; cp, cerebral plate; dc, dorsal cirrus; e, eye; el, elytra; ep, episphere; hyp, hyposhere; int, intestine; lo, lower lip; ltl, lateral tentacle; mt, median tentacle; mtr, metatroch; nep, neuropod; nop, notopod; ntr; neurotroch; oes, oesophagus; p, palp; pa, paratroch; pp, parapod; pr, prostomium; proc, proctodaeum; ptr, prototroch; py, pygium; st, stomodaeum; sub-umbr, subumbrella:tlc. tentacular cirrus; ttr, telotroch; umbr, umbrella; up, upper lip; vc, ventral cirrus

to the prostomium is the peristomium. These two anterior segments form the head. Eyes and antennae form in the prostomium. The mouth, palps, or tentacular cirri form in the peristomium. Larvae may be planktotrophic and develop a mouth and gut while in the plankton. Others are lecithotrophic and survive on yolky substances. Once a larva has developed setae, the segments with setae are called setigers. The most posterior segment is called the pygidium. New segments develop from the pygidium in a stepwise manner.

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Advanced larvae are called nectochaetes (see Fig. 1). Nectochaetes typically have many more setigers than metatrochophores. Depending on the species, the larval ciliation may have been lost and the larvae may no longer be pelagic. After settlement, larvae may go through a benthic stage called the erpochaete stage, or they may metamorphose to become juvenile worms (Strathmann, 1987).

Polychaete larvae are morphologically complex and diverse. Because of this complexity, a large number of technical terms have been coined to help describe larval anatomy. Fig. 1 and the following glossary should help to make sense of this "foreign language." The definitions in the glossary are from Lacalli (1980) and Bhaud and Cazaux (1987).

achaetous: without setae

**acicula (pl. aciculae):** a stout chitinous rod embedded in one or both parapodial lobes

acrotroch: circlet of cilia in front of the prototroch

**antenna:** sensory appendage arising from the anterior or the dorsal surface of the prostomium

anal cirrus: elongated appendage arising from the pygidium

**apical tuft:** bundle or group of a few cilia projecting from the anterior pole of the larva

capillary seta: hairlike bristle that may be ornamented

chevron: V-shaped teeth laterally on the proboscis

**ciliated pit:** small dorsal ciliated cavity in segment 1, 2, or 3, limiting posteriorly the neurotroch of the spionidae

cirrus: respiratory and tactile appendage of the setiger, without blood vessel

**compound seta:** jointed bristle most often composed of two parts: the base and the distal part

crest (nuchal crest): mediodorsal zone of the cephalic area in the Spionidae bounded by two ciliated grooves

**elytron (pl. elytra):** dorsal scale, inserted on the dorsal side of the parapodium, arising from transformation of some dorsal cirri

erpochaeta: creeping stage, moving on or in the sediment using its setae

**erposoma:** creeping stage, without setae, moving by undulation or contraction of the body

gastrotroch: ventral troch

geniculate seta: seta that is bent but not articulated

**hooded hook:** seta that is curved distally and covered with a transparent envelope

limbate seta (spatulate seta): seta with a bladelike flattened margin

melanophore: black pigment cell or group of cells

**meniscotroch:** crescent-shaped area of short cilia in the region anterior to the prototroch; the cilia of the central part are longer, bent, and form a pointed brush

**mesotroch:** transversal ciliated ring in the middle part of the larval body **metatrochophore:** larval planktonic stage with marked segmentation; if parapodia are not yet formed, stage 1; if parapodia are present but not yet functioning, stage 2

monostichious: troch made of one ring of long cilia

**nectochaeta:** developmental stage bearing functioning parapodia serving locomotion

neuropodium: ventral section of the parapodium

neuroseta: seta of the neuropodium

neurotroch: row of short cilia running along the ventral side of the larva

notopodium: dorsal section of the parapodium

notoseta: seta of the notopodium

**palea (pl. paleae):** simple and stout seta, often enlarged as an oar (palea of parapodium) or simple and stout seta, not enlarged, but regularly tapering (opercular palea)

**palpi:** paired projections arising from the prostomium, used for food gathering and tactile purposes

papilla (pl. papillae): conus-like projection

**parapodium:** lateral segment footlike projections, composed of two parts, neuropodia and notopodia, each bearing a cirrus and setae

peristomium: buccal segment

polystichious: troch made of several rings of long cilia

**proboscis:** eversible anterior portion of the alimentary tract; with or without papillae

**prostomium:** preoral lobe that contains the cerebral ganglia and bears the most important sense organs; posteriorly and ventrally, the prostomium is delimited by the peristomium

**prototroch:** on the equatorial part of the trochophore, a ring of cilia anterior to the mouth; the single ciliated ring of certain trochophores **pygidium:** posterior part of the body bearing the anus, always devoid of coelomic cavity

serrated seta: seta or part of seta with one or two edges notched like a saw

**seta (pl. setae):** slender chitinous structure projecting from the parapodium, used for locomotion and defense

setiger: segment bearing setae simple seta: unjointed bristle telotroch: ring of cilia near the anus

troch: ciliated ring of embryonic or larval stage, used in locomotion trochophore: free pelagic larval stage, top-shaped structure, without visible segmentation, bearing one or two equatorial ciliated rings; in the latter case, the metatroch is situated behind the mouth

uncini: stout spines curved at the distal part; referred to as hooded hooks when the distal part is protected by hoods (see hooded hook); when they are more or less rectangular, with numerous teeth, they are called uncini

# Description and Identification of Local Taxa

Key to larvae of polychaete families from the Pacific Northwest (adapted from Bhaud and Cazaux, 1987; Plate and Husemann, 1994)

Ia. Larval tube present
al. Tube transparent with diameter constant thoughout length,
larva bearing I-several large tentacles, always directed toward
body and equaling at least half its length
TEREBELLIDAE (p. 73)
a2. Tube nontransparent and conical, no large tentacles, externa
paleae directed forwardPECTINARIDAE (p. 73)
Ib. Larva without tube2
2a. Larva bearing   or several stout anterior tentacles
TEREBELLIDAE (p. 73)
2b. No stout anterior tentacles present
3a. Trochophore or metatrochophore with 2 large prototrochal-
ventral lobes fused into funnel, resulting in dorsoventral
asymmetryPECTINARIDAE (p. 73)
3b. Lateral lobes not present
4a. Presence of heavy projecting acicular sickle-shaped hooks on
notopodium of several segments PILARGIDAE (p. 60)

4b. Dorsal sickle-shaped acicular hooks not present5
5a. Body and prostomium covered with stout spherical papillae  SPHAERODORIDAE (p. 63)
5b. Spherical papillae not covering body6
6a. Larva with 50–100+ segmentsTROCHOCHAETIDAE (p. 72)
6b. Larva with <50 segments7
7a. Setae absent
8a. Prostomium broad with marked frontal notch
8b. Prostomium rounded or conical9
9a. Prototroch present with no other ciliary bands
10a. Ciliation present as broad and definitive prototroch, metatroch, telotroch, and neurotrochEUNICIDAE (p. 49) 10b. Ciliation in distinctly narrow bands or not in defined bands
I I a. Eyespots absent, or if present only in conjunction with segmentation
12a. Cilia arranged in narrow bands at segment definitions, with or without short cilia covering rest of larva
I 2b. Cilia long and nearly covering entire larvaLUMBRINERIDAE (p. 50)
13a. 2 bundles of well-delineated setae
14a. Pair of well-developed tentacles present   15     14b. No such pair of tentacles   17
15a. 2 long tentacles, covered with adhesive papillae and internal blood vessel, without ciliated groove, tentacles motile and often coiled in spiral
16a. Tentacles with 2 ciliated grooves, tentacles projecting dorsally and not laterally, conical prostomium without terminal antenna.  AMPHINOMIDAE (p. 46)  16b. 2 laterodorsal tentacles, each with 1 ciliated groove; 2 narrow lateral bundles of setae, slightly bent and armed with regular denticulated collars

17a.Top-shaped larva with roughly conical episphere1817b. Umbrella-, bell- or mushroom-shaped larva19
18a. Transparent body; 2 lateral bundles of slender setae that are smooth or slightly serrated and bright OPHELIDAE (p. 51) 18b. Opaque body; 2 lateral bundles of bent larval setae that are thick, dark, and coarsely serrated on the convex side; transversely striated notosetal spines and paleae; neither tentacles or palpi present
19a. Umbrella-shaped larva; main part of larval body a transparent, unpigmented umbrella; setal sacs set close together inside umbrella
20a. Compact, barrel-shaped larva nearly as wide as long
21a. I–2 narrow ciliated rings, I pygidial cirrus present, adult setae present in early planktonic stages
22a. Metatrochophore with broad ciliary bands covering most of larva; erpochaete with bulbous prostomiumLUMBRINERIDAE (p. 50) 22b. Metatrochophore with broad prototroch, remaining bands
narrow; prostomium rounded or spatulateEUNICIDAE (p. 49)
narrow; prostomium rounded or spatulate

# Polychaeta

28a. 2 pairs of anterior tentacles at extreme end of prostomium  ALCIOPIDAE (p. 53)
29b. Prostomial tentacles absent
29a. Prostomial collar extending posteriorly to third segment
30a. Trochophore and metatrochophore elongated with diameter not varying substantially along length of body; larva at least 3 to 4 times longer than wide
31a. Prostomium and pygidium with large zones of short cilia 32 31b. Prostomium and pygidium with only 1 ring of long cilia 35
32a. 2 pairs of eyes; 3 antennae, possibly short; poorly developed acrotroch, prototroch, and metatroch present, telotroch absent in early trochophore; peristomium achaetous, remaining segments uniramous
33a. Ciliated zones reduced to prototroch and telotroch; conical prostomium with rudiment of the first tentacle; no statocyst  TEREBELLIDAE (p. 73)  33b. Larva with multiple ciliary bands, including acrotroch (in early forms), prototroch, metatroch, telotroch, and neurotroch 34
34a. Larva lacking observable mouth; possessing 13 tufts of cilia on apical end of prostomium; larva with 3 setigerous segments bearing 1 pair of short and 1 pair of long notopodial setae on each segment
35a. Parapodia present
36a. Prostomium flat, rectangular, and short, without antennae; I unpaired anal cirrus; parapodia developed from first segment onward
37a. Parapodia uniramous; 0–4 eyes; parapodia of first segment reduced; erpochaeta bearing chevrons on proboscis; no jaws, opaque body, brown pigmentation

38a. Each setigerous segment bearing I pair of capillary and I pair of spatulate setae; prostomium the largest segment
38b. Each setigerous segment bearing I pair of capillary setae; posterior segments bearing hooked setae
39a.Trochophore with curved acrotroch and tuft of long, thin cilia pressed against hyposphere at level of prototroch; elytra
present on dorsal surface of metatrochophore
39b. Trochophore with equatorial ciliated ring; long tuft of cilia absent from hyposphere or present on episphere;
metatrochophore without elytra42
40a. Tentacular cirri present on peristomium
APHRODITIDAE (p. 53)
40b.Tentacular cirri absent on peristomium41
41a. 4–5 pairs of elytra; segments lacking elytra bearing dorsal cirri;
simple setae; 7–10 larval segments POLYNOIDAE (p. 61)
41b. Four pairs of elytra or elytrophores; dorsal cirri absent or
present only in third segment; setae of neuropodia compound, of notopodia simple; 5–6 larval segments
SIGALIONIDAE (p. 63)
42a. Trochophore with ventral menisotroch; metatrochophore with
2 pairs of eyes and segmentation indistinct; nectochaeta with 4-
5 antennae, unarmed proboscis, lamellate dorsal cirri, several
pairs of smooth and nonarticulate tentacular cirri, compound
and spinigerous notosetae and neurosetae
PHYLLODOCIDAE (p. 58)
42b. Trochophore lacking ventral menisotroch; metatrochophore
with >2 pairs of eyes; clearly delineated segments; anterior part broad; nectochaeta with long and cylindrical dorsal cirri, jointed
tentacular cirri, compound and bent neurosetae
HESIONIDAE (p. 55)

# Order Amphinomida

**Family Amphinomidae** (Local Species 2, Local Species with Described Larvae 0). Development of this family includes a rostraria larva (Fig. 2). Amphinomids are called fire worms because of the discomfort caused when coming in contact with the spines of the adult worms (Fauchald, 1977).

Chloeia entypa (Notopygos labiatus) Chloeia pinnata

**Family Euphrosinidae** (Local Species 2, Local Species with Described Larvae 0). Euphrosinids are considered closely related to, but distinct from, the Amphinomidae (Fauchald

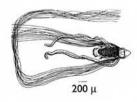


Fig. 2. Rostraria larva, family Amphinomidae. (From Mileikovskii, 1961)

1977). Adults are short and thick-bodied with one pair of antennae, tufts of neurosetae, and no palps. Development is not described.

Euphrosine bicirrata Euphrosine hortensis

## Order Capitellida

Family Arenicolidae (Local Species 4, Local Species with Described Larvae 2). Arenicolid worms spawn freely, brood larvae in burrows, or produce benthic egg masses. Larvae are non-feeding. Two local species (\*) are briefly pelagic during larval development prior to settling (Strathmann, 1987). Abarenicola pacifica develops in gelatinous masses that are brooded within adult tubes. Branchiomaldane vincenti undergoes direct development.

Abarenicola claparedi oceanica (A. vagabunda oceanica, Arenicola pusilla)\* Abarenicola pacifica (Arenicola pusilla, in part) Arenicola marina\* Branchiomaldane vincenti (B. simplex, Protocapitella)

## Key to pelagic arenicolid larvae (Fig. 3)

Family Capitellidae (Local Species 8, Local Species with Described Larvae 2). Intratubular brooding of larvae is quite common among the Capitellidae, though some species are known to broadcast spawn. Likewise, development ranges from direct within the parental tube to lecithotrophic and planktotrophic swimming larvae (Rouse, 1992). Capitella capitata appears to have two different developmental strategies: direct development from jelly masses, and a short pelagic phase. Capitella capitata may be a complex of six sibling species (see Strathmann, 1987), and therefore it is likely that different sibling species use different developmental strategies. Heteromastus filiformis egg masses are attached to the end of the female's tube, and larvae develop to the trochophore stage before release into the plankton (Rasmussen, 1956). Generalization of the development of capitellids based on trochophores is difficult because of generic differences (Lacalli, 1980).

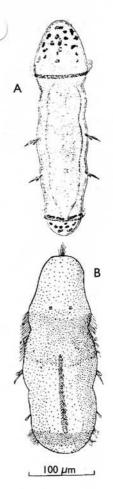
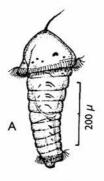


Fig. 3. (A) Arenicola marina and (B) Arenicola claparedi, Family Arenicolidae. (From Okuda, 1946; Newell, 1948)



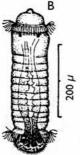


Fig. 4. (A) Heteromastus filiformis and (B) Capitella capitata, family Capitellidae. (From Lacalli, 1980)





Fig. 5. Example of a pelagic maldanid larva, *Clymenella torquata*; not a local species. (From Lacalli, 1980).

Capitella capitata

Heteromastus filiformis

Heteromastus filobranchus

Mediomastus californiensis

Notomastus giganteus (N. magnus)

Notomastus lineatus

Notomastus magnus

Notomastus tenuis (N. lineatus var. balanoglossi)

## Key to pelagic capitellid larvae (Fig. 4)

ded into several sections;
ot subdivided
Capitella capitato

Family Maldanidae (Local Species 13, Local Species with Described Larvae 0). Development for the maldanids was presumed to be primarily nonpelagic by Thorson (1946). Rouse (1992) cites two examples of free-spawned lecithotrophic larvae in the genera *Euclymene* and *Axiothella*. Wilson (1983) suggests that *Axiothella rubrocincta* is a sibling pair with one type brooding larvae and the other broadcasting demersal lecithotrophic larvae (Fig. 5).

Asychis disparidentata
Axiothella rubrocincta
Euclymene reticulata
Isocirrus longiceps
Maldane sarsi
Maldanella harai (Maldane robusta)
Nicomache lumbricalis
Nicomache personata
Notoproctus pacificus
Praxillella affinis var. pacifica
Praxillella gracilis
Rhodine bitorquata

#### Order Cossurida

**Family Cossuridae** (Local Species 1, Local Species with Described Larvae 0). Adult cossurids are common in sand and deep slope and abyssal muds, where they apparently feed on detritus through a pharynx (Fauchald, 1977). Development is not described.

Cossura modica

#### Order Ctenodrilida

Family Ctenodrilidae (Local Species 1, Local Species with Described Larvae 0). Fauchald (1977) describes adult worms of this family as small and grub-shaped. They are especially common in areas of aquaculture and may be commensal with Flabelliderma commensalis and Strongylocentrotus purpuratus. Bhaud and Cazaux (1987) warn that some species of this family are also holoplanktonic, and adults are often confused for larval stages.

Ctenodrilus serratus

#### Order Eunicida

Family Arabellidae (Local Species 4, Local Species with Described Larvae 0). Arabellids are often parasitic on other animals, especially polychaetes and echiurans. Parasitism may be a larval stage or a lifelong condition. For example, Allen (1952) found larvae through young worms of *Arabella iricolor* present in a single *Diopatra cuprea* host. Descriptions and figures of arabellid development are not known, though Pettibone (1957) indicates that *A. iricolor* spawns throughout the summer.

Arabella iricolor Drilonereis falcata Drilonereis filum Notocirrus californiensis

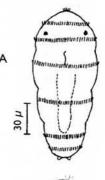
Dorvillea moniloceras

Family Dorvilleidae (Local Species 4, Local Species with Described Larvae 1). Adult dorvilleids are mainly small and common in shallow water (Fauchald, 1977). The described larva, *Schistomeringos longicornis* (Fig. 6), is free-spawned, lecithotrophic, and has been collected swimming at the water surface (Moore, 1903).

Dorvillea pseudorubrovittata Protodorvillea gracilis (Dorvillea, P. kefersteini, P. recuperata, Stauronereis) Schistomeringos longicornis (Dorvillea, Stauronereis, D.

Schistomeringos longicornis (Dorvillea, Stauronereis, D. rudolphi, D. atlantica, Stauronereis articulata)

Family Eunicidae (Local Species 3, Local Species with Described Larvae 0). Eunicids are large polychaetes, mostly associated with hard substrates and shallow water, mainly carnivorous, burrowers, and tube builders (Fauchald, 1977). Eunice valens is known to brood benthic non-feeding larvae (Akesson, 1967). Richards (1967) describes several species that produce benthic egg masses in which the larvae develop.



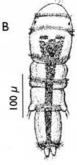


Fig. 6. Schistomeringos longicornis larvae, family Dorvilleidae: (A) late trochophore, (B) metatrochophore. (From Richards, 1967)

Eunice segregata Eunice valens (E. kobiensis) Marphysa stylobranchiata

Family Lumbrineridae (Local Species 8, Local Species with Described Larvae 2). Adults brood directly developing larvae in the two local species (\*) for which development has been described. Lumbrinerids are mostly free-living burrowers in sand and mud or in algal holdfasts. They occur in shallow and deep water (Fauchald, 1977).

Lumbrineris aff. abyssicola
Lumbrineris bicirrita (L. bifurcata)
Lumbrineris californiensis
Lumbrineris cruzensis
Lumbrineris japonica
Lumbrineris latreilli\*
Lumbrineris zonata\*
Ninoe gemmea

Family Onuphidae (Local Species 7, Local Species with Described Larvae 1). Onuphid development is either lecithotrophic or direct, with only one known example of planktotrophy (Sarsonuphis elegan, Fig. 7; Blake, 1975a). Diopatra cuprea broods larvae in egg masses attached to the adult tube and produces either nonpelagic (Monro, 1924) or swimming larvae (Allen, 1959).

Diopatra ornata (Onuphis longibranchiata)
Mooreonuphis stigmatis (Nothria)
Nothria occidentalis (Onuphis)
Nothria geophiliformis (Onuphis)
Nothria iridescens (Onuphis)
Sarsonuphis elegans (Onuphis, Nothria)
Sarsonuphis lepta (Onuphis, Nothria, N. abyssalis)

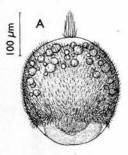
## Order Fauveliopsida

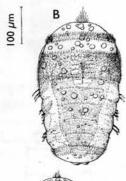
Family Fauveliopsidae (Local Species 1, Local Species with Described Larvae 0). This deep-water family is not well known. Adults are smooth-bodied without anterior appendages (Fauchald, 1977).

Fauveliopsis armata

#### Order Flabelligerida

**Family Flabelligeridae** (Local Species 3, Local Species with Described Larvae 0). Development has not been described for





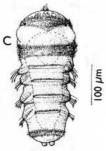


Fig. 7. Sarsonuphis elegans larvae, family Onuphidae: (A) trochophore, (B) metatrochophore, (C) nectochaete. (From

Blake, 1975a)

Polychaeta 51

any local species. Thorson (1946) suggests that Flabelligera affinis larvae are non-pelagic.

Flabelligera affinis (F. infundibularis) Pherusa inflata (Styleroides, Trophonia) Pherusa plumosa (Stylaroides, P. papillata, P. neopapillata)

## Order Ophelida

**Family Ophelidae** (Local Species 7, Local Species with Described Larvae 2). Ophelids have either direct or pelagic development. *Armandia brevis* is planktotrophic and settles at the 20 setiger stage (Hermans, 1978). *Euzonus mucronata* larvae are lecithotrophic and pelagic.

Armandia brevis (A. bioculata)
Euzonus mucronata (Thoracophelia)
Euzonus williamsi (Thoracophelia)
Ophelia limacina (O. borealis)
Ophelina acuminata (Ammotrypane aulogaster)
Travisia brevis
Travisia pupa (T. carnea)

#### Key to pelagic ophelid larvae (Fig. 8)

Family Scalibregmidae (Local Species 3, Local Species with Described Larvae 0). Thorson (1946) collected young polychaetes of this family at 2 m above the sea bottom and therefore assumes that larvae are able to swim or float. Further developmental characteristics are not available.

Asclerocheilus beringianus Hyboscolex pacificus (Oncoscolex) Scalibregma inflatum

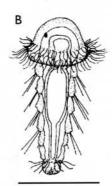
#### Order Orbiniida

Family Orbiniidae (Local Species 7, Local Species with Described Larvae 4). Three of the four described local species (\*) develop in gel masses and spend only a short time swimming as nectochaetes after hatching. *Scoloplos armiger* larvae have been described as nonpelagic (Blake, 1980) and pelagic (Plate and Husemann, 1994).

Leitoscoloplos elongatus (Scoloplos, Haploscoloplos, L. pugettensis)\*



100 μm



100 µm

Fig. 8. Larvae of (A)
Euzonus mucronata and
(B) Armandia brevis, family
Ophelidae. (From Dales,
1952; Hermans, 1977)

Leitoscoloplos panamensis (Scoloplos, Haploscoloplos, H. alaskensis)

Naineris dendritica (N. laevigata)\*

Naineris uncinata (N. berkeleyorum)

Phylo felix (Aricia michaelseni, Orbinia)

Scoloplos acmeceps

Scoloplos armiger\*

## Key to pelagic orbiniid larvae (Fig. 9)

**Family Paraonidae** (Local Species 5, Local Species with Described Larvae 0). Development of Oregon species is not known.

Aedicira pacifica Allia ramosa (Aricidea) Aricidea wassi Cirrophorus lyra (Paraonis) Tauberia gracilis (Paraonis, P. ivanovi)

family Orbiniidae.

(A) Scoloplos armiger;
(B) Naineris dendritica,
(left) metatrochophore,
(right) nectochaete.
(C) Leitoscoloplos
elongatus. (D) Scoloplos
acmeceps, (left)
metatrochophore, (right)
nectochaete. (From

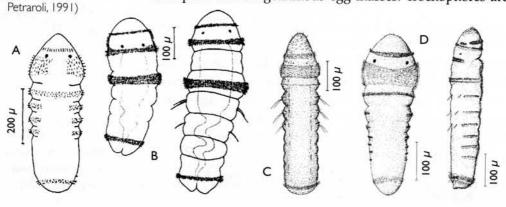
Anderson, 1959; Blake,

1980; Giangrande and

Fig. 9. Larvae from the

# Order Oweniida

Family Oweniidae (Local Species 2, Local Species with Described Larvae 2). Oweniidae worms live in tubes and spawn or deposit stalked gelatinous egg masses. Trochophores are



planktotrophic and develop into a characteristic stage called the mitraria, in which they have triangular bodies with undulating ciliated margins and numerous long flotation bristles (Strathmann, 1987).

Myriochele oculata (M. heeri) Owenia fusiformis (Ammochares)

### Key to pelagic oweniid larvae (Fig. 10)

## Order Phyllodocidae

Family Alciopidae (Local Species 2, Local Species with Described Larvae 0). Descriptions of larvae of local species are not known, though an undetermined alciopid larva is illustrated in Srikrishnadhas and Ramamoorthi (1975); see Fig. 11. Adults of this family are slender-bodied, exclusively pelagic, and known for their large and complex eyes (Fauchald, 1977). Because alciopids are holoplanktonic, adults are often confused with larval stages (Bhaud and Cazaux, 1987).

Alciopa reynaudi Alciopina tenuis (Plotohelmis)

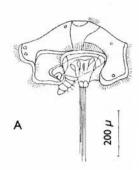
**Family Aphroditidae** (Local Species 5, Local Species with Described Larvae 0).

Aphrodita japonica Aphrodita longipalpa Aphrodita magellanica Aphrodita parva Aphrodita refulgida

Family Chrysopetalidae (Local Species 2,, Local Species with Described Larvae 1). The larvae of *Paleanotus bellis* (Fig. 12) are known for their sluggish behavior, presence of dorsal paleae, and red coloration of the gut (Blake, 1975b). Cazaux (1968) described similar features for *Chrysopetalum debile*, which may indicate that these characteristics are general for the family.

Paleanotus bellis (P. chrysolepis) Paleanotus occidentale (Chrysopetalum)

Family Glyceridae (Local Species 8, Local Species with Described Larvae 4). Glycerids adults are long, slender-bodied



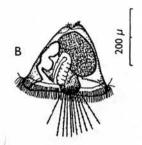


Fig. 10. Larvae of (A) Owenia fusiformis and (B) Myriochele oculata, family Oweniidae. (From Korn, 1960; Lacalli, 1980)

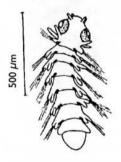
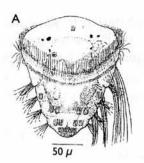
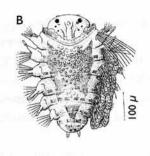


Fig. 11. General alciopid larva. (From Srikrishnadhas and Ramamoorthi, 1975)

Fig. 12. Paleanotus bellis, (A) late metatrochophore, (B) nectochaete, family Chrysopetalidae. (From Blake, 1975b)





with numerous segments, mainly carnivorous, have a long eversible pharynx with four black jaws at the tip, and live in soft sand or mud. Larvae are planktotrophic as trochophores and later, during a benthipelagic stage, they feed on detritus and algae until their jaws are formed (Strathmann, 1987).

Glycera americana Glycera capitata (G. nana) Glycera convoluta Glycera gigantea Glycera oxycephala (G. tenuis) Glycera robusta Glycera tesselata Hemipodus borealis

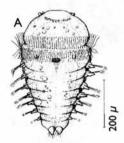




Fig. 13. (A) Hemipodus borealis, late metatrochophore, and (B) Glycera convoluta, late metatrochophore, family Glyceridae. (From Bhaud and Cazaux, 1987; Plate and Husemann, 1994)

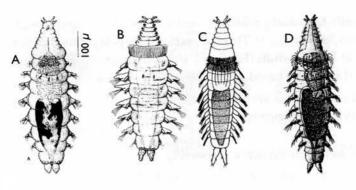
## Key to glycerid metatrochophores (Fig. 13)

Ia. Metatrochophore conical, narrow	ring posteriorly; menisotroch
present	Hemipodus borealis
1b. Metatrochophore barrel- or cigar	
	Glycera convoluta

#### Key to glycerid nectochaetes (Fig. 14)

Ia. Prototroch present2
Ib. Prototroch absent; weak annulations of prostomium
Glycera oxycephala
2a. Prototroch a double band of cilia3
2b. Prototroch a single band of ciliaHemipodus borealis
3a. Prostomium with distinct annulations; body clear with red
chromatophores near prototroch; 9 setigers Glycera capitata
3b. Annulations on prostomium less distinct; 8 setigers; possible
brownish pigment near prototroch

**Family Goniadidae** (Local Species 4, Local Species with Described Larvae 2). Goniadids resemble glycerids in form, but their pharyngeal organs are much larger. Goniadid species may also be distinguished by differences in pharyngeal teeth (Fauchald, 1977).



Glycinde armigera Glycinde polygnatha Glycinde picta Goniada brunnea

## Key to goniadid nectochaetes (Fig. 15)

- Ib. Body with numerous reddish granular pigment markings on each segment and on parapodial lobes of medial and posterior setigers; prostomium narrowly tapered ........... Glycinde polygnatha

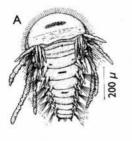
Family Hesionidae (Local Species 2, Local Species with Described Larvae 2). There are few descriptions of hesionid larvae. Direct development and short pelagic lecithotrophic larvae have been described for a few species. *Ophiodromus pugettensis* produces lecithotrophic larvae with a short pelagic phase (Blake, 1975). Adults are common in shallow water and hard substrates; they are fragile and fragment easily (Fauchald, 1977).

Ophiodromus pugettensis (Podarke) Podarkeopsis brevipalpa (Gyptis arenicola glabra; Gyptis)

## Key to hesionid metatrochophores (Fig. 16)

- Ia. Metatrochophore with several long tentacular cirri, some reaching nearly to posterior margin of body ......
  - Podarkeopsis brevipalpa
- 1b. Elongated tentacular cirri, none extending to posterior end of body; 2 lateral antennae on prostomium ......

.. Ophiodromus pugettensis



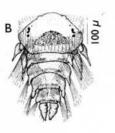


Fig. 14. (A) Glycera oxycephala,
(B) Hemipodus borealis,
(C) Glycera capitata, and
(D) Glycera convoluta,
family Glyceridae. (From Blake, 1975b; Bhaud and Cazaux, 1987; Plate and

Husemann, 1994)

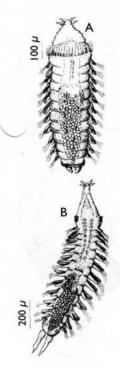
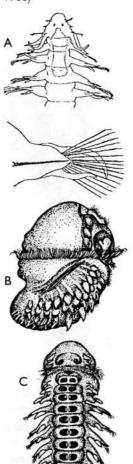


Fig. 15. Larvae of

(A) Glycinde armigera and
(B) G. polygnath, family
Goniadidae. (From Blake,
1975b)

Fig. 16. Larvae of
(A) Podarkeopsis
brevipalpa and
(B) Ophiodromus
pugettensis, family
Hesionidae. (From Blake,
1975b)

Fig. 17. (A) Pelagobia longicirrata nectochaete and cirri structure and (B, C; not to scale) Lopadorhyncus uncinatus metatrochophore and nectochaete, family Lopadorhynchidae. (From Muss, 1953; Akesson, 1966)



Family Lopadorhynchidae (Local Species 2, Local Species with Described Larvae 1). This is an exclusively pelagic family with short-bodied adults (Fauchald, 1977). Distinction between the local species is based on characteristics of the setae.

Lopadorhyncus uncinatus (L. varius) Pelagobia longicirrata

## Key to lopadorhynchid larvae (Fig. 17)

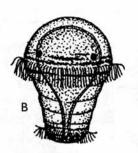
Ia. Dorsal and ventral cirri long and digitiform
Ib. Dorsal and ventral cirri thick and lanceolate
Lopadorhyncus uncinatu

Family Nephtyidae (Local Species 12, Local Species with Described Larvae 1). Worms of the genus *Nephtys* are predatory, free-spawn, and have pelagic development. Typical trochophores have a large dome-shaped episphere, one pair of eyes, and a barrel-shaped trunk with simple setae. Larvae are predatory as well (Strathmann, 1987). Lacalli (1980) suggests that the pigmentation pattern of irregular ruby red to redbrown bands on the larval tegument near the prototroch and pygidium is typical of larvae of the genus. *Nephtys caeca* (Fig. 18) has brownish pigmentation on the episphere and developing prostomium and pygidium, an olive-colored gut, and no blue pigmentation typical of other larvae in this family.

Nephtys assignis
Nephtys caeca
Nephtys caecoides
Nephtys californiensis
Nephtys cornuta cornuta
Nephtys cornuta franciscana
Nephtys ferruginea
Nephtys longosetosa
Nephtys paradoxa
Nephtys punctata
Nephtys rickettsi (N. discors)
Nephtys schmitti

Family Nereidae (Local Species 17, Local Species with Described Larvae 5). Nereid embryos develop in the plankton or in gelatinous benthic egg masses. Larvae do not feed until lipid drops in the gut are depleted and feeding structures have developed (Strathmann, 1987). Nereids either hatch as nectochaetes or proceed rapidly to the nectochaete stage if hatched at an earlier stage of development (Lacalli, 1980).





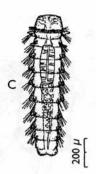


Fig. 18. (A, B) Nephtys caeca trochophore and metatrochophore and (C) general Nephtys nectochaete, family Nephtyidae. (From Lacalli, 1980)

Ceratonereis paucidentata

Cheilonereis cyclurus

Micronereis nanaimoensis (M. variegata, M. bodegae, Phyllodocella)

Nereis brandti (Neanthes)

Nereis eakini

Nereis grubei (N. callaona, N. mediator)

Nereis limnicola (N. lighti, N. diversicolor, N. japonica,

Neanthes)

Nereis natans

Nereis neoneanthes

Nereis pelagica

Nereis procera

Nereis vexillosa

Nereis virens (Neanthes)

Nereis zonata

Nicon moniloceras (Platynereis)

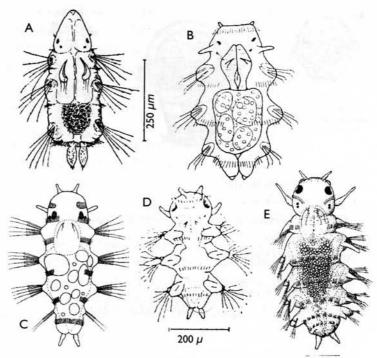
Perinereis monterea

Playtnereis bicanaliculata (P. dumerili var. agassizi)

# Key to nereid nectochaetes (Fig. 19)

Ta. Tentacular cirri present
2a. Prostomium rounded3  2b. Prostomium blunt or indented slightly at anterior margin
3a. Pygidium conical or bilobed; telotroch present but discontinuous
4a. Eyes posterior to prototroch at setiger stage 3, yolk reserves minimal; pygidium lacks posterior cleft

Fig. 19. Larvae of local species from the family Nereidae. (A) Micronereis nanaimoensis (B) Nereis limnicola, (C) Nereis pelagica, (C) Nereis virens (D) Platynereis bicanaliculata. (From Dales, 1950; Berkeley and Berkeley, 1953; Korn, 1960; Blake, 1975b; Plate and Husemann, 1994)



**Family Pholoididae** (Local Species 1, Local Species with Described Larvae 0). Though development of this family is not known, elytra may be seen in larval forms since the family is part of the scale worm group.

Pholoides aspera

Family Phyllodocidae (Local Species 19, Local Species with Described Larvae 6). Female phyllodocids spawn near the bottom or deposit strings of small eggs in benthic gelatinous masses. Larvae hatch and spend several weeks in the plankton. Nectochaetes are large and predatory (Strathmann, 1987). No illustrations of local *Phyllodoce* larvae are available, but descriptions of the genera from Bhaud and Cazaux (1987) are included in the key.

Anaitides groenlandica (Phyllodoce) Anaitides hartmanae (Phyllodoce) Anaitides medipapillata (Phyllodoce) Anaitides mucosa (Phyllodoce) Anaitides multiseriata (Phyllodoce) Anaitides williamsi (Phyllodoce)

Eteone californica

Eteone longa

Eteone pacifica (E. bistriata, E. maculata, E. spitsbergensis var. pacifica)

Eulalia bilineata

Eulalia levicornuta

Eulalia nigrimaculata (Bergstroemia, Eumida, Genetyllis, Phyllodoce)

Eulalia quadrioculata (E. aviculiseta)

Eulalia sanguinea (Eumida)

Eulalia viridis

Notophyllum imbricatum

Notophyllum tectum (Hesperophyllum)

Phyllodoce castanea (Genetyllis)

Phyllodoce polynoides (Paranaitis)

#### Key to phyllodocid metatrochopores (Fig. 20)

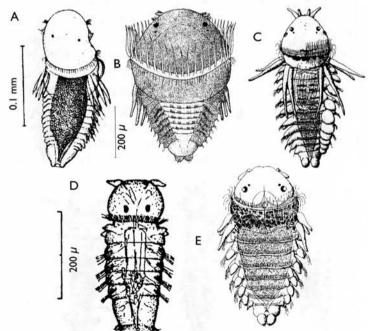


Fig. 20. (A) Anaitides groendlandica, (B) A. williamsi, (C) A. mucosa, (D) Eteone longa, and (E) Eùlalia sanguine, family Phyllodocidae. (From Thorson, 1946; Blake, 1975b; Lacalli, 1980; Bhaud and Cazaux, 1987; Plate and Husemann, 1994)

# Key to phyllodocid nectochaetes (Fig. 21)

Rounded or oval prostomium      Th. Triangular prostomium, no dorsal cirri on setiger 1; pigmented prostomium and pygidium
2a. Oval dorsal cirri
3a. I pair of red eyes; I pair of rounded anal cirri; slight olive-brow to dark green gut; olive-brown pygidium
4a. Dorsal cirri foliaceous and not glandular
5a. Prototroch strongly present or reduced
6a. Dorsal ciliation beginning on setiger 6Anaitides williams 6b. Dorsal ciliation beginning prior to setiger 6Anaitides mucoso

**Family Pilargiidae** (Local Species 4, Local Species with Described Larvae 0). Adults of this family are found in moderately coarse mixed sediments at shelf depths (Fauchald, 1977). Development of local species has not been described, but an unidentified *Ancistrosyllis* (Fig. 22) is described in Blake (1975b).

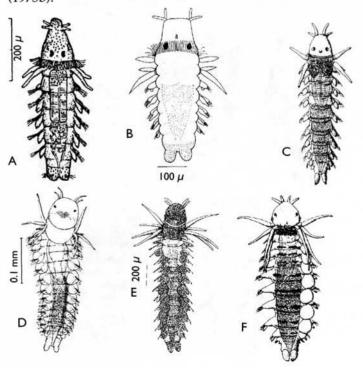


Fig. 21. (A) Eteone longa, (B) Eulalia viridis, (C) E. sanguinea, (E) A. williamsi, and (F) A. mucosa, family Phyllodocidae. (From Thorson, 1946; Blake, 1975b; Lacalli, 1980; Bhaud and Cazaux, 1987; Plate and Husemann, 1994)

Polychaeta

Ancistrosyllis aff. groenlandica Pilargus berkeleyi Sigambra tentaculata Syhelmis aff. klatti

Family Polynoidae (Local Species 21, Local Species with Described Larvae 5). This is the most common family of the scale worm complex. Those polynoids with smooth elytra are often commensal, whereas those with heavily ornamented elytra are free-living (Fauchald, 1977). Some species brood early embryos beneath dorsal elytra; other species spawn freely. Larvae are pelagic for long periods (Strathmann, 1987). Harmothoe brevisetosa larvae develop in the plankton, as do H. imbricata and Lepidonotus squamatus. Harmothoe extenuata and Harmothoe multisetosa larvae are brooded for a period of time between the parapodia (Pettibone, 1954, 1963).

Arctonoe fragilis

Arctonoe pulchra

Arctonoe vittata

Byglides macrolepida (Antinoella, Antinoe)

Eunoe nodosa

Eunoe oerstedi (E. barbata)

Eunoe senta (Gattyana)

Halosydna brevisetosa

Harmothoe extenuata (Lagisca, H. triannulata, L. rarispina)

Harmothoe fragilis

Harmothoe imbricata (H. hartmanae)

Harmothoe lunulata (Malmgrenia, M. nigralba)

Harmothoe multisetosa (Lagisca)

Harmothoe tenebricosa (H. pellucelytris)

Hermadion truncata

Hesperonoe adventor

Hesperonoe complanata

Lepidasthenia longicirrata

Lepidonotus squamatus (L. caekorus)

Polynoe canadensis (Enipo)

Polynoe gracilis (Enipo, Enipo cirrata)

#### Key to polynoid late trochophores (Fig. 23)

Ta. Episphere flattened and careened orally	Halosydna brevisetosa
1b. Episphere domed	
2a. Flat lenticular body, colorless tegument; ve	
green, diameter at prototroch 160 µm	Harmothoe extenuate
2b. Spherical lenticular body	

Figuere 22. Ancystrosyllis sp. nectochaete. (From Blake, 1975b)

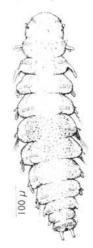


Fig. 23. Late trochophores of species in the family Polynoidae.

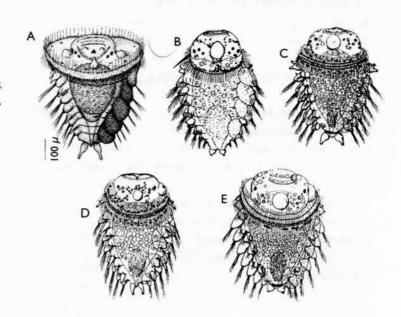
(A) Halosydna brevisetosa,
(B) Harmothoe extenuata,

(C) Harmothoe imbricata, (D) Harmothoe lunulata, and (E) Lepidonotus squamatus. (From Blake, 1975b; Lacalli, 1980; Bhaud and Cazaux, 1987; Plate and Husemann, 1994)

B 480 μm

Fig. 24. Metatro-chophores of species in the family Polynoidae.

(A) Halosydna brevisetosa,
(B) Harmothoe extenuata,
(C) Harmothoe imbricata,
(D) Harmothoe lunulata,
and (E) Lepidonotus
squamatus. (From Blake,
1975b; Lacalli, 1980;
Bhaud and Cazaux, 1987;
Plate and Husemann,
1994)



3a. Ventral side of intestine brown, diameter 180 µm
3b. Ventral side of intestine purplish violet, diameter 240 µm
Harmothoe lunulata
3c. Ventral side of intestine greenish blue, diameter 200 µm
Lepidonotus squamatus

# Key to polynoid metatrochophores (Fig. 24)

Ta. 8 segments	Lepidonotus squamatus
1b. 9 segments	2
2a. Prostomium broad and flattened	
2b. Prostomium rounded	3

a. 4 pairs of elytra4
b. 5 pairs of elytra, ventral side of intestine light yellow-green
a. Pale red-brown pigmentation on prostomium, peristomium, and mouth
b. Pale yellow spots, ventral side of intestine purple-brown
Cey to polynoid nectochaetes (Fig. 25)
a. 8 segments, 4 pairs of elytra, short dorsal cirri Lepidonotus squamatus
b. More than 8 segments, 5 pairs of elytra2
a. Dorsal cirri almost twice as long as neuropodia
a. Elytra with papillae; whitish intestine

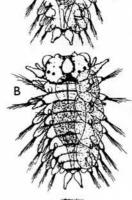
Family Sigalionidae (Local Species 4, Local Species with Described Larvae 1). Larvae of this family are quite similar to those of the Polynoidae. They are somewhat smaller, the elytra are less developed, and they have compound setae rather than simple setae (Lacalli, 1980). *Sthenelais* lack all dorsal elytra; *Pholoe* have dorsal elytra (Plate and Husemann, 1994). Blake (1975b) notes that nectochaetes of *Pholoe caeca* (Fig. 26) feed on algae in culture, though the yolky gut seen in early larval stages suggests that they are lecithotrophic as early larvae. Other members of this family are also described as planktotrophic.

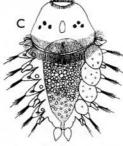
Neoleanira areolata (Leanira calcis) Pholoe caeca (Pholoe tuberculata, P. minuta) Sthenelais berkeleyi (S. fusca) Sthenelais verruculosa

**Family Sphaerodoridae** (Local Species 4, Local Species with Described Larvae 0). Development mode has been described only for *Sphaerodoropsis minuta*. Mileikovskii (1967) states that development is direct and nonpelagic for this species.

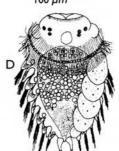
Sphaerodoropsis biserialis (Sphaerodorum, Sphaerodoridium) Sphaerodoropsis sphaerulifer (Ephesia, Sphaerodoridium) Sphaerodorum papillifer (Ephesia) Sphaerodoropsis minuta (Sphaerodorum, Ephesiella, Sphaerodoridium)

Fig. 25. Nectochaetes of species in the family Polynoidae. (A)
Harmothoe imbricata,
(B) Harmothoe lunulata,
(C) Lepidonotus squamatus, and
(D) Harmothoe extenuata. (From Blake, 1975b; Lacalli, 1980; Bhaud and Cazaux, 1987; Plate and Husemann, 1994)





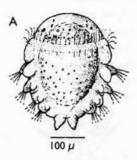
100 um

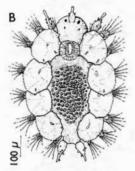


100 um

Fig. 26.

(A) Metatrochophore and (B) nectochaete of *Pholoe caeca*, family Sigalionidae. (From Blake, 1975b)





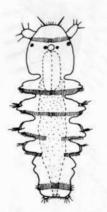


Fig. 27. Autolytus sp., family Syllidae (From Plate and Husemann, 1994)

**Family Syllidae** (Local Species 25, Local Species with Described Larvae 1). Several species are known to have direct development (\*), and Pocklington and Hutcheson (1983) suggest that all *Exogone* species are direct developers (Fig. 27).

Amblysyllis sp. (A. lineata var. alba)\*

Autolytus cornutus (A. prismaticus)

Autolytus varius

Ehlersia cornuta (Syllis, S. heterochaeta, S. alternata, S.

oerstedi, Langerhansia)

Eusyllis assimilis

Eusyllis blomstrandi

Exogone lourei (E. uniformis)\*

Exogone naidina (E. gemmifera)\*

Exogone verugera\*

Haplosyllis spongicola

Odontosyllis parva

Odontosyllis phosphorea (O. phosphorea var. nanaimoensis)

Pionosyllis gigantea

Sphaerosyllis californiensis

Sphaerosyllis hystrix\*

Sphaerosyllis pirifera

Syllis adamantea (S. spenceri, Typosyllis)

Syllis alternata (Typosyllis)

Syllis armillaris (Typosyllis)

Syllis elongata (Typosyllis)

Syllis hyalina (Typosyllis)

Syllis pulchra (Typosyllis)

Syllis variegata (Typosyllis)

Trypanosyllis gemmipara

Trypanosyllis ingens

Family Tomopteridae (Local Species 3, Local Species with Described Larvae 0). There are no descriptions of local tomopterid larvae. Adults of this family are holoplanktonic, transparent, and flattened (Fauchald, 1977).

Tomopteris cavalli

Tomopteris pacifica (T. elegans, T. renata)

Tompoteris septentrionalis

Family Typhloscolecidae (Local Species 2, Local Species with Described Larvae 0). This is a poorly known family. Adults are holoplanktonic, transparent, and fusiform (Fauchald, 1977).

Sagitella kowalevskii

Travisiopsis lobifera

### Order Sabellida

Family Sabellidae (Local Species 25, Local Species with Described Larvae 2). Patterns of development in this family include brooded lecithotrophs, direct developers, freely spawned eggs developing into lecithotrophic larvae, and those that develop in gel masses (reviewed by McEuen et al., 1983).

Chone ecaudata (C. minuta, C. gracilis)

Chone gracilis

Chone infundibuliformis (C. teres)

Chone magna

Chone mollis

Demonax media (Sabella, S. aulaconota, Distylia rugosa, Parasabella, P. maculata, Potamilla californica)

Euchone analis

Euchone sp. cf. hancocki

Euchone incolor (E. rosea, E. trisegmentata, E. barnardi)

Eudistylia polymorpha

Eudistylia vancouveri (E. tenella, E. plumosa, E. abbreviata)

Fabricia brunnea (F. sabella)

Fabricia oregonia

Fabricia sabella (F. dubia)

Fabriciola berkeleyi (F. pacifica, F. sabella)

Megalomma splendida (Branchiomma burrardum)

Myxicola aesthetica

Myxicola infundibulum

Oriopsis gracilis

Potamilla intermedia (Pseudopotamilla, P. reniformis)

Potamilla neglecta

Potamilla occelata (Pseudopotamilla)

Sabella crassicornis

Schizobranchia insignis

## Key to pelagic sabellidae larvae (Fig. 28)

Ia. 3 setigers at metatrochophore stag	ge; otocysts absent; telotroch
absent or reduced	Demonax media
1b. 3-4 setigers; otocysts and telotrock	n present
	Chone infundibuliformis

**Family Serpulidae** (Local Species 7, Local Species with Described Larvae 1). Adults of this family build calcareous tubes (Fig. 29).

Apomatus geniculatus

Apomatus timsi

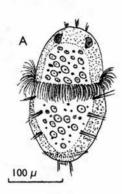
Crucigera irregularis

Crucigera zygophora

Fig. 28. Larvae

(A) Demonax media and
(B) Chone infundibuliformis, family Sabellidae.

(From Okuda, 1946;
McEuen et al., 1983)



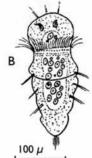
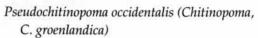
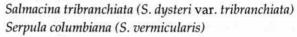




Fig. 29. (A) Late trochophore, (B) metatrochophore, and (C) late metatrochrophore larvae of Salmancia tribranchiata, family Serpulidae. (From Rullier, 1960)







Family Spirorbidae (Local Species 11, Local Species with Described Larvae 2). All spirorbid genera found in Pacific Northwest waters brood their larvae, either within the adult tube or externally (Fauchald, 1977). Larvae have a characteristic complex of gland cells in the ventral and lateral ectoderm of the thorax.

Circeis amoricana (C. spirillum, Spirorbis rugatus)
Circeis spirillum (Dexiospira, Spirorbis)
Janura rugata (Spirorbis, Dexiospira)
Paradexiospira violacea
Paradexiospira vitrea (Laeospira, Eulaeospira, Spirorbis, S. variabilis, S. semidentatus, S. racemosus)
Pileolaria langerhansi
Pileolaria potswaldi (Laeospira, Spirorbis, S. moerchi)
Pileolaria quadrangularis
Protolaeospira eximia (Spirorbis, S. ambilateralis)
Sinistrella media (Spirorbis, Laeospira, Romanchella)
Spirorbis bifurcatus

## Key to pelagic spirorbid larvae (Fig. 30)

## Order Spionida

**Family Acrocirridae** (Local Species 1, Local Species with Described Larvae 0). Little is known about the larval development in this family. It may be similar to that in the Cirratulidae and Flabelligeridae (Banse, 1969).

Acrocirrus heterochaetus

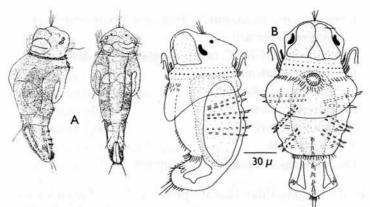


Fig. 30. Larvae of species in the family Spirorbidae. (A) Circeis spirillum, lateral and dorsal views, (B) Pileolaria potswaldi, lateral and ventral views. (From Okuda, 1946; Potswald, 1978)

Family Apistobranchidae (Local Species 1, Local Species with Described Larvae 0). Development in this family has not been described. Adults are tube dwellers but can exist outside tubes in loosely constructed burrows. They are common in shelly sands and muds (Fauchald, 1977).

Apistobranchus ornatus

**Family Chaetopteridae** (Local Species 4, Local Species with Described Larvae 2). Chaetopterid larvae are separated by comparison of the modified setae on the fourth anterior segment.

Chaetopterus variopedatus Mesochaetopterus taylori Phyllochaetopterus prolifica Spiochaetopterus costarum (Telepsavus)

#### Key to chaetopterid genera (Fig. 31)

la. I pair of setae on setiger 4	2
Ib. Several setae on setiger 4	3
2a. Modified setae with enlarged heart-shape	
	100
2b. Setae not enlarged distally	Phyllochaetopterus
3a. Distal part of setae blade-shaped	Mesochaetopterus
3b. Distal part of setae with lateral tip	Chaetopterus

Family Cirratulidae (Local Species 11, Local Species with Described Larvae 1). Development in this family includes many species that produce lecithotrophic larvae (\*) that hatch from eggs laid in a gel mass as well, as species that are direct developers (†). Christie (1985) notes that all larvae of this family are large and yolky (Fig. 32).

Caulleriella alata Caulleriella hamata (Tharyx) Chaetozone setosa\*

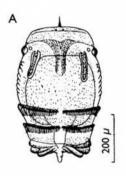




Fig. 31. (A) Chaetopterus variopedatus and (B) Spiochaetopterus costarum, family Chaetopteridae. (From Bhaud and Cazaux, 1987)

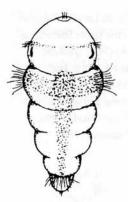


Fig. 32. Larva of Dodecaceria fewkesi, family Cirratulidae. (From Berkeley and Berkeley, 1954)

Cirratulus cirratus cirratust (Chaetozone berkeleyorum, Caulleriella gracilis)

Cirratulus spectabilis (C. cirratus cingulatus, C. robustus)

Cirriformia spirabranchia

Dodecaceria choncharum\*

Dodecaceria fewkesi

Dodecaceria fistulicola

Tharyx multifilis

Tharyx parvus (T. multifilus var. parvus)

Family Magelonidae (Local Species 1, Local Species with Described Larvae 0). An unknown magelonid, possibly *Magelona cerae*, is often found in plankton samples taken from Coos Bay, Oregon (K. Johnson, pers. comm.). Adults are common in sandy substrates and move through the sediment with a shovel-like prostomium and thread-like body. Larvae of this family have characteristically long larval tentacles that are flexible and often coiled. *Magelona alleni* illustrates characteristics of the family (Fig. 33), but it is not a species known to be in local waters.

Magelona sp.

**Family Spionidae** (Local Species 27, Local Species with Described Larvae 18). The spionids are a large and diverse group of polychaetes. Adult habitat and development modes vary across the family. A large number of larvae of local species have been described.

Boccardia columbiana (Polydora) Boccardia polybranchia Boccardia proboscidea (B. californica) Boccardiella hamata (Boccardia, B. uncata, Polydora) Laonice cirrata (Nerine) Paraprionospio pinnata (Prionospio, P. ornata)

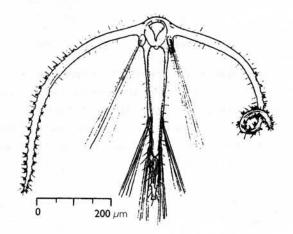
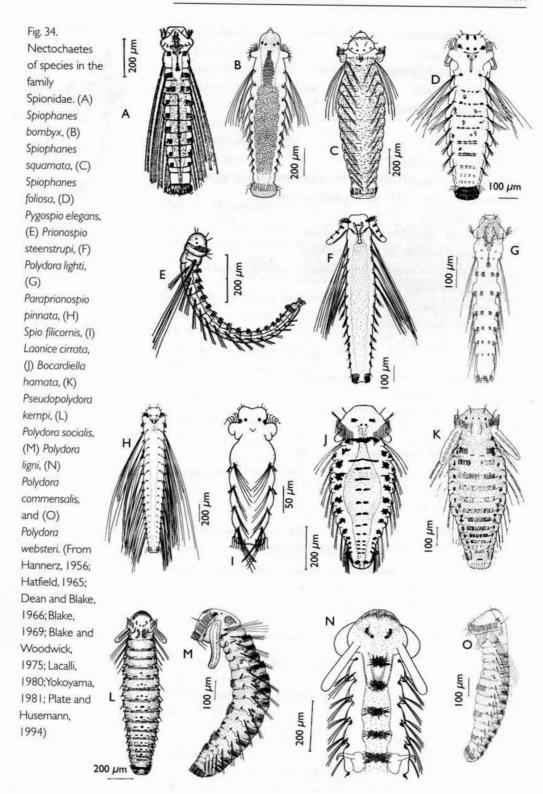


Fig. 33. Late metatrochophore of *Magelona alleni,* family Magelonidae. (From Wilson, 1982)

Polydora alloporis Polydora armata Polydora brachycephala Polydora cardalia Polydora commensalis Polydora giardi Polydora ligni Polydora pygidialis (P. ciliata) Polydora socialis (P. caeca, P. caeca var. mangna, P. plena) Polydora spongicola (P. ciliata var. spongicola) Polydora websteri Prionospio lighti (Minuspio cirrifera) Prionospio steenstrupi (P. malmgreni) Pseudopolydora kempi (Neopygospio laminifera, Polydora, Polydora kempi japonica) Pygospio elegans Scolelepis foliosa (Nerine) Scolelepis squamata (S. acuta) Spio filicornis Spiophanes berkeleyorum (S. cirrata) Spiophanes bombyx Streblospio benedicti

## Key to spionid nectochaetes (Fig. 34)

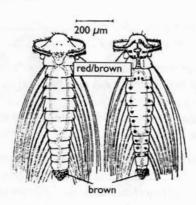
I a. Branchiae absent; setiger I with I-2 large curved neuropodial spines in addition to normal capillaries Spiophanes bombyx I b. Branchiae present; setiger I without specialized setae
2a. Setiger 5 modified, with specialized setae
3a. Prostomium distally pointed (may appear conical with rounded apex in extremely contracted specimens), with or without subdistal lateral horns
4a. Prostomium bell-shaped, orange-pigmented; gut blue-green
5a. Branchiae limited to middle and posterior setigers except for single pair on setiger 2 in males
6a. Branchiae concentrated in anterior setigers 1—2, absent posteriorly



7a. Branchiae beginning on setiger I9 7b. Branchiae beginning on setiger 28
8a. Peristomial palps short; I pair of anal cirri in early larvae, 2 pairs of anal cirri present in later larvae; eyes dark red Prionospio steenstrupi
8b. Persitomial palps differentiate only in late larval stages; I pair of anal cirri in late larvae
9a. I pair of cirriform branchiae; dorsal collar across setiger 2
Paraprionospio pinnata  10a. Branchiae beginning on setiger 1; hooded hooks only in neuropodia
I I a. Branchiae beginning on setiger 2
Najor spines of setiger 5 of one type: simple, falcate, with smaller companion setae
I 3a. Setiger 5 slightly to moderately modified, usually with prominent parapodia; major spines of two types: first simple, acicular or falcate, second pennoned with both types usually arranged in U- or J-shaped row. Hooded hooks with secondary tooth placed close to main fang
14a. Larvae with distinct ventral chromatophores (black, yellow-green, or iridescent)       15         14b. Larvae without ventral pigment, or if present not in distinct chromatophores       16
I 5a. Ventral pigment black
I 6a. Single dorsal row of melanophores present, at least posterior to segment 3
17a. Prostomium with yellow-brown pigment; gastrotrochs on segments 7, 10, 13, 15

<sup>a</sup>Levin (1991) illustrates lecithotrophic pelagic larvae that lack swimming setae. <sup>b</sup>Development of this species is described in Woodwick (1977).

Fig. 35. Representative trochochaetid larva, Disoma multisetosum. (From Hannerz, 1961)



Family Trochochaetidae (Local Species 1, Local Species with Described Larvae 0). This is a small family (nine species) of non-tubiculous worms that live in soft substrates in mainly shallow water. Adults are cylindrical with a slightly flattened body (Fauchald, 1977). An image of *Disoma multisetosum* is included for reference (Fig. 35).

Trochochaeta multisetosa (T. franciscanum)

## Order Sternaspida

Family Sternaspidae (Local Species 1, Local Species with Described Larvae 0). This small family has approximately ten recognized species. Adults are common in sand and mud at all depths, but usually at 100–200 m. Adults are rarely found in large numbers, are burrowers, and have dark yellow or reddish chitinized shields (Fauchald, 1977).

Sternaspis scutata (S. fossor, S. affinis)

#### Order Terebellidae

Family Ampharetidae (Local Species 14, Local Species with Described Larvae 0). Thorson (1946) suggests that several species (\*) in this family are direct developers. Nyholm (1950), however, describes *Melinna cristata* as a free-spawned lecithotrophic larva. This is a fairly large, deep-water family (Fauchald, 1977).

Amage anops
Ampharete acutifrons (A. grubei)\*
Ampharete finmarchica (A. arctica)\*
Ampharete goesi goesi
Amphicteis mucronata
Amphicteis scaphobranchiata
Amphisamytha bioculata
Anobothrus gracilis (Ampharete gagarae)\*
Asabellides lineata (Pseudosabellides)

Asabellides sibrica Hobsonia florida (Amphicteis gunneri floridus)\* Lysippe annectens Melinna cristata (M. denticulata) Schistocomus hiltoni

Family Pectinariidae (Local Species 3, Local Species with Described Larvae 3). Trochophores of this family are easily confused with nephtyid larvae (Lacalli, 1980). Pectinariids construct and live in fragile tusk-shaped tubes (Fauchald, 1977).

Amphictene moorei (A. auricoma, Pectinaria) Cistenides granulata (C. brevicoma, Pectinaria) Pectinaria californiensis (P. belgica)

#### Key to pectinariid metatrochophores (Fig. 36)

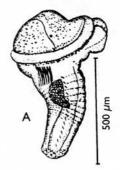
Family Sabellariidae (Local Species 4, Local Species with Described Larvae 0). Larval morphology is quite similar among sabellariid species (Ecklebarger, 1975, 1977). Opercular characteristics are useful for distinguishing genera according to Bhaud and Cazaux (1987). The key below is modified from Bhaud and Cazaux (1987) for the two local genera. *Sabellaria alveola* (Fig. 37) is not a local species.

Idanthyrsus armatus Idanthyrsus ornamentatus Sabellaria cementarium Sabellaria gracilis

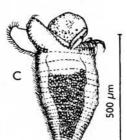
#### Key to sabellariid genera

**Family Terebellidae** (Local Species 20, Local Species with Described Larvae 2). Terebellids are common shallow-water polychaetes and are found in a diversity of environments. They inhabit permanent tubes from which they extend a crown of elongated extensile tentacles that they use to capture small food

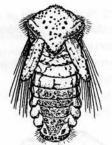
Fig. 36. (A) Pectinaria californiesis, (B) Cistenides granulata, and (C) Amphictene moorei, family Pectinariidae. (From Thorson, 1946; Korn, 1960; Lacalli, 1980)







200 μm



width 8  $\mu$ m; length 600  $\mu$ m

Fig. 37. Larvae of Sabellaria alveola, family Sabellaridae; this not a local species. (From Bhaud and Cazaux, 1987)



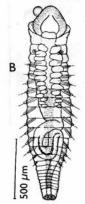


Fig. 38. (A) Lanice conchilega and (B) Amphitrite cirrata, outside tube, family Terebellidae. (From Thorson, 1946)

particles from the surface of the substrate or water (Fauchald, 1977).

Amphitrite cirrata (A. palmata)

Artacama coniferi

Eupolymnia heterobranchia (E. crescentis)

Lanice chonchilega

Loimia medusa

Neoamphitrite robusta (Amphitrite, A. scionides dux)

Neoleprea spiralis

Pista brevibranchiata (P. fasciata)

Pista cristata

Pista elongata

Pista fasciata

Pista fimbriata (P. brevibranchiata)

Pista moorei

Pista pacifica

Polycirrus californicus (P. caliendrum)

Scionella japonica

Thelepus cincinnatus (T. hamatus)

Thelepus crispus

Thelepus hamatus

Thelepus setosus

### Key to terebellidae larvae (Fig. 38)

**Family Trichobranchidae** (Local Species 3, Local Species with Described Larvae 0). Developmental patterns for *Artacamella* and *Trichobranchus* are not known. *Terebellides stroemi* develops directly in egg masses (Thorson, 1946).

Artacamella hancocki Terebellides stroemi Trichobranchus glacialis

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