

# Biodiversity and adaptations of CYP enzymes in the Amazon Loricariidae fishes

PI\_BR: Dr. Thiago E. Parente – FIOCRUZ, Rio de Janeiro, Brazil

PI\_US: Dr. Mark E. Hahn – WHOI, Woods Hole, MA, USA

[parente@ensp.fiocruz.br](mailto:parente@ensp.fiocruz.br) & [mhahn@whoi.edu](mailto:mhahn@whoi.edu)

## STAFF

MSc. Daniel A. Moreira  
Paula C.C. de Andrade (undergrad)  
Maithê G.P. Magalhães (undergrad)

## Colaborators

Dr. John Stegeman – WHOI, USA  
Dr. Francisco Paumgartten – FIOCRUZ, Brazil  
Dr. Paulo Buckup – MN, UFRJ, Brazil  
Dra. Carolina Furtado – INCA, Brazil  
Dr. Adalberto Val – INPA, Brazil  
Dr. Brian Haas – Broad Institute, USA

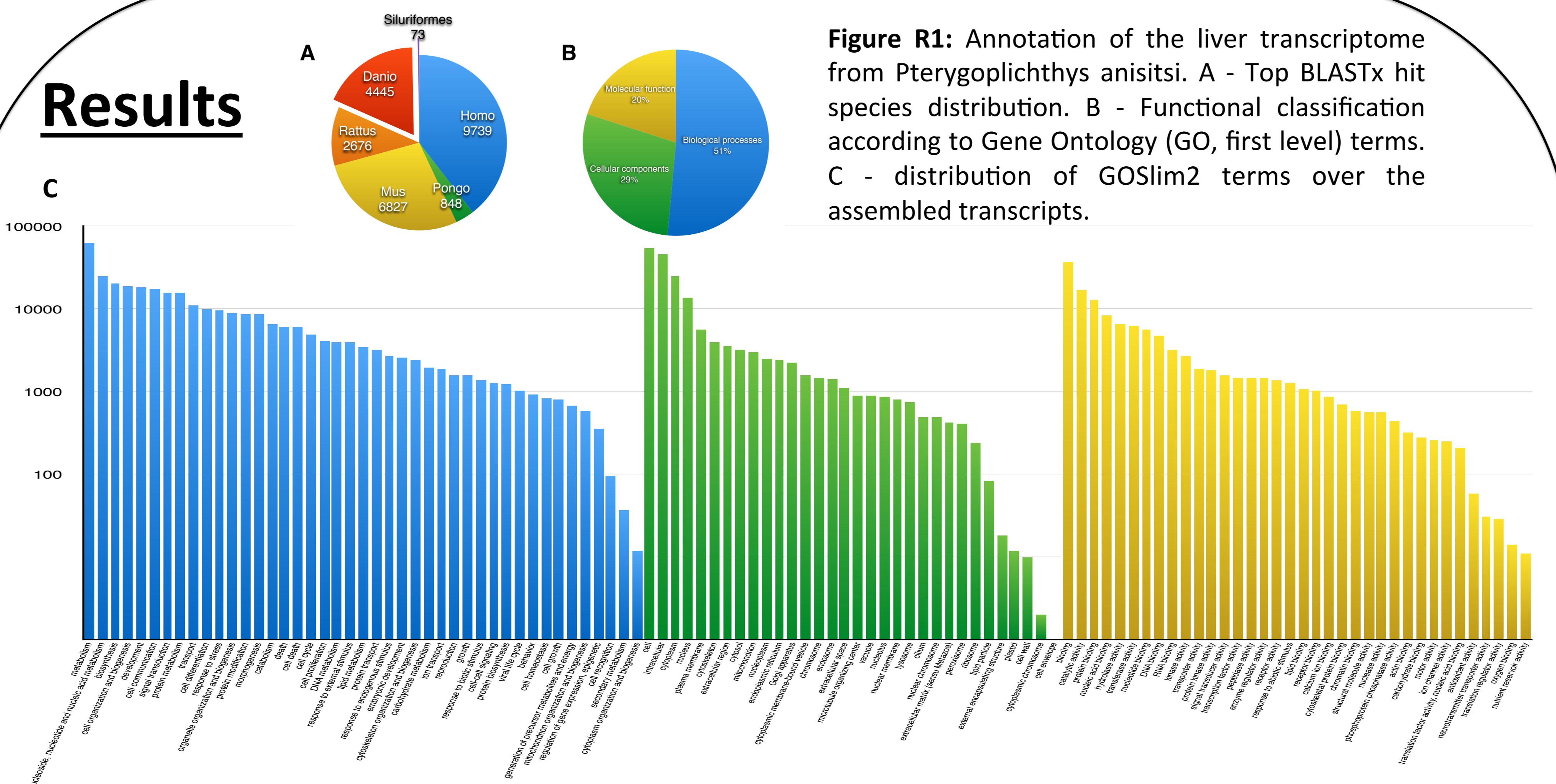


## Abstract

Loricariidae is the most diverse family of the fish order Siluriformes<sup>1</sup>. Currently, there are >700 valid species and this number is still growing due to discovery of new species and the resolution of cryptic ones<sup>2,3</sup>. Loricariids inhabit a wide spectrum of fresh water environments, have diverse ecological habitats and show high endemism rate<sup>4</sup>. This astonishing taxonomic and ecological diversity must rely on genetic, molecular and biochemical adaptations. In fact, our group has reported peculiar adaptations in loricariids, which might increase their susceptibility to the toxic effects of pollutants<sup>5,6,7</sup>. Here, the molecular biodiversity of Loricariidae fish is explored using Next-Generation Sequencing (NGS) technologies. In total, 101 species have been collected; 34 of these will have their liver transcriptome sequenced. For one species, *Hypancistrus zebra*, the transcriptome of brain, heart, kidney, gill, intestine and gonad will also be sequenced. The transcriptome of three species has been sequenced; *Pterygoplichthys anisitsi*, *Ancistrus* spp. and *Corydoras nattereri*. The results for *P. anisitsi* are shown below. Briefly, 34204 genes with homologs in zebrafish were sequenced; 189 of these contained the complete coding sequence of proteins involved in mechanisms of defense against toxins. Great expansions were detected in subfamilies of Cytochromes P450 and Sulfotransferases.

References:  
1: Covain & Fisch-Muller, 2007. Zootaxa 1462: 1-40  
2: Ferraris, 2007. Zootaxa 1435: 1-626  
3: Parente et al., 2013. Zootaxa 3641(4): 343-370  
4: Covain & Fisch-Muller, 2007. Zootaxa 1462: 1-40  
5: Parente et al., 2009. Comp. Bioch. Physiol. C 150: 252-260  
6: Parente et al., 2011. Genet. Appl. 43: 111-118  
7: Parente et al., 2014. Aquatic Toxicology 134: 193-199

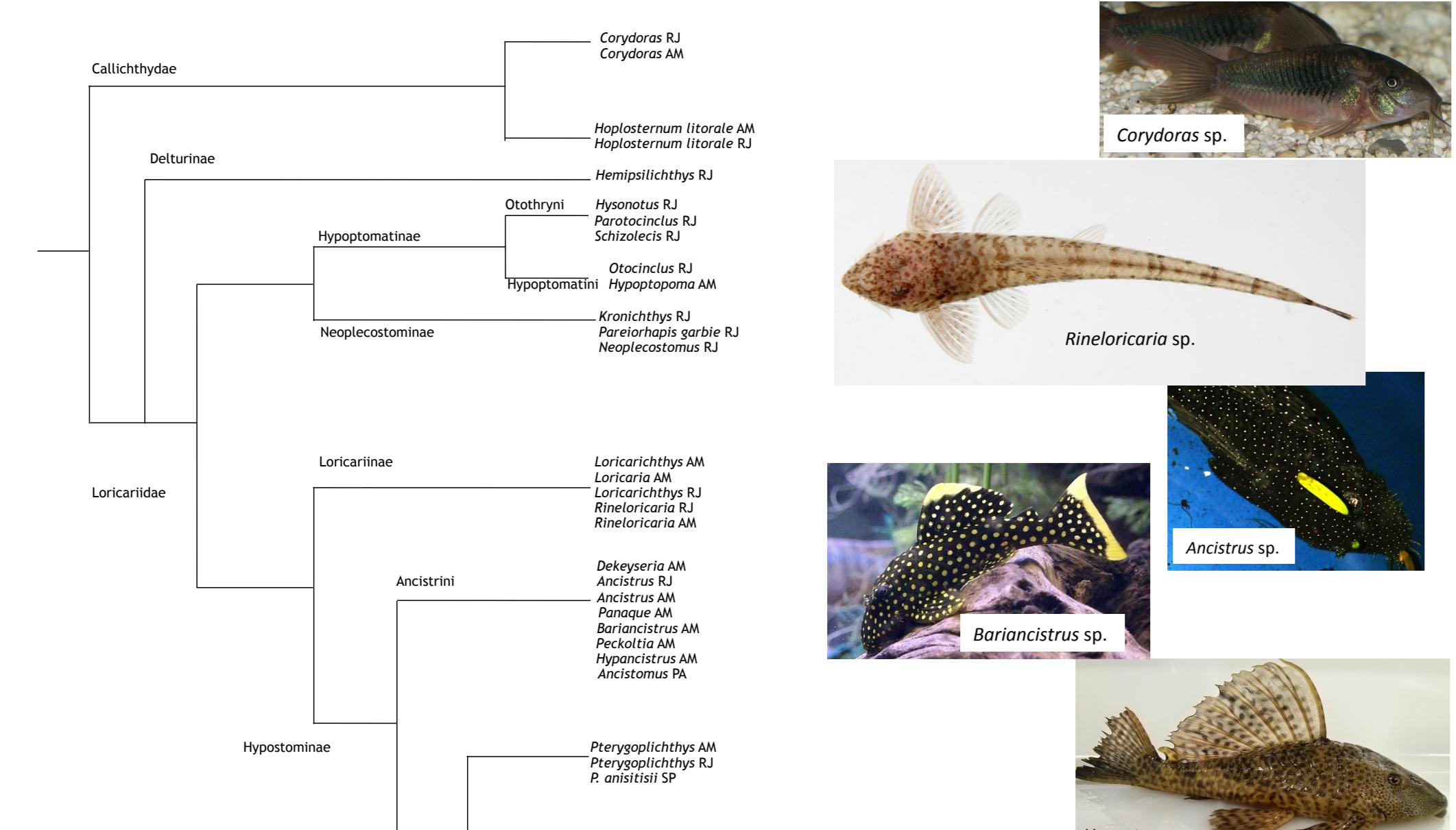
## Results



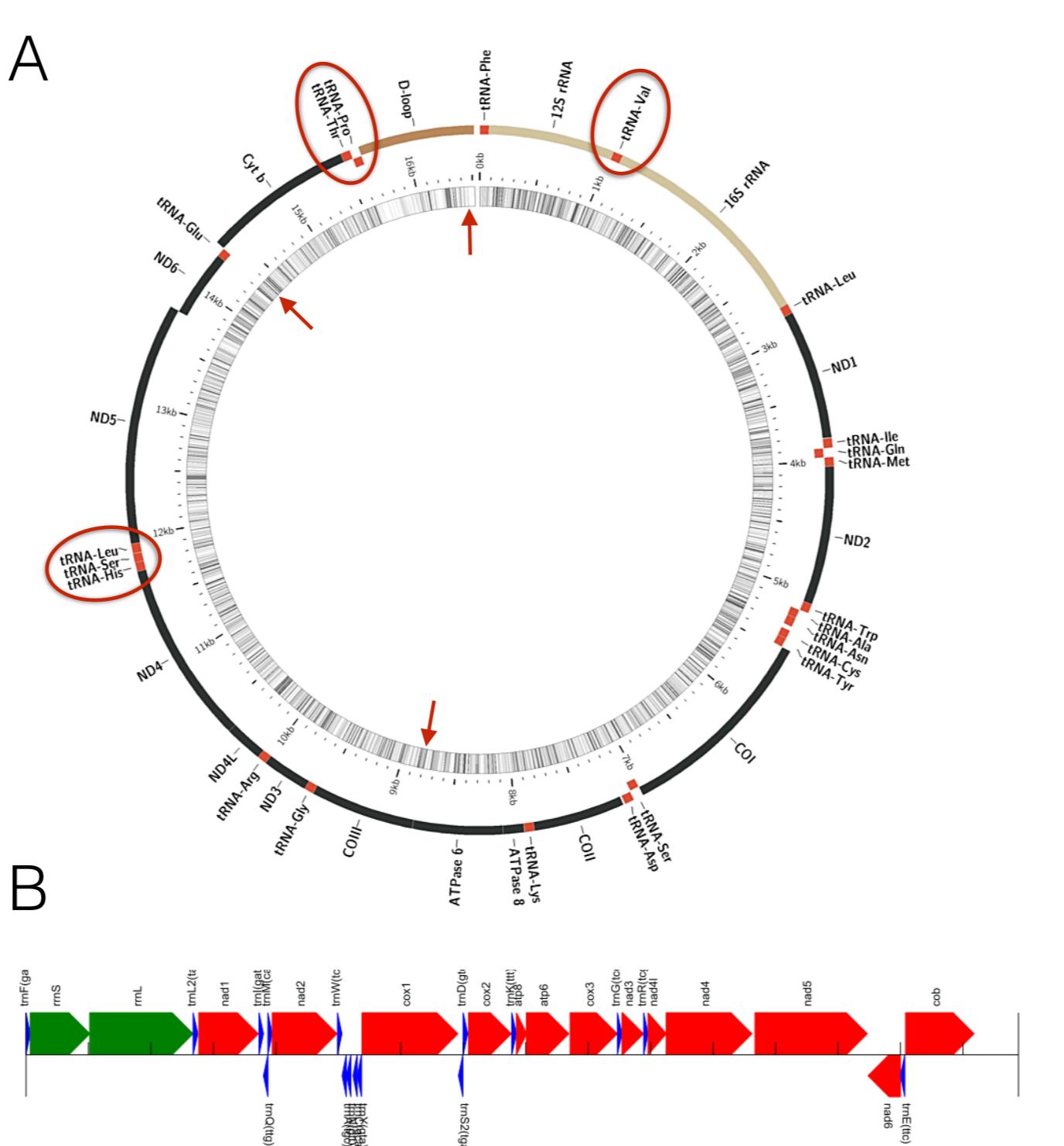
**Figure R1:** Annotation of the liver transcriptome from *Pterygoplichthys anisitsi*. A - Top BLASTx hit species distribution. B - Functional classification according to Gene Ontology (GO, first level) terms. C - distribution of GO Slim2 terms over the assembled transcripts.

## Material & Methods

### Fish sampling (FIOCRUZ, INPA and MN UFRJ)



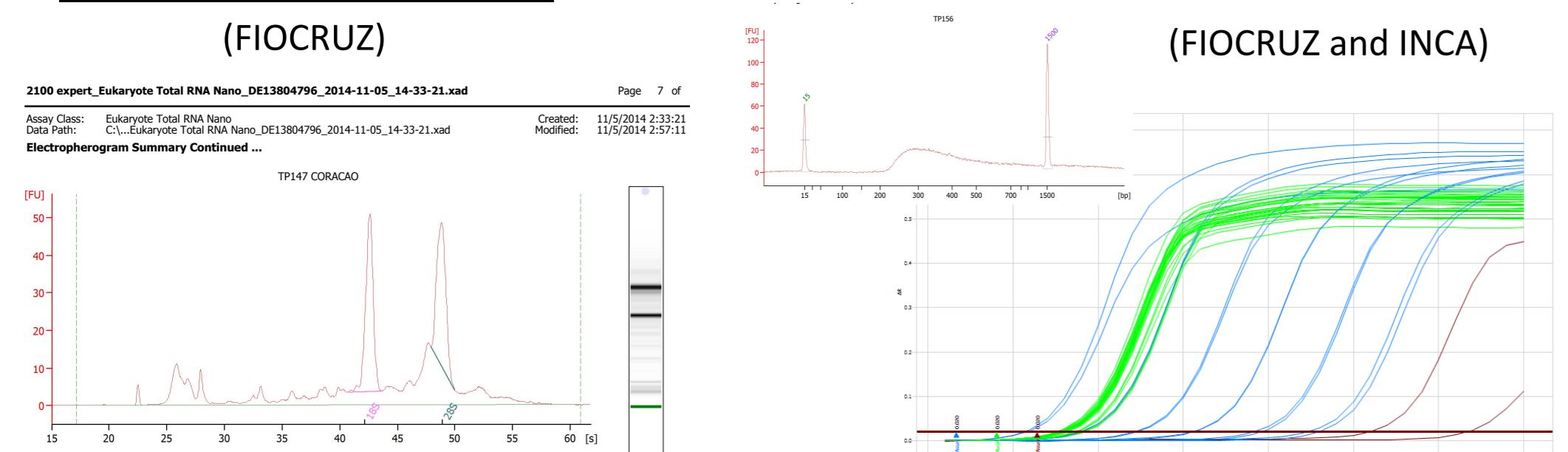
**Figure R2:** Mitochondrial genome of *Pterygoplichthys* spp.. A - Circular representation of *P. disjunctivus* mtDNA (G1: 339506171). Red circles indicate the tRNA not sequenced in *P. anisitsi* mtDNA and red arrows the approximate region of the three smallest gaps of 10, 24 and 31 nucleotides. B - Linear representation of the annotated mitochondrial genome of *P. anisitsi*.



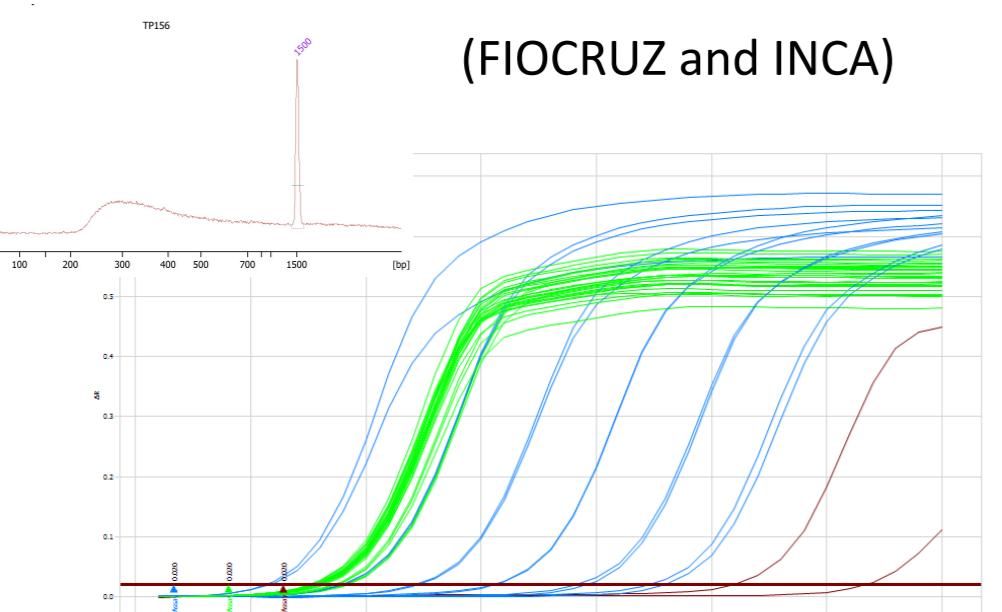
**Table R1:** Number of sequenced components in the hepatic transcriptome of *P. anisitsi* with complete coding sequence (CDS), >75% of the CDS, >50% of the CDS and the total number of contigs for each defensome gene family.

	Full length CDS	>75% CDS coverage	>50% CDS coverage	Contigs
AHR & ARNT	3	3	3	9
Aldo Keto Reductase	5	5	5	5
ATP Binding Cassette (ABC)	13	19	25	91
Basic leucine zipper	3	3	5	8
Catalase	1	1	1	1
Cytochrome P450	34	35	39	82
Epoxye hydroxilase	2	2	2	2
Glucuronosyltransferase	8	8	10	15
Glutathione Peroxidase	6	6	8	11
Glutathione-S-transferase	8	10	10	10
n-acetyl-transferases	10	11	11	14
Nuclear receptor	23	32	33	54
Sulfotransferases (SULT)	47	49	49	53
Superoxide desmutase	3	3	3	3
Thioredoxins	23	25	25	27
Total	189	212	229	385

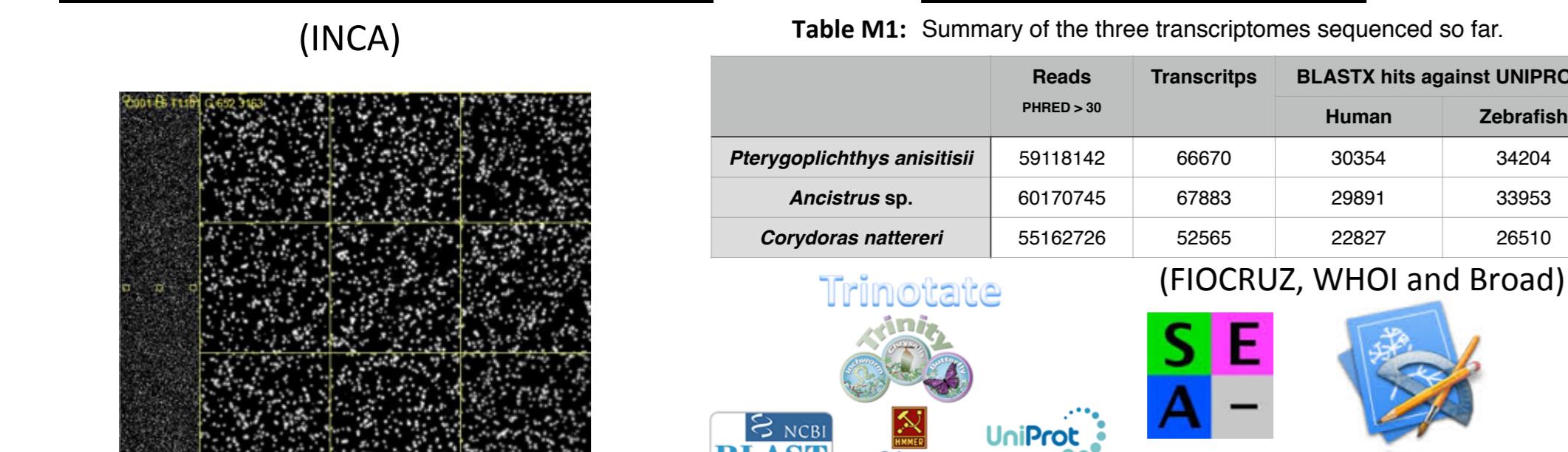
### RNA extraction (FIOCRUZ)



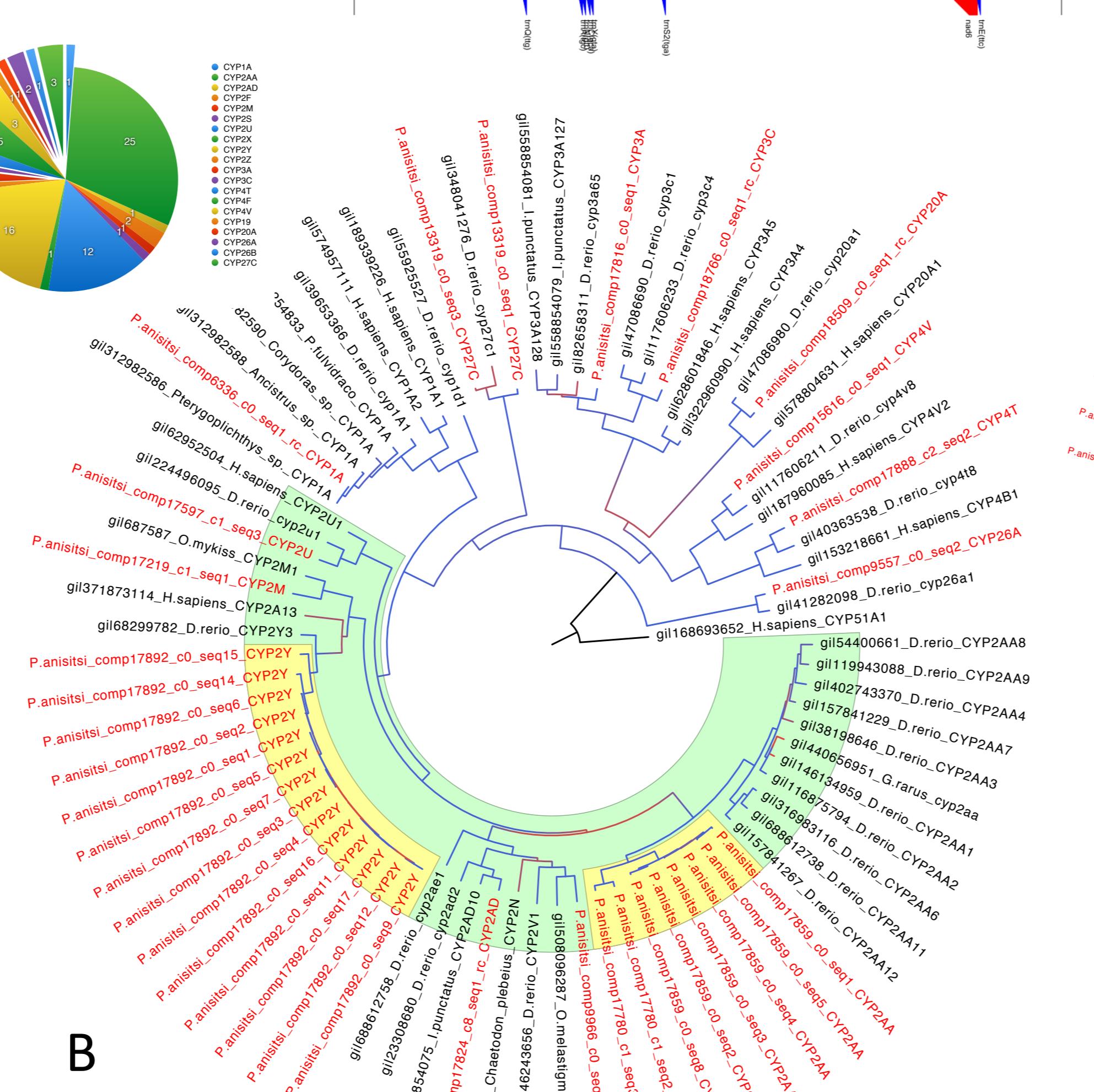
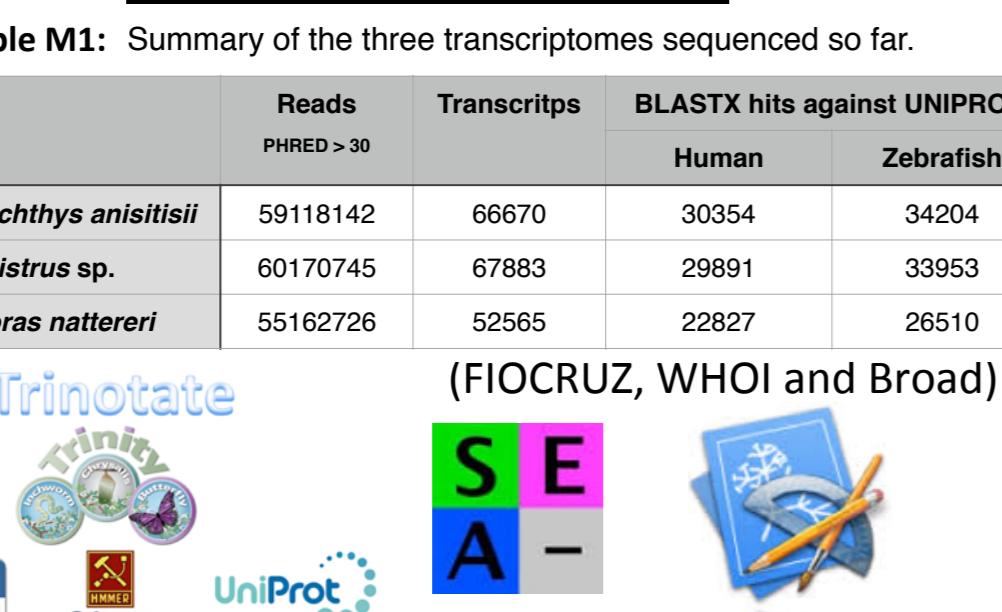
### Library preparation (FIOCRUZ and INCA)



### Illumina sequencing (INCA)



### Bioinformatic



**Figure R3:** Distribution of cytochromes P450 (CYP) into subfamilies (A); and the maximum likelihood phylogenetic tree of the sequenced CYP (B) and Sulfotransferases (SULT) genes (C).

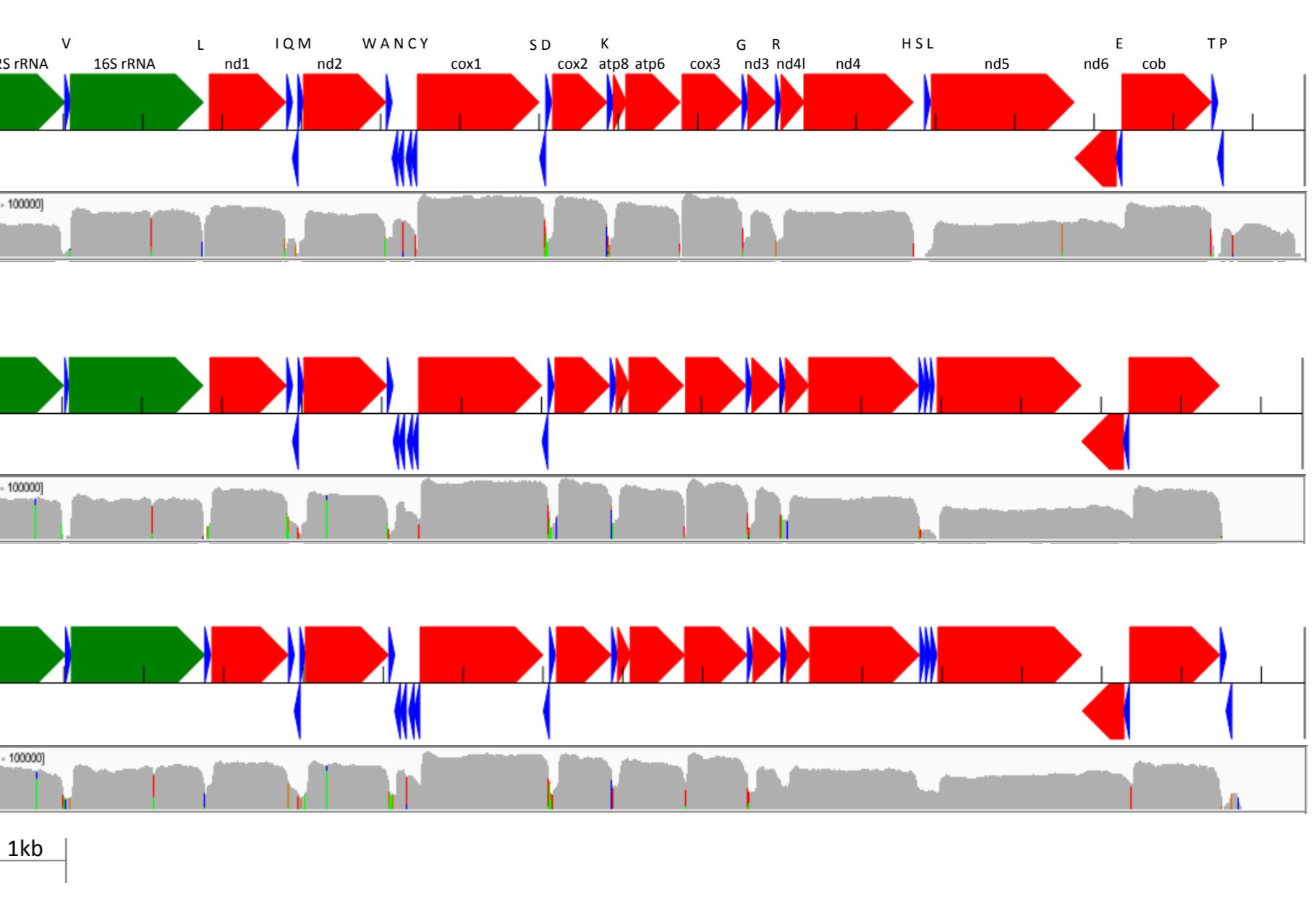
## Tangential project #1

### Mitogenomes assembled from transcriptome

We sequenced the transcriptome of three fish from the genus *Ancistrus* (Loricariidae, Siluriformes) using as start material total RNA isolated from the liver. The transcriptome data were used to assemble the mitogenome of each fish with 92%, 95% and 99% of the full length of their closest related species with a sequenced mitogenome. Taken the sequences of the three fish together, all the 13 protein-coding genes, two ribosomal RNAs, 22 tRNAs and the D-loop known in the mitogenomes of vertebrates were sequenced. The use of transcriptomic data also allowed the clear observation of the punctuation pattern of mtRNA editing, to analyze the transcriptional profile of mtRNA, and to detect heteroplasmic sites.

**Table TP1:** Summary of the transcriptome and mitogenome data for the three fish (*Ancistrus* spp.).

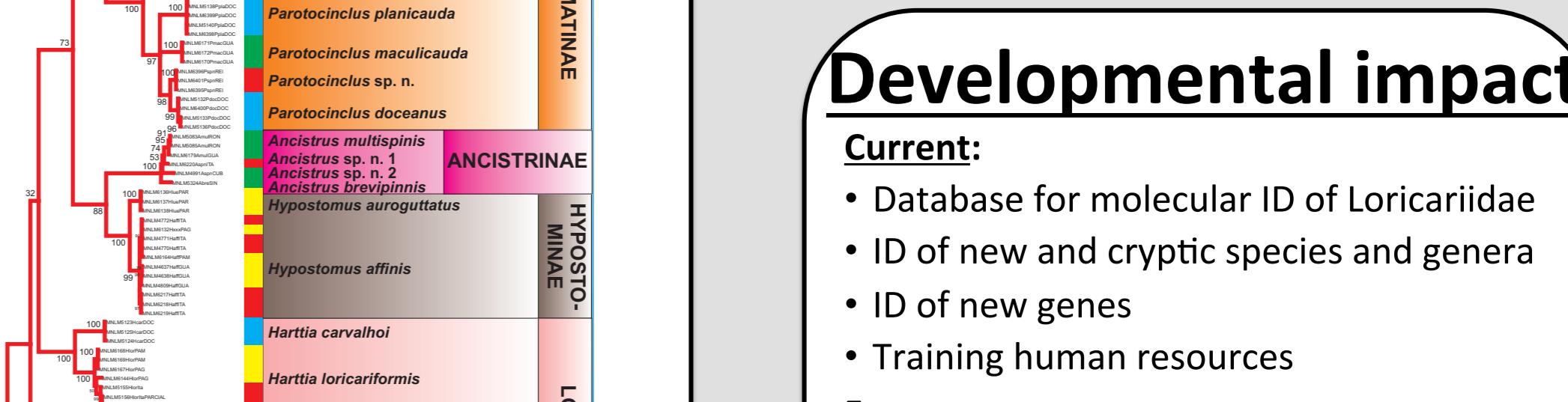
<i>Ancistrus</i> sp. #1	<i>Ancistrus</i> sp. #2a	<i>Ancistrus</i> sp. #2b
RIN	8.9	>7.00
Library size (bp)	230-800	285-370
Reads after QC	43502597	53961751
Transcripts	67988	67883
with BLASTx hit for mitogenome	35710	31886
mtRNA reads (%)	13	12
Mitogenome coverage (%)	99.2	92.5
Heteroplasmic sites	44	46



## Tangential project #2

### Barcode loricariids

- Sequence the transcriptome of 31 species
- Assemble the 31 transcriptomes
- Analyze CYPs and AHR genes in the 31 species
- Amplify and sequence CYP1A and AHR in the others sampled species
- Exposure of selected species to chemicals
- Analyze alterations in gene transcription



## Publications

- Paper published:**  
• Parente et al., 2015. Vigilância Sanitária em Debate 3(1): 88-93.  
• Felicio; Parente et al., 2015. Ecotoxicol. Environ. Safety 115: 26-32.  
• Parente et al., 2014. Aquatic Toxicology 134: 193-199.  
**Chapter published:**  
• Parente & Hahn, 2013: in: Pollution and fish health in tropical rivers. CCR Press.  
**Abstracts and posters in conferences:**  
• Parente et al., 2014. P450 Biodiversity and Biotechnology, Japan.  
• Magalhães; Andrade; Parente; Buckup, 2014; Congresso Brasileiro de Genética.  
• Buckup et al., 2015; Encontro Brasileiro de Ictiologia.  
**Manuscripts in preparation:**  
• Parente et al., The transcriptome and diversity of defensome genes in *P. anisitsi*.  
• Moreira, Furtado & Parente; Constructing mitogenomes from transcriptome.  
• Moreira et al.; The mitogenome of *Corydoras nattereri*.  
• Buckup et al.; Biogeography of Loricariidae from Rio de Janeiro state, Brazil.

