

Human Allergy Products and Services

Effective April 2014



Contents

| | |
|----|---|
| 3 | Ordering Information |
| 4 | Standardized Extracts |
| 5 | Pollen |
| 12 | Fungi and Smuts |
| 15 | Epithelia, Miscellaneous Inhalants, and Insects |
| 17 | Ingestant Extracts |
| 21 | Made-to-Order Items |
| 21 | Premium Allergens |
| 22 | Sterile Diluents |
| 25 | Sterile Empty Vials |
| 26 | Plastic Colored Caps |
| 26 | Dropper Vials |
| 26 | Safety Syringes |
| 27 | Syringes and Syringe Trays |
| 28 | Ancillary Products |
| 30 | Vial Racks |
| 31 | Skin Testing Devices |
| 34 | GREER® Service Team |
| 35 | Important Safety Information |
| 35 | Package Inserts |

Ordering Information

To set up a new account with GREER®, please contact a Customer Care Specialist.
800.378.3906 or 828.754.5327

Ordering

For the best possible service, specify account number, item number, product name, vial size and dilution, aqueous or glycerinated formulation, and scratch or multiple dose vial, as shown in the catalog.

Certain extracts and mixtures cannot be supplied in the 1:10 w/v strength due to excessive precipitation. These will be supplied at the strongest possible w/v strength. Glycerinated extracts can be supplied in 1:20 w/v or weaker dilutions. Extracts ordered in protein nitrogen units (PNU) are available in aqueous formulations only.

Physician license must be on file. Proper identification will be required from institutions.

Minimum Order

A minimum order of \$25.00 is required. Please combine orders to achieve minimum.

Made-to-Order Items

Specific non-stocked products have been identified as made-to-order items based on low utilization. Delivery will be a minimum of 4 to 6 weeks. Please consult your Customer Care Specialist for more information.

Premium Allergens

Specific products have been designated as premium allergens due to low utilization, decreased availability of raw materials, increased costs associated with the collection/extraction process, and/or market conditions. These items carry a surcharge. Please consult your Customer Care Specialist for more information.

Shipping

All extracts are shipped overnight for next-day delivery. All other products ship via UPS 3 Day Select® or Ground unless otherwise specified. Shipments with ice packs are available upon customer request, with an additional charge.

Terms

Net 30 days from date of invoice. FOB Lenoir, NC. Shipping and insurance charges are prepaid and added to the invoice. All prices are in US dollars. All prices are subject to change without notice. We accept payment via MasterCard®, Visa®, and American Express®.

Returns

Please contact a GREER® Customer Care Specialist at 800.378.3906 for *prior approval* of all product returns. Any shipping error must be reported within 7 days of receipt of the order. A Return Goods Authorization Number must accompany all product returns to GREER within 10 days of the return notification.

Allergenic extracts and sterile products cannot be returned unless due to a GREER error. Ancillary supplies must be returned in unopened, undamaged cases. Credit is not issued for duplications resulting from customer errors in ordering.

Warranty

We warrant only that solutions in unopened and undamaged vials are sterile and correctly labeled. Once shipment is made, we have no control over storage conditions, methods of usage, or dosage administered.

Telephone and Fax Orders

To order by telephone, call our Customer Care Specialists: **800.378.3906**

To order by fax, dial: **800.419.7302** or **828.754.5320**

For customers outside the United States, call: **828.754.5327**

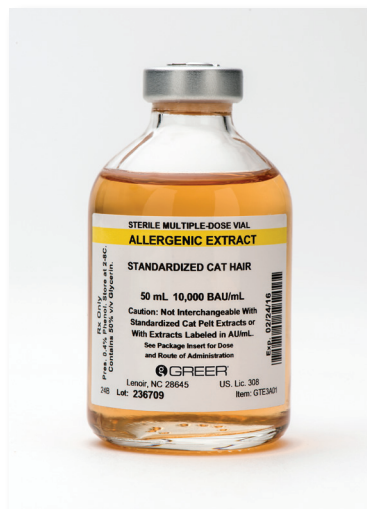
To email, write to: **customer care@greerlabs.com**

Our Customer Care Specialists are available Monday through Thursday 8 AM to 7 PM and Friday 8 AM to 5 PM Eastern Time.

For information on custom mixtures, please contact a GREER® Customer Care Specialist at 800.378.3906.

Standardized Extracts

All standardized products are available in 50% glycerin only.



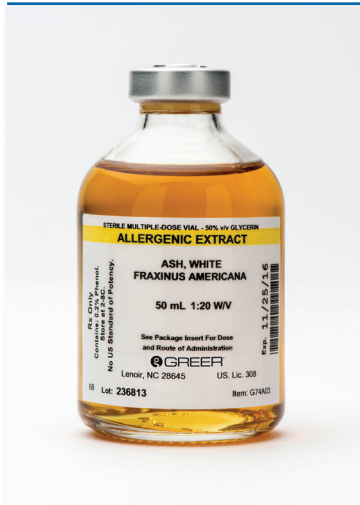
| Item # | Product Name | Testing 5mL Scratch | Treatment Glycerin |
|---|--|---------------------|--------------------|
| Standardized Cat Hair | | | |
| TE3 | Standardized Cat Hair <i>Felis catus (domesticus)</i> | ● | ● |
| Standardized Mites | | | |
| B64 | <i>Dermatophagoides farinae</i> | ● | ● |
| B70 | <i>Dermatophagoides pteronyssinus</i> | ● | ● |
| Standardized Mite Mix | | | |
| B03 | Mite Mix, Standardized Equal parts of: B64 <i>D. farinae</i> , B70 <i>D. pteronyssinus</i> | ● | ● |
| Standardized Grass Pollen | | | |
| T2 | Bermuda <i>Cynodon dactylon</i> | ● | ● |
| T16 | Kentucky Blue/June <i>Poa pratensis</i> | ● | ● |
| T14 | Meadow Fescue <i>Festuca elatior (pratensis)</i> | ● | ● |
| T20 | Orchard <i>Dactylis glomerata</i> | ● | ● |
| T22 | Red Top <i>Agrostis (alba) gigantea</i> | ● | ● |
| T24 | Ryegrass, Perennial <i>Lolium perenne</i> | ● | ● |
| T27 | Sweet Vernal* <i>Anthoxanthum odoratum</i> | ● | ● |
| T28 | Timothy <i>Phleum pratense</i> | ● | ● |
| Mixes Containing Standardized Grass Pollen | | | |
| TP20 | Timothy/Orchard Grass Mix 28 Timothy, 20 Orchard Grass | ● | ● |
| TP23 | K-O-T Grass Mix 16 Kentucky Blue/June, 20 Orchard, 28 Timothy | ● | ● |
| TP24 | K-O-R-T Grass Mix 16 Kentucky Blue/June, 20 Orchard, 22 Redtop, 28 Timothy | ● | ● |
| TP25 | K-O-R-T and Sweet Vernal Mix 16 Kentucky Blue/June, 20 Orchard, 22 Redtop, 28 Timothy, 27 Sweet Vernal | ● | ● |
| TP29 | T-O-S Grass Mix 28 Timothy, 20 Orchard, 27 Sweet Vernal | ● | ● |
| TP27 | 7 Grass Mix 16 Kentucky Blue/June, 14 Meadow Fescue, 20 Orchard, 24 Perennial Ryegrass, 22 Redtop, 27 Sweet Vernal, 28 Timothy | ● | ● |
| P19 | 9 Southern Grass Mix (Concentrate) Equal parts of: 2 Bermuda at 10,000 BAU/mL, P27 7 Grass Mix at 100,000 BAU/mL, 15 Johnson at 1:20 w/v | ● | ● |

Please see inside back cover for important safety information about GREER® Extracts™. In addition, please see accompanying package inserts for full prescribing information, including BOXED WARNING, in the pocket inside the back cover or go to www.greerlabs.com.

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- — Standardized or contains a standardized item
- (Blank space) — Not offered
- * — Premium offering

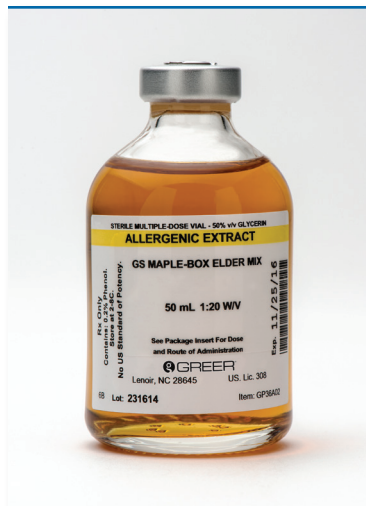
| Item # | Product Name | Testing 5mL Scratch | Treatment Glycerin |
|--|--|---------------------------|-----------------------|
| Standardized Weed Pollen | | | |
| 56 | Ragweed, Short <i>Ambrosia artemisiifolia</i> | ● | ● |
| Mixes Containing Standardized Weed Pollen | | | |
| P3 | National Weed Mix 33 Cocklebur, 55 Giant Ragweed, 43 Lamb's Quarter, 52 Rough/Redroot Pigweed, 56 Short Ragweed | ● | ● |
| P1 | Ragweed Mix 55 Giant, 56 Short | ● | ● |

Pollen



| Item # | Product Name | Testing | | Treatment | |
|-------------------------|--|------------|-----------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Trees and Shrubs | | | | | |
| 172 | Acacia* <i>Acacia</i> spp. | ● | ● | ● | ● |
| 217 | Alder, Red <i>Alnus rubra</i> | ● | ● | ● | ● |
| 69 | Alder, Hazel <i>Alnus serrulata</i> | ● | ● | ● | ● |
| 438 | Alder, White <i>Alnus rhombifolia</i> | ● | ● | ● | ● |
| 204 | Ash, Arizona* <i>Fraxinus velutina</i> | ● | ● | ● | ● |
| 73 | Ash, Oregon <i>Fraxinus latifolia</i> | ● | ● | ● | ● |
| 72 | Ash, Red/Green <i>Fraxinus pennsylvanica</i> | ● | ● | ● | ● |
| 74 | Ash, White <i>Fraxinus americana</i> | ● | ● | ● | ● |
| 75 | Aspen* <i>Populus tremuloides</i> | ● | ● | ● | ● |
| 173 | Bayberry/Wax Myrtle* <i>Morella cerifera</i> | ● | ● | ● | ● |
| 76 | Beech, American* <i>Fagus grandifolia</i> | | ● | ● | |
| 207 | Beefwood/Australian Pine* <i>Casuarina equisetifolia</i> | ● | ● | ● | ● |
| 81 | Birch, Black/Sweet <i>Betula lenta</i> | ● | ● | ● | ● |
| 79 | Birch, River <i>Betula nigra</i> | ● | ● | ● | ● |
| 426 | Birch, Spring <i>Betula occidentalis</i> | ● | ● | ● | ● |
| 80 | Birch, White <i>Betula populifolia</i> | ● | ● | ● | ● |
| 82 | Box Elder <i>Acer negundo</i> | ● | ● | ● | |
| 83 | Cedar, Mountain <i>Juniperus ashei</i> | ● | ● | ● | |
| 84 | Cedar, Red <i>Juniperus virginiana</i> | ● | ● | ● | ● |
| 242 | Cedar, Salt/Tamarisk* <i>Tamarix gallica</i> | ● | ● | ● | |
| 444 | Cottonwood, Black <i>Populus balsamifera</i> ssp. <i>trichocarpa</i> | ● | ● | ● | ● |

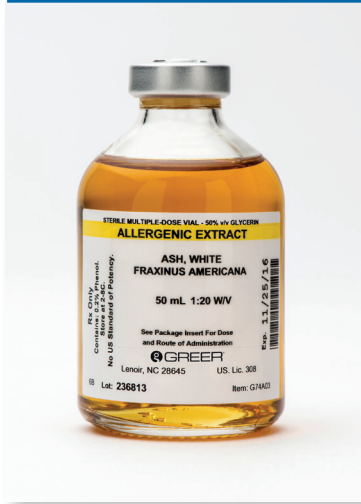
Pollen



| Item # | Product Name | Testing | | Treatment | |
|--|---|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Trees and Shrubs <i>Continued</i> | | | | | |
| 87 | Cottonwood, Eastern <i>Populus deltoides</i> | ● | ● | ● | |
| 255 | Cottonwood, Fremont* <i>Populus fremontii</i> | ● | ● | ● | ● |
| 88 | Cottonwood, Western* <i>Populus deltoides ssp. monilifera</i> | ● | ● | ● | |
| 244 | Cypress, Arizona <i>Cupressus arizonica</i> | ● | ● | ● | ● |
| 193 | Cypress, Bald* <i>Taxodium distichum</i> | ● | ● | ● | ● |
| 92 | Elm, American <i>Ulmus americana</i> | ● | ● | ● | |
| 179 | Elm, Cedar/Fall Blooming* <i>Ulmus crassifolia</i> | ● | ● | ● | ● |
| 93 | Elm, Siberian* <i>Ulmus pumila</i> | ● | ● | ● | ● |
| 196 | Eucalyptus* <i>Eucalyptus globulus</i> | ● | ● | ● | ● |
| 98 | Hackberry* <i>Celtis occidentalis</i> | ● | ● | ● | ● |
| 99 | Hazelnut, American* <i>Corylus americana</i> | ● | ● | ● | ● |
| 103 | Hickory, Shagbark* <i>Carya ovata</i> | ● | ● | ● | ● |
| 102 | Hickory, Shellbark* <i>Carya laciniosa</i> | ● | ● | ● | ● |
| 104 | Hickory, White <i>Carya alba</i> | ● | ● | ● | |
| 243 | Juniper, Oneseed <i>Juniperus monosperma</i> | ● | ● | ● | ● |
| 275 | Juniper, Pinchot <i>Juniperus pinchotii</i> | ● | ● | ● | ● |
| 280 | Juniper, Rocky Mountain <i>Juniperus scopulorum</i> | ● | ● | ● | ● |
| 233 | Juniper, Utah <i>Juniperus osteosperma</i> | ● | ● | ● | ● |
| 254 | Juniper, Western <i>Juniperus occidentalis</i> | ● | ● | ● | ● |
| 169 | Locust Blossom, Black* <i>Robinia pseudoacacia</i> | ● | ● | ● | ● |
| 257 | Mango Blossom* <i>Mangifera indica</i> | ● | ● | ● | ● |
| 108 | Maple, Red <i>Acer rubrum</i> | ● | ● | ● | ● |
| 109 | Maple, Soft/Silver <i>Acer saccharinum</i> | ● | ● | ● | ● |
| 110 | Maple, Sugar/Hard* <i>Acer saccharum</i> | ● | ● | ● | |
| 221 | Melaleuca* <i>Melaleuca quinquenervia</i> | ● | ● | ● | ● |
| 111 | Mesquite* <i>Prosopis velutina</i> | ● | ● | ● | |
| 113 | Mulberry, Paper* <i>Broussonetia papyrifera</i> | ● | ● | ● | ● |
| 112 | Mulberry, Red <i>Morus rubra</i> | ● | ● | ● | |
| 194 | Mulberry, White <i>Morus alba</i> | ● | ● | ● | ● |
| 256 | Oak, Arizona/Gambel* <i>Quercus gambelii</i> | ● | ● | ● | ● |
| 114 | Oak, Black <i>Quercus velutina</i> | | ● | ● | ● |
| 115 | Oak, Bur <i>Quercus macrocarpa</i> | ● | ● | ● | ● |

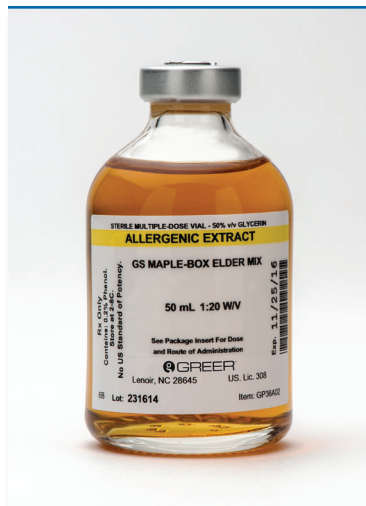
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| Item # | Product Name | Testing | | Treatment | |
|--|---|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Trees and Shrubs <i>Continued</i> | | | | | |
| 284 | Oak, California Black <i>Quercus kelloggii</i> | ● | ● | ● | ● |
| 195 | Oak, California Live <i>Quercus agrifolia</i> | ● | ● | ● | ● |
| 455 | Oak, California White <i>Quercus lobata</i> | ● | ● | ● | ● |
| 119 | Oak, Post <i>Quercus stellata</i> | | ● | ● | ● |
| 120 | Oak, Red <i>Quercus rubra</i> | ● | ● | ● | ● |
| 116 | Oak, Virginia Live <i>Quercus virginiana</i> | ● | ● | ● | ● |
| 198 | Oak, Water <i>Quercus nigra</i> | ● | ● | ● | ● |
| 285 | Oak, Western White <i>Quercus garryana</i> | ● | ● | ● | ● |
| 122 | Oak, White <i>Quercus alba</i> | ● | ● | ● | ● |
| 123 | Olive <i>Olea europaea</i> | ● | ● | ● | ● |
| 479 | Olive, Russian* <i>Elaeagnus angustifolia</i> | ● | ● | ● | ● |
| 247 | Orange Pollen* <i>Citrus sinensis</i> | ● | ● | ● | ● |
| 216 | Palm, Queen <i>Arecastrum romanzoffianum</i> | ● | ● | ● | ● |
| 127 | Pecan <i>Carya illinoensis</i> | ● | ● | ● | ● |
| 258 | Pepper Tree* <i>Schinus spp.</i> | ● | ● | ● | ● |
| 132 | Pine, Loblolly <i>Pinus taeda</i> | ● | ● | ● | ● |
| 129 | Pine, Longleaf <i>Pinus palustris</i> | ● | ● | ● | ● |
| 478 | Pine, Ponderosa <i>Pinus ponderosa</i> | ● | ● | ● | ● |
| 171 | Pine, Virginia Scrub <i>Pinus virginiana</i> | ● | ● | ● | ● |
| 130 | Pine, White/Eastern <i>Pinus strobus</i> | ● | ● | ● | ● |
| 480 | Pine, White/Western <i>Pinus monticola</i> | ● | ● | ● | ● |
| 131 | Pine, Yellow <i>Pinus echinata</i> | ● | ● | ● | ● |
| 177 | Poplar, Lombardy <i>Populus nigra</i> | ● | ● | ● | ● |
| 134 | Poplar, White <i>Populus alba</i> | ● | ● | ● | ● |
| 135 | Privet, Common* <i>Ligustrum vulgare</i> | ● | ● | ● | ● |
| 137 | Sweet Gum <i>Liquidambar styraciflua</i> | ● | ● | ● | ● |
| 138 | Sycamore, American/Eastern <i>Platanus occidentalis</i> | ● | ● | ● | ● |
| 228 | Sycamore, Western* <i>Platanus racemosa</i> | ● | ● | ● | ● |
| 140 | Walnut, Black <i>Juglans nigra</i> | ● | ● | ● | ● |
| 222 | Walnut, California Black <i>Juglans californica</i> | ● | ● | ● | ● |
| 210 | Walnut, English <i>Juglans regia</i> | ● | ● | ● | ● |
| 483 | Willow, Arroyo* <i>Salix lasiolepis</i> | ● | ● | ● | ● |
| 142 | Willow, Black* <i>Salix nigra</i> | ● | ● | ● | |

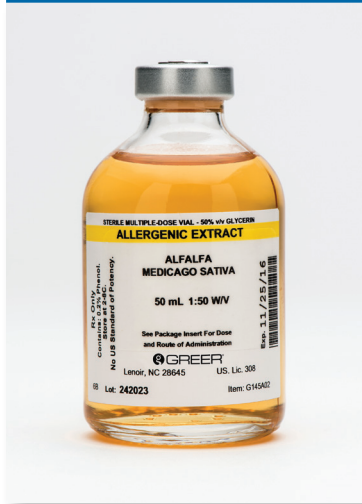
Pollen



| Item # | Product Name | Testing | | Treatment | |
|-------------------|---|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Tree Mixes | | | | | |
| P30 | Ash Mix 72 Red/Green, 74 White | ● | ● | ● | |
| P31 | Birch Mix 81 Black/Sweet, 79 River, 80 White | ● | ● | ● | ● |
| P32 | Elm Mix 92 American, 93 Siberian | ● | ● | ● | |
| P33 | Hickory Mix 101 Pignut, 103 Shagbark, 102 Shellbark, 104 White | ● | ● | ● | ● |
| P34 | Hickory-Pecan Mix 127 Pecan, 103 Shagbark Hickory | ● | ● | ● | ● |
| PO800 | Juniper Mix 243 Oneseed, 280 Rocky Mountain | ● | ● | ● | ● |
| P35 | 2 Maple Mix 108 Red, 110 Sugar/Hard | ● | ● | ● | |
| P53 | 3 Maple Mix 108 Red, 109 Soft/Silver, 110 Sugar/Hard | ● | ● | ● | |
| P36 | Maple-Box Elder Mix 82 Box Elder, 110 Sugar/Hard Maple | ● | ● | ● | |
| P37 | Pine Mix 132 Loblolly, 130 White/Eastern, 131 Yellow | ● | ● | ● | ● |
| P38 | Eastern Oak Mix 114 Black, 120 Red, 122 White | ● | ● | ● | ● |
| P46 | Eastern 6 Tree Mix 76 American Beech, 87 Eastern Cottonwood, 120 Red Oak, 79 River Birch, 103 Shagbark Hickory, 74 White Ash | ● | ● | ● | ● |
| P47 | Eastern 7 Tree Mix 92 American Elm, P46 Eastern 6 Tree Mix | ● | ● | ● | ● |
| P48 | Eastern 8 Tree Mix 92 American Elm, P46 Eastern 6 Tree Mix, 110 Maple Sugar/Hard | ● | ● | ● | |
| P50 | Eastern 10 Tree Mix 138 American/Eastern Sycamore, 92 American Elm, P46 Eastern 6 Tree Mix, 110 Sugar/Hard Maple, 137 Sweet Gum | ● | ● | ● | |
| P49 | Central/Eastern 4 Tree Mix 92 American Elm, 82 Box Elder, 127 Pecan, 116 Virginia Live Oak | ● | ● | ● | ● |
| P41 | Western Oak Mix 284 California Black, 195 California Live, 285 Western White | ● | ● | ● | ● |
| P56 | Western 3 Tree Mix 93 Siberian Elm, 228 Western Sycamore, 123 Olive | ● | ● | ● | |
| P39 | Western 10 Tree Mix 172 Acacia, 82 Box Elder, 255 Fremont Cottonwood, 123 Olive, 93 Siberian Elm, 426 Spring Birch, 254 Western Juniper, 228 Western Sycamore, 285 Western White Oak, 194 White Mulberry | ● | ● | ● | ● |
| P51 | Western Walnut Mix 222 California Black, 210 English | ● | ● | ● | ● |
| PO714 | 11 Tree Mix 76 American Beech, 138 American/Eastern Sycamore, 92 American Elm, 140 Black Walnut, 142 Black Willow, 87 Eastern Cottonwood, 120 Red Oak, 79 River Birch, 103 Shagbark Hickory, 110 Sugar/Hard Maple, 74 White Ash | ● | ● | ● | ● |
| PO3621 | Southern CA Tree Mix D 172 Acacia, 82 Box Elder, 222 California Black Walnut, P32 Elm Mix, 255 Fremont Cottonwood, 111 Mesquite, 123 Olive, 426 Spring Birch, P41 Western Oak Mix, 228 Western Sycamore, 194 White Mulberry | | ● | ● | |

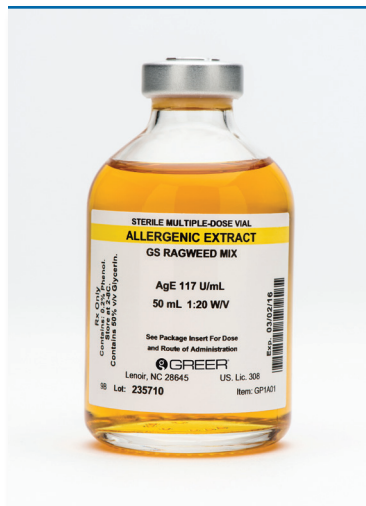
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| Item # | Product Name | Testing | | Treatment | |
|--------------------------|--|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Flowers | | | | | |
| 158 | Daisy, Ox-Eye* <i>Leucanthemum vulgare</i> | | ● | ● | ● |
| 159 | Dandelion <i>Taraxacum officinale</i> | ● | ● | ● | ● |
| 164 | Sunflower <i>Helianthus annuus</i> | ● | ● | ● | ● |
| Cultivated Plants | | | | | |
| 145 | Alfalfa* <i>Medicago sativa</i> | ● | ● | ● | ● |
| 174 | Mustard* <i>Brassica</i> spp. | ● | ● | ● | ● |
| 146 | Red Clover* <i>Trifolium pratense</i> | ● | ● | ● | ● |
| 63 | Sugar Beet <i>Beta vulgaris</i> | ● | ● | ● | ● |
| Weeds | | | | | |
| 202 | Allscale* <i>Atriplex polycarpa</i> | ● | ● | ● | ● |
| 225 | Baccharis* <i>Baccharis</i> spp. | ● | ● | ● | ● |
| 213 | Burrobrush* <i>Hymenoclea salsola</i> | ● | ● | ● | ● |
| 32 | Careless Weed, Amaranth/Green <i>Amaranthus hybridus</i> | ● | ● | ● | ● |
| 33 | Cocklebur* <i>Xanthium strumarium</i> | ● | ● | ● | |
| 35 | Dock, Yellow/Curly* <i>Rumex crispus</i> | ● | ● | ● | ● |
| 184 | Dog Fennel* <i>Eupatorium capillifolium</i> | ● | ● | ● | ● |
| 42 | Firebush/Kochia <i>Kochia scoparia</i> | ● | ● | ● | ● |
| 37 | Goldenrod* <i>Solidago</i> spp. | ● | ● | ● | ● |
| 40 | Hemp, Water* <i>Amaranthus rudis</i> | ● | ● | ● | |
| 212 | Iodine Bush* <i>Allenrolfea occidentalis</i> | ● | ● | ● | ● |
| 43 | Lamb's Quarter <i>Chenopodium album</i> | ● | ● | ● | |
| 240 | Lenscale/Quailbrush <i>Atriplex lentiformis</i> | ● | ● | ● | ● |
| 44 | Marsh Elder, Burweed/Giant Poverty* <i>Iva xanthifolia</i> | ● | ● | ● | |
| 45 | Marsh Elder, True/Rough <i>Iva annua</i> | ● | ● | ● | |
| 47 | Mugwort, Common* <i>Artemisia vulgaris</i> | ● | ● | ● | |
| 48 | Mugwort, Darkleaved/Sagebrush, Prairie* <i>Artemisia ludoviciana</i> | ● | ● | ● | ● |
| 36 | Nettle <i>Urtica dioica</i> | ● | ● | ● | ● |
| 49 | Palmer's Amaranth <i>Amaranthus palmeri</i> | ● | ● | ● | ● |
| 52 | Pigweed, Rough/Redroot <i>Amaranthus retroflexus</i> | ● | ● | ● | |
| 53 | Pigweed, Spiny* <i>Amaranthus spinosus</i> | | ● | ● | |

Pollen



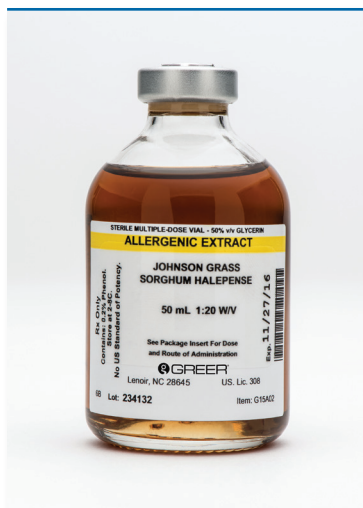
| Item # | Product Name | Testing | | Treatment | |
|-------------------------------|---|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Weeds <i>Continued</i> | | | | | |
| 54 | Plantain, English <i>Plantago lanceolata</i> | ● | ● | ● | ● |
| 238 | Rabbit Bush <i>Ambrosia deltoidea</i> | ● | ● | ● | ● |
| 272 | Ragweed, Desert <i>Ambrosia dumosa</i> | ● | ● | ● | ● |
| 185 | Ragweed, False* <i>Ambrosia acanthicarpa</i> | ● | ● | ● | ● |
| 55 | Ragweed, Giant <i>Ambrosia trifida</i> | ● | ● | ● | ● |
| 56 | Ragweed, Short <i>Ambrosia artemisiifolia</i> | | ● | ● | |
| 50 | Ragweed, Slender* <i>Ambrosia confertiflora</i> | ● | ● | ● | ● |
| 57 | Ragweed, Southern* <i>Ambrosia bidentata</i> | ● | ● | ● | ● |
| 58 | Ragweed, Western <i>Ambrosia psilostachya</i> | ● | ● | ● | ● |
| 59 | Russian Thistle* <i>Salsola kali</i> | ● | ● | ● | |
| 61 | Sagebrush, Common <i>Artemisia tridentata</i> | ● | ● | ● | ● |
| 199 | Saltbush, Annual* <i>Atriplex wrightii</i> | ● | ● | ● | ● |
| 64 | Sorrel, Sheep/Red <i>Rumex acetosella</i> | ● | ● | ● | ● |
| 206 | Wingscale <i>Atriplex canescens</i> | ● | ● | ● | ● |

Weed Mixes

| | | | | | |
|-------|--|---|---|---|---|
| P54 | 3 Weed Mix 33 Cocklebur, 43 Lamb's Quarter, 52 Rough/Redroot Pigweed | ● | ● | ● | ● |
| P14 | Central/Western Weed Mix 42 Firebush/Kochia, 43 Lamb's Quarter, 59 Russian Thistle | ● | ● | ● | ● |
| P15 | Common Weed Mix 33 Cocklebur, 54 English Plantain, 43 Lamb's Quarter, 52 Rough/Redroot Pigweed, 59 Russian Thistle | ● | ● | ● | ● |
| PO216 | Dock-Sorrel Mix 64 Sheep Sorrel, 35 Yellow/Curly Dock | ● | ● | ● | ● |
| P3 | National Weed Mix 33 Cocklebur, 55 Giant Ragweed, 43 Lamb's Quarter, 52 Rough/Redroot Pigweed, 56 Short Ragweed | | ● | ● | |
| P5 | Pigweed Mix 32 Careless Weed/Amaranth Green, 49 Palmer's Amaranth, 52 Rough/Redroot Pigweed | ● | ● | ● | ● |
| P6 | Plantain-Sorrel Mix 54 English Plantain, 64 Sheep/Red Sorrel | ● | ● | ● | ● |
| P1 | Ragweed Mix 55 Giant, 56 Short | | ● | ● | |
| P8 | Sage Mix 61 Common Sagebrush, 48 Darkleaved Mugwort/Prairie Sagebrush | ● | ● | ● | |
| P9 | Scale/Atriplex Mix 202 Allscale, 240 Lenscale/Quailbrush, 206 Wingscale | ● | ● | ● | ● |

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- (Blank space) — Not offered
- * — Premium offering



| Item # | Product Name | Testing | | Treatment | |
|------------------------------------|---|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Weed Mixes <i>Continued</i> | | | | | |
| P03138 | Southern CA Weed Mix B 32 Careless Weed, Amaranth/Green, P14 Central/Western Weed Mix, 33 Cocklebur, 61 Common Sagebrush, P0216 Dock-Sorrel Mix, 54 English Plantain, P9 Scale/Atriplex Mix, P2 Western Ragweed Mix | | ● | ● | |
| P2 | Western Ragweed Mix 185 False, 58 Western | ● | ● | ● | ● |
| Grasses | | | | | |
| 231 | Bahia Grass* <i>Paspalum notatum</i> | ● | ● | ● | ● |
| T2 | Bermuda Grass <i>Cynodon dactylon</i> | | ● | ● | |
| 8 | Brome Grass, Smooth* <i>Bromus inermis</i> | ● | ● | ● | ● |
| 9 | Canarygrass, Reed <i>Phalaris arundinacea</i> | ● | ● | ● | ● |
| 149 | Corn, Cultivated <i>Zea mays</i> | ● | ● | ● | ● |
| 11 | Couch/Quack Grass* <i>Elymus repens</i> | | ● | ● | |
| 15 | Johnson Grass <i>Sorghum halepense</i> | ● | ● | ● | |
| T16 | Kentucky Blue/June <i>Poa pratensis</i> | | ● | ● | |
| T14 | Meadow Fescue <i>Festuca elatior (pratensis)</i> | | ● | ● | |
| 21 | Oats, Common/Cultivated <i>Avena sativa</i> | ● | ● | ● | ● |
| T20 | Orchard <i>Dactylis glomerata</i> | | ● | ● | |
| T22 | Redtop <i>Agrostis (alba) gigantea</i> | | ● | ● | |
| 23 | Rye, Cultivated <i>Secale cereale</i> | ● | ● | ● | ● |
| 264 | Ryegrass, Giant Wild* <i>Leymus condensatus</i> | ● | ● | ● | ● |
| 25 | Ryegrass, Italian* <i>Lolium perenne ssp. multiflorum</i> | ● | ● | ● | ● |
| T24 | Ryegrass, Perennial <i>Lolium perenne</i> | | ● | ● | |
| T27 | Sweet Vernal* <i>Anthoxanthum odoratum</i> | | ● | ● | |
| T28 | Timothy <i>Phleum pratense</i> | | ● | ● | |
| 29 | Velvetgrass* <i>Holcus lanatus</i> | ● | ● | ● | ● |
| 31 | Wheat, Cultivated* <i>Triticum aestivum</i> | ● | ● | ● | ● |
| 186 | Wheatgrass, Western* <i>Pascopyrum smithii</i> | ● | ● | ● | ● |

Pollen



| Item # | Product Name | Testing | | Treatment | |
|--------------------|--|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Grass Mixes | | | | | |
| TP20 | Timothy/Orchard Grass Mix 28 Timothy, 20 Orchard Grass | | ● | ● | |
| TP23 | K-O-T Grass Mix 16 Kentucky Blue/June, 20 Orchard, 28 Timothy | | ● | ● | |
| TP24 | K-O-R-T Grass Mix 16 Kentucky Blue/June, 20 Orchard, 22 Redtop, 28 Timothy | | ● | ● | |
| TP25 | K-O-R-T and Sweet Vernal Mix 16 Kentucky Blue/June, 20 Orchard, 22 Redtop, 28 Timothy, 27 Sweet Vernal | | ● | ● | |
| TP29 | T-O-S Grass Mix 28 Timothy, 20 Orchard, 27 Sweet Vernal | | ● | ● | |
| TP27 | 7 Grass Mix 16 Kentucky Blue/June, 14 Meadow Fescue, 20 Orchard, 24 Perennial Ryegrass, 22 Redtop, 27 Sweet Vernal, 28 Timothy | | ● | ● | |
| P19 | 9 Southern Grass Mix (Concentrate) Equal parts of: 2 Bermuda at 10,000 BAU/mL, P27 7 Grass Mix at 100,000 BAU/mL, 15 Johnson at 1:20 w/v | | ● | ● | |

Fungi and Smuts

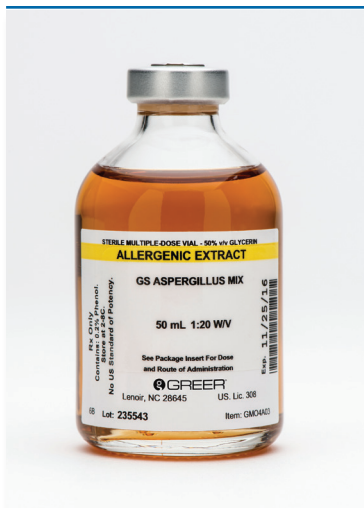


| Item # | Product Name | Testing | | Treatment | |
|--------------|---|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Fungi | | | | | |
| M6 | Acremonium strictum <i>Cephalosporium acremonium</i> | ● | ● | ● | |
| M1 | Alternaria alternata <i>Alternaria tenuis</i> | ● | ● | ● | ● |
| M4 | Aspergillus amstelodami <i>Aspergillus glaucus</i> | ● | ● | ● | ● |
| M2 | Aspergillus flavus | ● | ● | ● | ● |
| M3 | Aspergillus fumigatus | ● | ● | ● | ● |
| M35 | Aspergillus nidulans | ● | ● | ● | ● |
| M5 | Aspergillus niger | ● | ● | ● | ● |
| M21 | Aureobasidium pullulans <i>Pullularia pullulans</i> | ● | ● | ● | |
| M12 | Bipolaris sorokiniana <i>Helminthosporium sativum</i> | ● | ● | ● | ● |
| M30 | Botrytis cinerea | ● | ● | ● | |
| M15 | Candida albicans | ● | ● | ● | ● |
| M8 | Chaetomium globosum | ● | ● | ● | ● |
| M9 | Cladosporium herbarum | ● | ● | ● | ● |

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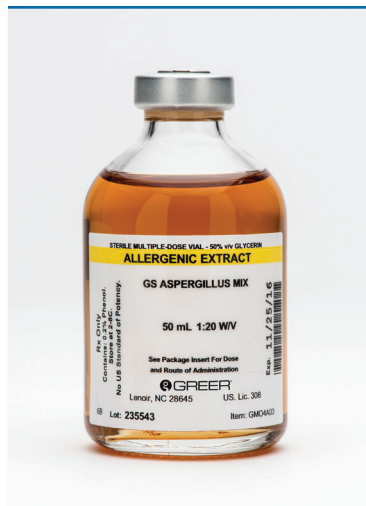
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- — Standardized or contains a standardized item
- (Blank space) — Not offered
- * — Premium offering

Fungi and Smuts



| Item # | Product Name | Testing | | Treatment | |
|------------------------|--|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Fungi Continued | | | | | |
| M13 | <i>Cladosporium sphaerospermum</i> <i>Hormodendrum hordei</i> | ● | ● | ● | |
| M45 | <i>Drechslera spicifera</i> <i>Curvularia spicifera</i> | ● | ● | ● | ● |
| M29 | <i>Epicoccum nigrum</i> <i>Epicoccum purpurascens</i> | ● | ● | ● | |
| M10 | <i>Epidermophyton floccosum</i> | ● | ● | ● | ● |
| M11 | <i>Fusarium moniliforme</i> | ● | ● | ● | ● |
| M48 | <i>Fusarium solani</i> | ● | ● | ● | ● |
| M63 | <i>Geotrichum candidum</i> | ● | ● | ● | |
| M41 | <i>Gliocladium viride*</i> <i>Gliocladium deliquescens</i> | ● | ● | ● | ● |
| M40 | <i>Helminthosporium solani</i> <i>Spondylocladium atrovirens</i> | ● | ● | ● | ● |
| M14 | <i>Microsporium canis*</i> | | ● | ● | |
| M52 | <i>Mucor circinelloides f. circinelloides</i> <i>Mucor mucedo</i> | ● | ● | ● | ● |
| M68 | <i>Mucor circinelloides f. lusitanicus</i> <i>Mucor racemosus</i> | ● | ● | ● | |
| M27 | <i>Mucor plumbeus</i> | ● | ● | ● | ● |
| M44 | <i>Mycogone perniciosa</i> | | ● | ● | |
| M16 | <i>Neurospora intermedia</i> <i>Monilia sitophila</i> | ● | ● | ● | ● |
| M43 | <i>Nigrospora oryzae</i> | | ● | ● | |
| M34 | <i>Paecilomyces variotii</i> | ● | ● | ● | ● |
| M28 | <i>Penicillium digitatum*</i> | ● | ● | ● | ● |
| M19 | <i>Penicillium chrysogenum</i> (<i>notatum</i>) | ● | ● | ● | ● |
| M39 | <i>Phoma betae</i> | ● | ● | ● | |
| M22 | <i>Rhizopus oryzae</i> <i>Rhizopus arrhizus</i> | ● | ● | ● | ● |
| M23 | <i>Rhizopus stolonifer</i> <i>Rhizopus nigricans</i> | ● | ● | ● | ● |
| M49 | <i>Rhodotorula mucilaginosa</i> <i>Rhodotorula rubra</i> | ● | ● | ● | ● |
| M67 | <i>Saccharomyces cerevisiae</i> | ● | ● | ● | ● |
| M33 | <i>Stemphylium solani</i> | ● | ● | ● | ● |
| M24 | <i>Trichoderma harzianum*</i> <i>Trichoderma viride</i> | ● | ● | ● | ● |
| M26 | <i>Trichophyton mentagrophytes</i> | ● | ● | ● | ● |
| M50 | <i>Trichophyton rubrum</i> | ● | ● | ● | |
| M7 | <i>Trichothecium roseum</i> <i>Cephalothecium roseum</i> | ● | ● | ● | ● |

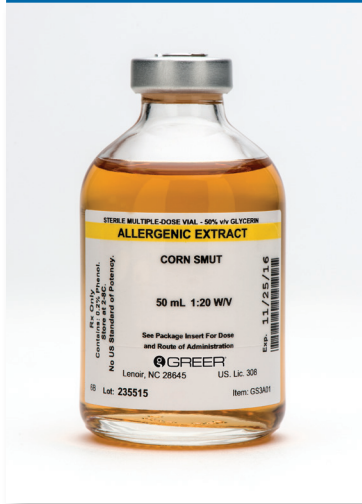
Fungi and Smuts



| Item # | Product Name | Testing | | Treatment | |
|--------------------|---|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Fungi Mixes | | | | | |
| M01 | Mold Mix #1 M1 <i>Alternaria alternata</i> , M5 <i>Aspergillus niger</i> , M12 <i>Bipolaris sorokiniana</i> , M13 <i>Cladosporium sphaerospermum</i> , M19 <i>Penicillium chrysogenum (notatum)</i> | ● | ● | ● | ● |
| M02 | Mold Mix #2 M21 <i>Aureobasidium pullulans</i> , M45 <i>Drechslera spicifera</i> , M11 <i>Fusarium moniliforme</i> , M27 <i>Mucor plumbeus</i> , M23 <i>Rhizopus stolonifer</i> | ● | ● | ● | ● |
| M03 | Mold Mix #3 M1 <i>Alternaria alternata</i> , M5 <i>Aspergillus niger</i> , M13 <i>Cladosporium sphaerospermum</i> , M19 <i>Penicillium chrysogenum (notatum)</i> | ● | ● | ● | ● |
| M064 | AHH Mix M1 <i>Alternaria alternata</i> , M12 <i>Bipolaris sorokiniana</i> , M13 <i>Cladosporium sphaerospermum</i> | ● | ● | ● | ● |
| M0126 | Alternaria/Hormodendrum Mix M1 <i>Alternaria alternata</i> , M13 <i>Cladosporium sphaerospermum</i> | ● | ● | ● | ● |
| M04 | Aspergillus Mix M4 <i>A. amstelodami</i> , M2 <i>A. flavus</i> , M3 <i>A. fumigatus</i> , M35 <i>A. nidulans</i> , M5 <i>A. niger</i> | ● | ● | ● | ● |
| M0464 | Dematiaceae Mix M1 <i>Alternaria alternata</i> , M21 <i>Aureobasidium pullulans</i> , M12 <i>Bipolaris sorokiniana</i> , M9 <i>Cladosporium herbarum</i> , M45 <i>Drechslera spicifera</i> , M40 <i>Helminthosporium solani</i> | ● | ● | ● | ● |
| M09 | Fusarium Mix M11 <i>F. moniliforme</i> , M48 <i>F. solani</i> | ● | ● | ● | ● |
| M010 | Monilia Mix M15 <i>Candida albicans</i> , M16 <i>Neurospora intermedia</i> | ● | ● | ● | ● |
| M011 | Mucor Mix M68 <i>M. circinelloides f. lusitanicus</i> , M27 <i>M. plumbeus</i> | ● | ● | ● | |
| M08 | New Stock Fungi Mix M6 <i>Acremonium strictum</i> , M1 <i>Alternaria alternata</i> , M5 <i>Aspergillus niger</i> , M21 <i>Aureobasidium pullulans</i> , M12 <i>Bipolaris sorokiniana</i> , M30 <i>Botrytis cinerea</i> , M15 <i>Candida albicans</i> , M8 <i>Chaetomium globosum</i> , M13 <i>Cladosporium sphaerospermum</i> , M29 <i>Epicoccum nigrum</i> , M11 <i>Fusarium moniliforme</i> , M27 <i>Mucor plumbeus</i> , M19 <i>Penicillium chrysogenum (notatum)</i> , M39 <i>Phoma betae</i> , M23 <i>Rhizopus stolonifer</i> , M26 <i>Trichophyton mentagrophytes</i> | ● | ● | ● | ● |
| M05 | Penicillium Mix M17 <i>P. camembertii</i> , M18 <i>P. chrysogenum</i> , M28 <i>P. digitatum</i> , M19 <i>P. chrysogenum (notatum)</i> , M42 <i>P. roquefortii</i> | ● | ● | ● | ● |
| M06 | Phycomycetes Mix M68 <i>Mucor circinelloides f. lusitanicus</i> , M23 <i>Rhizopus stolonifer</i> | ● | ● | ● | ● |
| M012 | Rhizopus Mix M22 <i>R. oryzae</i> , M23 <i>R. stolonifer</i> | ● | ● | ● | ● |

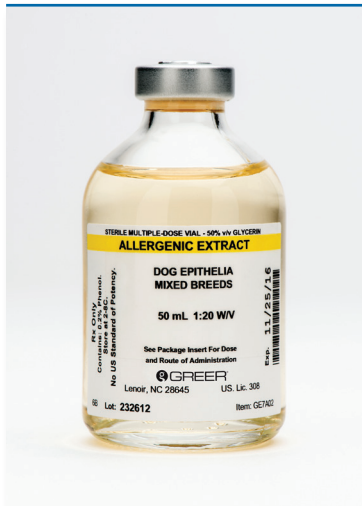
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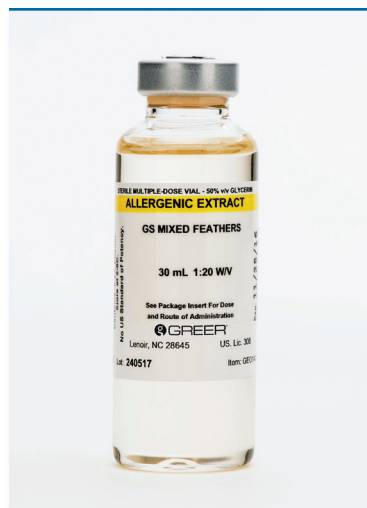
| Item # | Product Name | Testing | | Treatment | |
|---|---|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Smuts | | | | | |
| All smut products are available in 50% glycerin only. | | | | | |
| S1 | Loose Barley Smut <i>Ustilago nuda</i> | | ● | ● | |
| S2 | Bermuda Grass Smut <i>Ustilago cynodontis</i> | | ● | ● | |
| S3 | Corn Smut* <i>Ustilago maydis</i> | | ● | ● | |
| S4 | Johnson Grass Smut <i>Sporisorium cruentum</i> | | ● | ● | |
| S5 | Oat Smut <i>Ustilago avenae</i> | | ● | ● | |
| S6 | Loose Wheat Smut* <i>Ustilago tritici</i> | | ● | ● | |
| Smut Mixes | | | | | |
| All smut mixes are available in 50% glycerin only. | | | | | |
| S02 | Grain Smut Mix* S3 Corn Smut, S1 Loose Barley Smut, S6 Loose Wheat Smut, S5 Oat Smut | | ● | ● | |
| S01 | Grass Smut Mix* S2 Bermuda Grass Smut, S4 Johnson Grass Smut | | ● | ● | |

Epithelia, Miscellaneous Inhalants, and Insects



| Item # | Product Name | Testing | | Treatment | |
|------------------|---|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Epithelia | | | | | |
| TE3 | Standardized Cat Hair <i>Felis catus (domesticus)</i> | | ● | ● | |
| E4 | Cattle Epithelia* <i>Bos taurus</i> | ● | ● | ● | ● |
| E7 | Dog Epithelia <i>Canis sp.</i> | ● | ● | ● | ● |
| E50 | Gerbil Epithelia <i>Meriones unguiculatus</i> | ● | ● | ● | ● |
| E11 | Goat Epithelia <i>Capra hircus</i> | ● | ● | ● | ● |
| E14 | Guinea Pig Epithelia* <i>Cavia porcellus</i> | ● | ● | ● | ● |
| E44 | Hamster Epithelia* <i>Mesocricetus auratus</i> | ● | ● | ● | ● |
| E15 | Hog Epithelia <i>Sus scrofa</i> | ● | ● | ● | ● |
| E17 | Horse Epithelia* <i>Equus caballus</i> | ● | ● | ● | ● |
| E20 | Mouse Epithelia* <i>Mus musculus</i> | ● | ● | ● | ● |
| E24 | Rabbit Epithelia* <i>Oryctolagus cuniculus</i> | ● | ● | ● | ● |
| E25 | Rat Epithelia* <i>Rattus norvegicus</i> | ● | ● | ● | ● |

Epithelia, Miscellaneous Inhalants, and Insects

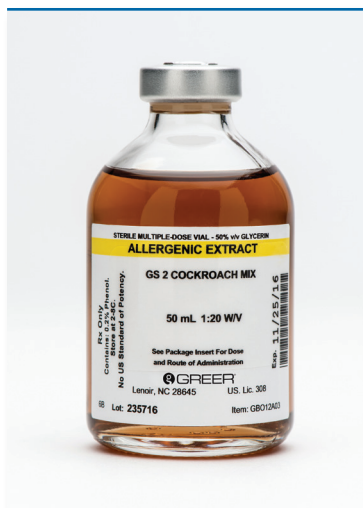


| Item # | Product Name | Testing | | Treatment | |
|--------------------|--|---------|--------------|-----------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Feathers | | | | | |
| E2 | Canary Feathers* <i>Serinus canaria</i> | ● | ● | ● | ● |
| E5 | Chicken Feathers <i>Gallus gallus</i> | ● | ● | ● | ● |
| E8 | Duck Feathers <i>Anas platyrhynchos</i> | ● | ● | ● | ● |
| E12 | Goose Feathers <i>Anser anser</i> | ● | ● | ● | ● |
| E22 | Parakeet Feathers* Psittacidae | ● | ● | ● | ● |
| Feather Mix | | | | | |
| E01 | Feather Mix E5 Chicken Feathers, E8 Duck Feathers, E12 Goose Feathers | ● | ● | ● | ● |

| Item # | Product Name | Testing | | Bulk Extract | |
|--------------------------------|---|---------|--------------|--------------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Miscellaneous Inhalants | | | | | |
| H21 | Cotton Linters <i>Gossypium hirsutum</i> | ● | ● | ● | ● |
| H1 | Cottonseed <i>Gossypium hirsutum</i> (For Diagnostic Use Only) | ● | ● | ● | ● |
| H2 | Flaxseed <i>Linum usitatissimum</i> (For Diagnostic Use Only) | ● | ● | ● | ● |
| H15 | Gum Arabic* <i>Acacia senegal</i> | ● | ● | ● | ● |
| H9 | Gum Karaya <i>Sterculia urens</i> | ● | ● | ● | ● |
| H18 | Gum Tragacanth* <i>Astragalus gummifer</i> | ● | ● | ● | ● |
| H4 | Kapok Seed <i>Ceiba pentandra</i> | ● | ● | ● | ● |
| H5 | Orris Root <i>Iris germanica var. florentina</i> | ● | ● | ● | ● |
| H6 | Pyrethrum <i>Chrysanthemum cinerariifolium</i> | ● | ● | ● | ● |
| H7 | Silk* <i>Bombyx mori</i> | ● | ● | ● | ● |
| H8 | Tobacco Leaf <i>Nicotiana tabacum</i> | ● | ● | ● | ● |
| Insects (Whole Body) | | | | | |
| B31 | Ant, Black/Carpenter* <i>Camponotus pennsylvanicus</i> | ● | ● | ● | ● |
| B47 | Ant, Fire* <i>Solenopsis invicta</i> | ● | ● | ● | |
| B14 | Ant, Fire* <i>Solenopsis richteri</i> | ● | ● | ● | |
| B28 | Caddisfly* Trichoptera | ● | ● | ● | ● |

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- (Blank space) — Not offered
- * — Premium offering



| Item # | Product Name | Testing | | Bulk Extract | |
|--|--|---------|--------------|--------------|---------|
| | | ID 5 mL | Scratch 5 mL | Glycerin | Aqueous |
| Insects (Whole Body) <i>Continued</i> | | | | | |
| B26 | Cockroach, American* <i>Periplaneta americana</i> | ● | ● | ● | |
| B46 | Cockroach, German* <i>Blattella germanica</i> | ● | ● | ● | |
| B39 | Deer Fly* <i>Chrysops</i> spp. | ● | ● | ● | ● |
| B22 | Flea* (Aqueous Only) <i>Ctenocephalides</i> spp. (For Diagnostic Use Only) | ● | | | ● |
| B8 | House Fly* <i>Musca domestica</i> (For Diagnostic Use Only) | ● | ● | ● | ● |
| B12 | Mayfly* Ephemeroptera | ● | ● | ● | ● |
| B10 | Mosquito* <i>Aedes taeniorhynchus</i> (For Diagnostic Use Only) | ● | ● | ● | ● |
| B11 | Moth* Lepidoptera (For Diagnostic Use Only) | ● | ● | ● | ● |
| Insect Mix (Whole Body) | | | | | |
| B012 | 2 Cockroach Mix B26 American, B46 German | ● | ● | ● | |

Ingestant Extracts

All ingestant products are available in 50% glycerin only.



| Item # | Product Name | Testing | | Bulk Extract |
|--------------------|---|---------|---------|--------------|
| | | 5mL | Scratch | Glycerin |
| Plant Foods | | | | |
| F48 | Apple <i>Malus pumila</i> | ● | | ● |
| F49 | Apricot <i>Prunus armeniaca</i> | ● | | ● |
| F55 | Banana <i>Musa x paradisiaca</i> var. <i>paradisiaca</i> | ● | | ● |
| F56 | Barley <i>Hordeum vulgare</i> (For Diagnostic Use Only) | ● | | ● |
| F59 | Bean, Lima <i>Phaseolus lunatus</i> | ● | | ● |
| F295 | Bean, Navy <i>Phaseolus vulgaris</i> | ● | | ● |
| F63 | Bean, String/Green <i>Phaseolus vulgaris</i> | ● | | ● |
| F70 | Blueberry <i>Vaccinium myrtilloides</i> | ● | | ● |
| F73 | Broccoli <i>Brassica oleracea</i> var. <i>botrytis</i> | ● | | ● |
| F75 | Buckwheat <i>Fagopyrum esculentum</i> | ● | | ● |
| F76 | Cabbage <i>Brassica oleracea</i> var. <i>capitata</i> | ● | | ● |
| F79 | Cantaloupe <i>Cucumis melo</i> | ● | | ● |
| F82 | Carrot <i>Daucus carota</i> | ● | | ● |

Ingestant Extracts

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| Item # | Product Name | Testing 5mL Scratch | Bulk Extract Glycerin |
|-------------------------------------|---|---------------------|-----------------------|
| Plant Foods <i>Continued</i> | | | |
| F87 | Cauliflower <i>Brassica oleracea</i> var. <i>botrytis</i> | ● | ● |
| F88 | Celery <i>Apium graveolens</i> var. <i>dulce</i> | ● | ● |
| F90 | Cherry <i>Prunus</i> sp. | ● | ● |
| F93 | Chocolate/Cocoa bean <i>Theobroma cacao</i> | ● | ● |
| F95 | Cinnamon <i>Cinnamomum verum</i> | ● | ● |
| F99 | Coffee <i>Coffea arabica</i> (For Diagnostic Use Only) | ● | ● |
| F102 | Corn <i>Zea mays</i> | ● | ● |
| F108 | Cranberry <i>Vaccinium macrocarpon</i> | ● | ● |
| F109 | Cucumber <i>Cucumis sativus</i> | ● | ● |
| F121 | Garlic <i>Allium sativum</i> | ● | ● |
| F122 | Ginger <i>Zingiber officinale</i> | ● | ● |
| F125 | Grape <i>Vitis</i> sp. | ● | ● |
| F128 | Grapefruit <i>Citrus x paradisi</i> | ● | ● |
| F132 | Hops <i>Humulus lupulus</i> | ● | ● |
| F139 | Lemon <i>Citrus limon</i> | ● | ● |
| F142 | Lettuce <i>Lactuca sativa</i> | ● | ● |
| F148 | Malt <i>Hordeum vulgare</i> | ● | ● |
| F150 | Mushroom <i>Agaricus bisporus</i> | ● | ● |
| F297 | Mustard <i>Brassica</i> spp. | ● | ● |
| F153 | Nutmeg <i>Myristica fragrans</i> | ● | ● |
| F154 | Oat <i>Avena sativa</i> (For Diagnostic Use Only) | ● | ● |
| F156 | Olive, Green <i>Olea europaea</i> | ● | ● |
| F158 | Onion <i>Allium cepa</i> var. <i>cepa</i> | ● | ● |
| F159 | Orange <i>Citrus sinensis</i> | ● | ● |
| F169 | Pea, Green (English) <i>Pisum sativum</i> | ● | ● |
| F170 | Peach <i>Prunus persica</i> | ● | ● |
| F172 | Pear <i>Pyrus communis</i> | ● | ● |
| F175 | Pepper, Black <i>Piper nigrum</i> | ● | ● |
| F177 | Pepper, Green <i>Capsicum annuum</i> var. <i>annuum</i> | ● | ● |

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All ingestant products are available in 50% glycerin only.



| Item # | Product Name | Testing 5mL Scratch | Bulk Extract Glycerin |
|-------------------------------------|--|------------------------|--------------------------|
| Plant Foods <i>Continued</i> | | | |
| F181 | Pineapple <i>Ananas comosus</i> (For Diagnostic Use Only) | ● | ● |
| F190 | Potato, Sweet <i>Ipomoea batatas</i> | ● | ● |
| F191 | Potato, White <i>Solanum tuberosum</i> | ● | ● |
| F198 | Raspberry <i>Rubus idaeus</i> var. <i>idaeus</i> | ● | ● |
| F200 | Rice <i>Oryza sativa</i> | ● | ● |
| F204 | Rye <i>Secale cereale</i> (For Diagnostic Use Only) | ● | ● |
| F208 | Sesame Seed <i>Sesamum orientale</i> (<i>indicum</i>) | ● | ● |
| F209 | Soybean <i>Glycine max</i> | ● | ● |
| F211 | Spinach <i>Spinacia oleracea</i> (For Diagnostic Use Only) | ● | ● |
| F212 | Squash, Yellow Summer Cucurbitaceae | ● | ● |
| F216 | Strawberry <i>Fragaria chiloensis</i> | ● | ● |
| F225 | Tomato <i>Solanum lycopersicum</i> var. <i>lycopersicum</i> | ● | ● |
| F228 | Vanilla Bean <i>Vanilla planifolia</i> | ● | ● |
| F232 | Watermelon <i>Citrullus lanatus</i> var. <i>lanatus</i> | ● | ● |
| F235 | Wheat, Whole <i>Triticum aestivum</i> (For Diagnostic Use Only) | ● | ● |
| Animal Foods | | | |
| F241 | Beef <i>Bos taurus</i> | ● | ● |
| F251 | Lamb <i>Ovis aries</i> | ● | ● |
| F258 | Pork <i>Sus scrofa</i> | ● | ● |
| Poultry Products | | | |
| F265 | Chicken Meat <i>Gallus gallus</i> | ● | ● |
| F272 | Egg White, Chicken <i>Gallus gallus</i> | ● | ● |
| F271 | Egg Whole, Chicken <i>Gallus gallus</i> | ● | ● |
| F273 | Egg Yolk, Chicken <i>Gallus gallus</i> | ● | ● |
| F346 | Turkey Meat <i>Meleagris gallopavo</i> | ● | ● |
| Dairy Products | | | |
| F293 | Milk, Cow <i>Bos taurus</i> | ● | ● |

Ingestant Extracts

All ingestant products are available in 50% glycerin only.



| Item # | Product Name | Testing 5mL Scratch | Bulk Extract Glycerin |
|---------------------------|---|---------------------|-----------------------|
| Fish and Shellfish | | | |
| F4 | Bass, Black <i>Centropristis striata</i> | ● | ● |
| F8 | Catfish <i>Ictalurus spp.</i> | ● | ● |
| F10 | Clam <i>Mercenaria spp.</i> | ● | ● |
| F11 | Codfish <i>Gadus spp.</i> | ● | ● |
| F11 | Codfish <i>Gadus spp.</i> | ● | ● |
| F12 | Crab <i>Callinectes spp.</i> | ● | ● |
| F16 | Flounder <i>Platichthys sp.</i> | ● | ● |
| F20 | Lobster <i>Homarus americanus</i> | ● | ● |
| F21 | Mackerel <i>Scomberomorus cavalla</i> | ● | ● |
| F23 | Oyster <i>Ostrea/Crassostrea</i> | ● | ● |
| F24 | Perch <i>Serranus scriba</i> | ● | ● |
| F30 | Salmon <i>Salmo salar</i> | ● | ● |
| F32 | Scallop <i>Placopecten magellanicus</i> | ● | ● |
| F34 | Shrimp, White/Brown/Pink <i>Litopaenaeus setiferus/Farfantepenaeus aztecus/Farfantepenaeus dourarum</i> | ● | ● |
| F40 | Trout, Lake <i>Oncorhynchus mykiss</i> | ● | ● |
| F41 | Tuna <i>Thunnus sp.</i> | ● | ● |
| Food Mixes | | | |
| F01 | Fish Mix F11 Codfish, F16 Flounder, F18 Halibut, F21 Mackerel, F41 Tuna | ● | ● |
| F02 | Shellfish Mix F10 Clam, F12 Crab, F23 Oyster, F32 Scallops, F34 Shrimp | ● | ● |
| Nuts | | | |
| F46 | Almond <i>Prunus dulcis</i> | ● | ● |
| F72 | Brazil Nut <i>Bertholletia excelsa</i> | ● | ● |
| F84 | Cashew Nut <i>Anacardium occidentale</i> | ● | ● |
| F98 | Coconut <i>Cocos nucifera</i> | ● | ● |
| F120 | Filbert/Hazelnut <i>Corylus americana</i> | ● | ● |
| F171 | Peanut <i>Arachis hypogaea</i> | ● | ● |
| F173 | Pecan <i>Carya illinoensis</i> | ● | ● |
| F229 | Walnut, Black <i>Juglans nigra</i> | ● | ● |
| F230 | Walnut, English <i>Juglans regia</i> | ● | ● |

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Made-to-Order Items

Specific non-stocked products have been identified as made-to-order items based on low utilization. Delivery will be a minimum of 4 to 6 weeks. Please consult your Customer Care Specialist for more information.

Premium Allergens

Specific products have been designated as premium allergens due to low utilization, decreased availability of raw materials, increased costs associated with the collection/extraction process, and/or market conditions. These items carry a surcharge. Please consult your Customer Care Specialist for more information.

Sterile Diluents

Diluents come in a variety of fills. 30 mL and 100 mL vials are overfilled to ensure that their labeled amount can be withdrawn. GREER® Sterile Diluents™ (Normal, Buffered, HSA, and 10% Glycerin Saline) contain 0.4% phenol, which acts as a preservative by inhibiting microbial growth.¹ Two neck sizes give a choice of stopper diameter. The second number after the vial size is the outer diameter of the top of the vial in millimeters.



| Item # | Fill/mL | Vial Size | Vial Seal Color | Qty/Pack |
|--|---------|-----------|-----------------|----------|
| Normal Saline | | | | |
| Also called Phenolated Saline. Water for injection (WFI) with 0.9% NaCl. | | | | |
| SD2507007 | 2.0 mL | 5-13 | Silver | 25 |
| SD2507001 | 2.0 mL | 5-20 | Silver | 25 |
| SD2507010 | 4.0 mL | 5-13 | Silver | 25 |
| SD2507041 | 4.0 mL | 5-13 | Red | 25 |
| SD2507042 | 4.0 mL | 5-13 | Blue | 25 |
| SD2507043 | 4.0 mL | 5-13 | Green | 25 |
| SD2507044 | 4.0 mL | 5-13 | Yellow | 25 |
| SD2507045 | 4.0 mL | 5-13 | Pink | 25 |
| SD2507046 | 4.0 mL | 5-13 | Purple | 25 |
| SD2507047 | 4.0 mL | 5-13 | Orange | 25 |
| SD2507020 | 4.5 mL | 5-13 | Silver | 25 |
| SD2507031 | 4.5 mL | 5-13 | Red | 25 |
| SD2507032 | 4.5 mL | 5-13 | Blue | 25 |
| SD2507033 | 4.5 mL | 5-13 | Green | 25 |
| SD2507034 | 4.5 mL | 5-13 | Yellow | 25 |
| SD2507035 | 4.5 mL | 5-13 | Pink | 25 |
| SD2507036 | 4.5 mL | 5-13 | Purple | 25 |
| SD2507028 | 4.0 mL | 5-20 | Silver | 25 |
| SD2507024 | 4.0 mL | 5-20 | Red | 25 |
| SD2507027 | 4.0 mL | 5-20 | Blue | 25 |
| SD2507025 | 4.0 mL | 5-20 | Green | 25 |
| SD2507136 | 4.0 mL | 5-20 | Yellow | 25 |
| SD2507038 | 4.0 mL | 5-20 | Pink | 25 |
| SD2507039 | 4.0 mL | 5-20 | Purple | 25 |
| SD2507037 | 4.0 mL | 5-20 | Orange | 25 |
| SD2507030 | 4.5 mL | 5-20 | Silver | 25 |
| SD2507040 | 4.5 mL | 5-20 | Red | 25 |
| SD2507060 | 4.5 mL | 5-20 | Blue | 25 |
| SD2507050 | 4.5 mL | 5-20 | Green | 25 |
| SD2507137 | 4.5 mL | 5-20 | Yellow | 25 |
| SD2507075 | 8.0 mL | 10-13 | Silver | 25 |
| SD2507131 | 8.0 mL | 10-20 | Silver | 25 |
| SD2507080 | 9.0 mL | 10-13 | Silver | 25 |
| SD2507090 | 9.0 mL | 10-20 | Silver | 25 |
| SD2507100 | 9.0 mL | 10-20 | Red | 25 |
| SD2507120 | 9.0 mL | 10-20 | Blue | 25 |
| SD2507110 | 9.0 mL | 10-20 | Green | 25 |
| SD2507138 | 9.0 mL | 10-20 | Yellow | 25 |
| SD1007139 | 24 mL | 30-20 | Silver | 10 |
| SD1007140 | 30 mL | 30-20 | Silver | 10 |
| SD07143 | 100 mL | 100-20 | Silver | 12 |

1. Cox L, Nelson H, Lockey R, et al. Allergen immunotherapy: a practice parameter third update. *J Allergy Clin Immunol*. January 2011;127(Suppl 1):S1-55.



| Item # | Fill/mL | Vial Size | Vial Seal Color | Qty/Pack |
|---|---------|-----------|-----------------|----------|
| HSA (Human Serum Albumin) Saline* | | | | |
| Normal Saline with 0.03% Human Serum Albumin added to enhance the stability of very dilute extracts. HSA helps maintain allergen potency by preventing adsorption of proteins to vial walls. ^{1,2} | | | | |
| SH2507260 | 1.8 mL | 5-20 | Silver | 25 |
| SH2507291 | 4.0 mL | 5-20 | Silver | 25 |
| SH2507292 | 4.0 mL | 5-20 | Red | 25 |
| SH2507294 | 4.0 mL | 5-20 | Blue | 25 |
| SH2507293 | 4.0 mL | 5-20 | Green | 25 |
| SH2507362 | 4.0 mL | 5-20 | Yellow | 25 |
| SH2507300 | 4.5 mL | 5-20 | Silver | 25 |
| SH2507310 | 4.5 mL | 5-20 | Red | 25 |
| SH2507330 | 4.5 mL | 5-20 | Blue | 25 |
| SH2507320 | 4.5 mL | 5-20 | Green | 25 |
| SH2507283 | 4.5 mL | 5-20 | Yellow | 25 |
| SH2507290 | 4.5 mL | 5-13 | Silver | 25 |
| SH2507301 | 4.5 mL | 5-13 | Red | 25 |
| SH2507302 | 4.5 mL | 5-13 | Blue | 25 |
| SH2507303 | 4.5 mL | 5-13 | Green | 25 |
| SH2507304 | 4.5 mL | 5-13 | Yellow | 25 |
| SH2507345 | 8.0 mL | 10-20 | Silver | 25 |
| SH2507350 | 9.0 mL | 10-13 | Silver | 25 |
| SH2507360 | 9.0 mL | 10-20 | Silver | 25 |
| SH2507361 | 9.0 mL | 10-20 | Red | 25 |
| SH2507359 | 9.0 mL | 10-20 | Blue | 25 |
| SH2507363 | 9.0 mL | 10-20 | Green | 25 |
| SH2507364 | 9.0 mL | 10-20 | Yellow | 25 |
| SH1007371 | 27 mL | 30-20 | Silver | 10 |
| SH1007370 | 30 mL | 30-20 | Silver | 10 |
| SH07380 | 100 mL | 100-20 | Silver | 12 |

*HSA Limit of Liability

If a lot of Human Serum Albumin (HSA) contained in this product is recalled by the manufacturer for any reason, GREER® hereby states our limit of liability to that of replacing the unused vials within the carton, and any unopened cartons of HSA diluent affected by the recall.

1. Cox L, Nelson H, Lockey R, et al. Allergen immunotherapy: a practice parameter third update. *J Allergy Clin Immunol*. January 2011;127(Suppl 1):S1-55.
 2. Nelson MR, Cox L. Allergen immunotherapy extract preparation manual. American Academy of Allergy Asthma & Immunology website. <http://www.aaaai.org/Aaaaai/media/MediaLibrary/PDF%20Documents/Practice%20Management/PM%20Resource%20Guide/Ch-9-Allergen-Immunotherapy-Extract-Preparation-Manual.pdf>. Published 2012. Accessed August 20, 2013.

Sterile Diluents



| Item # | Fill/mL | Vial Size | Vial Seal Color | Qty/Pack |
|--|---------|-----------|-----------------|----------|
| 50% Glycerin Saline | | | | |
| 50% Glycerin Saline is often used to enhance the stability of extracts. 50% Glycerin Saline also helps protect the structure of allergenic proteins that may be particularly susceptible to degradation in solution and may help protect against the effects of prolonged exposure to room temperature. ¹ | | | | |
| SA5007212 | 100 mL | 100-20 | Silver | 12 |
| 10% Glycerin Saline | | | | |
| Normal Saline with 10% glycerin is often used as an alternative to HSA to enhance the stability of very dilute extracts. (Not recommended for intradermal testing.) ² | | | | |
| SG2507183 | 4.0 mL | 5-13 | Silver | 25 |
| SG2507208 | 4.5 mL | 5-13 | Silver | 25 |
| SG2507174 | 4.5 mL | 5-13 | Red | 25 |
| SG2507175 | 4.5 mL | 5-13 | Blue | 25 |
| SG2507176 | 4.5 mL | 5-13 | Green | 25 |
| SG2507177 | 4.5 mL | 5-13 | Yellow | 25 |
| SG2507209 | 4.0 mL | 5-20 | Silver | 25 |
| SG2507207 | 4.0 mL | 5-20 | Red | 25 |
| SG2507198 | 4.0 mL | 5-20 | Blue | 25 |
| SG2507197 | 4.0 mL | 5-20 | Green | 25 |
| SG2507229 | 4.0 mL | 5-20 | Yellow | 25 |
| SG2507236 | 4.0 mL | 5-20 | Pink | 25 |
| SG2507237 | 4.0 mL | 5-20 | Purple | 25 |
| SG2507238 | 4.0 mL | 5-20 | Orange | 25 |
| SG2507200 | 4.5 mL | 5-20 | Silver | 25 |
| SG2507201 | 4.5 mL | 5-20 | Red | 25 |
| SG2507203 | 4.5 mL | 5-20 | Blue | 25 |
| SG2507202 | 4.5 mL | 5-20 | Green | 25 |
| SG2507230 | 4.5 mL | 5-20 | Yellow | 25 |
| SG2507211 | 8.0 mL | 10-13 | Silver | 25 |
| SG2507181 | 8.0 mL | 10-20 | Silver | 25 |
| SG2507228 | 9.0 mL | 10-13 | Silver | 25 |
| SG2507221 | 9.0 mL | 10-13 | Red | 25 |
| SG2507222 | 9.0 mL | 10-13 | Blue | 25 |
| SG2507223 | 9.0 mL | 10-13 | Green | 25 |
| SG2507224 | 9.0 mL | 10-13 | Yellow | 25 |
| SG2507191 | 9.0 mL | 10-20 | Silver | 25 |
| SG2507192 | 9.0 mL | 10-20 | Red | 25 |
| SG2507194 | 9.0 mL | 10-20 | Blue | 25 |
| SG2507193 | 9.0 mL | 10-20 | Green | 25 |
| SG2507231 | 9.0 mL | 10-20 | Yellow | 25 |
| SG1007210 | 30 mL | 30-20 | Silver | 10 |
| SG07213 | 100 mL | 100-20 | Silver | 12 |

1. Nelson MR. Joint Council of Allergy, Asthma and Immunology. Physician's instruction guide for the preparation of allergen extract: <http://www.aaaai.org/aaaai/media/MediaLibrary/PDF%20Documents/Practice%20Management/PM%20Resource%20Guide/Ch-9-Allergen-Immunotherapy-Extract-Preparation-Manual.pdf>. Published February 20, 2009. Accessed March 22, 2012.

2. Nelson HS. Effect of preservatives and conditions of storage on the potency of allergy extracts. *J Allergy Clin Immunol*. January 1991;67(1):64-69.



| Item # | Fill/mL | Vial Size | Vial Seal Color | Qty/Pack |
|---|---------|-----------|-----------------|----------|
| Buffered Saline | | | | |
| Buffered Saline maintains extract solutions at physiological pH levels. ¹ Contains sodium and potassium phosphates with 0.5% NaCl for isotonicity, in water for injection (WFI). | | | | |
| SB2507145 | 4.5 mL | 5-20 | Silver | 25 |
| SB2507146 | 4.5 mL | 5-20 | Red | 25 |
| SB2507147 | 4.5 mL | 5-20 | Blue | 25 |
| SB2507148 | 4.5 mL | 5-20 | Yellow | 25 |
| SB2507149 | 4.5 mL | 5-20 | Green | 25 |
| SB2507156 | 9.0 mL | 10-20 | Silver | 25 |
| SB07150 | 100 mL | 100-20 | Silver | 12 |

¹ Fasman GE. Measurement of pH: buffer solutions. *Practical Handbook of Biochemistry and Molecular Biology*, Boca Raton, FL: CRC Press; 1989: 544-549.

Sterile Empty Vials

Manufactured by GREER® in a wide range of convenient sizes.



| Item # | Vial Size | Vial Seal Color | Qty/Pack |
|-----------|-----------|-----------------|----------|
| SE2508020 | 2-13 | Silver | 25 |
| SE2508030 | 5-13 | Silver | 25 |
| SE2508031 | 5-13 | Red | 25 |
| SE2508032 | 5-13 | Blue | 25 |
| SE2508033 | 5-13 | Green | 25 |
| SE2508034 | 5-13 | Yellow | 25 |
| SE2508040 | 5-20 | Silver | 25 |
| SE2508050 | 5-20 | Red | 25 |
| SE2508060 | 5-20 | Green | 25 |
| SE2508070 | 5-20 | Blue | 25 |
| SE2508081 | 5-20 | Orange | 25 |
| SE2508082 | 5-20 | Pink | 25 |
| SE2508083 | 5-20 | Purple | 25 |
| SE2508084 | 5-20 | Yellow | 25 |
| SE2508090 | 10-13 | Silver | 25 |
| SE2508091 | 10-13 | Red | 25 |
| SE2508092 | 10-13 | Blue | 25 |
| SE2508093 | 10-13 | Green | 25 |
| SE2508094 | 10-13 | Yellow | 25 |
| SE2508100 | 10-20 | Silver | 25 |
| SE2508110 | 10-20 | Red | 25 |
| SE2508120 | 10-20 | Green | 25 |
| SE2508130 | 10-20 | Blue | 25 |
| SE2508150 | 10-20 | Yellow | 25 |
| SE1008150 | 30-20 | Silver | 10 |
| SE08155 | 50-20 | Silver | 12 |
| SE08160 | 100-20 | Silver | 12 |

Plastic Colored Caps

Snap-on color-coded caps can be used to differentiate strengths of treatment extracts.



| 13 mm Item # | Cap Color | Qty/Pack |
|--------------|-----------|----------|
| PCYEL13 | Yellow | 25 |
| PCORG13 | Orange | 25 |
| PCRED13 | Red | 25 |
| PCGRN13 | Green | 25 |
| PCPNK13 | Pink | 25 |
| PCPUR13 | Purple | 25 |
| PCBLU13 | Blue | 25 |
| 20 mm Item # | Cap Color | Qty/Pack |
| PCYEL20 | Yellow | 25 |
| PCORG20 | Orange | 25 |
| PCRED20 | Red | 25 |
| PCGRN20 | Green | 25 |
| PCWHT20 | White | 25 |
| PCPNK20 | Pink | 25 |
| PCPUR20 | Purple | 25 |
| PCBLU20 | Blue | 25 |

Dropper Vials



| Item # | Description | Qty/Pack |
|--------|---|----------|
| 5NSEVA | Nonsterile 5 mL vial with dropper cap, screw thread Type 1 glass | 25 |
| 10NSEV | Nonsterile 10 mL vial with dropper cap, screw thread Type 1 glass | 25 |

Safety Syringes



Photo courtesy of Magellan™

| Item # | Description |
|--|--|
| Magellan™ Tuberculin Safety Syringe Trays | |
| MAS27T | 1 cc 27 g x 1/2" regular bevel, 20 syringes/tray, 40 trays (800/case) |
| Becton-Dickinson | |
| BD5553 | Safety-Lok™ 1 cc 27 g x 1/2" regular bevel, individually wrapped (500/case) |
| BD5950 | SafetyGlide™ 1 cc 27 g x 1/2" regular bevel, 25 syringes/tray, 40 trays (1,000/case) |
| BD5951 | SafetyGlide™ 1 cc 26 g x 3/8" intradermal bevel, 25 syringes/tray, 40 trays (1,000/case) |

Syringes and Syringe Trays



| Item # | Description |
|---|--|
| Becton-Dickinson | |
| Cases consist of 25 syringes/tray, 40 trays (1,000/case). | |
| BD5535 | 1/2 cc 27 g x 1/2" regular bevel |
| BD5536 | 1/2 cc 27 g x 3/8" intradermal bevel |
| BD5537 | 1 cc 26 g x 1/2" regular bevel |
| BD5538 | 1 cc 26 g x 1/2" intradermal bevel |
| BD5539 | 1 cc 26 g x 3/8" intradermal bevel |
| BD5540 | 1 cc 27 g x 1/2" regular bevel |
| BD5541 | 1 cc 27 g x 3/8" intradermal bevel |
| BD5542 | 1 cc 27 g x 3/8" regular bevel |
| Terumo | |
| Cases consist of 25 syringes/tray, 40 trays (1,000/case). | |
| TB26T1 | 1 cc 26 g x 3/8" regular bevel |
| TB27T1 | 1 cc 27 g x 1/2" regular bevel |
| TB28T1 | 1 cc 27 g x 3/8" intradermal bevel |
| TB07T1 | 1 cc 27 g x 1/2" intradermal bevel |
| Syringe Trays | |
| RACHST | Rachman Syringe Tray: composed of sturdy, heavy-gauge ABS plastic that holds 18 syringes per tray. Measures 14 1/16" L x 9 1/4" W x 1 3/4" D |
| GRS025 | Plastic Syringe Tray Sets: 5 sets/pack, 2-layer tray, holds 25 syringes |
| GRSS025 | Safety Syringe Organizer Tray: 5 sets/pack, 2-layer tray, holds 25 syringes |

Ancillary Products



| Item # | Vial Size Type | Holds |
|--|--------------------------|---------|
| Mailing Containers | | |
| Inner compartments (trays) slide into outer cardboard cartons (boxes) for quick packaging. Sturdy material designed to help protect vials from breakage during transportation. | | |
| 5133B | 5 mL (13 mm neck) | 3 vials |
| 5133T | 5 mL (13 mm neck) | 3 vials |
| 5136B | 5 mL (13 mm neck) | 6 vials |
| 5136T | 5 mL (13 mm neck) | 6 vials |
| 5203B | 5mL (20mm neck) | 3 vials |
| 5203T | 5mL (20mm neck) | 3 vials |
| 5206B | 5mL (20mm neck) | 6 vials |
| 5206T | 5mL (20mm neck) | 6 vials |
| 10133B | 10 mL (13 mm neck) | 3 vials |
| 10133T | 10 mL (13 mm neck) | 3 vials |
| 10136B | 10 mL (13 mm neck) | 6 vials |
| 10136T | 10 mL (13 mm neck) | 6 vials |
| 5203C-100* | 5 and 10 mL (20 mm neck) | 3 vials |
| 5206C-100* | 5 and 10 mL (20 mm neck) | 6 vials |

Note: B in item number indicates boxes
 Note: T in item number indicates trays
 *These items include both boxes and trays



| Item # | Description |
|--|--|
| Styrofoam Containers | |
| Sturdy molded styrofoam conforms to each vial allowing protection against breakage and provides additional insulation when exposed to heat or cold. Lightweight design makes shipping easy and economical. | |
| STY1C | Holds any of our 5 mL or 10 mL vials or two 2 mL vials (250 case) |
| STY5C | Holds up to five 5 mL or 10 mL vials with space for instruction sheet (200 case) |



| Item # | Size | Qty/Case |
|--------|------|----------|
|--------|------|----------|

Jiffy™ Mailing Bags

Lightweight mailers are padded for added protection of extract vials. These versatile mailers allow for easy shipping of different materials.

| | | |
|------|--------------|-----|
| JB0C | 6" x 10" | 250 |
| JB1C | 7 1/4" x 12" | 100 |
| JB2C | 8 1/2" x 12" | 100 |

| Item # | Description | Qty |
|--------|-------------|-----|
|--------|-------------|-----|

Sharps Collectors

OSHA-approved containers for contaminated waste. Available in multiple sizes: 8.2 quart, 5 gallon, and 6 gallon.

| | | |
|--------|---|-------------|
| BD5490 | Sharps Collector, 8.2 quarts | 12 per case |
| BD5491 | Sharps Collector, 5 gallons | 8 per case |
| BD5465 | Sharps Collector, 6 gallons, funnel top | 12 per case |

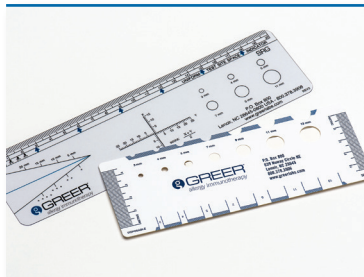
| Item # | Color | Size | Quantity |
|--------|-------|------|----------|
|--------|-------|------|----------|

Stock and Custom-Printed Labels

Pressure-sensitive labels adhere quickly and firmly to vials. Stock labels include space for vial labeling information recommended by allergy immunotherapy practice parameter.¹ For information about custom-printed labels, please contact your Customer Care Specialist.

| | | | |
|---------|--------|-------------|------------|
| L-PVLR | Red | 1.625" x 1" | 1,000/roll |
| L-PVLB | Blue | 1.625" x 1" | 1,000/roll |
| L-PVLG | Green | 1.625" x 1" | 1,000/roll |
| L-PVLY | Yellow | 1.625" x 1" | 1,000/roll |
| L-PVLBK | Black | 1.625" x 1" | 1,000/roll |

¹ Cox L, Nelson H, Lockey R, et al. Allergen immunotherapy: a practice parameter third update. *J Allergy Clin Immunol*. January 2011;127(Suppl 1):S1-S55.



| Item # | Description |
|--------|-------------|
|--------|-------------|

Skin Reaction Guides

Skin reaction guides help clinicians accurately measure the wheal and erythema response following percutaneous skin testing. Both plastic reusable and paper disposable skin reaction guides are available.

| | |
|------|--|
| SRG | Clear Plastic (1 pack contains 12) |
| SRG3 | Disposable Paper (1 pack contains 100) |

Vial Racks

GREER® Versa Vial Rack®

This one-color design features four easy-grip handles and tiered wells to hold multiple vial sizes. Racks are stackable with special drains in every well for quick cleanup. Three different base sizes allow for a better fit in most refrigerators.

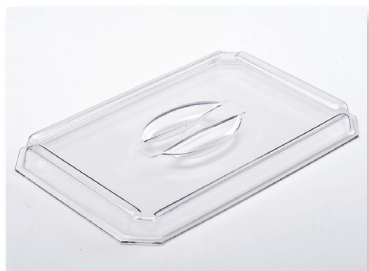
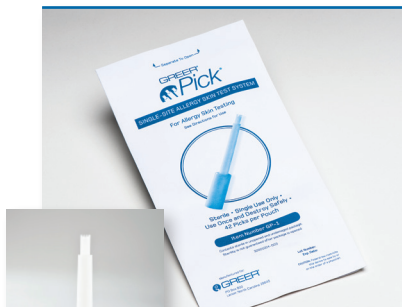


| Item # | Style | Vial Capacity | To Hold | Size |
|---------|--------------------------------------|--------------------|---|------------------|
| VRB001 | Bulk Versa Vial Rack | 39 | 50-20 vials 30-20 vials 10-13 vials | 9 1/8" x 13 1/8" |
| VRT002 | Testing Versa Vial Rack | 48 plus 2 controls | 5 mL scratch vials 5-13 vials Labeled negative and positive controls in two sizes | 8" x 10" |
| VRLT005 | Testing Versa Vial Rack (large) | 72 plus 2 controls | 5 mL scratch vials 5-13 vials Labeled negative and positive controls in two sizes | 8" x 14.5" |
| VRP003 | Prescription Versa Vial Rack | 48 | 5-20 vials 10-13 vials 10-20 vials | 8" x 10" |
| VRLP004 | Prescription Versa Vial Rack (large) | 72 | 5-20 vials 10-13 vials 10-20 vials | 8" x 14.5" |

Skin Testing Devices

The GREER® Pick® System

Versatile. Convenient. Easy to Use. GREER® Pick®, a self-loading, 6-tine, single site device, can average 1,800 tests from one 5 mL vial when the GREER® Pick® Well™ is filled with 0.125 mL of glycerinated extract.¹



For more information on GREER® Pick®, see accompanying directions for use in the pocket inside the back cover or visit www.greerlabs.com.

| Item # | Description |
|--|-------------------------|
| GREER® Pick® Packaged in 50 sterile pouches each containing 42 picks; 2,100 picks per case. | |
| GP-1C | 2,100 picks per case |
| GREER® Pick® Well™ Packaged in sterile pouches each containing 24 wells. | |
| GW-2 | 24 wells per pouch |
| GREER® Pick® Tray™ The clear GREER® Pick® Tray™ allows for easy inspection of extract levels and the closed-well system maintains extract integrity and provides for spill-free transporting. Each stackable tray can be labeled for easy identification. The GREER Pick Tray is available as a 60-well or 40-well tray. Upon request, customized, laminated color labels are available for all trays at no additional cost. | |
| GY-5 | Holds 60 wells per tray |
| GY-40 | Holds 40 wells per tray |
| GREER® Pick® Tray™ Lid The convenient tabbed lid helps pop the top effortlessly and provides spill-free transporting. | |
| GY-LD | 60-Well Tray Lid |

1. GREER. Data on file. Evaluation of Allergenic Extract Consumption for the GREER® Pick® and GREER® Pick® Well™ Skin Test Device Components. June 10, 2004.

Skin Testing Devices



(Limit 3 packages per customer.)



(Limit 1 package per customer.)

| Item # | Description |
|--|---|
| 40-Well GREER® Pick® Evaluation Package | |
| GP-10 | 40-Well GREER® Pick® Tray™ (bottom component) |
| | GREER Pick Tray Lid (lid component) |
| | Labels (1 blank set of 40) |
| | GREER Pick , 420 picks (10 pouches) |
| | GREER® Pick® Well™, 48 wells (2 pouches) |
| | GREER Pick Skin Testing Instructional DVD |
| | Skin Reaction Guide, Disposable Paper |
| | Sample GREER Pick Skin Testing Sheet |
| | GREER Pick Practical Guide |
| 60-Well GREER® Pick® Evaluation Package | |
| GP-11 | 60-Well GREER Pick Tray |
| | Labels (1 blank set of 60) |
| | GREER Pick, 630 picks (15 pouches) |
| | GREER Pick Well, 72 wells (3 pouches) |
| | GREER Pick Skin Testing Instructional DVD |
| | Skin Reaction Guide, Disposable Paper |
| | Sample GREER Pick Skin Testing Sheet |
| | GREER Pick Practical Guide |

For more information on GREER® Pick®, see accompanying directions for use in the pocket inside the back cover or visit www.greerlabs.com.



Skintestor OMNI™ is manufactured by QTI Corporation and distributed by GREER®.

| Item # | Description |
|--------|-------------|
|--------|-------------|

Skintestor OMNI™

The Skintestor OMNI™ is a self-loading, 10-head multiple-site skin testing device that offers flexibility and efficiency. When the wells of the Skintestor OMNI tray are filled with 0.250 mL of extract, an average of 1,031 tests can be performed from one 5 mL glycerinated extract vial.¹ The versatile design of the Skintestor OMNI allows for simple use, whether for skin testing on a patient's arm or back.

One shelf pack contains 10 sterile Skintestor OMNI devices. There are 10 shelf packs or 100 devices per box. Upon request, customized, laminated color labels are available for all trays at no additional cost.

¹ GREER. Data on file. DS043010: Comparisons of uptake of 50% glycerosaline by Skintestor OMNI devices using wells loaded with 0.250 mL and 0.500 mL fills at ambient and refrigerator temperatures. April 2010.

| | |
|--------|---|
| OM203C | 10 shelf packs per box; 100 devices per box |
|--------|---|

Skintestor OMNI™ Trays

The Skintestor OMNI System includes the Skintestor OMNI self-loading, multiple-site skin testing device for use in either 20- or 40- well labeled trays. Three 20-well trays fit into a carrier tray to create a 60-well option. The Skintestor OMNI device fits securely over its wells to maintain the extract integrity and shields the extract from evaporation.

| | |
|--------|---|
| OMT1 | Skintestor OMNI Tray (1 sterile 20-well tray) |
| OM303C | Skintestor OMNI Tray Pouch (5 individually wrapped sterile 20-well trays) |
| OM503 | Skintestor OMNI Carrier Tray (holds 3 sterile 20-well trays) |
| OM403C | Skintestor OMNI Tray (1 sterile 40-well tray) |

40-Well Skintestor OMNI™ Evaluation Package

| | |
|-------|--|
| OM-40 | 1 Skintestor OMNI Shelf Pack |
| | 1 Skintestor OMNI 40-Well Tray |
| | Skintestor OMNI Skin Testing Instructional DVD |
| | Skin Reaction Guide, Disposable Paper |
| | Sample Skintestor OMNI Skin Testing Sheet |
| | Skintestor OMNI Practical Guide |

60-Well Skintestor OMNI™ Evaluation Package

| | |
|-------|--|
| OM-EV | 1 Skintestor OMNI Shelf Pack |
| | 3 Skintestor OMNI 20-Well Trays |
| | 1 Skintestor OMNI Carrier Tray |
| | Skintestor OMNI Skin Testing Instructional DVD |
| | Skin Reaction Guide, Disposable Paper |
| | Sample Skintestor OMNI Skin Testing Sheet |
| | Skintestor OMNI Practical Guide |

GREER® Service Team



Your GREER® Service Team is devoted to providing continuous support for your practice. This team is comprised of:

- an **Allergy Sales Consultant**, an expert in assessing your allergy immunotherapy needs and helping you find the right solutions;
- the **Medical & Scientific Affairs Department**, an assembly of specialized individuals including physicians, nurse practitioners, medical sciences liaisons, and research scientists who assist your clinic with technical information and answer any medically-related questions; and
- **Customer Care Specialists**, knowledgeable, friendly, responsive professionals who are dedicated to your overall satisfaction.

With your GREER Service Team, you have GREER professionals ready to help you and your practice.

GREER. In Touch. Within Reach.®

The GREER® Service Team

Allergy Sales
Consultant



Customer
Care
Specialist



Medical &
Scientific Affairs



In Touch. Within Reach.®



Allergenic extracts are indicated for skin test diagnosis and treatment (immunotherapy) of patients with seasonal and perennial allergies.

IMPORTANT SAFETY INFORMATION:

- Do not inject intravenously.
- **Allergenic extracts may cause severe life-threatening systemic reactions, including the rare occurrence of anaphylaxis or death.** Systemic reactions include: generalized skin erythema, urticaria, pruritus, angioedema, rhinitis, wheezing, laryngeal edema, and hypotension. Other adverse reactions include: nausea, emesis, abdominal cramps, and diarrhea.
- Intended for use only by physicians who are experienced in the administration of allergenic extracts. Initial dose must be based on skin test.
- Observe patients in the office for at least 30 minutes following treatment. Emergency measures and personnel trained in their use must be available immediately in the event of a life-threatening reaction. Immunotherapy may not be suitable for patients with medical conditions that reduce their ability to survive a systemic reaction.

Please see Package Inserts for full prescribing information including **BOXED WARNING** or go to www.greerlabs.com/index.php/human_allergy/package_inserts/.



GREER Human Allergy

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Lenoir, NC 28645
800.378.3906
www.greerlabs.com

ALLERGENIC EXTRACTS

Standardized Grass Pollen Extracts

U.S. Government License No. 308



PO Box 800
Lenoir, NC 28645
USA

Revised 10/04

WARNING

THIS ALLERGENIC PRODUCT IS INTENDED FOR USE BY PHYSICIANS WHO ARE EXPERIENCED IN THE ADMINISTRATION OF ALLERGENIC EXTRACTS AND THE EMERGENCY CARE OF ANAPHYLAXIS, OR FOR USE UNDER THE GUIDANCE OF AN ALLERGY SPECIALIST.

THIS PRODUCT SHOULD NOT BE INJECTED INTRAVENOUSLY.

STANDARDIZED GRASS POLLEN EXTRACTS LABELED IN BIOEQUIVALENT ALLERGY UNITS (BAU)/ML ARE NOT INTERCHANGEABLE WITH GRASS POLLEN EXTRACTS LABELED IN ALLERGY UNITS (AU)/ML OR WITH NONSTANDARDIZED GRASS POLLEN EXTRACTS. FOR PREVIOUSLY UNTREATED PATIENTS OR PATIENTS PREVIOUSLY RECEIVING EXTRACTS FROM ANOTHER MANUFACTURER, THE INITIAL DOSE MUST BE BASED ON SKIN TESTING AS DESCRIBED IN THE DOSAGE AND ADMINISTRATION SECTION OF THIS INSERT. PATIENTS BEING SWITCHED FROM OTHER TYPES OF EXTRACTS TO STANDARDIZED EXTRACTS SHOULD BE INSTRUCTED TO RECOGNIZE ADVERSE REACTION SYMPTOMS AND CAUTIONED TO CONTACT THE PHYSICIAN'S OFFICE IF REACTION SYMPTOMS OCCUR. IN CERTAIN INDIVIDUALS THESE REACTIONS COULD BE FATAL. PATIENTS SHOULD BE OBSERVED FOR AT LEAST 20 MINUTES FOLLOWING TREATMENT. PATIENTS WITH LABILE OR STEROID-DEPENDENT ASTHMA ARE "HIGH RISK PATIENTS" WHO REQUIRE SPECIAL CAUTION IN DOSE ADMINISTRATION AND SHOULD REMAIN IN THE OFFICE FOR AT LEAST 30 MINUTES. AIRWAY OBSTRUCTION IN HIGH RISK PATIENTS CAN BE MONITORED BY PEAK FLOW MEASUREMENTS BEFORE AND AFTER ADMINISTRATION OF ALLERGENS. EMERGENCY MEASURES AS WELL AS PERSONNEL TRAINED IN THEIR USE SHOULD BE IMMEDIATELY AVAILABLE IN THE EVENT OF A LIFE THREATENING REACTION. TO REPORT SERIOUS ADVERSE EVENTS, THE FOOD AND DRUG ADMINISTRATION MED-WATCH NUMBER IS 1-800-332-1088. PATIENTS BEING SWITCHED FROM ONE LOT OF EXTRACT TO ANOTHER FROM THE SAME MANUFACTURER SHOULD HAVE THEIR DOSE REDUCED BY 75%.

RISK OF ANAPHYLAXIS SHOULD BE WEIGHED AGAINST BENEFITS: IN PATIENTS RECEIVING BETA BLOCKERS AS THEY MAY NOT BE RESPONSIVE TO BETA ADRENERGIC DRUGS SHOULD ANAPHYLAXIS OCCUR; IN PATIENTS WITH UNSTABLE OR STEROID-DEPENDENT ASTHMA; OR IN PATIENTS WITH CARDIOVASCULAR DISEASE.

REFER ALSO TO THE WARNINGS, PRECAUTIONS, ADVERSE REACTIONS AND OVERDOSAGE SECTIONS BELOW.

DESCRIPTION

Standardized Grass Pollen Allergenic Extracts are supplied as sterile solutions for intracutaneous or subcutaneous administration. Standardized Grass Pollen Allergenic Extracts include Bermuda (*Cynodon dactylon*), Kentucky Blue (June), (*Poa pratensis*), Meadow Fescue (*Festuca elatior*), Orchard (*Dactylis glomerata*), Perennial Rye (*Lolium perenne*), Redtop (*Agrostis alba*), Sweet Vernal (*Anthoxanthum odoratum*), and Timothy (*Phleum pratense*). Glycerinated concentrates contain the soluble extractants of the source material with 0.25% sodium chloride, 0.27% sodium bicarbonate, and 50% glycerin v/v. All extracts contain 0.4% phenol as the preservative. Source materials for each extract are the specific pollens collected from the respective plants.

Standardized Grass Pollen Extracts are labeled in Bioequivalent Allergy Units (BAU)/mL. STANDARDIZED GRASS POLLEN EXTRACTS LABELED IN BAU/ML ARE NOT INTERCHANGEABLE WITH GRASS POLLEN EXTRACTS LABELED IN AU/ML OR WITH NONSTANDARDIZED GRASS POLLEN EXTRACTS. Bioequivalent allergy units are assigned based on comparison by enzyme linked immunosorbent assay (ELISA) to references from the U. S. Food and Drug Administration, Center for Biologics Evaluation and Research (CBER). CBER References are assigned unitage based on quantitative skin testing.¹⁻⁴ CBER references which can be diluted 1:5,000,000 to intradermally elicit a 50 mm sum of erythema diameter response in highly puncture reactive subjects are assigned 100,000 BAU/mL, whereas references diluted 1:500,000 which elicit the same 50 mm sum of erythema diameter response are assigned 10,000 BAU/mL.

CLINICAL PHARMACOLOGY

The allergic reaction is dependent upon the presence of antigen-specific immunoglobulin E (IgE) antibodies that are bound to specific receptors on mast cells and basophils. The presence of IgE antibodies on mast cells and basophils sensitizes these cells and upon interaction with the appropriate allergen-histamine and other mediators are released. IgE antibody has been shown to correlate with atopic diseases such as allergic rhinitis and allergic asthma.⁵⁻⁸ In the skin these mediators are responsible for the characteristic wheal and flare (erythema) reactions upon allergenic extract skin testing in persons with the specific allergies.⁷⁻¹¹

Puncture test results with eight US reference extracts at 10,000 BAU/mL (15 grass-specific allergic subjects per extract) are shown in TABLE 1.³⁰ For the eight grass pollens, there was a mean sum-of-diameter wheal of 15.2 mm (SD = 1.8) and a mean sum-of-diameter erythema of 84.0 mm (SD = 5.8).

TABLE 1
Puncture Skin Tests with 10,000 BAU/mL Grass Extracts
(Bifurcated Needle)

| Reference Pollen | Puncture Sum of Wheal (mm) | | Puncture Sum of Erythema (mm) | |
|---------------------|-------------------------------|--------|----------------------------------|----------|
| | Mean | Range | Mean | Range |
| Bermuda | 15.7 | 7 - 31 | 90.3 | 43 - 123 |
| Kentucky Blue/June | 15.9 | 6 - 28 | 77.3 | 47 - 107 |
| Meadow Fescue | 11.9 | 7 - 22 | 81.1 | 57 - 115 |
| Orchard | 14.1 | 9 - 19 | 84.3 | 57 - 111 |
| Perennial Rye | 17.5 | 6 - 36 | 92.3 | 73 - 135 |
| Redtop | 14.1 | 8 - 19 | 77.1 | 42 - 98 |
| Sweet Vernal | 15.7 | 8 - 30 | 81.2 | 28 - 123 |
| Timothy | 16.9 | 8 - 40 | 88.3 | 51 - 109 |

Intradermal skin tests with eight U.S. reference extracts (TABLE 2) in highly puncture reactive subjects (TABLE 1) indicate that a calculated dose of 0.02 BAU/mL should yield an average sum of erythema reaction of 50 mm, as tested in subjects sensitive to the specific grass pollen extract. However in the more sensitive subjects, the dose was as low as 0.0003 BAU/mL for one grass to 0.002 BAU/mL for several others. Conversely, doses of from 0.1 to 1.9 BAU/mL were calculated to yield the same reaction in the least-sensitive subjects.

TABLE 2
Intradermal Skin Test Doses
(Calculated BAU/mL Required for 50 mm Sum of Erythema)

| Reference Pollen | Bioequivalent Allergy Units/mL | | | |
|---------------------|-----------------------------------|--------|---|-----|
| | Mean | Range | | |
| Bermuda | 0.02 | 0.0003 | - | 0.4 |
| Kentucky Blue/June | 0.02 | 0.004 | - | 0.1 |
| Meadow Fescue | 0.02 | 0.002 | - | 0.9 |
| Orchard | 0.02 | 0.002 | - | 1.9 |
| Perennial Rye | 0.02 | 0.002 | - | 0.7 |
| Redtop | 0.02 | 0.004 | - | 0.8 |
| Sweet Vernal | 0.02 | 0.002 | - | 1.0 |
| Timothy | 0.02 | 0.002 | - | 0.6 |

Specific immunotherapy with pollen extracts as employed for many years is helpful in reducing symptoms associated with exposure to the offending allergens. A summary of effectiveness by the Panel on Review of Allergenic Extracts, an advisory committee to the U.S. Food and Drug Administration, has been published.¹² Several mechanisms have been proposed to explain the effectiveness of immunotherapy: an increase in antigen-specific IgG antibodies is frequently associated with clinical effectiveness, although correlation is not

consistent in all studies; there is a decrease in specific IgE; and IgE production is suppressed during periods of seasonal or high exposure to the antigen.¹³ Other changes following immunotherapy have been noted including development of auto-anti-idiotypic antibodies, a decrease in blood basophil sensitivity to allergen, a decrease in lymphokine production and lymphocyte proliferation by cells exposed to allergen, and development of allergen-specific suppressor cells.¹⁴ The complete mechanisms of immunotherapy are not known and remain the subject of investigation.

Standardized versus nonstandardized extracts: Standardized grass pollen extracts cannot be directly compared to the previously marketed nonstandardized extract concentrates of the same grass pollens such as those labeled at 1:10 w/v or 1:20 w/v or at 20,000 to 40,000 PNU. The potency of the nonstandardized extracts vary from species to species. Some nonstandardized grass pollen concentrates have been from a few thousand BAU/mL to several hundred thousand BAU/mL as measured by in vitro ELISA testing. Extracts of some lots of Greer nonstandardized glycerinated 1:20 w/v extracts such as Meadow Fescue and Redtop have tested over 200,000 BAU/mL. Two Timothy aqueous nonstandardized 1:10 w/v aqueous extract lots were over 200,000 BAU/mL. Several lots of nonstandardized concentrates of Kentucky Blue/June, Orchard, Perennial Rye, and Sweet Vernal varied around 100,000 BAU/mL. Bermuda grass is not as potent. The FDA Bermuda reference is assigned 10,000 BAU/mL, a value similar to that found in several Greer lots of nonstandardized Bermuda. This is the maximum available strength of standardized Bermuda grass pollen extract. See TABLE 3 for examples of BAU potency by in vitro ELISA testing for nonstandardized grass pollen extracts.

TABLE 3
BAU/mL of Previously Marketed, Nonstandardized,
Grass Pollen Extracts BAU/mL Range by In Vitro ELISA*

| Pollen | # of Lots Tested | 1:10 w/v Aqueous | # of Lots Tested | 1:20 w/v |
|--------------------|------------------|--------------------|------------------|--------------------|
| | | | | glycerinated |
| Bermuda | 1 | 10,740 | 5 | 4,000 to 14,500 |
| Meadow Fescue | 3 | 287,300 to 666,000 | 4 | 169,200 to 378,200 |
| Kentucky Blue/June | 3 | 56,100 to 145,400 | 4 | 56,100 to 91,500 |
| Orchard | 2 | 134,000 to 139,200 | 5 | 71,200 to 110,500 |
| Redtop | 3 | 141,900 to 425,000 | 4 | 134,600 to 219,200 |
| Perennial Rye | 4 | 59,100 to 302,000 | 4 | 52,900 to 80,400 |
| Sweet Vernal | 2 | 171,900 to 234,800 | 5 | 63,900 to 201,200 |
| Timothy | 3 | 186,300 to 291,000 | 3 | 63,000 to 104,800 |

Extracts testing between 67,300 and 148,600 are not statistically different from 100,000 BAU/mL. Extracts which test between 6,730 to 14,860 are not statistically different from 10,000 BAU/mL.

***CAUTION:** Only a few lots of each nonstandardized pollen species have been tested by ELISA. The lots tested varied from fresh extracts to extracts more than three years old. Do not assume that these values apply to specific lots that are in distribution. In addition to age, storage temperatures influence potency.

Physicians must exercise care in switching patients from nonstandardized to standardized extracts. As with nonstandardized extracts, dosage with BAU extracts must be derived based on the patient's sensitivity to the specific pollen. Switching from an extract that was not standardized in BAU cannot be made by a calculated, numerical ratio, but TABLE 3 can be used as a guide. Dose selection can be confirmed by side-by-side testing of nonstandardized and standardized extracts at estimated equal doses. See WARNINGS section.

The potency of nonstandardized grass pollen extracts have varied enough so that the strength of any extract previously used in a specific patient cannot be related to a particular potency in switching to BAU extracts. Therefore, patients being switched from nonstandardized extracts from another manufacturer to extracts standardized in BAU can be reevaluated by diagnostic skin testing to judge the dose to start immunotherapy or to build up to new maintenance dosages.

INDICATIONS AND USAGE

Standardized Grass Pollen Extracts are indicated for the skin-test diagnosis of allergy and immunotherapy treatment of patients with a history of allergy to the respective pollen. The diagnosis of IgE-mediated allergy may be established by the allergy history, clinical evaluation, and skin test reactivity.^{8,11,15} Extracts at 10,000 BAU/mL are indicated for use in scratch, prick, or puncture skin test diagnosis. Extracts at 100,000 BAU/mL are indicated for use in scratch, prick, or puncture skin test diagnosis in less sensitive subjects, such as those negative or indeterminate upon scratch, prick, or puncture testing at 10,000 BAU/mL. Extracts at 10,000 BAU/mL or 100,000 BAU/mL are indicated for intradermal skin test diagnosis only when appropriately diluted.

Immunotherapy with Standardized Grass Pollen Extracts is indicated when testing and patient history have identified the offending allergens and when it is not possible or practical to avoid these allergens.¹⁶⁻¹⁸ Extracts at 10,000 BAU/mL or 100,000 BAU/mL are indicated for immunotherapy only when appropriately diluted. 10,000 BAU/mL extracts are indicated for immunotherapy on previously untreated patients. 100,000 BAU/mL extracts are indicated if a higher dose is needed. (See DOSAGE AND ADMINISTRATION) **STANDARDIZED GRASS POLLEN EXTRACTS LABELED IN BAU/mL ARE NOT INTERCHANGEABLE WITH GRASS POLLEN EXTRACTS LABELED IN AU/mL OR WITH NONSTANDARDIZED GRASS POLLEN EXTRACTS.** The use of Standardized Grass Pollen Extracts for the above purposes should be made only by physicians with special familiarity and knowledge of allergy. (See DOSAGE AND ADMINISTRATION)

CONTRAINDICATIONS

There are no known absolute contraindications to the use of Standardized Grass Pollen Extracts for immunotherapy. Immunotherapy with specific antigens is contraindicated in those individuals who do not exhibit skin test and clinical sensitivity to the particular antigens. (See WARNINGS and PRECAUTIONS)

Allergenic extract injections should not be administered in the presence of diseases characterized by a bleeding diathesis.

Children with nephrotic syndrome require careful consideration and probably should not receive injection therapy because a variety of seemingly unrelated events, such as immunization, can cause an exacerbation of their nephrotic disease.

General contraindications include:

EXTREME SENSITIVITY TO THE SPECIFIC ALLERGEN - Determined from previous anaphylaxis following exposure.

AUTOIMMUNE DISEASE - Individuals with autoimmune disease may be at risk, due to the possibility of routine immunizations exacerbating symptoms of the underlying disease.

WARNINGS

All concentrates of Standardized Grass Pollen Extracts are manufactured to assure high potency and have the ability during skin testing and immunotherapy to cause serious local and systemic reactions including death in extremely sensitive patients. Most reactions occur within 20 minutes after injection, but may occur later.¹⁹ To minimize the potential for local or systemic reactions, the relative sensitivity of the patient must be assessed from the allergic history and from clinical observations. Patients should be informed of these risks prior to skin testing and immunotherapy. (See PRECAUTIONS and ADVERSE REACTIONS)

Concentrated extracts at 10,000 and 100,000 BAU/mL, must be diluted with a sterile diluent prior to use in a patient for intradermal testing or for immunotherapy.

Skin testing should be initiated only with 10,000 BAU/mL extracts. If several concentrated extracts at 100,000 BAU/mL are administered concurrently to a sensitive patient, the additive effects of cross-reacting allergens may cause a systemic anaphylactic reaction.

Allergenic extracts should be temporarily withheld from patients or the dose adjusted downward if any of the following conditions exist:

1. severe symptoms of rhinitis and/or asthma
2. infection or flu accompanied by fever
3. exposure to excessive amounts of clinically relevant allergen prior to a scheduled injection
4. evidence of a local or systemic reaction to the preceding extract injection during a course of immunotherapy

The dosage must be reduced: 1) when starting a patient on fresh extract; 2) when transferring a patient from another form of extract to a BAU standardized extract; or 3) when modifying dosages or components in a mixture or an individual prescription, even though the labeled strength of the old and new vials may be the same. This reduction in dosage may be necessary: 1) due to the previously used extract having lost potency during storage; 2) due to the fact that standardized extracts labeled in BAU/mL differ in potency in comparison to nonstandardized extracts of the same species (see TABLE 3); or 3) due to different patient sensitivity to different components. The amount of new extract given should not exceed 25% of the last dose given from the old vial, assuming both extracts contain comparable amounts of allergen. Any evidence of a local or generalized reaction requires a reduction in dosage during the initial stages of immunotherapy, as well as during maintenance therapy. The information about nonstandardized extracts shown in TABLE 3 may be helpful in confirming the appropriateness of the initial dose. When a patient is first being administered a standardized extract labeled in BAU/mL, the new dose can be selected based on a side-by-side comparison with the previously used nonstandardized extract. The availability of 10,000 BAU/mL and 100,000 BAU/mL doses is intended to facilitate safe switching by providing the physicians access to lower and higher dosages.

Patients receiving beta blocker drugs may not be responsive to beta adrenergic drugs used to treat anaphylaxis. The risks of anaphylaxis in these patients should be carefully weighed against the benefits of immunotherapy.

PRECAUTIONS

GENERAL

Not for intravenous use!

Systemic allergic reactions may occur as a result of immunotherapy. The risk can be minimized by adherence to a careful injection schedule, which starts with a low concentration of extract and is increased slowly. Because of the danger of serious reactions, caution is needed in testing exquisitely sensitive patients or patients with labile or steroid-dependent asthma. Review the patient's history of reactions to previous injections and adjust dosages accordingly.

The physician must be prepared to treat anaphylaxis should it occur and have the necessary drugs and equipment on hand to do so.^{20,21,31} Extracts should not be administered by the patient or other individuals who are not prepared to treat anaphylaxis should it occur.

Patients receiving allergenic extracts should be kept under observation a minimum of twenty²⁰ minutes so that any adverse reaction can be observed and properly handled.²² This time should be extended to at least 30 minutes for high-risk patients such as those with labile or steroid-dependent asthma or those suffering an exacerbation of their symptoms.³² Airway obstruction in high risk patients can be monitored by peak flow measurements before and after administration of allergens.

Check the prescription or lot number, vial number, strength, and verify the dosage schedule of the prescription for the specific patient. Only after this verification has been made should an injection be given.

A separate, sterile syringe and needle or sterile disposable unit must be used for each patient to prevent the transmission of hepatitis or other infectious agents from person to person. Needles should not be recapped and should be disposed of properly.

Do not use the same syringe for different extracts, nor for the diluent after using it for an extract.

INFORMATION FOR PATIENTS

Most serious reactions following the administration of allergenic extracts occur within 30 minutes. The patient should remain under observation for this period of time or longer if instructed by the physician. The size of any local reaction should be recorded. Large local reactions may be indicative of subsequent systemic reactions as dosages increase. The patient should be instructed to report any unusual reactions. In particular, this includes unusual swelling and/or tenderness at the injection site or reactions such as rhinorrhea, sneezing, coughing, wheezing, shortness of breath, nausea, dizziness, or faintness. Reactions may occur some time after leaving the physician's office, in which case medical attention should be sought immediately.

DRUG INTERACTIONS

Skin test diagnosis with Allergenic Extracts may result in false negative responses when used within 3-10 days of H1 -Blockers such as cetirizine, loratadine, and terfenadine. The inhibitory effect of astemizole may last up to 60 days.³³ These products suppress histamine skin test reactions and could mask a positive response. The suppressive action of other drugs should be considered and emphasizes the need for a histamine positive-control test.

Patients receiving beta blocker drugs may not be responsive to beta adrenergic drugs used to treat anaphylaxis. The risks of anaphylaxis in these patients should be carefully weighed against the benefits of immunotherapy.

CARCINOGENESIS, MUTAGENESIS, IMPAIRMENT OF FERTILITY

There is no evidence of carcinogenicity, mutagenesis or impairment of fertility in humans from Standardized Grass Pollen Extracts. No long-term studies in animals have been performed to evaluate carcinogenic potential.

PREGNANCY

TERATOGENIC EFFECTS

PREGNANCY CATEGORY C - Animal reproduction studies have not been conducted with Standardized Grass Pollen Extracts. It is also not known whether Standardized Grass Pollen Extracts can cause fetal harm when administered to a pregnant woman or whether they can affect reproduction capacity. Standardized Grass Pollen Extracts should be given to a pregnant woman only if clearly needed.

There is no evidence of adverse effects of allergenic extracts on the fetus.¹² Studies have not been performed in animals to determine whether extracts affect fertility in males or females, have teratogenic potential, or have other adverse effects on the fetus. Caution should be exercised in testing or treating pregnant females because a systemic reaction may cause an abortion as a result of uterine muscle contractions.

LABOR AND DELIVERY

There is no known information of adverse effects during labor and delivery.

NURSING MOTHERS

It is not known whether allergenic extracts or their antigens are excreted in human milk. Because many drugs are excreted in human milk, caution should be exercised when extracts are administered to a nursing woman.

PEDIATRIC AND GERIATRIC USE

Although extracts have not been studied systematically in various age groups, older children and geriatric patients appear to tolerate injections of allergenic extracts well. Children less than five years of age on extract immunotherapy may have an increased risk of a severe reaction, but respond well to skin test diagnosis.²⁹ Studies with pollenosis and asthma have been conducted in children e.g. refs. 23-25. Extract usage in children should follow the same precautions as in adults.

ADVERSE REACTIONS

Adverse systemic reactions may occur within minutes upon use of an allergenic extract to which a person has specific sensitivity. These reactions consist primarily of allergic symptoms such as generalized skin erythema, urticaria, pruritus, angioedema, rhinitis, wheezing, laryngeal edema, and hypotension. Less commonly, nausea, emesis, abdominal cramps, diarrhea and uterine contractions may occur. Systemic reactions occur with varying frequency in different clinics and are usually less than 1%. To some extent, the reaction rate is related to the type and dose of administered extract and to the degree of sensitivity of the patient. Severe reactions may cause shock and loss of consciousness. Fatalities have occurred rarely.^{12,26,27} Fatal reactions are often associated with high-risk patients such as those with labile or steroid-dependent asthma, particularly in those suffering an exacerbation of their symptoms at the time of extract administration. In general, immunotherapy with allergenic extracts is considered to be safe.²⁸ Despite all precautions occasional reactions are unavoidable.

Adverse systemic / anaphylactic reactions should be treated as follows:

- A. If the injection site is an arm, a tourniquet should be immediately applied above the site. Release the tourniquet every few minutes for a few seconds.
- B. Epinephrine 1:1000 should be injected immediately in the opposite arm in amounts of 0.3 to 0.5 mL and 0.2 mL epinephrine should be administered at the site of injection. For children below the age of 6 years, adjust the initial dose of epinephrine to 0.005 mL per pound (0.01 mL/kg) of body weight. Repeat epinephrine dosage in 15 minutes if necessary and if symptoms persist.
- C. Adverse reactions not responding to epinephrine therapy may require other measures such as the use of inhaled, parenteral bronchodilators, vasopressors, oxygen, or volume replacement therapy. Proper equipment and trained personnel should be available.^{20,21,31}

Local reactions consisting of erythema, itching, swelling, tenderness and sometimes pain may occur at the injection site. These reactions may appear within a few minutes to hours and persist for several days. Local cold applications and oral antihistamines may be effective treatment. For marked and prolonged local reactions, steroids may be helpful.

Reporting of Adverse Events

Reporting of serious or unexpected adverse events occurring after extract administration is encouraged. MedWatch Forms, FDA Form 3500, are available from FDA, 1-800-332-1088. Health-care providers also should report these events to the Regulatory Affairs adverse reaction monitor, Greer, P.O. Box 800, Lenoir NC 28645-0800 or call 1-800-438-0088.

OVERDOSAGE

Systemic reactions are uncommon after injection, but if the patient receives more extract than can be tolerated at that particular time and begins to experience immediate hypersensitivity anaphylaxis, the procedures listed under ADVERSE REACTIONS should be instituted.

Overdosage may occur because of an error in the volume of extract injected, an incorrect dilution injected, or because the patient may be exposed to airborne or environmental antigens simultaneously with injection of the same antigens. In the event of a systemic reaction occurring, the dosage schedule should be carefully reviewed and if necessary adjusted as outlined above under WARNINGS.

DOSAGE AND ADMINISTRATION

1. DIAGNOSTIC TESTING

For the patient with a suspected diagnosis of allergy to more than one antigen, initial screening skin tests should include the individual extracts. If a screening skin test with a mixture is used, a positive response should be followed by testing with the individual extracts to determine the degree of sensitivity to each and to guide in the selection of extracts and their concentration for immunotherapy if indicated. However, because a negative skin test with a mixture may not be indicative of the absence of allergy to one or more of the components due to their

dilution, testing with individual extracts is more precise. False negative responses may occur if serum levels of antihistamines remain from prior medication administration. (See PRECAUTIONS) The use of a histamine positive control is especially recommended for patients on prior medications which may decrease the histamine skin test response.

Skin tests read after 15 to 20 minutes are graded in terms of the induration (wheal) and erythema (flare) response compared to the appropriate controls. Wheal and flare sizes may be recorded by actual measurements. The largest diameter of the wheal and flare may be recorded, or the sum of the largest diameter and the orthogonal (right angle) diameter wheal or flare may be used as in the studies in TABLE 1 and TABLE 2.

Scratch or Prick-puncture Skin Testing:

For puncture, prick, or scratch skin test, the 10,000 BAU/mL strength is recommended and will detect the more sensitive patients. Inconclusive results at 10,000 BAU/mL may be followed by a puncture, prick, or scratch skin test at 100,000. At the higher concentration, some nonspecific positives may occur.

Controls for Scratch, Prick-Puncture Testing:

As a positive control, glycerinated histamine phosphate 5 mg/mL (1.8 mg/mL histamine base) or aqueous histamine phosphate 2.75 mg/mL (1 mg/mL (1:1,000 w/v) histamine base) may be used as a positive control. A 50% glycerosaline solution may be used as the negative control.

Intradermal Skin Testing:

Extracts for intradermal testing must be prepared by diluting the concentrated extract with sterile diluent (such as normal or buffered saline, or normal saline with human serum albumin).

Intradermal skin tests with eight U.S. reference extracts (TABLE 2) indicate that a calculated dose of 0.02 BAU/mL should yield an average sum of erythema reaction of 50 mm, as tested in subjects with similar puncture reactivities described in TABLE 1 to that specific grass pollen extract. However in the more sensitive subjects, the dose was as low as 0.0003 BAU for one grass to 0.002 BAU for several others. Conversely, doses of from 0.1 to 1.9 BAU were calculated to yield the same reaction in the least-sensitive subjects.

Controls for Intradermal Testing:

As a positive control, use glycerinated histamine phosphate diluted to 0.5 mg/mL (0.18 mg/mL histamine base) or aqueous histamine phosphate 0.275 mg/mL (0.1 mg/mL histamine base). As a negative control, use 0.5% to 1% glycerin in 0.9% saline.

A. Patients with a negative scratch or prick-puncture test:

Patients who do not react to a scratch or prick-puncture test should be tested intradermally, using a 26 or 27 gauge 1/4 inch needle, with 0.02 to 0.05 mL of a 50 BAU/mL extract dilution. A negative test should be followed by repeat tests using progressively stronger concentrations until significant wheal and flare reaction sizes are attained or until the maximum

recommended strength of 200 BAU/mL is reached. As a positive control, use glycerinated histamine phosphate diluted to 0.5 mg/mL (0.18 mg/mL histamine base) or aqueous histamine phosphate 0.275 mg/mL (0.1 mg/mL histamine base). As a negative control use 0.5% to 1% glycerosaline solution.

B. Patients tested only by the intradermal method:

Since highly reactive individuals may react intracutaneously at doses even smaller than indicated above, it is recommended that intradermal testing be preceded by a puncture test and the dose adjusted accordingly. Other patients suspected of being moderately allergic may be tested with an intradermal test dose of 0.02 to 0.05 mL of a 0.05 BAU/mL dilution. A negative test should be followed by repeat tests using progressively stronger concentrations until the maximum recommended strength of 200 BAU/mL is reached. As a negative control use 0.5% to 1% glycerosaline solution. As a positive control, use glycerinated histamine phosphate diluted to 0.5 mg/mL (0.18 mg/mL histamine base) or aqueous histamine phosphate 0.275 mg/mL (0.1 mg/mL histamine base).

2. THERAPY

Standardized versus Nonstandardized Extracts:

Dosage with extracts standardized in BAU must be derived from a knowledge of the patient's sensitivity to the specific pollen. Switching from an extract not standardized in BAU cannot be made by a calculated ratio. There are no equivalent dosages in bioequivalent allergy units applicable to all the grass species that can be related to previously marketed nonstandardized extracts labeled in weight-to-volume (w/v), Protein Nitrogen Units (PNU), or Allergy Units (AU). The information about nonstandardized extracts shown in TABLE 3 may be helpful in selecting the initial dose for the side-by-side skin test comparison. Patients being switched from nonstandardized extracts to extracts standardized in BAU can be reevaluated by diagnostic skin testing to judge the dose for starting immunotherapy or building up to new maintenance dosages. When a patient is first being administered a standardized extract labeled in BAU/mL, the new dose can be selected based on a side-by-side comparison with the previously used nonstandardized extract.

Immunotherapy is administered by subcutaneous injection. Dosage is individualized according to the patient's sensitivity, the clinical response, and tolerance to the extract administered during the early phases of an injection regimen. Extracts for immunotherapy must be prepared by diluting the concentrate with sterile diluent (such as normal or buffered saline, or normal saline with human serum albumin).

The initial dose of an extract in BAU should be calculated based on the puncture test reactivity. Note in TABLE 1 and TABLE 2 the puncture and intradermal skin test reactivity of sensitive subjects evaluated with the US reference extracts.

The initial dose of the extract may be as low as 0.1 mL of a 0.005 to 0.05 BAU/mL dilution (0.0005 to 0.005 BAU) (dilution 5 or 6 in TABLE 4 below) or even less for the exquisitely sensitive patient. Patients with lesser sensitivity may be started at 0.1 mL of a 0.5 to 5 BAU/mL dilution (0.05 to 0.5 BAU).

The amount of allergenic extract is increased at each injection by no more than 50% of the previous amount, and the next increment is governed by the response to the last injection. Large local reactions which persist for longer than 24 hours are generally considered an indication for repeating the previous dose or reducing the dose at the next administration. Any evidence of systemic reaction is an indication for a reduction of 75% in the subsequent dose. The upper limits of dosage in BAU have not been established. Doses larger than 0.2 mL of an extract in 50% glycerin may cause discomfort upon injection. The dosages of allergenic extracts do not vary significantly with the allergic disease under treatment.

To prepare dilutions starting from a 100,000 BAU/mL concentrate, proceed as in TABLE 4. The 50,000 BAU/mL concentrate can be made by using equal parts of the 100,000 BAU/mL extract and the sterile diluent. The ten-fold dilution series uses 0.5 mL of concentrate to 4.5 mL of sterile diluent with additional dilutions made in the same manner.

TABLE 4
Ten-Fold Dilution Series*

| Dilution | Extract | Diluent | BAU/mL | BAU/mL |
|----------|--------------------|---------|---------|--------|
| 0 | Concentrate | | 100,000 | 10,000 |
| 1 | 0.5 mL concentrate | 4.5 mL | 10,000 | 1,000 |
| 2 | 0.5 mL dilution 1 | 4.5 mL | 1,000 | 100 |
| 3 | 0.5 mL dilution 2 | 4.5 mL | 100 | 10 |
| 4 | 0.5 mL dilution 3 | 4.5 mL | 10 | 1 |
| 5 | 0.5 mL dilution 4 | 4.5 mL | 1 | 0.1 |
| 6 | 0.5 mL dilution 5 | 4.5 mL | 0.1 | 0.01 |

*Due to differences such as source material, preservative, potency dilutions, storage conditions, and length of storage, there is no common potency correlation ratio between extracts standardized in Bioequivalent Allergy Units (BAU) and: 1) standardized extracts previously labeled in Allergy Units (AU); 2) nonstandardized extracts labeled weight-to-volume (w/v); 3) nonstandardized extracts labeled in Protein Nitrogen Units (PNU); or 4) alum-precipitated extracts.

The optimal interval between doses of allergenic extracts has not been established. Injections usually are given 1 or 2 times per week until the maintenance dose is reached. The injection interval is then increased to 2 weeks, then to 3 weeks and finally to 4 weeks. If the patient does not return for 6 to 8 weeks, the dose should be reduced to 25% of the last dose. If longer than 8 weeks, a dose reduction of one, two or three dilutions may be made considering the components and the patient's sensitivity. The dosage and the interval between injections may need to be modified according to the clinical response of the patient. When switching patients to fresh extract, the initial dose should be reduced to 25% of the previous dose.

The usual duration of treatment has not been established. A period of two or three years of injection therapy constitutes an average minimum course of treatment.

Parenteral drug products should be inspected visually for particulate matter and discoloration prior to administration, whenever solution and container permit. Some concentrated extracts naturally develop a cloudy appearance over time under refrigeration, the material settling to the bottom on standing.

HOW SUPPLIED

Stock concentrate extracts containing 10,000 BAU/mL and 100,000 BAU/mL are supplied in 10, 30 and 50 mL multiple-dose vials in 50% glycerin. (Standardized Bermuda grass pollen extract is supplied at a maximum strength of 10,000 BAU/mL.) Extracts for puncture, prick or scratch testing are supplied in 5 mL dropper vials at 10,000 BAU/mL or in multiple-dose vials at 100,000 BAU/mL (except Bermuda) in 50% glycerin. Intradermal strengths should be prepared by dilution of stock concentrates with normal saline or saline containing HSA.

STORAGE

All allergenic extracts should be stored at 2-8°C and kept in this temperature range during office use. Refer to vial labels for expiration dates. Clinicians should be aware that diluted extracts are inherently less stable than concentrates. Dilutions of glycerinated extracts which result in glycerin below 50% may also be less stable. Potency of a particular dilution can be checked by skin test in comparison to a fresh dilution of the extract on an individual known to be allergic to the specific antigen.

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ALLERGENIC EXTRACTS

STANDARDIZED CAT HAIR EXTRACT

Suggested Dosage Schedule
and Instructions

WARNING

THIS ALLERGENIC PRODUCT IS INTENDED FOR USE BY PHYSICIANS WHO ARE EXPERIENCED IN THE ADMINISTRATION OF ALLERGENIC EXTRACTS AND THE EMERGENCY CARE OF ANAPHYLAXIS, OR FOR USE UNDER THE GUIDANCE OF AN ALLERGY SPECIALIST.

STANDARDIZED CAT HAIR EXTRACT WITH POTENCY LABELING IN BIOEQUIVALENT ALLERGY UNITS per mL (BAU/mL) IS NOT INTERCHANGEABLE WITH OTHER CAT EXTRACTS. STANDARDIZED CAT HAIR EXTRACT IS NOT INTERCHANGEABLE WITH STANDARDIZED CAT PELT EXTRACTS LABELED IN BIOEQUIVALENT ALLERGY UNITS OR WITH OTHER CAT EXTRACTS LABELED IN ALLERGY UNITS (AU).

THE INITIAL DOSE OF STANDARDIZED CAT HAIR EXTRACT MUST BE BASED ON SKIN TESTING AS DESCRIBED IN THE DOSAGE AND ADMINISTRATION SECTION OF THIS INSERT. PATIENTS BEING SWITCHED FROM OTHER TYPES OF EXTRACTS TO STANDARDIZED CAT HAIR EXTRACT SHOULD BE INSTRUCTED TO RECOGNIZE ADVERSE REACTION SYMPTOMS AND CAUTIONED TO CONTACT THE PHYSICIAN'S OFFICE IF REACTION SYMPTOMS OCCUR. IN CERTAIN INDIVIDUALS THESE REACTIONS COULD BE FATAL. PATIENTS SHOULD BE OBSERVED FOR AT LEAST 20 MINUTES FOLLOWING IMMUNOTHERAPY. PATIENTS WITH LABILE OR STEROID-DEPENDENT ASTHMA ARE "HIGH RISK PATIENTS" WHO REQUIRE SPECIAL CAUTION IN DOSE ADMINISTRATION AND SHOULD REMAIN IN THE OFFICE FOR AT LEAST 30 MINUTES. AIRWAY OBSTRUCTION IN HIGH RISK PATIENTS CAN BE MONITORED BY PEAK FLOW MEASUREMENTS BEFORE AND AFTER ADMINISTRATION OF ALLERGENS. EMERGENCY MEASURES AS WELL AS PERSONNEL TRAINED IN THEIR USE SHOULD BE IMMEDIATELY AVAILABLE IN THE EVENT OF A LIFE THREATENING REACTION. PATIENTS BEING SWITCHED FROM ONE LOT OF EXTRACT TO ANOTHER FROM THE SAME MANUFACTURER SHOULD HAVE THEIR DOSE REDUCED BY 75%.

RISK OF ANAPHYLAXIS SHOULD BE WEIGHED AGAINST BENEFITS IN IMMUNOTHERAPY: IN PATIENTS RECEIVING BETA BLOCKERS AS THEY MAY NOT BE RESPONSIVE TO BETA ADRENERGIC DRUGS SHOULD ANAPHYLAXIS OCCUR; IN PATIENTS WITH UNSTABLE OR STEROID-DEPENDENT ASTHMA; OR IN PATIENTS WITH CARDIOVASCULAR DISEASE.

THIS PRODUCT SHOULD NOT BE INJECTED INTRAVENOUSLY.

REFER ALSO TO THE WARNINGS, PRECAUTIONS, ADVERSE REACTIONS AND OVERDOSAGE SECTION BELOW.

DESCRIPTION

Each vial contains a sterile extract of cat (*Felis domesticus*) pelt and cat dander, 0.50% sodium chloride, 0.25% sodium bicarbonate, 50% glycerin by volume, and 0.4% phenol as a preservative. Source materials for the extract are dry cat dander and dry defatted cat pelt.

Standardized Cat Hair Extract is a sterile solution for intracutaneous or subcutaneous administration. The extract is standardized by comparing potency in cat allergen 1 (*Fel d 1*) units measured by radial immunodiffusion against a reference

standard from the Center for Biologics Evaluation and Research of the U.S. Food and Drug Administration (FDA).⁽¹⁻³⁾ An extract with 10 to 19.9 *Fel d I* units per mL is designated as 10,000 Bioequivalent Allergy Units/mL (BAU/mL) by the FDA based on quantitative skin testing.⁽²⁾ Greer Standardized Cat Hair Extract concentrate is 10,000 BAU/mL.

Cat albumin is considered to be a minor allergen, but may be significant for certain patients.⁽⁴⁾ *Fel d I* is a relatively stable component while albumin is more labile and more easily destroyed by heat.⁽⁵⁻⁶⁾ For lot release, Standardized Cat Hair Extract is compared to FDA reference by isoelectric focusing (IEF) to differentiate it from Cat Pelt Extract.

CLINICAL PHARMACOLOGY

The allergic reaction is dependent upon the presence of antigen-specific IgE antibodies that are bound to specific receptors on mast cells and basophils and has been demonstrated for cat-allergic individuals.⁽⁷⁾ The presence of IgE antibodies on mast cells and basophils sensitizes these cells and upon interaction with the appropriate allergen-histamine and other mediators are released.⁽⁸⁾ In the skin these mediators are responsible for the characteristic wheal and flare reaction.⁽⁹⁾

An increase in cat antigen-specific IgG antibodies has been demonstrated as a result of immunotherapy.⁽¹⁰⁻¹¹⁾ The complete mechanisms of immunotherapy are not known and are still under investigation.

Immunotherapy with cat extract has been studied by several investigators. It is generally believed that hyposensitization with this product is helpful in reducing allergic symptoms associated with exposure to cat allergens in homes or the environment.⁽¹²⁻¹⁶⁾

INDICATIONS AND USAGE

Standardized Cat Hair Extract is indicated for the diagnosis and treatment (immunotherapy) of patients with a history of allergy to cats. The diagnosis of cat allergy is established by the allergy history, clinical evaluation, and skin test reactivity. Cat emanations are common causes of allergy and occur not only upon direct exposure to cats, but also occur in high levels in house dust and other environmental dusts.⁽¹⁷⁾ Persons suspected of having allergy to house dust should be tested for sensitivity to cat allergens. Immunotherapy is indicated when cat allergy is established and the patient cannot avoid exposure to cat allergens.

The use of cat extract for the above purposes should be made only by physicians with special familiarity with and knowledge of allergy. (SEE DOSAGE AND ADMINISTRATION)

CONTRAINDICATIONS

There are no known absolute contraindications to the use of Allergenic Extracts for immunotherapy. Immunotherapy with cat antigens is contraindicated in those individuals who do not exhibit skin test or clinical sensitivity to cat antigens. (See below under WARNINGS AND PRECAUTIONS)

Cat extract injections should not be administered in the presence of diseases characterized by a bleeding diathesis.

Children with nephrotic syndrome require careful consideration and probably should not receive injection therapy because a variety of seemingly unrelated events, such as immunization, can cause an exacerbation of their nephrotic disease.

Other contraindications include:

EXTREME SENSITIVITY TO CAT - Determined from previous anaphylaxis following skin testing.

AUTOIMMUNE DISEASE - Individuals with autoimmune disease may be at risk, due to the possibility of routine immunizations exacerbating symptoms of the underlying disease.

WARNINGS

Please also refer to warning box at beginning of package insert.

Concentrated extracts must be diluted with sterile diluent prior to first use on a patient for immunotherapy or intradermal testing. All concentrates of allergenic extracts are manufactured to assure high potency and therefore have the ability during skin testing and immunotherapy to cause serious local and systemic reactions including death in sensitive patients. Most reactions occur within 20 minutes after injection,⁽¹⁸⁾ but may occur later.⁽¹⁹⁾ To minimize the potential for local or systemic reactions, the relative sensitivity of the patient must be assessed from the allergic history and from clinical observations. Patients should be informed of this and the precautions should be discussed prior to immunotherapy (see PRECAUTIONS below).

Standardized Cat Hair Extract labeled in Bioequivalent Allergy Units is not interchangeable with Standardized Cat Pelt Extract or with cat extracts labeled in Allergy Units. Patient doses stated or calculated in Allergy Units should not be confused with Bioequivalent Allergy Units because the BAU is ten times as potent as the Allergy Unit used for cat extracts before September 1992.

The dosage must be reduced when starting a patient on fresh Standardized Cat Hair Extract or when transferring a patient from any other cat extract product to Standardized Cat Hair Extract (even though the labeled strength of the old and new vials may be the same). This reduction in dosage may be necessary due to a loss of extract potency during storage in the physician's office. **The cat allergen content of old and new extracts must be compared and adjusted by dosage reduction and/or dilution before the new extract is administered.** The amount of new extract given should not exceed 25% of the last dose given from the old vial, assuming both extracts contain comparable amounts of cat allergens. Any evidence of a local or generalized reaction requires a reduction in dosage during the initial stages of immunotherapy, as well as during maintenance therapy.

Allergenic extracts should be temporarily withheld from patients or the dose adjusted downward if any of the following conditions exist:

- (1) severe symptoms of rhinitis and/or asthma
- (2) infection or flu accompanied by fever
- (3) exposure to excessive amounts of clinically relevant allergen prior to a scheduled injection.

Risk of anaphylaxis should be weighed against benefits of immunotherapy: in patients receiving beta blockers as they may not be responsive to beta adrenergic drugs should anaphylaxis occur; in patients with unstable or steroid-dependent asthma; or in patients with cardiovascular disease.

Not for intravenous use!

PRECAUTIONS

GENERAL:

Systemic allergic reactions may occur as a result of immunotherapy. The risk can be minimized by adherence to a careful injection schedule, which starts with a low concentration of extract and is increased slowly. The physician must be prepared to treat anaphylaxis should it occur and have the necessary drugs and equipment on hand to do so.⁽²⁰⁾ Extracts should not be administered by the patient or other individuals who are not prepared to treat anaphylaxis should it occur.

Patients receiving allergenic extracts should be kept under observation a minimum of 20 minutes so that any adverse reaction can be observed and properly handled. This time should be extended to at least 30 minutes for high-risk patients such as those with labile or steroid-dependent asthma or those suffering an exacerbation of their symptoms.⁽²⁴⁾ Airway obstruction in high risk patients can be monitored by peak flow measurements before and after administration of allergens.

Check the prescription or lot number, vial number, strength, and verify the dosage schedule of the prescription for the specific patient. Only after this verification has been made should an injection be given.

A separate, sterile syringe and needle or sterile disposable unit must be used for each patient to prevent the transmission of hepatitis or other infectious agents from person to person. Needles should not be recapped and should be disposed of properly.

Do not use the same syringe for different extracts, nor for the diluent after using it for an extract.

INFORMATION FOR PATIENTS:

Most serious reactions following the administration of allergenic extracts occur within 20 minutes; the patient should remain under observation for this period of time or longer if instructed by the physician.⁽¹⁸⁾ The size of the local reaction should be recorded, because increasingly large local reactions may precede a subsequent systemic reaction with increasing dosage. In particular, this includes unusual swelling and/or tenderness at the injection site or reactions such as shortness of breath, rhinorrhea, sneezing, coughing, wheezing, nausea, dizziness or faintness. Reactions may occur some time after leaving the physician's office, in which case medical attention should be sought immediately.

The patient should be instructed to report any unusual reactions to the physician.

DRUG INTERACTIONS:

Skin test diagnosis with Allergenic Extracts may result in false negative responses when used within 3-10 days of H₁ -Blockers such as cetrizine, loratadine, and terfenadine. The inhibitory effect of astemizole may last up to 60 days.⁽²⁵⁾ These products suppress histamine skin test reactions and could mask a positive response.

The suppressive action of other drugs such as tricyclic antidepressants or topical steroids should be considered and emphasizes the need for a histamine positive-control test.

Patients receiving beta blocker drugs may not be responsive to beta adrenergic drugs used to treat anaphylaxis. The risks of anaphylaxis in these patients should be carefully weighed against the benefits of immunotherapy.

CARCINOGENESIS, MUTAGENESIS, IMPAIRMENT OF FERTILITY

There is no evidence of carcinogenicity, mutagenesis or impairment of fertility in humans from allergenic extracts. No long-term studies in animals have been performed to evaluate carcinogenic potential.

PREGNANCY

PREGNANCY CATEGORY C - Animal reproduction studies have not been conducted with cat extract. It is also not known whether cat extract can cause fetal harm when administered to a pregnant woman or whether it can affect reproduction capacity. Cat extract should be given to a pregnant woman only if clearly needed.

There is no evidence of adverse effects of allergenic extract on the fetus. Studies have not been performed in animals to determine whether this extract affects fertility in males or females, has teratogenic potential, or has other adverse effects on the fetus. Caution should be exercised in testing or treating pregnant females because a systemic reaction may cause an abortion as a result of uterine muscle contractions.

LABOR AND DELIVERY

There is no known information of adverse effects during labor and delivery.

NURSING MOTHERS

It is not known whether this product is excreted in human milk. Because many drugs are excreted in human milk, caution should be exercised when extracts are administered to a nursing woman.

PEDIATRIC AND GERIATRIC USE:

Although standardized cat extract has not been well-studied in children, children and geriatric patients appear to tolerate injections of allergenic extract well. Use in children under six years of age is not recommended. Cat extracts have been administered to children with adverse systemic responses occurring only when given in doses sufficiently high to induce an immediate hypersensitivity reaction.⁽¹⁴⁻¹⁵⁾

ADVERSE REACTIONS

SYSTEMIC REACTIONS

Adverse systemic reactions may occur within minutes with any allergenic extract, including cat. These reactions consist primarily of allergic symptoms such as generalized skin erythema, urticaria, pruritus, angioedema, rhinitis, wheezing, laryngeal edema and hypotension. Less commonly, nausea, emesis, abdominal cramps, diarrhea and uterine contractions may occur. Severe reactions may cause shock and loss of consciousness. Fatalities with allergenic extracts have occurred rarely.⁽²¹⁾ Anaphylaxis and deaths following the subcutaneous injection of extracts have also been

reported by the British Committee on Safety of Medicine.⁽²²⁾ Systemic reactions occur with varying frequency in different clinics and are usually less than 1%. To some extent, the reaction rate is related to the type and dose of administered extract and to the degree of sensitivity of the patient. In general, immunotherapy with allergenic extracts is considered to be safe.⁽²³⁾ Despite all precautions, occasional reactions are unavoidable.

Adverse reactions should be treated as follows:

A. A tourniquet should be immediately applied to the extremity above the site of injection. Release the tourniquet every few minutes for a few seconds.

B. Epinephrine 1:1000 should be injected immediately in the opposite arm in amounts of 0.3 to 0.5 mL and 0.2 mL epinephrine should be administered at the site of injection. For children below the age of 6 years, adjust the dosage of epinephrine to 0.005 mL per pound of body weight per dose. Repeat epinephrine dosage in 15 minutes if necessary and if symptoms persist.

C. Adverse reactions not responding to epinephrine therapy may require the use of parenteral bronchodilators, vasopressors, oxygen, or volume replacement therapy.

LOCAL REACTIONS

Local reactions consisting of erythema, itching, swelling, tenderness and sometimes pain may occur at the injection site. These reactions may appear within a few minutes to hours and persist for several days. Local cold applications and oral antihistamines may be effective immunotherapy. For marked and prolonged local reactions, steroids may be helpful.

OVERDOSAGE

Systemic reactions are uncommon after injection, but if the patient receives more extract than can be tolerated at that particular time, the procedures listed under ADVERSE REACTIONS should be instituted.

Overdosage may occur because of an error in the volume of extract injected, or an incorrect dilution injected, or because the patient may be exposed to airborne or environmental antigens simultaneously with injection of the same antigens. In the event of a systemic reaction occurring, the dosage schedule should be carefully reviewed and if necessary adjusted as outlined above under WARNINGS.

DOSAGE AND ADMINISTRATION

GENERAL

Concentrated standardized cat hair extracts should be diluted prior to intradermal testing or first use in immunotherapy.

Normal or buffered saline or normal saline with human serum albumin may be used to prepare appropriate dilutions. Vials should be visually inspected to ensure particulate free prior to use.

DIAGNOSTIC TESTING

For the patient with a suspected diagnosis of cat allergy, initial testing may be conducted with the concentrate by means of a puncture test employing a multiple puncture device or other appropriate instrument. Prick testing through a drop of extract or scratch testing with a drop of extract applied to the scratch may also be employed to

determine the degree of sensitivity. If the response is negative, this initial test may be followed by intradermal testing where the clinical history is strongly indicative of allergy to cats. Use of a positive and negative control is recommended.

The most frequently used test sites are the back and the volar surface of the forearms. The skin should be cleansed with alcohol and allowed to dry. A minimum of at least 1 inch should be allowed between test sites. A marking pencil may be used to indicate the site locations.

Skin tests read after 15 to 20 minutes are graded in terms of the induration (wheal) and erythema (flare) response compared to the appropriate controls. Wheal and flare sizes may be recorded by actual measurements. The largest diameter of the wheal and flare may be recorded, or the sum of the largest diameter and the orthogonal (right angle) diameter wheal or flare may be used.

Puncture, prick or scratch testing:

The skin test concentration of 10,000 BAU/mL is used for puncture, prick or scratch testing. Puncture tests with Standardized Cat Hair Extract performed on ten highly sensitive cat puncture-positive patients showed a mean diameter wheal of 6.0 mm \pm 2.2 mm and a mean erythema of 36.7 mm \pm 6.7 mm. Glycerinated histamine phosphate 5 mg/mL (1.8 mg/mL histamine base) or aqueous histamine phosphate 2.75 mg/mL (1 mg/mL histamine base; 1:1000 w/v) may be used as a positive control. A 50% glycerosaline solution may be used as the negative control.

A sterile puncture device, needle, scalpel blade, or scarifier is used. A separate sterile device must be used for each patient to prevent transmission of infectious agents. If the device contacts extracts, use a separate device for each antigen to prevent cross-contamination.

The skin is abraded only enough to enter the dermis without drawing blood. Follow the directions for the device being used. The antigen may be applied directly with a puncture device or is introduced by applying a drop of extract to the scratch or prick site, taking care not to touch the skin with the dropper tip.

Intradermal testing:

Extract for intradermal testing must be prepared by diluting the stock concentrate with sterile diluent (use normal or buffered saline or normal saline with human serum albumin) or obtained by ordering the appropriate dilutions ready made. As a positive control, use glycerinated histamine phosphate diluted to 0.5 mg/mL (0.18 mg/mL histamine base) or aqueous histamine phosphate 0.275 mg/mL (0.1 mg/mL histamine base). As a negative control, use 0.5% to 1% glycerosaline solution.

Intradermal skin tests with 0.05 mL of three-fold serial dilutions in ten highly sensitive cat puncture test positive (sum of erythema >57 mm) persons showed the results in Table I:

TABLE I
BAU/mL to Elicit 50mm
Sum of Diameter Erythema Reaction

| Geometric | |
|-------------|-----------------|
| <u>Mean</u> | <u>Range</u> |
| 0.0542 | 0.0050 - 0.4809 |

Intradermal extract is used as follows:

a. Patients with a negative scratch or prick-puncture test:

Patients who do not react to a scratch or prick-puncture test should be tested intradermally with 0.02 to 0.05 mL of a 50 BAU/mL extract dilution. If this test is negative, a second intradermal test may be performed using a 200 BAU/mL extract dilution.

b. Patients tested only by the intradermal method:

Since highly reactive individuals may react intracutaneously at 1:1 million or even 1:10 million dilutions, any intradermal injection should be preceded and the dose adjusted according to puncture test reactivity. Other patients suspected of being moderately allergic may be tested with an intradermal test dose of 0.02 to 0.05 mL of a 0.05 BAU/mL dilution. A negative test should be followed by repeat tests using progressively stronger concentrations until the maximum recommended strength of 200 BAU/mL is reached.

Skin tests are graded in terms of the wheal and erythema response noted at 15 to 20 minutes. Wheal and erythema size may be recorded by actual measurement of the extent of both responses.

IMMUNOTHERAPY

Immunotherapy is administered by subcutaneous injection. **Not for intravenous use!**

Dosage of allergenic extracts is individualized according to the patient's sensitivity, the clinical response, and tolerance to the extract administered during the early phases of an injection regimen. The initial dose of the extract should be calculated based on the puncture test reactivity. The initial dose of the extract may be as low as 0.1 mL of a 0.005 to 0.05 BAU/mL dilution (dilution 5 or 6 in Table II below) or even less for the exquisitely sensitive patient. Patients with lesser sensitivity may be started at 0.1 mL of a 0.5 to 5 BAU/mL dilution. The amount of allergenic extract is increased at each injection by no more than 50% of the previous amount; the next increment is governed by the response to the last injection. Large local reactions which persist for longer than 24 hours are generally considered an indication for repeating the previous dose or reducing the dose. Any evidence of systemic reaction is an indication for a significant reduction (at least 50%) in the subsequent dose. The upper limits of dosage have not been established; however, doses larger than 0.2 mL of 5,000 BAU/mL may cause discomfort upon injection because of the high glycerin content. The dosage of allergenic extract does not vary significantly with the respiratory allergic disease under treatment.

To prepare dilutions for intradermal and therapeutic use starting from a 5,000 or 10,000 BAU/mL stock concentrate proceed as follows: (Note add 1 mL of concentrate to 9.0 mL of sterile diluent and make additional dilutions in the same manner.)

TABLE II

| <u>Dilution</u> | <u>Extract</u> | <u>Diluent</u> | <u>BAU/mL</u> | |
|-----------------|------------------|----------------|---------------|--------|
| 0 | Concentrate | | 5,000 | 10,000 |
| 1 | 1 mL concentrate | 9.0 | 500 | 1,000 |
| 2 | 1 mL dilution 1 | 9.0 | 50 | 100 |
| 3 | 1 mL dilution 2 | 9.0 | 5 | 10 |
| 4 | 1 mL dilution 3 | 9.0 | 0.5 | 1.0 |
| 5 | 1 mL dilution 4 | 9.0 | 0.05 | 0.1 |
| 6 | 1 mL dilution 5 | 9.0 | 0.005 | 0.01 |

The optimal interval between doses of allergenic extract has not been definitely established. However, as is customarily practiced, injections are given 1, 2 or 3 times per week until the maintenance dose of extract is reached. At this time, the injection interval is increased to 2 weeks, then to 3 weeks and finally to 4 weeks. If the patient does not return for 6 to 8 weeks after the last injection, the dose should be reduced to 25% of the last dose. If longer than 8 weeks, a dose reduction of one, two or three dilutions may be made depending on a consideration of the components and the patient's sensitivity. The dosage and the interval between injections may need to be modified according to the clinical response of the patient. When switching patients to fresh extract, the initial dose should be reduced to one-quarter of the previous dose.

The usual duration of immunotherapy has not been established. A period of two or three years of injection therapy constitutes an average minimum course of treatment.

HOW SUPPLIED

Stock concentrate extract containing 10,000 Bioequivalent Allergy Units per mL (10,000 BAU/mL) in 50% glycerin v/v or other dilutions as requested by the physician are supplied in 5, 10, 30 and 50 mL vials. Cat extracts are also supplied in 5 mL dropper vials for puncture, prick or scratch testing. All dilutions of cat extracts are supplied in 50% Glycero-Coca's solution.

STORAGE

Standardized Cat Hair Extract should be stored at 2-8 degrees C and kept at this temperature range during office use. Refer to vial label for expiration date of concentrated extract. Dilutions should not be used past the expiry dating on the concentrate. Dilutions of concentrated extract result in a glycerin content of less than 50% which is less stable, particularly for the serum component. Potency of the dilution can be checked by skin test in comparison to a fresh dilution of the extract on a known cat allergic individual.

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ALLERGENIC EXTRACTS

Short Ragweed
and
G.S. Ragweed Mix

Suggested Dosage Schedule
and Instructions

WARNING

Allergenic extracts can elicit severe adverse reactions when improperly administered (see below under Warnings). Any person administering allergenic extracts should be experienced in their use, aware of the risk of adverse reactions, and capable of properly handling such reactions.

DESCRIPTION

Each vial contains an extract of Short Ragweed (**Ambrosia elatior**) pollen or of equal parts Short and Giant Ragweed (**A. trifida**) pollen extracts. Extracts are supplied in a buffered saline solution with or without glycerin (50%) added as a stabilizer. Extracts are supplied as a sterile solution intended for subcutaneous or intracutaneous administration.

Each lot of Short Ragweed extract or Mixed Ragweed extract is assayed for Antigen E content using a radial immunodiffusion test¹ employing known Antigen E standards and anti-Antigen E antisera. Extracts containing Short Ragweed at a concentration of 1:20 or stronger are assayed for Antigen E. Extracts more dilute than this are not assayed but the Antigen E content is obtained by calculation based on the Antigen E content of the concentrate.

Antigen E, a protein with a molecular weight of approximately 38,000, has been found to be a major antigenic component of Short Ragweed pollen.^{2,3} Antigen E content has been shown to correlate well with skin test reactivity of ragweed extracts.⁴ For this reason, Antigen E content is being used to standardize Short Ragweed extracts in addition to the protein nitrogen content or weight to volume ratios as have been used.

When transferring patients from non-standardized extracts to a standardized extract, it is advisable to compare the potency of the different lots by comparative skin testing using the same concentrations of each extract. Marked differences in skin reactivity will indicate differences in potency and the need to adjust dosages so as to avoid possible severe reaction.

CLINICAL PHARMACOLOGY

Controlled studies employing Ragweed extracts and Antigen E immunotherapy have demonstrated an increase in Ragweed antigen-specific blocking antibodies. These studies also demonstrated significant symptom amelioration in Ragweed allergic individuals.^{5,6} The total mechanism of immunotherapy is not yet known and is still being investigated.

INDICATIONS

Hyposensitization is indicated when careful testing and patient history can pinpoint allergens responsible for allergic symptoms, and when it is not possible or practical to avoid these allergens. Allergenic extracts are administered to reduce symptoms of allergy of a seasonal or perennial nature.

CONTRAINDICATIONS

Immunotherapy with Ragweed antigens is contraindicated in those individuals who do not exhibit skin test or clinical sensitivity to Ragweed antigens. (See below under Warnings and Precautions)

WARNINGS

Allergenic extracts can elicit adverse local and systemic reactions if initial dosage or rate of dosage increase is too high. These factors must be carefully evaluated by the physician based on history, degree of sensitivity, and skin test results prior to commencement or continuation of therapy. Any person administering a biological product should be aware of the risk of local or systemic reactions if improperly used and be capable of handling such reactions. (See below under Adverse Reactions.) Patients receiving allergenic extracts should be kept under observation a minimum of thirty (30) minutes so that any adverse reaction can be observed and properly handled. See information above in Description category for information on transferring patients from non-standardized to standardized extracts.

PRECAUTIONS

Check lot number and dosage schedule of patient to verify correctness of prescription number or vial number. Only after this verification has been made should an appropriate injection be given. A separate sterile needle and syringe should be used for each patient to prevent transmission of homologous serum hepatitis and other infectious agents.

Pregnancy Category C. Animal reproduction studies have not been conducted with allergenic extracts. It is also not known whether allergenic extracts can cause fetal harm when administered to a pregnant woman or whether it can affect reproduction capacity.

ADVERSE REACTIONS:

Allergic reactions following injections of allergens include generalized erythema, pruritis, rhinitis, asthma, and laryngeal edema. Syncope, shock, and hypotension have also been reported. Adverse reactions should be treated as follows:

- A. A tourniquet should be immediately applied to the extremity above the site of injection. Release tourniquet every few minutes for a few seconds.
- B. Epinephrine 1:1000 should be injected immediately in the opposite arm in amounts of 0.3 to 0.5 mL and 0.2 mL epinephrine should be administered at the site of injection. For children below the age of 6 years, adjust dosage of epinephrine to 0.005 mL per pound of body weight per dose. Repeat epinephrine dosage in 15 minutes if necessary and if symptoms persist.
- C. Adverse reactions not responding to epinephrine therapy may require the use of parenteral bronchodilators, vasopressors, oxygen, or volume replacement therapy.

DOSAGE AND ADMINISTRATION

A. Testing:

1. Scratch and Prick testing normally employ 1:20 glycerinated extracts, which contain 125-250 units Antigen E per mL (Short Ragweed) or 75-150 units Antigen E per mL (Ragweed Mix).
2. Intradermal testing with Short Ragweed and G.S. Ragweed Mix is routinely performed using 1:1000 w/v and 1,000 PNU/mL or more dilute material. It has been reported that as little as 10^{-6} unit (0.000001 μg) is sufficient to cause a positive skin test. In another study of 25 Antigen E sensitive individuals, the dose required for a 2+ reaction (8-10 mm wheal) ranged from 10^{-6} to 10^{-1} μg Antigen per mL using a 0.5 mL dose.³

Individual sensitivity to Ragweed extracts will vary greatly and it is recommended that intradermal testing be started at low concentrations (i.e. 0.05 mL at 0.1 $\mu\text{g}/\text{mL}$ - dose equals 0.005 μg Antigen E). If no reaction occurs, testing may be repeated with a stonger dilution.

B. Treatment:

Initial and subsequent treatment dosage must be based on careful testing procedures and evaluation of patient history. Studies have shown a definite correlation between symptom amelioration and maximum or cumulative dosage achieved. One study⁵ has shown significant clinical improvement with an average cumulative preseasonal dosage equivalent to 252 μg of Antigen E, but no significant improvement with an average cumulative dosage of 32 μg Antigen E. In another study⁸ a mean cumulative dosage of 84.9 μg Antigen E (as whole ragweed extract) was significantly effective in reducing allergenic symptoms compared to placebo.

The dosage schedule shown below, based on Antigen E content, will deliver approximately 85 μg Antigen E in 18 injections. Initial and subsequent treatment dosage must be based on careful testing procedures and patient history. Highly sensitive individuals will require lower initial dosage, more moderate dosage increase, and may not tolerate as high a maintenance dosage as the moderately sensitive individual. The suggested schedule shown below is for a moderately sensitive individual and must be modified by the physician to suit each patient if necessary.

**SUGGESTED DOSAGE SCHEDULE
OF SHORT RAGWEED AND G.S. RAGWEED MIX
ANTIGEN E STANDARD**

| Injection Number | Vial Number | Antigen E per mL | Volume (mL) | Dose AgE |
|------------------|-------------|---------------------|-------------|----------|
| 1 | 1 | 0.3 | 0.05 | 0.015 |
| 2 | 1 | 0.3 | 0.10 | 0.03 |
| 3 | 1 | 0.3 | 0.20 | 0.06 |
| 4 | 1 | 0.3 | 0.40 | 0.12 |
| 5 | 1 | 0.3 | 0.70 | 0.21 |
| 6 | 2 | 3.0 | 0.10 | 0.30 |
| 7 | 2 | 3.0 | 0.20 | 0.60 |
| 8 | 2 | 3.0 | 0.30 | 0.90 |
| 9 | 2 | 3.0 | 0.50 | 1.50 |
| 10 | 2 | 3.0 | 0.70 | 2.10 |
| 11 | 3 | 30.0 | 0.10 | 3.00 |
| 12 | 3 | 30.0 | 0.15 | 4.50 |
| 13 | 3 | 30.0 | 0.20 | 6.00 |
| 14 | 3 | 30.0 | 0.30 | 9.00 |
| 15 | 3 | 30.0 | 0.40 | 12.00 |
| 16 | 3 | 30.0 | 0.50 | 15.00* |
| 17 | 3 | 30.0 | 0.50 | 15.00* |
| 18 | 3 | 30.0 | 0.50 | 15.00* |

*Maintenance dosage

It is recommended that patients receive injections at five to seven day intervals until maintenance dosage is achieved. Maintenance dosage can be given at 2 to 4 week intervals. All doses of allergenic extract are administered subcutaneously in the lateral aspect of the upper arm or thigh. Avoid injecting directly into any blood vessel and use a 26 or 27 gauge needle $\frac{3}{8}$ " in length.

OVERDOSAGE

Systemic reactions are uncommon after injection, but if the patient receives more extract than can be tolerated at that particular time, the procedures listed under Adverse Reactions should be instituted.

Overdosage may occur because of an error in the volume of extract injected, an incorrect dilution injected, or because the patient may be exposed to airborne antigens simultaneously with injection of the same antigens. In the event of a systemic reaction occurring, the dosage schedule should be carefully reviewed and if necessary adjusted as outlined above.

STORAGE

Short Ragweed containing extracts should be stored at 2°-8°C at all times, even during use, to prolong the potency of the extracts. Our studies indicate that Antigen E in aqueous extracts decays approximately ten times more rapidly at room temperature (20°-25°C) than at refrigerator temperature (2°-8°C).

HOW SUPPLIED

Short Ragweed and G.S. Ragweed Mix extracts are supplied as sterile solutions 5, 10, 30 and 50 mL multiple dose vials and in 5 mL dropper vials for scratch testing. Various concentrations are available to suit the varying needs of the physician.

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- ⁶Arbesman, Carl E., et al., "Treatment of ragweed-sensitive patients with Antigen E and ragweed extract: Clinical and immunologic studies," **The Journal of Allergy**, 39:2, p. 124, 1967.
- ⁷Norman, Phillip S., "A rational approach to desensitization," **The Journal of Allergy**, 44:3, pp. 129-145, 1969.
- ⁸Van Metre, Thomas E., et al., "A comparative study of the effectiveness of the Rinkel method and the current standard method of immunotherapy for ragweed pollen hay fever," **The Journal of Allergy and Clinical Immunology**, 66:6, pp. 500-513, 1980.

OVERDOSAGE

Systemic reactions are uncommon after injection, but if the patient receives more extract than can be tolerated at that particular time and begins to experience immediate hypersensitivity anaphylaxis, the procedures listed under ADVERSE REACTIONS should be instituted.

Overdosage may occur because of an error in the volume of extract injected, or an incorrect dilution injected, or because the patient may be exposed to airborne or environmental antigens simultaneously with injection of the same antigens. In the event of a systemic reaction occurring, the dosage schedule should be carefully adjusted as outlined above under WARNINGS.

DOSAGE AND ADMINISTRATION

1. DIAGNOSTIC TESTING

For the patient with a suspected diagnosis of allergy to more than one antigen, initial skin testing should include the individual extracts. If a screening skin test with a mixture is used, a positive response should be followed by testing with the individual extracts to determine the degree of sensitivity to each and to guide in the selection of extracts and their concentration for immunotherapy if indicated. However, because a negative skin test with a mixture may not be indicative of the absence of allergy to one or more of the components due to their dilution, testing with individual extracts is more precise. False negative responses may occur if serum levels of antihistamines remain from prior medication administration (see CONTRAINDICATIONS). The use of a positive control is especially recommended for patients on prior medications which may decrease the histamine skin test response.

Scratch or Prick-Puncture Skin Testing:

Allergenic Extract concentrates may be used for scratch or prick-puncture testing or scratch tests in 50% glycerin 1:20 W/V or strongest available strength in 5 mL vials may be used. Prick-puncture tests with concentrated extracts in patients highly sensitive to the specific antigen should yield distinctive wheals with diameters of greater than 5 mm and with much larger erythema reactions. Glycerinated histamine phosphate 5 mg/mL (1.8 mg/mL histamine base) or aqueous histamine phosphate 2.75 mg/mL (1mg/mL histamine base; 1:1000W/V) may be used as a positive control.

Intradermal Skin Testing:

Extract for intradermal testing must be prepared by diluting the stock concentrate injection vials with sterile diluent (use normal or buffered saline, or normal saline with human serum albumin) or the appropriate dilutions may be purchased.

a. Patients with a negative scratch or prick-puncture test:

Patients who do not react to a scratch or prick-puncture test should be tested intradermally, using a 26 or 27 gauge 1/4 inch needle, with 0.02 to 0.05 mL of an appropriate extract dilution from 1/100 to 1/1000 of the concentrate. A negative test should be followed by a repeat test using a

higher concentration until significant wheal and flare reaction sizes are attained or until the responses remain negative. As a negative control use the diluent or, in the case of extracts in 50% glycerin, use 0.5% to 1% glycerosaline solution. As a positive control use glycerinated histamine phosphate diluted to 0.5 mg/mL (0.18 mg/mL histamine base) or aqueous histamine phosphate 0.275 mg/mL (0.1 mg/mL histamine base).

b. Patients tested only by the intradermal method:

Since highly reactive individuals may react intracutaneously at 1:1 million or even 1:10 million dilutions, any intradermal injection should be preceded by a puncture test and the dose adjusted accordingly. Other patients suspected of being moderately allergic should be tested with 0.02 to 0.05 mL of an appropriate extract dilution on the order of 1/10,000 to 1/100,000 of the concentrate. A negative test should be followed by repeat tests using progressively stronger ten-fold concentrations until significant wheal and flare reaction sizes are attained, or until skin test responses with the higher concentrations remain negative. As a negative control, use the diluent or, in the case of extracts in 50% glycerin, use 0.5% to 1% glycerosaline solution. As a positive control, use glycerinated histamine phosphate diluted to 0.5 mg/mL (0.18 mg/mL histamine base) or aqueous histamine phosphate 0.275 mg/mL (0.1 mg/mL histamine base).

Skin tests are graded in terms of the wheal and erythema response noted at 15 to 20 minutes, and compared to the appropriate controls. Wheal and erythema sizes may be recorded by actual measurement.

2. IMMUNOTHERAPY

Immunotherapy is administered by subcutaneous injection. Dosage of Allergenic Extracts is individualized according to the patient's sensitivity, the clinical response, and tolerance to the extract administered during the phases of an injection regimen. The initial dose of the extract should be determined based on the puncture test reactivity. In patients who appear to be exquisitely sensitive by history and skin test, the initial dose of the extract should be 0.05 to 0.1 mL of a low concentration, such as dilution number 5 or 6 in TABLE 1 below. Patients with lesser sensitivity may be started with 0.05 to 0.1 mL of the next higher concentration. The amount of Allergenic Extract is increased at each injection by no more than 50% of the previous amount, and the next increment is governed by the response to the last injection. Large local reactions which persist for longer than 24 hours are generally considered an indication for repeating the previous dose or reducing the dose at the next administration. Any evidence of systemic reaction is an indication for a reduction of 75% in the subsequent dose. The upper limits of dosage have not been established; however, doses larger than 0.2 mL of an extract in 50% glycerin may cause discomfort upon injection. The dosage of Allergenic Extract does not vary significantly with the allergic disease under treatment.

To prepare dilutions starting from a concentrate such as 1:10 W/V, 1:20 W/V OR 20,000 PNU/mL, proceed as in TABLE 1 below. (Note: Add 0.5 mL of concentrate to 4.5 mL of sterile diluent and make additional dilutions in the same manner.

TABLE 1

| Ten-Fold Dilution Series* | | | | | |
|---------------------------|--------------------|---------|--------------|--------------|--------|
| Dilution | Extract | Diluent | W/V | W/V | PNU/mL |
| 0 | Concentrate | | 1:10 | 1:20 | 20,000 |
| 1 | 0.5 mL concentrate | 4.5 mL | 1:100 | 1:200 | 2,000 |
| 2 | 0.5 mL dilution 1 | 4.5 mL | 1:1,000 | 1:2,000 | 200 |
| 3 | 0.5 mL dilution 2 | 4.5 mL | 1:10,000 | 1:20,000 | 20 |
| 4 | 0.5 mL dilution 3 | 4.5 mL | 1:100,000 | 1:200,000 | 2 |
| 5 | 0.5 mL dilution 4 | 4.5 mL | 1:1,000,000 | 1:2,000,000 | 0.2 |
| 6 | 0.5 mL dilution 5 | 4.5 mL | 1:10,000,000 | 1:20,000,000 | 0.02 |

*There is no direct potency correlation across the table between PNUs and W/V.

The optimal interval between doses of Allergenic Extract has not been established. Injections usually are given 1 or 2 times per week until the maintenance dose is reached. The injection interval then is increased to 2 weeks, then to 3 weeks, and finally to 4 weeks. If the patient does not return for 6 to 8 weeks, the dose should be reduced to 25% of the last dose. If longer than 8 weeks, a dose reduction of one, two, or three dilutions may be made considering the components and the patient's sensitivity. The dosage and the interval between injections may need to be modified according to the clinical response of the patient. When switching patients to fresh extract, the initial dose should be reduced to 25% of the previous dose.

The usual duration of treatment has not been established. A period of two or three years of injection therapy constitutes an average minimum course of treatment.

HOW SUPPLIED

Stock concentrate extracts containing up to 40,000 PNU/mL, or 1:10 W/V or other dilutions as requested by the physician are supplied in 5, 10, 30, and 50 mL in aqueous or 50% glycerin buffered saline. House dust extract is supplied in a 1:1 W/V concentrate, or a maximum of 10,000 PNU/mL. Extracts are also supplied in dropper vials for scratch or prick testing.

STORAGE

Allergenic Extracts should be stored at 2-8° C and kept at this temperature range during office use. Refer to vial labels for expiration dates. Diluted extracts are inherently less stable than concentrates. Dilutions of glycerinated extracts which result in glycerin below 50% are also less stable. The more dilute extracts in aqueous diluents should be replenished daily. Potency of a particular dilution can be checked by skin test in comparison to a fresh dilution of the extract on an individual known to be allergic to the specific antigen.

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ALLERGENIC EXTRACTS

POLLENS, MOLDS, EPIDERMALS, INSECTS, DUSTS, FOODS, AND

MISCELLANEOUS INHALANTS

Suggested Dosage Schedule
and Instructions

WARNING

THIS ALLERGENIC PRODUCT IS INTENDED FOR USE BY PHYSICIANS WHO ARE EXPERIENCED IN THE ADMINISTRATION OF ALLERGENIC EXTRACTS AND THE EMERGENCY CARE OF ANAPHYLAXIS, OR FOR USE UNDER THE GUIDANCE OF AN ALLERGY SPECIALIST.

ALLERGENIC EXTRACTS MAY CAUSE SEVERE OR FATAL ANAPHYLAXIS IN EXTREMELY SENSITIVE PATIENTS. THE INITIAL DOSE MUST BE BASED ON SKIN TESTING AS DESCRIBED IN THE DOSAGE AND ADMINISTRATION SECTION OF THIS INSERT. PATIENTS SHOULD BE INSTRUCTED TO RECOGNIZE ADVERSE REACTION SYMPTOMS AND CAUTIONED TO CONTACT THE PHYSICIAN'S OFFICE IF REACTION SYMPTOMS OCCUR. IN CERTAIN INDIVIDUALS, THESE REACTIONS COULD BE FATAL. PATIENTS SHOULD BE OBSERVED FOR AT LEAST 20 MINUTES FOLLOWING TREATMENT.

EMERGENCY MEASURES, AS WELL AS PERSONNEL TRAINED IN THEIR USE, SHOULD BE IMMEDIATELY AVAILABLE IN THE EVENT OF A LIFE-THREATENING REACTION. PATIENTS BEING SWITCHED FROM ONE LOT OF EXTRACT TO ANOTHER FROM THE SAME MANUFACTURER SHOULD HAVE THEIR DOSE REDUCED BY 75%. THIS PRODUCT SHOULD NOT BE INJECTED INTRAVENOUSLY.

REFER ALSO TO THE WARNINGS, PRECAUTIONS, ADVERSE REACTIONS AND OVERDOSAGE SECTIONS BELOW.

DESCRIPTION

Allergenic Extracts are supplied as a sterile solution for intracutaneous or subcutaneous administration. Concentrates contain the soluble extractants of the source material with 0.5% sodium chloride and 0.54% Sodium bicarbonate at a pH of 6.8 to 8.4 as aqueous extracts in water for injection or in 50% glycerin. Aqueous extracts contain 0.4% phenol as a preservative and 50% glycerinated extracts contain 0.2% phenol. Diluted aqueous extracts contain Buffered Saline with 0.5% sodium chloride, 0.04% potassium phosphate, 0.11% sodium phosphate heptahydrate, and 0.4% phenol in water for injection.

Source materials for these extracts are as follows: Pollens are collected from the respective grasses, weeds, trees, shrubs, cultured plants, and flowers. Mold extracts are produced from pure culture mycelial mats. Rusts and smuts are obtained from natural growths. Epidermal extracts are produced from the hide, hair, or feathers containing the natural dander, or from separated dander. Insects are the whole body insects. House dust is made from various dusts ordinarily found in the home with the extract dialyzed to remove low-molecular weight irritants and concentrated to an extraction ratio of 1:1. Food extracts are prepared from the edible portions of the respective foods, obtained fresh if possible. Certain diagnostic food extracts contain 0.1% sodium formaldehyde sulfoxylate as an antioxidant. Other miscellaneous inhalants involved in respiratory allergy are obtained in the naturally occurring form to which a patient may be exposed.

Extracts are labeled either by weight-to-volume (W/V) based on the weight of the source material to the volume of the extracting fluid, or in

protein nitrogen units (PNU) based on assay with one PNU representing 0.00001 mg of protein nitrogen.

CLINICAL PHARMACOLOGY

The allergic reaction is dependent upon the presence of antigen-specific immunoglobulin E (IgE) antibodies that are bound to specific receptors on mast cells and basophils. The presence of IgE antibodies on mast cells and basophils sensitizes these cells and, upon interaction with the appropriate allergen, histamine and other mediators are released. IgE antibody has been shown to correlate with atopic diseases such as allergic rhinitis and allergic asthma.(1-4) In the skin these mediators are responsible for the characteristic wheal and flare (erythema) reactions upon Allergenic Extract skin testing in persons with the specific allergies.(3-7)

Specific immunotherapy with Allergenic Extracts as employed for over 45 years is helpful in reducing symptoms associated with exposure to the offending allergens. A summary of effectiveness by the Panel on Review of Allergenic Extracts, an advisory committee to the U.S. Food and Drug Administration, has been published.(8) Several mechanisms have been proposed to explain the effectiveness of immunotherapy: an increase in antigen-specific IgG antibodies is frequently associated with clinical effectiveness, although correlation is not consistent in all studies; there is a decrease in specific IgE; and IgE production is suppressed during periods of seasonal or high exposure to the antigen.(9) Other changes following immunotherapy have been noted including development of auto-anti-idiotypic antibodies; a decrease in blood basophil sensitivity to allergen; a decrease in lymphokine production and lymphocyte proliferation by cells exposed to allergen; and development of allergen-specific suppressor cells.(10) The complete mechanisms of immunotherapy are not known and remain the subject of investigation.

INDICATIONS AND USAGE

Allergenic Extracts are indicated for the diagnosis and treatment of patients with immediate hypersensitivity allergy to the respective allergens, inhaled, ingested, or otherwise introduced into contact with sensitive tissues. The diagnosis of IgE-mediated allergy may be established by the allergy history, clinical evaluation, and skin test reactivity.(4,7,11) Immunotherapy with Allergenic Extracts is indicated when testing and patient history have identified the offending allergens and when it is not possible or practical to avoid these allergens.(12-14) Food extracts have not been proven effective in immunotherapy.

The use of Allergenic Extracts for the above purposes should be made only by physicians with special familiarity and knowledge of allergy. (See DOSAGE AND ADMINISTRATION.)

CONTRAINDICATIONS

There are no known absolute contraindications to the use of Allergenic Extracts for immunotherapy. Immunotherapy with specific antigens should not be done in those individuals who do not exhibit skin test or clinical sensitivity to the particular antigens. (See below under WARNINGS and PRECAUTIONS.)

Allergenic extract injections should not be administered in the presence of diseases characterized by a bleeding diathesis.

Children with nephrotic syndrome require careful consideration and probably should not receive injection therapy because a variety of seemingly unrelated events, such as immunization, can cause an exacerbation of their nephrotic disease.

General contraindications include:

EXTREME SENSITIVITY TO THE SPECIFIC ALLERGEN - Determined from previous anaphylaxis following exposure.

AUTOIMMUNE DISEASE - Individuals with autoimmune disease may be at risk, due to the possibility of routine immunizations exacerbating symptoms of the underlying disease.

WARNINGS

Concentrated extracts must be diluted with a sterile diluent prior to first use on a patient for treatment or intradermal testing. Allergenic Extracts are manufactured to assure high potency and have the ability during skin testing and immunotherapy to cause serious local and systemic reactions including death in sensitive patients. Most reactions occur within 20 minutes after injection.(15) but may occur later.(16) To minimize the potential for local or systemic reactions, the relative sensitivity of the patient must be assessed from the allergic history and from clinical observations. Patients should be informed of these risks prior to skin testing and immunotherapy (see PRECAUTIONS and ADVERSE REACTIONS below).

Allergenic Extract immunotherapy doses should be lowered or temporarily withheld from patients if any of the following conditions exist:

(1) severe symptoms of rhinitis and/or asthma

(2) infection or flu accompanied by fever

(3) exposure to excessive amounts of clinically relevant allergen prior to a scheduled injection

(4) evidence of a local or systemic reaction to the preceding extract injection during a course of immunotherapy.

The dosage must be reduced when modifying dosages or components in a mixture or an individual prescription, or when starting a patient on fresh extract, even though the labeled strength of the old and new vials may be the same. This reduction in dosage may be necessary due to the older vial losing potency during storage, or due to different sensitivities to different components. The amount of new extract given should not exceed 25% of the last dose given from the old vial, assuming both extracts contain comparable amounts of allergen. Any evidence of a local or generalized reaction requires a reduction in dosage during the initial stages of immunotherapy, as well as during maintenance therapy.

PRECAUTIONS

GENERAL:

Not for intravenous use!

Systemic allergic reactions may occur as a result of immunotherapy. The risk can be minimized by adherence to a careful injection schedule, which starts with a low concentration of extract and is increased slowly.

Because of the danger of serious reactions, caution is needed in testing exquisitely sensitive patients, particularly with potent allergens, e.g., peanut, cottonseed, and flaxseed.(8) Such extracts should be appropriately diluted before use.

The physician must be prepared to treat anaphylaxis should it occur and have the necessary drugs and equipment on hand to do so.(17-18) Extracts should not be administered by the patient or other individuals who are not prepared to treat anaphylaxis should it occur.

Patients receiving Allergenic Extracts should be kept under observation a minimum of twenty (20) minutes so that any adverse reaction can be observed and properly handled.(15) This time should be extended for high-risk patients such as those with unstable asthma or those suffering an exacerbation of their symptoms.

Patients receiving beta blockers may not be responsive to beta adrenergic drugs used to treat anaphylaxis. The risks of anaphylaxis in these patients should be carefully weighed against the benefits of immunotherapy.

Check the lot number and dosage schedule of the patient to verify correctness of a prescription number, a vial number, or strength. Only after this verification has been made should an injection be given.

A separate sterile needle and syringe should be used for each patient to prevent transmission of hepatitis or other infectious agents.

INFORMATION FOR PATIENTS:

Most serious reactions following the administration of Allergenic Extracts occur within 20 minutes; the patient should remain under observation for this period of time or longer if instructed by the physician. The size of any local reaction should be recorded, because increasingly large local reactions may precede a subsequent systemic reaction with increasing dosage. The patient should be instructed to report any unusual reactions. In particular, this includes unusual swelling and/or tenderness at the injection site, or reactions such as rhinorrhea, sneezing, coughing, wheezing, shortness of breath, nausea, dizziness, or faintness. Reactions may occur some time after leaving the physician's office, in which case medical attention should be sought immediately.

DRUG INTERACTIONS: Skin test diagnosis with Allergenic Extracts is contraindicated within 24 hours after the last dose of most antihistamines, within 48 hours after the last dose of terfenadine, and within 3 weeks or longer after the last dose of astemizole. These products suppress histamine skin test reactions and could mask a positive response.

CARCINOGENESIS, MUTAGENESIS, IMPAIRMENT OF FERTILITY: There is no evidence of carcinogenicity, mutagenesis, or impairment of fertility in humans from Allergenic Extracts. No long-term studies in animals have been performed to evaluate carcinogenic potential.

PREGNANCY: PREGNANCY CATEGORY C - Animal reproduction studies have not been conducted with Allergenic Extracts. It is also not known whether Allergenic Extracts can cause fetal harm when administered to a pregnant woman or whether they can affect reproduction capacity. Allergenic Extracts should be given to a pregnant woman only if clearly needed.

There is no evidence of adverse effects of Allergenic Extracts on the fetus.(8) Studies have not been performed in animals to determine

whether extracts affect fertility in males or females, have teratogenic potential, or have other adverse effects on the fetus. Caution should be exercised in testing or treating pregnant females because a systemic reaction may cause an abortion as a result of uterine muscle contractions.

LABOR AND DELIVERY: There is no known information of adverse effects during labor and delivery.

NURSING MOTHERS: It is not known whether this product is excreted in human milk. Because many drugs are excreted in human milk, caution should be exercised when extracts are administered to a nursing woman.

PEDIATRIC AND GERIATRIC USE: Although most extracts have not been studied systematically in children, children and geriatric patients appear to tolerate injections of Allergenic Extracts well. Studies with pollenosis and asthma have been conducted in children (e.g. Refs. 19-21). Extract usage in children should follow the same precautions as in adults.

ADVERSE REACTIONS

Adverse systemic reactions may occur within minutes upon use of an Allergenic Extract to which a person has specific sensitivity. These reactions consist primarily of allergic symptoms such as generalized skin erythema, urticaria, pruritis angioedema, rhinitis, wheezing, laryngeal edema, and hypotension. Less commonly, nausea, emesis, abdominal cramps, diarrhea and uterine contractions may occur. Severe reactions may cause shock and loss of consciousness. Fatalities have occurred rarely.(8,22,23) These systemic reactions occur with varying frequency in different clinics and are usually less than 1%. To some extent, the reaction rate is related to the type and dose of administered extract and to the sensitivity of the patient. In general, immunotherapy with Allergenic Extracts is considered to be safe.(24) Despite all precautions, occasional reactions are unavoidable.

Adverse systemic reactions should be treated as follows:

A. A tourniquet should be immediately applied to the extremity above the site of the injection. Release the tourniquet every few minutes for a few seconds.

B. Epinephrine 1:1000 should be injected immediately in the opposite arm in amounts of 0.3 to 0.5 mL and 0.2 mL epinephrine should be administered at the site of injection. For children below the age of 6 years, adjust the dosage of epinephrine to 0.005 mL per pound of body weight per dose. Repeat epinephrine dosage in 15 minutes if necessary and if symptoms persist.

C. Adverse reactions not responding to epinephrine therapy may require the use of parenteral bronchodilators, vasopressors, oxygen, or volume replacement therapy.

Local reactions consisting of erythema, itching, swelling, tenderness and sometimes pain may occur at the injection site. These reactions may appear within a few minutes to hours and persist for several days. Local cold applications and oral antihistamines may be effective treatment. For marked and prolonged local reactions, steroids may be helpful.

ALLERGENIC EXTRACTS FOR DIAGNOSTIC USE ONLY

Scratch, Prick, Puncture
or Intradermal Testing

U.S. Government License No. 308
Canadian License No. 547



PO Box 800
Lenoir, NC 28645
USA

Revised 04/04

WARNING

THIS ALLERGENIC PRODUCT IS INTENDED FOR DIAGNOSTIC USE ONLY. THIS PRODUCT IS INTENDED FOR USE BY PHYSICIANS WHO ARE EXPERIENCED IN THE ADMINISTRATION OF ALLERGENIC EXTRACTS AND THE EMERGENCY CARE OF ANAPHYLAXIS, OR FOR USE UNDER THE GUIDANCE OF AN ALLERGY SPECIALIST. ALLERGENIC EXTRACTS MAY CAUSE SEVERE OR FATAL ANAPHYLAXIS IN EXTREMELY SENSITIVE PATIENTS. THIS PRODUCT SHOULD NOT BE INJECTED INTRAVENOUSLY. REFER ALSO TO THE WARNINGS, PRECAUTIONS, ADVERSE REACTIONS AND OVERDOSAGE SECTION BELOW.

DESCRIPTION

Allergenic Extracts intended for diagnostic skin testing are for scratch, prick, puncture or intradermal use only. The extracts are supplied as sterile solutions which contain the soluble extractants of the allergen source material in a buffered saline solution with or without glycerin (50% v/v) added as a stabilizer. Concentrated aqueous extracts contain the soluble extractants of the source material with 0.5% sodium chloride and 0.54% sodium bicarbonate at a pH of 6.8 to 8.4 in water for injection. Dilutions of aqueous extracts are made with buffered saline (0.5% sodium chloride, 0.04% potassium phosphate, 0.11% sodium phosphate heptahydrate, and 0.4% phenol in water for injection). Glycerinated extracts are concentrated aqueous extracts diluted to 50% v/v with USP Glycerin. Aqueous extracts contain 0.4% phenol as a preservative (0.2% phenol if glycerinated). Certain food extracts contain 0.1% sodium formaldehyde sulfoxylate as an antioxidant. Extracted source material substances are obtained as near as possible in the naturally occurring form to which a patient may be exposed.

Extracts labeled for diagnostic use only include the following foods: Barley, Coffee, Oat, Pineapple, Rye, Spinach, and Wheat. Other extracts for diagnostic only are Flea, House Fly, Mosquito, Moth, Cottonseed, and Flax Seed.

Extracts are labeled either by weight-to-volume (w/v) based on the weight of the source material to the volume of the extracting fluid, or in protein nitrogen units (PNU) based on assay with one PNU representing 10 micrograms of protein nitrogen. (See separate package inserts for standardized extracts such as cat hair, mite, or ragweed.)

CLINICAL PHARMACOLOGY

Allergen specific skin test reactions are mediated by specific immunoglobulin E (IgE) antibodies on the surface of mast cells. Interaction of allergens with cell-bound IgE induces cross linking of the IgE receptor which triggers the release of inflammatory mediators, particularly histamine. The characteristic mediator-induced induration (wheal) and erythema (flare) reaction observed upon diagnostic skin testing is an indication of specific allergy.¹ Allergen pharmacokinetics have not been characterized.

INDICATIONS AND USAGE

FOR DIAGNOSTIC USE ONLY - Allergenic Extracts are indicated for skin test assessment of the allergic state in patients with suspected IgE-mediated, immediate hypersensitivity allergy to the respective allergens when inhaled, ingested, or otherwise introduced into contact with sensitive tissues. The diagnosis of allergy may be established by the allergy history, clinical evaluation, and skin test reactivity. There are no reported limitations such as age group or progression of disease.

Extracts labeled FOR DIAGNOSTIC USE ONLY have not been shown by adequate data to be safe and effective for therapeutic use.

The use of Allergenic Extracts for the above purposes should be made only by physicians with special familiarity and knowledge of allergy. (See DOSAGE AND ADMINISTRATION)

CONTRAINDICATIONS

There are no known absolute contraindications to the use of Allergenic Extracts for skin test diagnosis.

Use of diagnostic extracts may be contraindicated in individuals with bleeding disorders or skin disease.

Skin test reactions can be suppressed when patients have received antihistamine medications within 24 hours, within 48 hours for terfenadine, or within 21 days or longer for astemizole. See DRUG INTERACTIONS.

WARNINGS

Concentrated extracts must be diluted with a sterile diluent (such as normal saline, buffered saline, saline with human serum albumin, or saline with 10% glycerin) prior to use in a patient for intradermal testing. Concentrates of Allergenic Extracts are manufactured to assure high potency and have the ability to cause serious local and systemic reactions including death in sensitive patients. Most reactions occur within 20 minutes after injection, but may occur later. To minimize the potential for local or systemic reactions, the relative sensitivity of the patient must be assessed from the allergic history and from clinical observations. Patients should be informed of the possibility of these reactions and the precautions should be discussed prior to testing (see PRECAUTIONS and ADVERSE REACTIONS below).

PRECAUTIONS

GENERAL

Not for intravenous or intramuscular use!

Systemic allergic reactions may occur as a result of the use of Allergenic Extracts. The physician must be prepared to treat anaphylaxis should it occur and have the necessary drugs, such as epinephrine, and equipment on hand to do so. Extracts should not be administered by individuals who are not prepared to treat anaphylaxis should it occur.²

A separate, sterile testing device, or needle and syringe, must be used to prevent the transmission of hepatitis or other infectious agents from person to person. Needles should not be recapped and should be disposed of properly.

INFORMATION FOR PATIENTS

The patient should be adequately informed of typical reactions to be expected as a result of diagnostic testing. Because most serious reactions following the administration of allergenic extracts occur within 20 minutes, the patient should remain under observation for this period of time or to 30 minutes or longer if instructed by the physician, such as for high-risk patients with unstable asthma, those with a history of reactions on injection, or patients suffering an exacerbation of their symptoms.² The patient should be instructed to report any unusual reactions. In particular, this includes unusual swelling and/or tenderness at the test site or reactions such as rhinorrhea, sneezing, coughing, wheezing, shortness of breath, nausea, dizziness or faintness. Reactions may occur some time after leaving the physician's office, in which case medical attention should be sought immediately.

DRUG INTERACTIONS

Skin test diagnosis with Allergenic Extracts may result in false negative responses if used within 24 hours after the last dose of most antihistamines, within 48 hours after the last dose of terfenadine, and within 3 weeks or longer after the last dose of astemizole. These products suppress histamine skin test reactions and could mask a positive response. The suppressive action of other drugs should be considered and emphasizes the need for a histamine positive-control test.¹

Patients receiving beta blocker drugs may not be responsive to beta adrenergic drugs used to treat anaphylaxis. The risks of anaphylaxis in these patients should be weighed against the benefits of diagnostic skin testing.

CARCINOGENESIS, MUTAGENESIS, IMPAIRMENT OF FERTILITY

There is no evidence of carcinogenicity, mutagenesis or impairment of fertility in humans from allergenic extracts. No long-term studies in animals have been performed to evaluate carcinogenic potential.

PREGNANCY

TERATOGENIC EFFECTS

PREGNANCY CATEGORY C - Animal reproduction studies have not been conducted with Allergenic Extracts for Diagnostic Use Only. It is also not known whether Allergenic Extracts can cause fetal harm when administered to a pregnant woman or can affect reproduction capacity. Allergenic Extracts should be given to a pregnant woman only if clearly needed.

LABOR AND DELIVERY

There is no known information of adverse effects during labor and delivery. Caution should be exercised in testing pregnant females because a systemic reaction may cause an abortion as a result of uterine muscle contractions.

NURSING MOTHERS

It is not known whether this product is excreted in human milk. Because many

drugs are excreted in human milk, caution should be exercised when extracts are administered to a nursing woman.

PEDIATRIC AND GERIATRIC USE

Although most Allergenic Extracts have not been studied systematically in children, children and geriatric patients appear to tolerate injections of Allergenic Extracts well. Extract usage in children should follow the same precautions as in adults.

ADVERSE REACTIONS

Adverse systemic reactions usually occur within minutes and consist primarily of allergic symptoms such as generalized skin erythema, urticaria, pruritus, angioedema, rhinitis, wheezing, laryngeal edema and hypotension. Less commonly, nausea, emesis, abdominal cramps, diarrhea and uterine contractions may occur. Severe reactions may cause shock and loss of consciousness. Fatalities have occurred rarely. The reaction rate is related to the type and dose of administered extract and to the degree of sensitivity of the patient; severe systemic reactions are not common with skin test diagnosis, but do occur. Despite all precautions occasional reactions are unavoidable.

Adverse systemic reactions should be treated as follows:

- A. If the testing site was an extremity a tourniquet should be immediately applied above the site. Release the tourniquet every few minutes for a few seconds.
- B. Epinephrine 1:1000 should be injected immediately in the opposite arm in amounts of 0.3 to 0.5 mL and 0.2 mL epinephrine should be administered at the site of injection. For children below the age of 6 years, adjust the dosage of epinephrine to 0.005 mL per pound (0.01 mL/kg) of body weight per dose. Repeat epinephrine dosage in 15 minutes if necessary and if symptoms persist.
- C. Adverse reactions not responding to epinephrine therapy may require other measures such as the use of parenteral bronchodilators, vasopressors, oxygen, or volume replacement therapy. Proper equipment and trained personnel should be available.²

Local reactions consisting of erythema, itching, swelling, tenderness and sometimes pain may occur as a result of testing. These reactions may appear within a few minutes to hours and persist for several days. Local cold applications and oral antihistamines may be effective treatment. For marked and prolonged local reactions, steroids may be helpful.

OVERDOSAGE

Systemic reactions are uncommon after testing, but if the patient receives more extract than can be tolerated at that particular time and begins to experience immediate hypersensitivity anaphylaxis, the procedures listed under ADVERSE REACTIONS should be instituted.

Overdosage may occur because of an error in the volume of extract used, or an incorrect dilution, or because the patient may be exposed to antigens simultaneously to testing with the same antigens.

DOSAGE AND ADMINISTRATION

TECHNIQUES

The most frequently used test sites are the back and the volar surface of the forearms. The skin should be cleansed with alcohol and allowed to dry. A minimum of at least 1 inch should be allowed between test sites, and preferably 2 inches between sites for pollens. A marking pencil may be used to indicate the site locations.

A sterile puncture device, needle, scalpel blade, or scarifier is used. A separate sterile device must be used for each patient to prevent transmission of infectious agents. If the device contacts extracts, use a separate device for each antigen to prevent cross-contamination.

The skin is abraded only enough to enter the dermis without drawing blood. Follow the directions for the device being used. The antigen may be applied directly with a puncture device or is introduced by applying a drop of extract to the scratch or prick site, taking care not to touch the skin with the dropper tip.

The number of tests that can be performed at one sitting depends upon the sensitivity of the patient and other clinical aspects. Normally, 30 to 40 tests administered at one time should be sufficient. For patients of extreme sensitivity, it is advisable to limit the number of tests to not more than 10, and use the forearm only so that a tourniquet can be applied in the event of a severe reaction.

For the patient with a suspected diagnosis of allergy to more than one antigen, initial skin testing should include the individual extracts. If a screening skin test with a mixture is used, a positive response should be followed by testing with the individual extracts to determine the degree of sensitivity to each. However, because a negative skin test with a mixture may not be indicative of the absence of allergy to one or more of the components due to their dilution, testing with individual extracts is more precise. False negative responses may occur if serum levels of antihistamines remain from prior medication administration (See DRUG INTERACTIONS). The use of a positive control is especially recommended for patients on prior medications which may decrease the histamine skin test response.

Initial testing may be conducted with the concentrate by means of a puncture test employing a multiple puncture device or other appropriate instrument. Prick testing through a drop of extract or scratch testing with a drop of extract applied to the scratch may also be employed to determine the degree of sensitivity. If a large whealing reaction occurs prior to the 20-minute period, wipe the area free of extract with an alcohol sponge. After final readings, all antigens should be removed by gently wiping with an alcohol sponge or sterile cotton swab. If the response is negative, this initial test may be followed by intradermal testing which should be correlated with the clinical history.

1. Scratch or Prick-puncture Skin Testing:

Scratch and Prick-puncture testing normally is performed with 1:20 w/v extracts in 50% glycerin or the strongest available strength in 5 mL vials for diagnostic use. Prick-puncture tests with concentrated extracts in

patients highly sensitive to the specific antigen should yield distinctive wheals with diameters of greater than 5 mm and with much larger erythma reactions. Glycerinated histamine phosphate 5 mg/mL (1.8 mg/mL histamine base) or aqueous histamine phosphate 2.75 mg/mL (1 mg/mL histamine base; 1:1,000 w/v) may be used as a positive control.

2. Intradermal Skin Testing:

Extracts for intradermal testing may be purchased or be prepared by diluting the stock concentrate injection vials with sterile diluent (use normal or buffered saline, normal saline with human serum albumin, or normal saline with glycerin).

To prepare dilutions starting from a concentrate such as 1:10 w/v, 1:20 w/v, or 20,000 PNU/mL, proceed as in the table below. (Note: Add 0.5 mL of concentrate to 4.5 mL of sterile diluent and make additional dilutions in the same manner.)

TABLE 1
Ten-Fold Dilution Series*

| Dilution | Extract | Diluent | w/v | w/v | PNU/mL |
|----------|-------------------|---------|--------------|--------------|--------|
| 0 | Concentrate | | 1:10 | 1:20 | 20,000 |
| 1 | 0.5mL concentrate | 4.5 mL | 1:10 | 1:200 | 2,000 |
| 2 | 0.5mL dilution 1 | 4.5 mL | 1:1,000 | 1:2,000 | 200 |
| 3 | 0.5mL dilution 2 | 4.5 mL | 1:10,000 | 1:20,000 | 20 |
| 4 | 0.5mL dilution 3 | 4.5 mL | 1:100,000 | 1:200,000 | 2 |
| 5 | 0.5mL dilution 4 | 4.5 mL | 1:1,000,000 | 1:2,000,000 | 0.2 |
| 6 | 0.5mL dilution 5 | 4.5 mL | 1:10,000,000 | 1:20,000,000 | 0.02 |

*There is no direct potency correlation across the table between PNU and w/v.

A. Patients with a negative scratch or prick-puncture test:

Patients with a negative scratch or prick-puncture test should be tested intradermally, using a 26 or 27 gauge 1/4 inch needle, with 0.02 to 0.05mL of an appropriate extract dilution of 1/100 to 1/1000 of the concentrate.

B. Patients tested only by the intradermal method:

Since highly reactive individuals may react intracutaneously at high dilutions, it is normally recommended that any intradermal injection should be preceded by and the dose adjusted according to puncture test reactivity. Patients suspected of being highly allergic should be tested with 0.02 to 0.05 mL of an appropriate extract dilution on the order of 1/10,000 to 1/100,000 or higher dilution of the concentrate.

C. Skin test titration:

A negative test should be followed by repeat skin test titration dosages using progressively stronger ten-fold concentrations until significant wheal and flare reaction sizes are attained, or until skin test responses with the higher concentrations remain negative.

D. Controls:

As a negative control use the diluent or, in case of extracts in 50% glycerin, use 0.5% to 1% glycerol saline solution. As a positive control, use glycerinated histamine phosphate diluted to 0.5 mg/mL (0.18 mg/mL histamine base) or aqueous histamine phosphate 0.275 mg/mL (0.1 mg/mL histamine base).

RESPONSES

Tests should be read 15 to 20 minutes after application of the test antigens.

One means for reporting test results is the “plus” system:

- no reaction or reaction not greater than control
- 1+ erythema smaller than 21 mm
- 2+ erythema larger than 21 mm with no wheal
- 3+ erythema and wheal
- 4+ erythema and wheal with pseudopods

Other systems for reporting positive results include measuring the largest erythema or wheal diameter, or the sum of the orthogonal diameters of the erythema or wheal. Parenteral drug products should be inspected visually for particulate matter and discoloration prior to administration, whenever solution and container permit. Some concentrated extracts naturally develop a cloudy appearance over time under refrigeration, the material settling to the bottom on standing.

HOW SUPPLIED

Scratch tests in 50% glycerin (v/v) are supplied at 1:20 w/v (or lesser strengths for selected extracts) in 5 mL vials only. Concentrates (which must be diluted for intradermal testing) are supplied in aqueous or 50% v/v glycerin solutions containing 10,000, 20,000, or 40,000 PNU/mL and 1:10, 1:20, 1:40, or 1:100 w/v, in 10, 30, and 50 mL vials. Dilutions for intradermal testing are available in 5 mL vials at 1,000 PNU/mL or 1:1000 w/v. (See separate package inserts for standardized extracts such as cat hair, mite, or ragweed.)

STORAGE AND EXPIRATION DATING: Allergenic Extracts should be stored at 2-8 degrees C and kept at this range during office use. Refer to the vial label for the expiration date of the extract. Diluted extracts are inherently less stable than the concentrates. Dilutions of glycerinated extracts which result in a glycerin content of less than 50% are also less stable. Highly-diluted aqueous extracts should be replenished daily. Potency of a particular dilution can be checked by skin test in comparison to a fresh dilution of the extract on an individual known to be allergic to the specific antigen.

REFERENCES

- ¹ Bousquet, J., Michel, F.B.: In vivo methods for study of allergy. *“Allergy Principles & Practice,”* 4th ed., Eds: Middleton, E., Reed, C.E., Ellis, E.F., et al., Mosby, St. Louis, 1993, 573 ff.
- ² Board of Directors: American Academy of Allergy and Immunology. Guidelines to minimize the risk from systemic reactions caused by immunotherapy with allergenic extracts. (Position statement). *J. Allergy and Clin. Immunol.* 1994; 93:811-12.

Greer*Pick*[™]

Optimized Skin
Testing System



GREER[™]

PO Box 800
Lenoir, NC 28645
USA

ALLERGY SKIN TEST SYSTEM

The GreerPick™ system is an easy to use, economical system for skin testing of suspected allergic individuals to determine specific allergen sensitivity. The system is designed for use by allergy practitioners who are trained in the application and interpretation of allergy skin tests, and who are trained in the recognition and treatment of adverse allergic reactions should they occur. Before employing the system, users should fully familiarize themselves with the instructions contained herein, and any pertinent data and warnings regarding test antigens.

The system is comprised of the following components as illustrated in *Figure 1* below:



Figure 1

1. The 60-well Greer Tray™, a larger tray that has a maximum of 60 well positions for test antigens.
2. The 40-well Greer Tray™, a medium size tray that has a maximum of 40 well positions for test antigens.
3. The Greer Wells™, supplied in sterile packs and designed to contain the antigens for skin testing.
4. The Greer Picks™, supplied in sterile packs and designed for the application of test antigens to the skin, and to serve as a cap to the Greer Wells™ when not in use.

PREPARATION OF THE SYSTEM FOR USE

Each Greer *Tray*[™] is supplied with labels ruled to correspond to the well positions in the tray. Labels should be used to identify the antigens contained in the wells.

Greer *Wells*[™] are supplied in packs of 24. Using aseptic procedures, open the pack carefully and place the wells in each of the positions to be filled with skin test antigens in the Greer *Tray*[™]. Press the wells firmly into place.

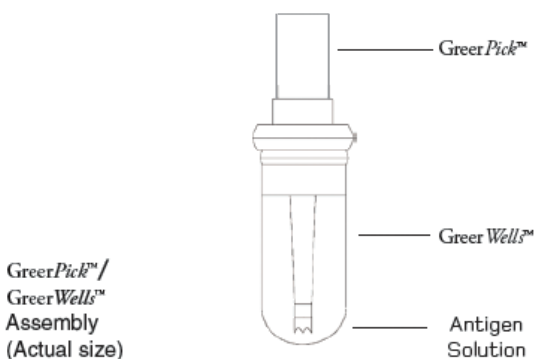


Figure 2 (diagram)

Fill each Greer *Well*[™] with approximately 0.125 mL of the appropriate extract as shown in Figure 2. Pay careful attention to assure that the antigens in the wells correspond with the labels on the tray. It is strongly recommended that glycerinated extracts containing a phenol preservative be used to inhibit microbial growth.

Greer *Picks*[™] are supplied sterile in trays of 42. Greer *Picks*[™] should be inserted into the Greer *Wells*[™] immediately after filling to prevent the entry of foreign matter. Handle the Greer *Picks*[™] only by the larger handle end and take care not to contaminate or damage the small end which contacts the patient's skin. Once the Greer *Tray*[™] is labeled, loaded with antigens, and the Greer *Picks*[™] are inserted, the system is ready for use. Store the system in the refrigerator at all times when not in use to maintain antigenic potency.

When the wells are emptied they should be discarded and replaced with new sterile wells loaded with the appropriate fresh antigens. This practice will help ensure antigen potency. Empty wells can be identified by viewing the Greer *Tray*[™] from beneath by holding the tray overhead. To remove empty Greer *Wells*[™] from the Greer *Tray*[™] simply lift upward and discard. Replace and refill as above.

SKIN TESTING WITH THE GREER *Pick*[™] SYSTEM

Allergy testing with the Greer *Pick*[™], as with any skin test device, requires that the proper technique be developed and used. Proper contact between the device and the skin is required to ensure that the antigen bearing tines of the device penetrate, scarify or prick the epidermis to deliver the antigen to the underlying reactive dermis layer of the skin. Proper test technique, however, will not cause bleeding. The prick technique is the recommended method for use of the Greer *Pick*[™].

Using The GreerPick™ as a Prick Test Device

Remove the GreerPick™ from the GreerWell™ and briefly inspect to ensure that a small droplet of antigen is present at the tip of the device. Holding the GreerPick™ at an approximate 45 degree angle to the skin at the test site, bring the tip of the GreerPick™ in contact with the skin in the direction the GreerPick™ is pointing. Press the tines making contact into the superficial skin, withdrawing with a slight lifting of the skin. A properly applied prick test will result in a small pricking of the epidermis, but will not result in bleeding or scarring. Discard the used device into a suitable biohazard container.

While the Prick method is the preferred method of use, Greer recognizes that health care providers may want to use other accepted techniques, which are described below:

Using The GreerPick™ as a Puncture Test Device

Remove the GreerPick™ from the GreerWell™ and briefly inspect to ensure that a small droplet of antigen is present at the tip of the device. Holding the GreerPick™ perpendicular to the skin surface at the test site, firmly press the tines of the device into the skin. A properly performed test will leave six small indentations corresponding to the tines of the device, but will not result in bleeding or scarring. Discard the used device into a suitable biohazard container.

Using The GreerPick™ as a Scratch Test Device

Remove the GreerPick™ from the GreerWell™ and briefly inspect to ensure that a small droplet of antigen is present at the tip of the device. Holding the GreerPick™ perpendicular to the skin at the test site, lightly touch the tip of the device to the skin ensuring that all six tines are in contact and simultaneously rotate the device approximately one-quarter turn. A properly applied scratch test will result in some abrasion of the epidermis but will not result in bleeding or scarring. Discard the used device into a suitable biohazard container.

It is recommended that technicians unfamiliar with the use of the device practice the different techniques using glycerinated histamine positive control and glycerinated negative control solution. A properly applied positive control should result in a wheal reaction of approximately 5 mm or greater, and an erythema (redness/flare) reaction of approximately 20 mm to 25 mm diameter in normally reactive individuals. As with any test, wide variations in patient reactivity are not uncommon.

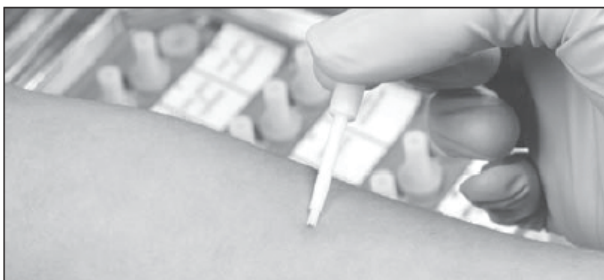


Figure 3

Skin testing is normally performed on the volar surface of the forearm and on the back. Sanitize the test sites with alcohol swabs, and mark the sites for reference. (See *Figure 3*) Test sites should be separated by approximately 3 cm (0.8 inches)* to prevent strong positive reactions from overlapping and causing difficulties in interpreting results.

* (*JACI Position Statement, 1993; 92:636-7*)

After the test sites have been sanitized and marked, you may begin testing using the *testing method of your choice*: prick, puncture or scratch. Upon the completion of each test, a small amount of antigen solution will remain on the skin at the test site. Continue testing with the desired number of antigens, discarding the used GreerPick™ into an appropriate biohazard container after each test. Place new GreerPicks™ in the GreerWells™ as soon as possible to avoid contamination of the antigen in the wells. It is recommended that each patient be tested with both a positive (50% Glycerinated Histamine), and negative (50% Glycerinated saline) control in addition to the test antigens.

GreerPick™ devices are single use only devices. To prevent the occurrence of antigen cross-contamination or the transfer of infectious agents, such as serum Hepatitis, DO NOT reuse the devices.

INTERPRETATION OF TEST RESULTS

Allergic individuals exhibit extreme variability in their reactions to test antigens, and it is important to judge the results of allergy skin tests in relation to both the positive and negative control tests. Careful consideration of patient history, current antigen exposure, and any recent use of interfering drugs such as antihistamines or beta-blockers should be considered. Readings of the test sites should be taken approximately 15 to 20 minutes after administration of the test antigens. Assessment of both the wheal (swelling and edema) and erythema (flare/redness) reactions should be conducted and recorded.

Reactions appearing less than or equal to the size of the negative control test are to be considered negative reactions. Reactions of wheal and/or erythema size exceeding those of the negative control test should be considered positive reactions. Various methods are in use for the grading of skin test reactions to allergens; a common scheme is presented below. It is important to note that individual patient reactivity can vary greatly with time, antigen potency, drug therapy and/or immunotherapy, as well as testing technique.

- 1+ Wheal, if present, equal to or larger than negative control reaction.
Erythema larger than negative control, but less than 10 mm diameter.
- 2+ Wheal up to 7 mm diameter, and erythema up to 20 mm diameter.
- 3+ Wheal diameter up to 10 mm, and erythema larger than 20 mm diameter; pseudopodia may or may not be present.
- 4+ Wheal diameter larger than 10 mm, with pseudopodia, erythema as for 3 +.

Alternatively, test results may be recorded by noting the largest diameter of both wheal and erythema as well as the diameter of the reaction perpendicular to the largest reaction diameter. Thus, an approximate area for both wheal and erythema can be calculated. While some correlation exists between the size of patient reaction and the degree of sensitivity, other factors as mentioned above must be considered in the diagnosis of allergy to specific antigens.

HOW SUPPLIED

| | | |
|--------------|---|--|
| Item # GP-1C | - | 50 packs of 42 (2,100) Greer <i>Picks</i> [™] (Sterile) |
| Item # GW-2 | - | 1 pack 24 Greer <i>Wells</i> [™] (Sterile) |
| Item # GY-5 | - | 1 ea. 60-Well Greer <i>Tray</i> [™] with labels |
| Item # GY-40 | - | 1 ea. 40-Well Greer <i>Tray</i> [™] with labels |

WE WELCOME YOUR QUESTIONS

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